CASE REPORT

Pseudomonas putida bacteremia in adult patients: five case reports and a review of the literature

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Abstract Pseudomonas putida belongs to the fluorescent group of Pseudomonas species, a group of opportunistic pathogens that primarily cause nosocomial infections. However, few cases of P. putida bacteremia in adult patients have been reported. We report five cases of P. putida bacteremia in adult patients and review 23 previously reported cases. Our five patients consisted of three cases of catheter-related bloodstream infection (CRBSI), one case of indwelling biliary drainage tube-related cholangitis, and one case of cholecystitis. Many of the 23 previously reported cases also included CRBSI. Of the clinical backgrounds, in all 28 reported cases including ours, 24 (85.7%) were immunocompromised. Of the clinical management, in CRBSI, devices were removed in almost all cases (92.9%). Antibiotic susceptibility data of our five cases and another previous case showed that patients with bacteremia had a high susceptibility of P. putida to anti-pseudomonal β -lactams. The prognosis for bacteremia with P. putida was good, as 26 (92.9%) of the total 28 cases were cured.

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Introduction

Pseudomonas spp. are aerobic Gram-negative bacteria. Due to their ability to metabolize a wide range of compounds, members of this species are able to colonize soil, fresh water and moist environments [1, 2]. These bacteria can act as opportunistic pathogens that primarily cause nosocomial infections. Pseudomonas aeruginosa is the most prevalent pathogen among the genus Pseudomonas, yet non-aeruginosa Pseudomonas have also been associated with clinical infections [3–5]. Pseudomonas putida, which belongs to the fluorescent group of Pseudomonas spp., has been recognized as a rare pathogen of bacteremia. Most reported cases of bacteremia with P. putida have been neonatal infections or outbreak infections due to transfusion of contaminated blood or fluid [6-8]. P. putida can acquire broad resistance to β -lactam antibiotics, and some isolates of this organism are capable of producing metallo- β -lactamases [9–12]. Despite this, there have been few reports about antimicrobial susceptibilities in bacteremia. Clinical courses of bacteremia with P. putida have not been precisely described due to the rarity of reported infections from P. putida, which comprises mostly immunocompromised patients and newborns [6–8, 13]. We report five adult cases of P. putida bacteremia occurring in our hospital between April 2003 and March 2007. Upon a brief review of the literature, we were able to identify clinical characteristics of P. putida bacteremia and antimicrobial susceptibility of P. putida in these bacteremic patients.

Cases

Case 1

A 79-year-old man was admitted for gastrectomy due to gastric cancer. After the surgery, cefmetazole was administered for 6 days because of postoperative prophylaxis. At 7 days post-operation, he became febrile. Peripheral catheter-associated bacteremia was suspected because he developed skin flare and pain at the insertion region with no other findings at the site of infection. The catheter was removed and cefmetazole was replaced with meropenem. *P. putida* was identified on blood culture. The patient fully recovered, and antibiotic treatment was continued for 10 days.

Case 2

A 76-year-old man was admitted for colectomy due to sigmoid colon adenocarcinoma. He had liver cirrhosis with hepatitis C virus and hepatocellular carcinoma. After the colectomy, cephazolin was administered for 5 days as a means of postoperative prophylaxis. At 6 days post-operation, he had a fever and diarrhea. As catheter-associated bacteremia and *Clostridium difficile*-associated diarrhea were suspected, the peripheral vein catheter was removed. Cephazolin was replaced with intravenous ceftazidime and oral vancomycin. *P. putida* was identified on cultures from blood and the catheter. The stool examination tested positive for *C. difficile* toxin. The patient fully recovered and antibiotic treatment was continued for 10 days.

Case 3

A 52-year-old woman with Burkitt lymphoma was admitted for induction chemotherapy. On day 15 post-chemotherapy, her neutrophil count reduced to $240/\mu l$ and she developed a fever of $39.5^{\circ}C$. For febrile neutropenia treatment, meropenem administration was initiated, and a central vein catheter was removed. *P. putida* was isolated on cultures from blood and the central vein catheter. She recovered from fever 3 days after antibiotic therapy, although meropenem was switched to ciprofloxacin on day 20 post-chemotherapy because the strain was resistant to almost all β -lactams. Antibiotic therapy with ciprofloxacin was continued for 10 days.

Case 4

A 48-year-old man with liver cirrhosis due to hepatitis B virus and hepatocellular carcinoma underwent liver transplantation. He was discharged with an indwelling biliary drainage tube and continuing to take 11 mg of tacrolimus

and 12 mg of methyl prednisolone. Two months after discharge, he visited the emergency room with a fever and shivering. On the blood test, his C reactive protein level was 5.63 mg/dl, and white blood cell count was 18,100/µl. Hepatobiliary enzyme levels were not elevated. Indwelling drainage tube was clogged. Abdominal ultrasonography showed slight dilation of the biliary duct, which was suggestive of device related acute cholangitis. The biliary drainage tube was removed and endoscopic retrograde biliary drainage was performed. Intravenous administration of ceftazidime and vancomycin was initiated. *P. putida* was identified on blood culture, although no organism was cultured from bile. He fully recovered and antibiotic therapy was continued for 10 days.

Case 5

A 27-year-old man was admitted to hospital with fever that had persisted for 4 weeks and cough that had persisted for 5 days. He had common variable immunodeficiency after developing autoimmune hemolytic anemia at the age of 24 years and has been taking 15 mg of prednisolone daily. On chest CT, ground glass-like infiltrates were identified in the bilateral lung, and because the blood $(1 \rightarrow 3)$ - β -Dglucan level was 293.1 pg/ml, Pneumocystis jirovecii pneumonia (PCP) was suspected. After 2 weeks of treatment with trimethoprim-sulfamethoxazole, the patient recovered. On day 4 after the termination of PCP treatment, however, he became febrile again. On the blood test, the aspartate aminotransferase level was 131 IU/I, the alanine aminotransferase level was 310 IU/l, and total bilirubin was 3.3 mg/dl. Panipenem-betamipron was initiated. Abdominal CT showed thickening of the gallbladder wall and fluid around the gall bladder, which was indicative of acute cholecystitis. Percutaneous transhepatic gallbladder drainage was performed. P. putida and Klebsiella oxytoca were identified on blood cultures. No organism was cultured from bile. He fully recovered and treatment was continued for 18 days.

Discussion

P. putida has been recognized as a rare pathogen of bacteremia in adult patients. The five reported cases of *P. putida* bacteremia in our hospital accounted for 0.22% of the 2307 cases of bacteremia that occurred from April 2003 to March 2007. In the literature, only 23 cases have been reported, excluding paediatric and outbreak cases due to transfusion of contaminated blood or fluid (Table 1) [14–19].

Among the five cases reported herein, four had a medical device as the primary infection site. In 21 cases of



Table 1 Review of the literature, including our five cases

| No. | Citation | Age/gender | Co-morbidities | Primary infection site | Indwelling device | Antibiotics | Outcome |
|------|-----------|------------------------------------------|---------------------------------------|------------------------|----------------------------------|-----------------------------------------------------|---------|
| 1 | [15] | 65/female | Surgery for maxillary sinus carcinoma | Acute cholecystitis | VC | Kanamycin, cephalothin | Cured |
| 2 | | 30/female | Surgery for ectopic pregnancy | Unknown | VC | Nothing | Cured |
| 3 | [18] | 19/female | ALL | CRBSI | CVC, removal | Moxalactam | Cured |
| 4 | | 45/female | CML | Unknown | CVC, removal | Ceftriaxone, amikacin | Cured |
| 5 | | 50/male | Lung cancer | Pneumonia | CVC | Cefoperazone, mezlocillin | Cured |
| 6 | | 72/male | Smoldering leukemia | Unknown | CVC | Piperacillin, ceftazidime | Cured |
| 7 | | 36/male | AML | CRBSI | CVC | Moxalactam, ticarcillin, tobramycin | Cured |
| 8 | | 25/male | AML | CRBSI | CVC, removal | Nothing | Cured |
| 9–14 | [14] | 18–61 ^a /male (3), female (3) | Lymphoma (4), AL (1), myeloma (1) | CRBSI (6) ^b | VC (6), removal (6) ^c | Nothing (5), antibiotic drug (1) ^d | Cured |
| 15 | [19] | 18/female | APL | Thrombophlebitis | Unknown | Imipenem, amikacin | Cured |
| 16 | | 62/male | RHD, CHF | Unknown | Unknown | Ceftazidime | Died |
| 17 | | 32/male | Infective endocarditis | Unknown | Unknown | Imipenem, amikacin | Cured |
| 18 | | 70/female | Cerebral infarction | Acute tonsillitis | Unknown | Piperacillin | Cured |
| 19 | | 70/female | Carcinoma of cervix | Pneumonia | Unknown | Piperacillin, gentamicin | Cured |
| 20 | | 23/female | CHF | CRBSI | CVC, removal | Cefotaxime, oxacillin | Cured |
| 21 | | 23/female | Trauma | Unknown | Unknown | Cefoperazone, netilmicin | Cured |
| 22 | | 79/male | Gastric cancer | Unknown | Unknown | Cefazolin, gentamicin | Died |
| 23 | [16] | 78/female | Nothing | Soft tissue infection | Nothing | Ceftazidime | Cured |
| 24 | Our cases | 79/male | Post-gastrectomy (gastric carcinoma) | CRBSI | PVC, removal | Meropenem | Cured |
| 25 | | 76/male | LC, post-colectomy (colon carcinoma) | CRBSI | PVC, removal | Ceftazidime | Cured |
| 26 | | 52/female | Burkitt lymphoma | CRBSI | CVC, removal | Meropenem, ciprofloxacin | Cured |
| 27 | | 48/male | LC, post-liver transplantation (HCC) | Acute cholangitis | Biliary drainage tube, removal | Ceftazidime | Cured |
| 28 | | 27/male | CVID, AIHA | Acute cholecystitis | Nothing | Panipenem, betamipron | Cured |

VC central venous catheter or peripheral venous catheter, PVC peripheral venous catheter, CVC central venous catheter, CRBSI catheter related blood stream infection, ALL acute lymphoblastic leukemia, CML chronic myelogenous leukemia, AML acute myelogenous leukemia, AL acute leukemia, RHD rheumatoid heart disease, CHF congestive heart failure, LC liver cirrhosis, HCC hepatocellular carcinoma, CVID common variable immunodeficiency, AIHA autoimmune hemolytic anemia



^a Age range

^b Definite (4) or probable (2)

^c As primary (5) or secondary (1) treatment

d Antibacterial drug was administrated in one case as a primary treatment, but the name of the drug was not mentioned

Table 2 Antibiotic susceptibilities of *P. putida* in our five cases

| Antibiotics | Susceptible | Intermediate | Resistant |
|---------------------|-------------|--------------|-----------|
| Piperacillin | 4 (80) | 1 (20) | 0 (0) |
| Ceftazidime | 4 (80) | 0 (0) | 1 (20) |
| Cefotaxime | 1 (20) | 2 (40) | 1 (20) |
| Aztreonam | 1 (20) | 2 (40) | 2 (40) |
| Meropenem | 4 (80) | 0 (0) | 1 (20) |
| Imipenem/cilastatin | 3 (60) | 0 (0) | 2 (40) |
| Gentamicin | 4 (80) | 1 (20) | 0 (0) |
| Amikacin | 5 (100) | 0 (0) | 0 (0) |
| Ciprofloxacin | 5 (100) | 0 (0) | 0 (0) |
| | | | |

Numbers shown in parentheses represent the percentage of cases that are susceptible or resistant to the different antibiotics

Antibacterial susceptibility assays to determine the minimum inhibitory concentrations (MICs) of antibacterial agents were performed by broth microdilution according to guidelines recommended by the Clinical Laboratory Standards Institute (CLSI)

identified primary infection sites including our cases, 13 (61.9%) were device related, and among these cases, 12 were CRBSI (92.9%). Of the device-associated infections, the ability of microorganisms to adhere to materials and to promote the formation of a biofilm appears to be the most important feature of their pathogenicity [20]. *P. putida* are able to form biofilms [21]. For treatment of *P. putida* bacteremia, the medical device was removed in 12 (92.3%) of the 13 cases. Thus, when *P. putida* is isolated on blood culture of a patient with a medical device, device-related bacteremia should be mostly suspected and the device should be removed.

In addition to device-related infection, acute cholecystitis was evident in 2t cases (9.5%) and pneumonia in 2 cases (9.5%). *P. putida* has been reported to cause urinary tract infection as well as pneumonia in several cases [19]. We thought that like *P. aeruginosa*, *P. putida* could also colonize the respiratory tract and urinary tract in immunocompromised patients. Since *P. putida* can be detected in the normal oropharyngeal flora [22, 23], it may also cause cholecystitis by colonizing the intestinal tract.

Considering the clinical background, 24 (85.7%) of the 28 cases were in an immunocompromised state (from the use of immunosuppressive drugs, liver cirrhosis, malignancy or other immunosuppressive diseases, or post-operation). Although this factor might be biased as the previous cases comprised those from a cancer center only, limiting the clinical background to cancer patients. Our five cases were all in an immunocompromised state. *P. putida*, therefore, could cause bacteremia, particularly in an immunocompromised patient, though the overall incidence of bacteremia with *P. putida* still remains low.

The rate of susceptibility of *P. putida* to antibiotics in our five cases is shown in Table 2. In case 3, *P. putida*

was found to be resistant to both ceftazidime, an antipseudomonal cephalosporin, and to carbapenem, while in the other four cases, P. putida was susceptible to those antibiotics. In case 4, P. putida was resistant to imipenem/ cilastatin but not to meropenem. We speculate that a long duration of imipenem/cilastatin use in this case would be related to resistance to imipenem/cilastatin, as in the case of P. aeruginosa which develops resistant strains that survive during antibiotic use [24]. In previous reports, P. putida isolated from the urinary tract, tracheal aspiration, and areas other than blood of bacteremia patients, has been found to acquire metallo- β -lactamases and were resistant to most β -lactams, including carbapenems [10–12]. On the other hand, Anaissie reported that of most of the clinical isolates, P. putida in bacteremic patients was susceptible to ceftazidime, imipenem, and ciprofloxacin [18]. We considered that in order to investigate the cause of high-rate susceptible strains in the blood, comparing susceptibilities of P. putida in patients with pneumonia or urinary tract infections to that in patients with bacteremia is imperative.

Prognosis of *P. putida* bacteremia was good, whereby 26 (92.9%) out of 28 cases were cured. Yang reported that in one of two cases where the patient died, inappropriate antibiotic therapy to other co-pathogens was a possible contributing factor [19]. Bacteremia with another *Pseudomonas sp.*, *P. aeruginosa* is known to result in poor prognosis of the disease. Mortality of *P. aeruginosa* bacteremia has been reported at over 30% [25, 26]. We believe the reason for a better prognosis from *P. putida* bacteremia was that the major cause of the bacteremia was device related, and infected devices were removed in most cases. However, we cannot exclude the possibility of bacteriological factors causing bacteremia, such as pyocyanin, exotoxin A, or the type III secretion system that can arise from *P. aeruginosa* infection but not from *P. putida* [27].

In conclusion, we report five adult cases of *P. putida* bacteremia. These cases displayed clinical characteristics: device relatedness as a major cause of bacteremia, an immunocompromised host, a high susceptibility of strains to β -lactams, and a good prognosis overall.

References

- Favero MS, Carson LA, Bond WW, Petersen NJ. *Pseudomonas aeruginosa*: growth in distilled water from hospitals. Science. 1971;173(999):836–8.
- Penna VT, Martins SA, Mazzola PG. Identification of bacteria in drinking and purified water during the monitoring of a typical water purification system. BMC Public Health. 2002;2:13.
- 3. Roig P, Orti A, Navarro V. Meningitis due to *Pseudomonas stutzeri* in a patient infected with human immunodeficiency virus. Clin Infect Dis. 1996;22(3):587–8.



- Hsueh PR, Teng LJ, Pan HJ, Chen YC, Sun CC, Ho SW, et al. Outbreak of *Pseudomonas fluorescens* bacteremia among oncology patients. J Clin Microbiol. 1998;36(10):2914–7.
- Rastogi S, Sperber SJ. Facial cellulitis and *Pseudomonas luteola* bacteremia in an otherwise healthy patient. Diagn Microbiol Infect Dis. 1998;32(4):303–5.
- Bouallegue O, Mzoughi R, Weill FX, Mahdhaoui N, Ben Salem Y, Sboui H, et al. Outbreak of *Pseudomonas putida* bacteraemia in a neonatal intensive care unit. J Hosp Infect. 2004;57(1):88–91.
- Ladhani S, Bhutta ZA. Neonatal *Pseudomonas putida* infection presenting as staphylococcal scalded skin syndrome. Eur J Clin Microbiol Infect Dis. 1998;17(9):642–4.
- Taylor M, Keane CT, Falkiner FR. Pseudomonas putida in transfused blood. Lancet. 1984;2(8394):107.
- Docquier JD, Riccio ML, Mugnaioli C, Luzzaro F, Endimiani A, Toniolo A, et al. IMP-12, a new plasmid-encoded metallo-betalactamase from a *Pseudomonas putida* clinical isolate. Antimicrob Agents Chemother. 2003;47(5):1522–8.
- Almuzara M, Radice M, de Garate N, Kossman A, Cuirolo A, Santella G, et al. VIM-2-producing *Pseudomonas putida*, Buenos Aires. Emerg Infect Dis. 2007;13(4):668–9.
- Bogaerts P, Huang TD, Rodriguez-Villalobos H, Bauraing C, Deplano A, Struelens MJ, et al. Nosocomial infections caused by multidrug-resistant *Pseudomonas putida* isolates producing VIM-2 and VIM-4 metallo-beta-lactamases. J Antimicrob Chemother. 2008;61(3):749–51.
- Lee K, Park AJ, Kim MY, Lee HJ, Cho JH, Kang JO, et al. Metallo-beta-lactamase-producing *Pseudomonas* spp. in Korea: high prevalence of isolates with VIM-2 type and emergence of isolates with IMP-1 type. Yonsei Med J. 2009;50(3):335–9.
- Centers for Disease Control (CDC). Reported contamination of heparin sodium with *Pseudomonas putida*. MMWR Morb Mortal Wkly Rep 1986;35(8):123–4.
- 14. Martino R, Martinez C, Pericas R, Salazar R, Sola C, Brunet S, et al. Bacteremia due to glucose non-fermenting gram-negative bacilli in patients with hematological neoplasias and solid tumors. Eur J Clin Microbiol Infect Dis. 1996;15(7):610–5.
- Von Graevenitz A, Weinstein J. Pathogenic significance of Pseudomonas fluorescens and Pseudomonas putida. Yale J Biol Med. 1971;44(3):265–73.
- Chen CH, Hsiu RH, Liu CE, Young TG. Pseudomonas putida bacteremia due to soft tissue infection contracted in a flooded

- area of central Taiwan: a case report. J Microbiol Immunol Infect. 2005;38(4):293–5.
- Spelman DW, Russo P, Harrington G, Davis BB, Rabinov M, Smith JA, et al. Risk factors for surgical wound infection and bacteraemia following coronary artery bypass surgery. Aust N Z J Surg. 2000;70(1):47–51.
- Anaissie E, Fainstein V, Miller P, Kassamali H, Pitlik S, Bodey GP, et al. *Pseudomonas putida*. Newly recognized pathogen in patients with cancer. Am J Med. 1987;82(6):1191–4.
- Yang CH, Young T, Peng MY, Weng MC. Clinical spectrum of Pseudomonas putida infection. J Formos Med Assoc. 1996; 95(10):754–61.
- von Eiff C, Jansen B, Kohnen W, Becker K. Infections associated with medical devices: pathogenesis, management and prophylaxis. Drugs. 2005;65(2):179–214.
- Tolker-Nielsen T, Brinch UC, Ragas PC, Andersen JB, Jacobsen CS, Molin S. Development and dynamics of *Pseudomonas* sp biofilms. J Bacteriol. 2000;182(22):6482–9.
- Koh AY, Priebe GP, Pier GB. Virulence of *Pseudomonas aeru-ginosa* in a murine model of gastrointestinal colonization and dissemination in neutropenia. Infect Immun. 2005;73(4): 2262–72.
- Carlson D, McKeen E, Mitchell M, Torres B, Parad R, Comeau AM, et al. Oropharyngeal flora in healthy infants: observations and implications for cystic fibrosis care. Pediatr Pulmonol. 2009;44(5):497–502.
- Mouton JW, den Hollander JG, Horrevorts AM. Emergence of antibiotic resistance amongst *Pseudomonas aeruginosa* isolates from patients with cystic fibrosis. J Antimicrob Chemother. 1993;31(6):919–26.
- 25. Kang CI, Kim SH, Kim HB, Park SW, Choe YJ, Oh MD, et al. Pseudomonas aeruginosa bacteremia: risk factors for mortality and influence of delayed receipt of effective antimicrobial therapy on clinical outcome. Clin Infect Dis. 2003;37(6):745–51.
- Osmon S, Ward S, Fraser VJ, Kollef MH. Hospital mortality for patients with bacteremia due to *Staphylococcus aureus* or *Pseu-domonas aeruginosa*. Chest. 2004;125(2):607–16.
- Nelson KE, Weinel C, Paulsen IT, Dodson RJ, Hilbert H, Martins dos Santos VA, et al. Complete genome sequence and comparative analysis of the metabolically versatile *Pseudomonas putida* KT2440. Environ Microbiol. 2002;4(12):799–808.

