Library Declaration Form



University of Otago Library

Author's full name and year of birth:	Simon Thomas Kelly,
(for cataloguing purposes)	24 February 1992

Title of thesis: A Bioinformatics Approach to Synthetic Lethal Interactions in Breast Cancer with Gene Expression Data

Degree: Doctor of Philosophy

Department: Department of Biochemistry

Permanent Address: 710 Cumberland Street, Dunedin, NZ

I agree that this thesis may be consulted for research and study purposes and that reasonable quotation may be made from it, provided that proper acknowledgement of its use is made.

I consent to this thesis being copied in part or in whole for

- i) a library
- ii) an individual

at the discretion of the University of Otago.

Signature:

Date:

A Bioinformatics Approach to Synthetic Lethal Interactions in Breast Cancer with Gene Expression Data

S. Thomas Kelly

a thesis submitted for the degree of Doctor of Philosophy at the University of Otago, Dunedin, New Zealand.

 $23~\mathrm{June}~2017$

Abstract

Background

Synthetic lethal genetic interactions are re-emerging in the post-genomics era due to their potential for use in precision medicine against cancers. Synthetic lethal drug design exploits the functional redundancy of genes disrupted in cancers (including tumour suppressors) to develop specific treatments against them. *CDH1*, which encodes E-cadherin, is a tumour supressor gene with loss of function in breast and stomach cancers. Experimental screens have identified candidate synthetic lethal interactions for drug target triage, which can be further supported with bioinformatics analysis. Furthermore, gene expression data enables investigation of synthetic lethal pathways and graph structure of synthetic lethal genes within them.

Approach

A computational methodology, the Synthetic Lethal Prediction Tool (SLIPT) has been developed to detect synthetic lethal interactions in gene expression data. This methodology was demonstrated on interactions with *CDH1* in breast and stomach cancer data from The Cancer Genome Atlas (TCGA) project. Synthetic lethal genes and pathways were further investigated with unsupervised clustering, gene set over-representation analysis, metagenes, and permutation resampling. In particular, analyses focused on comparing SLIPT gene candidates to an experimental siRNA screen Telford *et al.* (2015). Network analysis methods were applied to the most supported pathways to test for pathway structure among between synthetic lethal candidates. Simulation and modelling was used to assess the statistical performance of SLIPT, including simulated data with correlation structures derived from graph structures.

Findings

Many candidate synthetic lethal partners of CDH1 were detected in both TCGA breast cancer. These genes clustered into several distinct groups, with distinct biological functions and elevated expression in different clinical subtypes. While the number of genes detected by both approaches was not significant, these contained significantly enriched pathways. In particular, $G_{\alpha i}$ signalling, cytoplasmic microfibres, and extracellular fibrin clotting were robustly supported by both approaches, which is consistent with the known cytoskeletal and cell signalling roles of E-cadherin and validation of GPCR pathways performed by Telford $et\ al.\ (2015)$. Many of these pathways were replicated in stomach cancer data. The pathways supported only by SLIPT included regulation of immune signalling and translational elongation which were not expected to be detected in an isogenic cell line model but are still candidates for further investigation.

Synthetic lethal candidates detected by SLIPT and siRNA were compared within graph structures of the candidate synthetic lethal pathways. These genes did not differ with respect to network metrics of importance or connectivity in the pathway. There was little support, across pathways, that SLIPT gene candidates were consistently upstream or downstream of siRNA gene candidates within pathways.

A model of synthetic lethality was used to simulate gene expression data with synthetic lethal partners of a gene. The SLIPT methodology had high statistical performance, detecting few synthetic lethal partners, which diminished with more synthetic lethal partners or lower sample size. The SLIPT methodology performed better than Pearson correlation or the χ^2 -test. In particular, it performed well with high specificity for datasets containing thousands of genes or genes positively correlated with the query gene (as expected to occur in expression data). SLIPT was robust across correlation structures, including those derived from complex pathway structures and often distinguished synthetic lethal genes from those positively or negatively correlated with them. Therefore SLIPT is appropriate to identify synthetic lethal genes within pathways and use candidate synthetic lethal genes (and their correlates) to identify synthetic lethal pathways.

Summary

Thus this thesis has developed, evaluated, and refined a bioinformatics approach to discovery of synthetic lethal genes solely from gene expression data. This approach has been demonstrated to detect biologically informative and clinically relevant candidate partners for *CDH1* in breast and stomach cancers. These investigations have also involved the development of network analysis and simulation procedures which may be more widely applicable.

Acknowledgements

I thank my supervisors A/Prof. Mik Black and Prof. Parry Guilford for their support and guidance throughout this my postgraduate studies. It has been a great experience, I look forward to seeing what your research groups produce in the future, may this not be the end for us.

I am also thankful for the guidance and mentorship of Prof. Hamish Spencer for career advice throughout my studies and time in his research group.

I am also grateful to the past and current members of these research groups, and my peers at the laboratory benches and computers across campus. The peer support, camaraderie, and guidance of newer students has been an incredible part of my time at Otago and has made my thesis studies not just easier but possible at all. The postgraduate community is very special here and I have truly made some lifelong friends from all over the world. You are talented researchers and amazing people. May we meet again some day. Where-ever you may end up, its small world and there's always time to catch up. I'd be delighted to host some visits while working abroad.

I cannot thank my friends, flatmates, family, and diligent proofreaders enough for their patience and support during such as massive, challenging, and (I'm sure you've heard too often) stressful undertaking during both my PhD and the study leading up to it. There are too many of you to name everyone here without leaving someone out, so thank you all for everything you've done, both the good times and the tough. Thank you for at least pretending to understand complex math oft brought up at the wrong moment. Thank you for checking my writing or slides, even when sprung on you last minute. Thank for your time when what I really needed was a chat, a walk, a drink with "the guys", or a moment to think clearly.

I thank the various organisations that supported this research project:

- This thesis was supported by the Postgraduate Tassell Scholarship in Cancer Research, a University of Otago Doctoral Scholarship.
- The New Zealand eScience Infrastucture (NeSI) provided access to the Intel Pan high-performance computing cluster, support, and training to use it effectively. Various aspects of this thesis would not have been possible without access to such an incredible national resource.
- The Health Research Council (HRC) of New Zealand provided funding for experimental research in the Cancer Genetics Laboratory. Some aspects of this project would not have been possible without access to the data and findings funded by this grant.
- The Allan Wilson Centre and Otago School of Biomedical Sciences provided funding for summer research placements which was a valuable opportunity to gain experience and training used in this thesis project.

I thank the following organisations for support towards presenting findings in this thesis at conference and seminars:

- Google (eResearch 2014, Hamilton)
- NeSI (Software Carpentry training and Research Bazaar 2015, Melbourne)
- REANNZ, NZGL, and NeSI (eResearch 2016, Queenstown)
- Otago Division of Health Sciences, Oxford Global, and Maurice and Phyllis Paykel Trust (NGS Asia 2016, Singapore)
- RIKEN Division of Genomic Technologies and the Okinawa Institute of Science and Technology (seminar visits in Japan)

Thanks most of all to my fianceé, Dr Yui Kawagishi, you've been an inspiration. Thank you for your support and encouragement, every day, even from afar: it has always made a difference. It's been incredible to see you flourish in your career and I look forward to joining you again soon. May the next chapter of our adventures involve a bit less Skype across timezones.

どうもありがとう由ちゃん。頑張った!もうすぐ行きます。!また来月!

Contents

\mathbf{G}	Glossary		xvii		
Acronyms			xxiv		
1	Intr	oducti	ion and Literature Review		1
	1.1	Cance	er Research in the Post-Genomic Era		1
		1.1.1	Cancer is a Global Health Issue		2
			1.1.1.1 The Genetics and Molecular Biology of Cancers		3
		1.1.2	The Genomics Revolution in Cancer Research		3
			1.1.2.1 High-Throughput Technologies		4
			1.1.2.2 Bioinformatics and Genomic Data		5
		1.1.3	Genomics Projects		5
			1.1.3.1 The Cancer Genome Project		6
			1.1.3.2 The Cancer Genome Atlas Project		6
		1.1.4	Genomic Cancer Medicine		8
			1.1.4.1 Cancer Genes and Driver Mutations		8
			1.1.4.2 Precision Cancer Medicine		9
			1.1.4.3 Molecular Diagnostics and Pan-Cancer Medicine		9
			1.1.4.4 Targeted Therapeutics and Pharmacogenomics		10
		1.1.5	Systems and Network Biology		11
	1.2		etic Lethal Cancer Medicine		12
		1.2.1	Synthetic Lethal Genetic Interactions		13
		1.2.2	Synthetic Lethal Concepts in Genetics		13
		1.2.3	Synthetic Lethality in Model Systems		15
			1.2.3.1 Synthetic Lethal Pathways and Networks		15
			1.2.3.2 Evolution of Synthetic Lethality		16
		1.2.4	Synthetic Lethality in Cancer		17
		1.2.5	Clinical Impact of Synthetic Lethality in Cancer		18
		1.2.6	High-throughput Screening for Synthetic Lethality		20
			1.2.6.1 Synthetic Lethal Screens		21
		1.2.7	Computational Prediction of Synthetic Lethality		22
			1.2.7.1 Bioinformatics Approaches to Genetic Interactions .		22
			1.2.7.2 Comparative Genomics		23
			1.2.7.3 Analysis and Modelling of Protein Data		26
			1.2.7.4 Differential Gene Expression		28
			1.2.7.5 Data Mining and Machine Learning		29

			1.2.7.6 Mutual Exclusivity and Bimodality
			1.2.7.7 Rationale for Further Development
	1.3	E-cad	herin as a Synthetic Lethal Target
		1.3.1	The CDH1 gene and its Biological Functions
			1.3.1.1 Cytoskeleton
			1.3.1.2 Extracellular and Tumour Micro-environment 3
			1.3.1.3 Cell-Cell Adhesion and Signalling
		1.3.2	CDH1 as a Tumour (and Invasion) Suppressor
			1.3.2.1 Breast Cancers and Invasion
		1.3.3	Hereditary Diffuse Gastric (and Lobular Breast) Cancer 3
		1.3.4	Cell Line Models of <i>CDH1</i> Null Mutations
	1.4	Summ	ary and Research Direction of Thesis
		1.4.1	Thesis Aims
2	Met	thods a	and Resources 4
	2.1	Bioinf	ormatics Resources for Genomics Research
		2.1.1	Public Data and Software Packages
			2.1.1.1 Cancer Genome Atlas Data
			2.1.1.2 Reactome and Annotation Data 4
	2.2	Data 1	Handling
		2.2.1	Normalisation
		2.2.2	Sample Triage
		2.2.3	Metagenes and the Singular Value Decomposition
		2.2.4	Candidate Triage and Integration with Screen Data 4
	2.3	Techn	iques
		2.3.1	Statistical Procedures and Tests
		2.3.2	Gene Set Over-representation Analysis
		2.3.3	Clustering
		2.3.4	Heatmap
		2.3.5	Modelling and Simulations
			2.3.5.1 Receiver Operating Characteristic Curves 5
		2.3.6	Resampling Analysis
	2.4	Pathw	ray Structure Methods
		2.4.1	Network and Graph Analysis
		2.4.2	Sourcing Graph Structure Data
		2.4.3	Constructing Pathway Subgraphs
		2.4.4	Network Analysis Metrics
	2.5	Imple	mentation
		2.5.1	Computational Resources and Linux Utilities
		2.5.2	R Language and Packages
		2.5.3	High Performance and Parallel Computing
3	Met		Developed During Thesis 6
	3.1		thetic Lethal Detection Methodology 6
	3.2	Synthe	etic Lethal Simulation and Modelling 6
		3.2.1	A Model of Synthetic Lethality in Expression Data 6

		3.2.2 Simulation Procedure	67
	3.3	Detecting Simulated Synthetic Lethal Partners	70
		3.3.1 Binomial Simulation of Synthetic Lethality	70
		3.3.2 Multivariate Normal Simulation of Synthetic Lethality	72
		3.3.2.1 Multivariate Normal Simulation with Correlated Genes	74
		3.3.2.2 Specificity with Query-Correlated Pathways	82
	3.4	Graph Structure Methods	84
		3.4.1 Upstream and Downstream Gene Detection	84
		3.4.1.1 Permutation Analysis for Statistical Significance	85
		3.4.2 Simulating Gene Expression from Graph Structures	86
	3.5	Customised Functions and Packages Developed	90
		3.5.1 Synthetic Lethal Interaction Prediction Tool	90
		3.5.2 Data Visualisation	91
		3.5.3 Extensions to the iGraph Package	92
		3.5.3.1 Sampling Simulated Data from Graph Structures	92
		3.5.3.2 Plotting Directed Graph Structures	92
		3.5.3.3 Computing Information Centrality	94
		3.5.3.4 Testing Pathway Structure with Permutation Testing .	94
		3.5.3.5 Metapackage to Install iGraph Functions	95
4	Syn	hetic Lethal Analysis of Gene Expression Data	96
•	4.1	Synthetic Lethal Genes in Breast Cancer	97
	1.1	4.1.1 Synthetic Lethal Pathways in Breast Cancer	98
		· · · · · · · · · · · · · · · · · · ·	100
		· ·	103
	4.2		105
	1.2		105
		· · · · · · · · · · · · · · · · · · ·	105
			108
			110
		4.2.5 Comparison to Primary Screen at Pathway Level	
			113
			$\frac{118}{118}$
	4.3	v v	119
	4.4		121
	4.5	1	122
			122
		9	123
		v v	125
		•	125
		<u> </u>	$\frac{126}{126}$
	4.6	•	$\frac{126}{126}$

5	Syn	thetic Lethal Pathway Structure 12	28
	5.1		28
			29
			31
			34
			34
	5.2		36
		v v	37
			38
			38
			40
	5.3		41
			42
		U I	44
	5.4	- · · · · · · · · · · · · · · · · · · ·	46
	5.5		48
		· ·	
6		8	49
	6.1	V	50
		, v	51
			54
		v e	56
		V	57
		Ų 1	58
	6.2	<u>-</u>	59
		*	60
		1 1	60
		*	63
			65
		V I	71
		$oldsymbol{arphi}$	74
	6.3	Simulations in More Complex Graph Structures	79
		6.3.1 Simulations over Pathway-based Graphs	80
		6.3.2 Pathway Structures in a Large Simulated Datasets	83
	6.4	Discussion	86
		6.4.1 Simulation Procedure	86
		6.4.2 Comparing Methods with Simulated Data	87
		6.4.3 Design and Performance of SLIPT	88
		6.4.4 Simulations from Graph Structures	90
	6.5	Summary	91
7	D!:		ഹ
7			92
	7.1	Ų SV	92
			93
	7.2		93 94
	1.7.	appuncance .	94

	7.2.1 Synthetic Lethality in the Genomic Era	196 197
	Bibliography	201
A	Sample Quality A.1 Sample Correlation	
В	Software Used for Thesis	232
C	Mutation Analysis in Breast Cancer C.1 Synthetic Lethal Genes and Pathways	242 245 247
D	Metagene AnalysisD.1 Pathway Signature Expression	
\mathbf{E}	Intrinsic Subtyping	256
F	Stomach Expression AnalysisF.1 Synthetic Lethal Genes and PathwaysF.2 Comparison to Primary ScreenF.2.1 Resampling AnalysisF.3 Metagene Analysis	262 264
\mathbf{G}	Synthetic Lethal Genes in Pathways	267
Н	Network Analysis for Mutation SLIPT	274
Ι	Pathway Structure for Mutation SLIPT	277
J	Performance of SLIPT and χ^2 J.1 Correlated Query Genes affects Specificity	279 285
K	Simulations on Graph Structures K.0.1 Simulations from Inhibiting Graph Structures K.1 Simulation across Graph Structures K.2 Simulations from Complex Graph Structures K.2.1 Simulations from Complex Inhibiting Graphs K.3 Simulations from Pathway Graph Structures	295 299 302

List of Tables

1.1	Methods for predicting genetic interactions	23
1.2	Methods for predicting synthetic lethality in cancer	24
1.3	Methods used by Wu et al. (2014)	25
2.1	Excluded samples by batch and clinical characteristics	44
2.2	Computers used during thesis	54
2.3	Linux utilities and applications used during thesis	55
2.4	R installations used during thesis	56
2.5	R Packages used during thesis	56
2.6	R packages developed during thesis	58
4.1	Candidate synthetic lethal gene partners of <i>CDH1</i> from SLIPT	98
4.2	Pathways for <i>CDH1</i> partners from SLIPT	99
4.3	Pathways for clusters of <i>CDH1</i> partners from SLIPT	104
4.4	ANOVA for synthetic lethality and correlation with <i>CDH1</i>	107
4.5	Comparison of Synthetic Lethal Interaction Prediction Tool (SLIPT)	
	genes against secondary short interfering RNA (siRNA) screen	111
4.6	Pathways for <i>CDH1</i> partners from SLIPT and siRNA	112
4.7	Pathways for <i>CDH1</i> partners from SLIPT	115
4.8	Pathways for $CDH1$ partners from SLIPT and siRNA primary screen .	116
4.9	Examples of candidate metagenes synthetic lethal for $CDH1$ from SLIPT	120
5.1	ANOVA for synthetic lethality and vertex degree	138
5.2	ANOVA for synthetic lethality and information centrality	139
5.3	ANOVA for synthetic lethality and PageRank centrality	140
5.4	Resampling for pathway structure of synthetic lethal detection methods	145
B.1	Complete list of R packages used during this thesis	232
C.1	Candidate synthetic lethal gene partners of <i>CDH1</i> from mtSLIPT	241
C.2	Pathways for <i>CDH1</i> partners from mtSLIPT	242
C.3	Pathways for clusters of <i>CDH1</i> partners from mtSLIPT	244
C.4	Pathways for <i>CDH1</i> partners from mtSLIPT and siRNA	246
C.5	Pathways for <i>CDH1</i> partners from mtSLIPT	247
C.6	Pathways for $\mathit{CDH1}$ partners from mtSLIPT and siRNA primary screen	248
D.1	Candidate synthetic lethal metagenes against CDH1 from mtSLIPT	255

E.1	Comparison of intrinsic subtypes	256
F.1	Synthetic lethal gene partners of <i>CDH1</i> from SLIPT in stomach cancer	258
F.2	Pathways for <i>CDH1</i> partners from SLIPT in stomach cancer	259
F.3	Pathways for clusters of <i>CDH1</i> partners in stomach SLIPT	261
F.4	Pathways for <i>CDH1</i> partners from SLIPT and siRNA	263
F.5	Pathways for <i>CDH1</i> partners from SLIPT in stomach cancer	264
F.6	Pathways for $CDH1$ partners from SLIPT in stomach and siRNA	265
F.7	Synthetic lethal metagenes against <i>CDH1</i> in stomach cancer	266
H.1	ANOVA for synthetic lethality and vertex degree	276
H.2	ANOVA for synthetic lethality and information centrality	276
H.3	ANOVA for synthetic lethality and PageRank centrality	276
I.1	Resampling for pathway structure of synthetic lethal detection methods	278

List of Figures

1.1 1.2	v C	14 17
2.1 2.2	V	45 45
3.1	Framework for synthetic lethal prediction	61
3.2	Synthetic lethal prediction adapted for mutation	62
3.3	A model of synthetic lethal gene expression	64
3.4	Modelling synthetic lethal gene expression	65
3.5	Synthetic lethality with multiple genes	66
3.6	Simulating gene function	68
3.7	Simulating synthetic lethal gene function	68
3.8		69
3.9	Performance of binomial simulations	71
3.10		71
3.11	Performance of multivariate normal simulations	73
3.12	Simulating expression with correlated gene blocks	75
3.13	Simulating expression with correlated gene blocks	76
3.14	Synthetic lethal prediction across simulations	78
3.15	Performance with correlations	79
3.16	Comparison of statistical performance with correlation structure	80
3.17	Performance with query correlations	81
		82
3.19	Performance of directional criteria	83
3.20	Simulated graph structures	87
3.21	Simulating expression from a graph structure	88
3.22	Simulating expression from graph structure with inhibitions	89
3.23	Demonstration of violin plots with custom features	93
3.24	Demonstration of annotated heatmap	93
3.25	Simulating graph structures	94
4.1		01
4.2	•	06
4.3	1	06
4.4		08
4.5	Comparison of SLIPT and siRNA genes with screen viability	09

4.6 4.7	Comparison of SLIPT genes with siRNA screen viability Resampled intersection of SLIPT and siRNA candidate genes	109 114
5.1 5.2 5.3 5.4 5.5	Synthetic lethality in the PI3K cascade Synthetic lethality in Elastic Fibre Formation Synthetic lethality in Fibrin Clot Formation Synthetic lethality in the GPCRs Synthetic lethality and vertex degree	130 132 133 135 137
5.6 5.7 5.8	Synthetic lethality and centrality	139 141 143
6.1 6.2 6.3 6.4	Performance of χ^2 and SLIPT across quantiles	152 153 154 155
6.5 6.6 6.7 6.8	Performance of negative correlation and SLIPT	158 161 162 163
$6.11 \\ 6.12$	Performance of simulations on a pathway	164 166 168 169 170
6.14 6.15 6.16	Detection of synthetic lethality within a graph structure	170 172 176 177 178
6.18 6.19	Performance of simulations on the PI3K cascade	182 184 185
A.1 A.2 A.3 A.4 A.5	Correlation profiles of removed samples	226 227 228 229 230
C.1 C.2 C.3 C.4 C.5	Synthetic lethal expression profiles of analysed samples	243 245 249 249 250
D.1	Pathway metagene expression profiles	253

D.2	Expression profiles for estrogen receptor related genes	254
F.1 F.2	Synthetic lethal expression profiles of stomach samples	260 262
G.1 G.2 G.3 G.4 G.5 G.6 G.7	Synthetic lethality in the PI3K/AKT pathway	267 268 269 270 271 272 273
H.1 H.2 H.3	Synthetic lethality and vertex degree	274 275 275
I.1	Structure of synthetic lethality resampling	277
J.1 J.2 J.3 J.4 J.5 J.6	Performance of χ^2 and SLIPT across quantiles	279 281 283 285 287
K.1 K.2 K.3	Performance of simulations on a simple graph	291 292 293
	Performance of simulations on a constructed graph with inhibition Detection of synthetic lethality within a graph structure Detection of synthetic lethality within an inhibiting graph	294 295 297
K.7	Detection of synthetic lethality within an inhibiting graph	298 299
K.9	Performance of simulations on a large graph	300 301
	Performance of simulations on a branching graph with inhibition Performance of simulations on a branching graph with inhibition	302 303
K.14	Performance of simulations on a complex graph with inhibition Performance of simulations on a complex graph with inhibition	304 305
K.16	Performance of simulations on a large constructed graph with inhibition Performance of simulations on a large constructed graph with inhibition Performance of simulations on the $G_{\alpha i}$ signalling pathway	306 307 308
	Performance of simulations on the $G_{\alpha i}$ signalling pathway	309

Glossary

allele A gene variant with a specific sequence and

phenotype.

bioinformatics Statistical or computational approaches to

biological data or research tools.

bisulfite-Seq Epigenomic data from sequencing bisulfite

treated DNA.

CAGE-Seq Transcriptome data from cap analysis of gene

expression.

cancer A class of diseases, formally "malignant neo-

plasm", of abnormal cellular growth and

spread to other organs.

cancer gene A gene which is involved in the malignancy

of some cancers, encompassing oncogenes and tumour suppressors, which have molecular aberrations in cancer or variants which predis-

pose individuals to cancer.

centrality A network metric which identifies important

vertices.

chemoprevention The use of drugs to prevent early-stage can-

cers, generally applied to high-risk mutation

carriers.

chemotherapy The use of cytotoxic drugs to treat cancers, in

combinations, generally applied to advanced

stage cancers.

ChIP-Seq Epigenome data from chromatin immuno-

preciptation sequencing.

compound screen A high-throughput screen performed using a

library of chemical compounds.

computational biology Applying computational or mathematical

modelling to understanding biological systems

and relationships.

conditional essentiality A gene becoming essential to viability un-

der certain environmental conditions, including presence of compounds which inactivate

other genes.

copy number The number of copies of DNA, typically two

copies for diploid organisms but subject to

variation.

de novo A bioinformatics sequence assembly conduc-

ted entirely from raw genomics data without

a reference sequence.

diagnosis The identification of disease by clinical, cellu-

lar, and molecular characteristics.

driver mutation A mutation which promotes cancer growth.

E-cadherin Epithelial cadherin (calcium-dependent ad-

hesion), a cell-adhesion protein encoded by

CDH1.

edge or link A relationship connecting a pair of elements of

a graph structure or network, may be weighted

or directional.

epigenome An analysis of epigenetic modifications of all

genes in the genome.

epistasis (biological) The effects of a gene modifying or masking the

phenotype of another gene.

epistasis (statistical) A divergence of the observed double mutant

phenotype from that expected based on the respective phenotypes of single mutant (Fisher,

1919).

essential A gene which is required to be functional or

expressed for a cell or organism to be viable,

grow or develop.

exome A sequencing approach designed to generate

data enriched for coding genes within the gen-

ome.

familial A trait recurrently occurring in families, not

necessarily with a genetic cause.

functional redundancy Genes which perform a common function, also

known as genetic redundancy.

gene expression A measure of the relative expression of each

gene from the mRNA extracted from (pooled)

cells.

genetic robustness A system of biological pathways which (has

evolved to) continue to function as a whole under various conditions, including the inac-

tivation of various individual genes.

genome All of the DNA sequence in the genome.

genomic The use of data from all genes in the genome.

The use of genomic information to tailor medi-

cine treatment to the genetics of an individual.

germline mutation A mutation that occurred in germline cells and

is passed between generation.

graph or network A mathematical structure modelling or depict-

ing the relationships between elements.

hallmark of cancer An underlying characteristic of cancer as part

of a rational approach devised by (Hanahan

and Weinberg, 2000).

hereditary A trait or disease which has a genetic cause

and is inherited from family members.

high-throughput screen An experimental procedure to perform a large

scale series of chemical, genetic, or pharmaco-

logical tests.

hub A central or highly connected component of a

network.

in silico An investigation conducted using computa-

tions, typically simulations or analyses.

in vitro An investigation conducted using a controlled

experimental system to examine biomolecules.

in vivo An investigation conducted using in the con-

text of a biological cell or organism, including

pre-clincal models and clinical trials.

induced essentiality A gene becoming essential to viability under

certain conditions, including inactivation of a

synthetic lethal partner.

information centrality A network centrality metric which uses the im-

pact of removing a vertex or node on connec-

tions in the network.

intrinsic subtype Distinguishing cancer by molecular and ge-

netic features.

MCF10A cell line A non-tumorigenic epithelial cell line derived

from breast tissue.

metabolome All the metabolites and enzymes in the cell. metagene A consistent signal of expression for a collec-

tion of genes such as a biological pathway, de-

rived from singular value decomposition.

All of the genes and genomes in a community. metagenome A secondary growth of a tumour or spread of metastasis

cancer to other organs.

methylation A measure of the epigenetic regulation of DNA

at CpG dinucleotide (CpG) sites.

microarray A high-throughput technique to measure pres-

ence or abundance of nucleic acid sequences

from binding to probes.

Short RNA molecules generally regarded to microRNA

regulate gene expression by binding to mRNA.

molecular profile A combination of genetic and biochemical

measures which identifies characteristic traits

of a tumour.

A classification of cancers based on an identimolecular subtype

fication using molecular properties.

mutant A variant or dysfunctional phenotype arising

from a mutation in a gene.

mutation A change in DNA sequence that disrupts gene

function.

network biology The application mathematical and computa-

tional approaches to networks in understand-

ing biological relationships.

network medicine The use of network biology to understand, pre-

vent, or treat diseases.

non-oncogene addiction The dependence of a cancer cell on functioning

non-mutant genes.

'omics A combination of approaches to generating

> biological data with high-throughput procedures such as genomics, proteomics or meta-

bolomics.

oncogene A gene that potentially causes cancer, typic-

ally by over-expression or mutant gene vari-

ants.

oncogene addiction The dependence of a cancer cell on a specific

oncogenic pathway.

PageRank centrality A network centrality metric which uses eigen-

vectors with a scaling factor (Brin and Page,

1998).

pan cancer A focus on the molecular and genetic features

across cancers in different tissues.

passenger mutation A mutation that occurs in cancers but does

not affect the growth of cancers.

pathway A series of biomolecules that produces a par-

ticular product or biological function.

pleiotropy When a gene has multiple biological functions. polypharmacology The design of drugs to target multiple molecu-

lar targets or biological pathways.

precision medicine The application of prevention and treatment

measures to target diseases by molecular and

genetic features.

prognosis The estimation of disease progression and pa-

tient outcome.

proteome All the proteins expressed from the genome.

proto-oncogene The non-mutant variant or precursor to a

mutant oncogene.

recurrent mutation The repeated occurrence of mutations in a

particular gene across cancers.

RNAi screen A high-throughput screen performed using a

RNA interference (RNAi).

RNA-Seq The generation of transcriptome data from se-

quencing RNA.

Sanger sequencing A dideoxy chain termination method for DNA

sequencing (named after Fred Sanger).

scale-free A property of a network which has a power

law vertex degree distribution, that is several highly connected hub genes and many with

very few connections.

shortest path A path with the fewest possible edges which

connects two particular vertices.

small world A property of a network which is highly

connected and has a low characteristic path length, derived from the mean shortest path

length across all pairs of nodes.

somatic mutation A mutation that occurs in somatic cells, dur-

ing a patient's lifespan.

sporadic cancer Cancers which do occur in patients with a fam-

ily history or carry a high-risk genetic variant. When multiple drugs have more effect than

expected from the effect of each separately.

synthetic dosage lethal A synthetic genetic interaction (SGI) ana-

logous to synthetic lethality where where one gene is inactivated and the other over-

expressed.

synergy

synthetic lethal Genetic interactions where inactivation of

multiple genes is inviable (or deleterious) which are viable if inactivated separately.

synthetic lethal screen A high-throughput screen performed on iso-

genic cell lines to detect genes for which inhibition specifically deleterious to the null mutant

genotype.

synthetic rescue A synthetic genetic interaction when the com-

bined mutations restores the wild-type the

phenotype of one of the mutations.

synthetic sick Genetic interactions where inactivation of

multiple genes is deleterious which are viable

if inactivated separately.

synthetic suppression A synthetic genetic interaction when the com-

bined mutations (partially) suppresses the mutant phenotype of one of the mutations.

targeted therapy Cancer treatment that specifically acts against

a molecular target, in contrast to standard

chemotherapy.

transcriptome All of the genes expressed in the genome.

treatment Medical procedures for a disease to improve

patient outcomes.

tumour An abnormal lump of tissue or growth of cells,

may be cancerous.

tumour suppressor A gene potentially causes cancer, typically by

disruption of functions which protect the cell

from cancer.

vertex degree A network metric of connectivity of vertices

which uses the number of edges connected to

each vertex or node.

vertex or node An element of a graph structure or network.

wild-type A natural phenotype of a trait or the normally

functional allele which encodes it.

Acronyms

1KGP 1000 genomes project.

ADP Adenosine Diphosphate.

AMP Adenosine Monophosphate.

AMPK AMP-activated Protein Kinase.

ANOVA Analysis of Variance. ATP Adenosine Triphosphate.

AUROC Area Under the Receiver Operating Charac-

teristic (curve).

Bash Bourne Again Shell.

BioPAX Biological Pathway Exchange.BiSEp Bimodal Subsetting Expression.BMP Bone Morphogenic Protein.

cAMP Cylic AMP.

CCLE Cancer Cell Line Encyclopaedia. cDNA Complementary DNA (from mRNA).

CGP Cancer Genome Project.

ChIP Chromatin Immunopreciptation.

ChIP-Seq Chromatin Immunopreciptation Sequencing.

CNV Copy Number Variation.

COSMIC Catalogue Of Somatic Mutations In Cancer.

CpG 5'-C-phosphate-G-3'.

CPM Counts Per Million mapped reads.

CPU Central Processing Unit.

CRAN comprehensive R archive network.

CXCR Chemokine Receptor.

DAISY Data Mining Synthetic Lethal Identification

Pipeline.

DDBJ DNA Data Bank of Japan.
DNA Deoxyribonucleic Acid.

EMBL European Molecular Biology Laboratory.

EMT Epithelial-Mesenchymal Transition.
ENA The European Nucleotide Archive.
ENCODE Encyclopaedia of DNA Elements.

ER Estrogen Receptor.

exprSL Synthetic Lethality (expression).

FANTOM Functional Annotation Of Mammalian gen-

ome.

FDR False Discovery Rate.

GEO Gene Expression Omnibus.

GO Gene Ontology.

GPCR G Crotein Coupled Receptor.

HDAC Histone Deacetylase.

HDGC Hereditary Diffuse Gastric Cancer.

HLRCC Hereditary Leiomyomatosis and Renal Cell

Carcinoma.

HPC High Performance Computing.

ICGC International Cancer Genome Consortium.

IHC Immunohistochemistry.

InDel Insertion or Deletion (in DNA sequence).

JAK Janus Kinase.

lncRNA Long Non-Coding RNA.

METABRIC Molecular Taxonomy of Breast Cancer Inter-

national Consortium.

microRNA Micro RNA. mRNA Messenger RNA.

MSI Microsatellite Instability. mtSL synthetic Lethality (mutation).

mtSLIPT Synthetic Lethal Interaction Prediction Tool

(against mutation).

NCBI National Center for Biotechnology Informa-

tion (in the USA).

NCI National Cancer Institute (in the USA). NeSI New Zealand eScience Infrastructure. NGS Next-Generation Sequencing.

NHGRI National Human Genome Research Institute

(in the USA).

NIG National Institute of Genetics (in Japan).

NIH National Institutes of Health (in the USA).

NMD Nonsense-Mediated Decay.

PAM50 Prediction Analysis of Microarray 50.

PARP Poly-ADP-Ribose Polymerase.

PCR Polymerase Chain Reaction.

PDE Phosphodiesterase.

PI3K Phosphoinositide 3-kinase.

PIP₂ Phosphatidylinositol-(4,5)-bisphosphate. PIP₃ Phosphatidylinositol-(3,4,5)-trisphosphate.

PPI Protein-Protein Interaction.

PR Progesterone Receptor.

qPCR Quantitative (real-time) Polymerase Chain

Reaction.

RFLP Restriction Fragment Length Polymorphism.

RGS G-protein Signalling.

RHO Ras Homolog Family.

RMA Robust Multiarray Averaging (normalisation.

RNA Ribonucleic Acid.

RNAi RNA Interference.

ROC Reciever Operating Characteristic (curve).

RPKM Reads Per Kilobase per Million mapped reads.

RPPA Reverse Phase Protein Arrays.

RRBS Reduced Representation Bisulfite Sequencing.

rRNA Ribonucleic acid.

RSEM RNA-Seq by Expectation Maximization (nor-

malisation.

SGA Synthetic Gene Array (technique).

SGI Synthetic Genetic Interaction.

shRNA Short Hairpin RNA.

siRNA Short Interfering RNA.

SL Synthetic Lethal.

SLIPT Synthetic Lethal Interaction Prediction Tool.

Slurm Simple Linux Utility for Resource Manage-

ment.

SNP Single Nucleotide Polymorphism.

SOCKS Socket Secure.

SR Synthetic Rescue (or viability).

SS Synthetic Suppression.

SSL Synthetic Sick.

TCGA The Cancer Genome Atlas (genomics project).

TGF α Transforming Growth Factor α .

TMM Trimmed Mean of M values (normalisation.

tRNA Transfer RNA.

UCSC University of California, Santa Cruz. UTR Untranslated Region (of mRNA).

WNT Wingless-Related Integration Site.

Bibliography

- Aarts, M., Bajrami, I., Herrera-Abreu, M.T., Elliott, R., Brough, R., Ashworth, A., Lord, C.J., and Turner, N.C. (2015) Functional genetic screen identifies increased sensitivity to weel inhibition in cells with defects in fanconi anemia and hr pathways. Mol Cancer Ther, 14(4): 865–76.
- Abeshouse, A., Ahn, J., Akbani, R., Ally, A., Amin, S., Andry, C.D., Annala, M., Aprikian, A., Armenia, J., Arora, A., et al. (2015) The Molecular Taxonomy of Primary Prostate Cancer. Cell, 163(4): 1011–1025.
- Adler, D. (2005) vioplot: Violin plot. R package version 0.2.
- Akbani, R., Akdemir, K.C., Aksoy, B.A., Albert, M., Ally, A., Amin, S.B., Arachchi, H., Arora, A., Auman, J.T., Ayala, B., et al. (2015) Genomic Classification of Cutaneous Melanoma. Cell, 161(7): 1681–1696.
- Akobeng, A.K. (2007) Understanding diagnostic tests 3: receiver operating characteristic curves. *Acta Pædiatrica*, **96**(5): 644–647.
- American Cancer Society (2017) Genetics and cancer. https://www.cancer.org/cancer/cancer-causes/genetics.html. Accessed: 22/03/2017.
- Anjomshoaa, A., Lin, Y.H., Black, M.A., McCall, J.L., Humar, B., Song, S., Fukuzawa, R., Yoon, H.S., Holzmann, B., Friederichs, J., et al. (2008) Reduced expression of a gene proliferation signature is associated with enhanced malignancy in colon cancer. Br J Cancer, 99(6): 966–973.
- Araki, H., Knapp, C., Tsai, P., and Print, C. (2012) GeneSetDB: A comprehensive meta-database, statistical and visualisation framework for gene set analysis. *FEBS Open Bio*, **2**: 76–82.
- Ashburner, M., Ball, C.A., Blake, J.A., Botstein, D., Butler, H., Cherry, J.M., Davis, A.P., Dolinski, K., Dwight, S.S., Eppig, J.T., et al. (2000) Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. Nat Genet, 25(1): 25–29.
- Ashworth, A. (2008) A synthetic lethal therapeutic approach: poly(adp) ribose polymerase inhibitors for the treatment of cancers deficient in dna double-strand break repair. J Clin Oncol, 26(22): 3785–90.
- Ashworth, A., Lord, C.J., and Reis-Filho, J.S. (2011) Genetic interactions in cancer progression and treatment. *Cell*, **145**(1): 30–38.

- Audeh, M.W., Carmichael, J., Penson, R.T., Friedlander, M., Powell, B., Bell-McGuinn, K.M., Scott, C., Weitzel, J.N., Oaknin, A., Loman, N., et al. (2010) Oral poly(adp-ribose) polymerase inhibitor olaparib in patients with *BRCA1* or *BRCA2* mutations and recurrent ovarian cancer: a proof-of-concept trial. *Lancet*, **376**(9737): 245–51.
- Babyak, M.A. (2004) What you see may not be what you get: a brief, nontechnical introduction to overfitting in regression-type models. *Psychosom Med*, **66**(3): 411–21.
- Bamford, S., Dawson, E., Forbes, S., Clements, J., Pettett, R., Dogan, A., Flanagan, A., Teague, J., Futreal, P.A., Stratton, M.R., et al. (2004) The COSMIC (Catalogue of Somatic Mutations in Cancer) database and website. Br J Cancer, 91(2): 355–358.
- Barabási, A.L. and Albert, R. (1999) Emergence of scaling in random networks. *Science*, **286**(5439): 509–12.
- Barabási, A.L., Gulbahce, N., and Loscalzo, J. (2011) Network medicine: a network-based approach to human disease. *Nat Rev Genet*, **12**(1): 56–68.
- Barabási, A.L. and Oltvai, Z.N. (2004) Network biology: understanding the cell's functional organization. *Nat Rev Genet*, **5**(2): 101–13.
- Barrat, A. and Weigt, M. (2000) On the properties of small-world network models. The European Physical Journal B - Condensed Matter and Complex Systems, 13(3): 547–560.
- Barretina, J., Caponigro, G., Stransky, N., Venkatesan, K., Margolin, A.A., Kim, S., Wilson, C.J., Lehar, J., Kryukov, G.V., Sonkin, D., et al. (2012) The Cancer Cell Line Encyclopedia enables predictive modelling of anticancer drug sensitivity. Nature, 483(7391): 603–607.
- Barry, W.T. (2016) safe: Significance Analysis of Function and Expression. R package version 3.14.0.
- Baryshnikova, A., Costanzo, M., Dixon, S., Vizeacoumar, F.J., Myers, C.L., Andrews, B., and Boone, C. (2010a) Synthetic genetic array (sga) analysis in saccharomyces cerevisiae and schizosaccharomyces pombe. *Methods Enzymol*, **470**: 145–79.
- Baryshnikova, A., Costanzo, M., Kim, Y., Ding, H., Koh, J., Toufighi, K., Youn, J.Y., Ou, J., San Luis, B.J., Bandyopadhyay, S., et al. (2010b) Quantitative analysis of fitness and genetic interactions in yeast on a genome scale. Nat Meth, 7(12): 1017–1024.
- Bass, A.J., Thorsson, V., Shmulevich, I., Reynolds, S.M., Miller, M., Bernard, B., Hinoue, T., Laird, P.W., Curtis, C., Shen, H., et al. (2014) Comprehensive molecular characterization of gastric adenocarcinoma. Nature, **513**(7517): 202–209.

- Bates, D. and Maechler, M. (2016) *Matrix: Sparse and Dense Matrix Classes and Methods*. R package version 1.2-7.1.
- Bateson, W. and Mendel, G. (1909) Mendel's principles of heredity, by W. Bateson. University Press, Cambridge [Eng.].
- Becker, K.F., Atkinson, M.J., Reich, U., Becker, I., Nekarda, H., Siewert, J.R., and Höfler, H. (1994) E-cadherin gene mutations provide clues to diffuse type gastric carcinomas. *Cancer Research*, **54**(14): 3845–3852.
- Bell, D., Berchuck, A., Birrer, M., Chien, J., Cramer, D., Dao, F., Dhir, R., DiSaia, P., Gabra, H., Glenn, P., et al. (2011) Integrated genomic analyses of ovarian carcinoma. Nature, 474(7353): 609–615.
- Benjamini, Y. and Hochberg, Y. (1995) Controlling the false discovery rate: A practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society Series B (Methodological)*, **57**(1): 289–300.
- Berx, G., Cleton-Jansen, A.M., Nollet, F., de Leeuw, W.J., van de Vijver, M., Cornelisse, C., and van Roy, F. (1995) E-cadherin is a tumour/invasion suppressor gene mutated in human lobular breast cancers. *EMBO J*, **14**(24): 6107–15.
- Berx, G., Cleton-Jansen, A.M., Strumane, K., de Leeuw, W.J., Nollet, F., van Roy, F., and Cornelisse, C. (1996) E-cadherin is inactivated in a majority of invasive human lobular breast cancers by truncation mutations throughout its extracellular domain. *Oncogene*, **13**(9): 1919–25.
- Berx, G. and van Roy, F. (2009) Involvement of members of the cadherin superfamily in cancer. *Cold Spring Harb Perspect Biol*, **1**: a003129.
- Bitler, B.G., Aird, K.M., Garipov, A., Li, H., Amatangelo, M., Kossenkov, A.V., Schultz, D.C., Liu, Q., Shih Ie, M., Conejo-Garcia, J.R., et al. (2015) Synthetic lethality by targeting ezh2 methyltransferase activity in arid1a-mutated cancers. Nat Med, 21(3): 231–8.
- Blake, J.A., Christie, K.R., Dolan, M.E., Drabkin, H.J., Hill, D.P., Ni, L., Sitnikov, D., Burgess, S., Buza, T., Gresham, C., et al. (2015) Gene Ontology Consortium: going forward. *Nucleic Acids Res*, **43**(Database issue): D1049–1056.
- Boettcher, M., Lawson, A., Ladenburger, V., Fredebohm, J., Wolf, J., Hoheisel, J.D., Frezza, C., and Shlomi, T. (2014) High throughput synthetic lethality screen reveals a tumorigenic role of adenylate cyclase in fumarate hydratase-deficient cancer cells. *BMC Genomics*, **15**: 158.
- Boone, C., Bussey, H., and Andrews, B.J. (2007) Exploring genetic interactions and networks with yeast. *Nat Rev Genet*, 8(6): 437–49.
- Borgatti, S.P. (2005) Centrality and network flow. Social Networks, 27(1): 55 71.

- Boucher, B. and Jenna, S. (2013) Genetic interaction networks: better understand to better predict. *Front Genet*, 4: 290.
- Bozovic-Spasojevic, I., Azambuja, E., McCaskill-Stevens, W., Dinh, P., and Cardoso, F. (2012) Chemoprevention for breast cancer. *Cancer treatment reviews*, **38**(5): 329–339.
- Breiman, L. (2001) Random forests. Machine Learning, 45(1): 5–32.
- Brin, S. and Page, L. (1998) The anatomy of a large-scale hypertextual web search engine. Computer Networks and ISDN Systems, 30(1): 107 117.
- Brouxhon, S.M., Kyrkanides, S., Teng, X., Athar, M., Ghazizadeh, S., Simon, M., O'Banion, M.K., and Ma, L. (2014) Soluble E-cadherin: a critical oncogene modulating receptor tyrosine kinases, MAPK and PI3K/Akt/mTOR signaling. *Oncogene*, **33**(2): 225–235.
- Brückner, A., Polge, C., Lentze, N., Auerbach, D., and Schlattner, U. (2009) Yeast two-hybrid, a powerful tool for systems biology. *Int J Mol Sci*, **10**(6): 2763–2788.
- Bryant, H.E., Schultz, N., Thomas, H.D., Parker, K.M., Flower, D., Lopez, E., Kyle, S., Meuth, M., Curtin, N.J., and Helleday, T. (2005) Specific killing of *BRCA2*-deficient tumours with inhibitors of poly*adpribose* polymerase. *Nature*, **434**(7035): 913–7.
- Bussey, H., Andrews, B., and Boone, C. (2006) From worm genetic networks to complex human diseases. *Nat Genet*, **38**(8): 862–3.
- Butland, G., Babu, M., Diaz-Mejia, J.J., Bohdana, F., Phanse, S., Gold, B., Yang, W., Li, J., Gagarinova, A.G., Pogoutse, O., et al. (2008) esga: E. coli synthetic genetic array analysis. Nat Methods, 5(9): 789–95.
- cBioPortal for Cancer Genomics (cBioPortal) (2017) cBioPortal for Cancer Genomics. http://www.cbioportal.org/. Accessed: 26/03/2017.
- Cerami, E.G., Gross, B.E., Demir, E., Rodchenkov, I., Babur, O., Anwar, N., Schultz, N., Bader, G.D., and Sander, C. (2011) Pathway Commons, a web resource for biological pathway data. *Nucleic Acids Res*, **39**(Database issue): D685–690.
- Chen, A., Beetham, H., Black, M.A., Priya, R., Telford, B.J., Guest, J., Wiggins, G.A.R., Godwin, T.D., Yap, A.S., and Guilford, P.J. (2014) E-cadherin loss alters cytoskeletal organization and adhesion in non-malignant breast cells but is insufficient to induce an epithelial-mesenchymal transition. *BMC Cancer*, **14**(1): 552.
- Chen, S. and Parmigiani, G. (2007) Meta-analysis of BRCA1 and BRCA2 penetrance. J Clin Oncol, 25(11): 1329–1333.
- Chipman, K. and Singh, A. (2009) Predicting genetic interactions with random walks on biological networks. BMC Bioinformatics, $\mathbf{10}(1)$: 17.

- Christofori, G. and Semb, H. (1999) The role of the cell-adhesion molecule E-cadherin as a tumour-suppressor gene. *Trends in Biochemical Sciences*, **24**(2): 73 76.
- Ciriello, G., Gatza, M.L., Beck, A.H., Wilkerson, M.D., Rhie, S.K., Pastore, A., Zhang, H., McLellan, M., Yau, C., Kandoth, C., et al. (2015) Comprehensive Molecular Portraits of Invasive Lobular Breast Cancer. Cell, 163(2): 506–519.
- Clark, M.J. (2004) Endogenous Regulator of G Protein Signaling Proteins Suppress G o-Dependent -Opioid Agonist-Mediated Adenylyl Cyclase Supersensitization. Journal of Pharmacology and Experimental Therapeutics, 310(1): 215–222.
- Collingridge, D.S. (2013) A primer on quantitized data analysis and permutation testing. *Journal of Mixed Methods Research*, **7**(1): 81–97.
- Collins, F.S. and Barker, A.D. (2007) Mapping the cancer genome. Pinpointing the genes involved in cancer will help chart a new course across the complex landscape of human malignancies. *Sci Am*, **296**(3): 50–57.
- Collisson, E., Campbell, J., Brooks, A., Berger, A., Lee, W., Chmielecki, J., Beer, D., Cope, L., Creighton, C., Danilova, L., et al. (2014) Comprehensive molecular profiling of lung adenocarcinoma. *Nature*, **511**(7511): 543–550.
- Costanzo, M., Baryshnikova, A., Bellay, J., Kim, Y., Spear, E.D., Sevier, C.S., Ding, H., Koh, J.L., Toufighi, K., Mostafavi, S., et al. (2010) The genetic landscape of a cell. Science, 327(5964): 425–31.
- Costanzo, M., Baryshnikova, A., Myers, C.L., Andrews, B., and Boone, C. (2011) Charting the genetic interaction map of a cell. *Curr Opin Biotechnol*, **22**(1): 66–74.
- Courtney, K.D., Corcoran, R.B., and Engelman, J.A. (2010) The PI3K pathway as drug target in human cancer. *J Clin Oncol*, **28**(6): 1075–1083.
- Creighton, C.J., Morgan, M., Gunaratne, P.H., Wheeler, D.A., Gibbs, R.A., Robertson, A., Chu, A., Beroukhim, R., Cibulskis, K., Signoretti, S., et al. (2013) Comprehensive molecular characterization of clear cell renal cell carcinoma. Nature, 499(7456): 43–49.
- Croft, D., Mundo, A.F., Haw, R., Milacic, M., Weiser, J., Wu, G., Caudy, M., Garapati, P., Gillespie, M., Kamdar, M.R., et al. (2014) The Reactome pathway knowledge-base. Nucleic Acids Res, 42(database issue): D472–D477.
- Crunkhorn, S. (2014) Cancer: Predicting synthetic lethal interactions. *Nat Rev Drug Discov*, **13**(11): 812.
- Csardi, G. and Nepusz, T. (2006) The igraph software package for complex network research. *InterJournal*, **Complex Systems**: 1695.
- Dai, X., Li, T., Bai, Z., Yang, Y., Liu, X., Zhan, J., and Shi, B. (2015) Breast cancer intrinsic subtype classification, clinical use and future trends. *Am J Cancer Res*, **5**(10): 2929–2943.

- Davierwala, A.P., Haynes, J., Li, Z., Brost, R.L., Robinson, M.D., Yu, L., Mnaimneh, S., Ding, H., Zhu, H., Chen, Y., et al. (2005) The synthetic genetic interaction spectrum of essential genes. Nat Genet, 37(10): 1147–1152.
- De Leeuw, W.J., Berx, G., Vos, C.B., Peterse, J.L., Van de Vijver, M.J., Litvinov, S., Van Roy, F., Cornelisse, C.J., and Cleton-Jansen, A.M. (1997) Simultaneous loss of E-cadherin and catenins in invasive lobular breast cancer and lobular carcinoma in situ. *J Pathol*, **183**(4): 404–11.
- De Santis, G., Miotti, S., Mazzi, M., Canevari, S., and Tomassetti, A. (2009) E-cadherin directly contributes to PI3K/AKT activation by engaging the PI3K-p85 regulatory subunit to adherens junctions of ovarian carcinoma cells. *Oncogene*, **28**(9): 1206–1217.
- Demir, E., Babur, O., Rodchenkov, I., Aksoy, B.A., Fukuda, K.I., Gross, B., Sumer, O.S., Bader, G.D., and Sander, C. (2013) Using biological pathway data with Paxtools. *PLoS Comput Biol*, **9**(9): e1003194.
- Deshpande, R., Asiedu, M.K., Klebig, M., Sutor, S., Kuzmin, E., Nelson, J., Piotrowski, J., Shin, S.H., Yoshida, M., Costanzo, M., et al. (2013) A comparative genomic approach for identifying synthetic lethal interactions in human cancer. Cancer Res, 73(20): 6128–36.
- Dickson, D. (1999) Wellcome funds cancer database. *Nature*, **401**(6755): 729.
- Dijkstra, E.W. (1959) A note on two problems in connexion with graphs. *Numerische Mathematik*, **1**(1): 269–271.
- Dixon, S.J., Andrews, B.J., and Boone, C. (2009) Exploring the conservation of synthetic lethal genetic interaction networks. *Commun Integr Biol*, **2**(2): 78–81.
- Dixon, S.J., Fedyshyn, Y., Koh, J.L., Prasad, T.S., Chahwan, C., Chua, G., Toufighi, K., Baryshnikova, A., Hayles, J., Hoe, K.L., et al. (2008) Significant conservation of synthetic lethal genetic interaction networks between distantly related eukaryotes. Proc Natl Acad Sci U S A, 105(43): 16653-8.
- Dong, L.L., Liu, L., Ma, C.H., Li, J.S., Du, C., Xu, S., Han, L.H., Li, L., and Wang, X.W. (2012) E-cadherin promotes proliferation of human ovarian cancer cells in vitro via activating MEK/ERK pathway. *Acta Pharmacol Sin*, **33**(6): 817–822.
- Dorsam, R.T. and Gutkind, J.S. (2007) G-protein-coupled receptors and cancer. *Nat Rev Cancer*, **7**(2): 79–94.
- Erdős, P. and Rényi, A. (1959) On random graphs I. Publ Math Debrecen, 6: 290–297.
- Erdős, P. and Rényi, A. (1960) On the evolution of random graphs. In *Publ. Math. Inst. Hung. Acad. Sci*, volume 5, 17–61.
- Eroles, P., Bosch, A., Perez-Fidalgo, J.A., and Lluch, A. (2012) Molecular biology in breast cancer: intrinsic subtypes and signaling pathways. *Cancer Treat Rev*, **38**(6): 698–707.

- Farmer, H., McCabe, N., Lord, C.J., Tutt, A.N., Johnson, D.A., Richardson, T.B., Santarosa, M., Dillon, K.J., Hickson, I., Knights, C., et al. (2005) Targeting the dna repair defect in BRCA mutant cells as a therapeutic strategy. Nature, 434(7035): 917–21.
- Fawcett, T. (2006) An introduction to ROC analysis. *Pattern Recognition Letters*, **27**(8): 861 874. {ROC} Analysis in Pattern Recognition.
- Fece de la Cruz, F., Gapp, B.V., and Nijman, S.M. (2015) Synthetic lethal vulnerabilities of cancer. *Annu Rev Pharmacol Toxicol*, **55**: 513–531.
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D.M., Forman, D., and Bray, F. (2015) Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*, **136**(5): E359–386.
- Fisher, R.A. (1919) Xv.—the correlation between relatives on the supposition of mendelian inheritance. *Earth and Environmental Science Transactions of the Royal Society of Edinburgh*, **52**(02): 399–433.
- Fong, P.C., Boss, D.S., Yap, T.A., Tutt, A., Wu, P., Mergui-Roelvink, M., Mortimer, P., Swaisland, H., Lau, A., O'Connor, M.J., et al. (2009) Inhibition of poly(adpribose) polymerase in tumors from BRCA mutation carriers. N Engl J Med, 361(2): 123–34.
- Fong, P.C., Yap, T.A., Boss, D.S., Carden, C.P., Mergui-Roelvink, M., Gourley, C., De Greve, J., Lubinski, J., Shanley, S., Messiou, C., et al. (2010) Poly(adp)-ribose polymerase inhibition: frequent durable responses in BRCA carrier ovarian cancer correlating with platinum-free interval. *J Clin Oncol*, **28**(15): 2512–9.
- Forbes, S.A., Beare, D., Gunasekaran, P., Leung, K., Bindal, N., Boutselakis, H., Ding, M., Bamford, S., Cole, C., Ward, S., et al. (2015) COSMIC: exploring the world's knowledge of somatic mutations in human cancer. Nucleic Acids Res, 43(Database issue): D805–811.
- Fraser, A. (2004) Towards full employment: using RNAi to find roles for the redundant. *Oncogene*, **23**(51): 8346–52.
- Fromental-Ramain, C., Warot, X., Lakkaraju, S., Favier, B., Haack, H., Birling, C., Dierich, A., Doll e, P., and Chambon, P. (1996) Specific and redundant functions of the paralogous Hoxa-9 and Hoxd-9 genes in forelimb and axial skeleton patterning. *Development*, **122**(2): 461–472.
- Futreal, P.A., Coin, L., Marshall, M., Down, T., Hubbard, T., Wooster, R., Rahman, N., and Stratton, M.R. (2004) A census of human cancer genes. *Nat Rev Cancer*, 4(3): 177–183.
- Futreal, P.A., Kasprzyk, A., Birney, E., Mullikin, J.C., Wooster, R., and Stratton, M.R. (2001) Cancer and genomics. *Nature*, **409**(6822): 850–852.

- Gao, B. and Roux, P.P. (2015) Translational control by oncogenic signaling pathways. *Biochimica et Biophysica Acta*, **1849**(7): 753–65.
- Gatza, M.L., Kung, H.N., Blackwell, K.L., Dewhirst, M.W., Marks, J.R., and Chi, J.T. (2011) Analysis of tumor environmental response and oncogenic pathway activation identifies distinct basal and luminal features in HER2-related breast tumor subtypes. *Breast Cancer Res*, 13(3): R62.
- Gatza, M.L., Lucas, J.E., Barry, W.T., Kim, J.W., Wang, Q., Crawford, M.D., Datto, M.B., Kelley, M., Mathey-Prevot, B., Potti, A., et al. (2010) A pathway-based classification of human breast cancer. *Proc Natl Acad Sci USA*, **107**(15): 6994–6999.
- Gatza, M.L., Silva, G.O., Parker, J.S., Fan, C., and Perou, C.M. (2014) An integrated genomics approach identifies drivers of proliferation in luminal-subtype human breast cancer. *Nat Genet*, **46**(10): 1051–1059.
- Gentleman, R.C., Carey, V.J., Bates, D.M., Bolstad, B., Dettling, M., Dudoit, S., Ellis, B., Gautier, L., Ge, Y., Gentry, J., et al. (2004) Bioconductor: open software development for computational biology and bioinformatics. Genome Biol, 5(10): R80.
- Genz, A. and Bretz, F. (2009) Computation of multivariate normal and t probabilities. In *Lecture Notes in Statistics*, volume 195. Springer-Verlag, Heidelberg.
- Genz, A., Bretz, F., Miwa, T., Mi, X., Leisch, F., Scheipl, F., and Hothorn, T. (2016) mvtnorm: Multivariate Normal and t Distributions. R package version 1.0-5. URL.
- Glaire, M.A., Brown, M., Church, D.N., and Tomlinson, I. (2017) Cancer predisposition syndromes: lessons for truly precision medicine. *J Pathol*, **241**(2): 226–235.
- Globus (Globus) (2017) Research data management simplified. https://www.globus.org/. Accessed: 25/03/2017.
- Goodwin, S., McPherson, J.D., and McCombie, W.R. (2016) Coming of age: ten years of next-generation sequencing technologies. *Nat Rev Genet*, **17**(6): 333–351.
- Grady, W.M., Willis, J., Guilford, P.J., Dunbier, A.K., Toro, T.T., Lynch, H., Wiesner, G., Ferguson, K., Eng, C., Park, J.G., et al. (2000) Methylation of the CDH1 promoter as the second genetic hit in hereditary diffuse gastric cancer. Nat Genet, 26(1): 16–17.
- Graziano, F., Humar, B., and Guilford, P. (2003) The role of the E-cadherin gene (*CDH1*) in diffuse gastric cancer susceptibility: from the laboratory to clinical practice. *Annals of Oncology*, **14**(12): 1705–1713.
- Guaragnella, N., Palermo, V., Galli, A., Moro, L., Mazzoni, C., and Giannattasio, S. (2014) The expanding role of yeast in cancer research and diagnosis: insights into the function of the oncosuppressors p53 and BRCA1/2. *FEMS Yeast Res*, **14**(1): 2–16.

- Güell, O., Sagués, F., and Serrano, M. (2014) Essential plasticity and redundancy of metabolism unveiled by synthetic lethality analysis. *PLoS Comput Biol*, **10**(5): e1003637.
- Guilford, P. (1999) E-cadherin downregulation in cancer: fuel on the fire? *Molecular Medicine Today*, **5**(4): 172 177.
- Guilford, P., Hopkins, J., Harraway, J., McLeod, M., McLeod, N., Harawira, P., Taite, H., Scoular, R., Miller, A., and Reeve, A.E. (1998) E-cadherin germline mutations in familial gastric cancer. *Nature*, 392(6674): 402–5.
- Guilford, P., Humar, B., and Blair, V. (2010) Hereditary diffuse gastric cancer: translation of *CDH1* germline mutations into clinical practice. *Gastric Cancer*, **13**(1): 1–10.
- Guilford, P.J., Hopkins, J.B., Grady, W.M., Markowitz, S.D., Willis, J., Lynch, H., Rajput, A., Wiesner, G.L., Lindor, N.M., Burgart, L.J., *et al.* (1999) E-cadherin germline mutations define an inherited cancer syndrome dominated by diffuse gastric cancer. *Hum Mutat*, **14**(3): 249–55.
- Guo, J., Liu, H., and Zheng, J. (2016) SynLethDB: synthetic lethality database toward discovery of selective and sensitive anticancer drug targets. *Nucleic Acids Res*, 44(D1): D1011–1017.
- Hajian-Tilaki, K. (2013) Receiver Operating Characteristic (ROC) Curve Analysis for Medical Diagnostic Test Evaluation. *Caspian J Intern Med*, 4(2): 627–635.
- Hall, M., Frank, E., Holmes, G., Pfahringer, B., Reutemann, P., and Witten, I.H. (2009) The weka data mining software: an update. *SIGKDD Explor Newsl*, **11**(1): 10–18.
- Hammerman, P.S., Lawrence, M.S., Voet, D., Jing, R., Cibulskis, K., Sivachenko, A., Stojanov, P., McKenna, A., Lander, E.S., Gabriel, S., et al. (2012) Comprehensive genomic characterization of squamous cell lung cancers. Nature, 489(7417): 519–525.
- Hanahan, D. and Weinberg, R.A. (2000) The hallmarks of cancer. Cell, 100(1): 57–70.
- Hanahan, D. and Weinberg, R.A. (2011) Hallmarks of cancer: the next generation. *Cell*, **144**(5): 646–674.
- Hanna, S. (2003) Cancer incidence in new zealand (2003-2007). In D. Forman, D. Bray F Brewster, C. Gombe Mbalawa, B. Kohler, M. Piñeros, E. Steliarova-Foucher, R. Swaminathan, and J. Ferlay (editors), Cancer Incidence in Five Continents, volume X, 902–907. International Agency for Research on Cancer, Lyon, France. Electronic version http://ci5.iarc.fr Accessed 22/03/2017.
- Hansford, S., Kaurah, P., Li-Chang, H., Woo, M., Senz, J., Pinheiro, H., Schrader, K.A., Schaeffer, D.F., Shumansky, K., Zogopoulos, G., et al. (2015) Hereditary Diffuse Gastric Cancer Syndrome: CDH1 Mutations and Beyond. JAMA Oncol, 1(1): 23–32.

- Heiskanen, M.A. and Aittokallio, T. (2012) Mining high-throughput screens for cancer drug targets-lessons from yeast chemical-genomic profiling and synthetic lethality. Wiley Interdisciplinary Reviews: Data Mining and Knowledge Discovery, 2(3): 263–272.
- Hell, P. (1976) Graphs with given neighbourhoods i. problémes combinatorics at theorie des graphes. *Proc Coil Int CNRS, Orsay,* **260**: 219–223.
- Higgins, M.E., Claremont, M., Major, J.E., Sander, C., and Lash, A.E. (2007) CancerGenes: a gene selection resource for cancer genome projects. *Nucleic Acids Res*, **35**(Database issue): D721–726.
- Hillenmeyer, M.E. (2008) The chemical genomic portrait of yeast: uncovering a phenotype for all genes. *Science*, **320**: 362–365.
- Hoadley, K.A., Yau, C., Wolf, D.M., Cherniack, A.D., Tamborero, D., Ng, S., Leiserson, M.D., Niu, B., McLellan, M.D., Uzunangelov, V., et al. (2014) Multiplatform analysis of 12 cancer types reveals molecular classification within and across tissues of origin. Cell, 158(4): 929–944.
- Hoehndorf, R., Hardy, N.W., Osumi-Sutherland, D., Tweedie, S., Schofield, P.N., and Gkoutos, G.V. (2013) Systematic analysis of experimental phenotype data reveals gene functions. *PLoS ONE*, **8**(4): e60847.
- Holm, S. (1979) A simple sequentially rejective multiple test procedure. *Scandinavian Journal of Statistics*, **6**(2): 65–70.
- Hopkins, A.L. (2008) Network pharmacology: the next paradigm in drug discovery. *Nat Chem Biol*, **4**(11): 682–690.
- Hu, Z., Fan, C., Oh, D.S., Marron, J.S., He, X., Qaqish, B.F., Livasy, C., Carey, L.A., Reynolds, E., Dressler, L., et al. (2006) The molecular portraits of breast tumors are conserved across microarray platforms. BMC Genomics, 7: 96.
- Huang, E., Cheng, S., Dressman, H., Pittman, J., Tsou, M., Horng, C., Bild, A., Iversen, E., Liao, M., Chen, C., et al. (2003) Gene expression predictors of breast cancer outcomes. *Lancet*, **361**: 1590–1596.
- Hutchison, C.A., Chuang, R.Y., Noskov, V.N., Assad-Garcia, N., Deerinck, T.J., Ellisman, M.H., Gill, J., Kannan, K., Karas, B.J., Ma, L., et al. (2016) Design and synthesis of a minimal bacterial genome. *Science*, **351**(6280): aad6253.
- International HapMap 3 Consortium (HapMap) (2003) The International HapMap Project. *Nature*, **426**(6968): 789–796.
- Jeanes, A., Gottardi, C.J., and Yap, A.S. (2008) Cadherins and cancer: how does cadherin dysfunction promote tumor progression? *Oncogene*, **27**(55): 6920–6929.

- Jerby-Arnon, L., Pfetzer, N., Waldman, Y., McGarry, L., James, D., Shanks, E., Seashore-Ludlow, B., Weinstock, A., Geiger, T., Clemons, P., et al. (2014) Predicting cancer-specific vulnerability via data-driven detection of synthetic lethality. Cell, 158(5): 1199–1209.
- Joachims, T. (1999) Making large-scale support vector machine learning practical. In S. Bernhard, lkopf, J.C.B. Christopher, and J.S. Alexander (editors), Advances in kernel methods, 169–184. MIT Press.
- Kaelin, Jr, W. (2005) The concept of synthetic lethality in the context of anticancer therapy. *Nat Rev Cancer*, **5**(9): 689–98.
- Kaelin, Jr, W. (2009) Synthetic lethality: a framework for the development of wiser cancer therapeutics. *Genome Med*, 1: 99.
- Kamada, T. and Kawai, S. (1989) An algorithm for drawing general undirected graphs. *Information Processing Letters*, **31**(1): 7–15.
- Kawai, J., Shinagawa, A., Shibata, K., Yoshino, M., Itoh, M., Ishii, Y., Arakawa, T., Hara, A., Fukunishi, Y., Konno, H., et al. (2001) Functional annotation of a full-length mouse cDNA collection. Nature, 409(6821): 685–690.
- Kelley, R. and Ideker, T. (2005) Systematic interpretation of genetic interactions using protein networks. *Nat Biotech*, **23**(5): 561–566.
- Kelly, S.T. (2013) Statistical Predictions of Synthetic Lethal Interactions in Cancer. Dissertation, University of Otago.
- Keshava Prasad, T.S., Goel, R., Kandasamy, K., Keerthikumar, S., Kumar, S., Mathivanan, S., Telikicherla, D., Raju, R., Shafreen, B., Venugopal, A., et al. (2009) Human Protein Reference Database–2009 update. Nucleic Acids Res, 37(Database issue): D767–772.
- Kim, N.G., Koh, E., Chen, X., and Gumbiner, B.M. (2011) E-cadherin mediates contact inhibition of proliferation through Hippo signaling-pathway components. *Proc Natl Acad Sci USA*, **108**(29): 11930–11935.
- Koboldt, D.C., Fulton, R.S., McLellan, M.D., Schmidt, H., Kalicki-Veizer, J., McMichael, J.F., Fulton, L.L., Dooling, D.J., Ding, L., Mardis, E.R., et al. (2012) Comprehensive molecular portraits of human breast tumours. *Nature*, **490**(7418): 61–70.
- Kockel, L., Zeitlinger, J., Staszewski, L.M., Mlodzik, M., and Bohmann, D. (1997) Jun in drosophila development: redundant and nonredundant functions and regulation by two mapk signal transduction pathways. *Genes & Development*, **11**(13): 1748–1758.
- Kozlov, K.N., Gursky, V.V., Kulakovskiy, I.V., and Samsonova, M.G. (2015) Sequence-based model of gap gene regulation network. *BMC Genomics*, **15**(Suppl 12): S6.

- Kranthi, S., Rao, S., and Manimaran, P. (2013) Identification of synthetic lethal pairs in biological systems through network information centrality. *Mol BioSyst*, **9**(8): 2163–2167.
- Kroepil, F., Fluegen, G., Totikov, Z., Baldus, S.E., Vay, C., Schauer, M., Topp, S.A., Esch, J.S., Knoefel, W.T., and Stoecklein, N.H. (2012) Down-regulation of CDH1 is associated with expression of SNAI1 in colorectal adenomas. *PLoS ONE*, **7**(9): e46665.
- Lander, E.S. (2011) Initial impact of the sequencing of the human genome. *Nature*, 470(7333): 187–197.
- Lander, E.S., Linton, L.M., Birren, B., Nusbaum, C., Zody, M.C., Baldwin, J., Devon, K., Dewar, K., Doyle, M., FitzHugh, W., et al. (2001) Initial sequencing and analysis of the human genome. *Nature*, **409**(6822): 860–921.
- Langmead, B., Trapnell, C., Pop, M., and Salzberg, S.L. (2009) Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biol*, **10**(3): R25.
- Latora, V. and Marchiori, M. (2001) Efficient behavior of small-world networks. *Phys Rev Lett*, 87: 198701.
- Laufer, C., Fischer, B., Billmann, M., Huber, W., and Boutros, M. (2013) Mapping genetic interactions in human cancer cells with RNAi and multiparametric phenotyping. *Nat Methods*, **10**(5): 427–31.
- Law, C.W., Chen, Y., Shi, W., and Smyth, G.K. (2014) voom: precision weights unlock linear model analysis tools for RNA-seq read counts. *Genome Biol*, **15**(2): R29.
- Le Meur, N. and Gentleman, R. (2008) Modeling synthetic lethality. *Genome Biol*, **9**(9): R135.
- Le Meur, N., Jiang, Z., Liu, T., Mar, J., and Gentleman, R.C. (2014) Slgi: Synthetic lethal genetic interaction. r package version 1.26.0.
- Lee, A.Y., Perreault, R., Harel, S., Boulier, E.L., Suderman, M., Hallett, M., and Jenna, S. (2010a) Searching for signaling balance through the identification of genetic interactors of the rab guanine-nucleotide dissociation inhibitor gdi-1. *PLoS ONE*, **5**(5): e10624.
- Lee, I., Lehner, B., Vavouri, T., Shin, J., Fraser, A.G., and Marcotte, E.M. (2010b) Predicting genetic modifier loci using functional gene networks. *Genome Research*, **20**(8): 1143–1153.
- Lee, I. and Marcotte, E.M. (2009) Effects of functional bias on supervised learning of a gene network model. *Methods Mol Biol*, **541**: 463–75.
- Lee, M.J., Ye, A.S., Gardino, A.K., Heijink, A.M., Sorger, P.K., MacBeath, G., and Yaffe, M.B. (2012) Sequential application of anticancer drugs enhances cell death by rewiring apoptotic signaling networks. *Cell*, **149**(4): 780–94.

- Lehner, B., Crombie, C., Tischler, J., Fortunato, A., and Fraser, A.G. (2006) Systematic mapping of genetic interactions in caenorhabditis elegans identifies common modifiers of diverse signaling pathways. *Nat Genet*, **38**(8): 896–903.
- Li, B., Ruotti, V., Stewart, R.M., Thomson, J.A., and Dewey, C.N. (2010) RNA-Seq gene expression estimation with read mapping uncertainty. *Bioinformatics*, **26**(4): 493–500.
- Li, X.J., Mishra, S.K., Wu, M., Zhang, F., and Zheng, J. (2014) Syn-lethality: An integrative knowledge base of synthetic lethality towards discovery of selective anticancer therapies. *Biomed Res Int*, **2014**: 196034.
- Linehan, W.M., Spellman, P.T., Ricketts, C.J., Creighton, C.J., Fei, S.S., Davis, C., Wheeler, D.A., Murray, B.A., Schmidt, L., Vocke, C.D., et al. (2016) Comprehensive Molecular Characterization of Papillary Renal-Cell Carcinoma. N Engl J Med, 374(2): 135–145.
- Lokody, I. (2014) Computational modelling: A computational crystal ball. *Nature Reviews Cancer*, **14**(10): 649–649.
- Lord, C.J., Tutt, A.N., and Ashworth, A. (2015) Synthetic lethality and cancer therapy: lessons learned from the development of PARP inhibitors. *Annu Rev Med*, **66**: 455–470.
- Lu, X., Kensche, P.R., Huynen, M.A., and Notebaart, R.A. (2013) Genome evolution predicts genetic interactions in protein complexes and reveals cancer drug targets. *Nat Commun*, 4: 2124.
- Lu, X., Megchelenbrink, W., Notebaart, R.A., and Huynen, M.A. (2015) Predicting human genetic interactions from cancer genome evolution. *PLoS One*, **10**(5): e0125795.
- Lum, P.Y., Armour, C.D., Stepaniants, S.B., Cavet, G., Wolf, M.K., Butler, J.S., Hinshaw, J.C., Garnier, P., Prestwich, G.D., Leonardson, A., *et al.* (2004) Discovering modes of action for therapeutic compounds using a genome-wide screen of yeast heterozygotes. *Cell*, **116**(1): 121–137.
- Luo, J., Solimini, N.L., and Elledge, S.J. (2009) Principles of Cancer Therapy: Oncogene and Non-oncogene Addiction. *Cell*, **136**(5): 823–837.
- Machado, J., Olivera, C., Carvalh, R., Soares, P., Berx, G., Caldas, C., Sercuca, R., Carneiro, F., and Sorbrinho-Simoes, M. (2001) E-cadherin gene (*CDH1*) promoter methylation as the second hit in sporadic diffuse gastric carcinoma. *Oncogene*, **20**: 1525–1528.
- Markowetz, F. (2017) All biology is computational biology. *PLoS Biol*, **15**(3): e2002050.

- Masciari, S., Larsson, N., Senz, J., Boyd, N., Kaurah, P., Kandel, M.J., Harris, L.N., Pinheiro, H.C., Troussard, A., Miron, P., et al. (2007) Germline E-cadherin mutations in familial lobular breast cancer. J Med Genet, 44(11): 726–31.
- Mattison, J., van der Weyden, L., Hubbard, T., and Adams, D.J. (2009) Cancer gene discovery in mouse and man. *Biochim Biophys Acta*, **1796**(2): 140–161.
- McLachlan, J., George, A., and Banerjee, S. (2016) The current status of parp inhibitors in ovarian cancer. *Tumori*, **102**(5): 433–440.
- McLendon, R., Friedman, A., Bigner, D., Van Meir, E.G., Brat, D.J., Mastrogianakis, G.M., Olson, J.J., Mikkelsen, T., Lehman, N., Aldape, K., et al. (2008) Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature*, **455**(7216): 1061–1068.
- Miles, D.W. (2001) Update on HER-2 as a target for cancer therapy: herceptin in the clinical setting. *Breast Cancer Res*, **3**(6): 380–384.
- Muzny, D.M., Bainbridge, M.N., Chang, K., Dinh, H.H., Drummond, J.A., Fowler, G., Kovar, C.L., Lewis, L.R., Morgan, M.B., Newsham, I.F., et al. (2012) Comprehensive molecular characterization of human colon and rectal cancer. Nature, 487(7407): 330–337.
- Nagalla, S., Chou, J.W., Willingham, M.C., Ruiz, J., Vaughn, J.P., Dubey, P., Lash, T.L., Hamilton-Dutoit, S.J., Bergh, J., Sotiriou, C., et al. (2013) Interactions between immunity, proliferation and molecular subtype in breast cancer prognosis. Genome Biol, 14(4): R34.
- Novomestky, F. (2012) matrixcalc: Collection of functions for matrix calculations. R package version 1.0-3.
- Nowak, M.A., Boerlijst, M.C., Cooke, J., and Smith, J.M. (1997) Evolution of genetic redundancy. *Nature*, **388**(6638): 167–171.
- Oliveira, C., Senz, J., Kaurah, P., Pinheiro, H., Sanges, R., Haegert, A., Corso, G., Schouten, J., Fitzgerald, R., Vogelsang, H., et al. (2009) Germline *CDH1* deletions in hereditary diffuse gastric cancer families. *Human Molecular Genetics*, **18**(9): 1545–1555.
- Oliveira, C., Seruca, R., Hoogerbrugge, N., Ligtenberg, M., and Carneiro, F. (2013) Clinical utility gene card for: Hereditary diffuse gastric cancer (HDGC). Eur J Hum Genet, 21(8).
- Pandey, G., Zhang, B., Chang, A.N., Myers, C.L., Zhu, J., Kumar, V., and Schadt, E.E. (2010) An integrative multi-network and multi-classifier approach to predict genetic interactions. *PLoS Comput Biol*, **6**(9).
- Parker, J., Mullins, M., Cheung, M., Leung, S., Voduc, D., Vickery, T., Davies, S., Fauron, C., He, X., Hu, Z., et al. (2009) Supervised risk predictor of breast cancer based on intrinsic subtypes. *Journal of Clinical Oncology*, 27(8): 1160–1167.

- Pereira, B., Chin, S.F., Rueda, O.M., Vollan, H.K., Provenzano, E., Bardwell, H.A., Pugh, M., Jones, L., Russell, R., Sammut, S.J., et al. (2016) Erratum: The somatic mutation profiles of 2,433 breast cancers refine their genomic and transcriptomic landscapes. Nat Commun, 7: 11908.
- Perou, C.M., Sørlie, T., Eisen, M.B., van de Rijn, M., Jeffrey, S.S., Rees, C.A., Pollack, J.R., Ross, D.T., Johnsen, H., Akslen, L.A., et al. (2000) Molecular portraits of human breast tumours. Nature, 406(6797): 747–752.
- Polyak, K. and Weinberg, R.A. (2009) Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits. *Nat Rev Cancer*, **9**(4): 265–73.
- R Core Team (2016) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. R version 3.3.2.
- Ritchie, M.E., Phipson, B., Wu, D., Hu, Y., Law, C.W., Shi, W., and Smyth, G.K. (2015) limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Research*, **43**(7): e47.
- Roguev, A., Bandyopadhyay, S., Zofall, M., Zhang, K., Fischer, T., Collins, S.R., Qu, H., Shales, M., Park, H.O., Hayles, J., *et al.* (2008) Conservation and rewiring of functional modules revealed by an epistasis map in fission yeast. *Science*, **322**(5900): 405–10.
- Roychowdhury, S. and Chinnaiyan, A.M. (2016) Translating cancer genomes and transcriptomes for precision oncology. *CA Cancer J Clin*, **66**(1): 75–88.
- Rung, J. and Brazma, A. (2013) Reuse of public genome-wide gene expression data. *Nat Rev Genet*, **14**(2): 89–99.
- Ryan, C., Lord, C., and Ashworth, A. (2014) Daisy: Picking synthetic lethals from cancer genomes. *Cancer Cell*, **26**(3): 306–308.
- Schena, M. (1996) Genome analysis with gene expression microarrays. *Bioessays*, **18**(5): 427–431.
- Scheuer, L., Kauff, N., Robson, M., Kelly, B., Barakat, R., Satagopan, J., Ellis, N., Hensley, M., Boyd, J., Borgen, P., et al. (2002) Outcome of preventive surgery and screening for breast and ovarian cancer in BRCA mutation carriers. *J Clin Oncol*, **20**(5): 1260–1268.
- Semb, H. and Christofori, G. (1998) The tumor-suppressor function of E-cadherin. *Am J Hum Genet*, **63**(6): 1588–93.
- Sing, T., Sander, O., Beerenwinkel, N., and Lengauer, T. (2005) Rocr: visualizing classifier performance in r. *Bioinformatics*, **21**(20): 7881.
- Slurm development team (Slurm) (2017) Slurm workload manager. https://slurm.schedmd.com/. Accessed: 25/03/2017.

- Sørlie, T., Perou, C.M., Tibshirani, R., Aas, T., Geisler, S., Johnsen, H., Hastie, T., Eisen, M.B., van de Rijn, M., Jeffrey, S.S., et al. (2001) Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc Natl Acad Sci USA*, **98**(19): 10869–10874.
- Srihari, S., Singla, J., Wong, L., and Ragan, M.A. (2015) Inferring synthetic lethal interactions from mutual exclusivity of genetic events in cancer. *Biology Direct*, **10**(1): 57.
- Stajich, J.E. and Lapp, H. (2006) Open source tools and toolkits for bioinformatics: significance, and where are we? *Brief Bioinformatics*, **7**(3): 287–296.
- Stratton, M.R., Campbell, P.J., and Futreal, P.A. (2009) The cancer genome. *Nature*, **458**(7239): 719–724.
- Ström, C. and Helleday, T. (2012) Strategies for the use of poly(adenosine diphosphate ribose) polymerase (parp) inhibitors in cancer therapy. *Biomolecules*, **2**(4): 635–649.
- Tarazona, S., Garcia-Alcalde, F., Dopazo, J., Ferrer, A., and Conesa, A. (2011) Differential expression in RNA-seq: a matter of depth. *Genome Res*, **21**(12): 2213–2223.
- Telford, B.J., Chen, A., Beetham, H., Frick, J., Brew, T.P., Gould, C.M., Single, A., Godwin, T., Simpson, K.J., and Guilford, P. (2015) Synthetic lethal screens identify vulnerabilities in gpcr signalling and cytoskeletal organization in E-cadherin-deficient cells. *Mol Cancer Ther*, 14(5): 1213–1223.
- The 1000 Genomes Project Consortium (1000 Genomes) (2010) A map of human genome variation from population-scale sequencing. *Nature*, **467**(7319): 1061–1073.
- The Cancer Genome Atlas Research Network (TCGA) (2017) The Cancer Genome Atlas Project. https://cancergenome.nih.gov/. Accessed: 26/03/2017.
- The Catalogue Of Somatic Mutations In Cancer (COSMIC) (2016) Cosmic: The catalogue of somatic mutations in cancer. http://cancer.sanger.ac.uk/cosmic. Release 79 (23/08/2016), Accessed: 05/02/2017.
- The Comprehensive R Archive Network (CRAN) (2017) Cran. https://cran.r-project.org/. Accessed: 24/03/2017.
- The ENCODE Project Consortium (ENCODE) (2004) The ENCODE (ENCyclopedia Of DNA Elements) Project. *Science*, **306**(5696): 636–640.
- The National Cancer Institute (NCI) (2015) The genetics of cancer. https://www.cancer.gov/about-cancer/causes-prevention/genetics. Published: 22/04/2015, Accessed: 22/03/2017.
- The New Zealand eScience Infrastructure (NeSI) (2017) NeSI. https://www.nesi.org.nz/. Accessed: 25/03/2017.
- Tierney, L., Rossini, A.J., Li, N., and Sevcikova, H. (2015) snow: Simple Network of Workstations. R package version 0.4-2.

- Tiong, K.L., Chang, K.C., Yeh, K.T., Liu, T.Y., Wu, J.H., Hsieh, P.H., Lin, S.H., Lai, W.Y., Hsu, Y.C., Chen, J.Y., et al. (2014) Csnk1e/ctnnb1 are synthetic lethal to tp53 in colorectal cancer and are markers for prognosis. Neoplasia, 16(5): 441–50.
- Tischler, J., Lehner, B., and Fraser, A.G. (2008) Evolutionary plasticity of genetic interaction networks. *Nat Genet*, **40**(4): 390–391.
- Tomasetti, C. and Vogelstein, B. (2015) Cancer etiology. Variation in cancer risk among tissues can be explained by the number of stem cell divisions. *Science*, **347**(6217): 78–81.
- Tong, A.H., Evangelista, M., Parsons, A.B., Xu, H., Bader, G.D., Page, N., Robinson, M., Raghibizadeh, S., Hogue, C.W., Bussey, H., et al. (2001) Systematic genetic analysis with ordered arrays of yeast deletion mutants. Science, 294(5550): 2364–8.
- Tong, A.H., Lesage, G., Bader, G.D., Ding, H., Xu, H., Xin, X., Young, J., Berriz, G.F., Brost, R.L., Chang, M., et al. (2004) Global mapping of the yeast genetic interaction network. Science, 303(5659): 808–13.
- Tran, B., Dancey, J.E., Kamel-Reid, S., McPherson, J.D., Bedard, P.L., Brown, A.M., Zhang, T., Shaw, P., Onetto, N., Stein, L., et al. (2012) Cancer genomics: technology, discovery, and translation. J Clin Oncol, 30(6): 647–660.
- Travers, J. and Milgram, S. (1969) An experimental study of the small world problem. Sociometry, **32**(4): 425–443.
- Tunggal, J.A., Helfrich, I., Schmitz, A., Schwarz, H., Gunzel, D., Fromm, M., Kemler, R., Krieg, T., and Niessen, C.M. (2005) E-cadherin is essential for in vivo epidermal barrier function by regulating tight junctions. *EMBO J*, 24(6): 1146–1156.
- Tutt, A., Robson, M., Garber, J.E., Domchek, S.M., Audeh, M.W., Weitzel, J.N., Friedlander, M., Arun, B., Loman, N., Schmutzler, R.K., et al. (2010) Oral poly(adpribose) polymerase inhibitor olaparib in patients with *BRCA1* or *BRCA2* mutations and advanced breast cancer: a proof-of-concept trial. *Lancet*, **376**(9737): 235–44.
- University of California, Santa Cruz (UCSC) (2012) Ucsc cancer browser. Accessed 29/03/2012.
- van der Meer, R., Song, H.Y., Park, S.H., Abdulkadir, S.A., and Roh, M. (2014) RNAi screen identifies a synthetic lethal interaction between PIM1 overexpression and PLK1 inhibition. *Clinical Cancer Research*, **20**(12): 3211–3221.
- van der Post, R.S., Vogelaar, I.P., Carneiro, F., Guilford, P., Huntsman, D., Hoogerbrugge, N., Caldas, C., Schreiber, K.E., Hardwick, R.H., Ausems, M.G., et al. (2015) Hereditary diffuse gastric cancer: updated clinical guidelines with an emphasis on germline CDH1 mutation carriers. J Med Genet, 52(6): 361–374.
- van Steen, K. (2012) Travelling the world of gene-gene interactions. *Briefings in Bioin-formatics*, **13**(1): 1–19.

- van Steen, M. (2010) Graph Theory and Complex Networks: An Introduction. Maarten van Steen, VU Amsterdam.
- Vapnik, V.N. (1995) The nature of statistical learning theory. Springer-Verlag New York, Inc.
- Vizeacoumar, F.J., Arnold, R., Vizeacoumar, F.S., Chandrashekhar, M., Buzina, A., Young, J.T., Kwan, J.H., Sayad, A., Mero, P., Lawo, S., et al. (2013) A negative genetic interaction map in isogenic cancer cell lines reveals cancer cell vulnerabilities. Mol Syst Biol, 9: 696.
- Vogelstein, B., Papadopoulos, N., Velculescu, V.E., Zhou, S., Diaz, L.A., and Kinzler, K.W. (2013) Cancer genome landscapes. *Science*, **339**(6127): 1546–1558.
- Vos, C.B., Cleton-Jansen, A.M., Berx, G., de Leeuw, W.J., ter Haar, N.T., van Roy, F., Cornelisse, C.J., Peterse, J.L., and van de Vijver, M.J. (1997) E-cadherin inactivation in lobular carcinoma in situ of the breast: an early event in tumorigenesis. Br J Cancer, 76(9): 1131–3.
- Waldron, D. (2016) Cancer genomics: A multi-layer omics approach to cancer. *Nat Rev Genet*, **17**(8): 436–437.
- Wang, K., Singh, D., Zeng, Z., Coleman, S.J., Huang, Y., Savich, G.L., He, X., Mieczkowski, P., Grimm, S.A., Perou, C.M., et al. (2010) MapSplice: accurate mapping of RNA-seq reads for splice junction discovery. Nucleic Acids Res, 38(18): e178.
- Wang, X. and Simon, R. (2013) Identification of potential synthetic lethal genes to p53 using a computational biology approach. *BMC Medical Genomics*, **6**(1): 30.
- Wappett, M. (2014) Bisep: Toolkit to identify candidate synthetic lethality. r package version 2.0.
- Wappett, M., Dulak, A., Yang, Z.R., Al-Watban, A., Bradford, J.R., and Dry, J.R. (2016) Multi-omic measurement of mutually exclusive loss-of-function enriches for candidate synthetic lethal gene pairs. BMC Genomics, 17: 65.
- Warnes, G.R., Bolker, B., Bonebakker, L., Gentleman, R., Liaw, W.H.A., Lumley, T., Maechler, M., Magnusson, A., Moeller, S., Schwartz, M., et al. (2015) gplots: Various R Programming Tools for Plotting Data. R package version 2.17.0.
- Watts, D.J. and Strogatz, S.H. (1998) Collective dynamics of 'small-world' networks. *Nature*, **393**(6684): 440–2.
- Weinstein, I.B. (2000) Disorders in cell circuitry during multistage carcinogenesis: the role of homeostasis. *Carcinogenesis*, **21**(5): 857–864.
- Weinstein, J.N., Akbani, R., Broom, B.M., Wang, W., Verhaak, R.G., McConkey, D., Lerner, S., Morgan, M., Creighton, C.J., Smith, C., et al. (2014) Comprehensive molecular characterization of urothelial bladder carcinoma. *Nature*, **507**(7492): 315–322.

- Weinstein, J.N., Collisson, E.A., Mills, G.B., Shaw, K.R., Ozenberger, B.A., Ellrott, K., Shmulevich, I., Sander, C., Stuart, J.M., Chang, K., et al. (2013) The Cancer Genome Atlas Pan-Cancer analysis project. Nat Genet, 45(10): 1113–1120.
- Wickham, H. and Chang, W. (2016) devtools: Tools to Make Developing R Packages Easier. R package version 1.12.0.
- Wickham, H., Danenberg, P., and Eugster, M. (2017) roxygen2: In-Line Documentation for R. R package version 6.0.1.
- Wong, S.L., Zhang, L.V., Tong, A.H.Y., Li, Z., Goldberg, D.S., King, O.D., Lesage, G., Vidal, M., Andrews, B., Bussey, H., et al. (2004) Combining biological networks to predict genetic interactions. Proceedings of the National Academy of Sciences of the United States of America, 101(44): 15682–15687.
- World Health Organization (WHO) (2017) Fact sheet: Cancer. http://www.who.int/mediacentre/factsheets/fs297/en/. Updated February 2017, Accessed: 22/03/2017.
- Wu, M., Li, X., Zhang, F., Li, X., Kwoh, C.K., and Zheng, J. (2014) In silico prediction of synthetic lethality by meta-analysis of genetic interactions, functions, and pathways in yeast and human cancer. *Cancer Inform*, **13**(Suppl 3): 71–80.
- Yu, H. (2002) Rmpi: Parallel statistical computing in r. R News, 2(2): 10–14.
- Zhang, F., Wu, M., Li, X.J., Li, X.L., Kwoh, C.K., and Zheng, J. (2015) Predicting essential genes and synthetic lethality via influence propagation in signaling pathways of cancer cell fates. *J Bioinform Comput Biol*, **13**(3): 1541002.
- Zhang, J., Baran, J., Cros, A., Guberman, J.M., Haider, S., Hsu, J., Liang, Y., Rivkin, E., Wang, J., Whitty, B., et al. (2011) International cancer genome consortium data portal—a one-stop shop for cancer genomics data. Database: The Journal of Biological Databases and Curation, 2011: bar026.
- Zhong, W. and Sternberg, P.W. (2006) Genome-wide prediction of c. elegans genetic interactions. *Science*, **311**(5766): 1481–1484.
- Zweig, M.H. and Campbell, G. (1993) Receiver-operating characteristic (roc) plots: a fundamental evaluation tool in clinical medicine. *Clinical Chemistry*, **39**(4): 561–577.