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A Bioinformatics Approach to  
Synthetic Lethal Interactions in  
Breast Cancer with Gene  
Expression Data

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a thesis submitted for the degree of  
Doctor of Philosophy  
at the University of Otago, Dunedin,  
New Zealand.

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## **Abstract**

### **Background**

Synthetic lethal interactions are re-emerging in genetics research in the genomics era driven by potential applications in precision medicine against cancers. This approach aims to exploit functional redundancy at the genetic level against mutations in cancers for developing specific treatments against them, including loss of function events in tumour suppressors. Of particular interest is the targeting loss of function of E-cadherin, encoded by *CDH1*, a tumour suppressor gene involved in Breast and Stomach cancers. Experimental screens have been used to identify candidate synthetic lethal interactions and here bioinformatics analysis used to augment the triage drug target triage process. Furthermore the pathway composition of synthetic lethal candidates and the effect of pathway structure on their detection in genomics data.

### **Approach**

A computational statistics methodology, the Synthetic Lethal Prediction Tool (SLIPT) has been developed to detect synthetic lethal interactions in gene expression datasets. The methodology has been demonstrated on Breast and Stomach cancer datasets from The Cancer Genome Atlas (TCGA) database, testing for interactions with *CDH1*. Various analyses have been applied to further elucidate these candidates, including differential gene expression, correlation co-expression, unsupervised clustering, gene set over-representation analysis, singular-value decomposition “metagenes”, and permutation re-sampling analysis. A particular challenge of performing these analyses was to compare SLIPT gene candidates to the results of an experimental synthetic lethal siRNA screen of E-cadherin Telford

*et al.* (2015) at the pathway level. Graph theory methods including information centrality and shortest paths were applied to the most supported pathways from both the computational and experimental synthetic lethal candidates to test for graph structure among hits from each approach. Simulation and modelling was performed to test the statistical performance of the SLIPT methodology and further applied to datasets with simulated correlation structures, including those derived from known graph stuctures.

## Findings

A vast number of genes having expression consistent with being synthetic lethal partners of *CDH1* were detected in both TCGA Breast and Stomach cancer genes. For breast cancers, these genes clustered into several distinct groups, with distinct enriched biological functions and elevated expression in different clinical subclasses such as normal-like, basal, or estrogen receptor negative samples. While the number of genes detected by both computational and experimental approaches were not significant, there was significant pathway composition in the overlapping genes. In particular  $G_{\alpha i}$  signalling, cytoplasmic microfibres, and extracelluar fibrin clotting were supported by both approaches even after permutation testing. These findings are consistent with the known roles of E-cadherin in cytoskeletal or cell signalling roles and the proposed downstream targets of GPCR singalling of Telford *et al.* (2015). Many of these and related pathways were replicated in the separate stomach cancer dataset. Furthermore other candidate pathways uniquely supported by the computational predictions included regulation of immune signaling and translational elongation, both unlikely to have been detected with high dose siRNA in an isogenic cell line and these are still candidates for further testing in mouse xenograft models.

A number of approaches were adapted or developed to test whether there was a connection between synthetic lethal candidates in the graph structures of the pathways most supported by prior analyses. Network centrality measures were used to compare the importance or connectivity of genes in the pathway subnetworks but no significant difference was found between synthetic candidates and other genes within the same pathway. Another hypothesis was that computational synthetic lethal candidates would be

downstream of experimental candidates within a pathway but no evidence of directionality between the candidates was detected.

A model of synthetic lethality was developed and was sucessfully implemented to simulate gene expression datasets with known underlying synthetic lethal partners of a query gene. For small numbers of known synthetic lethal partners, the SLIPT methdology performed well respect to reciever operator characteristic curves. As the number of true partners to detect increases, the power to detect them diminishes. Increasing sample sizes, however, was able to mitigate this effect somewhat as expected. This finding was replicated in simulations up to a feasible number of human genes (20,000) with more true negatives and correlations structures. The SLIPT methdology performs similarly across these conditions and performs better than Pearson's correlation (for co-expression) or the  $\chi^2$ -test without a directional criterion. However, correlation structure of the dataset does impact on synthetic lethal predictions, genes correlated with (or in a pathway structure near to) true synthetic lethal partners having elevated test statistic values over other true negatives. A quadratic (second order polynomial) least squares linear regression methodology has been developed as a comparable alternative with the added benefit of conditioning against known partners (or strongest candidates prior analyses).

Thus my thesis has developed, evaluated and refined a bioinformatics approach to discovery of synthetic lethal genes solely from gene expression data.

## Research Contributions During Candidature

### Publications Under Peer-Review

Kelly, S. T. and Spencer, H. G. (2017) Population-Genetics Models of Sex-Limited Genomic Imprinting. *Theoretical Population Biology* **115**:35-44 doi:10.1016/j.tpb.2017.03.004

Kelly, S. T., Single, A. B., Telford, B. J., Beetham, H. G., Godwin, T. D., Chen, A., Black, M., A., and Guilford, P. J. (2017) Towards HDGC chemoprevention: vulnerabilities in E-cadherin-negative cells identified by genomic interrogation of isogenic cell lines and whole tumors. Submitted to *Cancer Prevention Research*.

Kelly, S. T., Chen, A., Guilford, P. J., and Black, M. A. (2017) Synthetic lethal interaction prediction of target pathways in E-cadherin deficient breast cancers. Submitted to *BMC Genomics*.

### Community Blog Posts

Black, M. A., Kelly, S. T., and Cadzow, M. Posted on the *Software Carpentry* website 2016 July 4<sup>th</sup>: “Software Carpentry workshop at the University of Otago, New Zealand” <https://software-carpentry.org/blog/2016/07/otago-workshop.html>

Kelly, S. T., Black, M., A., Bae, S., Hayek, W., and Pawlik, A. Posted on the *Software Carpentry* website 2016 September 28<sup>th</sup>: “Software Carpentry Workshop Attendance: a New Zealand Perspective” <https://software-carpentry.org/blog/2016/09/attendance-nz.html>

### Software Packages

Several software packages in the R language have been released on GitHub while preparing this thesis. Please see the appropriate GitHub repository for more information on installing and running these packages, on the following account: <https://github.com/TomKellyGenetics>

**slipt** is the Synthetic Lethal interaction Prediction Tool, released to accompany the synthetic lethal publication above. **slipt-app** contains an application developed in the R **shiny** environment as part of a related project.

Several plotting functions were customised for the Figures in this thesis (and the above publications), notably `heatmap.2x` and `vioplotx` have been prepared largely for use during this project but are also documented and available to other R users. These are enhancements to the CRAN `gplots` and `vioplot` packages respectively and are intended be user-friendly for those familiar with `heatmap.2` or `vioplot` and `boxplot` (base R) functions. These are backwards compatible with these functions, taking similar inputs as demonstrated in the appropriate vignettes.

The use of iGraph (the R `igraph` package) operations of graph-network structure in the analysis and simulations of pathways involved several original or customised functions to manipulate or plot `igraph` objects and adjacency matrices. These can be install separately from their respective repositories of with the metapackage: `igraph.extensions`. `plot.igraph` enables plotting graph networks with customised inhibitor arrow and node colours. `info.centrality` enables the calculation of additional node and network centrality metrics not available in the `igraph` package. `pathway.structure.permutation` enables testing of related states or node groups in a network by directionality of shortest paths. The `graphsim` package has been set up to simulate a multi-variate normal gene expression dataset with `mvtnorm` while deriving the correlation structure,  $\Sigma$ , from a graph structure. Note that these require various packages for graph theory, statistics and matrix operations and these will be installed as dependencies.

## Conference Participation

eResearch 2017 (Queenstown) Speaker February 20<sup>th</sup>-22<sup>nd</sup> “Detecting Synthetic Lethality from Cancer Gene Expression: A PhD project on genetic interactions with CDH1 inactivation in TCGA data”

RIKEN Division of Genomic Technologies (Yokohama, Japan) Seminar 2016 October 20<sup>th</sup>; National Institute of Genetics (Mishima, Japan) Seminar 2016 October 21<sup>st</sup>; Tokyo University Institute of Medical Science (Shirokanedai Campus, Japan) Seminar 2016 October 24<sup>th</sup>; Sokendai Graduate University (Hayama, Japan) Seminar 2016 October 25<sup>th</sup> “Analysis of Synthetic Lethal Pathways in Breast Cancer: A PhD project on genetic interactions with CDH1 inactivation in TCGA data”

Next Generation Sequencing Asia 2016 (Singapore) Poster October 11<sup>th</sup>-12<sup>th</sup> “Bioinformatic Investigations of Synthetic Lethal Interactions with E-cadherin in Breast Cancer” (Supported by the University of Otago Division of Health Sciences; Maurice and Phyllis Paykel Trust)

eResearch 2016 (Queenstown, New Zealand) Speaker February 9<sup>th</sup>-11<sup>th</sup> “Sifting the Needles in the Haystack: Permutation Resampling Biological Pathways in Cancer Genomic Interaction Data” (Supported by REANNZ)

Genetics Otago Symposium 2016 (Dunedin, New Zealand) Student Speaker March 7<sup>th</sup>-8<sup>th</sup> “A Bioinformatics approach to Genetic Interactions: Synthetic Lethal Pathways with E-cadherin in Breast Cancer Genomics Data”

Research Bazaar 2015 (Melbourne, Australia) February 16<sup>th</sup>-18<sup>th</sup> “My digital research toolkit” (Supported by the New Zealand eScience Infrastructure)

QMB Cancer Drugs Satellite 2014 (Queenstown, New Zealand) Poster August 24<sup>th</sup>-25<sup>th</sup>; Otago School of Medical Sciences 2015 (Dunedin, New Zealand) Poster Postgraduate Symposium April 28<sup>th</sup>-29<sup>th</sup> Bioinformatics Prioritisation of Synthetic Lethal Targets for Drug Activity Against E-Cadherin Deficient Cancers

DunDead: Zombie Science and Popular Culture Festival 2014 (Dunedin, New Zealand) Ignite Speaker August 16<sup>th</sup>-17<sup>th</sup> “Hidden in Plain Sight - The Genetics of Zombies”

eResearch 2014 (Waikato University, Hamilton, New Zealand) Ignite Speaker June 30<sup>th</sup>-July 2<sup>nd</sup> “Bioinformatic analysis of synthetic lethal genetic interactions in breast cancer” (Supported by Google)

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- REANNZ, NZGL, and NeSI (towards eResearch 2016 conference, Queenstown)
- Otago Division of Health Sciences, Department of Biochemistry, Oxford Global, and Maurice and Phyllis Paykel Trust (towards NGS Asia 2016, Singapore)
- RIKEN Division of Genomics Technologies and the Okinawa Institute of Science and Technology (for hosting seminars in Japan)

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# Glossary

RNA-Seq Transcriptome data from sequencing RNA.

# **Acronyms**

DNA Deoxyribonucleic acid.

qPCR Quantitative (real-time) polymerase chain reaction.

# Chapter 1

## Introduction

The thesis presents research into genetic interactions based on genomics data and bioinformatics approaches. This Chapter introduces the recent developments in genomics and bioinformatics, particularly in their application to cancer research. Synthetic lethal interactions are a long standing area of research in genetics in both model organisms and cancer biology. Various reasons why these interactions are of interest in fundamental and translational biology will be outlined but first these and similar interactions will be defined. A bioinformatics approach to synthetic lethal interactions enables much wider exploration of the inter-connected nature of genes and proteins within a cancer cell than previous candidate-based approaches. An alternative approach is experimental screening which will be presented and contrasted with bioinformatics approaches in more detail. An emerging application of synthetic lethality is the design of treatments with specificity against loss of function mutations in tumour suppressor genes. E-cadherin (encoded by *CDH1*) is a prime example of this which will be the focus of the analysis in this thesis and as such the role of this gene in cellular and cancer biology will be briefly reviewed.

### 1.1 Cancer Research in the Post-Genomic Era

Genomics technologies have the potential to vastly impact upon various areas including health and cancer medicine. Considering the progress in recent genomics research, it could soon impact greatly upon clinical and wider applications of genetics either directly or by enabling more focused genetics research from candidates selected from genomics or bioinformatics analysis. The completion of the draft Human Genome (Lander *et al.*, 2001) marks a major accomplishment in genetics research and raises new challenges to utilise this genomic scale information effectively. Technologies in this

area have rapidly developed since completion of the human genome project and many global large-scale projects have expanded upon the human genome, to populations (1000 Genomes, 2010), to cancers (Dickson, 1999; Zhang *et al.*, 2011), and to deeper functional understanding (Kawai *et al.*, 2001; ENCODE, 2004). However, impact on the clinic has been slower than initially anticipated following the completion of the “draft” genome with genomics technologies yet to become widely adopted in healthcare and oncology. Here we outline the genomics technologies and bioinformatics approaches which have led to availability of genomics data and techniques used in this thesis and potential for applications in cancer research or the clinic in the future.

### **1.1.1 Cancer as a Global Health Concern**

Cancer is a class of diseases involving malignant cellular growth, invasion of tissues, and spread to other organs. While there are also environmental factors, most cancers occur more frequently with age and family history so genetics is widely acknowledged to have an important role in cancer risk. Cancers arise from dysregulated cellular growth or differentiation from stem cells, these can occur through genetic mutations or alterations in gene regulation or expression.

Cancers are a major global health concern, being the second leading cause of death globally (WHO, 2017), with an estimated annual incidence of 14.1 million cases and annual mortality of 8.2 million people (Ferlay *et al.*, 2015). Breast and stomach cancers are among the 5 most frequent cancers globally, with breast cancer affecting women more than other cancer tissue types. Breast cancer has an estimated annual incidence of 1.6 million cases and mortality of 520 thousand people. Stomach cancer has an estimated annual incidence of 950 thousand cases and a mortality of 723 thousand people. Cancer is also a major health concern here in New Zealand, with 19.1 thousand people (including 2.5 thousand cases of breast cancer and 370 cases of stomach cancer) diagnosed annually (Hanna, 2003), among the highest incidence (age-standardised per capita) of cancer in the world (Ferlay *et al.*, 2015).

While the genetic contribution to cancer risk and many of the molecular changes occurring cancers are widely acknowledged (ASCO, 2017; Cancer Research UK, 2017; Cancer Society of NZ, 2017), much of these findings have yet to impact on clinical practice. Diagnostics are traditionally based on pathological examination of cancer cell and tissue samples, including histological staining for biomolecules and biomarkers, and continue to be widely used. The current standard of care is surgery, radiation, and cytotoxic chemotherapy, depending on whether the cancer is localised or has become

systemic (via metastasis) and spread to other organ systems. These approaches are effective against cancers, particularly in patients particular subtypes (such as acute myeloid leukaemia) or early stage cancers. Thus early intervention is important to patient survival and quality of life with national screening programs aiming to diagnose cancers early and subtypes more accurately, including identification of patients with genetic variants or family histories for high risk of particular cancers.

Chemotherapy is a treatment for advanced stage (systemic) cancers which is designed to inhibit the growth and spread of cancer throughout the body by targeting rapidly growing cells. However, this approach is notorious for adverse effects and a narrow therapeutic window and is not suitable for chemopreventative application in many cases (Kaelin, Jr, 2009). Thus high risk individuals are regularly monitored for cancers and offered preventative surgery (Guilford *et al.*, 2010; Scheuer *et al.*, 2002), although this is not completely effective at preventing cancers and may impact on quality of life, depending on the cancer tissue types they are at risk of. Alternative treatment strategies based on molecular biology and other fields are being investigated, including immunological, endocrine, and targeted therapeutics, with a particular interest in treatments with specificity against cancer cells and wider applications (i.e., tolerable effective doses in applications as a chemopreventative or against advanced stage cancers).

#### **1.1.1.1 Genetics and Molecular Biology in Cancers**

Cancers involves dysregulation of genes with both somatic mutations or regulatory disruptions which accumulate during a patient's lifetime and germline mutations which predispose individuals to high-risk early onset cancers (American Cancer Society, 2017; Guilford *et al.*, 1998; NCI, 2015). Cancer is widely viewed to be a genetic disease due to these familial cancer syndromes, hereditary risk factors, and the molecular changes occurring in cancers, including numerous cancer genes which have been identified Stratton *et al.* (2009); Vogelstein *et al.* (2013). Cancer genes are generally classified into two classes: "oncogenes" which are activated in cancers driving tumour growth and invasion or "tumour suppressors" which are inactivated in cancers removing cellular regulation and genomic maintenance functions. The mutations which cause cancers accumulate with age and have been suggested to be inevitably coupled with aging due to the association of cancer incidence with the stem cell divisions in which mutations could occur across tissue types (Tomasetti and Vogelstein, 2015).

Hanahan and Weinberg (2000) identified several key molecular and cellular traits shared across most cancers as a rational approach to the complex change that occur

cancer initiation and progression due to common molecular machinery underlying all cells. A cancer cell must possess limitless replication potential, modulate growth signals to grow indefinitely, and gain invasive or metastatic capabilities. In addition, cancers must evade apoptosis, the immune system, and sustain angiogenesis and energy metabolism in order to survive (Hanahan and Weinberg, 2000, 2011). In order to achieve this, cancer cells undergo changes to their genomes and the surrounding cells to create a tumour microenvironment. Thus genomic instability has a key role in the survival and proliferation of cancer cells and the progression of further disease, as these malignant characteristics are acquired. Identifying the mechanisms of these acquired traits and the underlying genetic mutation or dysregulation behind them, such as E-cadherin mutation in metastasis or p53 mutation in genomic instability (Hanahan and Weinberg, 2000), will be an important step in understanding and inhibiting cancer with the next generation of genomically-informed treatments.

Molecular biological processes have particular importance in characterising breast cancers. Gene expression and regulatory signals confer cell identity and response to the environment. Therefore gene expression has been investigated with microarray technologies Perou *et al.* (2000), with “intrinsic subtypes” identified characterised by estrogen receptor, *HER2*, and basal, epithelial signalling. The expression profiles were similar across independent samples of the same tumour and between primary and metastatic tumours of the same patient. Thus expression profiles represent the molecular state of a tumour rather than the sample and the molecular configuration of the cells regulation is carried through the cellular lineage of during metastasis preserving the molecular subtype. These molecular intrinsic subtypes “luminal A”, “luminal B”, “*HER2*-enriched”, “basal-like”, and “normal-like” have been replicated across microarray studies (Hu *et al.*, 2006), with their relevance to prognosis (including predicting survival and response to neoadjuvant chemotherapy) demonstrated and a 50-gene subtype predictor from microarray and qPCR analysis has been provided (Parker *et al.*, 2009; Sørlie *et al.*, 2001). This has been further updated with the “claudin-low” subtype (Herschkowitz *et al.*, 2007) and stimulated further investigations into subtyping of breast cancers by molecular properties. Despite differences in subtyping performed by different research groups and companies, there is widespread agreement that distinguishing luminal, *HER2*-enriched, and triple negative tumours can be performed with expression profiles and have value in our understanding of cancer progression and prognostic importance for patients Dai *et al.* (2015). High-throughput technologies have the potential to enable such subtyping on a vast scale in discovery of further sub-

types in breast cancer or other diseases and in identification of these subtypes along with mutations in routine clinical diagnostic and prognostic testing. The “Pan cancer” approaches by the cancer genome atlas project (as discussed in more detail in Section 1.1.5.2.1) expand on the importance of molecular differences between cancers by examining molecular profiles across cancer tissue types (Weinstein *et al.*, 2013).

Cancer is a major health concern with a well-established genetic contribution, in risk and in the molecular changes occurring during progression (Stratton *et al.*, 2009). Many genes have been discovered to be important in different cancers with molecular differences between cancers, including alterations across the genome, being of clinical importance. As such cancers were among the first samples investigated with genomics following the sequencing of the human genome Dickson (1999) and continue to be the subject of genomics and bioinformatics investigations.

### 1.1.2 The Human Genome Revolution

The advent of the Human Genome sequence (Lander *et al.*, 2001) has transformed genetics research including the study of health and disease (Lander, 2011; Peltonen and McKusick, 2001). Systematic, unbiased studies across all of the genes in the genome are viable in unprecedented ways. The successful undertaking of such an international scientific megaproject has set an example for numerous initiatives to follow, including many genomics investigations expanding to species, to the functional, or to the population level (Collins *et al.*, 2003). These projects serve as excellent resource for genetics research globally, particularly for cancers where genomics investigation have been widely applied to different tissues across molecular profiles Bamford *et al.* (2004); Weinstein *et al.* (2013); Zhang *et al.* (2011) . Genome sequencing technologies continue to improve, drop in price, and become feasible in more research and for clinical applications.

#### 1.1.2.1 The First Human Genome Sequence

The first human genome is a good example of a large-scale genomics project for its success as an international collaboration and releasing their data as a resource for the wider scientific community (Collins *et al.*, 2003; Lander *et al.*, 2001). This particular project generated significant public interest due to it being a landmark achievement, the first of its scale, and some controversial findings. Namely, the number of genes discovered (particularly those specific to vertebrates) was much lower than most estimates of a genome of its size and the number of repetitive transposon elements was

very high. Even the figure of 30–40,000 genes given by the original publication is now regarded to be an overestimate (Ezkurdia *et al.*, 2014; IHGSC, 2004).

Accounting for the “complexity” encoded by the human genome with so few genes has led to investigations into molecular function, expression profiling, and population variation. When announcing the draft genome, Lander *et al.* (2001) concede that genomic information alone is not sufficient for biological understanding and that many investigations remain to be done, with their objective being to share the raw genome data so that it was available for further inquiry rather than interpreting it themselves. While genomics technologies and genomics projects have flourished since then, the need in turn for systematic means of interpreting data of such scale and for the interdisciplinary expertise to do so has only grown.

The “whole genome shotgun” approach (now widely used in genomics sequencing) was pioneered by a competing private genome project completed shortly afterwards by Celera Genomics, demonstrating the power and speed of this approach by sequencing 27 million reads of the entire 2.91Gbp human genome ( $5.11 \times$  coverage) in only 9-months (Venter *et al.*, 2001). Assembly was assisted with the  $2.9 \times$  coverage public genome data, reduced to raw shotgun reads to remove cloning bias. While, repetitive sequences remained an issue for this project, more than 90% of the genome was able to be assembled into 100kbp scaffolds and 26,588 protein coding genes were identified, closer to the current consensus for the number of genes in the human genome. This project in particular emphasised the value of computational assembly methods in handling a large number of reads, reducing the time and cost of sequencing, and established the shotgun approach for wider adoption with more recent sequencing technologies with shorter reads.

### 1.1.2.2 Impact of Genomics

Genomics has stimulated investigations into many of these previously largely explored areas of functional genetics and thus been of immense value in genetics research, attracting high expectations for further applications. Genomics research has become anticipated for its potential for widespread applications in healthcare, agriculture, ecology, conservation, and evolutionary biology, although many of these are yet to come to fruition.

Cancer research is an area of particularly high expectations for the clinical impact of genomics in oncology. Genomics technologies have potential applications across cancer diagnostics, prognosis, management, and developing treatment. Cancers are often involve genetic mutation or dysregulated gene expression which can be detected in a

genome or transcriptome with potential to improve patient care. While direct impact of genomics on the clinic has been limited, compared to initial expectations following the publication of the human genome, diagnostic cancer genes and therapeutic targets identified with genomics research have begun to be introduced in the clinic (Stratton *et al.*, 2009).

### **1.1.3 Technologies to Enable Genetics Research**

#### **1.1.3.1 DNA Sequencing and Genotyping Technologies**

Genotyping was once commonly performed on variable regions of the genome with restriction fragment length polymorphisms (RFLP) or repetitious microsatellite regions. These exploited sequence variation at target sites of restriction enzymes or measured the length of repetitious regions, using polymerase chain reaction (PCR), restriction enzymes, and gel electrophoresis to measure deoxyribonucleic acid (DNA) genotypes at particular sites. This is laborious and limited to well characterised variable regions of the genome, generally genes or nearby marker regions.

The Sanger (dideoxy) chain termination method (Sanger and Coulson, 1975) enabled DNA sequencing and genotyping at a widespread scale, being less technically difficult than the Maxam-Gilbert sequencing by degradation method (Gilbert and Maxam, 1973; Maxam and Gilbert, 1977), which required more radioactive and toxic reactants. The Sanger methodology has relatively long read length (particularly compared to early versions of more recent technologies), with read lengths of 500–700 base pairs accurately sequenced in most applications, usually following targeted amplification with PCR. Sanger sequencing by gel electrophoresis takes around 6–8 hours and has been further refined with the “capillary” approach to 1–3 hours and requiring less input DNA and reactants. The capillary approach has been scaled up to run in parallel from a 96 well plate, at 166 kilobases per hour. The 96 well parallel capillary method was one of the main innovations which made the first Human Genome Project feasible and was used throughout (Lander *et al.*, 2001). Due to the quality of the Sanger sequence reads and low cost, it is still widely used in smaller scale applications, clinical testing, and to validate the findings of newer approaches.

#### **1.1.3.2 Microarrays and Quantitative Technologies**

Real-time or quantitative PCR (qPCR (qPCR)) is another adaptation of genetic technologies to quantitatively study nucleic acids, often reverse transcribed “” or messenger “” to measure (relative) gene expression or transcript abundance. While numerous

quality control measures are required to correctly interpret a qPCR experiment, these have similarly become widely adopted as are still used for smaller scale experiments and as a “gold standard” for measuring gene expression (Adamski *et al.*, 2014). This also represents a shift in the application of PCR and sequencing technology, where the primary interest is quantifying the amount of input material (by the rate of amplification to a certain level) rather than the qualitative nature of the sequence itself. The more recent technologies of microarrays and RNA-Seq have similarly embraced this application to quantify DNA copy number, RNA expression, and DNA methylation levels. Due to results of comparable or arguably better quality from these newer technologies (Beck *et al.*, 2016; Git *et al.*, 2010; McCourt *et al.*, 2013; Robin *et al.*, 2016), this “gold standard” status has started to come under scrutiny.

Microarrays represent a truly high-throughput molecular technique, reducing the cost, time, and labour required to study molecular factors such as genotype, expression, or methylation across many genes, making it feasible to do so over a statistically meaningful number of samples. Microarrays are manufactured with probes which measure binding of particular nucleotide sequences to either quantitatively detect the presence of a sequence such as a single nucleotide polymorphism (SNP) or quantify DNA copy number, gene expression, or DNA CpG methylation. Microarray technologies have popularised “genome scale” studies of genetic variation and expression.

In addition to being more versatile and higher-throughput than PCR based techniques, microarrays are considered cost-effective, particularly when scaled up to large number of probes. They are also available with established gene panels or customised probes from a number of commercial manufacturers. These remained popular during the introduction of newer technologies due to reliability and this relatively lower cost, especially in large-scale projects involving many samples. However, microarrays have issues with signal-to-noise ratio, with both sensitivity to low nucleic acid abundance and “saturation” of probes at high abundance, edge effects, and requiring more starting material than qPCR. Thus qPCR is still used for many small gene panel studies.

#### **1.1.3.3 Massively Parallel “Next Generation” Sequencing**

Similar to microarrays, the introduction massively parallel sequencing technologies have further expanded the availability of high-throughput molecular studies to researchers, with corresponding availability of genomics data from these studies. This “Next Generation Sequencing” (NGS) expands not only gene expression studies (compared to microarrays) but extends to genome sequencing *de novo* for previously unknown genome and transcriptome sequences at an unprecedented scale. This has been a particu-

larly important technological revolution in genomics, as the cost and time of genome sequencing has dropped dramatically and enabled sequencing projects of far more samples and applications beyond the Human Genome Project. Particularly, when dealing with variants in a species with an existing reference sequence such as humans, where the computational cost of mapping to a reference over a genome assembly. However, the cost of sequencing (RNA-Seq) for gene expression or DNA methylation studies is still considerably higher than a microarray study (limiting feasible sample sizes).

Compared with arrays, NGS studies have additional challenges, particularly with large data and compute requirements to handle the raw output data. Compared the the established methods to analyse microarray data, handling NGS data can be more technically difficult. While methods developed for analysing microarray data can be repurposed for sequence analysis in many cases, more bioinformatics expertise is required particularly to handle the raw read data and changing approaches for various changes in sequencing technologies. One of the main computational challenges is the assembly reads or mapping to a reference genome due to the inherently small reads of most NGS technologies compared to the Sanger methodology. Furthermore, there are fewer software releases and best practices established specifically RNA-Seq data, thus many analyses are still conducted with customised analysis approaches and command-line tools. Compared to existing graphical tools or pipelines for microarray analysis, this is a more active technology for bioinformatics research with many applications of genomics data have yet to be explored.

However, the methodology itself has challenges with the sample preparation, requiring a relatively high quantity of input material and “contamination” with over abundant ribosomal rRNA taking up the majority of the sequencing if not purified correctly. This abundance of rRNA is a particularly important issue in microarray and RNA experiments in Eukaryotes where it is commonplace target the mRNA by binding to the poly-A tail (RNA-Seq) or 5’ cap (CAGE-Seq). However, this has the potential to exclude microRNAs (miRNA) and long non-coding RNAs (lncRNA) of interest unless the sample is prepared specifically to study these. Similarly capturing a subsection of the genome for exome analysis or reduced representation bisulfite sequencing (RRBS), focuses on sequencing DNA sequences and methylation levels of CpG sites near known genes to reduce cost, noise, and incidental findings.

In many cases, the benefits of NGS technologies over microarrays still outweigh the additional cost. NGS technologies have the advantage of greater potential accuracy and sensitivity than microarrays, depending on the sequencing depth or “coverage”,

theoretically sensitive down to the exact number of molecules for each transcript. NGS experiments are regarded as “reproducible” with no need for technical replicates, although these are still performed for a subset of samples in many projects for quality assurance purposes. NGS has a wider dynamic range than microarrays and is able to detect SNPs, InDels, and splice variants in addition to quantifying DNA copy number or transcript abundance. NGS scales to all genes and beyond for these molecular applications without having to design new probes as required for a microarray. Thus NGS technologies are not limited to genes already characterised sequence or functions, do not need to be updated with new probes for each genome annotation release, and do not require a reference genome at all for new species. A “transcriptome” can be assembled *de novo* for an expression study in any organism by sequencing the mRNA extracted from a cell.

#### **1.1.3.3.1 Molecular Profiling with Genomics Technology**

NGS is highly adaptable to different applications: DNA sequencing (whole genome or exome), DNA methylation (bisulfite-Seq), RNA-Seq, miRNAs, lncRNA, or chromatin immunoprecipitation (CHIP-Seq). Employing RNA-Seq to the transcriptome are a common adaptation, RNAs are reverse transcribed and sequenced from the resulting complementary “cDNA”. This is utilised to quantify the levels of RNA and identify which regions of DNA are expressed. Similar bisulfite treatment converts cytosine residues to uracil (sequenced as thymidine), sparing methylated cytosine enabling it to be distinguished with bisulfite-Seq for high-throughput detection of the notable epigenetic mark and is a common procedure to generate an epigenome. Subsets of the nucleic acid may be extracted for sequencing such the coding regions of DNA (for the “exome”), the mRNA 5’cap (CAGE-Seq), mRNA 3’poly-A tail (RNA-Seq), microRNA, or an enriched subset of variable regions for DNA sequencing (“genotyping by sequencing”) and methylation studies (“reduced-representation bisulfite sequencing). High-throughput gel and mass spectrometry techniques have been employed to proteins and metabolites to generate the proteome and metabolome respectively. These “omics” technologies are applicable across a wide range of biomolecules in a cell and these “molecular profiles” are produced in many experimental laboratories.

#### **1.1.3.3.2 Established Sequencing Technologies**

454 sequencing (acquired by Roche) commercially released from 2005 to 2013 was the first NGS technology, generating a vast 1 million reads per day or 400–600Mbp in a 10

hour run. This technology used the “pyrosequencing” method of sequencing by synthesis, detecting phosphates released when a compatible nucleotide reacts and extends the DNA synthesis of a complementary strand. This technology popularised NGS with the first complete genome from a single individual (Wadman and Watson, 2008; Wheeler *et al.*, 2008) and Neanderthal ancient DNA studies (Green *et al.*, 2009; Noonan *et al.*, 2006). While this technology was capable of reads up to 1kb, reads of 400–500bp were more typical and the technology had difficulties with accurately processing runs of repeated bases (Rothberg and Leamon, 2008). These are still relatively long reads for an NGS technology but it has been discontinued due to competing short read technologies being more cost-effective with lower running costs.

SOLiD sequencing (acquired by Life Technologies and then Thermo Fisher) released in 2006 employed a vastly different approach to NGS, using labelled dinucleotide pairs for “sequencing by ligation” to produce a highly accurate sequence (99.94%) with built-in error correction by sequencing two reading frames and is unaffected by consecutive bases. This technology is also high-throughput, producing 1200–1400 million reads (66–120Gbp) in a 7–14 day run (ThermoFisher, 2017b). However, SOLiD sequencing does not cope well with palindromic sequences and SOLiD reads are very short only 35bp, making it more difficult to assemble them.

Illumina sequencing (developed by Solexa and later acquired by Illumina) was also released in 2006. It utilises reversible terminating dyes to sequence by synthesis with a lower accuracy (98%) and read lengths of 150–250bp. Illumina more than makes up for relatively short reads (along with improving the read length of the technology) and low accuracy with high-throughput and cost effectiveness, with a Hi-Seq 4000 platform producing up to 10 billion paired-end reads (1500Gbp) in a run of appropriately 3 days, capable of sequencing 12 human genomes ( $30\times$  coverage) or 100 human transcriptomes simultaneously (Illumina, 2017). Illumina has further reduced the cost of sequencing with the economies of scale with the Hi-Seq X 10 claiming to produce a human genome (with  $30\times$  coverage) for less than US\$1000, the first platform to achieve this long-standing goal in genomics. The high-throughput of Illumina sequencing also makes deep sequencing for high coverage, high quality consensus reads, and sensitive RNA-Seq experiments feasible. Illumina sequencing now has a dominating market share of the NGS technologies.

### 1.1.3.3.3 Emerging Sequencing Technologies

Ion Torrent (also acquired by Life Technologies) released in 2010 employs “sequencing by synthesis” but in a drastically different way with ion semiconductor sequencing, detecting  $H^+$  ions released when bases during DNA synthesis. Without the use of optical detection, the Ion Torrent system is compact offering rapid, cost-effective sequencing with the potential to scale with the future development of silicon semiconductors which have historically doubled in density every 2 years (Moore’s Law). It is capable of reads of 100–200bp in only an hour (as fast as 4 seconds per base) and up to 400bp in a 2 hour run with an accuracy of 99.6% (dropping to 98% for consecutive sequences of 5 bases). While fast, cost effective, and accurate, Ion Torrent has short reads and modest throughput (up to 10 Gp for the Ion Proton and 15 Gb for the Ion S5 XL systems) compared to other sequencing technologies (ThermoFisher, 2017a).

Pacific Biosciences (PacBio) released the RS and RS II platforms in 2010 and 2011 to make up for the short reads in NGS technologies with the single molecule real time (SMRT) approach capable of long read lengths, averaging between 2.5–7kb and up to 80kb PacBio (2017). The PacBio methodology traps each molecule in a zero mode waveguide (ZMW) and sequences it in real time. The RS II has 150,000 ZMW and an output of 500Mbp–1Gbp per SMRT cell (doubling that of the RS), with the capacity to run up to 16 concurrently for 0.5–6 hours. While the single molecule sequencing approach has strengths in sensitivity and potential to detect 3D structures, such as G-quadruplexes, this has the drawback of slowing down the sequencing and reducing the throughput of the platform. Another issue is sequence quality with the raw data as poor as 20–30%. However, PacBio recommends specific software to assemble as consensus with 99.999% for sequences with over 20 $\times$  coverage, regardless of sequence repeats or GC composition. Despite concerns over data quality and higher cost than other approaches, the long reads are appealing for genome assembly and in many genome studies combine PacBio reads with more accurate short read technologies. However, due to the poor separate quality of reads this technology may not be appropriate for RNA-Seq studies, while it does have the potential for high sensitivity and detecting alternative splicing were it be improved. PacBio has recently released the Sequel (2016) system, increasing the throughput of the SMRT Cells 7 $\times$  to 1 million ZMW holes with an output of 5–10Gb for each of 16 SMRT cells.

Nanopore sequencing is another technology capable of long reads in real time and direct single molecule sequencing, avoiding amplification bias, detecting modified bases and directly sequencing RNA molecules. This also reduces laboratory preparation

times. Nanopores work by measuring the ion current through a pore in a electrically insulating membrane as a nucleic moves through it. Oxford Nanopore has been developing this technology since 2005, launching the MinION in 2014 which employs biological nanopores: a transmembrane protein through which DNA or RNA passes, blocking ion current differently for each base. Each pore sequences in real time, capable of sequencing 450bp per second (Nanopore, 2017). However, there are quality issues with each individual read with quality estimates varying between 87–98%, with improvements to the quality of detection accounting for significant delays in the release of this technology. The MinION makes up for the is a capacity for extremely long reads, averaging 5.4kbp (Hayden, 2014) up to a maximum of 200Kbp and being a portable platform with very few overhead costs. While the MinION is limited in scale with only one flow cell of 512 pores (5–10Gbp), the PromethION being released in early access in 2016 scales this technology with flow cells of 3000 pores and the capacity to run 48 (up to 4 samples each) in parallel for 144,000 long reads with a versatile, modular system including built-in computing resources. One of the main issues with Oxford Nanopore systems is accuracy, with the manufacturer suggesting the use of consensus sequences for higher accuracy as PacBio does. The main source of this pore accuracy is the width of biological pores resulting in several bases being in the pore at any one time, inferring the sequence from the ion currents of each respective combination of bases and distinguishing them is a major technical challenge.

Quantum Biosystems in Japan is developing a synthetic nanopore system to address this issue. While the technology is still in development, it has the potential to produce similarly long reads, with a high-throughput, low running cost, and rapid run time (Quantum Biosystems, 2017). The technical challenges to develop a nanotechnology capable of this are immense but such developments serve as but one of example of how sequencing technologies may continue to improve, becoming more feasible for a wider variety of applications.

Due to such benefits of sequencing over previous technologies (and their continued refinement), this thesis has focused on gene expression data generated by RNA-Seq rather than microarrays. RNA-Seq data is widely available as a resource from large-scale cancer genomics projects and methods to make inferences from RNA-Seq experiments could feasibly be applied to many other studies based on these current (or similar future) technologies.

#### **1.1.3.4 Bioinformatics as Interdisciplinary Genomic Analysis**

Genomics technologies have given rise to data at a scale previously rarely encountered in molecular biology, making inference with conventional techniques difficult. Computational, Mathematical, and Statistical skills are required to handle this data effectively, in addition to biological background to frame and interpret research questions. Drawing upon these disciplines to handle biological data has become the field of “Bioinformatics”, focusing specifically on making inferences from genomics and high-throughput molecular data or developing the tools to do so. This contrasts with the existing fields of “theoretical” or “computational biology” which existed prior to genomics data, focusing on modelling and simulating aspects of biology without necessarily addressing the genomics data or detecting the phenomena in nature, extending beyond genetics to cell modelling, neuroscience, cancer development, ecology, and evolution.

In practice, many researchers identify with both bioinformatics and computational biology, or draw upon the findings and methods of the other field. This thesis uses many approaches in bioinformatics to biological research questions and established mathematical or bioinformatics resources.

Gene expression analysis is the focus of many bioinformatics research groups, drawing upon statistical approaches to appropriately handle microarray and RNA-Seq data along with making biological inferences from a large number of statistical tests. This presents various challenges from normalising sample data and accounting for batch effects to developing or applying statistical tests tailored to biological hypotheses and testing them at a genome-wide scale, generally across thousands of genes. There are numerous approaches for dealing with these challenges, some of which will be described in Chapter 2.

#### **1.1.4 Follow-up Large-Scale Genomics Projects**

A number of projects have attempted to follow up on the human genome project to varying degrees of success. The genomes have since been sequenced for a variety of model organisms, organisms of importance in health, agriculture, metagenomics of microorganisms (microbiome), ecology and conservation. Genomics projects have also been applied functional genetics (Kawai *et al.*, 2001; ENCODE, 2004) and to human populations with an interest variability between individuals and health or disease risk (HapMap, 2003; 1000 Genomes, 2010).

Other genomics databases have focused on facilitating distribution of genomic data generated by researchers, rather than generating it themselves. Genbank (NCBI) in the

US, EMBL in Europe, and the DDBJ (NIG) in Japan do so by serving as repositories of DNA sequence data. GEO (Clough and Barrett, 2016), arrayExpress (Rustici *et al.*, 2013), and caArray (Heiskanen *et al.*, 2014) serve a similar purpose as a resource for gene expression datasets, originally developed for microarray data but RNA-Seq data is now supported by some platforms. They are repositories for researchers to deposit, share, and access gene expression data, which serve as a resource to support ongoing research to utilise data for genes of interest to particular research groups and further to make inferences based on larger datasets than accessible to any individual laboratory (Rung and Brazma, 2013). These resources cover not only DNA sequence across the genome but also molecular profiles of other factors by adapting genomic sequencing or other high throughput technologies for quantifying gene expression or DNA methylation. Sharing the expression datasets generated in a publication is now required by some journals.

Similarly, international projects and consortiums have begun to release data gathered using common agreed upon protocols in laboratories across the world, often hosting public databases of these themselves, publishing their own investigations into the datasets as they are released, or offering basic searches and analytics of the data via a web portal. These databases include many of the genomics projects discussed above and the cancer-specific projects discussed below. In many ways, the quality, consistency, and accessibility of these international projects has become more appealing than accessing smaller studies, particularly for gene expression datasets where the more recent, larger projects have switched from microarray to RNA-Seq technologies. This distinction will also be discussed later.

### 1.1.5 Cancer Genomes

It's importance in the future of cancer research was noticed, even in the early days of genomics (Dickson, 1999). The Cancer Genome Project (CGP) based at Wellcome Trust Sanger Institute in the UK were among the first to launch investigations into cancer after the publication of the Human Genome, using this genome sequence, consensus across the cancer research literature, and sequencing the genes of cancers themselves. Initially, the Sanger Institute set out to sequence 20 genes across 378 samples while the Human Genome project was still ongoing (Collins and Barker, 2007), optimising sequencing and computation infrastructure for a larger project while doing so. The main aim of the Cancer Genome Project was to discover “cancer genes”, those frequently mutated in cancers by comparing the genes of cancer and normal tissue samples, both

“oncogenes” and “tumour suppressors” which are activated and inactivated respectively in cancers. This project is ongoing and the UK continues to be involved in international sequencing initiatives and those focused on particular tissue types.

The Sanger Institute also hosts the Catalogue of Somatic Mutations in Cancer (COSMIC, 2016), a database and website of cancer genes. This launched with 66,634 samples and 10,647 mutations from initial investigations into *BRAF*, *HRAS*, *KRAS*, and *NRAS* (Bamford *et al.*, 2004). It has since expanded to include 1,257,487 samples with 4,175,8787 gene mutations curated from 23,870 publications, including 29,112 whole genomes (COSMIC, 2016). This database now also identifies cancer genes from DNA copy number, differential gene expression and differential DNA methylation.

#### **1.1.5.1 The Cancer Genome Atlas Project**

Based in the US, the Cancer Genome Atlas (TCGA) project was established in 2005, a combined effort of the National Cancer Institute (NCI) and the National Human Genome Research Institute (NHGRI) of the National Institutes of Health (NIH) (TCGA, 2017). They first set out to demonstrate the pilot project on brain (McLendon *et al.*, 2008), ovarian (Bell *et al.*, 2011), and squamous cell lung (Hamerman *et al.*, 2012) cancers. In 2009, the project expanded aiming to analyse 500 samples each for 20-25 tumour tissue types. They have since exceeded that goal, with data available for 33 cancer types including 10 “rare” cancers, a total of over 10,000 samples.

The TCGA projects set out to generate a molecular “profile” of the tumour (and some matched normal tissue) samples: the genotype, somatic mutations, gene expression, DNA copy number, and RNA methylation levels. While these were originally performed largely with microarray technologies, exome and RNA-Seq has been since adopted and performed for many TCGA samples, with whole genomes being performed for some samples. Data which cannot be used to identify the patients (such as somatic mutation, expression, methylation, and various clinical factors) are publicly available.

#### **1.1.5.2 The International Cancer Genome Consortium**

TCGA and the Cancer Genome project in the UK are part of a larger International Cancer Genome Consortium (ICGC), now a concerted effort across 16 countries to sequence the genome, transcriptome, and epigenome of 50 tumour types from over 25,000 samples total (Zhang *et al.*, 2011). With some redundancy the following countries are profiling various tumour types: USA (including TCGA), China (16), France (10), Australia (4), South Korea (4), the UK (4), Germany (4), Canada (3), Japan (3), Mexico (3 in collaboration with the US), Singapore (2), Brazil, India, Italy, Saudi

Arabia, and Spain. This is inherently international and several projects are collaborations, such as between the USA and Mexico, Australia and Canada, Singapore and Japan, along with the UK and France representing the European Union (ICGC, 2017). In order to avoid competing the existing TCGA projects, some countries focus on a particular cancer they have health interest: Australia (melanoma), Brazil (melanoma), India (oral), Saudi Arabia (thyroid), and Spain (CML). Others focus on a particular tissue subtype with poor prognosis: The UK (triple negative or Her2+ breast cancer), France (clear cell kidney), Australia and Canada (ductal Pancreas). Another approach is to focus on rare or child cancers: Canada, Italy, France, Germany, Japan and Singapore, and the US (TARGET project). Particularly countries in Asia (China, Japan, Singapore, and South Korea) have emphasised the value of adding tumour data from non-Western countries or non-European populations in addition the data from Europe and the TCGA in the US. Data from 9 of these countries is already available on the ICGC website with the project ongoing.

#### 1.1.5.2.1 Findings from Cancer Genomes

The cancer genome atlas pilot projects (Bell *et al.*, 2011; Hamerman *et al.*, 2012; McLendon *et al.*, 2008) serve to demonstrate the power of applying genomics technologies to cancer research at such as scale. In addition to sequence the whole genome or a subset (exome), DNA copy number, gene expression, DNA methylation, and somatic mutations were also analysed. The initial projects used microarray technologies for expression and methylation data but these have since been replaced by RNA-Seq for expression. TCGA demonstrated the potential discovery of the molecular basis of cancer by analysing 206 glioblastoma brain cancer samples (McLendon *et al.*, 2008), highlighting the roles of *ERBB2*, *NF1*, *TP53*, and *PIK3R1* mutations, along with altered methylation of *MGMT*, and the core pathways of RTK, p53, and RB signaling in brain cancer. An analysis of 489 serious ovarian cancers (Bell *et al.*, 2011) similarly reported *TP53* mutations specifically over-represented in high grade tumours and reported 133 copy number variants, 168 differentially methylated regions, and recurrently somatic mutations in 9 genes in low grade tumours including *NF1*, *BRCA1*, *BRCA1*, *RB1*, and *CDK12*. Four transcriptional subtypes of ovarian cancers were identified, alterations in *BRCA1*, *BRCA2*, and *CCLE* had an impact of patient survival, and the homologous recombination, NOTCH and FOXM1 signaling pathways were involved in ovarian cancer growth. The genomics of 178 squamous cell lung cancers (Hamerman *et al.*, 2012) were highly complex, averaging at 360 mutations in coding regions. While

no targeted therapies existed for this cancer subtype, 11 recurrently mutated genes were identified including *TP53* and *HLA-A*. The pathways altered in various squamous cell lung cancers were NFE2L2, KEAP1, differentiation genes, PI3K, CDKN2A and RB1. These aberrant genes and pathways represent potential therapeutic targets which could be identified for most samples.

The TCGA breast cancer analysis (TCGA, 2012) consisted of 802 samples with exomes, copy number variants, RPPA protein quantification, and DNA methylation, mRNA, and microRNA arrays with 97 whole genomes sequenced. Four main molecular classes were identified to subtype the samples, despite considerable heterogeneity between samples. Recurrent mutations across more than 10% of samples were identified in *TP53*, *PIK3CA*, and *GATA*. TCGA further suggests subtypes by HER2 and EGFR protein levels. In a further analysis of 817 breast cancer samples including 127 invasive lobular breast and 88 mixed type samples (Ciriello *et al.*, 2015), 3 molecular subtypes of lobular breast cancer were identified. Lobular breast cancer was also characterised by recurrent mutations in *CDH1*, *PTEN*, *TBX2*, and *FOXA1*/

TCGA reported results of colon and rectal cancers in a combined analysis of 267 samples (Muzny *et al.*, 2012), finding no genomic distinction between colorectal cancers. Apart from 16% of hypermutated colorectal cancers, the remaining samples were very similar at the molecular level with 24 significantly recurrently mutated genes identified. These include the expected *APC*, *TP53*, *SMAD4*, *PIK3CA*, and *KRAS* genes. Additionally, novel recurrent mutations were identified in *ARID1A*, *SOX9*, and *FAM123* along with recurrent copy number alterations in *ERBB2* and *IFG2*. Thus the molecular findings of colon and rectal tumours can be applicable across colorectal cancers, including the known characteristics of microsatellite instability (MSI) and CpG island methylator phenotype (CIMP) found in some colorectal tumours.

The TCGA stomach cancer analysis of 295 samples (Bass *et al.*, 2014) identified 4 molecular subtypes of stomach cancers characterised by: the Epstein-Barr virus, MSI, genomics instability, and chromosomal instability. Aberrations in *PD-L1*, *PIK3CA*, and *JAK2* were also identified in stomach cancers which may present therapeutic targets.

#### 1.1.5.2.2 Genomic Comparisons Across Cancer Tissues

TCGA have identified various genes as recurrent, driver mutations across cancer types which are likely to have a role in driving the proliferation of these cancers and present a molecular target that could be applied across tissue types. These include *TP53*

(in brain, lung/head/neck squamous cell, breast, colorectal, uterine, and endometrial cancers), *ERBB2*/HER2/NEU (in brain, breast, colorectal, bladder, and lung cancers), *PIK3CA*, *PIK3R1* (in brain, breast, colorectal, endometrial, bladder, clear cell renal, and lung cancers), *BRCA1*/*BRCA2* (in breast and ovarian cancers), *NF1* (in brain, ovarian, and skin cancers), *ARID1A* (in colorectal, endometrial, and clear cell renal cancers), *KRAS* (in colorectal, endometrial, and skin cancers), *BRAF* (in colorectal, thyroid, and skin cancers), *EGFR* (in brain, breast, and lung cancers), and *PTEN* (in breast, endometrial, and uterine cancers) (Agrawal *et al.*, 2014; Akbani *et al.*, 2015; Bass *et al.*, 2014; Bell *et al.*, 2011; Burk *et al.*, 2017; Cherniack *et al.*, 2017; Ciriello *et al.*, 2015; Collisson *et al.*, 2014; Creighton *et al.*, 2013; Hammerman *et al.*, 2012; Kandoth *et al.*, 2013; Lawrence *et al.*, 2015; McLendon *et al.*, 2008; Muzny *et al.*, 2012; TCGA, 2012; Weinstein *et al.*, 2014). In addition to disregarding the distinct between colon and rectal cancers based on molecular similarity (Muzny *et al.*, 2012), the TCGA project have observed differences within tumour types and proposed molecular subtyping for breast, clear cell renal, papillary renal, stomach, skin, bladder, and prostate cancers (Abeshouse *et al.*, 2015; Akbani *et al.*, 2015; Bass *et al.*, 2014; Ciriello *et al.*, 2015; Creighton *et al.*, 2013; Hammerman *et al.*, 2012; Linehan *et al.*, 2016; Muzny *et al.*, 2012; TCGA, 2012; Weinstein *et al.*, 2014).

The “Pan cancer” project (Hoadley *et al.*, 2014; Weinstein *et al.*, 2013) analysed 3527 samples across 12 tissue types for DNA, RNA, protein, and epigenetic molecular profiles. This project was initiated in 2012 to perform a comprehensive analysis of molecular data across cancer types to identify molecular similarities and differences. Recurrent *TP53* mutations characterised high grade tumours across breast, ovarian, and endometrial cancers. HER2 was identified in brain, endometrial, bladder, and lung cancers, in addition to the known role of HER2 in breast cancers. *BRCA1* and *BRCA2* mutations were also detected across cancers, mainly breast and ovarian cancers as expected. Microsatellite instability characterised both endometrial and colorectal cancers. The Pan cancer project (Hoadley *et al.*, 2014) has identified 11 molecular subtypes across these tissues, 5 of corresponding to tissue cancer types and the remainder reassigned due to molecular similarities shared across cancer types. Squamous cell lung, head, and neck and a subset bladder cancers were grouped together by molecular similarities, characterised by a high frequency of *TP53* mutations. Conversely, bladder cancers were divided into 3 of these molecular subtypes with distinct profiles. This project further supports the genomic stratification of patients, demonstrated in breast cancer (Parker *et al.*, 2009; Pereira *et al.*, 2016; Perou *et al.*, 2000), which may apply

to other cancer types and to molecular characteristics across them targeting recurrent mechanisms of cancer growth and progression (Hanahan and Weinberg, 2000, 2011).

#### **1.1.5.2.3 Cancer Genomic Data Resources**

While the findings from the TCGA projects themselves are a considerable contribution to understanding cancer biology within and across tissue types, the main eventual benefit of such projects will be the availability of the data for the research community to analyse further and use to inform future investigations (McLendon *et al.*, 2008; TCGA, 2017; Weinstein *et al.*, 2013). These serve as a vast resource of common and rare cancer types and are publicly available to analyse further (cBioPortal, 2017; TCGA, 2017; Zhang *et al.*, 2011). This also applies to the Molecular Taxonomy of Breast Cancer International Consortium (METABRIC) project which focuses on breast cancer which also aimed to identify novel molecular subtypes (Curtis *et al.*, 2012). They performed an analysis of 2433 breast cancer samples with long-term clinical data, gene expression, copy number variants, and 173 genes sequenced which identified 40 driver mutations in breast cancer in addition to further support for molecular subtyping to identify patient groups with different clinical outcomes (Pereira *et al.*, 2016).

### **1.1.6 Genomic Cancer Medicine**

There is much anticipation in cancer research for genomics technologies to have a clinical impact in cancer medicine: from diagnosis and prognosis to treatment developments and strategies. These may result either from direct use of genome or RNA-Seq in clinical laboratories or indirectly from biomarkers and treatments developed with research facilitated by genomics. This second strategy is likely to have a more immediate patient benefit due to the cost of genome sequencing, particularly considering adoption in public healthcare systems with a limited budget.

#### **1.1.6.1 Cancer Genes and Driver Mutations**

There are two main categories of “cancer genes” (Futreal *et al.*, 2001). Oncogenes are those activated in cancers either by gain of function mutations in proto-oncogenes, amplification of DNA copies, or elevated gene expression. Their normal functions are typically to regulate stem cells or to promote cellular growth and recurrent mutations are typically concentrated to particular gene regions. Conversely, tumour suppressor genes are those inactivated in cancer either by loss of function mutations, deletion of DNA copies, repression of gene expression, or hypermethylation. Their normal functions are typically to regulate cell division, DNA repair, and cell signalling.

Detecting these cancer genes is a major challenge in cancer biology and has been revolutionised by genomic technologies. Recurrent mutations, or DNA copy number variants and differential gene expression or DNA methylation are all indicative of cancer genes (Mattison *et al.*, 2009), which can be detected in genomics data (Pereira *et al.*, 2016; Weinstein *et al.*, 2013). Important “driver” cancer genes (Stratton *et al.*, 2009) are difficult to detect from “passenger” mutations due to patient variation, tumour heterogeneity, and genomic instability. However, many cancer genes have been replicated from previous studies or well supported from genomics data. There remains the challenge of translating the identification of cancer genes to patient benefit with characterisation of variants of unknown significance, which mutation or gene expression markers can be used to monitor tumour progression or treatment response, and design of therapeutic intervention against many molecular targets for which they have yet to be developed or repurposed from other disease to cancers.

Driver mutations can be identified by whether they co-occur or are mutually exclusive with mutations in other genes in cancers, are recurrently mutated across a significant proportion of samples for a specific tissue type, or if mutations are recurrent across different cancer tissue types (cBioPortal, 2017; Pereira *et al.*, 2016; COSMIC, 2016; Weinstein *et al.*, 2013; Zhang *et al.*, 2011). Approximately 140 driver mutations have been identified, including many novel genes in particular cancers from genomics studies, with 2–8 in typically occurring in each tumour usually affecting cell fate, survival, or genome maintenance (Vogelstein *et al.*, 2013).

#### **1.1.6.2 Personalised or Precision Cancer Medicine**

The notion of using a patient’s genome to tailor healthcare to an individual has been appealing since the advent of genomics, popularised with the term “personalised medicine”. This approach was expected to span from preventative lifestyle advice to effective treatments. Personalised medicine was intended contrast with current strategies of health advice, screening, prognostics, and treatments based on what works well with the majority of the population, highlighting that adverse effects of treatments occur in a significant subpopulation and that many clinical studies are dominated by Western populations of European ancestry and may not generalise to other populations.

While the importance of genomics is still recognised in translational cancer research, its potential has been emphasised particularly in molecular diagnosis, prognosis, and treatments of patients already presenting with cancers in the clinic rather than preventative medicine. This is in part due to the vast number of variants of unknown clinical significance, the ethical issue of reporting on incidental findings, and the reg-

ulatory issues direct-to-consumer genetics companies have encountered offering health risk assessment.

More recently the term “genomic medicine” has been preferred to describe the paradigm of treating cancers by their genomic features, particularly grouping patients by the mutation, expression, or DNA methylation profiles of their cancers. Radical proponents advocate for these molecular subtypes to supersede tissue or cell type specific diagnosis of cancers. However, in practice they are often used in combination, with clinical and pathological factors being informative of prognosis and surgical training specialising by organ system. The related term of “precision medicine” also stems from this trend with the rationale to target these molecular subtypes with separate treatment strategies, particularly in developing and applying treatments targeted against a particular mutation specific to cancers. To this end much research in this field is focused on identifying mutations and gene expression signatures amenable to distinguishing cancers, particularly oncogenic driver mutations, and developing treatments against them.

#### **1.1.6.2.1 Molecular Diagnostics and Pan-Cancer Medicine**

There is growing support for the use of molecular tools such as mutations or gene expression signatures to diagnose tumour subtypes in replacement or addition to tissue of origin or histology. This is particularly important in breast cancer where analysis of molecular data detected several distinct “intrinsic subtypes” with differences in malignancy and patient outcome which were distinguished by molecular mechanisms rather than tissue or cellular phenotype (Parker *et al.*, 2009; Perou *et al.*, 2000). Conversely, common molecular mechanisms may be shared between cancers across tissue types as discovered by the “Pan cancer” studies, such as those conducted by the TCGA and ICGC projects, which combined molecular profiles across tissue types Weinstein *et al.* (2013). The molecular subtypes could feasibly be included in clinic testing as a panel of biomarkers for diagnostics and prognosis. Such biomarkers also have the potential to monitor drug response or risk of recurrence. This is also raises the need for development of treatments for targeting these molecular subtypes.

#### **1.1.6.3 Targeted Therapeutics and Pharmacogenomics**

Targeted therapies with specificity against a molecular target are emerging as precision cancer medicine. Molecular targets can be tested in laboratory conditions with RNA interference or pharmacological agents. Identification of molecular targets is important

for developing novel anti-cancer treatments along with validation and drug testing. For oncogenic mutations, the recurrent mutant variant or overexpressed gene is directly inhibited using structure-aided drug design or compound screening. However, oncogenes with high homology to other genes or tumour suppressor genes (where lost in cancers) are not amenable to direct targeting (Kaelin, Jr, 2009).

Despite controversy over their prohibitively high cost (PHARMAC, 2016), targeted therapeutics have been applied as monoclonal antibodies against oncogenes (such as *HER2*) with relative success in clinical trials (Miles, 2001), generating considerable interest in wider application of this approach. Targeted therapeutics have potential to have applications across cancer tissue types, specificity against tumour cells, wide therapeutic windows, and combination therapies (even in advanced disease or as a chemopreventative in high-risk individuals).

#### 1.1.6.3.1 Targeting Oncogenic Driver Mutations

Oncogene targeted therapies have also been developed with some examples of effective clinical application against cancers. However, they already begun to manifest problems with resistance, recurrence, tissue specificity, and design of inhibitors specific to oncogenic variants rather than proto-oncogene precursors. Targeted anticancer therapeutics can exploit complex interactions to distinguish normal and cancerous cells which may benefit from studies of gene regulation or interaction networks. The unexpected synergy between inhibitors of the oncogenes *BRAF*<sup>V600E</sup> and *EGFR* in colorectal cancer is an example of such a system Prahallad *et al.* (2012).

Despite successful application of vemurafenib against *BRAF*<sup>V600E</sup> in melanomas Dienstmann and Tabernero (2011); Ravn and Matalka (2012), colorectal cancers with *BRAF*<sup>V600E</sup> mutations have poor prognosis and lack drug response. Prahallad *et al.* (2012) used an RNAi screen and found that *EGFR* inhibition is synergistic with vemurafenib against *BRAF*<sup>V600E</sup> in colon cell lines and xenografts due feedback activation of *EGFR*. Vemurafenib which induced rapid reactivation of MAPK/ERK signalling via *EGFR* in colorectal cell lines in a tissue-specific manner Corcoran *et al.* (2012), although these may be relevant to acquired resistance in melanoma Sun *et al.* (2014). Thus combination therapies against several molecular pathways may be necessary to anticipate acquired resistance Ravn and Matalka (2012) and targeted therapeutics may be further refined from understanding the pathway structure and functional interactions cancer cells.

#### 1.1.6.4 Systems and Network Biology

It is also important to consider that driver mutations in oncogenes and tumour suppressor genes do not occur in isolation. The genetic interaction, regulatory and cellular signaling, and metabolic reactions of are all inter-related and may each be perturbed by aberrations in gene function occurring in cancers. These relationships can be represented by biological networks, mapping pairs of genes with a particular relationship. Due to the complexity of a cell, these molecular networks are very large consisting of thousands of nodes such as genes or proteins.

The properties of large networks were first studied by constructing random networks by randomly linking a fixed number of nodes (Erdős and Rényi, 1959, 1960). Despite the random nature of these networks, properties such as their connectivity were well characterised. The vertex degree (number of partners for each node) of random network follows a Poisson distribution, however this property does not hold in nature, suggesting that natural networks are non-random or not formed in this way Barabási and Oltvai (2004).

This work formed the foundation for studying complex networks (van Steen, 2010), which model features of real world networks not found in Erdős and Rényi's random networks (Erdős and Rényi, 1959, 1960). The small world property, made popular by findings in social networks (Travers and Milgram, 1969), is the remarkably short path lengths between any nodes in a small world network. A small world network is well-connected with a characteristic path length (the average length of shortest paths between all pairs of nodes) proportional to the logarithm of the number of nodes. Watts and Strogatz (1998) developed a model of random rewiring of a regular network to construct random networks with the small world property and a high clustering coefficient. While these properties are more representative of networks occurring in nature, their model is limited by the degree distribution which converges to a Poisson distribution as it is rewired Barrat and Weigt (2000).

The vertex degree distribution of naturally occurring networks often follows a power law distribution with the majority of nodes having far fewer connections than average and a small subset of highly connected network 'hubs' Barabási and Albert (1999). Hubs further differentiate into 'party' hubs (which interact simultaneously with many partners) and 'date' hubs (which interact with different partners in different conditions) Han *et al.* (2004). Network hubs can also be classed as associative or dissociative depending on whether they tend toward or away from connecting directly to other network hubs (van Steen, 2010). The associative and dissociative properties can also

be used to test whether nodes of a particular subgroup (e.g., gene function) associate with each other.

Barabási and Albert (1999) constructed a network model in an entirely different way to randomly generate scale-free networks which have a power law degree distribution. They constructed random networks by preferential attachment, modelling growth of a network by sequentially adding nodes with links to existing nodes. The scale-free nature of the random networks was ensured by adding new nodes with an increasing probability of attachment to an existing node if it has higher degree. These networks successfully capture the scale-free nature of many real world networks with short characteristic path length and low eccentricity resulting in super small worlds Barabási and Albert (1999). Scale-free networks are limited by a low clustering coefficient and lack of modular structure; however, they have enabled the study of scale-free network topology and served as a basis for modified scale-free models (Dorogovtsev and Mendes, 2003; Holme and Kim, 2002).

Han *et al.* (2004) observed dynamic modularity in biological networks and suggested the network structure may underpin genetic robustness and plasticity. They focus on network hubs which are more likely to be essential genes and define the subgroups of hubs based on correlation of gene expression with protein-protein interaction partners: ‘party’ hubs (which interact simultaneously with many partners) and ‘date’ hubs (which interact with different partners in different conditions). Party and date hubs occurred most frequently within and between network modules respectively. Party hubs were considered local regulators, whereas date hubs were considered important to network connectivity as global regulators. This distinction between classes of network hubs was supported by differences in tissue specificity and clinical relevance as a proposed predictor of clinical outcome in breast cancer with an AUROC of 0.784 Taylor *et al.* (2009). However, correlation between expression and protein interactions were not robustly reproduced. The importance of date hubs has been criticised for assuming a bimodal distribution and basing the global importance of data hubs on a small subset Agarwal *et al.* (2010). As an alternative interpretation, (Agarwal *et al.*, 2010) suggest the importance of interactions rather than network hubs as interactions important to the network were between functionally similar proteins. Network hubs can also be classed as associative or dissociative depending on whether they tend toward or away from connecting directly to other network hubs (van Steen 2010). The associative and dissociative properties can also be used to test whether nodes of a particular subgroup (e.g., gene function) associate with each other.

Applications of network theory are diverse, including uses in social sciences, engineering, and computer science. Due to their complexity and difficulty of gathering sufficient empirical data, biological applications of network theory are relatively unexplored. High-throughput technologies such as siRNA screens, two-hybrid screens, microarrays and massively parallel sequencing have made generating genome-scale molecular data feasible and enabled analysis of biological networks at the molecular level. Many types of inter-related molecular networks can be constructed and analysed, depending on the biological application. Genetic interaction networks will be the focus of this project because they are relatively unexplored compared to other molecular networks, have potential for applications in drug discovery (particularly cancer treatment), and may lead to better understanding of the role of genetics in cellular function and disease. Genetic interactions are usually studied at a high-throughput scale in simple model organisms such as bacteria, yeasts or the nematode worm; studies in humans, mammals, and non-model organisms (where applications would have the most societal impact) are limited by cost, time and labour constraints. Computational approaches with effective predictive models are the only feasible approach to study the connectivity of a biological network in a complex metazoan cell at the genome-scale.

#### **1.1.6.4.1 Network Medicine, and Polypharmacology**

Molecular networks are biological networks consisting on biological molecules including genes, transcripts (with non-coding and microRNAs), or proteins related by known interactions and gene regulatory or metabolic pathways. Targeted therapeutics have had some success for drug discovery, particularly in anticancer applications, including exploiting these molecular networks by designing combination therapies and applying a network pharmacology framework Hopkins (2008). Rational design of drugs selective to a single target has often failed to deliver clinical efficacy. Many existing effective drugs modulate multiple proteins, having been selected for biological effects or clinical outcome rather than molecular targets. Proponents of network biology and polypharmacology (specific binding to multiple targets) recommend to develop drugs with a desired target profile designed for the target topology Barabási and Oltvai (2004); Hopkins (2008). Multi-target treatments aim to achieve a clinical outcome through modulation of molecular networks since the genetic robustness of a cell often compensates for loss of a single molecular target.

While multi-target drugs may be more difficult to design, they are faster to test clinically than drug combinations which are usually required to be tested separately

first Hopkins (2008). Synthetic lethal treatments for cancer, drug combinations and multi-target drugs to combat resistance to chemotherapy and antibiotics can be informed by biological networks Barabási and Oltvai (2004); Hopkins (2008). Further optimisation of timing and dosing of drug combinations may increase efficacy and needs to be explored for combination effects with low efficacy as separate treatments. Low doses and drug holidays are other counter intuitive approaches which may increase clinical efficacy, reduce adverse effects, and reduce drug resistance (Sun *et al.*, 2014; Tsai *et al.*, 2012).

A molecular map of the interactions and pathways in the mammalian cellular network has the potential to impact upon drug design and clinical practice, particularly in treatment of cancer and infectious disease. Characterisation of the target system and impact of existing treatments, such as *BRAF*<sup>V600E</sup> and *EGFR* inhibitors, enable wider application of the mechanisms for such interventions exploiting genetic interactions or pathways. This could lead to development of more effective treatment interventions for these systems and prediction of similar molecular systems for development of novel drug targets and combinations.

## 1.2 A Synthetic Lethal Approach to Cancer Medicine

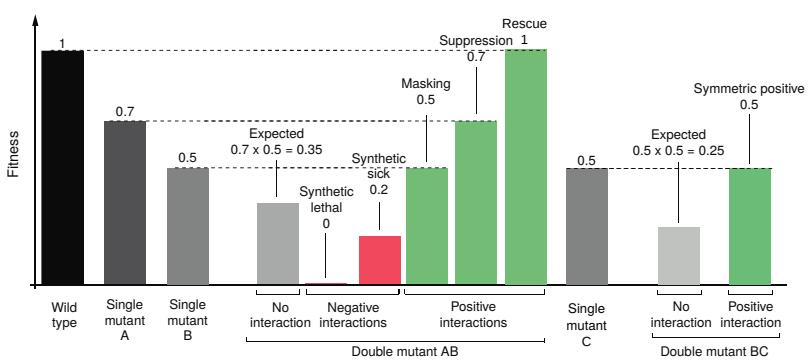
Synthetic lethality has vast potential to improve cancer medicine by expanding application of targeted therapeutics to include inactivation of tumour suppressors and genes that are difficult to target directly. Synthetic lethal interactions are also studied for gene function and drug mode-of-action in model organisms. This section introduces the concept of synthetic lethality as it was originally conceived and how it has been adopted conceptually in cancer research. Detecting these interactions at scale and interpreting them is the focus of this thesis, hence we start with an overview of the concepts involved, initial work on the interaction, and the rationale for applications to cancer. Specific investigations into synthetic lethality in cancer, detection by experimental screening, and prediction by computational analysis will then reviewed.

### 1.2.1 Synthetic Lethal Genetic Interactions

Genetic interactions are a core concept of molecular biology, discovered among earliest investigations of Mendelian genetics, and receiving revived interest with new technologies and potential applications. Biological epistasis is the effect of an allele at one locus “masking” the phenotype of another locus (Bateson and Mendel, 1909). Statistical epistasis is where there is significant disparity between the observed and expected

phenotype of a double mutant, compared to the respective phenotypes of single mutants and the wild-type (Fisher, 1919). Fisher's definition lends itself to quantitative traits and more broadly encompasses synthetic genetic interactions (SGIs). These have become popular for studies in yeast genetics and cancer drug design (Boone *et al.*, 2007; Kaelin, Jr, 2005).

Synthetic genetic interactions are substantial deviations of growth or viability from the expected null mutant phenotype (of an organism or cell) assuming additive (deleterious) effects of the single mutants. The double mutant does not necessarily have either single mutant phenotype (as shown for cellular growth phenotypes in Figure 1.1). Most SGIs are more viable than either single mutant or less viable than the expected double mutant. Mutations are “synergistic” in negative SGIs with more deviation from the wild-type than expected. Formally, “synthetic sick” (SSL) and “synthetic lethal” (SL) interactions are negative SGIs giving growth inhibition and inviability respectively. Synthetic lethality in cancer research more broadly describes any negative SGI with specific inhibition of a mutant cell, including SSL interactions. Mutations are “alleviating” in positive SGIs with less deviation from the wild-type than expected. For viability, “suppression” and “rescue” are positive SGIs giving at least partial restoration of wild-type growth from single mutants with growth impairment and lethal phenotypes respectively. Negative SGIs were markedly more common than positive SGIs in a number of studies in model systems Boucher and Jenna (2013); Tong *et al.* (2004).



**Figure 1.1: Synthetic genetic interactions.** Impact of various negative and positive SGIs: negative interactions involve deleterious (sick) or inviable (lethal) phenotypes whereas positive interactions involve restoring viability by masking or suppressing the other mutation or complete rescue of the wildtype phenotype. Figure adapted from (Costanzo *et al.*, 2011) concerning growth viability fitness in yeast.

### **1.2.2 Synthetic Lethal Concepts in Genetics**

Synthetic lethal genes are generally regarded to arise due to functional redundancy. Due to the functional level of SGIs, synthetic lethal genes do not need directly interact, nor be expressed in the same cell or at the same developmental stage: serving related functions is sufficient to affect cell (or organism) viability and be relevant to drug-mode-of-action cancer biology. Combined loss of genes performing an essential or important function in a cell are therefore deleterious. Synthetic lethal gene pairs are therefore pairwise essential with “induced essentiality”: each synthetic lethal gene becomes essential to the cell upon loss of the other.

Since synthetic lethal gene partners can be affected by extracellular stimuli and chemical, essentiality of synthetic lethal genes can be induced by the environment of a cell. An environmental stress conditions may inhibit one or the other synthetic lethal gene, such as exposure to chemicals, in which case the synthetic lethal partner gene is “conditionally essential” (Hillenmeyer, 2008). Thus the evolutionary rationale for the abundance of SGIs (compared to the surprisingly low number of essential genes) in a Eukaryotic genome attributed to genetic functional redundancy and network robustness of a cell which are advantageous to survival.

Biological functions are typically performed by a pathway of genes (or their products), may genes of the same pathway may be interchangeable as synthetic lethal partners of a particular gene since loss of the pathway is deleterious without the synthetic lethal partner gene. Therefore biological pathways can be subject to induced essentiality under loss of a gene and synthetic lethality be defined occur at pathway level or occur in a gene regulation network.

### **1.2.3 Studies of Synthetic Lethality**

Genetic high-throughput screens have identified unexpected, functionally informative, and clinically relevant synthetic lethal interactions; including synthetic lethal partners of genes recurrently mutated in cancer or attributed to familial early-onset cancers. While screening presents an appealing strategy for synthetic lethal discovery, computational approaches are becoming popular as an alternative or complement to experimental methods to overcome inherent bias and limitations of experimental screens. An array of recently developed computational methods (Jerby-Arnon *et al.*, 2014; Lu *et al.*, 2015; Tiong *et al.*, 2014; Wang and Simon, 2013; Wappett, 2014) show the need for synthetic lethal discovery in the fundamental genetics and translational cancer research community. However, existing computational methods are not suitable for queries of

genomic data for interacting partners of a particular gene: they have been applied pairwise across the genome, do not have software released to apply the methodology, or lack statistical measures of error for further analysis. A robust prediction of gene interactions is an effective and practical approach at a scale of the entire genome for ideal translational applications, analysis of biological systems, and constructing functional gene networks.

### 1.2.3.1 Synthetic Lethal Pathways and Networks

SGIs are very common in genomes, with a  $4\times$  more interactions detected with synthetic gene array mating screens than protein-protein interactions yeast-2-hybrid studies (Tong *et al.*, 2004). The SGI network is scale-free with power-law vertex degree distribution and low average shortest path length (3.3) as expected for a complex biological network (Barabási and Oltvai, 2004). Highly connected “hub” genes with the highest number of links (vertex degree) are functionally important with many negative SGI hubs involved in cell cycle regulation and many positive SGI hubs involved in translation (Baryshnikova *et al.*, 2010b; Costanzo *et al.*, 2010). Negative SGIs were far more common than positive SGIs, with synthetic gene loss being more likely to be deleterious to cell than advantageous which indicates than synthetic lethality may be comparably easier to detect than other SGIs.

Essential pathways are highly buffered with  $5\times$  more interactions than other SGIs, consistent with strong selection for survival, as found with conditional and partial mutations in essential genes (Davierwala *et al.*, 2005). This SGI network had scale-free topology and rarely shared interactions with the protein-protein interaction network. These networks are related by an “orthogonal” relationship: shared partners in one network tend to be themselves connected directly in the other network. Essential genes were likely to have closely related functions, whereas non-essential networks more relatively more inclined to have SGIs between distinct biological pathways.

#### 1.2.3.1.1 Evolution of Synthetic Lethality

There is poor conservation of specific SGIs between *S. cerevisiae* and *S. pombe* with 29% of the interactions tested in both distantly related species being conserved between them (Dixon *et al.*, 2008). The remaining interactions show high species-specific differences; however, many of the species specific interactions were still conserved between biological pathways, protein complexes, or protein-protein interaction modules. Similarly, conservation of pathway redundancy was also found between Eukaryotes (*S.*

*cerevisiae*) and prokaryotes (*E. coli*) (Butland *et al.*, 2008). Negative SGIs were more likely to be conserved between biological pathways, whereas positive SGIs were more likely to be conserved within a pathway or protein complex (Roguev *et al.*, 2008).

A modest 5% of interactions were conserved between unicellular (*S. cerevisiae*) and multicellular (*C. elegans*) organisms but the nematode SGI network had similar scale-free topology and modularity despite difficulties metazoan RNAi screens being incomplete knockouts compared to null mutations in yeast (Bussey *et al.*, 2006). The nematode SGI screen identified network hubs with important interactions to orthologues of known human disease genes (Lehner *et al.*, 2006). Despite the lack of direct conservation of SGIs between yeasts and nematode worms, genetic redundancy at the gene or pathway level may yet be consistent with an induced essentiality model of SGIs where gene functions are conserved with network restructuring over evolutionary change (Tischler *et al.*, 2008). While nematode models are more closely related to human cells, cancer cells can present growth and viability phenotypes more comparable to yeast models. Therefore findings from both SGA and RNAi models are relevant to understanding cellular network structure and in healthy and cancerous human cells. RNAi has also been applied to human and mouse cancer cells in cell culture and genetic screening experiments. These findings suggest that SGI network “rewiring” is a concern for identifying specific synthetic lethal interactions in cancer and a pathway approach may be more robust in the context of evolution, patient variation, tumour heterogeneity, and disease progression.

#### 1.2.4 Synthetic Lethal Concepts in Cancer

Loss of function occurs in many genes in cancers including tumour suppressors and yet few interventions target such mutations compared to targeted therapies for gain of function mutation in oncogenes (Kaelin, Jr, 2005). Synthetic lethality is a powerful design strategy for therapies selective against loss of gene function with potential for application against a range of genes and diseases (Fece de la Cruz *et al.*, 2015; Kaelin, Jr, 2009). Since synthetic lethality affects cellular viability by indirect functional relationships genes, it is suitable for indirectly targeting of mutations in cancers. Once synthetic lethal partners of cancer genes are identified, targeted therapeutics can be applied against them. When genes are disrupted in cancers, the induced essentiality of synthetic lethal partners is a vulnerability that may be exploited for anti-cancer therapy. This has the potential to be very specific against cancer cells (with the target mutation) over non-cancer cells (with a functional compensating gene). Analogous to

“oncogene addiction”, where cancer cells adapt to particular oncogenic growth signals and become reliant on them to remain viable (Luo *et al.*, 2009; Weinstein, 2000), synthetic lethal partners of inactivated tumour suppressors are required to maintain cancer cell viability and proliferation as such they are subject to “non-oncogene addiction” and are feasible anti-cancer drug targets.

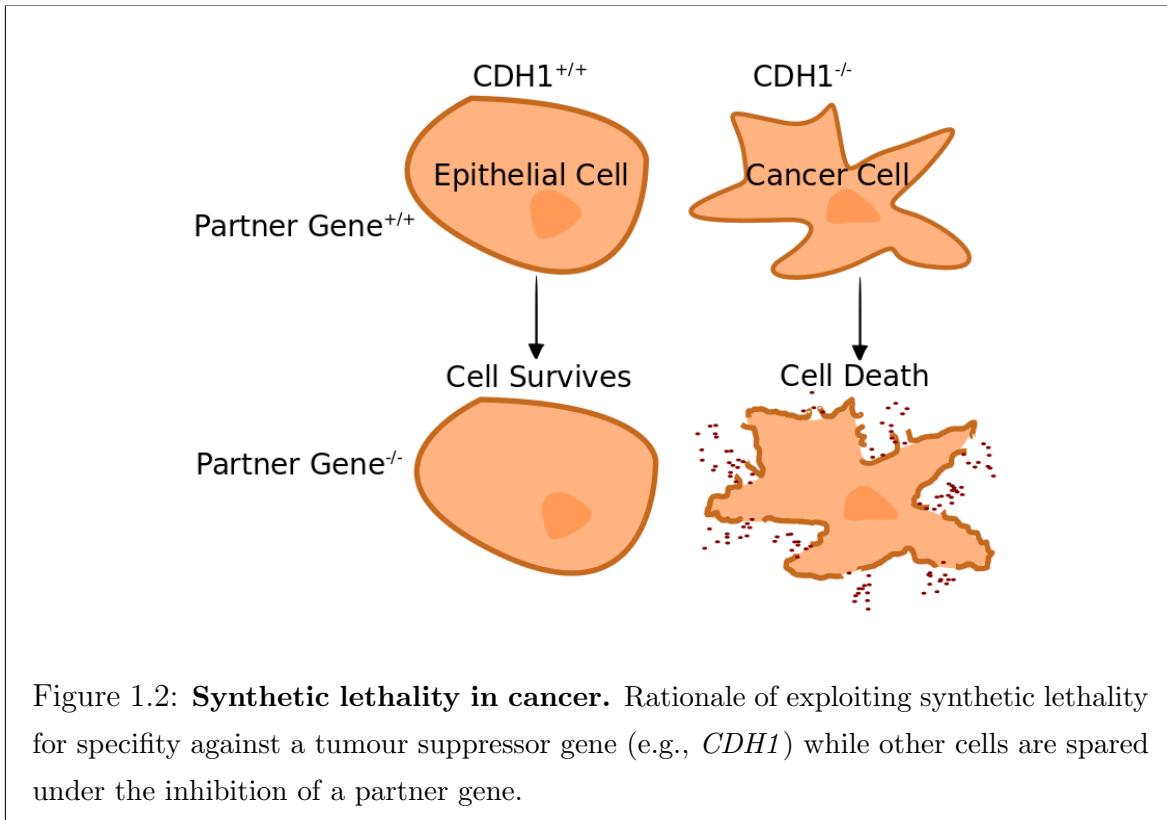


Figure 1.2: **Synthetic lethality in cancer.** Rationale of exploiting synthetic lethality for specificity against a tumour suppressor gene (e.g., *CDH1*) while other cells are spared under the inhibition of a partner gene.

The synthetic lethal approach to cancer medicine is most amenable to loss of function mutations in tumour suppressor genes, where it would feasibly be effective against any loss of function mutation across the tumour suppressor with a viable synthetic lethal partner gene (as shown in Figure 1.2). However, the approach may also be suitable for cases where cancer cells have mutations where the normal function of the gene is disrupted such as if it were overexpression (“synthetic dosage lethality”) or if an oncogene interfered with the function of the proto-oncogenic variant such as competitive inhibition. Thus synthetic lethality expands the range of cancer-specific mutations feasible to target with targeted therapeutics to absence of tumour suppressor genes and distinguishing highly homologous oncogenes by functional differences by targeting their synthetic lethal partners.

### 1.2.5 Clinical Impact of Synthetic Lethality in Cancer

The synthetic lethal interaction of *BRCA1* or *BRCA2* with *PARP1* in breast cancer is an example of how gene interactions are important in cancer, including translation to the clinic. These genetic interactions enable specific targeting of mutations in *BRCA1* or *BRCA2* tumour suppressor genes with PARP inhibitors by inducing synthetic lethality in breast cancer (Farmer *et al.*, 2005). PARP inhibitors are one of the first targeted therapeutics against a tumour suppressor mutation with success in clinical trials.

*BRCA1* or *BRCA2* and *PARP1* genes demonstrate the application of the synthetic lethal approach to cancer therapy Ashworth (2008); Kaelin, Jr (2005). *BRCA1* and *BRCA2* are homologous DNA repair genes, widely known as tumour suppressors; mutation carriers have substantially increased risk of breast (risk by age 70 of 57% for *BRCA1* and 59% for *BRCA2*) and ovarian cancers (risk by age 70 of 40% for *BRCA1* and 18% for *BRCA2*) (Chen and Parmigiani, 2007). The *BRCA1* or *BRCA2* genes, which usually repair DNA or destroy the cell if it cannot be repaired, have inactivating somatic mutations in some familial and sporadic cancers. Poly-ADP-ribose polymerase (PARP) genes are tumour suppressor genes involved in base excision DNA repair. Loss of PARP activity results in single-stranded DNA breaks. However, *PARP1*<sup>-/-</sup> knockout mice are viable and healthy indicating low toxicity from PARP inhibition (Bryant *et al.*, 2005).

Bryant *et al.* (2005) showed that *BRCA2* cells were sensitive to PARP inhibition by siRNA of *PARP1* or drug inhibition (which targets *PARP1* and *PARP2*) using Chinese hamster ovary cells, MCF7 and MDA-MB-231 breast cell lines. This effect was sufficient to kill mouse tumour xenografts and showed high specificity to *BRCA2* deficient cells in culture and xenografts. Farmer *et al.* (2005) replicated these results in embryonic stem cells and showed that *BRCA1* cells were also sensitive to PARP inhibition relative to the wild-type with siRNA and drug experiments in cell culture and drug activity against *BRCA1* or *BRCA2* deficient embryonic stem cell mouse xenografts. They found evidence that PARP inhibition causes DNA lesions, usually repaired in wild-type cells, which lead to chromosomal instability, cell cycle arrest, and induction of apoptosis in *BRCA1* or *BRCA2* deficient cells. Therefore, the pathways cooperate to repair DNA giving a plausible mechanism for combined loss as an effective anti-cancer treatment.

Thus PARP inhibitors have potential for clinical use against *BRCA1* or *BRCA2* mutations in hereditary and sporadic cancers (Ashworth 2008; Kaelin 2005). PARP inhibition has been found to be effective in cancer patients carrying *BRCA1* or *BRCA2* mu-

tations and some other ovarian cancers, suggesting synthetic lethality between PARP and other DNA repair pathways (Ström and Helleday, 2012). This supports the potential for PARP inhibition as a chemo-preventative alternative to prophylactic surgery for high risk individuals with *BRCA1* or *BRCA2* mutations (Ström and Helleday, 2012). Hormone-based therapy has also been suggested as a chemo-preventative in such high risk individuals and aromatase inhibitors have completed phase I clinical trials for this purpose (Bozovic-Spasojevic 2012). Ström and Helleday (2012) also postulate increased efficacy of PARP inhibitors in the hypoxic DNA-damaging tumour micro-environment.

A PARP inhibitor, olaparib, showed fewer adverse effects than cytotoxic chemotherapy and anti-tumour activity in phase I trials against *BRCA1* or *BRCA2* deficient familial breast, ovarian, and prostate cancers (Fong *et al.*, 2009) and sporadic ovarian cancer (Fong *et al.*, 2010). AstraZeneca has reported phase II trials showing the treatment is effective in *BRCA1* or *BRCA2* deficient breast (Tutt *et al.*, 2010) and ovarian cancers (Audeh *et al.*, 2010) with a favourable therapeutic window and similar toxicity between carriers of *BRCA1* or *BRCA2* mutations and sporadic cases. AstraZeneca announced that olaparib has begun phase III trials for breast and ovarian cancers in 2013. Mixed results in phase II trials in ovarian cancer are behind the delays addressed by retrospective analysis of the cohort subgroup with confirmed mutation of *BRCA1* or *BRCA2* genes in the tumour; unsurprisingly these patients, benefit most from the PARP inhibitor treatment and have increased platinum sensitivity in combination treatment. This demonstrates the clinical impact of a well characterised system of synthetic lethality with known cancer risk genes. Synthetic lethality has the benefit of being effective against inactivation of tumour suppressor genes by any means, broader than targeting a particular oncogenic mutation (Kaelin, Jr, 2005). The targeted therapy is effective in both sporadic and hereditary *BRCA1* or *BRCA2* deficient tumours acting against an oncogenic molecular aberration across several tissues.

[Update re. FDA approval for Ovarian]

These PARP inhibitors are FDA approved for some cancers McLachlan *et al.* (2016), are effective against germline and sporadic *BRCA1* or *BRCA2* mutations, and are a potential prevention alternative to prophylactic surgery for high risk mutation carriers Ström and Helleday (2012).

### **1.2.6 High-throughput Screening for Synthetic Lethality**

The function of signalling pathways and combinations of interacting genes are important in cancer research but classical genetics approaches have been limited to non-

redundant pathways (Fraser, 2004). The emerging RNAi technologies have vastly expanded the potential for studying genetic redundancy in mammalian experimental models including testing experimentally for synthetic lethality (Fraser, 2004). Identifying synthetic lethality is crucial to study gene function, drug mechanisms, and design novel therapies (Lum *et al.*, 2004). Candidate selection of synthetic lethal gene pairs relevant to cancer has shown some success but is limited because interactions are difficult to predict; they can occur between seemingly unrelated pathways in model organisms (Costanzo *et al.*, 2011). While biologically informed hypotheses have had some success in synthetic lethal discovery (Bitler *et al.*, 2015; Bryant *et al.*, 2005; Farmer *et al.*, 2005), interactions occurring indirectly between distinct pathways would be missed (Boone *et al.*, 2007; Costanzo *et al.*, 2011). Scanning the entire genome for interactions against a clinically relevant gene is an emerging strategy being explored with high-throughput screens (Fece de la Cruz *et al.*, 2015) and computational approaches (Boucher and Jenna, 2013; van Steen, 2012).

Experimental screening for synthetic lethality is an appealing strategy for wider discovery of functional interactions *in vivo* despite many potential sources of error which must be considered. The synthetic lethal concept has both genetic and pharmacological screening applications to cancer research. Genetic screens, with RNAi to discover the specific genes involved, inform development of targeted therapies with a known mode of action, anticipated mechanisms of resistance, and biomarkers for treatment response. RNAi is a transient knockdown of gene expression more similar to the effect of drugs than complete gene loss and makes comparison to screens in model organisms difficult (Bussey *et al.*, 2006). The RNAi gene knockdown process has inherent toxicity to some cells, potential off-target effects, and issues with a high false positive rate. Therefore, it is important to validate any candidates in a secondary screen and replicate knockdown experiments with a number of independent shRNAs. Alternative gene knockout procedures have also been proposed for synthetic lethal screening including a genome-wide application of the CRIPR/Cas9/sgRNA genome editing technology (Sander and Joung, 2014), episomal gene transfer (Vargas *et al.*, 2004), or RNAi with lentiviral transfection for delivery of shRNA (Telford *et al.*, 2015). Genetic screens have potential for quantitative gene disruption experiments to selectively target overexpressed genes in cancer via synthetic dosage lethality. While powerful for understanding fundamental cellular function, analysis of isogenic cell lines is inherently limited by assuming only a single mutation differs between them despite susceptibility to “genetic drift” and cannot account for diverse genetic backgrounds or tumour heterogeneity (Fece de la Cruz

*et al.*, 2015). Genetic screens thus identify targets to develop or repurpose targeted therapies for disease but alone will not directly identify a lead compound to develop for the market or clinical translation.

Chemical screens are immediately applicable to the clinic by directly screening for selective lead compounds with suitable pharmacological properties. However chemical screens lack a known mode of action, may affect many targets, and screen a narrow range of genes with existing drugs. With either approach there are many challenges translating candidates into the clinic such as finding targets relevant to a range of patients, validation of targets, accounting for a range of genetic (and epigenetic) contexts or tumour micro-environment, identifying effective synergistic combinations, enhancers of existing radiation or cytotoxic treatments, avoiding inherent or acquired drug resistance, and developing biomarkers for patients which will respond to synthetic lethal treatment, including integrating these into clinical trials and clinical practice. Identifying specific target genes is an effective way to anticipate such challenges, which can be approached with genetic screens, so we will focus on these and computational alternatives. Screening methods have proven a fruitful area of research, despite being costly, laborious, and having many different sources of error. These limitations suggest a need for complementary computational approaches to synthetic lethal discovery.

#### 1.2.6.1 Synthetic Lethal Screens

Overexpression of genes is another suitable application for synthetic lethality since overexpressed genes cannot be distinguished from the wild-type by direct sequence specific targeted therapy. Overexpression of oncogenes, such as *EGFR*, *MYC*, and *PIM1*, has been found to drive many cancers. *PIM1* is a candidate for synthetic lethal drug design in lymphomas and prostate cancers, where it interacts with *MYC* to drive cancer growth. van der Meer *et al.* (2014) performed an RNAi screen to for synthetic lethality between *PIM1* overexpression and gene knockdown in RWPE prostate cancer cell lines. *PLK1* gene knockdown and drug inhibition was an effective as a specific inhibitor of *PIM1* overexpressing prostate cells in cell culture and mouse tumour xenografts. *PLK1* inhibition reduced *MYC* expression in pre-clinical models, consistent with expression in human tumours which *PIM1* and *PLK1* are co-expressed and correlated with tumour grade. Thus RNAi screening was valuable to identify a therapeutic targets and biomarkers for patient response as demonstrated with the finding of *PLK1* as a candidate drug target against prostate cancer progression.

Heredity leiomyomatosis and renal cell carcinoma (HLRCC) is a cancer syndrome of predisposition to benign tumours in the uterus and risk of malignant cancer of the

kidney attributed to inherited mutations in fumarate hydratase (*FH*). Boettcher *et al.* (2014) performed an RNAi screen on HEK293T renal cells for synthetic lethality with *FH*. They found enrichment of haem metabolism (consistent with the literature) and adenylate cyclase pathways (consistent with cAMP dysregulation in *FH* mutant cells). Synthetic lethality between *FH* mutation and adenylate cyclases was validated with gene knockdown, drug experiments, and replicated across both HEK293T renal cells and VOK262 cells derived from a HLRCC patient, suggesting new potential treatments against the disease.

Similarly, hereditary diffuse gastric cancer (HDGC) is a cancer syndrome of predisposition to early-onset malignant stomach and breast cancers attributed to mutations in E-cadherin (*CDH1*). Telford *et al.* (2015) performed an RNAi screen on MCF10A breast cells for synthetic lethality with *CDH1*. They found enrichment of G-protein coupled receptors (GPCRs) and cytoskeletal gene functions. The results were consistent with a concurrent drug compound screen with a number of candidates validated by lentiviral shRNA gene knockdown and drug testing including inhibitors of Janus kinase, histone deacetylases, phosphoinositide 3-kinase, aurora kinase, and tyrosine kinases. Therefore the synthetic lethal strategy has potential for clinical impact against HDGC, with particular interest in interventions with low adverse effects for chemoprevention, including repurposing existing approved drugs for activity against *CDH1* deficient cancers.

RNAi screening for synthetic lethality is also useful for functional genetics to understand drug sensitivity. Aarts *et al.* (2015) screened WiDr colorectal cells for synthetic lethality between *WEE1* inhibitor treatment and an RNAi library of 1206 genes with functions known to be amenable to drug treatment or important in cancer such as kinases, phosphatases, tumour suppressors, and DNA repair (a pathway *WEE1* regulates). Screening identified a number of synthetic lethal candidates including genes involved in cell cycle regulation, DNA replication, repair, homologous recombination, and Fanconi anaemia. Synthetic lethality with cell-cycle and DNA repair genes was consistent with the literature and validation in a panel of breast and colorectal cell lines supported checkpoint kinases, Fanconi anaemia, and homologous recombination as synthetic lethal partners of *WEE1*. These results show that synthetic lethality can be used to improve drug sensitivity as a combination treatment, especially to exploit genomic instability and DNA repair, which are known to be clinically applicable from previous results with *BRCA1* or *BRCA2* genes and PARP inhibitors (Lord *et al.*, 2015). Therefore, *WEE1* inhibitors are an example of treatment which could be repurposed

with the synthetic lethal strategy and similar findings would be valuable to clinicians as a source of biomarkers and novel treatments. While using a panel of cell lines to replicate findings across genetic background is a promising approach to ensure wide clinical application of validated synthetic lethal partners, a computational approach may be more effective as it could account for wider patient variation than scaling up intensive experiments on a wide array of cell lines and could screen beyond limited candidates from an RNAi library.

Chemical genetic screens are also a viable strategy to identify therapeutically relevant synthetic lethal interactions. Bitler *et al.* (2015) investigated *ARID1A* mutations, aberrations in chromatin remodelling known to be common in ovarian cancers, for drug response. Ovarian RMG1 cells were screened for drug response specific to *ARID1A* knockdown cells. They used *ARID1A* gene knockdown for consistent genetic background, with control experiments and 3D cell culture to ensure relevance to drug activity in the tumour micro-environment. Screening a panel of commercially available drugs targeting epigenetic regulators found *ESH2* methyltransferase inhibitors effective and specific against *ARID1A* mutation with validation in a panel of ovarian cell lines. Synthetic lethality between *ARID1A* and *ESH2* was supported by decreases in H3K27Me3 epigenetic marks and markers of apoptosis in response to *ESH2* inhibitors. This was mechanistically supported with differential expression of *PIK3IP1* and association of both synthetic lethal genes with the *PIK3IP1* promoter identifying the PI3K-AKT signalling pathway as disrupted when both genes are inhibited. This successfully demonstrates the importance of synthetic lethality in epigenetic regulators, identifies a therapeutically relevant synthetic lethal interaction, and shows that chemical genetic screens could model drug response and combination therapy in cancer cells. However this approach is limited to finding synthetic lethal interactions between genes with known similar function, which may not be the most suitable for treatment. Further limiting experiments to genes with existing targeted drugs reduces the number of synthetic lethal interactions detected, assumes on their drug specificity to a particular target, and many of these drugs are not clinically available yet anyway as they are still in clinical trials for other diseases or are not supported by healthcare systems in many countries.

The examples above show that high-throughput screens are an effective approach to discover synthetic lethality in cancer with a wide range of applications. Screens are more comprehensive than hypothesis-driven candidate gene approaches and successfully find known and novel synthetic lethal interactions with potential for rapid clinical

application. They have the power to test mode of action of drugs, find unexpected synthetic lethal interactions between pathways, or identify effective treatment strategies without needing a clear mechanism. However, synthetic lethal screens are costly, labour-intensive, error-prone, and biased towards genes with effective RNAi knockdown libraries. Limited genetic background, lethality to wild-type cell during gene knockdown, off-target effects, and difficulty replicating synthetic lethality across different cell lines, tissues, laboratories, or conditions stems from a high false positive rate and a lack of standardised thresholds to identify synthetic lethality in a high-throughput screen. Therefore there is a need for replication, validation, and alternative approaches to identify synthetic lethal candidates. Varied conditions between experimental screens and differences between RNAi or drug screens makes meta-analysis difficult. Thus genome-scale synthetic lethal experiments are not feasible, even in model organisms, so a computational approach would be more suitable for this task.

## 1.2.7 Computational Prediction of Synthetic Lethality

### 1.2.7.1 Bioinformatics Approaches to Genetic Interactions

Prediction of gene interaction networks is a feasible alternative to high-throughput screening with biological importance and clinical relevance. There are many existing methods to predict gene networks, as reviewed by van Steen (2012) and Boucher and Jenna (2013) and summarised in Table 1.1. However, many of these methods have limitations including the requirement for existing SGI data, several data inputs, and reliability of gene function annotation. Many of the existing methods also assume conservation of individual interactions between species, which has been found not to hold in yeast studies (Dixon *et al.*, 2008). Tissue specificity is important in gene regulation and gene expression, which are used as predictors of genetic interaction. However, tissue specific of genetic interactions cannot be explored in yeast studies and has not been considered in many studies of multicellular model organisms, human networks, or cancers. Similarly, investigation into tissue specific of protein-protein interactions (PPIs), an important predictor of genetic interactions, is difficult given the high-throughput two-hybrid screens occur out of cellular context for multicellular organisms.

There are a number of existing computational methods for predicting synthetic lethal gene pairs in humans with a specific interest in cancer (as summarised in 1.2). While these demonstrate the power and need for predictions of synthetic lethality in human and cancer contexts, limitations of previous methods could be met with a

Table 1.1: Methods for Predicting Genetic Interactions

| Method                         | Input Data                                 | Species                                   | Source   | Tool Offered   |
|--------------------------------|--|---|--|----------------|
| Between Pathways Model         | PPI, SGI                                   | <i>S. cerevisiae</i>                      | Kelley and Ideker (2005)                                     |                |
| Within Pathways Model          | PPI, SGI                                   | <i>S. cerevisiae</i>                      | Kelley and Ideker (2005)                                     |                |
| Decision Tree                  | PPI, expression, phenotype                 | <i>S. cerevisiae</i>                      | Wong <i>et al.</i> (2004)                                    | 2 Hop          |
| Logistic Regression            | SGI, PPI, co-expression, phenotype         | <i>C. elegans</i>                         | Zhong and Sternberg (2006)                                   | Gene Orienteer |
| Network Sampling               | SGI, PPI, GO                               | <i>S. cerevisiae</i>                      | Le Meur and Gentleman (2008)<br>Le Meur <i>et al.</i> (2014) | SLGI(R)        |
| Random Walk                    | GO, PPI, expression                        | <i>S. cerevisiae</i><br><i>C. elegans</i> | Chipman and Singh (2009)                                     |                |
| Shared Function                | Co-expression, PPI, text mining, phylogeny | <i>C. elegans</i>                         | Lee <i>et al.</i> (2010b)                                    | WormNet        |
| Logistic Regression            | Co-expression, PPI, phenotype              | <i>C. elegans</i>                         | Lee <i>et al.</i> (2010a)                                    | GI Finder      |
| Jaccard Index                  | GO, SGI, PPI, phenotype                    | Eukarya                                   | Hoechndorf <i>et al.</i> (2013)                              |                |
| Machine Learning               |  |   | Pandey <i>et al.</i> (2010)                                  | MNMC           |
| Machine Learning Meta-Analysis |  |   | Wu <i>et al.</i> (2014)                                      | MetaSL         |
| Flux Variability Analysis      |  |   |  |                |
| Flux Balance Analysis          | Metabolism                                 | <i>E. coli</i><br><i>M. pneumoniae</i>    | Güell <i>et al.</i> (2014)                                   |                |
| Network Simulation             |  |   |  |                |

Table 1.2: Methods for Predicting Synthetic Lethality in Cancer

| Method                  | Input Data  | Source   | Tool Offered   |
|-------------------------|---|--|----------------|
| Network Centrality      | protein-protein interactions                        | Kranthi <i>et al.</i> (2013)   |                |
| Differential Expression | Expression<br>Mutation                              | Wang and Simon (2013)  |                |
| Comparative Genomics    | Yeast synthetic gene interactions                   | Heiskanen and Aittokallio (2012)   |                |
| Chemical-Genomics       | Homology  |  |                |
| Comparative Genomics    | Yeast synthetic gene interactions<br>Homology       | Deshpande <i>et al.</i> (2013)   |                |
| Machine Learning        |   | Discussed by Babyak (2004)<br>and Lee and Marcotte (2009)  |                |
| Differential Expression | Expression  | Tiong <i>et al.</i> (2014)   |                |
| Literature Database     |   | Li <i>et al.</i> (2014)  | Syn-Lethality  |
| Meta-Analysis           | Meta-Analysis<br>Machine Learning                   | Wu <i>et al.</i> (2014)  | MetaSL         |
| Pathway Analysis        |   | Zhang <i>et al.</i> (2015)   |                |
| Protein Domains         | Homology  | Kozlov <i>et al.</i> (2015)  |                |
| Data-Mining             | Expression  | Jerby-Arnon <i>et al.</i> (2014)   |                |
| Machine Learning        | Somatic mutation and DNA CNV<br>siRNA in cell lines | Ryan <i>et al.</i> (2014)<br>Crunkhorn (2014)<br>Lokody (2014)                                   | DAISY (method) |
| Genome Evolution        | Expression  | Lu <i>et al.</i> (2013)  |                |
| Hypothesis Test         | DNA CNV   | Lu <i>et al.</i> (2015)  |                |
| Machine Learning        | Known SL  |  |                |
| Bimodality              | Expression<br>DNA CNV<br>Somatic Mutation           | Wappett (2014)<br>Wappett <i>et al.</i> (2016)   | BiSEp          |
| Directional Chi-Square  | Expression (microarray)<br>Somatic mutation         | Kelly, S. T., Guilford, P. J., and Black, M. A.<br>Dissertation (Kelly, 2013) and developed here | SLIPT          |

different approach. Existing computational approaches to synthetic lethal prediction are often difficult to interpret, replicate for new genes, or reliant on are data types not available for a wider range of genes to test.

### 1.2.7.2 Comparative Genomics

A comparative genomics approach by Deshpande *et al.* (2013) used the results of well characterised high-throughput mutation screens in *S. cerevisiae* as candidates for synthetic lethality in humans (Baryshnikova *et al.*, 2010a; Costanzo *et al.*, 2010, 2011; Tong *et al.*, 2001, 2004). Yeast synthetic lethal partners were compared to human orthologues to find cancer relevant synthetic lethal candidate pairs with direct therapeutic potential. Proposed as a complementary approach to siRNA screens, approximately 24,000 of the 116,000 negative SGIs in yeast (Costanzo *et al.*, 2011) were matched to human orthologues, with over 500 involving a cancer gene (Futreal *et al.*, 2004). Under strict criteria of one-to-one orthologues, large effect size and significant interaction in yeast data ( $\epsilon < -0.2$ ,  $p < 0.05$ ), 1,522 interactions were identified with 70 involving cancer genes. Of the 21 gene interactions tested with pairs of siRNA in IMR1 fibroblast cells, 6 exhibited synthetic lethal effects. The two strongest interactions (*SMARCB1* with *PSMA4* and *ASPSCR1* with *PSMC2*) were successfully validated in by protein analysis of human cells and replication with tetrad analysis for yeast orthologues.

Another approach to systematic synthetic lethality discovery specific to human cancer (in contrast to the plethora of yeast synthetic lethality data) was to build a database as done by Li *et al.* (2014). In their relational database, called “Syn-lethality”, they have curated both known experimentally discovered synthetic lethal pairs in humans (113 pairs) from the literature and those predicted from synthetic lethality between orthologous genes in *S. cerevisiae* yeast (1114 pairs). This knowledge-based database is the first dedicated to human cancer synthetic lethal interactions and integrates gene functional, annotation, pathway and molecular mechanism data with experimental and predicted synthetic lethal gene pairs. This combination of data sources is intended to tackle the trade-off between more conclusive synthetic lethal experiments in yeast and more clinically relevant synthetic lethal experiments in human cancer models, such as RNAi, especially when high-throughput screens are costly and prone to false positives in either system and difficult to replicate across gene backgrounds. This database centralises a wealth of knowledge scattered in the literature including cancer relevant genes (*BRCA1*, *BRCA2*, *PARP1*, *PTEN*, *VHL*, *MYC*, *EGFR*, *MSH2*, *KRAS*, and *TP53*) and is publicly available as a Java App. These included the previously mentioned interactions of *BRCA1* and *BRCA2* with

*PARP1* and *TP53* with *WEE1* and *PLK1*. However, the computational methodology was not released, so it is not possible to replicate their results, nor to add to the findings with new datasets, which are limited to 647 human genes. Suggested future directions were promising, such as constructing networks of known synthetic lethality, applying known synthetic lethality to cancer treatment, data mining, replicating the approach for synthetic lethality in model organisms, signalling pathways, and develop a complete global network in human cancer or yeast (both of which are still incomplete with experimental data), some of which has been implemented in “SynLethDB” (Guo *et al.*, 2016).

Table 1.3: Machine Learning Methods used by Wu *et al.* (2014)

| Method                                  | Source                      | Tool Offered |
|---|-----------------------------|--------------|
| Random Forest                           | Breiman (2001)              |              |
| Random Forest                           |                             |              |
| J48 (decision tree)                     |                             |              |
| Bayes (Log Regression)                  |                             |              |
| Bayes (Network)                         | Hall <i>et al.</i> (2009)   | WEKA         |
| PART (Rule-based)                       |                             |              |
| RBF Network                             |                             |              |
| Bagging / Bootstrap                     |                             |              |
| Classification via Regression           |                             |              |
| Support Vector Machine (Linear)         | Vapnik (1995)               |              |
| Support Vector Machine (RBF – Gaussian) | Joachims (1999)             |              |
| Multi-Network Multi-Class (MNMC)        | Pandey <i>et al.</i> (2010) |              |
| MetaSL (Meta-Analysis)                  | Wu <i>et al.</i> (2014)     | MetaSL       |

Machine learning approaches have also been proposed for synthetic lethal discovery (Babyak, 2004; Lee and Marcotte, 2009). Due to concerns that these may be subject to overfitting or noise, Wu *et al.* (2014) developed a meta-analysis method (based on the machine learning methods in Table 1.3) for synthetic lethal gene pairs relevant to developing selective drugs against human cancer, building upon their previous database (Li *et al.*, 2014). The used training data of 10,885 synthetic lethal interactions from yeast experiments of which 7347 occurred between the 5,504 non-essential genes. Their “metaSL” approach utilises genomic, proteomic and annotation data (including GO terms Ashburner *et al.* (2000), PPI, protein complexes, and biological pathway) with strong statistical performance in yeast data (AUROC of 0.871). The predicted

orthologous synthetic lethal partners in human data were not experimentally validated but several would be relevant to cancer such as *EGFR* with *PRKCZ*. They note that computational approaches scale-up across the genome at lower cost than experimental screen and share their most supported interactions online. However, the method is not available for analysis of other genes studied by the cancer research community. While machine learning has great potential as a predictor, the results vary greatly depending on the predictive features selected and it is not clear which threshold should be used to report reliably detected genes. Syn-Lethality (Li *et al.*, 2014) and MetaSL (Wu *et al.*, 2014) demonstrate the value of computational approaches to synthetic lethality but omit many genes of importance in cancer, such as *CDH1*, and there remains a need to enable biological researchers to query such genes in a particular tissue or genetic background.

There is also concern for analyses based on yeast data that many synthetic lethal interactions may not be conserved between species Dixon *et al.* (2009), although interactions between pathways may be more comparable. It is unsurprising that many of the interactions identified were not experimentally validated. There have been many gene duplications in the separate evolutionary histories of humans and yeast which may lead to differences in genetic redundancy. Yeast are further not an ideal human cancer model because they do not have tissue specificity, multicellular gene regulation, or orthologues to a number of known cancer genes such as p53. Although these studies have tried to anticipate these issues with stringent criteria such as requiring one-to-one orthologues, there remains the possibility that changes in gene function may affect whether these are solely redundant such as if functions had coevolved without sequence homology. Many genes will also be excluded by lacking homologous gene in yeast, the corresponding experimental data, or having paralogues in either species. Thus conservation of yeast interactions is not an ideal strategy and analysis of human data directly for comparison with human experimental data will be the focus of this thesis.

#### 1.2.7.3 Analysis and Modelling of Protein Data

Kranthi *et al.* (2013) took a network approach to discovery of synthetic lethal candidate selection applying the concept to “centrality” to a human PPI network involving interacting partners of known cancer genes. The effect of removing pairs of genes on connectivity of the network was used as a surrogate for viability which is supported by observations that the PPI and synthetic lethal networks are orthogonal in *S. cerevisiae* studies (Tong *et al.*, 2004). They showed that the human cancer protein interaction

network (of 1539 proteins and 6471 interactions) exhibits the power law distribution expected of a scale-free synthetic lethal network with high connectivity (average vertex degree of 23.67 and network efficiency of 0.2952). Their top 100 candidate interactions included interactions of the tumour suppressor *TP53* with *BRCA1*, *CDKNA1*, *CDKNA2*, *MET*, and *RB1* which have been detected by prior studies. The gene pairs were often observed to be in the same or a plausible compensatory pathway. Thus the network structure is important in the biological functions of cancers and could be exploited for targeting *TP53* loss of function mutations.

However, their approach was limited to known cancer genes and is not applicable to genes that do not have PPI data. Other nucleotide sequencing data types are more commonly available for cancer studies at a genomic scale. Of further concern is that the results were enriched for p53 synthetic lethal partners which is relevant to many cancer researchers but this genome-wide approach did not detect many other cancer genes due to multiple testing. This enrichment may be due to the known drastic effect of removing p53 itself from the network as a master regulator, cancer driving tumour suppressor gene, and highly connected network “hub”. The focus on cancer genes is useful for translation into therapeutics but does not account for variable genetic backgrounds or effect of protein removal on the whole cellular network.

Focusing on the potential for synthetic lethality to be an effective anti-cancer drug target, Zhang *et al.* (2015) used modelling of signalling pathways to identify synthetic lethal interactions between known drug targets and cancer genes by simulating gene knockdowns. A computational approach applied to avoid the limitations of experimental RNAi screens such as scale, instability of knockdown, and off-target effects. This ‘hybrid’ method of a data-driven model and known signalling pathways showed potential as a means to predict cell death in single and combination gene knockouts. They used time series protein phosphorylation data (Lee *et al.*, 2012) for 28 signalling proteins and Gene Ontology (GO) pathways Ashburner *et al.* (2000); Blake *et al.* (2015). This approach successfully detected many known essential genes in the human gene essentiality database, known synthetic lethal partners in the Syn-Lethality database (Li *et al.*, 2014), and predicted novel synthetic lethal gene pairs. The strongest essential genes in single knockdowns were *AKT*, *TP53*, *CHK1*, *S6K1*, and *CYCLIND1*. Pairwise knockdowns identified 252 candidate synthetic lethal interactions including *TP53* with *CHK1*, *S6K1*, *WEE1*, *CYCLIND1*, and *CASP9*; *AKT* with *WEE1*; and *CDK1* with *CYCLIND1*. These novel results contained many *TP53* and *AKT* synthetic lethal partners, genes known to be important in many cancers, however these also have a

high impact on the signalling pathways in their essentiality analysis of single gene disruptions and large phenotypic changes in cancer. This approach is amenable to detect functionally related pathways and protein complexes across the molecular function, cellular component, and biological process annotations provided by GO. The results were consistent with the experimental results in the literature but the novel synthetic lethal interactions have yet to be validated. While the mathematical reasoning and algorithms are given, the code was not released to replicate the findings or apply the methodology beyond the signalling pathways analysed by Zhang *et al.* (2015). While this is an interesting approach, the analysis of this thesis will focus on gene expression and RNAi data which is available to test a wider range of candidate gene pairs.

#### 1.2.7.4 Differential Gene Expression

Differential gene expression has been explored to predict synthetic lethal pairs in cancer which would be widely applicable due to the availability of public gene expression data for a large number of samples and cancer types. Wang and Simon (2013) found differentially expressed genes (by the t-test, adjusted by FDR) between tumours with or without functional p53 mutations in TCGA (McLendon *et al.*, 2008) and Cell Line Encyclopaedia (CCLE) (Barretina *et al.*, 2012) RNA-Seq gene expression data as candidate synthetic lethal partner pathways of p53. They identified 2, 8, and 21 candidate synthetic lethal partner genes in 3 microarray datasets from the NCI60 cell lines, 31 partner genes from the CCLE RNA-Seq data, and 50 in TCGA RNA-Seq data. *PLK1* was replicated across 4 of these analyses and 17 other genes were replicated across 2 analyses (including *MTOR*, *PLK4*, *MAST2*, *MAP3K4*, *AURKA*, *BUB1* and 6 CDK genes) with many playing a role in cell cycle regulation. This was supported by a drug sensitivity experiment on the NCI60 cell lines which found that cells which lacked functional p53 were more sensitive to paclitaxel (which targets *PLK1*, *AURKA*, and *BUB1*). This demonstrated the potential of gene expression as a surrogate for gene function and use of public genomic data to predict synthetic lethal gene pairs in cancer. Wang and Simon (2013) advocated for pre-screening of expression profiles to augment future RNAi screens. However, the analyses were limited to kinase genes and focused on currently druggable genes, lacking wider application of synthetic lethal prediction methodology. This approach may not be feasible or applicable in cancer genes with a lower mutation rate than p53.

Tiong *et al.* (2014) also investigated gene expression as a predictor of synthetic lescale-freethal pairs with colorectal cancer microarrays from a Han Chinese population with a sample size of 70 tumour and 12 normal tissue samples. Simultaneously differ-

entially expression of “tumour dependent” gene pairs (which includes co-expression) between cancer and normal tissue was used to rank 663 candidate synthetic lethal interactions identified in cell line siRNA experiments. Of the top 20 genes, 17 were tested for testing differential expression at the protein level with immunohistochemistry staining and correlation with clinical characteristics, with 11 pairs exhibiting synergistic effects. Some of the predicted synthetic lethal pairs were consistent with the literature (including *TP53* with *S6K1* and partners of *KRAS*, *PTEN*, *BRCA1*, and *BRCA2*) and two novel synthetic lethal interactions (*TP53* with *CSNK1E* and *CTNNB1*) were validated in pre-clinical models. This serves a valuable proof-of-concept for integration of *in silico* approaches to synthetic lethal discovery in cancer demonstrating it’s utility to triage and identify synthetic lethal partners of p53 applicable to colorectal tissues. Although the experimental work was the focus of the paper, these findings show that bioinformatics synthetic lethal candidates can be validated in patient tissue samples (from a non-caucasian population) to find those applicable to colorectal cancers.

#### 1.2.7.5 Data Mining and Machine Learning

Recognising the utility of synthetic lethality to drug inhibition and specificity of anti-cancer treatments, Jerby-Arnon *et al.* (2014) also saw the need for effective prediction of gene essentiality and synthetic lethality to augment experimental studies of SL. They developed a data-driven pipeline called DAISY (data mining synthetic lethality identification pipeline) and tested for genome-wide analysis of synthetic lethality in public cancer genomics data from TCGA and CCLE (Barretina *et al.*, 2012). DAISY is intended to predict the candidate synthetic lethal partners of a query gene such as genes recurrently mutated in cancer.

Jerby-Arnon *et al.* (2014) combined a computational approach to triage candidates with a conventional RNAi screen to validate synthetic lethal partners. They screened a selection of computationally predicted candidates and randomly selected genes with RNAi against *VHL* loss of function mutation in RCC4 renal cell lines. The computational method had a high AUROC of 0.779 and predictions were enriched 4× for validated RNAi hits over randomly selected genes. This approach detected known synthetic lethal pairs such as *BRCA1* or *BRCA2* genes with *PARP1* and *MSH2* with *DHFR*. The synthetic lethal candidates identified with both RNAi screening and computational prediction formed an extensive network of 2077 genes with 2816 synthetic lethal interactions and similar network of 3158 genes with 3635 synthetic dosage lethal interactions (for synthetic lethality with over-expression). Each network was scale-free as expected of a biological network and was enriched for known cancer genes, essen-

tial genes in mice, and could be harnessed for predicting prognosis and drug response. While demonstrating the feasibility of combining experimental and computational approaches to synthetic lethality in cancer, there remain challenges in predicting synthetic lethal genes, novel drug targets, and translation into the clinic.

The DAISY methodology (Jerby-Arnon *et al.*, 2014) compares the results of analysis of several data types to predict synthetic lethality, namely: DNA copy number and somatic mutation for TCGA patient samples and CCLE cell lines. The cell lines were also analysed with gene expression and gene essentiality (shRNA screening) profiles. Genes were classed as inactivated by copy number deletion, somatic loss of function mutation, or low expression and tested for synthetic lethal gene partners which are either essential in screens or not deleted with copy number variants. Co-expression is also used for synthetic lethality prediction based on studies in yeast (Costanzo *et al.*, 2010; Kelley and Ideker, 2005). Copy number, gene expression and, essentiality analyses are stringently compared by adjusting each for multiple tests with Bonferroni correction and only taking hits which occur in all analyses. This methodology was also adapted for synthetic dosage lethality by testing for partner genes where genes are overactive with high copy number or expression. As discussed above, the predictions performed well and an RNAi screen for the example of *VHL* in renal cancer validated predicted synthetic lethal partners of *VHL* demonstrating the feasibility of combining approaches to synthetic lethal discovery in cancer and using computational predictions to enable more efficient high-throughput screening. DAISY performs well statistically with a AUROC of 0.779 on a set of gene pairs with experimental screen data, although co-expression and shRNA functional examination contributes much less of this than the mutation and copy number analysis (AUROC 0.683 alone). However, this methodology is very stringent, missing potentially valuable synthetic lethal candidates, may not be applicable to genes of interest to other groups and the software for the procedure is not publicly released for replication.

Although the DAISY procedure performs well and has been well received by the scientific community (Crunkhorn, 2014; Lokody, 2014; Ryan *et al.*, 2014), showing a need for such methodology, there is no indication of adoption of the methodology in the community yet. The co-expression analysis may not be the most effective way to test gene expression for directional synthetic lethal interactions (where inverse correlation would be expected). In the interests of a large sample size, tissue types were not tested separately despite tissue-specific synthetic lethality being likely since gene function (and by extension expression, isoforms, and clinical characteristics) in cancers may often be

tissue-dependent. Some data forms and analyses used, such as gene essentiality, may not be available for all cancers, genes, or tissues, and may not be reproduced.

Lu *et al.* (2015) critique the assumption of co-expression in the DAISY methodology and propose an alternative computational prediction of synthetic lethality based on machine learning methods and a cancer genome evolution hypothesis. Using DNA copy number and gene expression data from TCGA patient samples, a cancer genome evolution model assumes that synthetic lethal gene pairs behave in 2 distinct ways in response to an inactive synthetic lethal partner gene, either a “compensation” pattern where the other synthetic lethal partner is overactive or a “co-loss underrepresentation” pattern where the other synthetic lethal partner is less likely to be lost, since loss of both genes would cause death of the cancer cell. During the cancer genome evolution as the cell becomes addicted to the remaining synthetic lethal partner due to induced gene essentiality. These patterns would explain why DAISY detects only a small number of synthetic lethal pairs, compared to the large number expected based on model organism studies (Boone *et al.*, 2007), and the disparity between screening and computationally predicted synthetic lethal candidates due to testing different classes of synthetic lethal gene pairs.

Lu *et al.* (2015) compared a genome-wide computational model of genome evolution and gene expression patterns to the experimental data of Vizeacoumar *et al.* (2013) and Laufer *et al.* (2013). This simpler model performing well with an AUROC of 0.751 but was less than DAISY, although it did not rely on data from cell lines which may not represent patient disease. They predict a larger comprehensive list of 591,000 human synthetic lethal partners with a probability score threshold of 0.81, giving a precision of 67% and 14 $\times$  enrichment of synthetic lethal true positives compared to randomly selected gene pairs. Discovery of such a vast number of cancer-relevant synthetic lethal interactions in humans would not be feasible experimentally and is a valuable resource for research and clinical applications. These predictions are not limited by assuming co-expression of synthetic lethal partners or evolutionary conservation with model organisms enabling wider synthetic lethal discovery. However, there remains a lack of basis for an expectation of how many synthetic lethal partners a particular gene will have, how many pairs there are in the human genome, and whether pathways or correlation structure would influence predicted synthetic lethal partners.

Large scale, computational approaches have yet to determine whether synthetic lethal interactions are tissue-specific since Lu *et al.* (2015) used pan-cancer data for 14136 patients with 31 cancer types. Experimental data used for comparison was a

small training dataset specific to colorectal cancer, and based on screens for other phenotypes, which may limit performance of the model or application to other cancers. Proposed expansion of the computational approach to mutation, microRNA, or epigenetic modulation of gene function and tumour micro-environment or heterogeneity suggests that synthetic lethal discovery could be widely applied to the current challenges in cancer genomics. This approach was also based on machine learning methodology and not supported by a software released for the community to develop, contribute to, or reproduce beyond the gene pairs given in the supplementary results.

#### 1.2.7.6 Bimodality

Wappett *et al.* (2016) demonstrate a multi-omic approach to identification of synthetic lethality in cancer with a strategy to detect bimodal patterns in molecular profiles. They release this solution as the Bimodal Subsetting Expression (BiSEp) R package Wappett (2014) which aims to detect subtle bimodal and non-normal patterns in expression data. Since loss of gene function is not consistently genetic, Wappett *et al.* (2016) advocate the use of gene expression (loss of mRNA) and deletion (loss of copy number) data in addition to mutation. The BiSEp procedure was demonstrated on an analysis of 881 cell lines from CCLE (Barretina *et al.*, 2012), 442 cell lines from COSMIC (Forbes *et al.*, 2015), and RSEM normalised RNA-Seq data for 178 TCGA lung patient samples (Collisson *et al.*, 2014). BiSEp was demonstrated to have significant enrichment of validated tumour suppressor, synthetic lethal gene pairs (detecting 76 experimentally supported gene pairs) and was improved (detecting 420) with expression data rather than relying on detecting loss of gene function by mutation or deletion. They identified interactions with genes relevant to cancer with support in experimental screens including *ERCC4* with *XRCC1*, *BRCA1* with *PARP3*, and *SMARCA1* with *SMARCA4*.

Wappett *et al.* (2016) demonstrated that analysis of genomics data, particularly expression data, is relevant to augment the identification of synthetic lethal interactions with screening experiments. They further show that this is applicable in both genetically homogenous cell lines and heterogeneous cell population from patient samples. This approach is limited however to genes which exhibit bimodal expression patterns which do not commonly occur, particularly in normalised gene expression data, and other approaches may need to be considered for gene such as *CDH1* which were not identified by BiSEp.

### **1.2.7.7 Rationale for Further Development**

Many of the approaches discussed here aimed to identify the strongest synthetic lethal pairs across the yeast or human genome (Deshpande *et al.*, 2013; Lu *et al.*, 2015; Wappett *et al.*, 2016; Wu *et al.*, 2014), which may not be an ideal strategy to identify interactions in particular functions or relevance to particular cancers. These demonstrate a need for computational approaches to prioritise candidate gene pairs for validation but this thesis will focus on the interactions with *CDH1* with particular importance in breast and stomach cancers, although these partners may be applicable in other cancers. As such, this thesis presents a query-based method, amenable to identification of candidate partners for a selected gene of functional or translational importance such as *CDH1*.

## **1.3 E-cadherin as a Synthetic Lethal Target**

E-cadherin is a transmembrane protein (encoded by *CDH1*) with several characterised functions in the cytoskeleton and cell-to-cell signaling. Here we outline the key known functions of E-cadherin and its importance in cancer biology. *CDH1* is a tumour suppressor gene, with loss of function occurring in both familial (germline mutations) and sporadic (somatic mutations) cancers. As such, *CDH1* inactivation is a prime example of a genetic event that could be targeted by synthetic lethality for anti-cancer treatments. Most notably this includes patients at risk of developing hereditary breast and stomach cancers for which conventional surgical or cytotoxic chemotherapy is not ideal (due to impact of quality of life) and who have a known genetic aberration in their familial syndromic cancers. Effective treatments against *CDH1* inactivation would also benefit patients with sporadic diffuse gastric cancers since they often present with symptoms at a late stage.

### **1.3.1 The *CDH1* gene and its Biological Functions**

The tumour suppressor gene *CDH1* is implicated in hereditary and sporadic lobular breast cancers (Berx *et al.*, 1996; Berx and van Roy, 2009; De Leeuw *et al.*, 1997; Masciari *et al.*, 2007; Semb and Christofori, 1998; Vos *et al.*, 1997). The *CDH1* gene encodes the E-cadherin protein and is normally expressed in epithelial tissues, where it has also been identified as an invasion suppressor and loss of *CDH1* function has been implicated in breast cancer progression and metastasis (Becker *et al.*, 1994; Berx *et al.*, 1995; Christofori and Semb, 1999).

### **1.3.1.1 Cytoskeleton**

The primary function of *CDH1* is cell-cell adhesion forming the adherens junction, maintaining the cytoskeleton and mediating molecular signals between cells. The function of the adherens complex is particularly important for cell structure and regulation because it interacts with cytoskeletal actins and microtubules. The cytoskeletal role of E-cadherin maintains healthy cellular viability and growth in epithelial tissues including cellular polarity. E-cadherin is not essential to cellular viability but loss in epithelial cells does lead to defects in cytoskeletal structure and proliferation. In addition to a central role in the adherens complex, E-cadherin is involved in many other cellular functions and thus *CDH1* is regarded as a highly pleiotropic gene.

### **1.3.1.2 Extracellular and Tumour Micro-Environment**

As a transmembrane signaling protein E-cadherin also interacts with the extracellular environment and other cells, most notably forming tight junctions between cells. These junctions serve to both regulate movement of ion signals between cells and separate membrane proteins on the apical and basal surfaces of a cell, maintaining cell polarity. Thus E-cadherin is an important regulator of epithelial tissues by intercellular communication. It also has important roles in the extracellular matrix, including fibrin clot formation. The role of intercellular interactions and the tissue micro-environment are important themes in cancer research, being a potential mechanism for cancer progression and malignancy in addition to its potential for specifically targeting tumour cells.

### **1.3.1.3 Cell-Cell Adhesion and Signalling**

The signals mediated by tight junctions are also passed on to intracellular signalling pathways and thus E-cadherin also has a role in maintaining cellular function and growth. One such example is the regulation of  $\beta$ -catenin which interacts with both the actin cytoskeleton and acts as a transcription factor via the WNT pathway. Similarly, the HIPPO and PI3K/AKT pathways are implicated in being mediated by E-cadherin, having roles in promoting cell survival, proliferation, and repressing apoptosis. E-cadherin shares several downstream pathways with signaling pathways such as integrins and thus indirectly interacts with them, particularly since feedback loops may occur in such pathways. Conversely, the multifaceted roles of E-cadherin have been shown with differing overexpression in ovarian cells promoting tumour growth, while it maintains healthy cellular functions in other cells.

### **1.3.2 *CDH1* as a Tumour (and Invasion) Suppressor**

E-cadherin has key roles in maintaining cellular structure and regulating growth, consistent with *CDH1* being a tumour suppressor gene. Loss of *CDH1* in epithelial tissues leads to disrupted cell polarity, differentiation, and migration. E-cadherin loss has been identified as a recurrent driver tumour suppressor mutation in sporadic cancers of many tissues including breast, stomach, lung, colon, and pancreas tissue.

#### **1.3.2.1 Breast Cancers and Invasion**

E-cadherin loss in breast cancers has been shown to cause increased proliferation, lymph node invasion, and metastasis with poor cell-cell contact. Thus *CDH1* gene has also been implicated as an invasion suppressor, with a key role in the epithelial-mesenchymal transition (EMT), an established mechanism of cancer progression (Hanahan and Weinberg, 2011). The epithelial-mesenchymal transition is important during development and wound healing but such changes in cellular differentiation also occur in cancers. If *CDH1* is inactivated by mutation or DNA methylation (Berx *et al.*, 1996; Guilford, 1999; Machado *et al.*, 2001), it is likely that EMT will drive growth of E-cadherin deficient cancers (Berx and van Roy, 2009; Graziano *et al.*, 2003; Polyak and Weinberg, 2009). While loss of E-cadherin is not sufficient to cause EMT or tumourigenesis, it is an important step in this mechanism of tumour progression and a potential therapeutic intervention may therefore also impede cancer progression and have activity against advanced stage cancers.

### **1.3.3 Hereditary Diffuse Gastric Cancer and Lobular Breast Cancer**

*CDH1* loss of function mutations also causes familial cancers, including diffuse gastric cancer and lobular breast cancer (Graziano *et al.*, 2003; Guilford *et al.*, 2010, 1999; Oliveira *et al.*, 2009). Individuals carrying a null mutation in *CDH1* haave a syndromic predisposition to early-onset these cancers, known as Hereditary Diffuse Gastric Cancer (HDGC) (Guilford *et al.*, 1998). Due to the loss of an allele, these individuals are prone to carcinogenic lesion in the breast and stomach when the other allele is inactivated, occurring much more frequently and thus younger than in individuals without a second functional allele of *CDH1*. The loss of the second allele is most often hypermethylation suppressing expression rather than mutation, although loss of heterozygosity may also occur. Therefore HDGC is an autosomal dominant cancer syndrome with incomplete penetrance. The “lifetime” (until age 80 years) risk for mutation carriers of diffuse

gastric cancer is 70% in males and 56% in females. In addition, the lifetime risk of lobular breast cancer is 42% in female mutation carriers.

HDGC affects less than 1 in a million people globally (Ferlay *et al.*, 2015) and less than 1% of gastric cancers. However, HDGC is documented to affect several hundred families globally. E-cadherin mutations in the germline is implicated in 1-3% of gastric cancers presenting with a family history, varying between high and low incidence populations. E-cadherin is also mutated in 13% of sporadic gastric cancers.

While diagnostic testing for *CDH1* genotype has enabled more effective management of HDGC and improved patient outcomes, there are still limited options for clinical interventions (Guilford *et al.*, 2010). Individuals with a family history of HDGC are recommended to be tested for *CDH1* mutations in late adolescence and are offered prophylactic stomach surgery before the risk of developing cancers increases with age. Another option is annual endoscopic screening to diagnose early stage stomach cancers with surgical intervention once they are detected (Oliveira *et al.*, 2013). However, these early stage cancers are difficult to detect and may be missed in regular screening. Thus patients carrying *CDH1* mutations either have surgical interventions with a significant impact on quality of life and risk of complications or remain at risk of developing advanced stage stomach cancers. Due to the lower mortality rate due to stomach cancers, there is increasing concerns among these HDGC families on the elevated risk of lobular breast cancers for women later in life.

The current clinical management of HDGC still has significant risks for patients and therefore a greater understanding of the molecular and cellular function of *CDH1* is important for its role in these cancers. Such studies may lead to alternative treatment strategies such as pharmacological treatments with specificity against *CDH1* null cells, once they lose the second allele. While a loss of gene function cannot be targeted directly, designing a treatment with specificity against *CDH1* may also have activity in sporadic cancers in a range of epithelial cancers. Thus an effective treatment against *CDH1* mutant cancers would potentially have significant therapeutic and preventative applications in a large number of patients.

### 1.3.4 Somatic Mutations

#### 1.3.4.1 Mutation Rate

Estimates for the prevalence of *CDH1* somatic mutations in sporadic cancers varies. The Cancer Gene Census (Futreal *et al.*, 2004; Pleasance *et al.*, 2010) detected 994 distinct mutations in 10,143 tumour samples (at a rate of 7.52%), COSMIC (2016)

detected 632 distinct mutations in 43,865 tumour samples (at a rate of 1.71%), and the NCI60 detected mutations in 13.2% of 53 cancer cell lines. While there is no consensus on the prevalence of *CDH1* mutations, the vast variability of mutations is consistent with its role as a tumour suppressor and it has been found to be recurrently mutated in a wide range of cancers of epithelial tissues.

COSMIC (2016) reports *CDH1* mutations in 40 cancer tissue types including stomach (11.40% in N=1342), breast (10.29% in N=3343), large colon (2.87%), skin (2.83%), endometrial (2.81%), and bladder (1.9%) cancer. ICGC reports *CDH1* mutations in 29 cancer tissue types including skin (23.41% in N=598), breast (14.50% in N=1696), ovary (13.98%, N=93), and stomach (11.41% in N=289) cancer samples. *CDH1* mutations are reported at similar rates in breast and stomach cancer in other cancer genomics projects and studies across distinct populations. cBioPortal reports *CDH1* mutation prevalence in stomach cancer at 16.7% (Tokyo Univ., Kakiuchi, 2014, N=30), 15% (Pfizer/UHK, Wang, 2014, N=100), 14.1% (Tianjin Medical University, Chen, 2015, N=78), and 9.4% (TCGA , 2017 prov, N=393). cBioPortal also reports *CDH1* mutation prevalence in breast cancer at 12.7% (TCGA, 2017 prov, N=963) and 10.8% (METABRIC, 2012/2016, N=2051). The rare plasmacytoid bladder cancer subtype also has a high prevalence of *CDH1* mutations in COSMIC (2016) at a rate of 81.8% (N=33). These demonstrate that *CDH1* is important in many cancers and targeting *CDH1* may be widely applied against sporadic cancers in addition to hereditary cancers. However, some of these studies have focused on disease subgroups (such as lobular subtype or estrogen receptor negative breast cancers) with poor patient outcomes which may have inflated the prevalence of *CDH1* mutations which are more common in some of these subtypes.

#### 1.3.4.2 Co-occurring Mutations

Another concern is that *CDH1* mutations may co-occur with other known cancer driver genes such as highly prevalent tumour suppressor gene *TP53* or the proto-oncogene *PIK3CA*. cBioPortal reports the prevalence of the mutations in these genes at 10% for *CDH1*, 49% for *TP53*, 22% for *PIK3CA* in stomach cancer (TCGA, 2017 prov, N=393). There is no evidence of significant co-occurring mutations between *CDH1* and *PIK3CA* (mutex  $p = 0.231$ ) but there is evidence for significant mutually exclusive mutations for *CDH1* (mutex  $p = 0.002$ ) and *PIK3CA* (mutex  $p = 0.004$ ) with *TP53*. cBioPortal also reports the prevalence of the mutations in these genes at 13% for *CDH1*, 32% for *TP53*, 36% for *PIK3CA* in breast cancer (TCGA, 2017 prov, N=3963). There is evidence of significant co-occurring mutations with *CDH1* and *PIK3CA* (mutex  $p < 0.0001$ ) and

evidence for significant mutually exclusive mutations for *CDH1* (mutex  $p = 0.003$ ) and *PIK3CA* (mutex  $p = 0.032$ ) with *TP53*.

These cancer driver mutations have distinct molecular features, leading to disease progression in distinct ways which is a concern for drug resistance when several mutations may accumulate, particularly for sporadic cancers where this is common. Targeting *CDH1* specifically is most suitable for hereditary cancers and combination therapies may be required for sporadic cancers. However, *CDH1* and *TP53* mutant cancers appear to be distinct pathways of tumour progression so the high impact of *TP53* mutation on cancer cells need not be considered for the purposes of studying *CDH1*.

### 1.3.5 Models of *CDH1* loss in cell lines

Previous work our research group has published used a model of homozygous *CDH1*<sup>-/-</sup> null mutation in non-malignant MCF10A breast cells to show that loss of *CDH1* alone was not sufficient to induce EMT with compensatory changes in the expression of other cell adhesion genes occurring (Chen *et al.*, 2014). However, *CDH1* deficient cells did manifest changes in morphology, migration, and weaker cell adhesion (Chen *et al.*, 2014).

This *CDH1*<sup>-/-</sup> MCF10A model has been used in a genome-wide screen of 18,120 genes using small interfering RNAs (siRNA) and a complementary drug screen using 4,057 compounds to identify synthetic lethal partners to E-cadherin (Telford *et al.*, 2015). One of the strongest candidate pathways identified by Telford *et al.* (2015) were the GPCR signalling cascades, which were highly enriched by Gene Ontology analysis of the candidate synthetic lethal partners the primary siRNAs screen. This was supported by validation with Pertussis toxin, known to target G<sub>αi</sub> signalling (Clark, 2004), as were various candidate cytoskeletal pathways by inhibition of Janus kinase (JAK/STAT) and aurora kinase. The drug screen also produced candidates in histone deacetylase (HDAC) and phosphoinositide 3-kinase (PI3K) which were supported by validation and time course experiments.

## 1.4 Summary and Research Direction of Thesis

Genomics technologies and the data made available from them have great potential for understanding of genetics and improving healthcare, including identification of genes altered in cancer for molecular diagnosis, prognostic biomarkers, and therapeutic targets. This has been demonstrated with the identification of cancer genes in many cancers,

distinguishing tumour subtypes by expression profiles, and the development of targeted therapies against oncogenes (such as *BRAF* and tumour suppressors (such as *BRCA1*). Synthetic lethality is an important genetic interaction to study fundamental cellular functions and exploit them for biomarkers and cancer treatment. They present a means to target loss of function mutations and genetic dysregulation in tumour suppressor genes by identifying interacting partners with redundant or compensating molecular functions.

*CDH1* (encoding E-cadherin) is an example of a tumour suppressor gene implicated in sporadic breast and stomach cancers. Germline mutations in *CDH1* are also found in many patients with familial early onset cancers (HDGC). Discovery of synthetic lethal partners would contribute to an understanding on the molecular mechanisms driving the growth of *CDH1* deficient tumours and identification of potential therapeutic targets or chemopreventative agents for management of HDGC. The clinical potential of the synthetic lethal approach has been demonstrated with the application of olaparib against *BRCA1* and *BRCA2* mutations Lord *et al.* (2015) but there remains the need to systematically identify synthetic lethal partner genes for other tumour suppressors such as *CDH1*. A synthetic lethal screen has been conducted on breast cell lines Telford *et al.* (2015) but computational approaches to identification of synthetic lethal partners of *CDH1* remains to be done.

While there are a wide range of experimental and computational approaches to synthetic lethal discovery, many are limited to particular applications, prone to false positives, inconsistent across independent approaches, or enriched for particular genes of interest. Therefore synthetic lethal interactions are difficult to replicate or apply in the clinic. Computational approaches to synthetic lethality are not widely adopted by the cancer research community and experimental approaches cannot be combined to study synthetic lethality at a genome-wide scale. However, these show interest in synthetic lethal discovery in the community and the need for robust predictions of synthetic lethal interactions in cancer and human tissues.

Effective screening, prediction, and analysis of synthetic lethal interactions are a crucial part of developing next generation anti-cancer strategies. Therefore, we propose developing a computational statistical procedure to identify synthetic lethal interactions and construct gene networks. This will enable the development of personalised medicine targeted to particular molecular aberrations. Genetic tests and genomics have the potential to revolutionise cancer screening, diagnosis, and prognostics; tar-

geted therapeutics, similarly, have applications in prevention and therapy of sporadic or hereditary cancers with known molecular properties.

To address the concerns raised by recent computational approaches to synthetic lethal discovery in cancer (Jerby-Arnon *et al.*, 2014; Lu *et al.*, 2015; Wappett *et al.*, 2016), I present similar analysis using solely gene expression data which is widely available for a large number of samples in many different cancers. This uses a statistical methodology the Synthetic Lethal Interaction Prediction Tool (SLIPT) developed for this purpose. To further determine the limitations and implications of synthetic lethal predictions, modelling and simulation was performed upon the statistical behaviour of synthetic lethal gene pairs in genomics data. Comparison of synthetic lethal gene candidates from public data analysis and experimental candidates, pathway analysis, and networks structure will also be presented to investigate the relationships between synthetic lethal candidates. Release of R codes used for simulation, prediction, and analysis will enable adoption of the methodology in the cancer research community and comparison to existing methods.

My thesis aims to develop such predictions for synthetic lethal partner genes with a focus on the example of E-cadherin to compare to the findings of Telford *et al.* (2015), develop of network approaches for pathway structure, and simulate gene expression on pathway structure with the following bioinformatics and computational biology investigations:

- Developed a query-based synthetic lethal detection methodology (SLIPT) for use on gene expression data
- Adapt this methodology to utilise somatic mutation for query genes or candidate pathway metagenes
- Apply Synthetic lethal prediction to public breast cancer genomics data from TCGA (TCGA, 2012)
- Identify over-represented biological pathways using Reactome (Croft *et al.*, 2014) among synthetic lethal candidate partner genes
- Compare these at the gene and pathway level to experimental screen data in breast cell lines from Telford *et al.* (2015)
- Replicate these analyses in stomach cancer genomics data from TCGA (Bass *et al.*, 2014)

- Determine whether synthetic lethal candidates have importance in biological networks of candidate partner pathways
- Determine whether there are relationships within biological network structures between experimental and predicted gene candidate partners
- Develop a statistical model of synthetic lethal gene expression
- Simulate gene expression with synthetic lethal genes and pathway structures
- Evaluate the effects of modification to the SLIPT procedure on it's statistical performance
- Compare the statistical performance of the SLIPT procedure to alternative statistical methods
- Release a synthetic lethal prediction methodology (SLIPT) to the research community for wider application

## Thesis Aims

- To develop a statistical approach to detect synthetic lethal gene pairs in cancer from expression data
- To apply this methodology to public cancer gene expression data against *CDH1* and analyse pathway structure with comparisons to experimental screen data
- To construct a statistical model of synthetic lethality in multivariate normal expression data
- To develop a simulation pipeline of expression with pathway structure on a high-performance computing cluster
- To examine the statistical performance of the methodology with simulated expression including pathways and compare it to other approaches
- To release the synthetic lethal detection methodology and pathway simulation procedure as R software packages

# Chapter 2

## Methods and Resources

In this Chapter, I will outline the various existing resources and methods utilised throughout this project. This includes public data repositories, stable and development releases of software packages (mostly for the R programming environment), and implementation of bioinformatics methods and statistical concepts with Shell or R scripts developed for this purpose. Methods and packages developed specifically for this project will be covered in more detail along with preliminary data to demonstrate and support their use in Chapter 3.

### 2.1 Bioinformatics Resources for Genomics Research

#### 2.1.1 Public Data and Software Packages

Various bioinformatics resources, such as databases and methods, have become integral parts of genetics and genomics research. Reference genomes, genotyped variants, gene expression, and epigenetics profiles are among the most commonly used resources. Gene expression data in particular is widely available from many microarray and RNA-Seq studies, from repositories such as Gene Expression Omnibus (GEO) (Clough and Barrett, 2016), caArray (Heiskanen *et al.*, 2014), and ArrayExpress (Rustici *et al.*, 2013). Such profiles are excellent resources to examine the changes of gene expression occurring in cancers and the variation between samples. These microarray initiatives have set a precedent for data sharing, data mining, and the wider benefits of publicly available data for enabling the scientific community to further utilise the data rather than a single research group or consortium (Rung and Brazma, 2013). The practice of integrating findings from publicly available genomics data with the research questions and experimental results of individual research groups has carried over into RNA-Seq

datasets including the large-scale cancer genomics projects (Zhang *et al.*, 2011). This thesis is one such example of an investigation enabled by this wider movement and tools developed in various disciplines to generate, process, and disseminate genomic-scale data.

Along with databases, it is also becoming common practice for bioinformatics researchers to release their code as open-source or provide a software package to enable replication of the findings or further applications of the methods (Stajich and Lapp, 2006). This is part of a wider movement in software and data analysis with many tools to facilitate such work being released for use in Linux or the R programming environment (R Core Team, 2016). In addition to the R packages hosted on CRAN (CRAN, 2017), the Bioconductor repositories (Gentleman *et al.*, 2004) also contain many packages specifically for applications in bioinformatics, and the GitHub site hosts many packages in various stages of development and early release. Packages from these various sources have been used throughout this project and cited where-ever possible. Several R packages have been developed during this thesis project and either publicly released on GitHub or prepared to accompany a publication.

### 2.1.1.1 Cancer Genome Atlas Data

Molecular profile data from normal and tumour samples was downloaded from publicly available sources, using the TCGA (TCGA, 2012) and the International Cancer Genome Consortium (ICGC) web portals (Zhang *et al.*, 2011). These include gene expression (RNA-Seq), somatic mutations, and anonymous clinical data. These versions downloaded were on the 6<sup>th</sup> of August 2015 (Release 19) and the 2<sup>nd</sup> of May 2016 (Release 20) for breast and stomach cancer respectively via the ICGC data portal (<https://dcc.icgc.org/>).

Performing a genomic alignment remains a challenge in bioinformatics and methods to do so may yet be improved (Chen and Tompa, 2010). However, the statistical and biological aspects of bioinformatics are the focus of this thesis, comparing alignment methods is outside the scope of these investigations. The TCGA project (TCGA, 2012) used widely adopted tools: “Bowtie” for alignment (Langmead *et al.*, 2009), “mapsplice” to detect splice sites (Wang *et al.*, 2010), and the Reads Per Kilobase per Million mapped reads (RPKM) approach to qualify reads per transcript as a measure of gene expression (Mortazavi *et al.*, 2008). These are widely acceptable tools for processing RNA-Seq data which were used to produce the raw counts of mapped reads (tier 1) and normalised expression data (tier 3) publicly available from TCGA.

Raw count and RSEM normalised TCGA expression data from Illumina RNA-Seq protocols were available from 1,177 breast samples (113 normal, 1,057 primary tumour, and 7 metastases) for 20,501 genes. TCGA breast somatic mutation data for 981 samples (976 primary tumours and 5 metastases) across 25,836 genes were available including 969 samples (964 primary tumours and 5 metastases) with corresponding RNA-Seq expression data and 19,166 genes mapped from Ensembl identifiers to gene symbols, of which 16,156 had corresponding gene expression information. Unless otherwise stated, the raw counts were used for further processing rather than the RSEM normalised data (provided by TCGA tier 3). For the purposes of this analysis somatic mutations were reported if they were detected to non-synonymous substitutions, frameshifts, or truncations (by premature stop codons) which would likely disrupt the wild-type gene function. Normalised protein expression was used (as provided by TCGA tier 3), generated from reverse phase protein arrays (RPPA) for 142 antibodies targeting 115 genes for 298 TCGA breast samples.

Raw count TCGA expression data (TCGA tier 1) from Illumina RNA-Seq was also available for 450 stomach samples (35 normal, 415 primary tumour) for 20,501 genes. TCGA stomach mutation data was also available for 289 samples across 25807 genes, corresponding to 19436 genes with expression data. Normalised protein expression (RPPA) data was also sourced (from TCGA tier 3) for 201 antibodies targeting 158 genes for 357 stomach samples.

Cell line data was downloaded from the Cancer Cell Line Encyclopaedia (CCLE) on the 7<sup>th</sup> of November 2014 (Barretina *et al.*, 2012; CCLE, 2014). This includes expression data for 1037 cell lines across 19544 genes (last updated on the 18<sup>th</sup> of October 2012), DNA copy number, somatic mutation, drug response, and sample information. Samples include 59 breast cell lines and 38 stomach cell lines.

### 2.1.1.2 Reactome and Annotation Data

Unless otherwise specified, pathway analysis was performed for human pathway annotation from the Reactome database (version 52) with pathway gene sets derived from the `reactome.db` R package. Entrez identifiers were mapped to gene symbols or aliases to match to TCGA expression and mutation data using the `org.Hs.eg.db` R package. Further pathway analysis used breast cancer gene signatures from Gatza and colleagues (Gatza *et al.*, 2011; Gatza *et al.*, 2014). These gene symbols were matched to the relevant dataset and used to construct a matrix of category membership using the `safe` R package (Barry, 2016).

## 2.2 Data Handling

### 2.2.1 Normalisation

Apart from the PAM50 subtyping procedure (Parker *et al.*, 2009), which required RSEM normalised data (J.S. Parker personal communication), the analysis of the RNA-Seq data presented here was based on raw read count data. Raw read counts were log-scaled; samples removed for consistency (based on a Euclidean distance correlation matrix as described in Section 2.2.2); and the final dataset was TMM normalised (Robinson and Oshlack, 2010) then processed using the `voom` function (Law *et al.*, 2014) in the `limma` R package (Ritchie *et al.*, 2015). Protein expression data generated from RPPA was normalised to dilution curves using the `SuperCurve` R package (Ju *et al.*, 2015; Neeley *et al.*, 2009).

### 2.2.2 Sample Triage

The TCGA breast RNA-Seq data were assessed for batch effects using a correlation matrix of the log-transformed raw counts for which a heatmap (Euclidean distance, complete linkage) is shown in Figure A.2. While no major batch effects were detectable between the samples, 9 samples were excluded due to poor correlation with the remaining samples, as detailed in Table 2.1. These samples showed unusual density plots compared to the rest of the dataset, and exhibited low mean read count in Figures 2.1 and 2.2. A heatmap showing key clinical properties of these excluded samples and their correlation with the remainder of the samples is shown in Figure A.1, and a full correlation heatmap (Figure A.2) shows these samples as relatively poorly correlated outliers in the bottom rows and left columns. In addition to the clustering analysis (in Appendix A.1), replicate tumour samples were also examined for sample quality in Appendix A.2. After removal of these samples, the TCGA dataset used for analysis consisted of the remaining 1168 samples (from 1040 patients): 1049 tumour samples, 112 normal tissue for matched samples, and 7 metastases.

Table 2.1: Excluded Samples by Batch and Clinical Characteristics.

| Tissue Source    | Type   | Batch | Plate | Patient | Samples | p53      | Subtype   | Treatment (History)                | Clinical Subtypes (Stage) |     |             |     |
|------------------|--------|-------|-------|---------|---------|----------|-----------|------------------------------------|---------------------------|-----|-------------|-----|
| A7 Christiana    | Tumour | 47    | A227  | A0DB    | 1 of 3  | NA       | Luminal A | Mastectomy                         | (no)                      | ER+ | Ductal      | (2) |
| A7 Christiana    | Tumour | 96    | A220  | A13D    | 1 of 3  | Wildtype | Luminal A | Mastectomy                         | (no)                      | ER+ | Ductal      | (2) |
| A7 Christiana    | Tumour | 96    | A227  | A13E    | 1 of 3  | NA       | Basal     | Lumpectomy                         | (no)                      | ER- | Ductal      | (2) |
| A7 Christiana    | Tumour | 142   | A277  | A26E    | 1 of 3  | NA       | Basal     | Lumpectomy                         | (no)                      | ER+ | Ductal      | (2) |
| A7 Christiana    | Tumour | 47    | A277  | A0DC    | 1 of 2  | NA       | Luminal A | Mastectomy                         | (yes)                     | ER+ | Lobular     | (3) |
| A7 Christiana    | Tumour | 142   | A220  | A26I    | 1 of 2  | Mutant   | Basal     | Lumpectomy                         | (yes)                     | ER- | Ductal      | (2) |
| AC Intl Genomics | Tumour | 177   | A18M  | A2QH    | 2 of 2  | Mutant   | Basal     | Radical Mastectomy                 | (no)                      | ER- | Metaplastic | (2) |
| AC Intl Genomics | Tumour | 177   | A220  | A2QH    | 2 of 2  | Mutant   | Basal     | Radical Mastectomy                 | (no)                      | ER- | Metaplastic | (2) |
| GI ABS IUPUI     | Normal | 177   | A16F  | A2C8    | 1 of 1  | NA       | Luminal A | Radical Mastectomy and Neoadjuvant | (no)                      | ER+ | Ductal      | (2) |

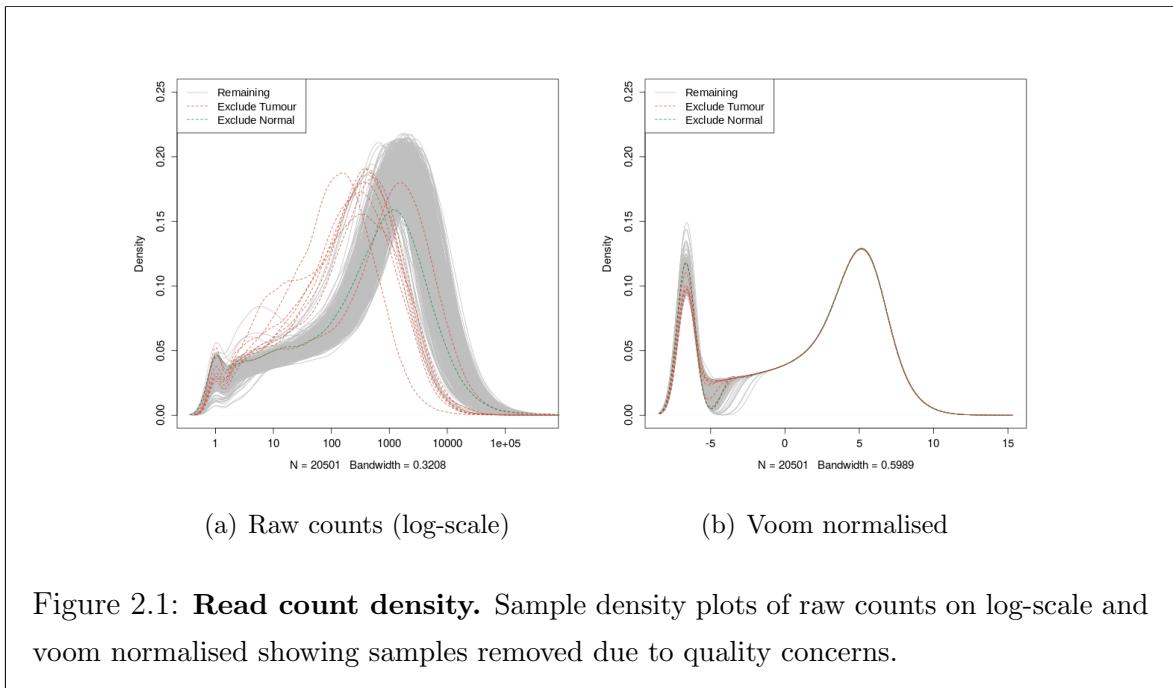


Figure 2.1: **Read count density.** Sample density plots of raw counts on log-scale and voom normalised showing samples removed due to quality concerns.

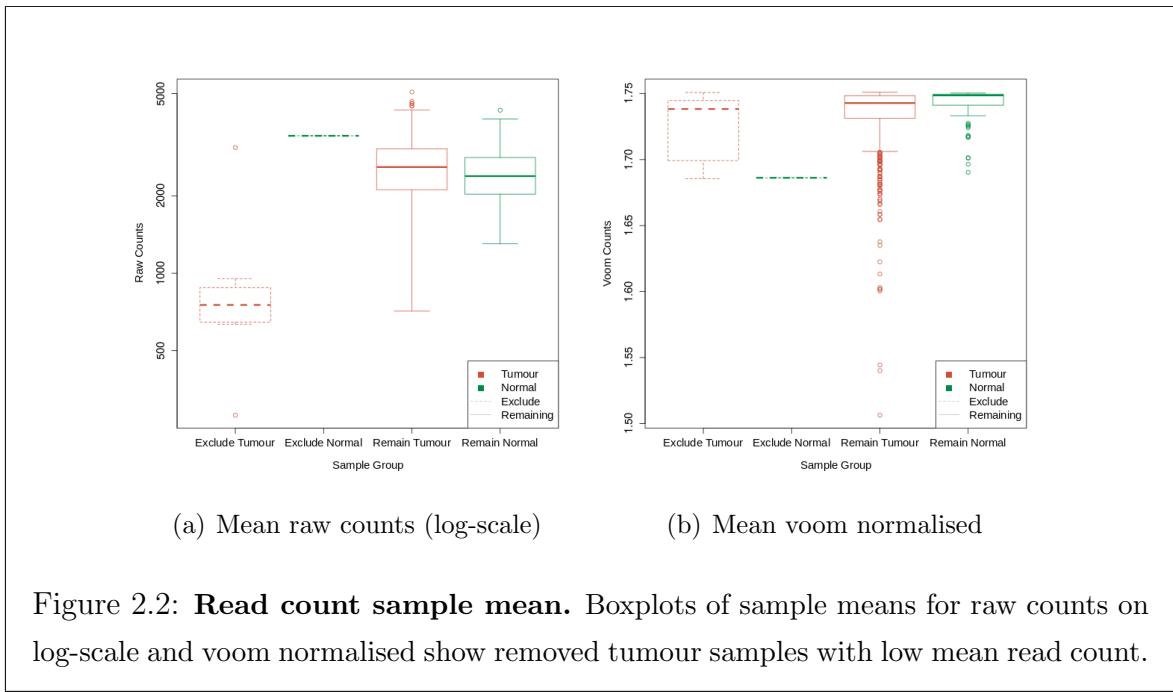


Figure 2.2: **Read count sample mean.** Boxplots of sample means for raw counts on log-scale and voom normalised show removed tumour samples with low mean read count.

Similarly, a correlation matrix of log-transformed raw counts was used to evaluate sample quality for TCGA stomach RNASeq. A tumour sample (patient 4294) was removed due to similar quality concerns leaving a final dataset for 449 samples (from 417 patients): 414 tumour samples and 35 normal tissue samples.

### 2.2.3 Metagenes and the Singular Value Decomposition

A “metagene” offers a consistent signal of pathway (expression) activation or inactivation by dimension reduction of a matrix, avoiding negatively correlated genes averaging out the signal of a mean-based centroid (Huang *et al.*, 2003). Constructing these pathway metagenes used gene sets for Reactome and Gatzka signatures (Gatzka *et al.*, 2011, 2014) as specified above (see Section 2.1.1.2). The singular-value decomposition was performed ( $X = U^T DV$  where  $X$  is the data matrix of the gene set with genes  $\times$  samples) and the leading eigenvector (first column of  $V$ ) corresponding to the largest singular value was used as a metagene for the pathway gene set. To ensure consistent directionality of metagene signals, the median of the gene set in each sample was calculated and correlated against the metagene with the (arbitrary) metagene sign adjusted as needed to conform with the majority of the gene set (i.e., positive correlation between metagene and the median-based centroid). To ensure that genes and pathways were weighted equally, metagenes were derived from a z-transformed dataset of gene expression and samples were scaled (by fractional ranking) for each metagene so that they were comparable on a [0, 1] scale.

#### 2.2.3.1 Candidate Triage and Integration with Screen Data

Candidate triage in combination with the experimental data was intended to integrate findings of the SLIPT analysis with an ongoing experiment project (Chen *et al.*, 2014; Telford *et al.*, 2015). The first procedure to compare the SLIPT gene candidates for *CDH1* with an siRNA experimental screen (Telford *et al.*, 2015) was a direct comparison of the overlapping candidates, presented in a Venn diagram and tested with the  $\chi^2$  test. Since these candidates modestly overlapped at the gene level (even when excluding genes not contained in both datasets), further gene set over-representation analysis was performed for pathways specific to each detection approach and the intersection of the two.

The pathway composition of the intersection was further verified by a permutation resampling analysis (as described in Section 2.3.6): the same number of genes detected by SLIPT were sampled randomly from the universe of genes tested by both approaches. These samplings were performed over 1 million iterations and the path-

way over-representation was compared for each of the 1,652 reactome pathways. These over-representation scores ( $\chi^2$ ) were compared the observed over-representation in the intersection of the SLIPT candidates, with the proportion of resamplings with higher  $\chi^2$  values used for empirical p-values of pathway composition. The  $\chi^2$  test was used as an appropriation of Fisher's exact test on a hypergeometric distribution for resampling to computationally scale pathway over-representation tests across iterations. Pathways for which no resamplings were occurred as high as the observed were reported as  $p < 10^{-6}$ . These empirical p-values were adjusted for multiple comparisons (FDR). Intersection size was not assumed to be constant across resamplings so similarly with the proportion of resamplings with higher or lower intersection size were used to evaluate significance of enrichment or depletion respectively (of siRNA candidate among SLIPT candidate genes).

## 2.3 Techniques

Various statistical, computational, and bioinformatics techniques were performed throughout this thesis. This section describes these techniques and gives the parameters used unless otherwise specified. Where relevant, the R package implementation which provided the technique will be acknowledged.

### 2.3.1 Statistical Procedures and Tests

As described in sections 2.3.4 and 2.2.3, the z-transform has been used to generate z-scores in various analyses in this thesis. Each row of dataset ( $x$ ) is transformed into a scores ( $z$ ) using the mean ( $\mu$ ) and standard deviation ( $\sigma$ ) of the data such that:

$$z = \frac{x - \mu(x)}{\sigma(x)}$$

This generates data where each row (gene) has a mean of 0 and standard deviation of 1. Where plotted as aa heatmap, any data more than 3 standard deviations above or below the mean is plotted as 3 or  $-3$  respectively.

Empirical Bayes differential expression analysis was performed using the `limma` R package (Ritchie *et al.*, 2015). Where specified, the Fisher's exact test,  $\chi^2$  test, and correlation were used to measure associations between variables (as implemented in the `stats` R package (R Core Team, 2016)). Unless otherwise specified, Pearson's correlation was used for correlation analyses ( $r$ ) and coefficient of determination ( $R^2$ ). Where these comparisons are discussed in more detail, Fisher's exact test and  $\chi^2$  tests are supported by a table or Venn diagram, rendered with the `limma` R package (Ritchie

*et al.*, 2015). In some analyses, correlation is further supported by a scatter plot and a line of best fit derived by least squares linear regression.

The `t.test` function (R Core Team, 2016) has also been used to implement the t-test to compare pairs of data. Where relevant, an analysis of variance (ANOVA) has been performed to report significance of multivariate predictors of outcomes, or least squares linear regression performed for the adjusted coefficient of determination ( $R^2$ ) and F-statistic p-value to evaluate the fit of the predictor variables. For some analyses these are supported by boxplot or violinplot visualisation (rendered in R).

Multiple comparisons are adjusted by the Benjamini-Hochberg procedure to control the false discovery rate (FDR) unless otherwise specified (Benjamini and Hochberg, 1995). This procedure adjusts p-values to achieve an average of the proportion of false-positives among significant tests below a threshold,  $\alpha$ . The more stringent Holm-Bonferroni (Holm, 1979) was also applied in some cases to adjust for multiple comparisons and control the family-wise error rate which adjusts p-values so that the probability that any one of the tests is a false-positive (type-1 error) below a threshold,  $\alpha$ .

### 2.3.2 Gene Set Over-representation Analysis

Gene set enrichment over-representation was performed to test whether there is an enrichment of a gene set (such as a biological pathway) among a group of input genes. Such input genes may be predicted synthetic lethal candidates or a subset defined by clustering (in Section 2.3.3) or comparison with experimental candidates (see Section 2.2.3.1). Initially, these tests were performed using the GeneSetDB web tool (Araki *et al.*, 2012) hosted by the University of Auckland on the Reactome pathways (Croft *et al.*, 2014). Since the GeneSetDB tool used an older version of Reactome (version 40), it was difficult to directly compare with the results of other analysis (see sections 2.2.3.1 and 2.3.6) performed on version 52 (as described in Section 2.1.1.2). Thus an implementation of the hypergeometric test in R (R Core Team, 2016) was used to test for over-representation against Reactome (version 52) pathways. Pathways containing less than 10 genes or more than 500 (as performed in GeneSetDB by Araki *et al.*, 2012) were excluded before adjusting for multiple comparisons.

### 2.3.3 Clustering

Clustering analysis when performed uses unsupervised hierarchical clustering with complete linkage (distance calculated from the furthest possible pairing). For correlation matrices or multivariate normal parameters (e.g.,  $\Sigma$ ), the distance metric used was

Euclidean distance. For empirical or simulated gene and pathway expression data correlation distance was used, calculated by  $distance = 1 - cor(t(x))$  where  $cor$  is Pearson's correlation and  $t(x)$  is the transpose of the expression matrix.

### 2.3.4 Heatmap

Standardised z-scores of the data were used to plot heatmaps on an appropriate scale. Raw (log-scale) read counts or voom normalised counts per gene (as specified) were plotted as normalised z-scores on a  $[-3, +3]$  blue-red scale. Similarly, correlations were plotted on a  $[-1, +1]$  blue-red scale. These heatmaps were performed using the linkage and distance specified for the clustering performed in Section 2.3.3. The `gplots` R package (Warnes *et al.*, 2015) was used to generate many of the heatmaps throughout this thesis, along with a customised heatmap function (released as `heatmap.2x`). Where clearly specified, data have been split into subsets with clustering performed separately on each subset with these plotted alongside each other.

### 2.3.5 Modeling and Simulations

Statistical modeling and simulations have been used to test various synthetic lethal detection procedures on simulated data. This involves constructing a statistical model of how synthetic lethality would appear in (continuous normally distributed) gene expression data. Where presented (in Section 3.2.1), the assumptions of the model are stated clearly. The model allows sampling from a multivariate normal distribution (using the `mvtnorm` R package (Genz and Bretz, 2009; Genz *et al.*, 2016)) to generate simulated data with known underlying synthetic lethal partners (detailed in Section 3.2.2). We can test whether statistical procedures, including those developed in this thesis (presented in Section 3.1), are capable of detecting them upon this simulated data. This multivariate normal simulation procedure also enables the inclusion of correlation structure which is either given as correlated blocks of genes or derived from pathway structures (as detailed in Section 3.4.2).

If this multivariate normal distribution was sampled once and the procedure to add known synthetic lethal partners was performed, it generates a simulated dataset. Performing this simulation procedure and testing with a synthetic lethal detection procedure iteratively, these simulations can be used to assess the statistical performance of the detection procedure. The number of iterations (`Reps`) will be given for each simulation result. Typically, these are performed 1000 or 10,000 times depending on computational feasibility of doing so on larger datasets.

Several measures of statistical performance were used to assess the simulations. The following measures used the final classification of the detection procedure, statistical significance for  $\chi^2$ , significance and directional criteria met for SLIPT (see Section 3.1), and an arbitrary threshold:  $< -0.2$  and  $> +0.2$  for negative correlation and correlation respectively. Sensitivity (or “true positive rate”) was measured as the proportion of known synthetic lethal partners predicted to be synthetic lethal. Specificity (or “true negative rate”) was measured as the proportion of known non-synthetic lethal partners predicted not to be synthetic lethal. The “false discovery rate” (also used in adjusting for multiple comparisons) was measured here as the proportion of known non-synthetic lethal partners out of all putative partners predicted by the detection procedure. Statistical “accuracy” is the proportion of true predictions for a detection procedure, which is both the correctly predicted known synthetic lethal partners and correctly negative known non-synthetic lethal partners.

### 2.3.5.1 Receiver Operating Characteristic (Performance)

A more general procedure to measure the statistical performance of a simulation is the Receiver Operating Characteristic (ROC) curve which does not assume a threshold for classification of synthetic lethality but demonstrates the trade-off of sensitivity and specificity (Akobeng, 2007; Fawcett, 2006; Zweig and Campbell, 1993). These curves (implemented with the `ROCR` R package (Sing *et al.*, 2005)) plot the True Positive Rate (sensitivity) against the False Positive Rate (1-specificity) as the prediction threshold is varied. An ideal detection method will have a true positive rate of 1 and a false positive rate of 0, hence the Area Under the ROC curve (AUC or AUROC) is a measure of statistical performance for a detection procedure accounting for this trade-off. AUROC values are typically range from 0.5 the value expected by random chance to 1 for an optimal detection method, however it is possible for an AUROC below 0.5 for a poor detection method that performs worse than random chance. In cancer biology, an AUROC of approximately 0.8 is a predictive biomarker suitable for publication (Hajian-Tilaki, 2013) but predictors with lower AUROC values may still be informative depending on the context. In this thesis, the AUROC values varies widely across simulation parameters and a primarily used for comparisons across these parameters, although they can also be used to refine thresholds for optimal classification.

### 2.3.6 Resampling Analysis

Resampling analyses (e.g., “permutation” analysis) are used to statistically test the significance of an observation without assuming the underlying distribution of expected

test statistics Collingridge (2013). Instead these are derived from randomly shuffling test statistics or randomly sampling predicted candidates. For the purposes of this thesis, this involved randomly sampling genes from those tested to be analysed as putative synthetic lethal candidates. This was performed both for testing the significance of pathway composition in the intersection with experimental gene candidates (Section 2.2.3.1) and for assessing the significance of pathway structure among synthetic lethal candidates (Section 3.4.1.1).

These were analysed to compare the observed synthetic lethal genes against values derived from randomly sampling the same number of genes as observed by synthetic lethal from among the genes tested. Sampling iteratively across many resampling procedures, these resampling-based values form a null distribution that would be observed if the null hypothesis were true. Thus the proportion of resampling-based values across these iterations that are greater than or equal to that observed, forms an empirically derived p-value to test significance.

Resampling was performed for comparison (in Section 2.2.3.1) with fixed experimental screen candidates (Telford *et al.*, 2015) both resampling the number of genes overlapping with the screen candidates and test statistics for pathway enrichment. Resampling analysis was also applied to shortest paths and network metrics (in Section 3.4.1.1) to test significance of directional relationships between synthetic lethal candidate genes within pathway structures.

The number of iterations determines the accuracy of these p-values. For pathway composition (in Section 2.2.3.1), a million iterations were performed using high performance computing (as detailed in Section 2.5.3) to provide sufficient accuracy after adjusting for multiple comparisons across pathways. For the purposes of network analysis (in Section 3.4.1.1), a thousand iterations were sufficient to reject the null hypothesis for the majority of pathways tested before adjusting for multiple comparisons, and thus further iterations were not performed.

## 2.4 Pathway Structure Methods

### 2.4.1 Network and Graph Analysis

Networks are important in considering the structure of relationships in molecular biology, including gene regulation, kinase cellular signaling, and metabolic pathways (Barabási and Oltvai, 2004). Network theory is an interdisciplinary field which combines the approaches of computer science with the metrics and fundamental principles

of graph theory, an area of pure mathematics dealing with relationships between sets of discrete elements. The vast amounts of molecular and cellular data from high-throughput technologies have enabled the application of network-based and genome-wide bioinformatics analysis to examine the complexity of a cell at the molecular level and understand aberrations in cancer. This thesis uses various metrics and analysis procedures developed in Graph and Network theory to analyse graph structure of biological pathways. Where feasible, these have been implemented using the `igraph` R package with such procedures described below (Csardi and Nepusz, 2006). Custom R functions to perform more complex analysis and visualisation of iGraph data objects will be described later.

Graph theory is a branch of pure mathematics which deals with the properties of sets of discrete objects (referred to as a ‘node’ or ‘vertex’) with some pairs are joined (by a ‘link’ or an ‘edge’). While a seemingly reductionist abstraction to mathematically study relationships, graph theory serves has applications in a wide range of studies including life sciences. Network theory is the sub-discipline of graph theory which deals with networks which has become popular due to the vast potential for applications of networks (van Steen, 2010).

Applications vary depending on the situation modelled, particularly in how the edges between vertices are defined, whether they are directed or weighted, and whether multiple redundant edges between a pair of vertices (referred to as ‘parallel edges’) or edges connecting a vertex to itself (referred to as ‘loops’) are permitted in the model. Networks are defined such that the edges represent a relationship between the vertices and may be directed, weighted, or contain parallel edges or loops depending on the application (van Steen, 2010). Unless otherwise stated, graph structures and networks in thesis will be unweighted and have no parallel edges or loops. Where a directional relationship is known or modelled, it will be represented with a directed edge in a directed graph.

## 2.4.2 Sourcing Graph Structure Data

Pathway Commons interaction data was sourced using the paxtools-4.3.0 Java application on October 6th 2015 (Cerami *et al.*, 2011; Demir *et al.*, 2013). This utility was used to source ‘sif’ format interaction data into R (R Core Team, 2016), from which the human Reactome (version 52) dataset of interactions was imported (Croft *et al.*, 2014), matching those used for pathway enrichment analysis. These interactions were

used to construct an adjacency matrix for the Reactome network and subnetworks corresponding to each relevant biological pathway.

### 2.4.3 Constructing Pathway Subgraphs

Subgraphs for each relevant pathway were constructed by matching the nodes in the complete Reactome network to the pathway gene sets (as derived in Section 2.1.1.2). A subgraph with adjacent nodes was constructed by adding nodes which have an edge with a gene in the pathway gene set. The pathways these adjacent nodes belong to were added to form a “meta-pathway” to account for the possibility for nodes within the pathway being linked by the surrounding graph structure.

### 2.4.4 Network Analysis Metrics

The existing network analysis measures applied in this thesis (as described below) used an implementation in the `igraph` R package where it was available (Csardi and Nepusz, 2006). Otherwise, custom features were developed for analysis of iGraph objects in R and released as `igraph.extensions` (as described in Section 3.5.3).

Vertex degree is the number of edges a node has and is a fundamental measure of the importance and connectivity of a network (van Steen, 2010). More connected nodes, such as network hubs, will have a higher vertex degree relative to other nodes. For the purposes of this thesis, vertex degree ignored edge direction with loops (edges with itself) and double edges to the same node excluded.

A fundamental concept in network analysis is a “shortest path”, that is the shortest route via edges between any two particular nodes in a network. These are computed by Dijkstra’s algorithm (Dijkstra, 1959) in the `igraph` R package (Csardi and Nepusz, 2006). Where applicable paths will only use directed edges in a particular direction. Shortest paths are a useful measure of how close nodes are in a network. This is used to compute information centrality, and for further analysis of pathway structure (as described in Section 3.4.1).

Network centrality is an alternative measure of the importance or influence of a node to the graph structure (Borgatti, 2005). Various strategies are used to derive centrality, typically based on how connected the node is or the impact of node removal on the connectivity of the network. One of the most notable is the “PageRank” algorithm, a refinement of eigenvector centrality based on the eigenvectors of the adjacency matrix (Brin and Page, 1998). This is implemented in the `igraph` R package (Csardi and Nepusz, 2006).

Another network centrality measure that has been previously applied to biological protein interaction networks (Kranthi *et al.*, 2013) is the “information centrality”. The information centrality of a node is the relative impact on efficiency (transmission of information via shortest paths) of the network when the node is removed. That is the centrality ( $C$ ) (Kranthi *et al.*, 2013) for node  $n$  in graph  $G$  is defined as:

$$C_n = \frac{E(G) - E(G')}{E(G)}$$

where  $G'$  is the subgraph with the node removed and  $E$  is the efficiency (Latora and Marchiori, 2001) derived from shortest paths ( $d_{ij}$  between nodes  $i$  and  $j$ ) as:

$$E(G) = \frac{2}{N(N-1)} \sum_{i < j \in G} \frac{1}{d_{ij}}$$

The efficiency of the network can be derived from shortest paths implemented in the `igraph` R package and the iterative network centrality computation of each node has been released as an R package (`info.centrality`) and included in the `igraph.extensions` package.

## 2.5 Implementation

### 2.5.1 Computational Resources and Linux Utilities

Several computers were used to process and store data during this thesis (as summarised in Table 2.2), running different versions of Linux operating systems, including a personal laptop computer, laboratory desktop machine, departmental server, and the New Zealand eScience Infrastructure Intel Pan high-performance computing cluster (a supercomputer based at the University of Auckland). Each of these systems support a 64-bit architecture. Current workflows on local machines use Elementary OS (based on the Ubuntu versions given in Table 2.2) and interacting with these via ZSH shell. However, Ubuntu OS and the Bourne Again SHell (bash) were used at the inception of this project and bash is continues to be used for running scripts. Various Linux applications and command-line utilities were used on these machines (as summarised in Table 2.3). As such, the workflows developed in this project should be backwards-compatible with Ubuntu Linux (and other derivatives). The majority of novel methodology and implementations were performed in R which is a cross-platform language, packages developed in R will be available for users of Linux, Mac, and Windows machines.

Table 2.2: Computers used during Thesis

|                       | Viao Laptop               | Lab Machine             | Biochem Server               | NeSI Pan Cluster           |
|-----------------------|---------------------------|-------------------------|------------------------------|----------------------------|
| Operating System (OS) | Elementary OS Freya 0.3.2 | Elementary OS Loki 0.4  | Red Hat Enterprise Maipo 7.2 | Cent OS Final 6.4          |
| Upstream OS           | Ubuntu LTS Trusty 14.04   | Ubuntu LTS Xenial 16.04 |                              |                            |
| Linux Kernel          | 3.19.0-65-generic         | 4.4.0-36-generic        | 3.10.0-327.36.2.el7.x86_64   | 2.6.32-504.16.2.el6.x86_64 |
| Shell: bash           | 4.3.11(1)                 | 4.3.46(1)               | 4.2.46(1)                    | 4.2.1(1)                   |
| Shell: zsh            | 5.0.2                     | 5.1.1                   | 5.0.2                        | 5.2                        |

Table 2.3: Linux Utilities and Applications used during Thesis

|                  |                           | Viao Laptop               | Lab Machine            | Biochem Server               | NeSI Pan Cluster           |
|------------------|---------------------------|---------------------------|------------------------|------------------------------|----------------------------|
|                  | OS                        | Elementary OS Freya 0.3.2 | Elementary OS Loki 0.4 | Red Hat Enterprise Maipo 7.2 | Cent OS Final 6.4          |
|                  | Linux Kernel              | 3.19.0-65-generic         | 4.4.0-36-generic       | 3.10.0-327.36.2.el7.x86_64   | 2.6.32-504.16.2.el6.x86_64 |
| Scripting        | Shell bash                | 4.3.11(1)                 | 4.3.46(1)              | 4.2.46(1)                    | 4.2.1(1)                   |
|                  | Shell zsh                 | 5.0.2                     | 5.1.1                  | 5.0.2                        | 5.2                        |
| Programming      | Python                    | 2.7.6                     | 2.7.12                 | 2.7.5                        |                            |
|                  | Java                      | 1.8.0_101                 | 9-ea                   | 1.8.0_101                    |                            |
|                  | C++                       | 4.8.4                     | 5.4.0                  | 4.8.5                        | 4.4.7                      |
| Text Editor      | nano                      | 2.2.6                     | 2.5.3                  | 2.3.1                        | 2.0.9                      |
|                  | kile ( $\text{\LaTeX}$ )  | 2.1.3                     | 2.1.3                  |                              |                            |
| Version Control  | git                       | 1.9.1                     | 2.11.0                 | 1.7.1                        | 1.8.3.1                    |
| Shell Utilities  | sed                       | 4.4.2                     | 4.4.2                  | 4.4.2                        | 4.4.1                      |
|                  | grep                      | 2.16-1                    | 2.25-1                 | 2.20                         | 2.6.3                      |
|                  | nohup                     | 8.21                      | 8.25                   | 8.22                         | 8.4                        |
| Typesetting      | T $\text{\TeX}$           | 3.1415926                 | 3.14159265             |                              |                            |
|                  | TeXLive ( $\text{\TeX}$ ) | 2013                      | 2015                   |                              |                            |
|                  | PDF $\text{\TeX}$         | 2.5-1                     | 2.6                    |                              |                            |
|                  | pandoc                    | 1.12.2.1                  | 1.16.0.2               |                              |                            |
| Remote Computing | slurm scheduler           |                           |                        |                              | 16.05.6                    |
|                  | OpenSSH                   | 7.2p2                     | 7.2p2                  | 6.6.1                        | 5.3p1                      |
|                  | OpenSSL                   | 1.0.2g                    | 1.0.2g                 | 1.0.01e-fips                 | 1.0.01e-fips               |
|                  | rsync                     | 3.1.0p31                  | 3.1.1p31               | 3.0.9p30                     |                            |
|                  | Globus Online Transfer    |                           |                        | 3.1                          | 3.1                        |
|                  | Cisco AnyConnect VPN      |                           | 3.1.05170              |                              |                            |
| Image Processing | Inkscape                  | 0.48.4                    | 0.91                   |                              |                            |
|                  | GIMP                      | 2.8.10                    | 2.8.16                 |                              |                            |
|                  | ImageMagick               | 6.7.7.10-6                |                        |                              |                            |

## 2.5.2 R Language and Packages

The R programming language has been used for the majority of this thesis. Current R installations across the machines used are given in Table 2.4. Local machines currently run the latest version of the R (at the time of writing) and remote machines run the versions and modules as managed by the system administrator.

Various scripts and packages in this thesis were developed or run in previous versions of RStudio and R but these run without error in the current version of R (and the older

versions on remote machines). The R packages which were used throughout this thesis (as detailed in Table 2.5 with versions specified) were installed from the Comprehensive R Archive Network (CRAN, 2017), Bioconductor (Gentleman *et al.*, 2004, version 3.4; BiocInstaller 1.24.0), or GitHub. These packages were not updated when they would change the functionality of scripts or functions in packages, in particular imported data from annotation packages (used to define gene sets) have been saved as local files to continue using stable versions of these pathway data (across machines).

This is a summary of the key packages which (in addition to their dependencies) have been used throughout this project. Where a package implementation has been central to the methods applied, they are described in more detail in the relevant section. A full table of packages used in this thesis can be found in Appendix B (Table B.1). The R packages developed during this thesis are given in Table 2.6 with the relevant sections describing their implementation and use where appropriate, in addition to further details on these functions in Section 3.5.

Table 2.4: R Installations used during Thesis

|             | Viao Laptop                  | Lab Machine               | Biochem Server                  | NeSI Pan Cluster     |
|-------------|------------------------------|---------------------------|---------------------------------|----------------------|
| OS          | Elementary OS<br>Freya 0.3.2 | Elementary OS<br>Loki 0.4 | Red Hat Enterprise<br>Maipo 7.2 | Cent OS<br>Final 6.4 |
| Programming | R<br>3.3.2                   | 3.3.2                     | 3.3.1                           | 3.3.0-intel (module) |
| Development | RStudio<br>1.0.136           | 1.0.136                   | 1.0.136 (server)                |                      |

Table 2.5: R Packages used during Thesis

| Package    | Version Used | Built | Repository |
|------------|--------------|-------|------------|
| colorspace | 1.3-2        | 3.3.1 | CRAN       |
| curl       | 2.3          | 3.3.1 | CRAN       |
| data.table | 1.9.6        | 3.3.1 | CRAN       |
| dendextend | 1.4.0        | 3.3.2 | CRAN       |
| DBI        | 0.5-1        | 3.3.1 | CRAN       |
| devtools   | 1.12.0       | 3.3.1 | CRAN       |
| dplyr      | 0.5.0        | 3.3.1 | CRAN       |
| ggplot2    | 2.2.1        | 3.3.1 | CRAN       |
| git2r      | 0.18.0       | 3.3.1 | CRAN       |
| gplots     | 3.0.1        | 3.3.1 | CRAN       |
| gtools     | 3.5.0        | 3.3.1 | CRAN       |
| igraph     | 1.0.1        | 3.3.1 | CRAN       |

|              |         |       |                 |
|--------------|---------|-------|-----------------|
| matrixcalc   | 1.0-3   | 3.3.1 | CRAN            |
| mclust       | 5.2.2   | 3.3.1 | CRAN            |
| mvtnorm      | 1.0-6   | 3.3.1 | CRAN            |
| org.Hs.eg.db | 3.1.2   | 3.1.2 | Bioconductor    |
| openssl      | 0.9.6   | 3.3.1 | CRAN            |
| plyr         | 1.8.4   | 3.3.1 | CRAN            |
| purrr        | 0.2.2   | 3.3.1 | CRAN            |
| reactome.db  | 1.52.1  | 3.2.1 | Bioconductor    |
| RColorBrewer | 1.1-2   | 3.3.1 | CRAN            |
| Rcpp         | 0.12.9  | 3.3.1 | CRAN            |
| ROCR         | 1.0-7   | 3.3.1 | CRAN            |
| roxygen2     | 6.0.1   | 3.3.2 | CRAN            |
| shiny        | 1.0.0   | 3.3.1 | CRAN            |
| snow         | 0.4-2   | 3.3.1 | CRAN            |
| testthat     | 1.0.2   | 3.3.2 | CRAN            |
| tidyverse    | 1.1.1   | 3.3.2 | GitHub (hadley) |
| sm           | 2.2-5.4 | 3.3.1 | CRAN            |
| Unicode      | 9.0.0-1 | 3.3.2 | CRAN            |
| vioplot      | 0.2     | 3.3.1 | CRAN            |
| viridis      | 0.3.4   | 3.3.2 | CRAN            |
| xml2         | 1.1.1   | 3.3.2 | CRAN            |
| xtable       | 1.8-2   | 3.3.1 | CRAN            |
| zoo          | 1.7-14  | 3.3.1 | CRAN            |
| graphics     | 3.3.2   | 3.3.2 | base            |
| grDevices    | 3.3.2   | 3.3.2 | base            |
| cluster      | 2.0.5   | 3.3.1 | base            |
| graphics     | 3.3.2   | 3.3.2 | base            |
| grDevices    | 3.3.2   | 3.3.2 | base            |
| Matrix       | 1.2-8   | 3.3.1 | base            |
| stats        | 3.3.2   | 3.3.2 | base            |

Table 2.6: R Packages Developed during Thesis

| Package Name       | Description and GitHub Repository   | Section |
|--------------------|---|---------|
| <code>slipt</code> | Synthetic lethal detection by SLIPT (to accompany publication)<br><a href="https://github.com/TomKellyGenetics/slipt">https://github.com/TomKellyGenetics/slipt</a>   | 3.1     |
| visualisation      | <code>vioplotx</code><br>Customised violin plots (based on <code>vioplot</code> )<br><a href="https://github.com/TomKellyGenetics/vioplotx">https://github.com/TomKellyGenetics/vioplotx</a>  | 3.4     |
|                    | <code>heatmap.2x</code><br>Customised heatmaps (based on <code>gplots</code> )<br><a href="https://github.com/TomKellyGenetics/heatmap.2x">https://github.com/TomKellyGenetics/heatmap.2x</a>   |         |
| igraph.extensions  | <code>igraph.extensions</code><br>Meta-package to install the follow iGraph functions<br><a href="https://github.com/TomKellyGenetics/igraph.extensions">https://github.com/TomKellyGenetics/igraph.extensions</a>                                    | 3.5.3   |
|                    | <code>plot.igraph</code><br>Custom plotting of directed graphs<br><a href="https://github.com/TomKellyGenetics/plot.igraph">https://github.com/TomKellyGenetics/plot.igraph</a>   | 2.4.4   |
|                    | <code>info.centrality</code><br>Computing information centrality from network efficiency<br><a href="https://github.com/TomKellyGenetics/info.centrality">https://github.com/TomKellyGenetics/info.centrality</a>                                     | 3.4.2   |
|                    | <code>pathway.structure.permutation</code><br>Testing pathway structure with resampling analysis<br><a href="https://github.com/TomKellyGenetics/pathway.structure.permutation">https://github.com/TomKellyGenetics/pathway.structure.permutation</a> | 3.4.1.1 |
|                    | <code>graphsim</code><br>Generating simulated expression from graph structures<br><a href="https://github.com/TomKellyGenetics/graphsim">https://github.com/TomKellyGenetics/graphsim</a>   | 3.4.2   |

### 2.5.3 High Performance and Parallel Computing

Another enabling technology for bioinformatics is parallel computing, performing independent operations in separate cores: this “multithreading” is widely used to increase the time to compute results. Bioinformatics is particularly amenable to this since performing multiple iterations of a simulation or testing separate genes is often “embarrassingly parallel”, being completely independent of the results of each other. As such parallel computing is offered by many high-performance “supercomputers” including national research infrastructure.

The New Zealand eScience Infrastructure (NeSI) is a computating resource providing the Intel Pan cluster hosted by the University of Auckland (NeSI, 2017). The Pan cluster used throughout this thesis project to optimise and perform computations which would have otherwise been infeasible in the timeframe of thesis. Such technological developments and infrastructure initiatives have enabled bioinformatics research including this project. High performance computing on the Pan cluster was used extensively in this project including for resampling analysis (in sections 2.3.6 and 3.4.1.1), calculating information centrality (in Section 2.4.4), and in simulations (in sections 2.3.5, 3.2, and 3.4.2)

Scripts and data were transferred between the Pan cluster and University of Otago computing resources by `rsync` or the Globus file transfer service (Globus, 2017). R scripts (R Core Team, 2016) were run in parallel with the “simple network of workstations” `snow` R package Tierney *et al.* (2015). This utilised the “message passing interface” (Yu, 2002) when it was feasible with memory requirements to run in par-

allel across multiple compute nodes, otherwise SOCKS was used to access multiple cores within an instance of R and pass input data to them. R jobs were submitted to queue for available resources and run on the Pan cluster via the Slurm (Simple Linux Utility for Resource Management) workload manager (Slurm, 2017). When running R scripts across many parameters or for memory-intensive jobs, Slurm array job submission and independent submission of different parameters via shell commands with arguments passed to R. In some cases, this submission was automated across a range of parameters with Bash scripts.

# Chapter 3

## Methods Developed During Thesis

In this Chapter, I will outline the rationale and development of various methods used throughout this thesis to examine synthetic lethality in gene expression data, graph structures, models and simulations. First by describing the Synthetic Lethal Interaction Prediction Tool (SLIPT), a bioinformatics approach to triage of synthetic lethal candidate genes. This is considered one of the main research outputs of the thesis, which is supported by comparisons to an experimental screen from a related project and performance on simulated data. These supporting data will be covered in further Chapters but preliminary data to support the use and design of SLIPT are provided alongside description of the method. This includes the construction of a statistical model of synthetic lethality in (continuous multivariate Gaussian) gene expression data, which enables testing SLIPT upon simulated data with known synthetic lethal partners. Another key component of the simulation pipeline used later is the generation of simulated data from a known graph structure or simulated biological pathway. The development of this simulation procedure and other statistical treatment of graph and network structures will also be covered. Various R packages have been developed to support this project, most notably the `slipt` package to implement the SLIPT methodology. The additional R packages for handling graph structures, simulations, and custom plotting features will also be described as research outputs of this thesis, methods applied throughout, and contributions to the open-source software community that made this project feasible.

### 3.1 A Synthetic Lethal Detection Methodology

The SLIPT methodology identifies gene expression patterns consistent with synthetic lethal interactions between a query gene and a panel of candidate interacting partners.

Gene expression is called low, medium, or high by separating samples into tertiles (3-quantiles) for each gene. Genes with insufficient expression across all samples were excluded by requiring that the first tertile of raw counts is above zero. Then a  $\chi^2$  test is performed between the query gene and each candidate partner, with the p-values for the  $\chi^2$  test being corrected for multiple testing using false discovery rate (FDR) error control to reduce false positives for large candidate gene panels (Benjamini and Hochberg, 1995). Significance was called only if FDR adjusted p-values were below the threshold  $p < 0.05$ . A synthetic lethal interaction is predicted (as shown in Figure 3.1) when (i) the  $\chi^2$  test is significant; (ii) observed low-query, low-candidate samples are less frequent than expected; and (iii) observed low-query, high-candidate and high-query, low-candidate samples are more frequent than expected.

The synthetic lethal prediction procedure has also been adapted to utilise somatic mutation data for the query gene. This is intended to utilise a query gene known to

|                                   |        | Candidate Gene              |        |                             |
|-----------------------------------|--------|-----------------------------|--------|-----------------------------|
|                                   |        | Low                         | Medium | High                        |
| Query Gene<br>(e.g. <i>CDH1</i> ) | Low    | Observed less than expected |        | Observed more than expected |
|                                   | Medium |                             |        |                             |
|                                   | High   | Observed more than expected |        |                             |

Figure 3.1: **Framework for synthetic lethal prediction.** Synthetic Lethal Interaction Prediction Tool (SLIPT) was designed to identify candidate interacting genes from gene expression data using the  $\chi^2$  test against a query gene. Samples are sorted into low, medium, and high expression quantiles for each gene to test for a directional shift. A sample being low in both genes of a synthetic lethal pair is unlikely, since loss of both genes will be deleterious, and is expected to be statistically under-represented in a gene expression dataset. We expect a corresponding (symmetric) increase in frequency of sample with low-high gene pairs. Synthetic lethal candidate (exprSL) partners of a gene are identified by running this procedure on all possible partner genes, selecting those with an FDR-adjusted  $\chi^2$  p-value of  $p < 0.05$ , and meeting the directional criteria. Since synthetic lethal genes are partners of each other commutatively, the symmetric direction criteria are all required such that synthetic lethal genes will be predicted to be partners of each other.

|                                   |           | Candidate Gene              |        |                             |
|-----------------------------------|-----------|-----------------------------|--------|-----------------------------|
|                                   |           | Low                         | Medium | High                        |
| Query Gene<br>(e.g. <i>CDH1</i> ) | Mutation  | Observed less than expected |        | Observed more than expected |
|                                   | Wild-type | Observed more than expected |        |                             |

Figure 3.2: **Synthetic lethal prediction adapted for mutation.** Synthetic Lethal Interaction Prediction Tool (SLIPT) was also adapted to identify candidate interacting genes using (somatic) mutation data of the query gene in the  $\chi^2$  test. Samples are sorted into low, medium, and high expression quantiles for each candidate gene and tested for a directional shift against mutation status of the query gene. A sample having low expression or mutation for the synthetic lethal pair is expected to be unlikely with a corresponding increase in frequency of sample with mutant-high or wild-type-low gene pairs. Synthetic lethal candidate (mtSL) partners of a gene are identified by running this procedure on all possible partner genes, selecting those with an FDR-adjusted  $\chi^2$  p-value of  $p < 0.05$ , and meeting the directional criteria.

be recurrently mutated in the disease (and dataset), with the majority of mutations inactivating gene function (such as null or frameshift mutations). A synthetic lethal interaction is predicted (as shown in Figure 3.2) when (i) the  $\chi^2$  test is significant; (ii) observed mutant-query, low-candidate samples are less frequent than expected; and (iii) observed mutant-query, high-candidate and wild-type-query, low-candidate samples are more frequent than expected. Unless otherwise specified, computationally predicted synthetic lethal gene candidates from SLIPT used expression data (exprSL) for both genes (as shown in Figure 3.1) rather than mutation data (mtSL) for the query gene (as shown in Figure 3.2).

The SLIPT methodology is amenable for use on expression data including pathway metagenes (as generated in Section 2.2.3). The suitability of the SLIPT methodology to application on public gene expression data will further be supported by simulation results in Section 3.3 and Chapter 6, including comparison to other statistical methods. SLIPT results for *CDH1* will also compare experimental screen results in a breast cell line (Telford *et al.*, 2015), primary screen results are discussed in Section 4.2 and secondary (validation) screen results are presented in Appendix C.

## 3.2 Synthetic Lethal Simulation and Modelling

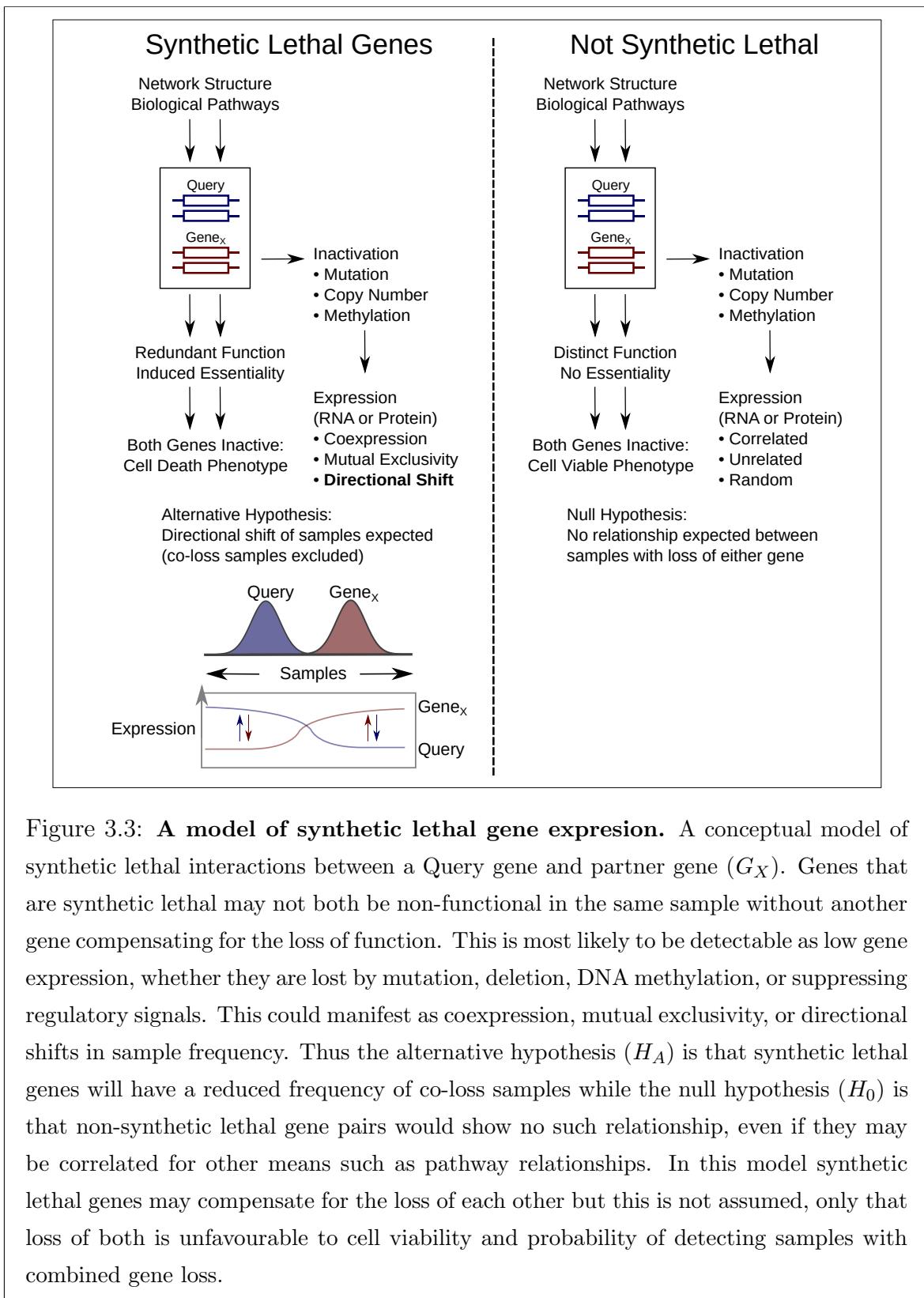
A statistical model of Synthetic Lethality was developed to generate simulated data to test the SLIPT procedure. This section will describe the synthetic lethal model and the simulation procedure for generating gene expression data with known synthetic lethal partners. Some preliminary results to support usage of the SLIPT methodology throughout this thesis will be presented here. The simulation procedure will be applied in more depth in Chapter 6, including in combination with simulations from graph structures.

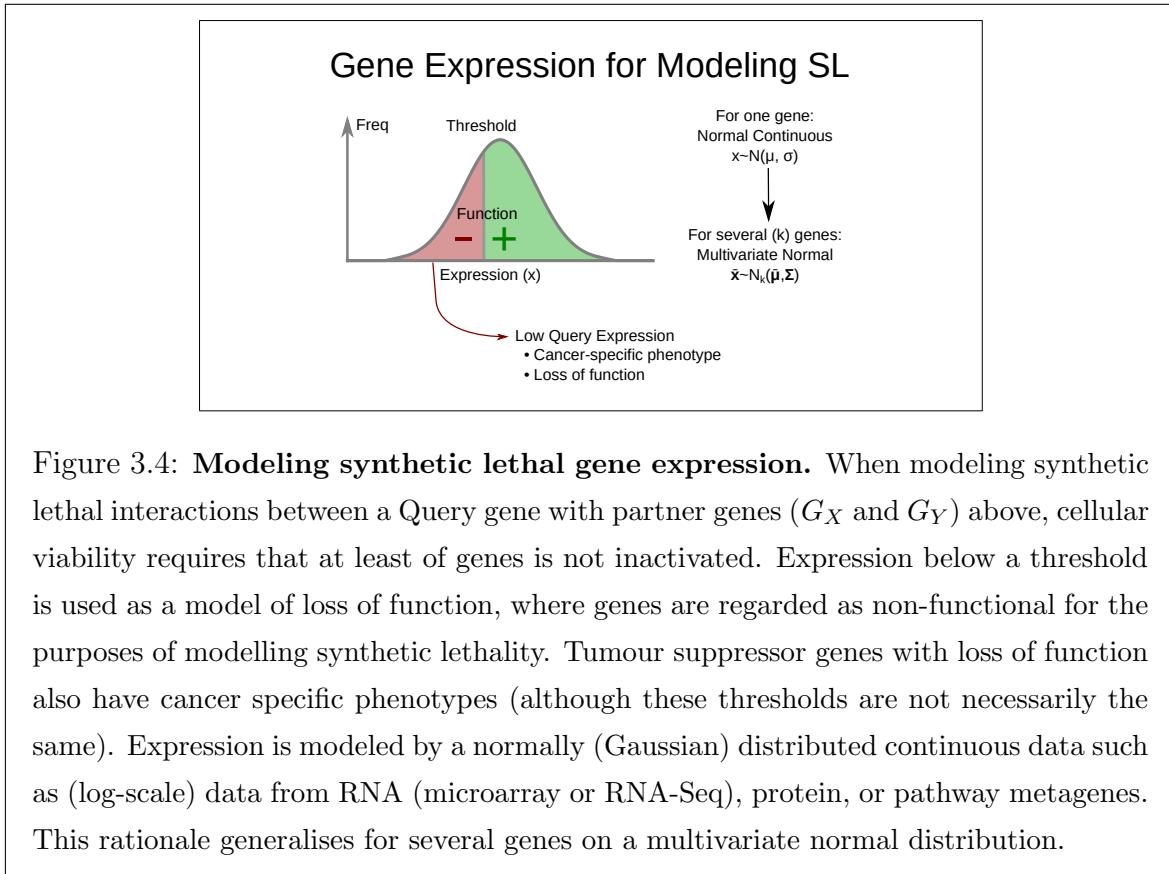
### 3.2.1 A Model of Synthetic Lethality in Expression Data

A conceptual model of synthetic lethality was constructed (see Figure 3.3), which will be used to build a statistical model of synthetic lethal gene expression from which to simulate expression data to on which test SLIPT and various potential synthetic lethal prediction methods. In the model, synthetic lethality arises between genes with related functions as a cell death phenotype when these functions are removed.

This model suggests that synthetic lethality is detectable in measures of gene inactivation across a sample population, namely mutation, DNA copy number, DNA methylation, and suppression of expression. While any of these mechanisms of gene inactivation could lead to synthetic lethality, expression data is readily available and changes in these alternative mechanisms are likely to impact on the amount of expressed (functional) RNA or protein detectable. There are several ways that functional relationships between genes could manifest in expression data, including coexpression, mutual exclusivity and directional shifts. Co-expression is overly simplistic and has previously performed poorly as a predictor of synthetic lethality (Jerby-Arnon *et al.*, 2014), although this will still be tested with correlation measures in later simulations. Here the alternative hypothesis is that synthetic lethality will lead to a detectable directional shift in the number of samples exhibiting low or high expression of either gene. This model does not preclude mutual exclusivity (Wappett *et al.*, 2016), compensating expression or co-loss under-representation (Lu *et al.*, 2015) as previously postulated to occur between synthetic lethal genes.

The first condition of the synthetic lethal model is that if there are only two synthetic lethal genes (e.g., *CDH1* and one SL partner), then they will not both be non-functional in the same sample (in an ideal model). Gene function is thus determined for each sample in a model of synthetic lethal with the proportion of samples with a functional or non-functional gene being arbitrary. Whether a gene is functional can





**Figure 3.4: Modeling synthetic lethal gene expression.** When modeling synthetic lethal interactions between a Query gene with partner genes ( $G_X$  and  $G_Y$ ) above, cellular viability requires that at least of genes is not inactivated. Expression below a threshold is used as a model of loss of function, where genes are regarded as non-functional for the purposes of modelling synthetic lethality. Tumour suppressor genes with loss of function also have cancer specific phenotypes (although these thresholds are not necessarily the same). Expression is modeled by a normally (Gaussian) distributed continuous data such as (log-scale) data from RNA (microarray or RNA-Seq), protein, or pathway metagenes. This rationale generalises for several genes on a multivariate normal distribution.

similarly be modelled by an arbitrary threshold of continuous and normally distributed gene expression data to define gene function (as shown in Figure 3.4). For the purposes of modeling synthetic lethality in breast cancer expression data, a threshold of the 30<sup>th</sup> percentile of the expression levels was used because approximately 30% of samples analysed had *CDH1* inactivation. This was generalised for a model of the proportion of samples inactivated for each gene. In this ideal case, no samples lowly expressing both of these genes are expected to be observed. While this is not observed, that is to be expected as it is unlikely that only 2 genes will have an exclusive synthetic lethal partnership. The threshold of the 0.3 quantile was used in simulations derived from this model throughout this thesis.

A synthetic lethal pair of genes is unlikely to act in isolation, therefore higher-order synthetic lethal interactions (i.e., 3 or more genes) must be considered in the model as shown in Figure 3.5. Even when testing pairwise interactions, modelling higher level interactions that may interfere is important. If there are additional synthetic lethal partners, there are two possibilities for adding these: 1) that they are independent partners of the query genes interacting pairwise (and not with each other) or 2) that

an addition partner gene interacts with both of the synthetic lethal genes already in the system and any of the three (or more) are required to be functional for the cell to survive.

The signal (in terms of gene expression data) will be weaker for this latter case and this model has the more stringent assumption that all synthetic lethal partner genes interact with each other: that only one of these must be expressed to satisfy the model of synthetic lethality. In this model any of the synthetic lethal genes in a higher-order interaction is able to provide the missing function of the others, allowing for higher-level synthetic lethal partners to compensate for loss a synthetic lethal gene pair. While samples expressing low levels of the synthetic lethal gene pairs will be under-represented, they may not be completely absent from the dataset due to these higher-level interactions.

In the example of 3 synthetic lethal genes 3.5, only one of genes involved in the higher-order synthetic lethal interaction is required for cell viability. For synthetic lethal pairs, only a subset of these samples will be inviable (i.e., removed from simulated data), leading to an under-representation.

In practice, samples are not removed from a simulated dataset, rather the expression and function of the query gene is generated across samples separately from the pool of potential partner genes. The query gene data is matched to simulated samples (as shown in Figure 3.7), satisfying the synthetic lethal condition with the procedure described in Section 3.2.2. This is performed to maintain a comparable samples size across simulations and the preserve the assumed (multivariate) normal distribution of the data.

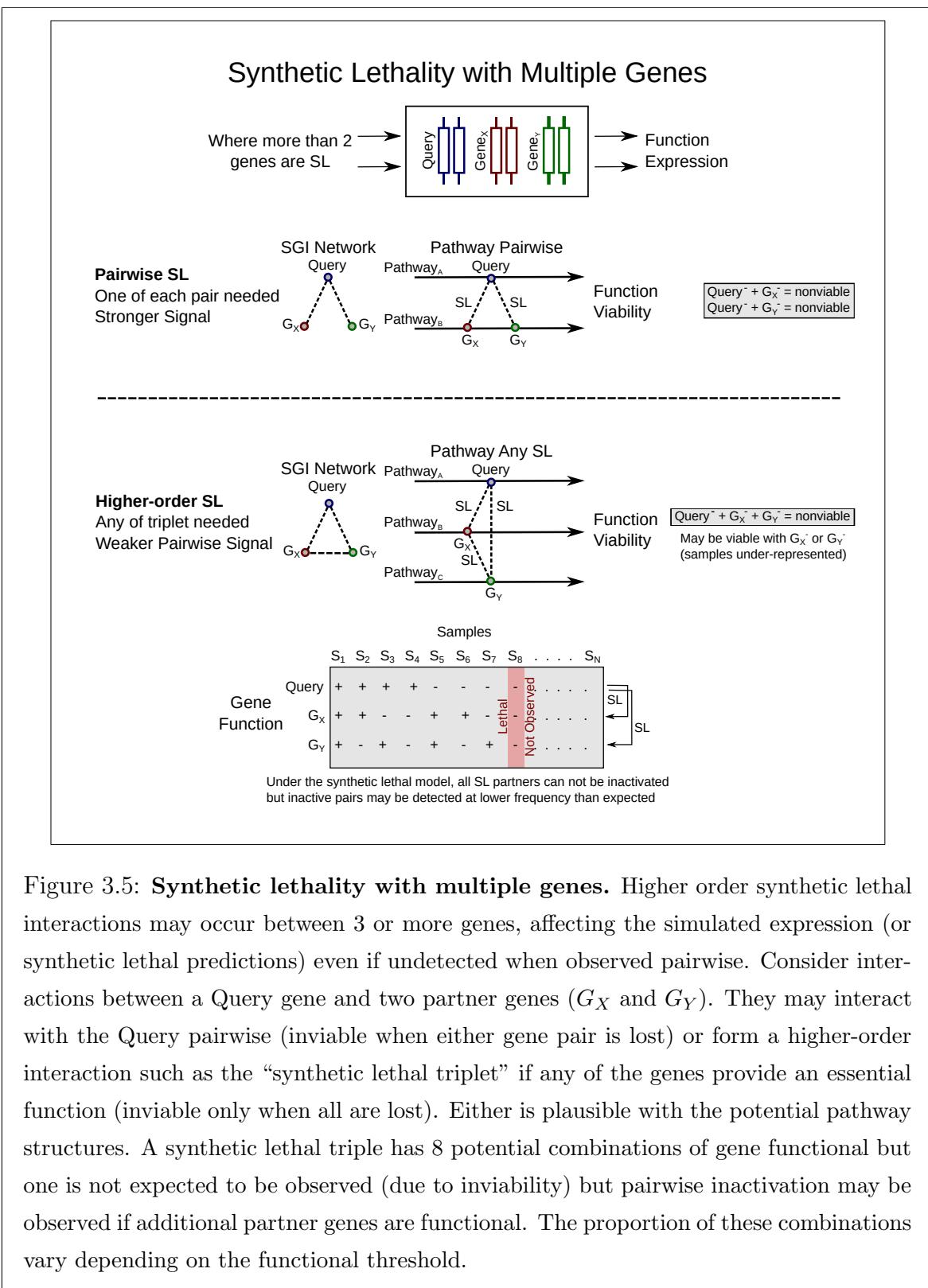


Figure 3.5: **Synthetic lethality with multiple genes.** Higher order synthetic lethal interactions may occur between 3 or more genes, affecting the simulated expression (or synthetic lethal predictions) even if undetected when observed pairwise. Consider interactions between a Query gene and two partner genes ( $G_X$  and  $G_Y$ ). They may interact with the Query pairwise (inviolate when either gene pair is lost) or form a higher-order interaction such as the “synthetic lethal triplet” if any of the genes provide an essential function (inviolate only when all are lost). Either is plausible with the potential pathway structures. A synthetic lethal triple has 8 potential combinations of gene functional but one is not expected to be observed (due to inviability) but pairwise inactivation may be observed if additional partner genes are functional. The proportion of these combinations vary depending on the functional threshold.

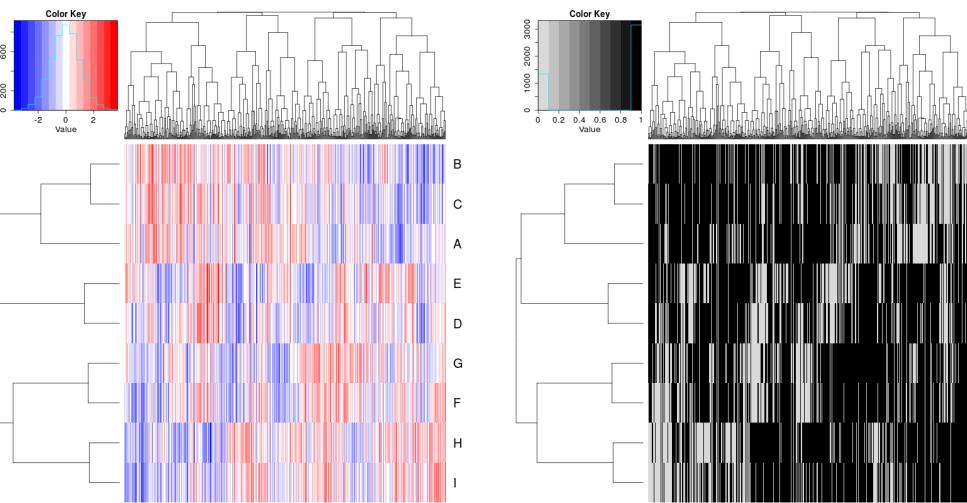
### 3.2.2 Simulation Procedure

Simulations were developed to simulate normal distributions of expression data and define function with a threshold cut-off. This is the reverse to the procedure of SLIPT to predict synthetic lethal partners (although the threshold is assumed to be unknown when testing upon simulated data). While gene function is used as an intermediary step in modelling synthetic lethal genes in expression data, the normal distribution is sampled for simulated data to represent normalised empirical gene expression data for which SLIPT (and other methods) will be applicable.

Sampling a distribution for expression profiles has the added advantage of being amenable to simulating correlation structures with the multivariate normal distribution (using the `mvtnorm` R package (Genz and Bretz, 2009; Genz *et al.*, 2016)). The parameter  $\Sigma$  is a covariance matrix defines the correlation structure between simulated genes being sampled. With a diagonal of one, this  $\Sigma$  matrix simulates genes with a standard deviation of one and the covariance parameters between them are the correlations between each gene. In Figure 3.6, an example of such a simulated multivariate normal dataset is shown with the functional threshold applied.

Once we have generated a simulated dataset, the samples are compared by gene function (as derived from a functional threshold). Known underlying synthetic lethal partners are selected within the dataset and a query gene is generated by sampling from the normal distribution. These are matched (as shown for 2 synthetic lethal partners in Figure 3.7) such that the synthetic lethal condition is met: that at least one of the synthetic partner genes and the query gene are functional in any particular cell. The samples are ordered by functional data (without assuming correlation of underlying expression values) with the query gene in one direction and the remaining dataset ordered by the selected synthetic lethal partner.

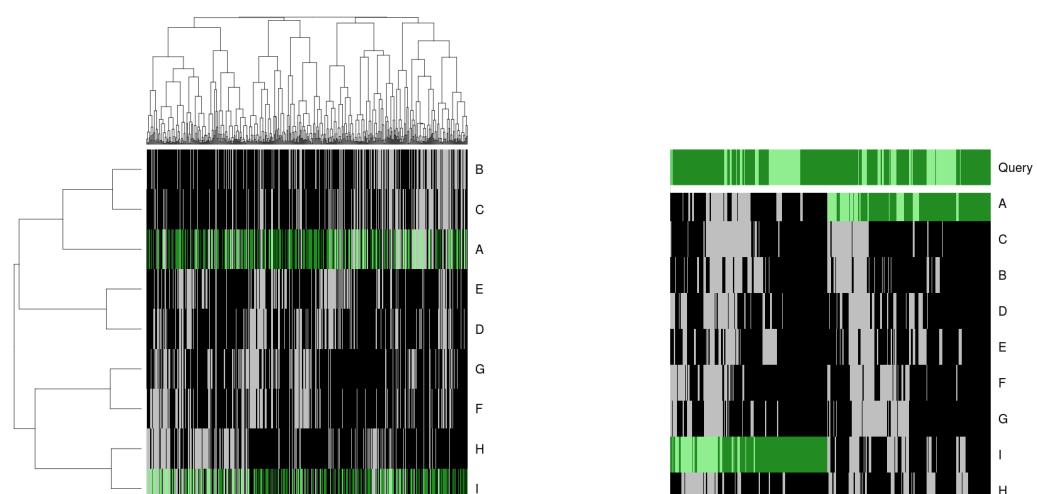
This results a simulated dataset where samples with non-functional query gene have at least one functional partner gene. Similarly, the query gene is functional in all samples where all of the synthetic lethal partner genes are non-functional. Therefore a dataset has been generated with known synthetic lethal partners (see Figure 3.8) by as few assumptions about the relationships between the each synthetic lethal pair as possible (and allowing compensating functions from higher-order interactions). This has been designed to have the most stringent (least detectable) synthetic lethal relationships where higher-order interactions are possible for the purposes of testing pairwise detection procedures such as SLIPT.



(a) Simulated expression matrix

(b) Corresponding gene function calls

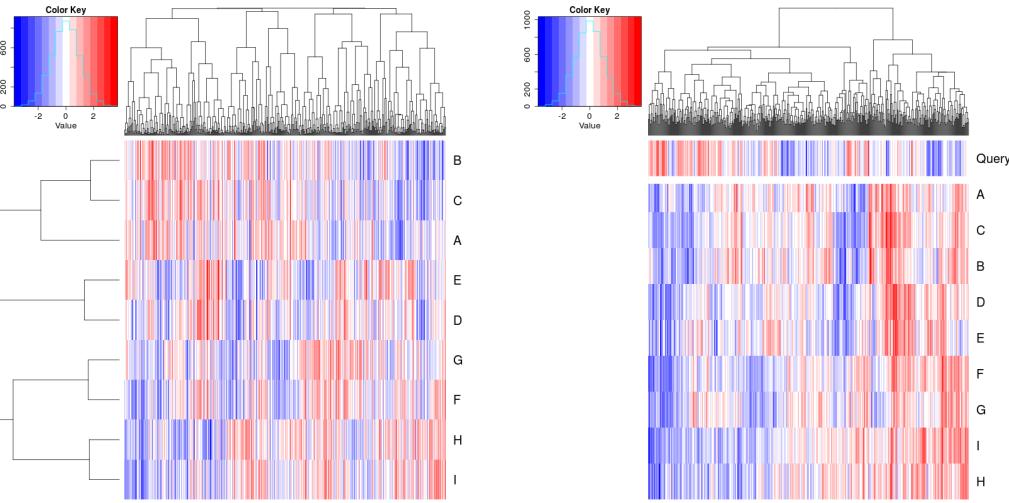
Figure 3.6: **Simulating gene function.** A simulated dataset with samples (columns) and genes A–H (rows) is transformed from a continuous (coloured blue–red) scale to a discrete matrix of gene function (black for functional levels and grey for non-functional).



(a) Simulated gene function with SL genes

(b) Query gene added with SL condition

Figure 3.7: **Simulating synthetic lethal gene function.** In a discrete simulated gene function dataset (shaded for functional levels and pale otherwise) with samples (columns) and genes (rows), genes A and I are SL partners of a “Query” gene. A partner is selected (highlighted in green) randomly in each sample for simulating synthetic lethality, then ordered such that the query gene or an SL partner are functional in each sample.



(a) Initial expression matrix

(b) Simulated synthetic lethal dataset

Figure 3.8: **Simulating synthetic lethal gene expression.** A simulated continuous expression dataset (blue-red scale) with samples (columns) and genes (rows) is matched to a query gene such that at least one synthetic lethal partner is above a functional threshold when the query gene is below it satisfying the synthetic lethal model.

### 3.3 Detecting Simulated Synthetic Lethal Partners

The synthetic lethal detection methodology (SLIPT), as described in Section 3.1, was tested on simulated data with known synthetic lethal partners, generated using the procedure described in Section 3.2.2. This section will present basic simulations to demonstrate the methodology and support its use throughout this thesis. These will be performed with sampling from basic statistical distributions as described, including multivariate normal distribution with correlated blocks of genes, with the  $\Sigma$  matrix shown in the plots where relevant. A more complex multivariate normal sampling procedure based on pathway graph structures, as described in section 3.4.2, will be applied in Chapter 6.

#### 3.3.1 Binomial Simulation of Synthetic lethality

A previous version of the synthetic lethal simulation procedure (described in Section 3.2.2), used gene function sampled directly from a binomial distribution using the binomial probability of observing functional gene levels ( $p = 0.3$ ) in one observa-

tion ( $n = 1$ ) for each samples:

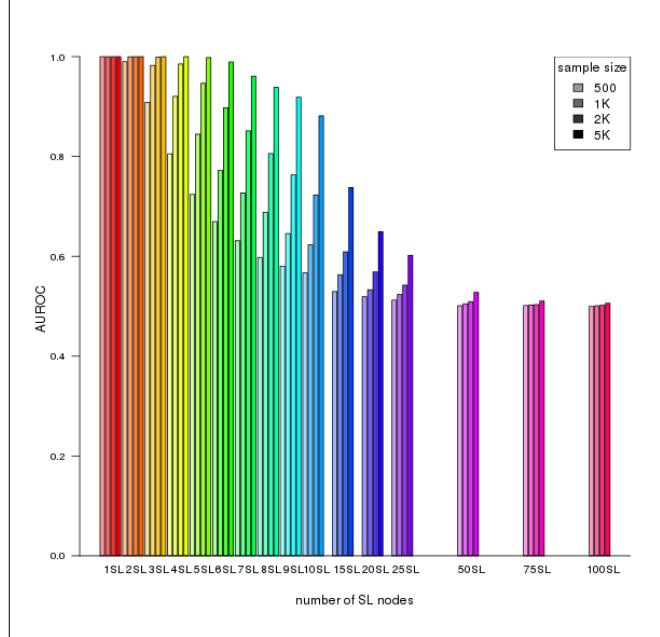
$$X \sim Bin(n, p)$$

Once a query gene consistent with synthetic lethality has been added, these functional levels were passed directly into SLIPT as “low” and “high” categories.

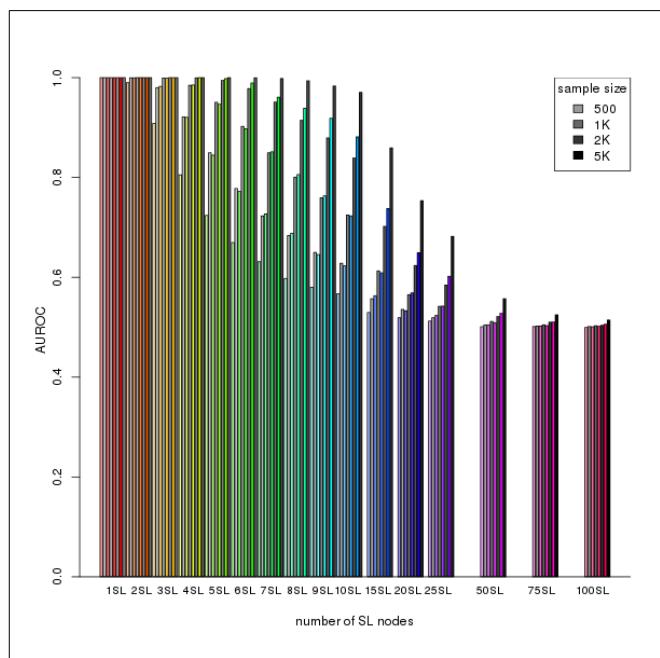
The simulation procedure was performed with 20,000 total genes (as feasible in the human genome and expression datasets) with a variable number of true synthetic lethal partners and sample sizes of 500, 1000, 2000, and 5000. Each ROC curve was derived from the results of 10,000 replicate simulations. The statistical performance (as shown in Figure 3.9) of such an approach based on the  $\chi^2$  p-value declines towards random predictions (an AUROC of 0.5) with an increasing number of underlying true synthetic lethal partners to detect. However, increased sample size mitigates this decline to some extent, as expected with a statistical predictor, particularly for moderate numbers of synthetic lethal partners.

Simulations based on a simple binomial model of synthetic lethality are limited but form a basis for building a more complex model including expression and correlation structures. While this does not represent the data that SLIPT will be applied to, binomial simulations do demonstrate that SLIPT is able to distinguish small numbers of synthetic lethal partners in a simplistic simulated system with behaviour expected with respect to sample size. This supported further development of the synthetic lethal model and simulation pipeline (as described in Section 3.2) using the multivariate normal distribution.

The multivariate normal simulation procedure is more representative of the (normalised) expression data SLIPT is intended for and enables the prediction procedure to be tested without changes to the methodology (presented in more detail in Section 3.3.2). Sampling continuous expression values from a normal distribution allows the expression threshold for gene function to differ from the categorical “low” and “high” expression binning performed by SLIPT (as discussed in Section 3.2.1) which represents that the SLIPT procedure does not assume a known threshold for expression but rather uses expression as an estimate of gene function. This functionality can be included in the multivariate normal simulation without compromising the statistical performance of the SLIPT, rather the performance estimates (shown in Figure 3.10) were a marked improvement over the binomial simulation procedure across simulation parameters in an equivalent simulation (without correlation structure). This improvement may be due to binomial model defining the synthetic lethal condition in a way that, while ensuring at least one synthetic lethal partner is active in query deficient sam-



**Figure 3.9: Performance of binomial simulations.** Gene function was simulated by binomial sampling and tested for synthetic lethal genes. Statistical performance declines with additional known synthetic partners but this is mitigated by increased sample sizes.



**Figure 3.10: Comparison of statistical performance.** Binomial simulation of synthetic lethality (in colour) is compared (in greyscale) to multivariate normal simulations (detailed below) which consistently outperforms binomial simulation across parameters.

ples, disrupts the number of samples with functional synthetic lethal genes compared to other genes affecting the expected sample proportions of  $\chi^2$  test.

### 3.3.2 Multivariate Normal Simulation of Synthetic lethality

The multivariate normal simulation procedure was initially performed using the `mvtnorm` R package (Genz and Bretz, 2009; Genz *et al.*, 2016) (as described in Section 3.2) without correlation structure.

Expression is sampled from multivariate normal distribution with a mean ( $\mu = 0$ ), standard deviation ( $\sigma = 1$ ), and no correlation between genes ( $r = 0$ ):

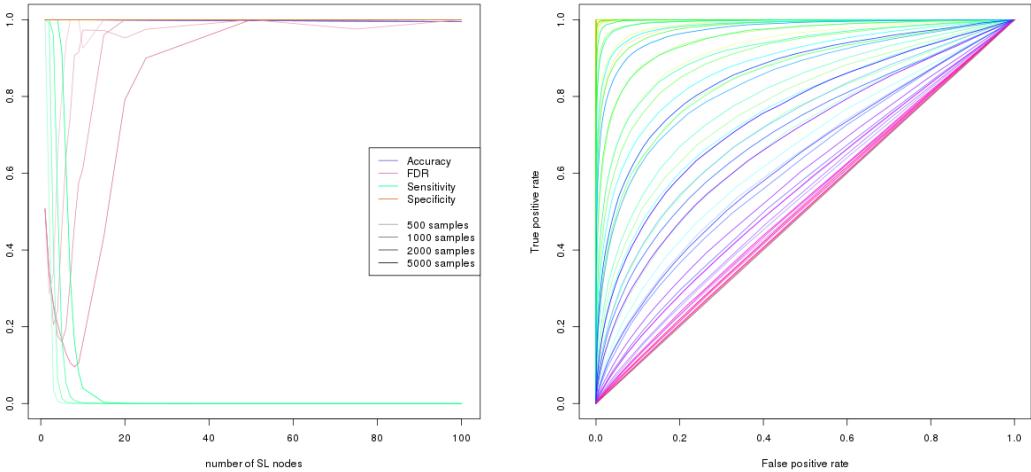
$$X \sim N(\bar{\mu}, \Sigma)$$

Once a query gene consistent with synthetic lethality has been added, the simulated expression values are tested by SLIPT exactly as described in Section 3.1.

As shown in Figure 3.11(a), the statistical accuracy of SLIPT as a binary classifier is considerably high across simulations of a full human dataset of 20,000 genes. However, with the  $\chi^2$  p-value as a threshold for prediction, this is largely to desirable specificity: the majority of non-SL genes are distinguished from the few underlying synthetic lethal genes. In this regard, the SLIPT methodology generally performs better with larger datasets with more expected negatives and thus the results of simulations of smaller numbers of genes (such as the graph structures analysed in Chapter 6) can be applied to larger datasets where they are expected to perform comparably or better with a lower false negative rate. Accordingly, key results will be supported by replication with larger numbers of non-SL genes added to the simulations.

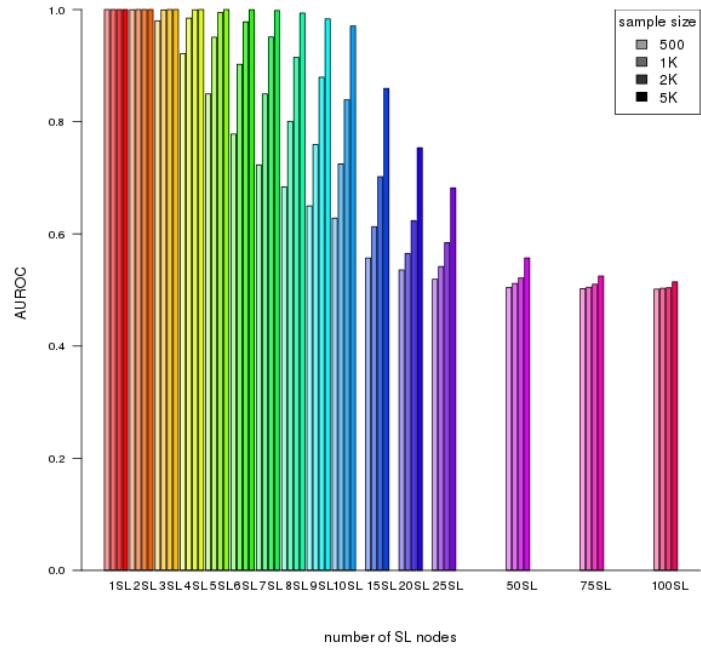
However, with higher numbers of synthetic lethal genes to detect, the sensitivity (in Figure 3.11(a)) of SLIPT as a binary classifier of synthetic lethality declines, although this is somewhat mitigated by higher sample sizes (shown in darker colours). Thus the minority of true synthetic lethal partners are more difficult to distinguish when there are more of them (and a weaker expression signal from each). While a reasonable reduction of the false discovery rate can be achieved for moderate numbers of underlying synthetic lethal partners, we can not be sure how many partners are expected to be detected in analyses of expression data. However this simulation procedure is amenable to assessing the performance of SLIPT across simulation parameters, graph structures and comparisons to other approaches (presented in more detail in Chapter 6).

Not all of the genes detected by SLIPT will be true synthetic lethals but these will be among the strongest candidates and it performs better with fewer underlying synthetic lethals to detect. This supports a focus on pathway analyses, in particular detecting pathways for further investigation. Since individually gene candidates are not necessarily gene synthetic lethal themselves, pathway over-representation analysis



(a) Statistical evaluation

(b) Receiver operating characteristic



(c) Statistical performance

**Figure 3.11: Performance of multivariate normal simulations.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (without correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (in darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

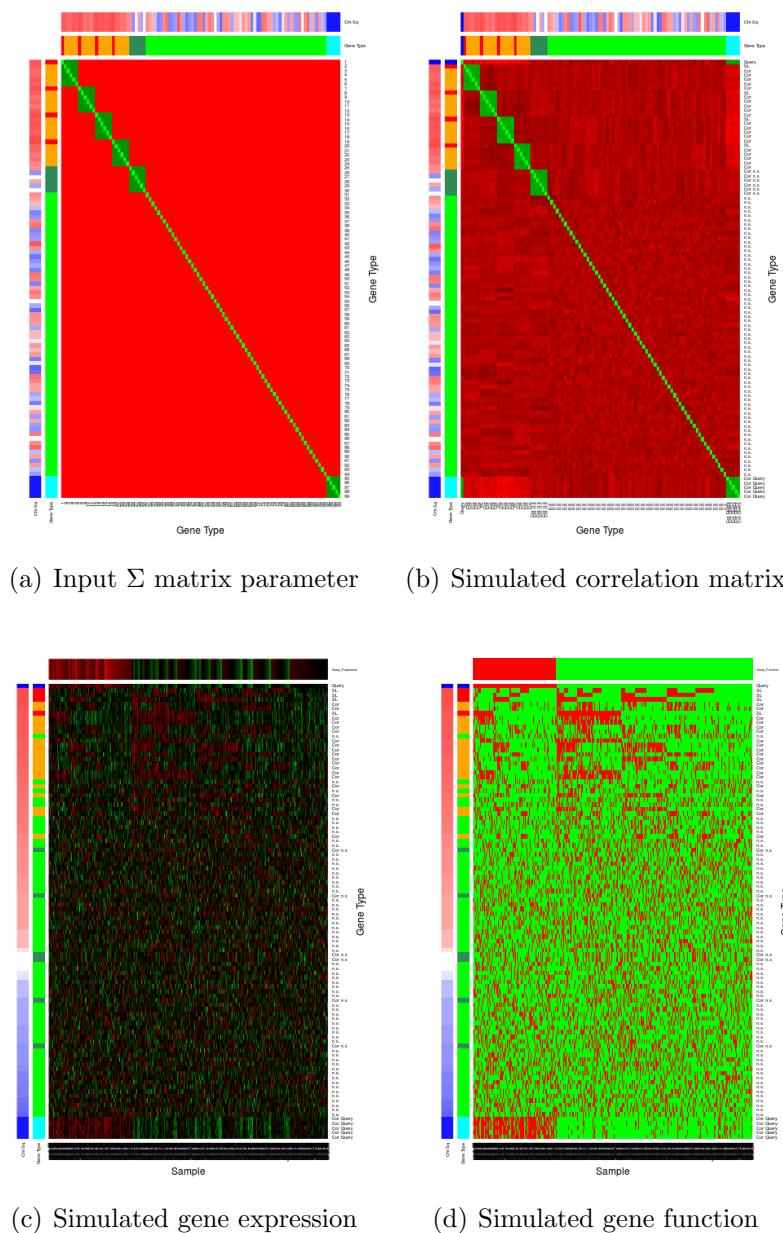
will be performed to detect functional groups recurrently detected by SLIPT as these detection of functionally related genes further support their role in synthetic lethal relationships in addition to being biologically informative. Alternatively, pathway metagenes will reduce the number of underlying synthetic lethals to identify synthetic lethal pathways. Both of these approaches will be applied in Chapter 4 to identify and replicate synthetic pathways of *CDH1*. Pathways are also more likely to replicate across experimental models as demonstrated by Dixon *et al.* (2008).

The receiver operating characteristic curves (in Figure 3.11(b)) demonstrate that SLIPT is subject to near equal trade-off between sensitivity and specificity across threshold values. The lower sensitivity and higher specificity with a binary classification (in Figure 3.11(a)) stems from stringent testing by SLIPT with (FDR) p-values adjusted for multiple tests. The area under these curves is also used to compare statistical performance (in Figure 3.11(c)), with declining performance across increased underlying synthetic lethal partners and increased performance with sample size in multivariate normal simulations.

### 3.3.2.1 Multivariate Normal Simulation with Correlated Genes

Correlation structures can be added to the simulation procedure (as discussed in Section 3.2), starting with simple correlated blocks of genes as the  $\Sigma$  parameter depicted in Figure 3.12(a). These correlated blocks represent genes with correlated expression such as that expected by coregulation or biological pathways. Figure 3.12 gives an example of 4 synthetic lethal genes (out of 100), each with 5 correlated genes that are not themselves synthetic lethal partners of the query gene. This serves to test whether synthetic lethal genes are distinguishable from correlated partners. This  $\Sigma$  matrix produces a similar correlation structure (Figure 3.12(b)) in the resulting expression profiles (Figure 3.12(c)) where apart from correlated blocks of genes ( $r = 0.8$ ), the remaining genes have only slight variations due to random sampling. The structure of the dataset, particularly between synthetic lethal genes and the query, is shown at the gene expression (Figure 3.12(c)) and function (Figure 3.12(d)). These are ordered by the SLIPT results and the synthetic lethal genes are ranked high, with the majority of them being distinguishable from highly correlated genes.

The use of correlation structures generalises to larger datasets, such as 1000 genes shown in Figure 3.13. Synthetic lethal genes are highly ranked by SLIPT and still largely distinguishable from correlated genes. As previously discussed in Section 3.3.2, these synthetic lethal genes are still detectable among a larger number of true negatives and the SLIPT methodology performs better on such datasets.



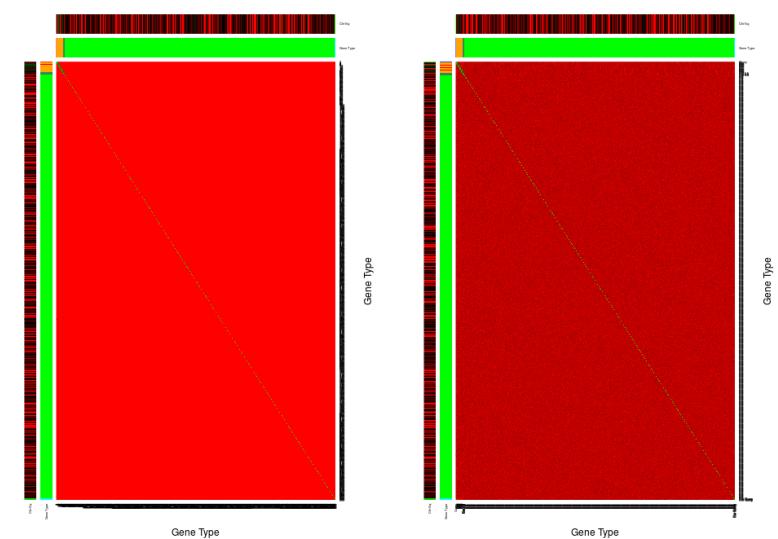
**Figure 3.12: Simulating expression with correlated gene blocks.** A  $\Sigma$  matrix (a) is used to generate a multivariate normal distribution with 100 genes in correlated blocks of genes (correlated by 0.8) with a comparable structure (b) to the input  $\Sigma$ , as shown by correlation on a red–green scale. The annotation bars for genes give the  $\chi^2$  (in blue if the direction of SLIPT is met or red otherwise) and the gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL). The simulated gene expression (c) and function (d) generated are ordered by  $\chi^2$  showing the functional structure of synthetic lethal genes and that they are among the strongest SLIPT results.

These plots (Figures 3.12 and 3.13) also show similar correlated blocks with a non-synthetic lethal gene (true negative) and the query gene (which is not synthetic lethal with itself). Neither of these should be synthetic lethal (or detected to be) but they may impact upon the performance of the model, particularly the specificity as correlated negative genes may be distinguishable from true synthetic lethals. The non-synthetic lethal correlated block has no impact on synthetic lethal detection but the impact of query correlated genes will be discussed in Section 3.3.2.2 and Chapter 6.

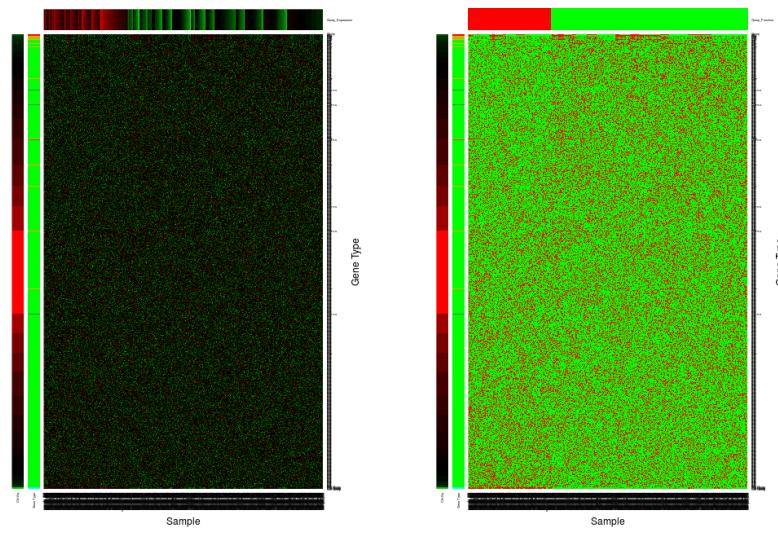
These simulations (on 100 genes) were repeated to examine the variation between detection on different samples and varying the number of underlying synthetic lethal partners, in simulated gene expression data with correlations structure. A small number (10 for each) simulations are shown in Figure 3.14 to demonstrate the variation between replicate simulations, with iterative sampling from the same multivariate normal distribution. These simulations show synthetic lethal genes are not only highly ranked by SLIPT when there are few of them but also that they are fairly consistent across replicate simulations. Whereas they become less consistent for increasing numbers of true synthetic lethal partners to detect and thus more difficult to distinguish from other genes, particularly those correlated with them. Similarly, the  $\chi^2$  values show a marked stepwise increase with clear thresholds for SL and correlated genes in simple simulations, whereas these become less evident for higher numbers of SL partners.

Whether the synthetic lethal genes detected in simple simulations (in Figure 3.14) are robustly detectable across greater number of simulations, in addition to further comparisons, was tested with a supporting ROC analysis. These results (in Figure 3.15) are very similar to simulations without correlation structure, with SLIPT as a binary classifier having a poor sensitivity with increasing numbers of synthetic lethal partners to detect but high specificity in a total of 20,000 genes with the vast majority being true negatives. This is reflected in a similar decline in statistical performance for increasing numbers of synthetic lethal partners and a compensating increase in performance with higher sample size. Overall, the statistical performance is very similar to simulations without correlation structure (as shown in Figure 3.16).

Thus SLIPT is robust across correlation structures and applicable to real gene expression data where pathway structures and correlations are a consideration. These correlation structures are not intended to model specific biological pathways or represent them, rather they serve to test the impact of correlation structure on the performance of SLIPT with an extreme example of closely correlated ( $r = 0.8$ ) gene blocks. More complex correlation structures, such as genes positively correlated with the query

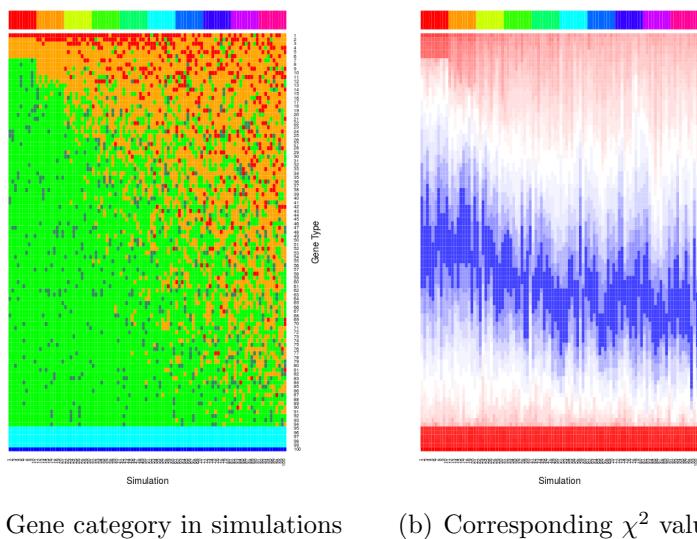


(a) Input  $\Sigma$  matrix parameter      (b) Simulated correlation matrix



(c) Simulated gene expression      (d) Simulated gene function

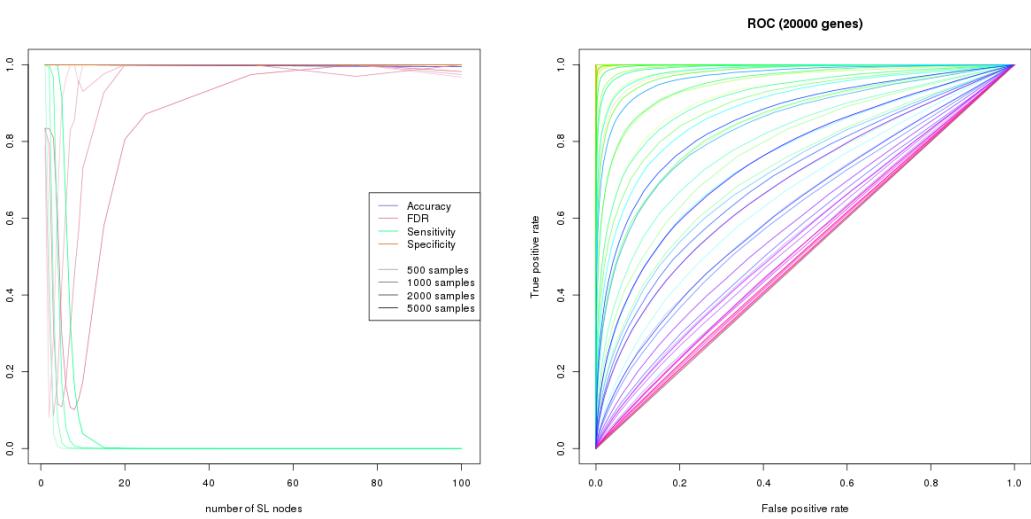
**Figure 3.13: Simulating expression with correlated gene blocks.** Using the (a)  $\Sigma$  matrix, sampling from a multivariate normal distribution with of 1000 genes produced (b) correlated blocks of genes (correlated by 0.8) on a red-green scale. The simulated gene expression (c) and function (d) generated are ordered by  $\chi^2$  and SLIPT direction show that synthetic lethal genes are among the strongest SLIPT results with high specificity against many potential false positives. These are annotated for  $\chi^2$  (on a red-green scale) and category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) for each gene.



**Figure 3.14: Synthetic lethal prediction across simulations.** The gene category (blue for query, cyan for query-correlated, red for SL, orange for SL-correlated, forest green for non-SL-correlated, and green for non-SL) ordered by  $\chi^2$  signed by the SLIPT directional condition is shown across simulations. For each of 1–10 SL partners, 10 simulations demonstrate that the increasing numbers of SL partners become harder detect. The  $\chi^2$  values show a clear threshold for SL and correlated genes when there are fewer of them, distinguishable from correlated genes in this case.

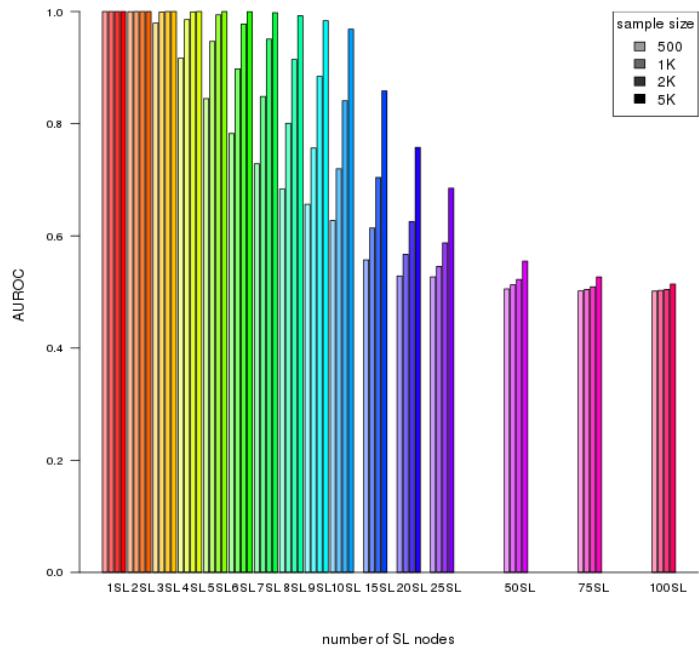
gene and derived from pathway graph structures (as described in 3.4.2) will be examined below (in Section 3.3.2.2) and in Chapter 6 respectively.

In particular, genes correlated with true synthetic lethal genes have little impact on the performance of SLIPT detection: synthetic lethal genes are as distinguishable from true negative genes as without correlated genes. Synthetic lethal correlated genes will not interfere detect of true synthetic lethals, although they may be ranked next below them and be biologically informative with related gene functions.



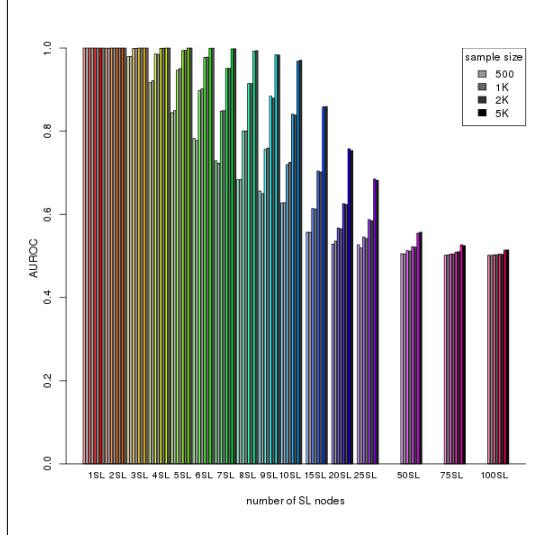
(a) Statistical evaluation

(b) Receiver operating characteristic

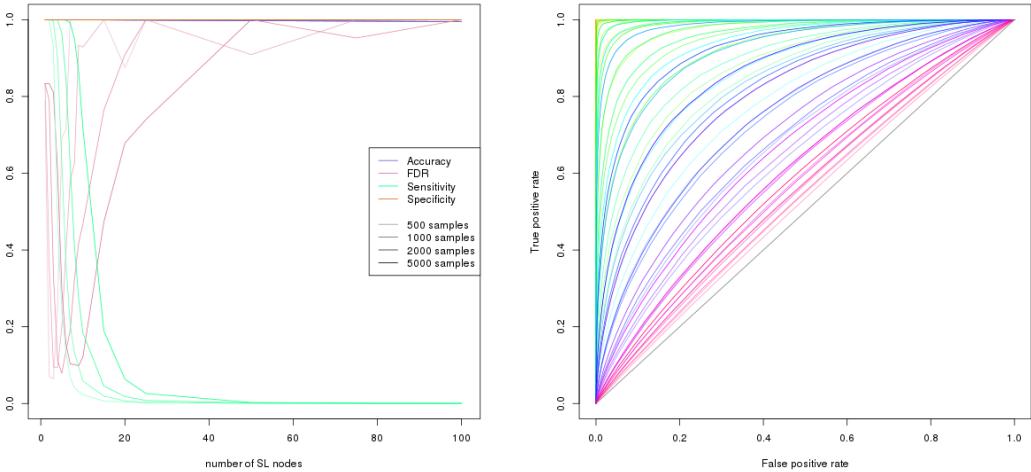


(c) Statistical performance

**Figure 3.15: Performance with correlations.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (with correlation structure). Performance of SLIPT declines for more synthetic partners but this is mitigated by increased sample sizes (darker colours). This generally occurs as the sensitivity decreases for a greater number of true positives to detect, leading to a trade off in accuracy as seen in a trough for false discovery rate and the ROC curves.

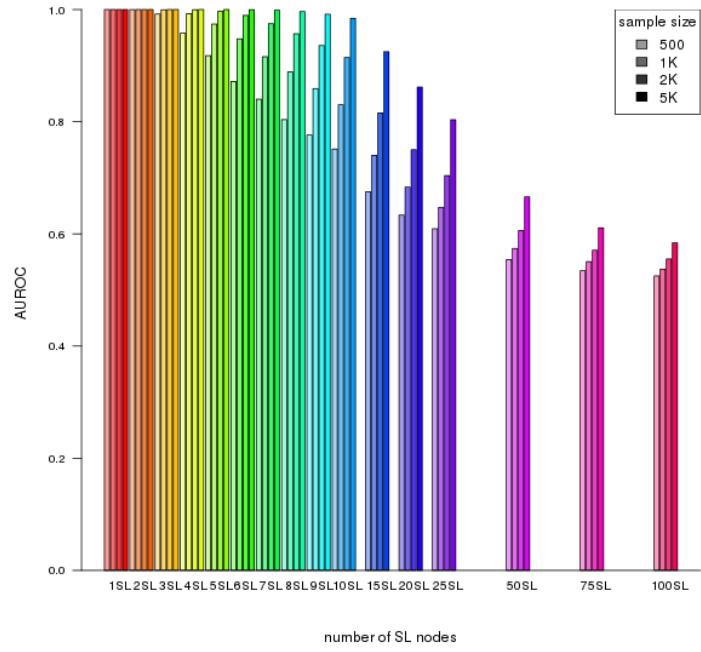


**Figure 3.16: Comparison of statistical performance with correlation structure.**  
 Multivariate simulation of synthetic lethality with correlation structure (in colour) has comparable performance to simulation without correlations (in greyscale) with known synthetic partners across parameters.



(a) Statistical evaluation

(b) Receiver operating characteristic



(c) Statistical performance

**Figure 3.17: Performance with query correlations.** Simulation of synthetic lethality was performed sampling from a multivariate normal distribution (with correlation structure including correlated genes with non-SL and query genes). As before, performance of SLIPT declines for more synthetic partners and is mitigated by increased sample sizes (darker colours)but the sensitivity remains higher for a greater number of true positives with corresponding improvements in ROC curves.

### 3.3.2.2 Specificity with Query-Correlated Pathways

Another consideration for correlation structures is positively correlated genes with the query that are not synthetic lethal. As described in Section 3.3.2.1, 5 highly correlated ( $r = 0.8$ ) with the query gene were added. These simulations perform similarly to before (in Figure 3.17) with a higher specificity and a lower false discovery rate being feasible (as shown in 3.17(a)).

#### 3.3.2.2.1 Importance of Directional Testing

It is important to notice here that the directional criteria of the SLIPT procedure is enhancing its performance, particularly in distinguishing positively correlated true negatives. The multivariate normal simulation results, with 20,000 genes including all of the correlation structures discussed (SL, non-SL, and query correlated genes), are compared here for SLIPT with and without ( $\chi^2$ ) directional testing. There is a marked improvement in statistical performance with directional criteria, particularly with increased sensitivity and lower false discovery rate (as shown in Figure 3.18).

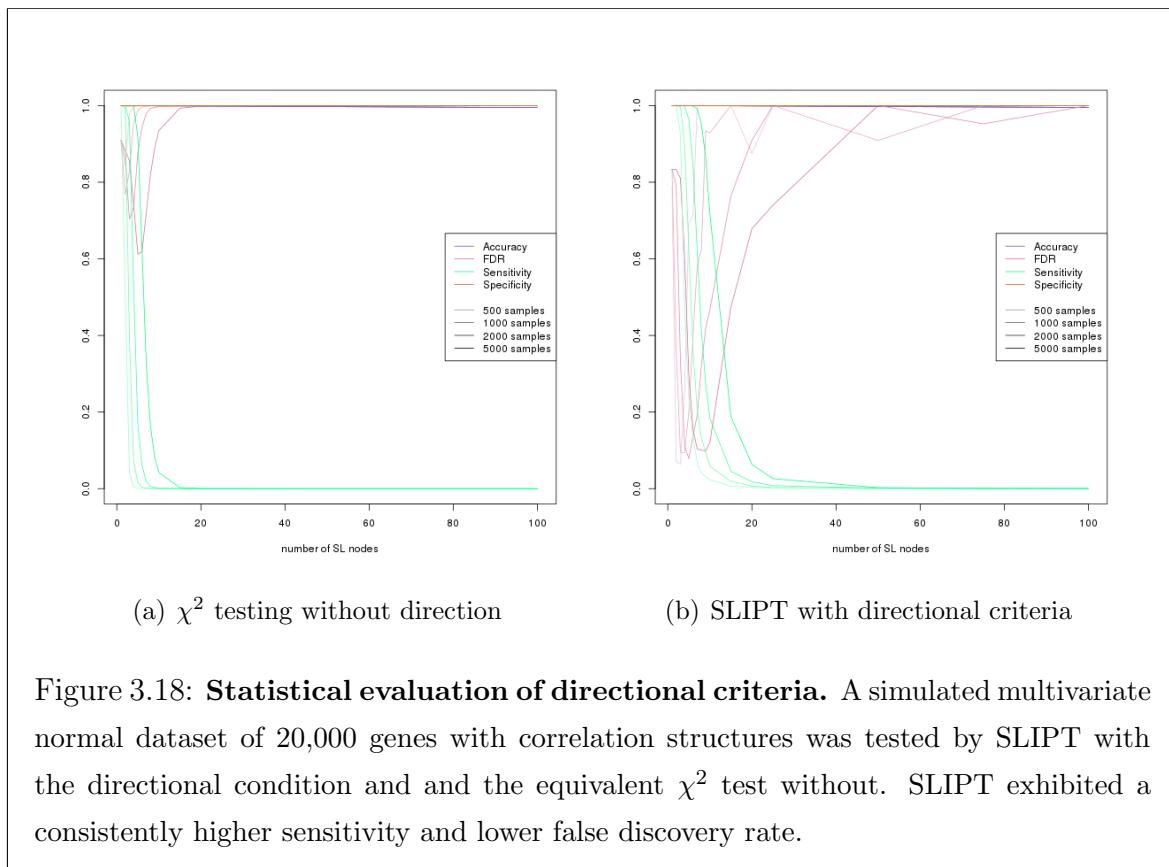
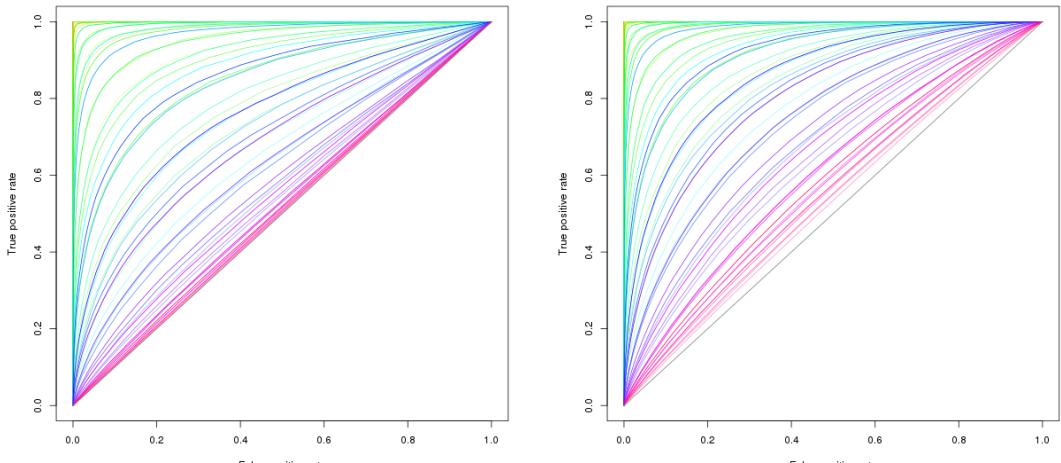
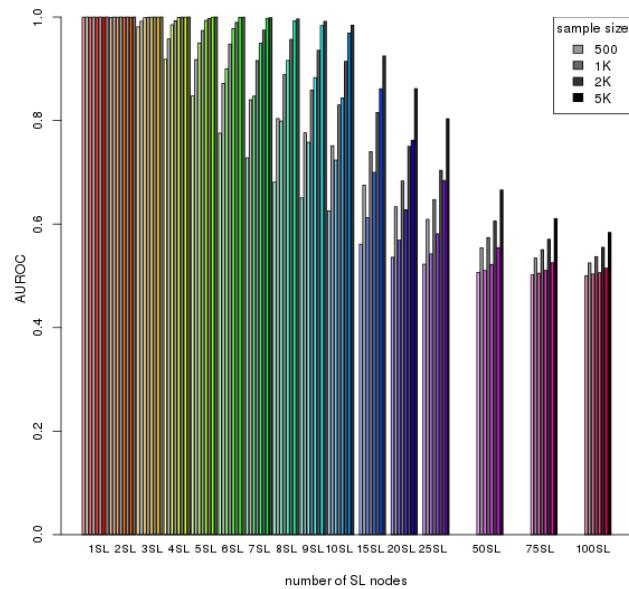


Figure 3.18: **Statistical evaluation of directional criteria.** A simulated multivariate normal dataset of 20,000 genes with correlation structures was tested by SLIPT with the directional condition and the equivalent  $\chi^2$  test without. SLIPT exhibited a consistently higher sensitivity and lower false discovery rate.



(a)  $\chi^2$  testing without direction

(b) SLIPT with directional criteria



(c) Statistical performance

Figure 3.19: **Performance with directional criteria.** A simulated multivariate normal dataset of 20,000 genes with correlation structures was tested by SLIPT with the directional condition and the equivalent  $\chi^2$  test without. SLIPT has higher performance across simulation parameters, clearly differing from random (grey diagonal) in ROC curves up to 100 SL genes (b). The performance (c) of SLIPT (in greyscale) was consistently higher than the  $\chi^2$  test (in color).

This is encouraging for the application of SLIPT to empirical expression datasets as positively correlated genes are likely to occur and the directional condition robustly improves the performance of SLIPT across simulation parameters. Without assuming the underlying number of synthetic lethal genes, SLIPT will perform better than the  $\chi^2$  test alone at detecting them. This is further supported irrespective of significance threshold for the  $\chi^2$  test by the ROC analysis in Figure 3.19. The directional SLIPT methodology outperforms the ordinary  $\chi^2$  test at detecting synthetic lethal partners with some predictive power (above random and AUROC of 0.5) even up to 100 synthetic lethal genes.

Together these simulation results support the application of the SLIPT methodology as it has been performed throughout Chapter 4 and 5. However, the methodology and simulation procedure will explored in more detail in Chapter 6, with the inclusion of graph structures and comparison to other synthetic lethal detection approaches.

## 3.4 Graph Structure Methods

Graph structures have been used in several ways in this project with novel approaches to analysis and simulations. Procedures were developed for statistical and network analysis of gene states in pathway structures. Specifically, the relationships between siRNA and SLIPT genes were tested within biological pathways in Chapter 5. These graph structures were also used in Chapter 6 for the simulation of synthetic lethality to derive correlation structure between simulated gene expression profiles in manner that resembles biological pathways.

### 3.4.1 Upstream and Downstream Gene Detection

Comparison of experimental and computational candidate synthetic lethal partner genes within pathway structures arose from the hypothesis that these sets of genes were related by pathway structure. Due to differences in how these candidates were generated, it should not be expected that they detect the identical genes within the candidate biological pathways, rather they may be related by being upstream or downstream of each other.

Using the Reactome version 52 data (Croft *et al.*, 2014) as described in Section 2.4.2, genes identified by each synthetic lethal discovery approach were mapped to the graph structure for the candidate pathways identified in Chapter 4 (with subgraphs defined as described in Section 2.4.3). To test whether siRNA candidate genes were upstream of SLIPT candidate genes, shortest paths were traced between each potential pair of

these genes in a directed network. The number of genes where the siRNA candidate was upstream were scored “up” and where the siRNA candidate was downstream were scored “down”. This procedure enabled counting the total number of shortest paths which supported siRNA genes being upstream or downstream of the SLIPT genes and measuring the difference between these to determine if there is an imbalance in a particular direction. While this difference is indicative of the number of paths between the gene candidate groups in either direction, alone it is not sufficient to statistically support structure or relationships between siRNA and SLIPT genes. However, it may be combined with a permutation resampling procedure (as described in Section 3.4.1.1) to test for directional relationships in either direction.

The original version of this procedure excluded gene detected by both approaches since they would count in both directions. Upon further consideration, the intersection genes were restored to being accounted for by the shortest paths counts since they may count unequally to being upstream or downstream of each gene set if there are unequal numbers above or below them in the pathway structure.

### 3.4.1.1 Permutation Analysis for Statistical Significance

A permutation procedure was developed to randomly assign members of the pathway to siRNA and/or SLIPT groups, with the same number of each candidate partner gene set as observed in the pathway. These permuted genes are measured for pathway structure between the permuted gene groups as performed for the observed candidates (as performed in Section 3.4.1). A distribution of pathway structure relationships expected by chance is generated by permuting iteratively over these pathways. This null distribution can be compared to the observed counts of relationships (in either direction), which yields a permutation p-value as the proportion of permutations in which had value or greater or more extreme magnitude than the observed value.

The null hypothesis is that there is no relationship between these gene groups that would not have occurred had the genes been selected at random. Thus we can test both the alternate hypothesis that the siRNA genes were upstream of the SLIPT genes or that they are downstream of them.

The permutation procedure does not assume the underlying distribution of the data under the null hypothesis and accounts for the total number of nodes, edges, siRNA, and SLIPT genes in each pathway network structure. The intersection size of the siRNA and SLIPT genes was originally not accounted for under the shortest path counts procedure that excluded them. A refined version of this procedure ensured that

the number of intersecting genes was equal to the number observed to test for pathway structure without changing the intersection size, the subject of prior analyses.

### 3.4.1.2 Ranking Based on Biological Context

An alternative approach to pathway structure was performed based on the biological context that genes at the upstream and downstream ends of a pathway perform different functions, such as a kinase signalling cascade receiving signals from external stimuli and passes these on ribosomes or the nucleus. The genes were ranked to determine if genes of either candidate group (or those with stronger support for either group) performed upstream or downstream functions disproportionately.

A network-based approach was used to determine the pathway ranking of genes in a computationally rational way when applied to different biological pathways with a directed graph structure,  $G$  (without loops). The diameter of the network (i.e., the length of longest possible shortest path between the most distant genes) was used to identify a gene ( $z$ ) at the downstream end of the pathway (at the end of a diameter spanning shortest path), assigned a rank of:

$$\text{rank}(z) = 1 + \text{diameter}(G)$$

Having identified the downstream end of the pathway, genes upstream (e.g., gene  $i$ ) of this are assigned a rank by the length of their shortest path to this gene,  $z$ .

$$\text{rank}(i) = \text{rank}(z) - d_{iz}$$

The remaining unassigned genes (e.g., gene  $j$ ) gain the rank of the length of the shortest path downstream from the nearest assigned gene if possible.

$$\text{rank}(j) = \text{rank}(i) + d_{ij}$$

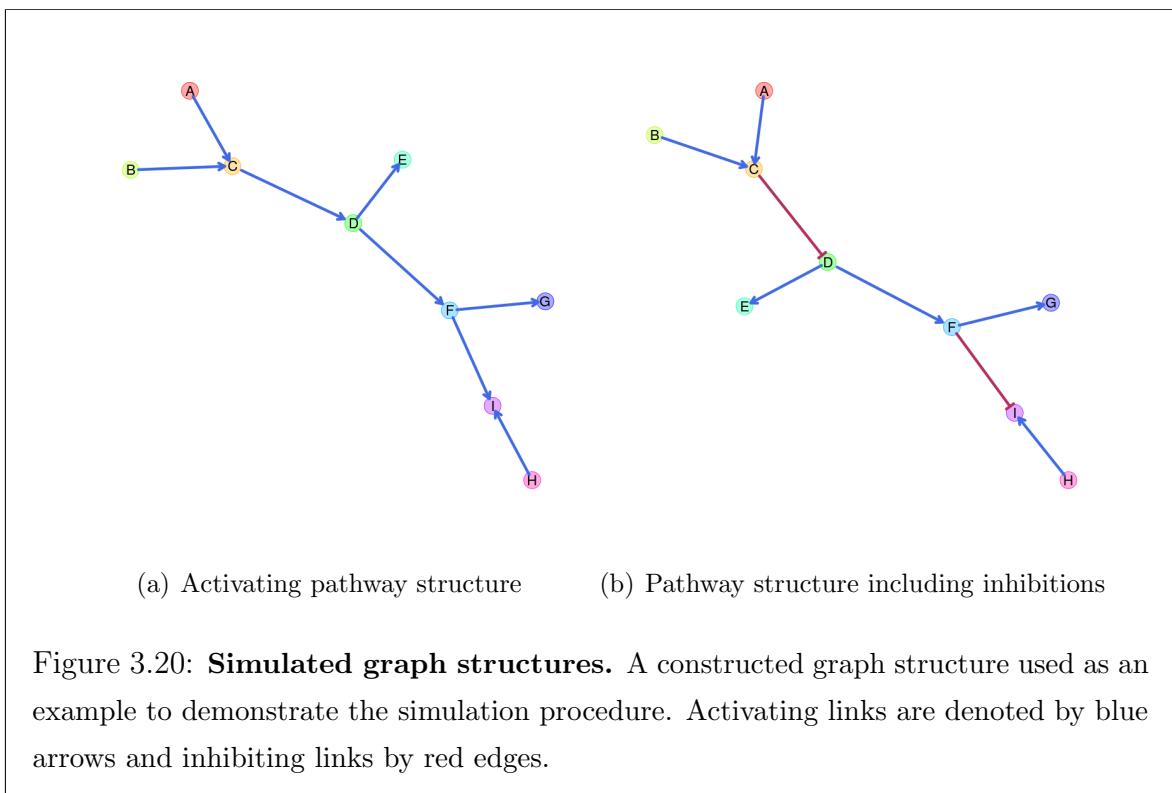
This process may be performed iteratively to fill in pathway ranking but it was not necessary to perform further iterations for the candidate synthetic lethal pathways investigated (amenable to this procedure) which exhibited clear directional structure and the small world property (with a low diameter). Thus genes in a pathway graph structure were assigned integer valued rankings upstream to downstream by this procedure:

$$\text{rank} \in \{1, 2, 3, \dots, 1 + \text{diameter}(G)\}$$

This ranking of pathway directionality can be used for comparison with measures of the number of genes of each candidate group and the support for being synthetic lethal partners with either approach.

### 3.4.2 Simulating Gene Expression from Graph Structures

A further refinement of the simulation procedure generated expression data with correlation structure, derived from a known graph structure. This enables modelling of synthetic lethal partners within a biological pathway and the investigation of impact of pathway structure on synthetic lethal prediction. A simulated pathway is first constructed as a graph structure, with the `igraph` R package Csardi and Nepusz (2006), with the added annotation of the state of the edges (i.e, whether they activate or inhibit downstream pathway members). This simulation procedure was intended for biological pathway members with correlated gene expression (higher than the background of genes in other pathways) but it may also be applicable to modelling protein levels (in a kinase regulation cascade) or substrates and products (in a metabolic pathway).



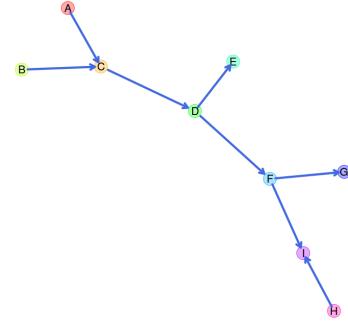
First, the graph structure is constructed for simulated data to be generated from (by sampling from a multivariate normal distribution using the `mvtnorm` R package (Genz and Bretz, 2009; Genz *et al.*, 2016)). Throughout this section, the simulation procedure will be demonstrated with the relatively simple constructed graph structure shown in Figure 3.20. This graph structure visualisation was specifically developed for (directed) iGraph objects in R and has been released in the `plot.igraph` package and

`igraph.extensions` library (see Table 2.6 and Section 3.5.3). The `plot_directed` function allows customisation of plot parameters for each node or edge and mixed (directed) edge types for indicating activation or inhibition. These inhibition links (which often occur in biological pathways) are demonstrated in Figure 3.20(b).

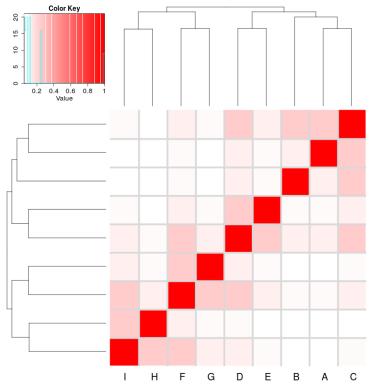
The simulation procedure is designed to use such graph structures to inform development of a “Sigma” variance-covariance matrix ( $\Sigma$ ) for sampling from a multivariate normal distribution (using the `mvtnorm` R package). Given a graph structure (or adjacency matrix), such as Figure 3.21(a), a relation matrix is calculated based on distance such that nearer nodes are given higher weight than farther nodes. For the purposes of this thesis a geometrically decreasing (relative) distance weighting is used, with each more distant node being related by  $1/2$  compared to the next nearest as shown in Figure 3.21(b). However, an arithmetically decreasing (absolute) distance weighting is also available in the `graphsim` R package release of this procedure.

A  $\Sigma$  matrix is derived from this distance weighting matrix, creating a matrix (with a diagonal of 1) where each node has a variance and standard deviation of 1. Thus covariances between adjacent nodes are assigned by a correlation parameter and the remaining matrix based on weighting these correlations with by the distance matrix (or the nearest “positive definite” matrix). For the purposes of this thesis, the correlation parameter is 0.8 unless otherwise specified (as used for the example in Figure 3.21(c)). This  $\Sigma$  matrix is used to sample from a multivariate normal distribution with each gene having a mean of 0, standard deviation 1, and covariance within the range [0, 1] such that they are correlations. This procedure generates a simulated (continuous normally distributed) expression profile for each node (as shown in Figure 3.21(e)) with corresponding correlation structure (Figure 3.21(d)). The simulated correlation structure closely resembles the expected correlation structure (Sigma in 3.21(c)) even for the relatively modest sample size ( $N = 100$ ) illustrated in 3.21. Once a simulated gene expression dataset has been generated (as in Figure 3.21(e)), then a discrete matrix of gene function can be constructed with a functional threshold quantile to simulate functional relationships of synthetic lethality (as shown in Figure 3.4). For the purposes of this thesis, this threshold is the 0.3 quantile (as discussed in Section 3.2.1) which generates functional discrete matrices such as those used for synthetic lethal simulation in Section 3.2.2 (as shown Figure 3.21(f))

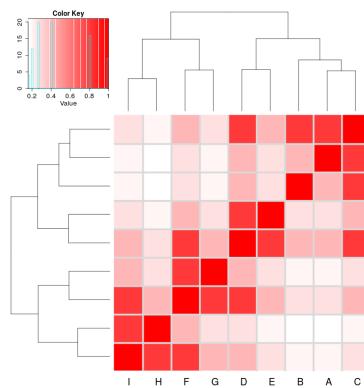
The simulation procedure (depicted in Figure 3.21) is amenable to pathways containing inhibition links (as shown in Figure 3.22) with several refinements. With the inhibition links (as shown in Figure 3.22(a)), distances are calculated in the same



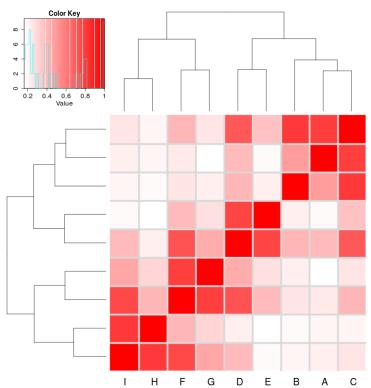
(a) Activating pathway structure



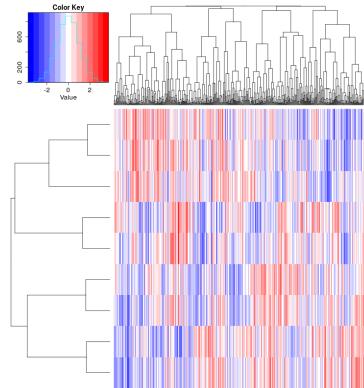
(b) Distance matrix



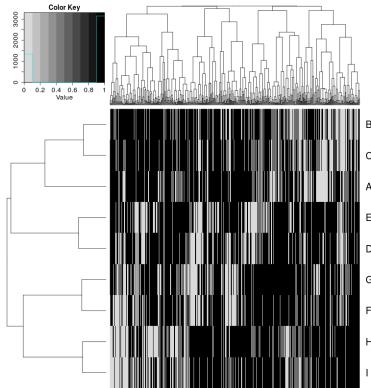
(c) Sigma,  $\Sigma$  (expected correlation)



(d) Simulated correlation structure

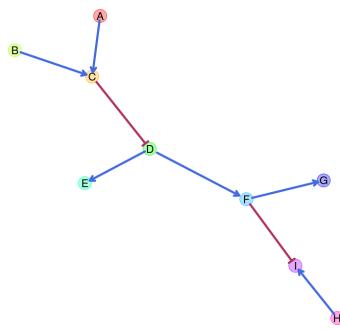


(e) Simulated expression data

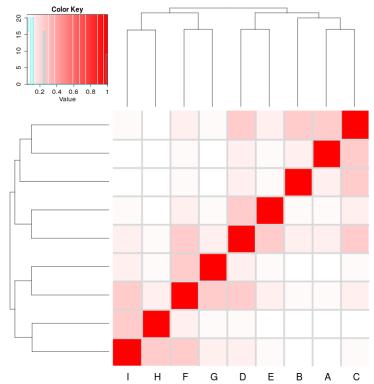


(f) Simulated gene function calls

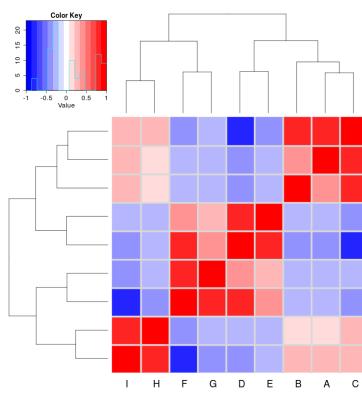
**Figure 3.21: Simulating expression from a graph structure.** An example graph structure is used to derive a correlation structure from the relative distances between nodes and simulate continuous gene expression with sampling from the multivariate normal distribution.



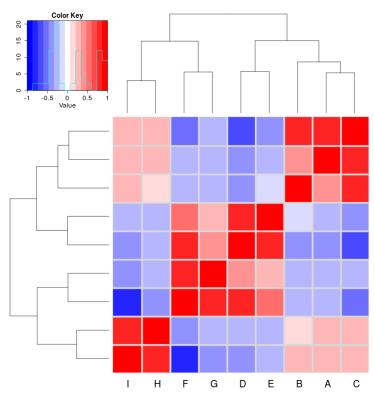
(a) Pathway structure with inhibition



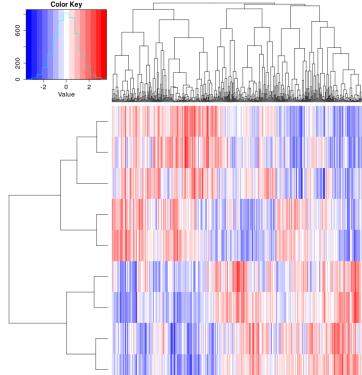
(b) Distance matrix



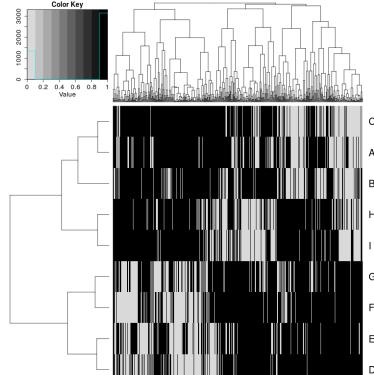
(c) Sigma,  $\Sigma$  (expected correlation)



(d) Simulated correlation structure



(e) Simulated expression data



(f) Simulated gene function calls

Figure 3.22: **Simulating expression from graph structure with inhibitions.** An example graph structure is used to derive a correlation structure from the relative distances between nodes and simulate continuous gene expression with sampling from the multivariate normal distribution.

manner as before (Figure 3.22(b)) with inhibitions accounted for by iteratively multiplying downstream nodes by  $-1$  to form blocks of negative correlations (as shown in Figures 3.22(c) and 3.22(d)). As before, a multivariate normal distribution with these negative correlations can be sampled to generate simulated data (as shown in Figures 3.22(e) and 3.22(f)).

These simulated datasets are amenable to simulating synthetic lethal partners of a query gene within a graph network. The query gene is assumed to be separate from the graph network pathway and is added to the dataset using the procedure in Section 3.2.2. Thus we can simulate known synthetic lethal partner genes within a synthetic lethal partner pathway structure.

## 3.5 Customised Functions and Packages Developed

[Move to Appendix?]

Various R packages have been developed throughout this thesis using `devtools` (Wickham and Chang, 2016) and `roxygen` (Wickham *et al.*, 2017) to enable reproducibility of customised analysis and visualisation. Many of these have the added benefit of the functions being documented, demonstrated in example vignettes, and released on GitHub to enable the research community to access utilise them in their own analysis. These are summarised in Table 2.6, along with the corresponding urls for their GitHub repository which contains a README file with instructions for installation with the `devtools` R package (Wickham and Chang, 2016) and links to the relevant vignette(s) where available.

### 3.5.1 Synthetic Lethal Interaction Prediction Tool

The statistical methodology for detection of synthetic lethality in gene expression data (SLIPT) is one of the main novel procedures developed in this thesis, as described in Section 3.1. The `slipt` R package has been prepared for release to accompany a publication demonstrating the applications of the methodology for identifying candidate interacting genes and pathways with *CDH1* in breast cancer (TCGA, 2012).

SLIPT is amenable to analysis of any effectively continuous measure of gene activity (e.g., microarray, RNA-Seq, protein abundance, or pathway metagenes). Executing `slipt` is straightforward: the `prep_data_for_SL` function scores samples as “low”, “medium”, or “high” for each gene, then the `detect_SL` function tests a given query gene against all potential partners by performing the chi-squared test and directional conditions. This function returns a table summarising the observed and expected sam-

ple numbers used for the directional criteria, the  $\chi^2$  values, and corresponding p-values including adjusting for multiple comparisons. The `count_of_SL` and `table_of_SL` functions serve to facilitate summary and extraction of the positive SLIPT hits, respectively, from the table of predictions of synthetic lethal partners.

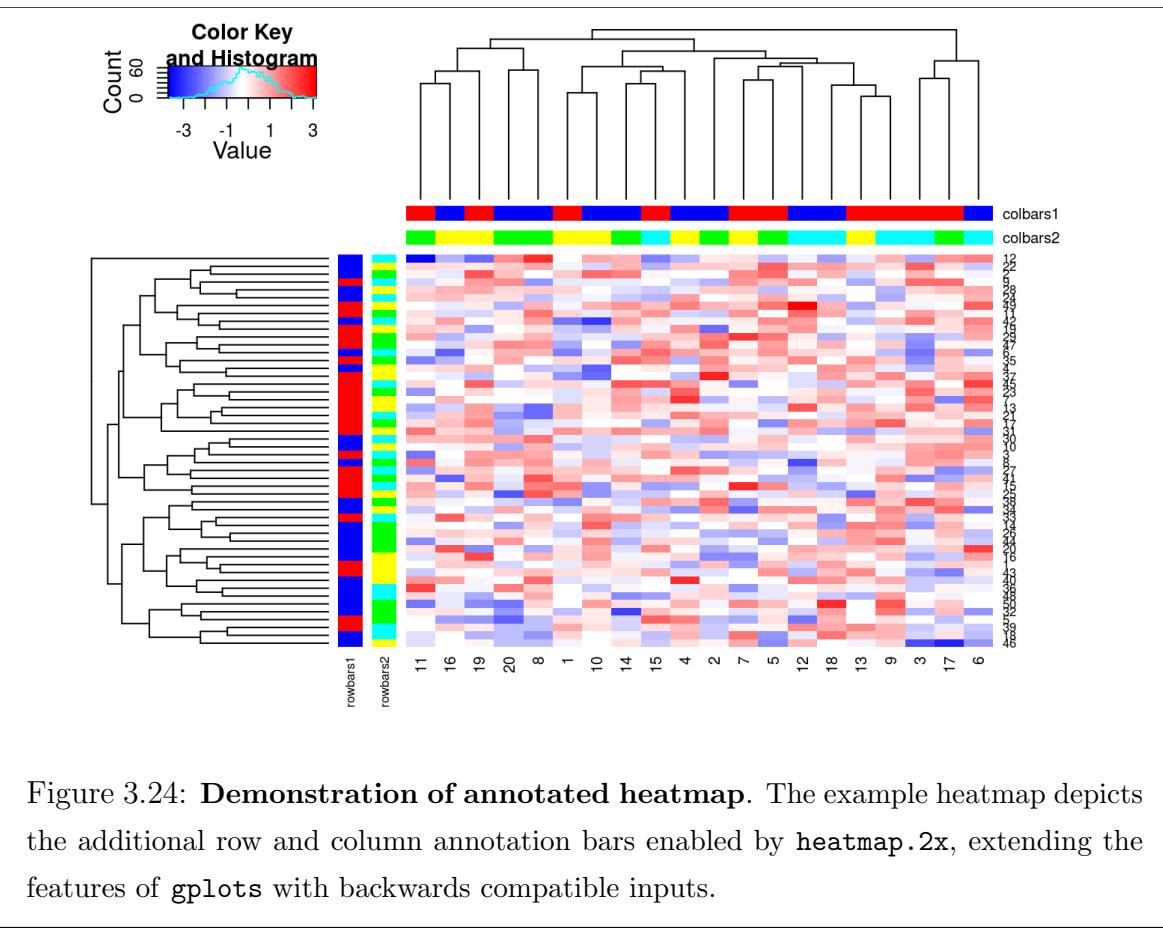
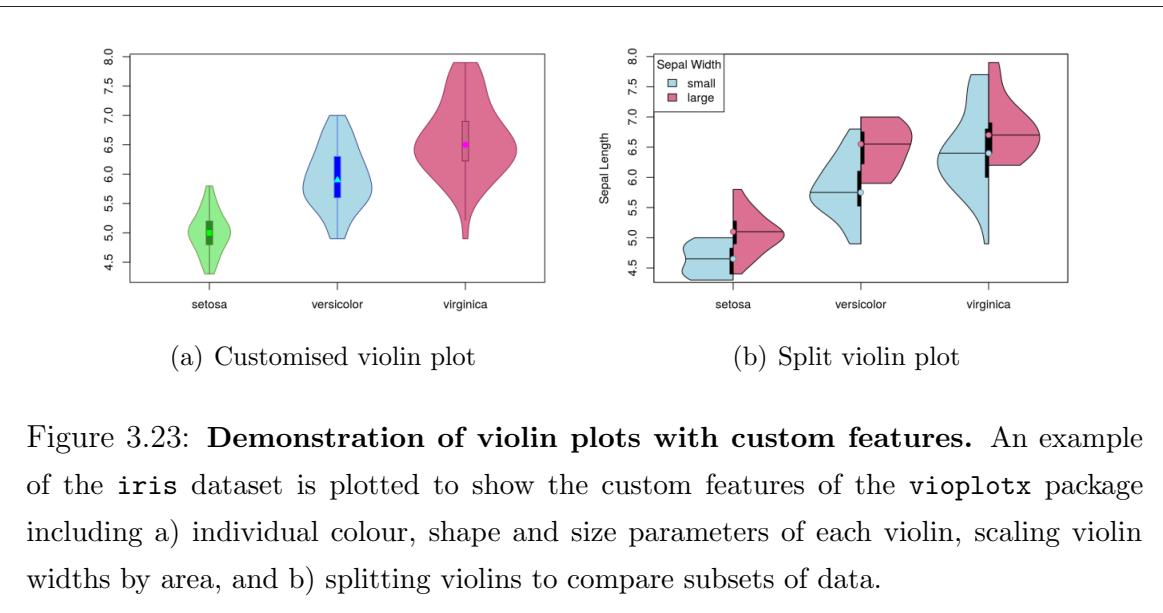
The SLIPT methodology in this package release has been used in later analyses rather than the corresponding source R code, including use on remote machines and upon simulated data. In particular, the functions in the package facilitate alterations to parameters, such as the proportion of samples called as exhibiting low or high gene activity. This release support reproducibility and enables wider use of SLIPT in future investigations into other disease genes.

### 3.5.2 Data Visualisation

Customisations to existing data visualisations in R have been developed to present data throughout this thesis. The `vioplotx` and `heatmap.2x` packages are enhancements of the `vioplot` package (Adler, 2005) and `heatmap.2` provided by the `gplots` package (Warnes *et al.*, 2015).

The `vioplotx` package provides an alternative visualisation (of continuous variables against categories) to the more familiar boxplot, showing variability of the data by the width of the plots. As demonstrated in Figure 3.23, the customised version enables separate plotting parameters for each violin with vector inputs for colour, shape, and size of various elements of the median point, central boxplot, borders, and fill colour for the violin. Scaling violin width to adjust violin area and splitting data by a second categorical variable is also enabled. This function is intended to be backwards compatible with the inputs of `vioplot` (applying scalar inputs across all violins) and `boxplot` (by enabling formula inputs as an S3 method). Each of these features is demonstrated with examples in respective vignettes on the package GitHub repository.

The `heatmap.2x` provides extensions for annotation colour bars for both the rows and columns (as shown in Figure 3.24). Multiple bars are enabled on both axes with matrix inputs (rather than single vector for `heatmap.2`) which facilitates additional plotting of gene and sample characteristics for comparison with correlation matrices, expression profiles, or pathway metagenes. Annotation bar inputs correspond to their orientation on the plot, each colour bar is provided as a column for the row annotation on the left of the heatmap and as a row for the column annotation on top of the heatmap. Row and column annotation bars are labelled with the column or row names respectively. Additional parameters enable resizing of these annotation bar labels and



control of reordering columns for if samples are ordered in advance (e.g., ranked by a metagene or split into groups clustered separately). These features were used through this thesis and are provided in a package GitHub repository.

### 3.5.3 Extensions to the iGraph Package

The following features were developed during this thesis using “iGraph” data objects, building upon the `igraph` package (Csardi and Nepusz, 2006). These have been released as separate packages for each respective procedure and can be installed together as a collection of extensions to the `igraph` package.

#### 3.5.3.1 Sampling Simulated Data from Graph Structures

The `graphsim` package implements the procedure for simulating gene expression from graph structures (as described in Section 3.4.2). By default, this derives a matrix with a geometrically decreasing weighting by distance (by shortest paths) between each pair of nodes with. An absolute decreasing weighting is also available with the option of to derive correlation structures from adjacency matrices or the number of links common partners (i.e., size of the shared “neighbourhood” (Hell, 1976)) between each pair of nodes. Functions to compute these are called directly by passing parameters to them when running the `generate_expression` or `make_sigma_mat` commands. This package enables simulating expression data directly from a graph structure (with the intermediate steps automated) or generating  $\Sigma$  parameters for `mvtnorm` from graph structures or matrices derived from them. These functions support assigning activating or inhibiting to each edge (with a `state` parameter).

#### 3.5.3.2 Plotting Directed Graph Structures

The `plot.igraph` package provides the `plot_directed` function specifically developed for directed graph structures and to plot activating or inhibiting for each edge (as described in Section 3.4.2). As shown in Figure 3.25, this function supports separate plotting parameters for each node, node label, and edge. This includes colours of node fill, border, label text, and edges and size of nodes, edge widths, arrowhead lengths, and font size of labels. The `state` parameter for assigning activating or inhibiting to each edge determines whether edges are depicted with 30° or 90° arrowheads. Colours are assigned separately and so they may be customised. Vectorised parameters are applied across each node or edge, whereas scalar parameters apply the same plotting parameters across them. The default layout function is `layout.fruchterman.reingold` but any layout function supported by `plot` function in `igraph` (Csardi and Nepusz, 2006)

is compatible such as `layout.kamada.kawai` used to implement the Kamada–Kawai algorithm (Kamada and Kawai, 1989) for graph plots throughout this thesis.

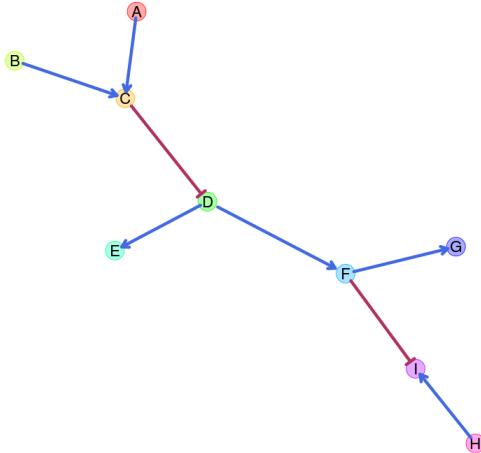


Figure 3.25: **Simulating graph structures.** An example graph structure which will be used throughout demonstrating the simulation procedure from graph structures. Here activating links are denoted by blue arrows and inhibiting links by red edges.

### 3.5.3.3 Computing Information Centrality

The shortest paths of a network are computed by the `igraph` package Csardi and Nepusz (2006) which can be extended to calculate the network efficiency but is not provided by the package itself (as described in Section 2.4.4). The “information centrality” of a vertex is computed as the relative change in the network efficiency when the vertex is removed. Information centrality is calculated iteratively for each node and the sum of information centrality for each vertex is the information centrality for the network. These metrics are provided by the `info.centrality` package.

### 3.5.3.4 Testing Pathway Structure with Permutation Testing

A network-based procedure developed was used for comparison of siRNA and SLIPT candidate genes in a pathway structure. Such pathway structure relationships were tested by computing the number of shortest paths between two different groups of nodes in either direction within a graph. This pathway relationship metric was implemented in the `pathway.structure.permutation` package with permutation testing (as described in sections 3.4.1 and 3.4.1.1).

### 3.5.3.5 Metapackage to Install iGraph Functions

These features may be installed together with `igraph.extensions` which can be accessed from a GitHub repository. This meta-package installs `igraph` (Csardi and Nepusz, 2006) and the packages described in Section 3.5.3 including their dependencies for matrix operations and statistical procedures: `Matrix`, `matrixcalc`, and `mvtnorm` (Bates and Maechler, 2016; Genz and Bretz, 2009; Genz *et al.*, 2016; Novomestky, 2012).

# Chapter 4

## Synthetic Lethal Analysis of Gene Expression Data

Having developed a statistical synthetic lethal detection methodology (SLIPT), it was applied to empirical (publicly available) cancer gene expression datasets in this Chapter. The analysis largely focuses findings from the TCGA breast cancer data (TCGA, 2012) which covers a range of clinical subtypes and is more closely modelled by siRNA data (Telford *et al.*, 2015) generated from screening experiments conducted in MCF10A breast cells. Although stomach cancer data will also be considered to replicate findings in an independent dataset and for its relevance to syndromic hereditary diffuse gastric cancer. The TCGA data also has the advantages of other clinical and molecular profiles (e.g., somatic mutation and DNA copy number) for many of the same samples, in addition to a considerable sample size for RNASeq expression data, treated with a rigorous procedure to minimise batch effects. Some findings will be replicated in the Cancer Cell Line Encyclopaedia (CCLE) (Barretina *et al.*, 2012) which may be more comparable to the cell line experiments.

Synthetic lethal candidate partners for *CDH1* will be described at both the gene and pathway level. SLIPT gene candidates will be analysed by cluster analysis for common expression profiles across samples and relationships with clinical factors and mutations in key breast cancer genes. These genes will also be compared to the gene candidates from a primary and secondary (validation) screens conducted by Telford *et al.* (2015) on isogenic cell lines. For comparison, an alternative SLIPT methodology which uses mutation data for *CDH1* against expression of candidate partners will also be presented which may better represent the null mutations in HDGC patients and the experiment cell model (Chen *et al.*, 2014). Pathways will be analysed by

over-representation analysis (with resampling for comparisons with siRNA data) and supported by a metagene analysis of pathway gene signatures. The pathway metagene expression profiles will be used to replicate known relationships between clinical and molecular characteristics for breast cancer and to demonstrate application of SLIPT directly on metagenes to detect synthetic lethal pathways.

Together these results will demonstrate the wide range of applications for SLIPT analysis and examine the synthetic lethal partners of *CDH1* in breast and stomach cancer. These synthetic lethal genes and pathways will be described in both context of the functional implications of novel synthetic lethal relationships and as potential actionable targets against *CDH1* deficient tumours, in addition to replication of established functions of E-cadherin. In particular, the focus of these analysis will be in comparisons with experimental screening data to explore the potential for SLIPT to augment such triage of candidate partners and support further experimental investigations. The key synthetic lethal partner pathways for *CDH1*, supported by both approaches, will be examined in more detail at the gene and pathway structure level in Chapter 5.

Some of the findings presented in this Chapter have also been included in manuscripts submitted for publication (Kelly *et al.*, 2017a,b) and may bear similarity to them, although the results in this thesis have been edited to cohesively fit with additional findings (including consistent data versions). These findings are the result of investigations conducted throughout this thesis project and only these contributions to the articles are included in this Chapter, not that conducted by co-authors.

## 4.1 Synthetic lethal genes in breast cancer

The SLIPT methodology (as described in Section 3.1) was applied to the normalised TCGA breast cancer gene expression dataset ( $n = 1168$ ). As shown in Table 4.1, the most significant genes had strong evidence of expression-based association with *CDH1* (high  $\chi^2$  values) with fewer samples exhibiting low expression of both genes than expected statistically. Eukaryotic translation gene were among the highest gene candidates, including initiation factors, elongation factors, and ribosomal proteins. These are clearly necessary for cancer cells to grow and proliferate, with sustained gene expression needed to maintain growth signaling pathways and resist apoptosis or immune factors translation may be subject to non-oncogene addiction for *CDH1*-deficient cells.

While these are among the strongest synthetic lethal candidates, translational genes are crucial to the viability of healthy cells and dosing for a selective synthetic lethal effect against these may be difficult compared to other biological functions which may also be supported among the SLIPT candidate genes. Furthermore, few known biological functions of *CDH1* were among the strongest SL candidates so the remaining candidate genes may also be informative since they are likely to contain these expected functions in addition to novel relationships for *CDH1*. Thus further pathway level analyses were also conducted to examine biological functions over-represented among synthetic candidate genes and identify synthetic lethal pathways.

Table 4.1: Candidate synthetic lethal gene partners of *CDH1* from SLIPT

| Gene             | Observed | Expected | $\chi^2$ value | p-value                | p-value (FDR)          |
|------------------|----------|----------|----------------|------------------------|------------------------|
| <i>TRIP10</i>    | 62       | 130      | 162            | $5.65 \times 10^{-34}$ | $1.84 \times 10^{-31}$ |
| <i>EEF1B2</i>    | 56       | 130      | 158            | $3.10 \times 10^{-33}$ | $9.45 \times 10^{-31}$ |
| <i>GBGT1</i>     | 61       | 131      | 156            | $1.08 \times 10^{-32}$ | $3.14 \times 10^{-30}$ |
| <i>ELN</i>       | 81       | 130      | 149            | $3.46 \times 10^{-31}$ | $8.82 \times 10^{-29}$ |
| <i>TSPAN4</i>    | 78       | 130      | 146            | $1.63 \times 10^{-30}$ | $3.79 \times 10^{-28}$ |
| <i>GLIPR2</i>    | 72       | 130      | 146            | $1.68 \times 10^{-30}$ | $3.86 \times 10^{-28}$ |
| <i>RPS20</i>     | 73       | 131      | 145            | $1.89 \times 10^{-30}$ | $4.28 \times 10^{-28}$ |
| <i>RPS27A</i>    | 80       | 130      | 143            | $5.53 \times 10^{-30}$ | $1.18 \times 10^{-27}$ |
| <i>EEF1A1P9</i>  | 63       | 130      | 141            | $1.91 \times 10^{-29}$ | $3.74 \times 10^{-27}$ |
| <i>C1R</i>       | 73       | 130      | 141            | $2.05 \times 10^{-29}$ | $3.97 \times 10^{-27}$ |
| <i>LYL1</i>      | 73       | 130      | 140            | $2.99 \times 10^{-29}$ | $5.74 \times 10^{-27}$ |
| <i>RPLP2</i>     | 71       | 130      | 139            | $4.88 \times 10^{-29}$ | $9.07 \times 10^{-27}$ |
| <i>C10orf10</i>  | 73       | 130      | 138            | $6.72 \times 10^{-29}$ | $1.20 \times 10^{-26}$ |
| <i>DULLARD</i>   | 74       | 131      | 138            | $9.29 \times 10^{-29}$ | $1.61 \times 10^{-26}$ |
| <i>PPM1F</i>     | 64       | 130      | 136            | $1.61 \times 10^{-28}$ | $2.65 \times 10^{-26}$ |
| <i>OBFC2A</i>    | 69       | 130      | 136            | $2.49 \times 10^{-28}$ | $3.93 \times 10^{-26}$ |
| <i>RPL11</i>     | 70       | 130      | 136            | $2.56 \times 10^{-28}$ | $3.97 \times 10^{-26}$ |
| <i>RPL18A</i>    | 70       | 130      | 135            | $3.08 \times 10^{-28}$ | $4.70 \times 10^{-26}$ |
| <i>MFNG</i>      | 76       | 131      | 133            | $7.73 \times 10^{-28}$ | $1.12 \times 10^{-25}$ |
| <i>RPS17</i>     | 77       | 131      | 133            | $8.94 \times 10^{-28}$ | $1.29 \times 10^{-25}$ |
| <i>MGAT1</i>     | 73       | 130      | 132            | $1.44 \times 10^{-27}$ | $2.03 \times 10^{-25}$ |
| <i>RPS12</i>     | 72       | 130      | 128            | $8.57 \times 10^{-27}$ | $1.12 \times 10^{-24}$ |
| <i>C10orf54</i>  | 73       | 130      | 127            | $1.37 \times 10^{-26}$ | $1.75 \times 10^{-24}$ |
| <i>LOC286367</i> | 72       | 130      | 126            | $2.20 \times 10^{-26}$ | $2.70 \times 10^{-24}$ |
| <i>GMFG</i>      | 70       | 130      | 126            | $2.20 \times 10^{-26}$ | $2.70 \times 10^{-24}$ |

Strongest candidate SL partners for *CDH1* by SLIPT with observed and expected samples with low expression of both genes

The modified mtSLIPT methodology (as described in Section 3.1) was also applied to the normalised TCGA breast cancer gene expression dataset, against somatic loss of function mutations in *CDH1*. As shown in Table D.1, the most significant genes also had strong evidence of expression associated with *CDH1* mutations (high  $\chi^2$  values) with fewer samples exhibiting both low expression and mutations of each gene than expected statistically. Although, these were not strongly supported as the expression analysis (in Table 4.1) nor were as many genes detected. This is unsurprising due to the lower sample size with matching somatic mutation data and the lower frequency of *CDH1* mutations compared to low expression by  $1/3$  quantiles.

The mtSLIPT candidates had more genes involved in cell and gene regulation, particularly DNA and RNA binding factors. The strongest candidates also include microtubule (*KIF12*), microfibril (*MFAP4*), and cell adhesion (*TENC1*) genes consistent with the established cytoskeletal role of *CDH1*. The elastin gene (*ELN*) was notably strongly supported by both expression and mutation SLIPT analysis of *CDH1* supporting interactions with extracellular proteins and the tumour microenvironment.

#### 4.1.1 Synthetic lethal pathways in breast cancer

Translational pathways were strongly over-represented in SLIPT partners, as shown in Table 4.2. These include ribosomal subunits, initiation, peptide elongation, and termination. Regulatory processes involving mRNA including 3' untranslated region (UTR) binding, L13a-mediated translational silencing, and nonsense-mediated decay were also implicated. These are consistent with protein translation being subject to “non-oncogene addiction” (Luo *et al.*, 2009), as a core process that is dysregulated to sustain cancer proliferation and survival (Gao and Roux, 2015).

Immune pathways, including the adaptive immune system and responses to infectious diseases were also strongly implicated as synthetic lethal with loss of E-cadherin. This is consistent with the alterations of immune response being a hallmark of cancer Hanahan and Weinberg (2000), since evading the immune system is necessary for cancer survival. Either of these systems are potential means to target *CDH1* deficient cells, although these were not detected in an isolated cell line experimental screen (Telford *et al.*, 2015) and the differences between findings in patient data will be described in more detail in Section 4.2.1.4.

It is also notable that the pathways over-represented in SLIPT candidate genes have strongly significant over-representation of Reactome pathways from the hypergeometric test (as described in Section 2.3.2). Even after adjusting stringently for multiple tests,

Table 4.2: Pathways for *CDH1* partners from SLIPT

| Pathways Over-represented   | Pathway Size | SL Genes | p-value (FDR)          |
|---|--------------|----------|------------------------|
| Eukaryotic Translation Elongation                                 | 86           | 81       | $1.3 \times 10^{-207}$ |
| Peptide chain elongation  | 83           | 78       | $5.6 \times 10^{-201}$ |
| Eukaryotic Translation Termination                                | 83           | 77       | $1.2 \times 10^{-196}$ |
| Viral mRNA Translation  | 81           | 76       | $1.2 \times 10^{-196}$ |
| Formation of a pool of free 40S subunits                          | 93           | 81       | $3.7 \times 10^{-194}$ |
| Nonsense Mediated Decay independent of the Exon Junction Complex  | 88           | 77       | $5.3 \times 10^{-187}$ |
| L13a-mediated translational silencing of Ceruloplasmin expression | 103          | 82       | $9.6 \times 10^{-183}$ |
| 3' -UTR-mediated translational regulation                         | 103          | 82       | $9.6 \times 10^{-183}$ |
| GTP hydrolysis and joining of the 60S ribosomal subunit           | 104          | 82       | $1.9 \times 10^{-181}$ |
| Nonsense-Mediated Decay   | 103          | 80       | $6.2 \times 10^{-176}$ |
| Nonsense Mediated Decay enhanced by the Exon Junction Complex     | 103          | 80       | $6.2 \times 10^{-176}$ |
| Adaptive Immune System  | 412          | 167      | $6.5 \times 10^{-174}$ |
| Eukaryotic Translation Initiation                                 | 111          | 82       | $5.7 \times 10^{-173}$ |
| Cap-dependent Translation Initiation                              | 111          | 82       | $5.7 \times 10^{-173}$ |
| SRP-dependent cotranslational protein targeting to membrane       | 104          | 79       | $2.0 \times 10^{-171}$ |
| Translation   | 141          | 91       | $6.1 \times 10^{-170}$ |
| Infectious disease  | 347          | 146      | $1.6 \times 10^{-166}$ |
| Influenza Infection   | 117          | 81       | $1.9 \times 10^{-163}$ |
| Influenza Viral RNA Transcription and Replication                 | 108          | 77       | $1.9 \times 10^{-160}$ |
| Influenza Life Cycle  | 112          | 77       | $2.5 \times 10^{-156}$ |

Gene set over-representation analysis (hypergeometric test) for Reactome pathways in SLIPT partners for *CDH1*

biologically related pathways give consensus support to these pathways. These pathways are further supported by testing for synthetic lethality against *CDH1* mutations (mtSLIPT) with many of these pathways also among the most strongly supported in this analysis (shown in Table D.2). This analysis more closely represents the null *CDH1* mutations in HDGC (Guilford *et al.*, 1998) and the experimental MCF10A cell model (Chen *et al.*, 2014). Although it still supports translational and immune pathways not detected in the isolated experimental system, G-protein-coupled receptors (GPCRs) were also among the most strongly supported pathways, supporting the experimental findings of Telford *et al.* (2015) for these intracellular signalling pathways already being targeted for other diseases.

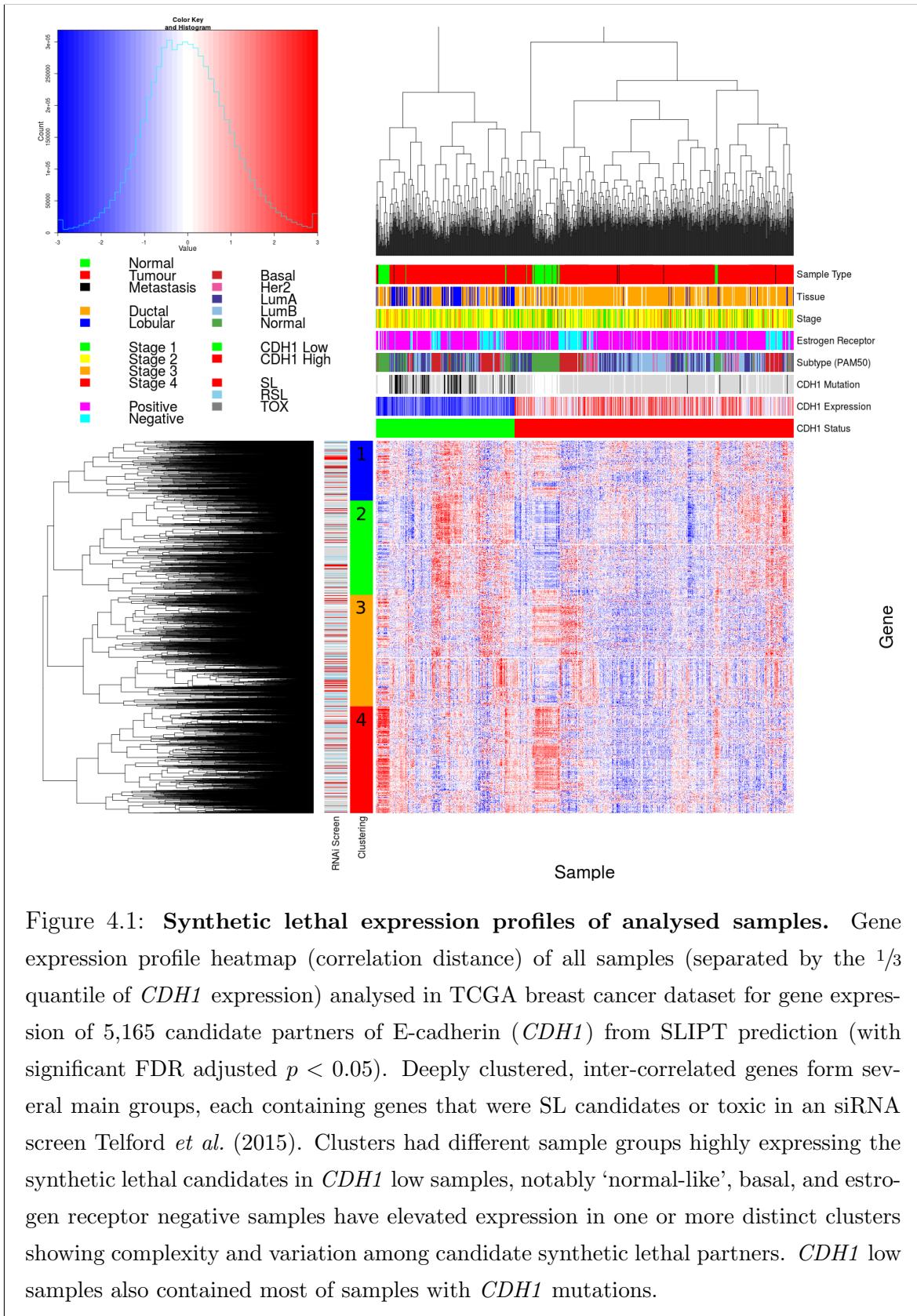
#### 4.1.2 Expression profiles of synthetic lethal partners

Due to the sheer number of gene candidates and to examine their expression patterns, investigations proceeded into correlation structure and pathway over-representation. This serves to explore the functional similarity of the synthetic lethal partners of *CDH1*, with the eventual aim to assess their utility as drug targets. As shown in Figure 4.1 (which clusters *CDH1* lowly expressing samples separately), there were several large

clusters of genes among the expression profiles of the *CDH1* synthetic lethal candidate partners. The clustering suggests co-regulation of genes or pathway correlation between partner gene candidates. A number of candidates from an experimental RNAi screen study performed by Telford *et al.* (2015) were also identified by this approach. In addition, we identified novel gene candidates, which had little effect on viability in isogenic cell line experiments.

In these expression profiles, a gene with a moderate or high signal across samples exhibiting low *CDH1* expression would represent a potential drug target. However, it appears that several molecular subtypes of cancer have elevation of different clusters of synthetic lethal candidates in samples with low *CDH1*. This clustering suggests that different targets or combinations could be effective in different patients suggesting potential utility for stratification. In particular, estrogen receptor negative, basal subtype, and “normal-like” samples Dai *et al.* (2015); Eroles *et al.* (2012); Parker *et al.* (2009) have elevation of genes specific to particular clusters which is indicative of some synthetic lethal interactions being specific to a particular molecular subtype or genetic background. Thus synthetic lethal drug therapy against these subtypes may be ineffective if it were designed against genes in another cluster.

A similar correlation structure was observed among the candidates tested against *CDH1* mutation (mtSLIPT), as shown in Figure D.1. This clustering analysis similarly identified several major clusters of putative synthetic lethal partner genes. Although in this case many partner genes had consistently high expression across most of the (predominantly lobular subtype) *CDH1* breast cancer samples. However, a major exception to this in the *CDH1* expression analysis were the normal samples which were excluded from the mutation data (as they were not tested for tumour-specific genotypes). This supports synthetic lethal interventions being more applicable to *CDH1* mutant tumours and genotyping tumours for loss of function will be essential for clinical application. There was still considerable correlation structure, particularly among *CDH1* wildtype samples, sufficient to distinguish gene clusters. In contrast to the expression analysis the (predominantly ductal *CDH1* wildtype) basal subtype and estrogen receptor negative samples have depleted expression among most candidate synthetic lethal partners. This is consistent with synthetic lethal interventions only being effective in lobular estrogen receptor positive breast cancers in which they are a more common, as recurrent (driver) mutation. However, the remaining samples are still informative for synthetic lethal analysis (by SLIPT) as it requires highly expressing *CDH1* samples for comparison.



The *CDH1* mutant samples (in Figure 4.1) were predominantly among the *CDH1* lowly expressing samples and distributed throughout *CDH1* samples with clustering analysis. Thus the molecular profiles of *CDH1* low samples are indistinguishable from *CDH1* mutant samples with the exception of normal samples (that do not have somatic mutation data as it is inferred from comparison to them to tumour-specific genotypes). Conversely, many of the *CDH1* mutant samples (in Figure D.1) had among the lowest *CDH1* expression and some of the synthetic lethal partners were also highly expressed in lowly expressing *CDH1* wildtype samples, despite these not being considered as “inactivated” by mtSLIPT analysis.

Together these results support the use for low *CDH1* expression as a strategy for detecting *CDH1* inactivation. This has the benefit of increasing sample size (including samples such as normal tissue which do not have somatic mutation data available) and increasing the expected number of mutually inactive (low-low) samples for the directional criteria of (mt)SLIPT which enabling it to better distinguish significant deviations below this (as discussed in Section 6.4). This also circumvents the assumption that all (detected) mutations are inactivating (although synonymous mutations were excluded from the analysis), which may not be the case for several highly expressing *CDH1* mutant samples that do not cluster together in Figures 4.1 or D.1. One of these exhibits among the lowest expression for many predicted synthetic lethal partners and would not be vulnerable to inactivation of these genes. As such correctly genotyping inactivating mutations will be essential in clinical practice for synthetic lethal targeting tumour suppressor genes, particularly for other genes such as *TP53* where oncogenic and tumour suppressor mutations (with different molecular consequences) are both common in cancers. Using expression as a measure of gene expression also avoids the assumptions that mutations are somatic rather than germline and that gene inactivation is by detectable mutations rather than other mechanisms such as epigenetic changes which is supported by many lowly expressing *CDH1* wildtype samples clustering with similar profiles to mutant samples.

#### 4.1.2.1 Subgroup pathway analysis

Synthetic lethal gene candidates for *CDH1* from SLIPT performed on RNA-Seq expression data were also used for pathway over-representation analyses (as described in Section 2.3.2). The correlation structure in the expression of candidates synthetic lethal genes in *CDH1* low tumours (lowest 1/3<sup>rd</sup> quantile of expression) was examined for distinct biological pathways in subgroups of genes elevated in different clusters of samples. These genes were highly expressed in different samples with their clinical fac-

Table 4.3: Pathway composition for clusters of *CDH1* partners from SLIPT

| Pathways Over-represented in Cluster 1                                       | Pathway Size | Cluster Genes | p-value (FDR)          |
|--|--------------|---------------|------------------------|
| Collagen formation   | 67           | 10            | $4.0 \times 10^{-11}$  |
| Extracellular matrix organisation  | 238          | 21            | $1.8 \times 10^{-9}$   |
| Collagen biosynthesis and modifying enzymes                                  | 56           | 8             | $1.8 \times 10^{-9}$   |
| Uptake and actions of bacterial toxins                                       | 22           | 5             | $9.5 \times 10^{-9}$   |
| Elastic fibre formation  | 37           | 6             | $1.9 \times 10^{-8}$   |
| Muscle contraction   | 62           | 7             | $2.4 \times 10^{-7}$   |
| Fatty acid, triacylglycerol, and ketone body metabolism                      | 117          | 10            | $4.9 \times 10^{-7}$   |
| XBP1(S) activates chaperone genes  | 51           | 6             | $6.6 \times 10^{-7}$   |
| IRE1alpha activates chaperones   | 54           | 6             | $1.2 \times 10^{-6}$   |
| Neurotoxicity of clostridium toxins  | 10           | 3             | $1.3 \times 10^{-6}$   |
| Retrograde neurotrophin signalling   | 10           | 3             | $1.3 \times 10^{-6}$   |
| Assembly of collagen fibrils and other multimeric structures                 | 40           | 5             | $1.9 \times 10^{-6}$   |
| Collagen degradation   | 58           | 6             | $2.0 \times 10^{-6}$   |
| Arachidonic acid metabolism  | 41           | 5             | $2.1 \times 10^{-6}$   |
| Synthesis of PA  | 26           | 4             | $3.0 \times 10^{-6}$   |
| Signaling by NOTCH   | 80           | 7             | $3.3 \times 10^{-6}$   |
| Signalling to RAS  | 27           | 4             | $3.7 \times 10^{-6}$   |
| Integrin cell surface interactions   | 82           | 7             | $4.2 \times 10^{-6}$   |
| Smooth Muscle Contraction  | 28           | 4             | $4.4 \times 10^{-6}$   |
| ECM proteoglycans  | 66           | 6             | $6.3 \times 10^{-6}$   |
| Pathways Over-represented in Cluster 2                                       | Pathway Size | Cluster Genes | p-value (FDR)          |
| Eukaryotic Translation Elongation  | 86           | 75            | $1.1 \times 10^{-181}$ |
| Viral mRNA Translation   | 81           | 72            | $9.8 \times 10^{-179}$ |
| Peptide chain elongation   | 83           | 72            | $1.9 \times 10^{-175}$ |
| Eukaryotic Translation Termination   | 83           | 72            | $1.9 \times 10^{-175}$ |
| Formation of a pool of free 40S subunits                                     | 93           | 75            | $1.9 \times 10^{-171}$ |
| Nonsense Mediated Decay independent of the Exon Junction Complex             | 88           | 72            | $9.9 \times 10^{-168}$ |
| L13a-mediated translational silencing of Ceruloplasmin expression            | 103          | 75            | $3.0 \times 10^{-159}$ |
| 3' -UTR-mediated translational regulation                                    | 103          | 75            | $3.0 \times 10^{-159}$ |
| Nonsense-Mediated Decay  | 103          | 75            | $3.0 \times 10^{-159}$ |
| Nonsense-Mediated Decay enhanced by the Exon Junction Complex                | 103          | 75            | $3.0 \times 10^{-159}$ |
| SRP-dependent cotranslational protein targeting to membrane                  | 104          | 75            | $3.2 \times 10^{-158}$ |
| GTP hydrolysis and joining of the 60S ribosomal subunit                      | 104          | 75            | $3.2 \times 10^{-158}$ |
| Eukaryotic Translation Initiation  | 111          | 75            | $4.5 \times 10^{-151}$ |
| Cap-dependent Translation Initiation   | 111          | 75            | $4.5 \times 10^{-151}$ |
| Influenza Infection  | 117          | 75            | $1.4 \times 10^{-145}$ |
| Influenza Viral RNA Transcription and Replication                            | 108          | 72            | $5.7 \times 10^{-145}$ |
| Translation  | 141          | 81            | $8.0 \times 10^{-143}$ |
| Influenza Life Cycle   | 112          | 72            | $2.3 \times 10^{-141}$ |
| Infectious disease   | 347          | 103           | $2.2 \times 10^{-95}$  |
| Formation of the ternary complex, and subsequently, the 43S complex          | 47           | 33            | $6.8 \times 10^{-80}$  |
| Pathways Over-represented in Cluster 3                                       | Pathway Size | Cluster Genes | p-value (FDR)          |
| Adaptive Immune System   | 412          | 90            | $6.1 \times 10^{-61}$  |
| Chemokine receptors bind chemokines  | 52           | 27            | $6.7 \times 10^{-56}$  |
| Generation of second messenger molecules                                     | 29           | 21            | $6.5 \times 10^{-55}$  |
| Immunoregulatory interactions between a Lymphoid and a non-Lymphoid cell     | 64           | 29            | $6.5 \times 10^{-55}$  |
| TCR signalling   | 62           | 27            | $8.9 \times 10^{-51}$  |
| Peptide ligand-binding receptors   | 161          | 40            | $1.5 \times 10^{-45}$  |
| Translocation of ZAP-70 to Immunological synapse                             | 16           | 14            | $3.1 \times 10^{-43}$  |
| Costimulation by the CD28 family   | 51           | 22            | $4.0 \times 10^{-43}$  |
| PD-1 signalling  | 21           | 15            | $4.0 \times 10^{-41}$  |
| Class A/I (Rhodopsin-like receptors)   | 258          | 50            | $6.7 \times 10^{-41}$  |
| Phosphorylation of CD3 and TCR zeta chains                                   | 18           | 14            | $1.3 \times 10^{-40}$  |
| Interferon gamma signalling  | 74           | 24            | $5.0 \times 10^{-39}$  |
| GPCR ligand binding  | 326          | 57            | $1.8 \times 10^{-38}$  |
| Cytokine Signaling in Immune system  | 268          | 48            | $8.9 \times 10^{-37}$  |
| Downstream TCR signalling  | 45           | 18            | $1.8 \times 10^{-35}$  |
| G <sub>αi</sub> signalling events  | 167          | 33            | $2.2 \times 10^{-33}$  |
| Cell surface interactions at the vascular wall                               | 99           | 21            | $1.3 \times 10^{-26}$  |
| Interferon Signalling  | 164          | 28            | $1.7 \times 10^{-26}$  |
| Extracellular matrix organisation  | 238          | 35            | $2.7 \times 10^{-25}$  |
| Antigen activates B Cell Receptor leading to generation of second messengers | 32           | 12            | $7.2 \times 10^{-25}$  |
| Pathways Over-represented in Cluster 4                                       | Pathway Size | Cluster Genes | p-value (FDR)          |
| Extracellular matrix organisation  | 238          | 48            | $8.0 \times 10^{-41}$  |
| Class A/I (Rhodopsin-like receptors)   | 258          | 47            | $2.8 \times 10^{-36}$  |
| GPCR ligand binding  | 326          | 54            | $2.1 \times 10^{-34}$  |
| G <sub>αi</sub> signalling events  | 83           | 22            | $1.4 \times 10^{-31}$  |
| GPCR downstream signalling   | 472          | 68            | $1.1 \times 10^{-29}$  |
| Haemostasis  | 423          | 61            | $3.3 \times 10^{-29}$  |
| Platelet activation, signalling and aggregation                              | 180          | 31            | $7.1 \times 10^{-28}$  |
| Binding and Uptake of Ligands by Scavenger Receptors                         | 40           | 14            | $9.9 \times 10^{-27}$  |
| RA biosynthesis pathway  | 22           | 11            | $2.5 \times 10^{-26}$  |
| Response to elevated platelet cytosolic Ca <sup>2+</sup>                     | 82           | 19            | $3.0 \times 10^{-26}$  |
| Developmental Biology  | 420          | 57            | $3.5 \times 10^{-26}$  |
| G <sub>αi</sub> signalling events  | 167          | 28            | $7.3 \times 10^{-26}$  |
| Platelet degranulation   | 77           | 18            | $1.6 \times 10^{-25}$  |
| Gastrin-CREB signalling pathway via PKC and MAPK                             | 171          | 28            | $2.5 \times 10^{-25}$  |
| Muscle contraction   | 62           | 16            | $4.7 \times 10^{-25}$  |
| G <sub>αq</sub> signalling events  | 150          | 25            | $3.2 \times 10^{-24}$  |
| Retinoid metabolism and transport  | 34           | 12            | $5.0 \times 10^{-24}$  |
| Phase 1 - Functionalisation of compounds                                     | 67           | 16            | $6.5 \times 10^{-24}$  |
| Signalling by Retinoic Acid  | 42           | 13            | $6.7 \times 10^{-24}$  |
| Degradation of the extracellular matrix                                      | 102          | 19            | $1.4 \times 10^{-22}$  |

tors including estrogen receptor status and intrinsic (PAM50) subtype (Parker *et al.*, 2009) shown in Figure 4.1.

As shown by the most over-represented pathways in Table 4.3, each correlated cluster of candidate synthetic lethal partners of *CDH1* contains functionally different genes. Cluster 1 contains genes with less evidence of over-represented pathways than other clusters, corresponding to less correlation between genes within the cluster, and to it being a relatively small group. While there is some indication that collagen biosynthesis, microfibril elastic fibres, extracellular matrix, and metabolic pathways may be over-represented in Cluster 1, these results are mainly based on small pathways containing few synthetic lethal genes. Genes in Cluster 2 exhibited low expression in normal tissue samples compared to tumour samples (see Figure 4.1) and show compelling evidence of over-representation of post-transcriptional gene regulation and protein translation processes. Similarly, Cluster 3 has over-representation of immune signalling pathways (including chemokines, secondary messenger, and TCR signaling) and downstream intracellular signalling cascades such as G protein coupled receptor (GPCR) and G<sub>αi</sub> signalling events. While pathway over-representation was weaker among genes in Cluster 4, they contained intracellular signalling pathways and were highly expressed in normal samples (in contrast to Cluster 2). Cluster 4 also involved extracellular factors and stimuli such as extracellular matrix, platelet activation, ligand receptors, and retinoic acid signalling.

Based on these results, potential synthetic lethal partners of *CDH1* include processes known to be dysregulated in cancer, such as translational, cytoskeletal, and immune processes. Intracellular signalling cascades such as the GPCRs and extracellular stimuli for these pathways were also implicated in potential synthetic lethality with *CDH1*.

Similar translational, cytoskeletal, and immune processes were identified among SLIPT partners with respect to *CDH1* mutation, shown in Table D.3. While GPCR signalling was replicated in mtSLIPT analysis, there was also stronger over-representation for NOTCH, ERBB2, and PI3K/AKT signalling in mutation analysis consistent with these signals being important for proliferation of *CDH1* deficient tumours. The GPCR and PI3K/AKT pathways are of particular interest as pathways with oncogenic mutations that can be targeted and downstream effects on translation (a strongly supported process across analyses). Extracellular matrix pathways (such as elastic fibre formation) were also supported across analyses (in Tables 4.3 and D.3) consistent with the

established cell-cell signalling role of *CDH1* and the importance of the tumour microenvironment for cancer proliferation.

## 4.2 Comparison of synthetic lethal gene candidates

### 4.2.1 Comparison with siRNA screen candidates

Gene candidates were compared between computational (SLIPT in TCGA breast cancer data) and experimental (the primary siRNA screen performed by Telford *et al.* (2015)) approaches in Figure 4.2. The number of genes detected by both methods did not produce a significant overlap but these may be difficult to compare due to vast differences between the detection methods. There were similar issues comparison of mtSLIPT genes tested against *CDH1* mutations (in Figure G.2), despite excluded genes not tested by both methods in either test. However, these intersecting genes may still be functionally informative or amenable to drug triage as they were replicated across both methods and pathway over-representation differed between the sections of the Venn diagram (see Figure 4.2).

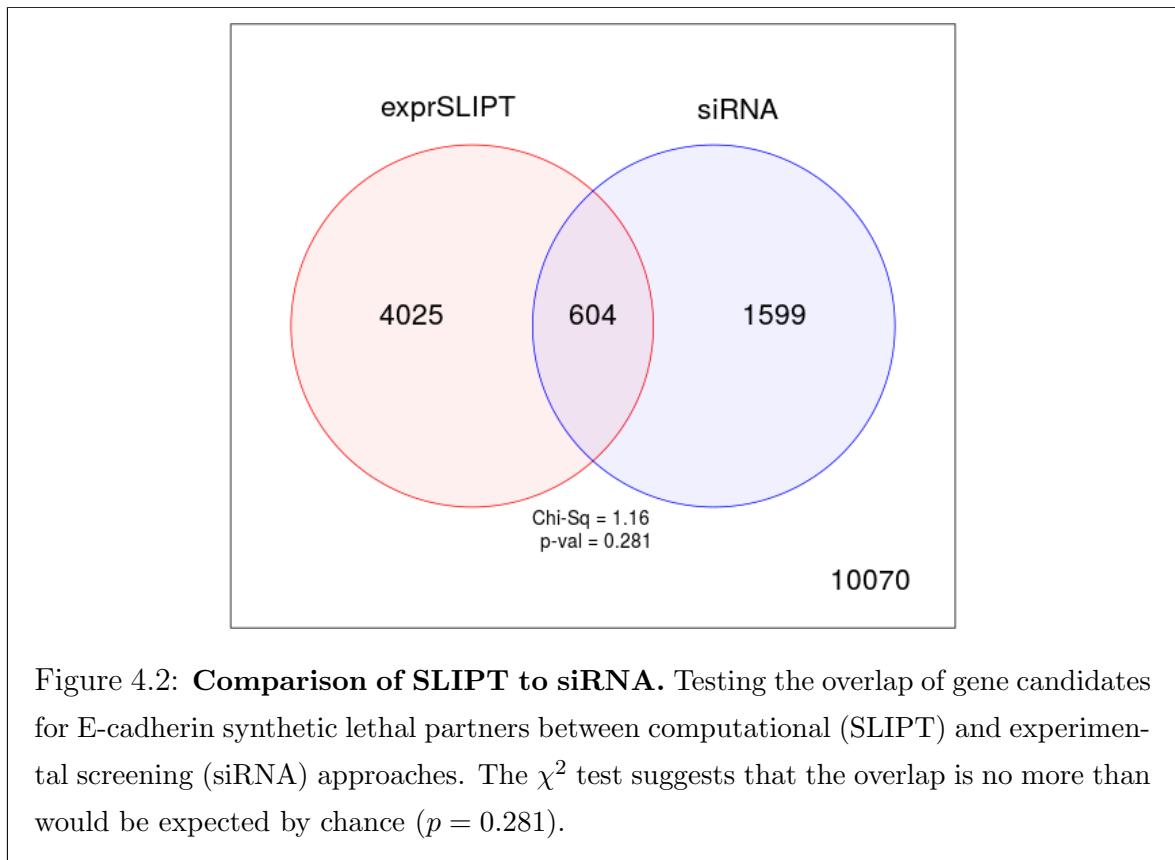
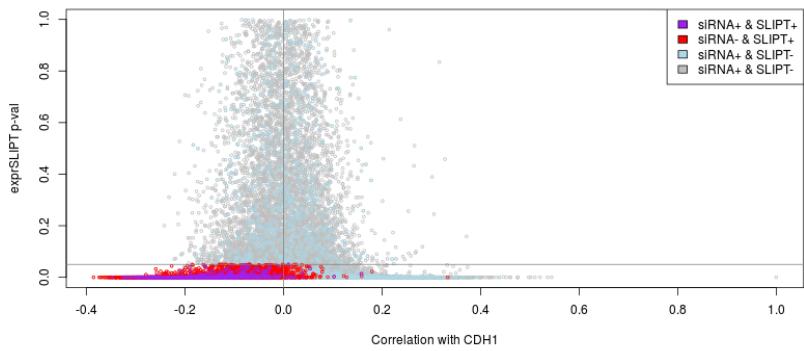
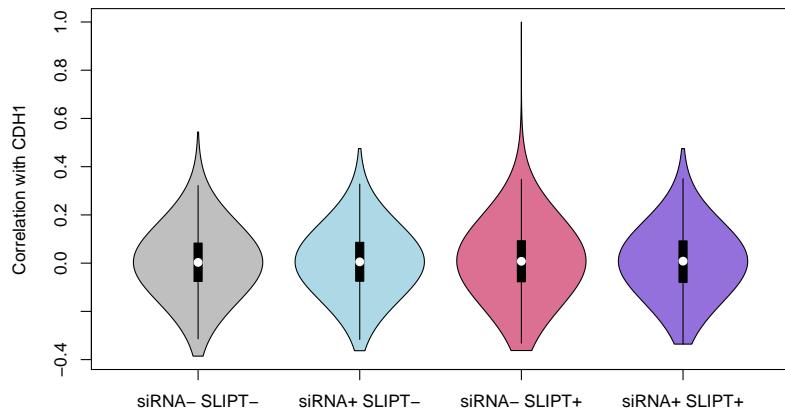


Figure 4.2: **Comparison of SLIPT to siRNA.** Testing the overlap of gene candidates for E-cadherin synthetic lethal partners between computational (SLIPT) and experimental screening (siRNA) approaches. The  $\chi^2$  test suggests that the overlap is no more than would be expected by chance ( $p = 0.281$ ).

#### 4.2.1.1 Comparison with correlation



**Figure 4.3: Compare SLIPT and siRNA genes with correlation.** The  $\chi^2$  p-values for genes tested by SLIPT (in TCGA breast cancer) expression analysis were compared against Pearson's correlation of gene expression with *CDH1*. Genes detected by SLIPT or siRNA are coloured according to the legend.



**Figure 4.4: Compare SLIPT and siRNA genes with correlation.** Genes detected as candidate synthetic lethal partners by SLIPT (in TCGA breast cancer) expression analysis and experimental screening (with siRNA) were compared against Pearson's correlation of gene expression with *CDH1*. There were no differences in correlation between gene groups detected by either approach.

Another potential means to triage drug target candidates is correlation of expression profiles with *CDH1*. Correlation with *CDH1* was compared to SLIPT and siRNA results in Figure 4.3. The genes not detected by SLIPT (including siRNA candidates) included genes with high (insignificant) SLIPT p-values. As expected, these genes were distributed around a correlation of zero and genes with higher correlation with *CDH1* (either direction) had more significant SLIPT p-values, although there were exceptions to this trend and larger positive correlations were negative correlations.

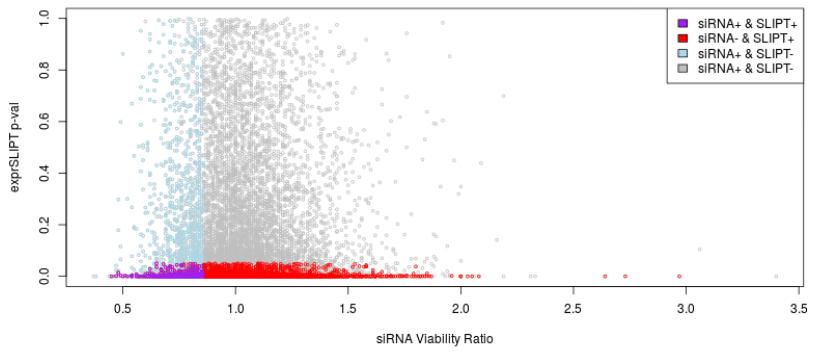
The majority of SLIPT candidates appeared to have negative correlations and moreso for those genes detected by both approaches, although these were typically weak correlations and are unlikely to be sufficient to detect such genes on their own. This is supported by simulation results in Section 6.4.

There were not strong positive correlations with *CDH1* among siRNA candidates, consistent with previous findings that co-expression is not predictive of synthetic lethality (Jerby-Arnon *et al.*, 2014; Lu *et al.*, 2015). Negative correlation may not be indicative of synthetic lethality either as many siRNA candidates also had positive correlations. The SLIPT methodology has shown to detect genes with both positive and negative correlations, although it does appear to preferentially detect negatively correlated genes to some extent. These findings were replicated with the mtSLIPT approach against *CDH1* mutation (in Figure D.3), although the range of the  $\chi^2$  p-values differ due to lower sample size for mutation analysis.

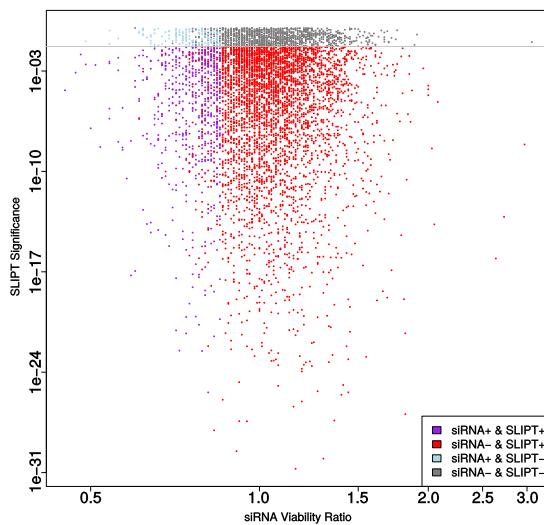
However, the apparent tendency for genes detected by SLIPT or siRNA to have negative correlations with *CDH1* expression may be due to the smaller number of genes in these groups. The distribution of *CDH1* correlations does not differ across these gene groups (as shown by Figures 4.4 and D.4). Therefore further triage of gene candidates by correlation is not suitable, nor is use of correlation itself to predict synthetic lethal partners in the first place.

#### 4.2.1.2 Comparison with viability

A similar comparison of SLIPT results was made with the viability ratio (of *CDH1* mutant to wildtype) in the primary siRNA screen performed by Telford *et al.* (2015). The significance and viability thresholds used for SLIPT and siRNA detection of synthetic lethal candidate partners of *CDH1* are clear in Figure 4.5. However note that not all of the genes below these thresholds are necessarily selected to be candidate partners as additional criteria were used in each case: directional criteria as for SLIPT (see Section 3.1) and minimum wildtype viability for siRNA (Telford *et al.*, 2015).



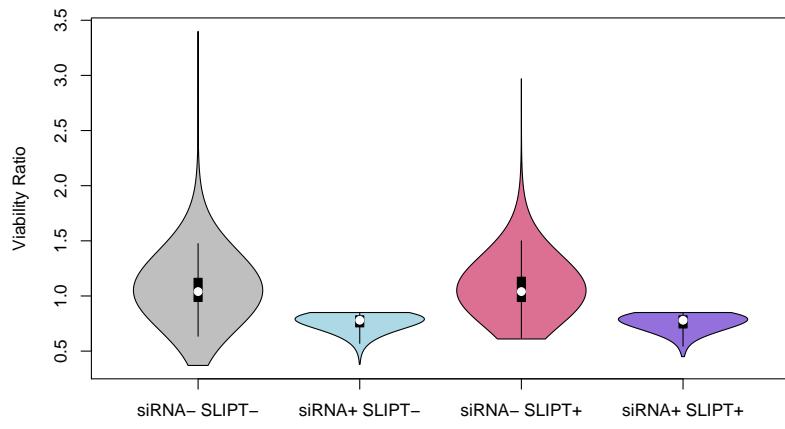
**Figure 4.5: Compare SLIPT and siRNA genes with siRNA viability.** The  $\chi^2$  p-values for genes tested by SLIPT (in TCGA breast cancer) expression analysis were compared against the viability ratio of *CDH1* mutant and wildtype cells in the primary siRNA screen. Genes detected by SLIPT or siRNA are coloured according to the legend.



**Figure 4.6: Compare SLIPT and siRNA genes with viability.** The  $\chi^2$  p-values for genes tested by SLIPT (in TCGA breast cancer) expression analysis were compared (on a log-scale) against the viability ratio of *CDH1* mutant and wildtype cells in the primary siRNA screen. Genes detected by SLIPT or siRNA are coloured according to the legend with a grey line for  $p = 0.05$ .

There does not appear to be a clear relationship between SLIPT and siRNA candidates. Many genes not detected by both approaches were numerous in Figures 4.2 and D.2. These genes detected by either are not necessarily near the thresholds for the other. In this respect the SLIPT approach with patient data and cell line experiments are independent means to identify synthetic lethal candidates. While genes detected by both approaches were not necessarily more strongly supported by either, the genes with a viability closer to 1 (no synthetic lethal effect) in siRNA included those with more significant SLIPT p-values whereas more extreme viability ratios tended to be less significant (as shown by a logarithmic plot in Figure 4.6). Although it should be noted that genes with more moderate viability ratios were more common and SLIPT was capable (despite adjusting for multiple testing) of detecting significant genes with extreme viability ratios, particularly those considerably lower than 1.

However, there was not support for SLIPT candidates or those detected by both approaches having considerably different viability ratios (as shown in Figures 4.7 and D.5). The difference between the gene groups stems largely from the viability thresholds used by Telford *et al.* (2015) to detect synthetic lethal candidates in the primary screen, rather than more extreme viability ratios for genes identified by SLIPT.



**Figure 4.7: Compare SLIPT and siRNA genes with siRNA viability.** Genes detected as candidate synthetic lethal partners by SLIPT (in TCGA breast cancer) expression analysis and experimental screening (with siRNA) were compared against the viability ratio of *CDH1* mutant and wildtype cells in the primary siRNA screen. There were clear no differences in viability between genes detected by SLIPT and those not with the differences being primarily due to viability thresholds being used to detect synthetic lethality by Telford *et al.* (2015).

#### **4.2.1.3 Comparison with secondary screen siRNA screen candidates**

However, it should be noted that genes with a lower viability ratio were not necessarily the strongest supported by experimental screening. The primary screen (with 4 pooled siRNAs) has been used for the majority of comparisons in this thesis because the genome-wide panel of target genes screened enables a large number of genes to be compared with SLIPT results from gene expression and somatic mutation analysis. A secondary screen was also performed by Telford *et al.* (2015) on the isogenic MCF10A breast cell lines to individually validate the siRNAs separately, with the strongest candidates being those exhibiting synthetic lethal viability ratios replicated across independently targeting siRNAs. This was performed for the top 500 candidates (with the lowest viability ratio) from the primary screen and the 482 of these genes also tested by SLIPT in breast cancer (and the 486 genes tested by SLIPT in stomach cancer).

The secondary screen results are given in Appendix C which show that SLIPT candidate genes are more significantly ( $p = 7.49 \times 10^{-3}$  by Fisher's exact test) more likely to be validated in the secondary screen and are thus informative of more robust partner genes, in addition to providing support that these interactions are consistent with expression profiles from heterogeneous patient samples across genetic backgrounds. While the individual genes detected by either approach do not necessarily match (and are potentially false-positives), the biological functions important in *CDH1* deficient cancers and potential mechanisms for specific targeting of them can be further supported by pathway analysis of the gene detected by either method. The genes detected by both approaches may therefore be more informative at the pathway level, where it is unlikely for a pathway to be consistently detected by chance. As the SLIPT candidates differ from the siRNA candidates (and are more likely to be validated), they can provide additional mechanisms by which *CDH1* deficient cancers proliferate and vulnerabilities that may be exploited against them by using the synthetic lethal pathways.

#### **4.2.1.4 Comparison of screen at pathway level**

These pathway over-representation analyses (performed as described in Section 2.3.2) correspond to genes separated into SLIPT or siRNA screen candidates unique to either method or detected by both (Table 4.4). The SLIPT-specific gene candidates were involved most strongly with translational and immune regulatory pathways, although extracellular matrix pathways were also supported. These pathways were largely consistent with those identified in Table 4.2 and in the clustering analysis (Table 4.3). The

Table 4.4: Pathway composition for *CDH1* partners from SLIPT and siRNA screening

| Predicted only by SLIPT (4025 genes)                              | Pathway Size | Genes Identified | p-value (FDR)          |
|---|--------------|------------------|------------------------|
| Eukaryotic Translation Elongation                                 | 80           | 75               | $1.5 \times 10^{-182}$ |
| Peptide chain elongation  | 77           | 72               | $2.9 \times 10^{-176}$ |
| Viral mRNA Translation  | 75           | 70               | $4.9 \times 10^{-172}$ |
| Eukaryotic Translation Termination                                | 76           | 70               | $5.9 \times 10^{-170}$ |
| Formation of a pool of free 40S subunits                          | 87           | 74               | $9.5 \times 10^{-166}$ |
| Nonsense Mediated Decay independent of the Exon Junction Complex  | 81           | 70               | $1.2 \times 10^{-160}$ |
| L13a-mediated translational silencing of Ceruloplasmin expression | 97           | 75               | $3.8 \times 10^{-155}$ |
| 3' -UTR-mediated translational regulation                         | 97           | 75               | $3.8 \times 10^{-155}$ |
| GTP hydrolysis and joining of the 60S ribosomal subunit           | 98           | 75               | $6.0 \times 10^{-154}$ |
| Nonsense-Mediated Decay   | 96           | 73               | $5.2 \times 10^{-150}$ |
| Nonsense Mediated Decay enhanced by the Exon Junction Complex     | 96           | 73               | $5.2 \times 10^{-150}$ |
| SRP-dependent cotranslational protein targeting to membrane       | 97           | 73               | $7.8 \times 10^{-149}$ |
| Eukaryotic Translation Initiation                                 | 105          | 75               | $4.7 \times 10^{-146}$ |
| Cap-dependent Translation Initiation                              | 105          | 75               | $4.7 \times 10^{-146}$ |
| Translation   | 133          | 83               | $4.0 \times 10^{-142}$ |
| Influenza Viral RNA Transcription and Replication                 | 102          | 71               | $2.9 \times 10^{-137}$ |
| Influenza Infection   | 111          | 74               | $3.7 \times 10^{-137}$ |
| Influenza Life Cycle  | 106          | 71               | $2.3 \times 10^{-133}$ |
| Infectious disease  | 326          | 125              | $4.2 \times 10^{-120}$ |
| Extracellular matrix organisation                                 | 189          | 77               | $5.4 \times 10^{-95}$  |

| Detected only by siRNA screen (1599 genes)       | Pathway Size | Genes Identified | p-value (FDR)         |
|--|--------------|------------------|-----------------------|
| Class A/1 (Rhodopsin-like receptors)             | 282          | 44               | $1.3 \times 10^{-27}$ |
| GPCR ligand binding                              | 363          | 52               | $5.8 \times 10^{-26}$ |
| G <sub>αq</sub> signalling events                | 159          | 26               | $6.7 \times 10^{-23}$ |
| Gastrin-CREB signalling pathway via PKC and MAPK | 180          | 27               | $2.0 \times 10^{-21}$ |
| G <sub>αi</sub> signalling events                | 184          | 27               | $5.3 \times 10^{-21}$ |
| Downstream signal transduction                   | 146          | 23               | $7.6 \times 10^{-21}$ |
| Signalling by PDGF                               | 172          | 25               | $4.0 \times 10^{-20}$ |
| Peptide ligand-binding receptors                 | 175          | 25               | $8.5 \times 10^{-20}$ |
| Signalling by ERBB2                              | 146          | 22               | $1.3 \times 10^{-19}$ |
| DAP12 interactions                               | 159          | 23               | $2.6 \times 10^{-19}$ |
| DAP12 signalling                                 | 149          | 22               | $2.7 \times 10^{-19}$ |
| Organelle biogenesis and maintenance             | 264          | 33               | $5.5 \times 10^{-19}$ |
| Signalling by NGF                                | 266          | 33               | $8.2 \times 10^{-19}$ |
| Downstream signalling of activated FGFR1         | 134          | 20               | $1.1 \times 10^{-18}$ |
| Downstream signalling of activated FGFR2         | 134          | 20               | $1.1 \times 10^{-18}$ |
| Downstream signalling of activated FGFR3         | 134          | 20               | $1.1 \times 10^{-18}$ |
| Downstream signalling of activated FGFR4         | 134          | 20               | $1.1 \times 10^{-18}$ |
| Signalling by FGFR                               | 146          | 21               | $1.3 \times 10^{-18}$ |
| Signalling by FGFR1                              | 146          | 21               | $1.3 \times 10^{-18}$ |
| Signalling by FGFR2                              | 146          | 21               | $1.3 \times 10^{-18}$ |

| Intersection of SLIPT and siRNA screen (604 genes)            | Pathway Size | Genes Identified | p-value (FDR)         |
|---|--------------|------------------|-----------------------|
| Visual phototransduction                                      | 54           | 9                | $6.9 \times 10^{-10}$ |
| G <sub>αs</sub> signalling events                             | 48           | 7                | $1.6 \times 10^{-7}$  |
| Retinoid metabolism and transport                             | 24           | 5                | $1.7 \times 10^{-7}$  |
| Acyl chain remodelling of PS                                  | 10           | 3                | $6.5 \times 10^{-6}$  |
| Transcriptional regulation of white adipocyte differentiation | 51           | 6                | $6.5 \times 10^{-6}$  |
| Chemokine receptors bind chemokines                           | 22           | 4                | $6.5 \times 10^{-6}$  |
| Signalling by NOTCH4  | 11           | 3                | $6.9 \times 10^{-6}$  |
| Defective EXT2 causes exostoses 2                             | 11           | 3                | $6.9 \times 10^{-6}$  |
| Defective EXT1 causes exostoses 1, TRPS2 and CHDS             | 11           | 3                | $6.9 \times 10^{-6}$  |
| Platelet activation, signalling and aggregation               | 146          | 12               | $6.9 \times 10^{-6}$  |
| Phase 1 - Functionalisation of compounds                      | 41           | 5                | $1.3 \times 10^{-5}$  |
| Amine ligand-binding receptors                                | 13           | 3                | $1.7 \times 10^{-5}$  |
| Acyl chain remodelling of PE                                  | 14           | 3                | $2.4 \times 10^{-5}$  |
| Signalling by GPCR  | 300          | 23               | $2.4 \times 10^{-5}$  |
| Molecules associated with elastic fibres                      | 29           | 4                | $2.6 \times 10^{-5}$  |
| DAP12 interactions  | 128          | 10               | $2.6 \times 10^{-5}$  |
| Cytochrome P <sub>450</sub> - arranged by substrate type      | 30           | 4                | $3.2 \times 10^{-5}$  |
| GPCR ligand binding   | 147          | 11               | $3.8 \times 10^{-5}$  |
| Acyl chain remodelling of PC                                  | 16           | 3                | $4.0 \times 10^{-5}$  |
| Response to elevated platelet cytosolic Ca <sup>2+</sup>      | 66           | 6                | $4.2 \times 10^{-5}$  |

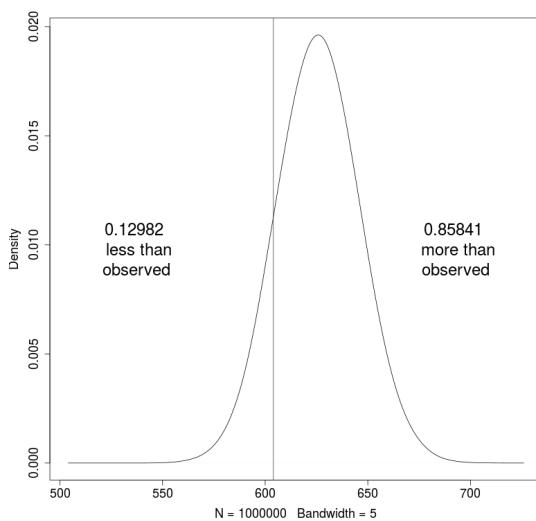
genes detected only by the siRNA screen had over-representation of cell signalling pathways, including many containing genes known to be involved in cancer (e.g., MAPK, PDGF, ERBB2, and FGFR), with the detection of Class A GPCRs supporting the independent analyses by Telford *et al.* Telford *et al.* (2015). The intersection of computational and experimental synthetic lethal partners of *CDH1* has stronger evidence for over-representation of GPCR pathways and more specific subclasses, such as visual phototransduction ( $p = 6.9 \times 10^{-10}$ ) and  $G_{\alpha s}$  signalling events ( $p = 1.7 \times 10^{-7}$ ), than other signalling pathways.

The pathway analysis for mtSLIPT against *CDH1* mutations (in Table D.4) had concordant results for both mtSLIPT-specific and siRNA-specific pathways. While the specific pathway composition of the intersection of these analyses differed from SLIPT against low *CDH1* expression, signalling pathways including GPCRs, NOTCH, EERB2, PDGF, and SCF-KIT. These findings indicate the signalling pathways are among the most suitable vulnerability to exploit in targeting *CDH1* deficient tumours as they can be detected in both a patient cohort (with TCGA expression data) and tested in a laboratory system. However, it is possible that the isolated experimental system is set up to preferentially detect kinase singalling pathways (which are amenable to pharmacological inhibition and translation to the clinic) and the other pathways identified by SLIPT may still be informative of the role of *CDH1* loss of function in cancers or mechanisms by which further gene loss leads to specific inviability.

#### 4.2.1.4.1 Resampling of genes for pathway enrichment

Comparing genes between experimental screen candidates and prediction from TCGA expression data has been less consistent than pathways. Although this is not unexpected since synthetic lethal pathways more robustly conserved (Dixon *et al.*, 2008) and the computational approach using patient samples from complex tumour microenvironment has considerably different strengths to an experimental screen (Telford *et al.*, 2015) based on genetically homogenous cell line models in an isolated laboratory environment. For instance, it is unlikely for immune signaling to be detected in an isolated cell culture system.

The overlap between synthetic lethal from bioinformatics SLIPT predictions and siRNA screening has raised other questions including whether the pathways over-represented would be expected by chance. This of particular concern since the siRNA candidate genes themselves are highly over-represented for particular pathways (such as GPCRs) so selecting any intersect with them would be enriched for these pathways.



**Figure 4.8: Resampled intersection of SLIPT and siRNA candidates.** Resampling analysis of intersect size from genes detected by SLIPT and siRNA screening approaches over 1 million replicates. The proportion of expected intersection sizes for random samples below or above the observed intersection size respectively, lacking significant over-representation or depletion of siRNA screen candidates within the SLIPT predictions for *CDH1*.

Another pathway approach is to test whether pathways are over-represented in randomly sampled genes, comparing many “resamplings” or “permutations” of these genes to the enrichment statistics observed for these pathways in the SLIPT candidates and their intersection with the siRNA hits shows whether we detect these pathways more than we expect by chance (as described in Section 2.3.6).

Of particular concern are the over-represented pathways in genes detected by both methods. Pathway over-representation alone does not detect whether SLIPT predicted genes or siRNA candidates are enriched within each other. This resampling analysis therefore detects whether over-represented pathways were detected by SLIPT independently of their over-representation among siRNA candidates (without assuming an underlying test statistic distribution).

A resampling approach is also applicable to testing whether the number of genes detected by each approach significantly intersected. As shown in Figure 4.8, resampling did not find evidence of significant depletion or over-representation for experimental synthetic lethal candidates in the computationally predicted synthetic lethal partners of *CDH1* and the overlap may be observed by chance. This is consistent with previous

findings (see Figure 4.2) and does not preclude pathway relationships being supported by resampling.

A permutation analysis was performed to resample the genes tested by both approaches to investigate whether the observed pathway over-representation could have occurred in a randomly selected sample of genes from the experimental candidates, that is, whether the pathway predictions from SLIPT could be expected by chance (as described in sections 2.2.3.1 and 2.3.6). While the number of siRNA candidate genes detected by SLIPT was not statistically significant ( $p = 0.281$ ), this may be due to the vastly different limitations of the approaches and the correlation structure of gene expression not being independent (as assumed for multiple testing procedures). The intersection may still be functionally relevant to *CDH1*-deficient cancers, such as the pathway data in Table 4.4. The resampling analysis for pathways was compared to the pathway over-representation for SLIPT predicted synthetic lethal partners in Table 4.5. Similarly, the pathway resampling for intersection between SLIPT predictions and experimental screen candidates was compared to pathway over-representation in Table 4.6 for intersection with siRNA data.

The pathway resampling approach for SLIPT-specific gene candidates (Table 4.5) replicates the gene set over-representation analysis for all SLIPT genes, detecting evidence of synthetic lethal pathways for *CDH1* in translational, immune, and cell signalling pathways including G<sub>αi</sub> signalling, GPCR downstream signalling, and chemokine receptor binding. While the immune and signal transduction pathways were not significantly over-represented in the resampling analysis, the results for the two approaches were largely consistent for translation and post-transcriptional gene regulation, supporting gene set over-representation of the SLIPT-specific pathways in Table 4.5. In particular, some of the most significantly over-represented pathways had higher observed  $\chi^2$  values than any of the 1 million random permutations. Similar pathways were also replicated by permutation analysis for mtSLIPT candidate partners against *CDH1* mutation (shown in Table D.5). This shows that many of the pathways detected specifically by SLIPT are replicated by permutation procedures and that the permutation approach is capable of detecting many of the most strongly over-represented pathways.

The permutation approach was then also applied to the intersection between computational and experimental candidates. Where the permutation analysis is testing for consistent detection of pathways independent of their pre-existing status as exper-

Table 4.5: Pathways for *CDH1* partners from SLIPT

| Reactome Pathway   | Over-representation    | Permutation              |
|--|------------------------|--------------------------|
| <b>Eukaryotic Translation Elongation</b>                           | $1.3 \times 10^{-207}$ | $< 1.241 \times 10^{-5}$ |
| Peptide chain elongation   | $5.6 \times 10^{-201}$ | $< 1.241 \times 10^{-5}$ |
| Viral mRNA Translation   | $1.2 \times 10^{-196}$ | $< 1.241 \times 10^{-5}$ |
| <b>Eukaryotic Translation Termination</b>                          | $1.2 \times 10^{-196}$ | $< 1.241 \times 10^{-5}$ |
| Formation of a pool of free 40S subunits                           | $3.7 \times 10^{-194}$ | $< 1.241 \times 10^{-5}$ |
| Nonsense Mediated Decay independent of the Exon Junction Complex   | $5.3 \times 10^{-187}$ | $< 1.241 \times 10^{-5}$ |
| L13a-mediated translational silencing of Ceruloplasmin expression  | $9.6 \times 10^{-183}$ | $< 1.241 \times 10^{-5}$ |
| 3' -UTR-mediated translational regulation                          | $9.6 \times 10^{-183}$ | $< 1.241 \times 10^{-5}$ |
| GTP hydrolysis and joining of the 60S ribosomal subunit            | $1.9 \times 10^{-181}$ | $< 1.241 \times 10^{-5}$ |
| Nonsense-Mediated Decay  | $6.2 \times 10^{-176}$ | $< 1.241 \times 10^{-5}$ |
| Nonsense Mediated Decay enhanced by the Exon Junction Complex      | $6.2 \times 10^{-176}$ | $< 1.241 \times 10^{-5}$ |
| Adaptive Immune System   | $6.5 \times 10^{-174}$ | 0.15753                  |
| <b>Eukaryotic Translation Initiation</b>                           | $5.7 \times 10^{-173}$ | $< 1.241 \times 10^{-5}$ |
| Cap-dependent Translation Initiation                               | $5.7 \times 10^{-173}$ | $< 1.241 \times 10^{-5}$ |
| <b>SRP-dependent cotranslational protein targeting to membrane</b> | $2.0 \times 10^{-171}$ | $< 1.241 \times 10^{-5}$ |
| Translation  | $6.1 \times 10^{-170}$ | $< 1.241 \times 10^{-5}$ |
| Infectious disease   | $1.6 \times 10^{-166}$ | 0.23231                  |
| <b>Influenza Infection</b>   | $1.9 \times 10^{-163}$ | $< 1.241 \times 10^{-5}$ |
| <b>Influenza Viral RNA Transcription and Replication</b>           | $1.9 \times 10^{-160}$ | $< 1.241 \times 10^{-5}$ |
| <b>Influenza Life Cycle</b>  | $2.5 \times 10^{-156}$ | $< 1.241 \times 10^{-5}$ |
| <i>Extracellular matrix organisation</i>                           | $1.1 \times 10^{-152}$ | 0.071761                 |
| GPCR ligand binding  | $1.1 \times 10^{-143}$ | 0.55801                  |
| Class A/1 (Rhodopsin-like receptors)                               | $1.5 \times 10^{-142}$ | 0.58901                  |
| <i>GPCR downstream signalling</i>                                  | $7.6 \times 10^{-140}$ | 0.098357                 |
| Haemostasis  | $1.9 \times 10^{-134}$ | 0.27059                  |
| Developmental Biology  | $2.0 \times 10^{-123}$ | 0.52737                  |
| Metabolism of lipids and lipoproteins                              | $3.3 \times 10^{-120}$ | 0.724                    |
| Cytokine Signalling in Immune system                               | $2.6 \times 10^{-119}$ | 0.39661                  |
| Peptide ligand-binding receptors                                   | $3.7 \times 10^{-109}$ | 0.61102                  |
| <b>G<sub>αs</sub> signalling events</b>                            | $8.9 \times 10^{-100}$ | $< 1.241 \times 10^{-5}$ |

Over-representation (hypergeometric test) and Permutation p-values adjusted for multiple tests across pathways (FDR). Significant pathways are marked in bold (FDR < 0.05) and italicics (FDR < 0.1).

imental candidates. The pathway results for these candidate partners (in Table 4.6) differed between over-representation and resampling analyses.

Namely, many of the over-represented pathways were not significant in the resampling analysis, including visual phototransduction and retinoic acid signalling, although pathways involving defective *EXT1* or *EXT2* genes approach significance after FDR adjustment for multiple tests. Of the highest over-represented pathways in the intersection, only G<sub>αs</sub> signalling events were supported by both over-representation and resampling analyses. Other pathways supported by both analyses were cytoplasmic elastic fibre formation, associated HS-GAG protein modification pathways, energy metabolism, and the fibrin clotting cascade.

Many of the pathways supported in the intersection by permutation analysis were also replicated in the mtSLIPT analysis of partners tested with *CDH1* mutation (in Table D.6), including G<sub>αs</sub>, elastic fibres, HS-GAG, and energy metabolism. While there were differences between the pathways Identified by over-representation analysis,

Table 4.6: Pathways for *CDH1* partners from SLIPT and siRNA primary screen

| Reactome Pathway  | Over-representation   | Permutation |
|---|-----------------------|-------------|
| Visual phototransduction  | $6.9 \times 10^{-10}$ | 0.91116     |
| <b>G<sub>as</sub> signalling events</b>                         | $1.6 \times 10^{-7}$  | 0.012988    |
| Retinoid metabolism and transport                               | $1.7 \times 10^{-7}$  | 0.20487     |
| Transcriptional regulation of white adipocyte differentiation   | $6.5 \times 10^{-6}$  | 0.38197     |
| Acylic chain remodelling of PS                                  | $6.5 \times 10^{-6}$  | 0.58485     |
| Chemokine receptors bind chemokines                             | $6.5 \times 10^{-6}$  | 0.97255     |
| <i>Defective EXT2 causes exostoses 2</i>                        | $6.9 \times 10^{-6}$  | 0.056437    |
| <i>Defective EXT1 causes exostoses 1, TRPS2 and CHDS</i>        | $6.9 \times 10^{-6}$  | 0.056437    |
| Signalling by NOTCH4  | $6.9 \times 10^{-6}$  | 0.15497     |
| Platelet activation, signalling and aggregation                 | $6.9 \times 10^{-6}$  | 0.53358     |
| Phase 1 - Functionalisation of compounds                        | $1.3 \times 10^{-5}$  | 0.24836     |
| Amine ligand-binding receptors                                  | $1.7 \times 10^{-5}$  | 0.3195      |
| Acylic chain remodelling of PE                                  | $2.4 \times 10^{-5}$  | 0.7307      |
| Signalling by GPCR  | $2.4 \times 10^{-5}$  | 0.9939      |
| <b>Molecules associated with elastic fibres</b>                 | $2.6 \times 10^{-5}$  | 0.0072929   |
| DAP12 interactions  | $2.6 \times 10^{-5}$  | 0.78273     |
| Cytochrome P <sub>450</sub> - arranged by substrate type        | $3.2 \times 10^{-5}$  | 0.87019     |
| GPCR ligand binding   | $3.8 \times 10^{-5}$  | 0.99417     |
| Acylic chain remodelling of PC                                  | $4.0 \times 10^{-5}$  | 0.65415     |
| Response to elevated platelet cytosolic Ca <sup>2+</sup>        | $4.2 \times 10^{-5}$  | 0.55461     |
| <i>Arachidonic acid metabolism</i>                              | $4.4 \times 10^{-5}$  | 0.060298    |
| Defective B4GALT7 causes EDS, progeroid type                    | $4.9 \times 10^{-5}$  | 0.15497     |
| Defective B3GAT3 causes JDSSDHD                                 | $4.9 \times 10^{-5}$  | 0.15497     |
| <b>Elastic fibre formation</b>                                  | $4.9 \times 10^{-5}$  | 0.0019227   |
| <b>HS-GAG degradation</b>                                       | $6.2 \times 10^{-5}$  | 0.017747    |
| Bile acid and bile salt metabolism                              | $6.2 \times 10^{-5}$  | 0.15497     |
| Netrin-1 signalling   | $7.1 \times 10^{-5}$  | 0.95056     |
| <b>Integration of energy metabolism</b>                         | $7.1 \times 10^{-5}$  | 0.0019287   |
| DAP12 signalling  | $7.9 \times 10^{-5}$  | 0.67835     |
| GPCR downstream signalling                                      | $8.1 \times 10^{-5}$  | 0.88678     |
| <b>Diseases associated with glycosaminoglycan metabolism</b>    | $8.7 \times 10^{-5}$  | 0.017747    |
| <b>Diseases of glycosylation</b>                                | $8.7 \times 10^{-5}$  | 0.017747    |
| Signalling by Retinoic Acid                                     | $8.7 \times 10^{-5}$  | 0.13592     |
| Signalling by Leptin  | $8.7 \times 10^{-5}$  | 0.15497     |
| Signalling by SCF-KIT   | $8.7 \times 10^{-5}$  | 0.73399     |
| Opioid Signalling   | $8.7 \times 10^{-5}$  | 0.99417     |
| Signalling by NOTCH   | 0.0001                | 0.26453     |
| Platelet homeostasis  | 0.0001                | 0.55912     |
| Signalling by NOTCH1  | 0.00011               | 0.13797     |
| Class B/2 (Secretin family receptors)                           | 0.00011               | 0.4659      |
| Diseases of Immune System                                       | 0.00013               | 0.15497     |
| Diseases associated with the TLR signalling cascade             | 0.00013               | 0.15497     |
| A tetrasaccharide linker sequence is required for GAG synthesis | 0.00013               | 0.33566     |
| Nuclear Receptor transcription pathway                          | 0.00016               | 0.22735     |
| <b>Formation of Fibrin Clot (Clotting Cascade)</b>              | 0.00016               | 0.0054639   |
| Syndecan interactions   | 0.00016               | 0.3974      |
| Class A/1 (Rhodopsin-like receptors)                            | 0.00016               | 0.99454     |
| HS-GAG biosynthesis   | 0.0002                | 0.37199     |
| Platelet degranulation  | 0.0002                | 0.39003     |
| EPH-ephrin mediated repulsion of cells                          | 0.00021               | 0.6193      |

Over-representation (hypergeometric test) and Permutation p-values adjusted for multiple tests across pathways (FDR). Significant pathways are marked in bold (FDR < 0.05) and italics (FDR < 0.1).

those replicated by permutation were highly concordant supported the combined use of these pathways approaches to identify synthetic lethal gene functions and targets.

While this indicates that  $G_{\alpha s}$  and GPCR class A/1 signalling events were significantly detected by both approaches, GPCR signalling pathways overall were not. It is likely that GPCRs were primarily over-represented in the intersection with the experimental candidates due to strong over-representation of these pathways in experimental candidates, rather than detection by SLIPT, which may be driven by these more specific constituent pathways.

However, several pathways, including some immune functions and neurotransmitters, were supported by the resampling analysis (in Tables 4.6 and D.6) when the initial pathway over-representation test was not significant. These functions appear to have been detected by both approaches more than expected by chance but must be interpreted with caution since they were still not common enough to be detected in pathway over-representation analysis.

Therefore computational and experimental approaches to synthetic lethal screening for *CDH1* lead to a broader functional characterisation and many candidate partners, when combined, despite different strengths and limitations. Compared to candidate gene approaches, experimental genome-wide screens are an appealing unbiased strategy for identifying synthetic lethal interactions. Since these screens are costly, laborious, and subject to genetic background, computational analysis can augment candidate triage to either reduce the initial panel of screened genes or prioritise validation.

GPCR pathways were detected among both computational and experimental synthetic lethal candidates, with more support in the experimental screen (Table 4.6). The homogeneous cell line model may be more likely to detect particular pathways. For instance, SLIPT identified immune pathways, not expected to be detected in isolated cell culture. GPCR signalling was supported in experimental models Telford *et al.* (2015) with some of these pathways replicated in varied genetic backgrounds of patient samples. These pathways require further investigation such as identification of more specific pathways, higher order interactions, and modes of resistance.

The pathway composition across computational and experimental synthetic lethal candidates was informative with over-representation (Table 4.4) and supported by resampling analysis (Table 4.6), despite a modest intersection of genes between them (Figure 4.2). Either approach may be significant for a pathway in this intersection without being supported by the other: resampling analysis may support pathways that were not over-represented due to small effect sizes, thus both tests are required

for a candidate pathway. The pathways detected by both over-representation and resampling are the strongest candidates for further investigation, such as  $G_{\alpha s}$  signalling, a strong candidate in prior analyses with a role in the regulation of translation in cancer Gao and Roux (2015), another function supported by SLIPT analysis.

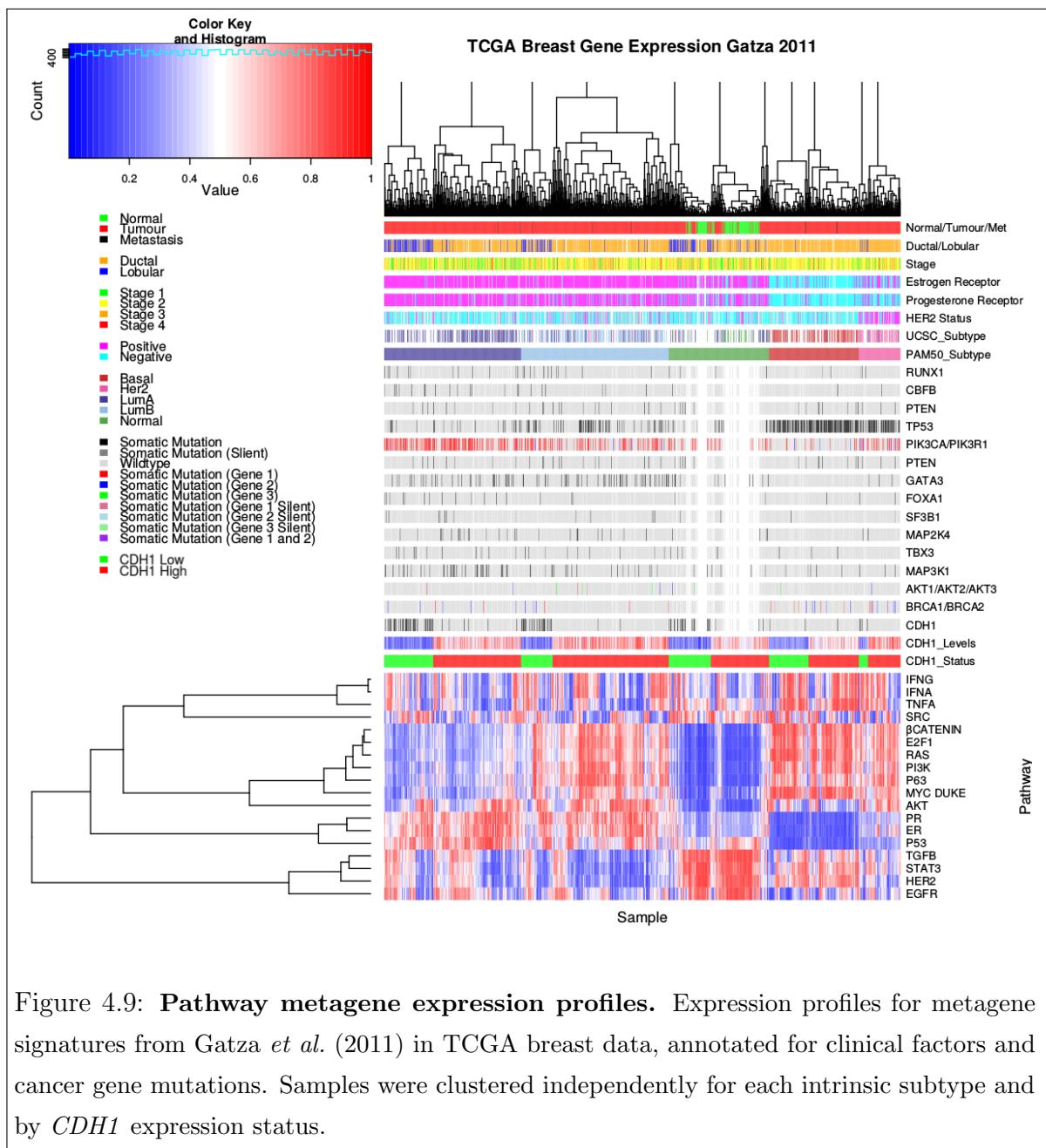
The predicted synthetic lethal partners occurred across functionally distinct pathways, including characterised functions of *CDH1*. This diversity is consistent with the wide ranging role of *CDH1* in cell-cell adhesion, cell signalling, and the cytoskeletal structure of epithelial tissues. Pathway structure may be relevant to identifying potential drug targets from gene expression signatures, indicating downstream effector genes and mechanisms leading to cell inviability. These distinct synthetic lethal gene clusters and pathways may further lead to the elucidation of drug resistance mechanisms.

## 4.3 Metagene Analysis

Metagenes serve as a consistent signal of pathway activity. The direction of metagenes is generally arbitrary but care has been taken to ensure that these occur in a direction which reflect overall activation of the pathway (as described in Section 2.2.3). This will be supported by examining the pathway expression of gene signatures in breast cancer to ensure they behave as expected in TCGA expression data. These metagenes were also compared to somatic mutation to show the limitations of mutation as a measure of gene activity. Having established that metagenes generated with this procedure reflect gene activity, these were then applied to the Reactome pathways for synthetic lethal analysis of pathways directly to provide an alternative approach to identifying synthetic lethal pathways with *CDH1*.

### 4.3.1 Pathway expression

Pathway metagenes (generated as described in Section 2.2.3) for gene signatures of key processes in breast cancer (Gatza *et al.*, 2011) were used to check that metagenes were generated in the correct direction to indicate pathway activation. These gene signatures were plotting in Figure 4.9 for comparison with clinical factors and somatic mutations. The “intrinsic subtype” was computed by performing the PAM50 procedure Parker *et al.* (2009) for RNASeq data which was highly concordant with the subtypes provided by UCSC for TCGA samples previously analysed by microarrays (TCGA, 2012). Somatic mutations were reported for recurrently mutated genes in breast cancer, as reported by TCGA (TCGA, 2012), related genes, and those previously discussed to be important in hereditary breast cancers (*BRCA1*, *BRCA2*, and *CDH1*).



These gene signatures reflect intrinsic subtypes as expected. In particular, the estrogen and progesterone receptor signatures are low in the ER– and PR– basal subtype tumours. These tumours also had the highest frequency of *TP53* mutations and a corresponding reduction of p53 metagene activity, as expected for loss of a tumour suppressor. The luminal A and luminal B tumour subtypes are the most similar, which is reflected in these metagenes signatures, although they are distinguishable molecular subtypes as shown by elevated PI3K, AKT, RAS, asn -catenin signalling in luminal B tumours. Although, these pathways were also elevated in Basal and HER2-enriched subtypes and lowly expressed in the “normal-like” subtype (which contained the normal samples). These intrinsic subtype specific gene signature profiles were further supported with metagenes for an extended set of signatures (Gatza *et al.*, 2014), as shown in Figure E.1.

*TP53* mutations were the most frequent and more common in basal subtype. Similarly, *GATA3* mutations were more common in luminal subtype tumours. PI3K mutations were more frequent across breast tumours, although these were less common in the basal subtype despite an elevated metagene (this discrepancy will be discussed further in Section 4.3.2). *CDH1* mutations similarly occurred across molecular subtypes with the exception of the basal subtype (as observed in gene expression with Figure 4.1). *CDH1* low samples occurred in all subtypes but were predominantly lobular subtype. Apart from these genes, mutations did not show clear specificity to a particular subtype and the variation between samples reflects the range of molecular cascades that can result in tumours with similar molecular profiles, supporting the use of expression for cancer diagnostics and identification of molecular targets.

The direction of the metagenes were also consistent with the clinical characteristics and formed a consensus of gene activity as shown in Figures E.2–E.5. In each of the examples for gene signatures for PI3K (Figure E.2), p53 (Figure E.3), estrogen receptor (Figure E.4), and BRCA (Figure E.5) genes (Gatza *et al.*, 2011, 2014), the expression of the majority of the genes were highly concordant with the metagene, being either positively or negatively correlated. These were generally consistent with established clinical and molecular subtypes of breast cancer and the recurrent mutations shown. However, the *PIK3CA* and *PIK3R1* mutant samples did not necessarily have elevated PI3K pathway metagene activity (as shown in Figure E.2).

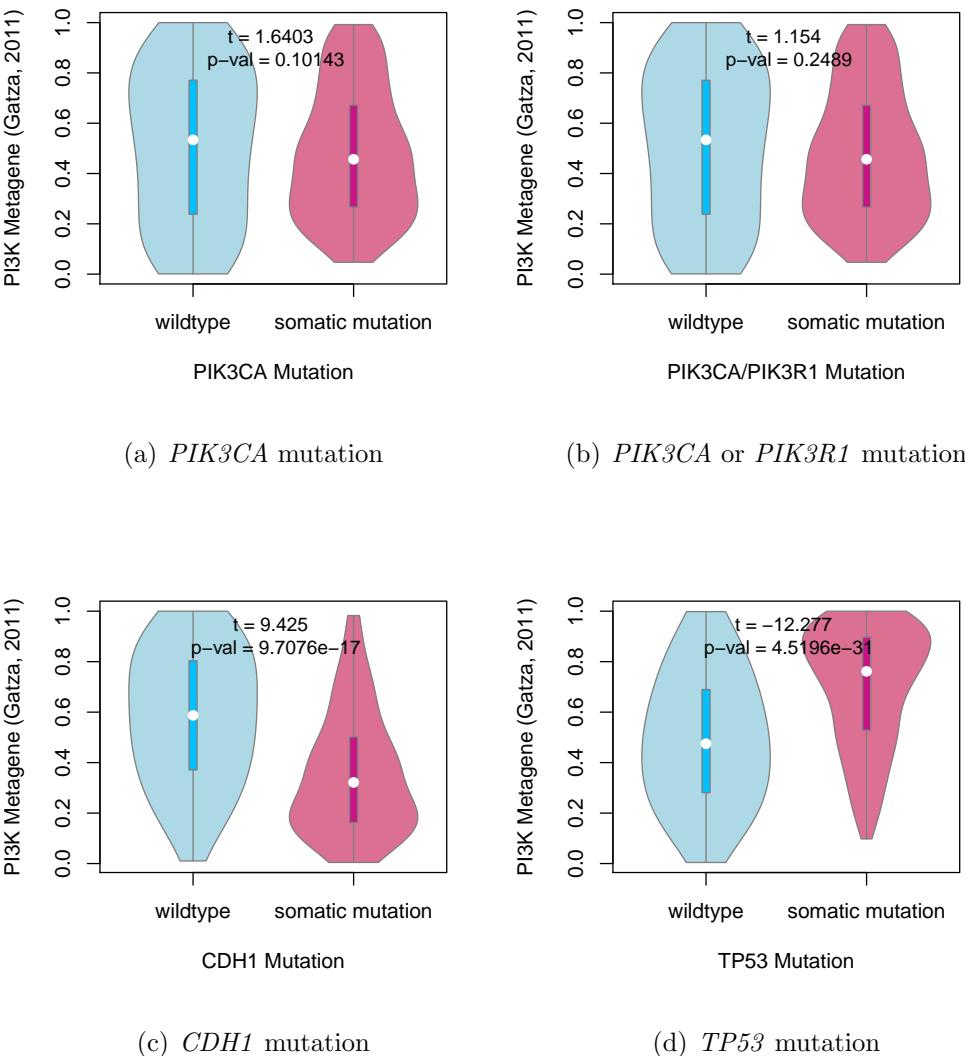


Figure 4.10: **Somatic mutation against PI3K metagene.** Mutations in *PIK3CA*, *PIK3R1*, *CDH1*, and *TP53* were examined in TCGA breast cancer for their effect on the PI3K (Gatza *et al.*, 2011) pathway metagene. The tumour suppressors *CDH1* and *TP53* showed an increase and decrease in the metagene respectively, whereas *PIK3CA* and *PIK3R1* mutations had little effect on the metagene levels.

### 4.3.2 Somatic mutation

It should be noted that metagenes, while consistent with the consensus of constituent expressed genes, were not necessarily reflecting the somatic mutation status. The PI3K (Gatza *et al.*, 2011) metagene levels in particular, were not statistically sig-

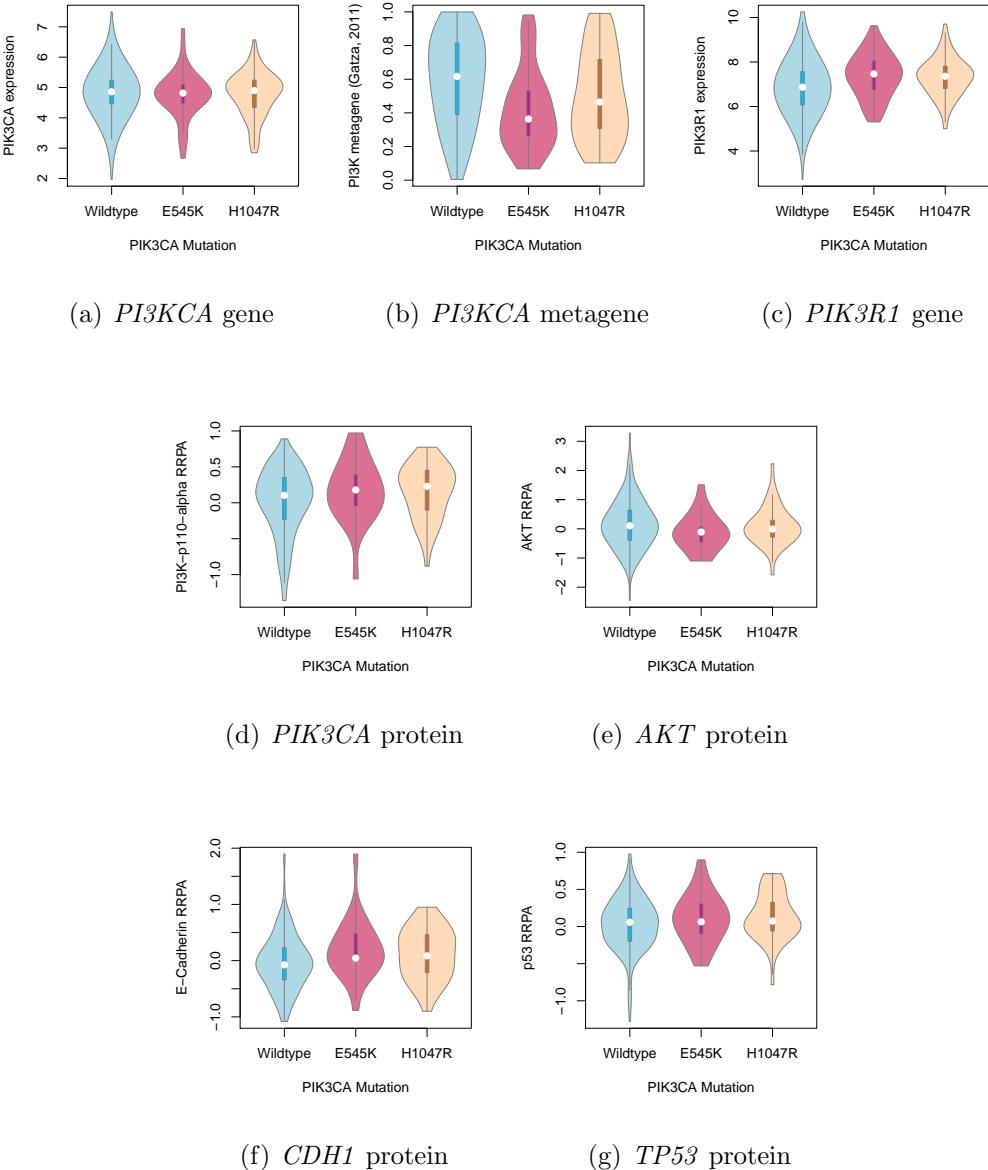
nificant in between mutant and wildtype *PIK3CA* samples (shown in Figure 4.10). Although the PI3K metagene differed across *CDH1* and *TP53* mutations, remarkably in opposite directions considering that PI3K is an oncogenic growth pathway and these are both most frequently tumour suppressors inactivated in cancers. This shows that *CDH1* and *TP53* deficient tumours have distinct molecular growth pathways and that synthetic lethal inactivations against *CDH1* inactivation may not be applicable to other cancers with driver mutations such as *TP53*, although these were kept in the analysis for comparison. These differences may be related to these mutations being more frequent in tumours with difference clinical characteristics (as observed in Section 4.3.1). Thus mutations do not necessarily have corresponding changes in pathway expression, particularly for oncogenes which may change in function rather than being upregulated.

While the more specific *PIK3CA* (Gatza *et al.*, 2014) metagene showed significant differences with *PIK3CA* and *PIK3R1* mutations (as shown in Figure D.7), this metagene replicated stronger differences for *CDH1* and *TP53*. These differences were less pronounced in the protein levels of p110 $\alpha$  (enocded by *PIK3CA*) and the downstream AKT gene (shown in Figures D.8 and D.9 respectively). Although this may be due to this regulatory cascade (kinases) being transmitted as a change in protein state (phosphorylation) rather than changes in expression levels. Another consideration is that mutations at different loci have different effects on protein function, particularly for oncogenes.

### 4.3.3 Mutation locus

The gene locus distribution of *PIK3CA* and it's receptor *PIK3R1* were consistent with oncogenic and tumour suppressor mutations, as shown in Figure D.6. *PIK3CA* is an has recurrent mutations in 2 hotspots, centered around the E545K and H1047R (shown in Figure D.6(a)), as expected for an oncogene. This contrasts with the tumour suppressors, *PIK3R1*, and *CDH1* (shown in Figures D.6(b) and D.6(c) respectively), which have low frequency inactivating mutations spread across them. A notable exception is *TP53* (shown in Figure D.6(d)) which displays both inactivating mutations throughout and recurrent (oncogenic) mutations at high frequency, consistent with the complex role of *TP53* in cancer biology which is outside of the scope of this thesis and shown for comparison.

These differences in gene locus may explain why mutations do not necessarily have corresponding changes in gene or metagene expression. Specifically, the recurrent



**Figure 4.11: Somatic mutation locus against expression.** The recurrent E545K and H1047R oncogene mutations in *PIK3CA* were examined in TCGA breast cancer to show the effect of mutation locus on gene, pathway, and protein expression. While neither of these mutations had an impact of *PIK3CA* mRNA expression, E545K had specifically lower PI3K (Gatza *et al.*, 2011) metagene levels and both mutations had higher *PIK3R1* mRNA expression. However, these differences were not reflected in the protein expression levels.

E545K and H1047R oncogene mutations in *PIK3CA* did not affect *PIK3CA* mRNA expression but E545K had specifically lower PI3K (Gatza *et al.*, 2011) metagene levels. Both mutations had higher *PIK3R1* mRNA expression but these differences differences were not reflected in the protein expression levels of p110 $\alpha$  protein (encoded by *PIK3CA*), it's downstream target AKT, E-cadherin (encoded by *CDH1*), or p53 (as shown in Figure 4.11).

While the complex effects of mutation in oncogenes such as *PIK3CA* are not necessarily detected in a pathway metagene, these do capture the consensus of pathway gene expression and account for other potential means of pathway activation. Thus metagenes are sufficient as a measure of gene activity for the purposes of synthetic lethal detection with SLIPT. This approach is more applicable to tumour suppressor genes with a relationship between gene expression and activity (rather than activation at the protein level) but this is not a major concern since synthetic lethality is more clinically relevant for targeting tumour suppressor mutations than oncogenes.

#### 4.3.4 Synthetic lethal metagenes

Pathway metagenes for Reactome pathways (generated as described in Section 2.2.3) were also used for testing synthetic lethal partner pathways with *CDH1* by SLIPT. Since the metagenes have are higher when the pathway as a whole is activated, they are amenable to SLIPT analysis using low metagene levels for inactivated pathways. These synthetic lethal metagenes differed to the over-represented pathways among synthetic lethal gene candidates. However, there were some similarities to previous findings, as shown in Tables 4.7. In particular, translational pathways were replicated as observed in Table 4.2. While the specific pathways differ, immune pathways (such as NF- $\kappa$ B) were also supported by metagene synthetic lethal analysis.

Signalling pathways were more strongly supported by mtSLIPT analysis of metagene pathway expression against *CDH1* mutation, as shown in Table D.7. Although these results were generally less statistically significant than expression analyses. Signalling pathways detected as synthetic lethal metagenes include G $\alpha$ z, insulin-related growth factor (IGF), GABA receptor, G $\alpha$ s, S6K1 and various toxin responses mediated by GPCRs. Metabolic processes including processing of carbohydrates and fatty acids were also implicated across these analyses.

The metagene analyses differ more between expression and *CDH1* mutation than previous analyses, with more specific signalling pathways identified in the mutation analysis. This supports the usage of a complete null mutant model in experimental

Table 4.7: Candidate synthetic lethal metagenes against *CDH1* from SLIPT

| Pathway  | ID      | Observed | Expected | $\chi^2$ value | p-value                | p-value (FDR)          |
|--|---------|----------|----------|----------------|------------------------|------------------------|
| Glycogen storage diseases  | 3229121 | 68       | 130      | 176            | $6.62 \times 10^{-37}$ | $1.82 \times 10^{-34}$ |
| Myoclonic epilepsy of Lafora   | 3785653 | 68       | 130      | 176            | $6.62 \times 10^{-37}$ | $1.82 \times 10^{-34}$ |
| Diseases of carbohydrate metabolism  | 5663084 | 68       | 130      | 176            | $6.62 \times 10^{-37}$ | $1.82 \times 10^{-34}$ |
| Arachidonic acid metabolism  | 2142753 | 81       | 130      | 157            | $8.13 \times 10^{-33}$ | $1.49 \times 10^{-30}$ |
| Translation initiation complex formation   | 72649   | 70       | 130      | 152            | $7.08 \times 10^{-32}$ | $1.17 \times 10^{-29}$ |
| Synthesis of 5-eicosatetraenoic acids  | 2142688 | 68       | 130      | 151            | $1.25 \times 10^{-31}$ | $1.88 \times 10^{-29}$ |
| SRP-dependent cotranslational protein targeting to membrane  | 1799339 | 69       | 130      | 150            | $2.01 \times 10^{-31}$ | $2.76 \times 10^{-29}$ |
| L13a-mediated translational silencing of Ceruloplasmin expression                                      | 156827  | 72       | 130      | 148            | $5.91 \times 10^{-31}$ | $6.44 \times 10^{-29}$ |
| 3' -UTR-mediated translational regulation  | 157279  | 72       | 130      | 148            | $5.91 \times 10^{-31}$ | $6.44 \times 10^{-29}$ |
| Activation of the mRNA upon binding of the cap-binding complex and eIFs, and subsequent binding to 43S | 72662   | 70       | 130      | 147            | $1.14 \times 10^{-30}$ | $9.28 \times 10^{-29}$ |
| Formation of the ternary complex, and subsequently, the 43S complex                                    | 72695   | 70       | 130      | 147            | $1.14 \times 10^{-30}$ | $9.28 \times 10^{-29}$ |
| Ribosomal scanning and start codon recognition   | 72702   | 70       | 130      | 147            | $1.14 \times 10^{-30}$ | $9.28 \times 10^{-29}$ |
| Eukaryotic Translation Elongation  | 156842  | 72       | 130      | 146            | $1.19 \times 10^{-30}$ | $9.28 \times 10^{-29}$ |
| Nonsense Mediated Decay independent of the Exon Junction Complex                                       | 975956  | 71       | 130      | 146            | $1.24 \times 10^{-30}$ | $9.28 \times 10^{-29}$ |
| Viral mRNA Translation   | 192823  | 70       | 130      | 146            | $1.51 \times 10^{-30}$ | $1.04 \times 10^{-28}$ |
| Eukaryotic Translation Termination   | 72764   | 70       | 130      | 146            | $1.51 \times 10^{-30}$ | $1.04 \times 10^{-28}$ |
| NF- $\kappa$ B is activated and signals survival   | 209560  | 71       | 130      | 145            | $1.90 \times 10^{-30}$ | $1.19 \times 10^{-28}$ |
| Peptide chain elongation   | 156902  | 72       | 130      | 145            | $1.91 \times 10^{-30}$ | $1.19 \times 10^{-28}$ |
| Influenza Life Cycle   | 168255  | 70       | 130      | 145            | $1.95 \times 10^{-30}$ | $1.19 \times 10^{-28}$ |
| Formation of a pool of free 40S subunits   | 72689   | 73       | 130      | 145            | $2.01 \times 10^{-30}$ | $1.19 \times 10^{-28}$ |
| Nonsense-Mediated Decay  | 927802  | 71       | 130      | 145            | $2.44 \times 10^{-30}$ | $1.34 \times 10^{-28}$ |
| Nonsense Mediated Decay enhanced by the Exon Junction Complex  | 975957  | 71       | 130      | 145            | $2.44 \times 10^{-30}$ | $1.34 \times 10^{-28}$ |
| GTP hydrolysis and joining of the 60S ribosomal subunit  | 72706   | 72       | 130      | 145            | $2.58 \times 10^{-30}$ | $1.37 \times 10^{-28}$ |
| Influenza Viral RNA Transcription and Replication  | 168273  | 72       | 130      | 144            | $4.01 \times 10^{-30}$ | $2.07 \times 10^{-28}$ |
| Signaling by NOTCH1 HD Domain Mutants in Cancer  | 2691230 | 79       | 130      | 143            | $5.99 \times 10^{-30}$ | $2.82 \times 10^{-28}$ |

Strongest candidate SL partners for *CDH1* by SLIPT with observed and expected samples with low expression of both genes

testing for synthetic lethality of signalling pathways against *CDH1* inactivation rather than a knockdown in expression. However, low expression of partners has been used in either case to be applicable to dose-dependent pharmacological inhibition and across genes where mutations have different functional consequences, including variants of unknown significance.

These results show an independent pathway approach to detecting synthetic lethal gene functions interacting with *CDH1*. Synthetic lethal metagenes, replicates support for these pathways independent of pathway size (as genes are weighted equally). The synthetic lethal analysis against low *CDH1* expression support prior findings in translational and immune pathways even if they were not able to detected in an experimental screen (Telford *et al.*, 2015). Together these findings support the role of *CDH1* loss in cancer disrupting cell signalling with wider effects on protein translation and metabolism necessary for the proliferation of cancer cells. This is consistent with the GPCR pathways such as *Gαs* signalling being supported by SLIPT gene candidates and the experimental primary siRNA screen, as shown by resampling in Section 4.2.1.4.1.

## 4.4 Replication in stomach cancer

The synthetic lethal analysis of genes and pathways (previously described for TCGA breast cancer data) was replicated in TCGA stomach cancer. The accompanying data for SLIPT and mtSLIPT analyses against *CDH1* expression and mutation are in Appendices F and G respectively.

The experimental screen (Telford *et al.*, 2015) was conducted in MCF10A breast cells so it may not be as comparable to stomach cancer. Nevertheless, *CDH1* is also important in stomach cancer biology as a driver tumour suppressor gene and as a germline mutation in many cases of hereditary diffuse gastric cancer.

While the sample size was lower for TCGA stomach cancer (particularly for mutations), these results serve to support the findings in breast cancer in an independent patient cohort and tissue samples. The molecular profiling, including RNA-Seq expression, were performed by TCGA using the sample procedures as for breast cancer and the findings reported here were performed used data analysis techniques identical to those presented previously. These procedures should ensure as close comparison as feasible across cancer types for those relevant to HDGC and recurrent *CDH1* mutations.

### 4.4.1 Synthetic Lethal Genes and Pathways

The strongest SLIPT genes for stomach cancer (shown in Table F.1) did not necessarily directly correspond to those observed in breast cancer (shown in Table 4.1). However, several gene functions were replicated in stomach cancer. Cell membrane genes including *EMP3*, *GYPC*, *LGALS1*, *PRR24*, and *FUNCD2* were among the strongest SL candidates. Similarly, cell signalling genes including *PLEKHO1*, *RARRES2*, *VEGFB*, *HSPB2*, and *CREM* were detected in stomach cancers. It is notable that several of these genes (*EMP3*, *PLEKHO1*, and *FUNCD2*) have a known role in cancer. Together these genes support the roles of *CDH1* in cell membrane and signalling functions (of epithelial tissues) which are perturbed in both breast and stomach cancers.

The strongest mtSLIPT genes tested against *CDH1* mutatoin for stomach cancer (shown in Table G.1) supported similar gene functions. Membrane and cell-adhesion genes including *KFBP6*, *THY1*, *CLELC2B*, *NISCH*, *TSPAN1*, and *KCTD12* and signalling genes including *ZEB2*, *CCND2*, *NEURL1B*, *KFBP6*, and *OGN* were detected. Similarly, these include cancer genes such as *VIM*, *ZEB2*, *BCL2*, *THY1*, and *RUNX1T1*. The mtSLIPT procedure also replicated several of the strongest candidates in breast cancer (shown in Table D.1) such as *NRIP2* and *NISCH*.

Together, these gene candidates indicate widespread functions of *CDH1* and strongly detectable synthetic lethality with many genes from a strategy that can be applied across cancer types. More specifically, the signalling genes included GPCR signalling genes (such as *GNG11*, *GNAI1*, *DZIP1*, *PTGFR*, and *KCTD12*), a growth signalling pathway which was one of the most supported synthetic lethal pathways in breast cancer analysis, the experimental screen (Telford *et al.*, 2015), and has many actionable drug targets which have been applied to other diseases.

These findings were further supported by the pathways over-represented in SLIPT candidates from TCGA stomach cancer (shown in Table 4.8) which were replicated the translational and immune pathways observed in TCGA breast cancer (shown in Table 4.2). Further support for GPCR signalling pathways including the class A/1 receptors. The extracellular matrix was also detected at the pathway level in stomach cancer SLIPT candidates and replicated in mtSLIPT analysis for *CDH1* mutation (shown in Table G.2), including elastic fibres, glycosylation, collagen, and integrin cell-surface interactions. Thus there was strong evidence for the role of extracellular matrix pathways and the tumour microenvironment in *CDH1* deficient stomach cancers, in addition to cell signalling and translation pathways important in tumour growth across breast and stomach cancer.

Table 4.8: Pathways for *CDH1* partners from SLIPT in stomach cancer

| Pathways Over-represented   | Pathway Size | SL Genes | p-value (FDR)          |
|---|--------------|----------|------------------------|
| Extracellular matrix organization                                 | 241          | 104      | $7.5 \times 10^{-140}$ |
| Hemostasis  | 445          | 138      | $1.8 \times 10^{-121}$ |
| Developmental Biology   | 432          | 125      | $9.2 \times 10^{-107}$ |
| Axon guidance   | 289          | 94       | $1.5 \times 10^{-102}$ |
| Eukaryotic Translation Termination                                | 84           | 49       | $1.9 \times 10^{-99}$  |
| GPCR ligand binding   | 373          | 108      | $3.8 \times 10^{-99}$  |
| Viral mRNA Translation  | 82           | 48       | $3.3 \times 10^{-98}$  |
| Formation of a pool of free 40S subunits                          | 94           | 51       | $3.3 \times 10^{-98}$  |
| Eukaryotic Translation Elongation                                 | 87           | 49       | $1.6 \times 10^{-97}$  |
| Peptide chain elongation  | 84           | 48       | $7.2 \times 10^{-97}$  |
| Class A/1 (Rhodopsin-like receptors)                              | 289          | 90       | $2.7 \times 10^{-96}$  |
| Nonsense Mediated Decay independent of the Exon Junction Complex  | 89           | 49       | $3.0 \times 10^{-96}$  |
| Infectious disease  | 349          | 100      | $2.6 \times 10^{-94}$  |
| GTP hydrolysis and joining of the 60S ribosomal subunit           | 105          | 52       | $3.4 \times 10^{-94}$  |
| L13a-mediated translational silencing of Ceruloplasmin expression | 104          | 51       | $2.8 \times 10^{-92}$  |
| 3' -UTR-mediated translational regulation                         | 104          | 51       | $2.8 \times 10^{-92}$  |
| Neuronal System   | 272          | 84       | $8.4 \times 10^{-92}$  |
| SRP-dependent cotranslational protein targeting to membrane       | 105          | 51       | $9.5 \times 10^{-92}$  |
| Eukaryotic Translation Initiation                                 | 112          | 52       | $2.0 \times 10^{-90}$  |
| Cap-dependent Translation Initiation                              | 112          | 52       | $2.0 \times 10^{-90}$  |

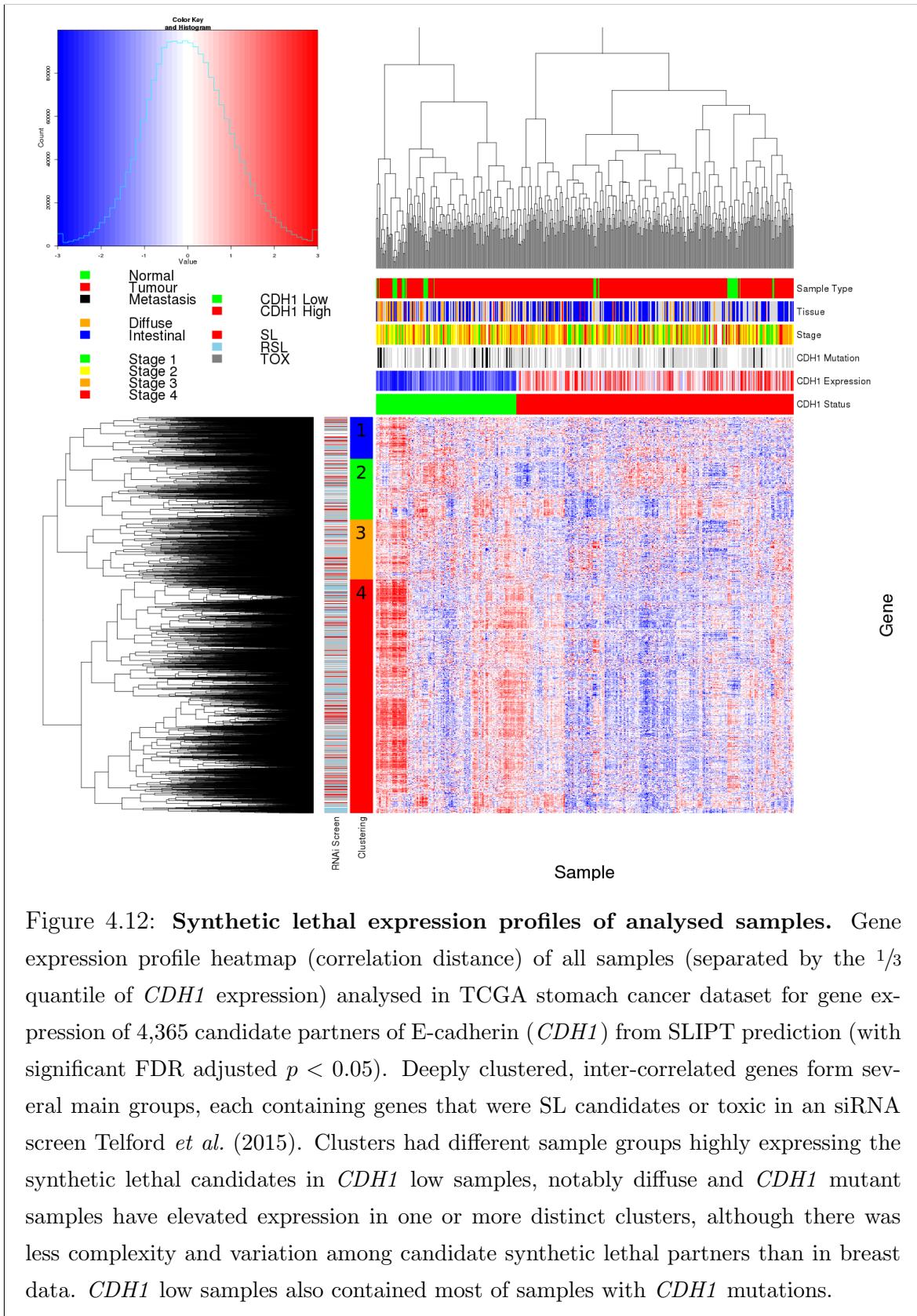
Gene set over-representation analysis (hypergeometric test) for Reactome pathways in SLIPT partners for *CDH1*

#### 4.4.2 Synthetic Lethal Expression Profiles

The expression profiles of candidate synthetic lethal partners detected by SLIPT and mtSLIPT in stomach cancer were plotted against clinical characteristics as described in section for breast cancer data in Section 4.1.2 (shown in Figures 4.12 and G.1 respectively). As expected the majority of *CDH1* mutant samples had low expression of *CDH1* and were the diffuse type of stomach cancer.

The SLIPT partners of *CDH1* exhibited similar clustering in stomach cancer to breast cancer, replicating the diverse roles of elevated partner genes in different clinical samples. Specifically (in Figure 4.12), the diffuse type stomach cancers had higher expression of the candidate synthetic lethal partners (where *CDH1* has a role as a driver mutation), despite an unbiased clustering. This is consistent with compensating expression of synthetic lethal partners under loss of *CDH1*, as suggested by Lu *et al.* (2015). The pathway composition of gene clusters for stomach cancer (shown in Table F.2) was also highly concordant with breast cancer findings (shown in Table 4.3). These included replicated of translation (Cluster 1), immune functions (Cluster 2), Gαs signalling (Cluster 3), and further support for the roles of GPCRs and the extracellular matrix (Cluster 4) in the synthetic lethal partners and functions of *CDH1*, replicated across stomach and breast cancers. Clusters 1 and 4, which had particularly high expression of SLIPT candidate partner genes in the diffuse subtype, also had the most significant over-representation of pathways.

There was less variation between the expression profiles of mtSLIPT partners of *CDH1* in stomach cancer, although clusters were still detectable (as shown in Figure G.1). While the genes and pathways detected was less significant (due to lower sample size), the composition of clusters was further indicative for the roles of extracellular matrix (including elastic fibres), immune functions, and the cell signalling.



#### 4.4.3 Comparison to Primary Screen

The number of genes detected by both SLIPT in TCGA stomach cancer data and siRNA in breast cell lines (shown in Figure F.1) was also not a significant overlap (as observed for breast cancer in Figure 4.2). This was particularly the case of mtSLIPT against *CDH1* mutation in stomach cancer which detected very few genes (as shown in Figure G.2) due to low sample size and mutation frequency.

This smaller overlap can also be attributed to the tissue-specific differences between the stomach cancers and the breast cells used for the experimental model (Chen *et al.*, 2014). Nevertheless, many genes were detected across SLIPT in stomach cancers and the experimental screen (Telford *et al.*, 2015) and the pathways detected were consistent with prior observations in breast cancer. Despite differences in the specific genes detected, the functions of *CDH1* were conserved across epithelial cancers in different tissues and synthetic lethal inhibition of interacting pathways may be effective against molecular targets such as *CDH1* inactivation across tissue types.

However, the pathway composition of SLIPT-specific genes and those replicated with the siRNA primary screen (Telford *et al.*, 2015) were highly concordant between the pathways identified by SLIPT in TCGA stomach cancer (shown in Table F.3) and pathways previously identified in TCGA breast cancer (shown in Table 4.4). In both cases, translation and immune pathways were highly over-represented in SLIPT-specific genes, which we would not expect to be detected by siRNA screening in cell lines, as discussed in Section 4.2.1.4. In addition, the extracellular matrix was supported by in stomach cancer. While the pathways identified by specifically by SLIPT in stomach cancer or siRNA screening were similar to those observed for breast cancer (in Table 4.4), the pathways over-represented in the intersection for stomach cancer SLIPT candidates and the siRNA primary screen (Telford *et al.*, 2015) also had a clear over-representation of signalling pathways, although they differed from those observed in breast cancer SLIPT candidates. GPCR signalling was supported in genes detected in both TCGA stomach cancer and screening, including  $G\alpha_q$ ,  $G\alpha_s$ , serotonin receptors, and class A signalling (shown in more detail in Table F.5). In addition MAPK and NOTCH signalling pathways were detected. These replicate the findings in breast cancer and show consistent detection of signalling pathways in stomach cancer despite less genes being detected by SLIPT and patient samples differing from the tissue in which the experiments were conducted.

Similarly, the SLIPT-specific gene candidates against *CDH1* mutation (shown in Table G.4) replicated pathways observed in breast cancer (shown in Table D.4), despite

a lower number of genes detected. In particular, the extracellular matrix and elastic fibres were over-represented. While the number of genes overlapping with the siRNA was too low to be amenable to pathway analysis, there is further indication that members of these genes replicated across mutation SLIPT analyses include cell-membrane, elastic fibre, and GPCR signalling genes.

#### 4.4.3.1 Resampling Analysis

Similarly, resampling for SLIPT specific candidates (shown in Tables F.4 and G.5) replicated many of the most highly over-represented pathways in stomach cancer. These include translational, immune, GPCR signalling, and elastic fibres, consistent with previous analyses in breast cancer (shown in Tables 4.5 and D.5).

While fewer pathways were supported by resampling for the intersection of SLIPT and experimental screen (Telford *et al.*, 2015) candidate partners in stomach cancer than breast cancer, many of those detected (shown in Table F.5) replicate those detected in breast cancer (shown in Tables 4.6 and D.6). The pathways detected by both permutation and over-representation were more likely to be replicated across stomach and breast cancer than those detected by over-representation alone, supporting the use of this procedure to detect synthetic lethal pathways applicable across cancer types. These include  $G\alpha_s$  signalling and elastic fibre formation as discussed for breast cancer (in Section 4.2.1.4.1)

While many pathways were detected by resampling for mtSLIPT against *CDH1* mutation in stomach cancer (shown in Table G.6), there were not enough genes detected by both mtSLIPT and the siRNA primary screen to determine over-represented pathways. Therefore this may be due to small numbers of genes which does not constitute support for pathway composition. However, this under-powered analysis does not preclude the replicated synthetic lethal pathways detected across SLIPT expression analyses in TCGA breast and stomach cancer data with an accompanying siRNA primary screen (Telford *et al.*, 2015). Rather this further supports the use of SLIPT to test against low expression of query genes as measure of gene inactivation to avoid this issue, despite mutation (which often produces similar results) being more indicative of complete gene inactivation.

#### 4.4.4 Metagene Analysis

Metagene analysis (as conducted in Section 4.3.4) was also performed for TCGA stomach cancer expression data, using Reactome pathways. These results (as shown in Table F.6) provided further support for signalling and extracellular processes as synthetic

lethal pathways across stomach and breast cancer. Namely, cell-cell communication, VEGF signalling, and various GPCR pathways were detected.

Signalling and immune pathways were also supported by mtSLIPT analysis of meta-gene pathway expression against *CDH1* mutation, as shown in Table G.7. Although these results were generally less statistically significant than expression analyses. Signalling pathways detected as synthetic lethal metagenes include prostacyclin, SCF-KIT, ERK, MAPK, NGF, VEGF, and PI3K/AKT. The innate immune response, the inflammasome, and integrin signaling were also implicated to be synthetic lethal with *CDH1* mutations. Cell surface interactions, cholesterol biosynthesis, and platelet homeostasis also support the role of extracellular processes in proliferation of *CDH1* deficient cancers and interactions of *CDH1* with the extracellular environment that was not tested in the cell line experimental screen.

## 4.5 Global Synthetic Lethality

Global levels of synthetic lethality were analysed to address concerns raised by the high numbers of synthetic lethal candidates for *CDH1*. The SLIPT procedure (as described in Section 3.1) was performed with each possible query gene from the TCGA breast cancer RNA-Seq dataset. Due to the computational demands of this procedure, it was performed on the New Zealand eScience Infrastructure Intel Pan supercomputer (as described in Section 2.5.3).

The observed number of SLIPT appears to be typical for most genes in the TCGA breast RNA- Seq dataset as shown in Figure 4.13. This figure was actually lower than the majority (95%) of genes tested, although *CDH1* was ranked higher for a similar in SLIPT analysis of TCGA stomach cancer data, shown in Figure H.1. The differences in sample size make these analyses difficult to compare but (in either case), the number of partners detected for *CDH1* is not unexpected, even when adjust for multiple comparisons across candidate partners.

The number of detected candidates reported here is higher than in Figures 4.2 and F.1 because these excluded genes not tested by the siRNA primary screen (Telford *et al.*, 2015) for comparison with it. For an statistically rigorous measure of global synthetic lethality, multiple comparison procedures would need to be performed for all pairs of genes tested. However, only partner genes for each query SLIPT analysis were performed for the purposes of comparing the number of partners predicted with those observed for *CDH1* throughout this thesis.

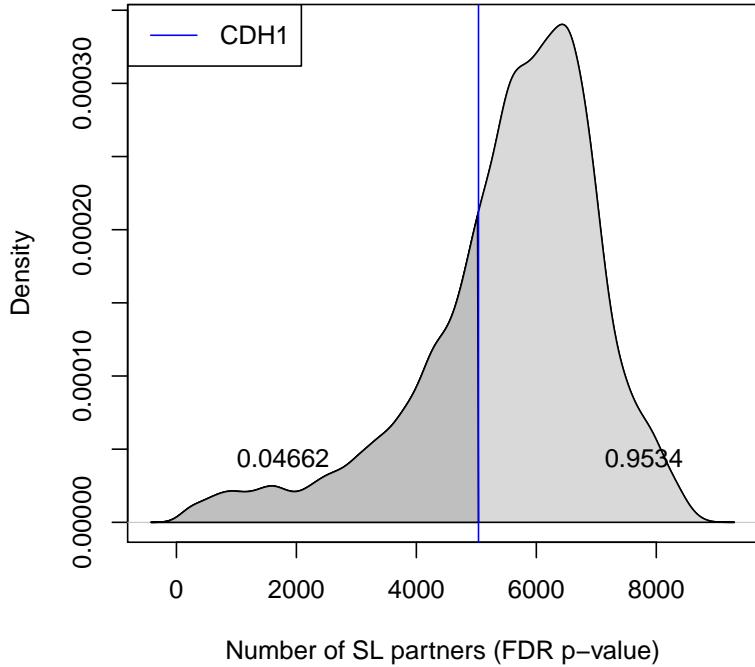


Figure 4.13: **Synthetic lethal partners across query genes.** Global synthetic lethal pairs were examined across the genome in TCGA breast expression data by applying SLIPT across query genes. The high number of predicted partners for *CDH1* was typical for a human gene and lower than many other genes.

#### 4.5.1 Hub Genes

The genes with the most synthetic lethal interactions by this SLIPT analysis are the “hub” genes of a synthetic lethal network. These genes with the highest number of candidate partners detected by SLIPT in TCGA breast cancer expression data are summarised in Table 4.9. These include several genes involved in cellular signalling such as *TGFBR2*, *PDGFRA*, *FAM126A*, *KCTD12*, *MAML2*, and *CAV1*. Gene regulation including chromatin, DNA, and RNA bindings genes were also observed as hub genes such as *CELF2*, *PLAGL1*, *TSHZ2*, *FOXO1*, and *SVEP1*. Genes involved in the cellular membrane such as *ANXA1* and *FAM171A1* were also observed in addition to genes specifically implicated in cell adhesion and tight junctions such as *TNS1*, *BOC*, *AMOTL1*, *FAT4*, and *EPB41L2*.

Table 4.9: Query synthetic lethal genes with the most SLIPT partners

| Gene            | Direction | raw p-value | p-value (FDR) | SLIPT raw p-value | SLIPT (FDR) |
|-----------------|-----------|-------------|---------------|-------------------|-------------|
| <i>TGFBR2</i>   | 8134      | 17982       | 17973         | 8007              | 8006        |
| <i>A2M</i>      | 8571      | 17605       | 17583         | 8345              | 8339        |
| <i>TNS1</i>     | 8019      | 17949       | 17934         | 7874              | 7873        |
| <i>PROS1</i>    | 8539      | 17668       | 17642         | 8317              | 8310        |
| <i>ANXA1</i>    | 9085      | 17330       | 17302         | 8689              | 8682        |
| <i>CELF2</i>    | 8665      | 17406       | 17368         | 8370              | 8355        |
| <i>BOC</i>      | 8694      | 17371       | 17348         | 8384              | 8381        |
| <i>PLAGL1</i>   | 8792      | 17361       | 17327         | 8448              | 8436        |
| <i>PDGFRA</i>   | 8296      | 17650       | 17621         | 8095              | 8087        |
| <i>FAM171A1</i> | 8874      | 17560       | 17533         | 8567              | 8562        |
| <i>FAM126A</i>  | 8510      | 17383       | 17356         | 8184              | 8178        |
| <i>TSHZ2</i>    | 7942      | 17983       | 17976         | 7787              | 7786        |
| <i>KCTD12</i>   | 8366      | 17651       | 17621         | 8115              | 8108        |
| <i>MAML2</i>    | 8336      | 17537       | 17503         | 8069              | 8061        |
| <i>FOXO1</i>    | 8027      | 17753       | 17737         | 7840              | 7836        |
| <i>AMOTL1</i>   | 8425      | 17388       | 17347         | 8147              | 8139        |
| <i>FAT4</i>     | 8111      | 17750       | 17732         | 7925              | 7919        |
| <i>CAV1</i>     | 8645      | 17491       | 17464         | 8342              | 8331        |
| <i>SVEP1</i>    | 7945      | 17859       | 17842         | 7791              | 7784        |
| <i>EPB41L2</i>  | 8415      | 17327       | 17296         | 8097              | 8092        |

Genes with the most candidate SL partners SLIPT in TCGA breast expression data with the number of partner genes predicted by direction criteria and  $\chi^2$  testing separately and combined as a SLIPT analysis.

Where specified, the p-values for the  $\chi^2$  test were adjusted for multiple tests (FDR).

Genes involved in adhesion and tight junctions were also hub genes in stomach cancer (shown in Table H.1) such as *HEG1*, *FAT4*, *NFASC*, *LAMA4*, *LAMC1*, *TNS1*, and *AMOTL1*. These also included cytoskeletal genes such as *ANK2*, *TTC28*, and *MACF1*. Cancer genes were also among hub genes across breast and stomach cancer such as *BOC*, *FAT4*, and *MRVI1*.

It is therefore unsurprising that signalling and regulatory genes have been detected throughout this thesis. Not only are they suitable targets for anti-cancer therapy, they are also highly interacting genes themselves and so it is plausible that their interactions would be detectable by SLIPT. This is consistent with the established role of aberrant signalling and gene regulation in proliferation and survival of tumours and the importance of these pathways in development with highly redundant functions across many genes under complex regulation. These are also highly amenable to detection by SLIPT analysis of expression data since their functions are dynamically regulated with corresponding changes in expression.

Cytoskeletal, membrane bound, and extracellular matrix genes are also among highly interacting synthetic lethal hubs, including focal adhesion, tight junctions, microtubules, and fibronectin. These support the use of synthetic lethal interactions to

target *CDH1*, as a tumour suppressor gene involved in these functions. Cellular structure and cell-cell interactions are thus important functions with highly redundant genes for which there are many feasible synthetic lethal interactions by which to understand regulation of cellular functions. These functions may also be exploited as vulnerabilities in cancer as they are frequently disrupted in cancers, including HDGC where loss of *CDH1* is a driver of cancer proliferation and malignancy.

#### 4.5.2 Hub Pathways

Pathways over-represented among TCGA breast cancer hub genes (as shown in Table 4.10) particularly support the importance of signalling pathways, such as the PI3K/AKT pathway, as synthetic lethal hubs. The highly redundant natures of cell-cell interaction and the extracellular matrix functions was also further supported.

Table 4.10: Pathways for genes with the most SLIPT partners

| Pathways Over-represented                                | Pathway Size | SL Genes | p-value               | p-value (FDR)         |
|--|--------------|----------|-----------------------|-----------------------|
| Constitutive Signaling by Aberrant PI3K in Cancer        | 56           | 10       | $8.4 \times 10^{-16}$ | $8.7 \times 10^{-13}$ |
| PI3K/AKT Signaling in Cancer                             | 78           | 11       | $2.1 \times 10^{-14}$ | $1.1 \times 10^{-11}$ |
| Role of LAT2/NTAL/LAB on calcium mobilization            | 96           | 12       | $7.7 \times 10^{-14}$ | $2.2 \times 10^{-11}$ |
| Complement cascade                                       | 33           | 7        | $1.2 \times 10^{-13}$ | $2.2 \times 10^{-11}$ |
| Cell surface interactions at the vascular wall           | 99           | 12       | $1.6 \times 10^{-13}$ | $2.2 \times 10^{-11}$ |
| PI3K events in ERBB4 signaling                           | 87           | 11       | $2.6 \times 10^{-13}$ | $2.2 \times 10^{-11}$ |
| PIP3 activates AKT signaling                             | 87           | 11       | $2.6 \times 10^{-13}$ | $2.2 \times 10^{-11}$ |
| PI3K events in ERBB2 signaling                           | 87           | 11       | $2.6 \times 10^{-13}$ | $2.2 \times 10^{-11}$ |
| PI-3K cascade:FGFR1                                      | 87           | 11       | $2.6 \times 10^{-13}$ | $2.2 \times 10^{-11}$ |
| PI-3K cascade:FGFR2                                      | 87           | 11       | $2.6 \times 10^{-13}$ | $2.2 \times 10^{-11}$ |
| PI-3K cascade:FGFR3                                      | 87           | 11       | $2.6 \times 10^{-13}$ | $2.2 \times 10^{-11}$ |
| PI-3K cascade:FGFR4                                      | 87           | 11       | $2.6 \times 10^{-13}$ | $2.2 \times 10^{-11}$ |
| Extracellular matrix organization                        | 238          | 22       | $4.7 \times 10^{-13}$ | $3.6 \times 10^{-11}$ |
| Muscle contraction                                       | 62           | 9        | $4.9 \times 10^{-13}$ | $3.6 \times 10^{-11}$ |
| PI3K/AKT activation                                      | 90           | 11       | $5.5 \times 10^{-13}$ | $3.8 \times 10^{-11}$ |
| GAB1 signalosome   | 91           | 11       | $7.1 \times 10^{-13}$ | $4.6 \times 10^{-11}$ |
| Smooth Muscle Contraction                                | 28           | 6        | $2.4 \times 10^{-12}$ | $1.5 \times 10^{-10}$ |
| Response to elevated platelet cytosolic Ca <sup>2+</sup> | 82           | 10       | $2.6 \times 10^{-12}$ | $1.5 \times 10^{-10}$ |
| Signaling by SCF-KIT                                     | 126          | 13       | $3.0 \times 10^{-12}$ | $1.6 \times 10^{-10}$ |
| Signaling by FGFR  | 143          | 14       | $5.0 \times 10^{-12}$ | $2.2 \times 10^{-10}$ |

Gene set over-representation analysis (hypergeometric test) for Reactome pathways in the top 500 “hub” genes with the most candidate synthetic lethal partners by SLIPT analysis of TCGA breast expression data

Pathway over-representation for synthetic lethal hub genes was replicated in TCGA stomach cancer expression data. However, these pathways differ considerably from breast cancer, as shown in Table H.2. Cell-cell interactions and extracellular matrix pathways, including elastic fibres, were also among the hub genes for stomach cancer.

The signalling pathways differ as expected in a different tissue type, although BMP and PAK signalling were detected as hub gene functions.

## 4.6 Replication in cell line encyclopaedia

As breast cancer cell lines are the experimental system in which many cancer genetics and drug targets are investigated, these were analysed in addition to patient samples from TCGA. The cancer cell line encyclopaedia (CCLE) is a resource for genomics profiles across a range of cell lines. These have also been used to generate synthetic lethal candidates for comparison to those in experimental screen and predictions from TCGA expression data.

Table 4.11: Pathways for *CDH1* partners from SLIPT in CCLE

| Pathways Over-represented            | Pathway Size | SL Genes | p-value (FDR)          |
|--------------------------------------|--------------|----------|------------------------|
| Cell Cycle                           | 442          | 207      | $1.2 \times 10^{-215}$ |
| Cell Cycle, Mitotic                  | 365          | 180      | $2.9 \times 10^{-209}$ |
| Signaling by Rho GTPases             | 311          | 136      | $9.4 \times 10^{-156}$ |
| M Phase                              | 212          | 104      | $8.8 \times 10^{-145}$ |
| Infectious disease                   | 289          | 123      | $1.3 \times 10^{-142}$ |
| RHO GTPase Effectors                 | 207          | 98       | $5.3 \times 10^{-135}$ |
| HIV Infection                        | 200          | 94       | $2 \times 10^{-130}$   |
| Separation of Sister Chromatids      | 140          | 77       | $5.6 \times 10^{-128}$ |
| Organelle biogenesis and maintenance | 258          | 107      | $1.4 \times 10^{-127}$ |
| Chromatin modifying enzymes          | 181          | 87       | $4.7 \times 10^{-126}$ |
| Chromatin organization               | 181          | 87       | $4.7 \times 10^{-126}$ |
| Mitotic Metaphase and Anaphase       | 149          | 78       | $1.2 \times 10^{-124}$ |
| Mitotic Anaphase                     | 148          | 77       | $6.3 \times 10^{-123}$ |
| Developmental Biology                | 421          | 142      | $1.6 \times 10^{-121}$ |
| RHO GTPases Activate Formins         | 94           | 60       | $5.3 \times 10^{-118}$ |
| Mitotic Prometaphase                 | 93           | 59       | $5.4 \times 10^{-116}$ |
| Hemostasis                           | 421          | 138      | $7.2 \times 10^{-116}$ |
| Adaptive Immune System               | 397          | 132      | $3.2 \times 10^{-115}$ |
| Assembly of the primary cilium       | 143          | 72       | $2.4 \times 10^{-114}$ |
| Transcription                        | 133          | 68       | $6.2 \times 10^{-111}$ |

Gene set over-representation analysis (hypergeometric test) for Reactome pathways in SLIPT partners for *CDH1*

The cancer cell line encyclopaedia provides further support for synthetic lethal genes and pathways that may be applicable across cell types and reproducible in experimental systems. In contrast to the homogeneous pooled cell samples of patients, the cell lines provide a genetically homogeneous cell population in which to examine molecular functions and as a preclinical model of cancerous disease. The complete set of 1037 cell lines was tested for synthetic lethality across tissues, in addition to the 59 breast cell lines and 38 stomach cell lines being tested separately for partners of *CDH1*. Synthetic lethal genes were detected by SLIPT (as described in Section 3.1) and over-represented synthetic lethal Reactome pathways (as described in Section 2.3.2).

Synthetic lethal gene candidates were detectable by SLIPT across each of these sample sets of cells lines (as shown in Tables I.1–I.3). Although these were most highly significant across the samples in the CCLE expression dataset (as shown in Table I.1) and included genes detected in prior analyses such as *VIM*, *ZEB2*, *EMP3*. Pathways were also highly over-represented among synthetic lethal candidates for the full CCLE dataset (as shown in Table 4.11) including Rho GTPase (GPCRs), immune, and gene regulation (chromatin and transcription). This is unexpected since immune pathways would not be expected to be detectable in isolated cell lines, although this could be attributed to cytokine and integrin signalling occurring the cancer cells in addition to interactions with immune cells in the tumour microenvironment (which could not be distinguished in patient samples). Cell cycle and mitosis were among the highest synthetic lethal pathways across cell lines supporting *CDH1* deficient cells having aberrant cell signalling and consequences for proliferation such as cancer cells. However, cell cycle genes were not as strongly supported in TCGA patient samples or the siRNA screen (Telford *et al.*, 2015) and they may not be applicable to epithelial tissues such as breast or stomach cancer or amenable to selective inhibition in experimental models.

Synthetic lethal pathways specific to SLIPT candidates from breast cell lines (as shown in Table 4.12) were more consistent with previous observations, particularly the established role of E-cadherin in cell junctions and the Adherens complex. Although the number of SLIPT candidate genes detected in stomach cell lines was insufficient to replicate the findings in breast cell lines to TCGA patient samples. However, SLIPT candidates across breast and stomach CCLE cell lines were over-represented (as shown in Table I.5) for similar pathways to breast cell lines with additional support for extracellular matrix pathways including elastic fibres which were replicated with resampling across breast and stomach TCGA analyses and the primary siRNA screen Telford *et al.* (2015).

Table 4.12: Pathways for *CDH1* partners from SLIPT in breast CCLE

| Pathways Over-represented  | Pathway Size | SL Genes | p-value (FDR) |
|--|--------------|----------|---------------|
| Cell junction organization   | 71           | 5        | 0.006         |
| Adherens junctions interactions  | 29           | 3        | 0.006         |
| Dermatan sulfate biosynthesis  | 11           | 2        | 0.006         |
| Non-integrin membrane-ECM interactions   | 52           | 4        | 0.006         |
| Regulation of pyruvate dehydrogenase (PDH) complex   | 12           | 2        | 0.0069        |
| Cell-extracellular matrix interactions   | 17           | 2        | 0.021         |
| Pyruvate metabolism  | 17           | 2        | 0.021         |
| Cell-cell junction organization  | 46           | 3        | 0.039         |
| Synthesis of substrates in N-glycan biosynthesis   | 50           | 3        | 0.057         |
| Detoxification of Reactive Oxygen Species  | 26           | 2        | 0.082         |
| Keratan sulfate biosynthesis   | 28           | 2        | 0.092         |
| Laminin interactions   | 28           | 2        | 0.092         |
| Cell-Cell communication  | 118          | 5        | 0.12          |
| Keratan sulfate/keratin metabolism   | 32           | 2        | 0.12          |
| Opioid Signalling  | 63           | 3        | 0.12          |
| Biosynthesis of the N-glycan precursor (dolichol lipid-linked oligosaccharide) and transfer to a nascent protein | 63           | 3        | 0.12          |
| Intraflagellar transport   | 34           | 2        | 0.14          |
| Signaling by Retinoic Acid   | 36           | 2        | 0.16          |
| Pyruvate metabolism and Citric Acid (TCA) cycle  | 36           | 2        | 0.16          |
| Nef mediated downregulation of MHC class I complex cell surface expression                                       | 10           | 1        | 0.22          |

Gene set over-representation analysis (hypergeometric test) for Reactome pathways in SLIPT partners for *CDH1*

## 4.7 Discussion

### 4.7.1 Strengths of the SLIPT Methodology

Synthetic lethal discovery with SLIPT used established statistical procedures identify putative partner genes from gene expression data. Such use of the  $\chi^2$ -value is amenable to pathway or permutation analyses and could feasibly be applied to other disease gene or pair-wise across the genome. Although genome-wide approaches were unable to find informative candidate genes for E-cadherin Lu *et al.* (2015). Synthetic lethal discovery in cancer has focused on genes with severe cellular mutant phenotypes, such as essential genes or the oncogenes *TP53* and *AKT* Lu *et al.* (2015); Tiong *et al.* (2014); Wang and Simon (2013), with other cancer genes, such as *CDH1*, requiring more focused investigations. Prior computational approaches for synthetic lethal discovery, in cancer, vary widely (Jerby-Arnon *et al.*, 2014; Lu *et al.*, 2015; Tiong *et al.*, 2014; Wappett *et al.*, 2016). There is no consensus for which for the approach is more appropriate and they are difficult to compare as they either do not have a released code implementation or do not make predictions solely from normalised expression data.

However, the query-based approach demonstrated by SLIPT analysis is suitable for wider application on expression data and augmenting experimental studies such as high-throughput screens. This approach has identified biologically plausible synthetic lethal pathways for *CDH1*, triaged candidates from experimental screening (Telford *et al.*, 2015), and replicates genes and pathways across breast and stomach cancers datasets. In addition, SLIPT avoids critical assumptions underlying the design of some prior approaches such as coexpression of synthetic candidates or that they will have known (annotated) similarities in function.

The DAISY methodology Jerby-Arnon *et al.* (2014), which took a similar query-based approach with the tumour suppressor *VHL*, has been critiqued for being too stringent Lu *et al.* (2015) which impedes pathway analysis. Since functional redundancy does not require genes to be expressed at the same time, the SLIPT approach does not assume co-expression of synthetic lethal genes which may enrich for synthetic lethal genes in established coregulated pathways. Rather, the interpretation of synthetic lethality for SLIPT was similar to other computational methods based on ‘co-loss under-representation’, ‘compensation’, or ‘simultaneous differential expression’ Lu *et al.* (2015); Tiong *et al.* (2014); Wang and Simon (2013).

Genomics analyses are prone to false-positives and require statistical caution, particularly where working with gene-pairs scale up the number of multiple tests drastically, at the expense of statistical power. Experimental screens for synthetic lethality are also error-prone, especially with false-positives, raising the need for understanding the expected behaviour and number of functional relationships and genetic interactions in the genome, or in discovery of synthetic lethal partners of a particular query gene. Thus analyses throughout this thesis have focused on querying for partners of a particular gene of interest. Statistical modelling and simulations (in Section 3.3 and Chapter 6) will further support the design decisions underlying SLIPT analysis and its strengths over other approaches.

#### 4.7.2 Synthetic Lethal Pathways for E-cadherin

As specific genes were difficult to replicate across experiments, gene expression profiles for synthetic lethal partners must be more complex than originally expected to directly compensate for loss of query gene or completely lack (or clearly under-representation) mutual loss (Jerby-Arnon *et al.*, 2014; Kelly, 2013; Lu *et al.*, 2015). The predicted synthetic lethal partners of *CDH1* (with FDR correction) were investigated with gene expression profiles and clinical variables to find relationships in gene expression, gene

function, and clinical characteristics. The large number of hits indicate that synthetic lethal detection is error-prone and identifying genes relevant for clinical application will be difficult without a supporting biological pathway rationale. As such, investigations into the genes identified by SLIPT, correlation structure between them, and those which were validated by experimental screening (Telford *et al.*, 2015) focused on the pathway level throughout this Chapter. Similarly, comparisons across analyses were largely made at the pathway level, including comparisons between expression and mutation, breast and stomach TCGA datasets, and patient sample data with cell line expression profiles.

Potential synthetic lethal partners of *CDH1* identified by SLIPT had many distinct functions, with each gene cluster highly expressed in different patient subgroups (Figure 4.1). The expression profiles of the SL partners of *CDH1* predicted from the TCGA breast cancer RNA-Seq data (expected to have compensating high or stable expression) and their corresponding functional enrichment found subgroups of genes with functional organisation particularly among *CDH1* low breast tumours. Ductal breast cancers show higher expression of synthetic lethal partners suggesting treatment would be more effective in this tumour subtype. However, there is consistently low expression of SL partners in estrogen receptor negative tumours, although this is independent of tumour stage and consistent with poor prognosis in these patients and could inform other treatment strategies or prevent ineffective treatment further impacting quality of life in these patients. These results suggest that synthetic lethal partner expression varies between patients; that these different tumour classes would react differently to the same treatment; that treatment of different pathways and combinations in different patients is the most effective approach to target genes compensating for *CDH1* gene loss; and the expression of synthetic partners could be a clinically important biomarker.

The pathways that synthetic lethal partners of *CDH1* identified by SLIPT were involved in a diverse range of biological functions and differed to those detected experimentally. This discrepancy may be accounted for by gene expression analyses detecting both synthetic lethal partners (as screened for experimentally Telford *et al.* (2015)) and their downstream targets (not detected by siRNA), capturing the wider pathways and mechanisms involved in synthetic lethality with *CDH1* inactivation. In particular, GPCR phosphorylation cascades (which regulates gene expression and translation in cancers Gao and Roux (2015)) were predicted to be synthetic lethal with *CDH1*. The predicted synthetic lethal partners occurred across functionally distinct pathways, including characterised functions of *CDH1*. The most consistently supported pathways

include elastic fibres in the extracellular matrix, GPCR signalling, and translation presenting vulnerabilities for *CDH1* deficient cancer cells from extracellular stimuli to the core growth mechanisms of a cell.

This diversity in synthetic lethal functions is consistent with the wide ranging role of *CDH1* in cell-cell adhesion, cell signalling, and the cytoskeletal structure of epithelial tissues. Pathway structure may be relevant to identifying potential drug targets from gene expression signatures, indicating downstream effector genes and mechanisms leading to cell inviability. Identification of distinct synthetic lethal gene clusters may further lead to the elucidation of drug resistance mechanisms. While these pathways are indicative of the main functions of E-cadherin and synthetic lethal partners, it remains to identify the genes within these pathways that are the most actionable or supported across SLIPT analysis in patient samples and detected by experiments in preclinical models (Chen *et al.*, 2014; Telford *et al.*, 2015). The specific genes within key pathways will be discussed in Chapter 5, along with further investigations into their relation to pathway structure. While these are important clinical implications, the synthetic lethal predictions lack enough confidence for direct translation into pre-clinical models or clinical applications leading to a need for statistical modelling and simulation of synthetic lethality in genomics expression data.

These synthetic lethal pathways have potential clinical implications, particularly those supported in pre-clinical models and in patient expression data. However, further validation of gene candidates will be necessary to ensure that these are able to be reproduced in further pre-clinical studies, they are applicable to tumours *in vivo*, and that effective inhibitory agents can be repurposed or designed against them.

### 4.7.3 Replication and Validation

#### 4.7.3.1 Integration with siRNA Screening

The pathway composition across computational and experimental synthetic lethal candidates was informative with over-representation (Table 4.4) and supported by resampling analysis (Table 4.6), despite a modest intersection of genes between them (Figure 4.2). Either approach may be significant for a pathway in this intersection without being supported by the other: resampling analysis may support pathways that were not over-represented due to small effect sizes, thus both tests are required for a candidate pathway.

The pathways detected by both over-representation and resampling are the strongest candidates for further investigation and the pathway structure analyses in Chapter 5

will focus on these pathways detected by both over-representation and resampling. Particularly, those replicated across datasets or with pathway metagenes. In addition to GPCR pathways detected across these analyses, the PI3K cascade will also be investigated in Chapter 5, this signalling pathway is a well characterised mediator between GPCR receptors and regulation of translation (Gao and Roux, 2015) (both detected throughout this Chapter) and exhibited unexpected behaviour with pathway the metagenes (in section 4.3). This pathway is activated by protein Phosphorylation states and thus inactivation may not be detectable with expression.

However, the SLIPT approach was shown to be predictive of which siRNA primary screen candidate partners of *CDH1* were validated in a secondary screen (as shown in Appendix C). These results further support SLIPT for identifying robust synthetic lethal candidates which can be validated and as a triage approach for interpreting screening experiments.

#### 4.7.3.2 Replication across Tissues and Cell lines

Furthermore, synthetic lethal partners identified by SLIPT were replicated across breast and stomach cancer. These were particularly concordant at the pathway level, as expected between tissues since synthetic lethal pathways have higher conservation between species (Dixon *et al.*, 2008). These findings support gene functions conserved across *CDH1* deficient cancers in breast and stomach tissues, presenting vulnerabilities that could be applied against molecular targets in both cancers. In addition, these analyses serve as a replication across independent patient cohorts from breast and stomach cancers, decreasing the likelihood of the synthetic lethal pathways detected being false positives or artifacts of either dataset.

Synthetic lethal pathways were also replicated across expression analyses of TCGA patient samples in heterogeneous tumours and homogeneous cell line isolates. This further supports that the subset of synthetic lethal functions detectable in experimental models (Chen *et al.*, 2014; Telford *et al.*, 2015) would be applicable tumours of patients with *CDH1* deficient cancers.

There are many gene functions replicated across breast cancer gene expression analyses. Many of these were also replicated with mutation analysis and with stomach cancer or cell line expression data. These pathways were more consistent across replication analyses than previous investigations with TCGA microarray data Kelly (2013).

## 4.8 Summary

We have developed a simple, interpretable, computational approach to predict synthetic lethal partners from genomics data. The analyses focus on gene expression data as it is widely available for applications in other cancers and other disease genes, particularly those with malignant loss of function.

This approach has been applied to robustly detect synthetic lethal pathways for the E-cadherin (*CDH1*) in TCGA breast cancer molecular profiles with comparisons to experimental screening (Telford *et al.*, 2015) in cell lines, and replication in TCGA stomach cancer molecular profiles and across cell types in the cancer cell line encyclopaedia. The pathway replicated across several analyses included extracellular matrix pathways (such as elastic fibres formation), cell signalling (including GPCRs), and core gene regulation and translation processes crucial for the growth and proliferation of cancer cells. These pathways show evidence of non-oncogene addiction for *CDH1* deficient cells and present vulnerabilities which may be exploited for specific treatment against *CDH1* mutations in HCGC and sporadic cancers. There was also support for synthetic lethal pathways with *CDH1* in cell adhesion and cytoskeletal processes to which *CDH1* belongs, supporting the finding that synthetic lethality occurs within biological pathways (Boone *et al.*, 2007; Kelley and Ideker, 2005).

While translational and immune pathways detected by SLIPT were not supported by primary siRNA screening (Telford *et al.*, 2015), these were replicated across various analyses. Due to the differences between an experimental cell line model (Barretina *et al.*, 2012; Chen *et al.*, 2014; Fece de la Cruz *et al.*, 2015) and patient molecular profiles (Bass *et al.*, 2014; TCGA, 2012), these would not be expected to be completely concordant. Furthermore, many pathways are difficult to test in an isolated experimental system. Nevertheless, many of the genes and pathways detected by SLIPT are suitable to inform further investigations and triage of therapeutic targets against *CDH1* deficient tumours in combination with experimental screening.

A characteristic of gene interaction networks is a scale-free topology leading to highly interacting hub genes, these represent important genes in a functional network. Cell surface interactions, the extracellular matrix, and cell signalling (particularly PI3K/AKT signalling) were also found to be synthetic lethal hubs with more interactions detected than other genes. This indicates that these pathways are functionally important to survival of cancer cells since they are subject to high functional redundancy, despite frequent disruptions in cancer. These pathways being involved in

a disproportionate number of synthetic lethal interactions is also consistent with their detection for *CDH1*.

Thus synthetic lethal pathways have been identified using TCGA patient molecular profiles, CCLE cancer cell line expression data, and experimental screening results. Some of these were robustly replicated across these datasets and against *CDH1* mutation or expression analysis. However, there remains the need to identify actionable genes within these pathways, relationships with experimental candidates, and how these pathways may affect viability when lost. While the genes identified between these analyses were less concordant the results of the TCGA breast cancer analysis will be used to test pathway structure relationships and further examine the synthetic lethal genes detected in the following Chapter.

## Aims

- Pathway Structure of Candidate Synthetic Lethal Genes for *CDH1* from TCGA breast data
- Comparisons to Experimental siRNA Screen Candidates
- Replication of Pathways across in TCGA Stomach data

## Summary

- We have developed a Synthetic Lethal detection method that generates a high number of synthetic lethal candidates
- Pathways in cell signalling, extracellular matrix, and cytoskeletal functions were supported with experimental candidates and the known functions of E-cadherin
- Several candidate pathways were supported by mutation analysis and replicated across breast and stomach cancer
- Translation and immune functions were uniquely detected by the computational approach which may be explained by differences between patient samples and cell line models
- There remains the need to identify actionable genes within these pathways, relationships with experimental candidates, and how these pathways may affect viability when lost

# Chapter 5

## Synthetic Lethal Pathway Structure

### Aims

- Synthetic Lethal Genes within a Biological Pathway Structure
- Importance and Connectivity of Synthetic Lethal Genes within Pathway Networks
- Upstream and Downstream Relationships between SLIPT and siRNA Candidates

### Summary

- Synthetic Lethal genes were explored within a graph structures for key pathways identified previously
- In some cases these graph structures appeared to have relationships between synthetic lethal genes
- However, no existing network metrics of importance and connectivity with the networks were elevated significantly for Synthetic Lethal genes
- Nor was there significant evidence of upstream and downstream relationships between SLIPT and siRNA Candidates in a shortest path permutation analysis

## **5.1 Reactome Network structure and Information Centrality as a measure of gene essentiality**

Network structure is another useful strategy to analyse gene function and this has been used to investigate network properties of a network constructed from of Reactome pathways imported with the paxtoolsr R package (Demir et al. 2010). Most notably, information centrality which has been proposed as a measure of gene essentiality was calculated as performed by Kranthi et al. (2013) using the efficiency and shortest path between each pair of nodes in the network before and after a node of interest is removed to test the importance of a node to network connectivity. Reactome contains substrates and cofactors in addition to genes or proteins, supporting the idea of centrality as a measure of essentiality, a number of nodes with the highest centrality were essential nutrients including  $Mg^{2+}$ ,  $Ca^{2+}$ ,  $Zn^{2+}$ , and  $Fe^{3+}$ .

## **5.2 Synthetic lethal genes in synthetic lethal pathways**

### **5.3 Centrality and connectivity of synthetic lethal genes**

### **5.4 Upstream or downstream synthetic lethal candidates**

### **5.5 Hierarchical approach**

### **5.6 Discussion**

### **5.7 Conclusion**

# Chapter 6

## Simulation and Modeling of Synthetic Lethal Pathways

### Aims

- A Model of Synthetic Lethal Genes in Gene Expression Data
- Simulations of Known Synthetic Lethal Genes within Pathway Networks
- Comparison of SLIPT to Alternative Approaches

### Summary

- We have designed a straight-forward rational query-based synthetic lethal detection method with the example of application to *CDH1* in cancer gene expression
- We have developed a simulation pipeline to generate continuous gene expression with pathway structure including a procedure to simulate synthetic lethality
- Our simulation procedure is robust across pathway structures and has desirable performance compared to other statistical techniques

Synthetic lethality (SL) is the death of a cell or organism with the combined loss of two non-essential genes. This phenomenon was originally used to study genetic interactions and functional redundancy in models organisms (Boone et al. 2007). While synthetic lethal experiments have been performed in *Drosophila melanogaster* (Dobzhansky 1946), *Caenorhabditis elegans* (Lehner et al. 2006), *Escherichia coli* (Butland et al.

2008), *Schizosaccharomyces pombe* (Roguev et al. 2007), and various mammalian cell lines (Kaelin 2005), the most extensive synthetic lethal screens have been performed with the synthetic gene array (SGA) technique in *Saccharomyces cerevisiae* (Boone et al. 2007; Costanzo et al. 2011; Tong et al. 2004).

Originally defined by double mutants, a range of mechanisms for gene inactivation of synthetic lethal partners can induce cell death including RNA interference and drug treatment where it is sometimes called induced essentiality or non-oncogene addiction in cancer research (Fece de la Cruz et al. 2015). Cellular viability is the main means to measure synthetic lethal effects experimentally because it is quantified and measured consistently (as shown in Figure 1), whereas qualitative measures of impaired organism viability are ambiguous and less relevant to yeast or cancer research.

The cancer genetics laboratory are currently working on developing a synthetic lethal approach to target the tumour suppressor gene CDH1 which has been found to cause predispose early-onset breast and stomach cancers in mutation carriers, including families of New Zealand Māori (Berx et al. 1995; Guilford et al. 1998). These families are currently closely monitored and offered drastic preventative surgery. If it were developed, a drug selective against CDH1 mutant tumours would serve not only as a chemopreventative alternative for these families but also benefit the wider community as a treatment for sporadic cases of CDH1 mutant cancer. To augment experimental work on CDH1 with isogenic cell lines (Telford et al. 2015), a computational methodology is explored here to exploit public cancer genomic databases.

Microarray and massively-parallel sequencing technologies are driving a revolution in molecular biological research, particularly with regard to cancer where the premise of genomic medicine is rapidly becoming feasible with the use of genomics to identify cancer genes, diagnose patients with actionable mutations, and use gene expression as a prognostic marker. Genomic data could also be used to identify novel drug targets and synthetic lethal partners of known cancer genes in particular. The Cancer Genome Atlas database (TCGA) and the overarching International Cancer Genome Consortium (ICGC) provide a valuable public cancer genome data resource because they support many different data types for the same samples, for many different cancer types, and for high sample sizes (Cancer Genome Atlas Research Network 2014; Cancer Genome Atlas Research Network et al. 2013; International Cancer Genome Consortium 2014). They host data of patient clinical factors, gene expression, somatic mutation, DNA copy number, and DNA methylation which could all serve to predict synthetic lethality from frequency of mutually exclusive gene inactivation and its impact on patient survival. A

number of other databases are given in the Table 6 which may be used to explore gene function, drug target feasibility, or replicate analyses but TCGA and ICGC datasets will be the focus of this project.

There is a growing need for a robust approach to cost-effective prediction of candidate synthetic lethal interaction, particularly in cancer research. Exploiting existing public genomic databases is an ideal way to utilise existing resources with suitable sample sizes, data types, and different limitations to those of laboratory experiments. A number of computation approaches to synthetic lethality have been developed but many of these rely on data not available to cancer researchers, methods that are difficult to replicate, over-fitted to a particular dataset, having mixed validation results, or do not have a software tool accessible to the research community. These methodologies are reviewed in detail in the accompanying literature review. They will still be considered to develop an improved synthetic lethal interaction prediction tool (SLIPT).

A bioinformatics approach has distinct limitations to experimental methods and would work well combined with genetic screen data and conventional molecular biology laboratory validation techniques to answer biological research questions. Compared with an experimental screen, a bioinformatics approach has the benefits of reduced costs, with the potential for automation, scaling up, and replication of the same gene across populations and cell types. Analysis of public genomic data accounts for real tumour variation showing detection with tumour heterogeneity and genomic instability. Compared with a cell line or xenograft experimental model we are limited by difficulties in establishing validity of a novel method, lack of mechanism, or potential for testing drug activity in the same system. However, computational methods may further miss useful therapeutic candidates from variable genetic background and be limited by the population sampled. This research builds on previous work in an Honours project and similar approaches in the literature (Jerby-Arnon et al. 2014; Kelly 2013; Lu et al. 2015).

## **6.1 Simulations and Modelling Synthetic Lethality in Expression Data**

Synthetic lethality was modelled for effects on expression levels and whether these are detectable in known interacting and non-interacting genes in simulated data. These were conducted for expression data but the nature of these simulations would be relevant to how synthetic lethality would manifest in other factors, particularly DNA copy number variation and DNA methylation. These simulations were discussed at length

in the previous meeting and showed that synthetic lethality was detectable with our approach in simple cases. While it was less effective, the methods were able to detect synthetic lethal genes in expression data with correlation structure (generated with the multi-variate normal distribution) and were distinguishable from correlated genes. Therefore the strongest (most significant) synthetic lethal genes are more likely to be true synthetic lethal partners and a high number of hits are expected from correlated genes and co-regulated pathways.

The power of the method to detect interactions depleted with increasing multiple tests, interactions, and cryptic (third party) interacting partners. Increased sample size counteracted these effects as expected. This led the idea that pathways would be more suitable as the focus of this project. Biological pathways led to fewer multiple tests, more relevant to understanding cancer biology, and are often drug targets in practice.

## **6.2 Simulations over simple graph structures**

### **6.2.1 Performance**

### **6.2.2 Synthetic lethality across graph stuctures**

### **6.2.3 Performance with inhibition links**

### **6.2.4 Performance with 20,000 genes**

## **6.3 Simulations over pathway-based graphs**

## **6.4 Comparing methods**

### **6.4.1 SLIPT and Chi-Squared**

#### **6.4.1.1 Correlated query genes**

#### **6.4.2 Correlation**

#### **6.4.3 Bimodality with BiSEp**

# Chapter 7

## Discussion

### Aims

- To develop a statistical approach to detect synthetic lethal gene pairs in cancer from expression data
- To apply this methodology to public cancer gene expression data against *CDH1* and analyse pathway structure with comparisons to experimental screen data
- To construct a statistical model of synthetic lethality in multivariate normal expression data
- To develop a simulation pipeline of expression with pathway structure on a high-performance computing cluster
- To examine the statistical performance of the methodology with simulated expression including pathways and compare it to other approaches
- To release the synthetic lethal detection methodology and pathway simulation procedure as R software packages

### Summary

- We have developed a Synthetic Lethal detection method that generates a high number of synthetic lethal candidates

- Pathways in cell signalling, extracellular matrix, and cytoskeletal functions were supported with experimental candidates and the known functions of E-cadherin
- Several candidate pathways were supported by mutation analysis and replicated across breast and stomach cancer
- Translation and immune functions were uniquely detected by the computational approach which may be explained by differences between patient samples and cell line models
- There remains the need to identify actionable genes within these pathways, relationships with experimental candidates, and how these pathways may affect viability when lost
- Synthetic Lethal genes were explored within a graph structures for key pathways identified previously
- In some cases these graph structures appeared to have relationships between synthetic lethal genes
- However, no existing network metrics of importance and connectivity with the networks were elevated significantly for Synthetic Lethal genes
- Nor was there significant evidence of upstream and downstream relationships between SLIPT and siRNA Candidates in a shortest path permutation analysis
- We have designed a straight-forward rational query-based synthetic lethal detection method with the example of application to *CDH1* in cancer gene expression
- We have developed a simulation pipeline to generate continuous gene expression with pathway structure including a procedure to simulate synthetic lethality
- Our simulation procedure is robust across pathway structures and has desirable performance compared to other statistical techniques

## 7.1 Significance

Development of an effective synthetic lethal discovery tool for bioinformatics analysis has a wide range of applications in genetics research including functional genomics, medical and agricultural applications. Of particular interest is a complementary approach to discovery of synthetic lethal drug targets for cancer therapy to aid the cancer research community which currently relies on cell line and mouse models for screening and validation experiments (Fece de la Cruz et al. 2015). The potential for synthetic lethal drug design against cancer mutations including gene loss or overexpression could lead to a revolution in cancer therapy and chemoprevention with personalised treatment of cancers and high risk individuals. Examples of the synthetic lethal strategy to cancer treatment have been shown to be clinically effective with many large-scale RNAi screens underway to discover more cancer gene function and drug targets for similar application.

However, there are limitations to both experimental screens and computational approaches, both known to be prone to false-positives. Modelling and simulation of synthetic lethal discovery in genomic data has been explored to address these concerns and ensure the validity of candidate synthetic lethal interactions, particularly given the recent emergence of a number of conflicting synthetic lethal screening and prediction approaches. Exploring synthetic lethality in simulated data will ensure the optimal performance of our prediction method with comparison to the distribution of test statistic distribution in empirical gene expression data, informed selection of thresholds for prediction, and estimated error rates. The model of gene expression with known synthetic lethal genes is limited by the assumption that it represents the distribution of gene expression when it may not. Having shown synthetic lethality is detectable in simple models and added correlation structure, the model still needs to be developed to better represent real data. However, the behaviour of synthetic lethal genes and effects of parameters explored so far remains important to inform future model design and interpretation of empirical data analysis. The synthetic lethal discovery strategy could be adapted to any form of gene inactivation or disruption such as changes to gene expression, regulation, epigenetics, DNA sequence, or copy number which could plausibly induce cell death due to SL interactions. Further applications of synthetic lethal interactions such as analysis of gene networks, tissue specificity, evolutionary conservation, or drug target feasibility are possible with synthetic lethal candidates predicted with confidence on a large scale.

Network analysis enables properties of the network and its connectivity to be measured and compared across datasets (Barabsi & Oltvai 2004). Tissue specificity is an important consideration, largely unexplored with synthetic lethal studies, since it has clinical importance to ensure targeted drug treatments are effective, predict adverse effects in other tissues, determine whether targeted treatments could be repurposed for other cancer types or diseases, and whether drug resistance mechanisms could emerge. Comparison of tissues, populations, and species can all ensure that synthetic lethal predictions are robust, that experimental candidates are clinically relevant, and treatments designed to exploit them would be specific to the disease in large patient cohort (with known biomarkers).

Drug targets must be feasible to have effective anti-cancer interventions designed against them, which raises the need for targets with existing drugs in the clinic, trials, or feasible to development with structural analysis or screening. Druggable targets could be selected by gene functions known to be amendable to drugs, with a structure amenable with development, with conserved specific sites without homology to other genes, or with known approval or developing drugs which could be repurposed from other disease applications.

## 7.2 Future Directions

Such a bioinformatically-informed synthetic lethal screening and validation strategy could be integrated into existing and future screens for synthetic lethality in cancer.

Possible improvements to the SLIPT method include developing a Bayesian inference method or simulations and modelling to account for pathway structure among synthetic lethal genes. Another extension would be to test for higher order synthetic lethal interactions, where 3 or more genes perform a redundant function.

Further development of the synthetic lethal model and simulation is needed to explore the parameters, ensure relevance to empirical data analysis, and understanding the implications of findings so far. An example of more complex correlation structure is shown in supplementary Figures S1 and S2 with genes correlated to the Query genes (showing need for directional synthetic lethal condition) and correlated with other non-synthetic lethal genes (showing the predictions are robust to other correlation structure). The impact of these modifications on model performance in a large number of genes or simulation replicates is yet to be seen or whether such correlation structure reflects the correlation structure of empirical data (as shown in Figure 3 with the row dendrogram for correlation distance between genes), known biological pathways, or

known synthetic lethal interactions. Correlation between synthetic lethal genes could also be considered.

Comparing the findings of modelling and simulation with public gene expression analysis and experimental screen targets is still needed to identify putative synthetic lethal interactions. This application will be tested with the example of CDH1 as a query gene in breast cancer for follow up to earlier results, relevance to ongoing research in the Cancer genetics Laboratory, and comparison to the experimental screen data of MCF10A cells by Telford et al. (2015). While this methodology is intended to be widely applicable, particularly to other cancer genes and will be made available to the research community (manuscript and code release in preparation).

There are several avenues for further research on synthetic lethality in breast cancer. The main alternative themes are network analysis with a focus on tissue specificity or drug feasibility with an emphasis on pharmacogenomics, biological pathways, and whether candidate targets could be inactivated by compounds with favourable pharmacokinetic properties. Either approach remains within the scope of the project, although each will require adoption of new computational tools, which is important topic for consideration in the meeting and changes to the project direction later in the year.

### 7.3 Conclusion

Synthetic lethal interactions are important for understanding gene function and development of targeted anti-cancer treatments. Synthetic lethal discovery with experimental screening is error prone and limited by the model systems in which it is performed. A bioinformatics tool to predict synthetic lethal interactions from genomics data would greatly benefit the cancer research community (and wider genetics research community). Several such tools exist, including one we have developed, but they have conflicting design and results are often inconsistent with experimental screen data. Therefore, modelling and simulation of synthetic lethality in gene expression data is needed to ensure the statistical validity of predictions. We have developed a model with correlation structure based on a Multivariate Normal distribution for which simulations detect synthetic lethality with high performance in simple cases and which has the potential to be developed to model complex correlation structure, biological pathways, or patterns observed in empirical gene expression data. The modelling, public data analysis, and experimental screen data approaches will be combined to further examine the example of CDH1 in breast cancer. Analysis of gene networks, tissue specificity,

biological pathways, or drug targets remain options to explore tool development and implications for synthetic lethal cancer research in the future.

# **Chapter 8**

## **Conclusion**

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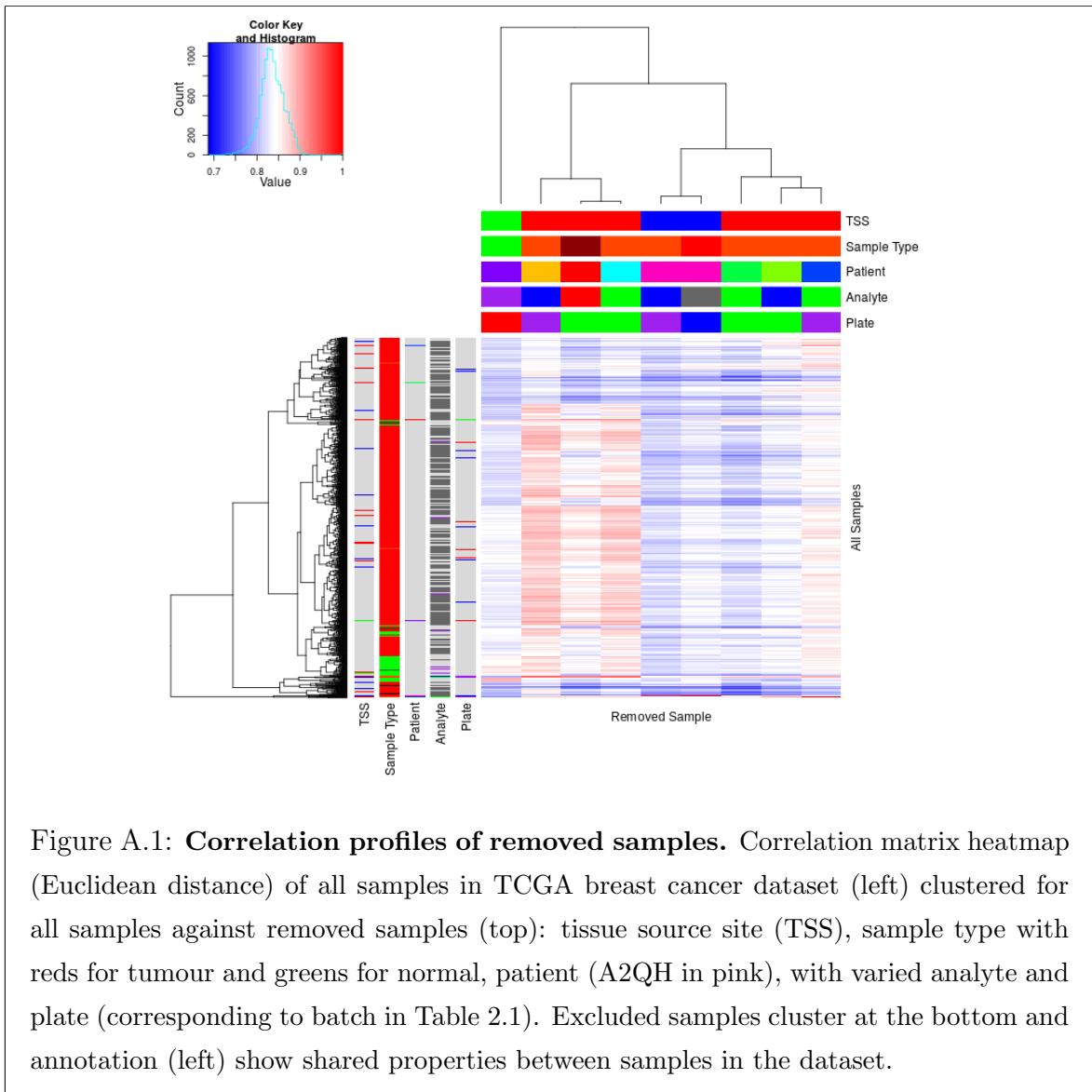
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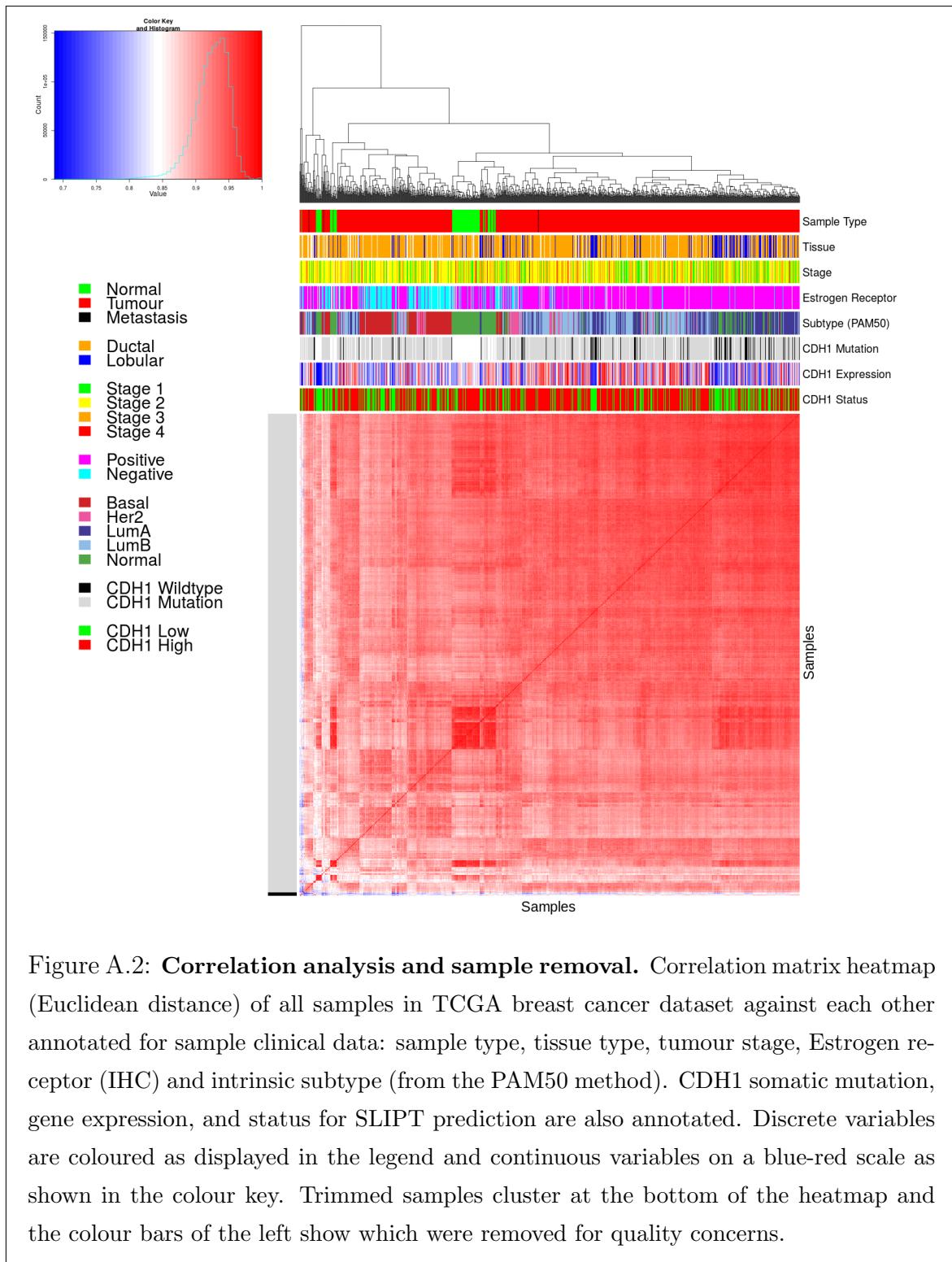
# **Appendix A**

## **Sample Quality**

### **A.1 Sample Correlation**

Samples were excluded from expression analysis based on sample correlations and the clustering analysis presented below, as described in Section 2.2.2.





## A.2 Replicate Samples in TCGA Breast

Replicate samples were picked where possible from the TCGA breast cancer gene expression data to examine for sample quality. Independent samples of the same tumour are expected to have very high Pearson's correlation between their expression profiles unless there were issues with sample collection or preparation and are thus an indicator of sample quality. The log-transformed raw read counts for replicate samples were examined in Figures A.3–A.5. These were examined before normalisation which would be expected to increase sample concordance.

Another consideration are the samples which were removed for quality concerns (in Section 2.2.2). While these were selected by unbiased hierarchical clustering (See Figure A.2), it is notable that many of the excluded (tumour) samples were performed in replicate despite relatively few replicate samples in the overall dataset. These samples correlate poorly with the rest of the dataset, in addition to with replicate samples.

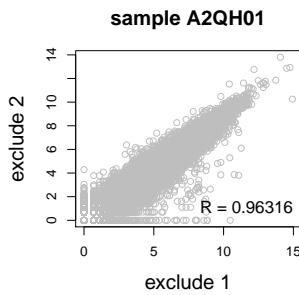
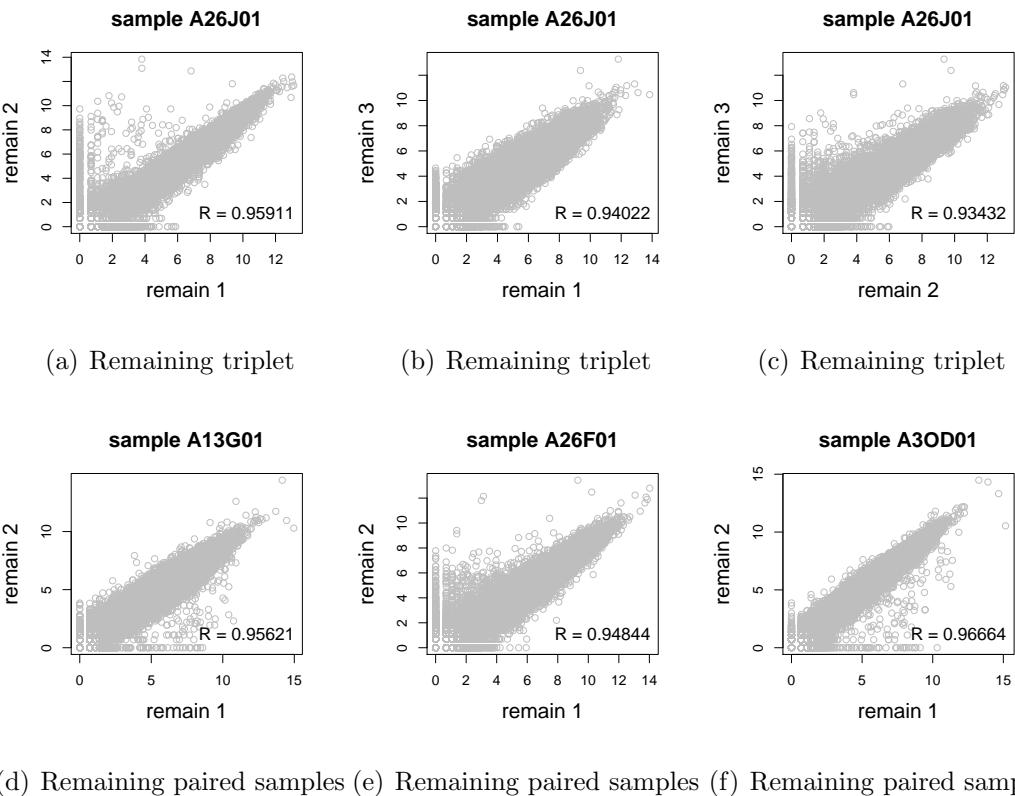
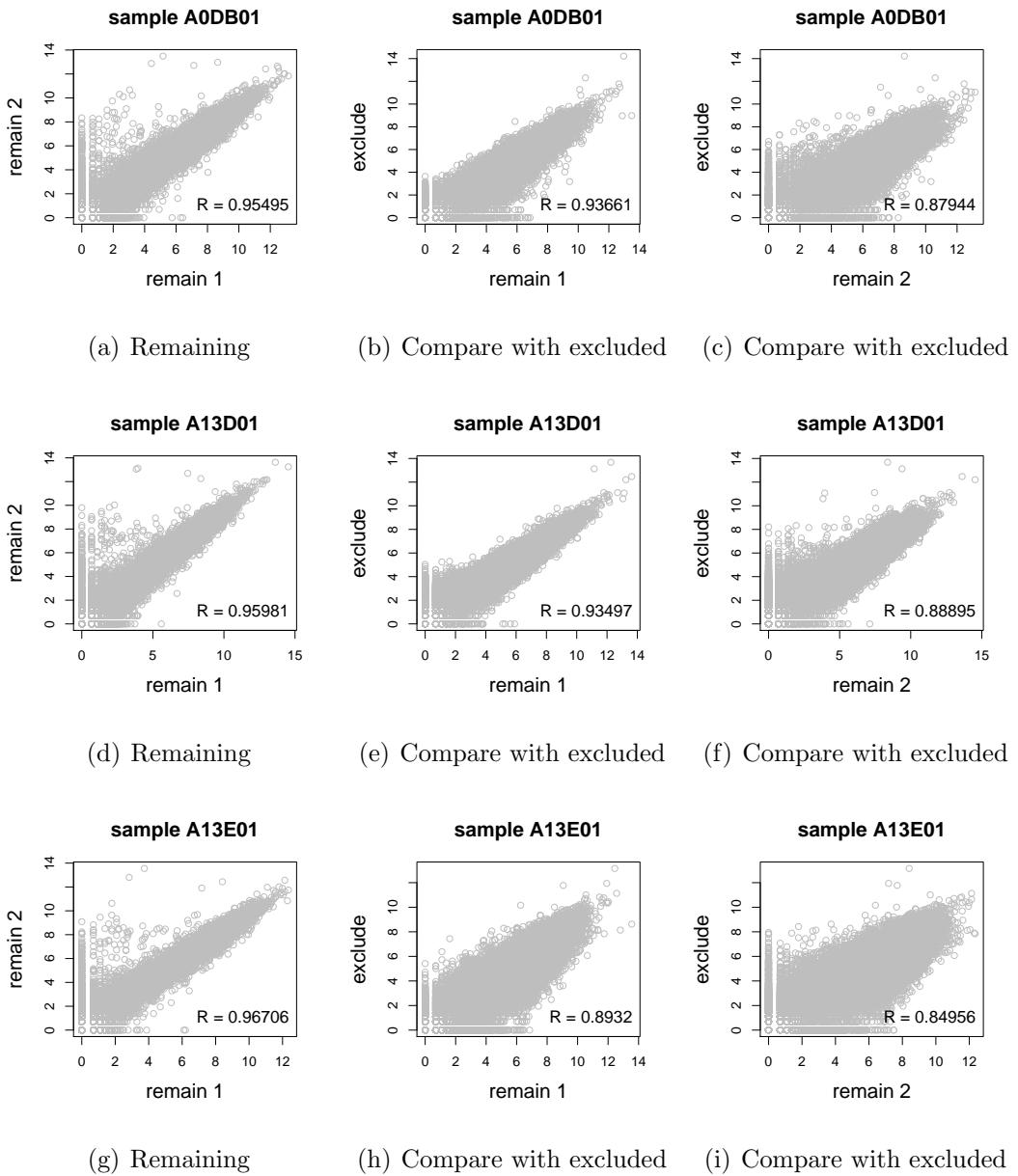


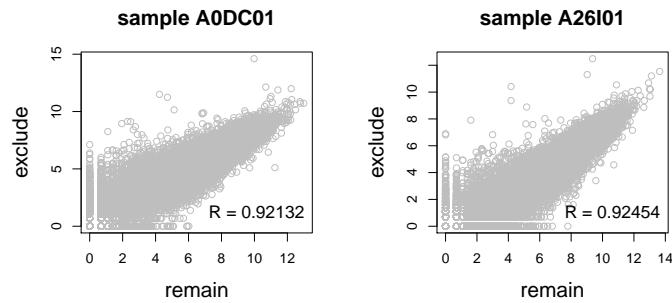
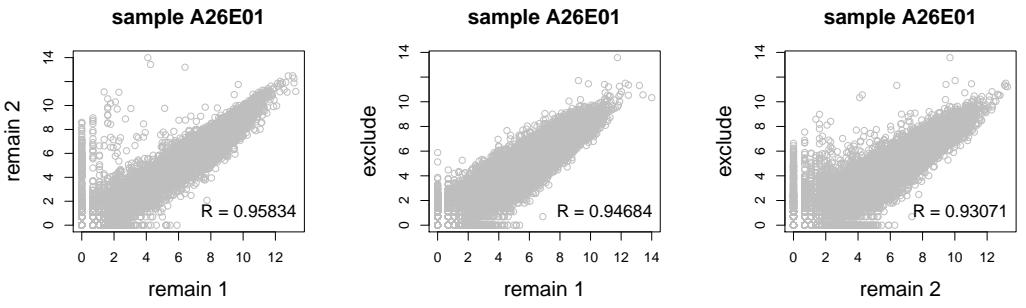
Figure A.3: **Replicate excluded samples.** Both tumour samples of patient A2QH were excluded as they were poorly correlated with other samples, although they are highly similar to each other as shown by Pearson's correlation of log-raw counts.



**Figure A.4: Replicate samples with all remaining.** Patient A26J was sampled 3 times and compared pairwise. Pairs of samples were also compared for other patients with replicate samples. In all cases, replicate samples remaining in the dataset were highly concordant as shown by Pearson's correlation of log-raw counts.



**Figure A.5: Replicate samples with some excluded.** Patients A0DB, A13D, A13E, and A26E were each sampled 3 times and compared pairwise. Pairs of samples were also compared for other patients with replicate samples. In all cases, the replicate samples remaining in the dataset more were highly concordant (as shown by Pearson's correlation of log-log counts) than those excluded from the analysis.



**Figure A.5: Replicate samples with some excluded.** Patients A0DB, A13D, A13E, and A26E were each sampled 3 times and compared pairwise. Pairs of samples were also compared for other patients with replicate samples. In all cases, the replicate samples remaining in the dataset more were highly concordant (as shown by Pearson's correlation of log-raw counts) than those excluded from the analysis.

# Appendix B

## Software Used for Thesis

Table B.1: R Packages used during Thesis

| Package       | Repository   | Laptop   | Lab      | Server   | NeSI     |
|---------------|--------------|----------|----------|----------|----------|
| base          | base         | 3.3.2    | 3.3.2    | 3.3.1    | 3.3.0    |
| abind         | CRAN         |          | 1.4-5    |          | 1.4-3    |
| acepack       | CRAN         |          | 1.4.1    |          | 1.3-3.3  |
| ade4          | CRAN         |          | 1.7-5    |          |          |
| annaffy       | Bioconductor |          | 1.46.0   |          |          |
| AnnotationDbi | Bioconductor |          | 1.36.0   | 1.36.0   | 1.34.4   |
| apComplex     | CRAN         |          | 2.40.0   |          |          |
| ape           | CRAN         |          | 4        |          | 3.4      |
| arm           | CRAN         |          | 1.9-3    |          |          |
| assertthat    | CRAN         | 0.1      | 0.1      | 0.1      | 0.1      |
| backports     | CRAN         | 1.0.5    | 1.0.4    | 1.0.5    | 1.0.2    |
| base64        | CRAN         |          |          | 2        | 2        |
| base64enc     | CRAN         |          | 0.1-3    |          | 0.1-3    |
| beanplot      | CRAN         |          | 1.2      | 1.2      | 1.2      |
| BH            | CRAN         | 1.60.0-2 | 1.62.0-1 | 1.62.0-1 | 1.60.0-2 |
| Biobase       | Bioconductor |          | 2.34.0   | 2.34.0   | 2.32.0   |
| BiocGenerics  | Bioconductor |          | 0.20.0   | 0.20.0   | 0.18.0   |
| BiocInstaller | Bioconductor |          | 1.24.0   | 1.20.3   | 1.22.3   |
| BiocParallel  | Bioconductor |          | 1.8.1    | 1.8.1    |          |
| Biostrings    | Bioconductor |          | 2.42.1   | 2.42.0   |          |
| BiSEp         | Bioconductor |          | 2.0.1    | 2.0.1    | 2.0.1    |

|            |      |        |         |        |        |
|------------|------|--------|---------|--------|--------|
| bitops     | CRAN | 1.0-6  | 1.0-6   | 1.0-6  | 1.0-6  |
| boot       | base | 1.3-18 | 1.3-18  | 1.3-18 | 1.3-18 |
| brew       | CRAN | 1.0-6  | 1.0-6   | 1.0-6  | 1.0-6  |
| broom      | CRAN | 0.4.1  |         |        |        |
| caTools    | CRAN | 1.17.1 | 1.17.1  | 1.17.1 | 1.17.1 |
| cgdssr     | CRAN |        | 1.2.5   |        |        |
| checkmate  | CRAN |        | 1.8.2   |        | 1.7.4  |
| chron      | CRAN | 2.3-47 | 2.3-48  | 2.3-50 | 2.3-47 |
| class      | base | 7.3-14 | 7.3-14  | 7.3-14 | 7.3-14 |
| cluster    | base | 2.0.5  | 2.0.5   | 2.0.5  | 2.0.4  |
| coda       | CRAN |        | 0.19-1  |        | 0.18-1 |
| codetools  | base | 0.2-15 | 0.2-15  | 0.2-15 | 0.2-14 |
| colorRamps | CRAN |        | 2.3     |        |        |
| colorspace | CRAN | 1.2-6  | 1.3-2   | 1.3-2  | 1.2-6  |
| commonmark | CRAN | 1.1    |         | 1.2    |        |
| compiler   | base | 3.3.2  | 3.3.2   | 3.3.1  | 3.3.0  |
| corpcor    | CRAN |        | 1.6.8   | 1.6.8  | 1.6.8  |
| Cprob      | CRAN |        | 1.2.4   |        |        |
| crayon     | CRAN | 1.3.2  | 1.3.2   | 1.3.2  | 1.3.2  |
| crop       | CRAN |        | 0.0-2   | 0.0-2  |        |
| curl       | CRAN | 1.2    | 2.3     | 2.3    | 0.9.7  |
| d3Network  | CRAN |        | 0.5.2.1 |        |        |
| data.table | CRAN | 1.9.6  | 1.10.0  | 1.10.1 | 1.9.6  |
| data.tree  | CRAN |        | 0.7.0   | 0.7.0  |        |
| datasets   | base | 3.3.2  | 3.3.2   | 3.3.1  | 3.3.0  |
| DBI        | CRAN | 0.5-1  | 0.5-1   | 0.5-1  | 0.5-1  |
| dendextend | CRAN | 1.4.0  | 1.4.0   | 1.4.0  |        |
| DEoptimR   | CRAN | 1.0-8  | 1.0-8   | 1.0-8  | 1.0-4  |
| desc       | CRAN | 1.1.0  |         | 1.1.0  |        |
| devtools   | CRAN | 1.12.0 | 1.12.0  | 1.12.0 | 1.12.0 |
| DiagrammeR | CRAN |        | 0.9.0   | 0.9.0  |        |
| dichromat  | CRAN | 2.0-0  | 2.0-0   | 2.0-0  | 2.0-0  |
| digest     | CRAN | 0.6.10 | 0.6.11  | 0.6.12 | 0.6.9  |
| diptest    | CRAN | 0.75-7 | 0.75-7  | 0.75-7 |        |
| doParallel | CRAN | 1.0.10 | 1.0.10  | 1.0.10 | 1.0.10 |

|                   |                            |        |        |        |        |
|-------------------|----------------------------|--------|--------|--------|--------|
| dplyr             | CRAN                       | 0.5.0  | 0.5.0  | 0.5.0  | 0.5.0  |
| ellipse           | CRAN                       |        | 0.3-8  | 0.3-8  | 0.3-8  |
| evaluate          | CRAN                       |        | 0.1    | 0.1    | 0.9    |
| fdrtool           | CRAN                       |        | 1.2.15 |        |        |
| fields            | CRAN                       |        | 8.1    |        |        |
| flexmix           | CRAN                       | 2.3-13 | 2.3-13 | 2.3-13 |        |
| forcats           | CRAN                       | 0.2.0  |        |        |        |
| foreach           | CRAN                       | 1.4.3  | 1.4.3  | 1.4.3  | 1.4.3  |
| foreign           | base                       | 0.8-67 | 0.8-67 | 0.8-67 | 0.8-66 |
| formatR           | CRAN                       |        | 1.4    | 1.4    | 1.4    |
| Formula           | CRAN                       |        | 1.2-1  |        | 1.2-1  |
| fpc               | CRAN                       | 2.1-10 | 2.1-10 | 2.1-10 |        |
| futile.logger     | CRAN                       |        | 1.4.3  | 1.4.3  | 1.4.1  |
| futile.options    | CRAN                       |        | 1.0.0  | 1.0.0  | 1.0.0  |
| gdata             | CRAN                       | 2.17.0 | 2.17.0 | 2.17.0 | 2.17.0 |
| geepack           | CRAN                       |        | 1.2-1  |        |        |
| GenomeInfoDb      | Bioconductor               |        | 1.10.2 | 1.10.1 |        |
| GenomicAlignments | Bioconductor               |        | 1.10.0 | 1.10.0 |        |
| GenomicRanges     | Bioconductor               |        | 1.26.2 | 1.26.1 |        |
| ggm               | CRAN                       |        | 2.3    |        |        |
| ggplot2           | CRAN                       | 2.1.0  | 2.2.1  | 2.2.1  | 2.1.0  |
| git2r             | CRAN                       | 0.15.0 | 0.18.0 | 0.16.0 | 0.15.0 |
| glasso            | CRAN                       |        | 1.8    |        |        |
| GO.db             | Bioconductor               |        | 3.4.0  | 3.2.2  | 3.3.0  |
| GOSemSim          | Bioconductor               |        | 2.0.3  | 1.28.2 | 1.30.3 |
| gplots            | CRAN                       | 3.0.1  | 3.0.1  | 3.0.1  | 3.0.1  |
| graph             | Bioconductor               |        | 1.52.0 |        |        |
| graphics          | base                       | 3.3.2  | 3.3.2  | 3.3.1  | 3.3.0  |
| graphsim          | GitHub<br>TomKellyGenetics | 0.1.0  | 0.1.0  | 0.1.0  | 0.1.0  |
| grDevices         | base                       | 3.3.2  | 3.3.2  | 3.3.1  | 3.3.0  |
| grid              | base                       | 3.3.2  | 3.3.2  | 3.3.1  | 3.3.0  |
| gridBase          | CRAN                       | 0.4-7  | 0.4-7  | 0.4-7  | 0.4-7  |
| gridExtra         | CRAN                       | 2.2.1  | 2.2.1  | 2.2.1  | 2.2.1  |
| gridGraphics      | CRAN                       |        | 0.1-5  |        |        |

|                   |                  |            |            |            |            |
|-------------------|------------------|------------|------------|------------|------------|
| gtable            | CRAN             | 0.2.0      | 0.2.0      | 0.2.0      | 0.2.0      |
| gtools            | CRAN             | 3.5.0      | 3.5.0      | 3.5.0      | 3.5.0      |
| haven             | CRAN             | 1.0.0      |            |            |            |
| heatmap.2x        | GitHub           |            | 0.0.0.9000 | 0.0.0.9000 | 0.0.0.9000 |
|                   | TomKellyGenetics |            |            |            | 0.0.0.9000 |
| hgu133plus2.db    | Bioconductor     |            | 3.2.3      |            |            |
| highr             | CRAN             |            | 0.6        | 0.6        | 0.6        |
| Hmisc             | CRAN             |            | 4.0-2      | 4.0-2      | 3.17-4     |
| hms               | CRAN             | 0.2        | 0.3        |            |            |
| htmlTable         | CRAN             |            | 1.8        | 1.9        |            |
| htmltools         | CRAN             | 0.3.5      | 0.3.5      | 0.3.5      | 0.3.5      |
| htmlwidgets       | CRAN             |            | 0.8        | 0.8        |            |
| httpuv            | CRAN             | 1.3.3      |            | 1.3.3      |            |
| httr              | CRAN             | 1.2.1      | 1.2.1      | 1.2.1      | 1.1.0      |
| huge              | CRAN             |            | 1.2.7      |            |            |
| hunspell          | CRAN             |            | 2.3        |            | 2          |
| hypergraph        | CRAN             |            | 1.46.0     |            |            |
| igraph            | CRAN             | 1.0.1      | 1.0.1      | 1.0.1      | 1.0.1      |
| igraph.extensions | GitHub           |            |            |            |            |
|                   | TomKellyGenetics | 0.1.0.9001 | 0.1.0.9001 | 0.1.0.9001 | 0.1.0.9001 |
| influenceR        | CRAN             |            | 0.1.0      | 0.1.0      |            |
| info.centrality   | GitHub           |            |            |            |            |
|                   | TomKellyGenetics | 0.1.0      | 0.1.0      | 0.1.0      | 0.1.0      |
| IRanges           | Bioconductor     |            | 2.8.1      | 2.8.1      | 2.6.1      |
| irlba             | CRAN             | 2.1.1      | 2.1.2      | 2.1.2      | 2.0.0      |
| iterators         | CRAN             | 1.0.8      | 1.0.8      | 1.0.8      | 1.0.8      |
| jpeg              | CRAN             |            | 0.1-8      |            |            |
| jsonlite          | CRAN             | 1.1        | 1.2        | 1.3        | 0.9.20     |
| KEGG.db           | Bioconductor     |            | 3.2.3      |            |            |
| kernlab           | CRAN             | 0.9-25     | 0.9-25     | 0.9-25     |            |
| KernSmooth        | base             | 2.23-15    | 2.23-15    | 2.23-15    | 2.23-15    |
| knitr             | CRAN             |            | 1.15.1     | 1.15.1     | 1.14       |
| labeling          | CRAN             | 0.3        | 0.3        | 0.3        | 0.3        |
| lambda.r          | CRAN             |            | 1.1.9      | 1.1.9      | 1.1.7      |
| lattice           | base             | 0.20-34    | 0.20-34    | 0.20-34    | 0.20-33    |

| latticeExtra | CRAN         |         | 0.6-28   |         | 0.6-28   |
|--------------|--------------|---------|----------|---------|----------|
| lava         | CRAN         |         | 1.4.6    |         |          |
| lavaan       | CRAN         |         | 0.5-22   |         |          |
| lazyeval     | CRAN         | 0.2.0   | 0.2.0    | 0.2.0   | 0.2.0    |
| les          | CRAN         |         | 1.24.0   |         |          |
| lgtdl        | CRAN         |         | 1.1.3    |         |          |
| limma        | Bioconductor |         | 3.30.7   | 3.30.3  |          |
| lme4         | CRAN         |         | 1.1-12   |         | 1.1-12   |
| lubridate    | CRAN         | 1.6.0   |          |         |          |
| magrittr     | CRAN         | 1.5     | 1.5      | 1.5     | 1.5      |
| maps         | CRAN         |         | 3.1.1    |         |          |
| markdown     | CRAN         |         | 0.7.7    | 0.7.7   | 0.7.7    |
| MASS         | base         | 7.3-45  | 7.3-45   | 7.3-45  | 7.3-45   |
| Matrix       | base         | 1.2-7.1 | 1.2-7.1  | 1.2-8   | 1.2-6    |
| matrixcalc   | CRAN         | 1.0-3   | 1.0-3    | 1.0-3   | 1.0-3    |
| mclust       | CRAN         | 5.2     | 5.2.1    | 5.2.2   | 5.2      |
| memoise      | CRAN         | 1.0.0   | 1.0.0    | 1.0.0   | 1.0.0    |
| methods      | base         | 3.3.2   | 3.3.2    | 3.3.1   | 3.3.0    |
| mgcv         | base         | 1.8-16  | 1.8-16   | 1.8-17  | 1.8-12   |
| mi           | CRAN         |         | 1        |         |          |
| mime         | CRAN         | 0.5     | 0.5      | 0.5     | 0.4      |
| minqa        | CRAN         |         | 1.2.4    |         | 1.2.4    |
| mnormt       | CRAN         | 1.5-5   | 1.5-5    |         | 1.5-4    |
| modelr       | CRAN         | 0.1.0   |          |         |          |
| modeltools   | CRAN         | 0.2-21  | 0.2-21   | 0.2-21  |          |
| multtest     | Bioconductor |         | 2.30.0   | 2.30.0  |          |
| munsell      | CRAN         | 0.4.3   | 0.4.3    | 0.4.3   | 0.4.3    |
| mvtnorm      | CRAN         | 1.0-5   | 1.0-5    | 1.0-6   | 1.0-5    |
| network      | CRAN         |         | 1.13.0   |         |          |
| nlme         | base         | 3.1-128 | 3.1-128  | 3.1-131 | 3.1-128  |
| nloptr       | CRAN         |         | 1.0.4    |         | 1.0.4    |
| NMF          | CRAN         | 0.20.6  | 0.20.6   | 0.20.6  | 0.20.6   |
| nnet         | base         | 7.3-12  | 7.3-12   | 7.3-12  | 7.3-12   |
| numDeriv     | CRAN         |         | 2016.8-1 |         | 2014.2-1 |
| openssl      | CRAN         | 0.9.4   | 0.9.6    | 0.9.6   | 0.9.4    |

|                   |                  |            |            |             |             |
|-------------------|------------------|------------|------------|-------------|-------------|
| org.Hs.eg.db      | Bioconductor     |            | 3.1.2      |             | 3.3.0       |
| org.Sc.sgd.db     | Bioconductor     |            | 3.4.0      |             |             |
| parallel          | base             | 3.3.2      | 3.3.2      | 3.3.1       | 3.3.0       |
| pathway.structure | GitHub           | 0.1.0      | 0.1.0      | 0.1.0       | 0.1.0       |
| .permutation      | TomKellyGenetics |            |            |             |             |
| pbivnorm          | CRAN             |            | 0.6.0      |             |             |
| PGSEA             | Bioconductor     |            | 1.48.0     |             |             |
| pkgmaker          | CRAN             | 0.22       | 0.22       | 0.22        | 0.22        |
| PKI               | CRAN             |            | 0.1-3      |             |             |
| plogr             | CRAN             |            | 0.1-1      | 0.1-1       |             |
| plot.igraph       | GitHub           | 0.0.0.9001 | 0.0.0.9001 | 0.0.0.9001  | 0.0.0.9001  |
|                   | TomKellyGenetics |            |            |             |             |
| plotrix           | CRAN             |            | 3.6-4      |             |             |
| plyr              | CRAN             | 1.8.4      | 1.8.4      | 1.8.4       | 1.8.3       |
| png               | CRAN             |            | 0.1-7      |             | 0.1-7       |
| prabclus          | CRAN             | 2.2-6      | 2.2-6      | 2.2-6       |             |
| praise            | CRAN             | 1.0.0      | 1.0.0      |             | 1.0.0       |
| pROC              | CRAN             |            | 1.8        | 1.9.1       |             |
| prodlim           | CRAN             |            | 1.5.7      |             |             |
| prof.tree         | CRAN             |            | 0.1.0      |             |             |
| protools          | CRAN             |            | 0.99-2     |             |             |
| progress          | CRAN             |            |            | 1.1.2       |             |
| psych             | CRAN             | 1.6.12     | 1.6.12     |             |             |
| purrr             | CRAN             | 0.2.2      | 0.2.2      | 0.2.2       | 0.2.2       |
| qgraph            | CRAN             |            | 1.4.1      |             |             |
| quadprog          | CRAN             |            | 1.5-5      | 1.5-5       | 1.5-5       |
| R.methodsS3       | CRAN             |            | 1.7.1      |             | 1.7.1       |
| R.oo              | CRAN             |            | 1.21.0     |             | 1.20.0      |
| R.utils           | CRAN             |            | 2.5.0      |             |             |
| R6                | CRAN             | 2.1.3      | 2.2.0      | 2.2.0       | 2.1.3       |
| RBGL              | CRAN             |            | 1.50.0     |             |             |
| RColorBrewer      | CRAN             | 1.1-2      | 1.1-2      | 1.1-2       | 1.1-2       |
| Rcpp              | CRAN             | 0.12.7     | 0.12.9     | 0.12.9      | 0.12.7      |
| RcppArmadillo     | CRAN             |            |            | 0.7.700.0.0 | 0.6.700.6.0 |
| RcppEigen         | CRAN             |            | 0.3.2.9.0  |             | 0.3.2.8.1   |

|              |                  |        |          |          |           |
|--------------|------------------|--------|----------|----------|-----------|
| RCurl        | CRAN             |        | 1.95-4.8 | 1.95-4.8 | 1.95-4.8  |
| reactome.db  | Bioconductor     |        | 1.52.1   | 1.52.1   |           |
| reactometree | GitHub           |        | 0.1      |          |           |
|              | TomKellyGenetics |        |          |          |           |
| readr        | CRAN             | 1.0.0  | 1.0.0    |          |           |
| readxl       | CRAN             | 0.1.1  |          |          |           |
| registry     | CRAN             | 0.3    | 0.3      | 0.3      | 0.3       |
| reshape2     | CRAN             | 1.4.1  | 1.4.2    | 1.4.2    | 1.4.1     |
| rgeff        | CRAN             |        | 0.15.3   | 0.15.3   |           |
| rgl          | CRAN             |        |          | 0.97.0   | 0.95.1441 |
| Rgraphviz    | CRAN             |        | 2.18.0   |          |           |
| rjson        | CRAN             |        | 0.2.15   |          |           |
| RJSONIO      | CRAN             |        | 1.3-0    |          |           |
| rmarkdown    | CRAN             |        | 1.3      | 1.3      | 1         |
| Rmpi         | CRAN             |        | 0.6-6    |          | 0.6-5     |
| rngtools     | CRAN             | 1.2.4  | 1.2.4    | 1.2.4    | 1.2.4     |
| robustbase   | CRAN             | 0.92-7 | 0.92-7   | 0.92-7   | 0.92-5    |
| ROCR         | CRAN             | 1.0-7  | 1.0-7    | 1.0-7    | 1.0-7     |
| Rook         | CRAN             |        | 1.1-1    | 1.1-1    |           |
| roxygen2     | CRAN             | 6.0.1  | 5.0.1    | 6.0.1    | 5.0.1     |
| rpart        | base             | 4.1-10 | 4.1-10   | 4.1-10   | 4.1-10    |
| rprojroot    | CRAN             | 1.2    | 1.1      | 1.2      |           |
| Rsamtools    | Bioconductor     |        | 1.26.1   | 1.26.1   |           |
| rsconnect    | CRAN             |        | 0.7      |          |           |
| RSQLite      | CRAN             |        | 1.1-2    | 1.1-2    | 1.0.0     |
| rstudioapi   | CRAN             | 0.6    | 0.6      | 0.6      | 0.6       |
| rvest        | CRAN             | 0.3.2  |          |          |           |
| S4Vectors    | Bioconductor     |        | 0.12.1   | 0.12.0   | 0.10.3    |
| safe         | Bioconductor     |        | 3.14.0   | 3.10.0   |           |
| scales       | CRAN             | 0.4.0  | 0.4.1    | 0.4.1    | 0.4.0     |
| selectr      | CRAN             | 0.3-1  |          |          |           |
| sem          | CRAN             |        | 3.1-8    |          |           |
| shiny        | CRAN             | 0.14   |          | 1.0.0    |           |
| slipt        | GitHub           |        | 0.1.0    | 0.1.0    | 0.1.0     |
|              | TomKellyGenetics |        |          |          |           |

|                       |                            |            |            |          |          |
|-----------------------|----------------------------|------------|------------|----------|----------|
| sm                    | CRAN                       | 2.2-5.4    | 2.2-5.4    |          |          |
| sna                   | CRAN                       |            | 2.4        |          |          |
| snow                  | CRAN                       | 0.4-1      | 0.4-2      | 0.4-2    | 0.3-13   |
| sourcetools           | CRAN                       | 0.1.5      |            | 0.1.5    |          |
| SparseM               | CRAN                       |            | 1.74       |          | 1.7      |
| spatial               | base                       | 7.3-11     | 7.3-11     | 7.3-11   | 7.3-11   |
| splines               | base                       | 3.3.2      | 3.3.2      | 3.3.1    | 3.3.0    |
| statnet.common        | CRAN                       |            | 3.3.0      |          |          |
| stats                 | base                       | 3.3.2      | 3.3.2      | 3.3.1    | 3.3.0    |
| stats4                | base                       | 3.3.2      | 3.3.2      | 3.3.1    | 3.3.0    |
| stringi               | CRAN                       | 1.1.1      | 1.1.2      | 1.1.2    | 1.0-1    |
| stringr               | CRAN                       | 1.1.0      | 1.1.0      | 1.2.0    | 1.0.0    |
| Summarized Experiment | Bioconductor               |            | 1.4.0      | 1.4.0    |          |
| survival              | base                       | 2.39-4     | 2.40-1     | 2.40-1   | 2.39-4   |
| tcltk                 | base                       | 3.3.2      | 3.3.2      | 3.3.1    | 3.3.0    |
| testthat              | CRAN                       | 1.0.2      | 1.0.2      |          | 1.0.2    |
| tibble                | CRAN                       | 1.2        | 1.2        | 1.2      | 1.2      |
| tidyverse             | GitHub<br>hadley           | 1.1.1      |            |          |          |
| timeline              | CRAN                       |            | 0.9        |          |          |
| tools                 | base                       | 3.3.2      | 3.3.2      | 3.3.1    | 3.3.0    |
| tpr                   | CRAN                       |            | 0.3-1      |          |          |
| trimcluster           | CRAN                       | 0.1-2      | 0.1-2      | 0.1-2    |          |
| Unicode               | CRAN                       | 9.0.0-1    | 9.0.0-1    | 9.0.0-1  |          |
| utils                 | base                       | 3.3.2      | 3.3.2      | 3.3.1    | 3.3.0    |
| vioplot               | CRAN                       |            | 0.2        |          |          |
| vioplotx              | GitHub<br>TomKellyGenetics | 0.0.0.9000 | 0.0.0.9000 |          |          |
| viridis               | CRAN                       | 0.3.4      | 0.3.4      | 0.3.4    |          |
| visNetwork            | CRAN                       |            | 1.0.3      | 1.0.3    |          |
| whisker               | CRAN                       | 0.3-2      | 0.3-2      | 0.3-2    | 0.3-2    |
| withr                 | CRAN                       | 1.0.2      | 1.0.2      | 1.0.2    | 1.0.2    |
| XML                   | base                       | 3.98-1.3   | 3.98-1.1   | 3.98-1.5 | 3.98-1.4 |

|          |              |        |        |        |
|----------|--------------|--------|--------|--------|
| xml2     | CRAN         | 1.1.1  | 1.1.1  | 1.0.0  |
| xtable   | CRAN         | 1.8-2  | 1.8-2  | 1.8-2  |
| XVector  | Bioconductor |        | 0.14.0 | 0.14.0 |
| yaml     | CRAN         |        | 2.1.14 | 2.1.14 |
| zlibbioc | CRAN         |        | 1.20.0 | 1.20.0 |
| zoo      | CRAN         | 1.7-13 | 1.7-14 | 1.7-13 |

# Appendix C

## Secondary Screen Data

A series of experimental genome-wide siRNA screens have been performed on synthetic lethal partners of *CDH1* (Telford *et al.*, 2015). The strongest candidates from a primary screen were subject to a further secondary screen for validation by independent replication with 4 gene knockdowns with different targeting siRNA. As shown in Table C.1, there is significant ( $p = 7.49 \times 10^{-3}$  by Fisher’s exact test) association between SLIPT candidates and stronger validations of siRNA candidates. Since there were more SLIPT– genes among those not validated and more SLIPT+ genes among those validated with several siRNAs, this supports the use of SLIPT as a synthetic lethal discovery procedure which may augment such screening experiments.

Table C.1: Comparing SLIPT genes against Secondary siRNA Screen in breast cancer

|        |          | Secondary Screen |     |     |     |     | Total |     |
|--------|----------|------------------|-----|-----|-----|-----|-------|-----|
|        |          | 0/4              | 1/4 | 2/4 | 3/4 | 4/4 |       |     |
| SLIPT+ | Observed | 70               | 46  | 31  | 8   | 2   | 157   |     |
|        | Expected | 85               | 44  | 10  | 4   | 2   |       |     |
| SLIPT– | Observed | 190              | 90  | 31  | 10  | 4   | 325   |     |
|        | Expected | 175              | 91  | 42  | 12  | 4   |       |     |
|        |          | Total            | 280 | 136 | 52  | 18  | 6     | 482 |

Similar analysis with mtSLIPT, comparing SLIPT against *CDH1* somatic mutation with siRNA validation results was not significant ( $p = 7.02 \times 10^{-1}$  by Fisher’s exact test). However, as shown in Table C.2, the observed and expected values were in a direction consistent with that observed above for SLIPT against low *CDH1* expression.

It is not unexpected that this result does not have comparable statistical support due to the lower sample size for mutation data.

Table C.2: Comparing mtSLIPT genes against Secondary siRNA Screen in breast cancer

|                 |          | Secondary Screen |     |     |     |     | <b>Total</b> |
|-----------------|----------|------------------|-----|-----|-----|-----|--------------|
|                 |          | 0/4              | 1/4 | 2/4 | 3/4 | 4/4 |              |
| <b>mtSLIPT+</b> | Observed | 54               | 35  | 17  | 4   | 6   | 111          |
|                 | Expected | 60               | 31  | 14  | 4   | 1   |              |
| <b>mtSLIPT-</b> | Observed | 206              | 101 | 45  | 14  | 5   | 371          |
|                 | Expected | 200              | 105 | 48  | 14  | 4   |              |
| <b>Total</b>    |          | 269              | 143 | 63  | 19  | 6   | 482          |

This analysis was replicated on a (smaller) stomach cancer dataset but it was less conclusive ( $p = 2.36 \times 10^{-1}$  by Fisher's exact test). As shown in Table C.3, fewer SLIPT candidates were validated than expected statistically. However, these results in stomach cancer may not be directly comparable to experiments in a breast cell line. Genes validated by 0 or 1 siRNA behave consistently with the results above.

Table C.3: Comparing SLIPT genes against Secondary siRNA Screen in stomach cancer

|               |          | Secondary Screen |     |     |     |     | <b>Total</b> |
|---------------|----------|------------------|-----|-----|-----|-----|--------------|
|               |          | 0/4              | 1/4 | 2/4 | 3/4 | 4/4 |              |
| <b>SLIPT+</b> | Observed | 67               | 47  | 13  | 4   | 1   | 132          |
|               | Expected | 71               | 37  | 17  | 5   | 2   |              |
| <b>SLIPT-</b> | Observed | 195              | 90  | 50  | 14  | 5   | 354          |
|               | Expected | 190              | 100 | 46  | 13  | 4   |              |
| <b>Total</b>  |          | 262              | 137 | 63  | 19  | 6   | 486          |

# **Appendix D**

## **Mutation Analysis in Breast Cancer**

### **D.1 Synthetic Lethal Genes and Pathways**

SLIPT expression analysis (described in Section 3.1) on TCGA breast cancer data ( $n = 969$ ) found the following genes and pathways, described in sections 4.1 and 4.1.1.

Table D.1: Candidate synthetic lethal gene partners of *CDH1* from mtSLIPT

| Gene            | Observed | Expected | $\chi^2$ value | p-value                | p-value (FDR)          |
|-----------------|----------|----------|----------------|------------------------|------------------------|
| <i>TFAP2B</i>   | 8        | 36.7     | 89.5           | $3.60 \times 10^{-20}$ | $8.37 \times 10^{-17}$ |
| <i>ZNF423</i>   | 15       | 36.7     | 78.8           | $7.89 \times 10^{-18}$ | $1.22 \times 10^{-14}$ |
| <i>CALCOCO1</i> | 11       | 36.7     | 76.8           | $2.09 \times 10^{-17}$ | $2.59 \times 10^{-14}$ |
| <i>RBM5</i>     | 13       | 36.7     | 75.7           | $3.65 \times 10^{-17}$ | $4.00 \times 10^{-14}$ |
| <i>BTG2</i>     | 7        | 36.7     | 71.7           | $2.72 \times 10^{-16}$ | $1.81 \times 10^{-13}$ |
| <i>RXRA</i>     | 6        | 36.7     | 70.5           | $5.00 \times 10^{-16}$ | $2.97 \times 10^{-13}$ |
| <i>SLC27A1</i>  | 11       | 36.7     | 70.3           | $5.42 \times 10^{-16}$ | $2.97 \times 10^{-13}$ |
| <i>MEF2D</i>    | 12       | 36.7     | 69.6           | $7.86 \times 10^{-16}$ | $3.95 \times 10^{-13}$ |
| <i>NISCH</i>    | 12       | 36.7     | 69.6           | $7.86 \times 10^{-16}$ | $3.95 \times 10^{-13}$ |
| <i>AVPR2</i>    | 9        | 36.7     | 69.2           | $9.36 \times 10^{-16}$ | $4.58 \times 10^{-13}$ |
| <i>CRY2</i>     | 13       | 36.7     | 68.9           | $1.07 \times 10^{-15}$ | $4.98 \times 10^{-13}$ |
| <i>RAPGEF3</i>  | 13       | 36.7     | 68.9           | $1.07 \times 10^{-15}$ | $4.98 \times 10^{-13}$ |
| <i>NRIP2</i>    | 10       | 36.7     | 68.2           | $1.58 \times 10^{-15}$ | $7.18 \times 10^{-13}$ |
| <i>DARC</i>     | 12       | 36.7     | 66.4           | $3.76 \times 10^{-15}$ | $1.54 \times 10^{-12}$ |
| <i>SFRS5</i>    | 12       | 36.7     | 66.4           | $3.76 \times 10^{-15}$ | $1.54 \times 10^{-12}$ |
| <i>NOSTRIN</i>  | 5        | 36.7     | 65.1           | $7.40 \times 10^{-15}$ | $2.70 \times 10^{-12}$ |
| <i>KIF13B</i>   | 12       | 36.7     | 63.4           | $1.69 \times 10^{-14}$ | $5.16 \times 10^{-12}$ |
| <i>TENC1</i>    | 10       | 36.7     | 62.5           | $2.67 \times 10^{-14}$ | $7.40 \times 10^{-12}$ |
| <i>MFAP4</i>    | 12       | 36.7     | 60.5           | $7.17 \times 10^{-14}$ | $1.67 \times 10^{-11}$ |
| <i>ELN</i>      | 13       | 36.7     | 59.7           | $1.07 \times 10^{-13}$ | $2.32 \times 10^{-11}$ |
| <i>SGK223</i>   | 14       | 36.7     | 59             | $1.51 \times 10^{-13}$ | $3.05 \times 10^{-11}$ |
| <i>KIF12</i>    | 11       | 36.7     | 58.8           | $1.74 \times 10^{-13}$ | $3.34 \times 10^{-11}$ |
| <i>SELP</i>     | 11       | 36.7     | 58.8           | $1.74 \times 10^{-13}$ | $3.34 \times 10^{-11}$ |
| <i>CIRBP</i>    | 9        | 36.7     | 58.7           | $1.83 \times 10^{-13}$ | $3.41 \times 10^{-11}$ |
| <i>CTDSP1</i>   | 9        | 36.7     | 58.7           | $1.83 \times 10^{-13}$ | $3.41 \times 10^{-11}$ |

Strongest candidate SL partners for *CDH1* by mtSLIPT with observed and expected mutant samples with low expression of partner genes

Table D.2: Pathways for *CDH1* partners from mtSLIPT

| Pathways Over-represented   | Pathway Size | SL Genes | p-value (FDR)          |
|---|--------------|----------|------------------------|
| Eukaryotic Translation Elongation                                 | 86           | 60       | $2.0 \times 10^{-128}$ |
| Peptide chain elongation  | 83           | 59       | $2.0 \times 10^{-128}$ |
| Eukaryotic Translation Termination                                | 83           | 58       | $2.3 \times 10^{-125}$ |
| Viral mRNA Translation  | 81           | 57       | $2.5 \times 10^{-124}$ |
| Nonsense Mediated Decay independent of the Exon Junction Complex  | 88           | 59       | $8.6 \times 10^{-124}$ |
| Nonsense-Mediated Decay   | 103          | 61       | $5.2 \times 10^{-117}$ |
| Nonsense Mediated Decay enhanced by the Exon Junction Complex     | 103          | 61       | $5.2 \times 10^{-117}$ |
| Formation of a pool of free 40S subunits                          | 93           | 58       | $1.6 \times 10^{-116}$ |
| L13a-mediated translational silencing of Ceruloplasmin expression | 103          | 59       | $1.3 \times 10^{-111}$ |
| 3' -UTR-mediated translational regulation                         | 103          | 59       | $1.3 \times 10^{-111}$ |
| GTP hydrolysis and joining of the 60S ribosomal subunit           | 104          | 59       | $6.2 \times 10^{-111}$ |
| SRP-dependent cotranslational protein targeting to membrane       | 104          | 58       | $2.9 \times 10^{-108}$ |
| Eukaryotic Translation Initiation                                 | 111          | 59       | $3.0 \times 10^{-106}$ |
| Cap-dependent Translation Initiation                              | 111          | 59       | $3.0 \times 10^{-106}$ |
| Influenza Viral RNA Transcription and Replication                 | 108          | 57       | $5.1 \times 10^{-103}$ |
| Influenza Infection   | 117          | 59       | $1.5 \times 10^{-102}$ |
| Translation   | 141          | 64       | $3.7 \times 10^{-101}$ |
| Influenza Life Cycle  | 112          | 57       | $1.4 \times 10^{-100}$ |
| GPCR downstream signaling   | 472          | 116      | $1.0 \times 10^{-80}$  |
| Hemostasis  | 422          | 105      | $1.4 \times 10^{-78}$  |

Gene set over-representation analysis (hypergeometric test) for Reactome pathways in mtSLIPT partners for *CDH1*

The genes and pathways identified in Tables D.1 and D.2 were derived from comparing the expression profiles of potential partners to the mutation status of *CDH1* (as shown in Figure 3.2). Thus the following analysis is only limited the samples for which TCGA provides both expression and somatic mutation data.

## D.2 Synthetic Lethal Expression Profiles

Similar to the analysis of synthetic lethal partners against low *CDH1* expression in 4.1.2, the partners detected from *CDH1* mutation were also examined for their expression profiles and the pathway composition of gene clusters. Hierarchical clustering was performed on mtSLIPT partners for *CDH1* as showing in Figure D.1. Overrepresentation for Reactome pathways for each of the gene clusters identified is given in Table D.3.

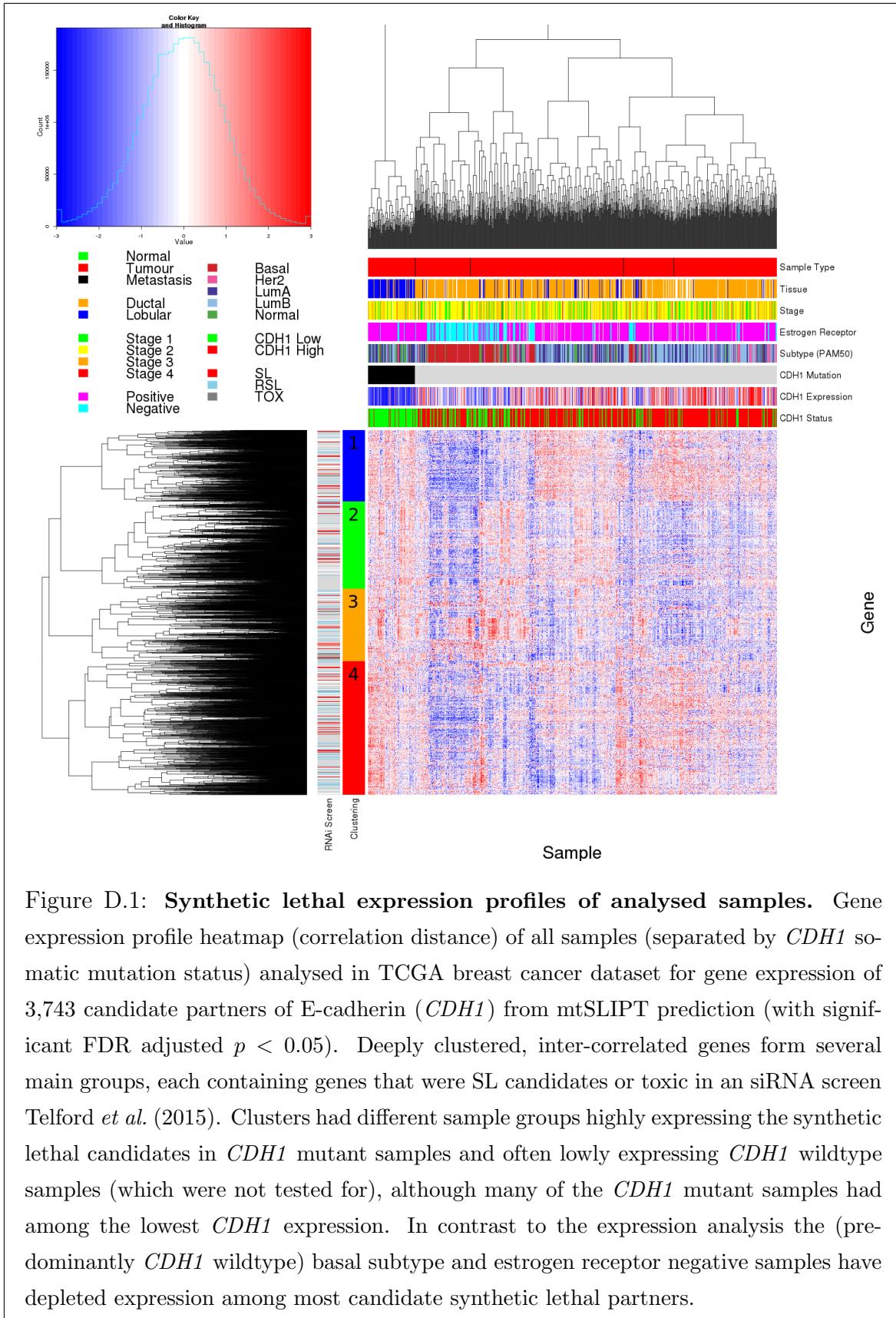


Table D.3: Pathway composition for clusters of *CDH1* partners from mtSLIPT

| Pathways Over-represented in Cluster 1        | Pathway Size | Cluster Genes | p-value (FDR)        |
|---|--------------|---------------|----------------------|
| Olfactory Signaling Pathway                   | 57           | 8             | $7.1 \times 10^{-9}$ |
| Assembly of the primary cilium                | 149          | 14            | $8.0 \times 10^{-9}$ |
| Sphingolipid metabolism                       | 62           | 8             | $9.6 \times 10^{-9}$ |
| Signaling by ERBB4                            | 133          | 12            | $5.1 \times 10^{-8}$ |
| PI3K Cascade                                  | 65           | 7             | $4.9 \times 10^{-7}$ |
| Circadian Clock                               | 33           | 5             | $4.9 \times 10^{-7}$ |
| Nuclear signaling by ERBB4                    | 34           | 5             | $4.9 \times 10^{-7}$ |
| Intraflagellar transport                      | 35           | 5             | $4.9 \times 10^{-7}$ |
| PI3K events in ERBB4 signaling                | 87           | 8             | $4.9 \times 10^{-7}$ |
| PIP3 activates AKT signaling                  | 87           | 8             | $4.9 \times 10^{-7}$ |
| PI3K events in ERBB2 signaling                | 87           | 8             | $4.9 \times 10^{-7}$ |
| PI-3K cascade:FGFR1                           | 87           | 8             | $4.9 \times 10^{-7}$ |
| PI-3K cascade:FGFR2                           | 87           | 8             | $4.9 \times 10^{-7}$ |
| PI-3K cascade:FGFR3                           | 87           | 8             | $4.9 \times 10^{-7}$ |
| PI-3K cascade:FGFR4                           | 87           | 8             | $4.9 \times 10^{-7}$ |
| Deadenylation of mRNA                         | 22           | 4             | $5.6 \times 10^{-7}$ |
| PI3K/AKT activation                           | 90           | 8             | $5.6 \times 10^{-7}$ |
| Cargo trafficking to the periciliary membrane | 38           | 5             | $5.6 \times 10^{-7}$ |
| Signaling by Hedgehog                         | 108          | 9             | $5.6 \times 10^{-7}$ |
| Downstream signal transduction                | 143          | 11            | $5.6 \times 10^{-7}$ |

| Pathways Over-represented in Cluster 2                        | Pathway Size | Cluster Genes | p-value (FDR)         |
|---|--------------|---------------|-----------------------|
| G <sub>αs</sub> signalling events                             | 83           | 19            | $5.1 \times 10^{-25}$ |
| Extracellular matrix organization                             | 238          | 30            | $1.4 \times 10^{-18}$ |
| Hemostasis  | 422          | 46            | $2.7 \times 10^{-16}$ |
| Aquaporin-mediated transport                                  | 32           | 9             | $2.7 \times 10^{-16}$ |
| Transcriptional regulation of white adipocyte differentiation | 56           | 11            | $1.7 \times 10^{-15}$ |
| Degradation of the extracellular matrix                       | 102          | 15            | $1.7 \times 10^{-15}$ |
| Integration of energy metabolism                              | 84           | 13            | $8.8 \times 10^{-15}$ |
| GPCR downstream signaling                                     | 472          | 48            | $2.8 \times 10^{-14}$ |
| G <sub>αs</sub> signalling events                             | 15           | 6             | $5.0 \times 10^{-14}$ |
| Molecules associated with elastic fibres                      | 33           | 8             | $5.4 \times 10^{-14}$ |
| Phase 1 - Functionalization of compounds                      | 67           | 11            | $5.6 \times 10^{-14}$ |
| Platelet activation, signaling and aggregation                | 179          | 20            | $5.6 \times 10^{-14}$ |
| Vasopressin regulates renal water homeostasis via Aquaporins  | 24           | 7             | $6.1 \times 10^{-14}$ |
| Elastic fibre formation                                       | 37           | 8             | $.03 \times 10^{-13}$ |
| Calmodulin induced events                                     | 27           | 7             | $3.3 \times 10^{-13}$ |
| CaM pathway   | 27           | 7             | $3.3 \times 10^{-13}$ |
| cGMP effects  | 18           | 6             | $3.6 \times 10^{-13}$ |
| G <sub>αs</sub> signalling events                             | 167          | 18            | $6.3 \times 10^{-13}$ |
| Ca-dependent events   | 29           | 7             | $8.2 \times 10^{-13}$ |
| Binding and Uptake of Ligands by Scavenger Receptors          | 40           | 8             | $8.2 \times 10^{-13}$ |

| Pathways Over-represented in Cluster 3                              | Pathway Size | Cluster Genes | p-value (FDR)          |
|---|--------------|---------------|------------------------|
| Eukaryotic Translation Elongation                                   | 86           | 55            | $1.1 \times 10^{-112}$ |
| Peptide chain elongation  | 83           | 54            | $1.3 \times 10^{-112}$ |
| Viral mRNA Translation  | 81           | 53            | $1.6 \times 10^{-111}$ |
| Eukaryotic Translation Termination                                  | 83           | 53            | $7.1 \times 10^{-110}$ |
| Nonsense Mediated Decay independent of the Exon Junction Complex    | 88           | 54            | $1.0 \times 10^{-108}$ |
| Formation of a pool of free 40S subunits                            | 93           | 53            | $4.1 \times 10^{-102}$ |
| Nonsense-Mediated Decay   | 103          | 54            | $3.9 \times 10^{-98}$  |
| Nonsense-Mediated Decay enhanced by the Exon Junction Complex       | 103          | 54            | $3.9 \times 10^{-98}$  |
| L13a-mediated translational silencing of Ceruloplasmin expression   | 103          | 53            | $1.2 \times 10^{-95}$  |
| 3' -UTR-mediated translational regulation                           | 103          | 53            | $1.2 \times 10^{-95}$  |
| SRP-dependent cotranslational protein targeting to membrane         | 104          | 53            | $4.3 \times 10^{-95}$  |
| GTP hydrolysis and joining of the 60S ribosomal subunit             | 104          | 53            | $4.3 \times 10^{-95}$  |
| Influenza Viral RNA Transcription and Replication                   | 108          | 53            | $9.6 \times 10^{-93}$  |
| Eukaryotic Translation Initiation                                   | 111          | 53            | $4.2 \times 10^{-91}$  |
| Cap-dependent Translation Initiation                                | 111          | 53            | $4.2 \times 10^{-91}$  |
| Influenza Life Cycle  | 112          | 53            | $1.4 \times 10^{-90}$  |
| Influenza Infection   | 117          | 53            | $6.2 \times 10^{-88}$  |
| Translation   | 141          | 55            | $3 \times 10^{-81}$    |
| Formation of the ternary complex, and subsequently, the 43S complex | 47           | 23            | $2.3 \times 10^{-48}$  |
| Translation initiation complex formation                            | 54           | 23            | $9.1 \times 10^{-45}$  |

| Pathways Over-represented in Cluster 4                                | Pathway Size | Cluster Genes | p-value (FDR)         |
|---|--------------|---------------|-----------------------|
| ECM proteoglycans   | 66           | 10            | $2.9 \times 10^{-11}$ |
| deactivation of the beta-catenin transactivating complex              | 38           | 7             | $5.1 \times 10^{-10}$ |
| Arachidonic acid metabolism   | 41           | 7             | $1.1 \times 10^{-9}$  |
| Gαq signalling events   | 149          | 14            | $4.0 \times 10^{-9}$  |
| HS-GAG degradation  | 21           | 5             | $4.5 \times 10^{-9}$  |
| Uptake and actions of bacterial toxins                                | 22           | 5             | $6.1 \times 10^{-9}$  |
| Gastrin-CREB signalling pathway via PKC and MAPK                      | 170          | 15            | $6.1 \times 10^{-9}$  |
| RNA Polymerase I, RNA Polymerase III, and Mitochondrial Transcription | 64           | 8             | $6.1 \times 10^{-9}$  |
| Non-integrin membrane-ECM interactions                                | 53           | 7             | $1.5 \times 10^{-8}$  |
| Syndecan interactions   | 25           | 5             | $1.5 \times 10^{-8}$  |
| NOTCH1 Intracellular Domain Regulates Transcription                   | 40           | 6             | $2.3 \times 10^{-8}$  |
| Synthesis of Leukotrienes and Exoxins                                 | 15           | 4             | $3.2 \times 10^{-8}$  |
| Signaling by NOTCH1   | 59           | 7             | $5.3 \times 10^{-8}$  |
| Regulation of insulin secretion                                       | 44           | 6             | $6.0 \times 10^{-8}$  |
| Metabolism of lipids and lipoproteins                                 | 471          | 37            | $8.2 \times 10^{-8}$  |
| Signaling by NOTCH  | 80           | 8             | $1.2 \times 10^{-7}$  |
| Platelet activation, signaling and aggregation                        | 179          | 14            | $1.2 \times 10^{-7}$  |
| Recruitment of mitotic centrosome proteins and complexes              | 64           | 7             | $1.2 \times 10^{-7}$  |
| Centrosome maturation   | 64           | 7             | $1.2 \times 10^{-7}$  |
| Biological oxidations   | 133          | 11            | $1.5 \times 10^{-7}$  |

### D.3 Comparison to Primary Screen

The mutation synthetic lethal partners with *CDH1* were also compared to siRNA primary screen data (Telford *et al.*, 2015), as performed in Section 4.2.1. These are expected to be more concordant with the experimental results performed on a null mutant, however this is not the case at the gene level: less genes overlapped with experimental candidates in Figure D.2. This may be affected by lower sample size for mutations in TCGA data or lower frequency (expected value) of *CDH1* mutations compared to low expression.

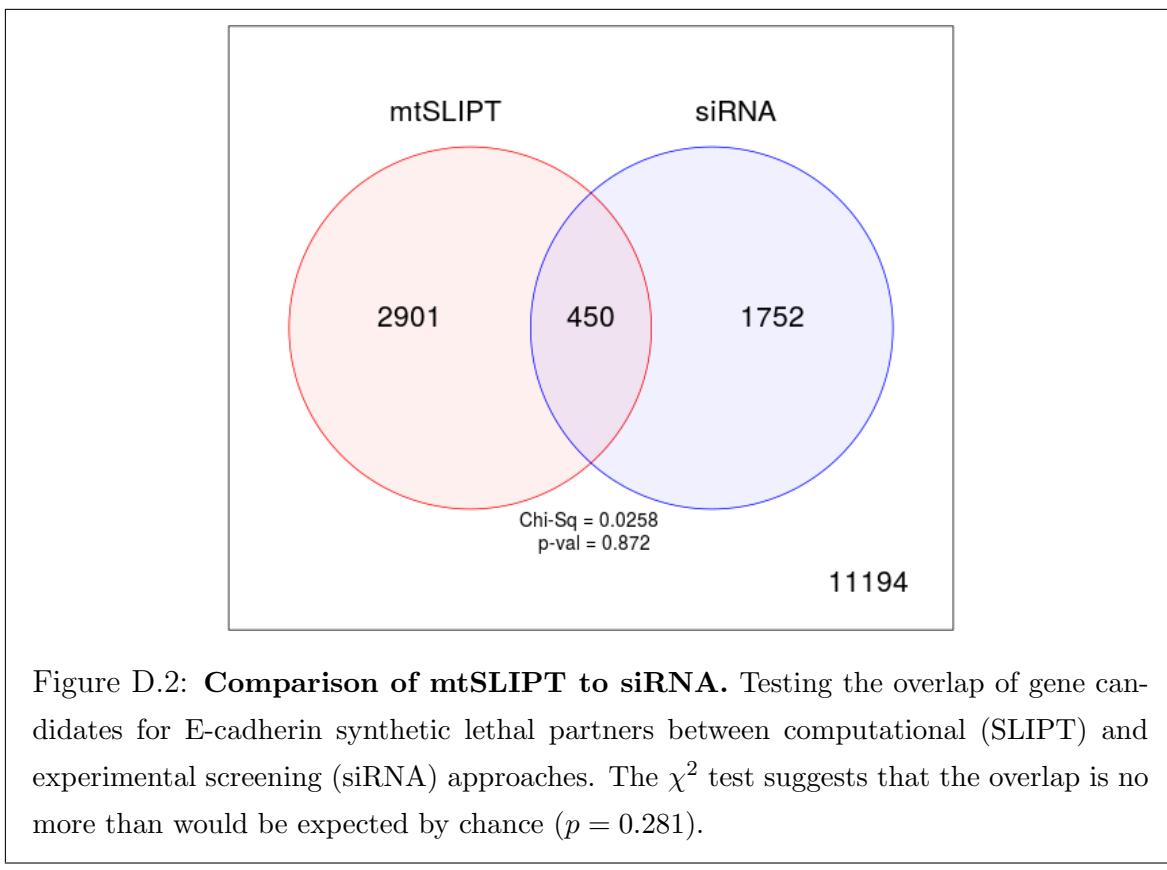


Figure D.2: **Comparison of mtSLIPT to siRNA.** Testing the overlap of gene candidates for E-cadherin synthetic lethal partners between computational (SLIPT) and experimental screening (siRNA) approaches. The  $\chi^2$  test suggests that the overlap is no more than would be expected by chance ( $p = 0.281$ ).

Despite a lower sample size (and low number of predicted partners) for mutation analysis, the pathway composition (Tables D.2 and D.4) is similar to expression analysis, as described in Section 4.2.1.4. In particular, the resampling analysis (Section ??) supported many of the results of expression analysis (Section 4.2.1.4.1) with Tables D.5 and D.6 detecting many of the same or functionally-related pathways.

Table D.4: Pathway composition for *CDH1* partners from mtSLIPT and siRNA

| Predicted only by SLIPT (2901 genes)                              | Pathway Size | Genes Identified | p-value (FDR)          |
|---|--------------|------------------|------------------------|
| Eukaryotic Translation Elongation                                 | 87           | 57               | $2.8 \times 10^{-120}$ |
| Peptide chain elongation  | 84           | 56               | $3.1 \times 10^{-120}$ |
| Eukaryotic Translation Termination                                | 84           | 55               | $2.8 \times 10^{-117}$ |
| Viral mRNA Translation  | 82           | 54               | $4.1 \times 10^{-116}$ |
| Nonsense Mediated Decay independent of the Exon Junction Complex  | 89           | 55               | $3.7 \times 10^{-113}$ |
| Formation of a pool of free 40S subunits                          | 94           | 55               | $2.8 \times 10^{-109}$ |
| Nonsense-Mediated Decay   | 104          | 57               | $8.4 \times 10^{-108}$ |
| Nonsense Mediated Decay enhanced by the Exon Junction Complex     | 104          | 57               | $8.4 \times 10^{-108}$ |
| L13a-mediated translational silencing of Ceruloplasmin expression | 104          | 56               | $3.4 \times 10^{-105}$ |
| 3' -UTR-mediated translational regulation                         | 104          | 56               | $3.4 \times 10^{-105}$ |
| GTP hydrolysis and joining of the 60S ribosomal subunit           | 105          | 56               | $1.4 \times 10^{-104}$ |
| Eukaryotic Translation Initiation                                 | 112          | 56               | $2.8 \times 10^{-100}$ |
| Cap-dependent Translation Initiation                              | 112          | 56               | $2.8 \times 10^{-100}$ |
| SRP-dependent cotranslational protein targeting to membrane       | 105          | 54               | $2.2 \times 10^{-99}$  |
| Influenza Viral RNA Transcription and Replication                 | 109          | 54               | $5.3 \times 10^{-97}$  |
| Influenza Life Cycle  | 113          | 54               | $9.6 \times 10^{-95}$  |
| Influenza Infection   | 118          | 55               | $1.7 \times 10^{-94}$  |
| Translation   | 142          | 60               | $3.5 \times 10^{-94}$  |
| Infectious disease  | 349          | 77               | $5.9 \times 10^{-62}$  |
| Extracellular matrix organization                                 | 241          | 54               | $3.0 \times 10^{-52}$  |

| Detected only by siRNA screen (1752 genes)       | Pathway Size | Genes Identified | p-value (FDR)         |
|--|--------------|------------------|-----------------------|
| Class A/1 (Rhodopsin-like receptors)             | 282          | 69               | $1.9 \times 10^{-59}$ |
| GPCR ligand binding                              | 363          | 78               | $2.7 \times 10^{-54}$ |
| Peptide ligand-binding receptors                 | 175          | 41               | $1.5 \times 10^{-42}$ |
| $G_{\alpha i}$ signalling events                 | 184          | 41               | $1.1 \times 10^{-40}$ |
| Gastrin-CREB signalling pathway via PKC and MAPK | 180          | 37               | $1.5 \times 10^{-35}$ |
| $G_{\alpha q}$ signalling events                 | 159          | 34               | $3.7 \times 10^{-35}$ |
| DAP12 interactions                               | 159          | 27               | $1.1 \times 10^{-24}$ |
| VEGFA-VEGFR2 Pathway                             | 91           | 19               | $1.0 \times 10^{-23}$ |
| Downstream signal transduction                   | 146          | 24               | $1.9 \times 10^{-22}$ |
| Signaling by VEGF                                | 99           | 19               | $2.6 \times 10^{-22}$ |
| DAP12 signaling                                  | 149          | 24               | $4.2 \times 10^{-22}$ |
| Organelle biogenesis and maintenance             | 264          | 34               | $4.3 \times 10^{-20}$ |
| Downstream signaling of activated FGFR1          | 134          | 21               | $4.3 \times 10^{-20}$ |
| Downstream signaling of activated FGFR2          | 134          | 21               | $4.3 \times 10^{-20}$ |
| Downstream signaling of activated FGFR3          | 134          | 21               | $4.3 \times 10^{-20}$ |
| Downstream signaling of activated FGFR4          | 134          | 21               | $4.3 \times 10^{-20}$ |
| Signaling by ERBB2                               | 146          | 22               | $5.3 \times 10^{-20}$ |
| Signaling by FGFR                                | 146          | 22               | $5.3 \times 10^{-20}$ |
| Signaling by FGFR1                               | 146          | 22               | $5.3 \times 10^{-20}$ |
| Signaling by FGFR2                               | 146          | 22               | $5.3 \times 10^{-20}$ |

| Intersection of SLIPT and siRNA screen (450 genes)       | Pathway Size | Genes Identified | p-value (FDR)        |
|--|--------------|------------------|----------------------|
| HS-GAG degradation                                       | 21           | 4                | $4.9 \times 10^{-6}$ |
| Retinoid metabolism and transport                        | 39           | 5                | $4.9 \times 10^{-6}$ |
| Platelet activation, signaling and aggregation           | 186          | 13               | $4.9 \times 10^{-6}$ |
| Signaling by NOTCH4                                      | 11           | 3                | $4.9 \times 10^{-6}$ |
| $G_{\alpha s}$ signalling events                         | 100          | 8                | $5.0 \times 10^{-6}$ |
| Defective EXT2 causes exostoses 2                        | 12           | 3                | $5.0 \times 10^{-6}$ |
| Defective EXT1 causes exostoses 1, TRPS2 and CHDS        | 12           | 3                | $5.0 \times 10^{-6}$ |
| Class A/1 (Rhodopsin-like receptors)                     | 289          | 18               | $2.2 \times 10^{-5}$ |
| Signaling by PDGF  | 173          | 11               | $2.9 \times 10^{-5}$ |
| Circadian Clock  | 34           | 4                | $2.9 \times 10^{-5}$ |
| Signaling by ERBB4                                       | 139          | 9                | $4.3 \times 10^{-5}$ |
| Role of LAT2/NTAL/LAB on calcium mobilization            | 99           | 7                | $4.4 \times 10^{-5}$ |
| Peptide ligand-binding receptors                         | 181          | 11               | $4.5 \times 10^{-5}$ |
| Defective B4GALT7 causes EDS, progeroid type             | 19           | 3                | $4.5 \times 10^{-5}$ |
| Defective B3GAT3 causes JDSSDHD                          | 19           | 3                | $4.5 \times 10^{-5}$ |
| Signaling by NOTCH                                       | 80           | 6                | $4.5 \times 10^{-5}$ |
| $G_{\alpha q}$ signalling events                         | 164          | 10               | $5.1 \times 10^{-5}$ |
| Response to elevated platelet cytosolic Ca <sup>2+</sup> | 84           | 6                | $7.1 \times 10^{-5}$ |
| Signaling by ERBB2                                       | 148          | 9                | $7.1 \times 10^{-5}$ |
| Signaling by SCF-KIT                                     | 129          | 8                | $8.3 \times 10^{-5}$ |

### D.3.1 Resampling Analysis

Table D.5: Pathways for *CDH1* partners from mtSLIPT

| Reactome Pathway  | Over-representation    | Permutation              |
|---|------------------------|--------------------------|
| <b>Eukaryotic Translation Elongation</b>                          | $3.2 \times 10^{-128}$ | $< 7.035 \times 10^{-4}$ |
| Peptide chain elongation  | $3.2 \times 10^{-128}$ | $< 7.035 \times 10^{-4}$ |
| <b>Eukaryotic Translation Termination</b>                         | $3.7 \times 10^{-125}$ | $< 7.035 \times 10^{-4}$ |
| Viral mRNA Translation  | $4.1 \times 10^{-124}$ | $< 7.035 \times 10^{-4}$ |
| Nonsense Mediated Decay independent of the Exon Junction Complex  | $1.4 \times 10^{-123}$ | $< 7.035 \times 10^{-4}$ |
| Nonsense-Mediated Decay   | $8.4 \times 10^{-117}$ | $< 7.035 \times 10^{-4}$ |
| Nonsense Mediated Decay enhanced by the Exon Junction Complex     | $8.4 \times 10^{-117}$ | $< 7.035 \times 10^{-4}$ |
| Formation of a pool of free 40S subunits                          | $2.6 \times 10^{-116}$ | $< 7.035 \times 10^{-4}$ |
| L13a-mediated translational silencing of Ceruloplasmin expression | $2.0 \times 10^{-111}$ | $< 7.035 \times 10^{-4}$ |
| 3' -UTR-mediated translational regulation                         | $2.0 \times 10^{-111}$ | $< 7.035 \times 10^{-4}$ |
| GTP hydrolysis and joining of the 60S ribosomal subunit           | $9.9 \times 10^{-111}$ | $< 7.035 \times 10^{-4}$ |
| SRP-dependent cotranslational protein targeting to membrane       | $4.7 \times 10^{-108}$ | $< 7.035 \times 10^{-4}$ |
| <b>Eukaryotic Translation Initiation</b>                          | $4.8 \times 10^{-106}$ | $< 7.035 \times 10^{-4}$ |
| Cap-dependent Translation Initiation                              | $4.8 \times 10^{-106}$ | $< 7.035 \times 10^{-4}$ |
| <b>Influenza Viral RNA Transcription and Replication</b>          | $8.1 \times 10^{-103}$ | $< 7.035 \times 10^{-4}$ |
| <b>Influenza Infection</b>  | $2.4 \times 10^{-102}$ | $< 7.035 \times 10^{-4}$ |
| <b>Translation</b>  | $6.0 \times 10^{-101}$ | $< 7.035 \times 10^{-4}$ |
| <b>Influenza Life Cycle</b>                                       | $2.2 \times 10^{-100}$ | $< 7.035 \times 10^{-4}$ |
| <b>Disease</b>  | $2.1 \times 10^{-90}$  | 0.013347                 |
| <b>GPCR downstream signaling</b>                                  | $1.6 \times 10^{-80}$  | 0.095478                 |
| Hemostasis  | $2.1 \times 10^{-78}$  | 0.2671                   |
| Signaling by GPCR   | $1.2 \times 10^{-73}$  | 0.44939                  |
| <i>Extracellular matrix organization</i>                          | $2.2 \times 10^{-67}$  | 0.054008                 |
| Metabolism of proteins  | $1.4 \times 10^{-66}$  | 0.9607                   |
| Signal Transduction   | $2.1 \times 10^{-66}$  | 0.48184                  |
| Developmental Biology   | $2.5 \times 10^{-66}$  | 0.54075                  |
| Innate Immune System  | $5.3 \times 10^{-66}$  | 0.9589                   |
| Infectious disease  | $9.6 \times 10^{-66}$  | 0.21075                  |
| Signalling by NGF   | $1.1 \times 10^{-62}$  | 0.43356                  |
| Immune System   | $2.8 \times 10^{-62}$  | 0.23052                  |

Over-representation (hypergeometric test) and Permutation p-values adjusted for multiple tests across pathways (FDR). Significant pathways are marked in bold (FDR < 0.05) and italics (FDR < 0.1).

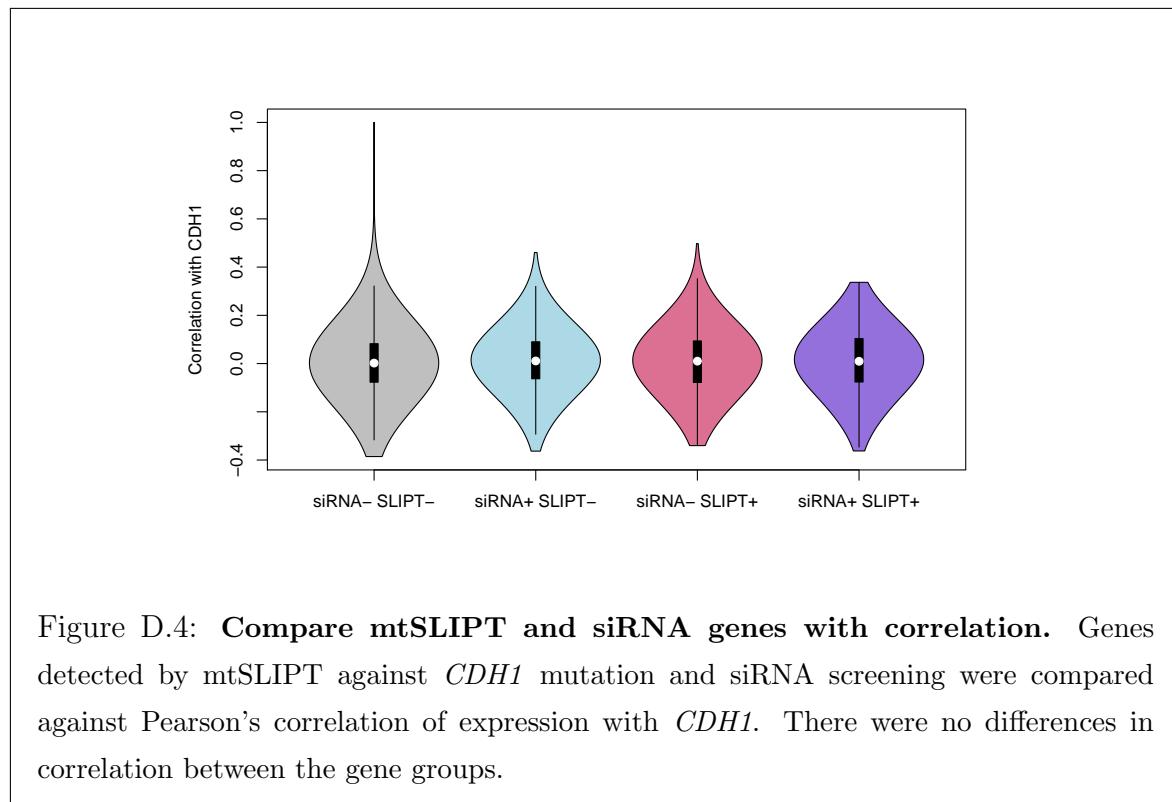
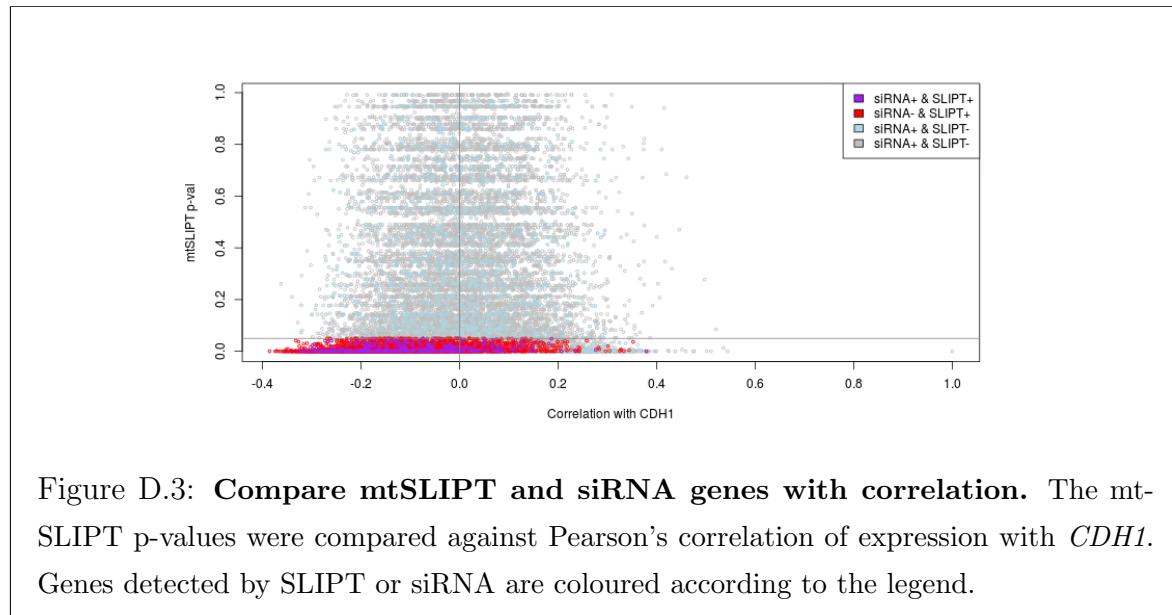
Table D.6: Pathways for *CDH1* partners from mtSLIPT and siRNA primary screen

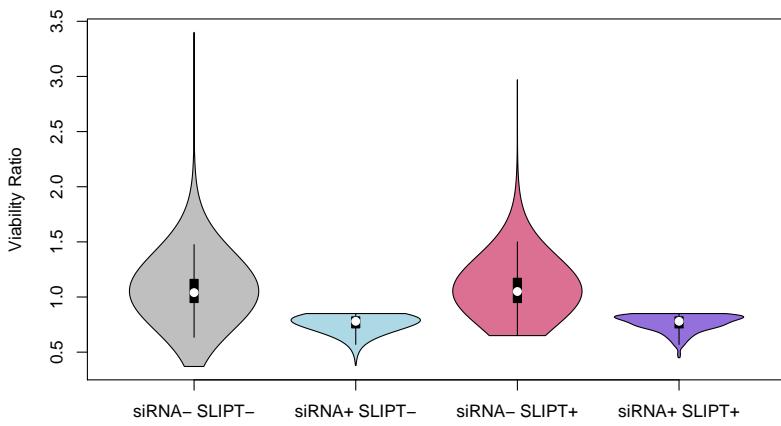
| Reactome Pathway  | Over-representation  | Permutation |
|---|----------------------|-------------|
| Visual phototransduction  | $1.2 \times 10^{-9}$ | 0.86279     |
| <b>G<sub>αs</sub> signalling events</b>                         | $2.9 \times 10^{-7}$ | 0.023066    |
| Retinoid metabolism and transport                               | $2.9 \times 10^{-7}$ | 0.299       |
| Acylic chain remodelling of PS                                  | $1.1 \times 10^{-5}$ | 0.42584     |
| Transcriptional regulation of white adipocyte differentiation   | $1.1 \times 10^{-5}$ | 0.53928     |
| Chemokine receptors bind chemokines                             | $1.1 \times 10^{-5}$ | 0.95259     |
| <i>Signaling by NOTCH4</i>                                      | $1.2 \times 10^{-5}$ | 0.079229    |
| Defective EXT2 causes exostoses 2                               | $1.2 \times 10^{-5}$ | 0.22292     |
| Defective EXT1 causes exostoses 1, TRPS2 and CHDS               | $1.2 \times 10^{-5}$ | 0.22292     |
| Platelet activation, signaling and aggregation                  | $1.2 \times 10^{-5}$ | 0.48853     |
| Serotonin receptors   | $1.4 \times 10^{-5}$ | 0.34596     |
| Nicotinamide salvaging  | $1.4 \times 10^{-5}$ | 0.70881     |
| Phase 1 - Functionalization of compounds                        | $2 \times 10^{-5}$   | 0.31142     |
| Amine ligand-binding receptors                                  | $2.5 \times 10^{-5}$ | 0.34934     |
| Acylic chain remodelling of PE                                  | $3.8 \times 10^{-5}$ | 0.42615     |
| Signaling by GPCR   | $3.8 \times 10^{-5}$ | 0.93888     |
| <b>Molecules associated with elastic fibres</b>                 | $3.9 \times 10^{-5}$ | 0.017982    |
| DAP12 interactions  | $3.9 \times 10^{-5}$ | 0.71983     |
| Beta defensins  | $3.9 \times 10^{-5}$ | 0.91458     |
| Cytochrome P <sub>450</sub> - arranged by substrate type        | $4.7 \times 10^{-5}$ | 0.83493     |
| GPCR ligand binding   | $5.7 \times 10^{-5}$ | 0.95258     |
| Acylic chain remodelling of PC                                  | $6.1 \times 10^{-5}$ | 0.42584     |
| Response to elevated platelet cytosolic Ca <sup>2+</sup>        | $6.4 \times 10^{-5}$ | 0.54046     |
| <b>Arachidonic acid metabolism</b>                              | $6.7 \times 10^{-5}$ | 0.026696    |
| Defective B4GALT7 causes EDS, progeroid type                    | $7.3 \times 10^{-5}$ | 0.24921     |
| Defective B3GAT3 causes JDSSDHD                                 | $7.3 \times 10^{-5}$ | 0.24921     |
| Hydrolysis of LPC   | $7.3 \times 10^{-5}$ | 0.80663     |
| <b>Elastic fibre formation</b>                                  | $7.4 \times 10^{-5}$ | 0.0058768   |
| <b>HS-GAG degradation</b>                                       | $9.4 \times 10^{-5}$ | 0.0083179   |
| <i>Bile acid and bile salt metabolism</i>                       | $9.4 \times 10^{-5}$ | 0.079905    |
| Netrin-1 signaling  | 0.00011              | 0.92216     |
| <b>Integration of energy metabolism</b>                         | 0.00011              | 0.011152    |
| Dectin-2 family   | 0.00012              | 0.10385     |
| Platelet sensitization by LDL                                   | 0.00012              | 0.34596     |
| DAP12 signaling   | 0.00012              | 0.62787     |
| Defensins   | 0.00012              | 0.77542     |
| GPCR downstream signaling                                       | 0.00012              | 0.79454     |
| <i>Diseases associated with glycosaminoglycan metabolism</i>    | 0.00013              | 0.065927    |
| <i>Diseases of glycosylation</i>                                | 0.00013              | 0.065927    |
| Signaling by Retinoic Acid                                      | 0.00013              | 0.22292     |
| Signaling by Leptin   | 0.00013              | 0.34596     |
| Signaling by SCF-KIT  | 0.00013              | 0.70881     |
| Opioid Signalling   | 0.00013              | 0.96053     |
| Signaling by NOTCH  | 0.00015              | 0.26884     |
| Platelet homeostasis  | 0.00015              | 0.4878      |
| Signaling by NOTCH1   | 0.00016              | 0.13043     |
| Class B/2 (Secretin family receptors)                           | 0.00016              | 0.13994     |
| <i>Diseases of Immune System</i>                                | 0.0002               | 0.0795      |
| <i>Diseases associated with the TLR signaling cascade</i>       | 0.0002               | 0.0795      |
| A tetrasaccharide linker sequence is required for GAG synthesis | 0.0002               | 0.42615     |

Over-representation (hypergeometric test) and Permutation p-values adjusted for multiple tests across pathways (FDR). Significant pathways are marked in bold (FDR < 0.05) and italics (FDR < 0.1).

## D.4 Compare SLIPT genes

The mutation synthetic lethal partners with *CDH1* were also compared to siRNA primary screen data (Telford *et al.*, 2015), by correlation and siRNA viability as described in sections 4.2.1.1 and 4.2.1.2.





**Figure D.5: Compare mtSLIPT and siRNA genes with siRNA viability.** Genes detected as candidate synthetic lethal partners by mtSLIPT (in TCGA breast cancer) expression analysis against *CDH1* mutation and experimental screening (with siRNA) were compared against the viability ratio of *CDH1* mutant and wildtype cells in the primary siRNA screen. There were clear no differences in viability between genes detected by mtSLIPT and those not with the differences being primarily due to viability thresholds being used to detect synthetic lethality by Telford *et al.* (2015).

## D.5 Metagene Analysis

Metagene analysis was also performed for synthetic lethal candidates for *CDH1* mutation. These are described and compared to expression analysis in Section 4.3.4.

Table D.7: Candidate synthetic lethal metagenes against *CDH1* from mtSLIPT

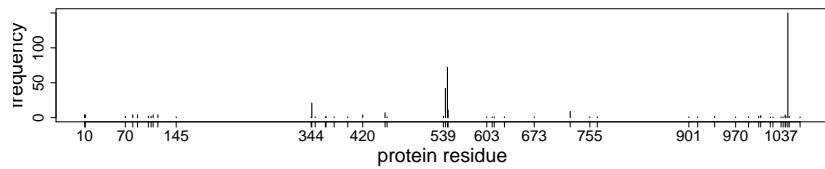
| Pathway  | ID      | Observed | Expected | $\chi^2$ value | p-value                | p-value (FDR)          |
|--|---------|----------|----------|----------------|------------------------|------------------------|
| Neurotoxicity of clostridium toxins                          | 168799  | 8        | 36.7     | 79.4           | $5.71 \times 10^{-18}$ | $3.14 \times 10^{-15}$ |
| Aquaporin-mediated transport                                 | 445717  | 8        | 36.7     | 76.3           | $2.73 \times 10^{-17}$ | $9.01 \times 10^{-15}$ |
| Toxicity of botulinum toxin type G (BoNT/G)                  | 5250989 | 8        | 36.7     | 76.3           | $2.73 \times 10^{-17}$ | $9.01 \times 10^{-15}$ |
| ABC-family proteins mediated transport                       | 382556  | 10       | 36.7     | 68.2           | $1.58 \times 10^{-15}$ | $1.86 \times 10^{-13}$ |
| G <sub>αz</sub> signalling events                            | 418597  | 10       | 36.7     | 59.9           | $9.97 \times 10^{-14}$ | $5.48 \times 10^{-12}$ |
| Regulation of IGF transport and uptake by IGFBPs             | 381426  | 9        | 36.7     | 56.3           | $5.88 \times 10^{-13}$ | $2.11 \times 10^{-11}$ |
| GP1b-IX-V activation signalling                              | 430116  | 8        | 36.7     | 55.7           | $8.20 \times 10^{-13}$ | $2.76 \times 10^{-11}$ |
| GABA receptor activation                                     | 977443  | 12       | 36.7     | 55.1           | $1.07 \times 10^{-12}$ | $3.26 \times 10^{-11}$ |
| Vasopressin regulates renal water homeostasis via Aquaporins | 432040  | 9        | 36.7     | 54.1           | $1.77 \times 10^{-12}$ | $4.88 \times 10^{-11}$ |
| Toxicity of botulinum toxin type D (BoNT/D)                  | 5250955 | 14       | 36.7     | 53.4           | $2.54 \times 10^{-12}$ | $6.64 \times 10^{-11}$ |
| Toxicity of botulinum toxin type F (BoNT/F)                  | 5250981 | 14       | 36.7     | 53.4           | $2.54 \times 10^{-12}$ | $6.64 \times 10^{-11}$ |
| STAT6-mediated induction of chemokines                       | 3249367 | 16       | 36.7     | 52.2           | $4.72 \times 10^{-12}$ | $1.13 \times 10^{-10}$ |
| Toxicity of botulinum toxin type B (BoNT/B)                  | 5250958 | 14       | 36.7     | 50.8           | $9.5 \times 10^{-12}$  | $1.98 \times 10^{-10}$ |
| S6K1 signalling  | 165720  | 12       | 36.7     | 50.2           | $1.24 \times 10^{-11}$ | $2.5 \times 10^{-10}$  |
| G <sub>αs</sub> signalling events                            | 418555  | 11       | 36.7     | 49.2           | $2.08 \times 10^{-11}$ | $3.85 \times 10^{-10}$ |
| RHO GTPases activate CIT                                     | 5625900 | 14       | 36.7     | 48.2           | $3.34 \times 10^{-11}$ | $5.9 \times 10^{-10}$  |
| NADE modulates death signalling                              | 205025  | 15       | 36.7     | 47.4           | $5.00 \times 10^{-11}$ | $8.32 \times 10^{-10}$ |
| Keratan sulfate degradation                                  | 2022857 | 10       | 36.7     | 46.6           | $7.5 \times 10^{-11}$  | $1.15 \times 10^{-9}$  |
| Signaling by Retinoic Acid                                   | 5362517 | 10       | 36.7     | 46.6           | $7.5 \times 10^{-11}$  | $1.15 \times 10^{-9}$  |
| Adenylate cyclase inhibitory pathway                         | 170670  | 14       | 36.7     | 45.9           | $1.11 \times 10^{-10}$ | $1.59 \times 10^{-9}$  |
| Inhibition of adenylate cyclase pathway                      | 997269  | 14       | 36.7     | 45.9           | $1.11 \times 10^{-10}$ | $1.59 \times 10^{-9}$  |
| Fatty acids  | 211935  | 6        | 36.7     | 45.7           | $1.21 \times 10^{-10}$ | $1.72 \times 10^{-9}$  |
| Ionotropic activity of Kainate Receptors                     | 451306  | 13       | 36.7     | 44.6           | $2.03 \times 10^{-10}$ | $2.58 \times 10^{-9}$  |
| Activation of Ca-permeable Kainate Receptor                  | 451308  | 13       | 36.7     | 44.6           | $2.03 \times 10^{-10}$ | $2.58 \times 10^{-9}$  |
| RA biosynthesis pathway                                      | 5365859 | 13       | 36.7     | 44.6           | $2.03 \times 10^{-10}$ | $2.58 \times 10^{-9}$  |

Strongest candidate SL partners for *CDH1* by mtSLIPT with observed and expected mutant samples with low expression of partner metagenes

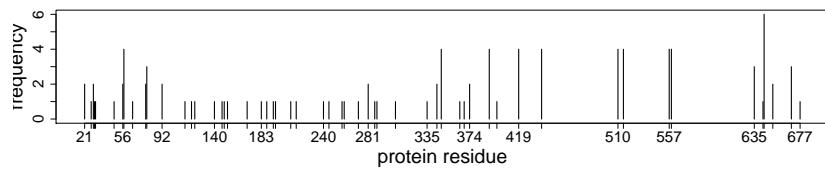
## D.6 Mutation Variation

Mutations have different effects as shown by the following examples in cancer genes.

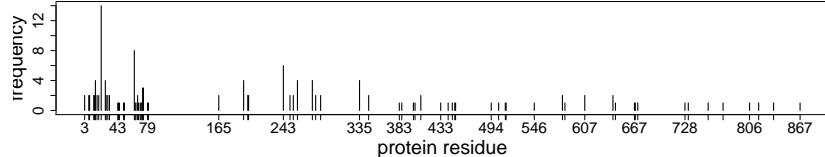
### D.6.1 Mutation Frequency



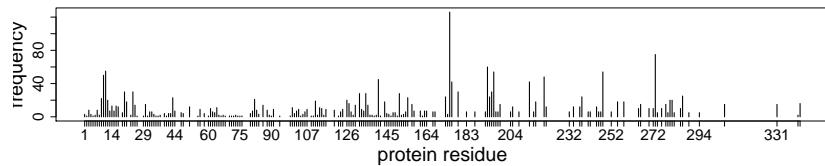
(a) *PI3KCA*



(b) *PI3KR1*



(c) *CDH1*



(d) *TP53*

Figure D.6: **Somatic mutation locus.** Mutation frequency at each locus in TCGA breast cancer. *PIK3CA* shows clear recurrent E545K and H1047R oncogene mutations consistent with it being an oncogene. *PIK3R1* and *CDH1* are tumour suppressors with inactivating mutations distributed throughout the gene, whereas *TP53* exhibits both of these properties and a very high mutation frequency compared to other genes.

## D.6.2 PI3K Mutation Expression

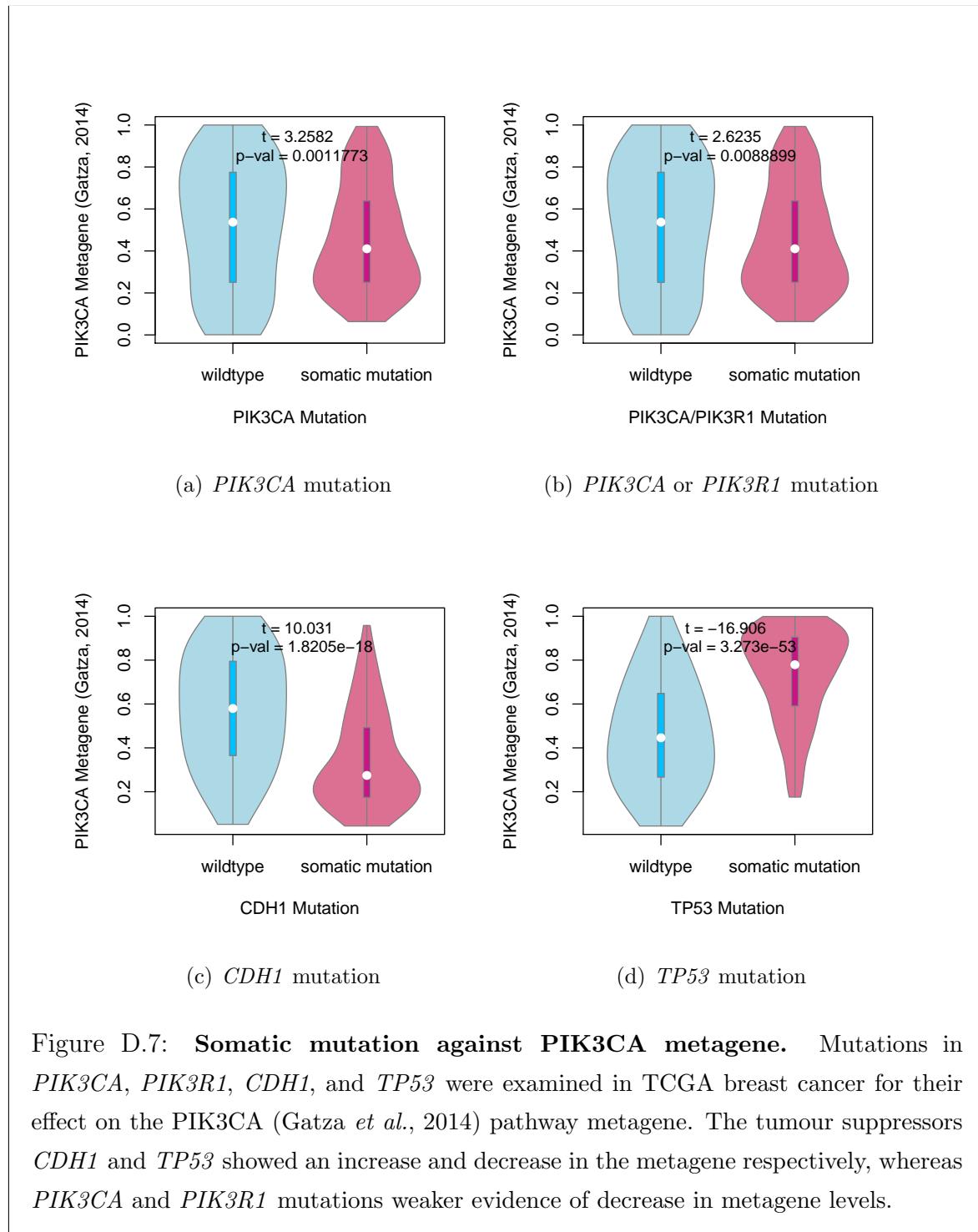
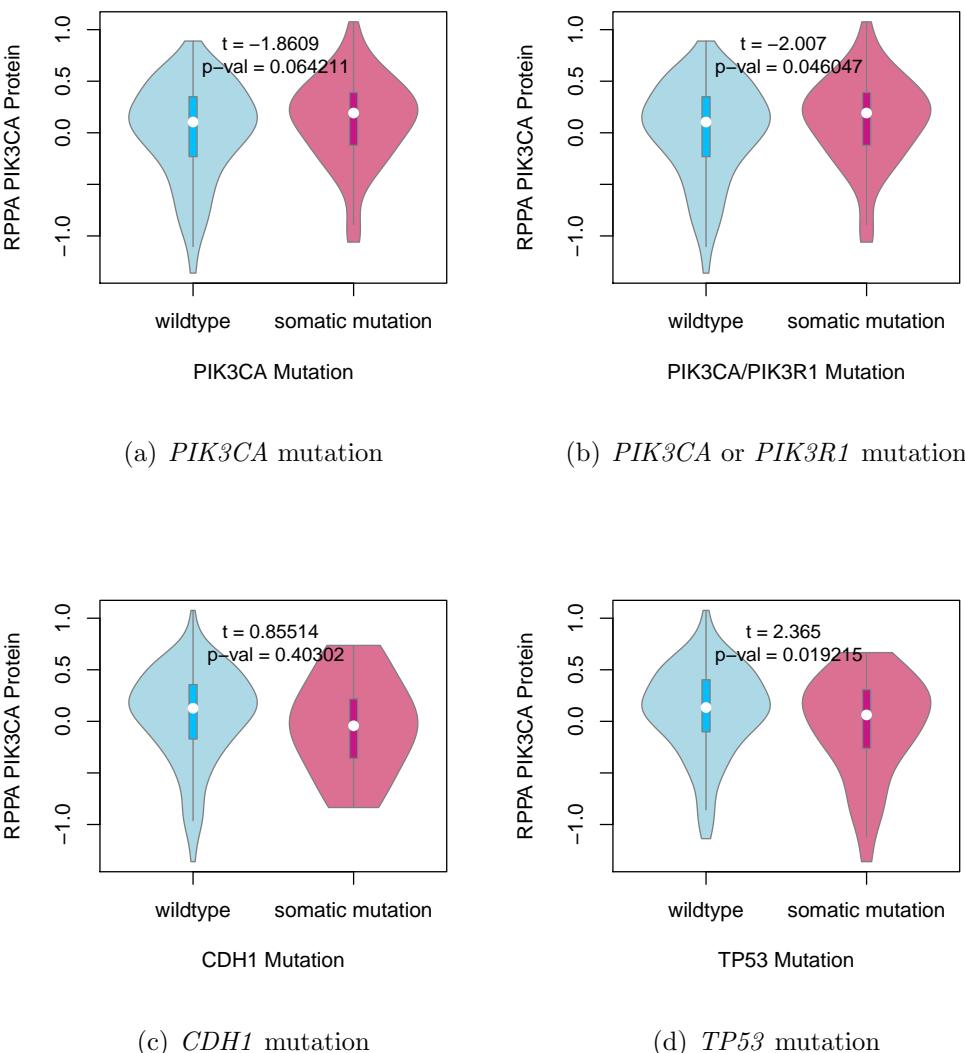
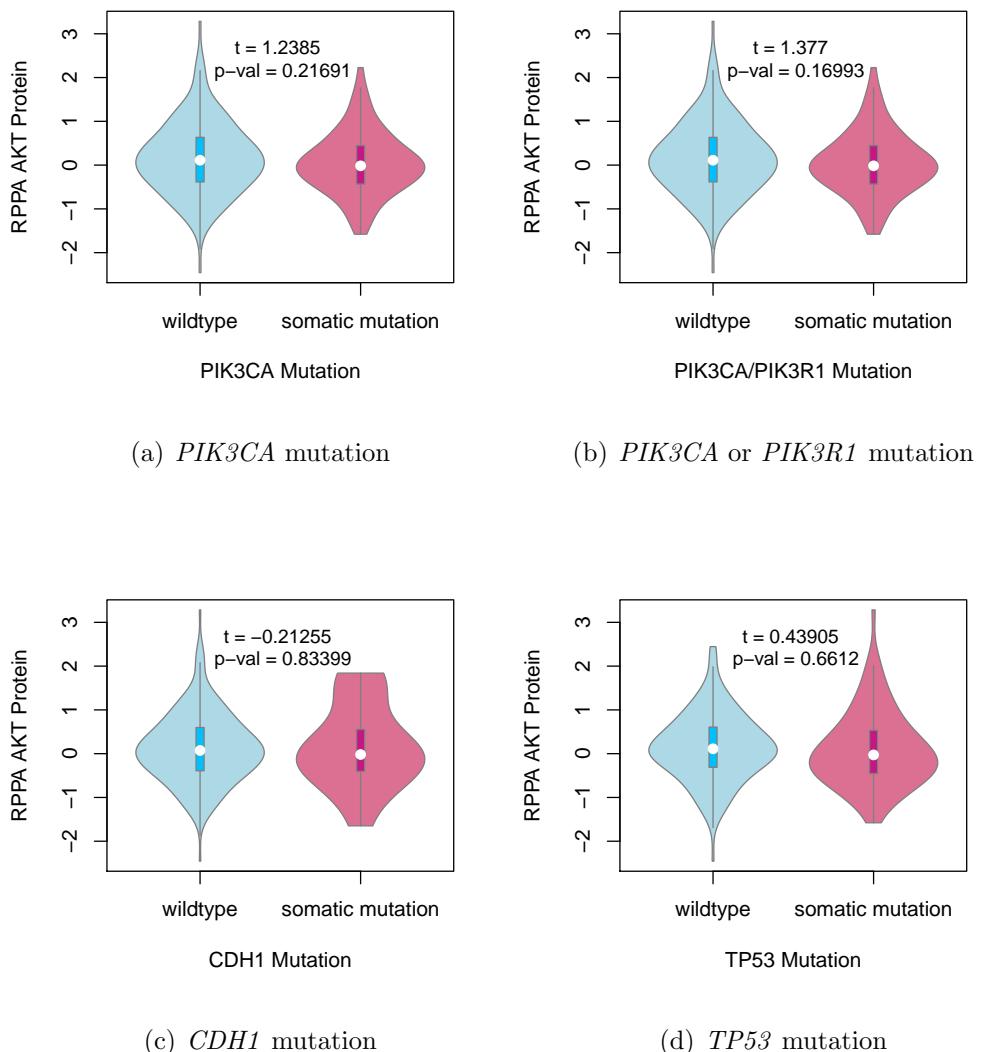


Figure D.7: **Somatic mutation against PIK3CA metagene.** Mutations in *PIK3CA*, *PIK3R1*, *CDH1*, and *TP53* were examined in TCGA breast cancer for their effect on the PIK3CA (Gatza *et al.*, 2014) pathway metagene. The tumour suppressors *CDH1* and *TP53* showed an increase and decrease in the metagene respectively, whereas *PIK3CA* and *PIK3R1* mutations weaker evidence of decrease in metagene levels.



**Figure D.8: Somatic mutation against PI3K protein.** Mutations in *PIK3CA*, *PIK3R1*, *CDH1*, and *TP53* were examined in TCGA breast cancer for their effect on the expression of the p110 $\alpha$  protein (encoded by *PIK3CA*). Protein levels were significantly elevated in samples with *PIK3CA* or *PIK3R1* mutations and lower in samples with *TP53* mutations.



**Figure D.9: Somatic mutation against AKT protein.** Mutations in *PIK3CA*, *PIK3R1*, *CDH1*, and *TP53* were examined in TCGA breast cancer for their effect on the expression of the AKT protein (a downstream target of *PIK3CA*). Protein levels were not significantly different in samples mutations in any of these cancer genes.

## **Appendix E**

### **Metagene Expression Profiles**

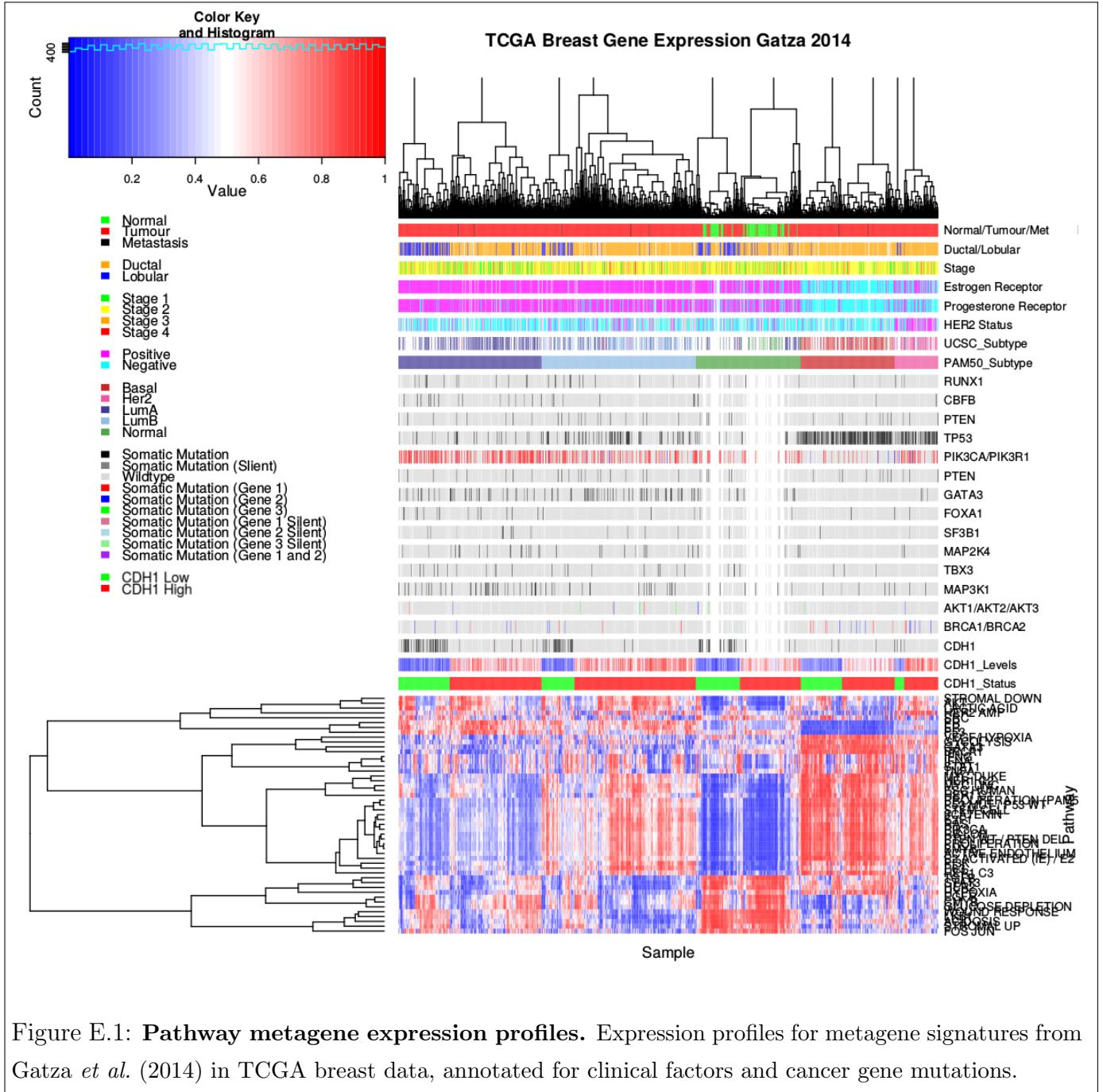
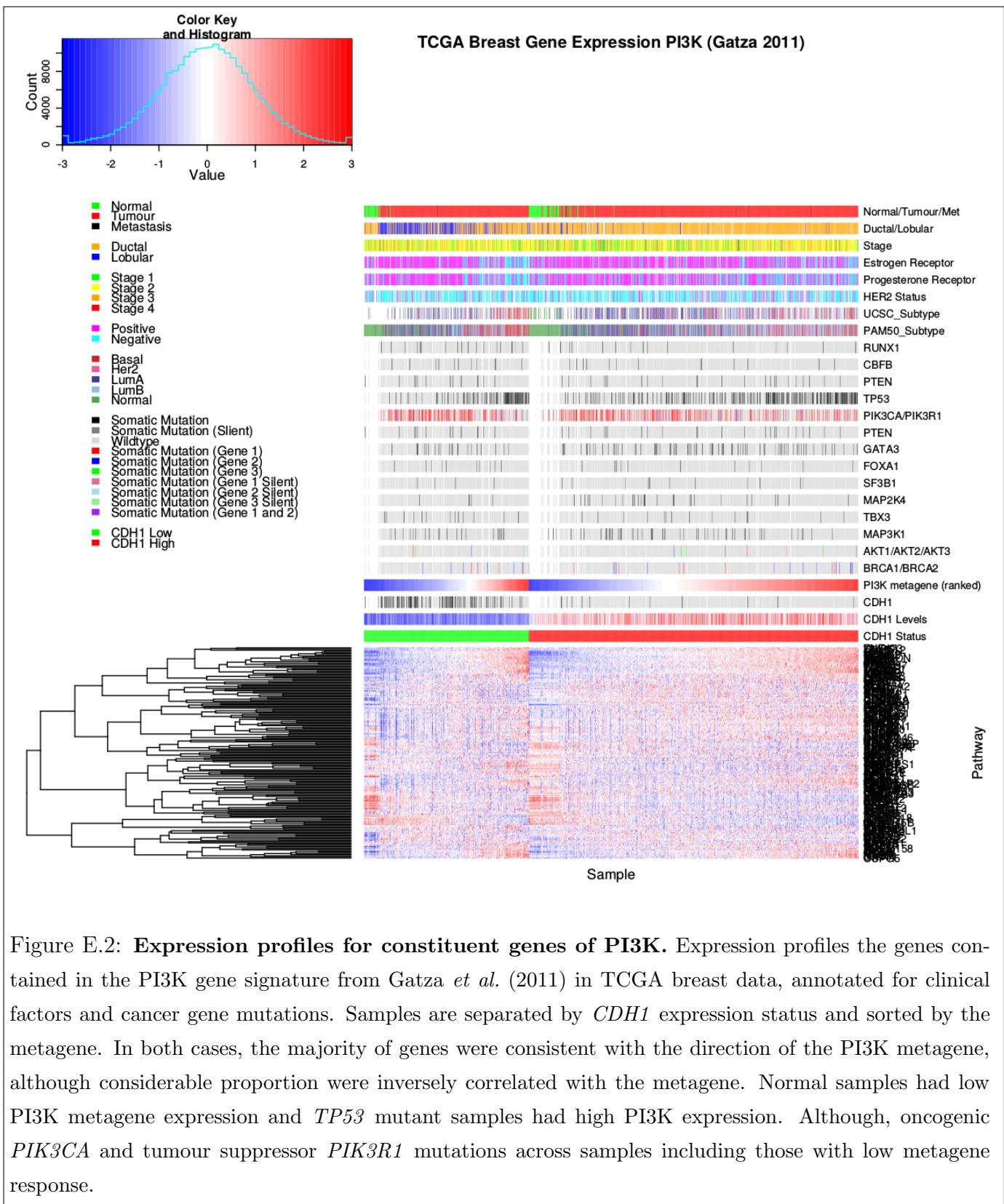
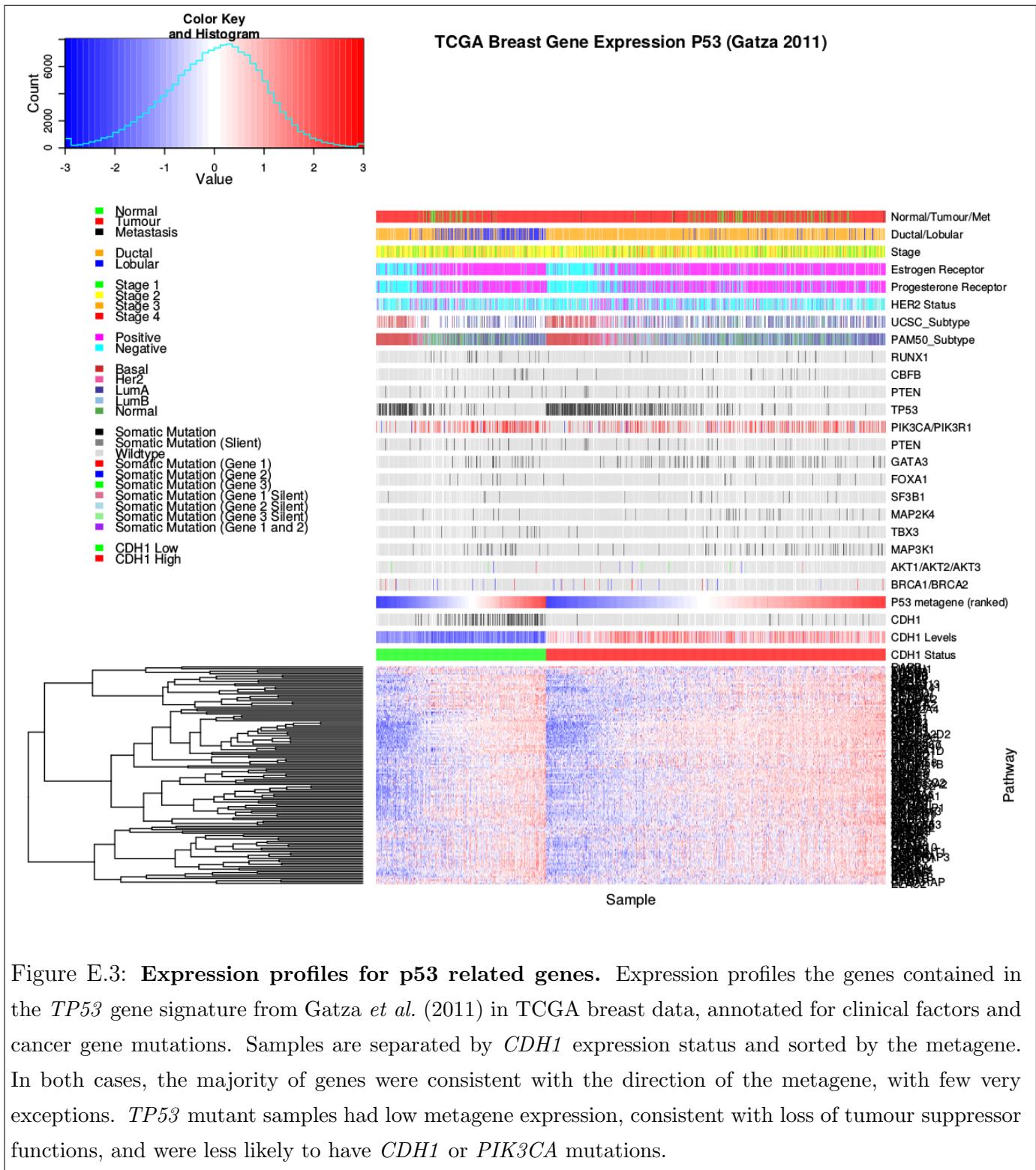


Figure E.1: **Pathway metagene expression profiles.** Expression profiles for metagene signatures from Gatz *et al.* (2014) in TCGA breast data, annotated for clinical factors and cancer gene mutations.





**Figure E.3: Expression profiles for p53 related genes.** Expression profiles the genes contained in the *TP53* gene signature from Gatza *et al.* (2011) in TCGA breast data, annotated for clinical factors and cancer gene mutations. Samples are separated by *CDH1* expression status and sorted by the metagene. In both cases, the majority of genes were consistent with the direction of the metagene, with few very exceptions. *TP53* mutant samples had low metagene expression, consistent with loss of tumour suppressor functions, and were less likely to have *CDH1* or *PIK3CA* mutations.

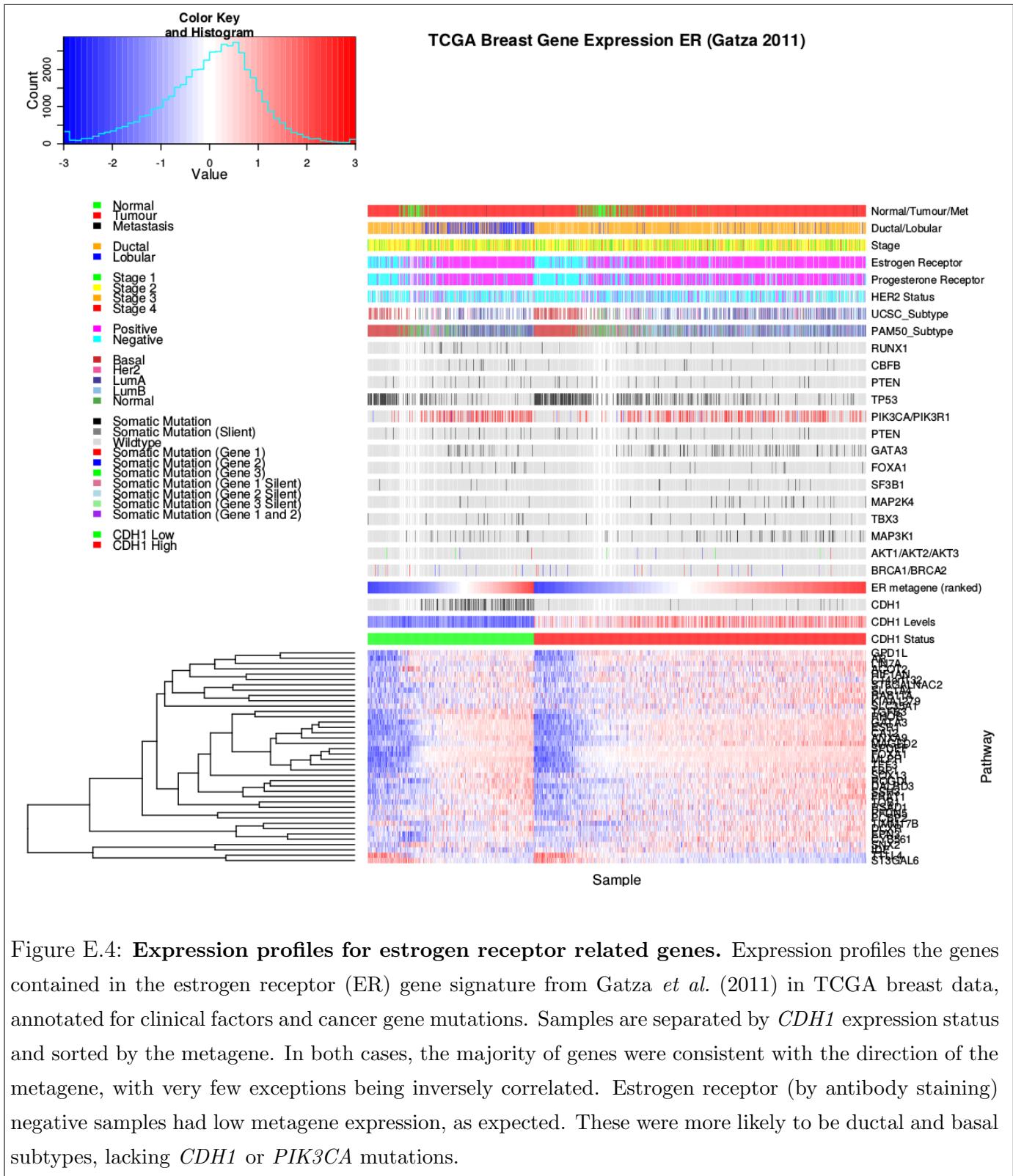
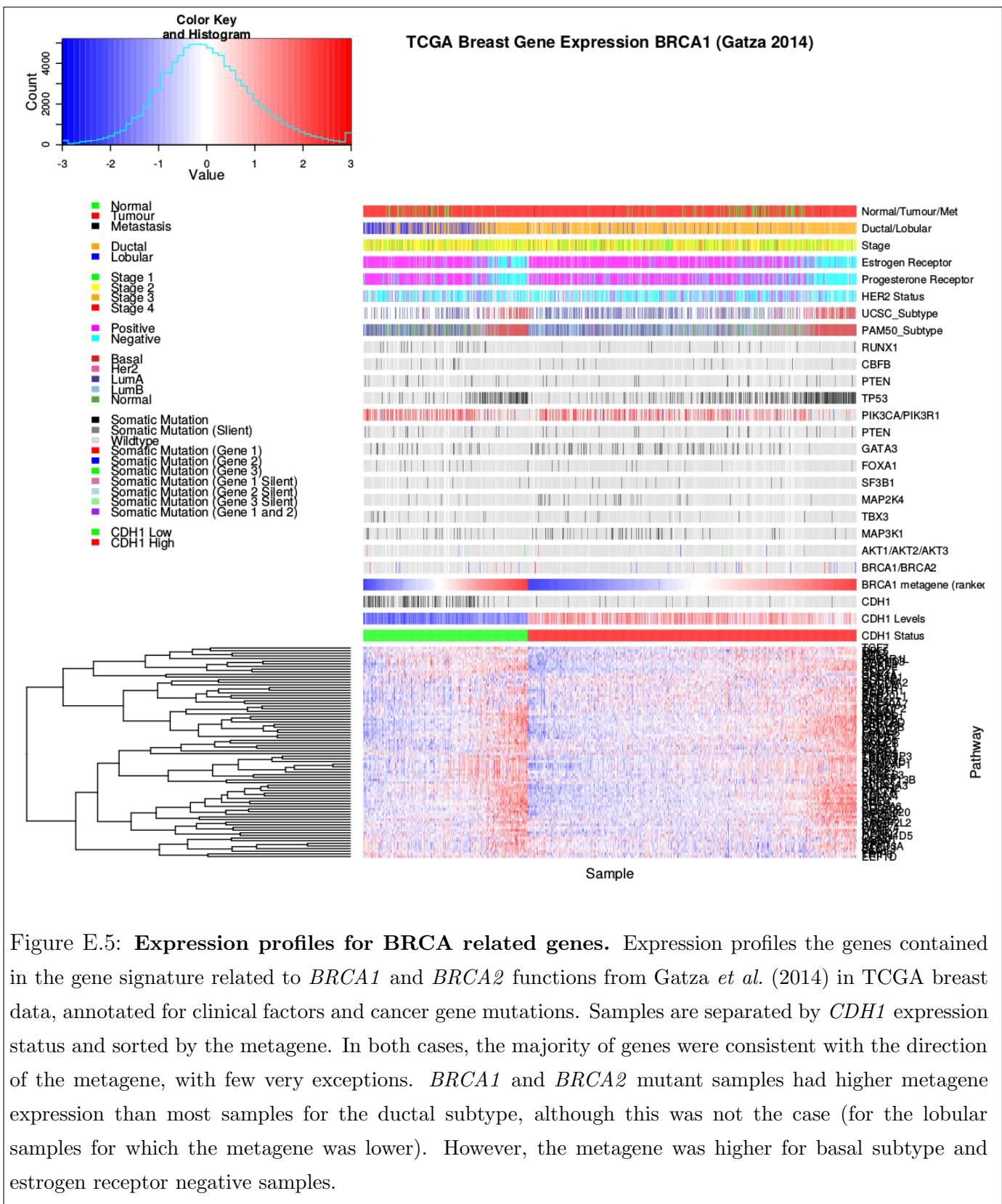


Figure E.4: **Expression profiles for estrogen receptor related genes.** Expression profiles the genes contained in the estrogen receptor (ER) gene signature from Gatza *et al.* (2011) in TCGA breast data, annotated for clinical factors and cancer gene mutations. Samples are separated by *CDH1* expression status and sorted by the metagene. In both cases, the majority of genes were consistent with the direction of the metagene, with very few exceptions being inversely correlated. Estrogen receptor (by antibody staining) negative samples had low metagene expression, as expected. These were more likely to be ductal and basal subtypes, lacking *CDH1* or *PIK3CA* mutations.



# **Appendix F**

## **Stomach Expression Analysis**

The following results are a replication of the TCGA results (in Chapter 4) with stomach cancer data, using synthetic lethality (SLIPT) against *CDH1* mutation.

### **F.1 Synthetic Lethal Genes and Pathways**

Table F.1: Synthetic lethal gene partners of *CDH1* from SLIPT in stomach cancer

| Gene             | Observed | Expected | $\chi^2$ value | p-value                | p-value (FDR)          |
|------------------|----------|----------|----------------|------------------------|------------------------|
| <i>PRAF2</i>     | 17       | 50.4     | 121            | $3.54 \times 10^{-25}$ | $1.45 \times 10^{-21}$ |
| <i>EMP3</i>      | 17       | 50.4     | 115            | $5.06 \times 10^{-24}$ | $1.48 \times 10^{-20}$ |
| <i>PLEKHO1</i>   | 22       | 50.4     | 112            | $2.14 \times 10^{-23}$ | $4.75 \times 10^{-20}$ |
| <i>SELM</i>      | 20       | 50.4     | 111            | $5.13 \times 10^{-23}$ | $8.09 \times 10^{-20}$ |
| <i>GYPC</i>      | 20       | 50.4     | 110            | $5.77 \times 10^{-23}$ | $8.45 \times 10^{-20}$ |
| <i>COX7A1</i>    | 18       | 50.4     | 109            | $1.15 \times 10^{-22}$ | $1.39 \times 10^{-19}$ |
| <i>TNFSF12</i>   | 20       | 50.4     | 106            | $4.06 \times 10^{-22}$ | $4.38 \times 10^{-19}$ |
| <i>SEPT4</i>     | 17       | 50.4     | 106            | $6.58 \times 10^{-22}$ | $5.91 \times 10^{-19}$ |
| <i>LGALS1</i>    | 19       | 50.4     | 105            | $6.64 \times 10^{-22}$ | $5.91 \times 10^{-19}$ |
| <i>RARRES2</i>   | 27       | 50.4     | 105            | $8.02 \times 10^{-22}$ | $6.85 \times 10^{-19}$ |
| <i>VEGFB</i>     | 16       | 50.4     | 104            | $1.19 \times 10^{-21}$ | $9.74 \times 10^{-19}$ |
| <i>PRR24</i>     | 22       | 50.4     | 102            | $2.96 \times 10^{-21}$ | $2.02 \times 10^{-18}$ |
| <i>SYNC</i>      | 19       | 50.4     | 102            | $3.73 \times 10^{-21}$ | $2.39 \times 10^{-18}$ |
| <i>MAGEH1</i>    | 17       | 50.4     | 100            | $9.52 \times 10^{-21}$ | $5.01 \times 10^{-18}$ |
| <i>HSPB2</i>     | 23       | 50.4     | 99.6           | $1.19 \times 10^{-20}$ | $5.82 \times 10^{-18}$ |
| <i>SMARCD3</i>   | 19       | 50.4     | 99             | $1.59 \times 10^{-20}$ | $7.57 \times 10^{-18}$ |
| <i>CREM</i>      | 13       | 50.4     | 98.1           | $2.48 \times 10^{-20}$ | $1.13 \times 10^{-17}$ |
| <i>GNG11</i>     | 20       | 50.4     | 97.3           | $3.68 \times 10^{-20}$ | $1.59 \times 10^{-17}$ |
| <i>GNAI2</i>     | 17       | 50.4     | 96.4           | $5.75 \times 10^{-20}$ | $2.36 \times 10^{-17}$ |
| <i>FUNDC2</i>    | 22       | 50.4     | 95.9           | $7.39 \times 10^{-20}$ | $2.91 \times 10^{-17}$ |
| <i>CNRIP1</i>    | 21       | 50.4     | 95.3           | $1.0 \times 10^{-19}$  | $3.66 \times 10^{-17}$ |
| <i>CALHM2</i>    | 22       | 50.4     | 93.1           | $2.94 \times 10^{-19}$ | $1.06 \times 10^{-16}$ |
| <i>ARID5A</i>    | 18       | 50.4     | 92.7           | $3.47 \times 10^{-19}$ | $1.22 \times 10^{-16}$ |
| <i>ST3GAL3</i>   | 27       | 50.4     | 92.2           | $4.49 \times 10^{-19}$ | $1.56 \times 10^{-16}$ |
| <i>LOC339524</i> | 21       | 50.4     | 92.1           | $4.8 \times 10^{-19}$  | $1.59 \times 10^{-16}$ |

SLIPT partners of *CDH1* with observed and expected mutant samples of both genes

Table F.2: Pathway composition for clusters of *CDH1* partners in stomach SLIPT

| Pathways Over-represented in Cluster 1                              | Pathway Size | Cluster Genes | p-value (FDR)         |
|---|--------------|---------------|-----------------------|
| Viral mRNA Translation  | 82           | 48            | $1.3 \times 10^{-97}$ |
| Formation of a pool of free 40S subunits                            | 94           | 51            | $1.3 \times 10^{-97}$ |
| Eukaryotic Translation Elongation                                   | 87           | 49            | $4.8 \times 10^{-97}$ |
| Peptide chain elongation  | 84           | 48            | $1.4 \times 10^{-96}$ |
| Eukaryotic Translation Termination                                  | 84           | 48            | $1.4 \times 10^{-96}$ |
| GTP hydrolysis and joining of the 60S ribosomal subunit             | 105          | 52            | $7.9 \times 10^{-94}$ |
| Nonsense Mediated Decay independent of the Exon Junction Complex    | 89           | 48            | $3.1 \times 10^{-93}$ |
| Li3a-mediated translational silencing of Ceruloplasmin expression   | 104          | 51            | $5.1 \times 10^{-92}$ |
| 3' UTR-mediated translational regulation                            | 104          | 51            | $5.1 \times 10^{-92}$ |
| SRP-dependent cotranslational protein targeting to membrane         | 105          | 51            | $1.7 \times 10^{-91}$ |
| Eukaryotic Translation Initiation                                   | 112          | 52            | $3.3 \times 10^{-90}$ |
| Cap-dependent Translation Initiation                                | 112          | 52            | $3.3 \times 10^{-90}$ |
| Translation   | 142          | 56            | $3.6 \times 10^{-85}$ |
| Nonsense-Mediated Decay   | 104          | 48            | $1.2 \times 10^{-84}$ |
| Nonsense Mediated Decay enhanced by the Exon Junction Complex       | 104          | 48            | $1.2 \times 10^{-84}$ |
| Influenza Viral RNA Transcription and Replication                   | 109          | 48            | $4.1 \times 10^{-82}$ |
| Influenza Life Cycle  | 113          | 48            | $3.4 \times 10^{-80}$ |
| Influenza Infection   | 118          | 48            | $6.4 \times 10^{-78}$ |
| Infections diseases   | 349          | 68            | $1.8 \times 10^{-50}$ |
| Formation of the ternary complex, and subsequently, the 43S complex | 48           | 21            | $3.7 \times 10^{-43}$ |

| Pathways Over-represented in Cluster 2                                   | Pathway Size | Cluster Genes | p-value (FDR)         |
|--|--------------|---------------|-----------------------|
| Immunoregulatory interactions between a Lymphoid and a non-Lymphoid cell | 65           | 12            | $1.3 \times 10^{-15}$ |
| Phosphorylation of CD3 and TCR zeta chains                               | 18           | 6             | $1.7 \times 10^{-12}$ |
| Generation of second messenger molecules                                 | 29           | 7             | $2.7 \times 10^{-12}$ |
| PD-1 signaling   | 21           | 6             | $7.4 \times 10^{-12}$ |
| TCR signaling  | 62           | 9             | $4.3 \times 10^{-11}$ |
| Translocation of ZAP-70 to Immunological synapse                         | 16           | 5             | $1.1 \times 10^{-10}$ |
| Interferon alpha/beta signaling  | 68           | 9             | $1.6 \times 10^{-10}$ |
| Initial triggering of complement   | 17           | 5             | $1.6 \times 10^{-10}$ |
| IKK complex recruitment mediated by RIP1                                 | 19           | 5             | $5.1 \times 10^{-10}$ |
| TRIF-mediated programmed cell death                                      | 10           | 4             | $6.2 \times 10^{-10}$ |
| Creation of C4 and C2 activators   | 11           | 4             | $1.3 \times 10^{-9}$  |
| RHO GTPases Activate NADPH Oxidases                                      | 11           | 4             | $1.3 \times 10^{-9}$  |
| Interferon Signaling   | 175          | 15            | $2.3 \times 10^{-9}$  |
| Chemokine receptors bind chemokines                                      | 52           | 7             | $4.0 \times 10^{-9}$  |
| Interferon gamma signaling   | 74           | 8             | $1.6 \times 10^{-8}$  |
| TRAF6 mediated induction of TAK1 complex                                 | 15           | 4             | $1.6 \times 10^{-8}$  |
| Activation of IRF3/IRF7 mediated by TBK1/IKK epsilon                     | 16           | 4             | $2.7 \times 10^{-8}$  |
| Downstream TCR signaling   | 45           | 6             | $3.5 \times 10^{-8}$  |
| Ligand-dependent caspase activation                                      | 17           | 4             | $4.2 \times 10^{-8}$  |
| Complement cascade   | 34           | 5             | $1.3 \times 10^{-7}$  |

| Pathways Over-represented in Cluster 3                            | Pathway Size | Cluster Genes | p-value (FDR)        |
|---|--------------|---------------|----------------------|
| Uptake and actions of bacterial toxins                            | 22           | 4             | $3.5 \times 10^{-6}$ |
| Neurotoxicity of clostridium toxins                               | 10           | 3             | $3.5 \times 10^{-6}$ |
| Activation of PPARGC1A (PGC-1alpha) by phosphorylation            | 10           | 3             | $3.5 \times 10^{-6}$ |
| SMAD2/SMAD3/SMAD4 heterotrimer regulates transcription            | 28           | 4             | $1.4 \times 10^{-5}$ |
| Assembly of the primary cilium                                    | 149          | 10            | $2.5 \times 10^{-5}$ |
| Serotonin Neurotransmitter Release Cycle                          | 15           | 3             | $2.5 \times 10^{-5}$ |
| Glycosaminoglycan metabolism                                      | 114          | 8             | $3.3 \times 10^{-5}$ |
| Platelet homeostasis  | 54           | 5             | $3.3 \times 10^{-5}$ |
| Norepinephrine Neurotransmitter Release Cycle                     | 17           | 3             | $3.3 \times 10^{-5}$ |
| Acetylcholine Neurotransmitter Release Cycle                      | 17           | 3             | $3.3 \times 10^{-5}$ |
| Gas signalling events   | 100          | 7             | $5.5 \times 10^{-5}$ |
| GABA synthesis, release, reuptake and degradation                 | 19           | 3             | $5.6 \times 10^{-5}$ |
| deactivation of the beta-catenin transactivating complex          | 39           | 4             | $6.7 \times 10^{-5}$ |
| Dopamine Neurotransmitter Release Cycle                           | 20           | 3             | $6.7 \times 10^{-5}$ |
| IRS-related events triggered by IGFIR                             | 83           | 6             | $7.1 \times 10^{-5}$ |
| Generic Transcription Pathway                                     | 186          | 11            | $7.1 \times 10^{-5}$ |
| Termination of O-glycan biosynthesis                              | 21           | 3             | $7.4 \times 10^{-5}$ |
| Kinesins  | 22           | 3             | $8.5 \times 10^{-5}$ |
| Signaling by Type 1 Insulin-like Growth Factor 1 Receptor (IGF1R) | 86           | 6             | $8.5 \times 10^{-5}$ |
| IGF1R signaling cascade   | 86           | 6             | $8.5 \times 10^{-5}$ |

| Pathways Over-represented in Cluster 4                   | Pathway Size | Cluster Genes | p-value (FDR)          |
|--|--------------|---------------|------------------------|
| Extracellular matrix organization                        | 241          | 97            | $8.8 \times 10^{-126}$ |
| Axon guidance  | 289          | 75            | $8.3 \times 10^{-72}$  |
| Hemostasis   | 445          | 101           | $8.3 \times 10^{-72}$  |
| Developmental Biology                                    | 432          | 95            | $3.0 \times 10^{-67}$  |
| Response to elevated platelet cytosolic Ca <sup>2+</sup> | 84           | 37            | $5.8 \times 10^{-67}$  |
| Platelet degranulation                                   | 79           | 36            | $5.8 \times 10^{-67}$  |
| Degradation of the extracellular matrix                  | 104          | 39            | $6.7 \times 10^{-63}$  |
| Platelet activation, signaling and aggregation           | 186          | 52            | $6.6 \times 10^{-62}$  |
| ECM proteoglycans  | 66           | 31            | $8.1 \times 10^{-61}$  |
| Neuronal System  | 272          | 64            | $5.1 \times 10^{-60}$  |
| Signaling by PDGF  | 173          | 47            | $9.7 \times 10^{-57}$  |
| Integrin cell surface interactions                       | 82           | 31            | $1.9 \times 10^{-53}$  |
| Collagen biosynthesis and modifying enzymes              | 56           | 26            | $1.1 \times 10^{-52}$  |
| Collagen formation                                       | 67           | 28            | $1.4 \times 10^{-52}$  |
| Class A/1 (Rhodopsin-like receptors)                     | 289          | 61            | $2.3 \times 10^{-52}$  |
| GPCR ligand binding                                      | 373          | 73            | $2.8 \times 10^{-52}$  |
| Elastic fibre formation                                  | 38           | 22            | $4.7 \times 10^{-52}$  |
| Non-integrin membrane-ECM interactions                   | 53           | 24            | $7.0 \times 10^{-49}$  |
| Glycosaminoglycan metabolism                             | 114          | 33            | $4.7 \times 10^{-47}$  |
| Platelet homeostasis                                     | 54           | 23            | $1.0 \times 10^{-45}$  |

## F.2 Comparison to Primary Screen

The synthetic lethal partners with *CDH1* expression in stomach cancers were also compared to siRNA primary screen data (Telford *et al.*, 2015), as performed in Section 4.2.1. These are expected to be more concordant with the experimental results performed on a null mutant, however this is not the case at the gene level: less genes overlapped with experimental candidates in Figure F.1. This may be affected by lower sample size for mutations in TCGA data or lower frequency (expected value) of *CDH1* mutations compared to low expression.

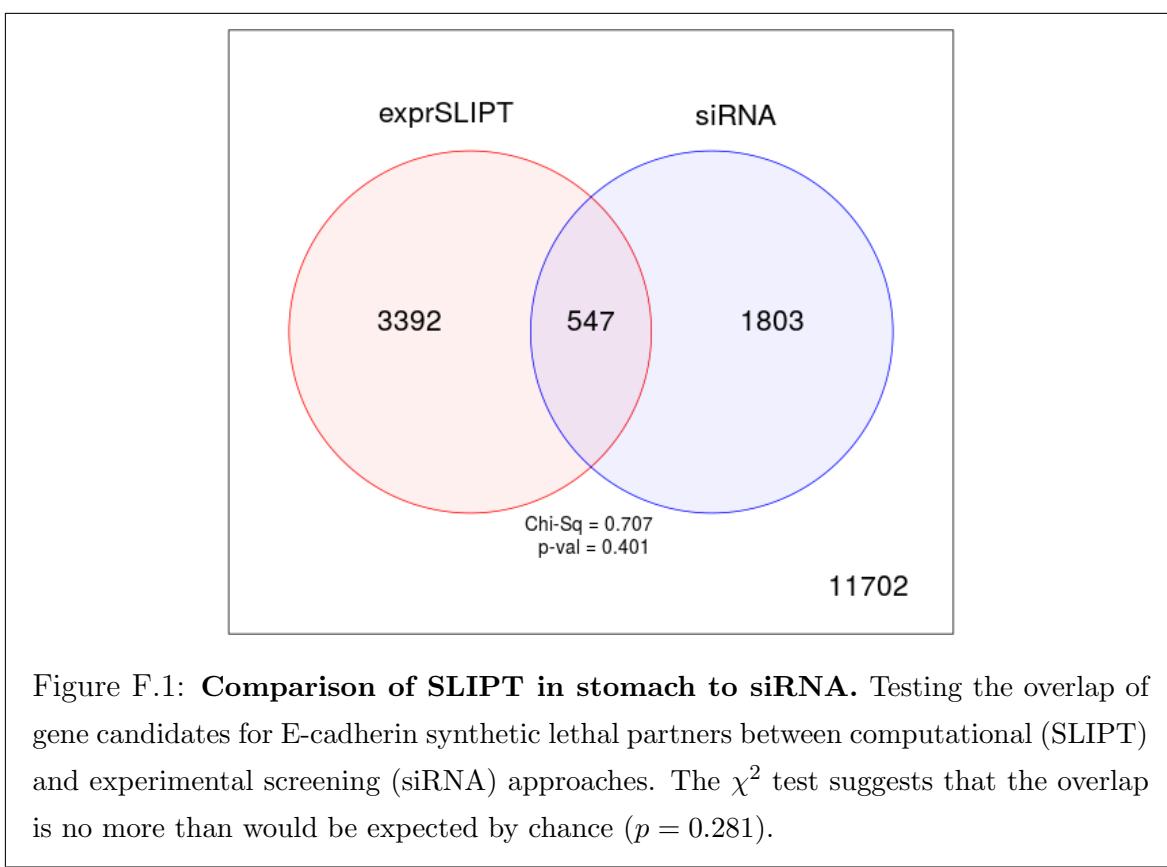


Figure F.1: **Comparison of SLIPT in stomach to siRNA.** Testing the overlap of gene candidates for E-cadherin synthetic lethal partners between computational (SLIPT) and experimental screening (siRNA) approaches. The  $\chi^2$  test suggests that the overlap is no more than would be expected by chance ( $p = 0.281$ ).

Table F.3: Pathway composition for *CDH1* partners from SLIPT and siRNA screening

| Predicted only by SLIPT (3392 genes)                              | Pathway | Size | Genes Identified | p-value (FDR)          |
|---|---------|------|------------------|------------------------|
| Extracellular matrix organization                                 |         | 238  | 90               | $3.4 \times 10^{-107}$ |
| Eukaryotic Translation Termination                                |         | 79   | 46               | $7.6 \times 10^{-91}$  |
| Viral mRNA Translation  |         | 77   | 45               | $1.2 \times 10^{-89}$  |
| Eukaryotic Translation Elongation                                 |         | 82   | 46               | $5.8 \times 10^{-89}$  |
| Peptide chain elongation  |         | 79   | 45               | $2.1 \times 10^{-88}$  |
| Nonsense Mediated Decay independent of the Exon Junction Complex  |         | 84   | 46               | $9.4 \times 10^{-88}$  |
| Formation of a pool of free 40S subunits                          |         | 89   | 47               | $3.3 \times 10^{-87}$  |
| GTP hydrolysis and joining of the 60S ribosomal subunit           |         | 100  | 48               | $3.2 \times 10^{-83}$  |
| Axon guidance   |         | 284  | 84               | $3.9 \times 10^{-82}$  |
| Developmental Biology   |         | 426  | 111              | $4.2 \times 10^{-82}$  |
| L13a-mediated translational silencing of Ceruloplasmin expression |         | 99   | 47               | $1.4 \times 10^{-81}$  |
| 3' -UTR-mediated translational regulation                         |         | 99   | 47               | $1.4 \times 10^{-81}$  |
| SRP-dependent cotranslational protein targeting to membrane       |         | 99   | 47               | $1.4 \times 10^{-81}$  |
| Nonsense-Mediated Decay   |         | 99   | 47               | $1.4 \times 10^{-81}$  |
| Nonsense Mediated Decay enhanced by the Exon Junction Complex     |         | 99   | 47               | $1.4 \times 10^{-81}$  |
| Hemostasis  |         | 438  | 112              | $1.2 \times 10^{-80}$  |
| Eukaryotic Translation Initiation                                 |         | 107  | 48               | $8.0 \times 10^{-80}$  |
| Cap-dependent Translation Initiation                              |         | 107  | 48               | $8.0 \times 10^{-80}$  |
| Infectious disease  |         | 338  | 90               | $1.6 \times 10^{-76}$  |
| Neuronal System   |         | 267  | 77               | $1.6 \times 10^{-76}$  |

| Detected only by siRNA screen (1803 genes)       | Pathway | Size | Genes Identified | p-value (FDR)         |
|--|---------|------|------------------|-----------------------|
| Class A/1 (Rhodopsin-like receptors)             |         | 282  | 62               | $8.1 \times 10^{-50}$ |
| GPCR ligand binding                              |         | 363  | 71               | $4.9 \times 10^{-46}$ |
| Peptide ligand-binding receptors                 |         | 175  | 38               | $7.9 \times 10^{-38}$ |
| <i>Gαi</i> signalling events                     |         | 184  | 37               | $1.1 \times 10^{-34}$ |
| Gastrin-CREB signalling pathway via PKC and MAPK |         | 180  | 35               | $1.4 \times 10^{-32}$ |
| <i>Gαq</i> signalling events                     |         | 159  | 32               | $4.8 \times 10^{-32}$ |
| DAP12 interactions                               |         | 159  | 29               | $1.4 \times 10^{-27}$ |
| Downstream signal transduction                   |         | 146  | 26               | $2.4 \times 10^{-25}$ |
| DAP12 signaling                                  |         | 149  | 26               | $6.4 \times 10^{-25}$ |
| VEGFA-VEGFR2 Pathway                             |         | 91   | 19               | $8.1 \times 10^{-24}$ |
| Signaling by PDGF                                |         | 172  | 27               | $5.7 \times 10^{-23}$ |
| Signaling by ERBB2                               |         | 146  | 24               | $1.4 \times 10^{-22}$ |
| Signaling by VEGF                                |         | 99   | 19               | $2.0 \times 10^{-22}$ |
| Visual phototransduction                         |         | 85   | 17               | $1.3 \times 10^{-21}$ |
| Downstream signaling of activated FGFR1          |         | 134  | 22               | $1.3 \times 10^{-21}$ |
| Downstream signaling of activated FGFR2          |         | 134  | 22               | $1.3 \times 10^{-21}$ |
| Downstream signaling of activated FGFR3          |         | 134  | 22               | $1.3 \times 10^{-21}$ |
| Downstream signaling of activated FGFR4          |         | 134  | 22               | $1.3 \times 10^{-21}$ |
| Signaling by FGFR                                |         | 146  | 23               | $2.0 \times 10^{-21}$ |
| Signaling by FGFR1                               |         | 146  | 23               | $2.0 \times 10^{-21}$ |

| Intersection of SLIPT and siRNA screen (547 genes)       | Pathway | Size | Genes Identified | p-value (FDR)        |
|--|---------|------|------------------|----------------------|
| Class A/1 (Rhodopsin-like receptors)                     |         | 282  | 25               | $3.9 \times 10^{-9}$ |
| Platelet activation, signaling and aggregation           |         | 182  | 17               | $3.9 \times 10^{-9}$ |
| Response to elevated platelet cytosolic Ca <sup>2+</sup> |         | 82   | 9                | $5.5 \times 10^{-8}$ |
| Platelet homeostasis                                     |         | 53   | 7                | $5.7 \times 10^{-8}$ |
| Nucleotide-like (purinergic) receptors                   |         | 16   | 4                | $1.8 \times 10^{-7}$ |
| Platelet degranulation                                   |         | 77   | 8                | $2.8 \times 10^{-7}$ |
| Peptide ligand-binding receptors                         |         | 175  | 14               | $3.8 \times 10^{-7}$ |
| Molecules associated with elastic fibres                 |         | 34   | 5                | $7.1 \times 10^{-7}$ |
| Amine ligand-binding receptors                           |         | 35   | 5                | $8.6 \times 10^{-7}$ |
| <i>Gαi</i> signalling events                             |         | 184  | 14               | $9.8 \times 10^{-7}$ |
| GPCR ligand binding                                      |         | 363  | 27               | $1.1 \times 10^{-6}$ |
| Elastic fibre formation                                  |         | 38   | 5                | $1.5 \times 10^{-6}$ |
| <i>Gαq</i> signalling events                             |         | 159  | 12               | $1.9 \times 10^{-6}$ |
| Serotonin receptors                                      |         | 12   | 3                | $3.8 \times 10^{-6}$ |
| P2Y receptors  |         | 12   | 3                | $3.8 \times 10^{-6}$ |
| Signal amplification                                     |         | 16   | 3                | $2.3 \times 10^{-5}$ |
| Gastrin-CREB signalling pathway via PKC and MAPK         |         | 180  | 12               | $2.3 \times 10^{-5}$ |
| Complement cascade                                       |         | 33   | 4                | $2.4 \times 10^{-5}$ |
| Glycosaminoglycan metabolism                             |         | 110  | 8                | $2.5 \times 10^{-5}$ |
| Glycogen breakdown (glycogenolysis)                      |         | 17   | 3                | $2.7 \times 10^{-5}$ |

## F.2.1 Resampling Analysis

Table F.4: Pathways for *CDH1* partners from SLIPT in stomach cancer

| Reactome Pathway  | Over-representation    | Permutation              |
|---|------------------------|--------------------------|
| <i>Extracellular matrix organization</i>                                | $7.5 \times 10^{-140}$ | 0.070215                 |
| Hemostasis  | $1.8 \times 10^{-121}$ | 0.25804                  |
| Developmental Biology   | $9.2 \times 10^{-107}$ | 0.53032                  |
| Axon guidance   | $1.5 \times 10^{-102}$ | 0.6704                   |
| <b>Eukaryotic Translation Termination</b>                               | $1.9 \times 10^{-99}$  | $> 1.031 \times 10^{-5}$ |
| GPCR ligand binding   | $3.8 \times 10^{-99}$  | 0.54914                  |
| <b>Viral mRNA Translation</b>   | $3.3 \times 10^{-98}$  | $> 1.031 \times 10^{-5}$ |
| Formation of a pool of free 40S subunits                                | $3.3 \times 10^{-98}$  | $> 1.031 \times 10^{-5}$ |
| <b>Eukaryotic Translation Elongation</b>                                | $1.6 \times 10^{-97}$  | $> 1.031 \times 10^{-5}$ |
| Peptide chain elongation  | $7.2 \times 10^{-97}$  | $> 1.031 \times 10^{-5}$ |
| Class A/1 (Rhodopsin-like receptors)                                    | $2.7 \times 10^{-96}$  | 0.58174                  |
| <b>Nonsense Mediated Decay independent of the Exon Junction Complex</b> | $3 \times 10^{-96}$    | $> 1.031 \times 10^{-5}$ |
| Infectious disease  | $2.6 \times 10^{-94}$  | 0.25484                  |
| GTP hydrolysis and joining of the 60S ribosomal subunit                 | $3.4 \times 10^{-94}$  | $> 1.031 \times 10^{-5}$ |
| L13a-mediated translational silencing of Ceruloplasmin expression       | $2.8 \times 10^{-92}$  | $> 1.031 \times 10^{-5}$ |
| <b>3' -UTR-mediated translational regulation</b>                        | $2.8 \times 10^{-92}$  | $> 1.031 \times 10^{-5}$ |
| Neuronal System   | $8.4 \times 10^{-92}$  | 0.53433                  |
| SRP-dependent cotranslational protein targeting to membrane             | $9.5 \times 10^{-92}$  | $> 1.031 \times 10^{-5}$ |
| <b>Eukaryotic Translation Initiation</b>                                | $2.0 \times 10^{-90}$  | $> 1.031 \times 10^{-5}$ |
| Cap-dependent Translation Initiation                                    | $2.0 \times 10^{-90}$  | $> 1.031 \times 10^{-5}$ |
| <b>Nonsense-Mediated Decay</b>  | $7.4 \times 10^{-90}$  | $> 1.031 \times 10^{-5}$ |
| Nonsense Mediated Decay enhanced by the Exon Junction Complex           | $7.4 \times 10^{-90}$  | $> 1.031 \times 10^{-5}$ |
| Adaptive Immune System  | $8.1 \times 10^{-88}$  | 0.14116                  |
| <b>Translation</b>  | $1.3 \times 10^{-87}$  | $> 1.031 \times 10^{-5}$ |
| Platelet activation, signaling and aggregation                          | $1.3 \times 10^{-86}$  | 0.28959                  |
| <b>Influenza Infection</b>  | $1 \times 10^{-82}$    | $> 1.031 \times 10^{-5}$ |
| <b>Influenza Viral RNA Transcription and Replication</b>                | $2.4 \times 10^{-82}$  | $> 1.031 \times 10^{-5}$ |
| <b>Influenza Life Cycle</b>   | $2 \times 10^{-80}$    | $> 1.031 \times 10^{-5}$ |
| Response to elevated platelet cytosolic Ca <sup>2+</sup>                | $4.9 \times 10^{-78}$  | 0.50817                  |
| Signalling by NGF   | $1.6 \times 10^{-75}$  | 0.38518                  |
| Rho GTPase cycle  | $5.1 \times 10^{-75}$  | 0.14864                  |
| Signaling by PDGF   | $7.4 \times 10^{-74}$  | 0.40493                  |
| <i>Signaling by Rho GTPases</i>   | $5.1 \times 10^{-73}$  | 0.077217                 |
| Glycosaminoglycan metabolism  | $1.4 \times 10^{-68}$  | 0.52984                  |
| Gαi signalling events   | $1.8 \times 10^{-66}$  | 0.9254                   |
| Metabolism of carbohydrates   | $1.1 \times 10^{-65}$  | 0.39501                  |
| <b>Gαs signalling events</b>  | $2.7 \times 10^{-65}$  | 0.0050293                |
| Potassium Channels  | $2.7 \times 10^{-65}$  | 0.53359                  |
| Transmission across Chemical Synapses                                   | $1.8 \times 10^{-64}$  | 0.81833                  |
| ECM proteoglycans   | $3.4 \times 10^{-64}$  | 0.083482                 |
| Peptide ligand-binding receptors  | $4.8 \times 10^{-64}$  | 0.62817                  |
| Degradation of the extracellular matrix                                 | $1.1 \times 10^{-63}$  | 0.80879                  |
| Platelet homeostasis  | $5.3 \times 10^{-63}$  | 0.53134                  |
| NGF signalling via TRKA from the plasma membrane                        | $6.1 \times 10^{-63}$  | 0.57117                  |
| Integration of energy metabolism  | $4.5 \times 10^{-61}$  | 0.10889                  |
| Collagen formation  | $5.4 \times 10^{-61}$  | 0.29896                  |
| Integrin cell surface interactions                                      | $7 \times 10^{-59}$    | 0.18167                  |
| Collagen biosynthesis and modifying enzymes                             | $7 \times 10^{-59}$    | 0.30208                  |
| Neurotransmitter Receptor Binding And Downstream Transmission           | $8.7 \times 10^{-57}$  | 0.82522                  |
| In The Postsynaptic Cell  |                        |                          |
| Signaling by Wnt  | $8.7 \times 10^{-57}$  | 0.25468                  |

Over-representation (hypergeometric test) and Permutation p-values adjusted for multiple tests across pathways (FDR). Significant pathways are marked in bold (FDR < 0.05) and italics (FDR < 0.1).

Table F.5: Pathways for *CDH1* partners from SLIPT in stomach and siRNA screen

| Reactome Pathway   | Over-representation  | Permutation               |
|--|----------------------|---------------------------|
| Platelet activation, signaling and aggregation                           | $3.9 \times 10^{-9}$ | 0.49557                   |
| Class A/1 (Rhodopsin-like receptors)                                     | $3.9 \times 10^{-9}$ | 0.98432                   |
| Response to elevated platelet cytosolic Ca <sup>2+</sup>                 | $5.5 \times 10^{-8}$ | 0.54349                   |
| Platelet homeostasis   | $5.7 \times 10^{-8}$ | 0.45017                   |
| Nucleotide-like (purinergic) receptors                                   | $1.8 \times 10^{-7}$ | 0.36966                   |
| Peptide ligand-binding receptors   | $3.8 \times 10^{-7}$ | 0.91294                   |
| <b>Molecules associated with elastic fibres</b>                          | $7.1 \times 10^{-7}$ | 0.0025868                 |
| Amine ligand-binding receptors   | $8.6 \times 10^{-7}$ | 0.43303                   |
| <i>Gαi</i> signalling events   | $9.8 \times 10^{-7}$ | 0.99626                   |
| GPCR ligand binding  | $1.1 \times 10^{-6}$ | 0.97733                   |
| <b>Elastic fibre formation</b>   | $1.5 \times 10^{-6}$ | 0.0025868                 |
| <i>Gαq</i> signalling events   | $1.9 \times 10^{-6}$ | 0.86089                   |
| P2Y receptors  | $3.8 \times 10^{-6}$ | 0.18795                   |
| Serotonin receptors  | $3.8 \times 10^{-6}$ | 0.37853                   |
| Signal amplification   | $2.3 \times 10^{-5}$ | 0.47856                   |
| Gastrin-CREB signalling pathway via PKC and MAPK                         | $2.3 \times 10^{-5}$ | 0.98567                   |
| <b>Complement cascade</b>  | $2.4 \times 10^{-5}$ | $> 3.4628 \times 10^{-6}$ |
| Glycosaminoglycan metabolism   | $2.5 \times 10^{-5}$ | 0.38953                   |
| Glycogen breakdown (glycogenolysis)                                      | $2.7 \times 10^{-5}$ | 0.83772                   |
| Defective B4GALT7 causes EDS, progeroid type                             | $4.9 \times 10^{-5}$ | 0.10792                   |
| Defective B3GAT3 causes JDSSDHD  | $4.9 \times 10^{-5}$ | 0.10792                   |
| Role of LAT2/NTAL/LAB on calcium mobilization                            | $5.6 \times 10^{-5}$ | 0.35373                   |
| Cell surface interactions at the vascular wall                           | $5.6 \times 10^{-5}$ | 0.47642                   |
| <i>Gαs</i> signalling events   | $6 \times 10^{-5}$   | 0.019858                  |
| Signaling by NOTCH   | $6 \times 10^{-5}$   | 0.19008                   |
| A tetrasaccharide linker sequence is required for GAG synthesis          | 0.00017              | 0.47642                   |
| <b>Extracellular matrix organization</b>                                 | 0.00018              | 0.0047308                 |
| Collagen formation   | 0.00018              | 0.19245                   |
| Effects of PIP2 hydrolysis   | 0.0002               | 0.37779                   |
| Syndecan interactions  | 0.0002               | 0.37779                   |
| <b>Diseases associated with glycosaminoglycan metabolism</b>             | 0.00023              | 0.01028                   |
| <b>Diseases of glycosylation</b>   | 0.00023              | 0.01028                   |
| <i>Chondroitin sulfate/dermatan sulfate metabolism</i>                   | 0.00023              | 0.085541                  |
| Integrin alphaIIb beta3 signaling  | 0.00028              | 0.76936                   |
| Keratan sulfate biosynthesis   | 0.00034              | 0.68744                   |
| Rho GTPase cycle   | 0.00034              | 0.15675                   |
| Creation of C4 and C2 activators   | 0.00035              | 0.12275                   |
| Abacavir transport and metabolism  | 0.00035              | 0.12443                   |
| Amine compound SLC transporters  | 0.00037              | 0.69773                   |
| FCER1 mediated NF-κB activation  | 0.00037              | 0.69846                   |
| Fc epsilon receptor (FCER1) signaling                                    | 0.00056              | 0.43303                   |
| Defective EXT2 causes exostoses 2  | 0.00067              | 0.16053                   |
| Defective EXT1 causes exostoses 1, TRPS2 and CHDS                        | 0.00067              | 0.16053                   |
| <i>Collagen biosynthesis and modifying enzymes</i>                       | 0.00071              | 0.052911                  |
| Keratan sulfate/keratin metabolism                                       | 0.00073              | 0.46533                   |
| G alpha (12/13) signalling events  | 0.00078              | 0.59164                   |
| <b>SEMA3A-Plexin repulsion signaling by inhibiting Integrin adhesion</b> | 0.00084              | 0.038504                  |
| Signal attenuation   | 0.00084              | 0.37779                   |
| Eicosanoid ligand-binding receptors                                      | 0.0011               | 0.11117                   |
| SOS-mediated signalling  | 0.0011               | 0.25387                   |

Over-representation (hypergeometric test) and Permutation p-values adjusted for multiple tests across pathways (FDR). Significant pathways are marked in bold (FDR < 0.05) and italicics (FDR < 0.1).

### F.3 Metagene Analysis

Metagene analysis was also performed for synthetic lethal candidates for *CDH1* expression in stomach cancer. These are described and compared to mutation analysis in Section G.4.

Table F.6: Candidate synthetic lethal metagenes against *CDH1* from SLIPT in stomach cancer

| Pathway   | ID      | Observed | Expected | $\chi^2$ value | p-value                | p-value (FDR)          |
|---|---------|----------|----------|----------------|------------------------|------------------------|
| Cell-Cell communication                               | 1500931 | 18       | 50.4     | 110            | $7.43 \times 10^{-23}$ | $1.53 \times 10^{-20}$ |
| VEGFR2 mediated vascular permeability                 | 5218920 | 19       | 50.4     | 109            | $1.36 \times 10^{-22}$ | $2.49 \times 10^{-20}$ |
| Sema4D in semaphorin signaling                        | 400685  | 20       | 50.4     | 104            | $1.62 \times 10^{-21}$ | $2.12 \times 10^{-19}$ |
| Ion transport by P-type ATPases                       | 936837  | 17       | 50.4     | 100            | $8.29 \times 10^{-21}$ | $8.06 \times 10^{-19}$ |
| Sialic acid metabolism                                | 4085001 | 19       | 50.4     | 95.3           | $9.95 \times 10^{-20}$ | $7.82 \times 10^{-18}$ |
| Synthesis of pyrophosphates in the cytosol            | 1855167 | 26       | 50.4     | 94             | $1.86 \times 10^{-19}$ | $1.23 \times 10^{-17}$ |
| Keratan sulfate/keratin metabolism                    | 1638074 | 25       | 50.4     | 93.5           | $2.36 \times 10^{-19}$ | $1.44 \times 10^{-17}$ |
| Ion channel transport                                 | 983712  | 19       | 50.4     | 92.8           | $3.37 \times 10^{-19}$ | $1.99 \times 10^{-17}$ |
| Keratan sulfate biosynthesis                          | 2022854 | 26       | 50.4     | 91.4           | $6.79 \times 10^{-19}$ | $3.62 \times 10^{-17}$ |
| Arachidonic acid metabolism                           | 2142753 | 22       | 50.4     | 90.6           | $9.81 \times 10^{-19}$ | $5.07 \times 10^{-17}$ |
| RHO GTPases activate CIT                              | 5625900 | 22       | 50.4     | 87             | $5.80 \times 10^{-18}$ | $2.66 \times 10^{-16}$ |
| Stimuli-sensing channels                              | 2672351 | 25       | 50.4     | 85.8           | $1.03 \times 10^{-17}$ | $4.58 \times 10^{-16}$ |
| Synthesis of PI                                       | 1483226 | 19       | 50.4     | 85.6           | $1.15 \times 10^{-17}$ | $4.89 \times 10^{-16}$ |
| G-protein activation                                  | 202040  | 19       | 50.4     | 85.3           | $1.34 \times 10^{-17}$ | $5.53 \times 10^{-16}$ |
| NrCAM interactions                                    | 447038  | 22       | 50.4     | 84.3           | $2.1 \times 10^{-17}$  | $8.27 \times 10^{-16}$ |
| Inwardly rectifying $K^+$ channels                    | 1296065 | 24       | 50.4     | 83.5           | $3.19 \times 10^{-17}$ | $1.22 \times 10^{-15}$ |
| Calcitonin-like ligand receptors                      | 419812  | 20       | 50.4     | 82.2           | $6.07 \times 10^{-17}$ | $2.13 \times 10^{-15}$ |
| Prostacyclin signalling through prostacyclin receptor | 392851  | 24       | 50.4     | 81.8           | $7.27 \times 10^{-17}$ | $2.5 \times 10^{-15}$  |
| Presynaptic function of Kainate receptors             | 500657  | 26       | 50.4     | 79.7           | $2.00 \times 10^{-16}$ | $6.34 \times 10^{-15}$ |
| ADP signalling through P2Y purinoceptor 12            | 392170  | 23       | 50.4     | 79.2           | $2.57 \times 10^{-16}$ | $7.71 \times 10^{-15}$ |
| regulation of FZD by ubiquitination                   | 4641263 | 22       | 50.4     | 78.8           | $3.15 \times 10^{-16}$ | $9.3 \times 10^{-15}$  |
| Toxicity of tetanus toxin (TeNT)                      | 5250982 | 27       | 50.4     | 78.7           | $3.36 \times 10^{-16}$ | $9.75 \times 10^{-15}$ |
| Gap junction degradation                              | 190873  | 21       | 50.4     | 78.5           | $3.66 \times 10^{-16}$ | $1.04 \times 10^{-14}$ |
| Nephrin interactions                                  | 373753  | 25       | 50.4     | 78.2           | $4.21 \times 10^{-16}$ | $1.14 \times 10^{-14}$ |
| GABA synthesis, release, reuptake and degradation     | 888590  | 26       | 50.4     | 77             | $7.69 \times 10^{-16}$ | $1.95 \times 10^{-14}$ |

Strongest candidate SL partners for *CDH1* by SLIPT with observed and expected samples with low expression of both genes

# **Appendix G**

## **Stomach Mutation Analysis**

The following results are a replication of the TCGA results (in Appendix D) with stomach cancer data, using synthetic lethality (mtSLIPT) against *CDH1* mutation.

### **G.1 Synthetic Lethal Genes and Pathways**

Table G.1: Synthetic lethal gene partners of *CDH1* from mtSLIPT in stomach cancer

| Gene             | Observed | Expected | $\chi^2$ value | p-value               | p-value (FDR) |
|------------------|----------|----------|----------------|-----------------------|---------------|
| <i>OLFML1</i>    | 5        | 10.1     | 29.2           | $4.53 \times 10^{-7}$ | 0.0031        |
| <i>NRIP2</i>     | 6        | 10.1     | 25.4           | $3.11 \times 10^{-6}$ | 0.00706       |
| <i>VIM</i>       | 3        | 10.1     | 24.7           | $4.29 \times 10^{-6}$ | 0.00706       |
| <i>TCF4</i>      | 5        | 10.1     | 24.7           | $4.33 \times 10^{-6}$ | 0.00706       |
| <i>ZEB2</i>      | 5        | 10.1     | 24.7           | $4.33 \times 10^{-6}$ | 0.00706       |
| <i>BCL2</i>      | 2        | 10.1     | 22             | $1.66 \times 10^{-5}$ | 0.0155        |
| <i>SMARCA2</i>   | 2        | 10.1     | 22             | $1.66 \times 10^{-5}$ | 0.0155        |
| <i>CCND2</i>     | 3        | 10.1     | 21.1           | $2.61 \times 10^{-5}$ | 0.0155        |
| <i>MMP19</i>     | 3        | 10.1     | 21.1           | $2.61 \times 10^{-5}$ | 0.0155        |
| <i>NEURL1B</i>   | 3        | 10.1     | 21.1           | $2.61 \times 10^{-5}$ | 0.0155        |
| <i>IGFBP6</i>    | 6        | 10.1     | 21.1           | $2.65 \times 10^{-5}$ | 0.0155        |
| <i>OGN</i>       | 6        | 10.1     | 21.1           | $2.65 \times 10^{-5}$ | 0.0155        |
| <i>THY1</i>      | 6        | 10.2     | 21             | $2.7 \times 10^{-5}$  | 0.0155        |
| <i>DZIP1</i>     | 4        | 10.1     | 20.6           | $3.29 \times 10^{-5}$ | 0.0155        |
| <i>LOC650368</i> | 4        | 10.1     | 20.6           | $3.29 \times 10^{-5}$ | 0.0155        |
| <i>PCOLCE</i>    | 4        | 10.1     | 20.6           | $3.29 \times 10^{-5}$ | 0.0155        |
| <i>PTGFR</i>     | 4        | 10.1     | 20.6           | $3.29 \times 10^{-5}$ | 0.0155        |
| <i>RUNX1T1</i>   | 4        | 10.1     | 20.6           | $3.29 \times 10^{-5}$ | 0.0155        |
| <i>CLEC2B</i>    | 5        | 10.1     | 20.6           | $3.3 \times 10^{-5}$  | 0.0155        |
| <i>MSC</i>       | 5        | 10.1     | 20.6           | $3.3 \times 10^{-5}$  | 0.0155        |
| <i>NISCH</i>     | 5        | 10.1     | 20.6           | $3.3 \times 10^{-5}$  | 0.0155        |
| <i>TSPAN11</i>   | 5        | 10.1     | 20.6           | $3.3 \times 10^{-5}$  | 0.0155        |
| <i>KCTD12</i>    | 2        | 10.1     | 19.1           | $7.19 \times 10^{-5}$ | 0.0246        |
| <i>LRRK55</i>    | 2        | 10.1     | 19.1           | $7.19 \times 10^{-5}$ | 0.0246        |
| <i>PCBP3</i>     | 2        | 10.1     | 19.1           | $7.19 \times 10^{-5}$ | 0.0246        |

mtSLIPT partners with observed and expected *CDH1* mutant samples with low expression

Table G.2: Pathways for *CDH1* partners from mtSLIPT in stomach cancer

| Pathways Over-represented                                       | Pathway Size | SL Genes | p-value (FDR)        |
|---|--------------|----------|----------------------|
| Extracellular matrix organization                               | 241          | 20       | $9.6 \times 10^{-9}$ |
| Elastic fibre formation   | 38           | 6        | $3.7 \times 10^{-8}$ |
| Diseases associated with glycosaminoglycan metabolism           | 26           | 5        | $3.7 \times 10^{-8}$ |
| Diseases of glycosylation                                       | 26           | 5        | $3.7 \times 10^{-8}$ |
| Nitric oxide stimulates guanylate cyclase                       | 24           | 4        | $3.1 \times 10^{-6}$ |
| Molecules associated with elastic fibres                        | 34           | 4        | $3.7 \times 10^{-5}$ |
| Platelet homeostasis  | 54           | 5        | $3.7 \times 10^{-5}$ |
| Initial triggering of complement                                | 17           | 3        | $3.7 \times 10^{-5}$ |
| Regulation of IGF transport and uptake by IGFBPs                | 17           | 3        | $3.7 \times 10^{-5}$ |
| Collagen degradation  | 58           | 5        | $5.6 \times 10^{-5}$ |
| Defective B4GALT7 causes EDS, progeroid type                    | 19           | 3        | $5.6 \times 10^{-5}$ |
| Defective B3GAT3 causes JDSSDHD                                 | 19           | 3        | $5.6 \times 10^{-5}$ |
| Degradation of the extracellular matrix                         | 104          | 7        | $8.0 \times 10^{-5}$ |
| ECM proteoglycans   | 66           | 5        | 0.00017              |
| A tetrasaccharide linker sequence is required for GAG synthesis | 25           | 3        | 0.00025              |
| RHO GTPases Activate WASPs and WAVEs                            | 29           | 3        | 0.00059              |
| Non-integrin membrane-ECM interactions                          | 53           | 4        | 0.00065              |
| Creation of C4 and C2 activators                                | 11           | 2        | 0.00079              |
| Dermatan sulfate biosynthesis                                   | 11           | 2        | 0.00079              |
| Integrin cell surface interactions                              | 82           | 5        | 0.00098              |

Gene set over-representation analysis (hypergeometric test) for Reactome pathways in mtSLIPT partners for *CDH1*

## G.2 Synthetic Lethal Expression Profiles

Similar to the analysis of synthetic lethal partners against low *CDH1* expression in F.1, the partners detected from *CDH1* mutation were also examined for their expression profiles and the pathway composition of gene clusters. Hierarchical clustering was performed on mtSLIPT partners for *CDH1* as showing in Figure G.1. Over-representation for Reactome pathways for each of the gene clusters identified is given in Table G.3.

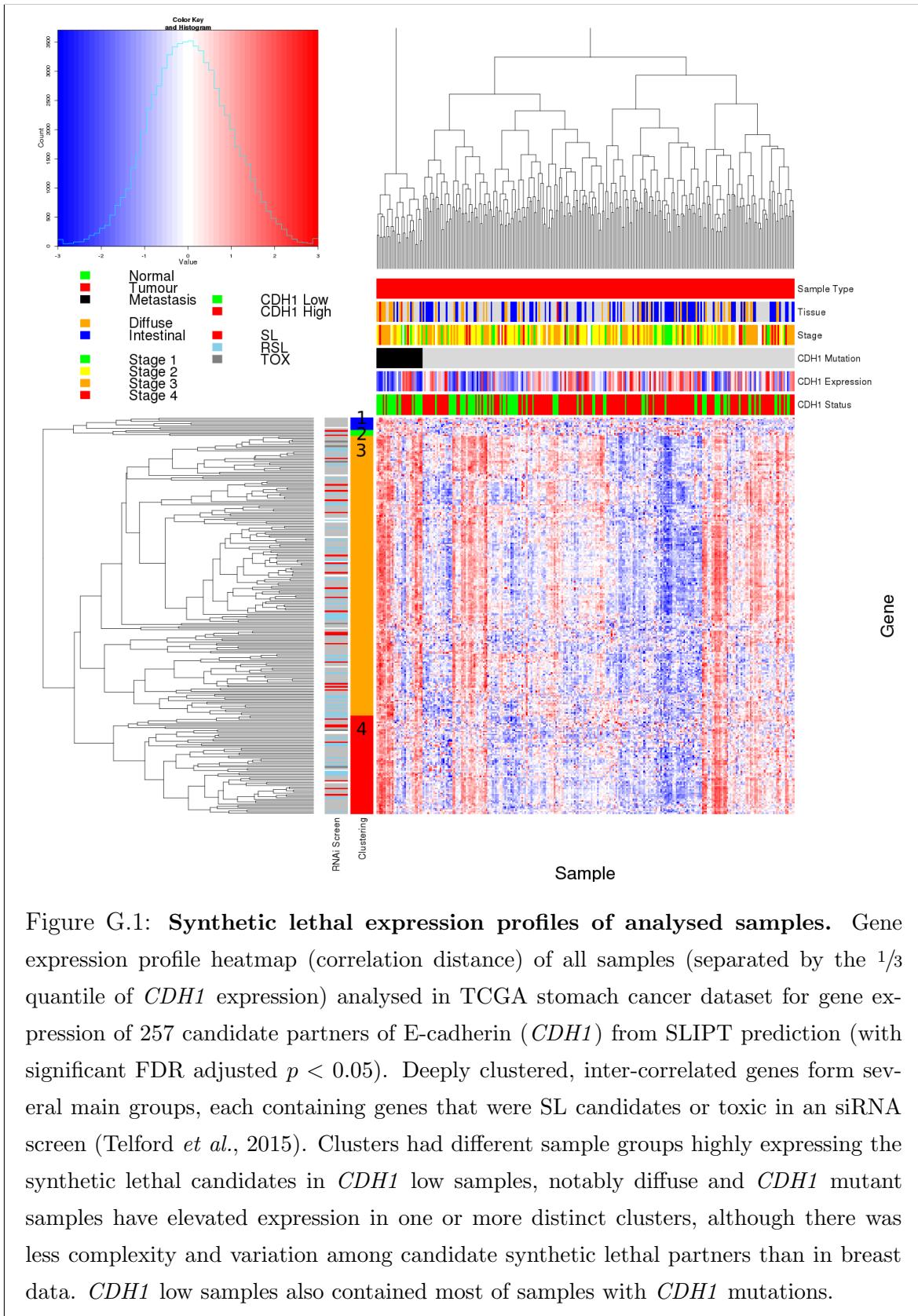


Table G.3: Pathway composition for clusters of *CDH1* partners in stomach mtSLIPT

| Pathways Over-represented in Cluster 1                             | Pathway Size | Cluster Genes | p-value (FDR) |
|--|--------------|---------------|---------------|
| CD28 dependent PI3K/Akt signaling                                  | 15           | 1             | 1             |
| Hormone-sensitive lipase (HSL)-mediated triacylglycerol hydrolysis | 19           | 1             | 1             |
| CD28 co-stimulation  | 26           | 1             | 1             |
| Lipid digestion, mobilization, and transport                       | 48           | 1             | 1             |
| Costimulation by the CD28 family                                   | 51           | 1             | 1             |
| Dectin-1 mediated noncanonical NF- $\kappa$ B signaling            | 58           | 1             | 1             |
| CLEC7A (Dectin-1) signaling  | 99           | 1             | 1             |
| C-type lectin receptors (CLRs)                                     | 123          | 1             | 1             |
| Adaptive Immune System   | 418          | 1             | 1             |
| Metabolism of lipids and lipoproteins                              | 494          | 1             | 1             |
| Interleukin-6 signaling  | 10           | 0             | 1             |
| Apoptosis  | 150          | 0             | 1             |
| Hemostasis   | 445          | 0             | 1             |
| Intrinsic Pathway for Apoptosis                                    | 36           | 0             | 1             |
| Cleavage of Growing Transcript in the Termination Region           | 33           | 0             | 1             |
| PKB-mediated events  | 28           | 0             | 1             |
| PI3K Cascade   | 68           | 0             | 1             |
| RAF/MAP kinase cascade   | 10           | 0             | 1             |
| Global Genomic NER (GG-NER)  | 35           | 0             | 1             |
| Repair synthesis for gap-filling by DNA polymerase in TC-NER       | 15           | 0             | 1             |

| Pathways Over-represented in Cluster 2                                | Pathway Size | Cluster Genes | p-value (FDR) |
|---|--------------|---------------|---------------|
| Kinesins  | 22           | 1             | 1             |
| O-linked glycosylation of mucins                                      | 49           | 1             | 1             |
| O-linked glycosylation  | 59           | 1             | 1             |
| MHC class II antigen presentation                                     | 85           | 1             | 1             |
| Factors involved in megakaryocyte development and platelet production | 120          | 1             | 1             |
| Post-translational protein modification                               | 303          | 1             | 1             |
| Adaptive Immune System  | 418          | 1             | 1             |
| Hemostasis  | 445          | 1             | 1             |
| Interleukin-6 signaling   | 10           | 0             | 1             |
| Apoptosis   | 150          | 0             | 1             |
| Intrinsic Pathway for Apoptosis                                       | 36           | 0             | 1             |
| Cleavage of Growing Transcript in the Termination Region              | 33           | 0             | 1             |
| PKB-mediated events   | 28           | 0             | 1             |
| PI3K Cascade  | 68           | 0             | 1             |
| RAF/MAP kinase cascade  | 10           | 0             | 1             |
| Global Genomic NER (GG-NER)   | 35           | 0             | 1             |
| Repair synthesis for gap-filling by DNA polymerase in TC-NER          | 15           | 0             | 1             |
| Gap-filling DNA repair synthesis and ligation in TC-NER               | 17           | 0             | 1             |
| Formation of transcription-coupled NER (TC-NER) repair complex        | 29           | 0             | 1             |
| Dual incision reaction in TC-NER                                      | 29           | 0             | 1             |

| Pathways Over-represented in Cluster 3                          | Pathway Size | Cluster Genes | p-value (FDR)        |
|---|--------------|---------------|----------------------|
| Extracellular matrix organization                               | 241          | 20            | $9.6 \times 10^{-9}$ |
| Elastic fibre formation   | 38           | 6             | $3.7 \times 10^{-8}$ |
| Diseases associated with glycosaminoglycan metabolism           | 26           | 5             | $3.7 \times 10^{-8}$ |
| Diseases of glycosylation                                       | 26           | 5             | $3.7 \times 10^{-8}$ |
| Molecules associated with elastic fibres                        | 34           | 4             | $4.8 \times 10^{-5}$ |
| Initial triggering of complement                                | 17           | 3             | $4.8 \times 10^{-5}$ |
| Regulation of IGF transport and uptake by IGFBPs                | 17           | 3             | $4.8 \times 10^{-5}$ |
| Collagen degradation  | 58           | 5             | $6.7 \times 10^{-5}$ |
| Defective B4GALT7 causes EDS, progeroid type                    | 19           | 3             | $6.7 \times 10^{-5}$ |
| Defective B3GAT3 causes JDSSDH                                  | 19           | 3             | $6.7 \times 10^{-5}$ |
| Degradation of the extracellular matrix                         | 104          | 7             | $9.5 \times 10^{-5}$ |
| ECM proteoglycans   | 66           | 5             | 0.0002               |
| A tetrasaccharide linker sequence is required for GAG synthesis | 25           | 5             | 0.00029              |
| Non-integrin membrane-ECM interactions                          | 53           | 4             | 0.00079              |
| Creation of C4 and C2 activators                                | 11           | 2             | 0.00093              |
| Dermatan sulfate biosynthesis                                   | 11           | 2             | 0.00093              |
| Integrin cell surface interactions                              | 82           | 5             | 0.0012               |
| Keratan sulfate degradation                                     | 12           | 2             | 0.0012               |
| Complement cascade  | 34           | 3             | 0.0013               |
| CS/DS degradation   | 13           | 2             | 0.0015               |

| Pathways Over-represented in Cluster 4                     | Pathway Size | Cluster Genes | p-value (FDR) |
|--|--------------|---------------|---------------|
| cGMP effects   | 18           | 2             | 0.11          |
| Nitric oxide stimulates guanylate cyclase                  | 24           | 2             | 0.19          |
| Neurotoxicity of clostridium toxins                        | 10           | 1             | 1             |
| Platelet homeostasis                                       | 54           | 2             | 1             |
| Eicosanoid ligand-binding receptors                        | 14           | 1             | 1             |
| Prolactin receptor signaling                               | 15           | 1             | 1             |
| Acyl chain remodelling of PI                               | 15           | 1             | 1             |
| Signaling by FGFR1 fusion mutants                          | 15           | 1             | 1             |
| PKA activation   | 16           | 1             | 1             |
| PKA-mediated phosphorylation of CREB                       | 17           | 1             | 1             |
| Synthesis of glycosylphosphatidylinositol (GPI)            | 17           | 1             | 1             |
| PKA activation in glucagon signalling                      | 17           | 1             | 1             |
| Butyrate Response Factor 1 (BRF1) destabilizes mRNA        | 17           | 1             | 1             |
| Other semaphorin interactions                              | 19           | 1             | 1             |
| Acyl chain remodelling of PE                               | 21           | 1             | 1             |
| Signaling by Leptin  | 21           | 1             | 1             |
| DARPP-32 events  | 22           | 1             | 1             |
| Glucagon-like Peptide-1 (GLP1) regulates insulin secretion | 22           | 1             | 1             |
| Uptake and actions of bacterial toxins                     | 22           | 1             | 1             |
| Acyl chain remodelling of PC                               | 23           | 1             | 1             |

### G.3 Comparison to Primary Screen

The mutation synthetic lethal partners with *CDH1* were also compared to siRNA primary screen data (Telford *et al.*, 2015), as performed in Section 4.2.1. These are expected to be more concordant with the experimental results performed on a null mutant, however this is not the case at the gene level: less genes overlapped with experimental candidates in Figure G.2. This may be affected by lower sample size for mutations in TCGA data or lower frequency (expected value) of *CDH1* mutations compared to low expression.

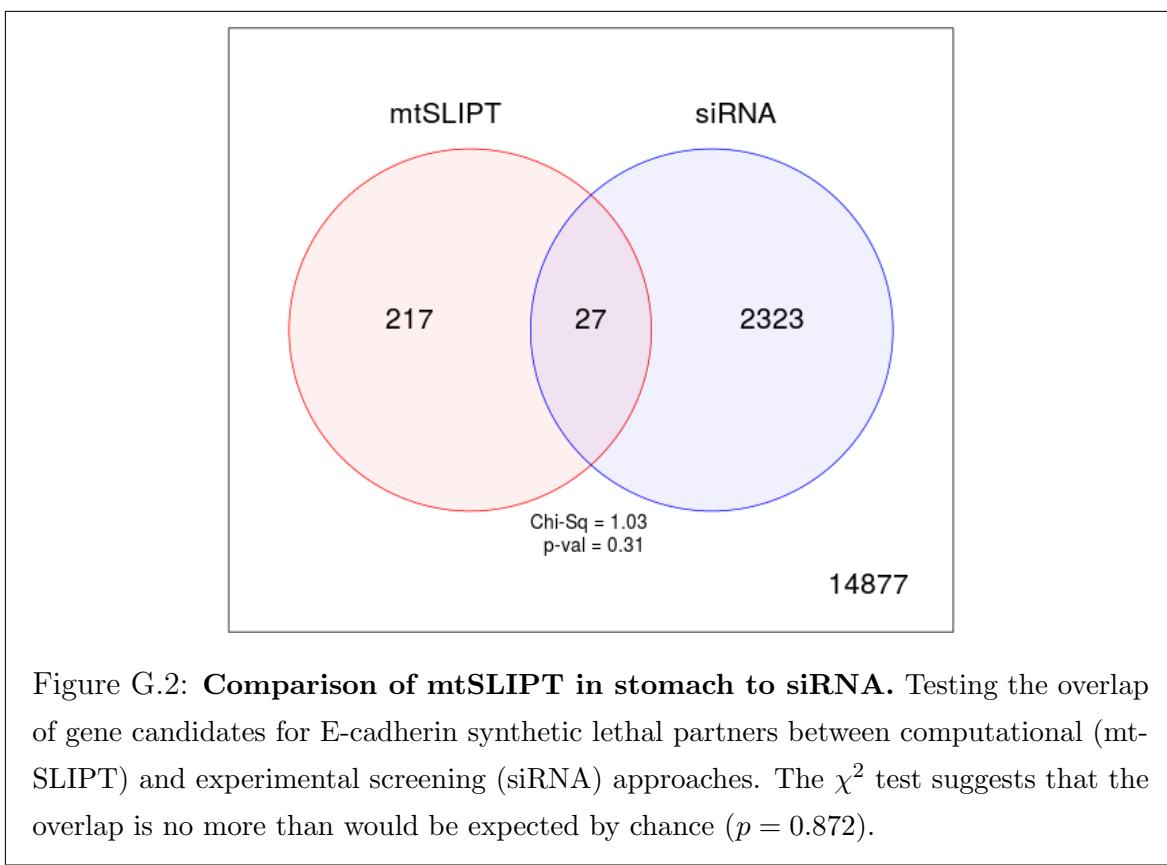


Table G.4: Pathway composition for *CDH1* partners from mtSLIPT and siRNA

| Predicted only by SLIPT (217 genes)                             | Pathway Size | Genes Identified | p-value (FDR)        |
|---|--------------|------------------|----------------------|
| Diseases associated with glycosaminoglycan metabolism           | 26           | 5                | $1.6 \times 10^{-7}$ |
| Diseases of glycosylation                                       | 26           | 5                | $1.6 \times 10^{-7}$ |
| Extracellular matrix organization                               | 238          | 18               | $1.7 \times 10^{-6}$ |
| Elastic fibre formation   | 38           | 5                | $4.6 \times 10^{-6}$ |
| Initial triggering of complement                                | 16           | 3                | $7.3 \times 10^{-5}$ |
| Regulation of IGF transport and uptake by IGFBPs                | 17           | 3                | $8.9 \times 10^{-5}$ |
| Defective B4GALT7 causes EDS, progeroid type                    | 19           | 3                | 0.00013              |
| Defective B3GAT3 causes JDSSDHD                                 | 19           | 3                | 0.00013              |
| Collagen degradation  | 57           | 5                | 0.00013              |
| ECM proteoglycans   | 65           | 5                | 0.00039              |
| A tetrasaccharide linker sequence is required for GAG synthesis | 24           | 3                | 0.00039              |
| Nitric oxide stimulates guanylate cyclase                       | 24           | 3                | 0.00039              |
| RHO GTPases Activate WASPs and WAVEs                            | 28           | 3                | 0.00094              |
| Creation of C4 and C2 activators                                | 10           | 2                | 0.00098              |
| Non-integrin membrane-ECM interactions                          | 52           | 4                | 0.0012               |
| Dermatan sulfate biosynthesis                                   | 11           | 2                | 0.0013               |
| Degradation of the extracellular matrix                         | 101          | 6                | 0.0016               |
| Keratan sulfate degradation                                     | 12           | 2                | 0.0016               |
| Complement cascade  | 33           | 3                | 0.0018               |
| Molecules associated with elastic fibres                        | 34           | 3                | 0.002                |

| Detected only by siRNA screen (2323 genes)       | Pathway Size | Genes Identified | p-value (FDR)         |
|--|--------------|------------------|-----------------------|
| Class A/1 (Rhodopsin-like receptors)             | 282          | 86               | $6.5 \times 10^{-85}$ |
| GPCR ligand binding                              | 363          | 97               | $9.2 \times 10^{-79}$ |
| Peptide ligand-binding receptors                 | 175          | 52               | $4.5 \times 10^{-61}$ |
| G <sub>αi</sub> signalling events                | 184          | 49               | $1.6 \times 10^{-53}$ |
| G <sub>αq</sub> signalling events                | 159          | 43               | $5.2 \times 10^{-50}$ |
| Gastrin-CREB signalling pathway via PKC and MAPK | 180          | 46               | $9.4 \times 10^{-50}$ |
| DAP12 interactions                               | 159          | 35               | $8.3 \times 10^{-37}$ |
| Platelet activation, signaling and aggregation   | 182          | 37               | $2.3 \times 10^{-35}$ |
| Hemostasis                                       | 438          | 71               | $3.3 \times 10^{-35}$ |
| Downstream signal transduction                   | 146          | 32               | $7.7 \times 10^{-35}$ |
| Signaling by PDGF                                | 172          | 35               | $2.1 \times 10^{-34}$ |
| DAP12 signaling                                  | 149          | 32               | $2.7 \times 10^{-34}$ |
| Signaling by ERBB2                               | 146          | 31               | $2.5 \times 10^{-33}$ |
| Signalling by NGF                                | 266          | 44               | $5.3 \times 10^{-31}$ |
| Downstream signaling of activated FGFR1          | 134          | 28               | $5.3 \times 10^{-31}$ |
| Downstream signaling of activated FGFR2          | 134          | 28               | $5.3 \times 10^{-31}$ |
| Downstream signaling of activated FGFR3          | 134          | 28               | $5.3 \times 10^{-31}$ |
| Downstream signaling of activated FGFR4          | 134          | 28               | $5.3 \times 10^{-31}$ |
| Signaling by FGFR                                | 146          | 29               | $2.0 \times 10^{-30}$ |
| Signaling by FGFR1                               | 146          | 29               | $2.0 \times 10^{-30}$ |

| Intersection of SLIPT and siRNA screen (23 genes)                              | Pathway Size | Genes Identified | p-value (FDR) |
|--|--------------|------------------|---------------|
| ADP signalling through P2Y purinoceptor 1                                      | 10           | 1                | 1             |
| G-protein beta:gamma signalling  | 11           | 1                | 1             |
| G-protein activation   | 12           | 1                | 1             |
| Eicosanoid ligand-binding receptors  | 14           | 1                | 1             |
| Platelet homeostasis   | 53           | 2                | 1             |
| G <sub>αz</sub> signalling events  | 15           | 1                | 1             |
| Signal amplification   | 16           | 1                | 1             |
| Activation of Kainate Receptors upon glutamate binding                         | 17           | 1                | 1             |
| Thrombin signalling through protease activated receptors (PARs)                | 17           | 1                | 1             |
| Nitric oxide stimulates guanylate cyclase                                      | 24           | 1                | 1             |
| Activation of G protein gated Potassium channels                               | 25           | 1                | 1             |
| G protein gated Potassium channels   | 25           | 1                | 1             |
| Inhibition of voltage gated Ca <sup>2+</sup> channels via Gbeta/gamma subunits | 25           | 1                | 1             |
| Laminin interactions   | 29           | 1                | 1             |
| Inwardly rectifying K <sup>+</sup> channels                                    | 31           | 1                | 1             |
| Glucagon signaling in metabolic regulation                                     | 33           | 1                | 1             |
| Molecules associated with elastic fibres                                       | 34           | 1                | 1             |
| Ca <sup>2+</sup> pathway   | 36           | 1                | 1             |
| Elastic fibre formation  | 38           | 1                | 1             |
| GABA B receptor activation   | 38           | 1                | 1             |

### G.3.1 Resampling Analysis

Table G.5: Pathways for *CDH1* partners from mtSLIPT in stomach cancer

| Reactome Pathway  | Over-representation  | Permutation               |
|---|----------------------|---------------------------|
| <i>Extracellular matrix organization</i>  | $9.6 \times 10^{-9}$ | 0.057678                  |
| <b>Elastic fibre formation</b>  | $3.7 \times 10^{-8}$ | 0.033817                  |
| <i>Diseases associated with glycosaminoglycan metabolism</i>  | $3.7 \times 10^{-8}$ | 0.049336                  |
| <i>Diseases of glycosylation</i>  | $3.7 \times 10^{-8}$ | 0.049336                  |
| <i>Nitric oxide stimulates guanylate cyclase</i>  | $3.1 \times 10^{-6}$ | 0.037904                  |
| <b>Initial triggering of complement</b>   | $3.7 \times 10^{-5}$ | 0.020828                  |
| <b>Molecules associated with elastic fibres</b>   | $3.7 \times 10^{-5}$ | 0.027865                  |
| <i>Regulation of IGF transport and uptake by IGFBPs</i>   | $3.7 \times 10^{-5}$ | 0.069102                  |
| <i>Platelet homeostasis</i>   | $3.7 \times 10^{-5}$ | 0.097294                  |
| <i>Defective B4GALT7 causes EDS, progeroid type</i>   | $5.6 \times 10^{-5}$ | 0.081505                  |
| <i>Defective B3GAT3 causes JDSSDHD</i>  | $5.6 \times 10^{-5}$ | 0.081505                  |
| <b>Collagen degradation</b>   | $5.6 \times 10^{-5}$ | 0.1104                    |
| <i>Degradation of the extracellular matrix</i>  | $8 \times 10^{-5}$   | 0.43477                   |
| <i>ECM proteoglycans</i>  | 0.00017              | 0.06469                   |
| <i>A tetrasaccharide linker sequence is required for GAG synthesis</i>  | 0.00025              | 0.10536                   |
| <i>RHO GTPases Activate WASPs and WAVES</i>   | 0.00059              | 0.053929                  |
| <i>Non-integrin membrane-ECM interactions</i>   | 0.00065              | 0.10424                   |
| <i>Creation of C4 and C2 activators</i>   | 0.00079              | 0.05461                   |
| <i>Dermatan sulfate biosynthesis</i>  | 0.00079              | 0.21163                   |
| <i>Integrin cell surface interactions</i>   | 0.00098              | 0.092405                  |
| <i>Glucagon signaling in metabolic regulation</i>   | 0.00098              | 0.13425                   |
| <i>Keratan sulfate degradation</i>  | 0.00098              | 0.22137                   |
| <b>Complement cascade</b>   | 0.0011               | 0.01552                   |
| <i>CS/DS degradation</i>  | 0.0012               | 0.065012                  |
| <i>Eicosanoid ligand-binding receptors</i>  | 0.0016               | 0.066128                  |
| <i>Nuclear signaling by ERBB4</i>   | 0.0016               | 0.15511                   |
| <i>Collagen formation</i>   | 0.0026               | 0.13447                   |
| <b>cGMP effects</b>   | 0.0041               | 0.020195                  |
| <i>Voltage gated Potassium channels</i>   | 0.0041               | 0.068923                  |
| <b>Chondroitin sulfate biosynthesis</b>   | 0.0059               | $> 1.5862 \times 10^{-5}$ |
| <i>Chondroitin sulfate/dermatan sulfate metabolism</i>  | 0.0065               | 0.087745                  |
| <i>Heparan sulfate/heparin (HS-GAG) metabolism</i>  | 0.0071               | 0.085622                  |
| <i>Synthesis of substrates in N-glycan biosynthesis</i>   | 0.0085               | 0.09456                   |
| <i>Regulation of actin dynamics for phagocytic cup formation</i>  | 0.0085               | 0.096227                  |
| <i>CDO in myogenesis</i>  | 0.01                 | 0.32599                   |
| <i>Myogenesis</i>   | 0.01                 | 0.32599                   |
| <i>Syndecan interactions</i>  | 0.012                | 0.10975                   |
| <i>Activation of Matrix Metalloproteinases</i>  | 0.012                | 0.33499                   |
| <i>Glycosaminoglycan metabolism</i>   | 0.012                | 0.29716                   |
| <i>Collagen biosynthesis and modifying enzymes</i>  | 0.013                | 0.10774                   |
| <i>Keratan sulfate biosynthesis</i>   | 0.016                | 0.12644                   |
| <i>O-linked glycosylation</i>   | 0.016                | 0.65101                   |
| <i>Laminin interactions</i>   | 0.021                | 0.12766                   |
| <i>Biosynthesis of the N-glycan precursor (dolichol lipid-linked oligosaccharide) and transfer to a nascent protein</i> | 0.027                | 0.065782                  |
| <i>Sialic acid metabolism</i>   | 0.027                | 0.13413                   |
| <i>Keratan sulfate/keratin metabolism</i>   | 0.029                | 0.15708                   |
| <i>Potassium Channels</i>   | 0.032                | 0.43477                   |
| <i>Fcgamma receptor (FCGR) dependent phagocytosis</i>   | 0.042                | 0.15851                   |
| <i>Ion transport by P-type ATPases</i>  | 0.048                | 0.66686                   |
| <i>Retinoid metabolism and transport</i>  | 0.051                | 0.058715                  |

Over-representation (hypergeometric test) and Permutation p-values adjusted for multiple tests across pathways (FDR).

Significant pathways are marked in bold (FDR < 0.05) and italics (FDR < 0.1).

Table G.6: Pathways for *CDH1* partners from mtSLIPT in stomach and siRNA screen

| Reactome Pathway   | Over-representation | Permutation               |
|--|---------------------|---------------------------|
| SLBP independent Processing of Histone Pre-mRNAs                         | 1                   | $> 1.2349 \times 10^{-5}$ |
| Mitochondrial protein import   | 1                   | $> 1.2349 \times 10^{-5}$ |
| Voltage gated Potassium channels   | 1                   | $> 1.2349 \times 10^{-5}$ |
| Tandem pore domain potassium channels                                    | 1                   | $> 1.2349 \times 10^{-5}$ |
| L13a-mediated translational silencing of Ceruloplasmin expression        | 1                   | $> 1.2349 \times 10^{-5}$ |
| Eukaryotic Translation Elongation  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Peptide chain elongation   | 1                   | $> 1.2349 \times 10^{-5}$ |
| 3' -UTR-mediated translational regulation                                | 1                   | $> 1.2349 \times 10^{-5}$ |
| Activation of Matrix Metalloproteinases                                  | 1                   | $> 1.2349 \times 10^{-5}$ |
| HIV Infection  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Cell Cycle   | 1                   | $> 1.2349 \times 10^{-5}$ |
| Influenza Infection  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Influenza Life Cycle   | 1                   | $> 1.2349 \times 10^{-5}$ |
| Influenza Viral RNA Transcription and Replication                        | 1                   | $> 1.2349 \times 10^{-5}$ |
| Neurotoxicity of clostridium toxins                                      | 1                   | $> 1.2349 \times 10^{-5}$ |
| p38MAPK events   | 1                   | $> 1.2349 \times 10^{-5}$ |
| SCF-beta-TrCP mediated degradation of Emi1                               | 1                   | $> 1.2349 \times 10^{-5}$ |
| SRP-dependent cotranslational protein targeting to membrane              | 1                   | $> 1.2349 \times 10^{-5}$ |
| Vpu mediated degradation of CD4  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Serotonin Neurotransmitter Release Cycle                                 | 1                   | $> 1.2349 \times 10^{-5}$ |
| Acetylcholine Binding And Downstream Events                              | 1                   | $> 1.2349 \times 10^{-5}$ |
| Viral mRNA Translation   | 1                   | $> 1.2349 \times 10^{-5}$ |
| Cobalamin (Cbl, vitamin B12) transport and metabolism                    | 1                   | $> 1.2349 \times 10^{-5}$ |
| ERK/MAPK targets   | 1                   | $> 1.2349 \times 10^{-5}$ |
| Vitamin B5 (pantothenate) metabolism                                     | 1                   | $> 1.2349 \times 10^{-5}$ |
| Signaling by BMP   | 1                   | $> 1.2349 \times 10^{-5}$ |
| Synthesis of Leukotrienes (LT) and Eoxins (EX)                           | 1                   | $> 1.2349 \times 10^{-5}$ |
| Separation of Sister Chromatids  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Mitotic Metaphase and Anaphase   | 1                   | $> 1.2349 \times 10^{-5}$ |
| TRP channels   | 1                   | $> 1.2349 \times 10^{-5}$ |
| Defects in cobalamin (B12) metabolism                                    | 1                   | $> 1.2349 \times 10^{-5}$ |
| Regulation by c-FLIP   | 1                   | $> 1.2349 \times 10^{-5}$ |
| Attenuation phase  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Autodegradation of the E3 ubiquitin ligase COP1                          | 1                   | $> 1.2349 \times 10^{-5}$ |
| Apoptotic cleavage of cell adhesion proteins                             | 1                   | $> 1.2349 \times 10^{-5}$ |
| Negative regulation of TCF-dependent signaling by WNT ligand antagonists | 1                   | $> 1.2349 \times 10^{-5}$ |
| PERK regulates gene expression   | 1                   | $> 1.2349 \times 10^{-5}$ |
| Regulation of the Fanconi anemia pathway                                 | 1                   | $> 1.2349 \times 10^{-5}$ |
| Passive transport by Aquaporins  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Lysosome Vesicle Biogenesis  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Zinc transporters  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Zinc influx into cells by the SLC39 gene family                          | 1                   | $> 1.2349 \times 10^{-5}$ |
| Asparagine N-linked glycosylation  | 1                   | $> 1.2349 \times 10^{-5}$ |
| AUF1 (hnRNP D0) destabilizes mRNA  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Asymmetric localization of PCP proteins                                  | 1                   | $> 1.2349 \times 10^{-5}$ |
| degradation of DVL   | 1                   | $> 1.2349 \times 10^{-5}$ |
| CASP8 activity is inhibited  | 1                   | $> 1.2349 \times 10^{-5}$ |
| Degradation of GLI1 by the proteasome                                    | 1                   | $> 1.2349 \times 10^{-5}$ |
| BBSome-mediated cargo-targeting to cilium                                | 1                   | $> 1.2349 \times 10^{-5}$ |
| Regulation of necroptotic cell death                                     | 1                   | $> 1.2349 \times 10^{-5}$ |

## G.4 Metagene Analysis

Metagene analysis was also performed for synthetic lethal candidates for *CDH1* mutation in stomach cancer. These are described and compared to expression analysis in Section F.3.

Table G.7: Candidate synthetic lethal metagenes against *CDH1* from mtSLIPT in stomach cancer

| Pathway  | ID      | Observed | Expected | $\chi^2$ value | p-value               | p-value (FDR) |
|--|---------|----------|----------|----------------|-----------------------|---------------|
| Prostacyclin signalling through prostacyclin receptor                              | 392851  | 1        | 10.1     | 26.5           | $1.73 \times 10^{-6}$ | 0.00286       |
| Cell surface interactions at the vascular wall                                     | 202733  | 3        | 10.1     | 21.1           | $2.61 \times 10^{-5}$ | 0.00642       |
| The NLRP1 inflammasome   | 844455  | 3        | 10.1     | 21.1           | $2.61 \times 10^{-5}$ | 0.00642       |
| Innate Immune System   | 168249  | 6        | 10.1     | 21.1           | $2.65 \times 10^{-5}$ | 0.00642       |
| Keratan sulfate/keratin metabolism   | 1638074 | 4        | 10.1     | 20.6           | $3.29 \times 10^{-5}$ | 0.00642       |
| Keratan sulfate biosynthesis   | 2022854 | 4        | 10.1     | 20.6           | $3.29 \times 10^{-5}$ | 0.00642       |
| Signaling by SCF-KIT   | 1433557 | 5        | 10.1     | 20.6           | $3.30 \times 10^{-5}$ | 0.00642       |
| VEGFA-VEGFR2 Pathway   | 4420097 | 5        | 10.1     | 20.6           | $3.30 \times 10^{-5}$ | 0.00642       |
| p130Cas linkage to MAPK signaling for integrins                                    | 372708  | 2        | 10.1     | 19.1           | $7.19 \times 10^{-5}$ | 0.00651       |
| cGMP effects   | 418457  | 8        | 10.1     | 19             | $7.46 \times 10^{-5}$ | 0.00651       |
| Regulation of cytoskeletal remodeling and cell spreading by IPP complex components | 446388  | 8        | 10.1     | 19             | $7.46 \times 10^{-5}$ | 0.00651       |
| Fcgamma receptor (FCGR) dependent phagocytosis                                     | 2029480 | 3        | 10.1     | 17.9           | 0.000127              | 0.00651       |
| A third proteolytic cleavage releases NICD   | 157212  | 7        | 10.1     | 17.9           | 0.00013               | 0.00651       |
| Signalling by NGF  | 166520  | 7        | 10.1     | 17.9           | 0.00013               | 0.00651       |
| Signaling by VEGF  | 194138  | 7        | 10.1     | 17.9           | 0.00013               | 0.00651       |
| Regulation of thyroid hormone activity   | 350864  | 7        | 10.1     | 17.9           | 0.00013               | 0.00651       |
| Nitric oxide stimulates guanylate cyclase  | 392154  | 7        | 10.1     | 17.9           | 0.00013               | 0.00651       |
| Platelet homeostasis   | 418346  | 7        | 10.1     | 17.9           | 0.00013               | 0.00651       |
| PI3K events in ERBB4 signaling   | 1250342 | 4        | 10.1     | 17.3           | 0.000179              | 0.00651       |
| PIP3 activates AKT signaling   | 1257604 | 4        | 10.1     | 17.3           | 0.000179              | 0.00651       |
| GAB1 signalosome   | 180292  | 4        | 10.1     | 17.3           | 0.000179              | 0.00651       |
| PI3K events in ERBB2 signaling   | 1963642 | 4        | 10.1     | 17.3           | 0.000179              | 0.00651       |
| PI3K/AKT Signaling in Cancer   | 2219528 | 4        | 10.1     | 17.3           | 0.000179              | 0.00651       |
| Rap1 signalling  | 392517  | 4        | 10.1     | 17.3           | 0.000179              | 0.00651       |
| Lysosphingolipid and LPA receptors   | 419408  | 4        | 10.1     | 17.3           | 0.000179              | 0.00651       |

Strongest candidate SL partners for *CDH1* by mtSLIPT with observed and expected mutant samples with low expression of partner metagenes

# **Appendix H**

## **Global Synthetic Lethality in Stomach Cancer**

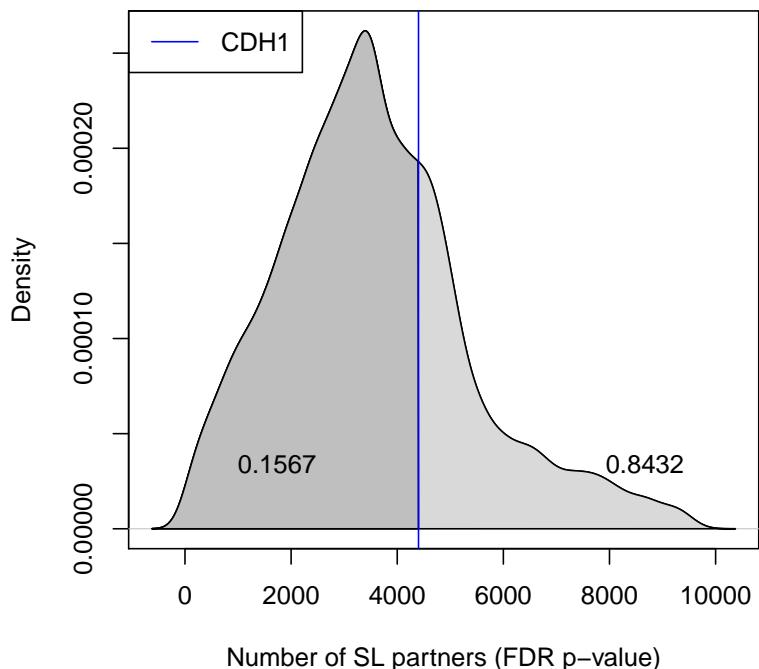


Figure H.1: **Synthetic lethal partners across query genes.** Global synthetic lethal pairs were examined across the genome in TCGA stomach expression data by applying SLIPT across query genes. The high number of predicted partners for *CDH1* was typical for a human gene and lower than many other genes.

## H.1 Hub Genes

Table H.1: Query synthetic lethal genes with the most SLIPT partners

| Gene          | Direction | raw p-value | p-value (FDR) | SLIPT raw p-value | SLIPT (FDR) |
|---------------|-----------|-------------|---------------|-------------------|-------------|
| <i>HEG1</i>   | 10719     | 16956       | 16724         | 9616              | 9532        |
| <i>SYNE1</i>  | 10755     | 17210       | 16984         | 9749              | 9676        |
| <i>A2M</i>    | 10743     | 16650       | 16378         | 9529              | 9433        |
| <i>ANK2</i>   | 11008     | 16616       | 16355         | 9764              | 9653        |
| <i>TTC28</i>  | 10757     | 16523       | 16248         | 9530              | 9429        |
| <i>FAT4</i>   | 10451     | 16286       | 15978         | 9225              | 9115        |
| <i>MRVI1</i>  | 10904     | 16967       | 16718         | 9775              | 9686        |
| <i>PAPLN</i>  | 10483     | 16405       | 16104         | 9305              | 9193        |
| <i>NFASC</i>  | 10773     | 16575       | 16307         | 9578              | 9475        |
| <i>MACF1</i>  | 9697      | 16378       | 16058         | 8620              | 8540        |
| <i>HMCN1</i>  | 10475     | 16101       | 15733         | 9156              | 9008        |
| <i>MPDZ</i>   | 10878     | 16550       | 16299         | 9599              | 9491        |
| <i>FLRT2</i>  | 10776     | 16760       | 16473         | 9590              | 9464        |
| <i>SETBP1</i> | 10869     | 16632       | 16349         | 9615              | 9489        |
| <i>LAMA4</i>  | 10463     | 16447       | 16121         | 9273              | 9151        |
| <i>IL1R1</i>  | 10611     | 16185       | 15803         | 9299              | 9174        |
| <i>ABCA6</i>  | 10499     | 16573       | 16318         | 9260              | 9158        |
| <i>LAMC1</i>  | 10238     | 15777       | 15392         | 8837              | 8691        |
| <i>TNS1</i>   | 10920     | 17038       | 16806         | 9836              | 9751        |
| <i>AMOTL1</i> | 10612     | 16458       | 16178         | 9367              | 9250        |

Genes with the most candidate SL partners SLIPT in TCGA stomach expression data with the number of partner genes predicted by direction criteria and  $\chi^2$  testing separately and combined as a SLIPT analysis. Where specified, the p-values for the  $\chi^2$  test were adjusted for multiple tests (FDR).

## H.2 Hub Pathways

Table H.2: Pathways for genes with the most SLIPT partners

| Pathways Over-represented                 | Pathway Size | SL Genes | p-value               | p-value (FDR)         |
|---|--------------|----------|-----------------------|-----------------------|
| Molecules associated with elastic fibres  | 34           | 10       | $4.6 \times 10^{-21}$ | $2.7 \times 10^{-18}$ |
| Extracellular matrix organization         | 241          | 29       | $5.3 \times 10^{-21}$ | $2.7 \times 10^{-18}$ |
| Smooth Muscle Contraction                 | 29           | 9        | $5.6 \times 10^{-20}$ | $1.6 \times 10^{-17}$ |
| Elastic fibre formation                   | 38           | 10       | $6 \times 10^{-20}$   | $1.6 \times 10^{-17}$ |
| Nitric oxide stimulates guanylate cyclase | 24           | 8        | $6.9 \times 10^{-19}$ | $1.4 \times 10^{-16}$ |
| Muscle contraction                        | 64           | 12       | $8.3 \times 10^{-19}$ | $1.4 \times 10^{-16}$ |
| Platelet homeostasis                      | 54           | 11       | $1.3 \times 10^{-18}$ | $1.9 \times 10^{-16}$ |
| cGMP effects                              | 18           | 6        | $3.3 \times 10^{-15}$ | $4.3 \times 10^{-13}$ |
| Laminin interactions                      | 30           | 7        | $1.3 \times 10^{-14}$ | $1.6 \times 10^{-12}$ |
| Axon guidance                             | 289          | 25       | $5 \times 10^{-13}$   | $5.2 \times 10^{-11}$ |
| Signaling by BMP                          | 23           | 5        | $3.7 \times 10^{-11}$ | $3.2 \times 10^{-9}$  |
| RHO GTPases activate PAKs                 | 23           | 5        | $3.7 \times 10^{-11}$ | $3.2 \times 10^{-9}$  |
| Non-integrin membrane-ECM interactions    | 53           | 7        | $7.2 \times 10^{-11}$ | $5.8 \times 10^{-9}$  |
| Rho GTPase cycle                          | 120          | 11       | $1.2 \times 10^{-10}$ | $8.7 \times 10^{-9}$  |
| Degradation of the extracellular matrix   | 104          | 10       | $1.3 \times 10^{-10}$ | $8.8 \times 10^{-9}$  |
| Netrin-1 signaling                        | 42           | 6        | $2.5 \times 10^{-10}$ | $1.6 \times 10^{-8}$  |
| Developmental Biology                     | 432          | 32       | $8.3 \times 10^{-10}$ | $5 \times 10^{-8}$    |
| L1CAM interactions                        | 80           | 8        | $8.7 \times 10^{-10}$ | $5 \times 10^{-8}$    |
| Semaphorin interactions                   | 64           | 7        | $1.1 \times 10^{-9}$  | $6.1 \times 10^{-8}$  |
| Cell-extracellular matrix interactions    | 18           | 4        | $1.3 \times 10^{-9}$  | $6.6 \times 10^{-8}$  |

Gene set over-representation analysis (hypergeometric test) for Reactome pathways in the top 500 “hub” genes with the most candidate synthetic lethal partners by SLIPT analysis of TCGA stomach expression data

## **Appendix I**

### **Replication in cell line encyclopaedia**

Table I.1: Candidate synthetic lethal gene partners of *CDH1* from SLIPT in CCLE

| Gene                 | Observed | Expected | $\chi^2$ value | p-value                 | p-value (FDR)           |
|----------------------|----------|----------|----------------|-------------------------|-------------------------|
| <i>ZEB1</i>          | 24       | 115      | 555            | $7.84 \times 10^{-119}$ | $3.62 \times 10^{-116}$ |
| <i>RP11-620J15.3</i> | 17       | 115      | 471            | $1.54 \times 10^{-100}$ | $3.68 \times 10^{-98}$  |
| <i>AP1S2</i>         | 20       | 115      | 462            | $1.38 \times 10^{-98}$  | $3.07 \times 10^{-96}$  |
| <i>VIM</i>           | 24       | 115      | 424            | $1.73 \times 10^{-90}$  | $3.06 \times 10^{-88}$  |
| <i>CCDC88A</i>       | 24       | 115      | 418            | $3.94 \times 10^{-89}$  | $6.86 \times 10^{-87}$  |
| <i>RECK</i>          | 28       | 115      | 416            | $8.23 \times 10^{-89}$  | $1.42 \times 10^{-86}$  |
| <i>AP1M1</i>         | 16       | 115      | 414            | $2.42 \times 10^{-88}$  | $4.06 \times 10^{-86}$  |
| <i>ZEB2</i>          | 23       | 115      | 396            | $2.32 \times 10^{-84}$  | $3.4 \times 10^{-82}$   |
| <i>WIPF1</i>         | 25       | 115      | 390            | $4.9 \times 10^{-83}$   | $6.74 \times 10^{-81}$  |
| <i>SLC35B4</i>       | 29       | 115      | 386            | $3.2 \times 10^{-82}$   | $4.38 \times 10^{-80}$  |
| <i>SACS</i>          | 28       | 115      | 373            | $2.13 \times 10^{-79}$  | $2.7 \times 10^{-77}$   |
| <i>ST3GAL2</i>       | 25       | 115      | 351            | $9.7 \times 10^{-75}$   | $1.08 \times 10^{-72}$  |
| <i>ATP8B2</i>        | 38       | 115      | 341            | $1.53 \times 10^{-72}$  | $1.61 \times 10^{-70}$  |
| <i>IFFO1</i>         | 39       | 115      | 332            | $1.66 \times 10^{-70}$  | $1.65 \times 10^{-68}$  |
| <i>EMP3</i>          | 38       | 115      | 329            | $5.04 \times 10^{-70}$  | $4.95 \times 10^{-68}$  |
| <i>LEPRE1</i>        | 40       | 115      | 325            | $5.4 \times 10^{-69}$   | $5.22 \times 10^{-67}$  |
| <i>STARD9</i>        | 39       | 115      | 311            | $4.52 \times 10^{-66}$  | $3.96 \times 10^{-64}$  |
| <i>DENND5A</i>       | 48       | 115      | 304            | $1.89 \times 10^{-64}$  | $1.59 \times 10^{-62}$  |
| <i>SYT11</i>         | 38       | 115      | 300            | $1.21 \times 10^{-63}$  | $9.89 \times 10^{-62}$  |
| <i>EID2B</i>         | 38       | 115      | 299            | $1.99 \times 10^{-63}$  | $1.61 \times 10^{-61}$  |
| <i>NXPE3</i>         | 35       | 115      | 294            | $1.71 \times 10^{-62}$  | $1.35 \times 10^{-60}$  |
| <i>STX2</i>          | 49       | 115      | 293            | $3.83 \times 10^{-62}$  | $3 \times 10^{-60}$     |
| <i>ARHGEF6</i>       | 43       | 115      | 289            | $2.2 \times 10^{-61}$   | $1.71 \times 10^{-59}$  |
| <i>KATNAL1</i>       | 50       | 115      | 283            | $4.45 \times 10^{-60}$  | $3.38 \times 10^{-58}$  |
| <i>ANXA6</i>         | 37       | 115      | 282            | $8.92 \times 10^{-60}$  | $6.67 \times 10^{-58}$  |

Strongest candidate SL partners for *CDH1* by SLIPT with observed and expected samples with low expression of both genes

Table I.2: Candidate synthetic lethal gene partners of *CDH1* from SLIPT in breast CCLE

| Gene             | Observed | Expected | $\chi^2$ value | p-value               | p-value (FDR) |
|------------------|----------|----------|----------------|-----------------------|---------------|
| <i>MIR155HG</i>  | 1        | 6.78     | 31.5           | $2.41 \times 10^{-6}$ | 0.00371       |
| <i>ENPP2</i>     | 1        | 6.78     | 30.7           | $3.47 \times 10^{-6}$ | 0.00383       |
| <i>DCLK2</i>     | 3        | 6.78     | 28.3           | $1.08 \times 10^{-5}$ | 0.0071        |
| <i>PID1</i>      | 1        | 6.78     | 27.8           | $1.34 \times 10^{-5}$ | 0.00791       |
| <i>SCFD2</i>     | 5        | 6.78     | 27.7           | $1.42 \times 10^{-5}$ | 0.00791       |
| <i>FAT4</i>      | 4        | 6.78     | 27.3           | $1.69 \times 10^{-5}$ | 0.00865       |
| <i>ILK</i>       | 1        | 6.78     | 26.9           | $2.04 \times 10^{-5}$ | 0.00884       |
| <i>RWDD1</i>     | 0        | 6.78     | 26.8           | $2.15 \times 10^{-5}$ | 0.00884       |
| <i>RIC8A</i>     | 2        | 6.78     | 26.8           | $2.2 \times 10^{-5}$  | 0.00884       |
| <i>F2RL2</i>     | 1        | 6.78     | 26.6           | $2.34 \times 10^{-5}$ | 0.00901       |
| <i>SDCBP</i>     | 5        | 6.78     | 25.9           | $3.26 \times 10^{-5}$ | 0.0108        |
| <i>PPM1F</i>     | 4        | 6.78     | 25.8           | $3.41 \times 10^{-5}$ | 0.0108        |
| <i>IKBIP</i>     | 5        | 6.78     | 25.8           | $3.49 \times 10^{-5}$ | 0.0108        |
| <i>SPRED1</i>    | 3        | 6.78     | 25.5           | $3.97 \times 10^{-5}$ | 0.0108        |
| <i>RNH1</i>      | 1        | 6.78     | 25.4           | $4.22 \times 10^{-5}$ | 0.0108        |
| <i>SYDE1</i>     | 3        | 6.78     | 25.4           | $4.22 \times 10^{-5}$ | 0.0108        |
| <i>LINC00968</i> | 1        | 6.78     | 25.2           | $4.63 \times 10^{-5}$ | 0.0109        |
| <i>ARHGEF10</i>  | 5        | 6.78     | 24.5           | $6.22 \times 10^{-5}$ | 0.0116        |
| <i>P4HA1</i>     | 0        | 6.78     | 24.5           | $6.34 \times 10^{-5}$ | 0.0116        |
| <i>AZI2</i>      | 2        | 6.78     | 24.5           | $6.34 \times 10^{-5}$ | 0.0116        |
| <i>TNFAIP6</i>   | 2        | 6.78     | 24.5           | $6.34 \times 10^{-5}$ | 0.0116        |
| <i>CD200</i>     | 4        | 6.78     | 24.5           | $6.37 \times 10^{-5}$ | 0.0116        |
| <i>SMPD1</i>     | 1        | 6.78     | 24.4           | $6.67 \times 10^{-5}$ | 0.0116        |
| <i>ATP6V1G2</i>  | 3        | 6.78     | 24.2           | $7.33 \times 10^{-5}$ | 0.0123        |
| <i>FGF2</i>      | 4        | 6.78     | 24.1           | $7.49 \times 10^{-5}$ | 0.0123        |

Strongest candidate SL partners for *CDH1* by SLIPT with observed and expected samples with low expression of both genes

Table I.3: Candidate synthetic lethal gene partners of *CDH1* from SLIPT in stomach CCLE

| Gene             | Observed | Expected | $\chi^2$ value | p-value               | p-value (FDR) |
|------------------|----------|----------|----------------|-----------------------|---------------|
| <i>ZEB1</i>      | 1        | 4.45     | 36             | $2.84 \times 10^{-7}$ | 0.00175       |
| <i>WDR47</i>     | 0        | 4.45     | 26.7           | $2.3 \times 10^{-5}$  | 0.013         |
| <i>KANK2</i>     | 1        | 4.45     | 25.1           | $4.81 \times 10^{-5}$ | 0.0222        |
| <i>LEPRE1</i>    | 0        | 4.45     | 24.5           | $6.26 \times 10^{-5}$ | 0.0228        |
| <i>KATNAL1</i>   | 0        | 4.45     | 24.3           | $6.88 \times 10^{-5}$ | 0.0231        |
| <i>TET1</i>      | 0        | 4.45     | 23.9           | $8.23 \times 10^{-5}$ | 0.0249        |
| <i>AP1S2</i>     | 1        | 4.45     | 23.1           | 0.00012               | 0.0273        |
| <i>CDKN2C</i>    | 1        | 4.45     | 22.8           | 0.000136              | 0.0292        |
| <i>ARMC4</i>     | 1        | 4.45     | 22.4           | 0.000164              | 0.0315        |
| <i>CSTF3</i>     | 1        | 4.45     | 22.4           | 0.000166              | 0.0315        |
| <i>FAM216A</i>   | 1        | 4.45     | 22.4           | 0.000166              | 0.0315        |
| <i>ANKRD32</i>   | 1        | 4.45     | 22.4           | 0.000166              | 0.0315        |
| <i>WDR35</i>     | 1        | 4.45     | 22.4           | 0.000169              | 0.0315        |
| <i>ECI2</i>      | 0        | 4.45     | 21.7           | 0.000232              | 0.0378        |
| <i>SAMD8</i>     | 0        | 4.45     | 21.7           | 0.000232              | 0.0378        |
| <i>CHST12</i>    | 0        | 4.45     | 21.7           | 0.000232              | 0.0378        |
| <i>RPL23AP32</i> | 0        | 4.45     | 21.7           | 0.000232              | 0.0378        |
| <i>STARD9</i>    | 1        | 4.45     | 21.7           | 0.000232              | 0.0378        |
| <i>MCM8</i>      | 0        | 4.45     | 21.5           | 0.000255              | 0.0379        |

Strongest candidate SL partners for *CDH1* by SLIPT with observed and expected samples with low expression of both genes

Table I.4: Pathways for *CDH1* partners from SLIPT in stomach CCLE

| Pathways Over-represented  | Pathway Size | SL Genes | p-value (FDR) |
|--|--------------|----------|---------------|
| Nef mediated downregulation of MHC class I complex cell surface expression                     | 10           | 1        | 1             |
| Unwinding of DNA   | 11           | 1        | 1             |
| Processing of Intronless Pre-mRNAs   | 13           | 1        | 1             |
| E2F mediated regulation of DNA replication   | 20           | 1        | 1             |
| Chondroitin sulfate biosynthesis   | 20           | 1        | 1             |
| Post-Elongation Processing of Intronless pre-mRNA  | 21           | 1        | 1             |
| Nef-mediates down modulation of cell surface receptors by recruiting them to clathrin adapters | 21           | 1        | 1             |
| Processing of Capped Intronless Pre-mRNA   | 21           | 1        | 1             |
| Post-Elongation Processing of Intron-Containing pre-mRNA                                       | 23           | 1        | 1             |
| Activation of the pre-replicative complex  | 23           | 1        | 1             |
| mRNA 3'-end processing   | 23           | 1        | 1             |
| Golgi Associated Vesicle Biogenesis  | 24           | 1        | 1             |
| Lysosome Vesicle Biogenesis  | 25           | 1        | 1             |
| Oncogene Induced Senescence  | 27           | 1        | 1             |
| The role of Nef in HIV-1 replication and disease pathogenesis                                  | 28           | 1        | 1             |
| Cyclin D associated events in G1   | 29           | 1        | 1             |
| G1 Phase   | 29           | 1        | 1             |
| Cleavage of Growing Transcript in the Termination Region                                       | 31           | 1        | 1             |
| Activation of ATR in response to replication stress  | 31           | 1        | 1             |
| DNA strand elongation  | 31           | 1        | 1             |

Gene set over-representation analysis (hypergeometric test) for Reactome pathways in SLIPT partners for *CDH1*

Table I.5: Pathways for *CDH1* partners from SLIPT in breast and stomach CCLE

| Pathways Over-represented   | Pathway Size | SL Genes | p-value (FDR)        |
|---|--------------|----------|----------------------|
| Collagen formation  | 66           | 8        | $1.1 \times 10^{-7}$ |
| Glycosaminoglycan metabolism                                      | 111          | 11       | $1.1 \times 10^{-7}$ |
| Extracellular matrix organization                                 | 236          | 20       | $1.1 \times 10^{-7}$ |
| Collagen biosynthesis and modifying enzymes                       | 55           | 7        | $1.7 \times 10^{-7}$ |
| Keratan sulfate biosynthesis                                      | 28           | 5        | $2.2 \times 10^{-7}$ |
| Keratan sulfate/keratin metabolism                                | 32           | 5        | $7.5 \times 10^{-7}$ |
| ECM proteoglycans   | 65           | 7        | $1.1 \times 10^{-6}$ |
| Non-integrin membrane-ECM interactions                            | 52           | 6        | $2.0 \times 10^{-6}$ |
| Cell junction organization  | 71           | 7        | $3.0 \times 10^{-6}$ |
| Assembly of collagen fibrils and other multimeric structures      | 39           | 5        | $3.6 \times 10^{-6}$ |
| Post-chaperonin tubulin folding pathway                           | 14           | 3        | $1.7 \times 10^{-5}$ |
| Adherens junctions interactions                                   | 29           | 4        | $1.7 \times 10^{-5}$ |
| Cell-Cell communication   | 118          | 9        | $1.7 \times 10^{-5}$ |
| Sialic acid metabolism  | 31           | 4        | $2.5 \times 10^{-5}$ |
| Synthesis and interconversion of nucleotide di- and triphosphates | 16           | 3        | $3.1 \times 10^{-5}$ |
| Transport to the Golgi and subsequent modification                | 34           | 4        | $4.8 \times 10^{-5}$ |
| Asparagine N-linked glycosylation                                 | 113          | 8        | $7.8 \times 10^{-5}$ |
| Elastic fibre formation   | 37           | 4        | $8.5 \times 10^{-5}$ |
| L1CAM interactions  | 77           | 6        | $9.5 \times 10^{-5}$ |
| Signal transduction by L1   | 20           | 3        | $9.5 \times 10^{-5}$ |

Gene set over-representation analysis (hypergeometric test) for Reactome pathways in SLIPT partners for *CDH1*