Contents

\mathbf{G}	Glossary		xiii	
A	crony	yms		$\mathbf{x}\mathbf{v}$
1	Intr	oducti	ion and Literature Review	1
	1.1	Cance	er Research in the Post-Genomic Era	. 1
		1.1.1	Cancer is a Global Health Issue	. 2
			1.1.1.1 The Genetics and Molecular Biology of Cancers	. 3
		1.1.2	The Genomics Revolution in Cancer Research	. 3
			1.1.2.1 High-Throughput Technologies	. 4
			1.1.2.2 Bioinformatics and Genomic Data	. 5
		1.1.3	Genomics Projects	. 5
			1.1.3.1 The Cancer Genome Project	. 6
			1.1.3.2 The Cancer Genome Atlas Project	
		1.1.4	Genomic Cancer Medicine	
			1.1.4.1 Cancer Genes and Driver Mutations	
			1.1.4.2 Precision Cancer Medicine	
			1.1.4.3 Molecular Diagnostics and Pan-Cancer Medicine	
			1.1.4.4 Targeted Therapeutics and Pharmacogenomics	
		1.1.5	Systems and Network Biology	
	1.2		etic Lethal Cancer Medicine	
		1.2.1	Synthetic Lethal Genetic Interactions	
		1.2.2	Synthetic Lethal Concepts in Genetics	
		1.2.3	Synthetic Lethality in Model Systems	
			1.2.3.1 Synthetic Lethal Pathways and Networks	
			1.2.3.2 Evolution of Synthetic Lethality	
		1.2.4	Synthetic Lethality in Cancer	
		1.2.5	Clinical Impact of Synthetic Lethality in Cancer	
		1.2.6	High-throughput Screening for Synthetic Lethality	
			1.2.6.1 Synthetic Lethal Screens	
		1.2.7	Computational Prediction of Synthetic Lethality	
			1.2.7.1 Bioinformatics Approaches to Genetic Interactions .	
			1.2.7.2 Comparative Genomics	
			1.2.7.3 Analysis and Modelling of Protein Data	
			1.2.7.4 Differential Gene Expression	
			1.2.7.5 Data Mining and Machine Learning	. 28

			1.2.7.6 Mutually Exclusive Bimodality	31
			1.2.7.7 Rationale for Further Development	32
	1.3	E-cadl	herin as a Synthetic Lethal Target	32
		1.3.1	The CDH1 gene and its Biological Functions	33
			1.3.1.1 Cytoskeleton	33
			1.3.1.2 Extracellular and Tumour Micro-environment	33
			1.3.1.3 Cell-Cell Adhesion and Signalling	34
		1.3.2	CDH1 as a Tumour (and Invasion) Suppressor	34
			1.3.2.1 Breast Cancers and Invasion	34
		1.3.3	Hereditary Diffuse Gastric (and Lobular Breast) Cancer	35
		1.3.4	Cell Line Models of <i>CDH1</i> Null Mutations	36
	1.4		nary and Research Direction of Thesis	37
		1.4.1	Thesis Aims	38
2	Mot	thode	and Resources	40
4	2.1		formatics Resources for Genomics Research	40
	2.1	2.1.1	Public Data and Software Packages	40
		2.1.1	2.1.1.1 Cancer Genome Atlas Data	41
			2.1.1.2 Reactome and Annotation Data	42
	2.2	Data l	Handling	42
	2.2	2.2.1	Normalisation	42
		2.2.2	Sample Triage	43
		2.2.3	Metagenes and the Singular Value Decomposition	43
		2.2.4	Candidate Triage and Integration with Screen Data	45
	2.3		iques	46
		2.3.1	Statistical Procedures and Tests	46
		2.3.2	Gene Set Over-representation Analysis	47
		2.3.3	Clustering	47
		2.3.4	Heatmap	47
		2.3.5	Modelling and Simulations	48
			2.3.5.1 Receiver Operating Characteristic Curves	49
		2.3.6	Resampling Analysis	49
	2.4	Pathw	vay Structure Methods	50
		2.4.1	Network and Graph Analysis	50
		2.4.2	Sourcing Graph Structure Data	51
		2.4.3	Constructing Pathway Subgraphs	51
		2.4.4	Network Analysis Metrics	52
	2.5	Implei	mentation	53
		2.5.1	Computational Resources and Linux Utilities	53
		2.5.2	R Language and Packages	54
		2.5.3	High Performance and Parallel Computing	57
3	Met	thods 1	Developed During Thesis	59
	3.1	A Syn	thetic Lethal Detection Methodology	59
	3.2	Synthe	etic Lethal Simulation and Modelling	61
			A Model of Synthetic Lethality in Expression Data	62

		3.2.2	Simulation Procedure	6
	3.3	Detect	ing Simulated Synthetic Lethal Partners	6
		3.3.1	Binomial Simulation of Synthetic Lethality	
		3.3.2	Multivariate Normal Simulation of Synthet	
			3.3.2.1 Multivariate Normal Simulation v	
			3.3.2.2 Specificity with Query-Correlated	
	3.4	Graph	Structure Methods	-
		3.4.1	Upstream and Downstream Gene Detection	
			3.4.1.1 Permutation Analysis for Statistic	
			3.4.1.2 Hierarchy Based on Biological Co	
		3.4.2	Simulating Gene Expression from Graph St	
	3.5	Custo	nised Functions and Packages Developed .	
		3.5.1	Synthetic Lethal Interaction Prediction Too	ol
		3.5.2	Data Visualisation	9
		3.5.3	Extensions to the iGraph Package	
			3.5.3.1 Sampling Simulated Data from G	raph Structures 9
			3.5.3.2 Plotting Directed Graph Structur	
			3.5.3.3 Computing Information Centrality	y 9.
			3.5.3.4 Testing Pathway Structure with F	Permutation Testing . 9
			3.5.3.5 Metapackage to Install iGraph Fu	nctions 9
	~			
4	-		Lethal Analysis of Gene Expression Da	
	4.1	•	etic Lethal Genes in Breast Cancer	
		4.1.1	Synthetic Lethal Pathways in Breast Cance	
		4.1.2	Expression Profiles of Synthetic Lethal Par	
	4.0		4.1.2.1 Subgroup Pathway Analysis	
	4.2		aring Synthetic Lethal Gene Candidates	
		4.2.1	Primary siRNA Screen Candidates	
		4.2.2	Comparison with Correlation	
		4.2.3	Comparison with Primary Screen Viability	
		4.2.4	Comparison with Secondary siRNA Screen	
		4.2.5	Comparison to Primary Screen at Pathway	
		4.0.0	4.2.5.1 Resampling Genes for Pathway E	
		4.2.6	Integrating Synthetic Lethal Pathways and	
		4.2.7	Synthetic Lethal Pathway Metagenes	
	4.0	4.2.8	Synthetic Lethality in Breast Cancer	
	4.3	-	ation in Stomach Cancer	
	4.4		sion	
		4.4.1	Strengths of the SLIPT Methodology	
		4.4.2	Synthetic Lethal Pathways for E-cadherin	
		4.4.3	Replication and Validation	
			4.4.3.1 Integration with short interfering F	
			ing	
	4 F	Summ	4.4.3.2 Replication across Tissues	
	/I 5	Summ	arv	1.7

5	\mathbf{Syn}	thetic	Lethal Pathway Structure 128
	5.1	Synthe	etic Lethal Genes in Reactome Pathways
		5.1.1	The PI3K/AKT Pathway
		5.1.2	The Extracellular Matrix
		5.1.3	G Protein Coupled Receptors
		5.1.4	Gene Regulation and Translation
	5.2	Netwo	ork Analysis of Synthetic Lethal Genes
		5.2.1	Gene Connectivity and Vertex Degree
		5.2.2	Gene Importance and Centrality
			5.2.2.1 Information Centrality
			5.2.2.2 PageRank Centrality
	5.3	Relati	onships between Synthetic Lethal Genes
		5.3.1	Hierarchical Pathway Structure
			5.3.1.1 Contextual Hierarchy of PI3K
			5.3.1.2 Testing Contextual Hierarchy of Synthetic Lethal Genes 141
		5.3.2	Upstream or Downstream Synthetic Lethality
			5.3.2.1 Measuring Structure of Candidates within PI3K 145
			5.3.2.2 Resampling for Synthetic Lethal Pathway Structure 147
	5.4	Discus	ssion
	5.5	Summ	ary
6	Sim	ulatio	n and Modelling of Synthetic Lethal Pathways 152
	6.1		etic Lethal Detection Methods
	0.1	6.1.1	
			6.1.1.1 Correlated Query Genes affects Specificity 157
		6.1.2	Alternative Synthetic Lethal Detection Strategies 159
			6.1.2.1 Correlation for Synthetic Lethal Detection 160
			6.1.2.2 Testing for Bimodality with BiSEp 161
	6.2	Simula	ations with Graph Structures
		6.2.1	•
			6.2.1.1 Simple Graph Structures
			6.2.1.2 Constructed Graph Structures
		6.2.2	Performance with Inhibitions
		6.2.3	Synthetic Lethality across Graph Structures
		6.2.4	Performance within a Simulated Human Genome 177
	6.3	Simula	ations in More Complex Graph Structures
		6.3.1	Simulations over Pathway-based Graphs
		6.3.2	Pathway Structures in a Simulated Human Genome 185
	6.4	Discus	ssion
		6.4.1	Simulation Procedure
		6.4.2	Comparing Methods with Simulated Data
		6.4.3	Design and Performance of SLIPT
		6.4.4	Simulations from Graph Structures
	6.5	Summ	•

7.1.1 Established Functions of CDH1 196 7.1.2 The Molecular Role of CDH1 in Cancer 196 7.2 Significance 197 7.2.1 Synthetic Lethality in the Genomic Era 197 7.2.2 Clinical Interventions based on Synthetic Lethality 198 7.3 Future Directions 200 7.4 Conclusions 202 References 204 A Sample Quality 228 A.1 Sample Correlation 228 A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 236 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 245 C.1 Synthetic Lethal Genes and Pathways 244 C.2 Synthetic Lethal Expression Profiles 244 C.3 Comparison to Primary Screen 245 C.4 Compare SLIPT genes 251 D Metagene Analysis 255 D.1 Pathway Expression 256 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 261 D.4 Expression of Somatic Mutations 262 E Intrinsic Subtyping 267 F Stomach Expression Analysis 276 <t< th=""><th>7</th><th>Discussion</th><th>195</th></t<>	7	Discussion	195
7.1.2 The Molecular Role of CDH1 in Cancer 196 7.2 Significance 197 7.2.1 Synthetic Lethality in the Genomic Era 197 7.2.2 Clinical Interventions based on Synthetic Lethality 198 7.3 Future Directions 200 7.4 Conclusions 202 References 204 A Sample Quality 228 A.1 Sample Correlation 228 A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 236 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 24 C.1 Synthetic Lethal Genes and Pathways 24 C.2 Synthetic Lethal Expression Profiles 24 C.3 Comparison to Primary Screen 24 C.3 Compares LIPT genes 25 D.1 Pathway Expression 25 D.2 Somatic Mutation 25 D.3 Synthetic Lethal Reactome Metagenes 26 D.4 Expression of Somatic Mutations 26 D.5 Metagene Expression Analysis 26 F.1 Synthetic Lethal Genes and Pathways 26 F.2 Comparison to Primary Screen 27 F.2 Comparison to Primary Screen 27		7.1 Synthetic Lethality and <i>CDH1</i> Biology	195
7.2 Significance 197 7.2.1 Synthetic Lethality in the Genomic Era 199 7.2.2 Clinical Interventions based on Synthetic Lethality 198 7.3 Future Directions 200 7.4 Conclusions 202 References 204 A Sample Quality 228 A.1 Sample Correlation 228 A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 236 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 24 C.1 Synthetic Lethal Genes and Pathways 24 C.2 Synthetic Lethal Expression Profiles 24 C.3 Comparison to Primary Screen 24 C.3.1 Resampling Analysis 24 C.4 Compare SLIPT genes 25 D Metagene Analysis 25 D.1 Pathway Expression 25 D.2 Somatic Mutation 25 D.3 Synthetic Lethal Reactome Metagenes 26 D.4 Expression of Somatic Mutations 26		7.1.1 Established Functions of <i>CDH1</i>	196
7.2.1 Synthetic Lethality in the Genomic Era 197 7.2.2 Clinical Interventions based on Synthetic Lethality 198 7.3 Future Directions 200 7.4 Conclusions 202 References 204 A Sample Quality 228 A.1 Sample Correlation 228 A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 236 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 245 C.1 Synthetic Lethal Genes and Pathways 246 C.2 Synthetic Lethal Expression Profiles 247 C.3 Comparison to Primary Screen 247 C.4 Compare SLIPT genes 255 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 260 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 266 E Intrinsic Subtyping 267 F Stomach Expression Analysis 268 F.1 Synthetic Lethal Genes and Pathways 269 F.2 Comparison to Primary Screen 277 <tr< td=""><td></td><td>7.1.2 The Molecular Role of <i>CDH1</i> in Cancer</td><td>196</td></tr<>		7.1.2 The Molecular Role of <i>CDH1</i> in Cancer	196
7.2.2 Clinical Interventions based on Synthetic Lethality 198 7.3 Future Directions 200 7.4 Conclusions 202 References 204 A Sample Quality 228 A.1 Sample Correlation 228 A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 236 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 245 C.1 Synthetic Lethal Genes and Pathways 244 C.2 Synthetic Lethal Expression Profiles 244 C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 248 C.4 Compare SLIPT genes 251 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 260 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 262 E Intrinsic Subtyping 267 F Stomach Expression Analysis 268 F.1 Synthetic Lethal Genes and Pathways 269 F.2 Comparison to Primary Screen 272 F.2 Res		7.2 Significance	197
7.3 Future Directions 200 7.4 Conclusions 202 References 204 A Sample Quality 228 A.1 Sample Correlation 228 A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 236 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 245 C.1 Synthetic Lethal Genes and Pathways 246 C.2 Synthetic Lethal Expression Profiles 244 C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 246 C.4 Compare SLIPT genes 251 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 266 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 265 E Intrinsic Subtyping 267 F Stomach Expression Analysis 268 F.1 Synthetic Lethal Genes and Pathways 268 F.2 Comparison to Primary Screen 272 F.3 Metagene Analysis 274 F.3 Metagene Analysis 276 <td></td> <td>7.2.1 Synthetic Lethality in the Genomic Era</td> <td>197</td>		7.2.1 Synthetic Lethality in the Genomic Era	197
7.4 Conclusions 202 References 204 A Sample Quality 228 A.1 Sample Correlation 228 A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 236 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 245 C.1 Synthetic Lethal Genes and Pathways 246 C.2 Synthetic Lethal Expression Profiles 246 C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 246 C.4 Compare SLIPT genes 251 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 260 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 263 E Intrinsic Subtyping 267 F Stomach Expression Analysis 269 F.1 Synthetic Lethal Genes and Pathways 269 F.2 Comparison to Primary Screen 275 F.2.1 Resampling Analysis 276 F.3 Metagene Analysis 277 G Synthetic Lethal Genes in Pathways <td></td> <td>7.2.2 Clinical Interventions based on Synthetic Lethality</td> <td>199</td>		7.2.2 Clinical Interventions based on Synthetic Lethality	199
References 204 A Sample Quality 228 A.1 Sample Correlation 228 A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 236 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 245 C.1 Synthetic Lethal Genes and Pathways 245 C.2 Synthetic Lethal Expression Profiles 246 C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 245 C.4 Compare SLIPT genes 251 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 260 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 266 E Intrinsic Subtyping 267 F Stomach Expression Analysis 267 F.2 Comparison to Primary Screen 27 F.2.1 Resampling Analysis 27 F.3 Metagene Analysis 27 G Synthetic Lethal Genes in Pathways 27 H Pathway Connectivity for Mutation SLIPT 285 I Information Centra		7.3 Future Directions	200
A Sample Quality 228 A.1 Sample Correlation 228 A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 230 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 245 C.1 Synthetic Lethal Genes and Pathways 244 C.2 Synthetic Lethal Expression Profiles 244 C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 245 C.4 Compare SLIPT genes 251 D Metagene Analysis 253 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 266 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 264 E Intrinsic Subtyping 267 F Stomach Expression Analysis 268 F.1 Synthetic Lethal Genes and Pathways 269 F.2 Comparison to Primary Screen 272 F.3 Metagene Analysis 274 F.3 Metagene Analysis 274 F.3 Metagene Analysis 276 G Synthetic Lethal Genes in Pathways 277 H Pathway C		7.4 Conclusions	202
A.1 Sample Correlation 228 A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 236 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 245 C.1 Synthetic Lethal Genes and Pathways 245 C.2 Synthetic Lethal Expression Profiles 246 C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 248 C.4 Compare SLIPT genes 251 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 260 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 264 E Intrinsic Subtyping 267 F Stomach Expression Analysis 267 F.2 Comparison to Primary Screen 272 F.2.1 Resampling Analysis 274 F.3 Metagene Analysis 276 G Synthetic Lethal Genes in Pathways 276 G Synthetic Lethal Genes in Pathways 277 H Pathway Connectivity for Mutation SLIPT 285 Information Centrality for Gene Essentiality		References	20 4
A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast 236 B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 245 C.1 Synthetic Lethal Genes and Pathways 246 C.2 Synthetic Lethal Expression Profiles 244 C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 248 C.4 Compare SLIPT genes 251 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 260 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 262 E Intrinsic Subtyping 263 F Stomach Expression Analysis 264 F.1 Synthetic Lethal Genes and Pathways 265 F.2 Comparison to Primary Screen 272 F.3 Metagene Analysis 274 G Synthetic Lethal Genes in Pathways 276 G Synthetic Lethal Genes in Pathways 277 H Pathway Connectivity for Mutation SLIPT 285 I Information Centrality for Gene Essentiality 285	A	Sample Quality	228
B Software Used for Thesis 234 C Mutation Analysis in Breast Cancer 245 C.1 Synthetic Lethal Genes and Pathways 246 C.2 Synthetic Lethal Expression Profiles 244 C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 246 C.4 Compare SLIPT genes 251 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 260 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 263 E Intrinsic Subtyping 267 F Stomach Expression Analysis 268 F.1 Synthetic Lethal Genes and Pathways 269 F.2 Comparison to Primary Screen 277 F.3 Metagene Analysis 274 F.3 Metagene Analysis 276 G Synthetic Lethal Genes in Pathways 277 H Pathway Connectivity for Mutation SLIPT 285 I Information Centrality for Gene Essentiality 285		A.1 Sample Correlation	228
C Mutation Analysis in Breast Cancer 243 C.1 Synthetic Lethal Genes and Pathways 244 C.2 Synthetic Lethal Expression Profiles 244 C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 248 C.4 Compare SLIPT genes 255 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 261 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 262 E Intrinsic Subtyping 263 F Stomach Expression Analysis 263 F.1 Synthetic Lethal Genes and Pathways 263 F.2 Comparison to Primary Screen 272 F.2.1 Resampling Analysis 274 F.3 Metagene Analysis 276 G Synthetic Lethal Genes in Pathways 276 H Pathway Connectivity for Mutation SLIPT 285 I Information Centrality for Gene Essentiality 285		A.2 Replicate Samples in The Cancer Genome Atlas (TCGA) Breast	230
C.1 Synthetic Lethal Genes and Pathways 24 C.2 Synthetic Lethal Expression Profiles 24 C.3 Comparison to Primary Screen 24 C.3.1 Resampling Analysis 24 C.4 Compare SLIPT genes 25 D Metagene Analysis 25 D.1 Pathway Expression 25 D.2 Somatic Mutation 25 D.3 Synthetic Lethal Reactome Metagenes 26 D.4 Expression of Somatic Mutations 26 D.5 Metagene Expression Profiles 26 E Intrinsic Subtyping 26 F Stomach Expression Analysis 26 F.1 Synthetic Lethal Genes and Pathways 26 F.2 Comparison to Primary Screen 27 F.2.1 Resampling Analysis 27 F.3 Metagene Analysis 27 G Synthetic Lethal Genes in Pathways 27 H Pathway Connectivity for Mutation SLIPT 28 I Information Centrality for Gene Essentiality 289	В	Software Used for Thesis	234
C.2 Synthetic Lethal Expression Profiles 244 C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 248 C.4 Compare SLIPT genes 253 D Metagene Analysis 253 D.1 Pathway Expression 253 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 260 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 264 E Intrinsic Subtyping 267 F Stomach Expression Analysis 268 F.1 Synthetic Lethal Genes and Pathways 268 F.2 Comparison to Primary Screen 272 F.2.1 Resampling Analysis 274 F.3 Metagene Analysis 276 G Synthetic Lethal Genes in Pathways 277 H Pathway Connectivity for Mutation SLIPT 285 I Information Centrality for Gene Essentiality 289	\mathbf{C}	· · · · · · · · · · · · · · · · · · ·	243
C.3 Comparison to Primary Screen 247 C.3.1 Resampling Analysis 248 C.4 Compare SLIPT genes 255 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 260 D.4 Expression of Somatic Mutations 261 D.5 Metagene Expression Profiles 262 E Intrinsic Subtyping 263 F Stomach Expression Analysis 263 F.1 Synthetic Lethal Genes and Pathways 263 F.2 Comparison to Primary Screen 272 F.2.1 Resampling Analysis 274 F.3 Metagene Analysis 276 G Synthetic Lethal Genes in Pathways 277 G Synthetic Lethal Genes in Pathways 278 I Information Centrality for Gene Essentiality 289			
C.3.1 Resampling Analysis 248 C.4 Compare SLIPT genes 255 D Metagene Analysis 255 D.1 Pathway Expression 255 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 260 D.4 Expression of Somatic Mutations 265 D.5 Metagene Expression Profiles 266 E Intrinsic Subtyping 267 F Stomach Expression Analysis 268 F.1 Synthetic Lethal Genes and Pathways 268 F.2 Comparison to Primary Screen 272 F.2.1 Resampling Analysis 274 F.3 Metagene Analysis 276 G Synthetic Lethal Genes in Pathways 277 H Pathway Connectivity for Mutation SLIPT 285 I Information Centrality for Gene Essentiality 285			
C.4 Compare SLIPT genes 25 D Metagene Analysis 25 D.1 Pathway Expression 25 D.2 Somatic Mutation 25 D.3 Synthetic Lethal Reactome Metagenes 26 D.4 Expression of Somatic Mutations 26 D.5 Metagene Expression Profiles 26 E Intrinsic Subtyping 26 F Stomach Expression Analysis 26 F.1 Synthetic Lethal Genes and Pathways 26 F.2 Comparison to Primary Screen 27 F.2.1 Resampling Analysis 27 F.3 Metagene Analysis 27 G Synthetic Lethal Genes in Pathways 27 G Synthetic Lethal Genes in Pathways 27 H Pathway Connectivity for Mutation SLIPT 28 I Information Centrality for Gene Essentiality 28		C.3 Comparison to Primary Screen	247
D Metagene Analysis 253 D.1 Pathway Expression 253 D.2 Somatic Mutation 256 D.3 Synthetic Lethal Reactome Metagenes 266 D.4 Expression of Somatic Mutations 265 D.5 Metagene Expression Profiles 266 E Intrinsic Subtyping 267 F Stomach Expression Analysis 268 F.1 Synthetic Lethal Genes and Pathways 268 F.2 Comparison to Primary Screen 277 F.2.1 Resampling Analysis 276 F.3 Metagene Analysis 276 G Synthetic Lethal Genes in Pathways 273 H Pathway Connectivity for Mutation SLIPT 285 I Information Centrality for Gene Essentiality 285		1 0 v	
D.1 Pathway Expression 25 D.2 Somatic Mutation 25 D.3 Synthetic Lethal Reactome Metagenes 26 D.4 Expression of Somatic Mutations 26 D.5 Metagene Expression Profiles 26 E Intrinsic Subtyping 267 F Stomach Expression Analysis 268 F.1 Synthetic Lethal Genes and Pathways 269 F.2 Comparison to Primary Screen 270 F.2.1 Resampling Analysis 270 F.3 Metagene Analysis 270 G Synthetic Lethal Genes in Pathways 270 G Synthetic Lethal Genes in Pathways 270 H Pathway Connectivity for Mutation SLIPT 285 I Information Centrality for Gene Essentiality 285		C.4 Compare SLIPT genes	251
D.2 Somatic Mutation	\mathbf{D}	Metagene Analysis	253
D.3 Synthetic Lethal Reactome Metagenes D.4 Expression of Somatic Mutations D.5 Metagene Expression Profiles E Intrinsic Subtyping 267 F Stomach Expression Analysis F.1 Synthetic Lethal Genes and Pathways F.2 Comparison to Primary Screen F.2.1 Resampling Analysis F.3 Metagene Analysis F.3 Metagene Analysis C Synthetic Lethal Genes in Pathways H Pathway Connectivity for Mutation SLIPT I Information Centrality for Gene Essentiality 268 269 269 269 269 269 269 269 269 269 269		D.1 Pathway Expression	253
D.4 Expression of Somatic Mutations D.5 Metagene Expression Profiles E Intrinsic Subtyping F Stomach Expression Analysis F.1 Synthetic Lethal Genes and Pathways F.2 Comparison to Primary Screen F.2.1 Resampling Analysis F.3 Metagene Analysis C Synthetic Lethal Genes in Pathways H Pathway Connectivity for Mutation SLIPT I Information Centrality for Gene Essentiality 267 268 268 269 270 271 270 270 271 271 271 271 271 271 271 271 271 271		D.2 Somatic Mutation	256
D.5 Metagene Expression Profiles 264 E Intrinsic Subtyping 265 F Stomach Expression Analysis 265 F.1 Synthetic Lethal Genes and Pathways 265 F.2 Comparison to Primary Screen 275 F.2.1 Resampling Analysis 274 F.3 Metagene Analysis 276 G Synthetic Lethal Genes in Pathways 277 H Pathway Connectivity for Mutation SLIPT 285 I Information Centrality for Gene Essentiality 285		D.3 Synthetic Lethal Reactome Metagenes	260
E Intrinsic Subtyping 267 F Stomach Expression Analysis 269 F.1 Synthetic Lethal Genes and Pathways 269 F.2 Comparison to Primary Screen 277 F.2.1 Resampling Analysis 274 F.3 Metagene Analysis 276 G Synthetic Lethal Genes in Pathways 277 H Pathway Connectivity for Mutation SLIPT 285 I Information Centrality for Gene Essentiality 285		D.4 Expression of Somatic Mutations	261
F Stomach Expression Analysis F.1 Synthetic Lethal Genes and Pathways F.2 Comparison to Primary Screen F.2.1 Resampling Analysis F.3 Metagene Analysis C Synthetic Lethal Genes in Pathways F Pathway Connectivity for Mutation SLIPT Information Centrality for Gene Essentiality 269 270 271 272 273 274 275 276 277 277 277 277 277 277 277 277 277		D.5 Metagene Expression Profiles	264
F.1 Synthetic Lethal Genes and Pathways	\mathbf{E}	Intrinsic Subtyping	267
F.2 Comparison to Primary Screen F.2.1 Resampling Analysis F.3 Metagene Analysis G Synthetic Lethal Genes in Pathways H Pathway Connectivity for Mutation SLIPT I Information Centrality for Gene Essentiality 272 285	\mathbf{F}	<u>.</u>	
F.2.1 Resampling Analysis		F.1 Synthetic Lethal Genes and Pathways	269
F.3 Metagene Analysis		•	
G Synthetic Lethal Genes in Pathways H Pathway Connectivity for Mutation SLIPT I Information Centrality for Gene Essentiality 289		F.2.1 Resampling Analysis	274
H Pathway Connectivity for Mutation SLIPT I Information Centrality for Gene Essentiality 289		F.3 Metagene Analysis	276
I Information Centrality for Gene Essentiality 289	\mathbf{G}	Synthetic Lethal Genes in Pathways	277
	Н	Pathway Connectivity for Mutation SLIPT	285
J Pathway Structure for Mutation SLIPT 292	Ι	Information Centrality for Gene Essentiality	289
	${f J}$	Pathway Structure for Mutation SLIPT	292

\mathbf{K}	Per	formance of SLIPT and χ^2	295
	K.1	Correlated Query Genes affects Specificity	301
${f L}$	Sim	ulations on Graph Structures	307
		L.0.1 Simulations from Inhibiting Graph Structures	308
	L.1	Simulation across Graph Structures	311
	L.2	Simulations from Complex Graph Structures	315
		L.2.1 Simulations from Complex Inhibiting Graphs	318
	L.3	Simulations from Pathway Graph Structures	324

List of Figures

1.1	Synthetic genetic interactions
1.2	Synthetic lethality in cancer
2.1	Read count density
2.2	Read count sample mean
3.1	Framework for synthetic lethal prediction
3.2	Synthetic lethal prediction adapted for mutation
3.3	A model of synthetic lethal gene expression
3.4	Modelling synthetic lethal gene expression
3.5	Synthetic lethality with multiple genes
3.6	Simulating gene function
3.7	Simulating synthetic lethal gene function
3.8	Simulating synthetic lethal gene expression
3.9	Performance of binomial simulations
3.10	Comparison of statistical performance
	Performance of multivariate normal simulations
	Simulating expression with correlated gene blocks
	Simulating expression with correlated gene blocks
	Synthetic lethal prediction across simulations
	Performance with correlations
	Comparison of statistical performance with correlation structure 79
	Performance with query correlations
	Statistical evaluation of directional criteria
	Performance of directional criteria
	Simulated graph structures
	Simulating expression from a graph structure
	Simulating expression from graph structure with inhibitions 88
3.23	Demonstration of violin plots with custom features
3.24	Demonstration of annotated heatmap
3.25	Simulating graph structures
4.1	Synthetic lethal expression profiles of analysed samples
4.2	Comparison of SLIPT with siRNA
4.3	Comparison of SLIPT and siRNA genes with correlation 100
4.4	Comparison of SLIPT and siRNA genes with correlation 108
4.5	Comparison of SLIPT and siRNA genes with screen viability 109

4.6 4.7	Comparison of SLIPT genes with siRNA screen viability Resampled intersection of SLIPT and siRNA candidate genes	109 114
5.1	synthetic lethality in the PI3K cascade	130
5.2	synthetic lethality in Elastic Fibre Formation	132
5.3	Synthetic lethality in Fibrin Clot Formation	133
5.4	Synthetic lethality and vertex degree	136
5.5	Synthetic lethality and centrality	139
5.6	Synthetic lethality and PageRank	140
5.7	Hierarchical structure of PI3K	142
5.8	Hierarchy score in PI3K against synthetic lethality in PI3K	143
5.9	Structure of synthetic lethality in PI3K	144
5.10	Structure of synthetic lethality resampling in PI3K	146
6.1	Performance of χ^2 and SLIPT across quantiles	155
6.2	Performance of χ^2 and SLIPT across quantiles with more genes	156
6.3 6.4	Performance of χ^2 and SLIPT across quantiles with query correlation . Performance of χ^2 and SLIPT across quantiles with query correlation	157
	and more genes	158
6.5	Performance of negative correlation and SLIPT	161
6.6	Simple graph structures	164
6.7	Performance of simulations on a simple graph	165
6.8	Performance of simulations is similar in simple graphs	166
6.9	Performance of simulations on a pathway	167
6.10	Performance of simulations on a simple graph with inhibition	169
	Performance is higher on a simple inhibiting graph	171
	Performance of simulations on a constructed graph with inhibition	172
	Performance is affected by inhibition in graphs	173
	Detection of synthetic lethality within a graph structure	175
	Performance of simulations including a simple graph	179
	Performance on a simple graph improves with more genes	180
	Performance on an inhibiting graph improves with more genes	181
	Performance of simulations on the PI3K cascade	184
	Performance of simulations including the PI3K cascade	186
6.20	Performance on pathways improves with more genes	187
A.1	Correlation profiles of removed samples	228
A.2	Correlation analysis and sample removal	229
A.3	Replicate excluded samples	230
A.4	Replicate samples with all remaining	231
A.5	Replicate samples with some excluded	232
C.1	Synthetic lethal expression profiles of analysed samples	245
C.2	Comparison of mtSLIPT to siRNA	247
C.3	Compare mtSLIPT and siRNA genes with correlation	251
C.4	Compare mtSLIPT and siRNA genes with correlation	251
C.5	Compare mtSLIPT and siRNA genes with siRNA viability	252

D.1	Pathway metagene expression profiles	255
D.2	Expression profiles for constituent genes of PI3K	257
D.3	Expression profiles for estrogen receptor related genes	258
D.4	Somatic mutation against the PI3K metagene	259
D.5	Somatic mutation against PIK3CA metagene	261
D.6	Somatic mutation against PI3K protein	262
D.7	Somatic mutation against AKT protein	263
D.8	Pathway metagene expression profiles	264
D.9	Expression profiles for p53 related genes	265
D.10	Expression profiles for BRCA related genes	266
F.1	Synthetic lethal expression profiles of stomach samples	271
F.2	Comparison of SLIPT in stomach to siRNA	272
G.1	Synthetic lethality in the PI3K/AKT pathway	277
G.2	Synthetic lethality in the PI3K/AKT pathway in cancer	278
G.3	Synthetic lethality in the Extracellular Matrix	279
G.4	Synthetic lethality in the GPCRs	280
G.5	Synthetic lethality in the GPCR Downstream	281
G.6	Synthetic lethality in the Translation Elongation	282
G.7	Synthetic lethality in the Nonsense-mediated Decay	283
G.8	Synthetic lethality in the 3' UTR	284
H.1	Synthetic lethality and vertex degree	285
H.2	Synthetic lethality and centrality	286
H.3	Synthetic lethality and PageRank	287
I.1	Information centrality distribution	291
Т 1		000
J.1	Synthetic lethality and heirarchy score in PI3K	292
J.2	Heirarchy score in PI3K against synthetic lethality in PI3K	293
J.3	Structure of synthetic lethality in PI3K	293
J.4	Structure of synthetic lethality resampling	294
K.1	Performance of χ^2 and SLIPT across quantiles	295
K.2	Performance of χ^2 and SLIPT across quantiles	297
K.3	Performance of χ^2 and SLIPT across quantiles with more genes	299
K.4	Performance of χ^2 and SLIPT across quantiles with query correlation .	301
K.5	Performance of χ^2 and SLIPT across quantiles with query correlation .	303
K.6	Performance of χ^2 and SLIPT across quantiles with query correlation and more genes	305
т 1		
L.1	Performance of simulations on a simple graph	307
L.2	Performance of simulations on an inhibiting graph	308
L.3	Performance of simulations on a constructed graph with inhibition	309
L.4	Performance of simulations on a constructed graph with inhibition	310
L.5	Detection of synthetic lethality within a graph structure	311
L.6	Detection of synthetic lethality within an inhibiting graph	313

L.7	Detection of synthetic lethality within an inhibiting graph	314
L.8	Performance of simulations on a branching graph	315
L.9	Performance of simulations on a complex graph	316
L.10	Performance of simulations on a large graph	317
L.11	Performance of simulations on a branching graph with inhibition	318
L.12	Performance of simulations on a branching graph with inhibition	319
L.13	Performance of simulations on a complex graph with inhibition	320
L.14	Performance of simulations on a complex graph with inhibition	321
L.15	Performance of simulations on a large constructed graph with inhibition	322
L.16	Performance of simulations on a large constructed graph with inhibition	323
L.17	Performance of simulations on the $G_{\alpha i}$ signalling pathway	324
L.18	Performance of simulations including the $G_{\alpha i}$ signalling pathway	325

List of Tables

1.1 1.2 1.3	Methods for predicting genetic interactions	22 23 25
2.1 2.2 2.3 2.4 2.5 2.6	Excluded samples by batch and clinical characteristics. Computers used during thesis	43 53 54 55 55 57
4.1 4.2 4.3 4.4 4.5 4.6 4.7 4.8 4.9	Candidate synthetic lethal gene partners of <i>CDH1</i> from SLIPT	98 99 104 107 111 112 115 116 119
5.1 5.2 5.3 5.4 5.5	ANOVA for synthetic lethality and vertex degree	137 139 141 144 148
B.1 C.1 C.2 C.3 C.4 C.5 C.6	Complete list of R packages used during this thesis	234 243 244 246 248 249 250
D.1	Candidate synthetic lethal metagenes against CDH1 from mtSLIPT	260

E.1	Comparison of intrinsic subtypes	267
F.1	Synthetic lethal gene partners of <i>CDH1</i> from SLIPT in stomach cancer	269
F.2	Pathways for <i>CDH1</i> partners from SLIPT in stomach cancer	270
F.4	Pathways for <i>CDH1</i> partners from SLIPT and siRNA	273
F.5	Pathways for <i>CDH1</i> partners from SLIPT in stomach cancer	274
F.6	Pathways for <i>CDH1</i> partners from SLIPT in stomach and siRNA	275
F.7	Synthetic lethal metagenes against <i>CDH1</i> in stomach cancer	276
H.1	ANOVA for synthetic lethality and vertex degree	288
H.2	ANOVA for synthetic lethality and information centrality	288
H.3	ANOVA for synthetic lethality and PageRank centrality	288
I.1	Information centrality for genes and molecules in the Reactome network	290
J.1 J.2	ANOVA for synthetic lethality and PI3K hierarchy Resampling for pathway structure of synthetic lethal detection methods	

Glossary

bioinformatics Statistical or computational approaches to bi-

ological data or research tools.

chemoprevention The use of drugs to prevent early-stage can-

cers, generally applied to high-risk mutation

carriers.

E-cadherin Epithelial cadherin (calcium-dependent ad-

hesion), a cell-adhesion protein encoded by

CDH1.

essential A gene which is required to be functional or

expressed for a cell or organism to be viable,

grow or develop.

familial A trait recurrently occurring in families, not

necessarily with a genetic cause.

functional redundancy Genes which perform a common function, also

known as genetic redundancy.

A measure of the relative expression of each gene expression

gene from the mRNA extracted from (pooled)

cells.

genome All of the DNA sequence in the genome.

genomic The use of data from all genes in the genome. graph or network

A mathematical structure modelling or depict-

ing the relationships between elements.

A consistent signal of expression for a collecmetagene

> tion of genes such as a biological pathway, derived from singular value decomposition.

A variant or dysfunctional phenotype arising mutant

from a mutation in a gene.

mutation A change in DNA sequence that disrupts gene

function.

oncogene A gene that potentially causes cancer, typi-

cally by over-expression or mutant gene vari-

ants.

pleiotropy When a gene has multiple biological functions.

sporadic cancer Cancers which do occur in patients with a fam-

ily history or carry a high-risk genetic variant.

synthetic lethal Genetic interactions where inactivation of

multiple genes is inviable (or deleterious) which are viable if inactivated separately.

targeted therapy Cancer treatment that specifically acts against

a molecular target, in contrast to standard

chemotherapy.

treatment Medical procedures for a disease to improve

patient outcomes.

tumour suppressor A gene potentially causes cancer, typically by

disruption of functions which protect the cell

from cancer.

Acronyms

ANOVA Analysis of Variance.

DNA Deoxyribonucleic Acid.

GPCR G Crotein Coupled Receptor.

HDGC Hereditary Diffuse Gastric Cancer.

mtSLIPT Synthetic Lethal Interaction Prediction Tool

(against mutation).

NMD Nonsense-Mediated Decay.

RNAi RNA Interference.

siRNA Short Interfering RNA.

SLIPT Synthetic Lethal Interaction Prediction Tool.

TCGA The Cancer Genome Atlas (genomics project).

UTR Untranslated Region (of mRNA).

Chapter 7

Discussion

This thesis combines analysis of gene expression data from TCGA with experimental screening results (Telford et al., 2015) to demonstrate synthetic lethal discovery for partners of CDH1. Together these findings further elucidate the functions of CDH1 in the cell, functional redundancy in cancer, and represent potential targets against loss of CDH1 function. These candidate synthetic lethal genes were further investigated for relationships within synthetic lethal pathways, developing a network-based approach to compare genes identified in genomics experiments and analyses in the process.

The synthetic lethal detection methodology, SLIPT, was applied to gene expression data throughout this thesis and was evaluated with simulated data. A simulation procedure was developed to stringently generate gene expression data from known synthetic lethal partners in simulated data. These simulations included simple and complex correlation structures and modelling synthetic lethal genes within pathways. Together, these results demonstrate SLIPT as a robust widely applicable gene expression analysis procedure (for which an R package has been released) for discovery of synthetic lethal partner genes. Performance of SLIPT on simulated data also highlights the strengths of the procedure and future directions to improve upon it.

7.1 Synthetic Lethality and *CDH1* Biology

The *CDH1* tumour suppressor gene was the focus of identifying synthetic lethal partners to demonstrate the novel SLIPT methodology. This gene is important in sporadic breast and stomach cancers, in addition to familial syndromes, such as hereditary diffuse gastric cancer (HDGC). The analysis of synthetic lethal partners of *CDH1* in breast and stomach cancers was enabled by the availability of molecular data (Bass

et al., 2014; Koboldt et al., 2012) and a synthetic lethal screen conducted in MCF10A breast cells (Chen et al., 2014; Telford et al., 2015).

Synthetic lethal interactions arise due to functional redundancy (Boone et al., 2007; Fece de la Cruz et al., 2015; Kaelin, Jr, 2005) and as such the synthetic lethal partners of CDH1 indicates the wide-ranging biological functions that E-cadherin is involved in. The diverse synthetic lethal pathways identified supports the known pleiotropic nature of the CDH1 gene by detecting established functions of CDH1, replicating candidates from an experimental screen (Telford et al., 2015), and identifying novel interactions with candidate genes and pathways for further investigation. The highly pleiotropic functions of E-cadherin was also consistent with CDH1 being a tumour suppressor gene.

7.1.1 Established Functions of *CDH1*

The *CDH1* has established functions in cell-cell communication and maintaining the cytoskeleton, specifically with cell-cell adhesion by forming tight junctions and the adherens complex. More recently, additional functions of *CDH1* in the extracellular matrix and fibrin clotting have also been identified. Synthetic lethal interactions within biological pathways (i.e., partners in the same pathway as the query gene) are expected according to previous synthetic lethal experiments (Boone *et al.*, 2007; Kelley and Ideker, 2005). Synthetic lethal interactions identified in these pathways are consistent with these being functions of *CDH1*, in addition to potentially actionable targets against cancers.

7.1.2 The Molecular Role of *CDH1* in Cancer

The involvement of *CDH1* in the extracellular matrix is important in cancers as it indicates a mechanism by which *CDH1* loss may affect the tumour microenvironment, contributing to its role as a tumour and invasion suppressor. Furthermore, perturbations in the extracellular matrix and tumour microenvironment present a means by which to specifically inhibit (cancerous) *CDH1*-deficient cells, in addition to those currently being considered. These may be further supported in further investigations with 3D cell culture, "organoid", or mouse xenograft cancer models.

In contrast, many of the pathways involved in cell signalling, including G protein coupled receptors, were identified by SLIPT in addition to the experimental screen (Telford *et al.*, 2015). These support the previous results in cell line models, that these pathways are essential to the growth of *CDH1*-deficient cancers and present a potential vulnerability specific to these (cancerous) cells. Furthermore, the replication of

synthetic lethality of *CDH1* with cell signalling pathways in TCGA data across cancer types and genetic backgrounds robustly supports these pathways being clinically applicable beyond the genetic background of the model system of *CDH1*-/- MCF10A cells (Chen *et al.*, 2014). While the specific synthetic lethal genes were not as consistently detected between the SLIPT analyses and siRNA screen (Telford *et al.*, 2015), they were sufficient to identify synthetic lethal pathways for further experimental investigation, which are more likely to be replicated between genetic backgrounds (Dixon *et al.*, 2008). Together these results demonstrate how SLIPT can be integrated with an experimental screen to triage potential therapeutic targets for further pre-clinical investigation.

The analysis of expression data with SLIPT is also indicative of additional biological mechanisms of synthetic lethality in pathways beyond those identified in screening experiments (Telford $et\ al.$, 2015). In particular, translation and regulatory pathways, involving 3' untranslated regions (UTRs) and nonsense-mediated decay (NMD), were identified as candidate synthetic lethal pathways with CDH1 by SLIPT. These pathways represent downstream targets regulated by the putative synthetic lethal signalling pathways which cancer cells are dependent on for sustained protein expression to proliferate and evade host defense processes such as apoptosis and immune responses (Gao and Roux, 2015) .

7.2 Significance

7.2.1 Synthetic Lethality in the Genomic Era

Development of an effective synthetic lethal discovery tool for bioinformatic analysis has a wide range of applications in genetics research including functional genomics, medical and agricultural applications. The SLIPT approach demonstrated in this thesis is widely applicable to other genes and biological questions. In addition to further query of cancer genes, including other tissues, synthetic lethal gene functions are also of wider interest for their implications for genetic redundancy. Highly redundant genes, and the genetically robust systems they give rise to, are of further relevance to evolutionary, developmental, and systems biology to understand how these change over time and play a role in fundamental development of cell types, in addition to cancers (Boone et al., 2007; Nowak et al., 1997; Tischler et al., 2008).

Developmental genes in particular, are highly evolutionary conserved and subject to high rates of redundancy (Fromental-Ramain *et al.*, 1996; Kockel *et al.*, 1997; Nowak

et al., 1997). These are often difficult to study with conventional functional genetics since individual knockouts of redundant genes do not necessarily have a mutant phenotype. Identifying genes with a common function is therefore also important to the study of developmental genes with unknown functions. Synthetic lethal discovery methods such as SLIPT provide a genomic approach to further systematic characterisation of gene function including such highly redundant developmental genes.

Similarly, variants of unknown significance and modifier loci are a major concerns in human genetics, including "monogenic" and "rare" diseases. Many of these could potentially be difficult to characterise individually due to synthetic lethal interactions where additional loci contribute to the disease (or only compensate for some variants). As such systematic identification of synthetic lethal interactions also has applications in the study of such "oligogenic" diseases along with similar applications in the study of heritability for traits including agricultural genomic selection.

Genetic redundancy is also a concern in pharmacology. Polypharmacology and network medicine are rationales to account for this by using drugs with multiple (known and specific) targets (Barabási et al., 2011; Hopkins, 2008). Further characterisation of synthetic lethal genes will be valuable to the design of effective multi-target drugs or combination therapies in a range of therapeutic applications including molecular targeted therapies against cancer for which combination therapies are a popular solution for acquired resistance against individual targeted therapies. Characterisation of genetic interactions and combination therapies also has the potential to expand pharmacogenomic investigations. These may elucidate the impact of genotypes at multiple loci, which lead to adverse effects in a subset of the population due to variants in synthetic lethal genes.

Furthermore, redundant functions and synthetic lethal interactions also present a means to expand upon the concept of the "minimal" genome (Hutchison *et al.*, 2016). It is important to account for essential gene functions that are performed by redundant genes (or in combination with pleiotropic genes), rather than simply those that are perturbed by individual genes. An essential gene approach is likely to produce an underestimate that does not account for synthetic lethal interactions.

Synthetic lethal interactions are fundamentally important throughout genetics. Further understanding of them in a genomic context, facilitated by methods such as SLIPT, would contribute towards deeper understanding of gene functions and their role in traits or diseases in the post-genomic era. Genes do not function in isolation and understanding them in the context of the complexity of a cell and across genetic

backgrounds is essential to further characterise their functions and ensure that findings can be validated or applied beyond experimental systems.

7.2.2 Clinical Interventions based on Synthetic Lethality

Synthetic lethal discovery with SLIPT is of particular interest in cancer research as a complementary approach to discovery of synthetic lethal drug targets. The cancer research community relies on cell line and mouse models for screening and validation experiments (Fece de la Cruz et al., 2015) which would benefit from integration with gene expression analysis as demonstrated for CDH1 and the screen conducted by Telford et al. (2015). Synthetic lethal drug design against cancer mutations, including gene loss or over-expression, could lead to a revolution in cancer therapy and chemoprevention. Such therapeutics would enable personalised treatment for cancer patients and high risk individuals. Examples of the synthetic lethal strategy (Bryant et al., 2005; Farmer et al., 2005) for cancer treatment have been shown to be clinically effective McLachlan et al. (2016). Many large-scale RNA interference (RNAi) screens have been conducted recently, aiming to discover gene function and drug targets for similar application with other cancer genes, including cancers in other tissues (Fece de la Cruz et al., 2015).

While SLIPT analysis and RNAi screens represent a significant step towards anticancer medicines, further validation is required to ensure that the synthetic lethal candidate genes and pathways identified for *CDH1* in breast and stomach cancer are applicable against *CDH1*-deficient cancers in the clinic. Validation with RNAi or pharmacological inhibitors is needed since false positives may occur in SLIPT analysis or siRNA screens. These candidates will need to be tested in pre-clinical models (cell lines and mouse xenografts) before proceeding to clinical trials. A therapeutic intervention will also require a targeted therapeutic to develop developed or repurposed against the synthetic lethal partner. Drug targets could be triaged from synthetic lethal genes by functions known to be amenable to drugs or structure with conserved specific sites that are not homologous to other genes, or those with existing drugs approved in trial for other applications. Both structure-aided drug design and compound screening are viable ways to target synthetic lethal partners.

Targeted therapeutics designed based on synthetic lethal interactions could expand the applications of "precision medicine" against molecular targets. Synthetic lethality expands the range of cancer genes which can be (indirectly) targeted to include tumour suppressor genes with loss of function, such as *CDH1*. Oncogenes with disrupted functions that are over-expressed or highly homologous to non-cancerous proto-oncogenes, such as MYC, EGFR or KRAS, may also be targeted by synthetic lethality. Applications against tumour suppressor genes is particularly important, as these cannot be approached by careful dosing. Synthetic lethal drug design has the benefit of being highly specific against a particular genotype (such as $CDH1^{-/-}$) with the potential for targeted therapies with a wide therapeutic index and few adverse effects, in contrast to many current anti-cancer drug regimens (Hopkins, 2008; Kaelin, Jr, 2009). These properties are highly desirable for chemoprevention applications, such as treatment against CDH1-deficient in HDGC patients (Guilford $et\ al.$, 2010), as an alternative to monitoring or surgery.

7.3 Future Directions

While further validation and pre-clinical testing is required to translate the findings for *CDH1* to cancer therapy or prevention, there are also further avenues for research into the detection of synthetic lethality in gene expression and other genomics data. The SLIPT methodology is amenable to wider application against a range of genes for which loss of function is deleterious, including other cancer genes in breast cancer or other tissues. Synthetic lethal interactions are functionally informative, particularly for mode-of-action of known drug targets, and are also relevant for identifying functions of newly characterised genes in genomics studies and designing specific interventions against cells with loss of function in cancer and other diseases. Thus synthetic lethal detection using SLIPT in expression data could be further used for many other genes, including others relevant to human health and disease.

These investigations do not need to be limited to expression data. While expression as a measure of gene function has been the focus of this thesis, other genomics data could be used for a similar purpose for SLIPT analysis. These include DNA copy number, DNA methylation, histone activation, mutation status, protein abundance, and protein activation state. For some applications or genes, these molecular profiles may be more informative of gene function and synthetic lethal relationships. However, expression was the focus of the investigations thus far as a widely accepted measure of gene function which has widely available genomics data. SLIPT is compatible with each of these data types (if the thresholds are selected appropriately) and may perform better for some applications with these molecular profiles or a weighted combination of these. As demonstrated, SLIPT is also suitable for future investigations with pathway metagenes and other summary data as well.

It may also be possible to improve the performance of SLIPT with refinements to the statistical or computational approach. This thesis has focused on rational query-based approach which computes relatively quickly, even in R (R Core Team, 2016), and is relatively intuitive to interpret. These computations are compatible with parallel computing and the computational resources may be further reduced by using a different computing language. The slipt R package has been documented and released open-source (as described in Section 3.5) to facilitate further development, wider adoption, or comparison with other scientific software for similar purposes.

Alternative methods may be also improve on the statistical performance of SLIPT. In particular, the sensitivity was generally as issue with higher numbers of synthetic lethal partners in simulated data. While approaches using continuous data such as Pearson correlation and linear regression did not perform as well as SLIPT, they could be improved. A least squares regression approach in particular, enables multiple measures of relationships such as the coefficients of the fitted curve and significance of the fit (computed from the residuals). A linear modelling approach using regression is also amenable to refinement such as extending from fitting a linear relationship to a polynomial or logistic regression. Another benefit to fitting linear models is that these would enable the conditioning of known synthetic lethal partners to identify subtle signatures of further interacting partners.

This approach could also be applied iteratively on the strongest candidates from previous synthetic lethal analyses in further rounds of prediction conditioned upon them. Similarly, synthetic lethal prediction could also be approached with a Bayesian framework which is also amenable to Bayesian priors on known or previously predicted synthetic lethal partners. Either of these approaches has the potential to improve upon the synthetic lethal predictions which have been demonstrated as possible and biologically relevant by SLIPT.

7.4 Conclusions

Synthetic lethal interactions are important for understanding gene function and development of highly specific targeted anti-cancer treatments. Synthetic lethality could expand the repertoire of applications for precision cancer medicine to indirectly targeting loss of function in tumour suppressor genes. Synthetic lethal discovery with experimental screening is error prone and limited by the model systems in which it is performed. There is a need for bioinformatics tool to predict synthetic lethal interactions from gene expression data facilitates rapid identification of synthetic lethal candidates to augment functional genetic screens and cancer drug target triage. I present the original Synthetic Lethal Interaction Prediction Tool (SLIPT) methodology as a statically robust procedure which performs this analysis.

The SLIPT methodology has been demonstrated to identify biologically relevant genes and pathways. An comprehensive analysis of synthetic lethal partners of the CDH1 was performed in TCGA breast cancer data (Koboldt et~al., 2012) with many of these findings replicated in stomach cancer data (Bass et~al., 2014). These genes clustered into several distinct groups, with distinct biological functions and elevated expression in different clinical subtypes. These analyses identified of synthetic lethal candidates in the $G_{\alpha i}$ signalling, cytoplasmic microfibres, and extracellular fibrin clotting pathways which were validated in an siRNA screen performed by Telford et~al. (2015) and consistent with the known cytoskeletal and cell signalling roles of E-cadherin. These findings support interventions against these pathways being applicable to specific cancer therapeutics beyond the pre-clinical cell line models in which they were validated. SLIPT has also identified synthetic lethal partners in novel pathways for CDH1 including the regulation of immune signalling and translational elongation which extend the range of pleiotropic functions of CDH1 and present further biological mechanisms to investigate the malignancy and vulnerabilities of CDH1-deficient cancers.

While some of these pathways are not expected to be detected in an isolated experimental cell line model, pathway structure may have accounted for this disparity. Thus synthetic lethal candidates detected by SLIPT and siRNA were compared within graph structures of the candidate synthetic lethal pathways. However, this did not generally account for differences between detection by these approaches. Neither synthetic lethal detection methodology preferentially detected genes of more importance or connectivity in pathway structures using established network metrics. Nor could it

be generally established that SLIPT gene candidates were upstream or downstream of siRNA gene candidates in pathway structures across biological pathways.

Pathway graph structures were also included in investigations with simulated data to ascertain whether the SLIPT procedure performed desirably in data with complex correlation structures derived based on biological pathways. A simulation procedure was developed based on a statistical model of synthetic lethality which generates multivariate normal data with known synthetic lethal partners and correlation structures. The SLIPT methodology had high statistical performance, particularly when detecting few synthetic lethal genes, with large sample sizes, and a background of many non synthetic lethal genes to distinguish true partners from. This method had high specificity, performed better than Pearson correlation or the χ^2 -test, and had had optimal performance across simulation parameter combinations for the thresholds used throughout this thesis. These findings were robust across correlation structures, including those derived from complex pathway structures containing strong positive and negative correlations between genes. Together, these findings support the release of the SLIPT software R packages and the application of the method to identify synthetic lethal genes within pathways and use candidate synthetic lethal genes to identify synthetic lethal pathways as demonstrated in this thesis.

Therefore, I present a widely applicable synthetic lethal procedure using gene expression data for wider use in genomics research, including the development of precision cancer medicine. This methodology is supported by the release of a software package in R, simulation results based on a statistical model of synthetic lethality, the demonstration of bioinformatics and network biology investigations into interactions with the *CDH1* gene in breast and stomach cancers.

References

- Abeshouse, A., Ahn, J., Akbani, R., Ally, A., Amin, S., Andry, C.D., Annala, M., Aprikian, A., Armenia, J., Arora, A., et al. (2015) The Molecular Taxonomy of Primary Prostate Cancer. Cell, 163(4): 1011–1025.
- Adler, D. (2005) vioplot: Violin plot. R package version 0.2.
- Akbani, R., Akdemir, K.C., Aksoy, B.A., Albert, M., Ally, A., Amin, S.B., Arachchi, H., Arora, A., Auman, J.T., Ayala, B., et al. (2015) Genomic Classification of Cutaneous Melanoma. *Cell*, **161**(7): 1681–1696.
- Akobeng, A.K. (2007) Understanding diagnostic tests 3: receiver operating characteristic curves. *Acta Pdiatrica*, **96**(5): 644–647.
- American Cancer Society (2017) Genetics and cancer. https://www.cancer.org/cancer/cancer-causes/genetics.html. Accessed: 22/03/2017.
- Anjomshoaa, A., Lin, Y.H., Black, M.A., McCall, J.L., Humar, B., Song, S., Fukuzawa, R., Yoon, H.S., Holzmann, B., Friederichs, J., et al. (2008) Reduced expression of a gene proliferation signature is associated with enhanced malignancy in colon cancer. Br J Cancer, 99(6): 966–973.
- Araki, H., Knapp, C., Tsai, P., and Print, C. (2012) GeneSetDB: A comprehensive meta-database, statistical and visualisation framework for gene set analysis. *FEBS Open Bio*, **2**: 76–82.
- Ashburner, M., Ball, C.A., Blake, J.A., Botstein, D., Butler, H., Cherry, J.M., Davis, A.P., Dolinski, K., Dwight, S.S., Eppig, J.T., et al. (2000) Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. Nat Genet, 25(1): 25–29.
- Ashworth, A. (2008) A synthetic lethal therapeutic approach: poly(adp) ribose polymerase inhibitors for the treatment of cancers deficient in dna double-strand break repair. J Clin Oncol, 26(22): 3785–90.

- Ashworth, A., Lord, C.J., and Reis-Filho, J.S. (2011) Genetic interactions in cancer progression and treatment. *Cell*, **145**(1): 30–38.
- Audeh, M.W., Carmichael, J., Penson, R.T., Friedlander, M., Powell, B., Bell-McGuinn, K.M., Scott, C., Weitzel, J.N., Oaknin, A., Loman, N., et al. (2010) Oral poly(adp-ribose) polymerase inhibitor olaparib in patients with *BRCA1* or *BRCA2* mutations and recurrent ovarian cancer: a proof-of-concept trial. *Lancet*, **376**(9737): 245–51.
- Babyak, M.A. (2004) What you see may not be what you get: a brief, nontechnical introduction to overfitting in regression-type models. *Psychosom Med*, **66**(3): 411–21.
- Bamford, S., Dawson, E., Forbes, S., Clements, J., Pettett, R., Dogan, A., Flanagan, A., Teague, J., Futreal, P.A., Stratton, M.R., et al. (2004) The COSMIC (Catalogue of Somatic Mutations in Cancer) database and website. Br J Cancer, 91(2): 355–358.
- Barabási, A.L. and Albert, R. (1999) Emergence of scaling in random networks. *Science*, **286**(5439): 509–12.
- Barabási, A.L., Gulbahce, N., and Loscalzo, J. (2011) Network medicine: a network-based approach to human disease. *Nat Rev Genet*, **12**(1): 56–68.
- Barabási, A.L. and Oltvai, Z.N. (2004) Network biology: understanding the cell's functional organization. *Nat Rev Genet*, **5**(2): 101–13.
- Barrat, A. and Weigt, M. (2000) On the properties of small-world network models. The European Physical Journal B - Condensed Matter and Complex Systems, 13(3): 547–560.
- Barretina, J., Caponigro, G., Stransky, N., Venkatesan, K., Margolin, A.A., Kim, S., Wilson, C.J., Lehar, J., Kryukov, G.V., Sonkin, D., et al. (2012) The Cancer Cell Line Encyclopedia enables predictive modelling of anticancer drug sensitivity. Nature, 483(7391): 603–607.
- Barry, W.T. (2016) safe: Significance Analysis of Function and Expression. R package version 3.14.0.

- Baryshnikova, A., Costanzo, M., Dixon, S., Vizeacoumar, F.J., Myers, C.L., Andrews, B., and Boone, C. (2010a) Synthetic genetic array (sga) analysis in saccharomyces cerevisiae and schizosaccharomyces pombe. *Methods Enzymol*, **470**: 145–79.
- Baryshnikova, A., Costanzo, M., Kim, Y., Ding, H., Koh, J., Toufighi, K., Youn, J.Y., Ou, J., San Luis, B.J., Bandyopadhyay, S., et al. (2010b) Quantitative analysis of fitness and genetic interactions in yeast on a genome scale. Nat Meth, 7(12): 1017–1024.
- Bass, A.J., Thorsson, V., Shmulevich, I., Reynolds, S.M., Miller, M., Bernard, B., Hinoue, T., Laird, P.W., Curtis, C., Shen, H., et al. (2014) Comprehensive molecular characterization of gastric adenocarcinoma. Nature, 513(7517): 202–209.
- Bates, D. and Maechler, M. (2016) Matrix: Sparse and Dense Matrix Classes and Methods. R package version 1.2-7.1.
- Bateson, W. and Mendel, G. (1909) Mendel's principles of heredity, by W. Bateson. University Press, Cambridge [Eng.].
- Becker, K.F., Atkinson, M.J., Reich, U., Becker, I., Nekarda, H., Siewert, J.R., and Hfler, H. (1994) E-cadherin gene mutations provide clues to diffuse type gastric carcinomas. *Cancer Research*, **54**(14): 3845–3852.
- Bell, D., Berchuck, A., Birrer, M., Chien, J., Cramer, D., Dao, F., Dhir, R., DiSaia, P., Gabra, H., Glenn, P., et al. (2011) Integrated genomic analyses of ovarian carcinoma. Nature, 474(7353): 609–615.
- Benjamini, Y. and Hochberg, Y. (1995) Controlling the false discovery rate: A practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society Series B (Methodological)*, **57**(1): 289–300.
- Berx, G., Cleton-Jansen, A.M., Nollet, F., de Leeuw, W.J., van de Vijver, M., Cornelisse, C., and van Roy, F. (1995) E-cadherin is a tumour/invasion suppressor gene mutated in human lobular breast cancers. *EMBO J*, **14**(24): 6107–15.
- Berx, G., Cleton-Jansen, A.M., Strumane, K., de Leeuw, W.J., Nollet, F., van Roy, F., and Cornelisse, C. (1996) E-cadherin is inactivated in a majority of invasive human lobular breast cancers by truncation mutations throughout its extracellular domain. *Oncogene*, **13**(9): 1919–25.

- Berx, G. and van Roy, F. (2009) Involvement of members of the cadherin superfamily in cancer. *Cold Spring Harb Perspect Biol*, **1**: a003129.
- Bitler, B.G., Aird, K.M., Garipov, A., Li, H., Amatangelo, M., Kossenkov, A.V., Schultz, D.C., Liu, Q., Shih Ie, M., Conejo-Garcia, J.R., et al. (2015) Synthetic lethality by targeting ezh2 methyltransferase activity in arid1a-mutated cancers. Nat Med, 21(3): 231–8.
- Blake, J.A., Christie, K.R., Dolan, M.E., Drabkin, H.J., Hill, D.P., Ni, L., Sitnikov, D., Burgess, S., Buza, T., Gresham, C., et al. (2015) Gene Ontology Consortium: going forward. Nucleic Acids Res, 43(Database issue): D1049–1056.
- Boone, C., Bussey, H., and Andrews, B.J. (2007) Exploring genetic interactions and networks with yeast. *Nat Rev Genet*, **8**(6): 437–49.
- Borgatti, S.P. (2005) Centrality and network flow. Social Networks, 27(1): 55 71.
- Boucher, B. and Jenna, S. (2013) Genetic interaction networks: better understand to better predict. *Front Genet*, 4: 290.
- Bozovic-Spasojevic, I., Azambuja, E., McCaskill-Stevens, W., Dinh, P., and Cardoso, F. (2012) Chemoprevention for breast cancer. *Cancer treatment reviews*, **38**(5): 329–339.
- Breiman, L. (2001) Random forests. Machine Learning, 45(1): 5–32.
- Brin, S. and Page, L. (1998) The anatomy of a large-scale hypertextual web search engine. *Computer Networks and ISDN Systems*, **30**(1): 107 117.
- Brouxhon, S.M., Kyrkanides, S., Teng, X., Athar, M., Ghazizadeh, S., Simon, M., O'Banion, M.K., and Ma, L. (2014) Soluble E-cadherin: a critical oncogene modulating receptor tyrosine kinases, MAPK and PI3K/Akt/mTOR signaling. *Oncogene*, **33**(2): 225–235.
- Brückner, A., Polge, C., Lentze, N., Auerbach, D., and Schlattner, U. (2009) Yeast two-hybrid, a powerful tool for systems biology. *Int J Mol Sci.*, **10**(6): 2763–2788.
- Bryant, H.E., Schultz, N., Thomas, H.D., Parker, K.M., Flower, D., Lopez, E., Kyle, S., Meuth, M., Curtin, N.J., and Helleday, T. (2005) Specific killing of *BRCA2*-deficient tumours with inhibitors of polyadpribose polymerase. *Nature*, **434**(7035): 913–7.

- Bussey, H., Andrews, B., and Boone, C. (2006) From worm genetic networks to complex human diseases. *Nat Genet*, **38**(8): 862–3.
- Butland, G., Babu, M., Diaz-Mejia, J.J., Bohdana, F., Phanse, S., Gold, B., Yang, W., Li, J., Gagarinova, A.G., Pogoutse, O., et al. (2008) esga: E. coli synthetic genetic array analysis. Nat Methods, 5(9): 789–95.
- cBioPortal for Cancer Genomics (cBioPortal) (2017) cBioPortal for Cancer Genomics. http://www.cbioportal.org/. Accessed: 26/03/2017.
- Cerami, E.G., Gross, B.E., Demir, E., Rodchenkov, I., Babur, O., Anwar, N., Schultz, N., Bader, G.D., and Sander, C. (2011) Pathway Commons, a web resource for biological pathway data. *Nucleic Acids Res*, 39(Database issue): D685–690.
- Chen, A., Beetham, H., Black, M.A., Priya, R., Telford, B.J., Guest, J., Wiggins, G.A.R., Godwin, T.D., Yap, A.S., and Guilford, P.J. (2014) E-cadherin loss alters cytoskeletal organization and adhesion in non-malignant breast cells but is insufficient to induce an epithelial-mesenchymal transition. *BMC Cancer*, **14**(1): 552.
- Chen, S. and Parmigiani, G. (2007) Meta-analysis of BRCA1 and BRCA2 penetrance. J Clin Oncol, 25(11): 1329–1333.
- Chipman, K. and Singh, A. (2009) Predicting genetic interactions with random walks on biological networks. BMC Bioinformatics, $\mathbf{10}(1)$: 17.
- Christofori, G. and Semb, H. (1999) The role of the cell-adhesion molecule E-cadherin as a tumour-suppressor gene. *Trends in Biochemical Sciences*, **24**(2): 73 76.
- Ciriello, G., Gatza, M.L., Beck, A.H., Wilkerson, M.D., Rhie, S.K., Pastore, A., Zhang, H., McLellan, M., Yau, C., Kandoth, C., et al. (2015) Comprehensive Molecular Portraits of Invasive Lobular Breast Cancer. Cell, 163(2): 506–519.
- Clark, M.J. (2004) Endogenous Regulator of G Protein Signaling Proteins Suppress G o-Dependent -Opioid Agonist-Mediated Adenylyl Cyclase Supersensitization.

 Journal of Pharmacology and Experimental Therapeutics, 310(1): 215–222.
- Collingridge, D.S. (2013) A primer on quantitized data analysis and permutation testing. *Journal of Mixed Methods Research*, **7**(1): 81–97.

- Collins, F.S. and Barker, A.D. (2007) Mapping the cancer genome. Pinpointing the genes involved in cancer will help chart a new course across the complex landscape of human malignancies. *Sci Am*, **296**(3): 50–57.
- Collisson, E., Campbell, J., Brooks, A., Berger, A., Lee, W., Chmielecki, J., Beer, D., Cope, L., Creighton, C., Danilova, L., et al. (2014) Comprehensive molecular profiling of lung adenocarcinoma. Nature, 511(7511): 543–550.
- Costanzo, M., Baryshnikova, A., Bellay, J., Kim, Y., Spear, E.D., Sevier, C.S., Ding, H., Koh, J.L., Toufighi, K., Mostafavi, S., et al. (2010) The genetic landscape of a cell. Science, 327(5964): 425–31.
- Costanzo, M., Baryshnikova, A., Myers, C.L., Andrews, B., and Boone, C. (2011) Charting the genetic interaction map of a cell. *Curr Opin Biotechnol*, **22**(1): 66–74.
- Courtney, K.D., Corcoran, R.B., and Engelman, J.A. (2010) The PI3K pathway as drug target in human cancer. *J Clin Oncol*, **28**(6): 1075–1083.
- Creighton, C.J., Morgan, M., Gunaratne, P.H., Wheeler, D.A., Gibbs, R.A., Robertson, A., Chu, A., Beroukhim, R., Cibulskis, K., Signoretti, S., et al. (2013) Comprehensive molecular characterization of clear cell renal cell carcinoma. Nature, 499(7456): 43–49.
- Croft, D., Mundo, A.F., Haw, R., Milacic, M., Weiser, J., Wu, G., Caudy, M., Garapati, P., Gillespie, M., Kamdar, M.R., et al. (2014) The Reactome pathway knowledge-base. *Nucleic Acids Res*, **42**(database issue): D472D477.
- Crunkhorn, S. (2014) Cancer: Predicting synthetic lethal interactions. *Nat Rev Drug Discov*, **13**(11): 812.
- Csardi, G. and Nepusz, T. (2006) The igraph software package for complex network research. *InterJournal*, Complex Systems: 1695.
- Dai, X., Li, T., Bai, Z., Yang, Y., Liu, X., Zhan, J., and Shi, B. (2015) Breast cancer intrinsic subtype classification, clinical use and future trends. *Am J Cancer Res*, **5**(10): 2929–2943.
- Davierwala, A.P., Haynes, J., Li, Z., Brost, R.L., Robinson, M.D., Yu, L., Mnaimneh, S., Ding, H., Zhu, H., Chen, Y., et al. (2005) The synthetic genetic interaction spectrum of essential genes. Nat Genet, 37(10): 1147–1152.

- De Leeuw, W.J., Berx, G., Vos, C.B., Peterse, J.L., Van de Vijver, M.J., Litvinov, S., Van Roy, F., Cornelisse, C.J., and Cleton-Jansen, A.M. (1997) Simultaneous loss of E-cadherin and catenins in invasive lobular breast cancer and lobular carcinoma in situ. *J Pathol*, **183**(4): 404–11.
- De Santis, G., Miotti, S., Mazzi, M., Canevari, S., and Tomassetti, A. (2009) E-cadherin directly contributes to PI3K/AKT activation by engaging the PI3K-p85 regulatory subunit to adherens junctions of ovarian carcinoma cells. *Oncogene*, **28**(9): 1206–1217.
- Demir, E., Babur, O., Rodchenkov, I., Aksoy, B.A., Fukuda, K.I., Gross, B., Sumer, O.S., Bader, G.D., and Sander, C. (2013) Using biological pathway data with Paxtools. *PLoS Comput Biol*, **9**(9): e1003194.
- Deshpande, R., Asiedu, M.K., Klebig, M., Sutor, S., Kuzmin, E., Nelson, J., Piotrowski, J., Shin, S.H., Yoshida, M., Costanzo, M., et al. (2013) A comparative genomic approach for identifying synthetic lethal interactions in human cancer. Cancer Res, 73(20): 6128–36.
- Dickson, D. (1999) Wellcome funds cancer database. *Nature*, **401**(6755): 729.
- Dijkstra, E.W. (1959) A note on two problems in connexion with graphs. *Numerische Mathematik*, **1**(1): 269–271.
- Dixon, S.J., Andrews, B.J., and Boone, C. (2009) Exploring the conservation of synthetic lethal genetic interaction networks. *Commun Integr Biol*, **2**(2): 78–81.
- Dixon, S.J., Fedyshyn, Y., Koh, J.L., Prasad, T.S., Chahwan, C., Chua, G., Toufighi, K., Baryshnikova, A., Hayles, J., Hoe, K.L., et al. (2008) Significant conservation of synthetic lethal genetic interaction networks between distantly related eukaryotes. Proc Natl Acad Sci U S A, 105(43): 16653–8.
- Dong, L.L., Liu, L., Ma, C.H., Li, J.S., Du, C., Xu, S., Han, L.H., Li, L., and Wang, X.W. (2012) E-cadherin promotes proliferation of human ovarian cancer cells in vitro via activating MEK/ERK pathway. Acta Pharmacol Sin, 33(6): 817–822.
- Dorsam, R.T. and Gutkind, J.S. (2007) G-protein-coupled receptors and cancer. *Nat Rev Cancer*, **7**(2): 79–94.
- Erdős, P. and Rényi, A. (1959) On random graphs I. Publ Math Debrecen, 6: 290–297.

- Erdős, P. and Rényi, A. (1960) On the evolution of random graphs. In *Publ. Math. Inst. Hung. Acad. Sci.*, volume 5, 17–61.
- Eroles, P., Bosch, A., Perez-Fidalgo, J.A., and Lluch, A. (2012) Molecular biology in breast cancer: intrinsic subtypes and signaling pathways. *Cancer Treat Rev*, **38**(6): 698–707.
- Farmer, H., McCabe, N., Lord, C.J., Tutt, A.N., Johnson, D.A., Richardson, T.B., Santarosa, M., Dillon, K.J., Hickson, I., Knights, C., et al. (2005) Targeting the dna repair defect in BRCA mutant cells as a therapeutic strategy. Nature, 434(7035): 917–21.
- Fawcett, T. (2006) An introduction to ROC analysis. *Pattern Recognition Letters*, **27**(8): 861 874. {ROC} Analysis in Pattern Recognition.
- Fece de la Cruz, F., Gapp, B.V., and Nijman, S.M. (2015) Synthetic lethal vulnerabilities of cancer. *Annu Rev Pharmacol Toxicol*, **55**: 513–531.
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D.M., Forman, D., and Bray, F. (2015) Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*, **136**(5): E359–386.
- Fisher, R.A. (1919) Xv.the correlation between relatives on the supposition of mendelian inheritance. *Earth and Environmental Science Transactions of the Royal Society of Edinburgh*, **52**(02): 399–433.
- Fong, P.C., Boss, D.S., Yap, T.A., Tutt, A., Wu, P., Mergui-Roelvink, M., Mortimer, P., Swaisland, H., Lau, A., O'Connor, M.J., et al. (2009) Inhibition of poly(adpribose) polymerase in tumors from BRCA mutation carriers. N Engl J Med, 361(2): 123–34.
- Fong, P.C., Yap, T.A., Boss, D.S., Carden, C.P., Mergui-Roelvink, M., Gourley, C., De Greve, J., Lubinski, J., Shanley, S., Messiou, C., et al. (2010) Poly(adp)-ribose polymerase inhibition: frequent durable responses in BRCA carrier ovarian cancer correlating with platinum-free interval. J Clin Oncol, 28(15): 2512–9.
- Forbes, S.A., Beare, D., Gunasekaran, P., Leung, K., Bindal, N., Boutselakis, H., Ding, M., Bamford, S., Cole, C., Ward, S., et al. (2015) COSMIC: exploring the world's

- knowledge of somatic mutations in human cancer. *Nucleic Acids Res*, **43**(Database issue): D805–811.
- Fraser, A. (2004) Towards full employment: using RNAi to find roles for the redundant. *Oncogene*, **23**(51): 8346–52.
- Fromental-Ramain, C., Warot, X., Lakkaraju, S., Favier, B., Haack, H., Birling, C., Dierich, A., Doll e, P., and Chambon, P. (1996) Specific and redundant functions of the paralogous Hoxa-9 and Hoxd-9 genes in forelimb and axial skeleton patterning. *Development*, **122**(2): 461–472.
- Futreal, P.A., Coin, L., Marshall, M., Down, T., Hubbard, T., Wooster, R., Rahman, N., and Stratton, M.R. (2004) A census of human cancer genes. *Nat Rev Cancer*, 4(3): 177–183.
- Futreal, P.A., Kasprzyk, A., Birney, E., Mullikin, J.C., Wooster, R., and Stratton, M.R. (2001) Cancer and genomics. *Nature*, **409**(6822): 850–852.
- Gao, B. and Roux, P.P. (2015) Translational control by oncogenic signaling pathways. Biochimica et Biophysica Acta, 1849(7): 753–65.
- Gatza, M.L., Kung, H.N., Blackwell, K.L., Dewhirst, M.W., Marks, J.R., and Chi, J.T. (2011) Analysis of tumor environmental response and oncogenic pathway activation identifies distinct basal and luminal features in HER2-related breast tumor subtypes. Breast Cancer Res, 13(3): R62.
- Gatza, M.L., Lucas, J.E., Barry, W.T., Kim, J.W., Wang, Q., Crawford, M.D., Datto, M.B., Kelley, M., Mathey-Prevot, B., Potti, A., et al. (2010) A pathway-based classification of human breast cancer. Proc Natl Acad Sci USA, 107(15): 6994–6999.
- Gatza, M.L., Silva, G.O., Parker, J.S., Fan, C., and Perou, C.M. (2014) An integrated genomics approach identifies drivers of proliferation in luminal-subtype human breast cancer. *Nat Genet*, **46**(10): 1051–1059.
- Gentleman, R.C., Carey, V.J., Bates, D.M., Bolstad, B., Dettling, M., Dudoit, S., Ellis, B., Gautier, L., Ge, Y., Gentry, J., et al. (2004) Bioconductor: open software development for computational biology and bioinformatics. Genome Biol, 5(10): R80.
- Genz, A. and Bretz, F. (2009) Computation of multivariate normal and t probabilities. In *Lecture Notes in Statistics*, volume 195. Springer-Verlag, Heidelberg.

- Genz, A., Bretz, F., Miwa, T., Mi, X., Leisch, F., Scheipl, F., and Hothorn, T. (2016) mvtnorm: Multivariate Normal and t Distributions. R package version 1.0-5. URL.
- Glaire, M.A., Brown, M., Church, D.N., and Tomlinson, I. (2017) Cancer predisposition syndromes: lessons for truly precision medicine. *J Pathol*, **241**(2): 226–235.
- Globus (Globus) (2017) Research data management simplified. https://www.globus.org/. Accessed: 25/03/2017.
- Goodwin, S., McPherson, J.D., and McCombie, W.R. (2016) Coming of age: ten years of next-generation sequencing technologies. *Nat Rev Genet*, **17**(6): 333–351.
- Grady, W.M., Willis, J., Guilford, P.J., Dunbier, A.K., Toro, T.T., Lynch, H., Wiesner, G., Ferguson, K., Eng, C., Park, J.G., et al. (2000) Methylation of the CDH1 promoter as the second genetic hit in hereditary diffuse gastric cancer. Nat Genet, 26(1): 16–17.
- Graziano, F., Humar, B., and Guilford, P. (2003) The role of the E-cadherin gene (*CDH1*) in diffuse gastric cancer susceptibility: from the laboratory to clinical practice. *Annals of Oncology*, **14**(12): 1705–1713.
- Guaragnella, N., Palermo, V., Galli, A., Moro, L., Mazzoni, C., and Giannattasio, S. (2014) The expanding role of yeast in cancer research and diagnosis: insights into the function of the oncosuppressors p53 and BRCA1/2. *FEMS Yeast Res*, **14**(1): 2–16.
- Güell, O., Sagus, F., and Serrano, M. (2014) Essential plasticity and redundancy of metabolism unveiled by synthetic lethality analysis. *PLoS Comput Biol*, **10**(5): e1003637.
- Guilford, P. (1999) E-cadherin downregulation in cancer: fuel on the fire? *Molecular Medicine Today*, **5**(4): 172 177.
- Guilford, P., Hopkins, J., Harraway, J., McLeod, M., McLeod, N., Harawira, P., Taite, H., Scoular, R., Miller, A., and Reeve, A.E. (1998) E-cadherin germline mutations in familial gastric cancer. *Nature*, 392(6674): 402–5.
- Guilford, P., Humar, B., and Blair, V. (2010) Hereditary diffuse gastric cancer: translation of *CDH1* germline mutations into clinical practice. *Gastric Cancer*, **13**(1): 1–10.

- Guilford, P.J., Hopkins, J.B., Grady, W.M., Markowitz, S.D., Willis, J., Lynch, H., Rajput, A., Wiesner, G.L., Lindor, N.M., Burgart, L.J., *et al.* (1999) E-cadherin germline mutations define an inherited cancer syndrome dominated by diffuse gastric cancer. *Hum Mutat*, **14**(3): 249–55.
- Guo, J., Liu, H., and Zheng, J. (2016) SynLethDB: synthetic lethality database toward discovery of selective and sensitive anticancer drug targets. *Nucleic Acids Res*, 44(D1): D1011–1017.
- Hajian-Tilaki, K. (2013) Receiver Operating Characteristic (ROC) Curve Analysis for Medical Diagnostic Test Evaluation. *Caspian J Intern Med*, 4(2): 627–635.
- Hall, M., Frank, E., Holmes, G., Pfahringer, B., Reutemann, P., and Witten, I.H. (2009) The weka data mining software: an update. *SIGKDD Explor Newsl*, **11**(1): 10–18.
- Hammerman, P.S., Lawrence, M.S., Voet, D., Jing, R., Cibulskis, K., Sivachenko, A., Stojanov, P., McKenna, A., Lander, E.S., Gabriel, S., et al. (2012) Comprehensive genomic characterization of squamous cell lung cancers. Nature, 489(7417): 519–525.
- Hanahan, D. and Weinberg, R.A. (2000) The hallmarks of cancer. Cell, 100(1): 57–70.
- Hanahan, D. and Weinberg, R.A. (2011) Hallmarks of cancer: the next generation. *Cell*, **144**(5): 646–674.
- Hanna, S. (2003) Cancer incidence in new zealand (2003-2007). In D. Forman, D. Bray
 F Brewster, C. Gombe Mbalawa, B. Kohler, M. Piñeros, E. Steliarova-Foucher,
 R. Swaminathan, and J. Ferlay (editors), Cancer Incidence in Five Continents,
 volume X, 902-907. International Agency for Research on Cancer, Lyon, France.
 Electronic version http://ci5.iarc.fr Accessed 22/03/2017.
- Hansford, S., Kaurah, P., Li-Chang, H., Woo, M., Senz, J., Pinheiro, H., Schrader, K.A., Schaeffer, D.F., Shumansky, K., Zogopoulos, G., et al. (2015) Hereditary Diffuse Gastric Cancer Syndrome: CDH1 Mutations and Beyond. JAMA Oncol, 1(1): 23–32.
- Heiskanen, M.A. and Aittokallio, T. (2012) Mining high-throughput screens for cancer drug targets-lessons from yeast chemical-genomic profiling and synthetic lethality.

- Wiley Interdisciplinary Reviews: Data Mining and Knowledge Discovery, 2(3): 263–272.
- Hell, P. (1976) Graphs with given neighbourhoods i. problémes combinatorics at theorie des graphes. *Proc Coil Int CNRS, Orsay,* **260**: 219–223.
- Higgins, M.E., Claremont, M., Major, J.E., Sander, C., and Lash, A.E. (2007) CancerGenes: a gene selection resource for cancer genome projects. *Nucleic Acids Res*, **35**(Database issue): D721–726.
- Hillenmeyer, M.E. (2008) The chemical genomic portrait of yeast: uncovering a phenotype for all genes. *Science*, **320**: 362–365.
- Hoadley, K.A., Yau, C., Wolf, D.M., Cherniack, A.D., Tamborero, D., Ng, S., Leiserson, M.D., Niu, B., McLellan, M.D., Uzunangelov, V., et al. (2014) Multiplatform analysis of 12 cancer types reveals molecular classification within and across tissues of origin. Cell, 158(4): 929–944.
- Hoehndorf, R., Hardy, N.W., Osumi-Sutherland, D., Tweedie, S., Schofield, P.N., and Gkoutos, G.V. (2013) Systematic analysis of experimental phenotype data reveals gene functions. *PLoS ONE*, **8**(4): e60847.
- Holm, S. (1979) A simple sequentially rejective multiple test procedure. *Scandinavian Journal of Statistics*, **6**(2): 65–70.
- Hopkins, A.L. (2008) Network pharmacology: the next paradigm in drug discovery. Nat Chem Biol, 4(11): 682–690.
- Hu, Z., Fan, C., Oh, D.S., Marron, J.S., He, X., Qaqish, B.F., Livasy, C., Carey, L.A., Reynolds, E., Dressler, L., et al. (2006) The molecular portraits of breast tumors are conserved across microarray platforms. BMC Genomics, 7: 96.
- Huang, E., Cheng, S., Dressman, H., Pittman, J., Tsou, M., Horng, C., Bild, A., Iversen, E., Liao, M., Chen, C., et al. (2003) Gene expression predictors of breast cancer outcomes. Lancet, 361: 1590–1596.
- Hutchison, C.A., Chuang, R.Y., Noskov, V.N., Assad-Garcia, N., Deerinck, T.J., Ellisman, M.H., Gill, J., Kannan, K., Karas, B.J., Ma, L., et al. (2016) Design and synthesis of a minimal bacterial genome. *Science*, **351**(6280): aad6253.

- International HapMap 3 Consortium (HapMap) (2003) The International HapMap Project. *Nature*, **426**(6968): 789–796.
- Jeanes, A., Gottardi, C.J., and Yap, A.S. (2008) Cadherins and cancer: how does cadherin dysfunction promote tumor progression? *Oncogene*, **27**(55): 6920–6929.
- Jerby-Arnon, L., Pfetzer, N., Waldman, Y., McGarry, L., James, D., Shanks, E., Seashore-Ludlow, B., Weinstock, A., Geiger, T., Clemons, P., et al. (2014) Predicting cancer-specific vulnerability via data-driven detection of synthetic lethality. Cell, 158(5): 1199–1209.
- Joachims, T. (1999) Making large-scale support vector machine learning practical. In S. Bernhard, lkopf, J.C.B. Christopher, and J.S. Alexander (editors), Advances in kernel methods, 169–184. MIT Press.
- Ju, Z., Liu, W., Roebuck, P.L., Siwak, D.R., Zhang, N., Lu, Y., Davies, M.A., Akbani, R., Weinstein, J.N., Mills, G.B., et al. (2015) Development of a robust classifier for quality control of reverse-phase protein arrays. Bioinformatics, 31(6): 912.
- Kaelin, Jr, W. (2005) The concept of synthetic lethality in the context of anticancer therapy. *Nat Rev Cancer*, **5**(9): 689–98.
- Kaelin, Jr, W. (2009) Synthetic lethality: a framework for the development of wiser cancer therapeutics. *Genome Med*, 1: 99.
- Kamada, T. and Kawai, S. (1989) An algorithm for drawing general undirected graphs. *Information Processing Letters*, **31**(1): 7–15.
- Kawai, J., Shinagawa, A., Shibata, K., Yoshino, M., Itoh, M., Ishii, Y., Arakawa, T., Hara, A., Fukunishi, Y., Konno, H., et al. (2001) Functional annotation of a full-length mouse cDNA collection. Nature, 409(6821): 685–690.
- Kelley, R. and Ideker, T. (2005) Systematic interpretation of genetic interactions using protein networks. *Nat Biotech*, **23**(5): 561–566.
- Kelly, S.T. (2013) Statistical Predictions of Synthetic Lethal Interactions in Cancer. Dissertation, University of Otago.
- Kelly, S.T., Single, A.B., Telford, B.J., Beetham, H.G., Godwin, T.D., Chen, A., Black, M.A., and Guilford, P.J. (unpublished) Towards HDGC chemoprevention:

- vulnerabilities in E-cadherin-negative cells identified by genome-wide interrogation of isogenic cell lines and whole tumors. Submitted to *Cancer Prev Res*.
- Keshava Prasad, T.S., Goel, R., Kandasamy, K., Keerthikumar, S., Kumar, S., Mathivanan, S., Telikicherla, D., Raju, R., Shafreen, B., Venugopal, A., et al. (2009) Human Protein Reference Database–2009 update. Nucleic Acids Res, 37(Database issue): D767–772.
- Kim, N.G., Koh, E., Chen, X., and Gumbiner, B.M. (2011) E-cadherin mediates contact inhibition of proliferation through Hippo signaling-pathway components. *Proc Natl Acad Sci USA*, **108**(29): 11930–11935.
- Koboldt, D.C., Fulton, R.S., McLellan, M.D., Schmidt, H., Kalicki-Veizer, J., McMichael, J.F., Fulton, L.L., Dooling, D.J., Ding, L., Mardis, E.R., et al. (2012) Comprehensive molecular portraits of human breast tumours. Nature, 490(7418): 61–70.
- Kockel, L., Zeitlinger, J., Staszewski, L.M., Mlodzik, M., and Bohmann, D. (1997) Jun in drosophila development: redundant and nonredundant functions and regulation by two maps signal transduction pathways. *Genes & Development*, **11**(13): 1748–1758.
- Kozlov, K.N., Gursky, V.V., Kulakovskiy, I.V., and Samsonova, M.G. (2015) Sequence-based model of gap gene regulation network. *BMC Genomics*, **15**(Suppl 12): S6.
- Kranthi, S., Rao, S., and Manimaran, P. (2013) Identification of synthetic lethal pairs in biological systems through network information centrality. *Mol BioSyst*, **9**(8): 2163–2167.
- Kroepil, F., Fluegen, G., Totikov, Z., Baldus, S.E., Vay, C., Schauer, M., Topp, S.A., Esch, J.S., Knoefel, W.T., and Stoecklein, N.H. (2012) Down-regulation of CDH1 is associated with expression of SNAI1 in colorectal adenomas. *PLoS ONE*, **7**(9): e46665.
- Lander, E.S. (2011) Initial impact of the sequencing of the human genome. *Nature*, **470**(7333): 187–197.
- Lander, E.S., Linton, L.M., Birren, B., Nusbaum, C., Zody, M.C., Baldwin, J., Devon, K., Dewar, K., Doyle, M., FitzHugh, W., et al. (2001) Initial sequencing and analysis of the human genome. Nature, 409(6822): 860–921.

- Langmead, B., Trapnell, C., Pop, M., and Salzberg, S.L. (2009) Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biol*, **10**(3): R25.
- Latora, V. and Marchiori, M. (2001) Efficient behavior of small-world networks. Phys Rev Lett, 87: 198701.
- Laufer, C., Fischer, B., Billmann, M., Huber, W., and Boutros, M. (2013) Mapping genetic interactions in human cancer cells with RNAi and multiparametric phenotyping. *Nat Methods*, **10**(5): 427–31.
- Law, C.W., Chen, Y., Shi, W., and Smyth, G.K. (2014) voom: precision weights unlock linear model analysis tools for RNA-seq read counts. *Genome Biol*, **15**(2): R29.
- Le Meur, N. and Gentleman, R. (2008) Modeling synthetic lethality. *Genome Biol*, **9**(9): R135.
- Le Meur, N., Jiang, Z., Liu, T., Mar, J., and Gentleman, R.C. (2014) Slgi: Synthetic lethal genetic interaction. r package version 1.26.0.
- Lee, A.Y., Perreault, R., Harel, S., Boulier, E.L., Suderman, M., Hallett, M., and Jenna, S. (2010a) Searching for signaling balance through the identification of genetic interactors of the rab guanine-nucleotide dissociation inhibitor gdi-1. *PLoS ONE*, **5**(5): e10624.
- Lee, I., Lehner, B., Vavouri, T., Shin, J., Fraser, A.G., and Marcotte, E.M. (2010b) Predicting genetic modifier loci using functional gene networks. *Genome Research*, **20**(8): 1143–1153.
- Lee, I. and Marcotte, E.M. (2009) Effects of functional bias on supervised learning of a gene network model. *Methods Mol Biol*, **541**: 463–75.
- Lee, M.J., Ye, A.S., Gardino, A.K., Heijink, A.M., Sorger, P.K., MacBeath, G., and Yaffe, M.B. (2012) Sequential application of anticancer drugs enhances cell death by rewiring apoptotic signaling networks. *Cell*, **149**(4): 780–94.
- Lehner, B., Crombie, C., Tischler, J., Fortunato, A., and Fraser, A.G. (2006) Systematic mapping of genetic interactions in caenorhabditis elegans identifies common modifiers of diverse signaling pathways. *Nat Genet*, **38**(8): 896–903.

- Li, B., Ruotti, V., Stewart, R.M., Thomson, J.A., and Dewey, C.N. (2010) RNA-Seq gene expression estimation with read mapping uncertainty. *Bioinformatics*, **26**(4): 493–500.
- Li, X.J., Mishra, S.K., Wu, M., Zhang, F., and Zheng, J. (2014) Syn-lethality: An integrative knowledge base of synthetic lethality towards discovery of selective anticancer therapies. *Biomed Res Int*, **2014**: 196034.
- Linehan, W.M., Spellman, P.T., Ricketts, C.J., Creighton, C.J., Fei, S.S., Davis, C., Wheeler, D.A., Murray, B.A., Schmidt, L., Vocke, C.D., et al. (2016) Comprehensive Molecular Characterization of Papillary Renal-Cell Carcinoma. N Engl J Med, 374(2): 135–145.
- Lokody, I. (2014) Computational modelling: A computational crystal ball. *Nature Reviews Cancer*, **14**(10): 649–649.
- Lord, C.J., Tutt, A.N., and Ashworth, A. (2015) Synthetic lethality and cancer therapy: lessons learned from the development of PARP inhibitors. *Annu Rev Med*, **66**: 455–470.
- Lu, X., Kensche, P.R., Huynen, M.A., and Notebaart, R.A. (2013) Genome evolution predicts genetic interactions in protein complexes and reveals cancer drug targets. *Nat Commun*, 4: 2124.
- Lu, X., Megchelenbrink, W., Notebaart, R.A., and Huynen, M.A. (2015) Predicting human genetic interactions from cancer genome evolution. *PLoS One*, **10**(5): e0125795.
- Lum, P.Y., Armour, C.D., Stepaniants, S.B., Cavet, G., Wolf, M.K., Butler, J.S., Hinshaw, J.C., Garnier, P., Prestwich, G.D., Leonardson, A., et al. (2004) Discovering modes of action for therapeutic compounds using a genome-wide screen of yeast heterozygotes. Cell, 116(1): 121–137.
- Luo, J., Solimini, N.L., and Elledge, S.J. (2009) Principles of Cancer Therapy: Oncogene and Non-oncogene Addiction. *Cell*, **136**(5): 823–837.
- Machado, J., Olivera, C., Carvalh, R., Soares, P., Berx, G., Caldas, C., Sercuca, R., Carneiro, F., and Sorbrinho-Simoes, M. (2001) E-cadherin gene (*CDH1*) promoter methylation as the second hit in sporadic diffuse gastric carcinoma. *Oncogene*, 20: 1525–1528.

- Markowetz, F. (2017) All biology is computational biology. *PLoS Biol*, **15**(3): e2002050.
- Masciari, S., Larsson, N., Senz, J., Boyd, N., Kaurah, P., Kandel, M.J., Harris, L.N., Pinheiro, H.C., Troussard, A., Miron, P., et al. (2007) Germline E-cadherin mutations in familial lobular breast cancer. J Med Genet, 44(11): 726–31.
- Mattison, J., van der Weyden, L., Hubbard, T., and Adams, D.J. (2009) Cancer gene discovery in mouse and man. *Biochim Biophys Acta*, **1796**(2): 140–161.
- McLachlan, J., George, A., and Banerjee, S. (2016) The current status of parp inhibitors in ovarian cancer. *Tumori*, **102**(5): 433–440.
- McLendon, R., Friedman, A., Bigner, D., Van Meir, E.G., Brat, D.J., Mastrogianakis, G.M., Olson, J.J., Mikkelsen, T., Lehman, N., Aldape, K., et al. (2008) Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature*, **455**(7216): 1061–1068.
- Miles, D.W. (2001) Update on HER-2 as a target for cancer therapy: herceptin in the clinical setting. *Breast Cancer Res*, **3**(6): 380–384.
- Muzny, D.M., Bainbridge, M.N., Chang, K., Dinh, H.H., Drummond, J.A., Fowler, G., Kovar, C.L., Lewis, L.R., Morgan, M.B., Newsham, I.F., et al. (2012) Comprehensive molecular characterization of human colon and rectal cancer. Nature, 487(7407): 330–337.
- Nagalla, S., Chou, J.W., Willingham, M.C., Ruiz, J., Vaughn, J.P., Dubey, P., Lash, T.L., Hamilton-Dutoit, S.J., Bergh, J., Sotiriou, C., et al. (2013) Interactions between immunity, proliferation and molecular subtype in breast cancer prognosis. Genome Biol, 14(4): R34.
- Neeley, E.S., Kornblau, S.M., Coombes, K.R., and Baggerly, K.A. (2009) Variable slope normalization of reverse phase protein arrays. *Bioinformatics*, **25**(11): 1384.
- Novomestky, F. (2012) matrixcalc: Collection of functions for matrix calculations. R package version 1.0-3.
- Nowak, M.A., Boerlijst, M.C., Cooke, J., and Smith, J.M. (1997) Evolution of genetic redundancy. *Nature*, **388**(6638): 167–171.

- Oliveira, C., Senz, J., Kaurah, P., Pinheiro, H., Sanges, R., Haegert, A., Corso, G., Schouten, J., Fitzgerald, R., Vogelsang, H., et al. (2009) Germline CDH1 deletions in hereditary diffuse gastric cancer families. Human Molecular Genetics, 18(9): 1545–1555.
- Oliveira, C., Seruca, R., Hoogerbrugge, N., Ligtenberg, M., and Carneiro, F. (2013) Clinical utility gene card for: Hereditary diffuse gastric cancer (HDGC). Eur J Hum Genet, 21(8).
- Pandey, G., Zhang, B., Chang, A.N., Myers, C.L., Zhu, J., Kumar, V., and Schadt, E.E. (2010) An integrative multi-network and multi-classifier approach to predict genetic interactions. *PLoS Comput Biol*, 6(9).
- Parker, J., Mullins, M., Cheung, M., Leung, S., Voduc, D., Vickery, T., Davies, S., Fauron, C., He, X., Hu, Z., et al. (2009) Supervised risk predictor of breast cancer based on intrinsic subtypes. *Journal of Clinical Oncology*, **27**(8): 1160–1167.
- Pereira, B., Chin, S.F., Rueda, O.M., Vollan, H.K., Provenzano, E., Bardwell, H.A., Pugh, M., Jones, L., Russell, R., Sammut, S.J., et al. (2016) Erratum: The somatic mutation profiles of 2,433 breast cancers refine their genomic and transcriptomic landscapes. Nat Commun, 7: 11908.
- Perou, C.M., Sørlie, T., Eisen, M.B., van de Rijn, M., Jeffrey, S.S., Rees, C.A., Pollack, J.R., Ross, D.T., Johnsen, H., Akslen, L.A., et al. (2000) Molecular portraits of human breast tumours. Nature, 406(6797): 747–752.
- Polyak, K. and Weinberg, R.A. (2009) Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits. *Nat Rev Cancer*, **9**(4): 265–73.
- R Core Team (2016) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. R version 3.3.2.
- Ritchie, M.E., Phipson, B., Wu, D., Hu, Y., Law, C.W., Shi, W., and Smyth, G.K. (2015) limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Research*, **43**(7): e47.
- Roguev, A., Bandyopadhyay, S., Zofall, M., Zhang, K., Fischer, T., Collins, S.R., Qu, H., Shales, M., Park, H.O., Hayles, J., et al. (2008) Conservation and rewiring of functional modules revealed by an epistasis map in fission yeast. Science, 322(5900): 405–10.

- Roychowdhury, S. and Chinnaiyan, A.M. (2016) Translating cancer genomes and transcriptomes for precision oncology. *CA Cancer J Clin*, **66**(1): 75–88.
- Rung, J. and Brazma, A. (2013) Reuse of public genome-wide gene expression data.

 Nat Rev Genet, 14(2): 89–99.
- Ryan, C., Lord, C., and Ashworth, A. (2014) Daisy: Picking synthetic lethals from cancer genomes. *Cancer Cell*, **26**(3): 306–308.
- Schena, M. (1996) Genome analysis with gene expression microarrays. *Bioessays*, **18**(5): 427–431.
- Scheuer, L., Kauff, N., Robson, M., Kelly, B., Barakat, R., Satagopan, J., Ellis, N., Hensley, M., Boyd, J., Borgen, P., et al. (2002) Outcome of preventive surgery and screening for breast and ovarian cancer in BRCA mutation carriers. *J Clin Oncol*, **20**(5): 1260–1268.
- Semb, H. and Christofori, G. (1998) The tumor-suppressor function of E-cadherin. *Am J Hum Genet*, **63**(6): 1588–93.
- Sing, T., Sander, O., Beerenwinkel, N., and Lengauer, T. (2005) Rocr: visualizing classifier performance in r. *Bioinformatics*, **21**(20): 7881.
- Slurm development team (Slurm) (2017) Slurm workload manager. https://slurm.schedmd.com/. Accessed: 25/03/2017.
- Sørlie, T., Perou, C.M., Tibshirani, R., Aas, T., Geisler, S., Johnsen, H., Hastie, T., Eisen, M.B., van de Rijn, M., Jeffrey, S.S., et al. (2001) Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proc Natl Acad Sci USA, 98(19): 10869–10874.
- Stajich, J.E. and Lapp, H. (2006) Open source tools and toolkits for bioinformatics: significance, and where are we? *Brief Bioinformatics*, **7**(3): 287–296.
- Stratton, M.R., Campbell, P.J., and Futreal, P.A. (2009) The cancer genome. *Nature*, **458**(7239): 719–724.
- Ström, C. and Helleday, T. (2012) Strategies for the use of poly(adenosine diphosphate ribose) polymerase (parp) inhibitors in cancer therapy. *Biomolecules*, **2**(4): 635–649.
- Tarazona, S., Garcia-Alcalde, F., Dopazo, J., Ferrer, A., and Conesa, A. (2011) Differential expression in RNA-seq: a matter of depth. *Genome Res*, **21**(12): 2213–2223.

- Telford, B.J., Chen, A., Beetham, H., Frick, J., Brew, T.P., Gould, C.M., Single, A., Godwin, T., Simpson, K.J., and Guilford, P. (2015) Synthetic lethal screens identify vulnerabilities in gpcr signalling and cytoskeletal organization in E-cadherin-deficient cells. *Mol Cancer Ther*, **14**(5): 1213–1223.
- The 1000 Genomes Project Consortium (1000 Genomes) (2010) A map of human genome variation from population-scale sequencing. *Nature*, **467**(7319): 1061–1073.
- The Cancer Genome Atlas Research Network (TCGA) (2017) The Cancer Genome Atlas Project. https://cancergenome.nih.gov/. Accessed: 26/03/2017.
- The Catalogue Of Somatic Mutations In Cancer (COSMIC) (2016) Cosmic: The catalogue of somatic mutations in cancer. http://cancer.sanger.ac.uk/cosmic. Release 79 (23/08/2016), Accessed: 05/02/2017.
- The Comprehensive R Archive Network (CRAN) (2017) Cran. https://cran.r-project.org/. Accessed: 24/03/2017.
- The ENCODE Project Consortium (ENCODE) (2004) The ENCODE (ENCyclopedia Of DNA Elements) Project. *Science*, **306**(5696): 636–640.
- The National Cancer Institute (NCI) (2015) The genetics of cancer. https://www.cancer.gov/about-cancer/causes-prevention/genetics. Published: 22/04/2015, Accessed: 22/03/2017.
- The New Zealand eScience Infrastructure (NeSI) (2017) NeSI. https://www.nesi.org.nz/. Accessed: 25/03/2017.
- Tierney, L., Rossini, A.J., Li, N., and Sevcikova, H. (2015) snow: Simple Network of Workstations. R package version 0.4-2.
- Tiong, K.L., Chang, K.C., Yeh, K.T., Liu, T.Y., Wu, J.H., Hsieh, P.H., Lin, S.H., Lai, W.Y., Hsu, Y.C., Chen, J.Y., et al. (2014) Csnk1e/ctnnb1 are synthetic lethal to tp53 in colorectal cancer and are markers for prognosis. Neoplasia, 16(5): 441–50.
- Tischler, J., Lehner, B., and Fraser, A.G. (2008) Evolutionary plasticity of genetic interaction networks. *Nat Genet*, **40**(4): 390–391.
- Tomasetti, C. and Vogelstein, B. (2015) Cancer etiology. Variation in cancer risk among tissues can be explained by the number of stem cell divisions. *Science*, **347**(6217): 78–81.

- Tong, A.H., Evangelista, M., Parsons, A.B., Xu, H., Bader, G.D., Page, N., Robinson, M., Raghibizadeh, S., Hogue, C.W., Bussey, H., et al. (2001) Systematic genetic analysis with ordered arrays of yeast deletion mutants. Science, 294(5550): 2364–8.
- Tong, A.H., Lesage, G., Bader, G.D., Ding, H., Xu, H., Xin, X., Young, J., Berriz, G.F., Brost, R.L., Chang, M., et al. (2004) Global mapping of the yeast genetic interaction network. Science, 303(5659): 808–13.
- Tran, B., Dancey, J.E., Kamel-Reid, S., McPherson, J.D., Bedard, P.L., Brown, A.M., Zhang, T., Shaw, P., Onetto, N., Stein, L., et al. (2012) Cancer genomics: technology, discovery, and translation. *J Clin Oncol*, **30**(6): 647–660.
- Travers, J. and Milgram, S. (1969) An experimental study of the small world problem. Sociometry, **32**(4): 425–443.
- Tunggal, J.A., Helfrich, I., Schmitz, A., Schwarz, H., Gunzel, D., Fromm, M., Kemler, R., Krieg, T., and Niessen, C.M. (2005) E-cadherin is essential for in vivo epidermal barrier function by regulating tight junctions. *EMBO J*, 24(6): 1146–1156.
- Tutt, A., Robson, M., Garber, J.E., Domchek, S.M., Audeh, M.W., Weitzel, J.N., Friedlander, M., Arun, B., Loman, N., Schmutzler, R.K., et al. (2010) Oral poly(adpribose) polymerase inhibitor olaparib in patients with *BRCA1* or *BRCA2* mutations and advanced breast cancer: a proof-of-concept trial. *Lancet*, **376**(9737): 235–44.
- University of California, Santa Cruz (UCSC) (2012) Ucsc cancer browser. Accessed 29/03/2012.
- van der Post, R.S., Vogelaar, I.P., Carneiro, F., Guilford, P., Huntsman, D., Hoogerbrugge, N., Caldas, C., Schreiber, K.E., Hardwick, R.H., Ausems, M.G., et al. (2015) Hereditary diffuse gastric cancer: updated clinical guidelines with an emphasis on germline CDH1 mutation carriers. J Med Genet, 52(6): 361–374.
- van Steen, K. (2012) Travelling the world of genegene interactions. *Briefings in Bioin*formatics, **13**(1): 1–19.
- van Steen, M. (2010) Graph Theory and Complex Networks: An Introduction. Maarten van Steen, VU Amsterdam.
- Vapnik, V.N. (1995) The nature of statistical learning theory. Springer-Verlag New York, Inc.

- Vizeacoumar, F.J., Arnold, R., Vizeacoumar, F.S., Chandrashekhar, M., Buzina, A., Young, J.T., Kwan, J.H., Sayad, A., Mero, P., Lawo, S., et al. (2013) A negative genetic interaction map in isogenic cancer cell lines reveals cancer cell vulnerabilities. Mol Syst Biol, 9: 696.
- Vogelstein, B., Papadopoulos, N., Velculescu, V.E., Zhou, S., Diaz, L.A., and Kinzler, K.W. (2013) Cancer genome landscapes. Science, 339(6127): 1546–1558.
- Vos, C.B., Cleton-Jansen, A.M., Berx, G., de Leeuw, W.J., ter Haar, N.T., van Roy, F., Cornelisse, C.J., Peterse, J.L., and van de Vijver, M.J. (1997) E-cadherin inactivation in lobular carcinoma in situ of the breast: an early event in tumorigenesis. Br J Cancer, 76(9): 1131–3.
- Waldron, D. (2016) Cancer genomics: A multi-layer omics approach to cancer. *Nat Rev Genet*, 17(8): 436–437.
- Wang, K., Singh, D., Zeng, Z., Coleman, S.J., Huang, Y., Savich, G.L., He, X., Mieczkowski, P., Grimm, S.A., Perou, C.M., et al. (2010) MapSplice: accurate mapping of RNA-seq reads for splice junction discovery. Nucleic Acids Res, 38(18): e178.
- Wang, X. and Simon, R. (2013) Identification of potential synthetic lethal genes to p53 using a computational biology approach. *BMC Medical Genomics*, **6**(1): 30.
- Wappett, M. (2014) Bisep: Toolkit to identify candidate synthetic lethality. r package version 2.0.
- Wappett, M., Dulak, A., Yang, Z.R., Al-Watban, A., Bradford, J.R., and Dry, J.R. (2016) Multi-omic measurement of mutually exclusive loss-of-function enriches for candidate synthetic lethal gene pairs. *BMC Genomics*, **17**: 65.
- Warnes, G.R., Bolker, B., Bonebakker, L., Gentleman, R., Liaw, W.H.A., Lumley, T., Maechler, M., Magnusson, A., Moeller, S., Schwartz, M., et al. (2015) gplots: Various R Programming Tools for Plotting Data. R package version 2.17.0.
- Watts, D.J. and Strogatz, S.H. (1998) Collective dynamics of 'small-world' networks. Nature, **393**(6684): 440–2.
- Weinstein, I.B. (2000) Disorders in cell circuitry during multistage carcinogenesis: the role of homeostasis. *Carcinogenesis*, **21**(5): 857–864.

- Weinstein, J.N., Akbani, R., Broom, B.M., Wang, W., Verhaak, R.G., McConkey, D., Lerner, S., Morgan, M., Creighton, C.J., Smith, C., et al. (2014) Comprehensive molecular characterization of urothelial bladder carcinoma. Nature, 507(7492): 315–322.
- Weinstein, J.N., Collisson, E.A., Mills, G.B., Shaw, K.R., Ozenberger, B.A., Ellrott, K., Shmulevich, I., Sander, C., Stuart, J.M., Chang, K., et al. (2013) The Cancer Genome Atlas Pan-Cancer analysis project. Nat Genet, 45(10): 1113–1120.
- Wickham, H. and Chang, W. (2016) devtools: Tools to Make Developing R Packages Easier. R package version 1.12.0.
- Wickham, H., Danenberg, P., and Eugster, M. (2017) roxygen2: In-Line Documentation for R. R package version 6.0.1.
- Wong, S.L., Zhang, L.V., Tong, A.H.Y., Li, Z., Goldberg, D.S., King, O.D., Lesage, G., Vidal, M., Andrews, B., Bussey, H., et al. (2004) Combining biological networks to predict genetic interactions. Proceedings of the National Academy of Sciences of the United States of America, 101(44): 15682–15687.
- World Health Organization (WHO) (2017) Fact sheet: Cancer. http://www.who.int/mediacentre/factsheets/fs297/en/. Updated February 2017, Accessed: 22/03/2017.
- Wu, M., Li, X., Zhang, F., Li, X., Kwoh, C.K., and Zheng, J. (2014) In silico prediction of synthetic lethality by meta-analysis of genetic interactions, functions, and pathways in yeast and human cancer. *Cancer Inform*, **13**(Suppl 3): 71–80.
- Yu, H. (2002) Rmpi: Parallel statistical computing in r. R News, 2(2): 10–14.
- Zhang, F., Wu, M., Li, X.J., Li, X.L., Kwoh, C.K., and Zheng, J. (2015) Predicting essential genes and synthetic lethality via influence propagation in signaling pathways of cancer cell fates. *J Bioinform Comput Biol*, **13**(3): 1541002.
- Zhang, J., Baran, J., Cros, A., Guberman, J.M., Haider, S., Hsu, J., Liang, Y., Rivkin, E., Wang, J., Whitty, B., et al. (2011) International cancer genome consortium data portala one-stop shop for cancer genomics data. Database: The Journal of Biological Databases and Curation, 2011: bar026.
- Zhong, W. and Sternberg, P.W. (2006) Genome-wide prediction of c. elegans genetic interactions. *Science*, **311**(5766): 1481–1484.

Zweig, M.H. and Campbell, G. (1993) Receiver-operating characteristic (roc) plots: a fundamental evaluation tool in clinical medicine. *Clinical Chemistry*, **39**(4): 561–577.