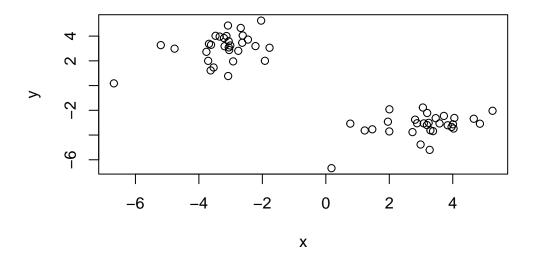
Class 7: machine learning for bioinformatics (part 1)

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k-means clustering

We will start with k-means clustering, one of the most prevalent of all clustering methods. A disadvantage is that you need to know how many clusters there should be (k = ?).

Generate random data:



The main function in R for k-means clustering is called kmeans():

```
k <- kmeans(data, centers = 2, nstart = 20)
k # output is a list of "components"</pre>
```

K-means clustering with 2 clusters of sizes 30, 30

Cluster means:

Clustering vector:

Within cluster sum of squares by cluster: [1] 66.84442 66.84442

(between_SS / total_SS = 89.9 %)

Available components:

```
[1] "cluster" "centers" "totss" "withinss" "tot.withinss" [6] "betweenss" "size" "iter" "ifault"
```

How many points are in each cluster?

```
k[7]
```

\$size

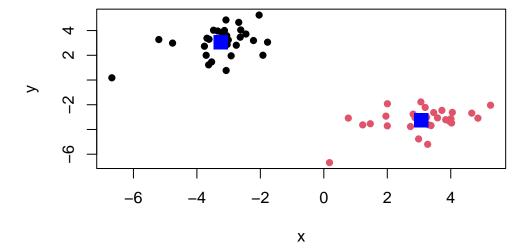
[1] 30 30

k\$size

[1] 30 30

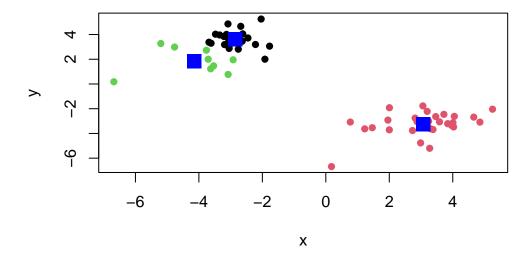
Make a plot of our data colored by clustering results with the cluster centers shown:

```
plot(data, col = k$cluster, pch = 16)
points(k$centers, col = "blue", pch = 15, cex = 2)
```



What if we did 3 clusters?

```
k <- kmeans(data, centers = 3, nstart = 20)
plot(data, col = k$cluster, pch = 16)
points(k$centers, col = "blue", pch = 15, cex = 2)</pre>
```



Hierarchial Clustering

Hierarchical clustering has the advantage that it can reveal the structure in your data rather than imposing a structure (k-means).

The main function in base R is hclust(). It requires a distance matrix as its input, not the data itself:

```
hc <- hclust(dist(data))
hc # output isn't very helpful</pre>
```

Call: hclust(d = dist(data))

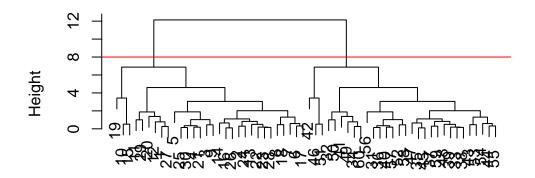
Cluster method : complete
Distance : euclidean

Number of objects: 60

Plot:

```
plot(hc)
# cut the tree
abline(h=8, col="red")
```

Cluster Dendrogram



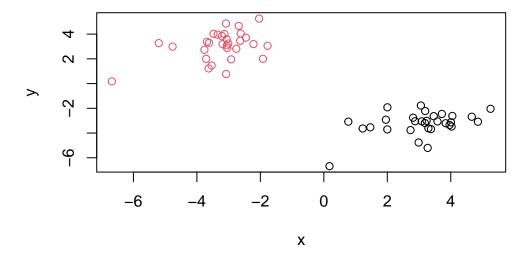
dist(data) hclust (*, "complete")

The function to get our clusters/groups from a hclust object is called cutree():

```
groups <- cutree(hc,h=8)</pre>
```

Plot helust results in terms of our data colored by cluster membership:

```
plot(data, col = groups)
```



Produced the same output, but we didn't need to know groups = 2 beforehand.

Principal Component Analysis (PCA)

(1) PCA of UK food data

Import data:

```
# load
url <- "https://tinyurl.com/UK-foods"
data <- read.csv(url, row.names = 1) # this is better than rownames(x) <- x[,1] approach
# examine data
str(data)

'data.frame': 17 obs. of 4 variables:
$ England : int 105 245 685 147 193 156 720 253 488 198 ...
$ Wales : int 103 227 803 160 235 175 874 265 570 203 ...
$ Scotland : int 103 242 750 122 184 147 566 171 418 220 ...
$ N.Ireland: int 66 267 586 93 209 139 1033 143 355 187 ...</pre>
```

head(data)

	England	Wales	Scotland	N.Ireland
Cheese	105	103	103	66
Carcass_meat	245	227	242	267
Other_meat	685	803	750	586
Fish	147	160	122	93
Fats_and_oils	193	235	184	209
Sugars	156	175	147	139

nrow(data)

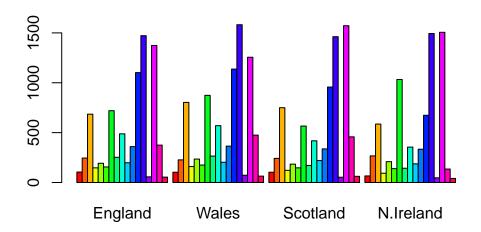
[1] 17

ncol(data)

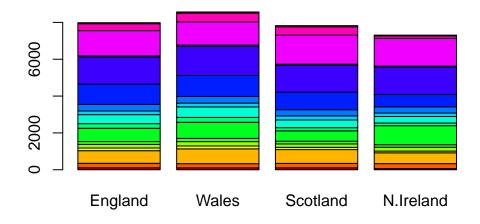
[1] 4

Try to visualize 17 variables with a bar plot:

```
barplot(as.matrix(data), beside=T, col=rainbow(nrow(data)))
```

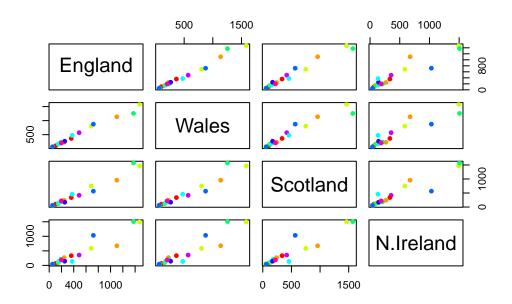


barplot(as.matrix(data), beside=F, col=rainbow(nrow(data)))



Try generating all pairwise plots:

```
pairs(data, col=rainbow(10), pch=16)
```



```
# point on the diagonal means those 2 countries have a similar value for that variable
# the colored points represent the 17 variables (ex: "cheese")
```

These plots are difficult to make sense of. PCA to the rescue!

The main function for PCA in base R is called prcomp():

```
pca <- prcomp(t(data)) # need to transpose because PCA expects variables to be columns
summary(pca)
```

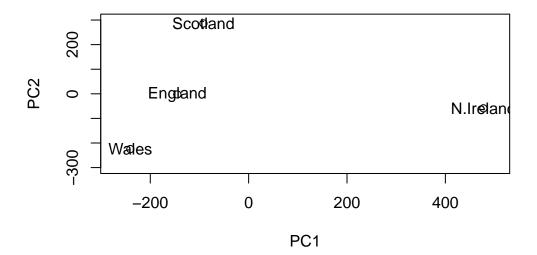
Importance of components:

```
PC1 PC2 PC3 PC4
Standard deviation 324.1502 212.7478 73.87622 2.921e-14
Proportion of Variance 0.6744 0.2905 0.03503 0.000e+00
Cumulative Proportion 0.6744 0.9650 1.00000 1.000e+00
```

PCA output (like k-means) is a list. One of the main PCA results people use is a 2D "score plot" (PC1 vs. PC2):

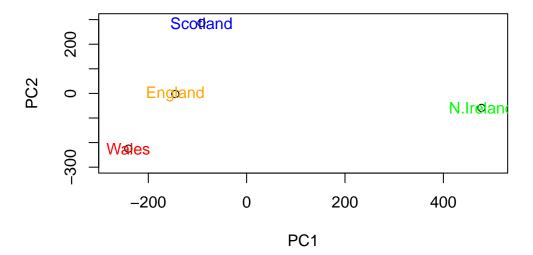
pca\$x

```
PC1
                             PC2
                                        PC3
                                                      PC4
England
          -144.99315
                       -2.532999 105.768945 -9.152022e-15
Wales
          -240.52915 -224.646925 -56.475555 5.560040e-13
Scotland
           -91.86934
                      286.081786 -44.415495 -6.638419e-13
N.Ireland 477.39164 -58.901862 -4.877895
                                             1.329771e-13
  # plot just PC1 vs. PC2 (represent 96.5% of the variance)
  plot(pca$x[,1], pca$x[,2], xlab="PC1", ylab="PC2", xlim=c(-270,500), ylim=c(-300,300))
  text(pca$x[,1], pca$x[,2], colnames(data))
```



Add color:

```
color <- c("orange", "red", "blue", "green")
plot(pca$x[,1], pca$x[,2], xlab="PC1", ylab="PC2", xlim=c(-270,500), ylim=c(-300,300))
text(pca$x[,1], pca$x[,2], colnames(data), col = color)</pre>
```



Variable loadings:

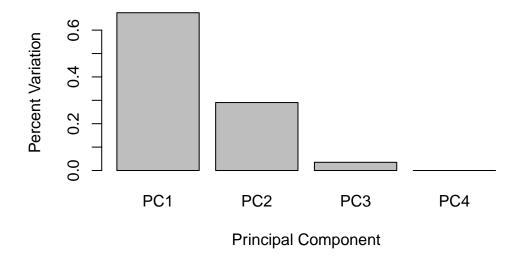
Calculate how much variation in the original data each PC accounts for:

```
s <- summary(pca)
s$importance</pre>
```

	PC1	PC2	PC3	PC4
Standard deviation	324.15019	212.74780	73.87622	2.921348e-14
Proportion of Variance	0.67444	0.29052	0.03503	0.000000e+00
Cumulative Proportion	0.67444	0.96497	1.00000	1.000000e+00

This information can be summarized in a plot of the variances (eigenvalues) with respect to the principal component number (eigenvector number). Plot:

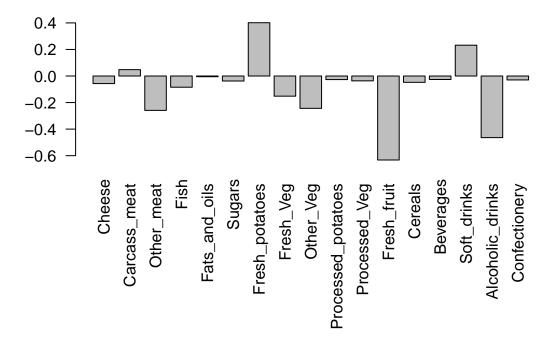
```
barplot(s$importance[2,], xlab="Principal Component", ylab="Percent Variation")
```



We can also consider the influence of each of the original variables upon the principal components (typically known as loading scores). This information can be obtained from the prcomp() returned \$rotation component.

Plot (focus on PC1 since it accounts for >90% of the variance):

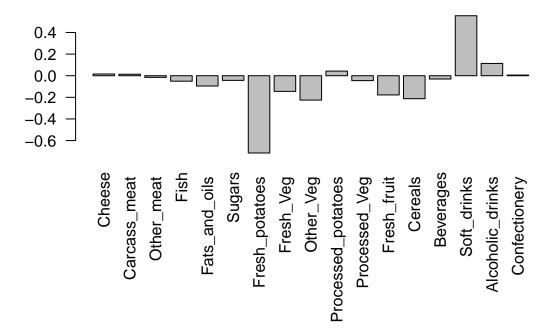
```
par(mar=c(10, 3, 0.35, 0))
barplot( pca$rotation[,1], las=2 )
```



Here we see variables (foods) with the largest positive / negative "loading scores" that effectively "push" observations (countries) to right / left side of the plot.

Generate a similar 'loadings plot' for PC2. What two food groups feature prominently and what does PC2 mainly tell us about?

```
par(mar=c(10, 3, 0.35, 0))
barplot( pca$rotation[,2], las=2 )
```



The largest positive loading score = soft drinks, pushing Scotland to the top of the plot. The largest negative loading score = fresh potatoes, pushing Wales to the bottom of the plot.

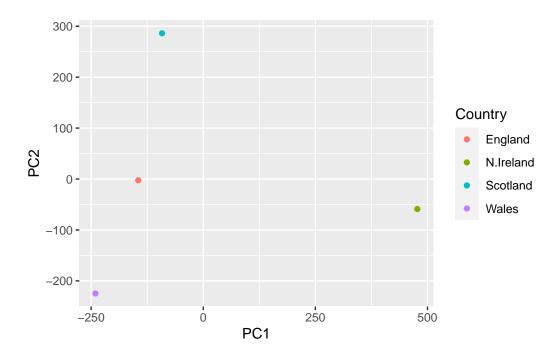
Let's make the score plot figure look nicer with ggplot:

```
library(ggplot2)

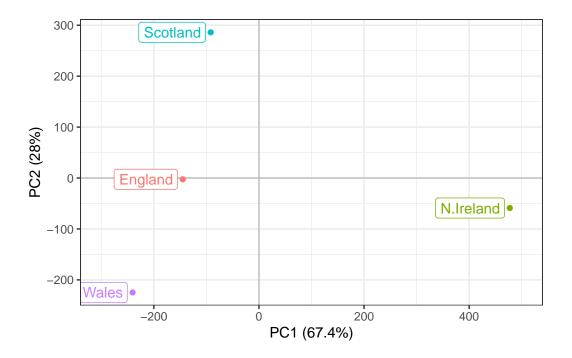
df <- as.data.frame(pca$x)

df_lab <- tibble::rownames_to_column(df, "Country")

# first basic plot
ggplot(df_lab) +
   aes(PC1, PC2, col=Country) +
   geom_point()</pre>
```



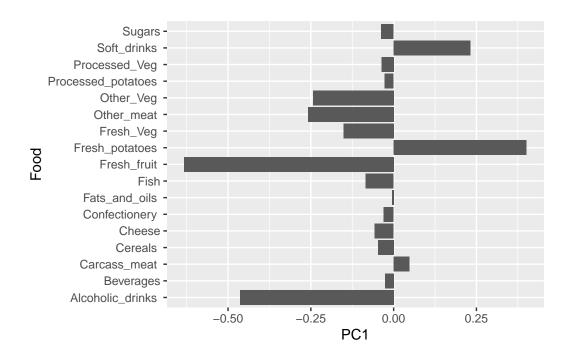
```
# nicer plot
ggplot(df_lab) +
  aes(PC1, PC2, col=Country, label=Country) +
  geom_hline(yintercept = 0, col="gray") +
  geom_vline(xintercept = 0, col="gray") +
  geom_point(show.legend = FALSE) +
  geom_label(hjust=1, nudge_x = -10, show.legend = FALSE) +
  expand_limits(x = c(-300,500)) +
  xlab("PC1 (67.4%)") +
  ylab("PC2 (28%)") +
  theme_bw()
```



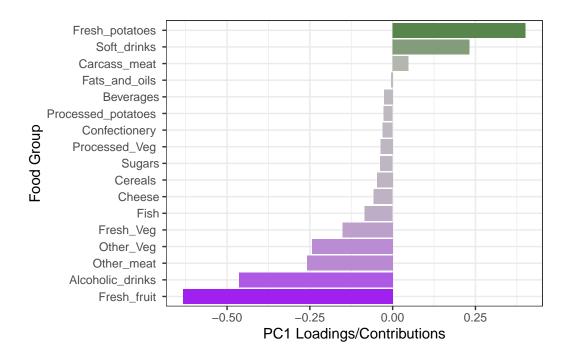
Let's do the same for our loadings/PC contributions figures:

```
ld <- as.data.frame(pca$rotation)
ld_lab <- tibble::rownames_to_column(ld, "Food")

# first basic plot
ggplot(ld_lab) +
   aes(PC1, Food) +
   geom_col()</pre>
```

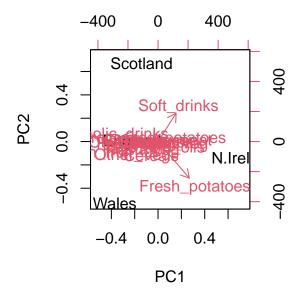


```
# nicer plot
ggplot(ld_lab) +
aes(PC1, reorder(Food, PC1), bg=PC1) +
geom_col() +
xlab("PC1 Loadings/Contributions") +
ylab("Food Group") +
scale_fill_gradient2(low="purple", mid="gray", high="darkgreen", guide=NULL) +
theme_bw()
```



Another way to see this information together with the main PCA plot is in a so-called biplot:

```
## can be useful for small datasets
biplot(pca)
```



Summary: PCA has the awesome ability to be able to make these associations for us. It has also successfully managed to reduce the dimensionality of our data set down from 17 to 2, allowing us to assert (using our figures above) that countries England, Wales and Scotland are 'similar' with Northern Ireland being different in some way. Furthermore, digging deeper into the loadings we were able to associate certain food types with each cluster of countries.

PCA of RNA-seq data