SNPsea Reference Manual

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Introduction

SNPsea is a general algorithm to identify cell types, tissues, and pathways likely to be affected by risk loci.

For example, with a gene expression matrix containing expression profiles for multiple cell types, we identify genes in linkage disequilibrium with trait-associated SNPs and score them for specificity to each cell type. We compare the score to a null distribution by sampling random SNP sets matched on the number of linked genes. To evaluate significance, we calculate an exact permutation p-value.

This implementation is generalized, so you may provide (1) a continuous gene matrix with gene expression (or any other values) or (2) a binary gene matrix with presence/absence 1/0 values.

The columns of the matrix could be tissues, cell types, GO annotation codes, or any other types of conditions. Continuous matrices must be normalized before running SNPsea so that columns are directly comparable to each other.

In general, this analysis is appropriate when you are interested in testing for enrichment of conditionspecificity of genes linked to a set of trait-associated SNPs.

If trait-associated alleles impact a small number of pathogenic tissues or cell types, we hypothesize that the subset of genes with critical functions in those pathogenic cell types are likely to be within trait-associated loci.

We assume that a gene's specificity to a given cell type or condition is a reasonable indicator of the gene's importance to its function.

If you benefit from this method, please cite:

Slowikowski, K. et al. SNPsea: an algorithm to identify cell types, tissues, and pathways affected by risk loci. Manuscript in progress.

See additional examples:

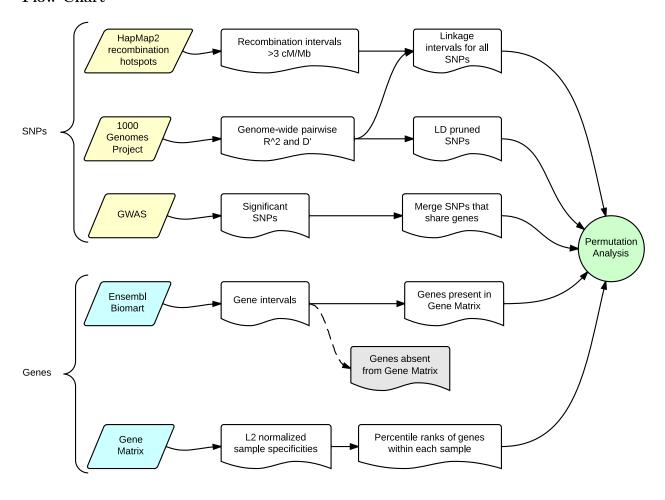
Hu, X. et al. Integrating autoimmune risk loci with gene-expression data identifies specific pathogenic immune cell subsets. The American Journal of Human Genetics 89, 496–506 (2011). PubMed

Contact

Please contact me with questions and comments: slowikow@broadinstitute.org

Visual Summary

Flow Chart



This flow chart shows the input data required to perform the analysis, and a summary of the intermediate steps.

Installation

Linux 64-bit

Download the binary and run it: https://github.com/slowkow/snpsea/releases

Mac

To compile C++ code with the required dependencies, you need XCode and MacPorts: http://guide.macports.org/#installing.xcode

All other platforms

The source code is available: https://github.com/slowkow/snpsea

Install the dependencies:

```
# Ubuntu
sudo apt-get install build-essential libopenmpi-dev libgs10-dev
# Mac
    Install MacPorts: http://www.macports.org/
    Then do this:
sudo port selfupdate
sudo port install gcc48 openmpi gsl
# Broad Institute
    You can add this to your .my.bashrc or .my.cshrc
use .gcc-4.8.1 .openmpi-1.4 .gsl-1.14
Download and compile the code:
    Clone with git, so you can get updates with 'git pull'
git clone https://github.com/slowkow/snpsea.git
cd snpsea
    or if you don't have git
curl -LOk https://github.com/slowkow/snpsea/archive/master.zip
unzip master.zip
cd snpsea-master
   Compile.
cd src
make
   Move the executables wherever you like.
mv ../bin/snpsea* ~/bin/
Download the required data and run SNPsea:
mkdir ../snpsea/data
cd ../snpsea/data
curl -LOk http://files.figshare.com/1307287/SNPsea_data_20131204.zip
unzip SNPsea_data_20131204.zip
snpsea
```

Data

Download the compressed archive with data required to perform this analysis here (138M): http://dx.doi.org/10.6084/m9.figshare.871430

GWAS SNPs

Celiac_disease-Trynka2011-35_SNPs.gwas HDL_cholesterol-Teslovich2010-46_SNPs.gwas Multiple_sclerosis-IMSGC-51_SNPs.gwas Red_blood_cell_count-Harst2012-45_SNPs.gwas

GeneAtlas2004.gct.gz # Gene Atlas 2004 gene expression matrix GO2013.gct.gz # Gene Ontology 2013 gene annotation matrix ImmGen2012.gct.gz # ImmGen 2012 gene expression matrix

NCBIgenes2013.bed.gz # NCBI gene intervals Lango2010.txt.gz # LD-pruned SNPs

TGP2011.bed.gz # 1000 Genomes Project SNP linkage intervals

Celiac_disease-Trynka2011-35_SNPs.gwas

35 SNPs associated with Celiac disease taken from Table 2. Positions are on hg19. All SNPs have P $\leq 5e-8$.

doi:10.1038/ng.998 PMID: 22057235

Trynka G, Hunt KA, Bockett NA, et al. Dense genotyping identifies and localizes multiple common and rare variant association signals in celiac disease. Nat Genet. 2011;43(12):1193-201.

http://www.ncbi.nlm.nih.gov/pubmed/22057235

HDL cholesterol-Tesolvich2010-46 SNPs.gwas

46 SNPs associated with HDL taken from Supplementary Table 2. Positions are on hg19. All SNPs have $P \le 5e-8$.

doi:10.1038/nature09270

PMID: 20686565

Teslovich TM, Musunuru K, Smith AV, et al. Biological, clinical and population relevance of 95 loci for blood lipids. Nature. 2010;466(7307):707-13.

http://www.ncbi.nlm.nih.gov/pubmed/20686565

Multiple_sclerosis-IMSGC-51_SNPs.gwas

51 SNPs associated with Multiple Sclerosis taken from Supplementary Table A. Positions are on hg19. All SNPs have $P \le 5e-8$.

doi:10.1038/nature10251

PMID: 21833088

Sawcer S, Hellenthal G, Pirinen M, et al. Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. Nature. 2011;476(7359):214-9.

http://www.ncbi.nlm.nih.gov/pubmed/21833088

$Red_blood_cell_count\text{-}Harst2012\text{-}45_SNPs.gwas$

45 SNPs associated with red blood cell count (RBC) taken from Table 1. Positions are on hg19. All SNPs have $P \le 5e-8$.

doi:10.1038/nature11677

PMID: 23222517

Van der harst P, Zhang W, Mateo leach I, et al. Seventy-five genetic loci influencing the human red blood cell. Nature. 2012;492(7429):369-75.

http://www.ncbi.nlm.nih.gov/pubmed/23222517

GeneAtlas 2004.gct.gz

Gene expression data for 79 human tissues from GSE1133. Replicates for each tissue profile were averaged. For each gene, the single probe with the largest minimum was selected.

Su AI et al. A gene atlas of the mouse and human protein-encoding transcriptomes. Proc Natl Acad Sci U S A, 2004 Apr 9;101(16):6062-7

http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE1133

GO2013.gct.gz

A GCT formatted gene matrix with 1s and 0s indicating presence or absence of genes in Gene Ontology annotations. 19,111 genes in 1,751 Gene Ontology annotations.

http://www.geneontology.org/

ImmGen2012.gct.gz

Gene expression data for 249 blood cell types from GSE15907. Replicates for each cell type profile were averaged. For each gene, the single probe with the largest minimum was selected.

Immunological Genome Project

http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE15907

NCBIgenes 2013.bed.gz

All human start and stop positions taken from:

ftp://ftp.ncbi.nlm.nih.gov/gene/DATA/gene2refseq.gz

Lango2010.txt.gz

A list of SNPs that span the whole genome, pruned by linkage disequilibrium (LD). SNPsea samples null SNP sets matched on the number of genes in the user's SNP set from this list. See this paper for more information:

Lango allen H, Estrada K, Lettre G, et al. Hundreds of variants clustered in genomic loci and biological pathways affect human height. Nature. 2010;467(7317):832-8.

http://www.ncbi.nlm.nih.gov/pubmed/20881960

TGP2011.bed.gz

Linkage intervals for a filtered set of SNPs from the 1000 Genomes Project Phase 1 (May 21, 2011). SNP genotypes were obtained from the BEAGLE release v3 website and processed to create linkage intervals for each SNP. The linkage intervals were extended to the nearest HapMap recombination hotspot with >3 cM/Mb recombination rate.

```
http://www.1000genomes.org/
http://bochet.gcc.biostat.washington.edu/beagle/1000_Genomes.phase1_release_v3/
http://hapmap.ncbi.nlm.nih.gov/downloads/
```

C++ Libraries

To compile SNPsea, you will need a modern C++ compiler that supports c++0x and the dependencies listed below.

See: Installation

intervaltree

a minimal C++ interval tree implementation

Eigen

Eigen is a C++ template library for linear algebra: matrices, vectors, numerical solvers, and related algorithms.

OpenMPI

MPI is a standardized API typically used for parallel and/or distributed computing. Open MPI is an open source, freely available implementation.

GSL - GNU Scientific Library

The GNU Scientific Library (GSL) is a numerical library for C and C++ programmers.

GCC, the GNU Compiler

The GNU Compiler Collection is a compiler system produced by the GNU Project supporting various programming languages.

I use c++0x features in my C++ code, so you must use a compiler that supports them. I compiled successfully with versions 4.6.3 (the default version for Ubuntu 12.04) and 4.8.1.

Python Packages

To plot visualizations of the results, you will need Python 2.7 and the packages listed below.

Instructions: Install with pip:

pip install docopt numpy pandas matplotlib

Note: The packages available on the Ubuntu repositories may be outdated and might fail to work. So, avoid using apt-get for these dependencies.

docopt

Command-line interface description language.

numpy

NumPy is the fundamental package for scientific computing with Python.

pandas

pandas is an open source, BSD-licensed library providing high-performance, easy-to-use data structures and data analysis tools for the Python programming language.

matplotlib

matplotlib is a python 2D plotting library which produces publication quality figures in a variety of hardcopy formats and interactive environments across platforms.

Note: On a server with no display, please edit your matplotlibre file to use the Agg backend:

```
perl -i -pe 's/^(\s*(backend).*)$/#$1\n$2:Agg/' ~/.matplotlib/matplotlibrc
```

Otherwise, you may see an error message like this:

```
_tkinter.TclError: no display name and no $DISPLAY environment variable
```

R Packages

Some visualizations use R and ggplot2 instead of Python and matplotlib.

Instructions: Start a session in R and run:

```
install.packages(c("data.table", "reshape2", "gap", "ggplot2"))
```

data.table

Extension of data.frame for fast indexing, fast ordered joins, fast assignment, fast grouping and list columns.

reshape2

Flexibly reshape data: a reboot of the reshape package.

gap

Genetic analysis package.

ggplot2

An implementation of the Grammar of Graphics.

Usage

Example

Here is a Bash script with a usage example:

```
options=(
                        Red_blood_cell_count-Harst2012-45_SNPs.gwas
    --snps
                         GeneAtlas2004.gct.gz
    --gene-matrix
    --gene-intervals
                        NCBIgenes2013.bed.gz
    --snp-intervals
                        TGP2011.bed.gz
    --null-snps
                        Lango2010.txt.gz
    --out
                        out
    --slop
                         10e3
    --threads
                        4
                        0
    --null-snpsets
    --min-observations
                         100
    --max-iterations
                         1e7
)
snpsea ${options[*]}
```

SNPs will test SNPs associated with Red blood cell count for tissue-specific expression of linked genes across 79 human tissues in the Gene Atlas expression matrix. Each tissue will be tested up to 10 million times with matched random SNP sets, or testing will stop for a tissue if 100 matched SNP sets achieve a higher specificity score than the user's SNPs.

Options

All input files may optionally be compressed with gzip.

Required

snps ARG	Text file with SNP identifiers in the first column. Instead of a file name, you may use 'randomN' with an integer N for a random SNP list of length N.
gene-matrix ARG	Gene matrix file in GCT format. The Name column must contain the same gene identifiers as ingene-intervals.
gene-intervals ARG	BED file with gene intervals. The fourth column must contain the same gene identifiers as ingene-matrix.

--snp-intervals ARG BED file with all known SNP intervals. The fourth

column must contain the same SNP identifiers as

in --snps and --null-snps.

--null-snps ARG Text file with names of SNPs to sample when

generating null matched or random SNP sets.

These SNPs must be a subset of --snp-intervals.

--out ARG Create output files in this directory. It will be

created if it does not already exist.

Optional

--condition ARG Text file with a list of columns in --gene-matrix

to condition on before calculating p-values. Each column in --gene-matrix is projected onto each column listed in this file and its projection is

subtracted.

--slop ARG If a SNP interval overlaps no gene intervals,

extend the SNP interval this many nucleotides

further and try again.

[default: 10000]

--threads ARG Number of threads to use.

[default: 1]

--null-snpsets ARG Test this many null matched SNP sets, so you can

compare your results to a distribution of null

results. [default: 0]

--min-observations ARG Stop testing a column in --gene-matrix after

observing this many null SNP sets with specificity scores greater or equal to those obtained with the SNP set in --snps. Increase this value to obtain more accurate p-values.

[default: 25]

--max-iterations ARG Maximum number of null SNP sets tested for each

column in --gene-matrix. Increase this value to

resolve smaller p-values.

[default: 10000]

Input File Formats

--snps ARG

You must provide one or more comma-separated text files. SNP identifiers must be listed one per line. Only the first column is used.

head Red_blood_cell_count-Harst2012-45_SNPs.gwas

- # Harst et al. 2012
- # doi:10.1038/nature11677
- # PMID: 23222517
- # 45 SNPs associated with red blood cell count (RBC) taken from Table 1.
- # Positions are on hg19. SNPs are included if P <= 5e-8.

CHR POS SNP P

chr1	40069939	rs3916164	3e-10
chr1	158575729	rs857684	4e-16
chr1	199007208	rs7529925	8e-09
chr1	248039451	rs3811444	5e-10

Instead of providing a file with SNPs, you may use "randomN" like this:

--snps random20

to sample 20 random SNPs from the --snp-intervals file.

--gene-matrix ARG

You must provide a single gene matrix that must be in GCT format.

zcat GeneAtlas2004.gct.gz | cut -f1-4 | head

#1.2

17581	79		
Name	Description	Colorectal_Adenocarcinoma	Whole_Blood
1	A1BG	115.5	209.5
2	A2M	85	328.5
9	NAT1	499	1578
10	NAT2	115	114
12	SERPINA3	419.5	387.5
13	AADAC	125	252.5
14	AAMP	2023	942.5

--condition ARG (Optional)

You may provide column names present in the **--gene-matrix** file, one per line. The matrix will be conditioned on these columns before the analysis is performed to help you identify secondary signals independent of these columns. Binary (0, 1) matrices will not be conditioned.

head conditions.txt

Whole_Blood

--gene-intervals ARG

You must provide gene intervals in BED format with a fourth column that contains the same gene identifiers as those present in the Name column of the **--gene-matrix** GCT file. Only the first four columns are used.

zcat NCBIgenes2013.bed.gz | head

chr1	10003485	10045555	64802	NMNAT1
chr1	100111430	100160096	54873	PALMD
chr1	100163795	100164756	100129320	HMGB3P10
chr1	100174205	100232185	391059	FRRS1
chr1	10027438	10027515	100847055	MIR5697
chr1	100308165	100308317	100270894	RPL39P9
chr1	100315632	100389578	178	AGL
chr1	100433941	100435837	730081	L0C730081
chr1	100435344	100492534	23443	SLC35A3
chr1	100503669	100548932	64645	HIAT1

--snp-intervals ARG

SNP linkage intervals must be specified in BED format and include a fourth column with the SNP identifiers. The linkage intervals assigned to the trait-associated SNPs you provide with **--snps** are taken from this file.

zcat TGP2011.bed.gz | head

```
chr1
        0
            254996 rs113759966
chr1
        0
           254996 rs114420996
chr1
        0
           254996 rs114608975
           254996 rs115209712
chr1
        0
        0
           254996 rs116400033
chr1
           254996 rs116504101
chr1
chr1
           254996 rs12184306
        0
chr1
        0
           254996 rs12184307
        0
           254996 rs138808727
chr1
        0
           254996 rs139113303
chr1
```

--null-snps ARG

The null SNPs file must have one SNP identifier per line. Only the first column is used. The identifiers must be a subset of the identifiers in **--snp-intervals**.

```
zcat Lango2010.txt.gz | head
rs58108140 chr1
                    10583
rs180734498 chr1
                    13302
rs140337953 chr1
                    30923
rs141149254 chr1
                    54490
rs2462492
            chr1
                    54676
rs10399749 chr1
                    55299
rs189727433 chr1
                    57952
rs149755937 chr1
                    59040
rs77573425 chr1
                    61989
rs116440577 chr1
                    63671
```

Output Files

The usage example shown above produces the following output files:

```
out/
    args.txt
    condition_pvalues.txt
    null_pvalues.txt
    snp_condition_scores.txt
    snp_genes.txt
```

args.txt

The command line arguments needed to reproduce the analysis.

```
cat args.txt
# SNPsea v1.0.2
                   Red_blood_cell_count-Harst2012-45_SNPs.gwas
--snps
--gene-matrix
                   GeneAtlas2004.gct.gz
--gene-intervals
                   NCBIgenes2013.bed.gz
--snp-intervals
                   TGP2011.bed.gz
--null-snps
                   Lango2010.txt.gz
--out
                   out
                   single
--score
--slop
                   100000
--threads
--null-snpsets
                   0
```

```
--min-observations 100
```

Repeat the analysis:

snpsea --args args.txt

condition_pvalues.txt

The p-values representing enrichment of condition-specificity for the given SNPs.

head condition_pvalues.txt | column -t

condition	pvalue	nulls_observed	nulls_tested
Colorectal_Adenocarcinoma	0.933555	280	300
Whole_Blood	0.521595	156	300
BM-CD33+Myeloid	0.159772	111	700
PB-CD14+Monocytes	0.103264	154	1500
PB-BDCA4+Dentritic_cells	0.0606256	187	3100
PB-CD56+NK_cells	0.194009	135	700
PB-CD4+T_cells	0.428571	128	300
PB-CD8+T_cells	0.531561	159	300
PB-CD19+B_cells	0.226819	158	700

null_pvalues.txt

If the argument for **--snps** is the name of a file, the p-values for null matched SNP sets. You can compare these null results to the results for your trait-associated SNPs.

If the argument for **--snps** is "randomN" where N is some integer, like "random20" the p-values for random unmatched SNP sets, each with N SNPs.

The fifth column is the replicate index. The number of replicates performed is specified with --null-snpsets INT.

head null_pvalues.txt | column -t

ColorectalAdenocarcinoma	0.056	84	1500	0
WholeBlood	0.236667	71	300	0
BM-CD33+Myeloid	0.55	55	100	0
PB-CD14+Monocytes	0.59	59	100	0
PB-BDCA4+Dentritic_Cells	0.59	59	100	0
PB-CD56+NKCells	0.71	71	100	0
PB-CD4+Tcells	0.383333	115	300	0
PB-CD8+Tcells	0.128571	90	700	0
PB-CD19+Bcells	0.168571	118	700	0
BM-CD105+Endothelial	0.386667	116	300	0

⁻⁻max-iterations 10000000

$snp_genes.txt$

Each SNP's linkage interval and overlapping genes. If a SNP is not found in the reference file specified with **--snp-intervals**, then the name of the SNP will be listed and the other columns will contain NA.

head snp_genes.txt | column -t

chrom	start	end	name	n_{genes}	genes
chr4	55364224	55408999	rs218238	0	NA
chr6	139827777	139844854	rs590856	0	NA
NA	NA	NA	rs99999999	NA	NA
chr6	109505894	109651220	rs1008084	2	8763,27244
chr10	71089843	71131638	rs10159477	1	3098
chr2	111807303	111856057	rs10207392	1	55289
chr16	88831494	88903796	rs10445033	4	353,2588,9780,81620
chr7	151396253	151417368	rs10480300	1	51422
chr12	4320955	4336783	rs10849023	2	894,57103
chr15	76129642	76397903	rs11072566	4	26263,92912,123591,145957

snp_condition_scores.txt

Each SNP, condition, gene with greatest specificity to that condition, and score for the SNP-condition pair, adjusted for the number of genes overlapping the given SNP's linkage interval.

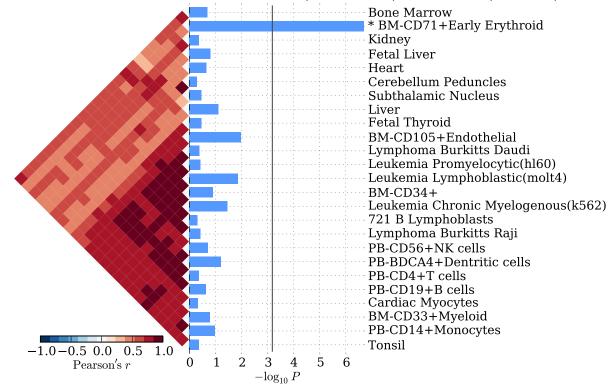
head snp_condition_scores.txt | column -t

snp	condition	gene	score
rs9349204	Colorectal_Adenocarcinoma	10817	0.693027
rs9349204	Whole_Blood	896	0.285864
rs9349204	BM-CD33+Myeloid	896	0.236487
rs9349204	PB-CD14+Monocytes	29964	0.340561
rs9349204	PB-BDCA4+Dentritic_cells	29964	0.411727
rs9349204	PB-CD56+NK_cells	896	0.0356897
rs9349204	PB-CD4+T_cells	896	0.38182
rs9349204	PB-CD8+T_cells	896	0.332008
rs9349204	PB-CD19+B_cells	29964	0.255196

Output Visualizations

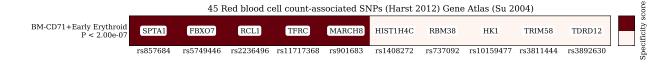
View enrichment of tissue-specific gene expression

45 Red blood cell count-associated SNPs (Harst 2012) Gene Atlas (Su 2004)



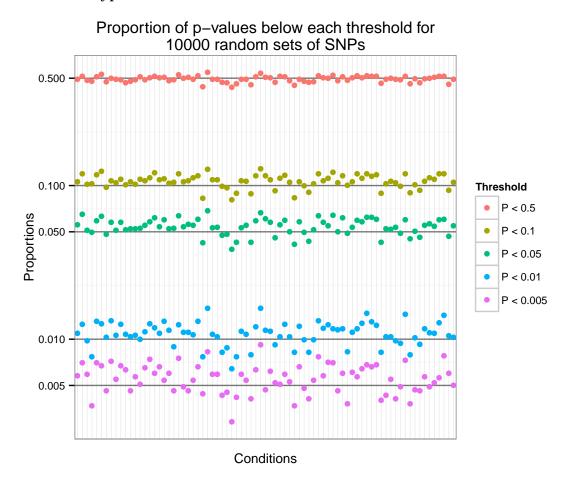
python bin/snpsea-barplot out

View the most specifically expressed gene for each SNP-tissue pair



python bin/snpsea-heatmap out

View the type 1 error rate estimates for each tissue



Rscript bin/snpsea-type1error out