NEURON Assignment

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1 Passive properties

1.1 Make a ball model and measure R_i , m_{τ} with current injection

We first define a function which injects current into a cell with passive properties, whilst recording voltage. The function returns the recorded voltage and time vectors.

```
| length of the soma (um)
somaLength
somaDiam
             / diameter of the soma
stimDur
              / duration of the stimulus (ms)
              / amplitude of current injection (pA)
stimAmp
stimDelay
               / delay before the stimulus is given (ms)
total\_time
              / total time of the simulation
printProperties | if true, print the properties of the cell
OUTPUT
voltage
time
cell.insert('pas')
cell.L = somaLength
cell.diam = somaDiam
cell.g_pas = g_pas
if printProperties is True:
   print('Cell properties')
   h.psection()
# add point process: current injection
stim = h.IClamp(cell(0.5))
stim.delay = stimDelay
stim.dur = stimDur # ms
stim.amp = stimAmp # nA
# measure voltage
voltage = h.Vector()
voltage.record(cell(0.5)._ref_v)
# measure time
time = h.Vector()
time.record(h._ref_t)
h.v_init = -70 # initial voltage
h.load_file('stdrun.hoc')
h.tstop = total_time
h.run() # ms
# time_np = time.as_numpy() # WARNING: don't output values with .as_numpy()
# voltage_np = voltage.as_numpy()
return voltage, time
```

We define a function (using Ohms law) which allows us to calculate the input resistance (R_i) from our cell recording using the returned voltage vector.

$$R_i = \frac{\Delta V}{\Delta I_c}$$

```
def cal_Ri(voltage_np, stimAmp = 0.05):
    delta_V = (max(voltage_np) - min(voltage_np)) * 10**(-3) # millivolts (mV)
    delta_Ie = stimAmp * 10**(-9) # nanoAmps
```

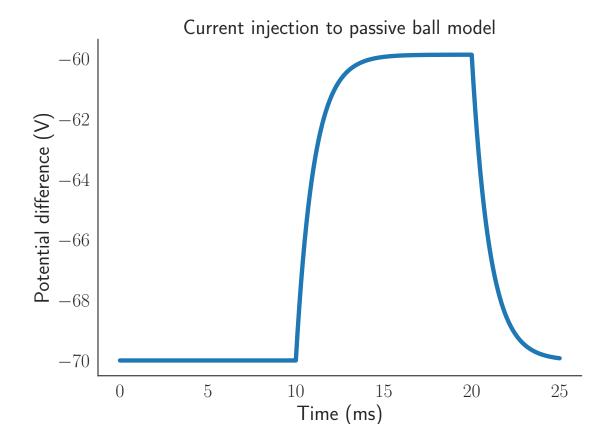
```
input_resistance = delta_V / delta_Ie
return input_resistance
```

We also define a function to calculate the membrane time constant τ_m from our recording. We are doing this by finding the time when the voltage has reached 63.2% of its peak voltage.

```
def find_nearest(array, value):
    array = np.asarray(array)
    idx = (np.abs(array - value)).argmin()
   return array[idx]
def cal TauM(voltage, time, stimDelay = 10):
    # convert voltage to numpy
   voltage_np = voltage.as_numpy()
   time_np = time.as_numpy()
    # from "experiment"
    special_v = (max(voltage_np) - min(voltage_np)) * (1 - 1/np.e) # about 2/3
    assert(special_v > 0)
    special_vv = special_v + min(voltage_np)
    # create trimmed voltage_np to ensure find_nearest finds only
    # within the 'charging' curve
   firstMaxIndex = np.where(voltage_np == max(voltage_np))
   firstMaxIndex = firstMaxIndex[0][0]
   voltage_np_trim = voltage_np[0:firstMaxIndex]
   tauIndex = np.where(voltage_np_trim == find_nearest(voltage_np_trim, special_vv))
    if tauIndex[0].size > 1: # more than one matching value
        tau_m_samp = tauIndex[0][0]
    else:
       tau_m_samp = tauIndex[0]
   tau_m_exp = time_np[tau_m_samp]
   TauM = tau_m_exp - stimDelay
   return TauM[0]
```

We run our current injection function on a cell which only contains a soma. According to the default settings of our function the soma diameter and length are the same, creating a ball. Below is a plot of the voltage recorded during current injection into our ball model.

```
voltage, time = inject_current(cell = h.Section(name='soma'))
# plt.figure(figsize=(8, 4))
# ax = sns.lineplot(time, voltage)
plt.figure()
plt.plot(time, voltage)
plt.title('Current injection to passive ball model')
plt.xlabel('Time (ms)')
plt.ylabel('Potential difference (V)')
plt.show()
# ax.set_title('Current injection to passive ball model')
# ax.set(xlabel='Time (ms)', ylabel='Potential difference (V)')
```



```
input_resistance = cal_Ri(voltage)
# print('Input resistance is %.2f ohms' % input_resistance)
print('Input resistance is %.2f Mohms' % (input_resistance / 10**6))

## Input resistance is 202.41 Mohms

TauM = cal_TauM(voltage, time)
print('TauM is %.2f ms' % TauM)
```

TauM is 1.00 ms

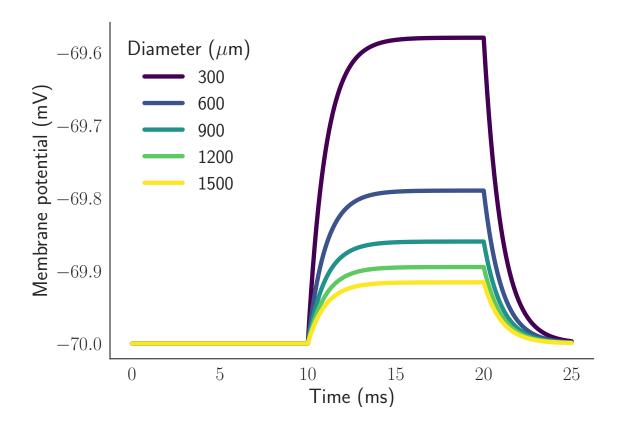
Due to the cells small size the input resistance is greater than we would normally expect. During our in vitro experiments this week we measured our input resistance to be around 5 Mohms. Equally, the time constant differs from our experimental observations (in our experiment the time constant \sim 20 ms). This is because the time constant is a product of the resistance and capacitance of the cell, and the membrane capacitance is proportional to the amount of membrane.

1.2 Varying neuron variables

Change diameter, R_m , R_a , C_m and see how it affects R_i and m_{τ}

We generate lists containing different values for the above mentioned variables. We first show how changes in diameter length effect the recorded voltage and then the effects of diameter on R_i and m_{τ} .

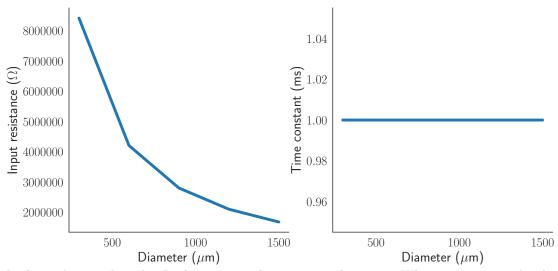
```
g_list = [0.001, 0.01, 0.1, 1, 10]
Ra_list = [10, 20, 30, 40, 50] # default is 35.4
diam_list = [300, 600, 900, 1200, 1500] # default is 500
Cm_list = [0.1, 0.3, 0.6, 1.0, 1.3] # default is 1
\# circum = soma.diam * np.pi
# length = soma.L
# A_list = (circum * 10**(-6)) * (length * 10 ** (-6))
input_resistance_list = list()
TauM_list = list()
# colormap
import matplotlib.pylab as pl
num_color = len(diam_list)
colors = pl.cm.viridis(np.linspace(0,1,num_color))
plt.figure()
for diam, n in zip(diam_list, np.arange(num_color)):
    voltage_np, time_np = inject_current(cell = h.Section(name='soma'),
                                         somaLength = 12.6157,
                                         somaDiam = diam,
                                         stimDur = 10,
                                         stimAmp = 0.05,
                                         stimDelay = 10)
    input_resistance_list.append(cal_Ri(voltage_np))
    TauM_list.append(cal_TauM(voltage_np, time_np))
    # plot voltage over time
    plt.plot(time_np, voltage_np,
             label = diam,
             color = colors[n])
plt.ylabel('Membrane potential (mV)')
plt.xlabel('Time (ms)')
plt.legend(frameon=False, title = 'Diameter ($\mu$m)')
plt.show()
```



From the above plot we can see that the voltage change following current injection is larger in a cell with a smaller diameter. This also means that according to Ohms law we will expect the cell with the smallest diameter to have the largest R_i .

1.2.1 Variation of input resistance and time constant with diameter

```
import seaborn as sns
aesthetics()
plt.figure()
plt.subplot(1, 2, 1)
ax = sns.lineplot(diam_list, input_resistance_list)
plt.xlabel('Diameter ($\mu$m)')
plt.ylabel('Input resistance ($\Omega$)')
plt.subplot(1, 2, 2)
# time constant vs. diameter
ax2 = sns.lineplot(diam_list, TauM_list)
plt.xlabel('Diameter ($\mu$m)')
plt.ylabel('Time constant (ms)')
plt.show()
```



Indeed we observe that the R_i decreases with increasing diameter. When we increase the diameter the

capacitance of the cell increases proportionally, however the input resistance decreases (as shown above) which means that the time constant (which is the product of membrane capacitance and resistance) is unaffected. ### Varying multiple variables simultaneously

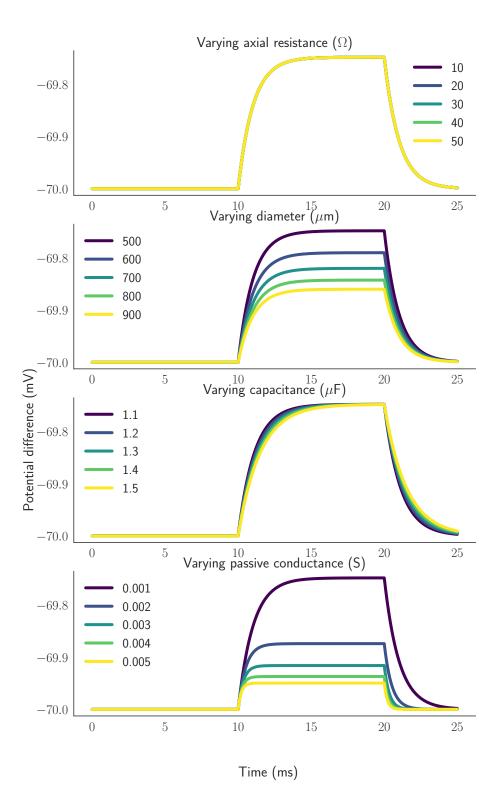
We iterate through each of these list, running our simulations and calculating input resistance and time constants as above.

1.2.1.1 Holding all variables except one

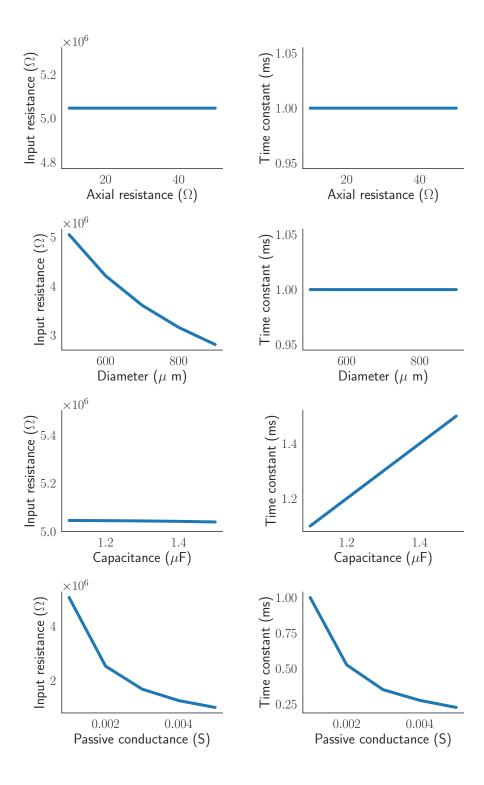
```
# second approach
# holding everything else in their default values, then varying one variable
\# use multiple y-axis for visualisation
Ra_default = 35.4 # uOhm
diam_default = 500 # uM
Cm_default = 1 # uF/cm^2
g_pas_default = 0.001
Ra_list = [10, 20, 30, 40, 50]
diam_list = [500, 600, 700, 800, 900]
Cm_list = [1.1, 1.2, 1.3, 1.4, 1.5]
gpas_list = [0.001, 0.002, 0.003, 0.004, 0.005]
# colormap
# this assumes all the list have the same length
import matplotlib.pylab as pl
num_color = len(Ra_list)
colors = pl.cm.viridis(np.linspace(0,1,num_color))
Ri_Ra_list = list()
Ri_diam_list = list()
Ri_Cm_list = list()
Ri_gpas_list = list()
mTau_Ra_list = list()
mTau_diam_list = list()
mTau Cm list = list()
mTau_gpas_list = list()
```

```
fig, ax = plt.subplots(nrows = 4, ncols = 1, figsize = (12, 20))
fig.text(0.5, 0.04, 'Time (ms)', ha='center')
fig.text(0.04, 0.5, 'Potential difference (mV)', ha='center', rotation = 'vertical')
# ideally, the plotting should be done in Object-oriented approach instead
plt.xlabel('Time (ms)')
ax1 = plt.subplot(4, 1, 1)
plt.title('Varying axial resistance ($\Omega$)')
# the effect of axial resistance on input resistance and time constant
for Ra, n in zip(Ra_list, np.arange(num_color)):
    cell = h.Section(name = 'cell')
    cell.Ra = Ra
   diam = diam_default
    cell.cm = Cm default
    voltage, time = inject_current(cell = cell, somaLength = 12.6157,
somaDiam = diam, stimDur = 10, stimAmp = 0.05, stimDelay = 10)
   Ri_Ra_list.append(cal_Ri(voltage))
   mTau_Ra_list.append(cal_TauM(voltage, time))
   plt.plot(time, voltage,
   label = Ra, color = colors[n])
   plt.legend(frameon = False)
ax1 = plt.subplot(4, 1, 2)
plt.title('Varying diameter ($\mu$m)')
# the effect of diameter on input resistance and time constant
for diam, n in zip(diam_list, np.arange(num_color)):
   cell = h.Section(name = 'cell')
   cell.Ra = Ra default
   diam = diam
    cell.cm = Cm_default
   voltage, time = inject_current(cell = cell, somaLength = 12.6157,
somaDiam = diam, stimDur = 10, stimAmp = 0.05, stimDelay = 10)
   Ri_diam_list.append(cal_Ri(voltage))
   mTau_diam_list.append(cal_TauM(voltage, time))
   plt.plot(time, voltage,
   label = diam, color = colors[n])
   plt.legend(frameon = False)
ax1 = plt.subplot(4, 1, 3)
plt.title('Varying capacitance ($\mu$F)')
# the effect of membrane capacitance on input resistance and time constant
for Cm, n in zip(Cm_list, np.arange(num_color)):
    cell = h.Section(name = 'cell')
   cell.Ra = Ra_default
   diam = diam_default
    cell.cm = Cm
   voltage, time = inject_current(cell = cell, somaLength = 12.6157,
somaDiam = diam, stimDur = 10, stimAmp = 0.05, stimDelay = 10)
   Ri_Cm_list.append(cal_Ri(voltage))
   mTau_Cm_list.append(cal_TauM(voltage, time))
   plt.plot(time, voltage,
   label = Cm, color = colors[n])
   plt.legend(frameon = False)
ax1 = plt.subplot(4, 1, 4)
plt.title('Varying passive conductance (S)')
# the effect of membrane conductance (resistance) on input resistance and time constant
```

```
for g_pas, n in zip(gpas_list, np.arange(num_color)):
    cell = h.Section(name = 'cell')
    cell.Ra = Ra_default
    diam = diam_default
    cell.cm = Cm_default
    voltage, time = inject_current(cell = cell, somaLength = 12.6157,
somaDiam = diam, stimDur = 10, stimAmp = 0.05, stimDelay = 10,
    g_pas = g_pas)
    Ri_gpas_list.append(cal_Ri(voltage))
    mTau_gpas_list.append(cal_TauM(voltage, time))
    plt.plot(time, voltage,
    label = g_pas, color = colors[n])
    plt.legend(frameon = False)
plt.show()
```

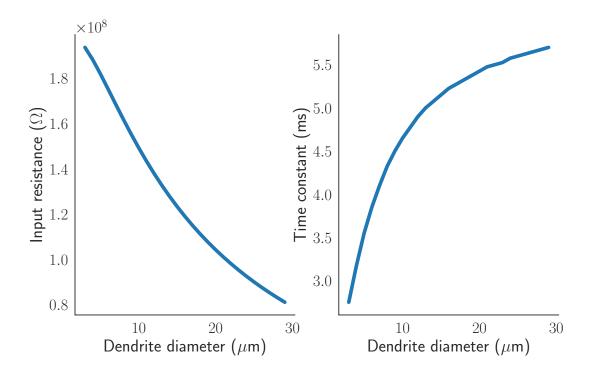


```
plt.figure(figsize = (12, 20))
plt.subplot(4, 2, 1)
plt.plot(Ra_list, Ri_Ra_list)
plt.xlabel('Axial resistance ($\Omega$)')
plt.ylabel('Input resistance ($\Omega$)')
plt.ticklabel_format(style='sci', axis='y', scilimits=(0,0))
plt.subplot(4, 2, 3)
plt.plot(diam_list, Ri_diam_list)
plt.xlabel('Diameter ($\mu$ m)')
plt.ylabel('Input resistance ($\Omega$)')
plt.ticklabel_format(style='sci', axis='y', scilimits=(0,0))
plt.subplot(4, 2, 5)
plt.plot(Cm_list, Ri_Cm_list)
plt.xlabel('Capacitance ($\mu$F)')
plt.ylabel('Input resistance ($\Omega$)')
# plt.ylim([1 * (10 ** 8), 2 * (10 ** 8)])
plt.ylim([5 * 10**6, 5.5 * 10**6])
plt.ticklabel_format(style='sci', axis='y', scilimits=(0,0))
plt.subplot(4, 2, 7)
plt.plot(gpas_list, Ri_gpas_list)
plt.xlabel('Passive conductance (S)')
plt.ylabel('Input resistance ($\Omega$)')
plt.ticklabel_format(style='sci', axis='y', scilimits=(0,0))
plt.subplot(4, 2, 2)
plt.plot(Ra list, mTau Ra list)
plt.xlabel('Axial resistance ($\Omega$)')
plt.ylabel('Time constant (ms)')
plt.ticklabel_format(style='sci', axis='y', scilimits=(0,0))
plt.subplot(4, 2, 4)
plt.plot(diam_list, mTau_diam_list)
plt.xlabel('Diameter ($\mu$ m)')
plt.ylabel('Time constant (ms)')
plt.ticklabel_format(style='sci', axis='y', scilimits=(0,0))
plt.subplot(4, 2, 6)
plt.plot(Cm_list, mTau_Cm_list)
plt.xlabel('Capacitance ($\mu$F)')
plt.ylabel('Time constant (ms)')
plt.ticklabel_format(style='sci', axis='y', scilimits=(0,0))
plt.subplot(4, 2, 8)
plt.plot(gpas_list, mTau_gpas_list)
plt.xlabel('Passive conductance (S)')
plt.ylabel('Time constant (ms)')
plt.ticklabel_format(style='sci', axis='y', scilimits=(0,0))
plt.subplots_adjust(hspace = 0.5, wspace = 0.5)
plt.show()
```



1.3 Adding dendrites and axons with increasing total area

```
from neuron import h, gui
# dendrite_diam_list = [3, 6, 9, 12, 15]
dendrite_diam_list = np.arange(3, 30)
axon_diam_list = [3, 6, 9, 12, 15]
input_resistance_list = list()
TauM_list = list()
theSoma = h.Section(name='theSoma')
dend = h.Section(name='dendrite')
# connect dendrite to soma
dend.connect(theSoma(1))
# h.psection()
# h.topology()
# axon = neuron.h.Section(name='axon')
# axon.connect(soma(0))
# inject current
for dend_diam in dendrite_diam_list:
   dend.diam = dend_diam
    # print(dend.psection())
   voltage, time = inject_current(cell = theSoma, somaLength = 12.6157, somaDiam = 12.6157,
                   stimDur = 10, stimAmp = 0.05, stimDelay = 10, total_time = 25,
                  printProperties = False)
    input_resistance = cal_Ri(voltage)
   TauM = cal_TauM(voltage, time)
    input_resistance_list.append(input_resistance)
    TauM_list.append(TauM)
import seaborn as sns
plt.figure(figsize = (12, 7))
plt.subplot(1, 2, 1)
ax = sns.lineplot(dendrite_diam_list, input_resistance_list)
ax.set(xlabel='Dendrite diameter ($\mu$m)',
       ylabel='Input resistance ($\Omega$)')
plt.subplot(1, 2, 2)
ax = sns.lineplot(dendrite_diam_list, TauM_list)
ax.set(xlabel='Dendrite diameter ($\mu$m)', ylabel='Time constant (ms)')
plt.show()
```



2 Active properties

2.1 Adding HH channels

Add HH Na and K voltage-gated channel to the ball model and inject current steps (add one at a time to see the effectof each one, and then combine).

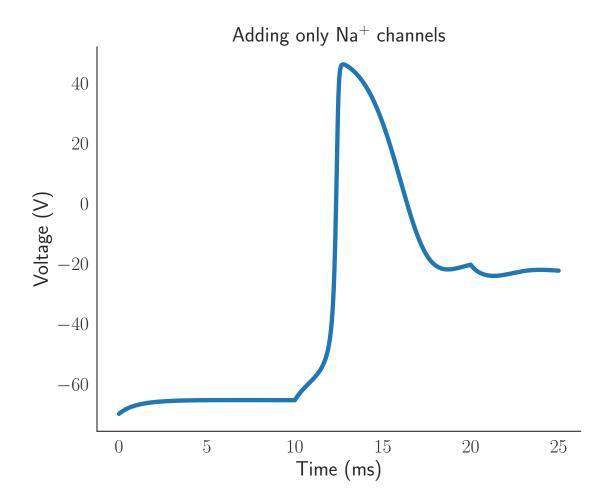
```
from neuron import h, gui
def make_hh_cell(cellName = 'cell',
                 gNa = 0.12, gK = 0.036, gL = 0.0003, eL = -54.3,
                 printProperties = False):
    11 11 11
    makes cell with specified channel conductances
    INPUT
    gNa | Sodium channel conductance (S / cm2)
   gK | Potassium channel conductance
       / Leak conductance
       / Reversal potential (mV)
   Note that default values set in this function are the same as in cell.insert('hh')
   Other implicitly defined properties:
    ena = 50
    ek = -77
    cm = 1
    diam = 500, L = 100, Ra = 35.4
```

```
cell = h.Section(name = cellName)
cell.insert('hh')
cell.gnabar_hh = gNa
cell.gkbar_hh = gK
cell.gl_hh = gL
cell.el_hh = eL

if printProperties is True:
    h.topology()
    h.psection()
```

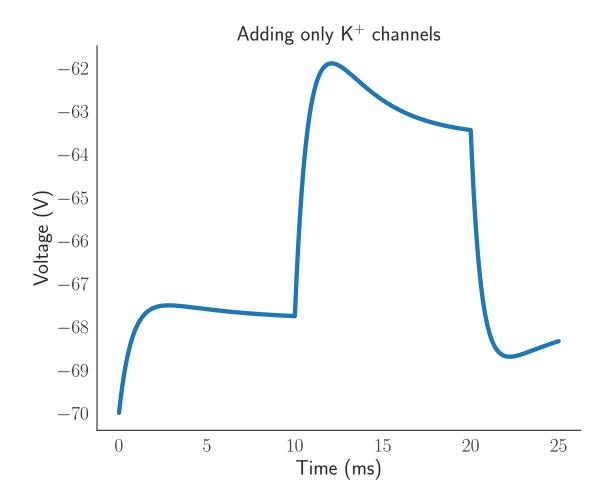
2.1.1 Only adding Na voltage-gated channel

```
from neuron import h, gui
import seaborn as sns
soma_hh = h.Section(name='soma_hh')
soma_hh.insert('hh')
soma_hh.gnabar_hh = 0.12 # peak sodium conductance
soma_hh.gkbar_hh = 0 # remove sodium conductance
soma_hh.el_hh = -54.3 # leaak conductance
# inject current
voltage_np, time_np = inject_current(cell = soma_hh, somaLength = 12.6157,
somaDiam = 12.6157, stimDur = 10, stimAmp = 0.05, stimDelay = 10, total_time = 25)
# plot
plt.figure(figsize = (10, 8))
ax = sns.lineplot(time_np, voltage_np)
ax.set_title('Adding only Na$^+$ channels')
ax.set(xlabel='Time (ms)', ylabel='Voltage (V)')
sns.despine()
plt.show()
# print(time_np)
```



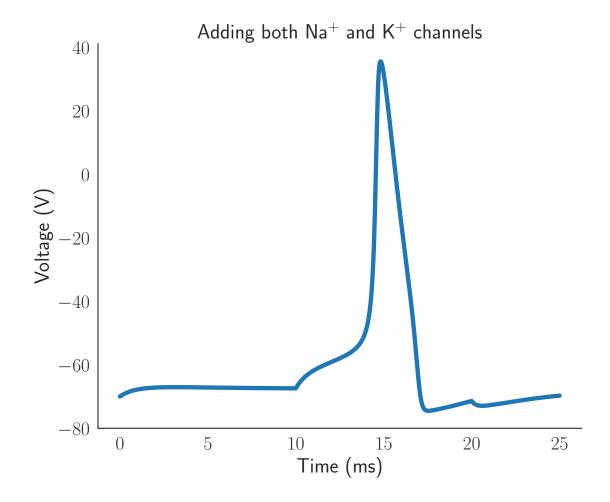
2.1.2 Only adding K+ voltage-gated channels

```
k_only_cell = make_hh_cell(cellName = 'cell', gNa = 0, gK = 0.036, gL = 0.0003)
voltage_np, time_np = inject_current(cell = k_only_cell, somaLength = 12.6157,
somaDiam = 12.6157, stimDur = 10, stimAmp = 0.05, stimDelay = 10, total_time = 25)
plt.figure(figsize = (10, 8))
ax = sns.lineplot(time_np, voltage_np)
ax.set_title('Adding only K$^+$ channels')
ax.set(xlabel='Time (ms)', ylabel='Voltage (V)')
sns.despine()
plt.show()
```



2.1.3 Adding both Na+ and K+ voltage-gated channels

```
hh_cell = make_hh_cell(cellName = 'hh_cell', gNa = 0.12, gK = 0.036, gL = 0.0003)
voltage_np, time_np = inject_current(cell = hh_cell, somaLength = 12.6157,
somaDiam = 12.6157, stimDur = 10, stimAmp = 0.05, stimDelay = 10, total_time = 25)
# aesthetics()
plt.figure(figsize = (10, 8))
ax = sns.lineplot(time_np, voltage_np)
ax.set_title('Adding both Na$^+$ and K$^+$ channels')
ax.set(xlabel='Time (ms)', ylabel='Voltage (V)')
sns.despine()
plt.show()
```



2.1.4 Measure gNa and gKv as a function of current injection at the soma and plot activation and inactivation curves

We first define a function to measure the conductance at the soma.

```
v_clamp_amp1 = 10, v_clamp_amp2 = 30, v_clamp_amp3 = 10,
                   v_clamp_dur1 = 10, v_clamp_dur2 = 20, v_clamp_dur3 = 20,
              printProperties = False,
                  injectCurrent = False):
# cell.insert('pas')
cell.L = somaLength
cell.diam = somaDiam
if printProperties is True:
   print('Cell properties')
   h.psection()
cell.insert('hh') # insert HH properties
# set Vm
# cell.el_hh = vInit
# set resting potential
# for seg in cell:
    seg.hh.el = vInit
if printProperties is True:
   h.psection(cell)
# add point process: current injection
if injectCurrent is True:
   stim = h.IClamp(cell(0.5))
    stim.delay = stimDelay
    stim.dur = stimDur # ms
    stim.amp = stimAmp # nA
v_clamp = h.SEClamp(cell(0.5))
v_clamp.amp1 = v_clamp_amp1
v_clamp.amp2 = v_clamp_amp2
# series resistance
v_{clamp.rs} = 0.01
v_clamp.dur1 = v_clamp_dur1
v_clamp.dur2 = v_clamp_dur2
v_clamp.dur3 = v_clamp_dur3
# measure voltage
voltage = h.Vector()
voltage.record(cell(0.5)._ref_v)
gNa = h.Vector()
gNa.record(cell(0.5)._ref_gna_hh)
gKv = h.Vector()
gKv.record(cell(0.5)._ref_gk_hh)
```

```
# measure time
time = h.Vector()
time.record(h._ref_t)

# h.v_init = vInit # initial voltage
# h.finitialize(vInit)

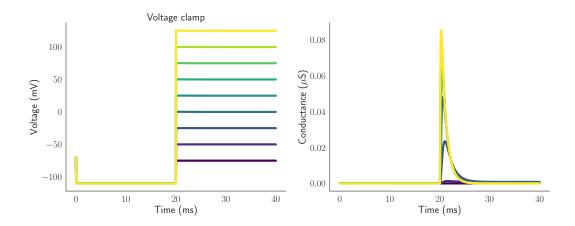
h.load_file('stdrun.hoc')
h.tstop = total_time
h.run() # ms

return gNa, gKv, voltage, time
```

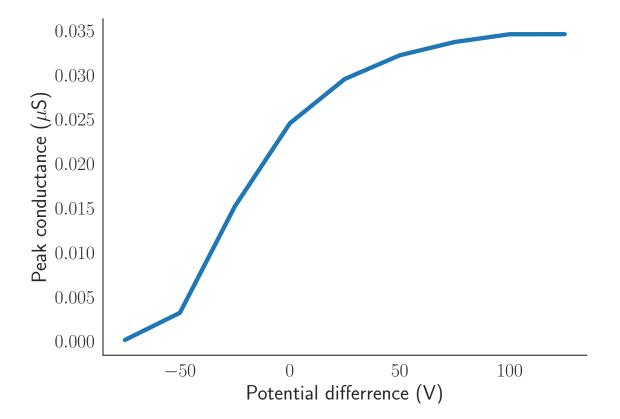
2.1.4.1 Activation curve

```
v clamp amp1 = -110
v_{clamp_amp2_list} = np.arange(-75, 150, 25)
v clamp dur1 = 20
v_clamp_dur2 = 30
curr_amp = 0.5
gNa_measured = list()
gKv_measured = list()
# colormap
import matplotlib.pylab as pl
num_color = len(v_clamp_amp2_list)
colors = pl.cm.viridis(np.linspace(0,1,num_color))
plt.figure(figsize = (20, 7))
ax1 = plt.subplot(1, 2, 1)
ax2 = plt.subplot(1, 2, 2)
for v_clamp_amp2, n in zip(v_clamp_amp2_list, np.arange(num_color)):
    conductance_cell = h.Section(name='conductance_cell')
    gNa, gKv, voltage, time = record_conductance(
    conductance_cell, stimAmp = curr_amp, vInit = v_clamp_amp2,
    stimDelay = 10, total_time = 40,
    v_clamp_amp1 = v_clamp_amp1, v_clamp_amp2 = v_clamp_amp2,
    v_clamp_dur1 = v_clamp_dur1, v_clamp_dur2 = v_clamp_dur2,
    printProperties = False)
    # print(qNa)
    gNa_measured.append(max(gNa))
    gKv_measured.append(max(gKv))
    ax1.plot(time, voltage,
    label = v_clamp_amp2,
             color = colors[n])
    # conductance over time
    ax2.plot(time, gNa,
    label = v_clamp_amp2,
             color = colors[n])
ax1.set_title('Voltage clamp')
ax1.set xlabel('Time (ms)')
ax1.set_ylabel('Voltage (mV)')
ax2.set_xlabel('Time (ms)')
```

```
ax2.set_ylabel('Conductance ($\mu$S)')
plt.show()
```

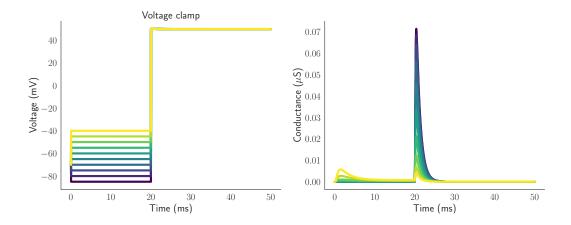


```
plt.figure()
ax = sns.lineplot(v_clamp_amp2_list, gKv_measured)
ax.set(xlabel = 'Potential differrence (V)', ylabel = 'Peak conductance ($\mu$S)')
plt.show()
```

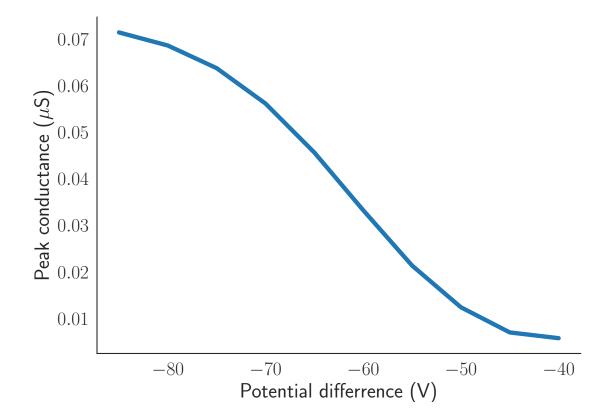


2.1.4.2 Inactivation curve

```
# Vm_list = [-70, -60, -50, -40, -30]
\# v_{clamp\_amp1\_list} = [-100, -90, -80, -70, -60, 20]
\# v\_clamp\_amp1\_list = np.arange(-150, 50, 5)
v_clamp_amp1_list = np.arange(-85, -35, 5)
v_{clamp_amp2} = 50
v_{clamp_dur1} = 20
v clamp dur2 = 30
curr amp = 0.5
gNa_measured = list()
gKv_measured = list()
aesthetics()
# colormap
import matplotlib.pylab as pl
num_color = len(v_clamp_amp1_list)
colors = pl.cm.viridis(np.linspace(0,1,num_color))
plt.figure(figsize = (20, 7))
ax1 = plt.subplot(1, 2, 1)
ax2 = plt.subplot(1, 2, 2)
for v_clamp_amp1, n in zip(v_clamp_amp1_list, np.arange(num_color)):
    conductance_cell = h.Section(name='conductance_cell')
    gNa, gKv, voltage, time = record_conductance(conductance_cell, stimAmp = curr_amp,
   vInit = -70, stimDelay = 10, total_time = 50,
   v_clamp_amp1 = v_clamp_amp1, v_clamp_amp2 = v_clamp_amp2,
   v_clamp_dur1 = v_clamp_dur1, v_clamp_dur2 = v_clamp_dur2,
   printProperties = False)
   gNa measured.append(max(gNa))
   gKv_measured.append(max(gKv))
   ax1.plot(time, voltage,
             label = v_clamp_amp1,
             color = colors[n])
    ax2.plot(time, gNa,
             label = v_clamp_amp1,
             color = colors[n])
ax1.set_title('Voltage clamp')
ax1.set_xlabel('Time (ms)')
ax1.set_ylabel('Voltage (mV)')
ax2.set_xlabel('Time (ms)')
ax2.set_ylabel('Conductance ($\mu$S)')
\# plt.legend(frameon = False, title = 'Initial holding voltage n (mV)')
plt.show()
```

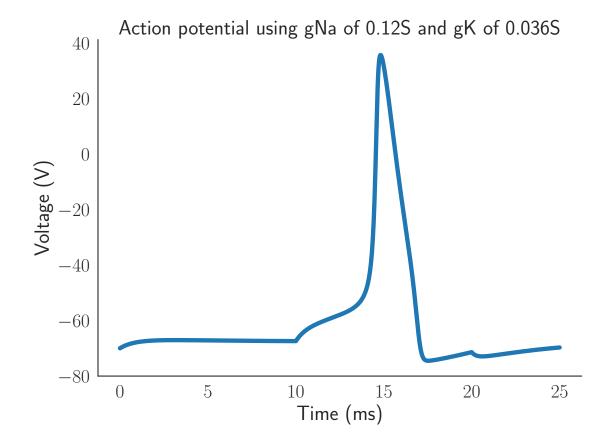


```
plt.figure()
ax = sns.lineplot(v_clamp_amp1_list, gNa_measured)
plt.xlabel('Potential differrence (V)')
plt.ylabel('Peak conductance ($\mu$S)')
plt.show()
```



2.2 Play with gNa and gK density to make an action potential

```
gNa_custom = 0.12
gK_custom = 0.036
ap_cell = make_hh_cell(cellName = 'ap_cell', gNa = gNa_custom, gK = gK_custom, gL = 0.0003, eL = -54.3,
voltage_np, time_np = inject_current(cell = ap_cell, somaLength = 12.6157,
somaDiam = 12.6157, stimDur = 10, stimAmp = 0.05, stimDelay = 10, total_time = 25)
aesthetics()
plt.figure()
ax = sns.lineplot(time_np, voltage_np)
ax.set_title('Action potential using gNa of %.2fS and gK of %.3fS' % (gNa_custom, gK_custom))
ax.set(xlabel='Time (ms)', ylabel='Voltage (V)')
sns.despine()
plt.show()
```



3 Syanptic integration

3.1 Add a single alpha excitatory synapse in the soma and then in the dendrite

Progressively far away, and record at the soma. What changes?

Background:

- A synapse (for the purpose of this simulation) is a location of transmitter release, which then binds to post-synaptic receptors at the soma to cause change in coductance for one or more ions
- The conductance change (usually increase) can be described by a mathematical function
- An alpha synapse is a synapse in which the conductance change in the post-synapse cell can be described by the *alpha function*
- In NEURON, the alpha synapse is a point process (ie. it is assumed to be a point source of current along the cell)

There seems to be multiple mathematical definition of the alpha function, and NEURON implements the alpha function (see ref 3) via:

$$g(t) = g_{\text{max}} \alpha t e^{1 - \alpha t}$$

References:

- AlphaSynapse
- Alpha function, Wolfram
- NEURON forum on alpha function

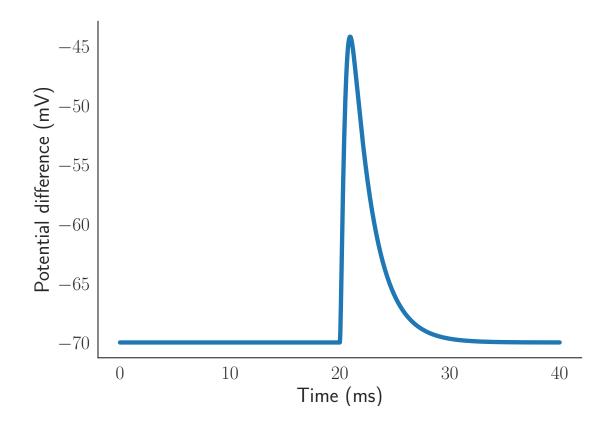
We first define functions to make a cell with alpha synapse directed to the dendrite and to record the potential at the soma.

```
from neuron import h, gui
import seaborn as sns
import matplotlib.pyplot as plt
def sim_alpha_synapse(alpha_loc = 0.5, record_loc = 0.5, simDur = 40, vInit = -70,
                     alpha_onset = 20, alpha_gmax = 1, printProperties = False,
                     nSomaSegment = 1, nDendSegment = 300,
                    somaLength = 1000, recordSegment = 'soma',
                     dendriteNum = 0):
    11 11 11
   Create alpha synapse, then simulate to record potential difference over time
    record_loc | location of recording the alpha synapse
             | duration of the simulation (ms)
   soma = h.Section(name='soma')
   soma.insert('pas')
   soma.L = 20 #somaLength
   soma.diam = 20 #
   # alpha synapse on soma
    \# soma.nseq = nSomaSegment
    # asyn = h.AlphaSynapse(soma(alpha_loc))
    # alpha synapse on dendrite
   dend = h.Section(name = 'dend')
   dend.L = 300
   dend.diam = 1
   dend.nseg = nDendSegment
   asyn = h.AlphaSynapse(dend(alpha_loc))
   dend.connect(soma(0))
```

```
asyn.onset = alpha_onset
asyn.gmax = alpha_gmax
if printProperties is True:
    h.psection(soma)
    h.psection(dend)
    # print(dir(asyn))
v_vec = h.Vector()
                               # Membrane potential vector
t_vec = h.Vector()
                               # Time stamp vector
if recordSegment == 'soma':
    # print('Recording at the soma')
    v_vec.record(soma(record_loc)._ref_v)
elif recordSegment == 'dendrite':
    # print('Recording at the dendrite')
    v_vec.record(dend(record_loc)._ref_v)
t_vec.record(h._ref_t)
h.v_init = vInit
h.tstop = simDur
h.run()
return t_vec, v_vec
```

Then we run the simulation:

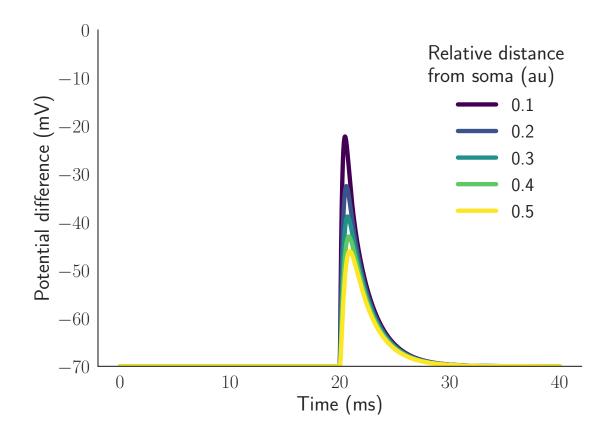
```
time, voltage = sim_alpha_synapse()
import matplotlib.pyplot as plt
plt.figure()
plt.plot(time, voltage)
plt.xlabel('Time (ms)')
plt.ylabel('Potential difference (mV)')
plt.show()
```



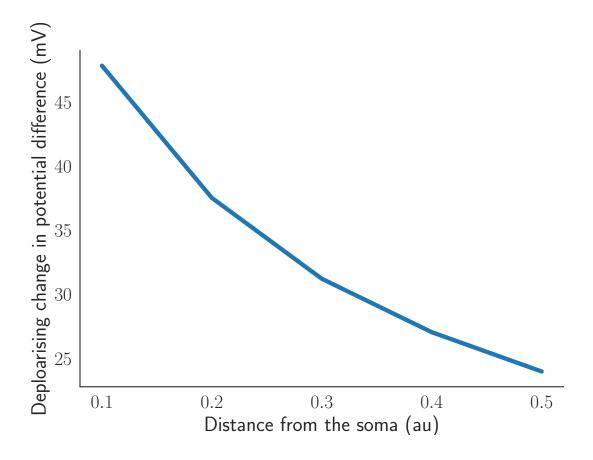
3.1.1 Moving the alpha synapse progressively far away

```
# progressively far away alpha synapse
location_list = [0.1, 0.2, 0.3, 0.4, 0.5]
alpha_onset_list = [1, 5, 10, 15, 20]
peak_voltage = []
# colormap
import matplotlib.pylab as pl
num_color = len(location_list)
colors = pl.cm.viridis(np.linspace(0,1,num_color))
plt.figure()
for loc, n in zip(location_list, np.arange(num_color)):
    time, voltage = sim_alpha_synapse(alpha_loc = loc, record_loc = 0.5, simDur = 40,
                                      nSomaSegment = 100, somaLength = 1000,
                                      alpha_gmax = 0.5,
                                      printProperties = False)
    sns.lineplot(time, voltage,
                 label = loc,
                 color = colors[n])
    peak_voltage.append(max(voltage) - min(voltage))
    plt.xlabel('Time (ms)')
```

```
plt.ylabel('Potential difference (mV)')
plt.ylim([-70, 0])
plt.legend(frameon=False, title = 'Relative distance \n from soma (au)')
plt.show()
```

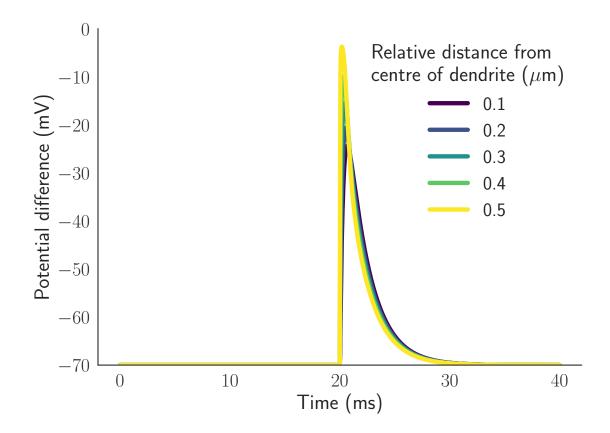


```
# plot the peak potential difference against the distance
plt.figure()
sns.lineplot(location_list, peak_voltage)
plt.ylabel('Deploarising change in potential difference (mV)')
plt.xlabel('Distance from the soma (au)')
plt.show()
```



As expected we see that the amplitude of the voltage change elicted by the alphasynapse decreases with distance due to the passive properties of the cell. As a result when the alphasynapse is further away from the some the depolarising change recorded as the soma is smaller then when it is closer to the soma.

3.2 Record at the dendrite where the synapse is whilst moving the alpha synapse progressively far away



When we record the voltage at the dendrite as the alphasynapse is activated we observe that the size of the voltage change is higher than when the synapse was on near the soma. The high voltage change is due to the dendrites smaller size.

4 Appendix

4.1 Varying multiple variables simultaneously

```
import itertools
import pandas as pd
# variable lists
```

```
diam_list = [3, 6, 9, 12, 15]
Cm_list = [0.1, 0.3, 0.6, 1.0, 1.3]
Ri_list = list()
mTau_list = list()
# using ranges
Ra_list = np.arange(10, 50, 2)
Cm_list = np.arange(0.1, 1.3, 0.1)
# varying all 3 variables
# variableCart, variableIter = itertools.tee(
# itertools.product(Ra_list, diam_list, Cm_list))
# cartesian product of the variables
# using tee to make two copies of the same iterator so I can use it twice
# varying just 2 variables
diam = 3
variableCart, variableIter = itertools.tee(itertools.product(Ra_list, Cm_list))
# df = pd.DataFrame(list(variableCart),
                    columns = ['Ra', 'Diam', 'Cm'])
df = pd.DataFrame(list(variableCart),
                  columns = ['Ra', 'Cm'])
for var_list in variableIter:
    cell = h.Section(name = 'cell')
    # 3 variables
    # cell.Ra = var_list[0] # axial resistance
    # diam = var_list[1]
    # cell.cm = var list[2]
    # 2 variables
    cell.Ra = var_list[0]
    cell.cm = var_list[1]
    diam = 50000 # default to 500
    voltage, time = inject_current(cell = cell, somaLength = 12.6157,
somaDiam = diam, stimDur = 10, stimAmp = 0.05, stimDelay = 10)
    Ri_list.append(cal_Ri(voltage))
    mTau_list.append(cal_TauM(voltage, time))
df['Ri'] = Ri_list
df['mTau'] = mTau_list
# print preview of the table
# df
Cm_list = np.arange(0.1, 2, 0.1)
diam_list = np.arange(250, 252, 0.1) # default diam = 500
# lists to store input resistance and time constant
Ri_list = list()
mTau list = list()
variableCart, variableIter = itertools.tee(itertools.product(Cm_list, diam_list))
df = pd.DataFrame(list(variableCart),
                  columns = ['Capacitance', 'Diameter'])
for var_list in variableIter:
    cell = h.Section(name = 'cell')
    # 2 variables
  # cell.insert('pas')
```

```
# cell.g_pas = var_list[0]
    # diam = var_list[1]
    cell.cm = var_list[0] # default capacitance
    cell.Ra = 35.4 # default axial resistance
   diam = var_list[1]
   voltage, time = inject current(cell = cell, somaLength = 12.6157,
somaDiam = diam, stimDur = 10, stimAmp = 0.05, stimDelay = 10)
   Ri list.append(cal Ri(voltage))
   mTau_list.append(cal_TauM(voltage, time))
df['Ri'] = Ri list
df['mTau'] = mTau list
# 3D surface plot to show the effect of the 2 variables on input resistance
from mpl_toolkits.mplot3d import Axes3D
import matplotlib.pyplot as plt
import pandas as pd
import seaborn as sns
# Make the plot
# fiq = plt.fiqure(fiqsize=(8, 6), dpi = 300)
fig = plt.figure(figsize=(10, 7))
ax = fig.gca(projection='3d')
ax.plot_trisurf(df['Capacitance'], df['Diameter'], df['Ri'],
                cmap=plt.cm.viridis, linewidth=0,
                edgecolor='none',
                antialiased=True)
ax.set(xlabel='Capacitance ($\mu$F/cm$^2$)', ylabel='Diameter ($\mu$ m)',
      zlabel = 'Input resistance ($\Omega$)')
# plt.show()
# to Add a color bar which maps values to colors.
# surf=ax.plot_trisurf(df['Capacitance'], df['Diameter'], df['Ri'],
                       cmap=plt.cm.viridis, linewidth=0.2)
# fiq.colorbar(surf, shrink=0.5, aspect=5)
# plt.show()
# padding for axis
ax.xaxis.labelpad = 20
ax.yaxis.labelpad = 20
ax.zaxis.labelpad = 20
# change the label so it doens't overlap
ax.set_zlabel(zlabel='$R_i$')
# padding for tick marks
ax.ticklabel_format(axis = 'z', style = 'sci', scilimits = [0, 0])
ax.tick_params(axis = 'z', pad = 10)
# Rotate it
ax.view_init(45, 45)
plt.show()
# 3D surface plot to show the effect of the 2 variables on input resistance
from mpl_toolkits.mplot3d import Axes3D
import matplotlib.pyplot as plt
import pandas as pd
import seaborn as sns
```

```
# Make the plot
fig = plt.figure(figsize=(8, 6))
fig = plt.figure()
ax = fig.gca(projection='3d')
ax.plot_trisurf(df['Ra'], df['Cm'], df['Ri'], cmap=plt.cm.viridis, linewidth=0.2)
ax.set(xlabel='Axial resistance ($\Omega$)',
       ylabel='Membrane capicitance ($\mu$F/cm$^2$)',
      zlabel = 'Input resistance ($\Omega$)')
# plt.show()
# to Add a color bar which maps values to colors.
surf=ax.plot_trisurf(df['Ra'], df['Cm'], df['Ri'], cmap=plt.cm.viridis, linewidth=0.2)
# fig.colorbar(surf, shrink=0.5, aspect=5) # toggle colorbar
# plt.show()
# padding for axis
ax.xaxis.labelpad = 20
ax.yaxis.labelpad = 20
ax.zaxis.labelpad = 20
# padding for tick mark
ax.tick_params(axis = 'z', pad = 10)
# Rotate it
ax.view_init(45, 45)
plt.show()
```

4.2 Holding all variables except one

```
# based on: https://matplotlib.org/gallery/ticks_and_spines/multiple_yaxis_with_spines.html
# but instead of twinx, I am using twiny, and so the top rather than right axis is reset
import matplotlib.pyplot as plt
aesthetics(font_scale = 2, line_width = 3) # 2, 3
def make_patch_spines_invisible(ax):
   ax.set_frame_on(True)
    ax.patch.set_visible(False)
   for sp in ax.spines.values():
        sp.set_visible(False)
fig, host = plt.subplots()
fig.subplots_adjust(right=0.75)
par1 = host.twiny()
par2 = host.twiny()
# Offset the right spine of par2. The ticks and label have already been
# placed on the right by twinx above.
# par2.spines["right"].set_position(("axes", 1.2))
par2.xaxis.set_ticks_position('bottom') # set the position of the second x-axis to bottom
par2.xaxis.set_label_position('bottom') # set the position of the second x-axis to bottom
par2.spines["bottom"].set_position(('axes', -0.2))
# Having been created by twinx, par2 has its frame off, so the line of its
# detached spine is invisible. First, activate the frame but make the patch
# and spines invisible.
make_patch_spines_invisible(par2)
# Second, show the right spine.
# par2.spines["right"].set_visible(True)
par2.spines["bottom"].set_visible(True)
```

```
p1, = host.plot(Ra_list, Ri_Ra_list, "b-", label="Axial resistance")
p2, = par1.plot(diam_list, Ri_diam_list, "r-", label="Diameter")
p3, = par2.plot(Cm_list, Ri_Cm_list, "g-", label="Membrane capacitance")
host.set_xlabel("Axial resistance ($\Omega$)")
host.set_ylabel("Input resistance ($\Omega$)")
par1.set_xlabel('Diameter ($\mu$m)')
par2.set_xlabel('Capacitance ($\mu$F)')
host.xaxis.label.set_color(p1.get_color())
par1.xaxis.label.set_color(p2.get_color())
par2.xaxis.label.set_color(p3.get_color())
tkw = dict(size=4, width=1.5)
host.tick_params(axis='x', colors=p1.get_color(), **tkw)
par1.tick_params(axis='x', colors=p2.get_color(), **tkw)
par2.tick_params(axis='x', colors=p3.get_color(), **tkw)
host.tick_params(axis='y', **tkw)
lines = [p1, p2, p3]
host.legend(lines, [l.get_label() for l in lines], frameon = False)
plt.subplots_adjust(bottom = 0.5) # adjust bottom margin to show second bottom x-axis
plt.show()
```