

## Library Declaration Form



University of Otago Library

Author's full name and year of birth: Simon Thomas Kelly,  
(for cataloguing purposes) 24 February 1992

Title of thesis: A Bioinformatics Approach to Synthetic Lethal Interactions in Breast Cancer with Gene Expression Data

Degree: Doctor of Philosophy

Department: Department of Biochemistry

Permanent Address: 710 Cumberland Street, Dunedin, NZ

I agree that this thesis may be consulted for research and study purposes and that reasonable quotation may be made from it, provided that proper acknowledgement of its use is made.

I consent to this thesis being copied in part or in whole for

- i) a library
- ii) an individual

at the discretion of the University of Otago.

Signature:

Date:

A Bioinformatics Approach to  
Synthetic Lethal Interactions in  
Breast Cancer with Gene  
Expression Data

S. Thomas Kelly

a thesis submitted for the degree of  
Doctor of Philosophy  
at the University of Otago, Dunedin,  
New Zealand.

22 June 2017

## Abstract

### Background

Synthetic lethal genetic interactions are re-emerging in the post-genomics era due to their potential for use in precision medicine against cancers. Synthetic lethal drug design exploits the functional redundancy of genes disrupted in cancers (including tumour suppressors) to develop specific treatments against them. *CDH1*, which encodes [E-cadherin](#), is a tumour suppressor gene with loss of function in breast and stomach cancers. Experimental screens have identified candidate synthetic lethal interactions for drug target triage, which can be further supported with bioinformatics analysis. Furthermore, gene expression data enables investigation of synthetic lethal pathways and graph structure of synthetic lethal genes within them.

### Approach

A computational methodology, the Synthetic Lethal Prediction Tool ([SLIPT](#)) has been developed to detect synthetic lethal interactions in gene expression data. This methodology was demonstrated on interactions with *CDH1* in breast and stomach cancer data from The Cancer Genome Atlas ([TCGA](#)) project. Synthetic lethal genes and pathways were further investigated with unsupervised clustering, gene set over-representation analysis, metagenes, and permutation resampling. In particular, analyses focused on comparing [SLIPT](#) gene candidates to an experimental [siRNA](#) screen [Telford \*et al.\* \(2015\)](#). Network analysis methods were applied to the most supported pathways to test for pathway structure among between synthetic lethal candidates. Simulation and modelling was used to assess the statistical performance of [SLIPT](#), including simulated data with correlation structures derived from graph structures.

## Findings

Many candidate synthetic lethal partners of *CDH1* were detected in both [TCGA](#) breast cancer. These genes clustered into several distinct groups, with distinct biological functions and elevated expression in different clinical subtypes. While the number of genes detected by both approaches was not significant, these contained significantly enriched pathways. In particular,  $G_{\alpha i}$  signalling, cytoplasmic microfibres, and extracellular fibrin clotting were robustly supported by both approaches, which is consistent with the known cytoskeletal and cell signalling roles of [E-cadherin](#) and validation of [GPCR](#) pathways performed by [Telford \*et al.\* \(2015\)](#). Many of these pathways were replicated in stomach cancer data. The pathways supported only by [SLIPT](#) included regulation of immune signalling and translational elongation which were not expected to be detected in an isogenic cell line model but are still candidates for further investigation.

Synthetic lethal candidates detected by [SLIPT](#) and [siRNA](#) were compared within graph structures of the candidate synthetic lethal pathways. These genes did not differ with respect to network metrics of importance or connectivity in the pathway. There was little support, across pathways, that [SLIPT](#) gene candidates were consistently upstream or downstream of [siRNA](#) gene candidates within pathways.

A model of synthetic lethality was used to simulate gene expression data with synthetic lethal partners of a gene. The [SLIPT](#) methodology had high statistical performance, detecting few synthetic lethal partners, which diminished with more synthetic lethal partners or lower sample size. The [SLIPT](#) methodology performed better than Pearson correlation or the  $\chi^2$ -test. In particular, it performed well with high specificity for datasets containing thousands of genes or genes positively correlated with the query gene (as expected to occur in expression data). [SLIPT](#) was robust across correlation structures, including those derived from complex pathway structures and often distinguished synthetic lethal genes from those positively or negatively correlated with them. Therefore [SLIPT](#) is appropriate to identify synthetic lethal genes within pathways and use candidate synthetic lethal genes (and their correlates) to identify synthetic lethal pathways.

## Summary

Thus this thesis has developed, evaluated, and refined a bioinformatics approach to discovery of synthetic lethal genes solely from gene expression data. This approach has been demonstrated to detect biologically informative and clinically relevant candidate partners for *CDH1* in breast and stomach cancers. These investigations have also involved the development of network analysis and simulation procedures which may be more widely applicable.

## Acknowledgements

I thank my supervisors A/Prof. Mik Black and Prof. Parry Guilford for their support and guidance throughout this my postgraduate studies. It has been a great experience, I look forward to seeing what your research groups produce in the future, may this not be the end for us.

I am also thankful for the guidance and mentorship of Prof. Hamish Spencer for career advice throughout my studies and time in his research group.

I am also grateful to the past and current members of these research groups, and my peers at the laboratory benches and computers across campus. The peer support, camaraderie, and guidance of newer students has been an incredible part of my time at Otago and has made my thesis studies not just easier but possible at all. The postgraduate community is very special here and I have truly made some lifelong friends from all over the world. You are talented researchers and amazing people. May we meet again some day. Where-ever you may end up, its small world and there's always time to catch up. I'd be delighted to host some visits while working abroad.

I cannot thank my friends, flatmates, family, and diligent proofreaders enough for their patience and support during such as massive, challenging, and (I'm sure you've heard too often) stressful undertaking during both my PhD and the study leading up to it. There are too many of you to name everyone here without leaving someone out, so thank you all for everything you've done, both the good times and the tough. Thank you for at least pretending to understand complex math oft brought up at the wrong moment. Thank you for checking my writing or slides, even when sprung on you last minute. Thank for your time when what I really needed was a chat, a walk, a drink with "the guys", or a moment to think clearly.

I thank the various organisations that supported this research project:

- This thesis was supported by the Postgraduate Tassell Scholarship in Cancer Research, a University of Otago Doctoral Scholarship.
- The New Zealand eScience Infrastructure (NeSI) provided access to the Intel Pan high-performance computing cluster, support, and training to use it effectively. Various aspects of this thesis would not have been possible without access to such an incredible national resource.
- The Health Research Council (HRC) of New Zealand provided funding for experimental research in the Cancer Genetics Laboratory. Some aspects of this project would not have been possible without access to the data and findings funded by this grant.
- The Allan Wilson Centre and Otago School of Biomedical Sciences provided funding for summer research placements which was a valuable opportunity to gain experience and training used in this thesis project.

I thank the following organisations for support towards presenting findings in this thesis at conference and seminars:

- Google (eResearch 2014, Hamilton)
- NeSI (Software Carpentry training and Research Bazaar 2015, Melbourne)
- REANNZ, NZGL, and NeSI (eResearch 2016, Queenstown)
- Otago Division of Health Sciences, Oxford Global, and Maurice and Phyllis Paykel Trust (NGS Asia 2016, Singapore)
- RIKEN Division of Genomic Technologies and the Okinawa Institute of Science and Technology (seminar visits in Japan)

Thanks most of all to my fiancé, Dr Yui Kawagishi, you've been an inspiration. Thank you for your support and encouragement, every day, even from afar: it has always made a difference. It's been incredible to see you flourish in your career and I look forward to joining you again soon. May the next chapter of our adventures involve a bit less Skype across timezones.

どうもありがとう由ちゃん。頑張った!もうすぐ行きます。!また来月!

# Contents

<b>Glossary</b>	<b>xix</b>
<b>Acronyms</b>	<b>xxvi</b>
<b>1 Introduction and Literature Review</b>	<b>1</b>
1.1 Cancer Research in the Post-Genomic Era . . . . .	1
1.1.1 Cancer is a Global Health Issue . . . . .	2
1.1.1.1 The Genetics and Molecular Biology of Cancers . . . . .	3
1.1.2 The Genomics Revolution in Cancer Research . . . . .	3
1.1.2.1 High-Throughput Technologies . . . . .	4
1.1.2.2 Bioinformatics and Genomic Data . . . . .	5
1.1.3 Genomics Projects . . . . .	5
1.1.3.1 The Cancer Genome Project . . . . .	6
1.1.3.2 The Cancer Genome Atlas Project . . . . .	6
1.1.4 Genomic Cancer Medicine . . . . .	8
1.1.4.1 Cancer Genes and Driver Mutations . . . . .	8
1.1.4.2 Precision Cancer Medicine . . . . .	9
1.1.4.3 Molecular Diagnostics and Pan-Cancer Medicine . . . . .	9
1.1.4.4 Targeted Therapeutics and Pharmacogenomics . . . . .	10
1.1.5 Systems and Network Biology . . . . .	11
1.2 Synthetic Lethal Cancer Medicine . . . . .	12
1.2.1 Synthetic Lethal Genetic Interactions . . . . .	13
1.2.2 Synthetic Lethal Concepts in Genetics . . . . .	13
1.2.3 Synthetic Lethality in Model Systems . . . . .	15
1.2.3.1 Synthetic Lethal Pathways and Networks . . . . .	15
1.2.3.2 Evolution of Synthetic Lethality . . . . .	16
1.2.4 Synthetic Lethality in Cancer . . . . .	17
1.2.5 Clinical Impact of Synthetic Lethality in Cancer . . . . .	18
1.2.6 High-throughput Screening for Synthetic Lethality . . . . .	20
1.2.6.1 Synthetic Lethal Screens . . . . .	21
1.2.7 Computational Prediction of Synthetic Lethality . . . . .	22
1.2.7.1 Bioinformatics Approaches to Genetic Interactions . . . . .	22
1.2.7.2 Comparative Genomics . . . . .	23
1.2.7.3 Analysis and Modelling of Protein Data . . . . .	26
1.2.7.4 Differential Gene Expression . . . . .	28
1.2.7.5 Data Mining and Machine Learning . . . . .	29



1.2.7.6	Mutual Exclusivity and Bimodality . . . . .	32
1.2.7.7	Rationale for Further Development . . . . .	33
1.3	E-cadherin as a Synthetic Lethal Target . . . . .	33
1.3.1	The <i>CDH1</i> gene and its Biological Functions . . . . .	34
1.3.1.1	Cytoskeleton . . . . .	34
1.3.1.2	Extracellular and Tumour Micro-environment . . . . .	34
1.3.1.3	Cell-Cell Adhesion and Signalling . . . . .	35
1.3.2	<i>CDH1</i> as a Tumour (and Invasion) Suppressor . . . . .	35
1.3.2.1	Breast Cancers and Invasion . . . . .	35
1.3.3	Hereditary Diffuse Gastric (and Lobular Breast) Cancer . . . . .	36
1.3.4	Cell Line Models of <i>CDH1</i> Null Mutations . . . . .	37
1.4	Summary and Research Direction of Thesis . . . . .	38
1.4.1	Thesis Aims . . . . .	39
<b>2</b>	<b>Methods and Resources</b>	<b>41</b>
2.1	Bioinformatics Resources for Genomics Research . . . . .	41
2.1.1	Public Data and Software Packages . . . . .	41
2.1.1.1	Cancer Genome Atlas Data . . . . .	42
2.1.1.2	Reactome and Annotation Data . . . . .	43
2.2	Data Handling . . . . .	43
2.2.1	Normalisation . . . . .	43
2.2.2	Sample Triage . . . . .	44
2.2.3	Metagenes and the Singular Value Decomposition . . . . .	44
2.2.4	Candidate Triage and Integration with Screen Data . . . . .	46
2.3	Techniques . . . . .	47
2.3.1	Statistical Procedures and Tests . . . . .	47
2.3.2	Gene Set Over-representation Analysis . . . . .	48
2.3.3	Clustering . . . . .	48
2.3.4	Heatmap . . . . .	48
2.3.5	Modelling and Simulations . . . . .	49
2.3.5.1	Receiver Operating Characteristic Curves . . . . .	50
2.3.6	Resampling Analysis . . . . .	50
2.4	Pathway Structure Methods . . . . .	51
2.4.1	Network and Graph Analysis . . . . .	51
2.4.2	Sourcing Graph Structure Data . . . . .	52
2.4.3	Constructing Pathway Subgraphs . . . . .	52
2.4.4	Network Analysis Metrics . . . . .	53
2.5	Implementation . . . . .	54
2.5.1	Computational Resources and Linux Utilities . . . . .	54
2.5.2	R Language and Packages . . . . .	55
2.5.3	High Performance and Parallel Computing . . . . .	58
<b>3</b>	<b>Methods Developed During Thesis</b>	<b>60</b>
3.1	A Synthetic Lethal Detection Methodology . . . . .	60
3.2	Synthetic Lethal Simulation and Modelling . . . . .	62
3.2.1	A Model of Synthetic Lethality in Expression Data . . . . .	63

3.2.2	Simulation Procedure . . . . .	67
3.3	Detecting Simulated Synthetic Lethal Partners . . . . .	70
3.3.1	Binomial Simulation of Synthetic Lethality . . . . .	70
3.3.2	Multivariate Normal Simulation of Synthetic Lethality . . . . .	72
3.3.2.1	Multivariate Normal Simulation with Correlated Genes . . . . .	74
3.3.2.2	Specificity with Query-Correlated Pathways . . . . .	80
3.4	Graph Structure Methods . . . . .	84
3.4.1	Upstream and Downstream Gene Detection . . . . .	84
3.4.1.1	Permutation Analysis for Statistical Significance . . . . .	85
3.4.1.2	Hierarchy Based on Biological Context . . . . .	85
3.4.2	Simulating Gene Expression from Graph Structures . . . . .	86
3.5	Customised Functions and Packages Developed . . . . .	91
3.5.1	Synthetic Lethal Interaction Prediction Tool . . . . .	91
3.5.2	Data Visualisation . . . . .	93
3.5.3	Extensions to the iGraph Package . . . . .	93
3.5.3.1	Sampling Simulated Data from Graph Structures . . . . .	94
3.5.3.2	Plotting Directed Graph Structures . . . . .	94
3.5.3.3	Computing Information Centrality . . . . .	95
3.5.3.4	Testing Pathway Structure with Permutation Testing . . . . .	95
3.5.3.5	Metapackage to Install iGraph Functions . . . . .	96
<b>4</b>	<b>Synthetic Lethal Analysis of Gene Expression Data</b>	<b>97</b>
4.1	Synthetic Lethal Genes in Breast Cancer . . . . .	98
4.1.1	Synthetic Lethal Pathways in Breast Cancer . . . . .	99
4.1.2	Expression Profiles of Synthetic Lethal Partners . . . . .	101
4.1.2.1	Subgroup Pathway Analysis . . . . .	104
4.2	Comparing Synthetic Lethal Gene Candidates . . . . .	106
4.2.1	Primary siRNA Screen Candidates . . . . .	106
4.2.2	Comparison with Correlation . . . . .	106
4.2.3	Comparison with Primary Screen Viability . . . . .	109
4.2.4	Comparison with Secondary siRNA Screen Validation . . . . .	111
4.2.5	Comparison to Primary Screen at Pathway Level . . . . .	112
4.2.5.1	Resampling Genes for Pathway Enrichment . . . . .	114
4.2.6	Integrating Synthetic Lethal Pathways and Screens . . . . .	119
4.3	Synthetic Lethal Pathway Metagenes . . . . .	120
4.4	Replication in Stomach Cancer . . . . .	122
4.5	Discussion . . . . .	123
4.5.1	Strengths of the SLIPT Methodology . . . . .	123
4.5.2	Synthetic Lethal Pathways for E-cadherin . . . . .	124
4.5.3	Replication and Validation . . . . .	126
4.5.3.1	Integration with siRNA Screening . . . . .	126
4.5.3.2	Replication across Tissues . . . . .	127
4.6	Summary . . . . .	127

<b>5</b>	<b>Synthetic Lethal Pathway Structure</b>	<b>129</b>
5.1	Synthetic Lethal Genes in Reactome Pathways . . . . .	129
5.1.1	The PI3K/AKT Pathway . . . . .	130
5.1.2	The Extracellular Matrix . . . . .	132
5.1.3	G Protein Coupled Receptors . . . . .	135
5.1.4	Gene Regulation and Translation . . . . .	135
5.2	Network Analysis of Synthetic Lethal Genes . . . . .	136
5.2.1	Gene Connectivity and Vertex Degree . . . . .	137
5.2.2	Gene Importance and Centrality . . . . .	138
	5.2.2.1 Information Centrality . . . . .	138
	5.2.2.2 PageRank Centrality . . . . .	140
5.3	Upstream or Downstream Synthetic Lethality . . . . .	142
5.3.1	Measuring Structure of Candidates within PI3K . . . . .	142
5.3.2	Resampling for Synthetic Lethal Pathway Structure . . . . .	144
5.4	Discussion . . . . .	146
5.5	Summary . . . . .	148
<b>6</b>	<b>Simulation and Modelling of Synthetic Lethal Pathways</b>	<b>153</b>
6.1	Synthetic Lethal Detection Methods . . . . .	154
6.1.1	Performance of SLIPT and $\chi^2$ across Quantiles . . . . .	155
	6.1.1.1 Correlated Query Genes affects Specificity . . . . .	158
6.1.2	Alternative Synthetic Lethal Detection Strategies . . . . .	160
	6.1.2.1 Correlation for Synthetic Lethal Detection . . . . .	161
	6.1.2.2 Testing for Bimodality with BiSEp . . . . .	162
6.2	Simulations with Graph Structures . . . . .	163
6.2.1	Performance over Graph Structures . . . . .	164
	6.2.1.1 Simple Graph Structures . . . . .	164
	6.2.1.2 Constructed Graph Structures . . . . .	167
6.2.2	Performance with Inhibitions . . . . .	169
6.2.3	Synthetic Lethality across Graph Structures . . . . .	175
6.2.4	Performance within a Simulated Human Genome . . . . .	178
6.3	Simulations in More Complex Graph Structures . . . . .	183
6.3.1	Simulations over Pathway-based Graphs . . . . .	184
6.3.2	Pathway Structures in a Simulated Human Genome . . . . .	186
6.4	Discussion . . . . .	189
	6.4.1 Simulation Procedure . . . . .	189
	6.4.2 Comparing Methods with Simulated Data . . . . .	190
	6.4.3 Design and Performance of SLIPT . . . . .	191
	6.4.4 Simulations from Graph Structures . . . . .	193
6.5	Summary . . . . .	194
<b>7</b>	<b>Discussion</b>	<b>196</b>
7.1	Synthetic Lethality and <i>CDH1</i> Biology . . . . .	196
	7.1.1 Established Functions of <i>CDH1</i> . . . . .	197
	7.1.2 The Molecular Role of <i>CDH1</i> in Cancer . . . . .	197
7.2	Significance . . . . .	198

7.2.1	Synthetic Lethality in the Genomic Era . . . . .	198
7.2.2	Clinical Interventions based on Synthetic Lethality . . . . .	200
7.3	Future Directions . . . . .	201
7.4	Conclusions . . . . .	203
	<b>Bibliography</b>	<b>205</b>
<b>A</b>	<b>Sample Quality</b>	<b>229</b>
A.1	Sample Correlation . . . . .	229
A.2	Replicate Samples in TCGA Breast Cancer Data . . . . .	232
<b>B</b>	<b>Software Used for Thesis</b>	<b>236</b>
<b>C</b>	<b>Mutation Analysis in Breast Cancer</b>	<b>245</b>
C.1	Synthetic Lethal Genes and Pathways . . . . .	245
C.2	Synthetic Lethal Expression Profiles . . . . .	246
C.3	Comparison to Primary Screen . . . . .	249
C.3.1	Resampling Analysis . . . . .	251
C.4	Compare SLIPT genes . . . . .	253
<b>D</b>	<b>Metagene Analysis</b>	<b>255</b>
D.1	Pathway Signature Expression . . . . .	255
D.2	Somatic Mutation . . . . .	264
D.3	Synthetic Lethal Reactome Metagenes . . . . .	265
D.4	Expression of Somatic Mutations . . . . .	267
<b>E</b>	<b>Intrinsic Subtyping</b>	<b>270</b>
<b>F</b>	<b>Stomach Expression Analysis</b>	<b>272</b>
F.1	Synthetic Lethal Genes and Pathways . . . . .	272
F.2	Comparison to Primary Screen . . . . .	276
F.2.1	Resampling Analysis . . . . .	278
F.3	Metagene Analysis . . . . .	280
<b>G</b>	<b>Synthetic Lethal Genes in Pathways</b>	<b>281</b>
<b>H</b>	<b>Pathway Connectivity for Mutation SLIPT</b>	<b>289</b>
<b>I</b>	<b>Pathway Structure for Mutation SLIPT</b>	<b>293</b>
<b>J</b>	<b>Performance of SLIPT and <math>\chi^2</math></b>	<b>295</b>
J.1	Correlated Query Genes affects Specificity . . . . .	301
<b>K</b>	<b>Simulations on Graph Structures</b>	<b>307</b>
K.0.1	Simulations from Inhibiting Graph Structures . . . . .	308
K.1	Simulation across Graph Structures . . . . .	311
K.2	Simulations from Complex Graph Structures . . . . .	315
K.2.1	Simulations from Complex Inhibiting Graphs . . . . .	318

K.3 Simulations from Pathway Graph Structures . . . . .	324
---	-----

# List of Tables

1.1	Methods for predicting genetic interactions . . . . .	23
1.2	Methods for predicting synthetic lethality in cancer . . . . .	24
1.3	Methods used by Wu <i>et al.</i> (2014) . . . . .	25
2.1	Excluded samples by batch and clinical characteristics. . . . .	44
2.2	Computers used during thesis . . . . .	54
2.3	Linux utilities and applications used during thesis . . . . .	55
2.4	R installations used during thesis . . . . .	56
2.5	R Packages used during thesis . . . . .	56
2.6	R packages developed during thesis . . . . .	58
4.1	Candidate synthetic lethal gene partners of <i>CDH1</i> from SLIPT . . . . .	99
4.2	Pathways for <i>CDH1</i> partners from SLIPT . . . . .	100
4.3	Pathways for clusters of <i>CDH1</i> partners from SLIPT . . . . .	105
4.4	ANOVA for synthetic lethality and correlation with <i>CDH1</i> . . . . .	108
4.5	Comparison of Synthetic Lethal Interaction Prediction Tool (SLIPT) genes against secondary short interfering RNA (siRNA) screen . . . . .	112
4.6	Pathways for <i>CDH1</i> partners from SLIPT and siRNA . . . . .	113
4.7	Pathways for <i>CDH1</i> partners from SLIPT . . . . .	116
4.8	Pathways for <i>CDH1</i> partners from SLIPT and siRNA primary screen .	117
4.9	Examples of candidate metagenes synthetic lethal for <i>CDH1</i> from SLIPT	121
5.1	ANOVA for synthetic lethality and vertex degree . . . . .	138
5.2	ANOVA for synthetic lethality and information centrality . . . . .	140
5.3	ANOVA for synthetic lethality and PageRank centrality . . . . .	142
5.4	Resampling for pathway structure of synthetic lethal detection methods	145
B.1	Complete list of R packages used during this thesis . . . . .	236
C.1	Candidate synthetic lethal gene partners of <i>CDH1</i> from mtSLIPT . . .	245
C.2	Pathways for <i>CDH1</i> partners from mtSLIPT . . . . .	246
C.3	Pathways for clusters of <i>CDH1</i> partners from mtSLIPT . . . . .	248
C.4	Pathways for <i>CDH1</i> partners from mtSLIPT and siRNA . . . . .	250
C.5	Pathways for <i>CDH1</i> partners from mtSLIPT . . . . .	251
C.6	Pathways for <i>CDH1</i> partners from mtSLIPT and siRNA primary screen	252
D.1	Candidate synthetic lethal metagenes against <i>CDH1</i> from mtSLIPT . .	266

E.1	Comparison of intrinsic subtypes . . . . .	270
F.1	Synthetic lethal gene partners of <i>CDH1</i> from SLIPT in stomach cancer	272
F.2	Pathways for <i>CDH1</i> partners from SLIPT in stomach cancer . . . . .	273
F.3	Pathways for clusters of <i>CDH1</i> partners in stomach SLIPT . . . . .	275
F.4	Pathways for <i>CDH1</i> partners from SLIPT and siRNA . . . . .	277
F.5	Pathways for <i>CDH1</i> partners from SLIPT in stomach cancer . . . . .	278
F.6	Pathways for <i>CDH1</i> partners from SLIPT in stomach and siRNA . . .	279
F.7	Synthetic lethal metagenes against <i>CDH1</i> in stomach cancer . . . . .	280
H.1	ANOVA for synthetic lethality and vertex degree . . . . .	292
H.2	ANOVA for synthetic lethality and information centrality . . . . .	292
H.3	ANOVA for synthetic lethality and PageRank centrality . . . . .	292
I.1	Resampling for pathway structure of synthetic lethal detection methods	294

# List of Figures

1.1	Synthetic genetic interactions . . . . .	14
1.2	Synthetic lethality in cancer . . . . .	17
2.1	Read count density . . . . .	45
2.2	Read count sample mean . . . . .	45
3.1	Framework for synthetic lethal prediction . . . . .	61
3.2	Synthetic lethal prediction adapted for mutation . . . . .	62
3.3	A model of synthetic lethal gene expression . . . . .	64
3.4	Modelling synthetic lethal gene expression . . . . .	65
3.5	Synthetic lethality with multiple genes . . . . .	66
3.6	Simulating gene function . . . . .	68
3.7	Simulating synthetic lethal gene function . . . . .	68
3.8	Simulating synthetic lethal gene expression . . . . .	69
3.9	Performance of binomial simulations . . . . .	71
3.10	Comparison of statistical performance . . . . .	71
3.11	Performance of multivariate normal simulations . . . . .	73
3.12	Simulating expression with correlated gene blocks . . . . .	75
3.13	Simulating expression with correlated gene blocks . . . . .	76
3.14	Synthetic lethal prediction across simulations . . . . .	78
3.15	Performance with correlations . . . . .	79
3.16	Comparison of statistical performance with correlation structure . . . . .	80
3.17	Performance with query correlations . . . . .	81
3.18	Statistical evaluation of directional criteria . . . . .	82
3.19	Performance of directional criteria . . . . .	83
3.20	Simulated graph structures . . . . .	87
3.21	Simulating expression from a graph structure . . . . .	88
3.22	Simulating expression from graph structure with inhibitions . . . . .	89
3.23	Demonstration of violin plots with custom features . . . . .	92
3.24	Demonstration of annotated heatmap . . . . .	92
3.25	Simulating graph structures . . . . .	95
4.1	Synthetic lethal expression profiles of analysed samples . . . . .	102
4.2	Comparison of SLIPT with siRNA . . . . .	107
4.3	Comparison of SLIPT and siRNA genes with correlation . . . . .	107
4.4	Comparison of SLIPT and siRNA genes with correlation . . . . .	109
4.5	Comparison of SLIPT and siRNA genes with screen viability . . . . .	110



4.6	Comparison of SLIPT genes with siRNA screen viability . . . . .	110
4.7	Resampled intersection of SLIPT and siRNA candidate genes . . . . .	115
5.1	synthetic lethality in the PI3K cascade . . . . .	131
5.2	synthetic lethality in Elastic Fibre Formation . . . . .	133
5.3	Synthetic lethality in Fibrin Clot Formation . . . . .	134
5.4	Synthetic lethality and vertex degree . . . . .	137
5.5	Synthetic lethality and centrality . . . . .	140
5.6	Synthetic lethality and PageRank . . . . .	141
5.7	Structure of synthetic lethality resampling in PI3K . . . . .	143
6.1	Performance of $\chi^2$ and SLIPT across quantiles . . . . .	156
6.2	Performance of $\chi^2$ and SLIPT across quantiles with more genes . . . . .	157
6.3	Performance of $\chi^2$ and SLIPT across quantiles with query correlation . . . . .	158
6.4	Performance of $\chi^2$ and SLIPT across quantiles with query correlation and more genes . . . . .	159
6.5	Performance of negative correlation and SLIPT . . . . .	162
6.6	Simple graph structures . . . . .	165
6.7	Performance of simulations on a simple graph . . . . .	166
6.8	Performance of simulations is similar in simple graphs . . . . .	167
6.9	Performance of simulations on a pathway . . . . .	168
6.10	Performance of simulations on a simple graph with inhibition . . . . .	170
6.11	Performance is higher on a simple inhibiting graph . . . . .	172
6.12	Performance of simulations on a constructed graph with inhibition . . . . .	173
6.13	Performance is affected by inhibition in graphs . . . . .	174
6.14	Detection of synthetic lethality within a graph structure . . . . .	176
6.15	Performance of simulations including a simple graph . . . . .	180
6.16	Performance on a simple graph improves with more genes . . . . .	181
6.17	Performance on an inhibiting graph improves with more genes . . . . .	182
6.18	Performance of simulations on the PI3K cascade . . . . .	185
6.19	Performance of simulations including the PI3K cascade . . . . .	187
6.20	Performance on pathways improves with more genes . . . . .	188
A.1	Correlation profiles of removed samples . . . . .	230
A.2	Correlation analysis and sample removal . . . . .	231
A.3	Replicate excluded samples . . . . .	232
A.4	Replicate samples with all remaining . . . . .	233
A.5	Replicate samples with some excluded . . . . .	234
C.1	Synthetic lethal expression profiles of analysed samples . . . . .	247
C.2	Comparison of mtSLIPT to siRNA . . . . .	249
C.3	Compare mtSLIPT and siRNA genes with correlation . . . . .	253
C.4	Compare mtSLIPT and siRNA genes with correlation . . . . .	253
C.5	Compare mtSLIPT and siRNA genes with siRNA viability . . . . .	254
D.1	Pathway metagene expression profiles . . . . .	257
D.2	Expression profiles for constituent genes of PI3K . . . . .	259

D.3	Expression profiles for estrogen receptor related genes . . . . .	260
D.4	Pathway metagene expression profiles . . . . .	261
D.5	Expression profiles for p53 related genes . . . . .	262
D.6	Expression profiles for BRCA related genes . . . . .	263
D.7	Somatic mutation against the PI3K metagene . . . . .	264
D.8	Somatic mutation against PIK3CA metagene . . . . .	267
D.9	Somatic mutation against PI3K protein . . . . .	268
D.10	Somatic mutation against AKT protein . . . . .	269
F.1	Synthetic lethal expression profiles of stomach samples . . . . .	274
F.2	Comparison of SLIPT in stomach to siRNA . . . . .	276
G.1	Synthetic lethality in the PI3K/AKT pathway . . . . .	281
G.2	Synthetic lethality in the PI3K/AKT pathway in cancer . . . . .	282
G.3	Synthetic lethality in the Extracellular Matrix . . . . .	283
G.4	Synthetic lethality in the GPCRs . . . . .	284
G.5	Synthetic lethality in the GPCR Downstream . . . . .	285
G.6	Synthetic lethality in the Translation Elongation . . . . .	286
G.7	Synthetic lethality in the Nonsense-mediated Decay . . . . .	287
G.8	Synthetic lethality in the 3' UTR . . . . .	288
H.1	Synthetic lethality and vertex degree . . . . .	289
H.2	Synthetic lethality and centrality . . . . .	290
H.3	Synthetic lethality and PageRank . . . . .	291
I.1	Structure of synthetic lethality resampling . . . . .	293
J.1	Performance of $\chi^2$ and SLIPT across quantiles . . . . .	295
J.2	Performance of $\chi^2$ and SLIPT across quantiles . . . . .	297
J.3	Performance of $\chi^2$ and SLIPT across quantiles with more genes . . . . .	299
J.4	Performance of $\chi^2$ and SLIPT across quantiles with query correlation . . . . .	301
J.5	Performance of $\chi^2$ and SLIPT across quantiles with query correlation . . . . .	303
J.6	Performance of $\chi^2$ and SLIPT across quantiles with query correlation and more genes . . . . .	305
K.1	Performance of simulations on a simple graph . . . . .	307
K.2	Performance of simulations on an inhibiting graph . . . . .	308
K.3	Performance of simulations on a constructed graph with inhibition . . . . .	309
K.4	Performance of simulations on a constructed graph with inhibition . . . . .	310
K.5	Detection of synthetic lethality within a graph structure . . . . .	311
K.6	Detection of synthetic lethality within an inhibiting graph . . . . .	313
K.7	Detection of synthetic lethality within an inhibiting graph . . . . .	314
K.8	Performance of simulations on a branching graph . . . . .	315
K.9	Performance of simulations on a complex graph . . . . .	316
K.10	Performance of simulations on a large graph . . . . .	317
K.11	Performance of simulations on a branching graph with inhibition . . . . .	318
K.12	Performance of simulations on a branching graph with inhibition . . . . .	319

K.13 Performance of simulations on a complex graph with inhibition . . . . .	320
K.14 Performance of simulations on a complex graph with inhibition . . . . .	321
K.15 Performance of simulations on a large constructed graph with inhibition	322
K.16 Performance of simulations on a large constructed graph with inhibition	323
K.17 Performance of simulations on the $G_{\alpha i}$ signalling pathway . . . . .	324
K.18 Performance of simulations including the $G_{\alpha i}$ signalling pathway . . . . .	325

# Glossary

allele	A gene variant with a specific sequence and phenotype.
bioinformatics	Statistical or computational approaches to biological data or research tools.
bisulfite-Seq	Epigenomic data from sequencing bisulfite treated DNA.
CAGE-Seq	Transcriptome data from cap analysis of gene expression.
cancer	A class of diseases, formally “malignant neoplasm”, of abnormal cellular growth and spread to other organs.
cancer gene	A gene which is involved in the malignancy of some cancers, encompassing <a href="#">oncogenes</a> and <a href="#">tumour suppressors</a> , which have molecular aberrations in cancer or variants which predispose individuals to cancer.
centrality	A network metric which identifies important <a href="#">vertices</a> .
chemoprevention	The use of drugs to prevent early-stage cancers, generally applied to high-risk mutation carriers.
chemotherapy	The use of cytotoxic drugs to treat cancers, in combinations, generally applied to advanced stage cancers.
ChIP-Seq	Epigenome data from chromatin immunoprecipitation sequencing.
compound screen	A <a href="#">high-throughput screen</a> performed using a library of chemical compounds.
computational biology	Applying computational or mathematical modelling to understanding biological systems and relationships.

conditional essentiality	A gene becoming essential to viability under certain environmental conditions, including presence of compounds which inactivate other genes.
copy number	The number of copies of DNA, typically two copies for diploid organisms but subject to variation.
<i>de novo</i>	A bioinformatics sequence assembly conducted entirely from raw genomics data without a reference sequence.
diagnosis	The identification of disease by clinical, cellular, and molecular characteristics.
driver mutation	A <a href="#">mutation</a> which promotes cancer growth.
E-cadherin	Epithelial cadherin (calcium-dependent adhesion), a cell-adhesion protein encoded by <i>CDH1</i> .
edge or link	A relationship connecting a pair of elements of a graph structure or network, may be weighted or directional.
epigenome	An analysis of epigenetic modifications of all genes in the genome.
epistasis (biological)	The effects of a gene modifying or masking the phenotype of another gene.
epistasis (statistical)	A divergence of the observed double <a href="#">mutant</a> phenotype from that expected based on the respective phenotypes of single <a href="#">mutant</a> (Fisher, 1919).
essential	A gene which is required to be functional or expressed for a cell or organism to be viable, grow or develop.
exome	A sequencing approach designed to generate data enriched for coding genes within the genome.
familial	A trait recurrently occurring in families, not necessarily with a genetic cause.
functional redundancy	Genes which perform a common function, also known as genetic redundancy.

gene expression	A measure of the relative expression of each gene from the mRNA extracted from (pooled) cells.
genetic robustness	A system of biological pathways which (has evolved to) continue to function as a whole under various conditions, including the inactivation of various individual genes.
genome	All of the DNA sequence in the genome.
genomic	The use of data from all genes in the genome.
genomic medicine	The use of genomic information to tailor medicine treatment to the genetics of an individual.
germline mutation	A <a href="#">mutation</a> that occurred in germline cells and is passed between generation.
graph or network	A mathematical structure modelling or depicting the relationships between elements.
hallmark of cancer	An underlying characteristic of cancer as part of a rational approach devised by ( <a href="#">Hanahan and Weinberg, 2000</a> ).
hereditary	A trait or disease which has a genetic cause and is inherited from family members.
high-throughput screen	An experimental procedure to perform a large scale series of chemical, genetic, or pharmacological tests.
hub	A central or highly connected component of a network.
<i>in silico</i>	An investigation conducted using computations, typically simulations or analyses.
<i>in vitro</i>	An investigation conducted using a controlled experimental system to examine biomolecules.
<i>in vivo</i>	An investigation conducted using in the context of a biological cell or organism, including pre-clinical models and clinical trials.
induced essentiality	A gene becoming <a href="#">essential</a> to viability under certain conditions, including inactivation of a synthetic lethal partner.
information centrality	A network <a href="#">centrality</a> metric which uses the impact of removing a <a href="#">vertex or node</a> on connections in the network.

intrinsic subtype	Distinguishing cancer by molecular and genetic features.
MCF10A cell line	A non-tumorigenic epithelial cell line derived from breast tissue.
metabolome	All the metabolites and enzymes in the cell.
metagene	A consistent signal of expression for a collection of genes such as a biological pathway, derived from singular value decomposition.
metagenome	All of the genes and genomes in a community.
metastasis	A secondary growth of a tumour or spread of cancer to other organs.
methylation	A measure of the epigenetic regulation of DNA at <a href="#">CpG dinucleotide (CpG)</a> sites.
microarray	A high-throughput technique to measure presence or abundance of nucleic acid sequences from binding to probes.
microRNA	Short RNA molecules generally regarded to regulate gene expression by binding to mRNA.
molecular profile	A combination of genetic and biochemical measures which identifies characteristic traits of a tumour.
molecular subtype	A classification of cancers based on an identification using molecular properties.
mutant	A variant or dysfunctional phenotype arising from a <a href="#">mutation</a> in a gene.
mutation	A change in DNA sequence that disrupts gene function.
network biology	The application mathematical and computational approaches to networks in understanding biological relationships.
network medicine	The use of <a href="#">network biology</a> to understand, prevent, or treat diseases.
non-oncogene addiction	The dependence of a cancer cell on functioning non-mutant genes.
'omics	A combination of approaches to generating biological data with high-throughput procedures such as genomics, proteomics or metabolomics.

oncogene	A gene that potentially causes cancer, typically by over-expression or mutant gene variants.
oncogene addiction	The dependence of a cancer cell on a specific oncogenic pathway.
PageRank centrality	A network <a href="#">centrality</a> metric which uses eigenvectors with a scaling factor ( <a href="#">Brin and Page, 1998</a> ).
pan cancer	A focus on the molecular and genetic features across cancers in different tissues.
passenger mutation	A <a href="#">mutation</a> that occurs in cancers but does not affect the growth of cancers.
pathway	A series of biomolecules that produces a particular product or biological function.
pleiotropy	When a gene has multiple biological functions.
polypharmacology	The design of drugs to target multiple molecular targets or biological pathways.
precision medicine	The application of prevention and treatment measures to target diseases by molecular and genetic features.
prognosis	The estimation of disease progression and patient outcome.
proteome	All the proteins expressed from the genome.
proto-oncogene	The non-mutant variant or precursor to a <a href="#">mutant oncogene</a> .
recurrent mutation	The repeated occurrence of mutations in a particular gene across cancers.
RNAi screen	A <a href="#">high-throughput screen</a> performed using a <a href="#">RNA interference (RNAi)</a> .
RNA-Seq	The generation of transcriptome data from sequencing RNA.
Sanger sequencing	A dideoxy chain termination method for DNA sequencing (named after Fred Sanger).
scale-free	A property of a network which has a power law <a href="#">vertex degree</a> distribution, that is several highly connected <a href="#">hub</a> genes and many with very few connections.



shortest path	A path with the fewest possible <b>edges</b> which connects two particular <b>vertices</b> .
small world	A property of a network which is highly connected and has a low characteristic path length, derived from the mean <b>shortest path</b> length across all pairs of nodes.
somatic mutation	A <b>mutation</b> that occurs in somatic cells, during a patient's lifespan.
sporadic cancer	Cancers which do occur in patients with a family history or carry a high-risk genetic variant.
synergy	When multiple drugs have more effect than expected from the effect of each separately.
synthetic dosage lethal	A <b>synthetic genetic interaction</b> (SGI) analogous to <b>synthetic lethality</b> where where one gene is inactivated and the other over-expressed.
synthetic lethal	Genetic interactions where inactivation of multiple genes is inviable (or deleterious) which are viable if inactivated separately.
synthetic lethal screen	A <b>high-throughput screen</b> performed on isogenic cell lines to detect genes for which inhibition specifically deleterious to the null <b>mutant</b> genotype.
synthetic rescue	A <b>synthetic genetic interaction</b> when the combined <b>mutations</b> restores the <b>wild-type</b> the phenotype of one of the <b>mutations</b> .
synthetic sick	Genetic interactions where inactivation of multiple genes is deleterious which are viable if inactivated separately.
synthetic suppression	A <b>synthetic genetic interaction</b> when the combined <b>mutations</b> (partially) suppresses the <b>mutant</b> phenotype of one of the <b>mutations</b> .
targeted therapy	Cancer treatment that specifically acts against a molecular target, in contrast to standard chemotherapy.
transcriptome	All of the genes expressed in the genome.
treatment	Medical procedures for a disease to improve patient outcomes.
tumour	An abnormal lump of tissue or growth of cells, may be cancerous.

tumour suppressor	A gene potentially causes cancer, typically by disruption of functions which protect the cell from cancer.
vertex degree	A network metric of connectivity of <a href="#">vertices</a> which uses the number of edges connected to each <a href="#">vertex</a> or <a href="#">node</a> .
vertex or node	An element of a graph structure or network.
wild-type	A natural phenotype of a trait or the normally functional <a href="#">allele</a> which encodes it.

# Acronyms

1KGP	1000 genomes project.
ADP	Adenosine Diphosphate.
AMP	Adenosine Monophosphate.
AMPK	<a href="#">AMP</a> -activated Protein Kinase.
ANOVA	Analysis of Variance.
ATP	Adenosine Triphosphate.
AUROC	Area Under the Receiver Operating Characteristic (curve).
Bash	Bourne Again Shell.
BioPAX	Biological Pathway Exchange.
BiSep	Bimodal Subsetting Expression.
BMP	Bone Morphogenic Protein.
cAMP	Cyclic <a href="#">AMP</a> .
CCL	Cancer Cell Line Encyclopaedia.
cDNA	Complementary DNA (from mRNA).
CGP	Cancer Genome Project.
ChIP	Chromatin Immunoprecipitation.
ChIP-Seq	Chromatin Immunoprecipitation Sequencing.
CNV	Copy Number Variation.
COSMIC	Catalogue Of Somatic Mutations In Cancer.
CpG	5'-C-phosphate-G-3'.
CPM	Counts Per Million mapped reads.
CPU	Central Processing Unit.
CRAN	comprehensive R archive network.
CXCR	Chemokine Receptor.
DAISY	Data Mining Synthetic Lethal Identification Pipeline.
DDBJ	DNA Data Bank of Japan.
DNA	Deoxyribonucleic Acid.
EMBL	European Molecular Biology Laboratory.

EMT	Epithelial-Mesenchymal Transition.
ENA	The European Nucleotide Archive.
ENCODE	Encyclopaedia of DNA Elements.
ER	Estrogen Receptor.
exprSL	Synthetic Lethality (expression).
FANTOM	Functional Annotation Of Mammalian genome.
FDR	False Discovery Rate.
GEO	Gene Expression Omnibus.
GO	Gene Ontology.
GPCR	G Protein Coupled Receptor.
HDAC	Histone Deacetylase.
HDGC	Hereditary Diffuse Gastric Cancer.
HLRCC	Hereditary Leiomyomatosis and Renal Cell Carcinoma.
HPC	High Performance Computing.
ICGC	International Cancer Genome Consortium.
IHC	Immunohistochemistry.
InDel	Insertion or Deletion (in <a href="#">DNA</a> sequence).
JAK	Janus Kinase.
lncRNA	Long Non-Coding RNA.
METABRIC	Molecular Taxonomy of Breast Cancer International Consortium.
microRNA	Micro RNA.
mRNA	Messenger RNA.
MSI	Microsatellite Instability.
mtSL	synthetic Lethality (mutation).
mtSLIPT	Synthetic Lethal Interaction Prediction Tool (against mutation).
NCBI	National Center for Biotechnology Information (in the USA).
NCI	National Cancer Institute (in the USA).
NeSI	New Zealand eScience Infrastructure.

NGS	Next-Generation Sequencing.
NHGRI	National Human Genome Research Institute (in the USA).
NIG	National Institute of Genetics (in Japan).
NIH	National Institutes of Health (in the USA).
NMD	Nonsense-Mediated Decay.
PAM50	Prediction Analysis of Microarray 50.
PARP	Poly-ADP-Ribose Polymerase.
PCR	Polymerase Chain Reaction.
PDE	Phosphodiesterase.
PI3K	Phosphoinositide 3-kinase.
PIP <sub>2</sub>	Phosphatidylinositol-(4,5)-bisphosphate.
PIP <sub>3</sub>	Phosphatidylinositol-(3,4,5)-trisphosphate.
PPI	Protein-Protein Interaction.
PR	Progesterone Receptor.
qPCR	Quantitative (real-time) Polymerase Chain Reaction.
RFLP	Restriction Fragment Length Polymorphism.
RGS	G-protein Signalling.
RHO	Ras Homolog Family.
RMA	Robust Multiarray Averaging (normalisation.
RNA	Ribonucleic Acid.
RNAi	RNA Interference.
ROC	Receiver Operating Characteristic (curve).
RPKM	Reads Per Kilobase per Million mapped reads.
RPPA	Reverse Phase Protein Arrays.
RRBS	Reduced Representation Bisulfite Sequencing.
rRNA	Ribonucleic acid.
RSEM	RNA-Seq by Expectation Maximization (normalisation.
SGA	Synthetic Gene Array (technique).
SGI	Synthetic Genetic Interaction.
shRNA	Short Hairpin RNA.
siRNA	Short Interfering RNA.
SL	Synthetic Lethal.
SLIPT	Synthetic Lethal Interaction Prediction Tool.
Slurm	Simple Linux Utility for Resource Management.

SNP	Single Nucleotide Polymorphism.
SOCKS	Socket Secure.
SR	Synthetic Rescue (or viability).
SS	Synthetic Suppression.
SSL	Synthetic Sick.
TCGA	The Cancer Genome Atlas (genomics project).
TGF $\alpha$	Transforming Growth Factor $\alpha$ .
TMM	Trimmed Mean of M values (normalisation.
tRNA	Transfer RNA.
UCSC	University of California, Santa Cruz.
UTR	Untranslated Region (of mRNA).
WNT	Wingless-Related Integration Site.

# Bibliography

- Aarts, M., Bajrami, I., Herrera-Abreu, M.T., Elliott, R., Brough, R., Ashworth, A., Lord, C.J., and Turner, N.C. (2015) Functional genetic screen identifies increased sensitivity to weel inhibition in cells with defects in fanconi anemia and hr pathways. *Mol Cancer Ther*, **14**(4): 865–76.
- Abeshouse, A., Ahn, J., Akbani, R., Ally, A., Amin, S., Andry, C.D., Annala, M., Aprikian, A., Armenia, J., Arora, A., *et al.* (2015) The Molecular Taxonomy of Primary Prostate Cancer. *Cell*, **163**(4): 1011–1025.
- Adler, D. (2005) *vioplot: Violin plot*. R package version 0.2.
- Akbani, R., Akdemir, K.C., Aksoy, B.A., Albert, M., Ally, A., Amin, S.B., Arachchi, H., Arora, A., Auman, J.T., Ayala, B., *et al.* (2015) Genomic Classification of Cutaneous Melanoma. *Cell*, **161**(7): 1681–1696.
- Akobeng, A.K. (2007) Understanding diagnostic tests 3: receiver operating characteristic curves. *Acta Pædiatrica*, **96**(5): 644–647.
- American Cancer Society (2017) Genetics and cancer. <https://www.cancer.org/cancer/cancer-causes/genetics.html>. Accessed: 22/03/2017.
- Anjomshoaa, A., Lin, Y.H., Black, M.A., McCall, J.L., Humar, B., Song, S., Fukuzawa, R., Yoon, H.S., Holzmann, B., Friederichs, J., *et al.* (2008) Reduced expression of a gene proliferation signature is associated with enhanced malignancy in colon cancer. *Br J Cancer*, **99**(6): 966–973.
- Araki, H., Knapp, C., Tsai, P., and Print, C. (2012) GeneSetDB: A comprehensive meta-database, statistical and visualisation framework for gene set analysis. *FEBS Open Bio*, **2**: 76–82.
- Ashburner, M., Ball, C.A., Blake, J.A., Botstein, D., Butler, H., Cherry, J.M., Davis, A.P., Dolinski, K., Dwight, S.S., Eppig, J.T., *et al.* (2000) Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat Genet*, **25**(1): 25–29.
- Ashworth, A. (2008) A synthetic lethal therapeutic approach: poly(adp) ribose polymerase inhibitors for the treatment of cancers deficient in dna double-strand break repair. *J Clin Oncol*, **26**(22): 3785–90.
- Ashworth, A., Lord, C.J., and Reis-Filho, J.S. (2011) Genetic interactions in cancer progression and treatment. *Cell*, **145**(1): 30–38.

- Audeh, M.W., Carmichael, J., Penson, R.T., Friedlander, M., Powell, B., Bell-McGuinn, K.M., Scott, C., Weitzel, J.N., Oaknin, A., Loman, N., *et al.* (2010) Oral poly(adp-ribose) polymerase inhibitor olaparib in patients with *BRCA1* or *BRCA2* mutations and recurrent ovarian cancer: a proof-of-concept trial. *Lancet*, **376**(9737): 245–51.
- Babyak, M.A. (2004) What you see may not be what you get: a brief, nontechnical introduction to overfitting in regression-type models. *Psychosom Med*, **66**(3): 411–21.
- Bamford, S., Dawson, E., Forbes, S., Clements, J., Pettett, R., Dogan, A., Flanagan, A., Teague, J., Futreal, P.A., Stratton, M.R., *et al.* (2004) The COSMIC (Catalogue of Somatic Mutations in Cancer) database and website. *Br J Cancer*, **91**(2): 355–358.
- Barabási, A.L. and Albert, R. (1999) Emergence of scaling in random networks. *Science*, **286**(5439): 509–12.
- Barabási, A.L., Gulbahce, N., and Loscalzo, J. (2011) Network medicine: a network-based approach to human disease. *Nat Rev Genet*, **12**(1): 56–68.
- Barabási, A.L. and Oltvai, Z.N. (2004) Network biology: understanding the cell’s functional organization. *Nat Rev Genet*, **5**(2): 101–13.
- Barrat, A. and Weigt, M. (2000) On the properties of small-world network models. *The European Physical Journal B - Condensed Matter and Complex Systems*, **13**(3): 547–560.
- Barretina, J., Caponigro, G., Stransky, N., Venkatesan, K., Margolin, A.A., Kim, S., Wilson, C.J., Lehar, J., Kryukov, G.V., Sonkin, D., *et al.* (2012) The Cancer Cell Line Encyclopedia enables predictive modelling of anticancer drug sensitivity. *Nature*, **483**(7391): 603–607.
- Barry, W.T. (2016) *safe: Significance Analysis of Function and Expression*. R package version 3.14.0.
- Baryshnikova, A., Costanzo, M., Dixon, S., Vizeacoumar, F.J., Myers, C.L., Andrews, B., and Boone, C. (2010a) Synthetic genetic array (sga) analysis in *saccharomyces cerevisiae* and *schizosaccharomyces pombe*. *Methods Enzymol*, **470**: 145–79.
- Baryshnikova, A., Costanzo, M., Kim, Y., Ding, H., Koh, J., Toufighi, K., Youn, J.Y., Ou, J., San Luis, B.J., Bandyopadhyay, S., *et al.* (2010b) Quantitative analysis of fitness and genetic interactions in yeast on a genome scale. *Nat Meth*, **7**(12): 1017–1024.
- Bass, A.J., Thorsson, V., Shmulevich, I., Reynolds, S.M., Miller, M., Bernard, B., Hinoue, T., Laird, P.W., Curtis, C., Shen, H., *et al.* (2014) Comprehensive molecular characterization of gastric adenocarcinoma. *Nature*, **513**(7517): 202–209.



- Bates, D. and Maechler, M. (2016) *Matrix: Sparse and Dense Matrix Classes and Methods*. R package version 1.2-7.1.
- Bateson, W. and Mendel, G. (1909) *Mendel's principles of heredity, by W. Bateson*. University Press, Cambridge [Eng.].
- Becker, K.F., Atkinson, M.J., Reich, U., Becker, I., Nekarda, H., Siewert, J.R., and Höfler, H. (1994) E-cadherin gene mutations provide clues to diffuse type gastric carcinomas. *Cancer Research*, **54**(14): 3845–3852.
- Bell, D., Berchuck, A., Birrer, M., Chien, J., Cramer, D., Dao, F., Dhir, R., DiSaia, P., Gabra, H., Glenn, P., *et al.* (2011) Integrated genomic analyses of ovarian carcinoma. *Nature*, **474**(7353): 609–615.
- Benjamini, Y. and Hochberg, Y. (1995) Controlling the false discovery rate: A practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society Series B (Methodological)*, **57**(1): 289–300.
- Berx, G., Cleton-Jansen, A.M., Nollet, F., de Leeuw, W.J., van de Vijver, M., Cornelisse, C., and van Roy, F. (1995) E-cadherin is a tumour/invasion suppressor gene mutated in human lobular breast cancers. *EMBO J*, **14**(24): 6107–15.
- Berx, G., Cleton-Jansen, A.M., Strumane, K., de Leeuw, W.J., Nollet, F., van Roy, F., and Cornelisse, C. (1996) E-cadherin is inactivated in a majority of invasive human lobular breast cancers by truncation mutations throughout its extracellular domain. *Oncogene*, **13**(9): 1919–25.
- Berx, G. and van Roy, F. (2009) Involvement of members of the cadherin superfamily in cancer. *Cold Spring Harb Perspect Biol*, **1**: a003129.
- Bitler, B.G., Aird, K.M., Garipov, A., Li, H., Amatangelo, M., Kossenkov, A.V., Schultz, D.C., Liu, Q., Shih, Ie, M., Conejo-Garcia, J.R., *et al.* (2015) Synthetic lethality by targeting ezh2 methyltransferase activity in arid1a-mutated cancers. *Nat Med*, **21**(3): 231–8.
- Blake, J.A., Christie, K.R., Dolan, M.E., Drabkin, H.J., Hill, D.P., Ni, L., Sitnikov, D., Burgess, S., Buza, T., Gresham, C., *et al.* (2015) Gene Ontology Consortium: going forward. *Nucleic Acids Res*, **43**(Database issue): D1049–1056.
- Boettcher, M., Lawson, A., Ladenburger, V., Fredebohm, J., Wolf, J., Hoheisel, J.D., Frezza, C., and Shlomi, T. (2014) High throughput synthetic lethality screen reveals a tumorigenic role of adenylate cyclase in fumarate hydratase-deficient cancer cells. *BMC Genomics*, **15**: 158.
- Boone, C., Bussey, H., and Andrews, B.J. (2007) Exploring genetic interactions and networks with yeast. *Nat Rev Genet*, **8**(6): 437–49.
- Borgatti, S.P. (2005) Centrality and network flow. *Social Networks*, **27**(1): 55 – 71.

- Boucher, B. and Jenna, S. (2013) Genetic interaction networks: better understand to better predict. *Front Genet*, **4**: 290.
- Bozovic-Spasojevic, I., Azambuja, E., McCaskill-Stevens, W., Dinh, P., and Cardoso, F. (2012) Chemoprevention for breast cancer. *Cancer treatment reviews*, **38**(5): 329–339.
- Breiman, L. (2001) Random forests. *Machine Learning*, **45**(1): 5–32.
- Brin, S. and Page, L. (1998) The anatomy of a large-scale hypertextual web search engine. *Computer Networks and ISDN Systems*, **30**(1): 107 – 117.
- Brouxhon, S.M., Kyrkanides, S., Teng, X., Athar, M., Ghazizadeh, S., Simon, M., O'Banion, M.K., and Ma, L. (2014) Soluble E-cadherin: a critical oncogene modulating receptor tyrosine kinases, MAPK and PI3K/Akt/mTOR signaling. *Oncogene*, **33**(2): 225–235.
- Brückner, A., Polge, C., Lentze, N., Auerbach, D., and Schlattner, U. (2009) Yeast two-hybrid, a powerful tool for systems biology. *Int J Mol Sci*, **10**(6): 2763–2788.
- Bryant, H.E., Schultz, N., Thomas, H.D., Parker, K.M., Flower, D., Lopez, E., Kyle, S., Meuth, M., Curtin, N.J., and Helleday, T. (2005) Specific killing of *BRCA2*-deficient tumours with inhibitors of polyadprbose polymerase. *Nature*, **434**(7035): 913–7.
- Bussey, H., Andrews, B., and Boone, C. (2006) From worm genetic networks to complex human diseases. *Nat Genet*, **38**(8): 862–3.
- Butland, G., Babu, M., Diaz-Mejia, J.J., Bohdana, F., Phanse, S., Gold, B., Yang, W., Li, J., Gagarinova, A.G., Pogoutse, O., *et al.* (2008) esga: E. coli synthetic genetic array analysis. *Nat Methods*, **5**(9): 789–95.
- cBioPortal for Cancer Genomics (cBioPortal) (2017) cBioPortal for Cancer Genomics. <http://www.cbioportal.org/>. Accessed: 26/03/2017.
- Cerami, E.G., Gross, B.E., Demir, E., Rodchenkov, I., Babur, O., Anwar, N., Schultz, N., Bader, G.D., and Sander, C. (2011) Pathway Commons, a web resource for biological pathway data. *Nucleic Acids Res*, **39**(Database issue): D685–690.
- Chen, A., Beetham, H., Black, M.A., Priya, R., Telford, B.J., Guest, J., Wiggins, G.A.R., Godwin, T.D., Yap, A.S., and Guilford, P.J. (2014) E-cadherin loss alters cytoskeletal organization and adhesion in non-malignant breast cells but is insufficient to induce an epithelial-mesenchymal transition. *BMC Cancer*, **14**(1): 552.
- Chen, S. and Parmigiani, G. (2007) Meta-analysis of BRCA1 and BRCA2 penetrance. *J Clin Oncol*, **25**(11): 1329–1333.
- Chipman, K. and Singh, A. (2009) Predicting genetic interactions with random walks on biological networks. *BMC Bioinformatics*, **10**(1): 17.

- Christofori, G. and Semb, H. (1999) The role of the cell-adhesion molecule E-cadherin as a tumour-suppressor gene. *Trends in Biochemical Sciences*, **24**(2): 73 – 76.
- Ciriello, G., Gatz, M.L., Beck, A.H., Wilkerson, M.D., Rhie, S.K., Pastore, A., Zhang, H., McLellan, M., Yau, C., Kandoth, C., *et al.* (2015) Comprehensive Molecular Portraits of Invasive Lobular Breast Cancer. *Cell*, **163**(2): 506–519.
- Clark, M.J. (2004) Endogenous Regulator of G Protein Signaling Proteins Suppress G  $\alpha$ -Dependent  $\mu$ -Opioid Agonist-Mediated Adenylyl Cyclase Supersensitization. *Journal of Pharmacology and Experimental Therapeutics*, **310**(1): 215–222.
- Collingridge, D.S. (2013) A primer on quantitized data analysis and permutation testing. *Journal of Mixed Methods Research*, **7**(1): 81–97.
- Collins, F.S. and Barker, A.D. (2007) Mapping the cancer genome. Pinpointing the genes involved in cancer will help chart a new course across the complex landscape of human malignancies. *Sci Am*, **296**(3): 50–57.
- Collisson, E., Campbell, J., Brooks, A., Berger, A., Lee, W., Chmielecki, J., Beer, D., Cope, L., Creighton, C., Danilova, L., *et al.* (2014) Comprehensive molecular profiling of lung adenocarcinoma. *Nature*, **511**(7511): 543–550.
- Costanzo, M., Baryshnikova, A., Bellay, J., Kim, Y., Spear, E.D., Sevier, C.S., Ding, H., Koh, J.L., Toufighi, K., Mostafavi, S., *et al.* (2010) The genetic landscape of a cell. *Science*, **327**(5964): 425–31.
- Costanzo, M., Baryshnikova, A., Myers, C.L., Andrews, B., and Boone, C. (2011) Charting the genetic interaction map of a cell. *Curr Opin Biotechnol*, **22**(1): 66–74.
- Courtney, K.D., Corcoran, R.B., and Engelman, J.A. (2010) The PI3K pathway as drug target in human cancer. *J Clin Oncol*, **28**(6): 1075–1083.
- Creighton, C.J., Morgan, M., Gunaratne, P.H., Wheeler, D.A., Gibbs, R.A., Robertson, A., Chu, A., Beroukhi, R., Cibulskis, K., Signoretti, S., *et al.* (2013) Comprehensive molecular characterization of clear cell renal cell carcinoma. *Nature*, **499**(7456): 43–49.
- Croft, D., Mundo, A.F., Haw, R., Milacic, M., Weiser, J., Wu, G., Caudy, M., Garapati, P., Gillespie, M., Kamdar, M.R., *et al.* (2014) The Reactome pathway knowledge-base. *Nucleic Acids Res*, **42**(database issue): D472–D477.
- Crunkhorn, S. (2014) Cancer: Predicting synthetic lethal interactions. *Nat Rev Drug Discov*, **13**(11): 812.
- Csardi, G. and Nepusz, T. (2006) The igraph software package for complex network research. *InterJournal*, **Complex Systems**: 1695.
- Dai, X., Li, T., Bai, Z., Yang, Y., Liu, X., Zhan, J., and Shi, B. (2015) Breast cancer intrinsic subtype classification, clinical use and future trends. *Am J Cancer Res*, **5**(10): 2929–2943.

- Davierwala, A.P., Haynes, J., Li, Z., Brost, R.L., Robinson, M.D., Yu, L., Mnaimneh, S., Ding, H., Zhu, H., Chen, Y., *et al.* (2005) The synthetic genetic interaction spectrum of essential genes. *Nat Genet*, **37**(10): 1147–1152.
- De Leeuw, W.J., Berx, G., Vos, C.B., Peterse, J.L., Van de Vijver, M.J., Litvinov, S., Van Roy, F., Cornelisse, C.J., and Cleton-Jansen, A.M. (1997) Simultaneous loss of E-cadherin and catenins in invasive lobular breast cancer and lobular carcinoma in situ. *J Pathol*, **183**(4): 404–11.
- De Santis, G., Miotti, S., Mazzi, M., Canevari, S., and Tomassetti, A. (2009) E-cadherin directly contributes to PI3K/AKT activation by engaging the PI3K-p85 regulatory subunit to adherens junctions of ovarian carcinoma cells. *Oncogene*, **28**(9): 1206–1217.
- Demir, E., Babur, O., Rodchenkov, I., Aksoy, B.A., Fukuda, K.I., Gross, B., Sumer, O.S., Bader, G.D., and Sander, C. (2013) Using biological pathway data with Pax-tools. *PLoS Comput Biol*, **9**(9): e1003194.
- Deshpande, R., Asiedu, M.K., Klebig, M., Sutor, S., Kuzmin, E., Nelson, J., Piotrowski, J., Shin, S.H., Yoshida, M., Costanzo, M., *et al.* (2013) A comparative genomic approach for identifying synthetic lethal interactions in human cancer. *Cancer Res*, **73**(20): 6128–36.
- Dickson, D. (1999) Wellcome funds cancer database. *Nature*, **401**(6755): 729.
- Dijkstra, E.W. (1959) A note on two problems in connexion with graphs. *Numerische Mathematik*, **1**(1): 269–271.
- Dixon, S.J., Andrews, B.J., and Boone, C. (2009) Exploring the conservation of synthetic lethal genetic interaction networks. *Commun Integr Biol*, **2**(2): 78–81.
- Dixon, S.J., Fedyshyn, Y., Koh, J.L., Prasad, T.S., Chahwan, C., Chua, G., Toufighi, K., Baryshnikova, A., Hayles, J., Hoe, K.L., *et al.* (2008) Significant conservation of synthetic lethal genetic interaction networks between distantly related eukaryotes. *Proc Natl Acad Sci U S A*, **105**(43): 16653–8.
- Dong, L.L., Liu, L., Ma, C.H., Li, J.S., Du, C., Xu, S., Han, L.H., Li, L., and Wang, X.W. (2012) E-cadherin promotes proliferation of human ovarian cancer cells in vitro via activating MEK/ERK pathway. *Acta Pharmacol Sin*, **33**(6): 817–822.
- Dorsam, R.T. and Gutkind, J.S. (2007) G-protein-coupled receptors and cancer. *Nat Rev Cancer*, **7**(2): 79–94.
- Erdős, P. and Rényi, A. (1959) On random graphs I. *Publ Math Debrecen*, **6**: 290–297.
- Erdős, P. and Rényi, A. (1960) On the evolution of random graphs. In *Publ. Math. Inst. Hung. Acad. Sci.*, volume 5, 17–61.
- Eroles, P., Bosch, A., Perez-Fidalgo, J.A., and Lluch, A. (2012) Molecular biology in breast cancer: intrinsic subtypes and signaling pathways. *Cancer Treat Rev*, **38**(6): 698–707.

- Farmer, H., McCabe, N., Lord, C.J., Tutt, A.N., Johnson, D.A., Richardson, T.B., Santarosa, M., Dillon, K.J., Hickson, I., Knights, C., *et al.* (2005) Targeting the dna repair defect in BRCA mutant cells as a therapeutic strategy. *Nature*, **434**(7035): 917–21.
- Fawcett, T. (2006) An introduction to ROC analysis. *Pattern Recognition Letters*, **27**(8): 861 – 874. {ROC} Analysis in Pattern Recognition.
- Fece de la Cruz, F., Gapp, B.V., and Nijman, S.M. (2015) Synthetic lethal vulnerabilities of cancer. *Annu Rev Pharmacol Toxicol*, **55**: 513–531.
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D.M., Forman, D., and Bray, F. (2015) Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*, **136**(5): E359–386.
- Fisher, R.A. (1919) Xv.—the correlation between relatives on the supposition of mendelian inheritance. *Earth and Environmental Science Transactions of the Royal Society of Edinburgh*, **52**(02): 399–433.
- Fong, P.C., Boss, D.S., Yap, T.A., Tutt, A., Wu, P., Mergui-Roelvink, M., Mortimer, P., Swaisland, H., Lau, A., O’Connor, M.J., *et al.* (2009) Inhibition of poly(adenosine diphosphate) polymerase in tumors from BRCA mutation carriers. *N Engl J Med*, **361**(2): 123–34.
- Fong, P.C., Yap, T.A., Boss, D.S., Carden, C.P., Mergui-Roelvink, M., Gourley, C., De Greve, J., Lubinski, J., Shanley, S., Messiou, C., *et al.* (2010) Poly(adenosine diphosphate)-ribose polymerase inhibition: frequent durable responses in BRCA carrier ovarian cancer correlating with platinum-free interval. *J Clin Oncol*, **28**(15): 2512–9.
- Forbes, S.A., Beare, D., Gunasekaran, P., Leung, K., Bindal, N., Boutselakis, H., Ding, M., Bamford, S., Cole, C., Ward, S., *et al.* (2015) COSMIC: exploring the world’s knowledge of somatic mutations in human cancer. *Nucleic Acids Res*, **43**(Database issue): D805–811.
- Fraser, A. (2004) Towards full employment: using RNAi to find roles for the redundant. *Oncogene*, **23**(51): 8346–52.
- Fromental-Ramain, C., Warot, X., Lakkaraju, S., Favier, B., Haack, H., Birling, C., Dierich, A., Dollé, P., and Chambon, P. (1996) Specific and redundant functions of the paralogous Hoxa-9 and Hoxd-9 genes in forelimb and axial skeleton patterning. *Development*, **122**(2): 461–472.
- Futreal, P.A., Coin, L., Marshall, M., Down, T., Hubbard, T., Wooster, R., Rahman, N., and Stratton, M.R. (2004) A census of human cancer genes. *Nat Rev Cancer*, **4**(3): 177–183.
- Futreal, P.A., Kasprzyk, A., Birney, E., Mullikin, J.C., Wooster, R., and Stratton, M.R. (2001) Cancer and genomics. *Nature*, **409**(6822): 850–852.

- Gao, B. and Roux, P.P. (2015) Translational control by oncogenic signaling pathways. *Biochimica et Biophysica Acta*, **1849**(7): 753–65.
- Gatza, M.L., Kung, H.N., Blackwell, K.L., Dewhirst, M.W., Marks, J.R., and Chi, J.T. (2011) Analysis of tumor environmental response and oncogenic pathway activation identifies distinct basal and luminal features in HER2-related breast tumor subtypes. *Breast Cancer Res*, **13**(3): R62.
- Gatza, M.L., Lucas, J.E., Barry, W.T., Kim, J.W., Wang, Q., Crawford, M.D., Datto, M.B., Kelley, M., Mathey-Prevot, B., Potti, A., *et al.* (2010) A pathway-based classification of human breast cancer. *Proc Natl Acad Sci USA*, **107**(15): 6994–6999.
- Gatza, M.L., Silva, G.O., Parker, J.S., Fan, C., and Perou, C.M. (2014) An integrated genomics approach identifies drivers of proliferation in luminal-subtype human breast cancer. *Nat Genet*, **46**(10): 1051–1059.
- Gentleman, R.C., Carey, V.J., Bates, D.M., Bolstad, B., Dettling, M., Dudoit, S., Ellis, B., Gautier, L., Ge, Y., Gentry, J., *et al.* (2004) Bioconductor: open software development for computational biology and bioinformatics. *Genome Biol*, **5**(10): R80.
- Genz, A. and Bretz, F. (2009) Computation of multivariate normal and t probabilities. In *Lecture Notes in Statistics*, volume 195. Springer-Verlag, Heidelberg.
- Genz, A., Bretz, F., Miwa, T., Mi, X., Leisch, F., Scheipl, F., and Hothorn, T. (2016) *mvtnorm: Multivariate Normal and t Distributions*. R package version 1.0-5. URL.
- Glaire, M.A., Brown, M., Church, D.N., and Tomlinson, I. (2017) Cancer predisposition syndromes: lessons for truly precision medicine. *J Pathol*, **241**(2): 226–235.
- Globus (Globus) (2017) Research data management simplified. <https://www.globus.org/>. Accessed: 25/03/2017.
- Goodwin, S., McPherson, J.D., and McCombie, W.R. (2016) Coming of age: ten years of next-generation sequencing technologies. *Nat Rev Genet*, **17**(6): 333–351.
- Grady, W.M., Willis, J., Guilford, P.J., Dunbier, A.K., Toro, T.T., Lynch, H., Wiesner, G., Ferguson, K., Eng, C., Park, J.G., *et al.* (2000) Methylation of the CDH1 promoter as the second genetic hit in hereditary diffuse gastric cancer. *Nat Genet*, **26**(1): 16–17.
- Graziano, F., Humar, B., and Guilford, P. (2003) The role of the E-cadherin gene (*CDH1*) in diffuse gastric cancer susceptibility: from the laboratory to clinical practice. *Annals of Oncology*, **14**(12): 1705–1713.
- Guaragnella, N., Palermo, V., Galli, A., Moro, L., Mazzoni, C., and Giannattasio, S. (2014) The expanding role of yeast in cancer research and diagnosis: insights into the function of the oncosuppressors p53 and BRCA1/2. *FEMS Yeast Res*, **14**(1): 2–16.

- Güell, O., Sagués, F., and Serrano, M. (2014) Essential plasticity and redundancy of metabolism unveiled by synthetic lethality analysis. *PLoS Comput Biol*, **10**(5): e1003637.
- Guilford, P. (1999) E-cadherin downregulation in cancer: fuel on the fire? *Molecular Medicine Today*, **5**(4): 172 – 177.
- Guilford, P., Hopkins, J., Harraway, J., McLeod, M., McLeod, N., Harawira, P., Taite, H., Scoular, R., Miller, A., and Reeve, A.E. (1998) E-cadherin germline mutations in familial gastric cancer. *Nature*, **392**(6674): 402–5.
- Guilford, P., Humar, B., and Blair, V. (2010) Hereditary diffuse gastric cancer: translation of *CDH1* germline mutations into clinical practice. *Gastric Cancer*, **13**(1): 1–10.
- Guilford, P.J., Hopkins, J.B., Grady, W.M., Markowitz, S.D., Willis, J., Lynch, H., Rajput, A., Wiesner, G.L., Lindor, N.M., Burgart, L.J., *et al.* (1999) E-cadherin germline mutations define an inherited cancer syndrome dominated by diffuse gastric cancer. *Hum Mutat*, **14**(3): 249–55.
- Guo, J., Liu, H., and Zheng, J. (2016) SynLethDB: synthetic lethality database toward discovery of selective and sensitive anticancer drug targets. *Nucleic Acids Res*, **44**(D1): D1011–1017.
- Hajian-Tilaki, K. (2013) Receiver Operating Characteristic (ROC) Curve Analysis for Medical Diagnostic Test Evaluation. *Caspian J Intern Med*, **4**(2): 627–635.
- Hall, M., Frank, E., Holmes, G., Pfahringer, B., Reutemann, P., and Witten, I.H. (2009) The weka data mining software: an update. *SIGKDD Explor Newsl*, **11**(1): 10–18.
- Hammerman, P.S., Lawrence, M.S., Voet, D., Jing, R., Cibulskis, K., Sivachenko, A., Stojanov, P., McKenna, A., Lander, E.S., Gabriel, S., *et al.* (2012) Comprehensive genomic characterization of squamous cell lung cancers. *Nature*, **489**(7417): 519–525.
- Hanahan, D. and Weinberg, R.A. (2000) The hallmarks of cancer. *Cell*, **100**(1): 57–70.
- Hanahan, D. and Weinberg, R.A. (2011) Hallmarks of cancer: the next generation. *Cell*, **144**(5): 646–674.
- Hanna, S. (2003) Cancer incidence in new zealand (2003-2007). In D. Forman, D. Bray F Brewster, C. Gombe Mbalawa, B. Kohler, M. Piñeros, E. Steliarova-Foucher, R. Swaminathan, and J. Ferlay (editors), *Cancer Incidence in Five Continents*, volume X, 902–907. International Agency for Research on Cancer, Lyon, France. Electronic version <http://ci5.iarc.fr> Accessed 22/03/2017.
- Hansford, S., Kaurah, P., Li-Chang, H., Woo, M., Senz, J., Pinheiro, H., Schrader, K.A., Schaeffer, D.F., Shumansky, K., Zogopoulos, G., *et al.* (2015) Hereditary Diffuse Gastric Cancer Syndrome: CDH1 Mutations and Beyond. *JAMA Oncol*, **1**(1): 23–32.

- Heiskanen, M.A. and Aittokallio, T. (2012) Mining high-throughput screens for cancer drug targets-lessons from yeast chemical-genomic profiling and synthetic lethality. *Wiley Interdisciplinary Reviews: Data Mining and Knowledge Discovery*, **2**(3): 263–272.
- Hell, P. (1976) Graphs with given neighbourhoods i. problèmes combinatoires at theorie des graphes. *Proc Coil Int CNRS, Orsay*, **260**: 219–223.
- Higgins, M.E., Claremont, M., Major, J.E., Sander, C., and Lash, A.E. (2007) CancerGenes: a gene selection resource for cancer genome projects. *Nucleic Acids Res*, **35**(Database issue): D721–726.
- Hillenmeyer, M.E. (2008) The chemical genomic portrait of yeast: uncovering a phenotype for all genes. *Science*, **320**: 362–365.
- Hoadley, K.A., Yau, C., Wolf, D.M., Cherniack, A.D., Tamborero, D., Ng, S., Leiserson, M.D., Niu, B., McLellan, M.D., Uzunangelov, V., *et al.* (2014) Multiplatform analysis of 12 cancer types reveals molecular classification within and across tissues of origin. *Cell*, **158**(4): 929–944.
- Hoehndorf, R., Hardy, N.W., Osumi-Sutherland, D., Tweedie, S., Schofield, P.N., and Gkoutos, G.V. (2013) Systematic analysis of experimental phenotype data reveals gene functions. *PLoS ONE*, **8**(4): e60847.
- Holm, S. (1979) A simple sequentially rejective multiple test procedure. *Scandinavian Journal of Statistics*, **6**(2): 65–70.
- Hopkins, A.L. (2008) Network pharmacology: the next paradigm in drug discovery. *Nat Chem Biol*, **4**(11): 682–690.
- Hu, Z., Fan, C., Oh, D.S., Marron, J.S., He, X., Qaqish, B.F., Livasy, C., Carey, L.A., Reynolds, E., Dressler, L., *et al.* (2006) The molecular portraits of breast tumors are conserved across microarray platforms. *BMC Genomics*, **7**: 96.
- Huang, E., Cheng, S., Dressman, H., Pittman, J., Tsou, M., Horng, C., Bild, A., Iversen, E., Liao, M., Chen, C., *et al.* (2003) Gene expression predictors of breast cancer outcomes. *Lancet*, **361**: 1590–1596.
- Hutchison, C.A., Chuang, R.Y., Noskov, V.N., Assad-Garcia, N., Deerinck, T.J., Ellisman, M.H., Gill, J., Kannan, K., Karas, B.J., Ma, L., *et al.* (2016) Design and synthesis of a minimal bacterial genome. *Science*, **351**(6280): aad6253.
- International HapMap 3 Consortium (HapMap) (2003) The International HapMap Project. *Nature*, **426**(6968): 789–796.
- Jeanes, A., Gottardi, C.J., and Yap, A.S. (2008) Cadherins and cancer: how does cadherin dysfunction promote tumor progression? *Oncogene*, **27**(55): 6920–6929.



- Jerby-Arnon, L., Pfotzer, N., Waldman, Y., McGarry, L., James, D., Shanks, E., Seashore-Ludlow, B., Weinstock, A., Geiger, T., Clemons, P., *et al.* (2014) Predicting cancer-specific vulnerability via data-driven detection of synthetic lethality. *Cell*, **158**(5): 1199–1209.
- Joachims, T. (1999) Making large-scale support vector machine learning practical. In S. Bernhard, Ikonf, J.C.B. Christopher, and J.S. Alexander (editors), *Advances in kernel methods*, 169–184. MIT Press.
- Ju, Z., Liu, W., Roebuck, P.L., Siwak, D.R., Zhang, N., Lu, Y., Davies, M.A., Akbani, R., Weinstein, J.N., Mills, G.B., *et al.* (2015) Development of a robust classifier for quality control of reverse-phase protein arrays. *Bioinformatics*, **31**(6): 912.
- Kaelin, Jr, W. (2005) The concept of synthetic lethality in the context of anticancer therapy. *Nat Rev Cancer*, **5**(9): 689–98.
- Kaelin, Jr, W. (2009) Synthetic lethality: a framework for the development of wiser cancer therapeutics. *Genome Med*, **1**: 99.
- Kamada, T. and Kawai, S. (1989) An algorithm for drawing general undirected graphs. *Information Processing Letters*, **31**(1): 7–15.
- Kawai, J., Shinagawa, A., Shibata, K., Yoshino, M., Itoh, M., Ishii, Y., Arakawa, T., Hara, A., Fukunishi, Y., Konno, H., *et al.* (2001) Functional annotation of a full-length mouse cDNA collection. *Nature*, **409**(6821): 685–690.
- Kelley, R. and Ideker, T. (2005) Systematic interpretation of genetic interactions using protein networks. *Nat Biotech*, **23**(5): 561–566.
- Kelly, S.T. (2013) *Statistical Predictions of Synthetic Lethal Interactions in Cancer*. Dissertation, University of Otago.
- Keshava Prasad, T.S., Goel, R., Kandasamy, K., Keerthikumar, S., Kumar, S., Mathivanan, S., Telikicherla, D., Raju, R., Shafreen, B., Venugopal, A., *et al.* (2009) Human Protein Reference Database–2009 update. *Nucleic Acids Res*, **37**(Database issue): D767–772.
- Kim, N.G., Koh, E., Chen, X., and Gumbiner, B.M. (2011) E-cadherin mediates contact inhibition of proliferation through Hippo signaling-pathway components. *Proc Natl Acad Sci USA*, **108**(29): 11930–11935.
- Koboldt, D.C., Fulton, R.S., McLellan, M.D., Schmidt, H., Kalicki-Veizer, J., McMichael, J.F., Fulton, L.L., Dooling, D.J., Ding, L., Mardis, E.R., *et al.* (2012) Comprehensive molecular portraits of human breast tumours. *Nature*, **490**(7418): 61–70.
- Kockel, L., Zeitlinger, J., Staszewski, L.M., Mlodzik, M., and Bohmann, D. (1997) Jun in drosophila development: redundant and nonredundant functions and regulation by two mapk signal transduction pathways. *Genes & Development*, **11**(13): 1748–1758.

- Kozlov, K.N., Gursky, V.V., Kulakovskiy, I.V., and Samsonova, M.G. (2015) Sequence-based model of gap gene regulation network. *BMC Genomics*, **15**(Suppl 12): S6.
- Kranthi, S., Rao, S., and Manimaran, P. (2013) Identification of synthetic lethal pairs in biological systems through network information centrality. *Mol BioSyst*, **9**(8): 2163–2167.
- Kroepil, F., Fluegen, G., Totikov, Z., Baldus, S.E., Vay, C., Schauer, M., Topp, S.A., Esch, J.S., Knoefel, W.T., and Stoecklein, N.H. (2012) Down-regulation of CDH1 is associated with expression of SNAIL in colorectal adenomas. *PLoS ONE*, **7**(9): e46665.
- Lander, E.S. (2011) Initial impact of the sequencing of the human genome. *Nature*, **470**(7333): 187–197.
- Lander, E.S., Linton, L.M., Birren, B., Nusbaum, C., Zody, M.C., Baldwin, J., Devon, K., Dewar, K., Doyle, M., FitzHugh, W., *et al.* (2001) Initial sequencing and analysis of the human genome. *Nature*, **409**(6822): 860–921.
- Langmead, B., Trapnell, C., Pop, M., and Salzberg, S.L. (2009) Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biol*, **10**(3): R25.
- Latora, V. and Marchiori, M. (2001) Efficient behavior of small-world networks. *Phys Rev Lett*, **87**: 198701.
- Laufer, C., Fischer, B., Billmann, M., Huber, W., and Boutros, M. (2013) Mapping genetic interactions in human cancer cells with RNAi and multiparametric phenotyping. *Nat Methods*, **10**(5): 427–31.
- Law, C.W., Chen, Y., Shi, W., and Smyth, G.K. (2014) voom: precision weights unlock linear model analysis tools for RNA-seq read counts. *Genome Biol*, **15**(2): R29.
- Le Meur, N. and Gentleman, R. (2008) Modeling synthetic lethality. *Genome Biol*, **9**(9): R135.
- Le Meur, N., Jiang, Z., Liu, T., Mar, J., and Gentleman, R.C. (2014) Slgi: Synthetic lethal genetic interaction. r package version 1.26.0.
- Lee, A.Y., Perreault, R., Harel, S., Boulier, E.L., Suderman, M., Hallett, M., and Jenna, S. (2010a) Searching for signaling balance through the identification of genetic interactors of the rab guanine-nucleotide dissociation inhibitor gdi-1. *PLoS ONE*, **5**(5): e10624.
- Lee, I., Lehner, B., Vavouri, T., Shin, J., Fraser, A.G., and Marcotte, E.M. (2010b) Predicting genetic modifier loci using functional gene networks. *Genome Research*, **20**(8): 1143–1153.
- Lee, I. and Marcotte, E.M. (2009) Effects of functional bias on supervised learning of a gene network model. *Methods Mol Biol*, **541**: 463–75.

- Lee, M.J., Ye, A.S., Gardino, A.K., Heijink, A.M., Sorger, P.K., MacBeath, G., and Yaffe, M.B. (2012) Sequential application of anticancer drugs enhances cell death by rewiring apoptotic signaling networks. *Cell*, **149**(4): 780–94.
- Lehner, B., Crombie, C., Tischler, J., Fortunato, A., and Fraser, A.G. (2006) Systematic mapping of genetic interactions in *caenorhabditis elegans* identifies common modifiers of diverse signaling pathways. *Nat Genet*, **38**(8): 896–903.
- Li, B., Ruotti, V., Stewart, R.M., Thomson, J.A., and Dewey, C.N. (2010) RNA-Seq gene expression estimation with read mapping uncertainty. *Bioinformatics*, **26**(4): 493–500.
- Li, X.J., Mishra, S.K., Wu, M., Zhang, F., and Zheng, J. (2014) Syn-lethality: An integrative knowledge base of synthetic lethality towards discovery of selective anti-cancer therapies. *Biomed Res Int*, **2014**: 196034.
- Linehan, W.M., Spellman, P.T., Ricketts, C.J., Creighton, C.J., Fei, S.S., Davis, C., Wheeler, D.A., Murray, B.A., Schmidt, L., Vocke, C.D., *et al.* (2016) Comprehensive Molecular Characterization of Papillary Renal-Cell Carcinoma. *N Engl J Med*, **374**(2): 135–145.
- Lokody, I. (2014) Computational modelling: A computational crystal ball. *Nature Reviews Cancer*, **14**(10): 649–649.
- Lord, C.J., Tutt, A.N., and Ashworth, A. (2015) Synthetic lethality and cancer therapy: lessons learned from the development of PARP inhibitors. *Annu Rev Med*, **66**: 455–470.
- Lu, X., Kensche, P.R., Huynen, M.A., and Notebaart, R.A. (2013) Genome evolution predicts genetic interactions in protein complexes and reveals cancer drug targets. *Nat Commun*, **4**: 2124.
- Lu, X., Megchelenbrink, W., Notebaart, R.A., and Huynen, M.A. (2015) Predicting human genetic interactions from cancer genome evolution. *PLoS One*, **10**(5): e0125795.
- Lum, P.Y., Armour, C.D., Stepaniants, S.B., Cavet, G., Wolf, M.K., Butler, J.S., Hinshaw, J.C., Garnier, P., Prestwich, G.D., Leonardson, A., *et al.* (2004) Discovering modes of action for therapeutic compounds using a genome-wide screen of yeast heterozygotes. *Cell*, **116**(1): 121–137.
- Luo, J., Solimini, N.L., and Elledge, S.J. (2009) Principles of Cancer Therapy: Oncogene and Non-oncogene Addiction. *Cell*, **136**(5): 823–837.
- Machado, J., Olivera, C., Carvalh, R., Soares, P., Berx, G., Caldas, C., Sercuca, R., Carneiro, F., and Sorbrinho-Simoes, M. (2001) E-cadherin gene (*CDH1*) promoter methylation as the second hit in sporadic diffuse gastric carcinoma. *Oncogene*, **20**: 1525–1528.

- Markowetz, F. (2017) All biology is computational biology. *PLoS Biol*, **15**(3): e2002050.
- Masciari, S., Larsson, N., Senz, J., Boyd, N., Kaurah, P., Kandel, M.J., Harris, L.N., Pinheiro, H.C., Troussard, A., Miron, P., *et al.* (2007) Germline E-cadherin mutations in familial lobular breast cancer. *J Med Genet*, **44**(11): 726–31.
- Mattison, J., van der Weyden, L., Hubbard, T., and Adams, D.J. (2009) Cancer gene discovery in mouse and man. *Biochim Biophys Acta*, **1796**(2): 140–161.
- McLachlan, J., George, A., and Banerjee, S. (2016) The current status of parp inhibitors in ovarian cancer. *Tumori*, **102**(5): 433–440.
- McLendon, R., Friedman, A., Bigner, D., Van Meir, E.G., Brat, D.J., Mastrogianakis, G.M., Olson, J.J., Mikkelsen, T., Lehman, N., Aldape, K., *et al.* (2008) Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature*, **455**(7216): 1061–1068.
- Miles, D.W. (2001) Update on HER-2 as a target for cancer therapy: herceptin in the clinical setting. *Breast Cancer Res*, **3**(6): 380–384.
- Muzny, D.M., Bainbridge, M.N., Chang, K., Dinh, H.H., Drummond, J.A., Fowler, G., Kovar, C.L., Lewis, L.R., Morgan, M.B., Newsham, I.F., *et al.* (2012) Comprehensive molecular characterization of human colon and rectal cancer. *Nature*, **487**(7407): 330–337.
- Nagalla, S., Chou, J.W., Willingham, M.C., Ruiz, J., Vaughn, J.P., Dubey, P., Lash, T.L., Hamilton-Dutoit, S.J., Bergh, J., Sotiriou, C., *et al.* (2013) Interactions between immunity, proliferation and molecular subtype in breast cancer prognosis. *Genome Biol*, **14**(4): R34.
- Neeley, E.S., Kornblau, S.M., Coombes, K.R., and Baggerly, K.A. (2009) Variable slope normalization of reverse phase protein arrays. *Bioinformatics*, **25**(11): 1384.
- Novomestky, F. (2012) *matrixcalc: Collection of functions for matrix calculations*. R package version 1.0-3.
- Nowak, M.A., Boerlijst, M.C., Cooke, J., and Smith, J.M. (1997) Evolution of genetic redundancy. *Nature*, **388**(6638): 167–171.
- Oliveira, C., Senz, J., Kaurah, P., Pinheiro, H., Sanges, R., Haegert, A., Corso, G., Schouten, J., Fitzgerald, R., Vogelsang, H., *et al.* (2009) Germline *CDH1* deletions in hereditary diffuse gastric cancer families. *Human Molecular Genetics*, **18**(9): 1545–1555.
- Oliveira, C., Seruca, R., Hoogerbrugge, N., Ligtenberg, M., and Carneiro, F. (2013) Clinical utility gene card for: Hereditary diffuse gastric cancer (HDGC). *Eur J Hum Genet*, **21**(8).

- Pandey, G., Zhang, B., Chang, A.N., Myers, C.L., Zhu, J., Kumar, V., and Schadt, E.E. (2010) An integrative multi-network and multi-classifier approach to predict genetic interactions. *PLoS Comput Biol*, **6**(9).
- Parker, J., Mullins, M., Cheung, M., Leung, S., Voduc, D., Vickery, T., Davies, S., Fauron, C., He, X., Hu, Z., *et al.* (2009) Supervised risk predictor of breast cancer based on intrinsic subtypes. *Journal of Clinical Oncology*, **27**(8): 1160–1167.
- Pereira, B., Chin, S.F., Rueda, O.M., Vollan, H.K., Provenzano, E., Bardwell, H.A., Pugh, M., Jones, L., Russell, R., Sammut, S.J., *et al.* (2016) Erratum: The somatic mutation profiles of 2,433 breast cancers refine their genomic and transcriptomic landscapes. *Nat Commun*, **7**: 11908.
- Perou, C.M., Sørlie, T., Eisen, M.B., van de Rijn, M., Jeffrey, S.S., Rees, C.A., Pollack, J.R., Ross, D.T., Johnsen, H., Akslen, L.A., *et al.* (2000) Molecular portraits of human breast tumours. *Nature*, **406**(6797): 747–752.
- Polyak, K. and Weinberg, R.A. (2009) Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits. *Nat Rev Cancer*, **9**(4): 265–73.
- R Core Team (2016) *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria. R version 3.3.2.
- Ritchie, M.E., Phipson, B., Wu, D., Hu, Y., Law, C.W., Shi, W., and Smyth, G.K. (2015) limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Research*, **43**(7): e47.
- Roguev, A., Bandyopadhyay, S., Zofall, M., Zhang, K., Fischer, T., Collins, S.R., Qu, H., Shales, M., Park, H.O., Hayles, J., *et al.* (2008) Conservation and rewiring of functional modules revealed by an epistasis map in fission yeast. *Science*, **322**(5900): 405–10.
- Roychowdhury, S. and Chinnaiyan, A.M. (2016) Translating cancer genomes and transcriptomes for precision oncology. *CA Cancer J Clin*, **66**(1): 75–88.
- Rung, J. and Brazma, A. (2013) Reuse of public genome-wide gene expression data. *Nat Rev Genet*, **14**(2): 89–99.
- Ryan, C., Lord, C., and Ashworth, A. (2014) Daisy: Picking synthetic lethals from cancer genomes. *Cancer Cell*, **26**(3): 306–308.
- Schena, M. (1996) Genome analysis with gene expression microarrays. *Bioessays*, **18**(5): 427–431.
- Scheuer, L., Kauff, N., Robson, M., Kelly, B., Barakat, R., Satagopan, J., Ellis, N., Hensley, M., Boyd, J., Borgen, P., *et al.* (2002) Outcome of preventive surgery and screening for breast and ovarian cancer in BRCA mutation carriers. *J Clin Oncol*, **20**(5): 1260–1268.

- Semb, H. and Christofori, G. (1998) The tumor-suppressor function of E-cadherin. *Am J Hum Genet*, **63**(6): 1588–93.
- Sing, T., Sander, O., Beerenwinkel, N., and Lengauer, T. (2005) ROCR: visualizing classifier performance in R. *Bioinformatics*, **21**(20): 7881.
- Slurm development team (Slurm) (2017) Slurm workload manager. <https://slurm.schedmd.com/>. Accessed: 25/03/2017.
- Sørlie, T., Perou, C.M., Tibshirani, R., Aas, T., Geisler, S., Johnsen, H., Hastie, T., Eisen, M.B., van de Rijn, M., Jeffrey, S.S., *et al.* (2001) Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc Natl Acad Sci USA*, **98**(19): 10869–10874.
- Srihari, S., Singla, J., Wong, L., and Ragan, M.A. (2015) Inferring synthetic lethal interactions from mutual exclusivity of genetic events in cancer. *Biology Direct*, **10**(1): 57.
- Stajich, J.E. and Lapp, H. (2006) Open source tools and toolkits for bioinformatics: significance, and where are we? *Brief Bioinformatics*, **7**(3): 287–296.
- Stratton, M.R., Campbell, P.J., and Futreal, P.A. (2009) The cancer genome. *Nature*, **458**(7239): 719–724.
- Ström, C. and Helleday, T. (2012) Strategies for the use of poly(adenosine diphosphate ribose) polymerase (parp) inhibitors in cancer therapy. *Biomolecules*, **2**(4): 635–649.
- Tarazona, S., Garcia-Alcalde, F., Dopazo, J., Ferrer, A., and Conesa, A. (2011) Differential expression in RNA-seq: a matter of depth. *Genome Res*, **21**(12): 2213–2223.
- Telford, B.J., Chen, A., Beetham, H., Frick, J., Brew, T.P., Gould, C.M., Single, A., Godwin, T., Simpson, K.J., and Guilford, P. (2015) Synthetic lethal screens identify vulnerabilities in gpcr signalling and cytoskeletal organization in E-cadherin-deficient cells. *Mol Cancer Ther*, **14**(5): 1213–1223.
- The 1000 Genomes Project Consortium (1000 Genomes) (2010) A map of human genome variation from population-scale sequencing. *Nature*, **467**(7319): 1061–1073.
- The Cancer Genome Atlas Research Network (TCGA) (2017) The Cancer Genome Atlas Project. <https://cancergenome.nih.gov/>. Accessed: 26/03/2017.
- The Catalogue Of Somatic Mutations In Cancer (COSMIC) (2016) Cosmic: The catalogue of somatic mutations in cancer. <http://cancer.sanger.ac.uk/cosmic>. Release 79 (23/08/2016), Accessed: 05/02/2017.
- The Comprehensive R Archive Network (CRAN) (2017) Cran. <https://cran.r-project.org/>. Accessed: 24/03/2017.
- The ENCODE Project Consortium (ENCODE) (2004) The ENCODE (ENCyclopedia Of DNA Elements) Project. *Science*, **306**(5696): 636–640.

- The National Cancer Institute (NCI) (2015) The genetics of cancer. <https://www.cancer.gov/about-cancer/causes-prevention/genetics>. Published: 22/04/2015, Accessed: 22/03/2017.
- The New Zealand eScience Infrastructure (NeSI) (2017) NeSI. <https://www.nesi.org.nz/>. Accessed: 25/03/2017.
- Tierney, L., Rossini, A.J., Li, N., and Sevcikova, H. (2015) *snow: Simple Network of Workstations*. R package version 0.4-2.
- Tiong, K.L., Chang, K.C., Yeh, K.T., Liu, T.Y., Wu, J.H., Hsieh, P.H., Lin, S.H., Lai, W.Y., Hsu, Y.C., Chen, J.Y., *et al.* (2014) Csnk1e/ctnnb1 are synthetic lethal to tp53 in colorectal cancer and are markers for prognosis. *Neoplasia*, **16**(5): 441–50.
- Tischler, J., Lehner, B., and Fraser, A.G. (2008) Evolutionary plasticity of genetic interaction networks. *Nat Genet*, **40**(4): 390–391.
- Tomasetti, C. and Vogelstein, B. (2015) Cancer etiology. Variation in cancer risk among tissues can be explained by the number of stem cell divisions. *Science*, **347**(6217): 78–81.
- Tong, A.H., Evangelista, M., Parsons, A.B., Xu, H., Bader, G.D., Page, N., Robinson, M., Raghibizadeh, S., Hogue, C.W., Bussey, H., *et al.* (2001) Systematic genetic analysis with ordered arrays of yeast deletion mutants. *Science*, **294**(5550): 2364–8.
- Tong, A.H., Lesage, G., Bader, G.D., Ding, H., Xu, H., Xin, X., Young, J., Berriz, G.F., Brost, R.L., Chang, M., *et al.* (2004) Global mapping of the yeast genetic interaction network. *Science*, **303**(5659): 808–13.
- Tran, B., Dancey, J.E., Kamel-Reid, S., McPherson, J.D., Bedard, P.L., Brown, A.M., Zhang, T., Shaw, P., Onetto, N., Stein, L., *et al.* (2012) Cancer genomics: technology, discovery, and translation. *J Clin Oncol*, **30**(6): 647–660.
- Travers, J. and Milgram, S. (1969) An experimental study of the small world problem. *Sociometry*, **32**(4): 425–443.
- Tunggal, J.A., Helfrich, I., Schmitz, A., Schwarz, H., Gunzel, D., Fromm, M., Kemler, R., Krieg, T., and Niessen, C.M. (2005) E-cadherin is essential for in vivo epidermal barrier function by regulating tight junctions. *EMBO J*, **24**(6): 1146–1156.
- Tutt, A., Robson, M., Garber, J.E., Domchek, S.M., Audeh, M.W., Weitzel, J.N., Friedlander, M., Arun, B., Loman, N., Schmutzler, R.K., *et al.* (2010) Oral poly(adenosine diphosphate) polymerase inhibitor olaparib in patients with *BRCA1* or *BRCA2* mutations and advanced breast cancer: a proof-of-concept trial. *Lancet*, **376**(9737): 235–44.
- University of California, Santa Cruz (UCSC) (2012) Uscs cancer browser. Accessed 29/03/2012.
- van der Meer, R., Song, H.Y., Park, S.H., Abdulkadir, S.A., and Roh, M. (2014) RNAi screen identifies a synthetic lethal interaction between PIM1 overexpression and PLK1 inhibition. *Clinical Cancer Research*, **20**(12): 3211–3221.

- van der Post, R.S., Vogelaar, I.P., Carneiro, F., Guilford, P., Huntsman, D., Hoogerbrugge, N., Caldas, C., Schreiber, K.E., Hardwick, R.H., Ausems, M.G., *et al.* (2015) Hereditary diffuse gastric cancer: updated clinical guidelines with an emphasis on germline CDH1 mutation carriers. *J Med Genet*, **52**(6): 361–374.
- van Steen, K. (2012) Travelling the world of gene–gene interactions. *Briefings in Bioinformatics*, **13**(1): 1–19.
- van Steen, M. (2010) *Graph Theory and Complex Networks: An Introduction*. Maarten van Steen, VU Amsterdam.
- Vapnik, V.N. (1995) *The nature of statistical learning theory*. Springer-Verlag New York, Inc.
- Vizeacoumar, F.J., Arnold, R., Vizeacoumar, F.S., Chandrashekhar, M., Buzina, A., Young, J.T., Kwan, J.H., Sayad, A., Mero, P., Lawo, S., *et al.* (2013) A negative genetic interaction map in isogenic cancer cell lines reveals cancer cell vulnerabilities. *Mol Syst Biol*, **9**: 696.
- Vogelstein, B., Papadopoulos, N., Velculescu, V.E., Zhou, S., Diaz, L.A., and Kinzler, K.W. (2013) Cancer genome landscapes. *Science*, **339**(6127): 1546–1558.
- Vos, C.B., Cleton-Jansen, A.M., Berx, G., de Leeuw, W.J., ter Haar, N.T., van Roy, F., Cornelisse, C.J., Peterse, J.L., and van de Vijver, M.J. (1997) E-cadherin inactivation in lobular carcinoma in situ of the breast: an early event in tumorigenesis. *Br J Cancer*, **76**(9): 1131–3.
- Waldron, D. (2016) Cancer genomics: A multi-layer omics approach to cancer. *Nat Rev Genet*, **17**(8): 436–437.
- Wang, K., Singh, D., Zeng, Z., Coleman, S.J., Huang, Y., Savich, G.L., He, X., Mieczkowski, P., Grimm, S.A., Perou, C.M., *et al.* (2010) MapSplice: accurate mapping of RNA-seq reads for splice junction discovery. *Nucleic Acids Res*, **38**(18): e178.
- Wang, X. and Simon, R. (2013) Identification of potential synthetic lethal genes to p53 using a computational biology approach. *BMC Medical Genomics*, **6**(1): 30.
- Wappett, M. (2014) Bisep: Toolkit to identify candidate synthetic lethality. r package version 2.0.
- Wappett, M., Dulak, A., Yang, Z.R., Al-Watban, A., Bradford, J.R., and Dry, J.R. (2016) Multi-omic measurement of mutually exclusive loss-of-function enriches for candidate synthetic lethal gene pairs. *BMC Genomics*, **17**: 65.
- Warnes, G.R., Bolker, B., Bonebakker, L., Gentleman, R., Liaw, W.H.A., Lumley, T., Maechler, M., Magnusson, A., Moeller, S., Schwartz, M., *et al.* (2015) *gplots: Various R Programming Tools for Plotting Data*. R package version 2.17.0.



- Watts, D.J. and Strogatz, S.H. (1998) Collective dynamics of 'small-world' networks. *Nature*, **393**(6684): 440–2.
- Weinstein, I.B. (2000) Disorders in cell circuitry during multistage carcinogenesis: the role of homeostasis. *Carcinogenesis*, **21**(5): 857–864.
- Weinstein, J.N., Akbani, R., Broom, B.M., Wang, W., Verhaak, R.G., McConkey, D., Lerner, S., Morgan, M., Creighton, C.J., Smith, C., *et al.* (2014) Comprehensive molecular characterization of urothelial bladder carcinoma. *Nature*, **507**(7492): 315–322.
- Weinstein, J.N., Collisson, E.A., Mills, G.B., Shaw, K.R., Ozenberger, B.A., Ellrott, K., Shmulevich, I., Sander, C., Stuart, J.M., Chang, K., *et al.* (2013) The Cancer Genome Atlas Pan-Cancer analysis project. *Nat Genet*, **45**(10): 1113–1120.
- Wickham, H. and Chang, W. (2016) *devtools: Tools to Make Developing R Packages Easier*. R package version 1.12.0.
- Wickham, H., Danenberg, P., and Eugster, M. (2017) *roxygen2: In-Line Documentation for R*. R package version 6.0.1.
- Wong, S.L., Zhang, L.V., Tong, A.H.Y., Li, Z., Goldberg, D.S., King, O.D., Lesage, G., Vidal, M., Andrews, B., Bussey, H., *et al.* (2004) Combining biological networks to predict genetic interactions. *Proceedings of the National Academy of Sciences of the United States of America*, **101**(44): 15682–15687.
- World Health Organization (WHO) (2017) Fact sheet: Cancer. <http://www.who.int/mediacentre/factsheets/fs297/en/>. Updated February 2017, Accessed: 22/03/2017.
- Wu, M., Li, X., Zhang, F., Li, X., Kwoh, C.K., and Zheng, J. (2014) In silico prediction of synthetic lethality by meta-analysis of genetic interactions, functions, and pathways in yeast and human cancer. *Cancer Inform*, **13**(Suppl 3): 71–80.
- Yu, H. (2002) Rmpi: Parallel statistical computing in r. *R News*, **2**(2): 10–14.
- Zhang, F., Wu, M., Li, X.J., Li, X.L., Kwoh, C.K., and Zheng, J. (2015) Predicting essential genes and synthetic lethality via influence propagation in signaling pathways of cancer cell fates. *J Bioinform Comput Biol*, **13**(3): 1541002.
- Zhang, J., Baran, J., Cros, A., Guberman, J.M., Haider, S., Hsu, J., Liang, Y., Rivkin, E., Wang, J., Whitty, B., *et al.* (2011) International cancer genome consortium data portal—a one-stop shop for cancer genomics data. *Database: The Journal of Biological Databases and Curation*, **2011**: bar026.
- Zhong, W. and Sternberg, P.W. (2006) Genome-wide prediction of c. elegans genetic interactions. *Science*, **311**(5766): 1481–1484.
- Zweig, M.H. and Campbell, G. (1993) Receiver-operating characteristic (roc) plots: a fundamental evaluation tool in clinical medicine. *Clinical Chemistry*, **39**(4): 561–577.