Knockout Prediction for Reaction Networks with Partial Kinetic Information

 $Mathias John^{12}$, $Mirabelle Nebut^{12}$, and $Joachim Niehren^{13}$

BioComputing, LIFL (CNRS UMR8022)
 University of Lille
 INRIA Lille

Abstract. In synthetic biology, a common application field for computational methods is the prediction of knockout strategies for reaction networks. Thereby, the major challenge is the lack of information on reaction kinetics. In this paper, we propose an approach, based on abstract interpretation, to predict candidates for reaction knockouts, relying only on partial kinetic information. We consider the usual deterministic steady state semantics of reaction networks and a few general properties of reaction kinetics. We introduce a novel abstract domain over pairs of real domain values to compute the differences between steady states that are reached before and after applying some knockout. We show that this abstract domain allows us to predict correct knockout strategy candidates independent of any particular choice of reaction kinetics. Our predictions remain candidates, since our abstract interpretation over-approximates the solution space. We provide an operational semantics for our abstraction in terms of constraint satisfaction problems and illustrate our approach on a realistic network.

Keywords: Abstract interpretation, deterministic semantics, steady state, constraint satisfaction, synthetic biology.

1 Introduction

Synthetic biology aims at creating artificial micro-organisms, either by constructing them from scratch or by modifying existing once [2,1,19]. To this end, computational modeling is applied to predict the dynamic behavior of the resulting organisms [22,30]. Thereby, it is a common task to abstract micro-organisms as sets of chemical reactions and to predict the effects of reaction deletion on the dynamics of the concentrations of chemical species [36,5]. Such knockout considerations are in particular applied to predict strategies that increase the rate of some target reactions of interest.

Kinetic information is essential to predict the dynamics of chemical systems [20]. In reaction networks, each reaction is endowed with a kinetic rate law, that is a function that defines a rate in dependence of the concentration of the reaction's reactants. Common examples of rate laws are the law of mass action and Michaelis-Menten kinetics. These depend on kinetic rate constants that,

however, remain mostly unknown. Moreover, in practice, many of a model's reactions combine several unknown reaction steps to one, see e.g. [17, 18]. In these cases, only basic properties of the kinetics function are given, e.g. that the rate increases or decreases with the concentration of some chemical species. Thus, the precise prediction of the dynamics of chemical systems remains out of reach.

The commonly accepted dynamic semantics of systems of chemical reactions describes species concentrations in terms of stochastic processes [16]. When dealing with high concentrations, a meaningful approximation is to ignore the moments of probability distributions with an order greater than one [38, 15]. Such, so called, deterministic semantics is then represented by systems of ordinary differential equations (ODEs) that describe the changes in the mean of species concentrations over time. Knockout predictions then usually regard fix points of ODEs when no more changes in concentrations occur (steady state). It is known that in nature systems of chemical reactions exist that have none or more than one steady state [35, 9, 12].

Abstract interpretation [6] has been introduced for the static analysis of program semantics. The idea is to approximate the state space of programs based on approximations of domains and computations. In the realm of chemical reaction systems, abstract interpretation has been applied to obtain different approximations of their dynamic, stochastic semantics [10, 11] or to decrease the size of (infinitely) large sets of ODEs representing their deterministic semantics [8].

In this paper we propose an approach based on abstract interpretation, that predicts candidates for reaction knockout, with only partial kinetic information. We focus on cases that are on one hand simple, since reaction systems are assumed to always reach a steady state as it is usually done when predicting reaction knockouts [37, 5]. On the other hand, they are complicated, since kinetics functions are completely unknown, except for a few basic properties. Our major idea is to compare steady states of reaction systems before and after a reaction knockout is applied. We use abstract interpretation to reason about the effects of knockouts in the absence of kinetic information. Therefore, we introduce a small abstract domain that works on relations of pairs of non negative real numbers and abstract operators that represent an over-approximation of real domain addition and multiplication. We then propose a small set of properties for kinetics functions that are general enough to be also fulfilled by mass action kinetics and Michaelis-Menten kinetics. As our central contribution, we show that the predictions obtained with our abstract interpretation are independent of any particular choice of kinetics functions, as long as these fulfill these properties. Our predictions remain candidates due to the over-approximating nature of our approach. We also present an operational semantics for our approach based on a mapping of the abstract reaction semantics to constraint satisfaction problems [31].

We start off by introducing the deterministic and steady state semantics of reaction systems (Section 2). Then we formalize the reaction knockout task in terms of a real domain constraint satisfaction problem (Section 3). To do so, in

the first instance, we restrict ourselves to reactions with mass action kinetics. Then we define our abstract domain and the abstract interpretation of our mass action knockout constraints yielding sets of finite domain constraints (Section 4). We show that our abstract interpretation is a correct over-approximation in that every solution of the real domain problem is reflected by the solutions of the abstract domain problem. We proceed by introducing properties of kinetics functions and show that these are fulfilled by mass action kinetics and Michaelis-Menten kinetics (Section 5). Then, we prove that every kinetics function with these properties yields the same results in our abstraction, which allows us to generally stick to mass action in our abstract interpretation. We next show how to solve the thus obtained table constraints (Section 6). As this may yield large sets of solutions, we propose to integrate Branch-And-Bound optimization w.r.t. different criteria. We then provide in Section 7 a real-world example based on a model from literature and our own implementation and discuss the obtained solutions. These, on one hand, confirm results already known in literature and on the other hand represent new interesting strategies that are currently evaluated in wet-lab experiments. To finish, we provide a discussion of existing approaches for prediction reaction knockouts in absence of kinetic information (Section 8).

2 Reaction Networks

Let M be a finite set of chemical species (molecule types). A chemical solution S with molecules in M is a tuple $(S_m)_{m\in M}$ with $S_m \in \mathbb{R}_{\geq 0}$. We call S_m the concentration of m in S, that is the number of molecules of species m in S divided by the volume of S.

A reaction network over M is a finite set R of chemical reactions with species in M. Thereby, a chemical reaction is a rewrite rule that states how chemical solutions change over time. More formally, a chemical reaction r with species in M is a tuple of the the following form:

$$(r) m_1, \ldots, m_j : m_{j+1}, \ldots, m_k \xrightarrow{\kappa} m_{k+1}, \ldots, m_l$$

where $m_1, \ldots, m_l \in M$ and $\kappa : \mathbb{R}^k_{\geq 0} \to \mathbb{R}_{\geq 0}$ is a function. Species m_{j+1}, \ldots, m_k are called the reactants of r, i.e. molecules of this type are consumed when r is applied. Species on the right hand side are called the products of r, which are produced at application time. Species m_1, \ldots, m_j are called enzymes of r. They are neither consumed nor produced but may increase the rate of reaction r, which is defined by the kinetics function κ . We assume that species can only play a single role in every reaction, i.e. they are either enzyme, reactant, or product. However, in every role they might occur more than once. We write $enz_r(m)$, $react_r(m)$, and $prod_r(m)$ for the number of occurrences of m as an enzyme, reactant, or product of r.

A kinetics function of arity $k, \kappa : \mathbb{R}^k_{\geq 0} \to \mathbb{R}_{\geq 0}$, defines the rate (propensity) at which k-tuples of molecules of S may react, in function of the concentrations of the reactant and enzyme types in the solution. If (m_1, \ldots, m_k) are the enzyme

$$(\text{SPEC}) \frac{m \in M}{\frac{dS_m}{dt}(t) = \sum_{r \in R} (prod_r(m) - react_r(m)) \cdot rate_{S,r}(t)}$$

$$(\text{RATE}) \frac{r \in R \quad enz(r) = (m_1, \dots, m_j) \quad react(r) = (m_{j+1}, \dots, m_k)}{rate_{S,r}(t) = \kappa_r(S_{m_1}(t), \dots, S_{m_k}(t))}$$

Fig. 1. Deterministic semantics of reaction network R over M.

and reactant types of a chemical reaction with kinetics function κ then the reaction rate for a chemical solution S is equal to $\kappa(S_{m_1}, \ldots, S_{m_k})$. We write κ_r for the kinetics function of $r \in R$.

Standard chemical reactions have two reactants, no enzymes, and follow mass action kinetics [16]. The kinetics of such standard reactions is then simply defined as the product of the concentration of its reactants times some rate constant (see Appendix A for more details). However, as a first approximation this reaction model can be extended to arbitrary many enzymes and reactants. We denote this generalized version of the mass action kinetics function as $ma_c : \mathbb{R}^k_{\geq 0} \to \mathbb{R}_{\geq 0}$ for some constant $c \in \mathbb{R}_{\geq 0}$, such that $ma_c(x_1, \ldots, x_k) = c \cdot x_1 \cdot \ldots \cdot x_k$.

Other kinds of kinetics functions give better models of enzymatic reactions. These are justified by compositions of several standard reactions with mass-action kinetics. The most frequent example is Michaelis-Menten kinetics, which accounts for a single-reactant reaction that is triggered by a single enzyme. It is given by the kinetics function $mm_{c_1,c_2}: \mathbb{R}^2_{\geq 0} \to \mathbb{R}_{\geq 0}$, such that $mm_{c_1,c_2}(a,e) = c_1 \cdot a \cdot e/(c_2+a)$, where a,e are the concentrations of the reactant and enzyme, respectively. Rates following Michaelis-Menten kinetics describe a saturation curve that steadily increases with the concentration of the reactant but approaches a limit depending on the enzyme concentration. Yet another interesting alternative are Hill kinetics.

Deterministic Semantics. The deterministic semantics of a reaction network R is a collection of functions $(S_m)_{m\in M}$ of type $S_m: \mathbb{R}_{\geq 0} \to \mathbb{R}_{\geq 0}$. The value of $S_m(t)$ defines the concentration of m in the solution at time point t, so that the solution at time point t is $(S_m(t))_{m\in M}$. If the initial solution at time point 0 is fixed then the solutions at all later time points t>0 are determined by a collection of ODEs (see below). However, we might not know the initial solution in practice.

The deterministic semantics of a reaction network R over M is defined by applying rule (SPEC) to each species in M. The resulting ODEs compute the change of S_m at time point t by substituting reactants by products for all chemical reactions $r \in R$ according to their rate, see Fig. 1. Rule (RATE) defines the rate of a reaction r at time point t by $rate_{S,r}(t) = \kappa_r(S_{m_1}(t), \ldots, S_{m_k}(t))$.

Steady-State Semantics. The steady state semantics assumes that reaction networks reach a fixed point in which all reactions continue to perform with constant

$$(\text{SPEC}_{st}) \frac{m \in M}{\sum_{r \in R} prod_r(m) \cdot rate_{S,r} = \sum_{r' \in R} react_{r'}(m) \cdot rate_{S,r'}}$$

$$(\text{RATE}_{st}) \frac{r \in R \quad enz(r) = (m_1, \dots, m_j) \quad react(r) = (m_{j+1}, \dots, m_k)}{rate_{S,r} = \kappa_r(S_{m_1}, \dots, S_{m_k})}$$

Fig. 2. Steady state semantics of reaction network R over M.

speed. This means that the changes for all $m \in M$ become zero:

$$\frac{dS_m}{dt}(t) = 0$$

The amounts $S_m(t)$ will thus become constant for all species $m \in M$, so that we can denote them by S_m . Furthermore, reactions $r \in R$ become constant fluxes with constant rates, so that $rate_{S,r}(t)$ becomes constant and can thus be denoted by $rate_{S,r}$.

The steady state semantics of a reaction network R is given by the system of arithmetic equations in Fig. 2. These equations relate molecule concentrations S_m to reaction rates $rate_{S,r}$. Rule (SPEC_{st}) states that the production and consumption must coincide for any molecule type. It should be noticed that these equations are only a necessary condition for steady states of the dynamic system (and that some systems may not have any steady state). However, as mentioned earlier, we assume that reaction networks always reach a steady state.

Example. We consider a reaction network where A's and B's are inputs from the environment that can react to a complex C which is then released into the environment. We assume that all inputs and outputs are done such that an equilibrium must be reached.

$$\begin{array}{cccc} (r_1) & \xrightarrow{ma_1} A & (r_2) & \xrightarrow{ma_2} B & (r_3) A, B \xrightarrow{ma_3} C \\ (r_4) A \xrightarrow{ma_4} & (r_5) B \xrightarrow{ma_5} & (r_6) & C \xrightarrow{ma_6} \end{array}$$

Note that we chose artificial rate constants i for reaction r_i . In practice the situation will be even worse in that most rate constants will be unknown. The deterministic semantics is given by the following system of ODEs:

$$\begin{split} \frac{dS_A}{dt}(t) &= 1 - 3 \cdot S_A(t) \cdot S_B(t) - 4 \cdot S_A(t), \\ \frac{dS_B}{dt}(t) &= 2 - 3 \cdot S_A(t) \cdot S_B(t) - 5 \cdot S_B(t), \\ \frac{dS_C}{dt}(t) &= 3 \cdot S_A(t) \cdot S_B(t) - 6 \cdot S_C(t). \end{split}$$

In order to determine the functions S_A , S_B , and S_C , it is sufficient to fix the initial solution. We will choose $S_A(0) = S_B(0) = S_C(0) = 0$ for illustration, which leads to the dynamics drawn in Fig. 3. It should be noticed that the concentrations of all molecules stabilize when time t tends to infinity, i.e., a steady

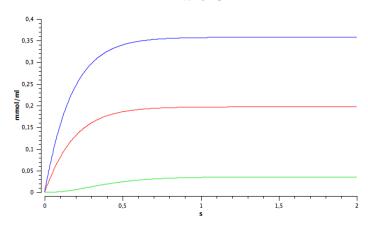


Fig. 3. Dynamics of example reaction network.

state solution is reached. The ODEs induce the following arithmetic equations for this steady state:

$$1 = 3 \cdot S_A \cdot S_B + 4 \cdot S_A, \qquad 2 = 3 \cdot S_A \cdot S_B + 5 \cdot S_B, \qquad 3 \cdot S_A \cdot S_B = 6 \cdot S_C.$$

One can now solve these quadratic equations to determine two solutions for S_A :

(1)
$$S_A = \frac{-23 + \sqrt{769}}{24}$$
 (2) $S_A = \frac{-23 - \sqrt{769}}{24}$

As only the second solution is positive, we obtain a single steady state at $S_A=0.197119,\,S_B=0.357695,\,{\rm and}\,\,S_C=0.0352542.$

3 Reaction Knockouts

We are now interested in modifying reaction networks, such that the rate of some reactions are increased or decreased in the steady state. The only modifications that we permit are reaction knockouts, i.e. inactivation of some reactions. As mentioned in introduction, we assume in this section that all reactions have mass action kinetics. In Section 5, we extend our approach to a more general, only partially known kinetics, relying on our abstract interpretation given in Section 4.

The knockout problem that we want to study is the following: we are given a reaction network R and some objective O. An objective compares the rate of reactions in steady states that R reaches before and after applying some reaction knockouts. O may state that the rate of some reactions r should be increased (denoted inc(r)), decreased (dec(r)), that a reaction may not be knocked out (on(r)), or conjunctions thereof. We then try to find a set of reactions $R' \subseteq R$

$$(\operatorname{SPEC}_{ko}) \frac{m \in M}{\sum_{r \in R} \operatorname{prod}_r(m) \cdot \operatorname{rate}_{S,r}^{on} = \sum_{r' \in R} \operatorname{react}_{r'}(m) \cdot \operatorname{rate}_{S,r'}^{on}}$$

$$(\operatorname{RATE}_{ko}) \frac{r \in R}{r} \frac{\kappa_r = \operatorname{ma}_{c_r} \operatorname{enz}(r) = (m_1, \dots, m_j) \operatorname{react}(r) = (m_1, \dots, m_k)}{\operatorname{rate}_{S,r}^{on} = c_r \cdot S_{m_1} \cdot \dots \cdot S_{m_k} \cdot \operatorname{on}_r}$$

Fig. 4. Knockout steady states semantics of mass action reaction networks R over M.

to knockout, i.e. a knockout strategy, such that there exist solutions S, S' that are steady states for R and $R \setminus R'$, respectively, and satisfy O. Notice that this knockout problem could also be defined to compare several steady states that are possibly reached before and after applying some knockout. We leave such extensions to future work, see Section 9.

In order to solve the above problem, we have to reason about the steady state semantics of a system of chemical reactions and all its subsets at the same time. We first introduce the notion of reaction knockout in the semantics (knockout steady state semantics) then we reduce the problem to reasoning with a single set of arithmetic constraints.

Knockout Steady State Semantics. We enrich our steady state semantics, such that it supports the knockout of a subset of the chemical reactions. The idea is to introduce a Boolean value $on_r \in \{0,1\}$ for all reactions $r \in R$, which expresses whether or not reaction r is switched on (that is $on_r = 1$). This leads us to the knockout steady state semantics in Fig. 4.

Language of Arithmetic Constraints. We introduce the following set of variables:

$$V = \{x_r, x_{on_r} \mid r \in R\} \cup \{x_m \mid m \in M\}$$

Variables x_m denote the unknown concentration S_m , variables x_r the unknown reaction rate $rate_{S,r}^{on}$, and variables x_{on_r} the unknown Boolean value on_r . We then consider the following language of arithmetic constraints where $x, y, z \in V$ and $c \in \mathbb{R}_{>0}$:

$$\phi ::= x + y = z \mid x \cdot y = z \mid x = c \mid x \in \{0, 1\} \mid \phi \land \phi' \mid \exists x. \phi$$

The conditions of the knockout semantics for a mass action reaction network R can now be expressed by the arithmetic constraint ϕ_R defined in Fig. 5. Notice that the constraint given there can be flattened easily, so that it belongs to the constraint language specified above. Notice further that equalities of the form x = y can be expressed by $\exists z.(x + z = y \land z = 0)$.

Let ν be a variable assignment into the domain $\mathbb{R}_{\geq 0}$. The constraint problem that we try to solve is now as follows: given a reaction network R, with its knockout constraint ϕ_R and an objective O, find variable assignments ν, ν' , such that ν, ν' satisfy ϕ_R , for all reactions $r \in R$ it holds that $\nu(x_{on_r}) = 1$, and (ν, ν') satisfies objective O. The reactions r for which it holds that $\nu'(x_{on_r}) = 0$ then define our reaction knockout strategy.

$$(\text{RATHM}_{con}) \ \phi_R = \bigwedge_{m \in M} \phi_m \wedge \bigwedge_{r \in R} \phi_r$$

$$(\text{SPEC}_{con}) \ \frac{m \in M}{\phi_m = \left(\sum_{r \in R} prod_r(m) \cdot x_r = \sum_{r' \in R} react_{r'}(m) \cdot x_{r'}\right)}$$

$$(\text{RATE}_{con}) \ \frac{r \in R}{\phi_r} \ \kappa_r = ma_{c_r} \ enz(r) = (m_1, \dots, m_j) \ react(r) = (m_1, \dots, m_k)}{\phi_r = (x_r = c_r \cdot x_{m_1} \cdot \dots \cdot x_{m_k} \cdot x_{on_r} \wedge x_{on_r} \in \{0, 1\})}$$

Fig. 5. Knockout constraint ϕ_R for mass action reaction networks R.

4 Abstract Interpretation

The next idea is to reason about changes in concentrations in steady states when switching off reactions. This is done by interpreting arithmetic constraints abstractly into finite domain table constraints.

Abstract Domain & Relations. We are interested in capturing the differences between pairs of nonnegative real numbers $(u, u') \in \mathbb{R}^2_{\geq 0}$. We distinguish the cases where u > u', u < u', and u = u', and in addition those cases where u or u' are equal to 0. More formally, we define the following set of difference relations:

$$\Delta = \{\uparrow, \downarrow, \sim, \uparrow, \downarrow, \approx\} \subseteq \mathbb{R}^2_{>0}$$

such that the following properties hold for all $u, u' \in \mathbb{R}_{>0}$:

$$\begin{array}{ll} u \uparrow u' \Leftrightarrow 0 < u < u' & u \uparrow u' \Leftrightarrow 0 = u < u' \\ u \downarrow u' \Leftrightarrow u > u' > 0 & u \downarrow u' \Leftrightarrow u > u' = 0 \\ u \sim u' \Leftrightarrow u = u' > 0 & u \approx u' \Leftrightarrow u = u' = 0 \end{array}$$

Lemma 1. For any $(u, u') \in \mathbb{R}^2_{\geq 0}$, there exists a unique $\delta \in \Delta$ such that $(u, u') \in \delta$.

Given an arithmetic relation $p \subseteq \mathbb{R}^n_{\geq 0}$, we next define an abstract relation $p^{\alpha} \subseteq \Delta^n$, such that for all difference relations $(\delta_1, \ldots, \delta_n) \in \Delta^n$:

$$(\delta_1,\ldots,\delta_n) \in p^{\alpha} \Leftrightarrow \exists (u_1,\ldots,u_n) \in p \ \exists (u'_1,\ldots,u'_n) \in p. \ \bigwedge_{i=1}^n (u_i,u'_i) \in \delta_i$$

In particular, we define abstract multiplication and addition functions \cdot^{α} , $+^{\alpha} \in \Delta^3$ from binary multiplication and addition functions, seen as ternary relations. The tables of these two relations are spelled out in Fig. 6.

Abstract Constraints. Abstract constraints are finite domain table constraints, whose variables have values in Δ , subject to constraints based on the abstract relations $+^{\alpha}$ and \cdot^{α} . We consider the following language of abstract constraints where $x, y, z \in V$ and $\Delta' \subseteq \Delta$:

$$\psi ::= +^{\alpha}(x, y, z) \mid \cdot^{\alpha}(x, y, z) \mid x \in \Delta' \mid \psi \wedge \psi'$$

$+^{\alpha}$	1	\uparrow	+	₩	~	\approx
\uparrow	1	1	\uparrow,\downarrow,\sim	\uparrow,\downarrow,\sim	1	\uparrow
\uparrow	sy.	1	\uparrow,\downarrow,\sim	\uparrow,\downarrow,\sim	\uparrow	\uparrow
\downarrow	sy.	sy.	↓	+	\downarrow	\downarrow
₩	sy.	sy.	sy.	₩	\downarrow	\Downarrow
~	sy.	sy.	sy.	sy.	2	7
\approx	sy.	sy.	sy.	sy.	sy.	\approx

\cdot^{α}	1	1	↓	₩	~	\approx
\uparrow	1	介	\uparrow,\downarrow,\sim	₩	1	\approx
1	sy.		1	\approx	1	\approx
\downarrow	sy.	sy.	+	₩		\approx
$ \downarrow $	sy.	sy.	sy.	₩	₩	\approx
\sim	sy.	sy.	sy.	sy.	~	\approx
\approx	sy.	sy.	sy.	sy.	sy.	\approx

Fig. 6. Abstraction of binary addition and multiplication functions (sy. = symmetric).

We first show who to compile objectives to abstract constraints:

The condition that initially all reactions are on is expressed by:

$$\bigwedge_{r \in R} x_{on_r} \in \Delta \setminus \{ \approx, \uparrow \}$$

We next use abstract interpretation in order to map arithmetic constraints to abstract constraints:

$$\begin{split} \llbracket x+y=z \rrbracket &= +^{\alpha}(x,y,z) & \llbracket x\cdot y=z \rrbracket = \cdot^{\alpha}(x,y,z) \\ \llbracket x=c \rrbracket &= x \in \{\sim\}, \text{with } c>0 & \llbracket x \in \{0,1\} \rrbracket = x \in \Delta \setminus \{\uparrow,\downarrow\} \\ \llbracket x=c \rrbracket &= x \in \{\approx\}, \text{with } c=0 & \llbracket \phi \wedge \phi' \rrbracket = \llbracket \phi \rrbracket \wedge \llbracket \phi' \rrbracket \\ \llbracket \exists x.\phi \rrbracket &= \exists x.\llbracket \phi \rrbracket \end{aligned}$$

Consider the constraint x=c which means that x is a constant that cannot be changed. Therefore, it is interpreted as $x\in\{\sim\}$, if c>0 or $x\in\{\approx\}$, else. Or consider the constraint $x+y=z\wedge y=0$. This is expressed by the corresponding abstract constraint $+^{\alpha}(x,y,z)\wedge y\in\{\approx\}$.

A pair (ν, ν') of two variable assignments into $\mathbb{R}_{\geq 0}$ induces a variable assignment μ into Δ , such that $\mu(x)$ is the difference relation between $\nu(x)$ and $\nu'(x)$ which is unique by Lemma 1. That is for all $\delta \in \Delta$ and $x \in V$:

$$\mu(x) = \delta \Leftrightarrow (\nu(x), \nu'(x)) \in \delta$$

We say that a pair (ν, ν') satisfies ψ if and only if μ is a solution of ψ .

We are now able to compile knockout constraint satisfaction problems into abstract domains. Consider, e.g., a simple knockout problem, where the reaction network is given as follows and the objective is to increase the rate of reaction r_4 :

$$(r_1) \xrightarrow{ma_2} A$$
 $(r_2) A \xrightarrow{ma_1} B$ $(r_4) B \xrightarrow{ma_1} C$ $(r_5) C \xrightarrow{ma_1} C$

Our objective is compiled with $x_{r_4} \in \{\uparrow, \uparrow\}$. Following our knockout semantics, we introduce the following constraint for our reaction network:

$$\begin{bmatrix} \bigwedge_{i=1}^{5} x_{on_i} \in \{0,1\} & x_{r_3} = x_A \cdot x_{on_3} & x_{r_1} = x_{r_2} + x_{r_3} \\ \exists x. (x_{r_1} = x \cdot x_{on_1} \wedge x = 2) & x_{r_4} = x_B \cdot x_{on_4} & x_{r_2} = x_{r_4} \\ x_{r_2} = x_A \cdot x_{on_2} & x_{r_5} = x_C \cdot x_{on_5} & x_{r_3} = x_{r_5} \end{bmatrix}$$

Solving the resulting constraint, we obtain that $x_{r_1} \in \{\sim, \downarrow\}$, such that for any solution μ also satisfying our objective it holds that $\mu(x_{r_1}) = \sim$. Thus, there exist only two solutions, either a knockout r_3 alone $(\mu(x_{on_3}) = \downarrow)$ or of both reactions r_3 and r_5 . Notice that a knockout of reaction r_5 only is not possible, since in this case, due to the constant production of $A(\mu(x_{r_1}) = \sim)$ and thus also C, no steady state is reached, i.e. we do not obtain a solution for our constraint problem.

Correctness. We now show that the abstract interpretation is correct in that every solution of the real domain problem is reflected by the solutions of the abstract domain problem.

Proposition 1 (Correctness of abstract interpretation). Let ϕ be an arithmetic constraint and ν, ν' variable assignments into $\mathbb{R}_{\geq 0}$. It holds that if ν, ν' satisfy ϕ then (ν, ν') satisfies $\llbracket \phi \rrbracket$.

Proof. By induction on the definition of arithmetic constraints.

- $\phi = x + y = z$ Let ν and ν' be both solutions of x + y = z. Then we have $\nu(x) + \nu(y) = \nu(z)$ and $\nu'(x) + \nu'(y) = \nu'(z)$. It follows from the definitions of μ and $+^{\alpha}$ that $(\mu(x), \mu(y), \mu(z)) \in +^{\alpha}$, i.e., μ is a solution of the abstract constraint $+^{\alpha}(x, y, z)$, i.e. of $\llbracket \phi \rrbracket$
- $\phi = x \cdot y = z$ analoguous to +.
- $\phi = (x = c)$ If c > 0 then $\nu(x) = c = \nu'(x)$ and thus $\mu(x) = \sim$, i.e., μ satisfies $\|\phi\| = x$. The case of ϕ being x = 0 is analogous.
- $\phi = x \in \{0,1\}$ It holds that $\nu(x)$ and $\nu'(x)$ belong to $\{0,1\}$. There are 4 possible cases, showing that the difference relation $\mu(x)$ between $\nu(x)$ and $\nu'(x)$ must be either of $\{\uparrow, \downarrow, \sim, \approx\}$ and thus μ satisfies $[\![\phi]\!] = x \in \{\uparrow, \downarrow, \sim, \approx\}$.
- $\phi = \exists x.\phi'$ We obtain that $\llbracket \phi \rrbracket = \exists x.\llbracket \phi' \rrbracket$. Since ν, ν' satisfy ϕ , there exist x and $u, u' \in \mathbb{R}_{\geq 0}$, such that $\nu \cup (x, u), \nu' \cup (x, u')$ satisfy ϕ' . Thus, by induction hypothesis, it holds that $(\nu \cup (x, u), \nu' \cup (x, u'))$ satisfy $\llbracket \phi' \rrbracket$ and thus that (ν, ν') satisfies $\llbracket \phi \rrbracket$.
- $\phi = \phi_1 \wedge \phi_2$ Since ν, ν' satisfy both ϕ_1 and ϕ_2 , the induction hypothesis provides that (ν, ν') satisfies $\llbracket \phi_1 \rrbracket$ and $\llbracket \phi_2 \rrbracket$. Thus, (ν, ν') also satisfies $\llbracket \phi_1 \rrbracket \wedge \llbracket \phi_2 \rrbracket$, which equals $\llbracket \phi \rrbracket$.

Proposition 1 states that every solution of a model in the real domain is reflected by its abstract interpretation. However, the converse does not hold. Consider, e.g., the constraint $\phi = \phi_1 \wedge \phi_2$, with $\phi_1 = (x_1 = x_2)$ and $\phi_2 = (x_1 = x_2 + x_3)$. For all ν that satisfy $\phi \wedge \phi'$, we obtain that $\nu(x_3) = 0$.

However, with our abstraction interpretation, μ satisfies $\llbracket \phi \rrbracket \land \llbracket \phi' \rrbracket$, even with $\mu(x_1) = \mu(x_2) = \mu(x_3) = \uparrow$. This is a correct approximation, since there exist pairs $(\nu_1, \nu'_1), (\nu_2, \nu'_2)$, such that ν_1, ν'_1 satisfy ϕ_1 and ν_2, ν'_2 satisfy ϕ_2 , and that, although differing, correspond both to μ . For example:

$$\begin{array}{ll} \nu_1 = \{(x_1,1),(x_2,1)\} & \nu_2 = \{(x_1,2),(x_2,1),(x_3,1)\} \\ \nu_1' = \{(x_1,2),(x_2,2)\} & \nu_2' = \{(x_1,4),(x_2,2),(x_3,2)\} \\ \nu_1(x_i) \uparrow \nu_1'(x_i), i \in \{1,2\} & \nu_2(x_i) \uparrow \nu_2'(x_i), i \in \{1,2,3\} \end{array}$$

Such a constraint results, e.g., from applying rule (SUBST $_{con}$) to species A and B, considering the following cyclic reaction network:

$$(r_1) A \xrightarrow{ma_1} B \quad (r_2) B \xrightarrow{ma_1} A \quad (r_3) B \xrightarrow{ma_1}$$

To this end, our approach, can be improved in different ways. On one hand, additional abstract relations could be defined, e.g. for commonly occuring patterns in reaction sets, like cycles of certain length. On the other hand, different methods could be applied to simplify equations. For example, one could use Gaussian elimination to symbolically solve the system of linear equations given by ϕ_1, ϕ_2 and account for the fact that $x_3 = 0$ by adding the constraint $x_3 \in \{\approx\}$. We leave such improvements as subject to future work.

5 Abstract Kinetics Functions

In the following, we extend our approach to more general kinetics. That is, we introduce properties of kinetics functions and show how kinetics functions fulfilling these properties are treated in our abstract interpretation.

The three properties of kinetics functions we consider are continuity, strict monotonicity, and conjunctiveness. Rates following strictly monotonic kinetics increase in the concentration of any reactant or enzyme. More formally, we call a kinetics function $\kappa: \mathbb{R}^n_{\geq 0} \to \mathbb{R}_{\geq 0}$ strictly monotonic if and only if for all $x_1, \ldots, x_n, x \in \mathbb{R}_{\geq 0}$ and all $i \in \{1, \ldots, n\}$ it holds:

$$x_i < x \Rightarrow \kappa(x_1, \dots, x_n) < \kappa(x_1, \dots, x_{i-1}, x, x_{i+1}, \dots, x_n)$$

Reactions that come with conjunctive kinetics can only perform if all its reactants and enzymes are present. More precisely, a kinetics function $\kappa : \mathbb{R}^n_{\geq 0} \to \mathbb{R}_{\geq 0}$ is called conjunctive if and only of for all $i \in \{1, ..., n\}$ it holds:

$$\bigwedge_{i=1}^{n} x_i \neq 0 \Leftrightarrow \kappa(x_1, \dots, x_n) \neq 0$$

In fact, the most widely used kinetics, mass action and Michaelis-Menten, are continuous, strictly monotonic, and conjunctive.

Lemma 2. Any mass action function ma_c (with c > 0) is strictly monotonic, continuous, and conjunctive.

Proof. Clear, since, by definition, we obtain $ma_c(x_1, \ldots, x_n) = c * \prod_{i=1}^n x_i$ for a reactions with order n.

Lemma 3. The Michaelis-Menten function $mm_{c,c'}$ (with c,c'>0) is strictly monotonic, continuous, and conjunctive.

Proof. By definition, we obtain $mm_{c,c'}(x_1,x_2) = c * x_1 * x_2/(c'+x_1)$. Strictly monotonic in x_2 , continuous, and conjunctive clear. Strictly monotonic in x_1 becomes obvious by:

$$\frac{c*x_1*x_2}{c'+x_1} = \frac{c*x_1*x_2}{x_1*(\frac{c'}{x_1}+1)} = \frac{c*x_2}{\frac{c'}{x_1}+1}$$

We obtain that in our abstract interpretation any two kinetics functions of the same arity provide the same results.

Proposition 2. Let $\kappa_1, \kappa_2 \subseteq \mathbb{R}^n_{\geq 0} \to \mathbb{R}_{\geq 0}$ be strictly monotonic, continuous, and conjunctive kinetics functions. It holds that $\kappa_1^{\alpha} = \kappa_2^{\alpha}$.

The proof is elaborated in Appendix B.

Since also mass action defines a strictly monotonic, continuous, and conjunctive kinetics function, we can represent any kinetics function by mass action in our abstract interpretation. In this way, we obtain complete independence from any kinetics information, except the three rather general properties listed above.

Corollary 1. Every strictly monotonic, continuous, and conjunctive, n-ary kinetics function can be abstracted as abstract mass action kinetics ma_c^{α} of reactions with order n, with any c.

Proof. By Lemma 2 and Proposition 2.

6 Constraint Solving

In the following, we discuss how to search for solutions of abstract constraints by constraint programming methods.

Solutions. The number of solutions would become huge if one tries to enumerate the values of all variables of the constraints $\llbracket \phi_R \rrbracket \land \llbracket O \rrbracket$. Indeed, we are only interested in knockout strategies, that is in the values of the variables x_{on_r} . All other variables define internal fluxes, so that they can be considered as existentially quantified. That is, for every choice of values for the variables $(x_{on_r})_{r \in R}$ we will only verify whether there exists one possible choice for the values of the other variables.

Optimal Solutions. Still, the number of solutions might be large since arbitrary subsets of reactions may be knocked out. The usual method to deal with this problem is to impose a quality measure on solutions and to search only for high quality solutions. First of all, the fewer reactions are knocked out the better, since knockouts in the wet lab impose high costs. Second, the fewer impact the modifications have on the input-output environment of the network, the better. Which reactions are to be considered as inputs and outputs is to be specified (and is usually evident in the applications). The speed of such reactions should change only if required by the objectives, but as few as possible otherwise.

Constraint Propagation. We apply the usual strategy of constraint programming to first propagate exhaustively and then distribute. We use the usual constraint propagation rules for table constraints. Let p be either of the relations $+^{\alpha}$ and \cdot^{α} and assume that we have a constraint $p(x_1, x_2, x_3)$. For all variables x_i we maintain a finite domain $\Delta_i \subseteq \Delta$ of possible values. We can then reduce the domain of variables x_j as follows:

$$\frac{j \in \{1, 2, 3\}}{x_j \in \{\delta_j \mid (\delta_1, \delta_2, \delta_3) \in p, \ \forall i \neq j. \ \delta_i \in \Delta_i\}}$$

Branch and Bound. During constraint solving we can always maintain a lower bound for the quality of the current presolution. As usual with branch and bound, we only search for solutions that are better or equally well as any solution found previously.

7 Leucine Overproduction: A Case Study

In this section we apply our approach to predict knockout strategies for the overproduction of Leucine in *B. subtilis* based on the model given by [17].

The reaction network for Leucine production in *B. subtilis* following [17] is given in Figure 7. Molecules types are notated by ovals and reactions by boxes, respectively. A reaction's products are denoted by outgoing arrows, reactants by continuous, and enzymes by dashed lines. For improved readability, some molecule types occur more than once in the picture, once in the role of a product of reactions (solid oval) and else in the role of an enzyme (dashed oval). Reactions marked by red boxes represent reactions that can be knocked out. We consider all reactions without reactants or without products to be input or output reactions, respectively. Our objective is to increase the rate of the Leucine secreting reaction 24. The reaction network in textual form and a legend for abbreviated species names is provided in Appendix C.

In a first experiment, the goal was to find knockout strategies that are optimized w.r.t. a minimal number of knockouts. Based on our implementation, we could solve this task in about 5s on a Dell Latitude E6320 machine (Intel Core i7-2640M CPU, 2.8 GHz, 8 GB of memory).

Several single knockout solutions were proposed. Explanations for three of them are graphically annotated in Figure 7 by orange, brown, and green boxed

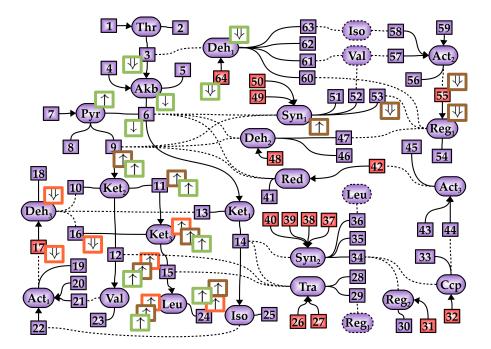


Fig. 7. A reaction network of Leucine production in *B. subtilis* following [17] in graphical form. Ovals graphically denote molecule types, rectangles reactions, arrows products and continuous and dashed lines reactants and enzymes, respectively. For improved readability, some molecule types occur more than once in the picture, once in the role of a product of reactions (solid oval) and else in the role of an enzyme (dashed oval). Red boxes mark reactions that may be knocked out. The objective is to increase the rate of reaction 24. Arrows in boxes of different colors represent different knockout strategies.

arrows, respectively. The first one is to knock out reaction 17 (orange). This leads to the removal of species Deh₃ and therefore to a stop of reaction 16, i.e. the secretion of species Ket₃. In this way the rate of reaction 15 increases and so do the concentration of species Leu and the rate of reaction 24.

Knocking out reaction 55 (brown) leads to a total lack of $\mathsf{Reg_1}$. Since this is involved in the consumption of $\mathsf{Syn_1}$ (reaction 53), the concentration of the latter increases. Consequently, reaction 9 is accelerated, such that more $\mathsf{Ket_2}$ is produced and thus also more $\mathsf{Ket_3}$ and Leu . This knockout strategy confirms what is presented in literature (cf. [4], knockout of gene codY).

The deletion of reaction 64 (green) also leads to an acceleration of reaction 9. When disabling reaction 64 no more Deh_1 is available, such that no more Thr is transformed to Akb (reaction 3). Thus, the concentration of Akb is decreased, such that reaction 6 is slowed down. Therefore, the concentration of Pyr is increased, augmenting the rate of reaction 9.

We performed a second experiment optimizing additionally w.r.t. the number of side effects. The computation took about 163s on the same machine. The orange knockout strategy and a combination of the green and the brown strategy turned out to be optimal. The latter is currently tested in wet-lab experiments.

8 Related Work

Most existing approaches consider the prediction of knockout strategies to be a two-level optimization problem [5, 25, 26, 24, 34, 21]. Thereby, the first level of optimization bases on the idea of flux balance analysis [37, 27, 3, 7]. That is, it captures only the equations stating that for any species the sum of the rates of the producing and the consuming reactions equal (Figure 1, rule (SPEC)). In this way, the problem of missing kinetics information is circumvented but at the price of losing all dependencies between species concentrations and reaction rates. Instead, determining the values of reaction rates is considered to be an optimization problem. Therefore, different kinds of optimization goals are applied, e.g. biomass production (optimal growth) [36,5] or ATP production (optimal energy) [28]. The intuition behind this first level of optimization is that, considering the background of evolution, organisms are assumed to be trimmed in a way, such that they always regulate their metabolism for optimal chances of survival. The second level of optimization is then concerned with finding the gene knockout strategy that yields the highest frequency value for the given target reaction, as determined by the first optimization level. Such two-level optimization problems are then solved by using e.g. integer linear programming approaches [5, 21] or evolutionary algorithms [24].

We see several drawbacks of applying two-level optimization approaches to predict gene knockout strategies. On one hand, since dependencies between concentrations and reaction rates are not considered, the determination of the effects of knockouts is strongly limited. In particular, negative feed-back loops that are a common theme in reaction networks cannot be taken into account.

On the other hand, whether the assumptions used to define optimization goals are appropriate is controversial [33, 29]. A major problem commonly listed is that artificially created organisms did in fact not face evolutionary pressure, such that they may control their metabolism in unexpected ways. To this end, on one hand, a reasoning is presented in [29] that is based on the maximal and minimal bounds of reaction rates. These are then obtained by a separate optimization procedure for each reaction. On the other hand, in [33], it is proposed to use the assumption of minimization of metabolic adjustment. This approach is similar to ours in that it compares reaction rates values before and after modification. The reaction rates before modifications are obtained by applying the optimal growth assumption. The rate values resulting from modification are then optimized to diverge as few as possible from their original value.

Finally, by using optimization approaches, it is not possible to apply a local reasoning that considers parts of metabolic networks as in Section 7, where only the production branched-chain amino acids (Leucine, Valine, Isoleucine) is

modeled. The reason is that optimization subjects apply to specific parts of metabolisms that thus always need to be considered, such as the Glycolysis pathway or the TCA cycle. This requires a reasoning on rather large models. Different models that aim to capture the entire metabolism of micro-organisms have been proposed so far [18, 23, 14]. However, this approach may favour predictions which apply to different parts of a metabolism and impact the reaction network in a more global manner with unwanted side effects [13]. Furthermore, a more local reasoning is favorable because resulting predictions are easier to trace (cf. explanations given in Figure 7 by arrows in colored boxed).

9 Conclusion and Future Work

We have introduced an approach for predicting knockout strategies in reaction networks with partial kinetic information, based on abstract interpretation and constraint solving. We showed that our approach is independent of any particular choice of kinetics functions, as long as these are continuous, strictly monotonic and conjunctive. Our predictions remain candidates due to the overapproximating nature of our abstraction.

A major subject for future work is to find ways to reduce the number of false solutions. On one hand, we plan to integrate methods to simplify systems of equation, linear and non-linear, as they result from our knockout semantics. On the other hand, we would like to come up with less aggressive abstract interpretations, so that one can predict weights of knockout effects. To this end, sources for more detailed kinetic information shall be developed, e.g. gene expression or flux data [32].

As a further subject, we also plan to integrate new optimization criteria for solutions. For example, one could consider, instead of only one, sets of solutions of the constraint satisfaction problem that correspond to the same knockout strategy. As each solution possibly represents a pair of steady states that are reached before and after a knockout is applied, it would make sense to favor, e.g., those knockout strategies that provide an optimal ratio between the numbers of solutions that fulfill an objective and those that do not.

We hope that the provided methods will help us to obtain better knockout results for wet-lab engineering. Concrete case studies are on the way. We also hope that better prediction methods will increase the interest in improving the quality of existing reactions networks in synthetic biology.

References

- 1. Ernesto Andrianantoandro, Subhayu Basu, David K. Karig, and Ron Weiss. Synthetic biology: new engineering rules for an emerging discipline. *Molecular systems biology*, 2(1):msb4100073–E1–msb4100073–E14, May 2006.
- 2. Steven A. Benner and A. Michael Sismour. Synthetic biology. *Nature Reviews Genetics*, 6(7):533–543, July 2005.

- 3. H. P. J. Bonarius, G. Schmid, and J. Tramper. Flux analysis of underdetermined metabolic networks: the quest for the missing constraints. *Trends in Biotechnology*, 15(8):308–314, August 1997.
- Shaun R. Brinsmade, Roelco J. Kleijn, Uwe Sauer, and Abraham L. Sonenshein. Regulation of CodY Activity through Modulation of Intracellular Branched-Chain Amino Acid Pools. J. Bacteriol., 192(24):6357–6368, December 2010.
- 5. Anthony P. Burgard, Priti Pharkya, and Costas D. Maranas. Optknock: a bilevel programming framework for identifying gene knockout strategies for microbial strain optimization. *Biotechnology and Bioengineering*, 84(6):647–657, 2003.
- Patrick Cousot and Radhia Cousot. Systematic design of program analysis frameworks. In POPL, pages 269–282, 1979.
- M. W. Covert, C. H. Schilling, and B. Palsson. Regulation of gene expression in flux balance models of metabolism. *Journal of theoretical biology*, 213(1):73–88, November 2001.
- 8. Vincent Danos, Jérôme Feret, Walter Fontana, Russell Harmer, and Jean Krivine. Abstracting the differential semantics of rule-based models: Exact and automated model reduction. In *LICS*, pages 362–381. IEEE Computer Society, 2010.
- 9. Michael B. Elowitz and Stanislas Leibler. A synthetic oscillatory network of transcriptional regulators. *Nature*, 403(6767):335–338, January 2000.
- 10. François Fages and Sylvain Soliman. Abstract interpretation and types for systems biology. *Theor. Comput. Sci.*, 403(1):52–70, 2008.
- 11. Jérôme Feret, Thomas Henzinger, Heinz Koeppl, and Tatjana Petrov. Lumpability abstractions of rule-based systems. *Theoretical Computer Science*, 2012.
- 12. James E. Ferrell. Feedback regulation of opposing enzymes generates robust, allor-none bistable responses. *Current biology: CB*, 18(6), March 2008.
- 13. Lope Florez, Katrin Gunka, Rafael Polania, Stefan Tholen, and Jorg Stulke. SPAB-BATS: A pathway-discovery method based on Boolean satisfiability that facilitates the characterization of suppressor mutants. *BMC Systems Biology*, 5(1):5+, 2011.
- 14. Jochen Förster, Iman Famili, Patrick Fu, Bernhard Ø. Palsson, and Jens Nielsen. Genome-scale reconstruction of the Saccharomyces cerevisiae metabolic network. *Genome research*, 13(2):244–253, February 2003.
- C. S. Gillespie. Moment-closure approximations for mass-action models. IET systems biology, 3(1):52–58, January 2009.
- Daniel T. Gillespie. Exact stochastic simulation of coupled chemical reactions. *Journal of Physical Chemistry*, 81:2340–2361, 1977.
- 17. Anne Goelzer, Fadia B. Brikci, Isabelle M. Verstraete, Philippe Noirot, Philippe Bessieres, Stephane Aymerich, and Vincent Fromion. Reconstruction and analysis of the genetic and metabolic regulatory networks of the central metabolism of Bacillus subtilis. *BMC Systems Biology*, 2(1):20+, 2008.
- 18. Christopher S. Henry, Jenifer F. Zinner, Matthew P. Cohoon, and Rick L. Stevens. iBsu1103: a new genome-scale metabolic model of Bacillus subtilis based on SEED annotations. *Genome biology*, 10(6):R69+, 2009.
- 19. Jay D. Keasling. Synthetic biology for synthetic chemistry. ACS chemical biology, 3(1):64-76, January 2008.
- Boris N. Kholodenko. Cell-signalling dynamics in time and space. Nature Reviews Molecular Cell Biology, 7:165–176, 2006.
- 21. Joonhoon Kim and Jennifer Reed. OptORF: Optimal metabolic and regulatory perturbations for metabolic engineering of microbial strains. *BMC Systems Biology*, 4(1):53+, 2010.

- 22. Tie Koide, Wyming Lee L. Pang, and Nitin S. Baliga. The role of predictive modelling in rationally re-engineering biological systems. *Nature reviews. Microbiology*, 7(4):297–305, April 2009.
- You-Kwan Oh, Bernhard O. Palsson, Sung M. Park, Christophe H. Schilling, and Radhakrishnan Mahadevan. Genome-scale Reconstruction of Metabolic Network in Bacillus subtilis Based on High-throughput Phenotyping and Gene Essentiality Data. *Journal of Biological Chemistry*, 282(39):28791–28799, September 2007.
- Kiran Raosaheb R. Patil, Isabel Rocha, Jochen Förster, and Jens Nielsen. Evolutionary programming as a platform for in silico metabolic engineering. BMC bioinformatics, 6(1):308+, 2005.
- Priti Pharkya, Anthony P. Burgard, and Costas D. Maranas. OptStrain: A computational framework for redesign of microbial production systems. Genome Research, 14(11):2367–2376, November 2004.
- Priti Pharkya and Costas D. Maranas. An optimization framework for identifying reaction activation/inhibition or elimination candidates for overproduction in microbial systems. *Metabolic engineering*, 8(1):1–13, January 2006.
- 27. Nathan D. Price, Jennifer L. Reed, and Bernhard Ø. Palsson. Genome-scale models of microbial cells: evaluating the consequences of constraints. *Nature reviews. Microbiology*, 2(11):886–897, November 2004.
- 28. R. Ramakrishna, J. S. Edwards, A. McCulloch, and B. O. Palsson. Flux-balance analysis of mitochondrial energy metabolism: consequences of systemic stoichiometric constraints. *American journal of physiology. Regulatory, integrative and comparative physiology*, 280(3):R695–704, March 2001.
- 29. Sridhar Ranganathan, Patrick F. Suthers, and Costas D. Maranas. OptForce: An Optimization Procedure for Identifying All Genetic Manipulations Leading to Targeted Overproductions. *PLoS Comput Biol*, 6(4):e1000744+, April 2010.
- 30. Guillermo Rodrigo, Javier Carrera, Thomas E. Landrain, and Alfonso Jaramillo. Perspectives on the automatic design of regulatory systems for synthetic biology. *FEBS Letters*, 586(15):2037–2042, July 2012.
- 31. F. Rossi, P. van Beek, and T. Walsh. *Handbook of Constraint Programming*. Elsevier, 2006.
- 32. Uwe Sauer. Metabolic networks in motion: 13C-based flux analysis. *Molecular systems biology*, 2(1), November 2006.
- 33. Daniel Segrè, Dennis Vitkup, and George M. Church. Analysis of optimality in natural and perturbed metabolic networks. *Proceedings of the National Academy of Sciences*, 99(23):15112–15117, November 2002.
- 34. Naama Tepper and Tomer Shlomi. Predicting metabolic engineering knockout strategies for chemical production: accounting for competing pathways. *Bioinformatics*, 26(4):536–543, February 2010.
- 35. R. Thomas. Boolean formalization of genetic control circuits. *Journal of theoretical biology*, 42(3):563–585, December 1973.
- 36. Amit Varma and Bernhard O. Palsson. Metabolic Capabilities of Escherichia coli II. Optimal Growth Patterns. *Journal of Theoretical Biology*, 165(4):503–522, December 1993.
- Amit Varma and Bernhard O. Palsson. Metabolic Flux Balancing: Basic Concepts,
 Scientific and Practical Use. Nature Biotechnology, 12(10):994–998, October 1994.
- 38. Olaf Wolkenhauer, Mukhtar Ullah, Walter Kolch, and Kwang-Hyun H. Cho. Modeling and simulation of intracellular dynamics: choosing an appropriate framework. *IEEE transactions on nanobioscience*, 3(3):200–207, September 2004.

A Mass action kinetics

In the following, we explain the derivation of Mass action kinetics in more detail, following the description in [16].

Intuitively, a chemical solution is a liquid in which molecules of different types are resolved. Thereby, the usual assumption is that the volume of each of these molecules is infinitesimally small, so that the volume of the chemical solution depends only on the volume of the liquid. Furthermore, the amount of the liquid is considered to remain unchanged over time, as well as its temperature and pressure. Therefore, the overall volume of the solution remains constant over time, independently of applications of any chemical reaction.

A further assumption is that chemical solutions are well stirred. In this way, the probability that two molecules meet at a given time point does not depend on their positions. In combination with the assumptions that molecules have no volume and that the volume of the solution is constant, this yields that the probability that two molecules meet at a given time point is constant, too.

Consider now a reaction with two reactants m_1 , m_2 and a solution S. As we argued above, the probability that some molecule of type m_1 meet another of type m_2 in S is constant over time and independent of the positions of the molecules. The rate with which they meet is thus proportional to their concentrations, i.e. equal to $c \cdot S_{m_1} \cdot S_{m_2}$ for some constant $c \in \mathbb{R}_{>0}$.

B Proof of Theorem

For our proof we introduce vector notation: $\mathbf{x} = (x_1, \dots, x_n)$. Thereby, the i-th vector element $\mathbf{x}[i]$ with $\mathbf{x} = (x_1, \dots, x_n)$ and $1 \le i \le n$ denotes value x_i . Substitution $\mathbf{x}[x/i]$ with $\mathbf{x} = (x_1, \dots, x_n)$ and $1 \le i \le n$ provides the vector $\mathbf{x} = (x_1, \dots, x_{i-1}, x, x_{i+1}, \dots, x_n)$. We introduce relations between vectors $\mathbf{x} \mathbf{p} \mathbf{x}'$, with $\mathbf{x} = (x_1, \dots, x_n), \mathbf{x}' = (x'_1, \dots, x'_n), \mathbf{p} = (p_1, \dots, p_n)$, and p_i as $x_i p_i x'_i$ for all i. We write $\kappa^{\alpha} \boldsymbol{\delta}$ instead of $\boldsymbol{\delta} \in \kappa^{\alpha}$, so that by definition of our abstract interpretation, $\kappa^{\alpha} \boldsymbol{\delta}$ iff $\exists \mathbf{y}, \mathbf{y}'$ such that $\kappa \mathbf{y}, \kappa \mathbf{y}'$ and $\mathbf{y} \delta \mathbf{y}'$, for $\boldsymbol{\delta} \in \Delta^{n+1}$ and $\mathbf{y}, \mathbf{y}' \in \mathbb{R}^{n+1}_{>0}$.

Proposition 2 Let $\kappa_1, \kappa_2 \subseteq \mathbb{R}^n_{\geq 0} \to \mathbb{R}_{\geq 0}$ be strictly monotonic, continuous, and conjunctive kinetics functions. It holds that $\kappa_1^{\alpha} = \kappa_2^{\alpha}$.

Proof. We need to show that for all $\boldsymbol{\delta} \in \Delta^{n+1}$, it holds that $\kappa_1^{\alpha} \boldsymbol{\delta}$ iff $\kappa_2^{\alpha} \boldsymbol{\delta}$. In the following, we only show \Rightarrow , since \Leftarrow is analogue. By the definition of the abstract interpretation, we need to show that for all $\boldsymbol{\delta}$, if $\kappa_1^{\alpha} \boldsymbol{\delta}$ then there exist $\boldsymbol{y}, \boldsymbol{y}' \in \mathbb{R}^{n+1}_{\geq 0}$, such that $\kappa_2 \boldsymbol{y}, \kappa_2 \boldsymbol{y}'$, and $\boldsymbol{y} \boldsymbol{\delta} \boldsymbol{y}'$. We distinguish the following cases:

- $\exists i \leq n. \boldsymbol{\delta}[i] = \approx$ Since κ_1 is conjunctive, we obtain $\boldsymbol{\delta}[n+1] = \approx$. Since κ_2 is also conjunctive, it holds for all $\boldsymbol{y}, \boldsymbol{y}'$, with $\boldsymbol{y} \boldsymbol{\delta} \boldsymbol{y}'$, that $\kappa_2 \boldsymbol{y}$ and $\kappa_2 \boldsymbol{y}'$.
- $\forall i \leq n. \boldsymbol{\delta}[i] \in \{\uparrow, \sim, \downarrow, \downarrow\}, \exists i \leq n. \boldsymbol{\delta}[i] = \downarrow$ Since κ_1 is conjunctive, we obtain $\boldsymbol{\delta}[n+1] = \downarrow$. Since κ_2 is also conjunctive it holds that there exists \boldsymbol{y} , such that for all \boldsymbol{y}' , with $\boldsymbol{y} \boldsymbol{\delta} \boldsymbol{y}'$, it is true that $\kappa_2 \boldsymbol{y}$ and $\kappa_2 \boldsymbol{y}'$.

- $\forall i \leq n. \boldsymbol{\delta}[i] \in \{\uparrow, \sim, \downarrow, \uparrow\}, \exists i \leq n. \boldsymbol{\delta}[i] = \uparrow$ Since κ_1 is conjunctive, we obtain $\boldsymbol{\delta}[n+1] = \uparrow$. Since κ_2 is also conjunctive it holds that there exists \boldsymbol{y}' , such that for all \boldsymbol{y} , with $\boldsymbol{y} \boldsymbol{\delta} \boldsymbol{y}'$, it is true that $\kappa_2 \boldsymbol{y}$ and $\kappa_2 \boldsymbol{y}'$.
- $\forall i \leq n. \boldsymbol{\delta}[i] \in \{\uparrow, \sim, \downarrow, \uparrow, \downarrow\}, \exists i, i' \leq n. \boldsymbol{\delta}[i] = \uparrow, \delta[i]' = \downarrow$ Since κ_1 is conjunctive, we obtain $\boldsymbol{\delta}[n+1] = \approx$. Since κ_2 is also conjunctive, it holds for all $\boldsymbol{y}, \boldsymbol{y}'$, with $\boldsymbol{y} \boldsymbol{\delta} \boldsymbol{y}'$, that $\kappa_2 \boldsymbol{y}$ and $\kappa_2 \boldsymbol{y}'$.
- $\forall i \leq n. \boldsymbol{\delta}[i] = \sim \text{ We obtain } \boldsymbol{\delta}[n+1] = \sim. \text{ It holds for all } \boldsymbol{y}, \text{ with } \kappa_2 \boldsymbol{y}, \text{ that } \boldsymbol{y} \boldsymbol{\delta} \boldsymbol{y}.$ $\forall i \leq n. \boldsymbol{\delta}[i] \in \{\sim, \downarrow\}, \exists i \leq n. \boldsymbol{\delta}[i] = \downarrow \text{ Since } \kappa_1 \text{ is strictly monotonic and conjunctive } \kappa_1^{\alpha} \boldsymbol{\delta} \text{ implies that } \boldsymbol{\delta}[n+1] = \downarrow. \text{ Since } \kappa_2 \text{ is also strictly monotonic and conjunctive it holds that there exist } \boldsymbol{y}, \boldsymbol{y}', \text{ with } \boldsymbol{y} \boldsymbol{\delta} \boldsymbol{y}', \text{ such that } \kappa_2 \boldsymbol{y} \text{ and } \kappa_2 \boldsymbol{y}'.$
- $\forall i \leq n. \boldsymbol{\delta}[i] \in \{\sim, \uparrow\}, \exists i \leq n. \boldsymbol{\delta}[i] = \uparrow \text{ Since } \kappa_1 \text{ is strictly monotonic and conjunctive } \kappa_1^{\alpha} \boldsymbol{\delta} \text{ implies that } \boldsymbol{\delta}[n+1] = \uparrow. \text{ Since } \kappa_2 \text{ is also strictly monotonic and conjunctive it holds that there exist } \boldsymbol{y}, \boldsymbol{y}', \text{ with } \boldsymbol{y} \boldsymbol{\delta} \boldsymbol{y}', \text{ such that } \kappa_2 \boldsymbol{y} \text{ and } \kappa_2 \boldsymbol{y}'.$
- $\forall i \leq n. \boldsymbol{\delta}[i] \in \{\uparrow, \sim, \downarrow\}, \exists i, i' \leq n. \boldsymbol{\delta}[i] = \uparrow, \delta[i]' = \downarrow \text{ Since } \kappa_1 \text{ is conjunctive, we obtain } \boldsymbol{\delta}[n+1] \in \{\uparrow, \downarrow, \sim\}. \text{ Let } \boldsymbol{y}, \boldsymbol{y}', \text{ such that } \kappa_2 \boldsymbol{y}, \kappa_2 \boldsymbol{y}', \text{ and for all } i \leq n \text{ it holds that } \boldsymbol{y}[i] \boldsymbol{\delta}[i] \boldsymbol{y}'[i]. \text{ We distinguish the following cases:}$
 - $\delta[n+1] = \downarrow$ If $\mathbf{y}[n+1] > \mathbf{y}'[n+1]$ it holds that $\mathbf{y}\delta\mathbf{y}'$. If $\mathbf{y}[n+1] \leq \mathbf{y}'[n+1]$, let $j \leq n$, such that $\delta[j] = \downarrow$, w.l.o.g, and thus $\mathbf{y}'[j] < \mathbf{y}[j]$. Since κ_2 is continuous and conjunctive there exist $y < \mathbf{y}'[j]$ and $y' < \mathbf{y}[n+1]$, such that $\kappa_2 \mathbf{y}'[y/j][y'/n+1]$. It holds that $\mathbf{y}\delta\mathbf{y}'[y/j][y'/n+1]$.
 - $\delta[n+1] = \uparrow$ If $\mathbf{y}'[n+1] > \mathbf{y}[n+1]$ it holds that $\mathbf{y}\delta\mathbf{y}'$. If $\mathbf{y}'[n+1] \leq \mathbf{y}[n+1]$, let $j \leq n$, such that $\mathbf{y}[j] = \uparrow$, w.l.o.g, and thus $\mathbf{y}[j] < \mathbf{y}'[j]$. Since κ_2 is continuous and conjunctive there exist $y < \mathbf{y}[j]$ and $y' < \mathbf{y}'[n+1]$, such that $\kappa_2 \mathbf{y}[y/j][y'/n+1]$. It holds that $\mathbf{y}[y/j][y'/n+1]\delta\mathbf{y}'$.
 - $\boldsymbol{\delta}[n+1] = \sim \text{ If } \boldsymbol{y}[n+1] = \boldsymbol{y}'[n+1] \text{ it holds that } \boldsymbol{y}\boldsymbol{\delta}\boldsymbol{y}'. \text{ The cases that } \boldsymbol{y}[n+1] > \boldsymbol{y}'[n+1] \text{ and } \boldsymbol{y}[n+1] < \boldsymbol{y}'[n+1] \text{ are analogue to above.}$

C Supplementary Information Example

```
Val: \longrightarrow Act_1
                                                                                                                                                        \longrightarrow Act<sub>3</sub>
  (1)

ightarrow Thr
                                                              (21)
                                                                                                                     (43)
  (2)
                              Thr \longrightarrow
                                                              (22)
                                                                                     Iso : \longrightarrow Act<sub>1</sub>
                                                                                                                                            Ccp : \longrightarrow Act_3
                                                                                                                     (44)
  (3)
                                                                                      \mathsf{Val} \longrightarrow
                                                              (23)
                \mathsf{Deh}_1 : \mathsf{Thr} \longrightarrow \mathsf{Akb}
                                                                                                                     (45)
                                                                                                                                             Act_3 \longrightarrow
                                                                                      \mathsf{Leu} \longrightarrow
  (4)
                                         \rightarrow \mathsf{Akb}
                                                              (24)
                                                                                                                     (46)
                                                                                                                                            \mathsf{Deh}_2 \longrightarrow
   (5)
                              \mathsf{Akb} \longrightarrow
                                                              (25)
                                                                                       Iso \longrightarrow
                                                                                                                     (47) \operatorname{Reg}_1 : \operatorname{Deh}_2 \longrightarrow
  (6)
               Syn_1, Deh_2,
                                                              (26)
                                                                                               \longrightarrow \mathsf{Tra}
                                                                                                                                                       \longrightarrow \mathsf{Deh}_2
                                                                                                                     (48)
          \mathsf{Red} : \mathsf{Akb}, \mathsf{Pyr} \longrightarrow \mathsf{Ket}_1
                                                              (27)
                                                                                               \longrightarrow Tra
                                                                                                                     (49)
                                                                                                                                                       \longrightarrow \mathsf{Syn}_1
                                                                                      \mathsf{Tra} \longrightarrow
  (7)
                                       \longrightarrow \mathsf{Pyr}
                                                              (28)
                                                                                                                     (50)
                                                                                                                                                       \longrightarrow \mathsf{Syn}_1
   (8)
                              \mathsf{Pyr} \longrightarrow
                                                                                                                                             \mathsf{Syn}_1 \longrightarrow
                                                              (29)
                                                                         \mathsf{Reg}_1 : \mathsf{Tra} \longrightarrow
                                                                                                                     (51)
   (9)
               \mathsf{Syn}_1, \mathsf{Deh}_2,
                                                              (30)
                                                                                    \mathsf{Reg}_2 \longrightarrow
                                                                                                                     (52)
                                                                                                                                  Val: Syn_1 \longrightarrow
                                                                                               \longrightarrow \mathsf{Reg}_2
                   \mathsf{Red} : \mathsf{Pyr} \longrightarrow \mathsf{Ket}_2
                                                                                                                     (53) \operatorname{Reg}_1 : \operatorname{Syn}_1 \longrightarrow
                                                              (31)
(10)
               \mathsf{Deh}_3 : \mathsf{Ket}_2 \longrightarrow
                                                              (32)
                                                                                              \longrightarrow \mathsf{Ccp}
                                                                                                                     (54)
                                                                                                                                            \mathsf{Reg}_1 \longrightarrow
(11)
                            \mathsf{Ket}_2 \longrightarrow \mathsf{Ket}_3
                                                              (33)
                                                                                     \mathsf{Ccp} \longrightarrow
                                                                                                                     (55)
                                                                                                                                           \mathsf{Act}_2: \longrightarrow \mathsf{Reg}_1
                  \mathsf{Tra}: \mathsf{Ket}_2 \longrightarrow \mathsf{Val}
(12)
                                                              (34) Ccp, Reg<sub>2</sub>:
                                                                                                                     (56)
                                                                                                                                             \mathsf{Act}_2 \longrightarrow
                                                                                                                                             \mathsf{Val}: \longrightarrow \mathsf{Act}_2
(13)
               \mathsf{Deh}_3 : \mathsf{Ket}_1 \longrightarrow
                                                                                                                     (57)
                                                                                    \mathsf{Syn}_2 \longrightarrow
                 \mathsf{Tra}, \mathsf{Syn}_2:
                                                                                                                                              \mathsf{Iso}: \longrightarrow \mathsf{Act}_2
                                                              (35)
                                                                                    \mathsf{Syn}_2 \longrightarrow
(14)
                                                                                                                     (58)
                                                                                                                                                       \longrightarrow \mathsf{Act}_2
                            \mathsf{Ket}_1 \longrightarrow \mathsf{Iso}
                                                              (36) Leu: Syn_2 \longrightarrow
                                                                                                                     (59)
                  \mathsf{Tra}: \mathsf{Ket}_3 \longrightarrow \mathsf{Leu}
                                                                                               \longrightarrow \mathsf{Syn}_2
(15)
                                                              (37)
                                                                                                                     (60) \operatorname{Reg}_1 : \operatorname{Deh}_1 \longrightarrow
(16)
               \mathsf{Deh}_3 : \mathsf{Ket}_3 \longrightarrow
                                                              (38)
                                                                                               \longrightarrow \mathsf{Syn}_2
                                                                                                                     (61)
                                                                                                                                 Val : Deh_1 \longrightarrow
(17)
                          \mathsf{Act}_1: \longrightarrow \mathsf{Deh}_3
                                                              (39)
                                                                                              \longrightarrow \mathsf{Syn}_2
                                                                                                                     (62)
                                                                                                                                            Deh_1 \longrightarrow
                                                                                                                                   \mathsf{Iso}:\mathsf{Deh}_1\longrightarrow
(18)
                           \mathsf{Deh}_3 \longrightarrow
                                                              (40)
                                                                                              \longrightarrow \mathsf{Syn}_2
                                                                                                                     (63)
                            \mathsf{Act}_1 \longrightarrow
                                                              (41)
                                                                                     \mathsf{Red} \longrightarrow
                                                                                                                     (64)
                                                                                                                                                        \longrightarrow \mathsf{Deh}_1
(19)
(20)
                                       \longrightarrow \mathsf{Act}_1
                                                              (42)
                                                                                 \mathsf{Act}_3: \longrightarrow \mathsf{Red}
                                                                                                            Pyruvate
              Transcriptional Activator 1
                                                                                                Pyr
              Transcriptional Activator 2
                                                                                                            ketol-acid reductoisomerase
              Transcriptional Activator 3
                                                                                                            Transcriptional Pleiotropic
  Act_3
  Akb
              L-2-amino-acetoacetate
                                                                                                            Regulator 1
                                                                                               {
m Reg}_2 Transcriptional Pleiotropic
  Сср
               Carbon Catabolite Control Protein A
  Deh<sub>1</sub> Threonine Dehydratase
                                                                                                            Regulator 2
  Deh<sub>2</sub> Dihydroxy-acid Dehydratase
                                                                                               Syn_1
                                                                                                           Acetolactate synthase
  Deh<sub>3</sub> 2-oxoisovalerate Dehydrogenase
                                                                                                Syn<sub>2</sub> 2-isopropylmalate synthase
               Isoleucine
                                                                                                            Threonine
              2-keto-3-methylvalerate
 \text{Ket}_1
                                                                                                Tra
                                                                                                            Branched-chain Amino Acid
  Ket<sub>2</sub> 2-ketoisovalerate
                                                                                                            Aminotransferase
                                                                                                            Valine
 Ket<sub>3</sub> 2-ketoisocaproate
                                                                                                Val
 Leu Leucine
```

Fig. 8. The reactions of the network in Figure 7 in textual form (top) and a legend for abbreviated species names (bottom).