

R Training Documentation

2023-10-17

Introduction

Source files, directories, important functions in R

The working directory is the folder your R session ‘lives in.’ In general, it is good practice to have your working directory to be set to the folder which houses the open file. `getwd()` displays the current working directory of the session. You can set the working directory to the path of the desired folder by using `setwd(...)`.

If ever you are confused about a package or function in R studio, typing ‘?’ in front of the argument will display the documentation for that function in your session. For information on Vectorsurv specific functions, please see the documentation below.

```
#To get current directory  
getwd()
```

```
## [1] "C:/Users/Christina/Desktop/R-tutorial"
```

```
#To get information on function in R documentation  
?getwd()  
  
#'VS_functions.R' should be in the same directory as the .rmd  
#'files which access its functions, you can set the working directory  
  
#Path to folder on my computer: "C:\Users\Christina\Desktop\R-tutorial"  
#this path will be slightly different for you depending on the file location.  
  
#Set below to the location of the downloaded files on your machine,  
#make sure to change the direction of the slash from "\" to "/" to indicate opening of the folder.  
  
setwd("/Users/Christina/Desktop/R-tutorial")
```

There is one R files. The .R file is a source file that contains all of the functions you will need to run the .RMarkdown files. You do not need to touch the source file. The .RMarkdown is a pre-coded sample report that you can customize to suit your needs.

```
#loads relevant packages and functions  
source("VS_functions.R")
```

Retriving Data

getArthroCollections(...)

Description

getArthroCollections(...) obtains collections data on a year range (start_year, end_year) and agency_id. It prompts the user for their Gateway username and password before retrieving the associated data. Agency id is the number associated with your agency through Vectorsurv. If you have access to multiple agencies, id can be used to specific what data you wish to retrieve. You can only retrieve data from agencies linked to your Gateway account.

Usage

```
getArthroCollections(start_year, end_year, agency_code)
```

Arguments

- start_year: Beginning of year range
- end_year: End of year range
- agency_id: Agency ID number, if left blank, the default will return data for all agencies linked to you account. The majority of users are linked to one agency

#Example

```
collections = getArthroCollections(2022,2023, 55)
```

getPools(...)

Description

getPools(...) similar to getArthroCollections() obtains pools on a year range (start_year, end_year) and agency_id. It prompts the user for their Gateway username and password before retrieving the associated data. getPools() retrieve data for both mosquito and tick pools.

Usage

```
getPools(start_year, end_year, agency_id)
```

Arguments

- start_year: Beginning of year range
- end_year: End of year range
- agency_id: Agency ID number

#Example

```
pools = getPools(2022,2023, 55)
```

Write Data to file

You can save retrieved data as a .csv file in your current directory using write.csv(). That same data can be retrieved using read.csv(). Writing data to a .csv can make the rendering process more efficient when generating reports in R. We recommend that you write the data pulled from our API into a csv and then load that data when generating reports.

```
read.csv(...)
```

```
#creates a file named "collections_18_23.csv" in your current directory
write.csv(x = collections, file = "collections_18_23.csv")

#loads collections data
collections = read.csv("collections_18_23.csv")
```

Basic subsetting, filtering, grouping, pivoting

Data can be subset to contain columns of interest. Sub-setting can also be used to reorder the columns in a dataframe. Do not subset collections or pools data before inputting them into Vectorsurv calculator functions to avoid losing essential columns. It is recommended to subset after calculations are complete and before inputting into a table generator. Remember, subsetting, filtering, grouping and summarizing will not change the value of the data unless it is reassigned to the same variable name. We recommend creating a new variable for processed data.

Subsetting

```
#Subset using column names or index number

colnames(collections) #displays column names and associated index
```

```
## [1] "X" "collection_id"
## [3] "collection_num" "collection_date"
## [5] "collection_date_date_only" "comments"
## [7] "identified_by" "num_trap"
## [9] "site" "surv_year"
## [11] "trap_nights" "trap_problem_bit"
## [13] "user" "add_date"
## [15] "deactive_date" "updated"
## [17] "id" "num_count"
## [19] "sex_id" "sex_type"
## [21] "sex_name" "species_id"
## [23] "species_full_name" "species_display_name"
## [25] "agency_id" "agency_code"
## [27] "agency_name" "trap_id"
## [29] "trap_acronym" "trap_name"
## [31] "trap_presence"
```

```
#Subsetting by name
head(collections[c("collection_date", "species_display_name", "num_count")])
```

```
##      collection_date species_display_name num_count
## 1 2023-10-20T07:00:00.000Z Ae melanimon      52
## 2 2023-10-20T07:00:00.000Z Cx pipiens      16
## 3 2023-10-20T07:00:00.000Z Cx tarsalis      42
## 4 2023-10-20T07:00:00.000Z Ae melanimon       4
## 5 2023-10-20T07:00:00.000Z Cx tarsalis       1
## 6 2023-10-20T07:00:00.000Z Ae melanimon      43
```

```
#by index
head(collections[c(3,23,17)])
```

```
##   collection_num species_full_name      id
## 1           8332   Aedes melanimon 6226776
## 2           8332     Culex pipiens 6226777
## 3           8332     Culex tarsalis 6226778
## 4           8333   Aedes melanimon 6226779
## 5           8333     Culex tarsalis 6226780
## 6           8334   Aedes melanimon 6226872
```

```
#to save a subset
collections_subset = collections[c(3,23,17)]
```

Filtering and subsetting in dplyr

Dplyr is a powerful package for filtering and sub-setting data. It follows logic similar to SQL queries.

For more information on data manipulation using dplyr [Click Here](#)

Dplyr utilizes the pipe operator ‘%>%’ to send data into functions.

```
#Subsetting columns with dplyr 'select'
collections %>%
  select(collection_date,species_display_name,num_count)%>%
  head()
```

```
##           collection_date species_display_name num_count
## 1 2023-10-20T07:00:00.000Z      Ae melanimon         52
## 2 2023-10-20T07:00:00.000Z       Cx pipiens          16
## 3 2023-10-20T07:00:00.000Z       Cx tarsalis          42
## 4 2023-10-20T07:00:00.000Z      Ae melanimon           4
## 5 2023-10-20T07:00:00.000Z       Cx tarsalis           1
## 6 2023-10-20T07:00:00.000Z      Ae melanimon          43
```

```
#filtering with dplyr 'filter'
collections %>%
  filter(species_display_name=="Cx pipiens") %>%
  head()
```

```
##   X collection_id collection_num      collection_date
## 1 2           3033442           8332 2023-10-20T07:00:00.000Z
## 2 9           3033489           8334 2023-10-20T07:00:00.000Z
## 3 17          3033497           8337 2023-10-20T07:00:00.000Z
## 4 19          3033498           8338 2023-10-20T07:00:00.000Z
## 5 23          3033525           8347 2023-10-20T07:00:00.000Z
## 6 24          3033610           8348 2023-10-20T07:00:00.000Z
##   collection_date_date_only
## 1                      TRUE
## 2                      TRUE
## 3                      TRUE
## 4                      TRUE
```

```

## 5 TRUE
## 6 TRUE
##
##                                     comments
## 1 Submitted using API - MVTrapID# 82801\n\t\t\t\tRandomNumber - 788422245
## 2 Submitted using API - MVTrapID# 82796\n\t\t\t\tRandomNumber - 811478541
## 3 Submitted using API - MVTrapID# 82799\n\t\t\t\tRandomNumber - 643517194
## 4 Submitted using API - MVTrapID# 82804\n\t\t\t\tRandomNumber - 582396875
## 5 Submitted using API - MVTrapID# 82811\n\t\t\t\tRandomNumber - 181335976
## 6 Submitted using API - MVTrapID# 82813\n\t\t\t\tRandomNumber - 685144888
##   identified_by num_trap   site surv_year trap_nights trap_problem_bit user
## 1 Anna;Cutshall      1  13000     2023         1         FALSE 331
## 2 Anna;Cutshall      1  13007     2023         1         FALSE 331
## 3 Anna;Cutshall      1  13010     2023         1         FALSE 331
## 4 Anna;Cutshall      1  17554     2023         1         FALSE 331
## 5 James;Brodigan     1 125844     2023         1         FALSE 331
## 6 James;Brodigan     1 125846     2023         1         FALSE 331
##   add_date deactivate_date      updated      id
## 1 2023-10-20T18:47:24.194Z      NA 2023-10-20T18:47:25.753Z 6226777
## 2 2023-10-20T19:09:08.137Z      NA 2023-10-20T19:09:09.287Z 6226874
## 3 2023-10-20T19:15:44.615Z      NA 2023-10-20T19:15:45.726Z 6226895
## 4 2023-10-20T19:15:47.298Z      NA 2023-10-20T19:15:48.502Z 6226899
## 5 2023-10-20T19:33:20.393Z      NA 2023-10-20T19:33:21.363Z 6226953
## 6 2023-10-20T19:35:26.652Z      NA 2023-10-20T19:35:27.644Z 6227157
##   num_count sex_id sex_type      sex_name species_id species_full_name
## 1      16     4   female Females - Mixed      65      Culex pipiens
## 2      25     4   female Females - Mixed      65      Culex pipiens
## 3      21     4   female Females - Mixed      65      Culex pipiens
## 4       2     4   female Females - Mixed      65      Culex pipiens
## 5       1     4   female Females - Mixed      65      Culex pipiens
## 6       1     4   female Females - Mixed      65      Culex pipiens
##   species_display_name agency_id agency_code      agency_name trap_id
## 1      Cx pipiens      55      SAYO Sacramento-Yolo MVCD      2
## 2      Cx pipiens      55      SAYO Sacramento-Yolo MVCD      2
## 3      Cx pipiens      55      SAYO Sacramento-Yolo MVCD      2
## 4      Cx pipiens      55      SAYO Sacramento-Yolo MVCD      3
## 5      Cx pipiens      55      SAYO Sacramento-Yolo MVCD     13
## 6      Cx pipiens      55      SAYO Sacramento-Yolo MVCD     13
##   trap_acronym      trap_name trap_presence
## 1      CO2 Carbon dioxide baited trap      FALSE
## 2      CO2 Carbon dioxide baited trap      FALSE
## 3      CO2 Carbon dioxide baited trap      FALSE
## 4      GRVD      Gravid trap      FALSE
## 5      BGSENT      BG Sentinel      FALSE
## 6      BGSENT      BG Sentinel      FALSE

```

```

#filtering multiple arguments using '%in%'
collections %>%
  filter(species_display_name %in% c("Cx pipiens", "Cx tarsalis"),
         num_trap %in% c(1:5))%>%
  head()

```

```

##   X collection_id collection_num      collection_date
## 1 2      3033442      8332 2023-10-20T07:00:00.000Z
## 2 3      3033442      8332 2023-10-20T07:00:00.000Z

```

```

## 3 5      3033443      8333 2023-10-20T07:00:00.000Z
## 4 9      3033489      8334 2023-10-20T07:00:00.000Z
## 5 10     3033489      8334 2023-10-20T07:00:00.000Z
## 6 17     3033497      8337 2023-10-20T07:00:00.000Z
## collection_date_date_only
## 1      TRUE
## 2      TRUE
## 3      TRUE
## 4      TRUE
## 5      TRUE
## 6      TRUE
##
##                                     comments
## 1 Submitted using API - MVTrapID# 82801\n\t\t\t\tRandomNumber - 788422245
## 2 Submitted using API - MVTrapID# 82801\n\t\t\t\tRandomNumber - 788422245
## 3 Submitted using API - MVTrapID# 82802\n\t\t\t\tRandomNumber - 456498256
## 4 Submitted using API - MVTrapID# 82796\n\t\t\t\tRandomNumber - 811478541
## 5 Submitted using API - MVTrapID# 82796\n\t\t\t\tRandomNumber - 811478541
## 6 Submitted using API - MVTrapID# 82799\n\t\t\t\tRandomNumber - 643517194
## identified_by num_trap site surv_year trap_nights trap_problem_bit user
## 1 Anna;Cutshall      1 13000      2023      1      FALSE 331
## 2 Anna;Cutshall      1 13000      2023      1      FALSE 331
## 3 Anna;Cutshall      1 13000      2023      1      FALSE 331
## 4 Anna;Cutshall      1 13007      2023      1      FALSE 331
## 5 Anna;Cutshall      1 13007      2023      1      FALSE 331
## 6 Anna;Cutshall      1 13010      2023      1      FALSE 331
##
## add_date deactivate_date updated id
## 1 2023-10-20T18:47:24.194Z      NA 2023-10-20T18:47:25.753Z 6226777
## 2 2023-10-20T18:47:24.194Z      NA 2023-10-20T18:47:25.753Z 6226778
## 3 2023-10-20T18:47:26.712Z      NA 2023-10-20T18:47:27.486Z 6226780
## 4 2023-10-20T19:09:08.137Z      NA 2023-10-20T19:09:09.287Z 6226874
## 5 2023-10-20T19:09:08.137Z      NA 2023-10-20T19:09:09.287Z 6226875
## 6 2023-10-20T19:15:44.615Z      NA 2023-10-20T19:15:45.726Z 6226895
## num_count sex_id sex_type sex_name species_id species_full_name
## 1      16      4 female Females - Mixed      65 Culex pipiens
## 2      42      4 female Females - Mixed      70 Culex tarsalis
## 3       1      3 male Males      70 Culex tarsalis
## 4      25      4 female Females - Mixed      65 Culex pipiens
## 5      35      4 female Females - Mixed      70 Culex tarsalis
## 6      21      4 female Females - Mixed      65 Culex pipiens
## species_display_name agency_id agency_code agency_name trap_id
## 1 Cx pipiens      55 SAYO Sacramento-Yolo MVCD      2
## 2 Cx tarsalis      55 SAYO Sacramento-Yolo MVCD      2
## 3 Cx tarsalis      55 SAYO Sacramento-Yolo MVCD      3
## 4 Cx pipiens      55 SAYO Sacramento-Yolo MVCD      2
## 5 Cx tarsalis      55 SAYO Sacramento-Yolo MVCD      2
## 6 Cx pipiens      55 SAYO Sacramento-Yolo MVCD      2
## trap_acronym trap_name trap_presence
## 1 CO2 Carbon dioxide baited trap FALSE
## 2 CO2 Carbon dioxide baited trap FALSE
## 3 GRVD Gravid trap FALSE
## 4 CO2 Carbon dioxide baited trap FALSE
## 5 CO2 Carbon dioxide baited trap FALSE
## 6 CO2 Carbon dioxide baited trap FALSE

```

Group by

In addition to filtering and sub-setting, data can be group by variables and summarized.

#groups by species and collection date and sums the number counted

```
collections %>%
  group_by(collection_date, species_display_name) %>%
  summarise(sum_count = sum(num_count, na.rm=T))%>%
  head()
```

```
## # A tibble: 6 x 3
## # Groups:   collection_date [1]
##   collection_date      species_display_name sum_count
##   <chr>                <chr>                <int>
## 1 2022-01-04T08:00:00.000Z An freeborni          1
## 2 2022-01-04T08:00:00.000Z Cs incidens           2
## 3 2022-01-04T08:00:00.000Z Cs inornata           2
## 4 2022-01-04T08:00:00.000Z Cx pipiens           22
## 5 2022-01-04T08:00:00.000Z Cx stigmatosoma       2
## 6 2022-01-04T08:00:00.000Z Cx tarsalis           1
```

#groups by species and collection date and takes the average the number counted

```
collections %>%
  group_by(collection_date, species_display_name) %>%
  summarise(sum_count = mean(num_count, na.rm=T))%>%
  head()
```

```
## # A tibble: 6 x 3
## # Groups:   collection_date [1]
##   collection_date      species_display_name sum_count
##   <chr>                <chr>                <dbl>
## 1 2022-01-04T08:00:00.000Z An freeborni          1
## 2 2022-01-04T08:00:00.000Z Cs incidens           1
## 3 2022-01-04T08:00:00.000Z Cs inornata           1
## 4 2022-01-04T08:00:00.000Z Cx pipiens           2.44
## 5 2022-01-04T08:00:00.000Z Cx stigmatosoma       1
## 6 2022-01-04T08:00:00.000Z Cx tarsalis           1
```

Pivoting

Data can be manipulated into long and wide (spread sheet) forms using `pivot_wider` and `pivot_longer`. By default data from the API is in long form. Here we pivot on species and sex condition names using `num_count` as values. The end result is data with `num_count` values in the columns named `species_sex`. For more on pivoting see `??pivot_longer` and `??pivot_wider`

```
#l
collections_wide = pivot_wider(collections,
                              names_from = c("species_display_name", "sex_name"),
                              values_from = "num_count")
```

Calculations

Abundance

getAbundance(...)

Description

getAbundance(...) uses any amount of arthro collections data to calculate the abundance for the specified parameters. The function calculates using the methods of the Gateway Abundance calculator.

Usage

```
getAbundance(collections,interval, species_list = NULL, trap_list = NULL, species_separate = FALSE)
```

Arguments

- collections: Collections data retrieved from getArthroCollections(...)
- interval: Calculation interval for abundance, accepts “collection_date”, “Biweek”, “Week”, and “Month.
- species_list: Species filter for calculating abundance. Species_display_name is the accepted notation. To see a list of species present in your data run unique(collections\$species_display_name). If species is unspecified, the default NULL will return data for all species in data.
- trap_list: Trap filter for calculating abundance. Trap_acronym is the is the accepted notation. Run unique(collections\$trap_acronym) to see trap types present in your data. If trap_list is unspecified, the default NULL will return data for all trap types.
- species_separate: Should the species in species_list have abundance calculated separately? Setting to FALSE calculates the combined abundance. The same result can be performed by calculating on one species at the time.

```
getAbundance(collections,
              interval = "Biweek",
              species_list = c("Cx tarsalis", "Cx pipiens"),
              trap_list = "C02",
              species_separate = FALSE)
```

##	EPIYEAR	Biweek	Count	Trap_Events	Abundance
## 1	2023	10	882	65	13.57
## 2	2023	11	3254	142	22.92
## 3	2023	12	4395	153	28.73
## 4	2023	13	15803	182	86.83
## 5	2023	14	24939	226	110.35
## 6	2023	15	24113	217	111.12
## 7	2023	16	19062	255	74.75
## 8	2023	17	12865	226	56.92
## 9	2023	18	10088	213	47.36
## 10	2023	19	7161	211	33.94
## 11	2023	20	5934	211	28.12
## 12	2023	21	2780	140	19.86
## 13	2022	4	9	3	3.00
## 14	2022	9	1358	126	10.78
## 15	2022	10	1202	133	9.04
## 16	2022	11	1969	145	13.58
## 17	2022	12	3503	159	22.03
## 18	2022	13	5630	159	35.41
## 19	2022	14	10444	154	67.82

##	20	2022	15	9722	178	54.62
##	21	2022	16	7949	186	42.74
##	22	2022	17	6501	180	36.12
##	23	2022	18	6038	166	36.37
##	24	2022	19	3798	163	23.30
##	25	2022	20	1869	120	15.57
##	26	2022	21	1189	84	14.15

Abundance Anomaly (comparison to 5 year average)

getAbundanceAnomaly()

Description

getAbundanceAnomaly(..) requires at least five years prior to the target_year of arthro collections data to calculate for the specified parameters. The function uses the methods of the Gateway Abundance Anomaly calculator, and will not work if there is fewer than five years of data present.

Usage

```
getAbundanceAnomaly(collections,interval,target_year, species_list = NULL, trap_list = NULL,
species_seperate = FALSE)
```

Arguments

- collections: Collections data retrieved from getArthroCollections(...)
- interval: Calculation interval for abundance, accepts "collection_date", "Biweek", "Week", and "Month."
- target_year: Year to calculate analysis on. Collections data must have a year range of at least (target_year - 5, target_year).
- species_list: Species filter for calculating abundance. Species_display_name is the accepted notation. To see a list of species present in your data run unique(collections\$species_display_name). If species is unspecified, the default NULL will return data for all species in data.
- trap_list: Trap filter for calculating abundance. Trap_acronym is the is the accepted notation. Run unique(collections\$trap_acronym) to see trap types present in your data. If trap_list is unspecified, the default NULL will return data for all trap types.
- species_seperate: Should the species in species_list have abundance calculated separately? Setting to FALSE calculates the combined abundance. The same result can be performed by calculating on one species at the time.

```
collections_18_23 = getArthroCollections(2018,2023, 55)

getAbundanceAnomaly(collections_18_23,
                     interval = "Biweek",
                     target_year = 2023,
                     species_list = c("Cx tarsalis", "Cx pipiens"),
                     trap_list = "CO2",
                     species_seperate = FALSE)
```

##	Biweek	EPIYEAR	Count	Trap_Events	Abundance	Five_Year_Avg	Delta
## 1	10	2023	882	65	13.57	12.926	4.98
## 2	11	2023	3254	142	22.92	19.666	16.55
## 3	12	2023	4395	153	28.73	37.988	-24.37
## 4	13	2023	15803	182	86.83	54.496	59.33
## 5	14	2023	24939	226	110.35	81.972	34.62
## 6	15	2023	24113	217	111.12	75.588	47.01

## 7	16	2023	19062	255	74.75	78.528	-4.81
## 8	17	2023	12865	226	56.92	66.406	-14.28
## 9	18	2023	10088	213	47.36	61.704	-23.25
## 10	19	2023	7161	211	33.94	51.736	-34.40
## 11	20	2023	5934	211	28.12	32.970	-14.71
## 12	21	2023	2780	140	19.86	20.082	-1.11

Infection Rate

getInfectionRate()

Description

getInfectionRate(.) requires at least five years prior to the target_year of arthro collections data to calculate for the specified parameters. The function uses the methods of the Gateway Abundance Anomaly calculator, and will not work if there is fewer than five years of data present.

Usage

```
getInfectionRate(pools,interval, target_year, target_disease,pt_estimate, species_list = c(NULL), trap_list = c(NULL))
```

Arguments

- pools: Pools data retrieved from getPools(...)
- interval: Calculation interval for abundance, accepts "collection_date", "Biweek", "Week", and "Month."
- target_year: Year to calculate infection rate for. This year must be present in the data.
- target_disease: The disease to calculate infection rate for—i.e. "WNV". Disease acronyms are the accepted input. To see a list of disease acronyms, run unique(pools\$target_acronym).
- pt_estimate: The estimation type for infection rate. Options include: "mle", "bc-mle", "mir."
- species_list: Species filter for calculating abundance. Species_display_name is the accepted notation. To see a list of species present in your data run unique(pools\$species_display_name). If species is unspecified, the default NULL will return data for all species in data.
- trap_list: Trap filter for calculating abundance. Trap_acronym is the accepted notation. Run unique(pools\$trap_acronym) to see trap types present in your data. If trap_list is unspecified, the default NULL will return data for all trap types.

```
IR = getInfectionRate(pools,
  interval = "Week",
  target_year = 2023,
  target_disease = "WNV",
  pt_estimate = "mle",
  species_list = c("Cx pipiens"),
  trap_list = c("CO2","GRVD") )
IR
```

##	Year	Week	Disease	Point_Estimate	Lower_CI	Upper_CI
## 1	2023	20	WNV	0.0000000	0.00000000	4.617179
## 2	2023	21	WNV	0.0000000	0.00000000	4.119261
## 3	2023	22	WNV	0.0000000	0.00000000	3.156551
## 4	2023	23	WNV	0.5727378	0.03289081	2.738134
## 5	2023	24	WNV	0.0000000	0.00000000	1.747176
## 6	2023	25	WNV	0.5095548	0.02921417	2.445178
## 7	2023	26	WNV	2.8603431	1.45376937	5.081510
## 8	2023	27	WNV	5.8779801	3.18495627	9.940152

##	9	2023	28	WNV	7.3108593	4.66480012	10.887089
##	10	2023	29	WNV	11.5973679	7.96664790	16.265986
##	11	2023	30	WNV	11.9602412	7.42715801	18.168158
##	12	2023	31	WNV	9.9743699	6.40363989	14.753549
##	13	2023	32	WNV	12.6118506	7.38382600	20.046595
##	14	2023	33	WNV	9.9985982	5.74908449	16.099310
##	15	2023	34	WNV	16.5956182	10.93686159	24.017062
##	16	2023	35	WNV	8.4449005	3.45657161	17.297528
##	17	2023	36	WNV	8.7729202	4.48933276	15.456824
##	18	2023	37	WNV	3.5282350	1.31243740	7.706321
##	19	2023	38	WNV	2.7297894	0.72200878	7.245097
##	20	2023	39	WNV	0.0000000	0.00000000	2.223982
##	21	2023	40	WNV	1.8502043	0.33078143	5.950046
##	22	2023	41	WNV	1.0256410	0.05906777	4.851477
##	23	2023	42	WNV	0.0000000	0.00000000	20.603165

Vector Index

getVectorIndex()

Description

getVectorIndex()(..) requires at least five years prior to the target_year of arthro collections data to calculate for the specified parameters. The function uses the methods of the Gateway Abundance Anomaly calculator, and will not work if there is fewer than five years of data present.

Usage

```
getVectorIndex(collections, pools, interval, target_year, target_disease, pt_estimate, species_list=NULL, trap_list = NULL)
```

Arguments - collections: collections data retrieved from getCollections(...) - pools: Pools data retrieved from getPools(...)

Note: Years from pools and collections data must match

- interval: Calculation interval for abundance, accepts “collection_date”, “Biweek”, “Week”, and “Month.
- target_year: Year to calculate infection rate for. This year must be present in the data.
- target_disease: The disease to calculate infection rate. Disease acronyms are the accepted input. To see a list of disease acronyms, run unique(pools\$target_acronym).
- pt_estimate: The estimation type for infection rate. Options include: “mle”, “bc-”mle”, “mir.”
- species_list: Species filter for calculating abundance. Species_display_name is the accepted notation. To see a list of species present in your data run unique(pools\$species_display_name). If species is unspecified, the default NULL will return data for all species in data.
- trap_list: Trap filter for calculating abundance. Trap_acronym is the is the accepted notation. Run unique(pools\$trap_acronym) to see trap types present in your data. If trap_list is unspecified, the default NULL will return data for all trap types.

```
source("VS_functions.R")
pools=getPools(2023,2023,55)
collections= getArthroCollections(2023,2023,55)
```

```
getVectorIndex(collections, pools, interval = "Biweek",
               2023,
               target_disease = "WNV", pt_estimate = "bc-mle",
               species_list=c("Cx tarsalis"),

               trap_list = c("C02"))
```

##	Biweek	EPIYEAR	Count	Trap_Events	Abundance	Year	Disease	Point_Estimate
## 1	10	2023	509	65	7.83	2023	WNV	0.0000000
## 2	11	2023	1910	142	13.45	2023	WNV	0.5205304
## 3	12	2023	2343	153	15.31	2023	WNV	0.4359300
## 4	13	2023	12226	182	67.18	2023	WNV	0.9791292
## 5	14	2023	21573	226	95.46	2023	WNV	3.2518724
## 6	15	2023	20979	217	96.68	2023	WNV	7.2460033
## 7	16	2023	17215	255	67.51	2023	WNV	10.4738708
## 8	17	2023	11019	226	48.76	2023	WNV	9.4266562
## 9	18	2023	8184	213	38.42	2023	WNV	5.9380352
## 10	19	2023	4625	211	21.92	2023	WNV	4.3159178
## 11	20	2023	3213	211	15.23	2023	WNV	0.7772615
## 12	21	2023	1479	140	10.56	2023	WNV	0.0000000

##	Lower_CI	Upper_CI	VectorIndex
## 1	0.00000000	6.735594	0.000000
## 2	0.02996660	2.523691	7.001133
## 3	0.02501275	2.117993	6.674089
## 4	0.43252140	1.931931	65.777899
## 5	2.37760030	4.355111	310.423742
## 6	5.77210610	9.004829	700.543600
## 7	8.34799912	13.020608	707.091020
## 8	7.47048950	11.776108	459.643757
## 9	4.32273418	7.990012	228.139314
## 10	2.61019696	6.773983	94.604917
## 11	0.13915932	2.544228	11.837692
## 12	0.00000000	3.338641	0.000000

Tables

getPoolsComparisionTable()

Description

getPoolsComparisionTable() produces a frequency table for positive and negative pools counts by year and species. The more years present in the data, the larger the table.

Usage

```
getPoolsComparisionTable(pools,target_disease, species_seperate=F)
```

Arguments

- pools: Pools data retrieved from getPools(...)
- target_disease: The disease to calculate infection rate for—i.e. “WNV”. Disease acronyms are the accepted input. To see a list of disease acronyms, run unique(pools\$target_acronym).
- species_seperate: Should the pools comparison be split by species of each pool. Default is FALSE.

```
getPoolsComparisionTable(pools, "WNV", species_seperate = T)
```

```
## # A tibble: 3 x 6
## # Groups:   surv_year, species_display_name [3]
##   surv_year species_display_name Negative Confirmed Total PercentPositive
##     <int> <chr>                <int>      <int> <int>          <dbl>
## 1     2023 An freeborni             1         0     1           0
## 2     2023 Cx pipiens            3820       247   4067          6.07
## 3     2023 Cx tarsalis           3392       393   3785         10.4
```

Styling Dataframes with kable

Professional looking tables can be produced using the kable and kableExtra packages.

```
AbAnOutput = getAbundance(collections,
                          interval = "Biweek",

                          species_list = c("Cx tarsalis", "Cx pipiens"),
                          trap_list = "CO2",
                          species_seperate = FALSE)
head(AbAnOutput)
```

```
##   EPIYEAR Biweek Count Trap_Events Abundance
## 1     2023     10   882         65     13.57
## 2     2023     11  3254        142     22.92
## 3     2023     12  4395        153     28.73
## 4     2023     13 15803        182     86.83
## 5     2023     14 24939        226    110.35
## 6     2023     15 24113        217    111.12
```

#As a kable table where column names, font_size, type and much more can be customized

```
AbAnOutput %>%
  kbl(col.names = c("Disease Year", "Biweek", "Count", "Trap Events", "Abundance")) %>%
  kable_styling(bootstrap_options = "striped",
               font_size = 14,
               latex_options="scale_down") %>%
  footnote(general = "Table X: Combined biweekly Abundance Calculation for Cx. tarsalis, pipiens in CO2")
```

Data using datatables

Interactive html only tables can be produced using the DT package. DT tables allow for sorting and filtering with in a webpage. These are ideal for viewing data but are not compatable with pdf or word formats.

```
#AbAnOutput %>%
#datatable(colnames = c("Disease Year", "Biweek", "Count", "Trap Events", "Abundance"))
```

Disease Year	Biweek	Count	Trap Events	Abundance
2023	10	882	65	13.57
2023	11	3254	142	22.92
2023	12	4395	153	28.73
2023	13	15803	182	86.83
2023	14	24939	226	110.35
2023	15	24113	217	111.12
2023	16	19062	255	74.75
2023	17	12865	226	56.92
2023	18	10088	213	47.36
2023	19	7161	211	33.94
2023	20	5934	211	28.12
2023	21	2780	140	19.86

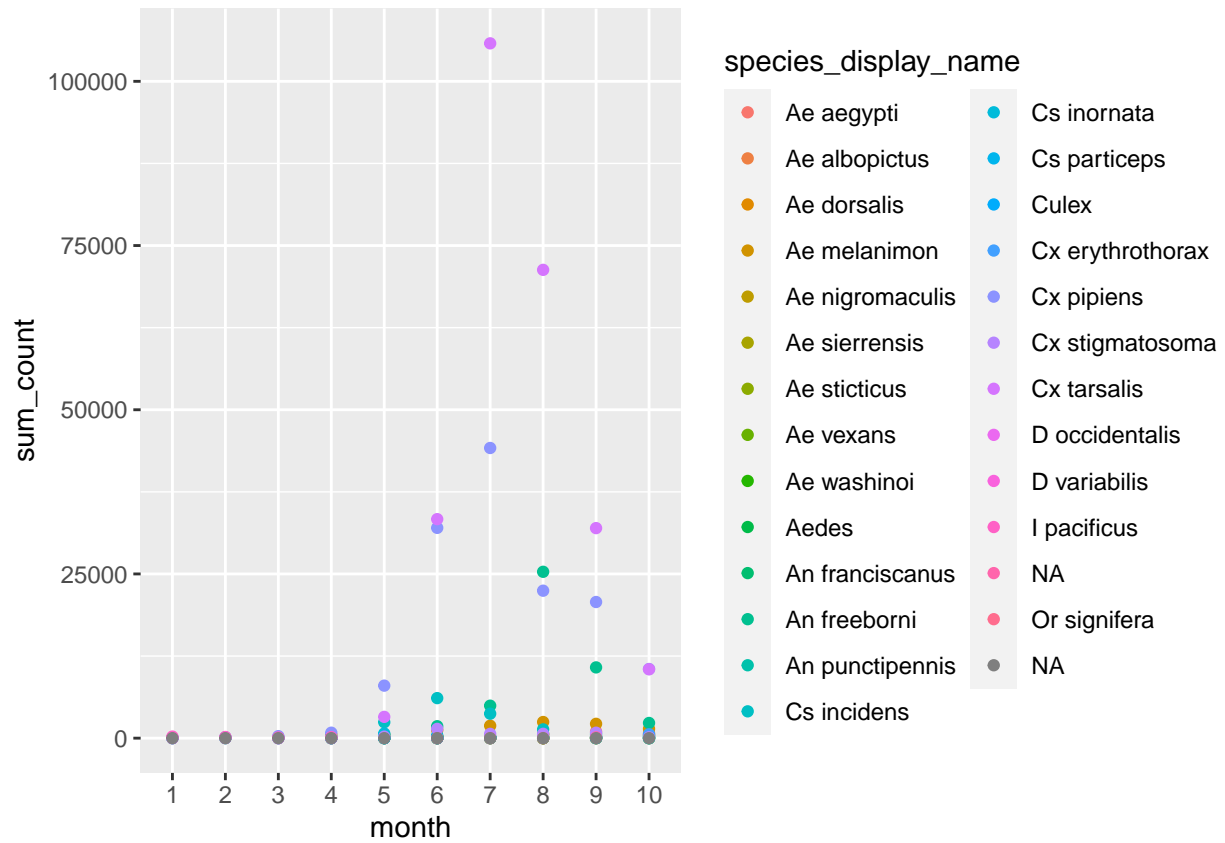
Table X: Combined biweekly Abundance Calculation for Cx. tarsalis, pipiens in CO2 traps

Charts and Graphs

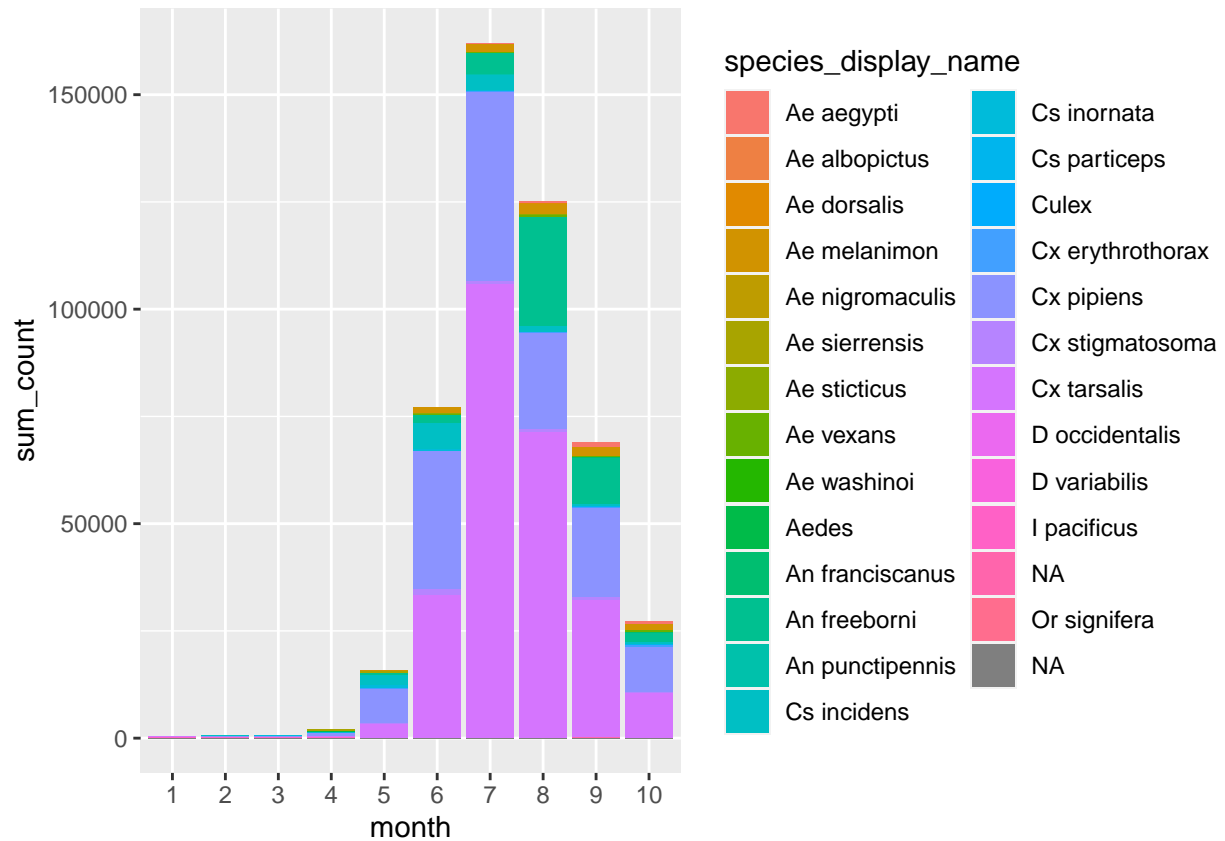
Ggplot is a easy to use plotting library in R. Gplot syntax consists of creating a ggplot object with a dataframe and adding subsequent arguments to that object. Aesthetics (aes) in ggplot represents the data mapping aspect of the plot. A simple example using collections is shown below.

```
#creates a month column and translates numerics
collections$month = as.factor(month(collections$collection_date))
collections_sums = collections %>%
  group_by(month, species_display_name) %>%
  summarise(sum_count = sum(num_count, na.rm=T))

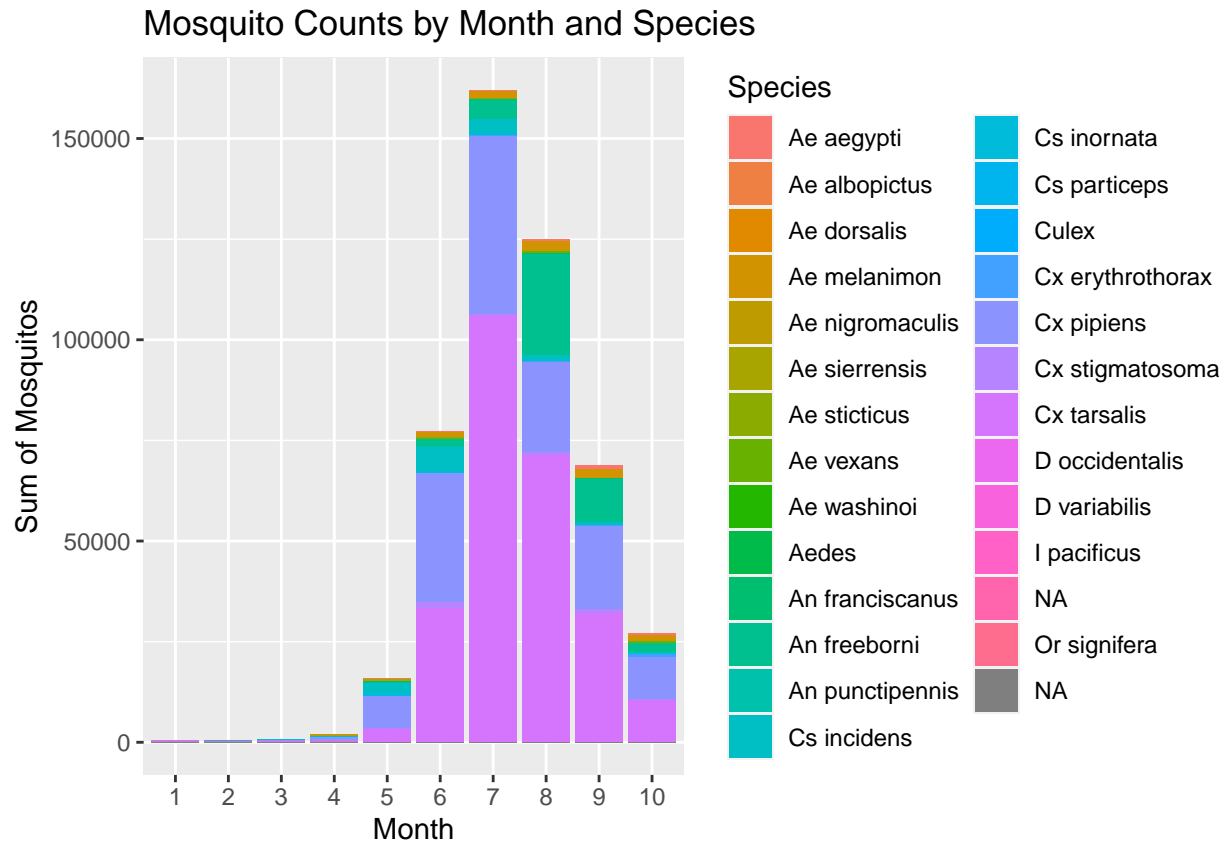
#ggplot with dots a values for each species
ggplot(data=collections_sums,
  aes(x = month, y = sum_count, color = species_display_name))+
  geom_point()
```



```
#bar chart
ggplot(data=collections_sums,
  aes(x = month, y = sum_count, fill = species_display_name))+
  geom_bar(stat="identity")
```



```
#adding labels
ggplot(data=collections_sums,
  aes(x = month, y = sum_count, fill = species_display_name))+
  geom_bar(stat="identity") +
  labs(title = "Mosquito Counts by Month and Species",x = "Month",
    y = "Sum of Mosquitos",
    fill = "Species")
```

When plotting with libraries in R, it is easiest when the data is prepared in long form. Most calculator outputs from our functions are in wide form. The following wrapper functions help process and plot this data.

ProcessAbunAnom()

Description

ProcessAbunAnom() processes the output returned from getAbundanceAnomaly() into a long form suitable for plotting in ggplot.

Usage

```
ProcessAbunAnom(AbAnomOutput)
```

Arguments

- AbAnomOutput: Output from returned getAbundanceAnomaly()

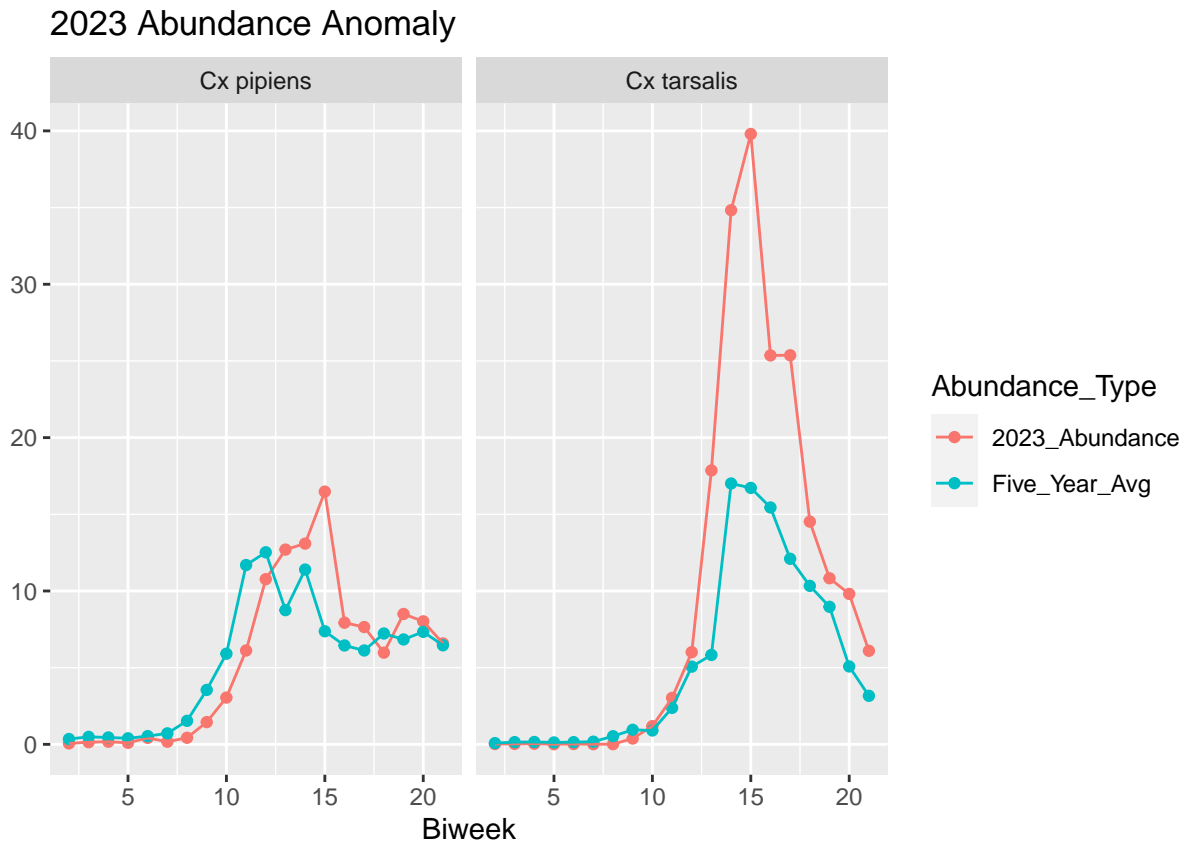
```
AbAnOut = getAbundanceAnomaly(collections_18_23,
                              interval = "Biweek",
                              target_year = 2023,
                              species_list = c("Cx tarsalis", "Cx pipiens"),
                              species_seperate = TRUE) #species_seperate set to true will allow for us
```

```
AbAnOut_L = ProcessAbunAnom(AbAnOut)
```

#Example using ggplot

```
AbAnOut_L %>% filter(Abundance_Type %in% c("2023_Abundance",
                                             "Five_Year_Avg"))%>%

  ggplot(aes(x=Biweek,
             y= Abundance_Calculation,
             color = Abundance_Type)) +
  geom_point()+
  geom_line() +
  facet_wrap(~species_display_name)+
  labs(title="2023 Abundance Anomaly",y = "")
```



plotInfectionRate()

Description

plotInfectionRate() plots the output returned from getInfectionRate() with confidence intervals using ggplot

Usage

```
plotInfectionRate(InfRtOutput)
```

Arguments

- InfRtOutput: Output from returned getInfectionRate()

```
IR = getInfectionRate(pools,
                      interval = "Week",
                      target_year = 2023,
```

```

target_disease = "WNV",
pt_estimate = "mle",
species_list = c("Cx pipiens"),
trap_list = c("CO2", "GRVD") )

```

```
plotInfectionRate(InfRtOutput = IR)
```

