## Contents

1	Intr	oducti	on	1
	1.1	Cance	r Research in the Post-Genomic Era	1
		1.1.1	Cancer as a Global Health Concern	2
			1.1.1.1 The Genetics and Molecular Biology of Cancers	3
		1.1.2	The Human Genome Revolution	6
			1.1.2.1 The First Human Genome Sequence	6
			1.1.2.2 Impact of Genomics	7
		1.1.3	Technologies to Enable Genetics Research	7
			1.1.3.1 DNA Sequencing and Genotyping Technologies	7
			1.1.3.2 Microarrays and Quantitative Technologies	8
			1.1.3.3 Massively Parallel "Next Generation" Sequencing	9
			1.1.3.3.1 Molecular Profiling with Genomics Technology .	11
			1.1.3.3.2 Sequencing Technologies	11
			1.1.3.4 Bioinformatics as Interdisciplinary Genomic Analysis .	12
		1.1.4	Follow-up Large-Scale Genomics Projects	13
		1.1.5	Cancer Genomes	14
			1.1.5.1 The Cancer Genome Atlas Project	15
			1.1.5.1.1 Findings from Cancer Genomes	15
			1.1.5.1.2 Genomic Comparisons Across Cancer Tissues .	17
			1.1.5.1.3 Cancer Genomic Data Resources	18
		1.1.6	Genomic Cancer Medicine	18
			1.1.6.1 Cancer Genes and Driver Mutations	18
			1.1.6.2 Personalised or Precision Cancer Medicine	19
			1.1.6.2.1 Molecular Diagnostics and Pan-Cancer Medicine	20
			1.1.6.3 Targeted Therapeutics and Pharmacogenomics	21
			1.1.6.3.1 Targeting Oncogenic Driver Mutations	21
			1.1.6.4 Systems and Network Biology	22
			1.1.6.4.1 Network Medicine, and Polypharmacology	24
	1.2	A Syn	thetic Lethal Approach to Cancer Medicine	25
		1.2.1	Synthetic Lethal Genetic Interactions	26
		1.2.2	Synthetic Lethal Concepts in Genetics	26
		1.2.3	Studies of Synthetic Lethality	27
			1.2.3.1 Synthetic Lethal Pathways and Networks	28
			1.2.3.1.1 Evolution of Synthetic Lethality	29
		1.2.4	Synthetic Lethal Concepts in Cancer	29

		1.2.5	Clinical Impact of Synthetic Lethality in Cancer	31
		1.2.6	High-throughput Screening for Synthetic Lethality	33
			1.2.6.1 Synthetic Lethal Screens	34
		1.2.7	Computational Prediction of Synthetic Lethality	37
			1.2.7.1 Bioinformatics Approaches to Genetic Interactions	37
			1.2.7.2 Comparative Genomics	38
			1.2.7.3 Analysis and Modelling of Protein Data	41
			1.2.7.4 Differential Gene Expression	43
			1.2.7.5 Data Mining and Machine Learning	44
			1.2.7.6 Bimodality	47
			1.2.7.7 Rationale for Further Development	48
	1.3	F-cad	herin as a Synthetic Lethal Target	48
	1.0	1.3.1	The <i>CDH1</i> gene and it's Biological Functions	48
		1.0.1	1.3.1.1 Cytoskeleton	49
			1.3.1.2 Extracellular and Tumour Micro-Environment	49
			1.3.1.3 Cell-Cell Adhesion and Signalling	49
		1.3.2	CDH1 as a Tumour (and Invasion) Suppressor	50
		1.0.2	1.3.2.1 Breast Cancers and Invasion	50
		1.3.3	Hereditary Diffuse Gastric Cancer and Lobular Breast Cancer .	50
		1.3.4	Somatic Mutations	52
		1.5.4	1.3.4.1 Mutation Rate	52
		1.3.5	1.3.4.2 Co-occurring Mutations	52 53
	1.4			54
	1.4	Summ	nary and Research Direction of Thesis	04
2	Met	thods	and Resources	58
	2.1		Formatics Resources for Genomics Research	58
		2.1.1	Public Data and Software Packages	58
			2.1.1.1 Cancer Genome Atlas Data	59
			2.1.1.2 Reactome and Annotation Data	60
	2.2	Data	Handling	61
			Normalisation	61
		2.2.2	Sample Triage	61
		2.2.3	Metagenes and the Singular Value Decomposition	63
		2.2.0	2.2.3.1 Candidate Triage and Integration with Screen Data	63
	2.3	Tochn	iques	64
		2.3.1	Statistical Procedures and Tests	64
		2.3.1 2.3.2	Statistical Procedures and Tests	64 65
		2.3.1 2.3.2 2.3.3	Statistical Procedures and Tests	64 65 66
		2.3.1 2.3.2 2.3.3 2.3.4	Statistical Procedures and Tests	64 65 66 66
		2.3.1 2.3.2 2.3.3	Statistical Procedures and Tests	64 65 66 66 66
		2.3.1 2.3.2 2.3.3 2.3.4 2.3.5	Statistical Procedures and Tests Gene Set Over-representation Analysis Clustering Heatmap Modeling and Simulations 2.3.5.1 Receiver Operating Characteristic (Performance)	64 65 66 66 67
		2.3.1 2.3.2 2.3.3 2.3.4 2.3.5	Statistical Procedures and Tests Gene Set Over-representation Analysis Clustering Heatmap Modeling and Simulations 2.3.5.1 Receiver Operating Characteristic (Performance) Resampling Analysis	64 65 66 66 67 68
	2.4	2.3.1 2.3.2 2.3.3 2.3.4 2.3.5 2.3.6 Pathw	Statistical Procedures and Tests  Gene Set Over-representation Analysis  Clustering  Heatmap  Modeling and Simulations  2.3.5.1 Receiver Operating Characteristic (Performance)  Resampling Analysis  vay Structure Methods	64 65 66 66 67 68 69
		2.3.1 2.3.2 2.3.3 2.3.4 2.3.5	Statistical Procedures and Tests Gene Set Over-representation Analysis Clustering Heatmap Modeling and Simulations 2.3.5.1 Receiver Operating Characteristic (Performance) Resampling Analysis	64 65 66 66 67 68

		2.4.3 Constructing Pathway Subgraphs	70 70
	2.5	Implementation	71
	2.0	2.5.1 Computational Resources and Linux Utilities	71
		2.5.2 R Language and Packages	73
		2.5.3 High Performance and Parallel Computing	75
		2.5.5 Ingh i chormance and i aranci companing	10
3		thods Developed During Thesis	77
	3.1	A Synthetic Lethal Detection Methodology	77
	3.2	Synthetic Lethal Simulation and Modelling	80
		3.2.1 A Model of Synthetic Lethality in Expression Data	80
		3.2.2 Simulation Procedure	84
	3.3	Detecting Simulated Synthetic Lethal Partners	87
		3.3.1 Binomial Simulation of Synthetic lethality	87
		3.3.2 Multivariate Normal Simulation of Synthetic lethality	89
		3.3.2.1 Multivariate Normal Simulation with Correlated Genes	92
		3.3.2.2 Specificity with Query-Correlated Pathways	99
		3.3.2.2.1 Importance of Directional Testing	99
	3.4	Graph Structure Methods	101
		3.4.1 Upstream and Downstream Gene Detection	101
		3.4.1.1 Permutation Analysis for Statistical Significance	102
		3.4.1.2 Hierarchy Based on Biological Context	103
		3.4.2 Simulating Gene Expression from Graph Structures	104
	3.5	Customised Functions and Packages Developed	108
		3.5.1 Synthetic Lethal Interaction Prediction Tool	108
		3.5.2 Data Visualisation	109
		3.5.3 Extensions to the iGraph Package	111
		3.5.3.1 Sampling Simulated Data from Graph Structures	111
		3.5.3.2 Plotting Directed Graph Structures	111
		3.5.3.3 Computing Information Centrality	112
		3.5.3.4 Testing Pathway Structure with Permutation Testing .	112
		3.5.3.5 Metapackage to Install iGraph Functions	113
4	Syn	thetic Lethal Analysis of Gene Expression Data	114
	4.1	Synthetic lethal genes in breast cancer	115
		4.1.1 Synthetic lethal pathways in breast cancer	117
		4.1.2 Expression profiles of synthetic lethal partners	118
		4.1.2.1 Subgroup pathway analysis	121
	4.2	Comparison of synthetic lethal gene candidates	124
		4.2.1 Comparison with siRNA screen candidates	124
		4.2.1.1 Comparison with correlation	125
		4.2.1.2 Comparison with viability	126
		4.2.1.3 Comparison with secondary siRNA screen candidates .	130
		4.2.1.4 Comparison of screen at pathway level	130
		4.2.1.4.1 Resampling of genes for pathway enrichment	132
	4.3	Metagene Analysis	138

		4.3.1	Pathway expression	38
		4.3.2	Somatic mutation	41
		4.3.3	Mutation locus	42
		4.3.4	Synthetic lethal metagenes	44
	4.4	Replic	eation in stomach cancer	45
		4.4.1	Synthetic Lethal Genes and Pathways	46
		4.4.2	Synthetic Lethal Expression Profiles	48
		4.4.3	Comparison to Primary Screen	<b>5</b> 0
			4.4.3.1 Resampling Analysis	51
		4.4.4	Metagene Analysis	
	4.5	Globa	l Synthetic Lethality	52
		4.5.1	Hub Genes	53
		4.5.2	Hub Pathways	
	4.6	_	eation in cell line encyclopaedia	
	4.7	Discus	ssion	
		4.7.1	Strengths of the SLIPT Methodology	
		4.7.2	Syntheic Lethal Pathways for E-cadherin	
		4.7.3	Replication and Validation	
			4.7.3.1 Integration with siRNA Screening	
			4.7.3.2 Replication across Tissues and Cell lines 16	
	4.8	Summ	ary	33
5	Svn	thetic	Lethal Pathway Structure 16	32
_	5.1		etic Lethal Genes in Reactome Pathways	
		5.1.1	The PI3K/AKT Pathway	
		5.1.2	The Extracellular Matrix	<sub>35</sub>
		5.1.3	G Protein Coupled Receptors	<sub>38</sub>
		5.1.4	Gene Regulation and Translation	<sub>38</sub>
	5.2	Netwo	ork Analysis of Synthetic Lethal Genes	39
		5.2.1	Gene Connectivity and Vertex Degree	<b>3</b> 9
		5.2.2	Gene Importance and Centrality	71
			5.2.2.1 Information Centrality	71
			5.2.2.2 PageRank Centrality	73
	5.3	Testin	g Pathway Structure of Synthetic Lethal Genes	74
		5.3.1	Hierarchical Pathway Structure	74
			5.3.1.1 Contextual Hierarchy of PI3K	74
			5.3.1.2 Testing Contextual Hierarchy of Synthetic Lethal Genes 17	75
		5.3.2	Upstream or Downstream Synthetic Lethality	78
			5.3.2.1 Measuring Structure of Candidates within PI3K 17	78
			5.3.2.2 Resampling for Synthetic Lethal Pathway Structure 18	80
			5.5.2.2 Resampling for Synthetic Lethar Lathway Structure 10	
	5.4	Discus	ssion	

$\operatorname{Sim}$	ulation and Modeling of Synthetic Lethal Pathways	186
6.1	Comparing methods	187
	6.1.1 Performance of SLIPT and $\chi^2$ across Quantiles	187
	6.1.1.1 Correlated Query Genes affects Specificity	189
	6.1.2 Correlation as a Synthetic Lethal Detection Strategy	191
	6.1.3 Testing for Bimodality with BiSEp	193
	6.1.4 Testing Synthetic Lethal Genes with Linear Models	193
6.2	Simulations with Graph Structures	193
	6.2.1 Performance over a Graph Structure	193
	6.2.2 Performance with Inhibitions	198
	6.2.3 Synthetic Lethality across Graph Structures	203
	6.2.4 Performance with feasible gene numbers (20,000)	207
6.3	Simulations over pathway-based graphs	216
6.4	Discussion	218
6.5	Summary	218
Disc	cussion	220
	8	
		226
Con	iciusion	220
Refe	erences	227
Sam	aple Quality	252
A.1	Sample Correlation	252
A.2	Replicate Samples in TCGA Breast	255
Soft	tware Used for Thesis	259
Seco	ondary Screen Data	268
Mui	tation Analysis in Breast Cancer	270
	· ·	
D.3	-	
	· v	
D.4		280
D.5		282
D.6	e · · · · · · · · · · · · · · · · · · ·	283
	D.6.2 PI3K Mutation Expression	
Met	tagene Expression Profiles	287
	6.1 6.2 6.3 6.4 6.5 Disc 7.1 7.2 7.3 Cor Ref San A.1 A.2 Soft Sec Mu D.1 D.2 D.3 D.4 D.5 D.6	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

${f F}$	Stomach Expression Analysis	293
	F.1 Synthetic Lethal Genes and Pathways	293
	F.2 Comparison to Primary Screen	296
	F.2.1 Resampling Analysis	
	F.3 Metagene Analysis	300
$\mathbf{G}$	Stomach Mutation Analysis	301
	G.1 Synthetic Lethal Genes and Pathways	301
	G.2 Synthetic Lethal Expression Profiles	304
	G.3 Comparison to Primary Screen	307
	G.3.1 Resampling Analysis	309
	G.4 Metagene Analysis	311
Н	Global Synthetic Lethality in Stomach Cancer	312
	H.1 Hub Genes	314
	H.2 Hub Pathways	315
Ι	Replication in cell line encyclopaedia	316
J	Synthetic Lethal Genes in Pathways	321
K	Pathway Connectivity for Mutation SLIPT	329
${f L}$	Information Centrality for Gene Essentiality	333
$\mathbf{M}$	Pathway Structure for Mutation SLIPT	336
N	Performance of SLIPT and $\chi^2$	339
- '	N.0.1 Correlated Query Genes affects Specificity	346
$\cap$	Charle Standards	352
U	Graph Structures	332
P	Graph Simulations	362
Q	Graph Simulations with Inhibition	367
$\mathbf{R}$	Simulation across Graph Structures	377
$\mathbf{S}$	Graph Simulations 20K genes	381
$\mathbf{T}$	Graph Simulations 20K genes with Inhibition	386
U	Pathway Simulations	396

# List of Figures

1.1	Synthetic genetic interactions
1.2	Synthetic lethality in cancer
2.1	Read count density
2.2	Read count sample mean
3.1	Framework for synthetic lethal prediction
3.2	Synthetic lethal prediction adapted for mutation
3.3	A model of synthetic lethal gene expression
3.4	Modeling synthetic lethal gene expression
3.5	Synthetic lethality with multiple genes
3.6	Simulating gene function
3.7	Simulating synthetic lethal gene function
3.8	Simulating synthetic lethal gene expression
3.9	Performance of binomial simulations
3.10	Comparison of statistical performance
3.11	Performance of multivariate normal simulations
3.12	Simulating expression with correlated gene blocks
3.13	Simulating expression with correlated gene blocks
3.14	Synthetic lethal prediction across simulations
3.15	Performance with correlations
3.16	Comparison of statistical performance with correlation structure 97
3.17	Performance with query correlations
3.18	Statistical evaluation of directional criteria
3.19	Performance of directional criteria
3.20	Simulated graph structures
	Simulating expression from a graph structure
3.22	Simulating expression from graph structure with inhibitions 107
3.23	Demonstration of violin plots with custom features
3.24	Demonstration of annotated heatmap
3.25	Simulating graph structures
4.1	Synthetic lethal expression profiles of analysed samples
4.2	Comparison of SLIPT to siRNA
4.3	Compare SLIPT and siRNA genes with correlation
4.4	Compare SLIPT and siRNA genes with correlation
4.5	Compare SLIPT and siRNA genes with siRNA viability

4.6	Compare SLIPT and siRNA genes with viability	127
4.7	Compare SLIPT and siRNA genes with siRNA viability	129
4.8	Resampled intersection of SLIPT and siRNA candidates	133
4.9	Pathway metagene expression profiles	139
4.10	Somatic mutation against PI3K metagene	141
4.11		143
4.12	Synthetic lethal expression profiles of stomach samples	149
	Synthetic lethal partners across query genes	153
F 1		101
5.1	Synthetic Lethality in the PI3K Cascade	164
5.2	Synthetic Lethality in the Elastic Fibre Formation Pathway	166
5.3	Synthetic Lethality in the Fibrin Clot Formation	167
5.4	Synthetic Lethality and Vertex Degree	170
5.5	Synthetic Lethality and Centrality	172
5.6	Synthetic Lethality and PageRank	174
5.7	Structure of PI3K Ranking	175
5.8	Synthetic Lethality and Hierarchy Score in PI3K	176
5.9	Hierarchy Score in PI3K against Synthetic Lethality in PI3K	176
5.10	v	178
5.11	Structure of Synthetic Lethality Resampling in PI3K	179
6.1	Performance on Graph Structures	188
6.2	Performance on Graph Structures	189
6.3	Performance on Graph Structures	190
6.4	Performance on Graph Structures	190
6.5	Performance on Graph Structures	192
6.6	Performance of simulations on a simple graph	194
6.7	Performance of simulations is similar in simple graphs	195
6.8	Performance of simulations on a graph	196
6.9	Performance of simulations on a large graph	197
6.10	Performance of simulations on a simple inhibiting graph	199
	Performance is higher on a simple inhibiting graph	200
	Performance of simulations on an inhibiting graph	201
	Performance is affected by inhibition in graphs	202
	Performance is affected by inhibition in graphs	203
	Performance is affected by inhibition in graphs	204
	Performance is affected by inhibition in graphs	205
	Performance is affected by inhibition in graphs	206
	Performance of simulations including a simple graph	208
	Performance on a simple graph improves with more genes	209
	Performance of simulations including a graph structure	210
	Performance of simulations including an inhibiting graph	211
	Performance on an inhibiting graph improves with more genes	212
	Performance of simulations including a graph structure	213
	Performance of simulations including an inhibiting graph	214
	Performance on an inhibiting graph improves with more genes	215

6.27	Performance of simulations on the PI3K cascade	216 217
6.28	Performance on pathways improves with more genes	218
A.1 A.2	Correlation profiles of removed samples	253 254
A.3 A.4	Replicate excluded samples	<ul><li>255</li><li>256</li></ul>
A.5 A.5	Replicate samples with some excluded	<ul><li>257</li><li>258</li></ul>
D.1 D.2	Synthetic lethal expression profiles of analysed samples	274 276
D.3	Compare mtSLIPT and siRNA genes with correlation	280
D.4 D.5	Compare mtSLIPT and siRNA genes with correlation	280 281
D.6 D.7	Somatic mutation locus	283 284
D.8 D.9	Somatic mutation against PI3K protein	285 286
E.1	Pathway metagene expression profiles	288
E.2 E.3	Expression profiles for constituent genes of PI3K	289 290
E.4 E.5	Expression profiles for estrogen receptor related genes	291 292
F.1	Comparison of SLIPT in stomach to siRNA	296
G.1 G.2	Synthetic lethal expression profiles of stomach samples	305 307
H.1	Synthetic lethal partners across query genes	313
J.1	Synthetic Lethality in the PI3K/AKT Pathway	321
J.2 J.3	Synthetic Lethality in the PI3K/AKT Pathway in Cancer	322 323
J.4 J.5	Synthetic Lethality in the GPCRs	$\frac{324}{325}$
J.6 J.7	Synthetic Lethality in the Translation Elongation	326 327
J.8	Synthetic Lethality in the 3' UTR	328
K.1 K.2	Synthetic Lethality and Vertex Degree	329 330
K.3	Synthetic Lethality and PageRank	331
T. 1	Information centrality distribution	335

M.1	Synthetic Lethality and Heirarchy Score in PI3K
M.2	Heirarchy Score in PI3K against Synthetic Lethality in PI3K 337
M.3	Structure of Synthetic Lethality in PI3K
M.4	Structure of Synthetic Lethality Resampling
N.1	Performance is affected by inhibition in graphs
N.1	Performance on Graph Structures
N.2	Performance is affected by inhibition in graphs
N.2	Performance on Graph Structures
N.3	Performance is affected by inhibition in graphs
N.3	Performance on Graph Structures
N.4	Performance is affected by inhibition in graphs
N.4	Performance on Graph Structures
N.5	Performance is affected by inhibition in graphs
N.5	Performance on Graph Structures
N.6	Performance is affected by inhibition in graphs
N.6	Performance on Graph Structures
0.1	Simulated graph structures
O.2	Simulated graph structures
O.3	Simulated graph structures
0.4	Simulated graph structures
O.5	Simulated graph structures
0.6	Simulated graph structures
O.7	Simulated graph structures
0.8	Simulated graph structures
0.9	Simulated graph structures
O.10	Simulated graph structures
	Simulated graph structures
	Simulated graph structures
O.13	Simulated graph structures
	Simulated graph structures
O.15	Simulated graph structures
P.1	Performance of multivariate normal simulations
P.2	Performance of multivariate normal simulations
P.3	Performance of multivariate normal simulations
P.4	Performance of multivariate normal simulations
Q.1	Performance of multivariate normal simulations
Q.2	Performance of multivariate normal simulations
Q.3	Performance of multivariate normal simulations
Q.4	Performance of multivariate normal simulations
Q.5	Performance of multivariate normal simulations
Q.6	Performance of multivariate normal simulations
Q.7	Performance of multivariate normal simulations
•	Performance of multivariate normal simulations 375

Q.9	Performance of multivariate normal simulations	376
R.1	Performance is affected by inhibition in graphs	377
R.1	Performance is affected by inhibition in graphs	378
R.2		379
R.3	Performance is affected by inhibition in graphs	380
S.1	Performance of multivariate normal simulations	382
S.2		383
S.3		384
S.4		385
T.1	Performance of multivariate normal simulations	387
T.2		388
T.3		389
T.4		390
T.5		391
T.6		392
T.7		393
T.8		394
T.9	Performance of multivariate normal simulations	395
U.1		
U.2	Performance of multivariate normal simulations	398

## List of Tables

1.1	Methods for Predicting Genetic Interactions	38
1.2	Methods for Predicting Synthetic Lethality in Cancer	39
1.3	Methods used by Wu et al. (2014)	40
2.1	Excluded Samples by Batch and Clinical Characteristics	63
2.2	Computers used during Thesis	72
2.3	Linux Utilities and Applications used during Thesis	72
2.4	R Installations used during Thesis	73
2.5	R Packages used during Thesis	73
2.6	R Packages Developed during Thesis	75
4.1	v O I	116
4.2	Pathways for <i>CDH1</i> partners from SLIPT	118
4.3	Pathway composition for clusters of <i>CDH1</i> partners from SLIPT	122
4.4	Pathway composition for CDH1 partners from SLIPT and siRNA screen-	
	ing	131
4.5	Pathways for <i>CDH1</i> partners from SLIPT	135
4.6	Pathways for CDH1 partners from SLIPT and siRNA primary screen .	136
4.7	Candidate synthetic lethal metagenes against CDH1 from SLIPT	145
4.8	Pathways for <i>CDH1</i> partners from SLIPT in stomach cancer	147
4.9	Query synthetic lethal genes with the most SLIPT partners	154
4.10	Pathways for genes with the most SLIPT partners	155
4.11	Pathways for <i>CDH1</i> partners from SLIPT in CCLE	156
4.12	Pathways for <i>CDH1</i> partners from SLIPT in breast CCLE	158
5.1	analysis of variance (ANOVA) for Synthetic Lethality and Vertex Degree	171
5.2	ANOVA for Synthetic Lethality and Information Centrality	173
5.3	ANOVA for Synthetic Lethality and PageRank Centrality	173
5.4	v	177
5.5	Resampling for pathway structure of synthetic lethal detection methods	181
B.1	R Packages used during Thesis	259
C.1 C.2	Comparing SLIPT genes against Secondary siRNA Screen in breast cancer Comparing mtSLIPT genes against Secondary siRNA Screen in breast	
~ .		269
C.3	Comparing SLIPT genes against Secondary siRNA Screen in stomach cancer	269

D.1 D.2 D.3 D.4 D.5 D.6 D.7	Candidate synthetic lethal gene partners of $CDH1$ from mtSLIPT Pathways for $CDH1$ partners from mtSLIPT	271 272 275 277 278 279 282
F.1 F.2 F.3	Synthetic lethal gene partners of <i>CDH1</i> from SLIPT in stomach cancer Pathway composition for clusters of <i>CDH1</i> partners in stomach SLIPT Pathway composition for <i>CDH1</i> partners from SLIPT and siRNA screening	294 295 297
F.4 F.5 F.6	Pathways for <i>CDH1</i> partners from SLIPT in stomach cancer Pathways for <i>CDH1</i> partners from SLIPT in stomach and siRNA screen Candidate synthetic lethal metagenes against <i>CDH1</i> from SLIPT in stomach cancer	298 299 300
G.1 G.2 G.3 G.4 G.5 G.6 G.7	Synthetic lethal gene partners of <i>CDH1</i> from mtSLIPT in stomach cancer Pathways for <i>CDH1</i> partners from mtSLIPT in stomach cancer Pathway composition for clusters of <i>CDH1</i> partners in stomach mtSLIPT Pathway composition for <i>CDH1</i> partners from mtSLIPT and siRNA Pathways for <i>CDH1</i> partners from mtSLIPT in stomach cancer Pathways for <i>CDH1</i> partners from mtSLIPT in stomach and siRNA screen Candidate synthetic lethal metagenes against <i>CDH1</i> from mtSLIPT in stomach cancer	303 306 308 309
H.1 H.2	Query synthetic lethal genes with the most SLIPT partners Pathways for genes with the most SLIPT partners	314 315
I.1 I.2 I.3	Candidate synthetic lethal gene partners of <i>CDH1</i> from SLIPT in CCLE Candidate synthetic lethal gene partners of <i>CDH1</i> from SLIPT in breast CCLE	318 319
I.4 I.5	Pathways for <i>CDH1</i> partners from SLIPT in stomach CCLE Pathways for <i>CDH1</i> partners from SLIPT in breast and stomach CCLE	320 320
K.1 K.2 K.3	ANOVA for Synthetic Lethality and Vertex Degree	332 332 332
L.1	Information centrality for genes and molecules in the Reactome network	334
	ANOVA for Synthetic Lethality and PI3K Hierarchy Resampling for pathway structure of synthetic lethal detection methods	336 338

## Chapter 6

# Simulation and Modeling of Synthetic Lethal Pathways

Simulation and modelling of synthetic lethality in gene expression will be revisited in greater detail in this chapter, building upon the results provided to support the use of Synthetic Lethal Interaction Prediction Tool (SLIPT) in Section 3.3. A simulation procedure for generating simulated data with underlying (known) synthetic lethal partners of a query gene, such as CDH1, was developed (as described in Section 3.2.2) by sampling from a Multivariate normal distribution based on a statistical model of synthetic lethality in expression data (as described in Section 3.2.1). This simulation framework was applied to simulated data (in Section 3.3), including simple correlation structures to assess the statistical performance of the SLIPT methodology and support it's use a computational approach for detecting synthetic lethal candidates from expression data throughout this thesis (in Chapters 4 and 5).

While this basic framework was sufficent to support the use of SLIPT in prior Chapters, further investigations with simulations were conducted to assess the strengths and limitations of the SLIPT methodology, compare it to alternative statistical approaches to synthetic lethal detection, and assess it's performance upon more complex correlation structures. Together these simulation investigations assess the performance of the SLIPT methodology, including on pathway graph structures (such as those discussed in Chapter 5) and determine whether the SLIPT methodology (or similar refined bioinformatics strategies) are statistically rigourous or suitable for wider genomics applications.

These simulation investigations continue to utilise the Multivariate Normal procedure (as applied in Section 3.3) with further refinements. The SLIPT methodology (and it's equivalent  $\chi^2$  test) were applied across a range of parameters (including al-

tering the quantiles for detecting synthetic lethal direction and compared correlation. This was also applied to with query correlated genes (as performed in Section 3.3).

A refined simulation procedure was developed specifically to extend the simulation procedure (described in Section 3.2) to utilise pathway graph structures for the correlation structures of simulated datasets (as described in Section 3.4.2). This methology can be applied to simulated correlation structures across simple graph structures to test specific network modules or use pathway structures based on biological pathways (as discussed in Chapter 5). Thus graph structure and simulation approaches were combined to test whether a gene locus in a pathway affects detection by SLIPT and whether SLIPT performance is affected by pathway structure. The simulation procedure based on graph structures were applied in a computational pipeline across many parameters with high-performance computing (as discussed in Section 2.5.3) and the core simulation functions have been released as a software package for wider use to test bioinformatics and statistical methods on graph structures (as described in Section 3.5.3).

#### 6.1 Comparing methods

Methods were compared ...

### 6.1.1 Performance of SLIPT and $\chi^2$ across Quantiles

Text

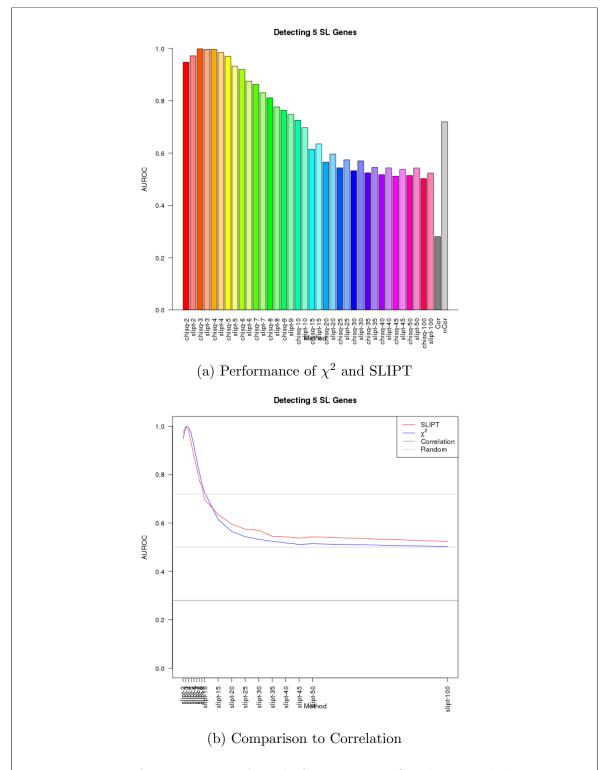


Figure 6.1: **Performance on Graph Structures**. Synthetic Lethal Detection Compared with  $\chi$ .

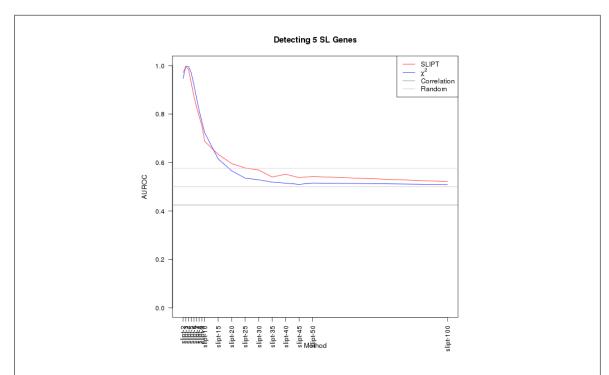


Figure 6.2: **Performance on Graph Structures**. Synthetic Lethal Detection Compared with  $\chi$  with more genes.

#### 6.1.1.1 Correlated Query Genes affects Specificity

Text

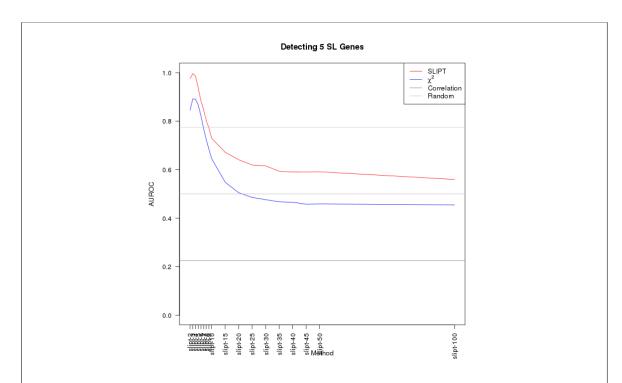


Figure 6.3: **Performance on Graph Structures**. Synthetic Lethal Detection Compared with  $\chi$  with query correlation genes.

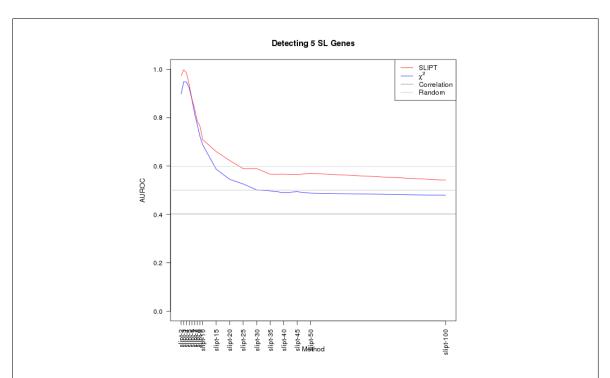


Figure 6.4: **Performance on Graph Structures**. Synthetic Lethal Detection Compared with  $\chi$  with query correlation and more genes.

6.1.2	Correlation as a Synthetic Lethal Detection Stra	$\mathbf{ategy}$
Text		

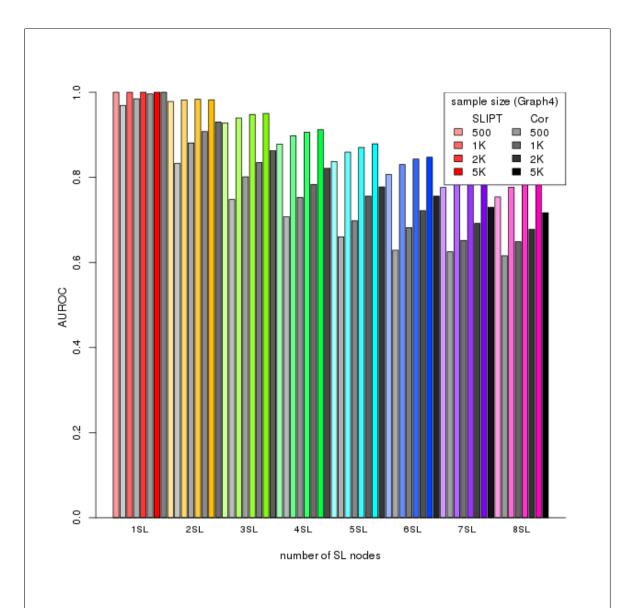


Figure 6.5: **Performance on Graph Structures**. Synthetic Lethal Detection Compared with negative correlation.

#### 6.1.3 Testing for Bimodality with BiSEp

Computer says No

### 6.1.4 Testing Synthetic Lethal Genes with Linear Models

Text

## 6.2 Simulations with Graph Structures

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### 6.2.1 Performance over a Graph Structure

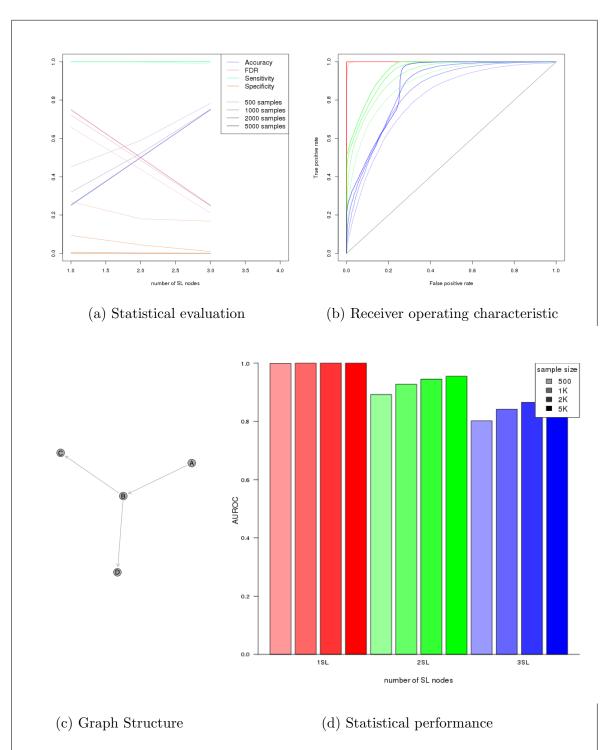
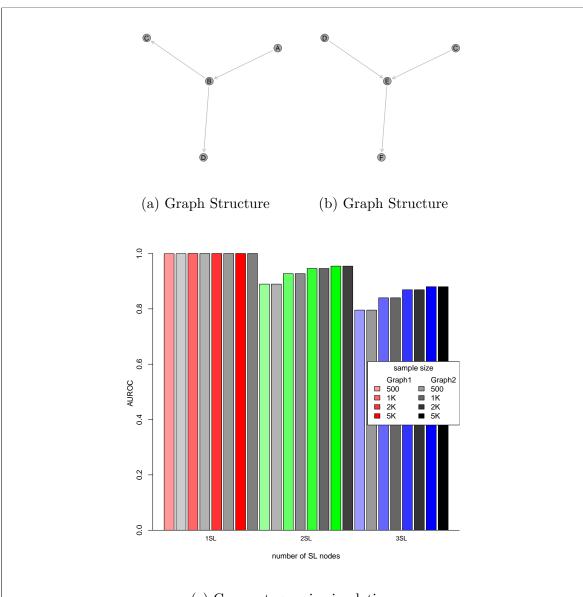


Figure 6.6: **Performance of simulations on a simple graph.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.



(c) Gene category in simulations

Figure 6.7: **Performance of simulations is similar in simple graphs.** The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

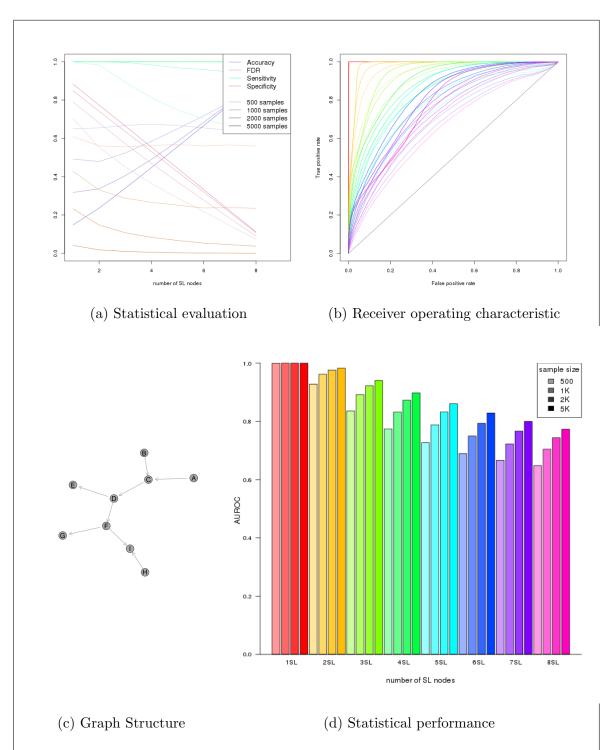


Figure 6.8: **Performance of simulations on a graph.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

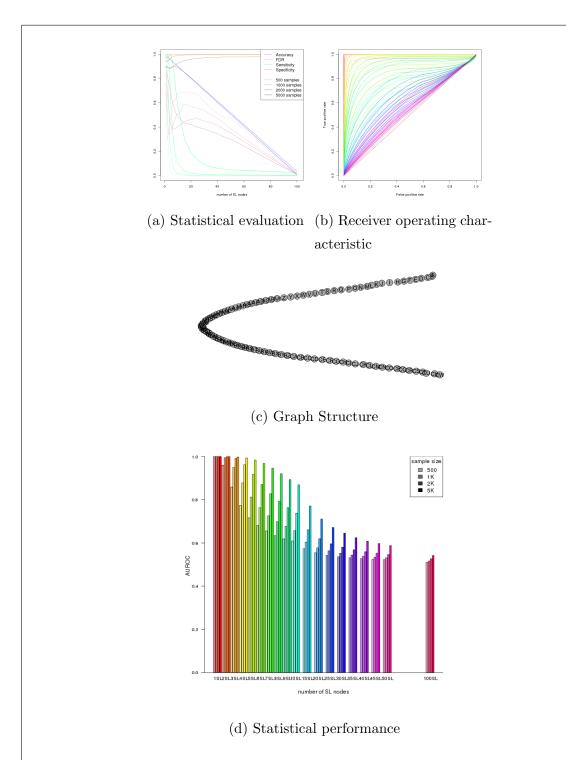


Figure 6.9: **Performance of simulations on a large graph.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

## 6.2.2 Performance with Inhibitions

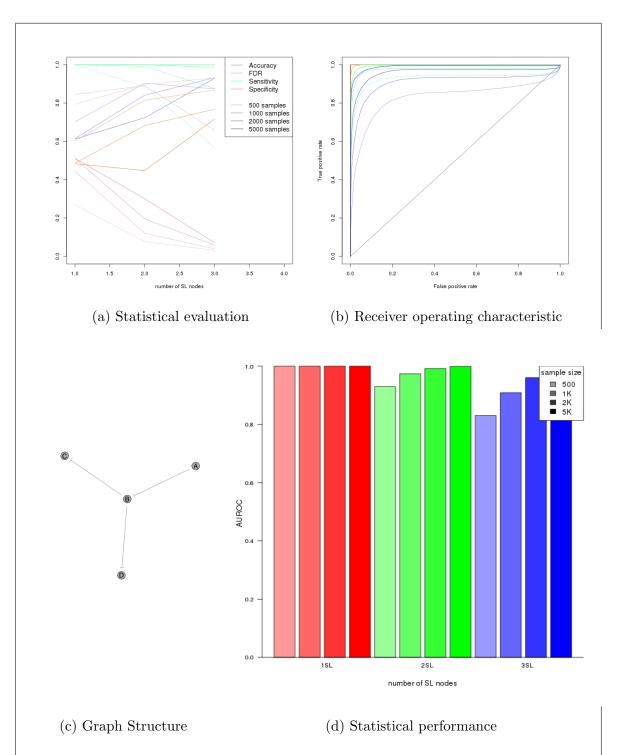
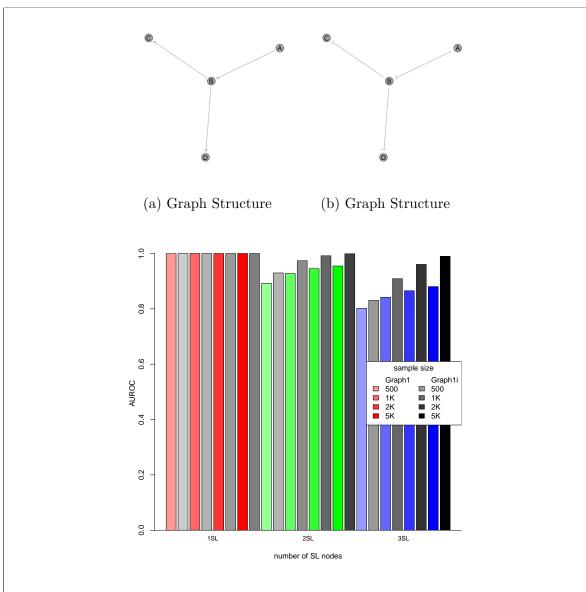
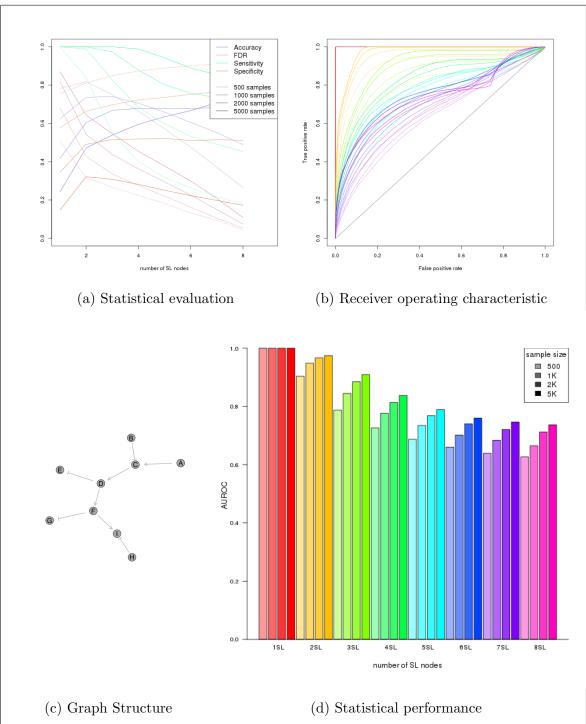


Figure 6.10: Performance of simulations on a simple inhibiting graph. Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.



(c) Gene category in simulations

Figure 6.11: **Performance is higher on a simple inhibiting graph.** The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.



(c) Graph Structure

Figure 6.12: **Performance of simulations on an inhibiting graph.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

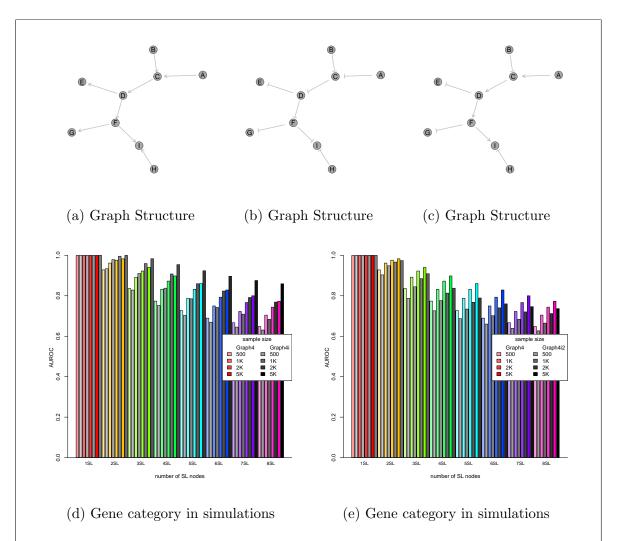


Figure 6.13: **Performance is affected by inhibition in graphs.** The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

#### 6.2.3 Synthetic Lethality across Graph Structures

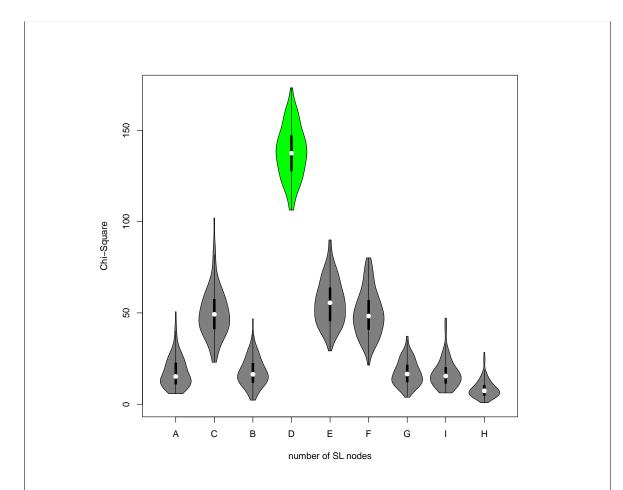


Figure 6.14: **Performance is affected by inhibition in graphs.** The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

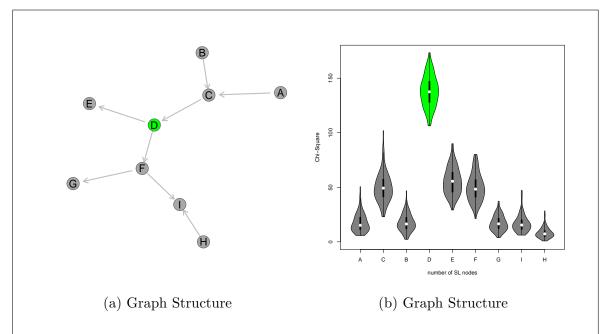


Figure 6.15: **Performance is affected by inhibition in graphs.** The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

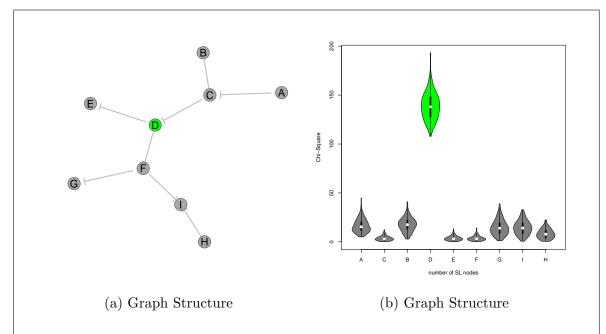


Figure 6.16: **Performance is affected by inhibition in graphs.** The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

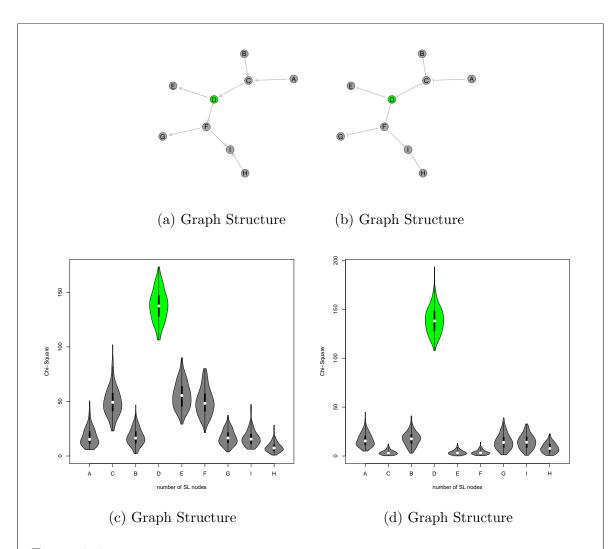


Figure 6.17: **Performance is affected by inhibition in graphs.** The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

6.2.4 Performance with feasible gene numbers (20,000)

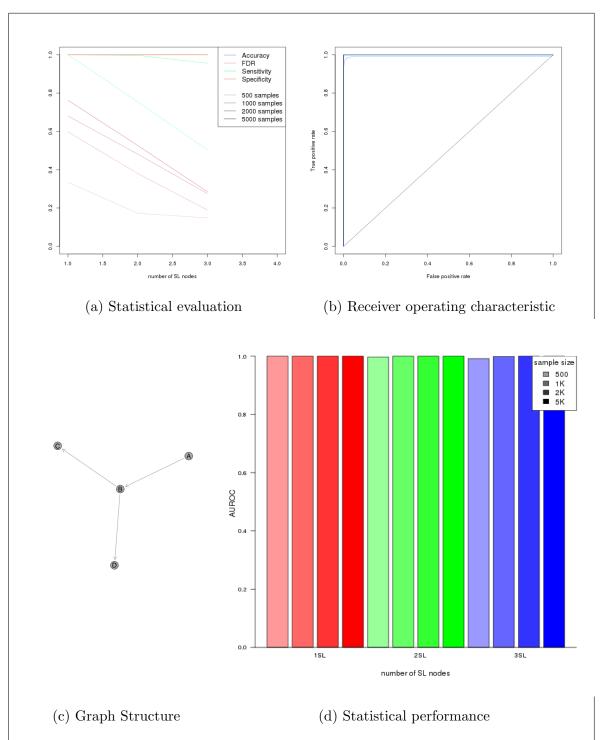


Figure 6.18: **Performance of simulations including a simple graph.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

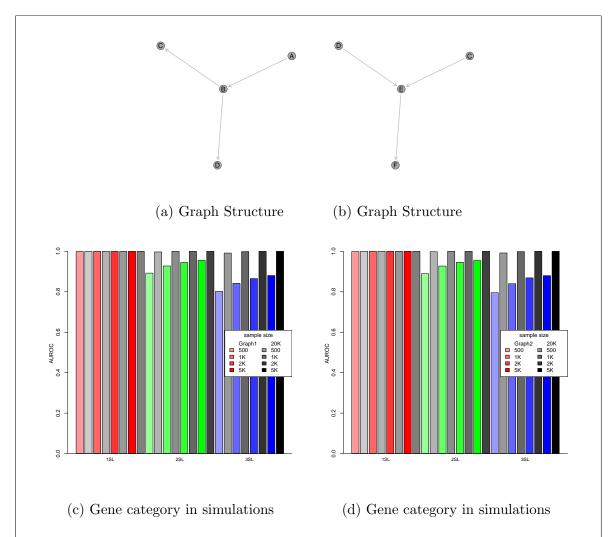


Figure 6.19: Performance on a simple graph improves with more genes. The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

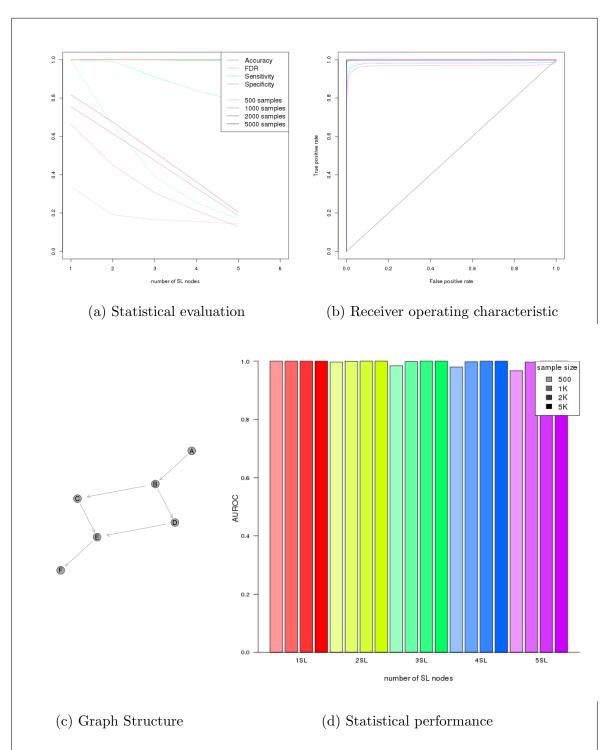


Figure 6.20: Performance of simulations including a graph structure. Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

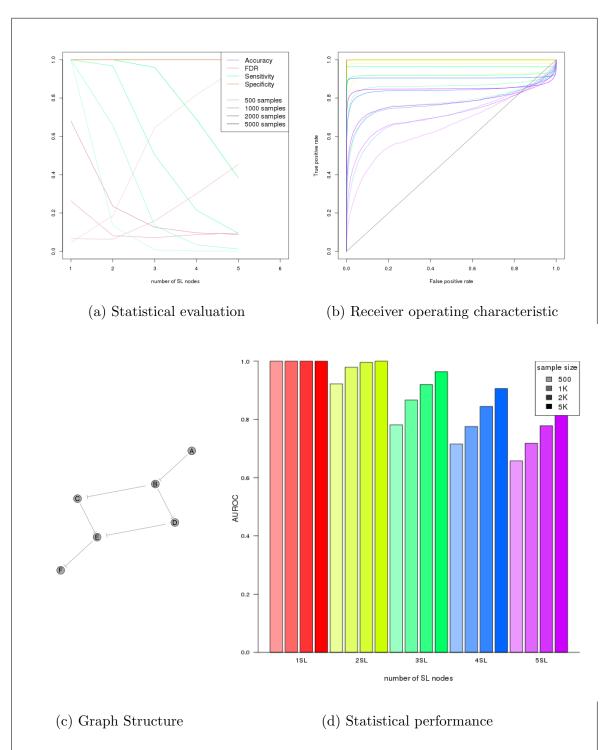


Figure 6.21: **Performance of simulations including an inhbiting graph.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

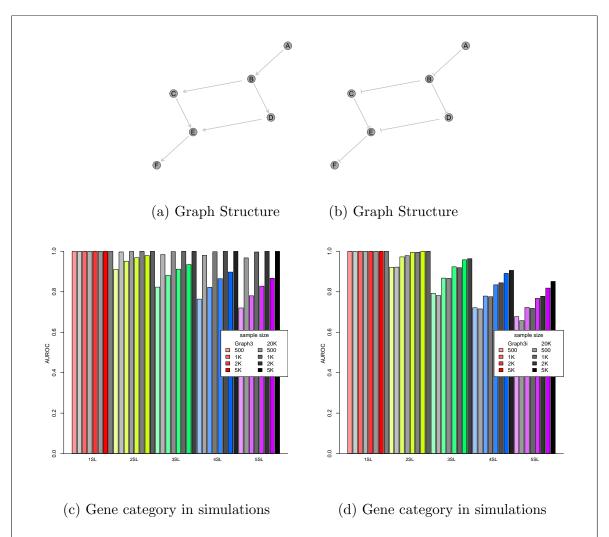


Figure 6.22: Performance on an inhibiting graph improves with more genes. The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

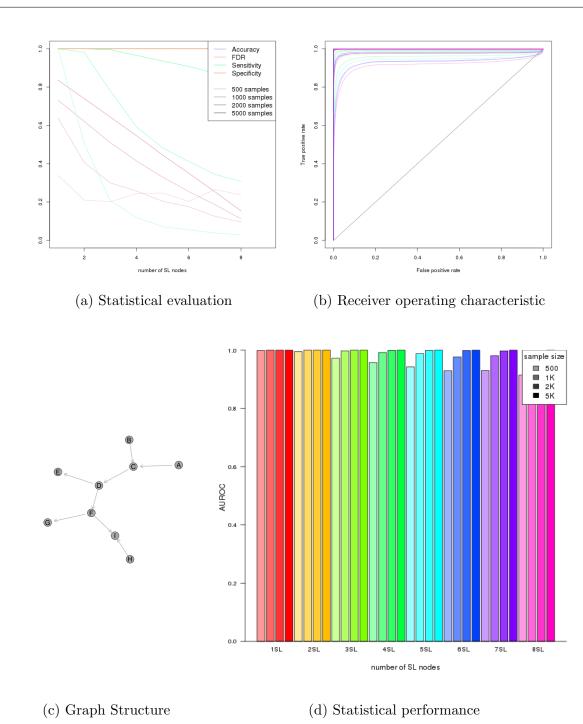


Figure 6.23: Performance of simulations including a graph structure. Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

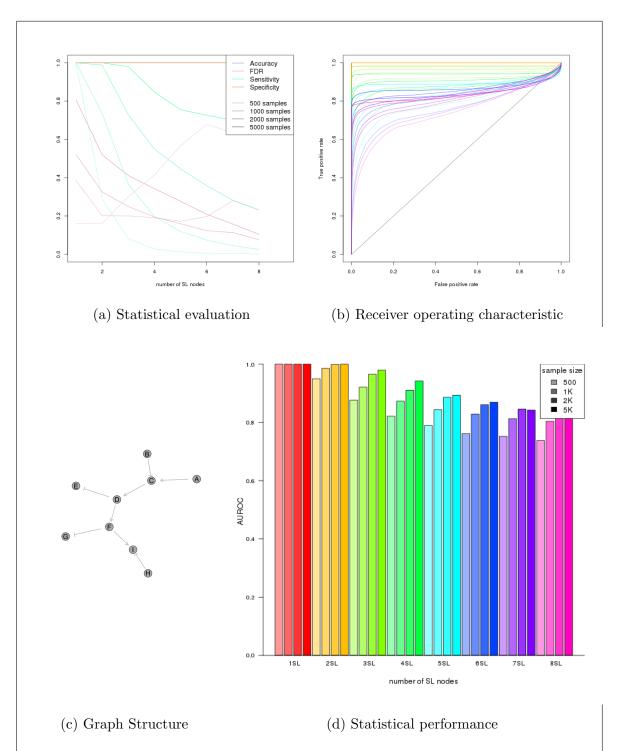


Figure 6.24: **Performance of simulations including an inhibiting graph.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

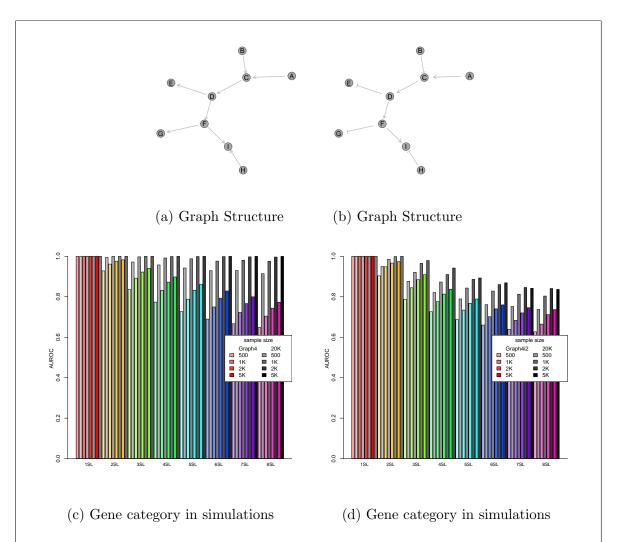


Figure 6.25: Performance on an inhibiting graph improves with more genes. The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

### 6.3 Simulations over pathway-based graphs

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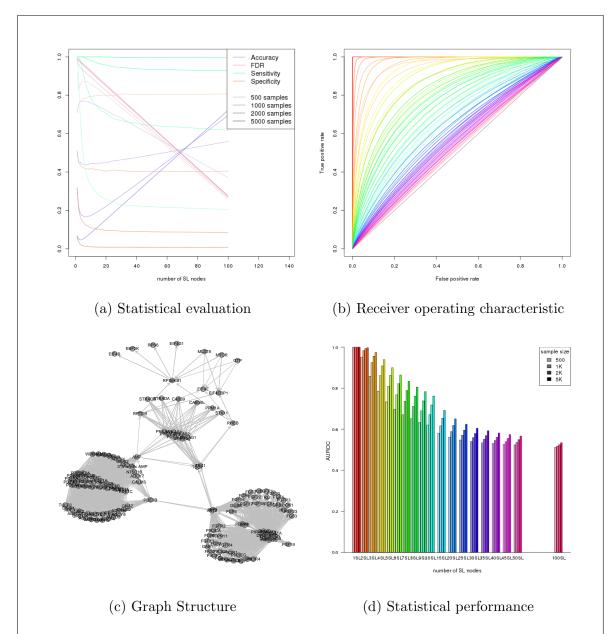


Figure 6.26: **Performance of simulations on the PI3K cascade.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

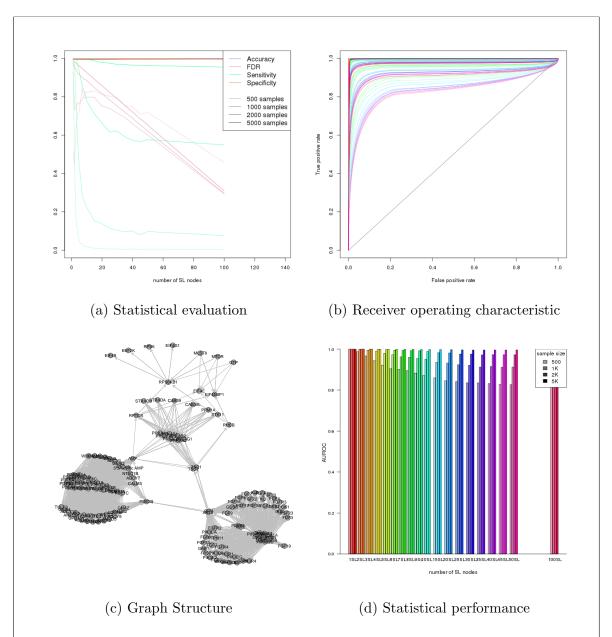


Figure 6.27: **Performance of simulations including the PI3K cascade.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

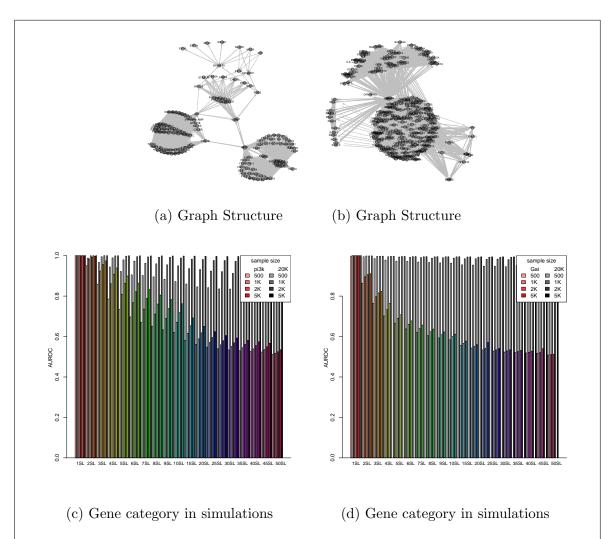


Figure 6.28: Performance on pathways improves with more genes. The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

## 6.4 Discussion

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## 6.5 Summary

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#### Aims

- A Model of Synthetic Lethal Genes in Gene Expression Data
- Comparison of SLIPT to Alternative Approaches
- Simulations of Known Synthetic Lethal Genes within Pathway Networks

#### Summary

- We have designed a straight-forward rational query-based synthetic lethal detection method with the example of application to *CDH1* in cancer gene expression
- I have developed a simulation pipeline to generate continuous gene expression with pathway structure including a procedure to simulate synthetic lethality
- The simulation procedure shows that SLIPT is robust across pathway structures and has desirable performance compared to other statistical techniques

# References

- Aarts, M., Bajrami, I., Herrera-Abreu, M.T., Elliott, R., Brough, R., Ashworth, A., Lord, C.J., and Turner, N.C. (2015) Functional genetic screen identifies increased sensitivity to weel inhibition in cells with defects in fanconi anemia and hr pathways. *Mol Cancer Ther*, **14**(4): 865–76.
- Abeshouse, A., Ahn, J., Akbani, R., Ally, A., Amin, S., Andry, C.D., Annala, M., Aprikian, A., Armenia, J., Arora, A., et al. (2015) The Molecular Taxonomy of Primary Prostate Cancer. Cell, 163(4): 1011–1025.
- Adamski, M.G., Gumann, P., and Baird, A.E. (2014) A method for quantitative analysis of standard and high-throughput qPCR expression data based on input sample quantity. *PLoS ONE*, **9**(8): e103917.
- Adler, D. (2005) vioplot: Violin plot. R package version 0.2.
- Agarwal, S., Deane, C.M., Porter, M.A., and Jones, N.S. (2010) Revisiting date and party hubs: Novel approaches to role assignment in protein interaction networks. *PLoS Comput Biol*, **6**(6): e1000817.
- Agrawal, N., Akbani, R., Aksoy, B.A., Ally, A., Arachchi, H., Asa, S.L., Auman, J.T., Balasundaram, M., Balu, S., Baylin, S.B., et al. (2014) Integrated genomic characterization of papillary thyroid carcinoma. Cell, 159(3): 676–690.
- Akbani, R., Akdemir, K.C., Aksoy, B.A., Albert, M., Ally, A., Amin, S.B., Arachchi, H., Arora, A., Auman, J.T., Ayala, B., et al. (2015) Genomic Classification of Cutaneous Melanoma. Cell, 161(7): 1681–1696.
- Akobeng, A.K. (2007) Understanding diagnostic tests 3: receiver operating characteristic curves. *Acta Pdiatrica*, **96**(5): 644–647.
- American Cancer Society (2017) Genetics and cancer. https://www.cancer.org/cancer/cancer-causes/genetics.html. Accessed: 22/03/2017.

- American Society for Clinical Oncology (ASCO) (2017) The genetics of cancer. http://www.cancer.net/navigating-cancer-care/cancer-basics/genetics-cancer. Accessed: 22/03/2017.
- Anjomshoaa, A., Lin, Y.H., Black, M.A., McCall, J.L., Humar, B., Song, S., Fukuzawa, R., Yoon, H.S., Holzmann, B., Friederichs, J., et al. (2008) Reduced expression of a gene proliferation signature is associated with enhanced malignancy in colon cancer. Br J Cancer, 99(6): 966–973.
- Araki, H., Knapp, C., Tsai, P., and Print, C. (2012) GeneSetDB: A comprehensive meta-database, statistical and visualisation framework for gene set analysis. *FEBS Open Bio*, **2**: 76–82.
- Ashburner, M., Ball, C.A., Blake, J.A., Botstein, D., Butler, H., Cherry, J.M., Davis, A.P., Dolinski, K., Dwight, S.S., Eppig, J.T., et al. (2000) Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. Nat Genet, 25(1): 25–29.
- Ashworth, A. (2008) A synthetic lethal therapeutic approach: poly(adp) ribose polymerase inhibitors for the treatment of cancers deficient in dna double-strand break repair. *J Clin Oncol*, **26**(22): 3785–90.
- Audeh, M.W., Carmichael, J., Penson, R.T., Friedlander, M., Powell, B., Bell-McGuinn, K.M., Scott, C., Weitzel, J.N., Oaknin, A., Loman, N., et al. (2010) Oral poly(adp-ribose) polymerase inhibitor olaparib in patients with *BRCA1* or *BRCA2* mutations and recurrent ovarian cancer: a proof-of-concept trial. *Lancet*, **376**(9737): 245–51.
- Babyak, M.A. (2004) What you see may not be what you get: a brief, nontechnical introduction to overfitting in regression-type models. *Psychosom Med*, **66**(3): 411–21.
- Bamford, S., Dawson, E., Forbes, S., Clements, J., Pettett, R., Dogan, A., Flanagan, A., Teague, J., Futreal, P.A., Stratton, M.R., et al. (2004) The COSMIC (Catalogue of Somatic Mutations in Cancer) database and website. Br J Cancer, 91(2): 355–358.
- Barabási, A.L. and Albert, R. (1999) Emergence of scaling in random networks. *Science*, **286**(5439): 509–12.

- Barabási, A.L. and Oltvai, Z.N. (2004) Network biology: understanding the cell's functional organization. *Nat Rev Genet*, **5**(2): 101–13.
- Barrat, A. and Weigt, M. (2000) On the properties of small-world network models. The European Physical Journal B - Condensed Matter and Complex Systems, 13(3): 547–560.
- Barretina, J., Caponigro, G., Stransky, N., Venkatesan, K., Margolin, A.A., Kim, S., Wilson, C.J., Lehar, J., Kryukov, G.V., Sonkin, D., et al. (2012) The Cancer Cell Line Encyclopedia enables predictive modelling of anticancer drug sensitivity. Nature, 483(7391): 603–607.
- Barry, W.T. (2016) safe: Significance Analysis of Function and Expression. R package version 3.14.0.
- Baryshnikova, A., Costanzo, M., Dixon, S., Vizeacoumar, F.J., Myers, C.L., Andrews, B., and Boone, C. (2010a) Synthetic genetic array (sga) analysis in saccharomyces cerevisiae and schizosaccharomyces pombe. *Methods Enzymol*, **470**: 145–79.
- Baryshnikova, A., Costanzo, M., Kim, Y., Ding, H., Koh, J., Toufighi, K., Youn, J.Y., Ou, J., San Luis, B.J., Bandyopadhyay, S., et al. (2010b) Quantitative analysis of fitness and genetic interactions in yeast on a genome scale. Nat Meth, 7(12): 1017–1024.
- Bass, A.J., Thorsson, V., Shmulevich, I., Reynolds, S.M., Miller, M., Bernard, B., Hinoue, T., Laird, P.W., Curtis, C., Shen, H., et al. (2014) Comprehensive molecular characterization of gastric adenocarcinoma. Nature, 513(7517): 202–209.
- Bates, D. and Maechler, M. (2016) Matrix: Sparse and Dense Matrix Classes and Methods. R package version 1.2-7.1.
- Bateson, W. and Mendel, G. (1909) Mendel's principles of heredity, by W. Bateson. University Press, Cambridge [Eng.].
- Beck, T.F., Mullikin, J.C., and Biesecker, L.G. (2016) Systematic Evaluation of Sanger Validation of Next-Generation Sequencing Variants. *Clin Chem*, **62**(4): 647–654.
- Becker, K.F., Atkinson, M.J., Reich, U., Becker, I., Nekarda, H., Siewert, J.R., and Hfler, H. (1994) E-cadherin gene mutations provide clues to diffuse type gastric carcinomas. *Cancer Research*, **54**(14): 3845–3852.

- Bell, D., Berchuck, A., Birrer, M., Chien, J., Cramer, D., Dao, F., Dhir, R., DiSaia, P., Gabra, H., Glenn, P., et al. (2011) Integrated genomic analyses of ovarian carcinoma. Nature, 474(7353): 609–615.
- Benjamini, Y. and Hochberg, Y. (1995) Controlling the false discovery rate: A practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society Series B (Methodological)*, **57**(1): 289–300.
- Berx, G., Cleton-Jansen, A.M., Nollet, F., de Leeuw, W.J., van de Vijver, M., Cornelisse, C., and van Roy, F. (1995) E-cadherin is a tumour/invasion suppressor gene mutated in human lobular breast cancers. *EMBO J*, **14**(24): 6107–15.
- Berx, G., Cleton-Jansen, A.M., Strumane, K., de Leeuw, W.J., Nollet, F., van Roy, F., and Cornelisse, C. (1996) E-cadherin is inactivated in a majority of invasive human lobular breast cancers by truncation mutations throughout its extracellular domain. *Oncogene*, **13**(9): 1919–25.
- Berx, G. and van Roy, F. (2009) Involvement of members of the cadherin superfamily in cancer. *Cold Spring Harb Perspect Biol*, **1**: a003129.
- Bitler, B.G., Aird, K.M., Garipov, A., Li, H., Amatangelo, M., Kossenkov, A.V., Schultz, D.C., Liu, Q., Shih Ie, M., Conejo-Garcia, J.R., *et al.* (2015) Synthetic lethality by targeting ezh2 methyltransferase activity in arid1a-mutated cancers. *Nat Med*, **21**(3): 231–8.
- Blake, J.A., Christie, K.R., Dolan, M.E., Drabkin, H.J., Hill, D.P., Ni, L., Sitnikov, D., Burgess, S., Buza, T., Gresham, C., et al. (2015) Gene Ontology Consortium: going forward. Nucleic Acids Res, 43(Database issue): D1049–1056.
- Boettcher, M., Lawson, A., Ladenburger, V., Fredebohm, J., Wolf, J., Hoheisel, J.D., Frezza, C., and Shlomi, T. (2014) High throughput synthetic lethality screen reveals a tumorigenic role of adenylate cyclase in fumarate hydratase-deficient cancer cells. *BMC Genomics*, **15**: 158.
- Boone, C., Bussey, H., and Andrews, B.J. (2007) Exploring genetic interactions and networks with yeast. *Nat Rev Genet*, **8**(6): 437–49.
- Borgatti, S.P. (2005) Centrality and network flow. Social Networks, 27(1): 55-71.
- Boucher, B. and Jenna, S. (2013) Genetic interaction networks: better understand to better predict. *Front Genet*, **4**: 290.

- Breiman, L. (2001) Random forests. Machine Learning, 45(1): 5–32.
- Brin, S. and Page, L. (1998) The anatomy of a large-scale hypertextual web search engine. Computer Networks and ISDN Systems, **30**(1): 107 117.
- Bryant, H.E., Schultz, N., Thomas, H.D., Parker, K.M., Flower, D., Lopez, E., Kyle, S., Meuth, M., Curtin, N.J., and Helleday, T. (2005) Specific killing of *BRCA2*-deficient tumours with inhibitors of poly*adpribose* polymerase. *Nature*, **434**(7035): 913–7.
- Burk, R.D., Chen, Z., Saller, C., Tarvin, K., Carvalho, A.L., Scapulatempo-Neto, C., Silveira, H.C., Fregnani, J.H., Creighton, C.J., Anderson, M.L., et al. (2017) Integrated genomic and molecular characterization of cervical cancer. Nature, **543**(7645): 378–384.
- Bussey, H., Andrews, B., and Boone, C. (2006) From worm genetic networks to complex human diseases. *Nat Genet*, **38**(8): 862–3.
- Butland, G., Babu, M., Diaz-Mejia, J.J., Bohdana, F., Phanse, S., Gold, B., Yang, W., Li, J., Gagarinova, A.G., Pogoutse, O., et al. (2008) esga: E. coli synthetic genetic array analysis. Nat Methods, 5(9): 789–95.
- Cancer Research UK (2017) Family history and cancer genes. http://www.cancerresearchuk.org/about-cancer/causes-of-cancer/inherited-cancer-genes-and-increased-cancer-risk/family-history-and-inherited-cancer-genes. Accessed: 22/03/2017.
- Cancer Cell Line Encyclopedia (CCLE) (2014) Broad-Novartis Cancer Cell Line Encyclopedia. http://www.broadinstitute.org/ccle. Accessed: 07/11/2014.
- cBioPortal for Cancer Genomics (cBioPortal) (2017) cBioPortal for Cancer Genomics. http://www.cbioportal.org/. Accessed: 26/03/2017.
- Cerami, E.G., Gross, B.E., Demir, E., Rodchenkov, I., Babur, O., Anwar, N., Schultz, N., Bader, G.D., and Sander, C. (2011) Pathway Commons, a web resource for biological pathway data. *Nucleic Acids Res*, 39(Database issue): D685–690.
- Chen, A., Beetham, H., Black, M.A., Priya, R., Telford, B.J., Guest, J., Wiggins, G.A.R., Godwin, T.D., Yap, A.S., and Guilford, P.J. (2014) E-cadherin loss alters cytoskeletal organization and adhesion in non-malignant breast cells but is insufficient to induce an epithelial-mesenchymal transition. *BMC Cancer*, **14**(1): 552.

- Chen, K., Yang, D., Li, X., Sun, B., Song, F., Cao, W., Brat, D.J., Gao, Z., Li, H., Liang, H., et al. (2015) Mutational landscape of gastric adenocarcinoma in Chinese: implications for prognosis and therapy. Proc Natl Acad Sci USA, 112(4): 1107–1112.
- Chen, S. and Parmigiani, G. (2007) Meta-analysis of BRCA1 and BRCA2 penetrance. J Clin Oncol, 25(11): 1329–1333.
- Chen, X. and Tompa, M. (2010) Comparative assessment of methods for aligning multiple genome sequences. *Nat Biotechnol*, **28**(6): 567–572.
- Cherniack, A.D., Shen, H., Walter, V., Stewart, C., Murray, B.A., Bowlby, R., Hu, X., Ling, S., Soslow, R.A., Broaddus, R.R., et al. (2017) Integrated Molecular Characterization of Uterine Carcinosarcoma. Cancer Cell, 31(3): 411–423.
- Chipman, K. and Singh, A. (2009) Predicting genetic interactions with random walks on biological networks. BMC Bioinformatics,  $\mathbf{10}(1)$ : 17.
- Christofori, G. and Semb, H. (1999) The role of the cell-adhesion molecule E-cadherin as a tumour-suppressor gene. *Trends in Biochemical Sciences*, **24**(2): 73 76.
- Ciriello, G., Gatza, M.L., Beck, A.H., Wilkerson, M.D., Rhie, S.K., Pastore, A., Zhang, H., McLellan, M., Yau, C., Kandoth, C., et al. (2015) Comprehensive Molecular Portraits of Invasive Lobular Breast Cancer. Cell, 163(2): 506–519.
- Clark, M.J. (2004) Endogenous Regulator of G Protein Signaling Proteins Suppress G o-Dependent -Opioid Agonist-Mediated Adenylyl Cyclase Supersensitization.

  Journal of Pharmacology and Experimental Therapeutics, 310(1): 215–222.
- Clough, E. and Barrett, T. (2016) The Gene Expression Omnibus Database. *Methods Mol Biol*, **1418**: 93–110.
- Collingridge, D.S. (2013) A primer on quantitized data analysis and permutation testing. *Journal of Mixed Methods Research*, **7**(1): 81–97.
- Collins, F.S. and Barker, A.D. (2007) Mapping the cancer genome. Pinpointing the genes involved in cancer will help chart a new course across the complex landscape of human malignancies. *Sci Am*, **296**(3): 50–57.
- Collins, F.S., Morgan, M., and Patrinos, A. (2003) The Human Genome Project: lessons from large-scale biology. *Science*, **300**(5617): 286–290.

- Collisson, E., Campbell, J., Brooks, A., Berger, A., Lee, W., Chmielecki, J., Beer, D., Cope, L., Creighton, C., Danilova, L., et al. (2014) Comprehensive molecular profiling of lung adenocarcinoma. Nature, 511(7511): 543–550.
- Corcoran, R.B., Ebi, H., Turke, A.B., Coffee, E.M., Nishino, M., Cogdill, A.P., Brown, R.D., Della Pelle, P., Dias-Santagata, D., Hung, K.E., et al. (2012) Egfr-mediated reactivation of mapk signaling contributes to insensitivity of BRAF-mutant colorectal cancers to raf inhibition with vemurafenib. Cancer Discovery, 2(3): 227–235.
- Costanzo, M., Baryshnikova, A., Bellay, J., Kim, Y., Spear, E.D., Sevier, C.S., Ding, H., Koh, J.L., Toufighi, K., Mostafavi, S., et al. (2010) The genetic landscape of a cell. Science, 327(5964): 425–31.
- Costanzo, M., Baryshnikova, A., Myers, C.L., Andrews, B., and Boone, C. (2011) Charting the genetic interaction map of a cell. *Curr Opin Biotechnol*, **22**(1): 66–74.
- Creighton, C.J., Morgan, M., Gunaratne, P.H., Wheeler, D.A., Gibbs, R.A., Robertson, A., Chu, A., Beroukhim, R., Cibulskis, K., Signoretti, S., et al. (2013) Comprehensive molecular characterization of clear cell renal cell carcinoma. Nature, 499(7456): 43–49.
- Croft, D., Mundo, A.F., Haw, R., Milacic, M., Weiser, J., Wu, G., Caudy, M., Garapati, P., Gillespie, M., Kamdar, M.R., et al. (2014) The Reactome pathway knowledge-base. Nucleic Acids Res, 42(database issue): D472D477.
- Crunkhorn, S. (2014) Cancer: Predicting synthetic lethal interactions. *Nat Rev Drug Discov*, **13**(11): 812.
- Csardi, G. and Nepusz, T. (2006) The igraph software package for complex network research. *InterJournal*, **Complex Systems**: 1695.
- Curtis, C., Shah, S.P., Chin, S.F., Turashvili, G., Rueda, O.M., Dunning, M.J., Speed, D., Lynch, A.G., Samarajiwa, S., Yuan, Y., et al. (2012) The genomic and transcriptomic architecture of 2,000 breast tumours reveals novel subgroups. Nature, 486(7403): 346–352.
- Dai, X., Li, T., Bai, Z., Yang, Y., Liu, X., Zhan, J., and Shi, B. (2015) Breast cancer intrinsic subtype classification, clinical use and future trends. *Am J Cancer Res*, **5**(10): 2929–2943.

- Davierwala, A.P., Haynes, J., Li, Z., Brost, R.L., Robinson, M.D., Yu, L., Mnaimneh, S., Ding, H., Zhu, H., Chen, Y., et al. (2005) The synthetic genetic interaction spectrum of essential genes. Nat Genet, 37(10): 1147–1152.
- De Leeuw, W.J., Berx, G., Vos, C.B., Peterse, J.L., Van de Vijver, M.J., Litvinov, S., Van Roy, F., Cornelisse, C.J., and Cleton-Jansen, A.M. (1997) Simultaneous loss of E-cadherin and catenins in invasive lobular breast cancer and lobular carcinoma in situ. *J Pathol*, **183**(4): 404–11.
- Demir, E., Babur, O., Rodchenkov, I., Aksoy, B.A., Fukuda, K.I., Gross, B., Sumer, O.S., Bader, G.D., and Sander, C. (2013) Using biological pathway data with Paxtools. *PLoS Comput Biol*, **9**(9): e1003194.
- Deshpande, R., Asiedu, M.K., Klebig, M., Sutor, S., Kuzmin, E., Nelson, J., Piotrowski, J., Shin, S.H., Yoshida, M., Costanzo, M., et al. (2013) A comparative genomic approach for identifying synthetic lethal interactions in human cancer. Cancer Res, 73(20): 6128–36.
- Dickson, D. (1999) Wellcome funds cancer database. Nature, 401(6755): 729.
- Dienstmann, R. and Tabernero, J. (2011) *BRAF* as a target for cancer therapy. *Anti*cancer Agents Med Chem, **11**(3): 285–95.
- Dijkstra, E.W. (1959) A note on two problems in connexion with graphs. *Numerische Mathematik*, **1**(1): 269–271.
- Dixon, S.J., Andrews, B.J., and Boone, C. (2009) Exploring the conservation of synthetic lethal genetic interaction networks. *Commun Integr Biol*, **2**(2): 78–81.
- Dixon, S.J., Fedyshyn, Y., Koh, J.L., Prasad, T.S., Chahwan, C., Chua, G., Toufighi, K., Baryshnikova, A., Hayles, J., Hoe, K.L., et al. (2008) Significant conservation of synthetic lethal genetic interaction networks between distantly related eukaryotes. Proc Natl Acad Sci U S A, 105(43): 16653–8.
- Dorogovtsev, S.N. and Mendes, J.F. (2003) Evolution of networks: From biological nets to the Internet and WWW. Oxford University Press, USA.
- Erdős, P. and Rényi, A. (1959) On random graphs I. Publ Math Debrecen, 6: 290–297.
- Erdős, P. and Rényi, A. (1960) On the evolution of random graphs. In *Publ. Math. Inst. Hung. Acad. Sci*, volume 5, 17–61.

- Eroles, P., Bosch, A., Perez-Fidalgo, J.A., and Lluch, A. (2012) Molecular biology in breast cancer: intrinsic subtypes and signaling pathways. *Cancer Treat Rev*, **38**(6): 698–707.
- Ezkurdia, I., Juan, D., Rodriguez, J.M., Frankish, A., Diekhans, M., Harrow, J., Vazquez, J., Valencia, A., and Tress, M.L. (2014) Multiple evidence strands suggest that there may be as few as 19 000 human protein-coding genes. *Human Molecular Genetics*, **23**(22): 5866.
- Farmer, H., McCabe, N., Lord, C.J., Tutt, A.N., Johnson, D.A., Richardson, T.B., Santarosa, M., Dillon, K.J., Hickson, I., Knights, C., et al. (2005) Targeting the dna repair defect in BRCA mutant cells as a therapeutic strategy. *Nature*, **434**(7035): 917–21.
- Fawcett, T. (2006) An introduction to ROC analysis. *Pattern Recognition Letters*, **27**(8): 861 874. {ROC} Analysis in Pattern Recognition.
- Fece de la Cruz, F., Gapp, B.V., and Nijman, S.M. (2015) Synthetic lethal vulnerabilities of cancer. *Annu Rev Pharmacol Toxicol*, **55**: 513–531.
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D.M., Forman, D., and Bray, F. (2015) Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*, **136**(5): E359–386.
- Fisher, R.A. (1919) Xv.the correlation between relatives on the supposition of mendelian inheritance. Earth and Environmental Science Transactions of the Royal Society of Edinburgh, **52**(02): 399–433.
- Fong, P.C., Boss, D.S., Yap, T.A., Tutt, A., Wu, P., Mergui-Roelvink, M., Mortimer, P., Swaisland, H., Lau, A., O'Connor, M.J., et al. (2009) Inhibition of poly(adpribose) polymerase in tumors from BRCA mutation carriers. N Engl J Med, 361(2): 123–34.
- Fong, P.C., Yap, T.A., Boss, D.S., Carden, C.P., Mergui-Roelvink, M., Gourley, C., De Greve, J., Lubinski, J., Shanley, S., Messiou, C., et al. (2010) Poly(adp)-ribose polymerase inhibition: frequent durable responses in BRCA carrier ovarian cancer correlating with platinum-free interval. J Clin Oncol, 28(15): 2512–9.

- Forbes, S.A., Beare, D., Gunasekaran, P., Leung, K., Bindal, N., Boutselakis, H., Ding, M., Bamford, S., Cole, C., Ward, S., et al. (2015) COSMIC: exploring the world's knowledge of somatic mutations in human cancer. Nucleic Acids Res, 43(Database issue): D805–811.
- Fraser, A. (2004) Towards full employment: using RNAi to find roles for the redundant. Oncogene, 23(51): 8346–52.
- Futreal, P.A., Coin, L., Marshall, M., Down, T., Hubbard, T., Wooster, R., Rahman, N., and Stratton, M.R. (2004) A census of human cancer genes. *Nat Rev Cancer*, 4(3): 177–183.
- Futreal, P.A., Kasprzyk, A., Birney, E., Mullikin, J.C., Wooster, R., and Stratton, M.R. (2001) Cancer and genomics. *Nature*, **409**(6822): 850–852.
- Gao, B. and Roux, P.P. (2015) Translational control by oncogenic signaling pathways. Biochimica et Biophysica Acta, 1849(7): 753–65.
- Gatza, M.L., Kung, H.N., Blackwell, K.L., Dewhirst, M.W., Marks, J.R., and Chi, J.T. (2011) Analysis of tumor environmental response and oncogenic pathway activation identifies distinct basal and luminal features in HER2-related breast tumor subtypes. *Breast Cancer Res*, **13**(3): R62.
- Gatza, M.L., Lucas, J.E., Barry, W.T., Kim, J.W., Wang, Q., Crawford, M.D., Datto, M.B., Kelley, M., Mathey-Prevot, B., Potti, A., et al. (2010) A pathway-based classification of human breast cancer. Proc Natl Acad Sci USA, 107(15): 6994–6999.
- Gatza, M.L., Silva, G.O., Parker, J.S., Fan, C., and Perou, C.M. (2014) An integrated genomics approach identifies drivers of proliferation in luminal-subtype human breast cancer. *Nat Genet*, **46**(10): 1051–1059.
- Gentleman, R.C., Carey, V.J., Bates, D.M., Bolstad, B., Dettling, M., Dudoit, S., Ellis, B., Gautier, L., Ge, Y., Gentry, J., et al. (2004) Bioconductor: open software development for computational biology and bioinformatics. Genome Biol, 5(10): R80.
- Genz, A. and Bretz, F. (2009) Computation of multivariate normal and t probabilities. In *Lecture Notes in Statistics*, volume 195. Springer-Verlag, Heidelberg.
- Genz, A., Bretz, F., Miwa, T., Mi, X., Leisch, F., Scheipl, F., and Hothorn, T. (2016) mvtnorm: Multivariate Normal and t Distributions. R package version 1.0-5. URL.

- Gilbert, W. and Maxam, A. (1973) The nucleotide sequence of the lac operator. *Proceedings of the National Academy of Sciences*, **70**(12): 3581–3584.
- Git, A., Dvinge, H., Salmon-Divon, M., Osborne, M., Kutter, C., Hadfield, J., Bertone, P., and Caldas, C. (2010) Systematic comparison of microarray profiling, real-time PCR, and next-generation sequencing technologies for measuring differential microRNA expression. RNA, 16(5): 991–1006.
- Globus (Globus) (2017) Research data management simplified. https://www.globus.org/. Accessed: 25/03/2017.
- Graziano, F., Humar, B., and Guilford, P. (2003) The role of the E-cadherin gene (*CDH1*) in diffuse gastric cancer susceptibility: from the laboratory to clinical practice. *Annals of Oncology*, **14**(12): 1705–1713.
- Güell, O., Sagus, F., and Serrano, M. (2014) Essential plasticity and redundancy of metabolism unveiled by synthetic lethality analysis. *PLoS Comput Biol*, **10**(5): e1003637.
- Guilford, P. (1999) E-cadherin downregulation in cancer: fuel on the fire? *Molecular Medicine Today*, **5**(4): 172 177.
- Guilford, P., Hopkins, J., Harraway, J., McLeod, M., McLeod, N., Harawira, P., Taite, H., Scoular, R., Miller, A., and Reeve, A.E. (1998) E-cadherin germline mutations in familial gastric cancer. *Nature*, 392(6674): 402–5.
- Guilford, P., Humar, B., and Blair, V. (2010) Hereditary diffuse gastric cancer: translation of *CDH1* germline mutations into clinical practice. *Gastric Cancer*, **13**(1): 1–10.
- Guilford, P.J., Hopkins, J.B., Grady, W.M., Markowitz, S.D., Willis, J., Lynch, H., Rajput, A., Wiesner, G.L., Lindor, N.M., Burgart, L.J., *et al.* (1999) E-cadherin germline mutations define an inherited cancer syndrome dominated by diffuse gastric cancer. *Hum Mutat*, **14**(3): 249–55.
- Guo, J., Liu, H., and Zheng, J. (2016) SynLethDB: synthetic lethality database toward discovery of selective and sensitive anticancer drug targets. *Nucleic Acids Res*, 44(D1): D1011–1017.
- Hajian-Tilaki, K. (2013) Receiver Operating Characteristic (ROC) Curve Analysis for Medical Diagnostic Test Evaluation. *Caspian J Intern Med*, 4(2): 627–635.

- Hall, M., Frank, E., Holmes, G., Pfahringer, B., Reutemann, P., and Witten, I.H. (2009) The weka data mining software: an update. *SIGKDD Explor Newsl*, **11**(1): 10–18.
- Hammerman, P.S., Lawrence, M.S., Voet, D., Jing, R., Cibulskis, K., Sivachenko, A., Stojanov, P., McKenna, A., Lander, E.S., Gabriel, S., et al. (2012) Comprehensive genomic characterization of squamous cell lung cancers. Nature, 489(7417): 519–525.
- Han, J.D.J., Bertin, N., Hao, T., Goldberg, D.S., Berriz, G.F., Zhang, L.V., Dupuy, D., Walhout, A.J.M., Cusick, M.E., Roth, F.P., et al. (2004) Evidence for dynamically organized modularity in the yeast protein-protein interaction network. Nature, 430(6995): 88–93.
- Hanahan, D. and Weinberg, R.A. (2000) The hallmarks of cancer. Cell, 100(1): 57–70.
- Hanahan, D. and Weinberg, R.A. (2011) Hallmarks of cancer: the next generation. *Cell*, **144**(5): 646–674.
- Hanna, S. (2003) Cancer incidence in new zealand (2003-2007). In D. Forman, D. Bray
  F Brewster, C. Gombe Mbalawa, B. Kohler, M. Piñeros, E. Steliarova-Foucher,
  R. Swaminathan, and J. Ferlay (editors), Cancer Incidence in Five Continents,
  volume X, 902-907. International Agency for Research on Cancer, Lyon, France.
  Electronic version http://ci5.iarc.fr Accessed 22/03/2017.
- Heiskanen, M., Bian, X., Swan, D., and Basu, A. (2014) caArray microarray database in the cancer biomedical informatics grid<sup>TM</sup> (caBIG<sup>TM</sup>). Cancer Research, **67**(9 Supplement): 3712–3712.
- Heiskanen, M.A. and Aittokallio, T. (2012) Mining high-throughput screens for cancer drug targets-lessons from yeast chemical-genomic profiling and synthetic lethality. Wiley Interdisciplinary Reviews: Data Mining and Knowledge Discovery, 2(3): 263–272.
- Hell, P. (1976) Graphs with given neighbourhoods i. problémes combinatorics at theorie des graphes. *Proc Coil Int CNRS*, *Orsay*, **260**: 219–223.
- Herschkowitz, J.I., Simin, K., Weigman, V.J., Mikaelian, I., Usary, J., Hu, Z., Rasmussen, K.E., Jones, L.P., Assefnia, S., Chandrasekharan, S., et al. (2007) Identifi-

- cation of conserved gene expression features between murine mammary carcinoma models and human breast tumors. *Genome Biol*, **8**(5): R76.
- Hillenmeyer, M.E. (2008) The chemical genomic portrait of yeast: uncovering a phenotype for all genes. *Science*, **320**: 362–365.
- Hoadley, K.A., Yau, C., Wolf, D.M., Cherniack, A.D., Tamborero, D., Ng, S., Leiserson, M.D., Niu, B., McLellan, M.D., Uzunangelov, V., et al. (2014) Multiplatform analysis of 12 cancer types reveals molecular classification within and across tissues of origin. Cell, 158(4): 929–944.
- Hoehndorf, R., Hardy, N.W., Osumi-Sutherland, D., Tweedie, S., Schofield, P.N., and Gkoutos, G.V. (2013) Systematic analysis of experimental phenotype data reveals gene functions. *PLoS ONE*, **8**(4): e60847.
- Holm, S. (1979) A simple sequentially rejective multiple test procedure. *Scandinavian Journal of Statistics*, **6**(2): 65–70.
- Holme, P. and Kim, B.J. (2002) Growing scale-free networks with tunable clustering. *Physical Review E*, **65**(2): 026107.
- Hopkins, A.L. (2008) Network pharmacology: the next paradigm in drug discovery. Nat Chem Biol, 4(11): 682–690.
- Hu, Z., Fan, C., Oh, D.S., Marron, J.S., He, X., Qaqish, B.F., Livasy, C., Carey, L.A., Reynolds, E., Dressler, L., et al. (2006) The molecular portraits of breast tumors are conserved across microarray platforms. BMC Genomics, 7: 96.
- Huang, E., Cheng, S., Dressman, H., Pittman, J., Tsou, M., Horng, C., Bild, A., Iversen, E., Liao, M., Chen, C., et al. (2003) Gene expression predictors of breast cancer outcomes. *Lancet*, **361**: 1590–1596.
- Illumina, Inc (Illumina) (2017) Sequencing and array-based solutions for genetic research. https://www.illumina.com/. Accessed: 26/03/2017.
- International HapMap 3 Consortium (HapMap) (2003) The International HapMap Project. *Nature*, **426**(6968): 789–796.
- Internation Human Genome Sequencing Consortium (IHGSC) (2004) Finishing the euchromatic sequence of the human genome. *Nature*, **431**(7011): 931–945.

- Jerby-Arnon, L., Pfetzer, N., Waldman, Y., McGarry, L., James, D., Shanks, E., Seashore-Ludlow, B., Weinstock, A., Geiger, T., Clemons, P., et al. (2014) Predicting cancer-specific vulnerability via data-driven detection of synthetic lethality. Cell, 158(5): 1199–1209.
- Joachims, T. (1999) Making large-scale support vector machine learning practical. In S. Bernhard, lkopf, J.C.B. Christopher, and J.S. Alexander (editors), Advances in kernel methods, 169–184. MIT Press.
- Ju, Z., Liu, W., Roebuck, P.L., Siwak, D.R., Zhang, N., Lu, Y., Davies, M.A., Akbani, R., Weinstein, J.N., Mills, G.B., et al. (2015) Development of a robust classifier for quality control of reverse-phase protein arrays. Bioinformatics, 31(6): 912.
- Kaelin, Jr, W. (2005) The concept of synthetic lethality in the context of anticancer therapy. *Nat Rev Cancer*, **5**(9): 689–98.
- Kaelin, Jr, W. (2009) Synthetic lethality: a framework for the development of wiser cancer therapeutics. *Genome Med*, **1**: 99.
- Kakiuchi, M., Nishizawa, T., Ueda, H., Gotoh, K., Tanaka, A., Hayashi, A., Yamamoto, S., Tatsuno, K., Katoh, H., Watanabe, Y., et al. (2014) Recurrent gain-of-function mutations of RHOA in diffuse-type gastric carcinoma. Nat Genet, 46(6): 583–587.
- Kamada, T. and Kawai, S. (1989) An algorithm for drawing general undirected graphs. *Information Processing Letters*, **31**(1): 7–15.
- Kandoth, C., Schultz, N., Cherniack, A.D., Akbani, R., Liu, Y., Shen, H., Robertson, A.G., Pashtan, I., Shen, R., Benz, C.C., et al. (2013) Integrated genomic characterization of endometrial carcinoma. Nature, 497(7447): 67–73.
- Kawai, J., Shinagawa, A., Shibata, K., Yoshino, M., Itoh, M., Ishii, Y., Arakawa, T., Hara, A., Fukunishi, Y., Konno, H., et al. (2001) Functional annotation of a full-length mouse cDNA collection. Nature, 409(6821): 685–690.
- Kelley, R. and Ideker, T. (2005) Systematic interpretation of genetic interactions using protein networks. *Nat Biotech*, **23**(5): 561–566.
- Kelly, S., Chen, A., Guilford, P., and Black, M. (2017a) Synthetic lethal interaction prediction of target pathways in E-cadherin deficient breast cancers. Submitted to *BMC Genomics*.

- Kelly, S.T. (2013) Statistical Predictions of Synthetic Lethal Interactions in Cancer. Dissertation, University of Otago.
- Kelly, S.T., Single, A.B., Telford, B.J., Beetham, H.G., Godwin, T.D., Chen, A., Black, M.A., and Guilford, P.J. (2017b) Towards HDGC chemoprevention: vulnerabilities in E-cadherin-negative cells identified by genome-wide interrogation of isogenic cell lines and whole tumors. Submitted to Cancer Prev Res.
- Kozlov, K.N., Gursky, V.V., Kulakovskiy, I.V., and Samsonova, M.G. (2015) Sequence-based model of gap gene regulation network. *BMC Genomics*, **15**(Suppl 12): S6.
- Kranthi, S., Rao, S., and Manimaran, P. (2013) Identification of synthetic lethal pairs in biological systems through network information centrality. *Mol BioSyst*, **9**(8): 2163–2167.
- Lander, E.S. (2011) Initial impact of the sequencing of the human genome. *Nature*, 470(7333): 187–197.
- Lander, E.S., Linton, L.M., Birren, B., Nusbaum, C., Zody, M.C., Baldwin, J., Devon, K., Dewar, K., Doyle, M., FitzHugh, W., et al. (2001) Initial sequencing and analysis of the human genome. *Nature*, **409**(6822): 860–921.
- Langmead, B., Trapnell, C., Pop, M., and Salzberg, S.L. (2009) Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biol*, **10**(3): R25.
- Latora, V. and Marchiori, M. (2001) Efficient behavior of small-world networks. *Phys Rev Lett*, **87**: 198701.
- Laufer, C., Fischer, B., Billmann, M., Huber, W., and Boutros, M. (2013) Mapping genetic interactions in human cancer cells with RNAi and multiparametric phenotyping. *Nat Methods*, **10**(5): 427–31.
- Law, C.W., Chen, Y., Shi, W., and Smyth, G.K. (2014) voom: precision weights unlock linear model analysis tools for RNA-seq read counts. *Genome Biol*, **15**(2): R29.
- Lawrence, M.S., Sougnez, C., Lichtenstein, L., Cibulskis, K., Lander, E., Gabriel, S.B., Getz, G., Ally, A., Balasundaram, M., Birol, I., et al. (2015) Comprehensive genomic characterization of head and neck squamous cell carcinomas. Nature, 517(7536): 576–582.

- Le Meur, N. and Gentleman, R. (2008) Modeling synthetic lethality. *Genome Biol*, **9**(9): R135.
- Le Meur, N., Jiang, Z., Liu, T., Mar, J., and Gentleman, R.C. (2014) Slgi: Synthetic lethal genetic interaction. r package version 1.26.0.
- Lee, A.Y., Perreault, R., Harel, S., Boulier, E.L., Suderman, M., Hallett, M., and Jenna, S. (2010a) Searching for signaling balance through the identification of genetic interactors of the rab guanine-nucleotide dissociation inhibitor gdi-1. *PLoS ONE*, **5**(5): e10624.
- Lee, I., Lehner, B., Vavouri, T., Shin, J., Fraser, A.G., and Marcotte, E.M. (2010b) Predicting genetic modifier loci using functional gene networks. *Genome Research*, **20**(8): 1143–1153.
- Lee, I. and Marcotte, E.M. (2009) Effects of functional bias on supervised learning of a gene network model. *Methods Mol Biol*, **541**: 463–75.
- Lee, M.J., Ye, A.S., Gardino, A.K., Heijink, A.M., Sorger, P.K., MacBeath, G., and Yaffe, M.B. (2012) Sequential application of anticancer drugs enhances cell death by rewiring apoptotic signaling networks. *Cell*, **149**(4): 780–94.
- Lehner, B., Crombie, C., Tischler, J., Fortunato, A., and Fraser, A.G. (2006) Systematic mapping of genetic interactions in caenorhabditis elegans identifies common modifiers of diverse signaling pathways. *Nat Genet*, **38**(8): 896–903.
- Li, X.J., Mishra, S.K., Wu, M., Zhang, F., and Zheng, J. (2014) Syn-lethality: An integrative knowledge base of synthetic lethality towards discovery of selective anticancer therapies. *Biomed Res Int*, **2014**: 196034.
- Linehan, W.M., Spellman, P.T., Ricketts, C.J., Creighton, C.J., Fei, S.S., Davis, C., Wheeler, D.A., Murray, B.A., Schmidt, L., Vocke, C.D., et al. (2016) Comprehensive Molecular Characterization of Papillary Renal-Cell Carcinoma. N Engl J Med, 374(2): 135–145.
- Lokody, I. (2014) Computational modelling: A computational crystal ball. *Nature Reviews Cancer*, **14**(10): 649–649.
- Lord, C.J., Tutt, A.N., and Ashworth, A. (2015) Synthetic lethality and cancer therapy: lessons learned from the development of PARP inhibitors. *Annu Rev Med*, **66**: 455–470.

- Lu, X., Kensche, P.R., Huynen, M.A., and Notebaart, R.A. (2013) Genome evolution predicts genetic interactions in protein complexes and reveals cancer drug targets. *Nat Commun*, 4: 2124.
- Lu, X., Megchelenbrink, W., Notebaart, R.A., and Huynen, M.A. (2015) Predicting human genetic interactions from cancer genome evolution. *PLoS One*, **10**(5): e0125795.
- Lum, P.Y., Armour, C.D., Stepaniants, S.B., Cavet, G., Wolf, M.K., Butler, J.S., Hinshaw, J.C., Garnier, P., Prestwich, G.D., Leonardson, A., et al. (2004) Discovering modes of action for therapeutic compounds using a genome-wide screen of yeast heterozygotes. Cell, 116(1): 121–137.
- Luo, J., Solimini, N.L., and Elledge, S.J. (2009) Principles of Cancer Therapy: Oncogene and Non-oncogene Addiction. *Cell*, **136**(5): 823–837.
- Machado, J., Olivera, C., Carvalh, R., Soares, P., Berx, G., Caldas, C., Sercuca, R., Carneiro, F., and Sorbrinho-Simoes, M. (2001) E-cadherin gene (*CDH1*) promoter methylation as the second hit in sporadic diffuse gastric carcinoma. *Oncogene*, **20**: 1525–1528.
- Masciari, S., Larsson, N., Senz, J., Boyd, N., Kaurah, P., Kandel, M.J., Harris, L.N., Pinheiro, H.C., Troussard, A., Miron, P., et al. (2007) Germline E-cadherin mutations in familial lobular breast cancer. J Med Genet, 44(11): 726–31.
- Mattison, J., van der Weyden, L., Hubbard, T., and Adams, D.J. (2009) Cancer gene discovery in mouse and man. *Biochim Biophys Acta*, **1796**(2): 140–161.
- Maxam, A.M. and Gilbert, W. (1977) A new method for sequencing DNA. *Proceedings* of the National Academy of Science, **74**(2): 560–564.
- McCourt, C.M., McArt, D.G., Mills, K., Catherwood, M.A., Maxwell, P., Waugh, D.J., Hamilton, P., O'Sullivan, J.M., and Salto-Tellez, M. (2013) Validation of next generation sequencing technologies in comparison to current diagnostic gold standards for BRAF, EGFR and KRAS mutational analysis. *PLoS ONE*, 8(7): e69604.
- McLachlan, J., George, A., and Banerjee, S. (2016) The current status of parp inhibitors in ovarian cancer. *Tumori*, **102**(5): 433–440.

- McLendon, R., Friedman, A., Bigner, D., Van Meir, E.G., Brat, D.J., Mastrogianakis, G.M., Olson, J.J., Mikkelsen, T., Lehman, N., Aldape, K., et al. (2008) Comprehensive genomic characterization defines human glioblastoma genes and core pathways. Nature, 455(7216): 1061–1068.
- Miles, D.W. (2001) Update on HER-2 as a target for cancer therapy: herceptin in the clinical setting. *Breast Cancer Res*, **3**(6): 380–384.
- Mortazavi, A., Williams, B.A., McCue, K., Schaeffer, L., and Wold, B. (2008) Mapping and quantifying mammalian transcriptomes by RNA-Seq. *Nat Methods*, **5**(7): 621–628.
- Muzny, D.M., Bainbridge, M.N., Chang, K., Dinh, H.H., Drummond, J.A., Fowler, G., Kovar, C.L., Lewis, L.R., Morgan, M.B., Newsham, I.F., et al. (2012) Comprehensive molecular characterization of human colon and rectal cancer. Nature, 487(7407): 330–337.
- Nagalla, S., Chou, J.W., Willingham, M.C., Ruiz, J., Vaughn, J.P., Dubey, P., Lash, T.L., Hamilton-Dutoit, S.J., Bergh, J., Sotiriou, C., et al. (2013) Interactions between immunity, proliferation and molecular subtype in breast cancer prognosis. Genome Biol, 14(4): R34.
- Neeley, E.S., Kornblau, S.M., Coombes, K.R., and Baggerly, K.A. (2009) Variable slope normalization of reverse phase protein arrays. *Bioinformatics*, **25**(11): 1384.
- Novomestky, F. (2012) matrixcalc: Collection of functions for matrix calculations. R package version 1.0-3.
- Oliveira, C., Senz, J., Kaurah, P., Pinheiro, H., Sanges, R., Haegert, A., Corso, G., Schouten, J., Fitzgerald, R., Vogelsang, H., et al. (2009) Germline CDH1 deletions in hereditary diffuse gastric cancer families. Human Molecular Genetics, 18(9): 1545–1555.
- Oliveira, C., Seruca, R., Hoogerbrugge, N., Ligtenberg, M., and Carneiro, F. (2013) Clinical utility gene card for: Hereditary diffuse gastric cancer (HDGC). Eur J Hum Genet, 21(8).
- Pandey, G., Zhang, B., Chang, A.N., Myers, C.L., Zhu, J., Kumar, V., and Schadt, E.E. (2010) An integrative multi-network and multi-classifier approach to predict genetic interactions. *PLoS Comput Biol*, **6**(9).

- Parker, J., Mullins, M., Cheung, M., Leung, S., Voduc, D., Vickery, T., Davies, S., Fauron, C., He, X., Hu, Z., et al. (2009) Supervised risk predictor of breast cancer based on intrinsic subtypes. *Journal of Clinical Oncology*, 27(8): 1160–1167.
- Peltonen, L. and McKusick, V.A. (2001) Genomics and medicine. Dissecting human disease in the postgenomic era. *Science*, **291**(5507): 1224–1229.
- Pereira, B., Chin, S.F., Rueda, O.M., Vollan, H.K., Provenzano, E., Bardwell, H.A., Pugh, M., Jones, L., Russell, R., Sammut, S.J., et al. (2016) Erratum: The somatic mutation profiles of 2,433 breast cancers refine their genomic and transcriptomic landscapes. Nat Commun. 7: 11908.
- Perou, C.M., Sørlie, T., Eisen, M.B., van de Rijn, M., Jeffrey, S.S., Rees, C.A., Pollack, J.R., Ross, D.T., Johnsen, H., Akslen, L.A., et al. (2000) Molecular portraits of human breast tumours. Nature, 406(6797): 747–752.
- Pleasance, E.D., Cheetham, R.K., Stephens, P.J., McBride, D.J., Humphray, S.J., Greenman, C.D., Varela, I., Lin, M.L., Ordonez, G.R., Bignell, G.R., et al. (2010) A comprehensive catalogue of somatic mutations from a human cancer genome. Nature, 463(7278): 191–196.
- Polyak, K. and Weinberg, R.A. (2009) Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits. *Nat Rev Cancer*, **9**(4): 265–73.
- Prahallad, A., Sun, C., Huang, S., Di Nicolantonio, F., Salazar, R., Zecchin, D., Beijersbergen, R.L., Bardelli, A., and Bernards, R. (2012) Unresponsiveness of colon cancer to *BRAF* (v600e) inhibition through feedback activation of egfr. *Nature*, **483**(7387): 100–3.
- R Core Team (2016) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. R version 3.3.2.
- Ravnan, M.C. and Matalka, M.S. (2012) Vemurafenib in patients with *BRAF* v600e mutation-positive advanced melanoma. *Clin Ther*, **34**(7): 1474–86.
- Ritchie, M.E., Phipson, B., Wu, D., Hu, Y., Law, C.W., Shi, W., and Smyth, G.K. (2015) limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Research*, **43**(7): e47.

- Robin, J.D., Ludlow, A.T., LaRanger, R., Wright, W.E., and Shay, J.W. (2016) Comparison of DNA Quantification Methods for Next Generation Sequencing. *Sci Rep*, 6: 24067.
- Robinson, M.D. and Oshlack, A. (2010) A scaling normalization method for differential expression analysis of RNA-seq data. *Genome Biol*, **11**(3): R25.
- Roguev, A., Bandyopadhyay, S., Zofall, M., Zhang, K., Fischer, T., Collins, S.R., Qu, H., Shales, M., Park, H.O., Hayles, J., et al. (2008) Conservation and rewiring of functional modules revealed by an epistasis map in fission yeast. Science, 322(5900): 405–10.
- Rung, J. and Brazma, A. (2013) Reuse of public genome-wide gene expression data.

  Nat Rev Genet, 14(2): 89–99.
- Rustici, G., Kolesnikov, N., Brandizi, M., Burdett, T., Dylag, M., Emam, I., Farne, A., Hastings, E., Ison, J., Keays, M., et al. (2013) ArrayExpress update—trends in database growth and links to data analysis tools. Nucleic Acids Res, 41(Database issue): D987–990.
- Ryan, C., Lord, C., and Ashworth, A. (2014) Daisy: Picking synthetic lethals from cancer genomes. *Cancer Cell*, **26**(3): 306–308.
- Sander, J.D. and Joung, J.K. (2014) Crispr-cas systems for editing, regulating and targeting genomes. *Nat Biotechnol*, **32**(4): 347–55.
- Sanger, F. and Coulson, A. (1975) A rapid method for determining sequences in dna by primed synthesis with dna polymerase. *Journal of Molecular Biology*, **94**(3): 441 448.
- Scheuer, L., Kauff, N., Robson, M., Kelly, B., Barakat, R., Satagopan, J., Ellis, N., Hensley, M., Boyd, J., Borgen, P., et al. (2002) Outcome of preventive surgery and screening for breast and ovarian cancer in BRCA mutation carriers. *J Clin Oncol*, **20**(5): 1260–1268.
- Semb, H. and Christofori, G. (1998) The tumor-suppressor function of E-cadherin. *Am J Hum Genet*, **63**(6): 1588–93.
- Sing, T., Sander, O., Beerenwinkel, N., and Lengauer, T. (2005) Rocr: visualizing classifier performance in r. *Bioinformatics*, **21**(20): 7881.

- Slurm development team (Slurm) (2017) Slurm workload manager. https://slurm.schedmd.com/. Accessed: 25/03/2017.
- Sørlie, T., Perou, C.M., Tibshirani, R., Aas, T., Geisler, S., Johnsen, H., Hastie, T., Eisen, M.B., van de Rijn, M., Jeffrey, S.S., et al. (2001) Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proc Natl Acad Sci USA, 98(19): 10869–10874.
- Stajich, J.E. and Lapp, H. (2006) Open source tools and toolkits for bioinformatics: significance, and where are we? *Brief Bioinformatics*, **7**(3): 287–296.
- Stratton, M.R., Campbell, P.J., and Futreal, P.A. (2009) The cancer genome. *Nature*, **458**(7239): 719–724.
- Ström, C. and Helleday, T. (2012) Strategies for the use of poly(adenosine diphosphate ribose) polymerase (parp) inhibitors in cancer therapy. *Biomolecules*, **2**(4): 635–649.
- Sun, C., Wang, L., Huang, S., Heynen, G.J.J.E., Prahallad, A., Robert, C., Haanen, J., Blank, C., Wesseling, J., Willems, S.M., et al. (2014) Reversible and adaptive resistance to BRAF(v600e) inhibition in melanoma. Nature, 508(7494): 118–122.
- Taylor, I.W., Linding, R., Warde-Farley, D., Liu, Y., Pesquita, C., Faria, D., Bull, S., Pawson, T., Morris, Q., and Wrana, J.L. (2009) Dynamic modularity in protein interaction networks predicts breast cancer outcome. *Nat Biotechnol*, 27(2): 199–204.
- Telford, B.J., Chen, A., Beetham, H., Frick, J., Brew, T.P., Gould, C.M., Single, A., Godwin, T., Simpson, K.J., and Guilford, P. (2015) Synthetic lethal screens identify vulnerabilities in gpcr signalling and cytoskeletal organization in E-cadherin-deficient cells. *Mol Cancer Ther*, **14**(5): 1213–1223.
- The 1000 Genomes Project Consortium (1000 Genomes) (2010) A map of human genome variation from population-scale sequencing. *Nature*, **467**(7319): 1061–1073.
- The Cancer Genome Atlas Research Network (TCGA) (2012) Comprehensive molecular portraits of human breast tumours. *Nature*, **490**(7418): 61–70.
- The Cancer Genome Atlas Research Network (TCGA) (2017a) The Cancer Genome Atlas Project. https://cancergenome.nih.gov/. Accessed: 26/03/2017.

- The Cancer Genome Atlas Research Network (TCGA) (2017b) The Cancer Genome Atlas Project Data Portal. https://tcga-data.nci.nih.gov/. Accessed: 06/02/2017 (via cBioPortal.
- The Cancer Society of New Zealand (Cancer Society of NZ) (2017) What is cancer? https://otago-southland.cancernz.org.nz/en/cancer-information/other-links/what-is-cancer-3/. Accessed: 22/03/2017.
- The Catalogue Of Somatic Mutations In Cancer (COSMIC) (2016) Cosmic: The catalogue of somatic mutations in cancer. http://cancer.sanger.ac.uk/cosmic. Release 79 (23/08/2016), Accessed: 05/02/2017.
- The Comprehensive R Archive Network (CRAN) (2017) Cran. https://cran.r-project.org/. Accessed: 24/03/2017.
- The ENCODE Project Consortium (ENCODE) (2004) The ENCODE (ENCyclopedia Of DNA Elements) Project. *Science*, **306**(5696): 636–640.
- The Internation Cancer Genome Consortium (ICGC) (2017) ICGC Data Portal. https://dcc.icgc.org/. Accessed: 06/02/2017.
- The National Cancer Institute (NCI) (2015) The genetics of cancer. https://www.cancer.gov/about-cancer/causes-prevention/genetics. Published: 22/04/2015, Accessed: 22/03/2017.
- The New Zealand eScience Infrastructure (NeSI) (2017) NeSI. https://www.nesi.org.nz/. Accessed: 25/03/2017.
- The Pharmaceutical Management Agency (PHARMAC) (2016) Approval of multiproduct funding proposal with roche.
- Tierney, L., Rossini, A.J., Li, N., and Sevcikova, H. (2015) snow: Simple Network of Workstations. R package version 0.4-2.
- Tiong, K.L., Chang, K.C., Yeh, K.T., Liu, T.Y., Wu, J.H., Hsieh, P.H., Lin, S.H., Lai, W.Y., Hsu, Y.C., Chen, J.Y., et al. (2014) Csnk1e/ctnnb1 are synthetic lethal to tp53 in colorectal cancer and are markers for prognosis. Neoplasia, 16(5): 441–50.
- Tischler, J., Lehner, B., and Fraser, A.G. (2008) Evolutionary plasticity of genetic interaction networks. *Nat Genet*, **40**(4): 390–391.

- Tomasetti, C. and Vogelstein, B. (2015) Cancer etiology. Variation in cancer risk among tissues can be explained by the number of stem cell divisions. *Science*, **347**(6217): 78–81.
- Tong, A.H., Evangelista, M., Parsons, A.B., Xu, H., Bader, G.D., Page, N., Robinson, M., Raghibizadeh, S., Hogue, C.W., Bussey, H., et al. (2001) Systematic genetic analysis with ordered arrays of yeast deletion mutants. Science, 294(5550): 2364–8.
- Tong, A.H., Lesage, G., Bader, G.D., Ding, H., Xu, H., Xin, X., Young, J., Berriz, G.F., Brost, R.L., Chang, M., et al. (2004) Global mapping of the yeast genetic interaction network. *Science*, **303**(5659): 808–13.
- Travers, J. and Milgram, S. (1969) An experimental study of the small world problem. Sociometry, **32**(4): 425–443.
- Tsai, H.C., Li, H., Van Neste, L., Cai, Y., Robert, C., Rassool, F.V., Shin, J.J., Harbom, K.M., Beaty, R., Pappou, E., et al. (2012) Transient low doses of dnademethylating agents exert durable antitumor effects on hematological and epithelial tumor cells. Cancer Cell, 21(3): 430–46.
- Tutt, A., Robson, M., Garber, J.E., Domchek, S.M., Audeh, M.W., Weitzel, J.N., Friedlander, M., Arun, B., Loman, N., Schmutzler, R.K., et al. (2010) Oral poly(adpribose) polymerase inhibitor olaparib in patients with *BRCA1* or *BRCA2* mutations and advanced breast cancer: a proof-of-concept trial. *Lancet*, **376**(9737): 235–44.
- van der Meer, R., Song, H.Y., Park, S.H., Abdulkadir, S.A., and Roh, M. (2014) RNAi screen identifies a synthetic lethal interaction between PIM1 overexpression and PLK1 inhibition. *Clinical Cancer Research*, **20**(12): 3211–3221.
- van Steen, K. (2012) Travelling the world of genegene interactions. *Briefings in Bioinformatics*, **13**(1): 1–19.
- van Steen, M. (2010) Graph Theory and Complex Networks: An Introduction. Maarten van Steen, VU Amsterdam.
- Vapnik, V.N. (1995) The nature of statistical learning theory. Springer-Verlag New York, Inc.
- Vargas, J.J., Gusella, G., Najfeld, V., Klotman, M., and Cara, A. (2004) Novel integrase-defective lentiviral episomal vectors for gene transfer. *Hum Gene Ther*, 15: 361–372.

- Vizeacoumar, F.J., Arnold, R., Vizeacoumar, F.S., Chandrashekhar, M., Buzina, A., Young, J.T., Kwan, J.H., Sayad, A., Mero, P., Lawo, S., et al. (2013) A negative genetic interaction map in isogenic cancer cell lines reveals cancer cell vulnerabilities. Mol Syst Biol, 9: 696.
- Vogelstein, B., Papadopoulos, N., Velculescu, V.E., Zhou, S., Diaz, L.A., and Kinzler, K.W. (2013) Cancer genome landscapes. Science, 339(6127): 1546–1558.
- Vos, C.B., Cleton-Jansen, A.M., Berx, G., de Leeuw, W.J., ter Haar, N.T., van Roy, F., Cornelisse, C.J., Peterse, J.L., and van de Vijver, M.J. (1997) E-cadherin inactivation in lobular carcinoma in situ of the breast: an early event in tumorigenesis. Br J Cancer, 76(9): 1131–3.
- Wang, K., Singh, D., Zeng, Z., Coleman, S.J., Huang, Y., Savich, G.L., He, X., Mieczkowski, P., Grimm, S.A., Perou, C.M., et al. (2010) MapSplice: accurate mapping of RNA-seq reads for splice junction discovery. Nucleic Acids Res, 38(18): e178.
- Wang, K., Yuen, S.T., Xu, J., Lee, S.P., Yan, H.H., Shi, S.T., Siu, H.C., Deng, S., Chu, K.M., Law, S., et al. (2014) Whole-genome sequencing and comprehensive molecular profiling identify new driver mutations in gastric cancer. Nat Genet, 46(6): 573–582.
- Wang, X. and Simon, R. (2013) Identification of potential synthetic lethal genes to p53 using a computational biology approach. *BMC Medical Genomics*, **6**(1): 30.
- Wappett, M. (2014) Bisep: Toolkit to identify candidate synthetic lethality. r package version 2.0.
- Wappett, M., Dulak, A., Yang, Z.R., Al-Watban, A., Bradford, J.R., and Dry, J.R. (2016) Multi-omic measurement of mutually exclusive loss-of-function enriches for candidate synthetic lethal gene pairs. *BMC Genomics*, **17**: 65.
- Warnes, G.R., Bolker, B., Bonebakker, L., Gentleman, R., Liaw, W.H.A., Lumley, T., Maechler, M., Magnusson, A., Moeller, S., Schwartz, M., et al. (2015) gplots: Various R Programming Tools for Plotting Data. R package version 2.17.0.
- Watts, D.J. and Strogatz, S.H. (1998) Collective dynamics of 'small-world' networks. *Nature*, **393**(6684): 440–2.
- Weinstein, I.B. (2000) Disorders in cell circuitry during multistage carcinogenesis: the role of homeostasis. *Carcinogenesis*, **21**(5): 857–864.

- Weinstein, J.N., Akbani, R., Broom, B.M., Wang, W., Verhaak, R.G., McConkey, D., Lerner, S., Morgan, M., Creighton, C.J., Smith, C., et al. (2014) Comprehensive molecular characterization of urothelial bladder carcinoma. Nature, 507(7492): 315–322.
- Weinstein, J.N., Collisson, E.A., Mills, G.B., Shaw, K.R., Ozenberger, B.A., Ellrott, K., Shmulevich, I., Sander, C., Stuart, J.M., Chang, K., et al. (2013) The Cancer Genome Atlas Pan-Cancer analysis project. Nat Genet, 45(10): 1113–1120.
- Wickham, H. and Chang, W. (2016) devtools: Tools to Make Developing R Packages Easier. R package version 1.12.0.
- Wickham, H., Danenberg, P., and Eugster, M. (2017) roxygen2: In-Line Documentation for R. R package version 6.0.1.
- Wong, S.L., Zhang, L.V., Tong, A.H.Y., Li, Z., Goldberg, D.S., King, O.D., Lesage, G., Vidal, M., Andrews, B., Bussey, H., et al. (2004) Combining biological networks to predict genetic interactions. Proceedings of the National Academy of Sciences of the United States of America, 101(44): 15682–15687.
- World Health Organization (WHO) (2017) Fact sheet: Cancer. http://www.who.int/mediacentre/factsheets/fs297/en/. Updated February 2017, Accessed: 22/03/2017.
- Wu, M., Li, X., Zhang, F., Li, X., Kwoh, C.K., and Zheng, J. (2014) In silico prediction of synthetic lethality by meta-analysis of genetic interactions, functions, and pathways in yeast and human cancer. *Cancer Inform*, **13**(Suppl 3): 71–80.
- Yu, H. (2002) Rmpi: Parallel statistical computing in r. R News, 2(2): 10–14.
- Zhang, F., Wu, M., Li, X.J., Li, X.L., Kwoh, C.K., and Zheng, J. (2015) Predicting essential genes and synthetic lethality via influence propagation in signaling pathways of cancer cell fates. *J Bioinform Comput Biol*, **13**(3): 1541002.
- Zhang, J., Baran, J., Cros, A., Guberman, J.M., Haider, S., Hsu, J., Liang, Y., Rivkin, E., Wang, J., Whitty, B., et al. (2011) International cancer genome consortium data portala one-stop shop for cancer genomics data. Database: The Journal of Biological Databases and Curation, 2011: bar026.
- Zhong, W. and Sternberg, P.W. (2006) Genome-wide prediction of c. elegans genetic interactions. *Science*, **311**(5766): 1481–1484.

Zweig, M.H. and Campbell, G. (1993) Receiver-operating characteristic (roc) plots: a fundamental evaluation tool in clinical medicine. *Clinical Chemistry*, **39**(4): 561–577.