

Lab 10 part 2: structural bioinformatics

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Today we are going to finish Lab 10 on analyzing protein structures (starting at section 4).

Comparative structure analysis of Adenylate Kinase

Starting from only one Adk PDB identifier (PDB ID: 1AKE) we will search the entire PDB for related structures using BLAST, fetch, align, and superpose the identified structures, perform PCA, and finally calculate the normal modes of each individual structure in order to probe for potential differences in structural flexibility.

We will use the `bio3d` package for this analysis that starts with a single sequence. We will also use the `msa` package from BioConductor. First, we need to install the `BiocManager` package from CRAN. We use `BiocManager::install()` to install any other BioConductor packages in the future.

Setup:

```
# Install packages in the R console, NOT your Rmd/Quarto file.
# install.packages("bio3d")
# install.packages("devtools")
# install.packages("BiocManager") # from CRAN, manages BioConductor packages
# BiocManager::install("msa") # install packages from BioConductor
# devtools::install_bitbucket("Grantlab/bio3d-view")
library(bio3d)
```

Q10. Which of the packages above is found only on BioConductor and not CRAN? `msa`

Q11. Which of the above packages is not found on BioConductor or CRAN? `bio3d-view`

Q12. True or False? Functions from the `devtools` package can be used to install packages from GitHub and BitBucket? TRUE

Search and retrieve ADK structures. Below we perform a blast search of the PDB database to identify related structures to our query Adenylate kinase (ADK) sequence. In this particular example we use function `get.seq()` to fetch the query sequence for chain A of the PDB ID 1AKE

and use this as input to `blast.pdb()`. Note that `get.seq()` would also allow the corresponding UniProt identifier.

Sequence of interest = lake_A:

```
aa <- get.seq("lake_A")
```

Warning in `get.seq("lake_A")`: Removing existing file: `seqs.fasta`

Fetching... Please wait. Done.

```
aa
```

```

      1      .      .      .      .      .      60
pdb|1AKE|A  MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAAVKSGSELGKQAKDIMDAGKLVT
      1      .      .      .      .      .      60

      61      .      .      .      .      .      120
pdb|1AKE|A  DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDRI
      61      .      .      .      .      .      120

     121      .      .      .      .      .      180
pdb|1AKE|A  VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
     121      .      .      .      .      .      180

     181      .      .      .      214
pdb|1AKE|A  YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
     181      .      .      .      214
```

Call:

```
read.fasta(file = outfile)
```

Class:

```
fasta
```

Alignment dimensions:

```
1 sequence rows; 214 position columns (214 non-gap, 0 gap)
```

```
+ attr: id, ali, call
```

Q13. How many amino acids are in this sequence, i.e. how long is this sequence? 214

I want to search for all related structures in the PDB database. Blast search:

```
#b <- blast.pdb(aa)
# hits <- plot(b) # top scoring hits are black
```

Save results thus far so we don't have to run blast again:

```
# save(hits, b, file = "blast_results.Rds")
```

Read this file:

```
load("blast_results.Rds")
```

Investigate hits:

```
head(hits)
```

\$hits

	pdb.id	acc	group
1	"1AKE_A"	"1AKE_A"	"1"
2	"8BQF_A"	"8BQF_A"	"1"
3	"4X8M_A"	"4X8M_A"	"1"
4	"6S36_A"	"6S36_A"	"1"
5	"6RZE_A"	"6RZE_A"	"1"
6	"4X8H_A"	"4X8H_A"	"1"
7	"3HPR_A"	"3HPR_A"	"1"
8	"1E4V_A"	"1E4V_A"	"1"
9	"5EJE_A"	"5EJE_A"	"1"
10	"1E4Y_A"	"1E4Y_A"	"1"
11	"3X2S_A"	"3X2S_A"	"1"
12	"6HAP_A"	"6HAP_A"	"1"
13	"6HAM_A"	"6HAM_A"	"1"
14	"4K46_A"	"4K46_A"	"1"
15	"4NP6_A"	"4NP6_A"	"1"
16	"3GMT_A"	"3GMT_A"	"1"
17	"4PZL_A"	"4PZL_A"	"1"

\$pdb.id

```
[1] "1AKE_A" "8BQF_A" "4X8M_A" "6S36_A" "6RZE_A" "4X8H_A" "3HPR_A" "1E4V_A"
[9] "5EJE_A" "1E4Y_A" "3X2S_A" "6HAP_A" "6HAM_A" "4K46_A" "4NP6_A" "3GMT_A"
[17] "4PZL_A"
```

```
$acc
[1] "1AKE_A" "8BQF_A" "4X8M_A" "6S36_A" "6RZE_A" "4X8H_A" "3HPR_A" "1E4V_A"
[9] "5EJE_A" "1E4Y_A" "3X2S_A" "6HAP_A" "6HAM_A" "4K46_A" "4NP6_A" "3GMT_A"
[17] "4PZL_A"
```

```
$inds
[1] TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE
[13] TRUE TRUE TRUE TRUE TRUE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[25] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[37] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[49] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[61] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[73] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
```

```
hits$pdb.id
```

```
[1] "1AKE_A" "8BQF_A" "4X8M_A" "6S36_A" "6RZE_A" "4X8H_A" "3HPR_A" "1E4V_A"
[9] "5EJE_A" "1E4Y_A" "3X2S_A" "6HAP_A" "6HAM_A" "4K46_A" "4NP6_A" "3GMT_A"
[17] "4PZL_A"
```

Now we will download all these related structures from the database with `get.pdb()`:

```
# download PDB files
files <- get.pdb(hits$pdb.id, path="pdb", split=TRUE, gzip=TRUE)
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):
pdb/1AKE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):
pdb/8BQF.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):
pdb/4X8M.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):
pdb/6S36.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):
pdb/6RZE.pdb.gz exists. Skipping download
```

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4X8H.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3HPR.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4V.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/5EJE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4Y.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3X2S.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAP.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAM.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4K46.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4NP6.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3GMT.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4PZL.pdb.gz exists. Skipping download

	0%
	6%
====	12%
	18%
=====	24%
	29%
=====	35%
	41%
=====	47%
	53%
=====	59%
	65%
=====	71%
	76%
=====	82%
	88%
=====	94%
	100%

```
# create folder so files don't get put in the project directory
# view all these structures in Mol*
```

Next we will use the `pdbaln()` function to align and also optionally fit (i.e. superpose) the identified PDB structures.

```
# Align related PDBs
pdbs <- pdbaln(files, fit = TRUE, exefile="msa")
```

Reading PDB files:

```
pdbs/split_chain/1AKE_A.pdb
pdbs/split_chain/8BQF_A.pdb
pdbs/split_chain/4X8M_A.pdb
pdbs/split_chain/6S36_A.pdb
pdbs/split_chain/6RZE_A.pdb
pdbs/split_chain/4X8H_A.pdb
pdbs/split_chain/3HPR_A.pdb
pdbs/split_chain/1E4V_A.pdb
pdbs/split_chain/5EJE_A.pdb
pdbs/split_chain/1E4Y_A.pdb
pdbs/split_chain/3X2S_A.pdb
pdbs/split_chain/6HAP_A.pdb
pdbs/split_chain/6HAM_A.pdb
pdbs/split_chain/4K46_A.pdb
pdbs/split_chain/4NP6_A.pdb
pdbs/split_chain/3GMT_A.pdb
pdbs/split_chain/4PZL_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
.... PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
....
```

Extracting sequences

```
pdb/seq: 1   name: pdbs/split_chain/1AKE_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 2   name: pdbs/split_chain/8BQF_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 3   name: pdbs/split_chain/4X8M_A.pdb
pdb/seq: 4   name: pdbs/split_chain/6S36_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 5   name: pdbs/split_chain/6RZE_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
```

```

pdb/seq: 6   name: pdbc/split_chain/4X8H_A.pdb
pdb/seq: 7   name: pdbc/split_chain/3HPR_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 8   name: pdbc/split_chain/1E4V_A.pdb
pdb/seq: 9   name: pdbc/split_chain/5EJE_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 10  name: pdbc/split_chain/1E4Y_A.pdb
pdb/seq: 11  name: pdbc/split_chain/3X2S_A.pdb
pdb/seq: 12  name: pdbc/split_chain/6HAP_A.pdb
pdb/seq: 13  name: pdbc/split_chain/6HAM_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 14  name: pdbc/split_chain/4K46_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 15  name: pdbc/split_chain/4NP6_A.pdb
pdb/seq: 16  name: pdbc/split_chain/3GMT_A.pdb
pdb/seq: 17  name: pdbc/split_chain/4PZL_A.pdb

```

pdbc

```

[Truncated_Name:1] 1AKE_A.pdb      1          .          .          .          40
[Truncated_Name:2] 8BQF_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:3] 4X8M_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:4] 6S36_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:5] 6RZE_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:6] 4X8H_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:7] 3HPR_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:8] 1E4V_A.pdb      -----MRIILLGAPVAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:9] 5EJE_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:10] 1E4Y_A.pdb      -----MRIILLGALVAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:11] 3X2S_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:12] 6HAP_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:13] 6HAM_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:14] 4K46_A.pdb      -----MRIILLGAPGAGKGTQAQFIMAKFGIPQIS
[Truncated_Name:15] 4NP6_A.pdb      -----NAMRIILLGAPGAGKGTQAQFIMEKFGIPQIS
[Truncated_Name:16] 3GMT_A.pdb      -----MRLILLGAPGAGKGTQANFIKEKFGIPQIS
[Truncated_Name:17] 4PZL_A.pdb      TENLYFQSNAMRIILLGAPGAGKGTQAKIIEQKYNIAHIS
                                **^*****  *  *^ *  **
1          .          .          .          40
41          .          .          .          80

```


[Truncated_Name:1] 1AKE_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:2] 8BQF_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:3] 4X8M_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:4] 6S36_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:5] 6RZE_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:6] 4X8H_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:7] 3HPR_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:8] 1E4V_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:9] 5EJE_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDACKLVTDDELVIALVKE
[Truncated_Name:10] 1E4Y_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:11] 3X2S_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDCGKLVTDDELVIALVKE
[Truncated_Name:12] 6HAP_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVRE
[Truncated_Name:13] 6HAM_A.pdb	TGDMRLRAAIKSGSELGKQAKDIMDAGKLVTDDEIIIALVKE
[Truncated_Name:14] 4K46_A.pdb	TGDMRLRAAIKAGTELGKQAKSVIDAGQLVSDDIILGLVKE
[Truncated_Name:15] 4NP6_A.pdb	TGDMRLRAAIKAGTELGKQAKAVIDAGQLVSDDIILGLIKE
[Truncated_Name:16] 3GMT_A.pdb	TGDMRLRAAVKAGTPLGVEAKTYMDEGKLVPSLIIGLVKE
[Truncated_Name:17] 4PZL_A.pdb	TGDMIRETIKSGSALGQELKKVLDAGELVSDEFIIKIVKD
	****~* ~* *~ ** * ~* ** * ^^ ~~~~
	41 . . . 80
	81 . . . 120
[Truncated_Name:1] 1AKE_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:2] 8BQF_A.pdb	RIAQE---GFLLDGFPR TIPQADAMKEAGINVDYVIEFD
[Truncated_Name:3] 4X8M_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:4] 6S36_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:5] 6RZE_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:6] 4X8H_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:7] 3HPR_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:8] 1E4V_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:9] 5EJE_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:10] 1E4Y_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:11] 3X2S_A.pdb	RIAQEDSRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:12] 6HAP_A.pdb	RICQEDSRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:13] 6HAM_A.pdb	RICQEDSRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:14] 4K46_A.pdb	RIAQDDCAKGFLLDGFPR TIPQADGLKEVGVVVDYVIEFD
[Truncated_Name:15] 4NP6_A.pdb	RIAQADCEKGFLLDGFPR TIPQADGLKEMGINVDYVIEFD
[Truncated_Name:16] 3GMT_A.pdb	RLKEADCANGYLFDFPR TIPQADAMKEAGVAIDYVLEID
[Truncated_Name:17] 4PZL_A.pdb	RISKNDCNNGFLLDGVPR TIPQAQELDKLGVNIDYIVEVD
	*~ *~* ** ***** ** ^ *~ ^**~* *
	81 . . . 120
	121 . . . 160
[Truncated_Name:1] 1AKE_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHVKNPPKVEGKDDVTG


```

[Truncated_Name:3] 4X8M_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:4] 6S36_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:5] 6RZE_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:6] 4X8H_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:7] 3HPR_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:8] 1E4V_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:9] 5EJE_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:10] 1E4Y_A.pdb     T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:11] 3X2S_A.pdb     T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:12] 6HAP_A.pdb     T--KYAKVDGTKPVCEVRADLEKILG-
[Truncated_Name:13] 6HAM_A.pdb     T--KYAKVDGTKPVCEVRADLEKILG-
[Truncated_Name:14] 4K46_A.pdb     T--QYLKFDGTKAVAESAELEKALA-
[Truncated_Name:15] 4NP6_A.pdb     T--QYLKFDGTKQVSEVSADIAKALA-
[Truncated_Name:16] 3GMT_A.pdb     E-----NGLKAPA-----YRKISG-
[Truncated_Name:17] 4PZL_A.pdb     KIPKYIKINGDQAVEKVSQDIFDQLNK
                                     *
                                201      .      .      227

```

Call:

```
pdbaln(files = files, fit = TRUE, exefile = "msa")
```

Class:

```
pdbs, fasta
```

Alignment dimensions:

```
17 sequence rows; 227 position columns (199 non-gap, 28 gap)
```

```
+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

```

# Vector containing PDB codes for figure axis
ids <- basename.pdb(pdbs$id)
# Draw schematic alignment
# plot(pdbs, labels=ids) # error message

```

This is a schematic representation of the alignment. Grey regions depict aligned residues, while white depict gap regions. The red bar at the top depict sequence conservation.

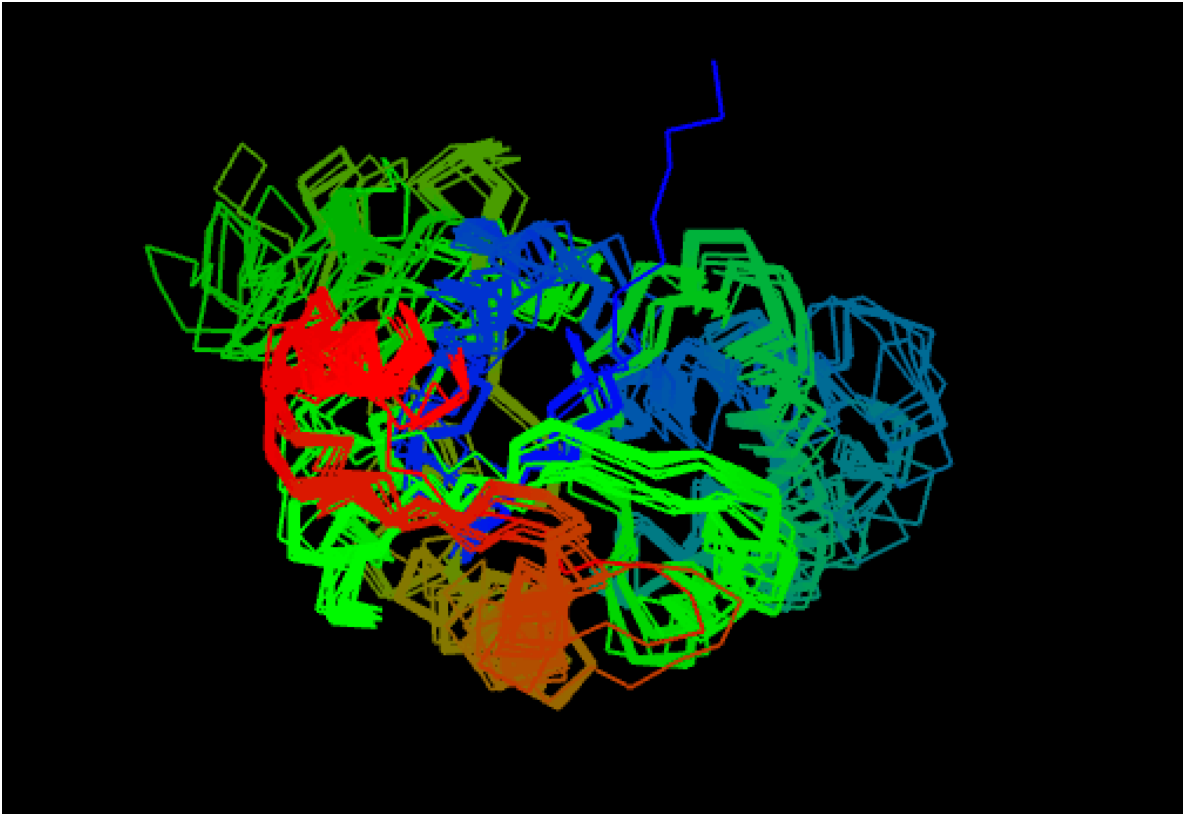
Viewing our superposed structures with `view.pdbs()`:

```

library(bio3d.view)
library(rgl)

```

```
view.pdbs(pdbs)
```



3D view of superposed ADK structures available in the PDB

Annotate collected PDB structures: The function `pdb.annotate()` provides a convenient way of annotating the PDB files we have collected. Below we use the function to annotate each structure to its source species. This will come in handy when annotating plots later on.

```
anno <- pdb.annotate(ids)
unique(anno$source)
```

```
[1] "Escherichia coli"
[2] "Escherichia coli K-12"
[3] "Escherichia coli O139:H28 str. E24377A"
[4] "Escherichia coli str. K-12 substr. MDS42"
[5] "Photobacterium profundum"
[6] "Vibrio cholerae O1 biovar El Tor str. N16961"
[7] "Burkholderia pseudomallei 1710b"
[8] "Francisella tularensis subsp. tularensis SCHU S4"
```

```
# view all annotation data
anno
```

	structureId	chainId	macromoleculeType	chainLength	experimentalTechnique
1AKE_A	1AKE	A	Protein	214	X-ray
8BQF_A	8BQF	A	Protein	234	X-ray
4X8M_A	4X8M	A	Protein	214	X-ray
6S36_A	6S36	A	Protein	214	X-ray
6RZE_A	6RZE	A	Protein	214	X-ray
4X8H_A	4X8H	A	Protein	214	X-ray
3HPR_A	3HPR	A	Protein	214	X-ray
1E4V_A	1E4V	A	Protein	214	X-ray
5EJE_A	5EJE	A	Protein	214	X-ray
1E4Y_A	1E4Y	A	Protein	214	X-ray
3X2S_A	3X2S	A	Protein	214	X-ray
6HAP_A	6HAP	A	Protein	214	X-ray
6HAM_A	6HAM	A	Protein	214	X-ray
4K46_A	4K46	A	Protein	214	X-ray
4NP6_A	4NP6	A	Protein	217	X-ray
3GMT_A	3GMT	A	Protein	230	X-ray
4PZL_A	4PZL	A	Protein	242	X-ray

	resolution	scopDomain	pfam
1AKE_A	2.000	Adenylate kinase	Adenylate kinase, active site lid (ADK_lid)
8BQF_A	2.050	<NA>	Adenylate kinase, active site lid (ADK_lid)
4X8M_A	2.600	<NA>	Adenylate kinase, active site lid (ADK_lid)
6S36_A	1.600	<NA>	Adenylate kinase, active site lid (ADK_lid)
6RZE_A	1.690	<NA>	Adenylate kinase, active site lid (ADK_lid)
4X8H_A	2.500	<NA>	Adenylate kinase, active site lid (ADK_lid)
3HPR_A	2.000	<NA>	Adenylate kinase, active site lid (ADK_lid)
1E4V_A	1.850	Adenylate kinase	Adenylate kinase, active site lid (ADK_lid)
5EJE_A	1.900	<NA>	Adenylate kinase, active site lid (ADK_lid)
1E4Y_A	1.850	Adenylate kinase	Adenylate kinase, active site lid (ADK_lid)
3X2S_A	2.800	<NA>	Adenylate kinase, active site lid (ADK_lid)
6HAP_A	2.700	<NA>	Adenylate kinase, active site lid (ADK_lid)
6HAM_A	2.550	<NA>	Adenylate kinase, active site lid (ADK_lid)
4K46_A	2.010	<NA>	Adenylate kinase, active site lid (ADK_lid)
4NP6_A	2.004	<NA>	Adenylate kinase, active site lid (ADK_lid)
3GMT_A	2.100	<NA>	Adenylate kinase, active site lid (ADK_lid)
4PZL_A	2.100	<NA>	Adenylate kinase, active site lid (ADK_lid)

	ligandId
1AKE_A	AP5
8BQF_A	AP5

4X8M_A	<NA>
6S36_A	CL (3),NA,MG (2)
6RZE_A	NA (3),CL (2)
4X8H_A	<NA>
3HPR_A	AP5
1E4V_A	AP5
5EJE_A	AP5,CO
1E4Y_A	AP5
3X2S_A	JPY (2),AP5,MG
6HAP_A	AP5
6HAM_A	AP5
4K46_A	ADP,AMP,PO4
4NP6_A	<NA>
3GMT_A	SO4 (2)
4PZL_A	CA,FMT,GOL

	ligandName
1AKE_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
8BQF_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
4X8M_A	<NA>
6S36_A	CHLORIDE ION (3),SODIUM ION,MAGNESIUM ION (2)
6RZE_A	SODIUM ION (3),CHLORIDE ION (2)
4X8H_A	<NA>
3HPR_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
1E4V_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
5EJE_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE,COBALT (II) ION
1E4Y_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
3X2S_A	N-(pyren-1-ylmethyl)acetamide (2),BIS(ADENOSINE)-5'-PENTAPHOSPHATE,MAGNESIUM ION
6HAP_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
6HAM_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
4K46_A	ADENOSINE-5'-DIPHOSPHATE,ADENOSINE MONOPHOSPHATE,PHOSPHATE ION
4NP6_A	<NA>
3GMT_A	SULFATE ION (2)
4PZL_A	CALCIUM ION,FORMIC ACID,GLYCEROL

	source
1AKE_A	Escherichia coli
8BQF_A	Escherichia coli
4X8M_A	Escherichia coli
6S36_A	Escherichia coli
6RZE_A	Escherichia coli
4X8H_A	Escherichia coli
3HPR_A	Escherichia coli K-12
1E4V_A	Escherichia coli
5EJE_A	Escherichia coli 0139:H28 str. E24377A

1E4Y_A Escherichia coli
 3X2S_A Escherichia coli str. K-12 substr. MDS42
 6HAP_A Escherichia coli O139:H28 str. E24377A
 6HAM_A Escherichia coli K-12
 4K46_A Photobacterium profundum
 4NP6_A Vibrio cholerae O1 biovar El Tor str. N16961
 3GMT_A Burkholderia pseudomallei 1710b
 4PZL_A Francisella tularensis subsp. tularensis SCHU S4

1AKE_A STRUCTURE OF THE COMPLEX BETWEEN ADENYLATE KINASE FROM ESCHERICHIA COLI AND THE INHIB

8BQF_A
 4X8M_A
 6S36_A
 6RZE_A
 4X8H_A
 3HPR_A
 1E4V_A
 5EJE_A
 1E4Y_A
 3X2S_A
 6HAP_A
 6HAM_A
 4K46_A
 4NP6_A
 3GMT_A
 4PZL_A

Cryst

The crys

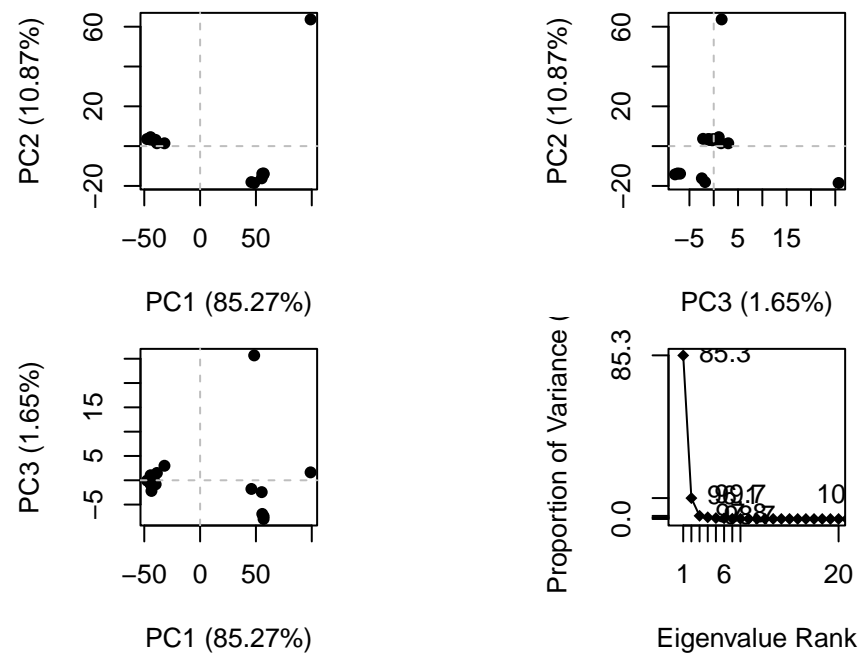
		citation	rObserved	rFree
1AKE_A	Muller, C.W., et al. J Mol Biol (1992)	0.19600	NA	
8BQF_A	Scheerer, D., et al. Proc Natl Acad Sci U S A (2023)	0.22073	0.25789	
4X8M_A	Kovermann, M., et al. Nat Commun (2015)	0.24910	0.30890	
6S36_A	Rogne, P., et al. Biochemistry (2019)	0.16320	0.23560	
6RZE_A	Rogne, P., et al. Biochemistry (2019)	0.18650	0.23500	
4X8H_A	Kovermann, M., et al. Nat Commun (2015)	0.19610	0.28950	
3HPR_A	Schrank, T.P., et al. Proc Natl Acad Sci U S A (2009)	0.21000	0.24320	
1E4V_A	Muller, C.W., et al. Proteins (1993)	0.19600	NA	
5EJE_A	Kovermann, M., et al. Proc Natl Acad Sci U S A (2017)	0.18890	0.23580	
1E4Y_A	Muller, C.W., et al. Proteins (1993)	0.17800	NA	
3X2S_A	Fujii, A., et al. Bioconjug Chem (2015)	0.20700	0.25600	
6HAP_A	Kantaev, R., et al. J Phys Chem B (2018)	0.22630	0.27760	
6HAM_A	Kantaev, R., et al. J Phys Chem B (2018)	0.20511	0.24325	
4K46_A	Cho, Y.-J., et al. To be published	0.17000	0.22290	
4NP6_A	Kim, Y., et al. To be published	0.18800	0.22200	
3GMT_A	Buchko, G.W., et al. Biochem Biophys Res Commun (2010)	0.23800	0.29500	

4PZL_A						Tan, K., et al. To be published	0.19360	0.23680
	rWork	spaceGroup						
1AKE_A	0.19600	P	21	2	21			
8BQF_A	0.21882	P	2	21	21			
4X8M_A	0.24630	C	1	2	1			
6S36_A	0.15940	C	1	2	1			
6RZE_A	0.18190	C	1	2	1			
4X8H_A	0.19140	C	1	2	1			
3HPR_A	0.20620	P	21	21	2			
1E4V_A	0.19600	P	21	2	21			
5EJE_A	0.18630	P	21	2	21			
1E4Y_A	0.17800	P	1	21	1			
3X2S_A	0.20700	P	21	21	21			
6HAP_A	0.22370	I	2	2	2			
6HAM_A	0.20311	P			43			
4K46_A	0.16730	P	21	21	21			
4NP6_A	0.18600	P			43			
3GMT_A	0.23500	P	1	21	1			
4PZL_A	0.19130	P			32			

Principle component analysis: The function `pca()` provides principal component analysis (PCA) of the structure data. PCA is a statistical approach used to transform a data set down to a few important components that describe the directions where there is most variance. In terms of protein structures PCA is used to capture major structural variations within an ensemble of structures.

We can do a PCA on the coordinate data of all structures:

```
# Perform PCA
pc.xray <- pca(pdbx)
plot(pc.xray)
```

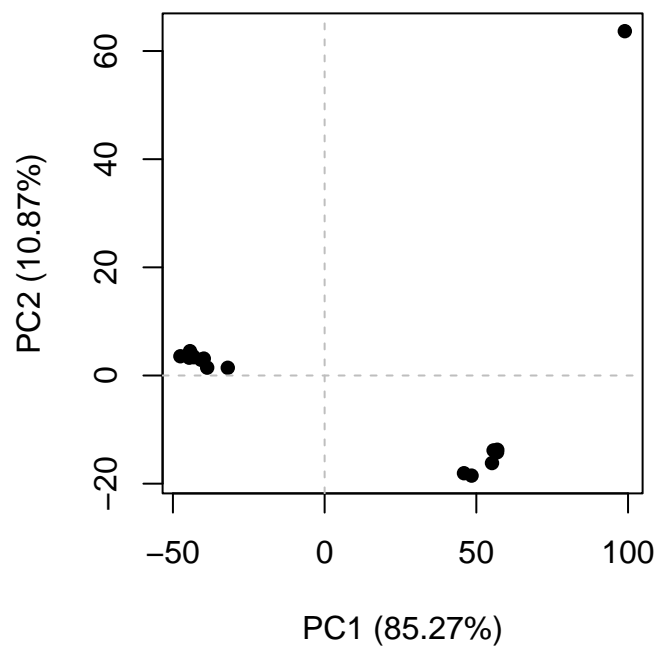



Scree plot: 3 new variables (PC1-3) capture ~98% of the variance. Reduced from 17 to 3 variables!

```
dim(pdb$xyz)
```

```
[1] 17 681
```

```
plot(pc.xray, 1:2)
```



Results of PCA on Adenylate kinase X-ray structures. Each dot represents one PDB structure.

View dynamics (structure -> function):

```
mktrj(pc.xray, file = "pca_results.pdb") # view in Mol*
```

There's not just one structure!