

Package ‘ADaM’

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Type Package

Title Adaptive Daisy Model

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Description The ADaM package implements a semi-supervised algorithm for computing a fuzzy-intersection of non-fuzzy sets by adaptively determining the minimal number of sets to which an element should belong in order to be a member of the fuzzy-intersection (the membership threshold). This threshold maximises the deviance from expectation of the cardinality of the resulting fuzzy-intersection, as well as the coverage of predefined elements. This method can be used to identify the minimal number of cell lines from a given tissue in which the inactivation of a gene (for example via CRISPR-Cas9 targeting) should exert a reduction of viability (or fitness effect) in order for that gene to be considered a core-fitness essential gene for the tissue under consideration. This method is used to discriminate between core-fitness and context-specific essential genes in a study describing a large scale genome-wide CRISPR-Cas9 pooled drop-out screening (Behan FM & Iorio F & Picco G et al., Prioritisation of cancer therapeutic targets using CRISPR-Cas9 screens. Nature, In press).

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LazyData true

Depends stats

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ADAM.empiricalOdds	<i>Empirical odds of fitness genes per number of cell lines</i>
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Description

This function calculates log10 odd ratios of observed vs. expected profiles of cumulative number of fitness genes in fixed number of cell lines. Expected values are the mean of those observed across randomised version of the observed binary matrix.

Usage

```
ADAM.empiricalOdds(observedCumSum, simulatedCumSum)
```

Arguments

observedCumSum	Observed profile of cumulative sum of numbers of fitness genes in fixed number of cell lines. This is generated by the ADAM.panessprofile function.
simulatedCumSum	Random profiles of cumulative sum of fitness genes in fixed number of cell lines. This is generated by the function ADAM.generateNullModel.

Value

A named vector:

odds	log base 10 odd ratios of observed versus expected cumulative sums of number of fitness genes across fixed numbers of cell lines.
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Author(s)

C. Pacini & F. Iorio

See Also

ADAM.panessprofile, ADAM.generateNullModel

Examples

```
data(exampleDepMat)
observed<-ADAM.panessprofile(depMat=exampleDepMat)
null_m<-ADAM.generateNullModel(depMat=exampleDepMat)
logOdds <- ADAM.empiricalOdds(observed$CUMsums, null_m$nullCumSUM)

logOdds
```

ADAM.generateNullModel

Generate null profile of number of fitness genes across fixed numbers of cell lines and cumulative sums.

Description

This function randomly perturbs the binary dependency matrix to generate a null distribution of profiles of fitness genes across fixed number of cell lines, and corresponding null distribution of cumulative sums.

Usage

```
ADAM.generateNullModel(depMat, ntrials=100, display=TRUE)
```

Arguments

depMat	Binary dependency matrix, rows are genes and columns are samples. 1 in position $[i,j]$ indicates that inactivation of the i -th gene exerts a significant loss of fitness in the j -th sample, 0 otherwise.
ntrials	Integer, default =100. How many times to randomly perturb dependency matrix to generate the null distributions.
display	Boolean, default is TRUE. Should bar plots of the null profiles be plotted

Details

For a number of trials specified in (ntrials) the inputted binary dependency matrix is randomised, keeping its column marginal sums. The profiles of fitness genes across fixed number of cell lines, and corresponding cumulative sums, are returned for each random perturbation.

Value

A list with the following two matrices

nullProf	Matrix of number of fitness genes for fixed number of cell lines from. Each rows of matrix corresponds to a random trial.
nullCumSum	Matrix of profile of cumulative number of fitness genes in fixed number of cell lines. Each row of matrix is one random trial.

Author(s)

C. Pacini & F. Iorio

Examples

```
data(exampleDepMat)
pprofile <- ADAM.generateNullModel(depMat = exampleDepMat, ntrials=100)
```

ADAM.panessprofile	<i>Calculate profile of number of fitness genes across fixed numbers of cell lines and cumulative sums.</i>
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Description

This function calculates the numbers (and cumulative numbers) of genes whose inactivation exerts a fitness effect in n cell lines, varying n from 1 to the number of cell lines in the dataset in input.

Usage

```
ADAM.panessprofile(depMat,
                   display=TRUE,
                   main_suffix='fitness genes in at least 1 cell line',
                   xlab='n. dependent cell lines')
```

Arguments

depMat	Binary dependency matrix, rows are genes and columns are samples. 1 in position $[i,j]$ indicates that inactivation of the i -th gene exerts a significant loss of fitness in the j -th sample, 0 otherwise.
display	Boolean, default is TRUE. Should bar plots of the dependency profiles be plotted
main_suffix	If display=TRUE, title suffix to give to plot of number of genes depleted in a give number of cell lines, default is 'genes depleted in at least 1 cell line'
xlab	If display=TRUE, label to give to x-axis of the plots, default is 'n. cell lines'

Value

A list with the following two named vectors:

panessprof	Number of genes that are depleted for a number of cell lines
CUMsums	Cumulative number of genes depleted in at least x cell lines

Author(s)

C. Pacini & F. Iorio

Examples

```
data(exampleDepMat)
pprofile <- ADAM.panessprofile(depMat = exampleDepMat)
```

ADAM.randomisedepMat *Binary matrix randomisation preserving column totals*

Description

This function takes in input a matrix and shuffles its entries column wisely. If the matrix is binary then the matrix resulting from this shuffling will have the same column marginal totals of the inputted one.

Usage

```
ADAM.randomisedepMat(depMat)
```

Arguments

depMat A numerical matrix.

Value

The matrix given in input with entries shuffled column wisely.

Author(s)

C. Pacini & F. Iorio

Examples

```
data(exampleDepMat)
rnd_exampleDepMat<-ADAM.randomisedepMat(exampleDepMat)
```

ADAM.tradeoffEO_TPR *Calculate ADaM threshold*

Description

This function finds the minimum number of cell lines in which a gene needs to be fitness in order to be called core-fitness. This is defined as the n providing the best trade-off between i) coverage of priori-known essential genes in the resulting set of predicted core-fitness genes, i.e. fitness in at least n cell lines, and ii) deviance from expectation of the number of fitness genes in n cell lines.

Usage

```
ADAM.tradeoffEO_TPR(E0, TPR, test_set_name)
```

Arguments

E0 Profile of empirical odds values. Computed with the ADAM.empiricalOdds function.

TPR Profile of True positive rates for across number of cell line. Computed with the ADAM.truePositiveRate function.

test_set_name Name to give to the analysis, used for plotting titles.

Details

Compare and plot the log10 odds ratios with the true positive rates to find the cross over point where the true positive rate falls below the odds ratio.

Value

point	Number of cell lines for which a gene needs to be a fitness gene in order to be predicted as core-fitness gene.
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Author(s)

C. Pacini & F. Iorio

See Also

[ADAM.empiricalOdds](#), [ADAM.truePositiveRate](#)

Examples

```
data(exampleDepMat)

pprofile<-ADAM.panessprofile(depMat=exampleDepMat)

nullmodel<-ADAM.generateNullModel(depMat=exampleDepMat,ntrials = 1000)

data(curated_BAGEL_essential)

EO<-ADAM.empiricalOdds(observedCumSum = pprofile$CUMsums,
                      simulatedCumSum =nullmodel$nullCumSUM)

TPR<-ADAM.truePositiveRate(exampleDepMat,curated_BAGEL_essential)

crossoverpoint<-ADAM.tradeoffEO_TPR(EO,TPR$TPR,
                                   test_set_name = 'curated BAGEL essential')
```

ADAM.truePositiveRate *Profile of True Positive Rates*

Description

This function calculates a profile of True Positive Rates for fitness genes in at least n cell lines, with positive cases from a reference set of essential genes.

Usage

```
ADAM.truePositiveRate(depMat,essentialGeneSet)
```

Arguments

depMat	Binary dependency matrix, rows are genes and columns are samples. 1 in position $[i,j]$ indicates that inactivation of the i -th gene exerts a significant loss of fitness in the j -th sample, 0 otherwise.
essentialGeneSet	Reference set of predefined essential genes. This is used to define positive cases.

Details

This function calculates true positive rates for fitness genes in at least n cell lines (for each n). First, this function calculates the number of cell lines each gene is a fitness gene. Second, for a given number of cell lines, the set of genes that are fitness genes in at least that number of cell lines is determined. Finally, this set of genes is then compared to the reference set of essential genes to calculate a true positive rate.

Value

A list of the following vectors:

P	Vector of number of genes that are depleted for a number of cell lines.
TP	Vector of number of genes in sets of P are true positives, i.e. in the essentialGeneSet.
TPR	TP divided by number of genes in set essentialGeneSet to give the true positive rate.

Author(s)

C. Pacini & F. Iorio

Examples

```
data(exampleDepMat)

pprofile<-ADAM.panessprofile(depMat=exampleDepMat)

nullmodel<-ADAM.generateNullModel(depMat=exampleDepMat,ntrials = 1000)

data(curated_BAGEL_essential)

EO<-ADAM.empiricalOdds(observedCumSum = pprofile$CUMsums,simulatedCumSum =nullmodel$nullCumSUM )

TPR<-ADAM.truePositiveRate(exampleDepMat,curated_BAGEL_essential)
```

curated_BAGEL_essential

Referecence set of core-fitness essential genes

Description

Reference set of predefined core-fitness essential genes used in [1], derived from [2] and further curated as follows. In order to avoid their status (essential/non-essential) being defined a priori, a set of high-confidence cancer driver genes from [3] were filtered out.

Usage

```
data(curated_BAGEL_essential)
```

Format

A vector of string with a gene symbol in each entry

References

- [1] Behan FM & Iorio F & Picco G et al., Prioritisation of cancer therapeutic targets using CRISPR-Cas9 screens. *Nature*, In press.
- [2] Hart T, Chandrashekhara M, Aregger M, Steinhart Z, Brown KR, MacLeod G, et al. High-Resolution CRISPR Screens Reveal Fitness Genes and Genotype-Specific Cancer Liabilities. *Cell*. 2015;163:1515–26.
- [3] Iorio F, Knijnenburg TA, Vis DJ, Bignell GR, Menden MP, Schubert M, et al. A Landscape of Pharmacogenomic Interactions in Cancer. *Cell*. 2016;166:740–54.

Examples

```
data(curated_BAGEL_essential)
head(curated_BAGEL_essential)
```

exampleDepMat

Example dependency matrix data object

Description

A binary matrix summarising the fitness effect status (1 = cellular fitness reduced upon gene inactivation via CRISPR-Cas9 targeting) of all the genes (at a genome scale) across 32 human colorectal cancer cell lines, derived from [1].

Usage

```
data("exampleDepMat")
```

Format

The format is: num [1:17995, 1:32] 0 0 0 0 0 0 0 0 0 0 ... - attr(*, "dimnames")=List of 2 ..\$: chr [1:17995] "A1BG" "A1CF" "A2M" "A2ML1"\$: chr [1:32] "Cell_line_1" "Cell_line_2" "Cell_line_3" "Cell_line_4" ...

References

- [1] Behan FM & Iorio F & Picco G et al., *Prioritisation of cancer therapeutic targets using CRISPR-Cas9 screens*. **Nature**, In press.

Examples

```
data(exampleDepMat)
head(exampleDepMat)
```


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