

Mutational signatures

Code ▾

Hide

```
library(readr)
```

Warning: package 'readr' was built under R version 4.1.2

Load VCF samples

Hide

```
# Load InDels
indel_grl <- read_vcfs_as_granges(indel_vcf_files, group_names, ref_genome, type = "i
ndel")
indel_grl
```

GRangesList object of length 84:

\$`WT-day0_chr1`

GRanges object with 449 ranges and 5 metadata columns:

seqnames			ranges	strand	paramRangeID
REF	ALT	QUAL			
		<Rle>	<IRanges>	<Rle>	<factor> <DNAStrings
et> <DNAStringSetList>	<numeric>				
chr1:3555815_GTC/G	chr1	3555815-3555817	*	NA	
GTC	G	55.12			
chr1:4009853_G/GA	chr1	4009853	*	NA	
G	GA	66.12			
chr1:4624693_CA/C	chr1	4624693-4624694	*	NA	
CA	C	47.12			
chr1:4851624_TA/T	chr1	4851624-4851625	*	NA	
TA	T	47.12			
chr1:4894076_TTG/T	chr1	4894076-4894078	*	NA	
TTG	T	61.12			
...
...			
chr1:194095071_GGT/G	chr1	194095071-194095073	*	NA	
GGT	G	48.12			
chr1:194301892_TTC/T	chr1	194301892-194301894	*	NA	
TTC	T	44.12			
chr1:194365483_CT/C	chr1	194365483-194365484	*	NA	
CT	C	43.12			
chr1:194846624_GC/G	chr1	194846624-194846625	*	NA	
GC	G	41.02			

```

chr1:194852291_AAC/A      chr1 194852291-194852293      * |      NA
AAC                        A      50.02
                        FILTER
                        <character>
chr1:3555815_GTC/G      PASS
chr1:4009853_G/GA      PASS
chr1:4624693_CA/C      PASS
chr1:4851624_TA/T      PASS
chr1:4894076_TTG/T      PASS
                        ...
chr1:194095071_GGT/G      PASS
chr1:194301892_TTC/T      PASS
chr1:194365483_CT/C      PASS
chr1:194846624_GC/G      PASS
chr1:194852291_AAC/A      PASS
-----
seqinfo: 21 sequences from mm10 genome

...
<83 more elements>

```

[Hide](#)

```

sample_names <- c("WT-day0", "WT-day77", "Pms1-day0", "Pms1-day77")

snpcvcf_files <- c("./cov_x5/WT-day0_tk39_filtered_snps_x5.vcf",
  "./cov_x5/WT-day77_tk39_filtered_snps_x5.vcf",
  "./cov_x5/Pms1-day0_tk39_filtered_snps_x5.vcf",
  "./cov_x5/Pms1-day77_tk39_filtered_snps_x5.vcf")
indelvcf_files <- c("./cov_x5/WT-day0_tk39_filtered_indels_x5.vcf",
  "./cov_x5/WT-day77_tk39_filtered_indels_x5.vcf",
  "./cov_x5/Pms1-day0_tk39_filtered_indels_x5.vcf",
  "./cov_x5/Pms1-day77_tk39_filtered_indels_x5.vcf")
group_names <- c("WT-day0", "WT-day77", "Pms1-day0", "Pms1-day77")

ref_genome <- "BSgenome.Mmusculus.UCSC.mm10"
library(ref_genome, character.only = TRUE)

groups <- c("WT-day0", "WT-day77", "Pms1-day0", "Pms1-day77")
genotype <- c(rep("WT", 2), rep("Pms1", 2))
time <- c(rep("day0", 1), rep("day77", 1), rep("day0", 1), rep("day77", 1))

# Load SNPs
snp_grl <- read_vcfs_as_granges(snpvcf_files, group_names, ref_genome)

```

Any neighbouring SNVs will be merged into DBS/MBS variants.
Set the 'predefined_dbs_mbs' to 'TRUE' if you don't want this.

Hide

snp_grl

GRangesList object of length 4:
\$`WT-day0`
GRanges object with 537 ranges and 5 metadata columns:

ALT	QUAL	seqnames	ranges	strand	paramRangeID	REF
		<Rle>	<IRanges>	<Rle>	<factor>	<DNAStrngSet> <DNAStrin
		gSetList> <numeric>				
	chr1:19143048_T/A	chr1	19143048	*	NA	T
A	44.75					
	chr1:26687461_G/A	chr1	26687461	*	NA	G
A	83.03					
	chr1:33853419_A/G	chr1	33853419	*	NA	A
G	41.75					
	chr1:33853432_A/T	chr1	33853432	*	NA	A
T	47.82					
	chr1:53208190_A/C	chr1	53208190	*	NA	A
C	41.75					

...	...					
	chrY:1364719_C/A	chrY	1364719	*	NA	C
A	50.75					
	chrY:90744547_G/C	chrY	90744547	*	NA	G
C	446.10					
	chrY:90744549_G/C	chrY	90744549	*	NA	G
C	446.10					
	chrY:90744585_G/A	chrY	90744585	*	NA	G
A	206.08					
	chrY:90744601_G/A	chrY	90744601	*	NA	G
A	119.18					
		FILTER				
		<character>				
	chr1:19143048_T/A	PASS				
	chr1:26687461_G/A	QDlow				
	chr1:33853419_A/G	PASS				
	chr1:33853432_A/T	PASS				
	chr1:53208190_A/C	PASS				
				
	chrY:1364719_C/A	PASS				
	chrY:90744547_G/C	PASS				

```
chrY:90744549_G/C      PASS
chrY:90744585_G/A      PASS
chrY:90744601_G/A      PASS
-----
seqinfo: 21 sequences from mm10 genome

...
<3 more elements>
```

Hide

```
# Load InDels
indel_grl <- read_vcfs_as_granges(indel_vcf_files, group_names, ref_genome, type = "i
ndel")
indel_grl
```

GRangesList object of length 4:
\$`WT-day0`
GRanges object with 686 ranges and 5 metadata columns:

	seqnames	ranges	strand	paramRangeID
REF	<Rle>	<IRanges>	<Rle>	<factor> <DNAS
tringSet>				
GA	chr1:6923386_GA/G	chr1 6923386–6923387	*	NA
T	chr1:6923627_T/TA	chr1 6923627	*	NA
T	chr1:16437642_T/TGAGGAGGAG	chr1 16437642	*	NA
TTGC	chr1:16496611_TTGC/T	chr1 16496611–16496614	*	NA
GAAA	chr1:16648011_GAAA/G	chr1 16648011–16648014	*	NA
...
GGA	chrX:113064791_GGA/G	chrX 113064791–113064793	*	NA
TA	chrX:139823014_TA/T	chrX 139823014–139823015	*	NA
G	chrX:157341920_G/GT	chrX 157341920	*	NA
TC	chrY:90744554_TC/T	chrY 90744554–90744555	*	NA
CCCTAG	chrY:90744588_CCCTAG/C	chrY 90744588–90744593	*	NA
		ALT	QUAL	FILTER

```

                                <DNAStringSetList> <numeric> <character>
      chr1:6923386_GA/G                G      44.84      PASS
      chr1:6923627_T/TA                TA      61.72      PASS
chr1:16437642_T/TGAGGAGGAG          TGAGGAGGAG    214.28      PASS
      chr1:16496611_TTGC/T              T       40.95      PASS
      chr1:16648011_GAAA/G              G       35.73      PASS
      ...                               ...         ...         ...
chrX:113064791_GGA/G                G      159.71      PASS
chrX:139823014_TA/T                 T       45.28      PASS
chrX:157341920_G/GT                  GT       59.75      PASS
chrY:90744554_TC/T                  T      443.85      PASS
chrY:90744588_CCCTAG/C              C      204.85      PASS
-----
seqinfo: 21 sequences from mm10 genome

...
<3 more elements>

```

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```

sample_names <- c("WT-day0", "WT-day77", "Pms1-day0", "Pms1-day77")

snp_vcf_files <- c("./cov_x2/WT-day0_tk39_filtered_snps.vcf",
  "./cov_x2/WT-day77_tk39_filtered_snps.vcf",
  "./cov_x2/Pms1-day0_tk39_filtered_snps.vcf",
  "./cov_x2/Pms1-day77_tk39_filtered_snps.vcf")
indel_vcf_files <- c("./cov_x2/WT-day0_tk39_filtered_indels.vcf",
  "./cov_x2/WT-day77_tk39_filtered_indels.vcf",
  "./cov_x2/Pms1-day0_tk39_filtered_indels.vcf",
  "./cov_x2/Pms1-day77_tk39_filtered_indels.vcf")
group_names <- c("WT-day0", "WT-day77", "Pms1-day0", "Pms1-day77")

ref_genome <- "BSgenome.Mmusculus.UCSC.mm10"
library(ref_genome, character.only = TRUE)

groups <- c("WT-day0", "WT-day77", "Pms1-day0", "Pms1-day77")
genotype <- c(rep("WT", 2), rep("Pms1", 2))
time <- c(rep("day0", 1), rep("day77", 1), rep("day0", 1), rep("day77", 1))

# Load SNPs
snp_grl <- read_vcfs_as_granges(snp_vcf_files, group_names, ref_genome)

```

Any neighbouring SNVs will be merged into DBS/MBS variants.
 Set the 'predefined_dbs_mbs' to 'TRUE' if you don't want this.

[Hide](#)

snp_grl

\$`WT-day0`

		seqnames	ranges	strand	paramRangeID	REF
ALT	QUAL					
		<Rle>	<IRanges>	<Rle>	<factor>	<DNAStringSet>
	<numeric>					<DNAStringSet>
A	40.18	chr1:3425962_C/A	chr1 3425962	*	NA	C
C	40.18	chr1:3512366_T/C	chr1 3512366	*	NA	T
C	40.18	chr1:3559604_T/C	chr1 3559604	*	NA	T
T	40.18	chr1:3932140_G/T	chr1 3932140	*	NA	G
T	40.18	chr1:4108374_C/T	chr1 4108374	*	NA	C
...
...
C	446.10	chrY:90744547_G/C	chrY 90744547	*	NA	G
C	446.10	chrY:90744549_G/C	chrY 90744549	*	NA	G
A	206.08	chrY:90744585_G/A	chrY 90744585	*	NA	G
A	119.18	chrY:90744601_G/A	chrY 90744601	*	NA	G
C	37.07	chrY:90805307_G/C	chrY 90805307	*	NA	G

```
<character>
```

• • • • •

```
seqinfo: 21 sequences from mm10 genome

...
<3 more elements>
```

Hide

```
# Load InDels
indel_grl <- read_vcfs_as_granges(indel_vcf_files, group_names, ref_genome, type = "indel")
indel_grl
```

```
GRangesList object of length 4:
$`WT-day0`
GRanges object with 6029 ranges and 5 metadata columns:
```

	seqnames	ranges	strand	paramRangeID	
REF	<Rle>	<IRanges>	<Rle>	<factor>	<DNAStringS
et>					
GTC	chr1:3555815_GTC/G	chr1 3555815-3555817	*	NA	
G	chr1:4009853_G/GA	chr1 4009853	*	NA	
CA	chr1:4624693_CA/C	chr1 4624693-4624694	*	NA	
TA	chr1:4851624_TA/T	chr1 4851624-4851625	*	NA	
TTG	chr1:4894076_TTG/T	chr1 4894076-4894078	*	NA	
...
AAC	chrY:39960747_AAC/A	chrY 39960747-39960749	*	NA	
CTG	chrY:76250767_CTG/C	chrY 76250767-76250769	*	NA	
GGA	chrY:86118528_GGA/G	chrY 86118528-86118530	*	NA	
TC	chrY:90744554_TC/T	chrY 90744554-90744555	*	NA	
TAG	chrY:90744588_CCCTAG/C	chrY 90744588-90744593	*	NA	CCC

	ALT	QUAL	FILTER
	<DNAStringSetList>	<numeric>	<character>
chr1:3555815_GTC/G	G	55.12	PASS
chr1:4009853_G/GA	GA	66.12	PASS
chr1:4624693_CA/C	C	47.12	PASS

```

chr1:4851624_TA/T      T      47.12      PASS
chr1:4894076_TTG/T     T      61.12      PASS
...
chrY:39960747_AAC/A    A      60.98      PASS
chrY:76250767_CTG/C    C      44.12      PASS
chrY:86118528_GGA/G    G      58.12      PASS
chrY:90744554_TC/T     T      443.85     PASS
chrY:90744588_CCCTAG/C C      204.85     PASS
-----
seqinfo: 21 sequences from mm10 genome

```

```

...
<3 more elements>

```

Remove filtered sites

Hide

```

snp_grl_pass <- list()
indel_grl_pass <- list()
for (i in names(snp_grl)){
  snp_grl_pass[[i]] <- subset(snp_grl[[i]], FILTER == "PASS")
  indel_grl_pass[[i]] <- subset(indel_grl[[i]], FILTER == "PASS")
}

```

SNVs

Base substitution types

You can retrieve base substitution types from the VCF GRanges object as “REF>ALT” using `mutations_from_vcf`:

Hide

```

muts <- mutations_from_vcf(snp_grl_pass[[1]])
head(muts, 12)

```

```
[1] "C>A" "T>C" "T>C" "G>T" "C>T" "T>C" "C>A" "G>T" "C>A" "C>A" "C>A" "C>T"
```

Hide

```

types <- mut_type(snp_grl_pass[[1]])
head(types, 12)

```

```
[1] "C>A" "T>C" "T>C" "C>A" "C>T" "T>C" "C>A" "C>A" "C>A" "C>A" "C>A" "C>T"
```

Hide

Show

```
context <- mut_context(snp_grl_pass[[1]], ref_genome)
head(context, 12)
```

```
chr1 chr1 chr1 chr1 chr1 chr1 chr1 chr1 chr1 chr1 chr1 chr1
"GCT" "CTG" "ATT" "GGA" "TCA" "TTC" "ACA" "TGA" "GCG" "GCT" "ACA" "TCA"
```

Hide

```
type_context <- type_context(snp_grl_pass[[1]], ref_genome)
lapply(type_context, head, 12)
```

```
$types
[1] "C>A" "T>C" "T>C" "C>A" "C>T" "T>C" "C>A" "C>A" "C>A" "C>A" "C>A" "C>T"

$context
chr1 chr1 chr1 chr1 chr1 chr1 chr1 chr1 chr1 chr1 chr1 chr1
"GCT" "CTG" "ATT" "TCC" "TCA" "TTC" "ACA" "TCA" "GCG" "GCT" "ACA" "TCA"
```

Hide

```
type_occurrences <- mut_type_occurrences(snp_grl_pass, ref_genome)
type_occurrences
```

	C>A <int>	C>G <int>	C>T <int>	T>A <int>	T>C <int>	T>G <int>	C>T at CpG <int>	C>T other <int>
WT-day0	8279	1299	5691	2030	9821	697	666	5025
WT-day77	9463	1357	5496	1745	9431	752	628	4868
Pms1-day0	6040	1113	5037	1830	9214	642	601	4436
Pms1-day77	9843	1475	5994	2094	10428	810	749	5245
4 rows								

Hide

```
NA
```

Fisher’s exact test of mutation types

Hide

```

type_occurrences_norm_t <- t(type_occurrences_norm)
my_fisher_p <-c()
my_conf_int_dn <-c()
my_conf_int_up <-c()
my_odds <- c()
my_fdr <- c()

for (mut in 1:nrow(type_occurrences_norm_t)){
  my_matrix <- t(matrix(type_occurrences_norm_t[mut,], nrow = 2, dimnames = list( Timepoint = c("d0", "d77"),
                                                                                   Genotype = c("WT", "Pms1"))))
  my_mut_name <- rownames(type_occurrences_norm_t)[mut]
  print(my_mut_name)
  print(my_matrix)
  my_test <- fisher.test(my_matrix)
  print(my_test)
  my_fisher_p <- c(my_fisher_p, my_test$p.value)
  my_conf_int_dn <- c(my_conf_int_dn, my_test$conf.int[1])
  my_conf_int_up <- c(my_conf_int_up, my_test$conf.int[2])
  my_odds <- c(my_odds, my_test$estimate)
  my_fdr <- c(my_fdr, p.adjust(p = my_test$p.value, method = "BH", n = nrow(type_occurrences_norm_t)))
}

```

```

[1] "C>A"
      Timepoint
Genotype  d0  d77
WT       2471 2805
Pms1     2089 2687

Fisher's Exact Test for Count Data

data:  my_matrix
p-value = 0.001877
alternative hypothesis: true odds ratio is not equal to 1
95 percent confidence interval:
 1.046534 1.226850
sample estimates:
odds ratio
 1.133086

[1] "C>G"
      Timepoint
Genotype  d0  d77

```

```
WT    388 402
```

```
Pms1  385 403
```

Fisher's Exact Test for Count Data

```
data:  my_matrix
```

```
p-value = 0.9199
```

```
alternative hypothesis: true odds ratio is not equal to 1
```

```
95 percent confidence interval:
```

```
0.8252233 1.2368935
```

```
sample estimates:
```

```
odds ratio
```

```
1.010294
```

```
[1] "C>T"
```

```
Timepoint
```

```
Genotype  d0 d77
```

```
WT    1698 1629
```

```
Pms1  1742 1636
```

Fisher's Exact Test for Count Data

```
data:  my_matrix
```

```
p-value = 0.6779
```

```
alternative hypothesis: true odds ratio is not equal to 1
```

```
95 percent confidence interval:
```

```
0.8884766 1.0786029
```

```
sample estimates:
```

```
odds ratio
```

```
0.9789266
```

```
[1] "T>A"
```

```
Timepoint
```

```
Genotype  d0 d77
```

```
WT    606 517
```

```
Pms1  633 572
```

Fisher's Exact Test for Count Data

```
data:  my_matrix
```

```
p-value = 0.5061
```

```
alternative hypothesis: true odds ratio is not equal to 1
```

```
95 percent confidence interval:
```

```
0.8968763 1.2508951
```

```
sample estimates:
```

```
odds ratio
```

```
1.059143
```

```
[1] "T>C"
      Timepoint
Genotype d0 d77
WT      2931 2795
Pms1    3187 2846

Fisher's Exact Test for Count Data

data: my_matrix
p-value = 0.07631
alternative hypothesis: true odds ratio is not equal to 1
95 percent confidence interval:
 0.8704766 1.0074501
sample estimates:
odds ratio
 0.9364641
```

```
[1] "T>G"
      Timepoint
Genotype d0 d77
WT      208 223
Pms1    222 221

Fisher's Exact Test for Count Data

data: my_matrix
p-value = 0.589
alternative hypothesis: true odds ratio is not equal to 1
95 percent confidence interval:
 0.7058685 1.2214320
sample estimates:
odds ratio
 0.9286132
```

```
[1] "C>T at CpG"
      Timepoint
Genotype d0 d77
WT      199 186
Pms1    208 204

Fisher's Exact Test for Count Data

data: my_matrix
p-value = 0.7768
alternative hypothesis: true odds ratio is not equal to 1
95 percent confidence interval:
```

```
0.7869958 1.3991067
```

```
sample estimates:
```

```
odds ratio
```

```
1.049221
```

```
[1] "C>T other"
```

```
Timepoint
```

```
Genotype d0 d77
```

```
WT 1500 1443
```

```
Pms1 1534 1432
```

```
Fisher's Exact Test for Count Data
```

```
data: my_matrix
```

```
p-value = 0.567
```

```
alternative hypothesis: true odds ratio is not equal to 1
```

```
95 percent confidence interval:
```

```
0.8750866 1.0760213
```

```
sample estimates:
```

```
odds ratio
```

```
0.9703735
```

[Hide](#)

```
type_occurrence_report <- tibble(as.data.frame(cbind(mutation = rownames(type_occurrences_norm_t), type_occurrences_norm_t, p.value = round(my_fisher_p, digits = 4), p.adj = round(my_fdr, digits = 4), conf.int.down = round(my_conf_int_dn, digits = 2), conf.int.up = round(my_conf_int_up, digits = 2), odds.ratio = round(my_odds, digits = 2))))
```

```
filter(type_occurrence_report, type_occurrence_report$p.value <= 0.1)
```

mutation <chr>	WT- day0 <chr>	WT- day77 <chr>	Pms1- day0 <chr>	Pms1- day77 <chr>	p.value <chr>	p.adj <chr>	conf.int.down <chr>	conf.int.u <chr>
C>A	2471	2805	2089	2687	0.0019	0.015	1.05	1.23
T>C	2931	2795	3187	2846	0.0763	0.6105	0.87	1.01

```
2 rows
```

[Hide](#)

```
type_occurrence_report
```

WT-

WT-

Pms1-

Pms1-

mutation <chr>	day0 <chr>	day77 <chr>	day0 <chr>	day77 <chr>	p.value <chr>	p.adj <chr>	conf.int.down <chr>	conf.in <chr>
C>A	2471	2805	2089	2687	0.0019	0.015	1.05	1.23
C>G	388	402	385	403	0.9199	1	0.83	1.24
C>T	1698	1629	1742	1636	0.6779	1	0.89	1.08
T>A	606	517	633	572	0.5061	1	0.9	1.25
T>C	2931	2795	3187	2846	0.0763	0.6105	0.87	1.01
T>G	208	223	222	221	0.589	1	0.71	1.22
C>T at CpG	199	186	208	204	0.7768	1	0.79	1.4
C>T other	1500	1443	1534	1432	0.567	1	0.88	1.08

8 rows | 1-9 of 10 columns

Hide

```

library(ggplot2)
library(cowplot)
library(extrafont)

type_occurrence_report$odds.ratio <- as.numeric(type_occurrence_report$odds.ratio)
type_occurrence_report$conf.int.down <- as.numeric(type_occurrence_report$conf.int.do
wn)
type_occurrence_report$conf.int.up <- as.numeric(type_occurrence_report$conf.int.up)

p.occurrence.coi <- ggplot(data = type_occurrence_report, aes(x = mutation, y = odds.
ratio, ymin = conf.int.down, ymax = conf.int.up)) +
  geom_hline(yintercept = 1, color = "blue" ) +
  geom_pointrange(colour=ifelse(type_occurrence_report$p.adj <= 0.05,"darkgreen",ifel
se(type_occurrence_report$p.value <= 0.05,"orange","darkgrey"))) +
  ggtitle("Odds ratio of normalized mutation-type counts between Pms1.d77/Pms1.d0 and
WT.d77/WT.d0") +
  theme_classic(base_size = 8) +
  coord_flip()

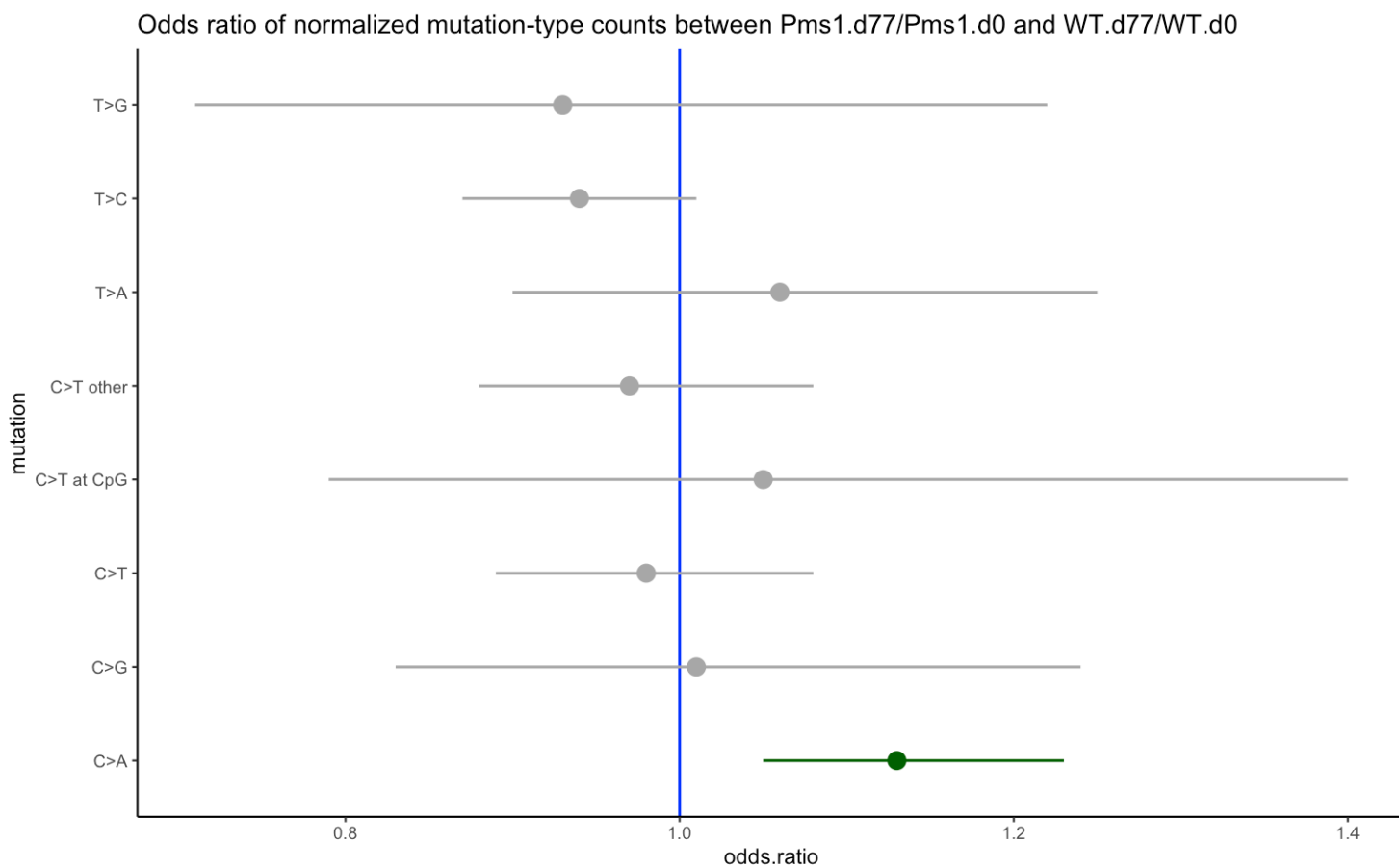
ggsave2(filename = "mut_ocurrence_report.pdf", plot = p.occurrence.coi)

```

Saving 7 x 7 in image

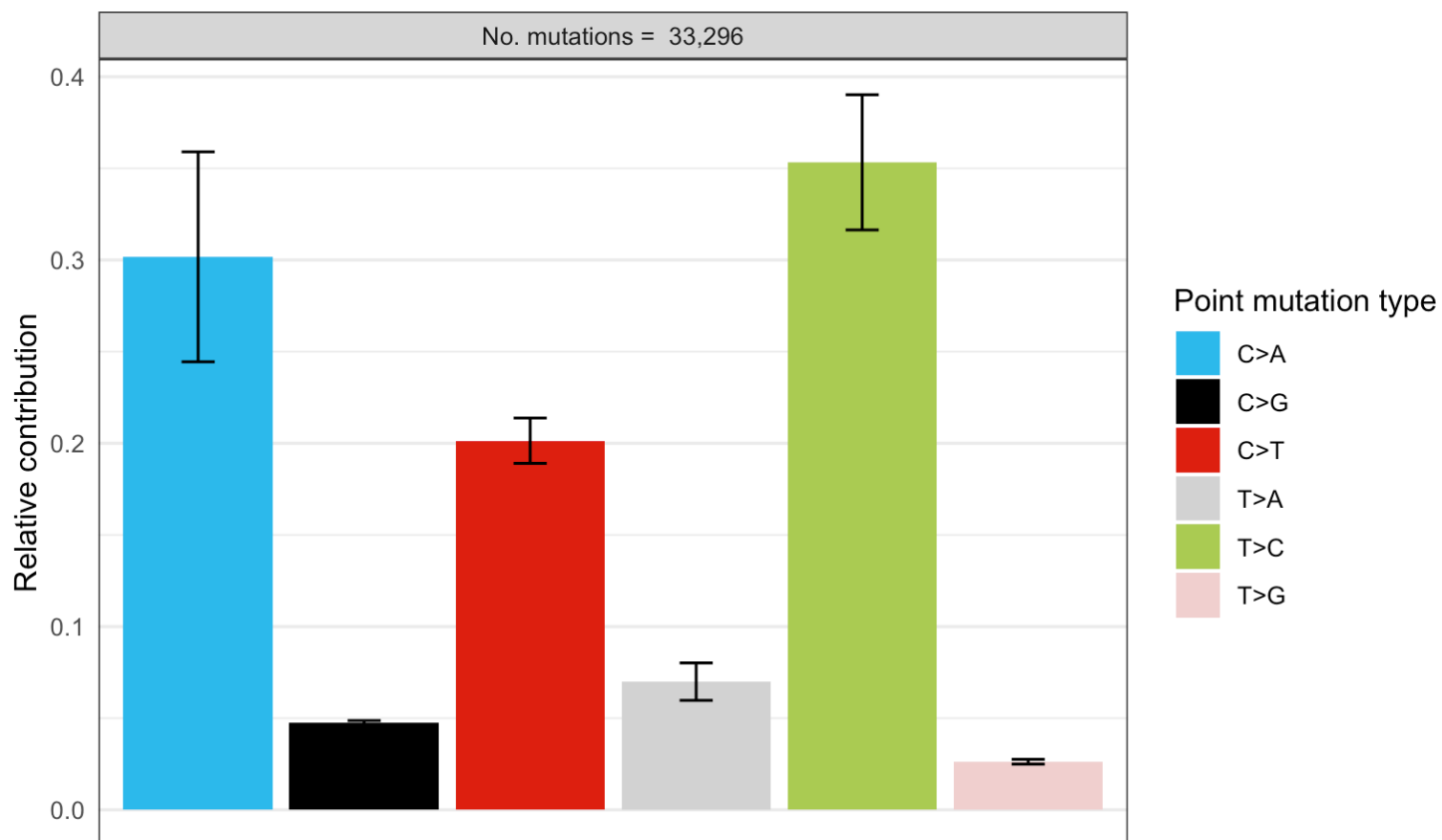
Hide

p.occurrence.coi

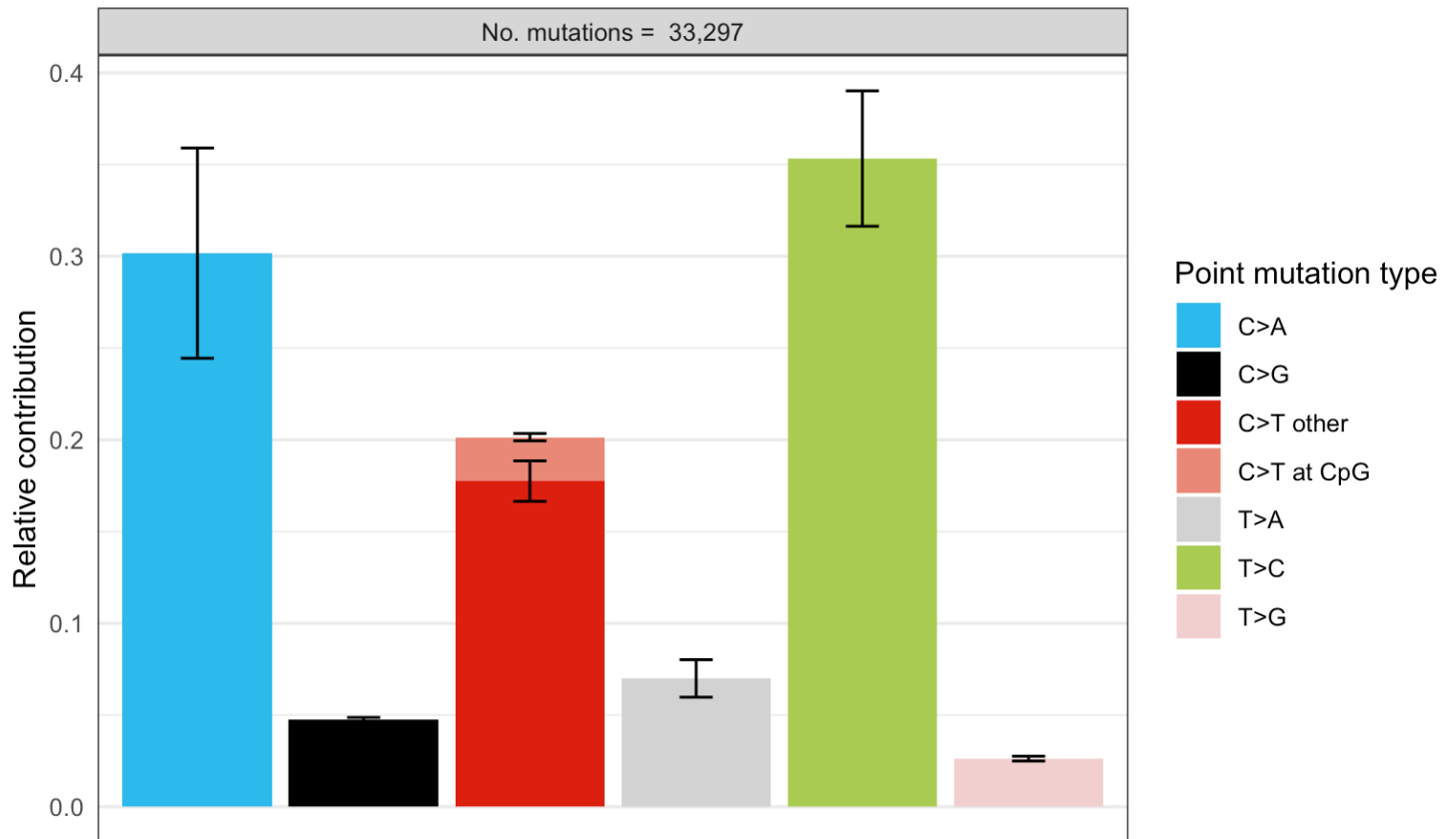


Hide

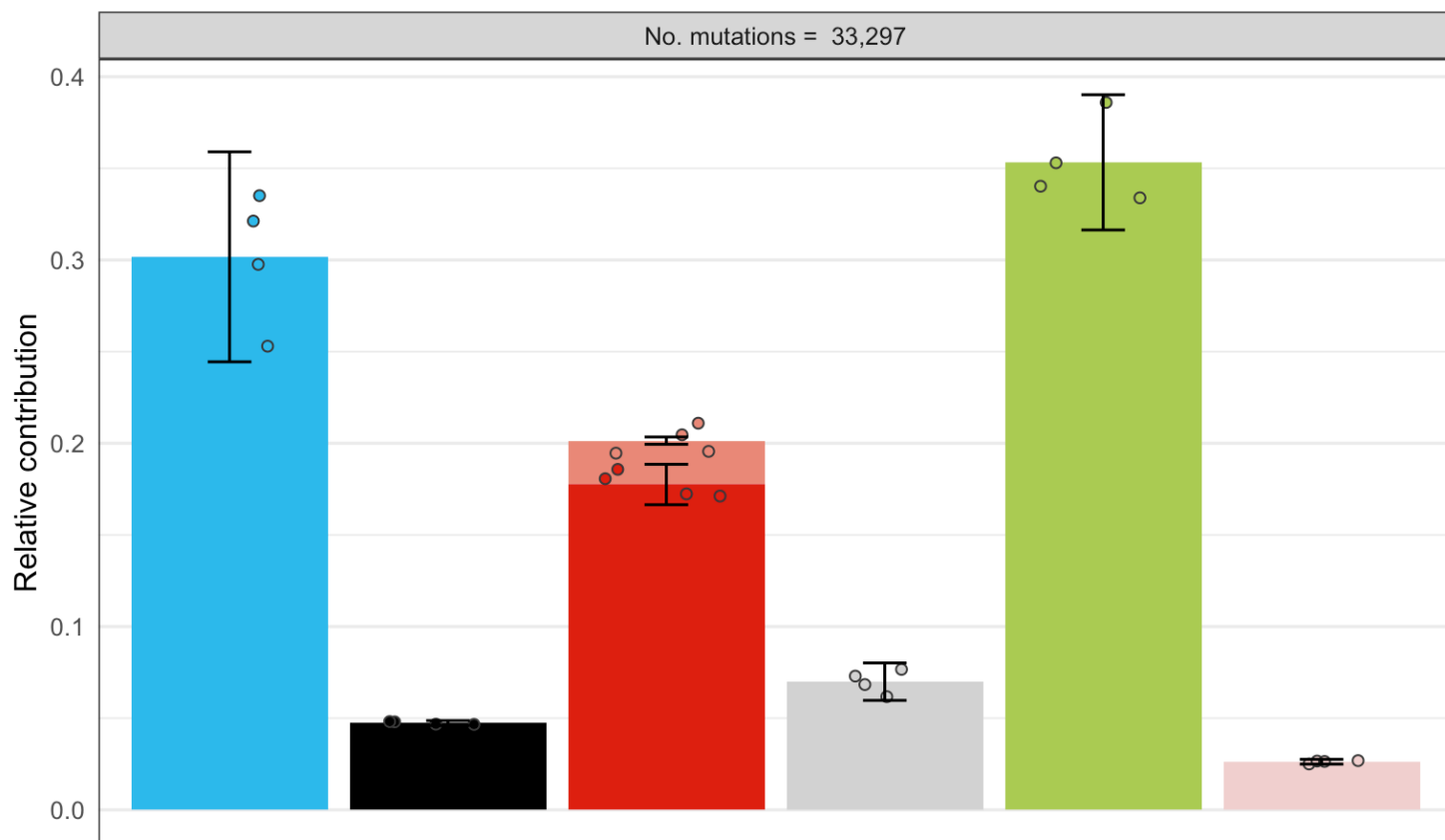
```
p1 <- plot_spectrum(type_occurrences_norm)
p1
```

[Hide](#)

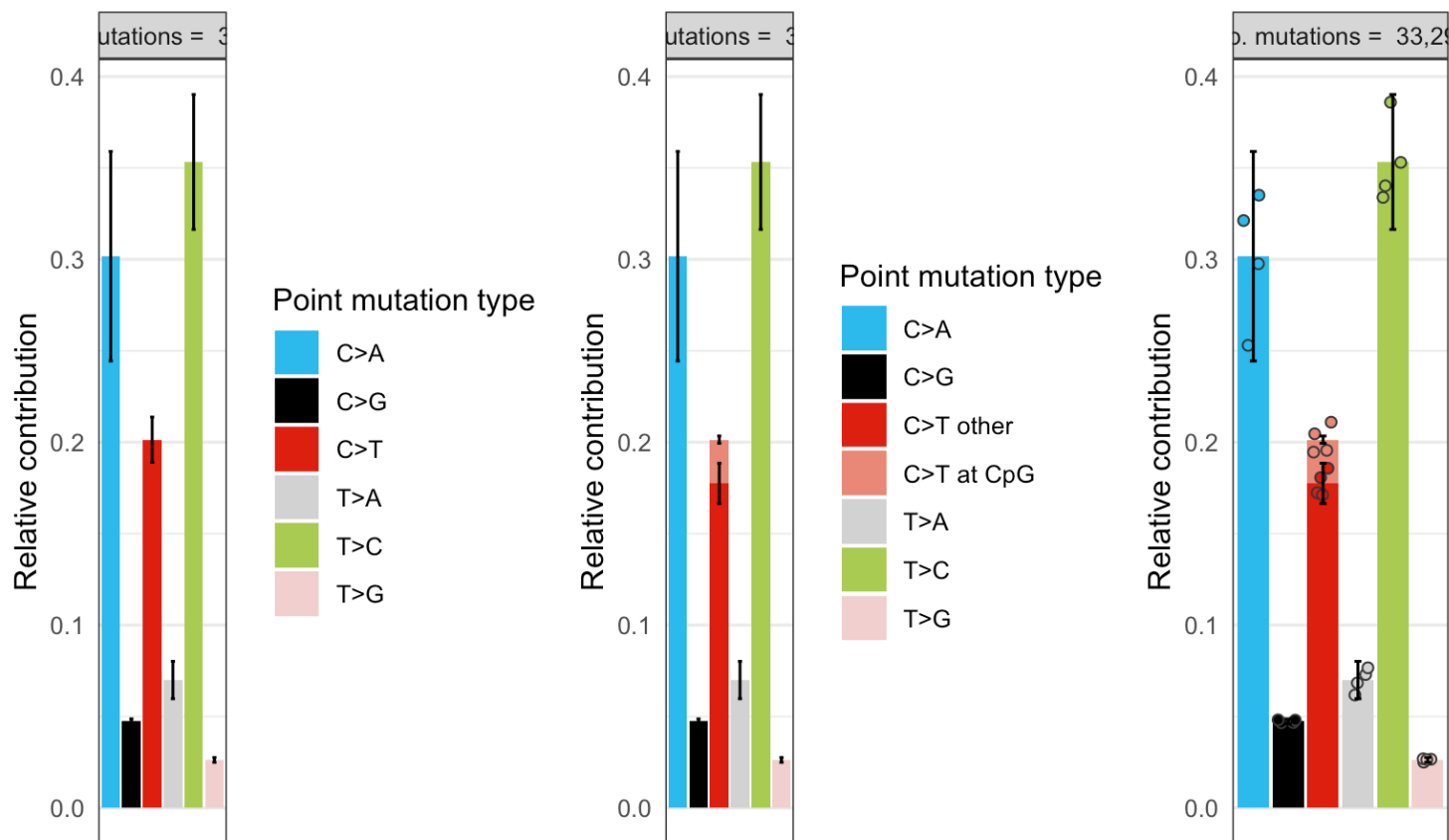
```
p2 <- plot_spectrum(type_occurrences_norm, CT = TRUE)
p2
```


[Hide](#)

```
p3 <- plot_spectrum(type_occurrences_norm, CT = TRUE,  
                    indv_points = TRUE, legend = FALSE)  
p3
```

[Hide](#)

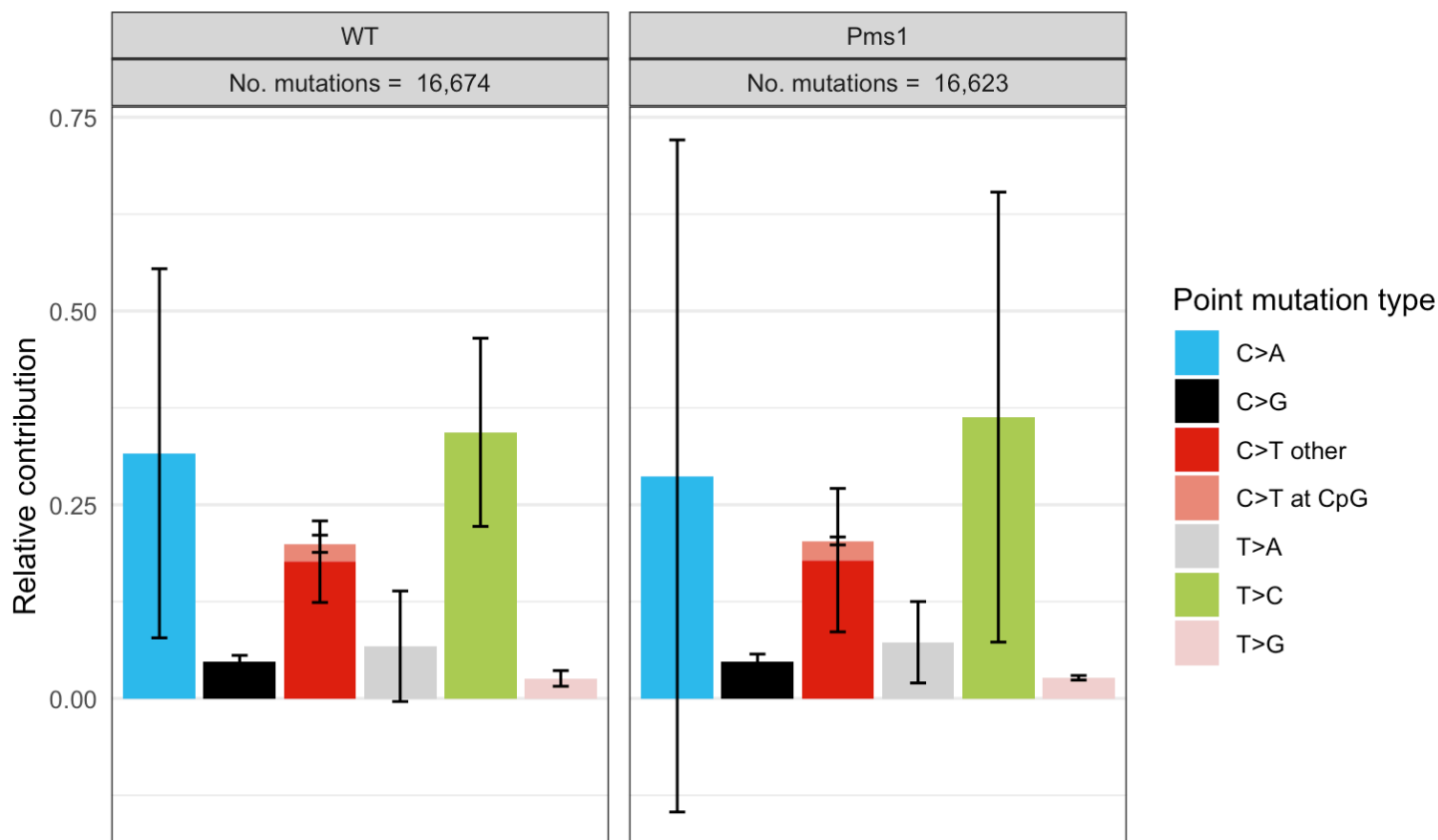
```
library("gridExtra")  
grid.arrange(p1, p2, p3, ncol = 3, widths = c(3, 3, 1.75))
```



It's also possible to create a facet per sample group, e.g. plot the spectrum for each tissue separately:

Hide

```
p4 <- plot_spectrum(type_occurrences_norm, by = genotype, CT = TRUE, legend = TRUE)
p4
```



Or you could use the standard deviation instead of a 95% confidence interval:

Hide

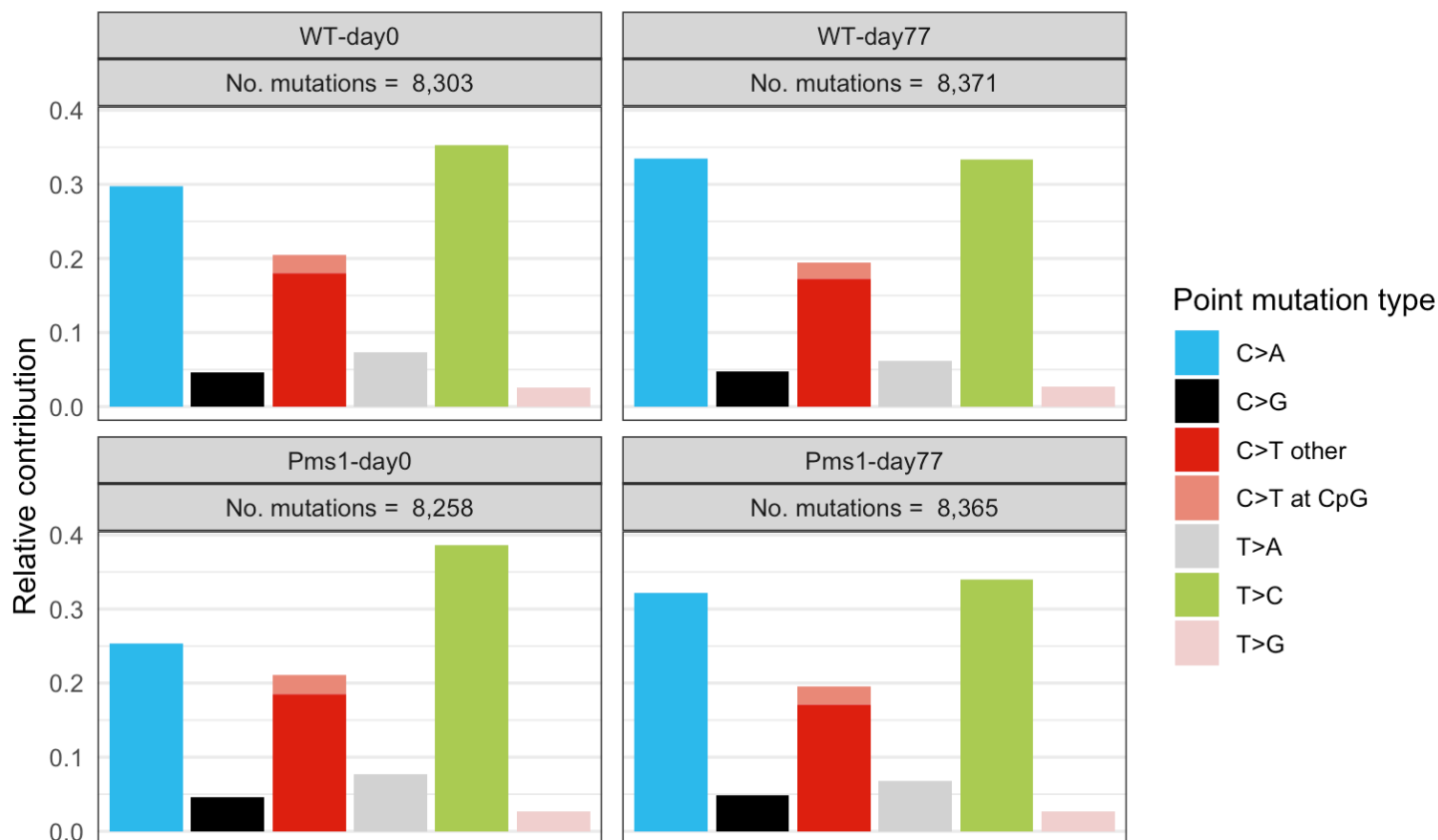
```
p5 <- plot_spectrum(type_occurrences_norm, by = groups, CT = TRUE,
  legend = TRUE, errorBars = "stdev")
```

Warning: No error bars can be plotted, because there is only one sample per mutation spectrum.

Use the argument: ``errorBars = 'none'``, if you want to avoid this warning.

Hide

p5



96 mutational profile

First you should make a 96 trinucleotide mutation count matrix. (In contrast to previous versions this also works for single samples.)

[Hide](#)

```
write.tsv(mut_mat_report, file = "mut_mat_report.txt")
```

```
Error in write.tsv(mut_mat_report, file = "mut_mat_report.txt") :
  could not find function "write.tsv"
```

Plot confidence intervals and odda.ratios

[Hide](#)

```

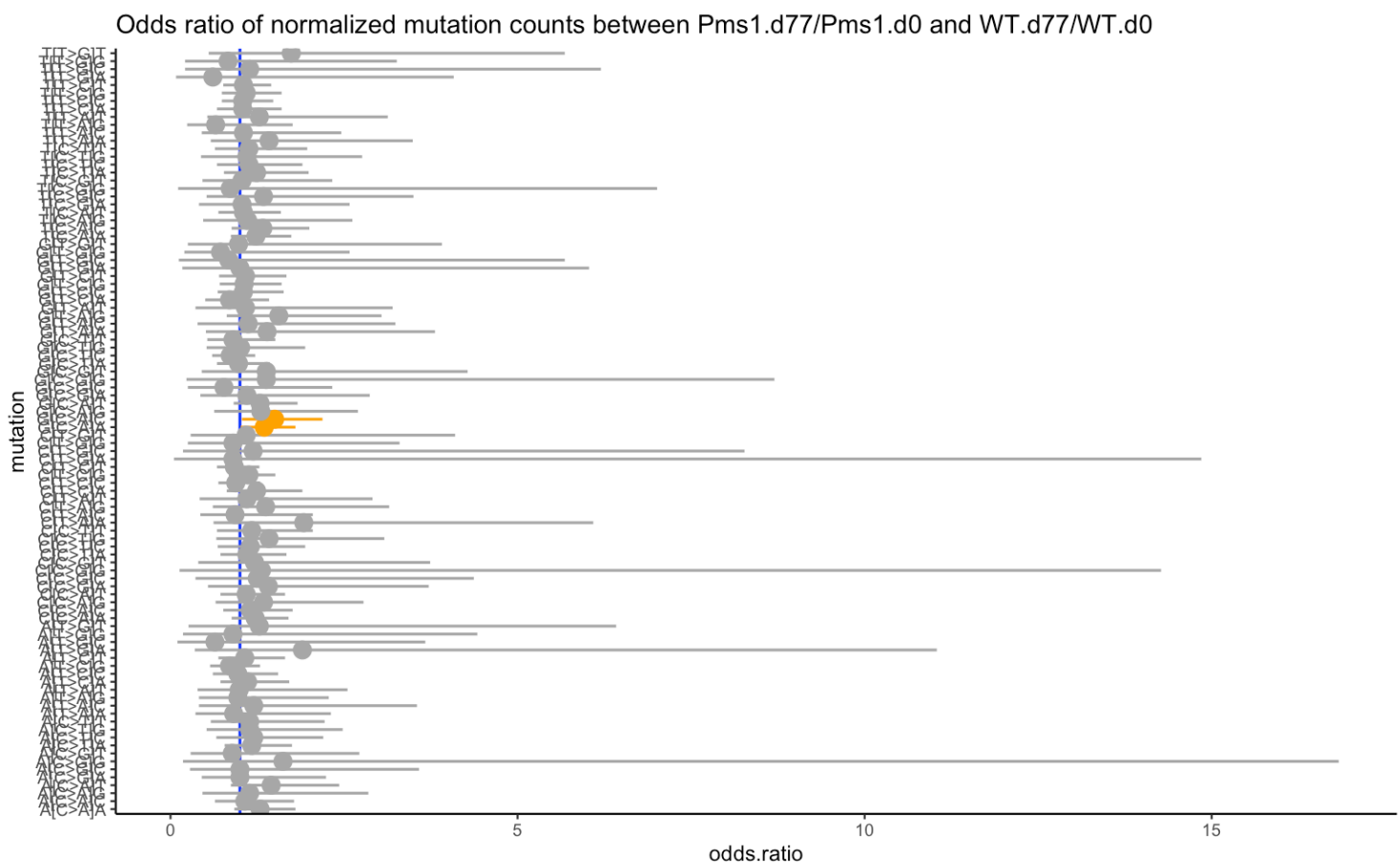
library(ggplot2)
library(cowplot)
library(extrafont)

mut_mat_report$odds.ratio <- as.numeric(mut_mat_report$odds.ratio)
mut_mat_report$conf.int.down <- as.numeric(mut_mat_report$conf.int.down)
mut_mat_report$conf.int.up <- as.numeric(mut_mat_report$conf.int.up)

p.coi <- ggplot(data = mut_mat_report, aes(x = mutation, y = odds.ratio, ymin = conf.
int.down, ymax = conf.int.up)) +
  geom_hline(yintercept = 1, color = "blue" ) +
  geom_pointrange(colour=ifelse(mut_mat_report$p.adj <= 0.05,"darkgreen",ifelse(mut_m
at_report$p.value <= 0.05,"orange","darkgrey"))) +
  ggtitle("Odds ratio of normalized mutation counts between Pms1.d77/Pms1.d0 and WT.d
77/WT.d0") +
  theme_classic(base_size = 8) +
  coord_flip()

ggsave2(filename = "mut_matrix_report.pdf", plot = p.coi, height = 11, width = 8)
p.coi

```



Green denotes mutations having a p.value ≤ 0.05 . In all cases adjusted p.values are > 0.05 .

Next, you can use this matrix to plot the 96 profile of samples. In this example we do this for 2 samples:

[Hide](#)

```
p96_profile <- plot_96_profile(mut_mat_norm[, c(1:4)], condensed = TRUE, ymax = 0.05)
+ scale_y_continuous(breaks = c(0.01,0.02,0.03,0.04,0.05))
```

Scale for y is already present.
Adding another scale for y, which will replace the existing scale.

[Hide](#)

```
ggsave2(filename = "p96_profile_norm.pdf", plot = p96_profile)
```

Saving 7 x 7 in image

It's also possible to look at larger mutational contexts. However, this is only usefull if you have a large number of mutations.

[Hide](#)

```
mut_mat_ext_context_norm
```

	WT-day0	WT-day77	Pms1-day0	Pms1-day77
AA[C>A]AA	14	14	9	11
AA[C>A]AC	9	8	6	10
AA[C>A]AG	9	10	10	12
AA[C>A]AT	7	8	8	11
AA[C>A]CA	10	8	5	7
AA[C>A]CC	7	6	4	6
AA[C>A]CG	1	1	0	1
AA[C>A]CT	5	5	6	6
AA[C>A]GA	1	1	1	2
AA[C>A]GC	2	1	1	1
AA[C>A]GG	1	1	1	1
AA[C>A]GT	2	1	1	2
AA[C>A]TA	5	4	4	4
AA[C>A]TC	5	3	5	5
AA[C>A]TG	7	6	4	7
AA[C>A]TT	4	3	4	3
AC[C>A]AA	14	15	9	13
AC[C>A]AC	11	15	8	18

AC[C>A]AG	13	14	9	17
AC[C>A]AT	8	13	9	13
AC[C>A]CA	10	12	10	15
AC[C>A]CC	6	8	5	8
AC[C>A]CG	1	2	2	1
AC[C>A]CT	8	9	6	8
AC[C>A]GA	2	4	2	3
AC[C>A]GC	4	3	1	2
AC[C>A]GG	2	3	2	3
AC[C>A]GT	3	3	3	5
AC[C>A]TA	4	5	3	5
AC[C>A]TC	8	9	3	8
AC[C>A]TG	8	11	7	9
AC[C>A]TT	5	7	5	5
AG[C>A]AA	23	20	19	24
AG[C>A]AC	16	19	17	21
AG[C>A]AG	26	24	23	29
AG[C>A]AT	20	19	16	19
AG[C>A]CA	20	20	16	21
AG[C>A]CC	15	13	9	15
AG[C>A]CG	2	2	2	2
AG[C>A]CT	14	14	10	17
AG[C>A]GA	3	3	4	4
AG[C>A]GC	5	4	1	3
AG[C>A]GG	4	2	2	4
AG[C>A]GT	3	3	2	2
AG[C>A]TA	10	10	7	11
AG[C>A]TC	12	14	9	12
AG[C>A]TG	19	19	12	19
AG[C>A]TT	10	12	6	11
AT[C>A]AA	6	8	6	11
AT[C>A]AC	9	13	7	13
AT[C>A]AG	9	10	11	14
AT[C>A]AT	9	10	6	11
AT[C>A]CA	11	10	6	10
AT[C>A]CC	9	9	6	9
AT[C>A]CG	1	1	1	1
AT[C>A]CT	9	8	7	10
AT[C>A]GA	1	2	1	2
AT[C>A]GC	2	1	2	1
AT[C>A]GG	1	2	1	2
AT[C>A]GT	2	2	2	2
AT[C>A]TA	3	5	4	5
AT[C>A]TC	6	8	4	7
AT[C>A]TG	7	9	5	10
AT[C>A]TT	5	7	4	7
CA[C>A]AA	12	11	7	8

CA[C>A]AC	21	17	18	20
CA[C>A]AG	13	15	10	15
CA[C>A]AT	11	9	9	14
CA[C>A]CA	6	10	7	8
CA[C>A]CC	5	5	6	4
CA[C>A]CG	1	1	1	0
CA[C>A]CT	5	5	7	6
CA[C>A]GA	2	1	1	1
CA[C>A]GC	2	2	1	2
CA[C>A]GG	3	3	3	3
CA[C>A]GT	2	3	3	3
CA[C>A]TA	3	3	3	4
CA[C>A]TC	4	3	3	4
CA[C>A]TG	10	7	7	10
CA[C>A]TT	5	7	2	7
CC[C>A]AA	6	9	8	9
CC[C>A]AC	12	12	7	10
CC[C>A]AG	12	14	11	14
CC[C>A]AT	5	10	6	7
CC[C>A]CA	10	12	7	10
CC[C>A]CC	5	7	4	7
CC[C>A]CG	1	3	1	2
CC[C>A]CT	5	6	4	6
CC[C>A]GA	2	3	2	3
CC[C>A]GC	3	2	2	2
CC[C>A]GG	3	3	2	3
CC[C>A]GT	2	2	2	2
CC[C>A]TA	3	5	2	4
CC[C>A]TC	7	8	6	5
CC[C>A]TG	8	10	8	8
CC[C>A]TT	5	7	5	5
CG[C>A]AA	1	1	0	1
CG[C>A]AC	3	1	2	1
CG[C>A]AG	2	1	2	2
CG[C>A]AT	1	1	1	1
CG[C>A]CA	1	1	1	2
CG[C>A]CC	2	2	0	3
CG[C>A]CG	0	0	0	0
CG[C>A]CT	1	1	0	1
CG[C>A]GA	0	0	0	0
CG[C>A]GC	1	1	0	1
CG[C>A]GG	1	1	0	0
CG[C>A]GT	0	1	0	1
CG[C>A]TA	1	1	0	1
CG[C>A]TC	1	1	1	1
CG[C>A]TG	1	1	2	2
CG[C>A]TT	1	1	1	0

CT[C>A]AA	7	7	5	7
CT[C>A]AC	10	8	8	8
CT[C>A]AG	12	15	9	14
CT[C>A]AT	5	8	4	8
CT[C>A]CA	6	12	7	11
CT[C>A]CC	8	9	7	9
CT[C>A]CG	1	2	1	2
CT[C>A]CT	6	9	6	7
CT[C>A]GA	2	2	1	2
CT[C>A]GC	2	1	2	3
CT[C>A]GG	2	4	1	3
CT[C>A]GT	1	3	1	2
CT[C>A]TA	2	4	5	4
CT[C>A]TC	4	9	8	8
CT[C>A]TG	10	10	8	11
CT[C>A]TT	4	6	5	6
GA[C>A]AA	7	8	6	9
GA[C>A]AC	9	8	5	8
GA[C>A]AG	10	13	10	13
GA[C>A]AT	7	7	5	9
GA[C>A]CA	9	6	4	5
GA[C>A]CC	5	5	4	4
GA[C>A]CG	1	1	0	1
GA[C>A]CT	5	4	6	6
GA[C>A]GA	2	2	2	1
GA[C>A]GC	1	1	1	2
GA[C>A]GG	1	1	1	1
GA[C>A]GT	2	1	1	1
GA[C>A]TA	2	2	4	3
GA[C>A]TC	4	4	3	6
GA[C>A]TG	5	6	4	6
GA[C>A]TT	4	3	4	5
GC[C>A]AA	7	9	5	8
GC[C>A]AC	5	11	6	8
GC[C>A]AG	14	14	8	11
GC[C>A]AT	6	7	4	8
GC[C>A]CA	8	9	6	10
GC[C>A]CC	6	6	4	6
GC[C>A]CG	2	1	1	2
GC[C>A]CT	4	7	4	7
GC[C>A]GA	1	1	1	1
GC[C>A]GC	1	2	1	2
GC[C>A]GG	2	2	2	2
GC[C>A]GT	2	3	0	2
GC[C>A]TA	3	4	3	3
GC[C>A]TC	7	7	6	9
GC[C>A]TG	10	10	8	10

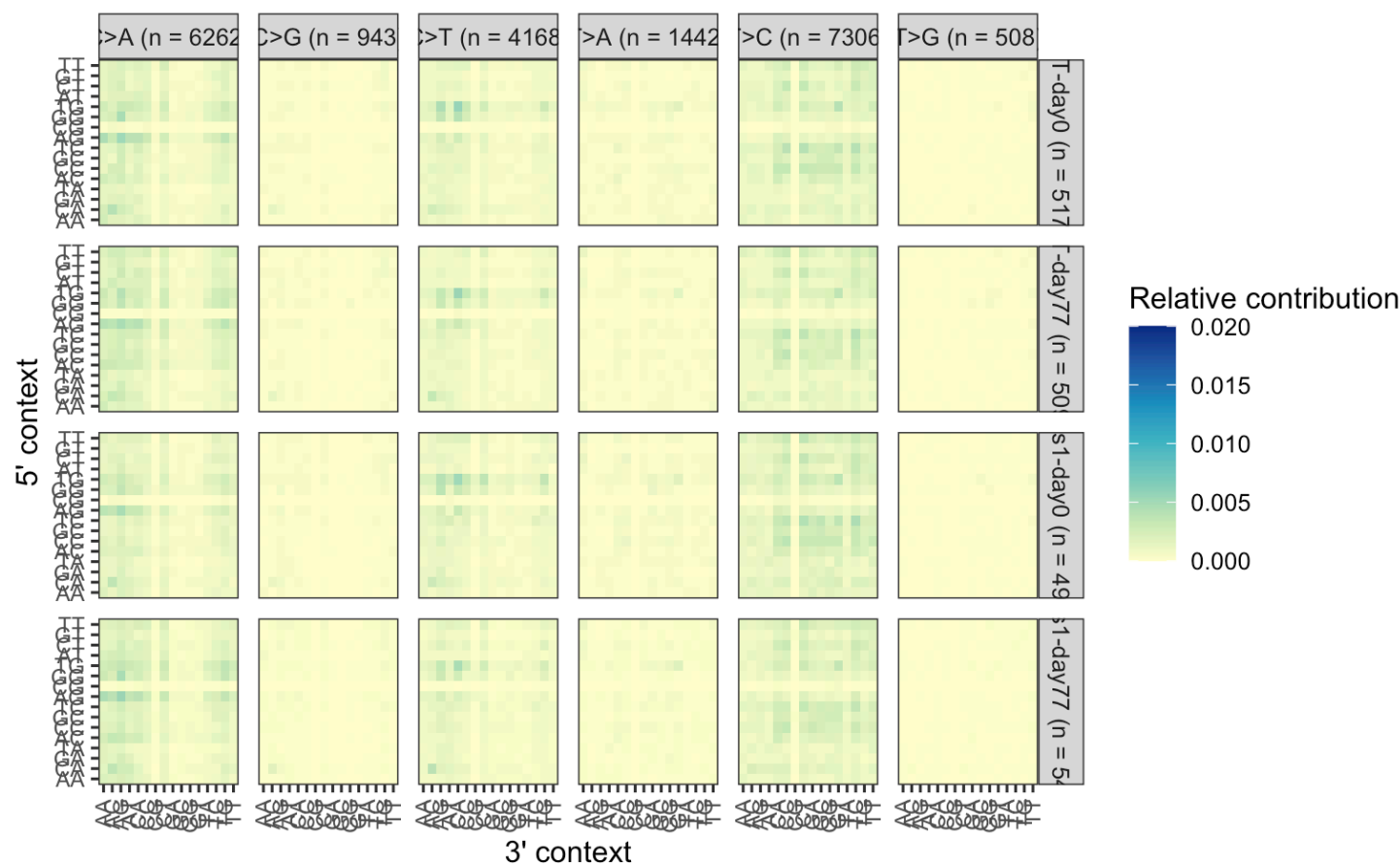
GC[C>A]TT	6	7	4	5
GG[C>A]AA	11	12	9	16
GG[C>A]AC	12	10	10	12
GG[C>A]AG	24	18	17	25
GG[C>A]AT	9	9	11	11
GG[C>A]CA	10	13	12	14
GG[C>A]CC	8	8	5	9
GG[C>A]CG	2	2	2	1
GG[C>A]CT	10	13	7	13
GG[C>A]GA	2	3	3	2
GG[C>A]GC	2	1	3	3
GG[C>A]GG	4	3	3	4
GG[C>A]GT	2	2	3	2
GG[C>A]TA	8	10	6	7
GG[C>A]TC	11	13	7	11
GG[C>A]TG	12	16	11	19
GG[C>A]TT	10	10	8	11
GT[C>A]AA	7	7	5	7
GT[C>A]AC	10	10	7	7
GT[C>A]AG	10	12	6	10
GT[C>A]AT	5	7	5	9
GT[C>A]CA	7	6	5	5
GT[C>A]CC	7	6	3	6
GT[C>A]CG	1	1	1	1
GT[C>A]CT	6	8	5	8
GT[C>A]GA	1	1	1	2
GT[C>A]GC	1	2	2	1
GT[C>A]GG	1	1	2	1
GT[C>A]GT	2	2	1	2
GT[C>A]TA	4	4	2	3
GT[C>A]TC	8	8	5	6
GT[C>A]TG	10	7	6	6
GT[C>A]TT	5	5	4	6
TA[C>A]AA	7	9	5	9
TA[C>A]AC	7	6	5	7
TA[C>A]AG	6	8	8	8
TA[C>A]AT	7	7	7	5
TA[C>A]CA	4	4	3	4
TA[C>A]CC	3	6	3	4
TA[C>A]CG	0	1	1	1
TA[C>A]CT	3	5	3	5
TA[C>A]GA	1	1	1	0
TA[C>A]GC	1	0	0	1
TA[C>A]GG	0	1	1	1
TA[C>A]GT	0	1	0	2
TA[C>A]TA	1	2	2	2
TA[C>A]TC	4	2	2	2

TA[C>A]TG	3	4	3	5
TA[C>A]TT	3	4	4	4
TC[C>A]AA	7	9	5	7
TC[C>A]AC	8	11	6	10
TC[C>A]AG	14	14	10	15
TC[C>A]AT	7	12	6	9
TC[C>A]CA	12	12	8	9
TC[C>A]CC	7	8	6	7
TC[C>A]CG	1	2	1	1
TC[C>A]CT	6	12	6	7
TC[C>A]GA	1	2	2	2
TC[C>A]GC	2	2	2	2
TC[C>A]GG	2	3	2	2
TC[C>A]GT	2	2	1	3
TC[C>A]TA	2	4	2	6
TC[C>A]TC	6	9	6	9
TC[C>A]TG	9	11	5	9
TC[C>A]TT	6	10	6	7
TG[C>A]AA	13	15	13	15
TG[C>A]AC	10	9	11	16
TG[C>A]AG	16	20	15	19
TG[C>A]AT	15	12	11	16
TG[C>A]CA	13	12	8	14
TG[C>A]CC	11	9	7	11
TG[C>A]CG	2	1	1	2
TG[C>A]CT	13	14	10	16
TG[C>A]GA	1	2	2	2
TG[C>A]GC	2	2	2	4
TG[C>A]GG	3	3	1	2
TG[C>A]GT	3	2	3	3
TG[C>A]TA	8	7	8	11
TG[C>A]TC	10	14	9	13
TG[C>A]TG	15	15	13	20
TG[C>A]TT	10	12	8	14
TT[C>A]AA	7	10	8	9
TT[C>A]AC	9	13	6	10
TT[C>A]AG	11	14	8	12
TT[C>A]AT	8	9	8	9
TT[C>A]CA	10	10	8	12
TT[C>A]CC	8	12	7	11
TT[C>A]CG	2	2	0	2
TT[C>A]CT	12	11	8	12
TT[C>A]GA	2	3	0	1
TT[C>A]GC	1	2	1	1

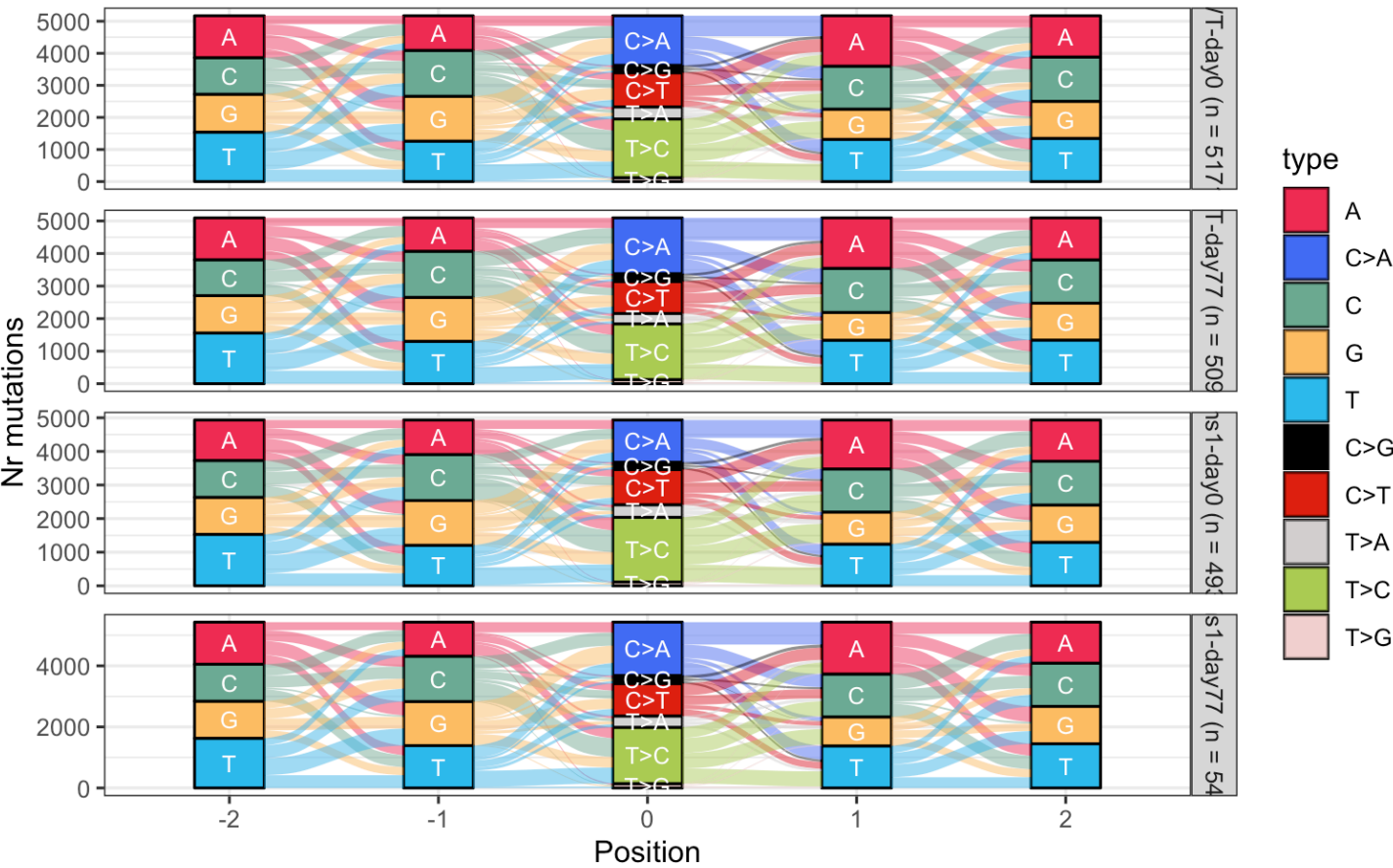
[reached getOption("max.print") -- omitted 1286 rows]

The extension argument also works for the `mut_context` and `type_context` functions.

You can visualize this matrix with a heatmap.



You can also visualize this with a riverplot.



Indels

First you should get the COSMIC indel contexts. This is done with `get_indel_context`, which adds the columns `muttype` and `muttype_sub` to the `GRangesList`. The `muttype` column contains the main type of indel. The `muttype_sub` column shows the number of repeat units. For microhomology (mh) deletions the mh length is shown.

Hide

```
head(indel_grl, n = 4)
```

```
$`WT-day0`  
GRanges object with 5971 ranges and 7 metadata columns:  
      seqnames      ranges strand | paramRangeID  
REF  
      <Rle>          <IRanges> <Rle> |      <factor> <DNAStringS  
et>  
chr1:3555815_GTC/G    chr1    3555815-3555817    * |      NA  
GTC  
chr1:4009853_G/GA     chr1    4009853      * |      NA  
G  
chr1:4624693_CA/C     chr1    4624693-4624694    * |      NA
```

```

CA
    chr1:4851624_TA/T    chr1    4851624-4851625    * |    NA
TA
    chr1:4894076_TTG/T    chr1    4894076-4894078    * |    NA
TTG
    ...
...
    chrY:39960747_AAC/A    chrY    39960747-39960749    * |    NA
AAC
    chrY:76250767_CTG/C    chrY    76250767-76250769    * |    NA
CTG
    chrY:86118528_GGA/G    chrY    86118528-86118530    * |    NA
GGA
    chrY:90744554_TC/T    chrY    90744554-90744555    * |    NA
TC
    chrY:90744588_CCCTAG/C    chrY    90744588-90744593    * |    NA    CCC
TAG
                                ALT    QUAL    FILTER    muttype muttyp
e_sub
                                <DNAStringSetList> <numeric> <character> <character> <num
eric>
    chr1:3555815_GTC/G    G    55.12    PASS    2bp_deletion
16
    chr1:4009853_G/GA    GA    66.12    PASS    T_insertion
1
    chr1:4624693_CA/C    C    47.12    PASS    T_deletion
11
    chr1:4851624_TA/T    T    47.12    PASS    T_deletion
12
    chr1:4894076_TTG/T    T    61.12    PASS    2bp_deletion
12
    ...
...
    chrY:39960747_AAC/A    A    60.98    PASS    2bp_deletion
11
    chrY:76250767_CTG/C    C    44.12    PASS    2bp_deletion
26
    chrY:86118528_GGA/G    G    58.12    PASS    2bp_deletion
14
    chrY:90744554_TC/T    T    443.85    PASS    C_deletion
1
    chrY:90744588_CCCTAG/C    C    204.85    PASS    5bp_deletion
2
-----
seqinfo: 21 sequences from mm10 genome

$`WT-day77`

```

GRanges object with 6568 ranges and 7 metadata columns:

REF	seqnames	ranges	strand	paramRangeID
	<Rle>	<IRanges>	<Rle>	<factor> <D
NAStrngSet>				
chr1:4351880_T/TA	chr1	4351880	*	NA
T				
chr1:4611421_TTC/T	chr1	4611421-4611423	*	NA
TTC				
chr1:4777182_TTC/T	chr1	4777182-4777184	*	NA
TTC				
chr1:5455397_AGTGTGT/A	chr1	5455397-5455403	*	NA
AGTGTGT				
chr1:5779323_T/TA	chr1	5779323	*	NA
T				
...
...				
chrY:82534962_TTCTC/T	chrY	82534962-82534966	*	NA
TTCTC				
chrY:90739415_G/GAGAGTTTAAAAAGA	chrY	90739415	*	NA
G				
chrY:90739472_AG/A	chrY	90739472-90739473	*	NA
AG				
chrY:90739474_CG/C	chrY	90739474-90739475	*	NA
CG				
chrY:90800381_TC/T	chrY	90800381-90800382	*	NA
TC				
	ALT	QUAL	FILTER	mut
type	<DNAStrngSetList>	<numeric>	<character>	<charac
ter>				
chr1:4351880_T/TA	TA	44.99	PASS	T_inser
tion				
chr1:4611421_TTC/T	T	69.71	PASS	2bp_dele
tion				
chr1:4777182_TTC/T	T	52.98	PASS	2bp_dele
tion				
chr1:5455397_AGTGTGT/A	A	81.12	PASS	6bp_dele
tion				
chr1:5779323_T/TA	TA	38.12	PASS	T_inser
tion				
...
...				
chrY:82534962_TTCTC/T	T	81.12	PASS	4bp_dele
tion				
chrY:90739415_G/GAGAGTTTAAAAAGA	GAGAGTTTAAAAAGA	119.40	PASS	14bp_inser
tion				


```

tion      chrY:90739472_AG/A      A      76.30      PASS      C_dele
tion      chrY:90739474_CG/C      C      76.30      PASS      C_dele
tion      chrY:90800381_TC/T      T      79.98      PASS      C_dele

```

```

muttype_sub
<numeric>

```

```

      chr1:4351880_T/TA      11
      chr1:4611421_TTC/T      27
      chr1:4777182_TTC/T      17
      chr1:5455397_AGTGTGT/A      7
      chr1:5779323_T/TA      16
      ...
      chrY:82534962_TTCTC/T      7
chrY:90739415_G/GAGAGTTTAAAAAGA      1
      chrY:90739472_AG/A      1
      chrY:90739474_CG/C      1
      chrY:90800381_TC/T      5

```

```

-----

```

```

seqinfo: 21 sequences from mm10 genome

```

```

$`Pms1-day0`

```

```

GRanges object with 5294 ranges and 7 metadata columns:

```

	seqnames	ranges	strand	paramRangeID	R
EF					
	<Rle>	<IRanges>	<Rle>	<factor>	<DNAStringSe
t>					
CA	chr1:3045468_TCA/T	chr1 3045468-3045470	*	NA	T
AC	chr1:4702032_AAC/A	chr1 4702032-4702034	*	NA	A
TG	chr1:4759375_TG/T	chr1 4759375-4759376	*	NA	
TG	chr1:6085755_TTG/T	chr1 6085755-6085757	*	NA	T
GT	chr1:6996884_GT/G	chr1 6996884-6996885	*	NA	
..
TA	chrY:3824549_TA/T	chrY 3824549-3824550	*	NA	
C	chrY:3826886_C/CTGTTT	chrY 3826886	*	NA	
GC	chrY:29299377_GC/G	chrY 29299377-29299378	*	NA	
	chrY:90813687_AAG/A	chrY 90813687-90813689	*	NA	A

```

AG
chrY:90813695_TAC/T      chrY 90813695-90813697      * |      NA      T
AC
                                ALT      QUAL      FILTER      mutty
pe muttype_sub
                                <DNAStringSetList> <numeric> <character>      <characte
r> <numeric>
chr1:3045468_TCA/T      T      49.03      PASS      2bp_deleti
on      14
chr1:4702032_AAC/A      A      44.12      PASS      2bp_deleti
on      28
chr1:4759375_TG/T      T      60.12      PASS      C_deleti
on      4
chr1:6085755_TTG/T      T      44.13      PASS      2bp_deleti
on      23
chr1:6996884_GT/G      G      55.33      PASS      T_deleti
on      2
...      ...      ...      ...      .
..      ...
on      chrY:3824549_TA/T      T      55.12      PASS      T_deleti
6
chrY:3826886_C/CTGTTT      CTGTTT      81.12      PASS      5bp_inserti
on      9
chrY:29299377_GC/G      G      62.12      PASS      C_deleti
on      3
chrY:90813687_AAG/A      A      38.03      PASS      2bp_deleti
on      3
chrY:90813695_TAC/T      T      38.03      PASS 2bp_deletion_with_mi
..      1
-----
seqinfo: 21 sequences from mm10 genome

$`Pms1-day77`
GRanges object with 5789 ranges and 7 metadata columns:
                                seqnames      ranges strand | paramRangeID
REF
                                <Rle>      <IRanges> <Rle> |      <factor> <DNAString
Set>
chr1:4095024_ATG/A      chr1      4095024-4095026      * |      NA
ATG
chr1:4850769_CA/C      chr1      4850769-4850770      * |      NA
CA
chr1:4941825_CGTGT/C      chr1      4941825-4941829      * |      NA      C
GTGT
chr1:6108783_GA/G      chr1      6108783-6108784      * |      NA
GA
chr1:6706414_C/CTG      chr1      6706414      * |      NA

```

C
...					
chrY:89038622_TTG/T	chrY 89038622-89038624	*		NA	
TTG					
chrY:90800540_GGGAGAT/G	chrY 90800540-90800546	*		NA	GGG
AGAT					
chrY:90805299_CAGAG/C	chrY 90805299-90805303	*		NA	C
AGAG					
chrY:90813518_GACAGAC/G	chrY 90813518-90813524	*		NA	GAC
AGAC					
chrY:90813555_A/AGGTTAG	chrY 90813555	*		NA	
A					
	ALT	QUAL	FILTER		mut
type					
	<DNAStringSetList>	<numeric>	<character>		<character>
chr1:4095024_ATG/A	A	50.12	PASS		2bp_deletion
chr1:4850769_CA/C	C	66.12	PASS		T_deletion
chr1:4941825_CGTGT/C	C	80.00	PASS		4bp_deletion
chr1:6108783_GA/G	G	47.12	PASS		T_deletion
chr1:6706414_C/CTG	CTG	50.12	PASS		2bp_insertion
...		
chrY:89038622_TTG/T	T	54.12	PASS		2bp_deletion
chrY:90800540_GGGAGAT/G	G	76.02	PASS	6bp_deletion_with_	
mi..					
chrY:90805299_CAGAG/C	C	37.02	PASS		4bp_deletion
chrY:90813518_GACAGAC/G	G	73.02	PASS	6bp_deletion_with_	
mi..					
chrY:90813555_A/AGGTTAG	AGGTTAG	73.02	PASS		6bp_insertion
tion					
	muttype_sub				
	<numeric>				
chr1:4095024_ATG/A	19				
chr1:4850769_CA/C	2				
chr1:4941825_CGTGT/C	10				
chr1:6108783_GA/G	12				
chr1:6706414_C/CTG	17				
...	...				

```
chrY:89038622_TTG/T      17
chrY:90800540_GGGAGAT/G   1
chrY:90805299_CAGAG/C     2
chrY:90813518_GACAGAC/G   5
chrY:90813555_A/AGGTTAG   1
-----
seqinfo: 21 sequences from mm10 genome
```

Next count the number of indels per type. This results in a matrix that is similar to the mut_mat matrix.

[Hide](#)

```
indel_counts <- count_indel_contexts(indel_grl)
head(indel_counts)
```

	WT-day0	WT-day77	Pms1-day0	Pms1-day77
C_deletion_1	99	78	93	101
C_deletion_2	132	99	129	143
C_deletion_3	104	92	103	93
C_deletion_4	48	39	55	55
C_deletion_5	39	24	21	20
C_deletion_6+	244	309	161	223

[Hide](#)

```
# Normalize Indel counts
total_indel <- colSums(indel_counts)
indel_counts_norm <- t(apply(indel_counts, 1, function(y) round(y*10000/total_indel))
)
indel_counts_norm
```

	WT-day0	WT-day77	Pms1-day0	Pms1-day77
C_deletion_1	166	119	176	174
C_deletion_2	221	151	244	247
C_deletion_3	174	140	195	161
C_deletion_4	80	59	104	95
C_deletion_5	65	37	40	35
C_deletion_6+	409	470	304	385
T_deletion_1	189	111	149	161
T_deletion_2	298	269	278	297
T_deletion_3	154	108	159	138
T_deletion_4	60	52	64	78
T_deletion_5	32	35	42	36
T_deletion_6+	1015	1558	667	1273
C_insertion_0	40	40	55	73
C_insertion_1	39	46	34	31

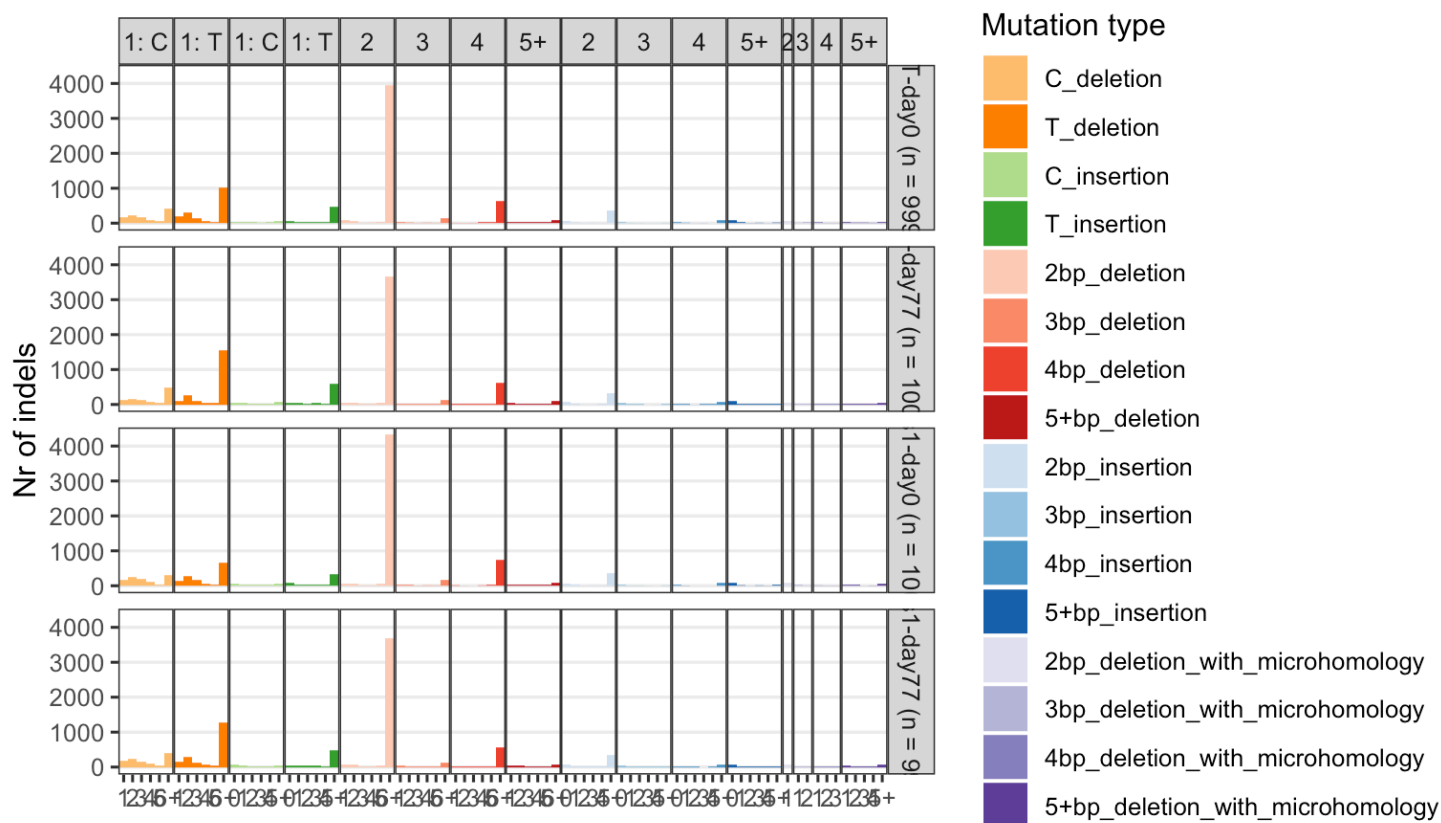
C_insertion_2	20	20	28	24
C_insertion_3	10	17	25	10
C_insertion_4	25	15	17	22
C_insertion_5+	54	64	59	47
T_insertion_0	55	43	83	55
T_insertion_1	35	41	36	43
T_insertion_2	28	18	36	31
T_insertion_3	28	32	32	36
T_insertion_4	18	18	23	28
T_insertion_5+	471	601	323	485
2bp_deletion_1	99	53	64	76
2bp_deletion_2	57	43	57	69
2bp_deletion_3	7	14	15	14
2bp_deletion_4	13	12	15	10
2bp_deletion_5	44	40	51	38
2bp_deletion_6+	3957	3665	4318	3691
3bp_deletion_1	18	23	26	33
3bp_deletion_2	17	17	19	17
3bp_deletion_3	7	6	6	5
3bp_deletion_4	12	12	13	19
3bp_deletion_5	8	15	11	10
3bp_deletion_6+	154	131	161	124
4bp_deletion_1	7	15	13	19
4bp_deletion_2	5	18	2	7
4bp_deletion_3	7	15	0	16
4bp_deletion_4	23	20	11	16
4bp_deletion_5	20	24	26	26
4bp_deletion_6+	640	624	748	556
5+bp_deletion_1	27	40	34	54
5+bp_deletion_2	37	12	42	33
5+bp_deletion_3	22	29	34	19
5+bp_deletion_4	27	29	28	26
5+bp_deletion_5	25	26	43	24
5+bp_deletion_6+	85	93	96	81
2bp_insertion_0	52	61	55	74
2bp_insertion_1	17	12	17	16
2bp_insertion_2	5	2	4	7
2bp_insertion_3	8	3	6	5
2bp_insertion_4	3	12	6	14
2bp_insertion_5+	362	314	363	333
3bp_insertion_0	37	38	21	43
3bp_insertion_1	8	12	8	9
3bp_insertion_2	5	5	6	5
3bp_insertion_3	2	0	2	2
3bp_insertion_4	2	2	0	2
3bp_insertion_5+	7	14	9	21
4bp_insertion_0	22	23	21	26

4bp_insertion_1	12	12	6	5
4bp_insertion_2	2	3	0	5
4bp_insertion_3	3	6	0	0
4bp_insertion_4	0	5	0	7
4bp_insertion_5+	74	79	87	81
5+bp_insertion_0	85	94	83	78
5+bp_insertion_1	23	15	15	17
5+bp_insertion_2	5	9	2	5
5+bp_insertion_3	10	6	11	9
5+bp_insertion_4	3	5	4	3
5+bp_insertion_5+	15	27	25	21
2bp_deletion_with_microhomology_1	59	35	77	69
3bp_deletion_with_microhomology_1	13	9	28	16
3bp_deletion_with_microhomology_2	18	9	11	12
4bp_deletion_with_microhomology_1	22	14	11	21
4bp_deletion_with_microhomology_2	7	11	6	9
4bp_deletion_with_microhomology_3	8	8	6	7
5+bp_deletion_with_microhomology_1	23	27	17	36
5+bp_deletion_with_microhomology_2	15	23	23	14
5+bp_deletion_with_microhomology_3	15	8	2	14
5+bp_deletion_with_microhomology_4	10	15	6	5
5+bp_deletion_with_microhomology_5+	42	44	60	60

Now you can plot the Indel spectra. The facets at the top show the indel types. First the C and T deletions. Then the C and T insertions. Next are the multi base deletions and insertions. Finally the deletions with microhomology are shown. The x-axis at the bottom shows the number of repeat units. For mh deletions the microhomology length is shown.

[Hide](#)

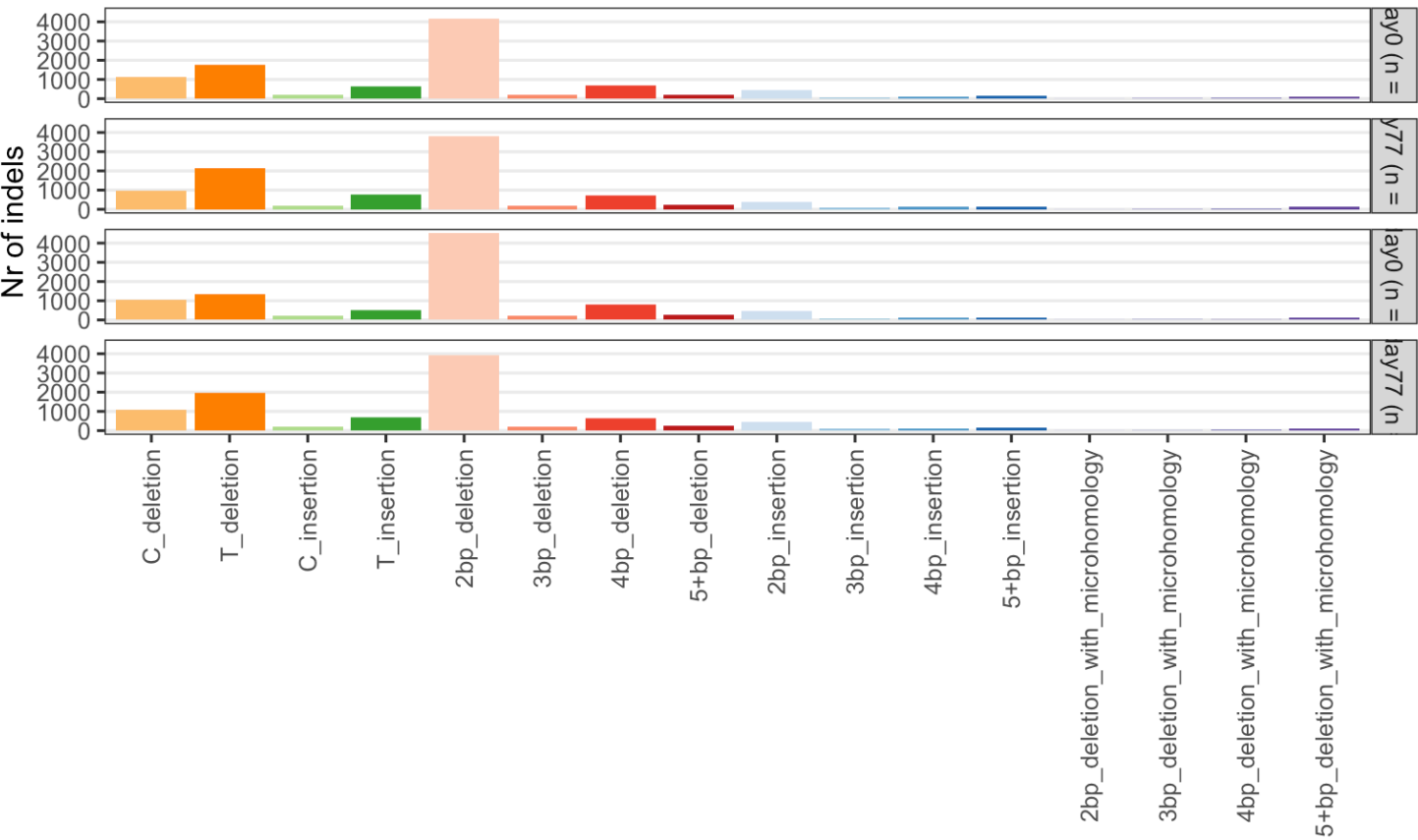
```
p_indel_context <- plot_indel_contexts(indel_counts_norm[,c(1:4)], condensed = TRUE,
same_y = TRUE)
ggsave2(filename = "p_indel_context_norm.pdf", plot = p_indel_context, width = 11, height = 8 )
p_indel_context
```



You can also choose to only plot the main contexts, without taking the number of repeat units or microhomology length into account.

Hide

```
plot_main_indel_contexts(indel_counts_norm[,c(1:4)], same_y = TRUE)
```



Fisher's exact test for normalized Indels

indel <chr>	WT- day0 <chr>	WT- day77 <chr>	Pms1- day0 <chr>	Pms1- day77 <chr>	p.value <chr>	p.adj <chr>	conf.int.down <chr>
T_deletion_1	189	111	149	161	2e-04	0.0201	1.31
T_deletion_6+	1015	1558	667	1273	5e-04	0.0413	1.1
2bp_deletion_1	99	53	64	76	0.001	0.08	1.35
4bp_deletion_6+	640	624	748	556	7e-04	0.0547	0.65

4 rows | 1-9 of 10 columns

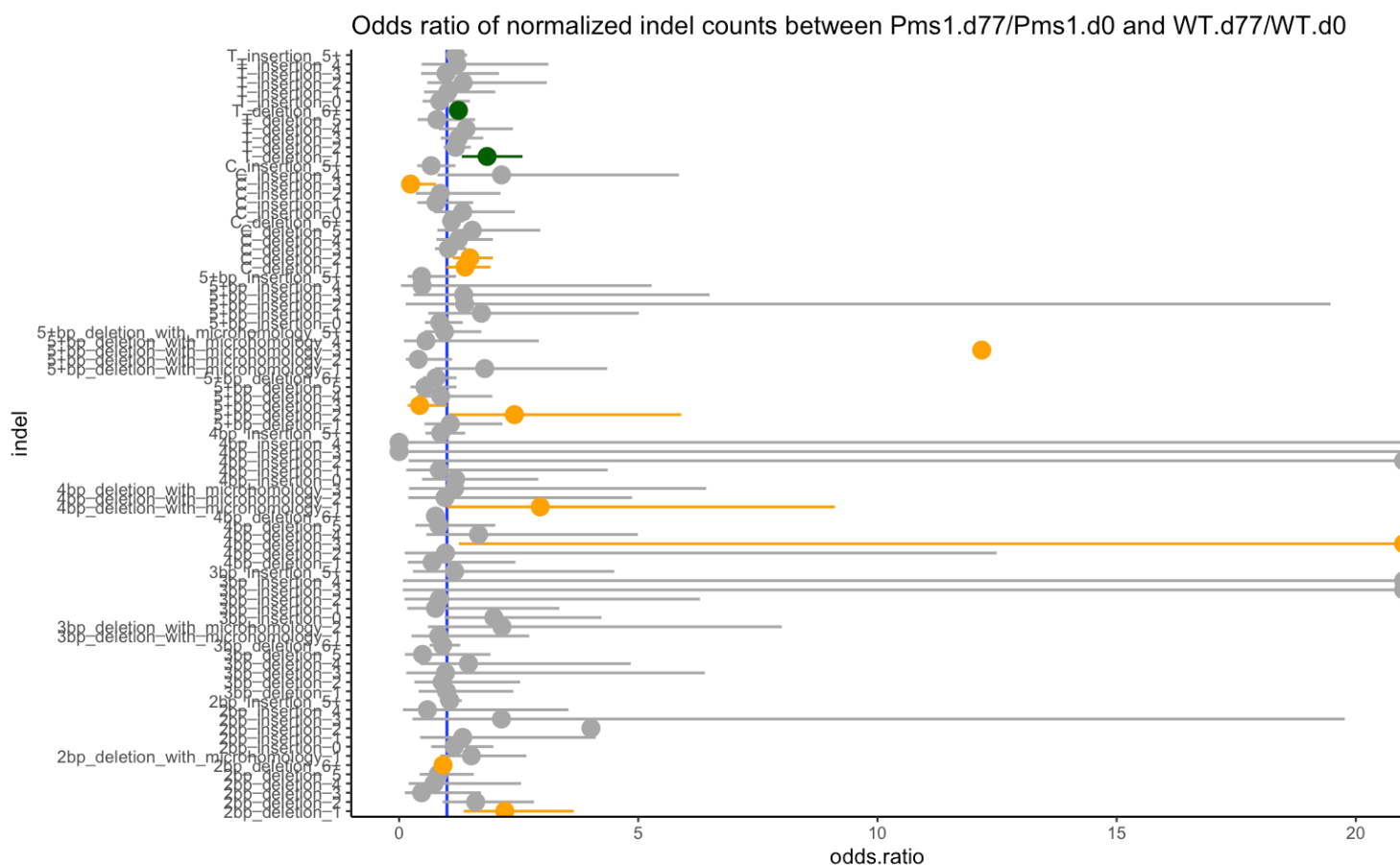
Plot COI for Indels

Hide


```
indel_mat_report$odds.ratio <- as.numeric(indel_mat_report$odds.ratio)
indel_mat_report$conf.int.down <- as.numeric(indel_mat_report$conf.int.down)
indel_mat_report$conf.int.up <- as.numeric(indel_mat_report$conf.int.up)

p.indel.coi <- ggplot(data = indel_mat_report, aes(x = indel, y = odds.ratio, ymin =
conf.int.down, ymax = conf.int.up)) +
  geom_hline(yintercept = 1, color = "blue" ) +
  geom_pointrange(colour=ifelse(indel_mat_report$p.adj <= 0.05,"darkgreen",ifelse(ind
el_mat_report$p.value <= 0.05,"orange","darkgrey"))) +
  ggtitle("Odds ratio of normalized indel counts between Pms1.d77/Pms1.d0 and WT.d77/
WT.d0") +
  theme_classic(base_size = 8) +
  coord_flip() +
  ylim(0,20)

ggsave2(filename = "indel_mat_report.pdf", plot = p.indel.coi, height = 11, width = 8
)
p.indel.coi
```


[Hide](#)

```
library("TxDb.Mmusculus.UCSC.mm10.knownGene")
genes_mm10 <- genes(TxDb.Mmusculus.UCSC.mm10.knownGene)
```

66 genes were dropped because they have exons located on both strands of the same reference sequence or on more than one reference sequence, so cannot be represented by a single genomic range.

Use 'single.strand.genes.only=FALSE' to get all the genes in a GRangesList object, or use suppressMessages() to suppress this message.

[Hide](#)

```
mut_strand(snp_grl[[1]], genes_mm10, mode = "transcription")
```

```
[1] untranscribed -          untranscribed untranscribed untranscribed untranscribed untranscribed
[8] transcribed   transcribed   transcribed   transcribed   transcribed   untranscribed
[15] transcribed   untranscribed transcribed   untranscribed untranscribed -
-
[22] -             -             untranscribed transcribed   untranscribed transcribed
[29] untranscribed untranscribed transcribed   transcribed   transcribed   transcribed
[36] transcribed   transcribed   untranscribed untranscribed untranscribed transcribed
[43] transcribed   transcribed   untranscribed transcribed   transcribed   transcribed
[50] transcribed   untranscribed transcribed   untranscribed untranscribed untranscribed
[57] untranscribed -             -             -             -             -
-
[64] untranscribed transcribed   transcribed   untranscribed transcribed   -
-
[71] -             untranscribed untranscribed untranscribed untranscribed transcribed
[78] untranscribed transcribed   untranscribed untranscribed -             -
-
[85] untranscribed -             -             -             -             -
-
[92] -             -             -             -             -             -
-
[99] -             -             -             untranscribed transcribed   untranscribed
```

ibed transcribed						
[106] transcribed	untranscribed	untranscribed	-		transcribed	transcribed
ed -						
[113] untranscribed	-	-	-	-	-	-
-						
[120] -	-	-	-	-	transcribed	transcribed
ed transcribed						
[127] -	untranscribed	transcribed	-	-	-	untranscribed
ibed -						
[134] -	untranscribed	-	-	-	-	-
-						
[141] -	-	-	-	-	-	-
-						
[148] -	-	-	-	-	-	-
-						
[155] -	-	-	-	-	-	-
-						
[162] -	-	-	-	-	-	-
-						
[169] -	-	-	-	-	-	-
-						
[176] -	transcribed	untranscribed	transcribed	transcribed	transcribed	untranscribed
ibed transcribed						
[183] transcribed	untranscribed	untranscribed	transcribed	transcribed	transcribed	untranscribed
ibed -						
[190] -	-	-	-	-	-	-
-						
[197] -	-	untranscribed	untranscribed	transcribed	transcribed	transcribed
ed transcribed						
[204] transcribed	untranscribed	transcribed	untranscribed	-	-	-
-						
[211] -	-	-	untranscribed	untranscribed	untranscribed	untranscribed
ibed transcribed						
[218] -	-	-	untranscribed	transcribed	transcribed	transcribed
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[225] -	-	-	-	-	-	transcribed
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[232] -	-	-	-	-	-	transcribed
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[239] transcribed	untranscribed	transcribed	transcribed	untranscribed	transcribed	transcribed
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[246] transcribed	transcribed	untranscribed	untranscribed	transcribed	transcribed	transcribed
ed transcribed						
[253] untranscribed	untranscribed	-	-	-	-	-
-						
[260] transcribed	untranscribed	untranscribed	untranscribed	transcribed	-	-
-						

[267]	transcribed ibed untranscribed	transcribed	transcribed	untranscribed	untranscribed	untranscribed
[274]	untranscribed ed -	transcribed	transcribed	transcribed	transcribed	transcribed
[281]	untranscribed -	transcribed	-	-	-	-
[288]	- -	-	-	-	-	-
[295]	- -	-	-	-	-	-
[302]	- -	-	-	-	-	-
[309]	- -	-	-	-	-	-
[316]	- -	-	-	-	-	-
[323]	- ed transcribed	-	-	-	transcribed	transcribed
[330]	untranscribed -	transcribed	transcribed	untranscribed	-	-
[337]	- -	transcribed	transcribed	untranscribed	transcribed	-
[344]	untranscribed ibed transcribed	transcribed	transcribed	transcribed	untranscribed	untranscribed
[351]	untranscribed ed transcribed	transcribed	untranscribed	transcribed	transcribed	transcribed
[358]	transcribed ed transcribed	untranscribed	transcribed	transcribed	untranscribed	transcribed
[365]	transcribed ed transcribed	transcribed	transcribed	transcribed	untranscribed	transcribed
[372]	- untranscribed	-	-	-	-	-
[379]	untranscribed untranscribed	transcribed	transcribed	untranscribed	-	-
[386]	untranscribed -	transcribed	-	untranscribed	-	-
[393]	untranscribed -	untranscribed	-	-	-	-
[400]	transcribed ibed untranscribed	transcribed	untranscribed	transcribed	untranscribed	untranscribed
[407]	transcribed -	-	transcribed	-	-	-
[414]	- -	-	-	-	-	-
[421]	- -	-	-	-	-	-
[428]	-	-	-	-	-	-

```

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[435] - - - - -
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ibed untranscribed
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[456] transcribed transcribed untranscribed transcribed transcribed transcrib
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[463] - - untranscribed - - -
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[470] - - - - untranscribed transcrib
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[477] untranscribed untranscribed transcribed untranscribed untranscribed untranscr
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[484] transcribed - transcribed untranscribed transcribed transcrib
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[491] untranscribed untranscribed untranscribed untranscribed untranscribed -
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[498] - - - - transcribed transcrib
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[505] - transcribed - - - -
-
[512] - - untranscribed - untranscribed untranscr
ibed -
[519] transcribed untranscribed untranscribed transcribed - transcrib
ed transcribed
[526] - - - - - untranscr
ibed transcribed
[533] transcribed - - - -
Levels: untranscribed transcribed -

```