# Package 'signeR'

June 17, 2016

Type Package

Title signeR
<b>Version</b> 0.99.1
Author Rafael Rosales, Rodrigo Drummond, Renan Valieris, Israel Tojal da Silva
Maintainer Renan Valieris < renan.valieris@cipe.accamargo.org.br>
<b>Description</b> The signeR package provides an empirical Bayesian approach to mutational signature discovery. It is designed to analyze single nucleotide variation (SNV) counts in cancer genomes, but can also be applied to other features as well. Functionalities to characterize signatures or genome samples according to exposure patterns are also provided.
License GPL-3
Imports BiocGenerics, Biostrings, BSgenome (>= 1.36.3), class, graphics, grDevices, GenomicRanges, nloptr, NMF, R.methodsS3, R.oo, stats, tensorA, utils, VariantAnnotation
LinkingTo Rcpp, RcppArmadillo
SystemRequirements C++11
NeedsCompilation yes
ByteCompile TRUE
biocViews GenomicVariation, SomaticMutation, StatisticalMethod, Visualization
Suggests knitr, rtracklayer, base64
VignetteBuilder knitr
R topics documented:
signeR-package         2           Classify         2
DiffExp
generateMatrix
plots
signeR
SignExp
Index 10
1

2 Classify

signeR-package

signeR

# **Description**

The signeR package provides an empirical Bayesian approach to mutational signature discovery. It is designed to analyze single nucleotide variation (SNV) counts in cancer genomes, but can also be applied to other features as well. Functionalities to characterize signatures or genome samples according to exposure patterns are also provided.

#### **Details**

signeR package focus on the characterization and analysis of mutational processes. Its functionalities can be divided in three steps. Firstly, it provides tools to process VCF files and generate matrices of SNV mutation counts and mutational opportunities, both divided according to a 3bp context (mutation site and its neighboring bases). Secondly, the main part of the package takes those matrices as input and applies a Bayesian approach to estimate the number of underlying signatures and their mutational profiles. Thirdly, the package provides tools to correlate the activities of those signatures with other relevant information, e.g. clinical data, in order to infer conclusions about the analyzed genome samples, which can be useful for clinical applications.

#### Author(s)

Rodrigo Drummond, Rafael Rosales, Renan Valieris, Israel Tojal da Silva Maintainer: Renan Valieris <a href="mailto:renan.valieris@cipe.accamargo.org.br">renan.valieris@cipe.accamargo.org.br</a>

#### References

here be references

# **Examples**

```
vignette(package="signeR")
```

Classify

Classify unknown samples

#### **Description**

Classify: Assign unknown samples to previously defined groups.

# Usage

```
## S3 method for class 'SignExp'
Classify(this, labels, method="knn", k=3,
    plotfile="Classification_barplot.pdf", colors=NA, min_agree=0.75, ...)
```

Classify 3

#### **Arguments**

this a SignExp object returned by signeR function.

labels sample labels. Every sample labeled as NA will be classified according to its

mutational profile and the profiles of labeled samples.

method Classification algorithm used. Default is k-Nearest Neighbors (kNN). Any other

algorithm may be used, as long as it is customized to satisfy the following con-

ditions:

Input: a matrix of labeled samples, with one sample per line and one feature per column; a matrix of unlabeled samples to classify, with the same structure; an

array of labels, with one entry for each labeled sample.

Output: an array of assigned labels, one for each unlabeled sample.

k number of nearest neighbors considered for classification, used only if method="kNN".

Default is 3.

plotfile file that will be generated with classification graphic output.

colors Array of color names, one for each sample class. Colors will be recycled if the

length of this array is less than the number of classes.

min\_agree Minimum frequency of agreement among individual classifications. Samples

showing a frequency of agreement below this value are considered as "unde-

fined". Default is 0.75.

... additional parameters for classification algorithm (defined by "method" above).

#### Value

A list with the following items:

class The assigned classes for each unlabeled sample.

freq Classification agreement for each unlabeled sample: the relative frequency of

assignment of each sample to the group specified in "class".

allfreqs Matrix with one column for each unlabeled sample and one row for each group

label. Contains the assignment frequencies of each sample to each group.

# **Examples**

```
# assuming signatures is the return value of signeR()

## Not run:
    my_labels <- c("a","a",NA,"b","b",NA)
    Class <- Classify(signatures$SignExposures, labels=my_labels, plotfile="Sample_classification.pdf")

## End(Not run)
# see also
vignette(package="signeR")</pre>
```

4 DiffExp

DiffExp Differential Exposure Analysis	
--	--

#### **Description**

DiffExp: Identify signatures with significantly different activities among sample groups.

#### Usage

```
## S3 method for class 'SignExp'
DiffExp(this, labels, method='kw', contrast="all", quant=0.5,
   cutoff=0.05, plotfile="Diffexp_boxplot.pdf", colored=TRUE, ...)
```

a SignExp object returned by signeR function.

# **Arguments**

this

labels sample labels used to define sample groups. method algorithm used to compare each signature exposures among sample groups. Default is "kw", which leads to the use of Kruskal-Wallis Rank Sum Test. contrast defines which sample groups will be considered in the analysis. Default is "all", which leads the algorithm to evaluates the null hypothesis of exposure levels beeing constant in all groups. Instead, if this parameter contains a list of group labels, the algorithm will evaluate the null hypothesis of exposure levels beeing constant among those groups. quant the p-values quantile which, after log-transform, will be used as DES (Differental Exposure Score). Deafult is 0.5, which means the median log-transformed p-value will be considered as DES. cutoff

threshold for p-values quantile for signatures to be considered as showing differential exposure.

plotfile Output file to export p-values boxplot.

colored Boolean variable, if TRUE boxplots of differentially exposed signatures will be

> colored in green, cutoff line will be colored in red and line segments showing the transformed p-value quantile used for DE evaluation will be colored in blue.

Otherwise the plot will be black & white.

additional parameters for test algorithm defined by the method parameter.

#### Value

A list with the following items:

boolean array with one entry for each signature, indicating whether it shows Pvquant

differential exposure.

**Pvalues** matrix containing all computed p-values, with one row for each signature.

MostExposed for each differentially exposed signature, this array contains the label of the

group where it showed higher levels of exposure. Contains NA for signatures

not showing differential exposure.

generateMatrix 5

#### **Examples**

# **Description**

```
genCountMatrixFromVcf: generate count matrix from a VCF file. genOpportunityFromGenome: generate opportunity matrix from a target regions set.
```

# Usage

```
genCountMatrixFromVcf(bsgenome, vcfobj)
genOpportunityFromGenome(bsgenome, target_regions, nsamples=1)
```

# **Arguments**

bsgenome A BSgenome object, equivalent to the genome used for the variant call.

vcfobj A VCF object. See VCF-class from the VariantAnnotation package.

target\_regions A GRanges object, describing the target region analyzed by the variant caller.

Number of samples to generate the matrix, should be the same number as rows of the count matrix.

#### Value

```
A matrix of samples x (96 features). Each feature is a SNV change with a 3bp context.
```

# **Examples**

```
## Not run:
    library(BSgenome.Hsapiens.UCSC.hg19)
    library(rtracklayer)
    library(VariantAnnotation)

vcfobj <- readVcf("/path/to/a/file.vcf", "hg19")
    mut <- genCountMatrixFromVcf(BSgenome.Hsapiens.UCSC.hg19, vcfobj)

target_regions <- import(con="/path/to/a/target.bed", format="bed")</pre>
```

6 plots

```
opp <- genOpportunityFromGenome(BSgenome.Hsapiens.UCSC.hg19, target_regions, nsamples=nrow(mut))
## End(Not run)
# see also
vignette(package="signeR")</pre>
```

plots

signeR plot functions

#### **Description**

SignPlot: Plot the mutational signatures in a barchart, with error bars according to the variation of individual entries along generated Gibbs samples.

SignHeat: Plot the mutations signatures in a heatmap.

ExposureBoxplot: Boxplot of exposure values, showing their variation along generated Gibbs samples.

Paths: Plot the convergence of the Gibbs sampler for signatures and exposures on separate charts.

BICboxplot: Plot the measured values of the Bayesian Information Criterion (BICs) for tested model dimensions.

# Usage

#### **Arguments**

this a SignExp object returned by signeR function. e.g.: sig\$SignExposures signeRout the value returned by the signeR function.

Output pdf file of the plots.

Color palette used. Options are: "brew","lba","bcr1","bcr2","bw".

signeR 7

threshold entries below this value will be rounded to 0. Default is 0 (all entries are kept). plots\_per\_page How many plots in a single page, default is 4. Distance between consecutive bars on the plot. gap Order of signatures for plotting. Should be a permutation of 1:nsig, where nsig reord is the number of signatures. By default, signatures are ordered by the total exposure, in decreasing order. nbins The range of signature entries is divided in this number of bins for plotting, each bin corresponding to a different color. file\_suffix The suffix of the output file. col Single color name for boxplots.

#### Value

The plot result is saved in the file defined by the file argument.

# **Examples**

```
# assuming signatures is the return value of signeR()
## Not run:
    SignPlot(signatures$SignExposures)
    Paths(signatures$SignExposures)
## End(Not run)
vignette(package="signeR")
```

signeR

signeR

# Description

Generates the signatures.

# Usage

```
signeR(M, Mheader = TRUE, samples = "rows", Opport = NA,
   Oppheader = FALSE, nsig = NA, nlim = c(NA, NA),
   try_all = FALSE, ap = NA, bp = NA, ae = NA, be = NA,
   lp = NA, le = NA, var.ap = 10, var.ae = 10,
   testing_burn = 1000, testing_eval = 1000, main_burn =
   10000, main_eval = 2000, start = "lee", estimate_hyper
   = FALSE, EMit_lim=100)
```

8 signeR

# Arguments

į	guments	
	М	mutation counts matrix of samples x features.
	Mheader	If M have colnames defined use TRUE, if FALSE a default order will be assumed.
	samples	If the samples are row-wise or column-wise in M, default is "row".
	Opport	context count matrix of samples x features in the target genome or region.
	Oppheader	If Opport have header defined.
	nsig	Number of signatures, which can be provided or estimated by the algorithm.
	nlim	Define a interval to search for the optimal number of signatures.
	try_all	If true, all possible values for nsig will be tested
	ар	shape parameter of the gamma distribution used to generate the entries of a matrix of rate parameters of the gamma distributions which generate signatures.
	bp	rate parameter of the gamma distribution used to generate the entries of a matrix of rate parameters of the gamma distributions which generate signatures.
	ae	shape parameter of the gamma distribution used to generate the entries of a matrix of rate parameters of the gamma distributions which generate exposures.
	be	rate parameter of the gamma distribution used to generate the entries of a matrix of rate parameters of the gamma distributions which generate exposures.
	lp	parameter of the exponential distribution used to generate the entries of a matrix of shape parameters of the gamma distributions which generate signatures.
	le	parameter of the exponential distribution used to generate the entries of a matrix of shape parameters of the gamma distributions which generate exposures.
	var.ap	variance of the gamma distribution used to generate proposals for shape parameters of signatures
	var.ae	variance of the gamma distribution used to generate proposals for shape parameters of exposures
	testing_burn	number of burning iterations of the Gibbs sampler used to estimate the number of signatures in data.
	testing_eval	number of iterations of the Gibbs sampler used to estimate the number of signatures in data.
	main_burn	number of burning iterations of the final Gibbs sampler.
	main_eval	number of iterations of the final Gibbs sampler.
	start	NMF algorithm used to generate initial values for signatures and exposures, options: "brunet", "KL", "lee", "Frobenius", "offset", "nsNMF", "ls-nmf", "pe-nmf", "siNMF", "snmf/r' or "snmf/l".
	estimate_hyper	if TRUE, algorithm estimates optimal values of ap,bp,ae,be,lp,le. Start values can still be provided.
	EMit_lim	Limit of EM iterations for the estimation of hyper-hyperparameters ap,bp,ae,be,lp,le.

Default is 100.

SignExp 9

# Value

signeR output is a list with the following items:

Nsign selected number of signatures.

tested\_n array containing the numbers of signatures tested by the algorithm.

Test\_BICs list of measured BIC values when testing different numbers of signatures.

Phat Estimated signatures, median of P samples.

Ehat Estimated exposures, median of E samples.

SignExposures SignExp object which contain the set of samples for the model parameters.

Bics measured BIC values on the final run of the sampler.

HyperParam evolution of estimated hyperparameters when testing different numbers of sig-

natures.

# **Examples**

vignette(package="signeR")

SignExp SignExp class

# **Description**

Keep samples for signature and exposure matrices.

# Value

# Object fields:

. Sign tensor of signature matrix samples.
.Exp tensor of exposure matrix samples.

. sigSums Signature sums for each sample, organized by row. Normalizing factors.

. samples Genome sample IDs.
.mutations mutation names.

. normalized boolean variable, indicating whether Sign tensor has been normalized.

# **Index**

```
*Topic package
    signeR-package, 2
BICboxplot (plots), 6
{\tt Classify}, {\color{red} 2}
DiffExp, 4
ExposureBoxplot (plots), 6
genCountMatrixFromVcf(generateMatrix),
generateMatrix, 5
{\tt genOpportunityFromGenome}
         (generateMatrix), 5
Paths (plots), 6
plots, 6
signeR, 7
\verb|signeR-package|, 2
SignExp, 9
SignHeat (plots), 6
SignPlot (plots), 6
```