**Supplementary Text**

**AGEseq (Analysis of Genome Editing by Sequencing) v1.0 User guide**

Liangjiao Xue and Chung-Jui Tsai, University of Georgia

March 20, 2015

Email Liangjiao Xue (lxue@uga.edu) for help or to report bugs

AGEseq compares amplicon sequences with expected target sequences and finds the insertion/deletion sites in the amplicon sequences. It is written in Perl and calls BLAT from the working directory or environment PATH.

AGEseq is available at AspenDB (<http://aspendb.uga.edu/downloads>) and GitHub (<https://github.com/liangjiaoxue/AGEseq>).

Download the “AGEseq.zip” file and unzip it on your local system, the AGEseq directory is the **working directory** for the analysis.

**1. Download necessary software**

1. Perl

a) Windows:  
Download and install ActivePerl ( <http://www.activestate.com/activeperl/downloads>) .

If you have difficulty installing the program on Windows 8 by double clicking the downloaded file, try the following command line:   
msiexec /i C:\dirctory\to\ActivePerl-XXX-MSWin32XXX.msi TARGETDIR="c:\perl" PERL\_PATH="Yes"

Hint: search for “cmd” APP to open the command line console. Type or copy the command line to the console, making sure the full address to the downloaded file name “.msi” is correct.

For more information, see: <http://docs.activestate.com/activeperl/5.20/install.html>

b) Other systems:

Perl is functional by default.

1. BLAT

a) Windows:  
Download blat\_windows.zip from <http://aspendb.uga.edu/downloads>

Unzip the directory and copy the blat.exe and cygwin1.dll file into the AGEseq working directory.

b) Other systems:  
Select an appropriate platform from: <http://hgdownload.cse.ucsc.edu/admin/exe/>  
Scroll down the application list, select “blat/”, and download “blat”

Copy the **executable file** to the working directory or put blat holding directory in the PATH:  
export PATH=$PATH:/usr/blat\_dir  
Change the permission of blat by typing: chmod 770 blat

To run the program with the test data provided with the package, go to **step 4** directly.

**2. Prepare the “reads” folder(s)**

The amplicon sequences need to be put into the “reads” folder, which can be found in the working directory. Two testing files are provided, which need to be deleted or moved before running real jobs.

The following file types are supported:

1. fastq (.fastq or .fq). The fq.gz files need to be unzipped first.
2. fasta (.fasta, .fas, .fa, .seq or .txt): either single or multiple sequences per file.

**3. Prepare the “design file”**

Open the text-delimited file named “targets” in the working directory using Excel or similar software.

The contents look like this:

|  |  |
| --- | --- |
| target | sequence |
| gene1\_1 | AGTCACCTGATCTTGACAAGCATGACTTGTCTTCTTTGAGGATGATAAAATCTGGAGGGGCTCCA |
| gene1\_2 | GGTCACCTGATCTTGACAAGCATGACTTGTCTTCTTTGAGGATGATAAAATCTGGAGGGGCTCCA |
| gene2 | CATATCTCGGAAGGTCGCTGCTGGGTTATCCAATTTGGGCATCAAGAAAGGCGATGT |

Replace the target (gene/allele) names and sequences with your own. The sequence usually spans 30-40 bp at each end of the predicted editing site. Using shorter sequences (rather than the entire amplicon length) usually results in more precise annotation. If allele sequences are included, they should be of the same length, or the BLAT engine may favor the longer allele for read assignment. Sequences of potential off-target sites can also be included, if your amplicon primers were designed to amplify homologous genes. Save the file.

**4. Run AGEseq**

1. Default settings:

Windows:

**Double click** the **Run\_AGEseq.bat** file.

Mac:

Change the permission of “**run\_AGEseq.sh**” by typing “chmod 770 run\_AGEseq.sh” in the working directory. Then **Double click** the **Run\_AGEseq.sh** file.

Other systems:

Change the permission of “**run\_AGEseq.sh**” by typing “chmod 770 run\_AGEseq.sh” in the working directory. Then run the program by typing:

./run\_AGEseq.sh

1. Advanced options:

The command to run AGEseq is as follows:

cd /dir/to/AGEseq

**perl AGEseq.pl reads targets.txt AGE\_output.txt**

# 1 read directory # 2 design file # 3 output file

The first two inputs are required and the output file (the third augment) is optional. A text-delimited output file named “AGE\_output.txt” will be generated by default if the output name is not provided. The filename extension “.txt” may be not shown on your system. The Perl script calls BLAT within the same directory to execute the analysis; please make sure BLAT is copied in the working directory or PATH environment. BLAT parameters can be customized to adjust the read matching sensitivity.

**5. User-configurable options:**

These parameters can be changed to customize the analysis strigency and reporting format.

Open the file “AGEseq.pl” with text editor like Notepad, and find the following lines (top 20 lines).

## setting for reports

my $mismatch\_cutoff = 0.05 ; # mismatch rate to filter low quality alignment, default 0.05(5%)

my $min\_cutoff = 0 ; # cutoff to filter reads with low abundance, default 0

my $wt\_like\_report = 20 ; # report top xx WT like records, default 20

my $indel\_report = 50 ; # report top xx records with indel, default 50

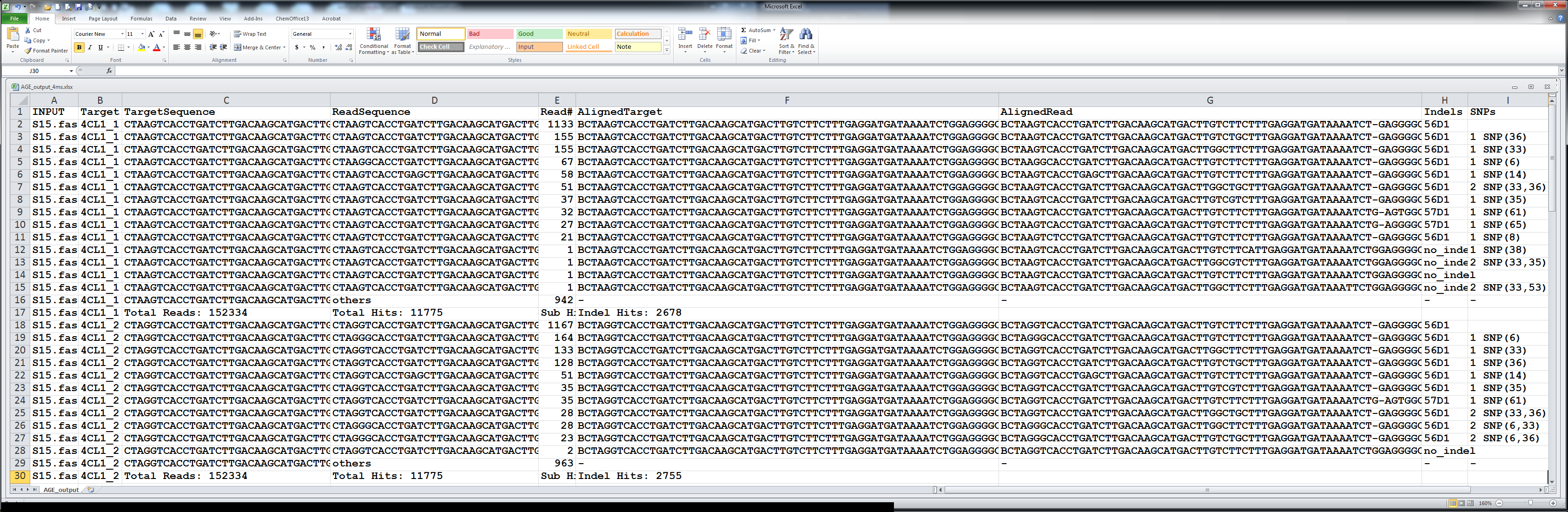
Modify the numbers shown in red color to re-set the values.

Reads with mismatches larger than $mismatch\_cutoff are dropped before read counting.  
Reads with abundance less than $min\_cutoff are ignored from individual cases, however they are counted in the summary section.  
$wt\_like\_report and $indel\_report are used to set total number of cases shown in the output file. The “ignored” reads are still counted in the summary section.

**6. Understanding the output file**

AGEseq results are summarized in a text-delimited output file which can be opened in Excel. Select a monospaced font (such as Courier or Courier New) to display sequence data.

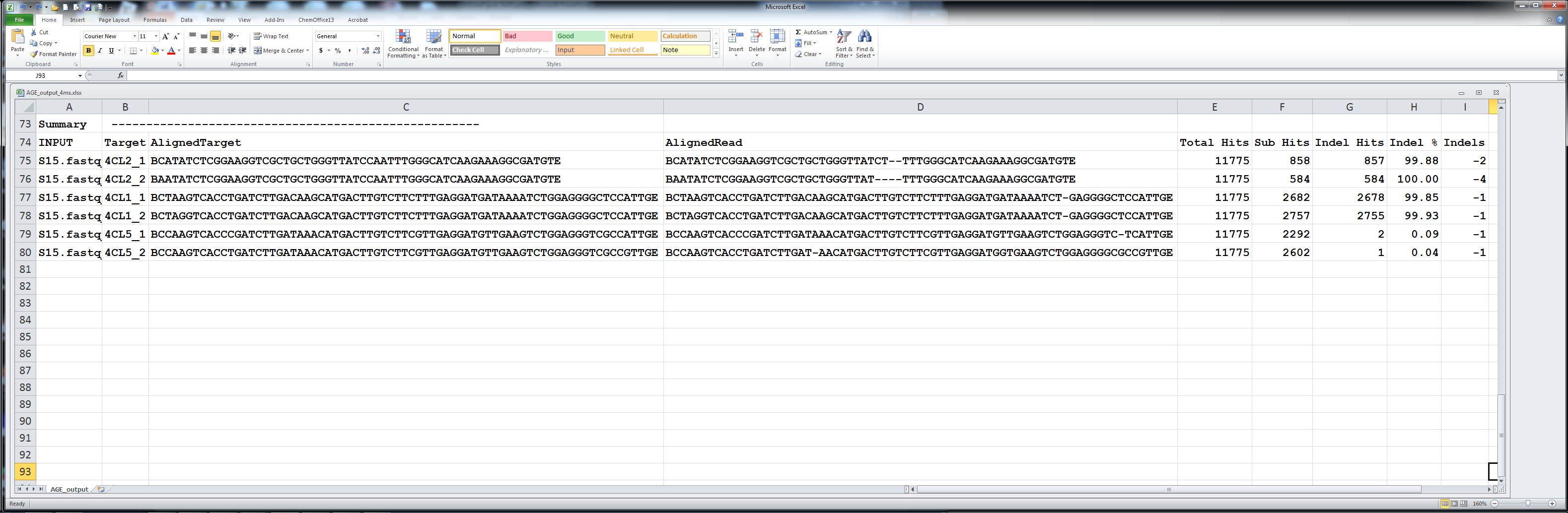
There are 9 columns (A through I) in the file (see the screenshot below).



1. INPUT: Name of the input read file. For Sanger data, individual read files are merged into one file and displayed with one name.
2. Target: Gene or allele name as provided in the design file.
3. TargetSequence: Target sequence as provided in the design file that has the best match to D.
4. ReadSequence: Amplicon sequence mapped to the target sequence in C.
5. Read#: Number of reads shown in D.
6. AlignedTarget: Target sequence displayed according to its alignment with the read sequence.
7. AlignedRead: Amplicon sequence displayed according to its alignment with the target sequence. Dashes are introduced to denote indels between the alignment pair (F and G).
8. Indels: Pattern of indels, if any, denoted by position (1st integer) of insertion (I) or deletion (D), followed by the number of indels (2nd integer). For example, 56D1 means 1-nt deletion at position 56.
9. SNPs: Patterns of SNPs, if any, denoted by the nucleotide position(s).

When multiple targets and/or samples are included in the analysis, each group (e.g., one target from one transgenic line) is separated by read statistics of the group. Within each group, only the top 20 non-indel reads and the top 50 indel reads, if present, are reported by default.

A summary is provided for each sequence input file at the end of the Excel file. The indel pattern, represented sequence and detection frequency for each target gene/allele are summarized in 9 columns.



1. INPUT: Name of the input read file.
2. Target: Gene or allele name as provided in the design file.
3. AlignedTarget: Target sequence displayed according to its alignment with the read sequence.
4. AlignedRead: Edited sequence displayed according to its alignment with the target sequence. By default, sequence from the predominant editing event is shown. Dashes are introduced to denote indels between the alignment pair (C and D).
5. Total hits: Total number of reads matching all target sequences in the file.
6. Sub hits: Number of reads matching the specific target/allele sequence.
7. Indel hits: Number of reads with indels relative to the target sequence.
8. Indel %: Fraction of indel hits (G) over the gene/allele-specific sub-hits (F).
9. Indels: Patterns of indels, if any, denoted by + (insertion) or – (deletion) and the number of nucleotides affected

**Known limitations and suggestions:**

1. Due to sequence alignment artifacts with BLAT or other similar programs concerning poly-nucleotides, it is highly recommended that the user manually inspects the AGE output and makes necessary adjustments. The first screenshot on the previous page shows an example of inconsistent indel-calling due to alignment artifacts (56D1 and 57D1 for both 4CL1\_1 and 4CL1\_2).

2) Sequence errors can be introduced during amplicon library preparation and sequencing that involve PCR, or by base-calling algorithms. These errors will appear as SNPs/indels in the AGE report. We suggest only indel reads with a coverage of 3-5 be considered further in the analysis. By default, AGE computes the “indel %”using the total number of indel reads regardless of the indel pattern. In the example above for 4CL1\_1, we detected 2682 hits matching this allele, 2678 with a 1-nt deletion and 4 with no indels (4 different SNPs, each with 1 hit). Of the 2678 indel hits, the predominant event (1133 hits) has perfect sequence match with the target sequence, except for the 1-nt deletion. The remaining 554 indel events contained distinct SNPs ranging from 1 to 3 nt, likely derived from sequence errors. User inspection and adjustment is therefore highly recommended. Alternatively, sequence matching stringency can be customized, or consider only reads without SNPs.

1. For analysis that includes both alleles, the indel detection frequency for each allele should be re-calculated by summing the total read counts from both alleles as the denominator. See the table below as an example (information re-calculated from read summary shown in the second screenshot above).

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Plant | Target | Sequence | Total Read # | Indel% | Indels |
| WT | 4CL2 | CATATCTCGGAAGGTCGCTGCTGGGTTATCCAATTTGGGCATCAAGAAAGGCGATGT  AATATCTCGGAAGGTCGCTGCTGGGTTATCCAATTTGGGCATCAAGAAAGGCGATGT | - | - | - |
| 201 | 4CL2 | CATATCTCGGAAGGTCGCTGCTGGGTTATCT--TTTGGGCATCAAGAAAGGCGATGT  AATATCTCGGAAGGTCGCTGCTGGGTTAT----TTTGGGCATCAAGAAAGGCGATGT | 857  584 | 59%  41% | -2  -4 |
| WT | 4CL1 | CTAAGTCACCTGATCTTGACAAGCATGACTTGTCTTCTTTGAGGATGATAAAATCTGGAGGGGCTCCATTG  CTAGGTCACCTGATCTTGACAAGCATGACTTGTCTTCTTTGAGGATGATAAAATCTGGAGGGGCTCCATTG | - | - | - |
| 105 | 4CL1 | CTAAGTCACCTGATCTTGACAAGCATGACTTGTCTTCTTTGAGGATGATAAAATCT-GAGGGGCTCCATTG  CTAGGTCACCTGATCTTGACAAGCATGACTTGTCTTCTTTGAGGATGATAAAATCT-GAGGGGCTCCATTG | 2682  2757 | 49%  51% | -1  -1 |
| 105 | 4CL5 | CCAAGTCACCCGATCTTGATAAACATGACTTGTCTTCGTTGAGGATGTTGAAGTCTGGAGGGTCGCCATTG  CCAAGTCACCTGATCTTGATAAACATGACTTGTCTTCGTTGAGGATGTTGAAGTCTGGAGGGTCGCCGTTG | 2290  2601 | 47%  53% | -  - |