

# Package ‘coMethDMR’

March 4, 2019

**Title** An unsupervised approach for identifying differentially methylated regions in Illumina arrays

**Version** 0.0.0.9000

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**Description** coMethDMR identifies genomic regions associated with continuous phenotypes by optimally leverages covariations among CpGs within predefined genomic regions. Instead of testing all CpGs within a genomic region, coMethDMR carries out an additional step that selects co-methylated sub-regions first without using any outcome information. Next, coMethDMR tests association between methylation within the sub-region and continuous phenotype using a random coefficient mixed effects model, which models both variations between CpG sites within the region and differential methylation simultaneously.

**Depends** R (>= 3.1.0), IlluminaHumanMethylation450kanno.ilmn12.hg19, IlluminaHumanMethylationEPICanno.ilm10b2.hg19

**License** GPL-3

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 6.1.1

**Imports** bumphunter, psych, pathwayPCA, lmerTest, GenomicRanges, IRanges

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betaMatrixChr22_df	<i>Prefrontal Cortex (PFC) Methylation Data from Alzheimer’s Disease subjects</i>
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**Description**

Subset of an Alzheimer’s methylation dataset, with beta values for CpGs.

**Usage**

betaMatrixChr22\_df

**Format**

A data frame containing beta values for 8552 CpGs in Chr22 for a subset of 20 subjects.

**Source**

GEO accession: GSE59685

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betaMatrix_ex1	<i>Alzheimer’s Prefrontal Cortex (PFC) Methylation Data</i>
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**Description**

Subset of an Alzheimer’s Disease methylation data set, with beta values for CpGs.

**Usage**

betaMatrix\_ex1

**Format**

A data frame containing beta values for 4 CpGs in one CpG islands for 110 subjects. Each column is a sample, each row is a CpG.

**Source**

GEO accession: GSE59685

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betaMatrix\_ex2*Alzheimer's Prefrontal Cortex (PFC) Methylation Data*

---

**Description**

Subset of an Alzheimer's Disease methylation data set, with beta values for CpGs.

**Usage**

betaMatrix\_ex2

**Format**

A data frame containing beta values for 4 CpGs in one CpG islands for 110 subjects. Each column is a sample, each row is a CpG.

**Source**

GEO accession: GSE59685

---

betaMatrix\_ex3*Alzheimer's Prefrontal Cortex (PFC) Methylation Data*

---

**Description**

Subset of an Alzheimer's Disease methylation data set, with beta values for CpGs.

**Usage**

betaMatrix\_ex3

**Format**

A data frame containing beta values for 6 CpGs in one CpG islands for 110 subjects. Each column is a sample, each row is a CpG.

**Source**

GEO accession: GSE59685

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betaMatrix_ex4	<i>Alzheimer's Prefrontal Cortex (PFC) Methylation Data</i>
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**Description**

Subset of an Alzheimer's Disease methylation data set, with beta values for CpGs.

**Usage**

```
betaMatrix_ex4
```

**Format**

A data frame containing beta values for 52 CpGs in one CpG islands for 110 subjects. Each column is a sample, each row is a CpG.

**Source**

GEO accession: GSE59685

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CloseBySingleRegion	<i>Extract clusters of CpGs located closely in a genomic region.</i>
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**Description**

Extract clusters of CpGs located closely in a genomic region.

**Usage**

```
CloseBySingleRegion(CpGs_char, arrayType = c("450k", "EPIC"),
  maxGap = 200, minCpGs = 3)
```

**Arguments**

CpGs_char	a list of CpG IDs
arrayType	Type of array, 450k or EPIC
maxGap	an integer, genomic locations within maxGap from each other are placed into the same cluster
minCpGs	an integer, minimum number of CpGs for the resulting CpG cluster

**Details**

Note that this function depends only on CpG locations, and not on any methylation data. The algorithm is based on the [clusterMaker](#) function in the bumhunter R package. Each cluster is essentially a group of CpG locations such that two consecutive locations in the cluster are separated by less than maxGap.

**Value**

a list, each item in the list is a character vector of CpG IDs located closely (i.e. in the same cluster)

**Examples**

```
CpGs_char <- c(
  "cg02505293", "cg03618257", "cg04421269", "cg17885402", "cg19890033",
  "cg20566587", "cg27505880"
)

cluster_ls <- CloseBySingleRegion(
  CpGs_char, arrayType = "450k", maxGap = 100, minCpGs = 3
)
```

---

CoMethAllRegions	<i>Extract contiguous co-methylated genomic regions from a list of pre-defined genomic regions</i>
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---

**Description**

Extract contiguous co-methylated genomic regions from a list of pre-defined genomic regions

**Usage**

```
CoMethAllRegions(betaMatrix, regionType = c("ISLAND", "NSHORE", "NSHELF",
  "SSHORE", "SSHELF", "TSS1500", "TSS200", "UTR5", "EXON1", "GENEBODY",
  "UTR3"), arrayType = c("450k", "EPIC"), file = NULL,
  fileType = c("gmt", "RDS"), rDropThresh_num = 0.4,
  returnAllCpGs = FALSE, ...)
```

**Arguments**

betaMatrix	matrix (or data frame) of beta values, with row names = CpG IDs, column names = sample IDs. This is typically genome-wide methylation beta values.
regionType	Type of input genomic regions (e.g. "ISLAND" for CpG island)
arrayType	Type of array, can be "450k" or "EPIC"
file	an RDS or gmt file with clusters of CpG locations (i.e. CpGs located closely to each other on the genome). This file can be generated by the <a href="#">WriteCloseByAllRegions</a> function. If RDS file, it would contain a list, where each item is a character vector of CpGs IDs.
fileType	file extension for input file, can be "gmt" or "RDS"
rDropThresh_num	threshold for min correlation between a cpG with sum of the rest of the CpGs
returnAllCpGs	When there is not a contiguous comethylated region in the inputing pre-defined region, returnAllCpGs = 1 indicates outputting all the CpGs in the input regions, while returnAllCpGs = 0 indicates not returning any CpG.
...	Dots for internal arguments. Currently unused.

**Value**

A list of two components:

- **Contiguous\_Regions** : A data frame with CpG (CpG name), Chr (chromosome number), MAPINFO (genomic position), r\_drop(correlation between the CpG with rest of the CpGs), keep (indicator for co-methylated CpG), keep\_contiguous (index for contiguous comethylated subregions)
- **CpGsSubregions** : results from all the regions, each item is a list of CpGs in the contiguous co-methylated subregion

**Examples**

```
data(betaMatrixChr22_df)

exampleFile <- readRDS(
  system.file ("extdata",
              "CpGislandsChr22_ex.RDS",
              package = 'coMethDMR',
              mustWork = TRUE
            )
)

CoMethAllRegions (
  betaMatrix = betaMatrixChr22_df,
  file = exampleFile,
  fileType = "RDS",
  arrayType = "450k",
  returnAllCpGs = FALSE
)

## Not run:

CoMethAllRegions (
  betaMatrix = betaMatrixChr22_df,
  regionType = "ISLAND",
  arrayType = "450k",
  returnAllCpGs = FALSE
)

## End(Not run)
```

---

CoMethSingleRegion	<i>Wrapper function to find contiguous and comethylated sub-regions within a pre-defined genomic region</i>
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**Description**

Wrapper function to find contiguous and comethylated sub-regions within a pre-defined genomic region

**Usage**

```
CoMethSingleRegion(CpGs_char, betaMatrix, rDropThresh_num = 0.4,
  minCpGs = 3, arrayType = c("450k", "EPIC"), returnAllCpGs = FALSE)
```

**Arguments**

CpGs_char	vector of CpGs in the inputting pre-defined genomic region.
betaMatrix	matrix (or data frame) of beta values, with row names = CpG ids, column names = sample ids. This should include the CpGs in CpGs_char, as well as additional CpGs.
rDropThresh_num	threshold for min correlation between a cpG with sum of the rest of the CpGs
minCpGs	minimum number of CpGs to be considered a "region". Only regions with more than minCpGs will be returned.
arrayType	Type of array, can be "450k" or "EPIC"
returnAllCpGs	When there is not a contiguous comethylated region in the inputting pre-defined region, returnAllCpGs = 1 indicates outputting all the CpGs in the input region, while returnAllCpGs = 0 indicates not returning any CpG.

**Value**

A list with two components:

- **Contiguous\_Regions**: a data frame with CpG (CpG ID), Chr (chromosome number), MAPINFO (genomic position), r\_drop (correlation between the CpG with rest of the CpGs), keep (indicator for co-methylated CpG), keep\_contiguous (index for contiguous comethylated subregion)
- **CpGs\_subregions**: lists of CpGs in each contiguous co-methylated subregion

**Examples**

```
data(betaMatrixChr22_df)
CpGsChr22_char <- c(
  "cg02953382", "cg12419862", "cg24565820", "cg04234412", "cg04824771",
  "cg09033563", "cg10150615", "cg18538332", "cg20007245", "cg23131131",
  "cg25703541"
)
CoMethSingleRegion(
  CpGs_char = CpGsChr22_char,
  betaMatrix = betaMatrixChr22_df
)

data(betaCluster_mat_example3)
betaMatrix_ex3 <- t(betaMatrix_ex3)
CpGsEx3_char <- c(
  "cg14221598", "cg02433884", "cg07372974", "cg13419809", "cg26856676",
  "cg25246745"
)
CoMethSingleRegion(
  CpGs_char = CpGsEx3_char,
  betaMatrix = betaMatrix_ex3,
  returnAllCpGs = TRUE
)
```

---

CpGsInfoAllRegions	<i>Test associations of individual CpGs in multiple genomic regions with a continuous phenotype</i>
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---

## Description

Test associations of individual CpGs in multiple genomic regions with a continuous phenotype

## Usage

```
CpGsInfoAllRegions(AllRegionNames_char, betas_df, pheno_df, contPheno_char,
  covariates_char, arrayType = c("450k", "EPIC"))
```

## Arguments

AllRegionNames_char	vector of character strings with location info for all the genomic regions. Each region should be specified in this format: "chrxx:xxxxxx-xxxxxx"
betas_df	data frame of beta values for all genomic regions, with row names = CpG IDs, column names = sample IDs
pheno_df	a data frame with phenotype and covariate variables, with variable "Sample" for sample IDs.
contPheno_char	character string of the continuous phenotype, to be tested against methylation values
covariates_char	character vector of covariate variables names
arrayType	Type of array, can be "450k" or "EPIC"

## Value

a data frame with locations of the genomic region (Region), CpG ID (cpg), chromosome (chr), position (pos), and results for testing association of methylation in individual CpGs with continuous phenotype (slopeEstimate, slopePval)

## Examples

```
data(betaMatrixChr22_df)
data(pheno_df)
AllRegionNames_char <- c("chr22:18267969-18268249", "chr22:18531243-18531447")

CpGsInfoAllRegions(
  AllRegionNames_char,
  betas_df = betaMatrixChr22_df,
  pheno_df, contPheno_char = "stage",
  covariates_char = c("age.brain", "sex")
)
```



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CpGsInfoOneRegion	<i>Test associations of individual CpGs in a genomic region with a continuous phenotype</i>
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---

## Description

Test associations of individual CpGs in a genomic region with a continuous phenotype

## Usage

```
CpGsInfoOneRegion(regionName_char, betas_df, pheno_df, contPheno_char,
  covariates_char, arrayType = c("450k", "EPIC"))
```

## Arguments

regionName_char	character string of location information for a genomic region, specified in the format of "chrxx:xxxxxx-xxxxxx"
betas_df	data frame of beta values with row names = CpG IDs, column names = sample IDs
pheno_df	a data frame with phenotype and covariate variables, with variable "Sample" for sample IDs.
contPheno_char	character string of the continuous phenotype, to be tested against methylation values
covariates_char	character vector of covariate variables names
arrayType	Type of array, can be "450k" or "EPIC"

## Details

This function implements linear models that test association between methylation values in a genomic region with a continuous phenotype. Note that methylation M values are used as regression outcomes in these models. The model for each CpG is:

methylation M value ~ contPheno\_char + covariates\_char

## Value

a data frame with location of the genomic region (Region), CpG ID (cpg), chromosome (chr), position (pos), results for testing association of methylation in individual CpGs with continuous phenotype (slopeEstimate, slopePval) and annotations for the regions

## Examples

```
data(betaMatrixChr22_df)
data(pheno_df)

CpGsInfoOneRegion(
  regionName_char = "chr22:19709548-19709755",
  betas_df = betaMatrixChr22_df,
  pheno_df, contPheno_char = "stage",
  covariates_char = c("age.brain", "sex"),
```

```

    arrayType = "450k"
  )

  # not adjusting for covariates
  CpGsInfoOneRegion(
    regionName_char = "chr22:18267969-18268249",
    betas_df = betaMatrixChr22_df,
    pheno_df, contPheno_char = "stage",
    covariates_char = NULL
  )

```

---

## FindComethylatedRegions

*Find contiguous comethylated regions based on output file from function MarkComethylatedCpGs*

---

### Description

Find contiguous comethylated regions based on output file from function MarkComethylatedCpGs

### Usage

```
FindComethylatedRegions(CpGs_df, minCpGs_int = 3)
```

### Arguments

CpGs_df	an output dataframe from function MarkComethylatedCpGs, with variables CpG, keep, ind, r_drop. See details in documentation for MarkComethylatedCpGs.
minCpGs_int	an integer, indicates minimum nubmer of CpGs for output genomic regions

### Value

A data frame with variables ProbeID and Subregion (index for each output contiguous comethylated regions)

### Examples

```

data(betaMatrix_ex4)

CpGs_df <- MarkComethylatedCpGs(betaCluster_mat = betaMatrix_ex4)

FindComethylatedRegions(CpGs_df)

```

---

GetCpGsInRegion	<i>Extract probe IDs for CpGs located in a genomic region</i>
-----------------	---

---

**Description**

Extract probe IDs for CpGs located in a genomic region

**Usage**

```
GetCpGsInRegion(regionName_char, arrayType = c("450k", "EPIC"))
```

**Arguments**

regionName_char	character string with location information for one region in this format: "chrxx:xxxxxx-xxxxxx"
arrayType	Type of array, 450k or EPIC

**Value**

vector of CpG probe IDs mapped to the genomic region

**Examples**

```
GetCpGsInRegion(
  regionName_char = "chr22:18267969-18268249",
  arrayType = "450k"
)
```

---

lmmTest	<i>Fit mixed model to methylation values in one genomic region</i>
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---

**Description**

Fit mixed model to methylation values in one genomic region

**Usage**

```
lmmTest(betaOne_df, pheno_df, contPheno_char, covariates_char,
  modelType = c("randCoef", "simple"), arrayType = c("450k", "EPIC"))
```

**Arguments**

betaOne_df	matrix of beta values for one genomic region, with row names = CpG IDs, column names = sample IDs
pheno_df	a data frame with phenotype and covariates, with variable Sample indicating sample IDs.
contPheno_char	character string of the main effect (a continuous phenotype) to be tested for association with methylation values in the region

covariates_char	character vector for names of the covariate variables
modelType	type of mixed model, can be randCoef for random coefficient mixed model, or simple for simple linear mixed model.
arrayType	Type of array, can be "450k" or "EPIC"

## Details

This function implements a mixed model to test association between methylation values in a genomic region with a continuous phenotype.

When randCoef is selected, the model is

methylation M value ~ contPheno\_char + covariates\_char + (1|Sample) + (contPheno\_char|CpG).  
The last two terms are random intercepts and slopes for each CpG.

When simple is selected, the model is

methylation M value ~ contPheno\_char + covariates\_char + (1|Sample)

In our simulation studies, we found both models are conservative, so p-values are estimated from normal distributions instead of t-distributions.

## Value

A dataframe with one row for association result of one region: Estimate, StdErr, and pvalue for the association of methylation values in the genomic region tested vs. continuous phenotype contPheno\_char

## Examples

```
data(betaMatrixChr22_df)

CpGsChr22_char<-c("cg02953382", "cg12419862", "cg24565820", "cg04234412",
  "cg04824771", "cg09033563", "cg10150615", "cg18538332", "cg20007245",
  "cg23131131", "cg25703541")

coMethCpGs <- CoMethSingleRegion(CpGsChr22_char, betaMatrixChr22_df)

# test only the first co-methylated region
coMethBetaMatrix <- betaMatrixChr22_df[coMethCpGs$CpGsSubregions[[1]], ]

data(pheno_df)

res <- ImmTest (betaOne_df = coMethBetaMatrix,
  pheno_df,
  contPheno_char = "stage",
  covariates_char = c("age.brain", "sex"),
  modelType = "randCoef",
  arrayType = "450k")
```

---

ImmTestAllRegions	<i>Fit mixed model to test association between a continuous phenotype and methylation values in a list of genomic regions</i>
-------------------	---

---

## Description

Fit mixed model to test association between a continuous phenotype and methylation values in a list of genomic regions

## Usage

```
ImmTestAllRegions(beta_df, region_ls, pheno_df, contPheno_char,
  covariates_char, modelType = c("randCoef", "simple"),
  arrayType = c("450k", "EPIC"), outFile = NULL)
```

## Arguments

beta_df	data frame of beta values for all genomic regions, with row names = CpG IDs, column names = sample IDs. This is often the genome-wide array data.
region_ls	a list of genomic regions, each item is a vector of CpG IDs within a genomic region. The co-methylated regions can be obtained by function CoMethAllRegions.
pheno_df	a data frame with phenotype and covariates, with variable Sample indicating sample IDs.
contPheno_char	character string of the main effect (a continuous phenotype) to be tested for association with methylation values in each region
covariates_char	character vector for names of the covariate variables
modelType	type of mixed model, can be randCoef for random coefficient mixed model, or simple for simple linear mixed model.
arrayType	Type of array, can be "450k" or "EPIC"
outFile	output .csv file with the results for the mixed model analysis

## Value

csv file with location of the genomic region (chrom, start, end), number of CpGs (nCpGs), Estimate, Standard error (StdErr) of the test statistic, p-value and False Discovery Rate (FDR) for association between methylation values in each genomic region with phenotype (pValue).

## Examples

```
data(betaMatrixChr22_df)

data(pheno_df)

CpGisland_ls <- system.file(
  "extdata", "CpGislandsChr22_ex.RDS",
  package = 'coMethDMR', mustWork = TRUE
)

coMeth_ls <- CoMethAllRegions(
```

```

        betaMatrix = betaMatrixChr22_df,
        file = CpGisland_ls,
        fileType = "RDS",
        arrayType = "450k",
        rDropThresh_num = 0.4,
        returnAllCpGs = FALSE
    )

lmmTestAllRegions(
  beta_df = betaMatrixChr22_df,
  region_ls = coMeth_ls$CpGsSubregions,
  pheno_df,
  contPheno_char = "stage",
  covariates_char = "age.brain",
  modelType = "randCoef",
  arrayType = "450k"
)

```

---

MarkComethylatedCpGs    *Mark CpGs in contiguous and co-methylated region*

---

## Description

Mark CpGs in contiguous and co-methylated region

## Usage

```
MarkComethylatedCpGs(betaCluster_mat, rDropThresh_num = 0.4)
```

## Arguments

**betaCluster\_mat**  
matrix of beta values, with rownames = sample ids, column names = CpG ids. Note that the CpGs need to be ordered by their genomic positions, this can be accomplished by the `OrderCpGbyLocation` function.

**rDropThresh\_num**  
threshold for min correlation between a cpG with sum of the rest of the CpGs

## Details

An outlier CpG in a genomic region will typically have low correlation with the rest of the CpGs in a genomic region. On the other hand, in a cluster of co-methylated CpGs, we expect each CpG to have high correlation with the rest of the CpGs. The `r.drop` statistic is used to identify these co-methylated CpGs here.

## Value

A data frame with the following columns:

- `CpG` : CpG ID
- `keep` : The CpGs with `keep = 1` belong to the contiguous and co-methylated region
- `ind` : Index for the CpGs
- `r_drop` : The correlation between each CpG with the sum of the rest of the CpGs

**Examples**

```

data(betaMatrix_ex1)
MarkComethylatedCpGs(betaCluster_mat = betaMatrix_ex1)

data(betaMatrix_ex2)
MarkComethylatedCpGs(betaCluster_mat = betaMatrix_ex2)

data(betaMatrix_ex3)
MarkComethylatedCpGs(betaCluster_mat = betaMatrix_ex3)

data(betaMatrix_ex4)
MarkComethylatedCpGs(betaCluster_mat = betaMatrix_ex4, rDropThresh_num = 0.6)

```

---

NameRegion	<i>Name a region with several CpGs based on its genomic location</i>
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---

**Description**

Name a region with several CpGs based on its genomic location

**Usage**

```
NameRegion(CpGsOrdered_df)
```

**Arguments**

CpGsOrdered\_df dataframe with columns for Probe IDs as character (cpg), chromosome number as character (chr) and genomic location as integer (pos)

**Value**

genome location of the CpGs, in the format of "chrxx:xxxxxx-xxxxxx"

**Examples**

```

CpGs_char <- c("cg04677227", "cg07146435", "cg11632906", "cg20214853")
CpGsOrdered_df <- OrderCpGsByLocation(CpGs_char, arrayType=c("EPIC"), output = "dataframe")
NameRegion(CpGsOrdered_df)

```

---

OrderCpGsByLocation	<i>Order CpGs by genomic location</i>
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---

**Description**

Order CpGs by genomic location

**Usage**

```
OrderCpGsByLocation(CpGs_char, arrayType = c("450k", "EPIC"),
  output = c("vector", "dataframe"))
```

**Arguments**

CpGs_char	vector of CpGs
arrayType	Type of array, 450k or EPIC
output	vector of CpGs or dataframe with CpGs, CHR, MAPINFO

**Value**

vector of CpGs ordered by location or dataframe with CpGs ordered by location (cpg), chromosome (chr), position (pos)

**Examples**

```
CpGs_char <- c("cg04677227", "cg07146435", "cg11632906", "cg20214853")
OrderCpGsByLocation(CpGs_char, arrayType=c("EPIC"), output = "dataframe")
```

---

pheno_df	<i>Example phenotype data file from Prefrontal Cortex (PFC) Methylation Data of Alzheimer's Disease subjects</i>
----------	--

---

**Description**

Subset of phenotype information for Alzheimer's methylation dataset.

**Usage**

```
pheno_df
```

**Format**

A data frame containing variables for Braak stage (stage), subject.id, Batch (Mplate), Sex, Sample, age of brain sample (age.brain)

**Source**

GEO accession: GSE59685



RegionsToRanges

*Convert genomic regions in a data frame to GRanges format***Description**

Convert genomic regions in a data frame to GRanges format

**Usage**

```
RegionsToRanges(regionName_char)
```

**Arguments**

regionName\_char

a character vector of regions, in this format: "chrxx:xxxxxx-xxxxxx"

**Value**

genomic regions in GRanges format

**Examples**

```
regions = c("chr22:19709548-19709755", "chr2:241721922-241722113")
RegionsToRanges (regions)
```

WriteCloseByAllRegions

*Extract clusters of close by CpGs from a list of pre-defined genomic regions***Description**

Extract clusters of close by CpGs from a list of pre-defined genomic regions

**Usage**

```
WriteCloseByAllRegions(file, regionType = c("ISLAND", "NSHORE", "NSHELF",
      "SSHORE", "SSHELF", "TSS1500", "TSS200", "UTR5", "EXON1", "GENEBODY",
      "UTR3"), arrayType = c("450k", "EPIC"), maxGap = 200, minCpGs = 3,
      fileType = c("gmt", "RDS"), ...)
```

**Arguments**

file	file where the output genomic regions will be saved. File extension should not be supplied, it is automatically added via the fileType argument.
regionType	Type of input genomic regions (e.g. "ISLAND" for CpG island)
arrayType	Type of array, can be "450k" or "EPIC"
maxGap	an integer, genomic locations within maxGap from each other are placed into the same cluster

<code>minCpGs</code>	an integer, minimum number of CpGs for each resulting region
<code>fileType</code>	the output files can be saved as .gmt or .RDS.
<code>...</code>	Dots for internal arguments. Currently unused.

### Details

For `maxGap = 200` and `minCpGs = 3`, we already calculated the clusters of CpGs. They are saved in folder `/inst/extdata/`.

Note that for output files, .gmt files can be opened as flat text file. .RDS files are half the size of .gmt files, but they can only be read in the R environment.

Creating and writing the file for one type of genomic region (`regionType = "ISLAND"`) took about 25 minutes.

### Value

a file with the genomic regions containing CpGs located closely within each inputting pre-defined genomic region

### Examples

```
## Not run:
  CloseByAllRegions(
    regionType = "ISLAND", arrayType = "450k", maxGap = 50,
    minCpGs = 3, fileType = "gmt", file = "closeByRegions"
  )

## End(Not run)
```

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