ACMG-ClinVar Penetrance RMarkdown

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Contents

Working Directory: /Users/jamesdiao/Documents/Kohane_Lab/2016-paper-ACMG-penetrance/ACMG_Penetrance

1 Download, Transform, and Load Data

1.1 Collect ACMG Gene Panel

http://www.ncbi.nlm.nih.gov/clinvar/docs/acmg/

```
## Processed Table from ACMG Website 64 x 4 (selected rows):
```

	Disease_Name	Disease_MIM	Gene_Name	Gene_MIM
N1	Adenomatous polyposis coli	175100	APC	611731
N2	Aortic aneurysm, familial thoracic 4	132900	MYH11	160745
N5	Arrhythmogenic right ventricular cardiomyopathy, type 5	604400	TMEM43	612048
N10	Breast-ovarian cancer, familial 1	604370	BRCA1	113705
N11	Breast-ovarian cancer, familial 2	612555	BRCA2	600185
N12	Brugada syndrome 1	601144	SCN5A	600163
N13	Catecholaminergic polymorphic ventricular tachycardia	604772	RYR2	180902
N14	Dilated cardiomyopathy 1A	115200	LMNA	150330
N16	Ehlers-Danlos syndrome, type 4	130050	COL3A1	120180
N17	Fabry's disease	301500	GLA	300644
N18	Familial hypercholesterolemia	143890	APOB	107730
N20	Familial hypertrophic cardiomyopathy 1	192600	MYH7	160760
N28	Familial medullary thyroid carcinoma	155240	RET	164761
N30	Left ventricular noncompaction 6	601494	TNNT2	191045
N31	Li-Fraumeni syndrome 1	151623	TP53	191170
N32	Loeys-Dietz syndrome type 1A	609192	TGFBR1	190181
N37	Long QT syndrome 1	192500	KCNQ1	607542
N40	Lynch syndrome	120435	MLH1	120436
N44	Malignant hyperthermia	145600	RYR1	180901
N46	Marfan's syndrome	154700	FBN1	134797
N48	Multiple endocrine neoplasia, type 1	131100	MEN1	613733
N51	MYH-associated polyposis	608456	MUTYH	604933
N52	Neurofibromatosis, type 2	101000	NF2	607379
N53	Paragangliomas 1	168000	SDHD	602690
N57	Peutz-Jeghers syndrome	175200	STK11	602216
N58	Pilomatrixoma	132600	MUTYH	604933
N59	PTEN hamartoma tumor syndrome	153480	PTEN	601728
N60	Retinoblastoma	180200	RB1	614041
N61	Tuberous sclerosis 1	191100	TSC1	605284
N63	Von Hippel-Lindau syndrome	193300	VHL	608537
N64	Wilms' tumor	194070	WT1	607102

```
## ACMG-56 Genes:

##  [1] APC      MYH11    ACTA2    MYLK     TMEM43   DSP      PKP2     DSG2
##  [9] DSC2     BRCA1    BRCA2    SCN5A    RYR2     LMNA     MYBPC3   COL3A1
## [17] GLA      APOB     LDLR     MYH7     TPM1     PRKAG2   TNNI3    MYL3
## [25] MYL2     ACTC1    RET      PCSK9    TNNT2    TP53     TGFBR1   TGFBR2
## [33] SMAD3    KCNQ1    KCNH2    MLH1     MSH2     MSH6     PMS2     RYR1
## [41] CACNA1S  FBN1     MEN1     MUTYH    NF2      SDHD     SDHAF2   SDHC
## [49] SDHB     STK11    PTEN     RB1      TSC1     TSC2     VHL      WT1
```

1.2 Download ClinVar VCF

ftp://ftp.ncbi.nlm.nih.gov/pub/clinvar/vcf_GRCh37/clinvar.vcf.gz
ClinVar is the central repository for variant interpretations. Relevant information from the VCF includes:
(a) CLNSIG = "Variant Clinical Significance, 0 - Uncertain, 1 - Not provided, 2 - Benign, 3 - Likely benign, 4 - Likely pathogenic, 5 - Pathogenic, 6 - Drug response, 7 - Histocompatibility, 255 - Other"
(b) CLNDBN = "Variant disease name"
(c) CLNDSDBID = "Variant disease database ID"
(d) INTERP = Pathogenicity (likely pathogenic or pathogenic; CLNSIG = 4 or 5)

```
## Processed ClinVar data frame 126349 x 14 (selected rows/columns):
```

VAR_ID	CHROM	POS	ID	REF	ALT	CLNSIG
1_949523_C_T	1	949523	rs786201005	C	T	5
1_949739_G_T	1	949739	rs672601312	G	T	5
1_955597_G_T	1	955597	rs115173026	G	T	2
1_955619_G_C	1	955619	rs201073369	G	C	255
1_957568_A_G	1	957568	rs115704555	A	G	2
1_957605_G_A	1	957605	rs756623659	G	A	5

Table continues below

CLNDBN	CLNDSDBID	INTERP
Immunodeficiency_38_with_basal_ganglia_calcification	CN221808:616126	TRUE
Immunodeficiency_38_with_basal_ganglia_calcification	CN221808:616126	TRUE
not_specified	CN169374	FALSE
not_specified	CN169374	FALSE
not_specified	CN169374	FALSE
Congenital_myasthenic_syndrome	C0751882:ORPHA590	TRUE

1.3 Download 1000 Genomes VCFs

ftp://ftp.1000genomes.ebi.ac.uk/vol1/ftp/release/20130502/ALL.[chrom].phase3_[version].20130502.genotypes.vcf.gz
Downloaded 1000 Genomes VCFs are saved in: /Users/jamesdiao/Documents/Kohane_Lab/2016-paper-ACMG-penetrance/1000G/

```
## Download report: region and successes: 56 x 6 (selected rows):
```

gene	name	chrom	start	end	downloaded
APC	NM_001127511	5	112043201	112181936	TRUE
MYH11	NM_001040113	16	15796991	15950887	TRUE
ACTA2	NM_001141945	10	90694830	90751154	TRUE
MYLK	NM_001321309	3	123331142	123603149	TRUE
TMEM43	NM_024334	3	14166439	14185180	TRUE

```
## File saved as download_output.txt in Supplementary_Files
```

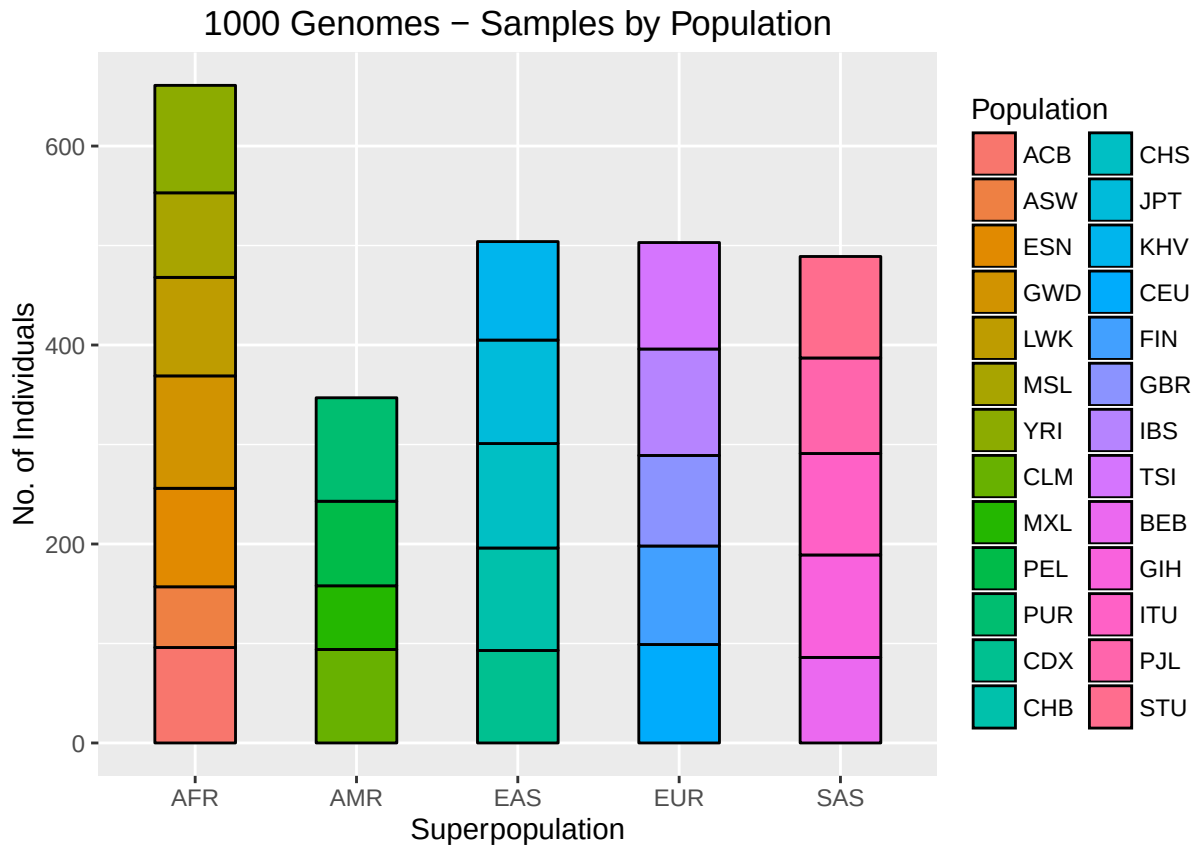
1.4 Collect 1000 Genomes Phase 3 Populations Map

This will allow us to assign genotypes from the 1000 Genomes VCF to ancestral groups.
From: ftp://ftp.1000genomes.ebi.ac.uk/vol1/ftp/release/20130502/integrated_call_samples_v3.20130502.ALL.panel

```
## Phase 3 Populations Map Table: 2504 x 4 (selected rows)
```

sample	pop	super_pop	gender
HG02881	GWD	AFR	male
NA18489	YRI	AFR	female
HG02623	GWD	AFR	male
HG02131	KHV	EAS	male
NA20832	TSI	EUR	female
NA20778	TSI	EUR	male
NA20810	TSI	EUR	male
NA12761	CEU	EUR	female
HG00339	FIN	EUR	female
HG03999	STU	SAS	male

```
## Population Distribution
```



1.5 Import and Process 1000 Genomes VCFs

(a) Unnest the data frames to 1 row per variant_ID key (CHROM_POSITION_REF_ALT).
(b) Remove all insertions, deletions, CNV, etc, and keep only missense variants (1 REF, 1 ALT)
(c) For 1000 Genomes: convert genomes to allele counts. For example: (0|1) becomes 1, (1|1) becomes 2. Multiple alleles are unnested into multiple counts. For example: (0|2) becomes 0 for the first allele (no 1s) and 1 for the second allele (one 2).

```
## Processed 1000 Genomes VCFs: 139335 x 2516 (selected rows/columns):
```

GENE	AF_1000G	VAR_ID	CHROM	POS	ID	REF	ALT
APC	0.000199681	5_112043211_A_G	5	112043211	rs554351451	A	G
APC	0.000199681	5_112043231_G_A	5	112043231	rs575784409	G	A
APC	0.005391370	5_112043234_C_T	5	112043234	rs115658307	C	T
APC	0.000199681	5_112043252_G_A	5	112043252	rs558562104	G	A
APC	0.008785940	5_112043263_C_T	5	112043263	rs138386816	C	T

Table continues below

HG00096	HG00097	HG00099	HG00100	HG00101	HG00102
0	0	0	0	0	0
0	0	0	0	0	0
0	0	0	0	0	0
0	0	0	0	0	0
0	0	0	0	0	0

1.6 Import and Process ExAC VCFs

(a) Unnest the data frames to 1 row per variant_ID key (CHROM_POSITION_REF_ALT).
(b) Remove all insertions, deletions, CNV, etc, and keep only missense variants (1 REF, 1 ALT)
(c) Collect superpopulation-level allele frequencies: African = AFR, Latino = AMR, European (Finnish + Non-Finnish) = EUR, East.Asian = EAS, South.Asian = SAS.

```
## Processed ExAC VCFs: 58873 x 45 (selected rows/columns):
```

GENE	AF_EXAC	AF_EXAC_AFR	AF_EXAC_AMR	AF_EXAC_EAS	AF_EXAC_EUR
APC	0.00008130	0.00000000	0.0000000	0	0.0000000
APC	0.00008131	0.00000000	0.0000000	0	0.0000000
APC	0.11120000	0.07978723	0.1021505	0	0.1063298
APC	0.00008131	0.00000000	0.0000000	0	0.0000000
APC	0.00008134	0.00000000	0.0000000	0	0.0000000

Table continues below

AF_EXAC_SAS	VAR_ID	CHROM	POS	ID	REF	ALT
0.0001313370	5_112043365_G_C	5	112043365	.	G	C
0.0001313025	5_112043382_A_G	5	112043382	.	A	G
0.1184659837	5_112043384_T_G	5	112043384	rs78429131	T	G
0.0001313025	5_112043392_C_T	5	112043392	.	C	T
0.0001313025	5_112043412_C_G	5	112043412	.	C	G

1.7 Merge ClinVar with 1000 Genomes and ExAC

```
## Breakdown of ClinVar Variants
```

Subset_ClinVar	Number_of_Variants
Total ClinVar	126349
LP/P-ClinVar	33033
LP/P-ClinVar & ACMG	6252
LP/P-ClinVar & ACMG & ExAC	826
LP/P-ClinVar & ACMG & 1000 Genomes	122

```
## Breakdown of ACMG-1000 Genomes Variants
```

Subset_1000_Genomes	Number_of_Variants
Total 1000_Genomes & ACMG	139335
1000_Genomes & ACMG & ClinVar	4891
1000_Genomes & ACMG & LP/P-ClinVar	122

```
## Breakdown of ACMG-ExAC Variants
```

Subset_ExAC	Number_of_Variants
Total ExAC & ACMG	58873
ExAC & ACMG & ClinVar	10043
ExAC & ACMG & LP/P-ClinVar	826

1.8 Comparison with ClinVar Browser Query Results

`clinvar_query.txt` contains all results matched by the search query: "(APC[GENE] OR MYH11[GENE]... OR WT1[GENE]) AND (clinsig_pathogenic[prop] OR clinsig_likely_pathogenic[prop])" from the ClinVar website. The exact query is saved in /Supplementary_Files/query_input.txt
This presents another way of collecting data from ClinVar.

Intermediate step: convert hg38 locations to hg19 using the Batch Coordinate Conversion tool (liftOver) from UCSC Genome Browser Utilities.

```
## ClinVar Query Results Table (substitutions only): 6714 x 13 (selected rows/columns)
```

VAR_ID	Gene(s)	Condition(s)	Frequency
X_100652891_C_G	GLA	Fabry disease	GMAF:0.00050(G)
11_47374186_C_G	MYBPC3	Primary familial hypertrophic cardiomyopathy	GMAF:0.00020(G)
11_47355233_C_G	MYBPC3	Familial hypertrophic cardiomyopathy 4	GMAF:0.00020(G)
11_47364162_C_G	MYBPC3	Familial hypertrophic cardiomyopathy 4	GMAF:0.00020(G)
14_23886482_G_C	MYH7	not specified	GMAF:0.00020(C)
14_23893148_C_G	MYH7	Primary dilated cardiomyopathy	GO-ESP:0.00046(G)
1_17355075_A_T	SDHB	Gastrointestinal stromal tumor	GMAF:0.00120(T)
1_17380507_G_C	SDHB	Cowden syndrome 2	GO-ESP:0.01323(C)

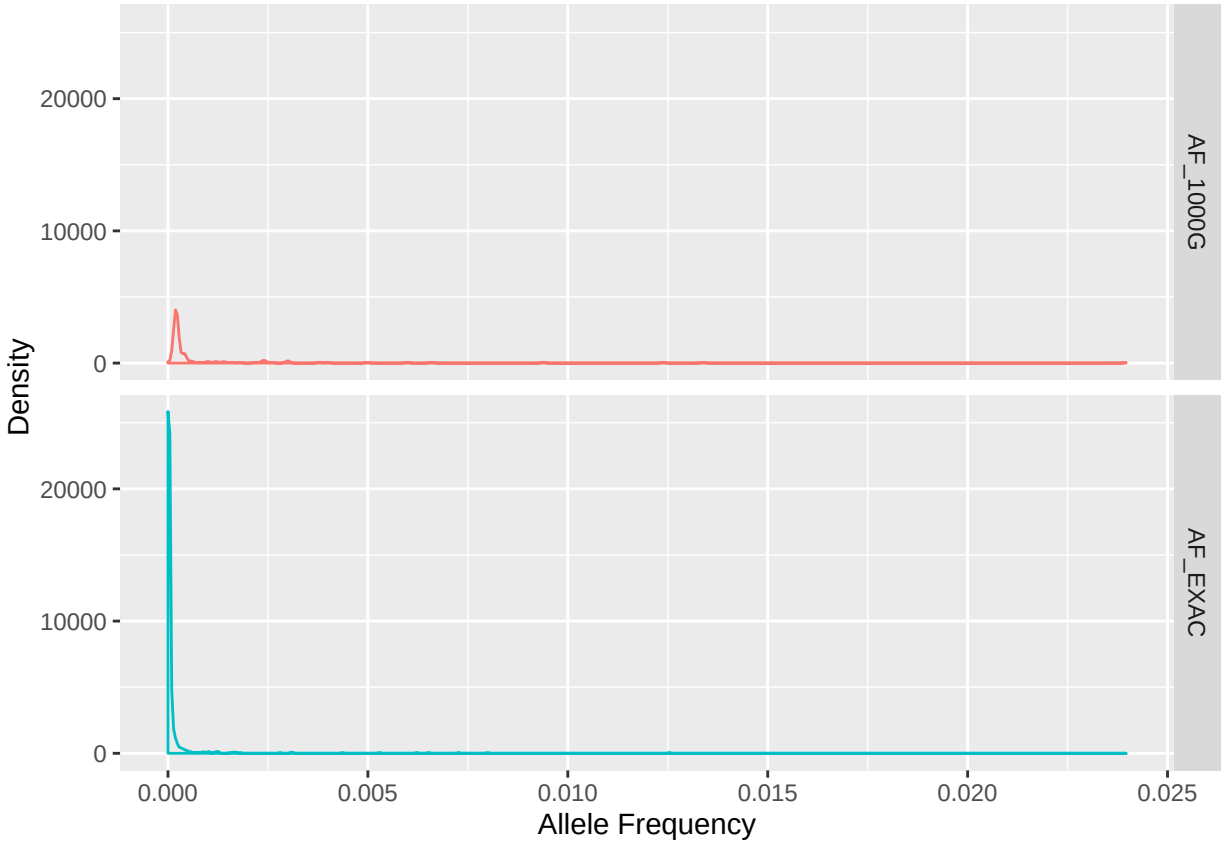
```
## Breakdown of ClinVar Query Results Table:
```

Subset	Number_of_Variants
Initial Count	12525
Filter Substitutions (N>N')	6732
Filter Coupling/Bad-Locations	6714
In ClinVar VCF	509
In LP/P-ClinVar VCF	503
^ & ACMG & ExAC	49
^ & ACMG & 1000 Genomes	9
^ & ACMG & ExAC & 1000 Genomes	8

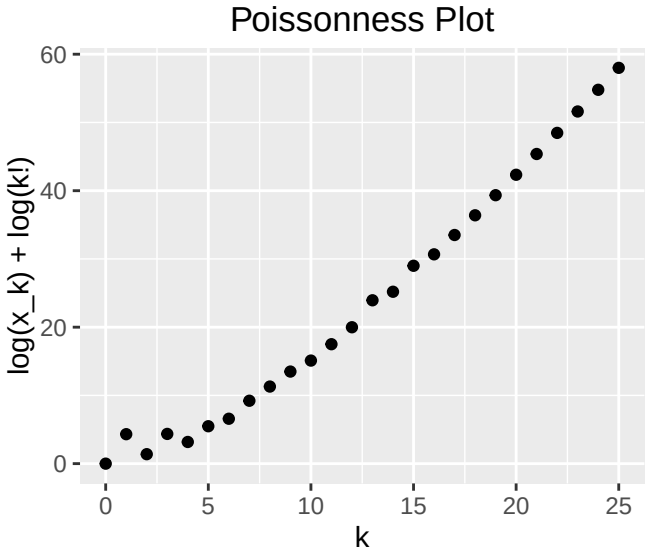
```
## Note the 12-fold reduction after merging the online query results with the VCF.
```

2 Plot Summary Statistics Across Populations

2.1 Distribution of Allele Frequencies



The distribution of allele frequencies is approximately Poisson, with "Poissonness plot" correlation = 0.99. The Poissonness plot (Hoaglin 1980) is defined as the plot of $log(x_k) + log(k!)$ vs. k, as shown below:



2.2 Overall Non-Reference Sites

2.2.0.1 For 1000 Genomes

Each individual has n non-reference sites, which can be found by counting. The mean number is computed for each population.

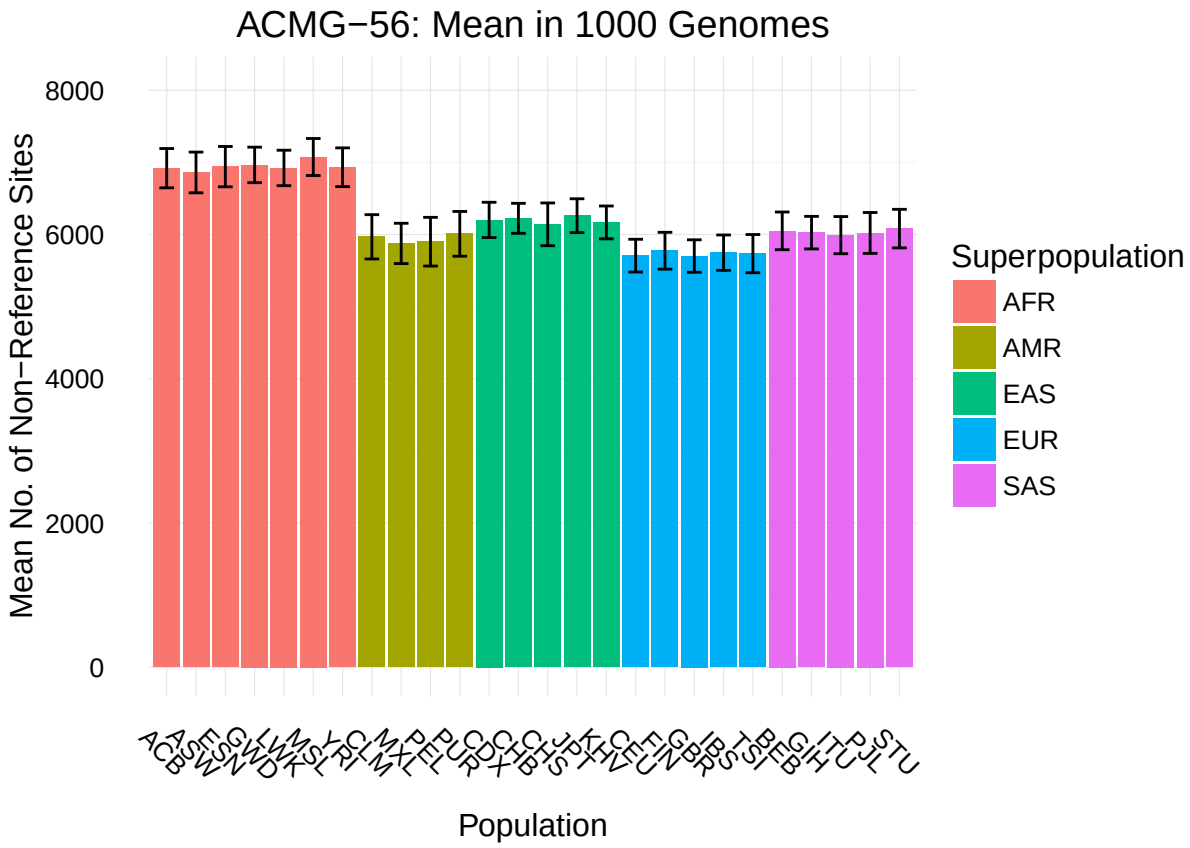
Ex: the genotype of 3 variants in 3 people looks like this:

	HG00097	HG00099	HG00100
Variant 1	0	2	1
Variant 2	0	0	1
Variant 3	0	0	1

Count the number of non-reference sites per individual:

HG00097	HG00099	HG00100
0	1	3

```
## Mean = 1.33
```



Note: the error bars denote standard deviation, not standard error.

2.2.0.2 For ExAC

The mean number of non-reference sites is $E(V)$, where $V = \sum_{i=1}^{n} v_i$ is the number of non-reference sites at all variant positions v_1 through v_n.

At each variant site, the probability of having at least 1 non-reference allele is $P(v_i) = P(v_{i,a} \cup v_{i,b})$, where a and b indicate the 1st and 2nd allele at each site.

If the two alleles are independent, $P(v_{i,a} \cup v_{i,b}) = 1 - (1 - P(v_{i,a}))(1 - P(v_{i,b})) = 1 - (1 - AF(v_i))^2$

If all variants are independent, $E(V) = \sum_{i=1}^{n} 1 - (1 - AF(v_i))^2$ for any set of allele frequencies.

Ex: the allele frequencies of 3 variants across the 5 superpopulations looks like this:

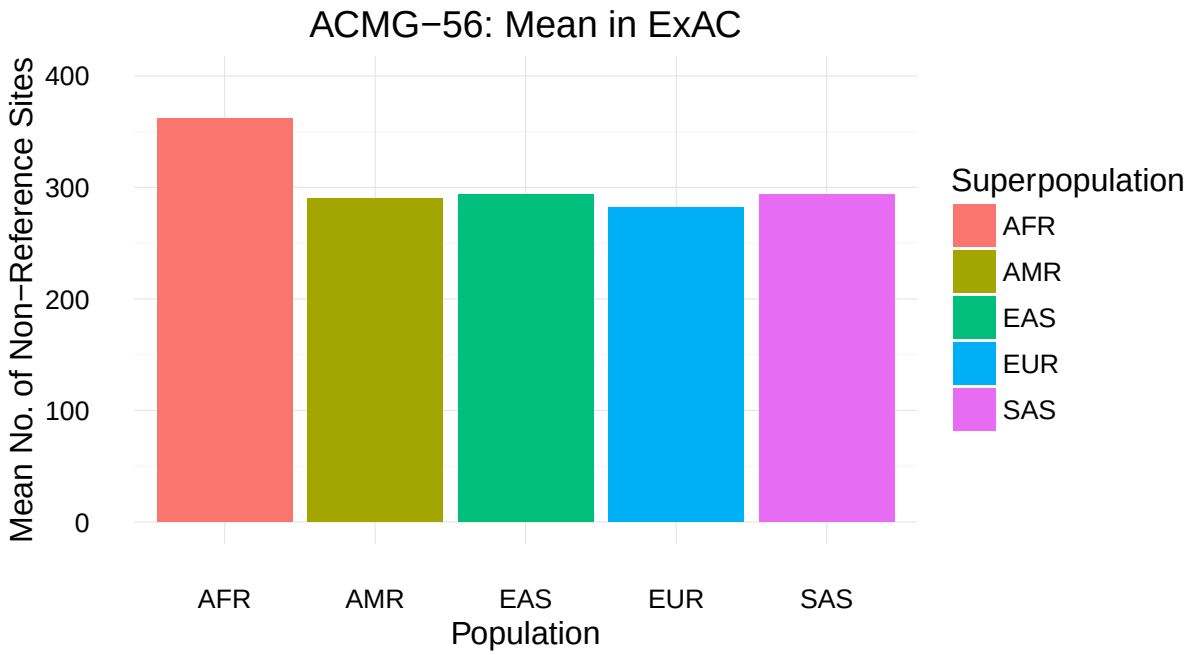
	AFR	AMR	EAS	EUR	SAS
Variant 1	0.1	0.2	0	0	0.3
Variant 2	0.2	0	0.3	0	0.1

The probability of having at least 1 non-reference site at each variant - (0|1) (1|0) or (1|1) is given by $1 - (1 - AF)^2$. Note that this is approximately $2 * AF$ when AF is small:

	AFR	AMR	EAS	EUR	SAS
Variant 1	0.19	0.36	0	0	0.51
Variant 2	0.36	0	0.51	0	0.19

By linearity of expectation, the expected (mean) number of non-reference sites is $\sum E(V_i) = \sum(columns)$.

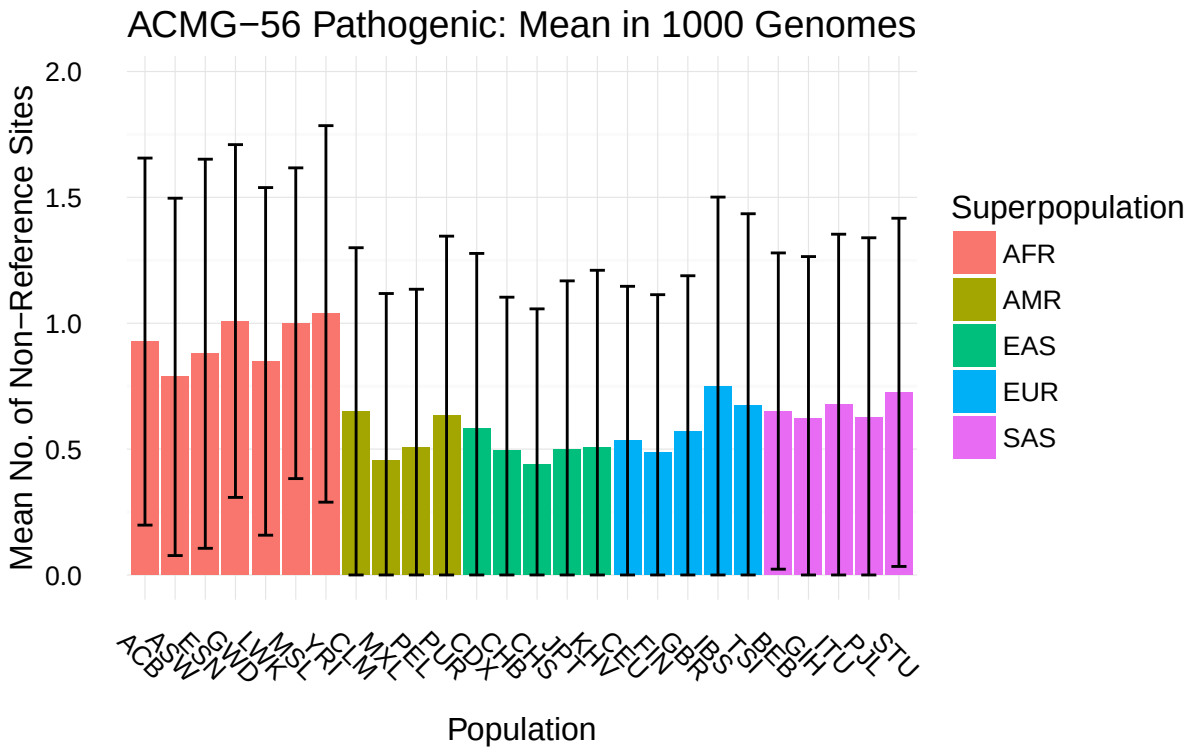
AFR	AMR	EAS	EUR	SAS
0.55	0.36	0.51	0	0.7

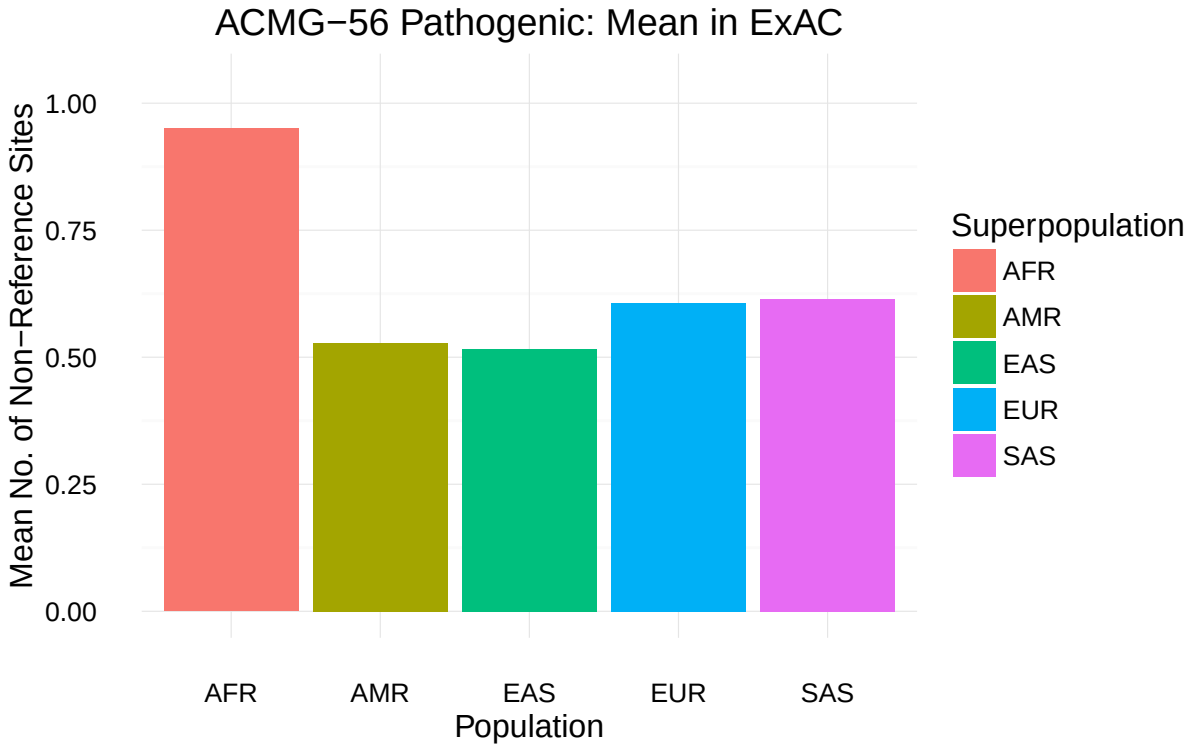


2.3 Pathogenic Non-Reference Sites

2.3.0.1 For 1000 Genomes and ExAC

This is the same procedure as above, but performed only on the subset of variants that are pathogenic.





2.4 Fraction of Individuals with Pathogenic Sites

2.4.0.1 For 1000 Genomes

We can count up the fraction of individuals with 1+ non-reference site(s) in each population. This is the fraction of individuals who would receive a positive genetic test result in at least 1 of the ACMG-56 genes.

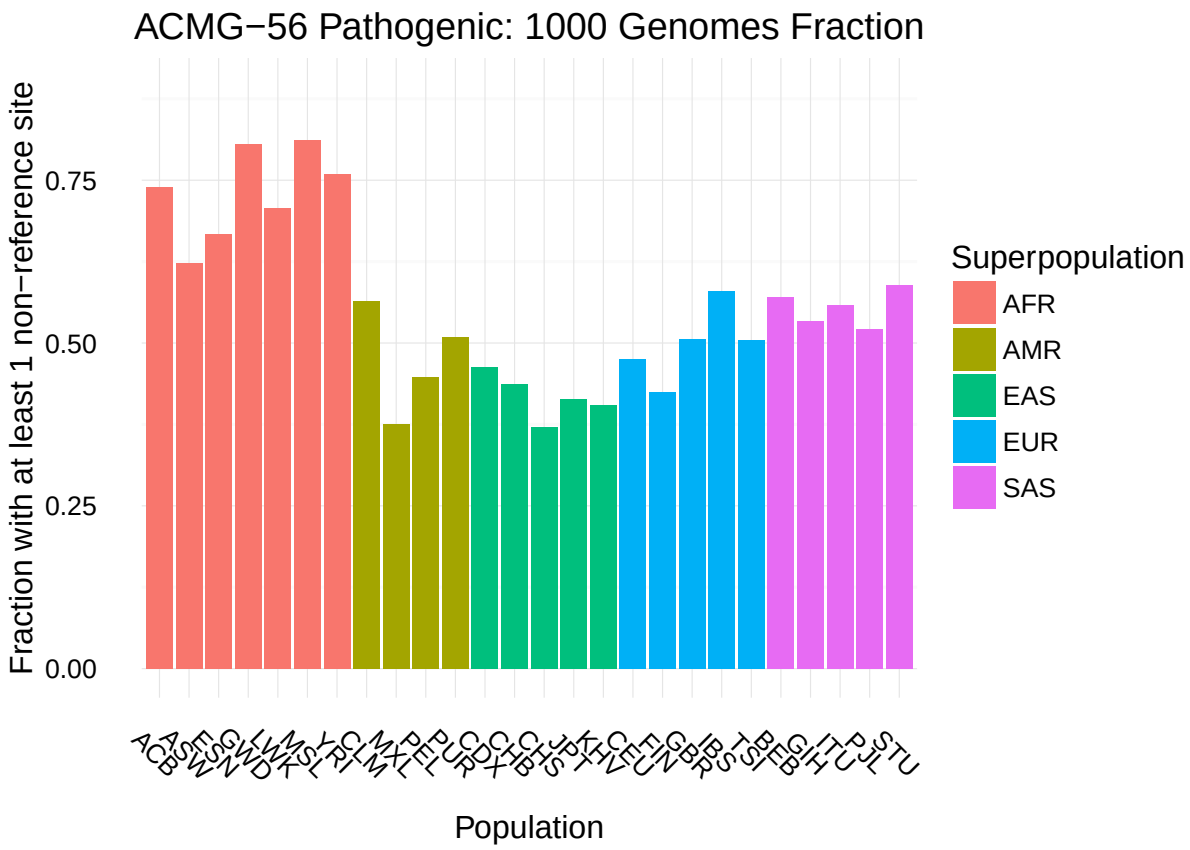
Ex: the genotype of 3 variants in 3 people looks like this:

	HG00097	HG00099	HG00100
Variant 1	0	2	1
Variant 2	0	0	1
Variant 3	0	0	1

Count each individual as having a non-reference site (1) or having only reference sites (0):

HG00097	HG00099	HG00100
0	1	1

```
## Mean = 0.667
```



2.4.0.2 For ExAC

The probability of having at least 1 non-reference site is $P(X)$, where X indicates a non-reference site at any variant position v_1 through v_n.

Recall that $P(v_i) = P(v_{i,a} \cup v_{i,b}) = 1 - (1 - AF(v))^2$ when alleles are independent.

If all alleles are independent, $P(X) = P(\bigcup_{i=1}^{n} v_i) = 1 - \prod_{i=1}^{n}(1 - AF(v_i))^2$

Ex: the allele frequencies of 3 variants across the 5 superpopulations looks like this:

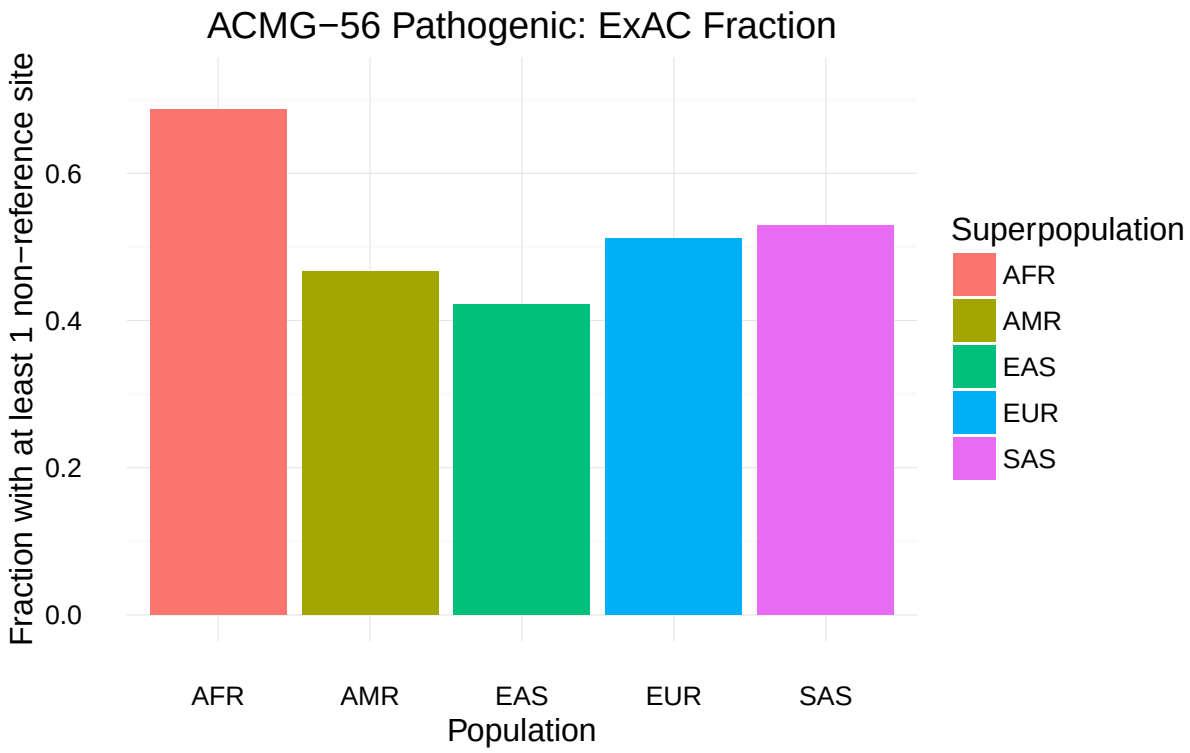
	AFR	AMR	EAS	EUR	SAS
Variant 1	0.1	0.2	0	0	0.3
Variant 2	0.2	0	0.3	0	0.1

The probability of having at least 1 non-reference site at each variant - (0|1) (1|0) or (1|1) is given by $1 - (1 - AF)^2$. Note that this is approximately $2 * AF$ when AF is small:

	AFR	AMR	EAS	EUR	SAS
Variant 1	0.19	0.36	0	0	0.51
Variant 2	0.36	0	0.51	0	0.19

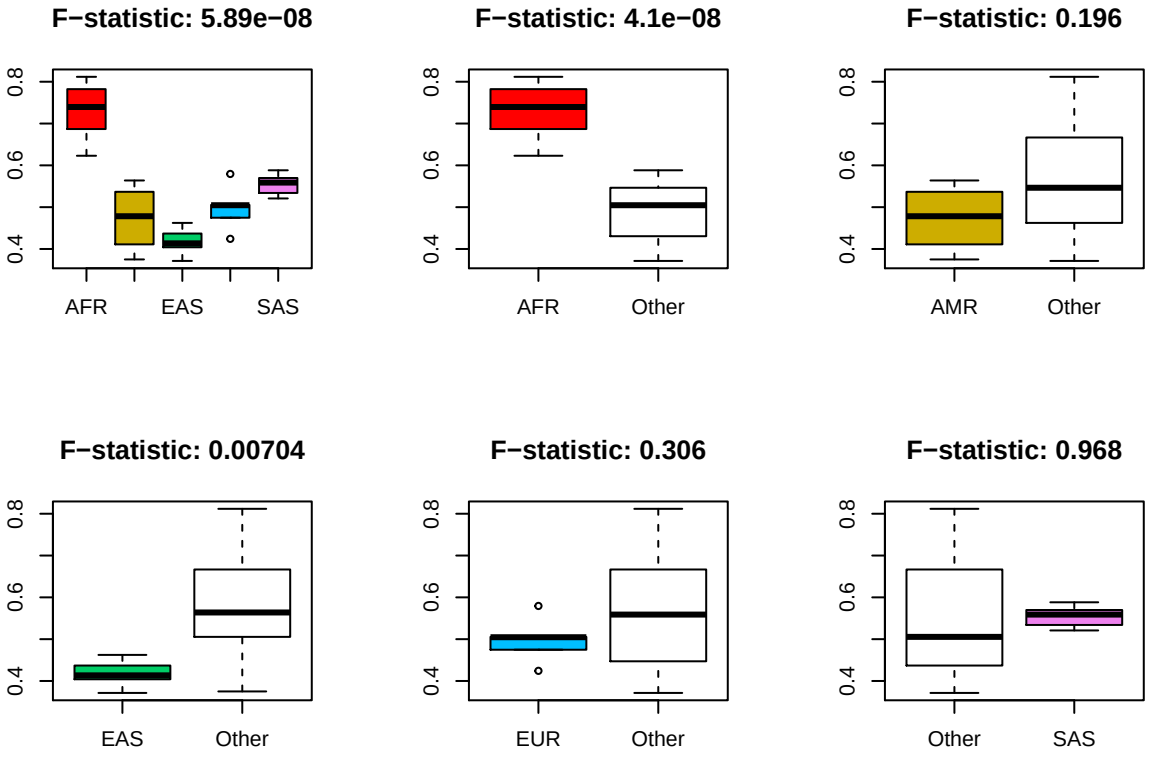
The expected (mean) number of non-reference sites is given by $1 - \prod(1 - AF)^2$.

AFR	AMR	EAS	EUR	SAS
0.4816	0.36	0.51	0	0.6031

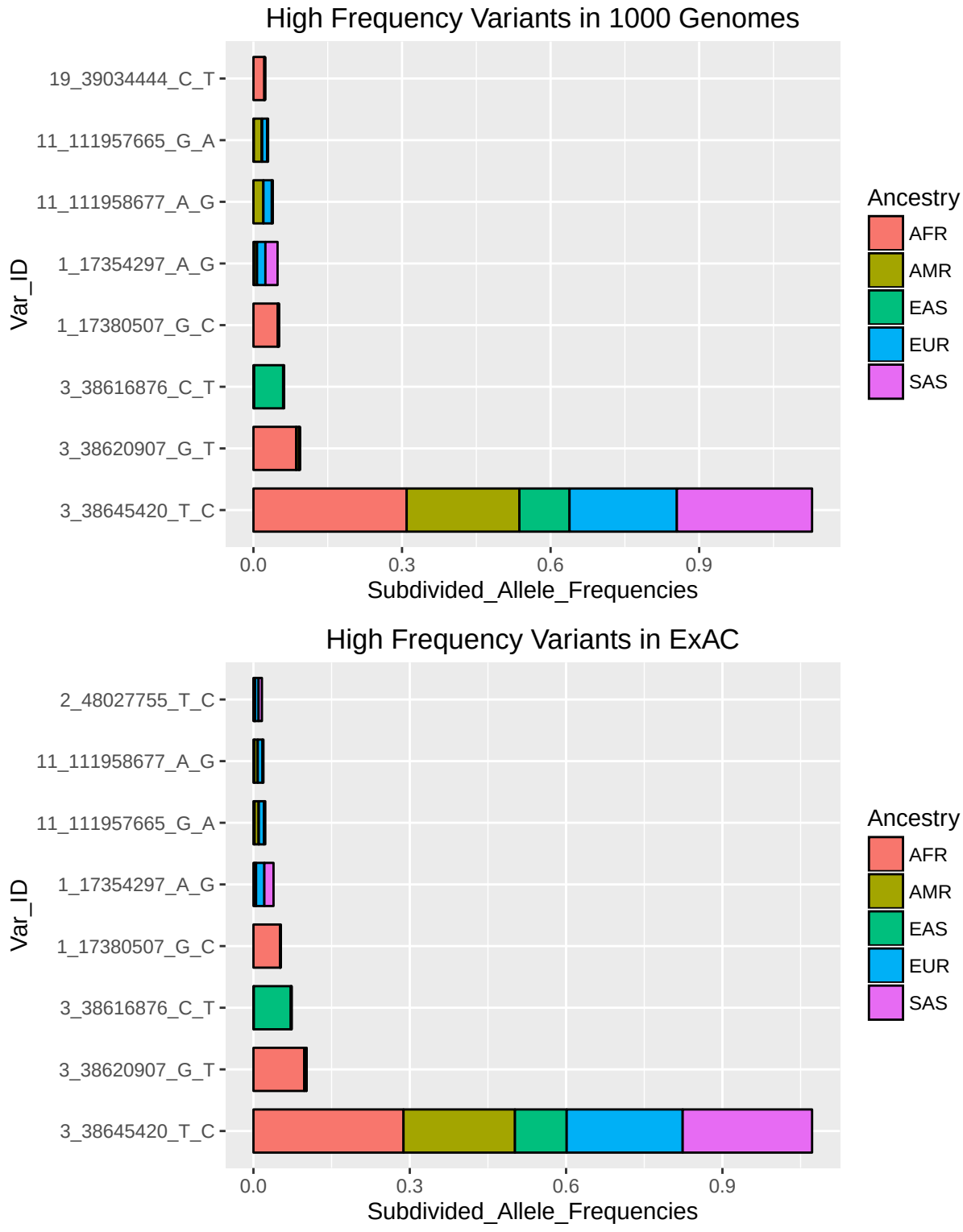


2.5 Test Statistics for Ancestral Differences

F-statistic/T-statistic: probability that the different groups are sampled from distributions with the same mean. These plots are from 4(a) - 1000 Genomes Fraction with 1+ Non-Reference Site, but can be replicated for plots 2(ab) and 3(ab) as well.



2.6 Common Pathogenic Variants by Ancestry



3 Penetrance Estimates

3.1 Bayes' Rule as a Model for Estimating Penetrance

Let V_x be the event that an individual has 1 or more variant related to disease x,
and D_x be the event that the individual is later diagnosed with disease x.

In this case, we can define the following probabilities:
1. Prevalence = $P(D_x)$
2. Allele Frequency = $P(V_x)$
3. Allelic Heterogeneity = $P(V_x|D_x)$
4. Penetrance = $P(D_x|V_x)$

By Bayes' Rule, the penetrance of a variant related to disease x may be defined as:

$$P(D_x|V_x) = \frac{P(D_x) * P(V_x|D_x)}{P(V_x)} = \frac{Prevalence * Allelic.Heteogeneity}{Allele.Frequency}$$

To compute penetrance estimates for each of the diseases related to the ACMG-56 genes, we will use the prevalence data we collected into `Literature_Prevalence_Estimates.csv`, allele frequency data from 1000 Genomes and ExAC, and a broad range of values for allelic heterogeneity.

3.2 Import Literature-Based Disease Prevalence Data

Data Collection: 1. Similar disease subtypes were grouped together (e.g., the 8 different types of familial hypertrophic cardiomyopathy), resulting in 30 disease categories across 56 genes.
2. The search query "[disease name] prevalence" was used to find articles using Google Scholar.
3. Prevalence estimates were recorded along with URL, journal, region, publication year, sample size, first author, population subset (if applicable), date accessed, and potential issues. Preference was given to studies with PubMed IDs, more citations, and larger sample sizes.

Prevalence was recorded as reported: either a point estimate or a range. Values of varying quality were collected across all diseases.

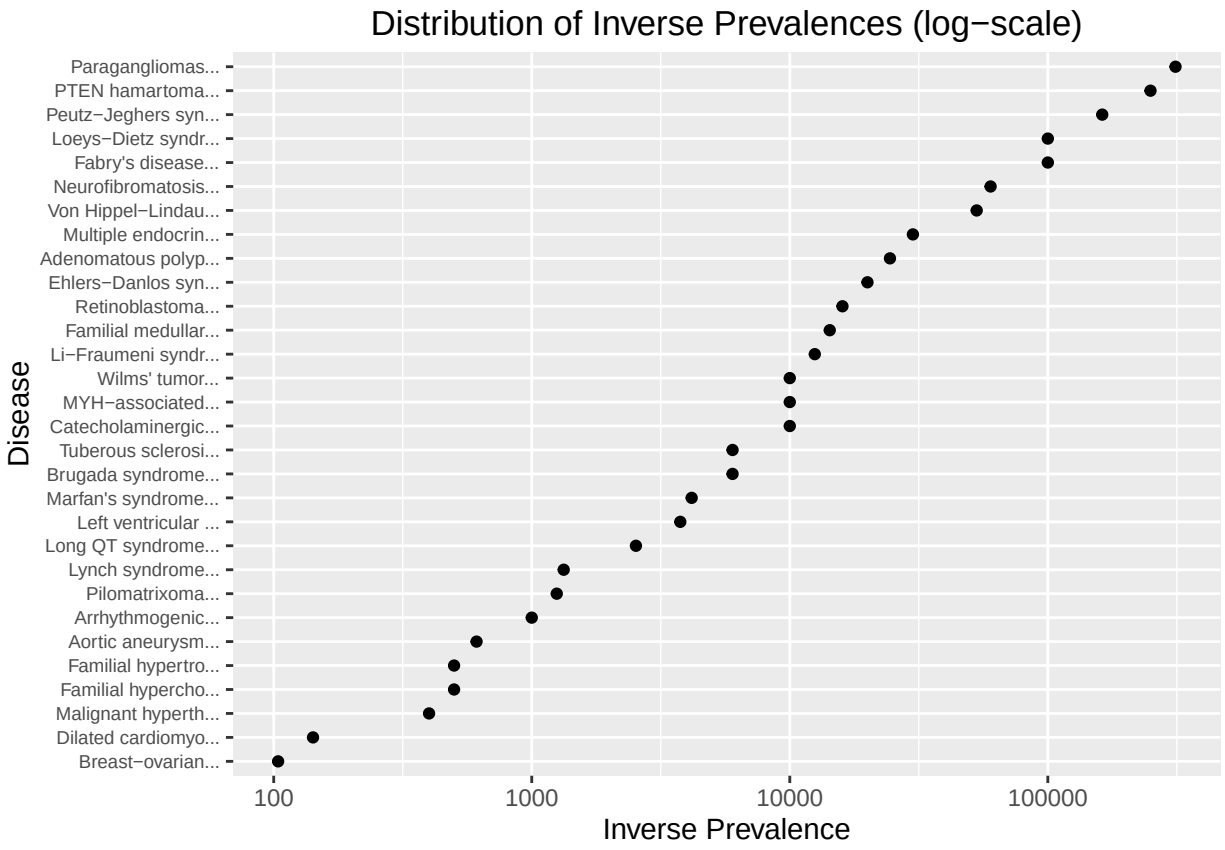
```
## Table of Literature-Based Estimates of Disease Prevalence 30 x 16 (selected rows/columns):
```

Gene	Disease	Disease_MIM	Tags
BRCA1;BRCA2	Breast-ovarian cancer familial	604370;612555	breast;ovarian
SCN5A	Brugada syndrome	601144	brugada
COL3A1	Ehlers-Danlos syndrome	130050	ehler;danlos
TP53	Li-Fraumeni syndrome	151623	fraumeni

Table continues below

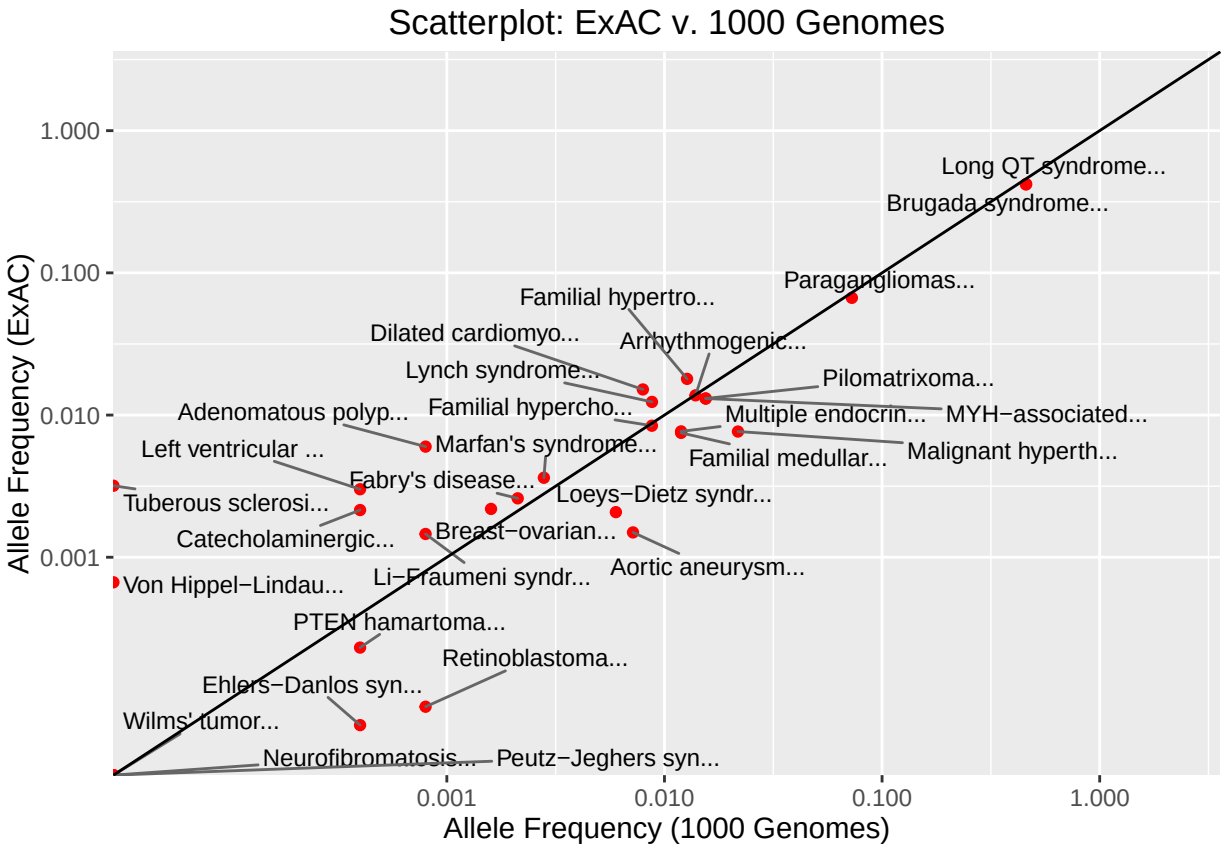
Inverse.Prevalence.1	Inverse.Prevalence.2	year	first.author	citations
104	NA	2013	NA	NA
10000	2000	2006	Antzelevitch	11
20000	NA	2010	Malfait	116
20000	5000	1999	Schneider	47

3.3 Distribution of Prevalences



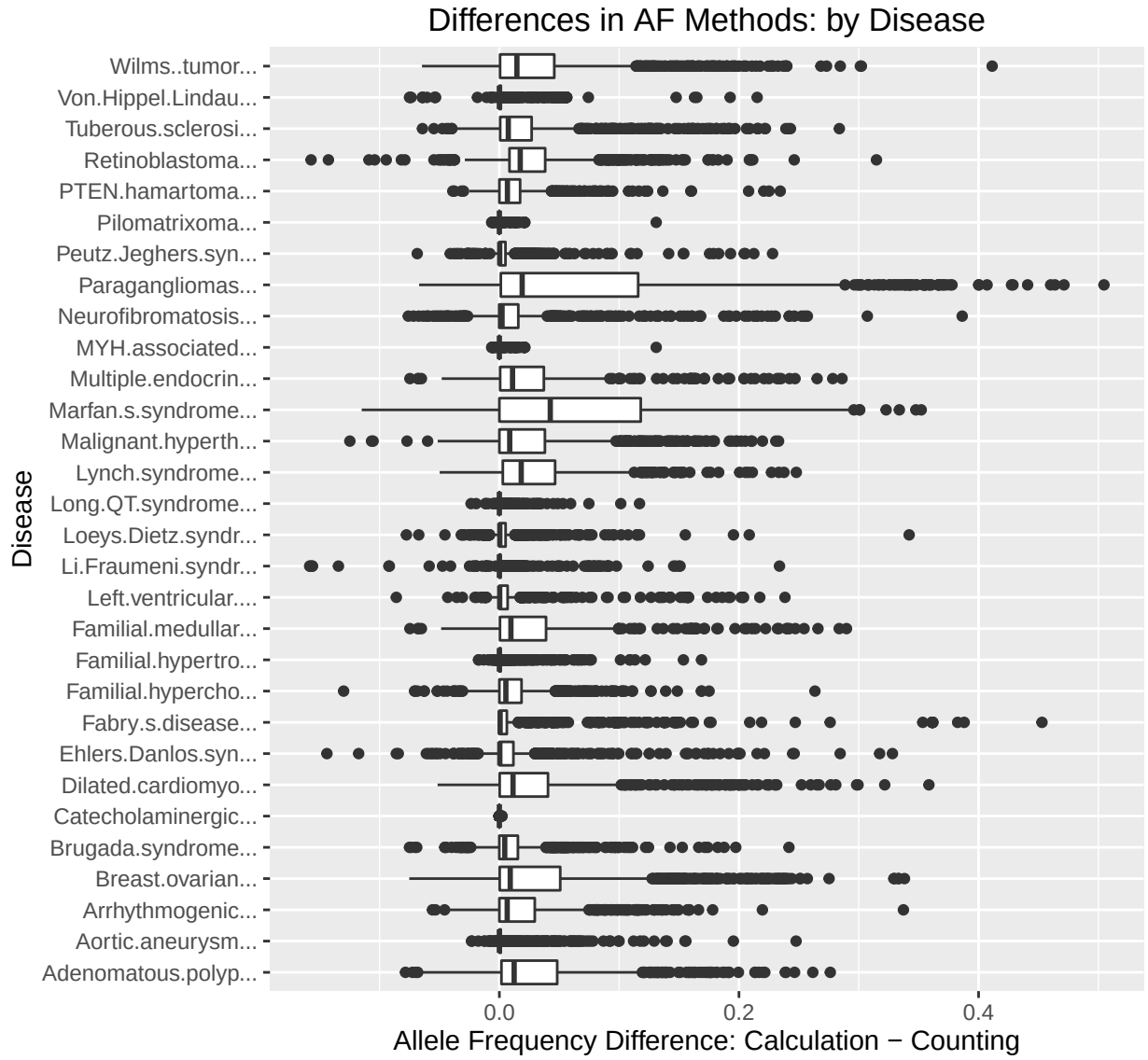
3.4 Collect and Aggregate Allele Frequencies at the Disease-Level

We define AF(disease) as the probability of having at least 1 variant associated with the disease.
The frequencies across the relevant variants can be aggregated in two ways:
(1) By direct counting, from genotype data in 1000 Genomes.
(2) AF(disease) $= 1 - \prod_{variant}(1 - AF_{variant})$, from population data in ExAC (assumes independence).

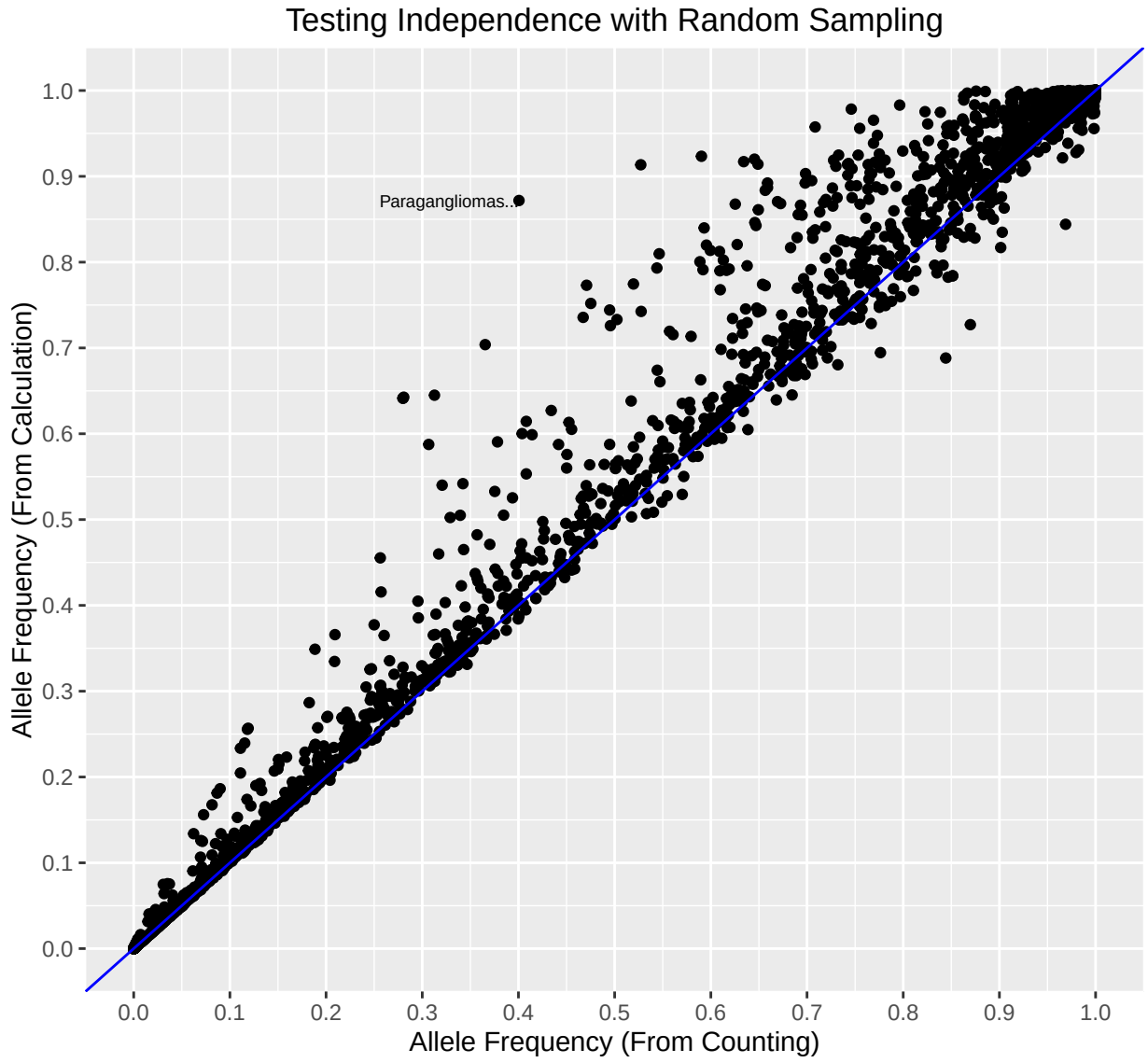


Ratio_1000G (red, top) computes AF(calculation in 1000 Genomes) / AF(counting in 1000 Genomes).
Ratio_ExAC (blue, bottom) computes AF(calculation in ExAC) / AF(counting in 1000 Genomes).

Sampling 1000 variants from all variants in 1000 Genomes to test deviations from independence assumptions. Repeat for 1000 trials and plot the distribution of disease-level allele frequencies (1000 points per disease).



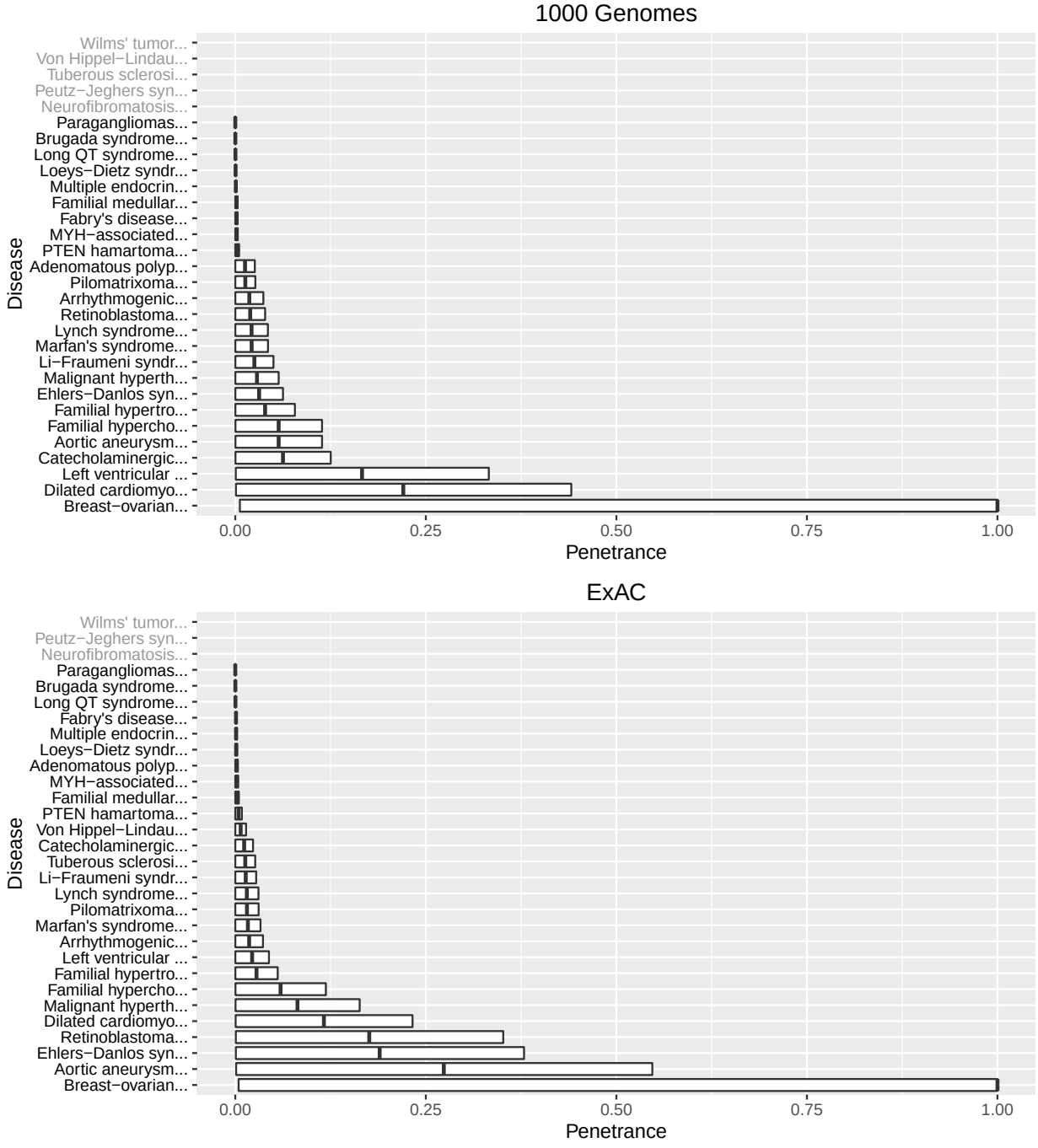

```
## 30 diseases x 1000 points = 30,000 points. This plot has been downsampled 10x and contains 3,000 poi
```



Pearson correlation: 0.99
Mean ratio (Calculation/Counting): 1.07

3.5 Penetrance as a Function of P(V|D)

The left end of the boxplot indicates P(V|D) = 0.001,
the bold line in the middle indicates P(V|D) = 0.25,
the right end of the boxplot indicates P(V|D) = 0.5.

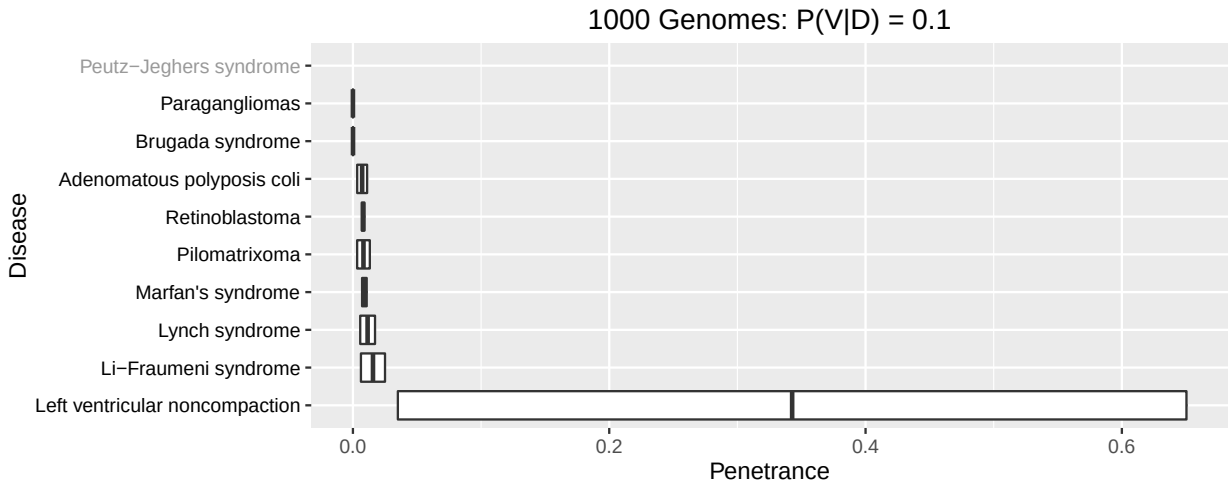


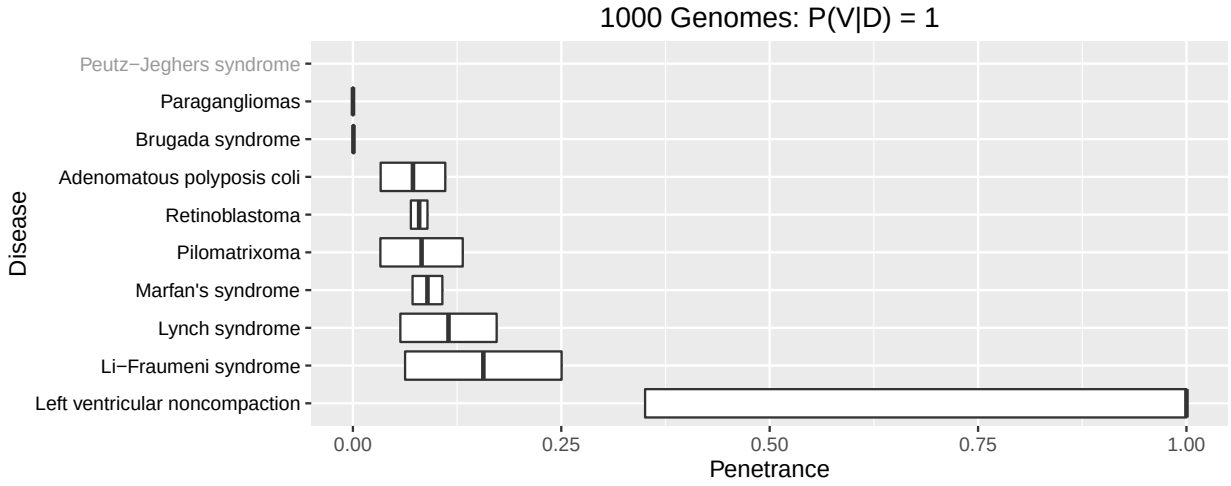
Note 1: the grayed-out empty lines at the top all indicate no allele frequency (disease_AF) data.
Note 2: For breast-ovarian cancer, mean theoretical penetrance > 1. This is because the assumed allelic heterogeneity (0.25) is greater than is possible, given the empirical prevalence and allele frequencies.

3.6 Penetrance as a Function of P(D)

Disease	Prevalence_Ratio
Retinoblastoma	1.3
Marfan's syndrome	1.5
Lynch syndrome	3.0
Adenomatous polyposis coli	3.3
Li-Fraumeni syndrome	4.0
Paragangliomas	4.0
Pilomatrixoma	4.0
Brugada syndrome	5.0
Peutz-Jeghers syndrome	12.0
Left ventricular noncompaction	18.6

The left end of the boxplot indicates P(D) = upper value,
the bold line in the middle indicates P(D) = mean(values),
the right end of the boxplot indicates P(D) = lower value.

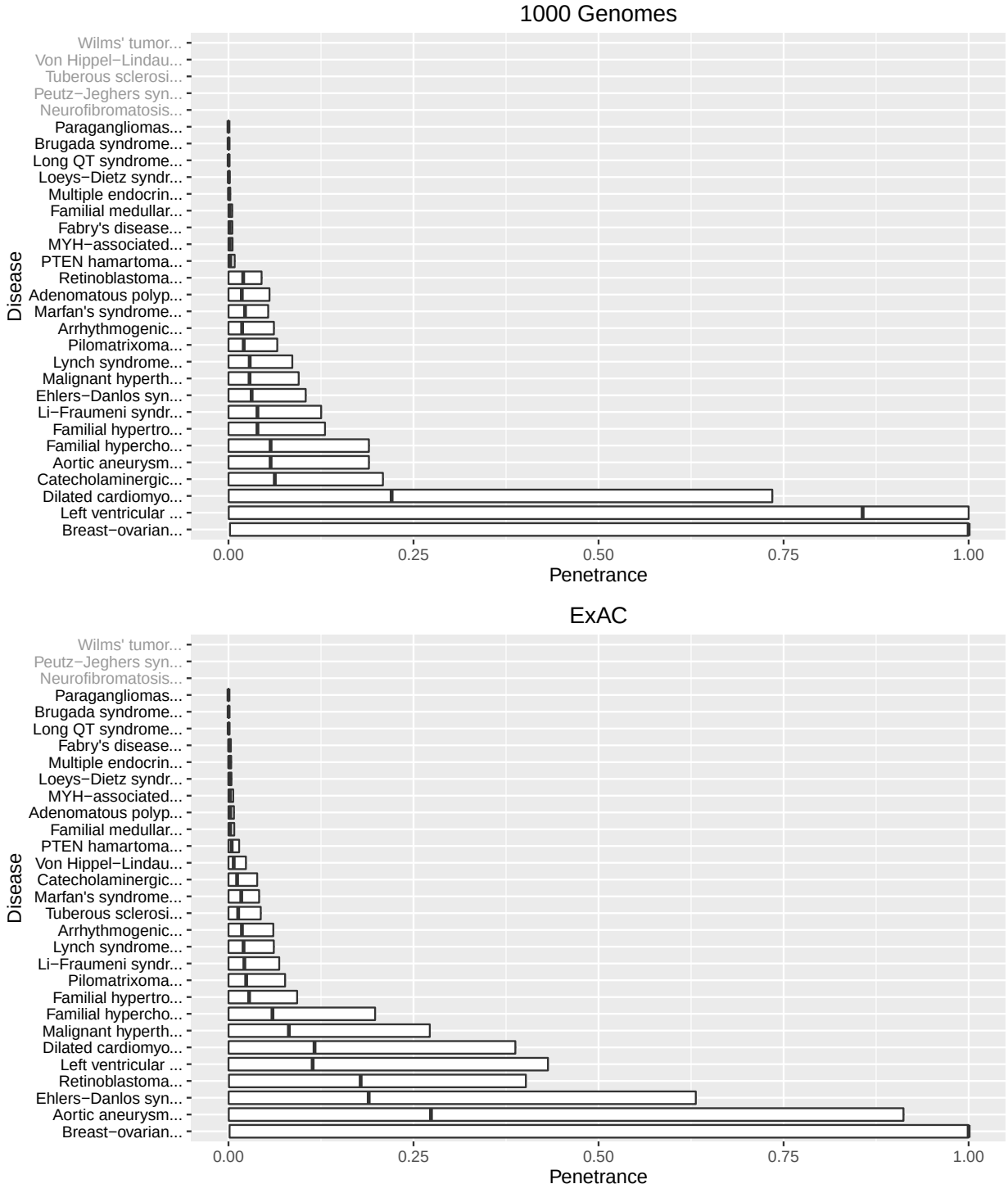




This can only be computed in the 9 cases where a prevalence range was given (rather than a point estimate) and the disease-level allele frequency is known (in this plot: all of them except Puetz-Jeghers).

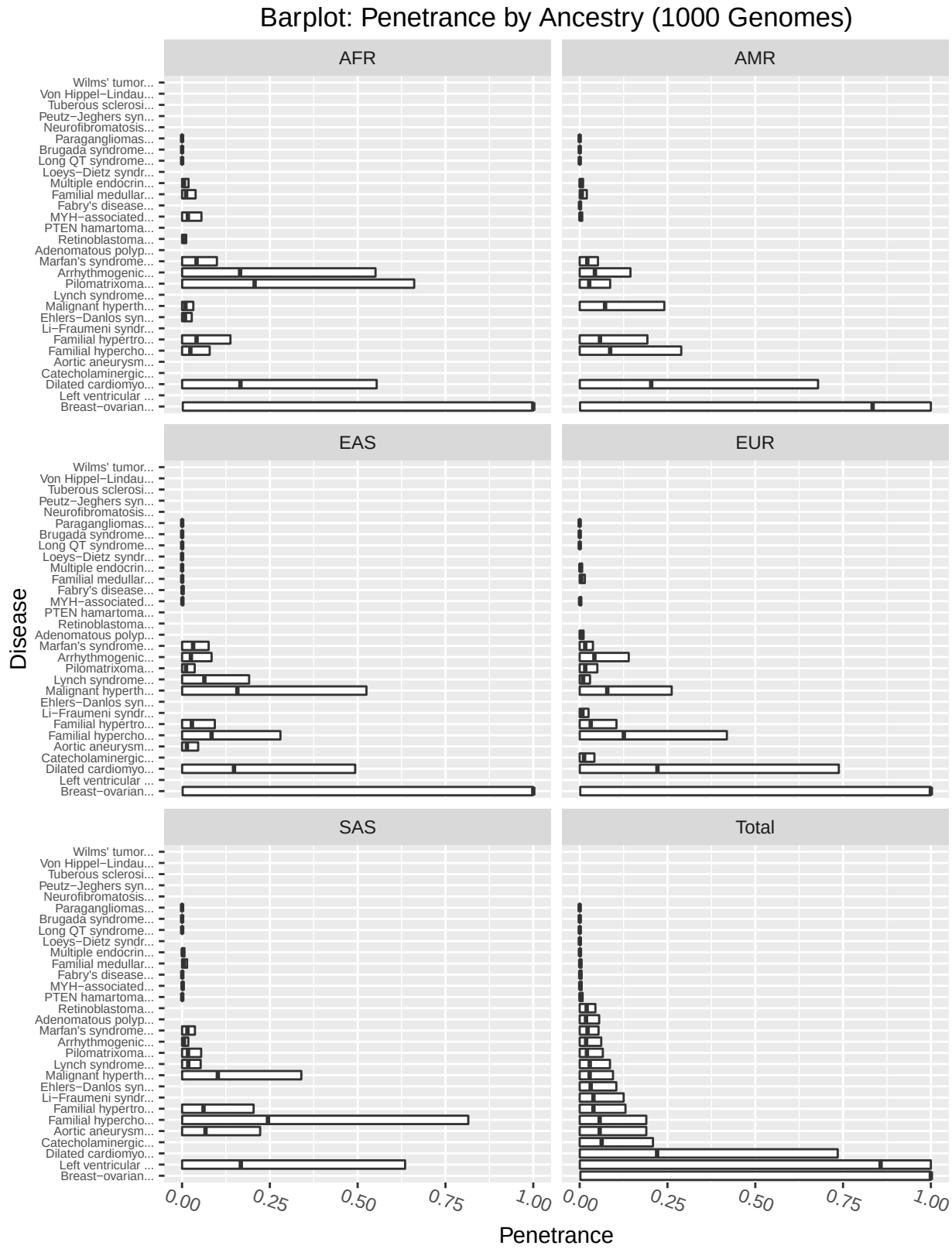
3.7 Max/Min Penetrance as a Function of P(D) and P(V|D)

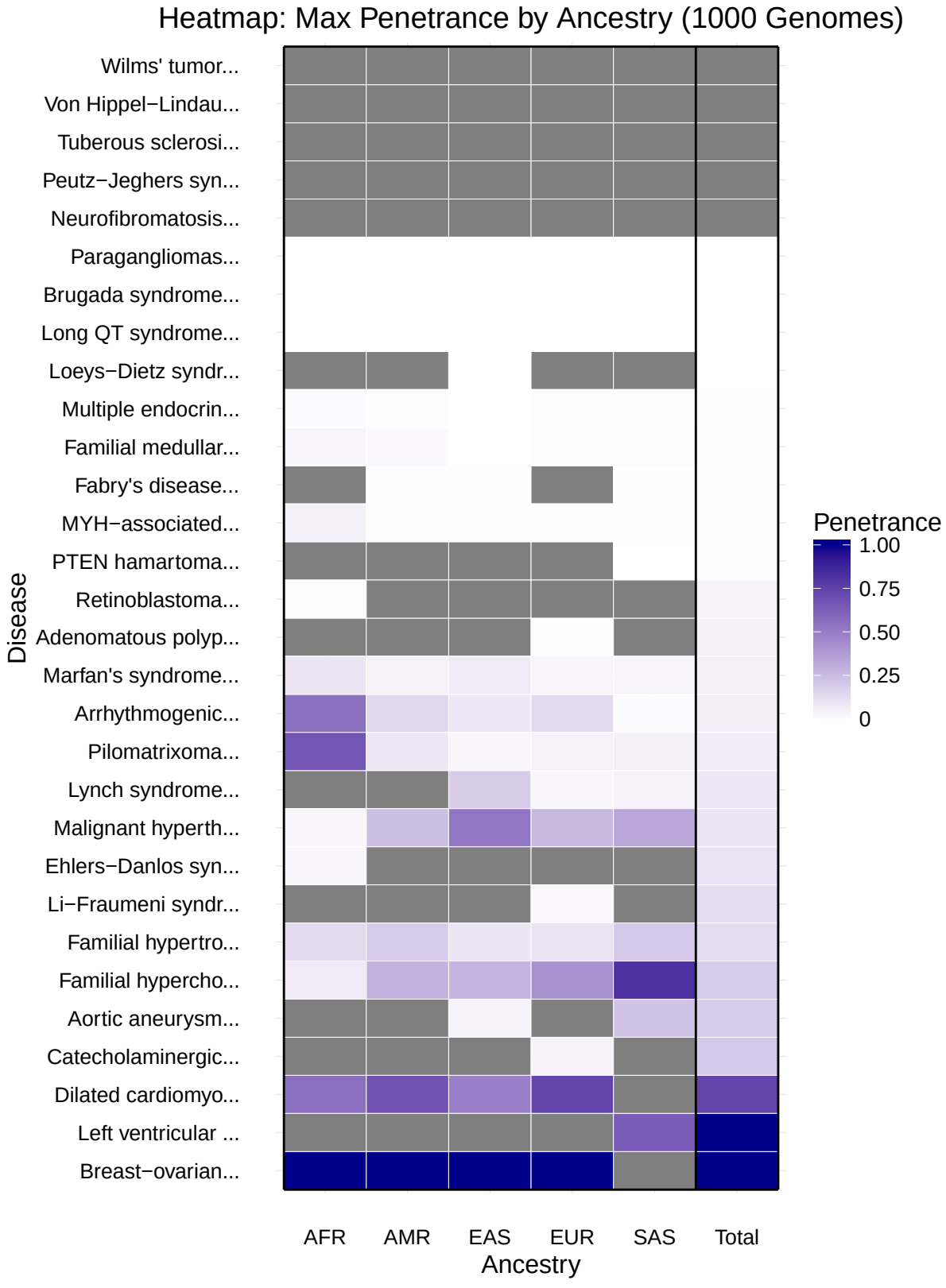
The left end of the boxplot indicates P(D) AND P(V|D) = lower value,
the bold line in the middle indicates P(D) AND P(V|D) = mean(values),
the right end of the boxplot indicates P(D) AND P(V|D) = upper value.



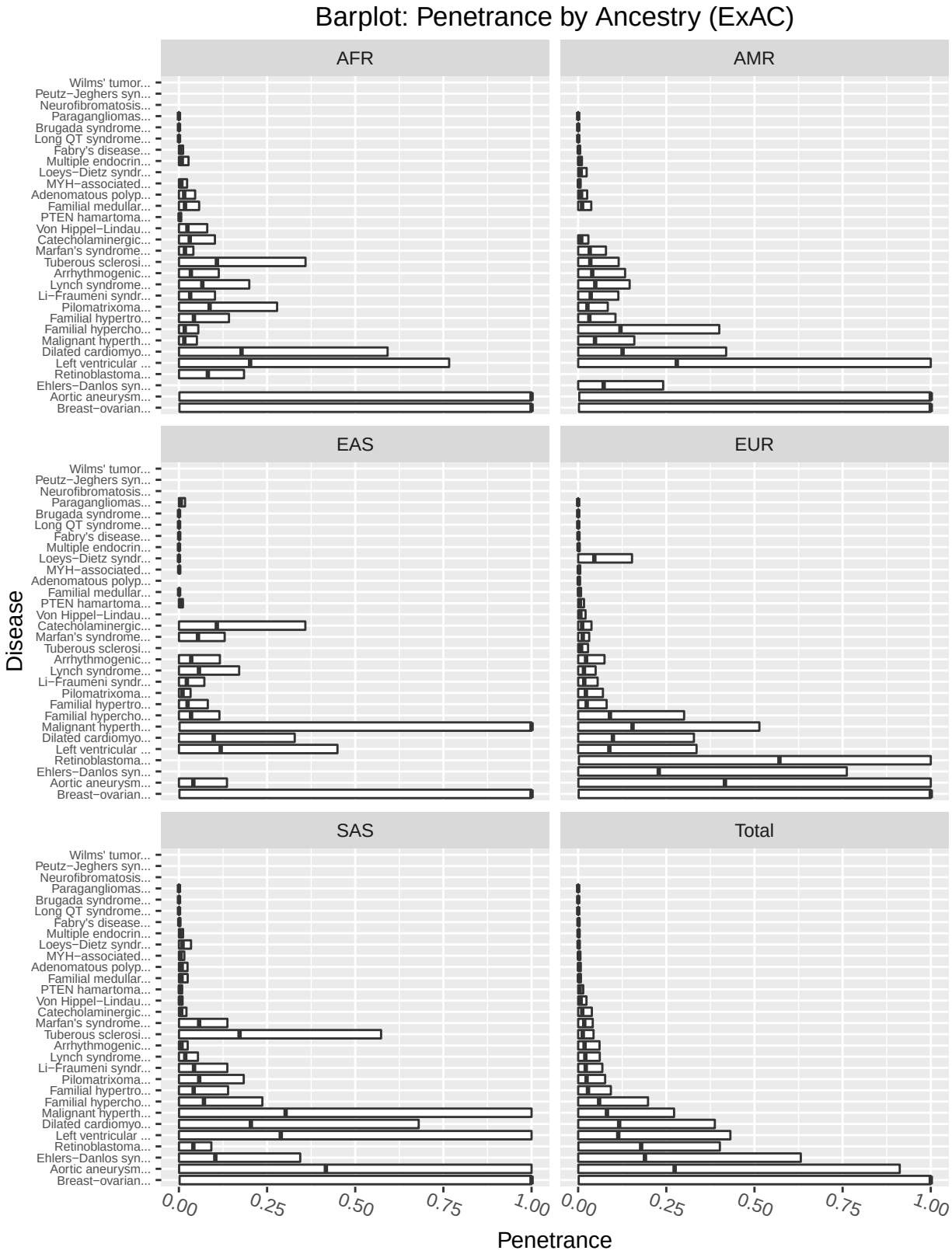
Note: Prevalence ranges of 5x were assumed for all point estimates of prevalence.
For example: a point estimate of 0.3 would be given the range [0.1, 0.5].

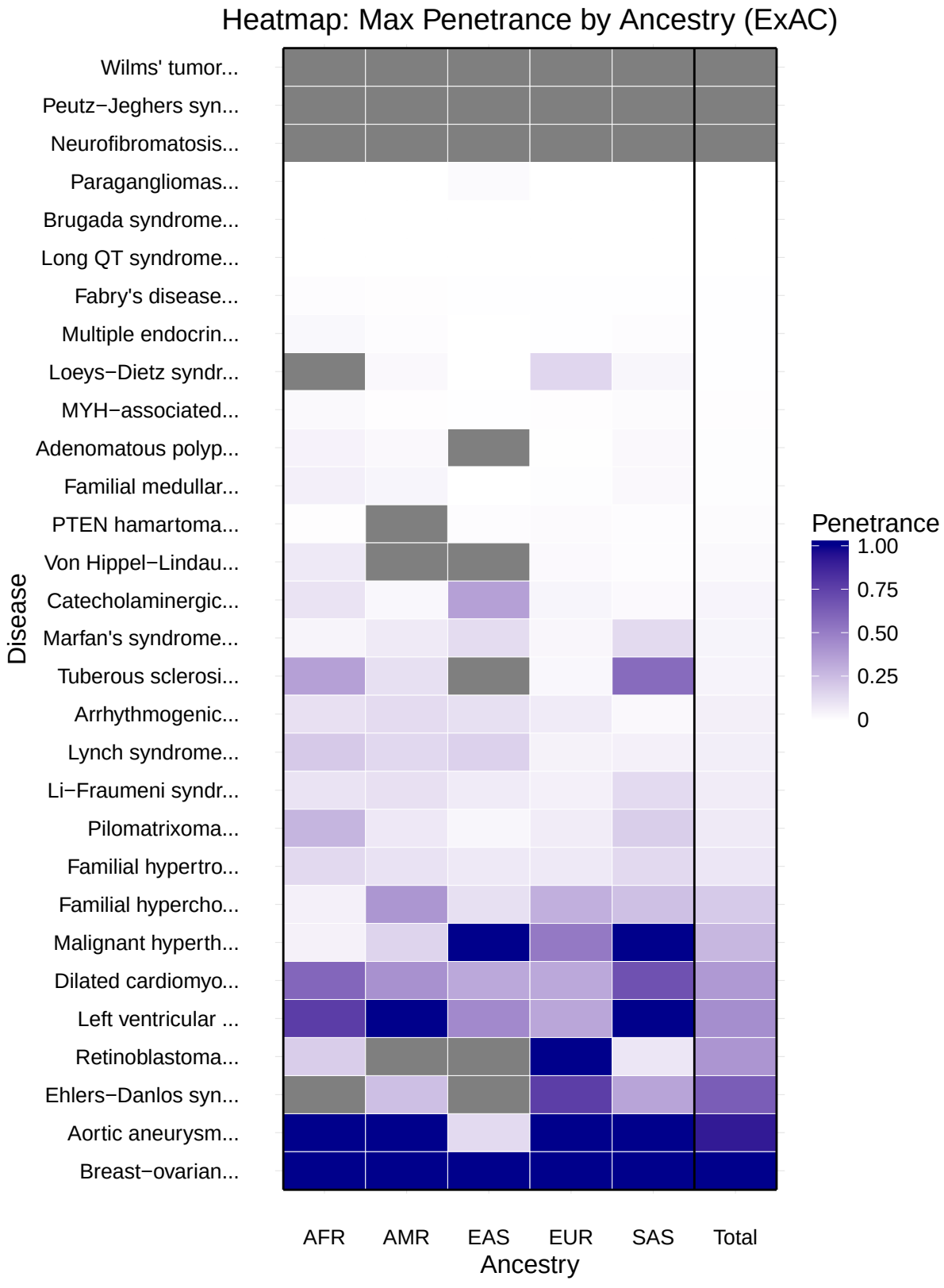
3.8 Penetrance Estimates by Ancestry





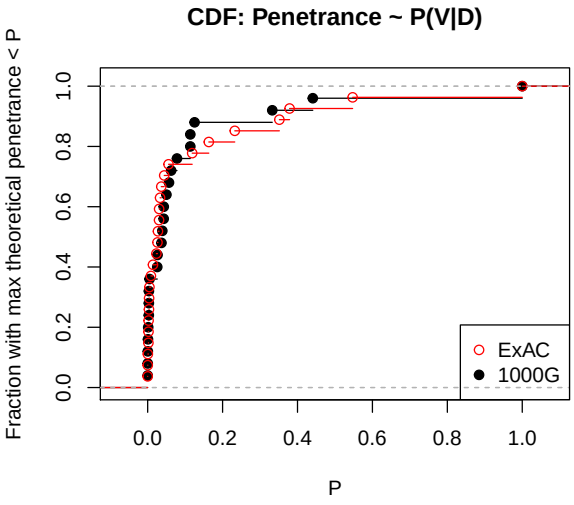
Dark gray boxes are NA: no associated variants discovered in that ancestral population.

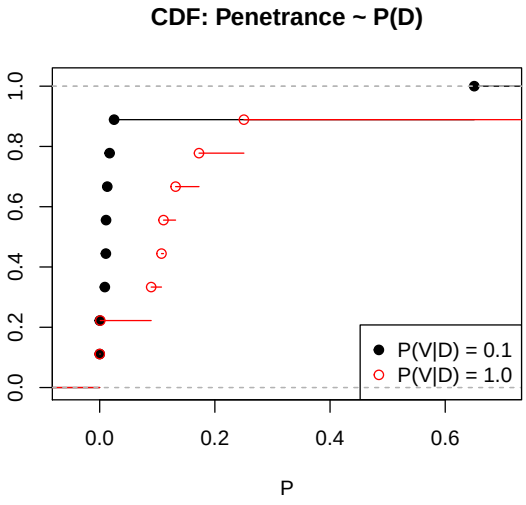

Barplot: Penetrance by Ancestry (ExAC)
AFR
AMR
EAS
EUR
SAS
Total
Disease
Wilms' tumor...
Peutz−Jeghers syn...
Neurofibromatosis...
Paragangliomas...
Brugada syndrome...
Long QT syndrome...
Fabry's disease...
Multiple endocrin...
Loeys−Dietz syndr...
MYH−associated...
Adenomatous polyp...
Familial medullar...
PTEN hamartoma...
Von Hippel−Lindau...
Catecholaminergic...
Marfan's syndrome...
Tuberous sclerosi...
Arrhythmogenic...
Lynch syndrome...
Li−Fraumeni syndr...
Pilomatrixoma...
Familial hypertro...
Familial hypercho...
Malignant hyperth...
Dilated cardiomyo...
Left ventricular ...
Retinoblastoma...
Ehlers−Danlos syn...
Aortic aneurysm...
Breast−ovarian...
0.00
0.25
0.50
0.75
1.00
Penetrance

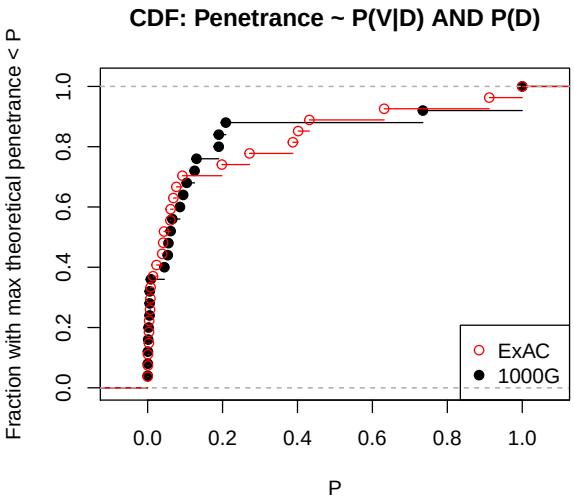


Dark gray boxes are NA: no associated variants discovered in that ancestral population.

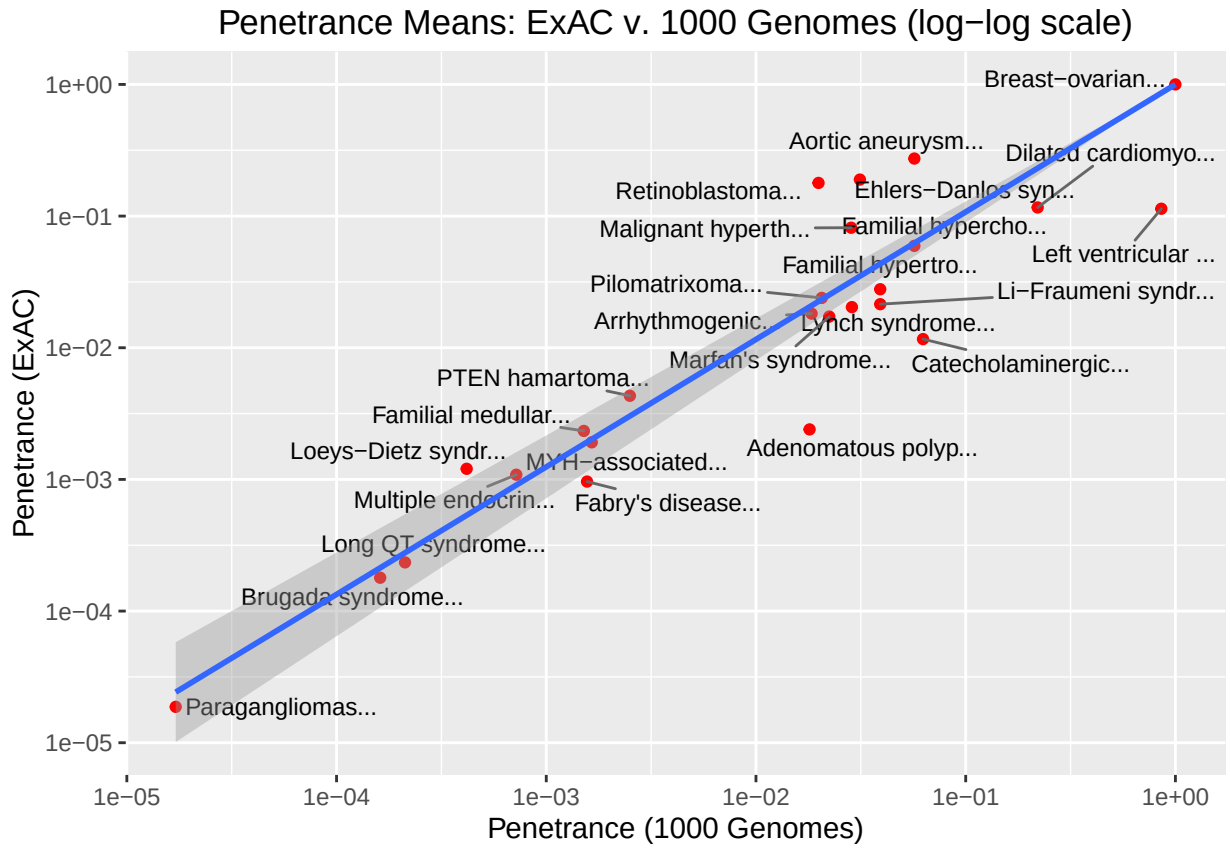
3.9 Empirical CDFs for All Penetrance Plots







3.10 Comparing Mean Penetrance between ExAC and 1000 Genomes



The Pearson correlation is 0.76.
Max penetrance values computed using 1000 Genomes are 0.944-fold larger than those computed using ExAC.