

Mutations in RNA Splicing Machinery in Human Cancers

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Massively parallel sequencing of cancer genomes is revealing a panoramic view of the genetic drivers of human neoplasms. In this issue of the *Journal*, Wang et al.¹ describe an analysis of the coding sequences of samples from 91 patients with chronic lymphocytic leukemia. The disease is characterized by the accumulation of mature B lymphocytes, and its genetic basis is being rapidly elucidated.^{2,3}

Wang et al. leveraged the large number of samples studied to identify sets of genes that are critical to the development of chronic lymphocytic leukemia. By sequencing both the malignant lymphocytes (CD19+CD5+) and matched nonmalignant control DNA from each patient, the authors pinpointed mutations that occurred somatically. By identifying genes and pathways with recurrent mutations, they highlighted the

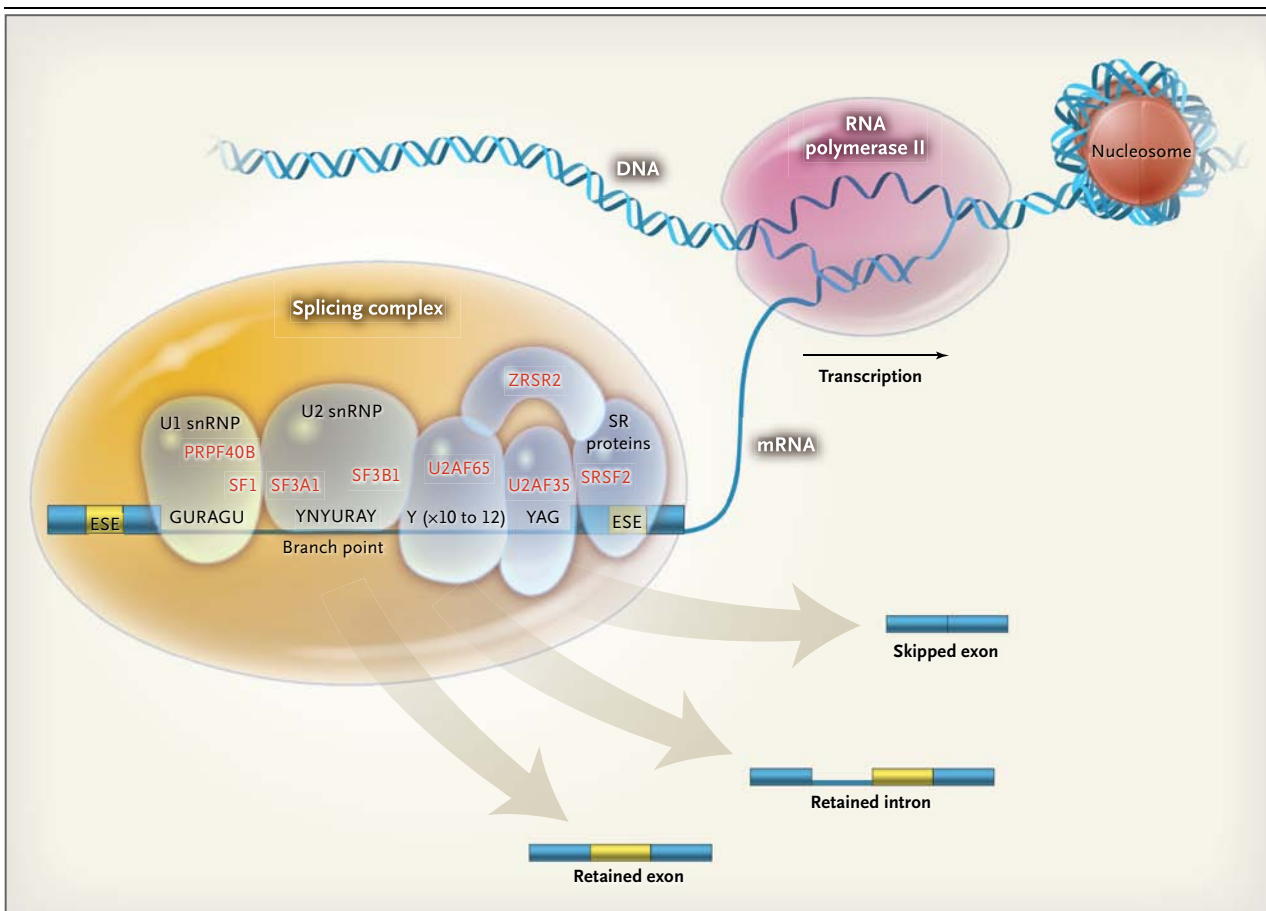


Figure 1. Schematic Representation of RNA Splicing.

After the transcription of nucleosome-free DNA into pre-messenger RNA (pre-mRNA) by RNA polymerase II, the RNA is processed and spliced into mRNA. Components of the early splicing complex are illustrated. Serine- and arginine-rich (SR) proteins bind to the exonic splice enhancer (ESE). The U1 small nuclear ribonucleoprotein (snRNP) binds to the 5' splice site, and the U2 snRNP binds to the branch point. The U2 auxiliary factor (U2AF) binds to the pyrimidine stretch and the 3' splice site. The genes encoding SF3B1, SF1, SF3A1, SRSF2, U2AF35, U2AF65, ZRSR2, and PRPF40B are mutated in myeloid cancers or in chronic lymphocytic leukemia; these proteins are shown in red. Three splicing outcomes are illustrated (retained exon, skipped exon, and retained intron), each of which could be affected by mutations that alter the splicing complex. Y denotes a pyrimidine nucleotide, N any nucleotide, and Y (x10 to 12) a stretch of 10 to 12 pyrimidines.

developmental drivers of chronic lymphocytic leukemia. Finally, the authors elegantly integrated the mutation data with known biologic information to define five key pathways of chronic lymphocytic leukemia that are affected by mutation: DNA damage and cell-cycle control, Notch signaling, inflammatory pathways, Wnt signaling, and RNA splicing.

The identification of mutations in genes involved in RNA splicing was highly unexpected, but it converges remarkably with recently published studies making use of genome sequencing in myelodysplastic syndromes.^{4,5} *SF3B1*, which encodes a core member of the U2 small nuclear ribonucleoprotein (U2 snRNP) complex, was mutated in 15% of the 91 patients in the current study, primarily as a recurrent, heterozygous missense mutation, K700E. A separate study published recently in the *Journal* reported *SF3B1* mutations in 20% of patients with myelodysplastic syndromes and 65% of patients with refractory anemia and ring sideroblasts.⁴ Moreover, mutations have been reported in multiple components of the spliceosome in 45 to 85% of patients with myelodysplastic syndrome.⁵ *SF3B1* mutations also occur in 1 to 5% of samples from a wide range of tumor types, which indicates that mutations in RNA splicing factors are a widespread cause of oncogenic transformation.⁴

The vast majority of human genes undergo RNA splicing after transcription, which means that mutations in the RNA splicing machinery could potentially alter the maturation of messenger RNA for most genes and the subsequent production of protein (Fig. 1). In addition, RNA splicing is linked to the epigenetic regulation of gene expression. In particular, *SF3B1* has been reported to interact with the polycomb repressive complex, an important regulator of hematopoiesis.⁶ Genes encoding members of these complexes are mutated in hematologic cancers.

The finding of *SF3B1* mutations in both chronic lymphocytic leukemia and myelodysplastic syndromes resonates with the recent finding of *TET2* mutations in both lymphoid and myeloid cancers.⁷ These developments raise the provocative possibility that *SF3B1* mutations might in some cases occur initially in hematopoietic stem cells, with additional mutations then being acquired in either the lymphoid or the myeloid lineages and causing chronic lymphocytic leukemia or myelodysplastic syndromes, respectively.

Consistent with this hypothesis, stem cells from patients with chronic lymphocytic leukemia have recently been reported to be abnormally lymphoid-primed, a finding that suggests that chronic lymphocytic leukemia could also derive from a stem-cell defect.⁸

The genetic characterization of chronic lymphocytic leukemia has the potential to refine the molecular classification and estimation of prognosis for this disease. Patterns of genetic lesions, such as the association of *SF3B1* mutations with deletion in chromosome 11q and with *ATM* mutations, provide clues about the molecular circuitry of chronic lymphocytic leukemia cells. The identification of mutations in genes encoding the RNA splicing machinery raises the intriguing possibility that the spliceosome could be a therapeutic target for the treatment of chronic lymphocytic leukemia and myelodysplastic syndromes.^{9,10}

Disclosure forms provided by the authors are available with the full text of this article at NEJM.org.

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1. Wang L, Lawrence MS, Wan Y, et al. *SF3B1* and other novel cancer genes in chronic lymphocytic leukemia. *N Engl J Med* 2011;365:2497-506.
2. Fabbri G, Rasi S, Rossi D, et al. Analysis of the chronic lymphocytic leukemia coding genome: role of NOTCH1 mutational activation. *J Exp Med* 2011;208:1389-401.
3. Puente XS, Pinyol M, Quesada V, et al. Whole-genome sequencing identifies recurrent mutations in chronic lymphocytic leukaemia. *Nature* 2011;475:101-5.
4. Papaemmanuil E, Cazzola M, Boultonwood J, et al. Somatic *SF3B1* mutation in myelodysplasia with ring sideroblasts. *N Engl J Med* 2011;365:1384-95.
5. Yoshida K, Sanada M, Shiraishi Y, et al. Frequent pathway mutations of splicing machinery in myelodysplasia. *Nature* 2011;478:64-9.
6. Isono K, Mizutani-Koseki Y, Komori T, Schmidt-Zachmann MS, Koseki H. Mammalian polycomb-mediated repression of Hox genes requires the essential spliceosomal protein *Sf3b1*. *Genes Dev* 2005;19:536-41.
7. Quivoron C, Couronné L, Della Valle V, et al. *TET2* inactivation results in pleiotropic hematopoietic abnormalities in mouse and is a recurrent event during human lymphomagenesis. *Cancer Cell* 2011;20:25-38.
8. Kikushige Y, Ishikawa F, Miyamoto T, et al. Self-renewing hematopoietic stem cell is the primary target in pathogenesis of human chronic lymphocytic leukemia. *Cancer Cell* 2011;20:246-59.
9. Kaida D, Motoyoshi H, Tashiro E, et al. Spliceostatin A targets *SF3b* and inhibits both splicing and nuclear retention of pre-mRNA. *Nat Chem Biol* 2007;3:576-83.
10. Yokoi A, Kotake Y, Takahashi K, et al. Biological validation that *SF3b* is a target of the antitumor macrolide pladienolide. *FEBS J* 2011;278:4870-80.

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