

Longitudinal analysis of Shigella

0.1 Data description

- Step 1: Load the Dataset

```
df <- read_excel("3.8.2024 Compiled Shigella datav2.xlsx",  
                 sheet = "Compiled")
```

- Step 2: Get a Quick Overview of the Data Structure

Table 1: Data summary

Name	df
Number of rows	2374
Number of columns	31
Column type frequency:	
character	9
numeric	22
Group variables	None

Variable type: character

skim_variable	n_missing	complete_rate	min	max	empty	n_unique	whitespace
isotype_name	0	1.00	3	3	0	2	0
dilution_name	0	1.00	6	6	0	1	0
study_name	0	1.00	4	22	0	5	0
sampleID	0	1.00	6	11	0	1017	0
sid	0	1.00	6	11	0	1017	0
unq_id	0	1.00	22	27	0	2374	0
cohort_name	0	1.00	3	11	0	12	0

skim_variable	n_missing	complete_rate	min	max	empty	n_unique	whitespace
site_name	0	1.00	5	12	0	5	0
treatment	1126	0.53	7	7	0	2	0

Variable type: numeric

skim_variable	n_missing	complete_rate	mean	sd	p0	p25	p50	p75	p100	hist
isotype	0	1.00	1.50	0.50	1.00	1.00	1.50	2.00	2.00	
dilution	0	1.00	1.00	0.00	1.00	1.00	1.00	1.00	1.00	
cohort	0	1.00	8.49	1.66	4.00	9.00	9.00	9.00	15.00	
site	0	1.00	5.33	2.22	1.00	2.50	7.00	7.00	7.00	
age	0	1.00	6.58	8.66	0.08	2.00	3.67	7.00	50.00	
timepoint	0	1.00	969.01	901.92	0.00	0.00	1825.00	1825.00	1825.00	
Actual day	1942	0.18	53.06	62.90	2.00	7.00	30.00	90.00	204.00	
ipab_MFI	0	1.00	43525.84	59594.93	-	818.58	4759.92	86098.81	189355.75	
					1.25					
sf3aospbsa_MFI	0	1.00	3651.27	8563.89	-	183.25	753.71	2956.48	83828.92	
					9.00					
sf2aospbsa_MFI	0	1.00	4681.13	14202.95	-	100.06	489.04	2694.00	214007.67	
					12.00					
sf6ospbsa_MFI	0	1.00	4043.35	9996.03	-	278.25	1036.29	3714.58	166878.50	
					7.75					
sonneiospbsa_MFI	0	1.00	3007.76	12898.10	-	28.08	118.12	887.46	184109.50	
					14.50					
n_ipab_MFI	0	1.00	24971.08	34729.71	-	361.53	1903.47	51093.85	116048.97	
					0.63					
n_sf3aospbsa_MFI	0	1.00	5702.38	12801.36	-	320.15	1273.00	4743.07	112888.86	
					12.10					
n_sf2aospbsa_MFI	0	1.00	5308.11	15272.92	-	135.66	621.84	3295.17	222195.19	
					11.82					
n_sf6ospbsa_MFI	0	1.00	2991.34	6997.96	-	231.92	845.01	2826.64	108831.02	
					6.84					
n_sonneiospbsa_MFI	0	1.00	3747.64	14652.84	-	38.78	183.09	1335.17	219024.17	
					28.75					
ipab_BC	0	1.00	80.03	29.48	17.00	59.00	73.00	91.00	260.00	
sf3aospbsa_BC	0	1.00	103.91	22.52	57.00	88.00	100.00	115.00	226.00	
sf2aospbsa_BC	0	1.00	67.03	18.96	23.00	54.00	63.00	76.00	155.00	
sf6ospbsa_BC	0	1.00	56.18	13.04	27.00	47.00	53.00	63.00	115.00	
sonneiospbsa_BC	0	1.00	58.37	14.60	17.00	49.00	58.00	68.00	114.00	

- Step 3: Identify and Summarize Missing Values (for the entire dataset and the SOSAR study)

```
# A tibble: 31 x 2
  variable      missing_count
  <chr>          <int>
1 Actual day      1942
2 treatment       1126
3 isotype          0
4 isotype_name     0
5 dilution        0
6 dilution_name   0
7 study_name       0
8 sampleID         0
9 sid              0
10 unq_id          0
# i 21 more rows
```

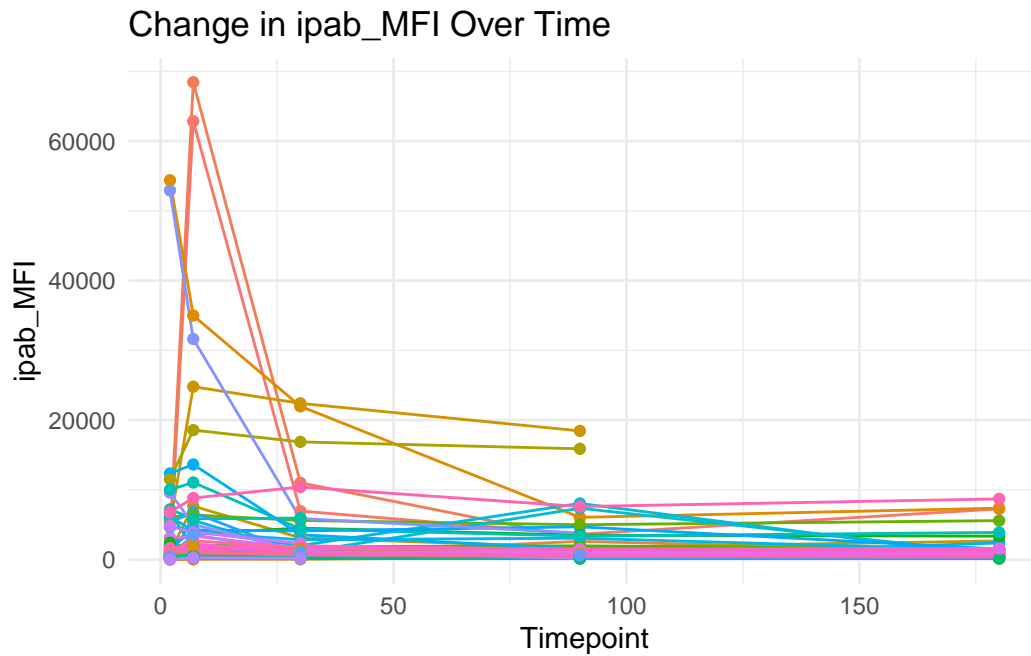
```
# A tibble: 31 x 2
  variable      missing_count
  <chr>          <int>
1 treatment       436
2 Actual day        4
3 isotype          0
4 isotype_name     0
5 dilution        0
6 dilution_name   0
7 study_name       0
8 sampleID         0
9 sid              0
10 unq_id          0
# i 21 more rows
```

0.2 Exploratory Data Analysis

- Step 4: Filter Data for the SOSAR Study and Split Data by Isotype

Verifying the unique sample identification numbers (SIDs) is crucial. There are 48 SIDs in this SOSAR study.

- Step 5: Plot Quantitative Response Change Over Time



0.3 Longitudinal Antibody Decay Modeling

- Step 6: Prepare Data for JAGS Model
- Step 7: Subset Data for Model Testing

```
#subset data for checking
dL_sub <- dL %>%
  filter(index_id %in% sample(unique(index_id), 20))
```

- Step 8: Prepare Data and Priors for JAGS
- Step 9: Run the JAGS Model

```
dump_and_quit <- function() {
  dump.frames(to.file = TRUE)
  q(status = 1)
}
options(error = dump_and_quit)
```

```
[1] "Starting run.jags execution"
```

List of 5

```
$ smpl.t          : num [1:21, 1:5] 2 2 2 2 2 2 2 2 2 2 ...
..- attr(*, "dimnames")=List of 2
.. ..$ subjects   : chr [1:21] "SOSAR-11001" "SOSAR-12017" "SOSAR-11012" "SOSAR-12005" ..
.. ..$ visit_number: chr [1:5] "V1" "V2" "V3" "V4" ...
$ logy           : num [1:21, 1:5, 1:2] -4.61 -4.61 -4.61 -4.61 -4.61 ...
..- attr(*, "dimnames")=List of 3
.. ..$ subjects   : chr [1:21] "SOSAR-11001" "SOSAR-12017" "SOSAR-11012" "SOSAR-12005" ..
.. ..$ visit_number: chr [1:5] "V1" "V2" "V3" "V4" ...
.. ..$ antigens    : chr [1:2] "IgG" "IgA"
$ n_antigen_isos: int 2
$ nsmpl          : num [1:21] 5 4 5 5 5 5 5 5 5 5 ...
$ nsubj          : num 21
- attr(*, "antigens")= chr [1:2] "IgG" "IgA"
- attr(*, "n_antigens")= int 2
- attr(*, "ids")= chr [1:21] "SOSAR-11001" "SOSAR-12017" "SOSAR-11012" "SOSAR-12005" ...
NULL
```

List of 6

```
$ n_params       : num 5
$ mu.hyp         : num [1:2, 1:5] 1 1 7 7 1 1 -4 -4 -1 -1
$ prec.hyp       : num [1:2, 1:5, 1:5] 1 1 0 0 0 0 0 0 0 0 ...
$ omega          : num [1:2, 1:5, 1:5] 1 1 0 0 0 0 0 0 0 0 ...
$ wishdf         : num [1:2] 20 20
$ prec.logy.hyp: num [1:2, 1:2] 4 4 1 1
NULL
```

Calling 4 simulations using the parallel method...

Following the progress of chain 1 (the program will wait for all chains to finish before continuing):

Welcome to JAGS 4.3.2 on Wed Aug 28 00:47:59 2024

JAGS is free software and comes with ABSOLUTELY NO WARRANTY

Loading module: basemod: ok

Loading module: bugs: ok

. . Reading data file data.txt

. Compiling model graph

Resolving undeclared variables

Allocating nodes

Graph information:

Observed stochastic nodes: 186

Unobserved stochastic nodes: 54

Total graph size: 3201

```

. Reading parameter file inits1.txt
. Initializing model
. Adapting 100
-----| 100
+++++ 100%
Adaptation incomplete.
. Updating 100
-----| 100
***** 100%
. . . . . Updating 100
-----| 100
***** 100%
. . . . . Updating 0
. Deleting model
.
All chains have finished

```

Warning: The adaptation phase of one or more models was not completed in 100 iterations, so the current samples may not be optimal - try increasing the number of iterations to the "adapt" argument

```

Simulation complete. Reading coda files...
Coda files loaded successfully
Finished running the simulation

```

```
[1] "run.jags completed successfully"
```

- Step 10: Covert JAGS Output to MCMC List

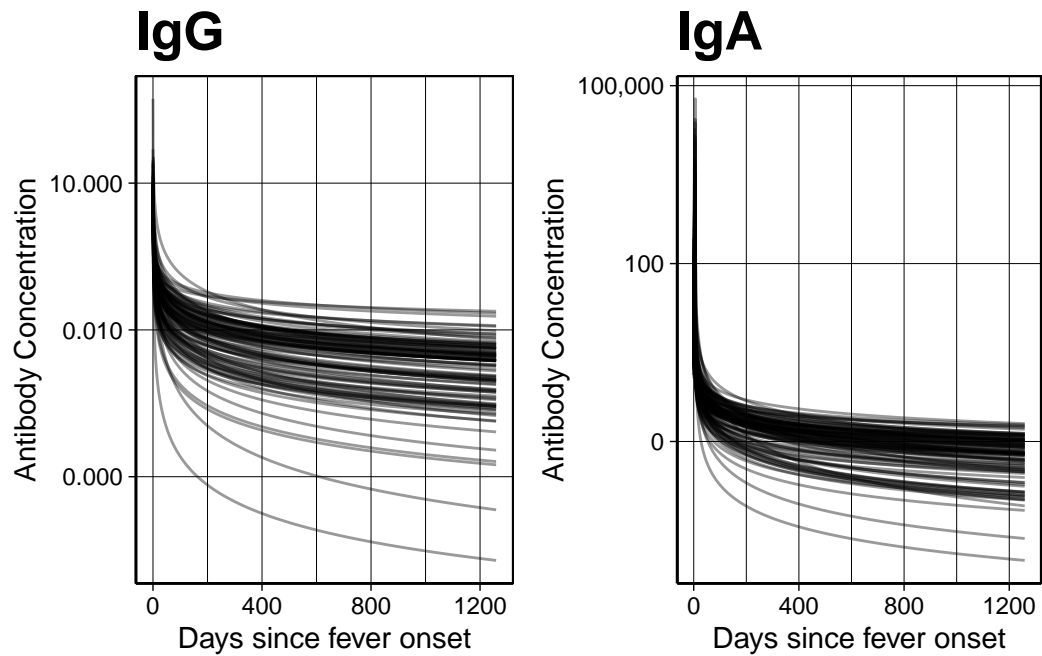
```

mcmc_list <- as.mcmc.list(jags.post)
mcmc_df <- ggs(mcmc_list)

```

- Step 11: Prepare Data for Serocalculator Package

- Step 12: Plot Longitudinal Antibody Decay



- Step 13: Define the Antibody Calculation Function

```
ab <- function(t, y0, y1, t1, alpha, shape) {
  beta <- log(y1/y0) / t1
  if (t <= t1) {
    yt <- y0 * exp(beta * t)
  } else {
    yt <- (y1^(1 - shape) - (1 - shape) * alpha * (t - t1))^(1 / (1 - shape))
  }
  return(yt)
}
```

- Step 14: Generate Time Points for Antibody Levels
- Step 15: Calculate Antibody Levels Over Time

- Step 16: Summarize and Plot Antibody Decay Data

