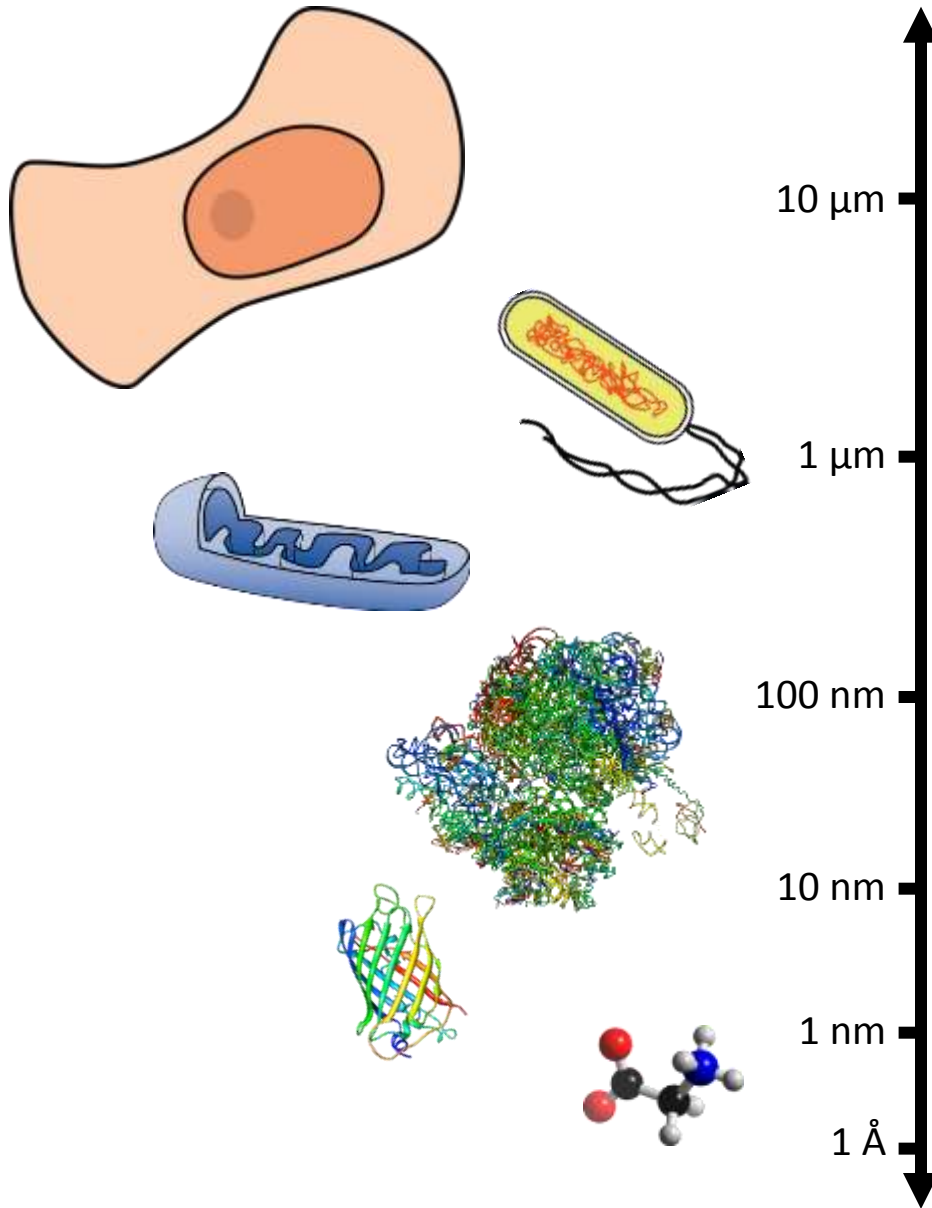


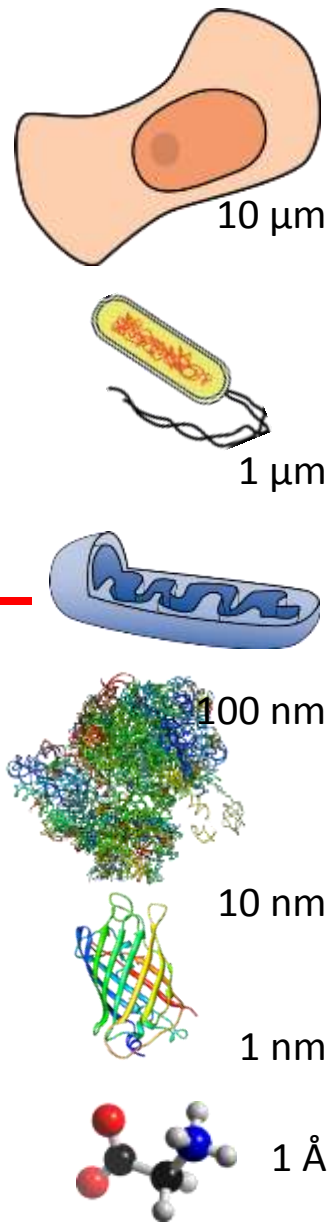
# Super-Resolution Microscopy Structured Illumination



Bo Huang

# Looking into microscopic world of life...

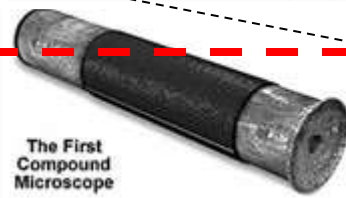




Naked eye:  $\sim 50\text{-}100\ \mu\text{m}$

★ 1595, Zaccharias and Hans Janssen  
First microscope, 9x magnification

★ Antonie van Leeuwenhoek  
(1632-1723), 200x



Compound microscope  
 $>1000\times$

Ernst Abbe (1840-1905)  
The "physical" diffraction limit



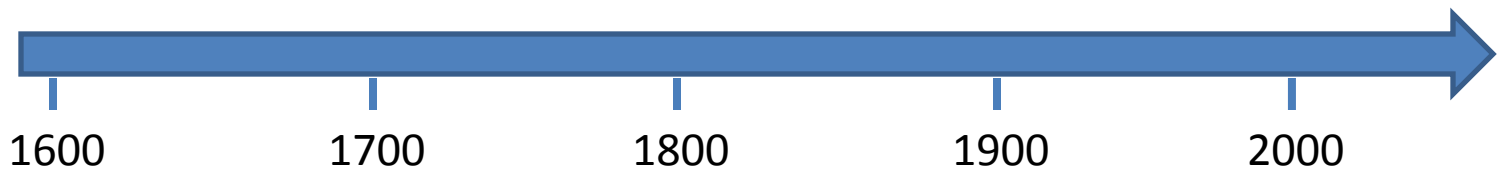
PLATE XXIV

fig: A

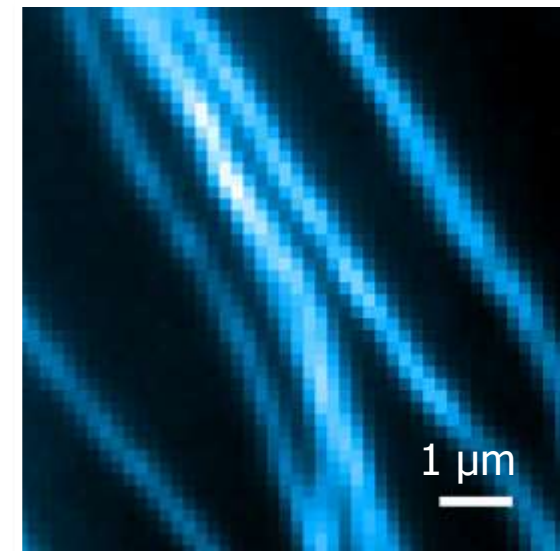
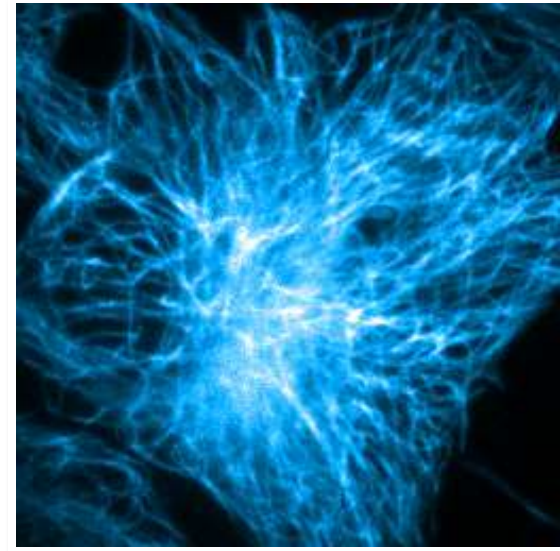
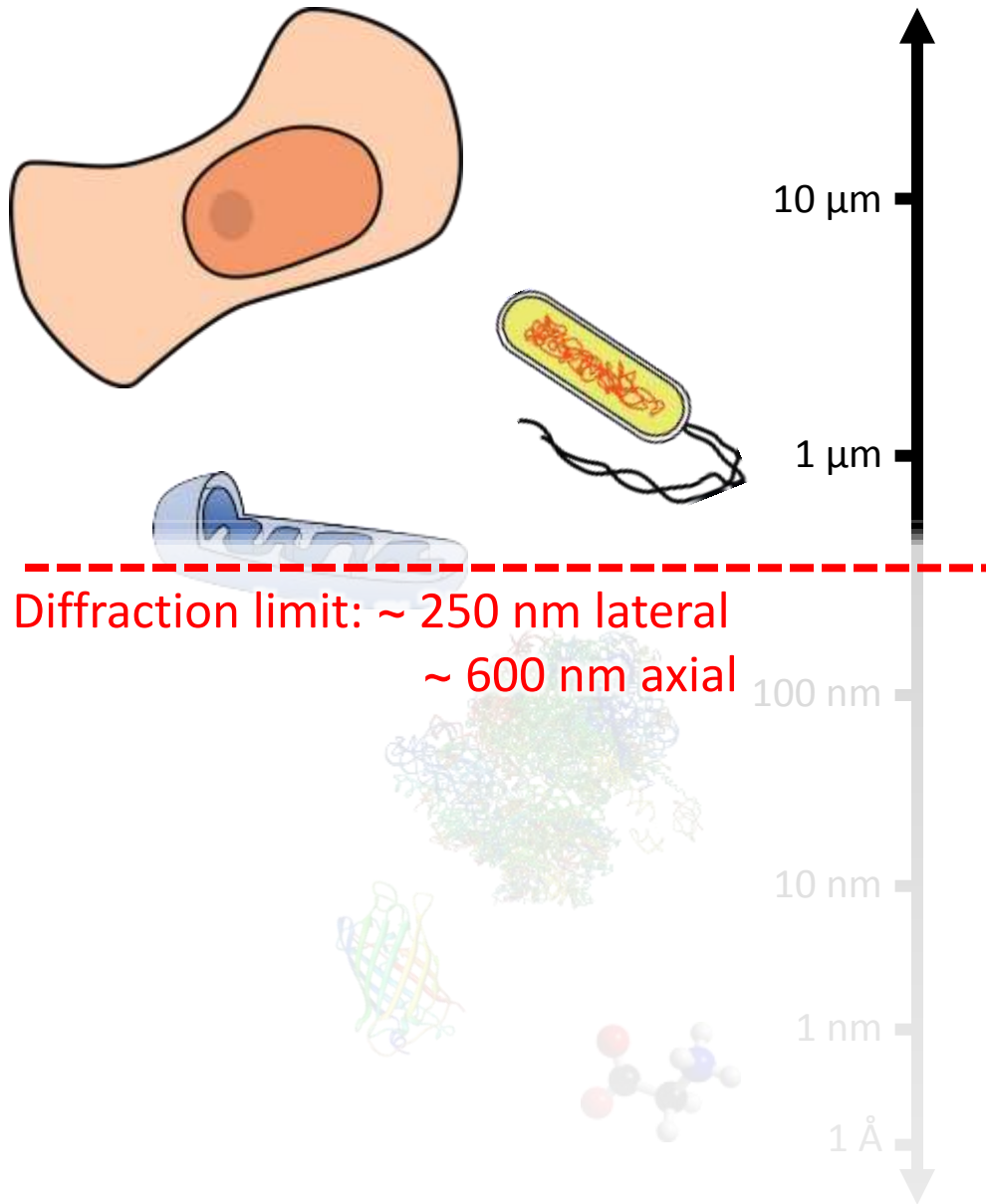
fig: B

fig: C

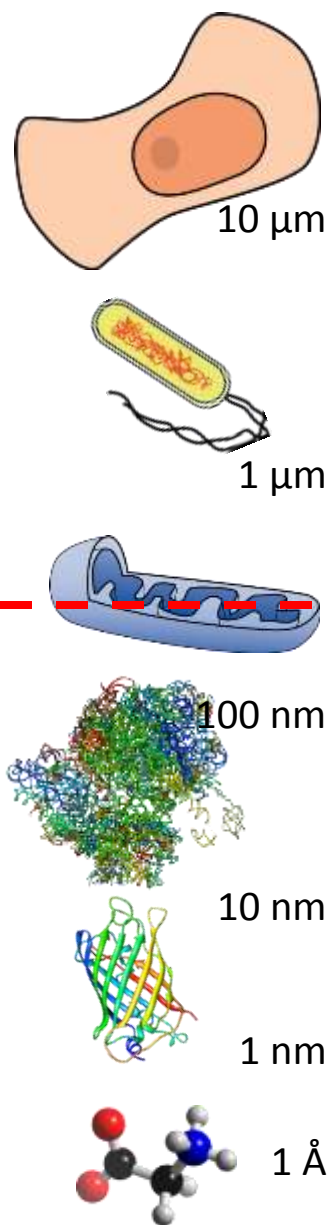
fig: r



# The diffraction barrier



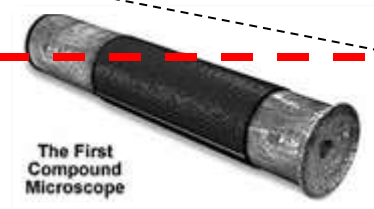
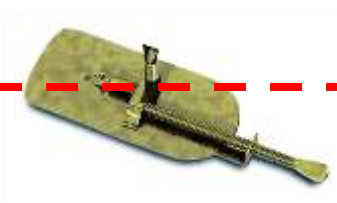




Naked eye:  $\sim 50\text{-}100\ \mu\text{m}$

★ 1595, Zaccharias and Hans Janssen  
First microscope, 9x magnification

★ Antony Van Leeuwenhoek  
(1632-1723), 200x

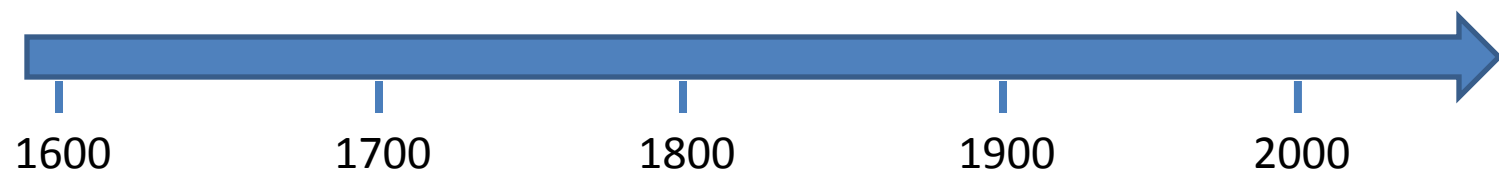
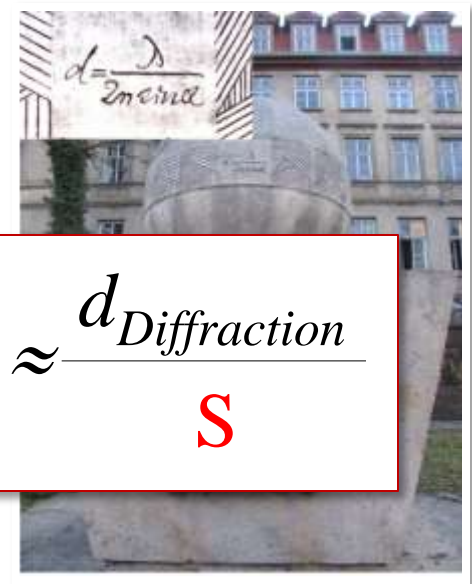


Compound microscope  
>1000x

Ernst Abbe (1840-1905)  
The "physical" diffraction limit

**Super-resolution**  
Deconvolution  
**Optical Microscopy**  
4-Pi microscopy  
...

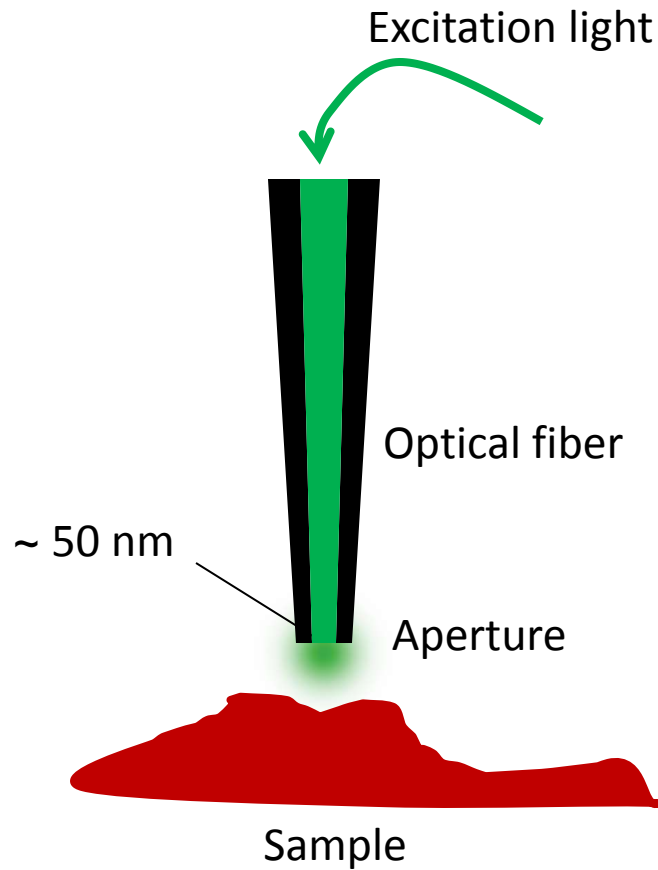
$$D \approx \frac{d_{\text{Diffraction}}}{S}$$



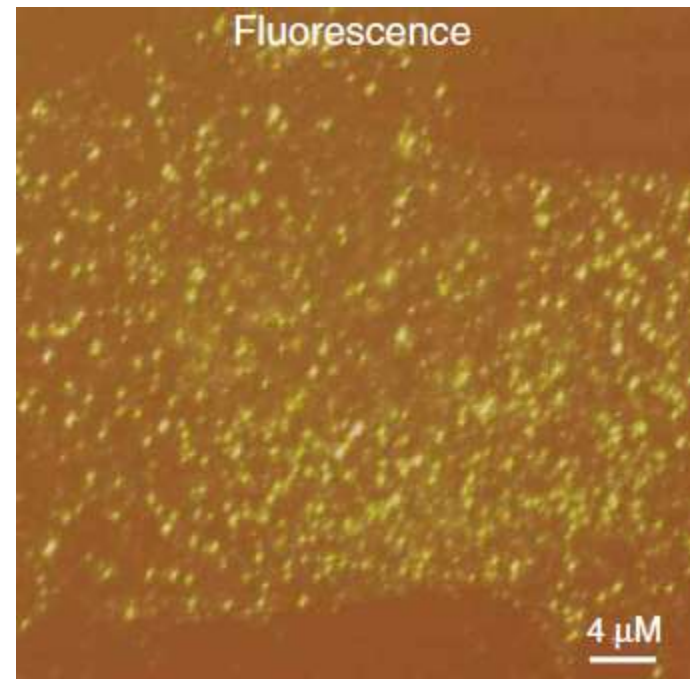
# 50 years to extend the resolution

- Confocal microscopy (1957)
- Near-field scanning optical microscopy (1972/1984)
- Multiphoton microscopy (1990)
- 4-Pi microscopy / I<sup>5</sup>M (1991-1995)
- Structured illumination microscopy (2000)
- Negative refractive index (2006)

# Near-field scanning optical microscopy

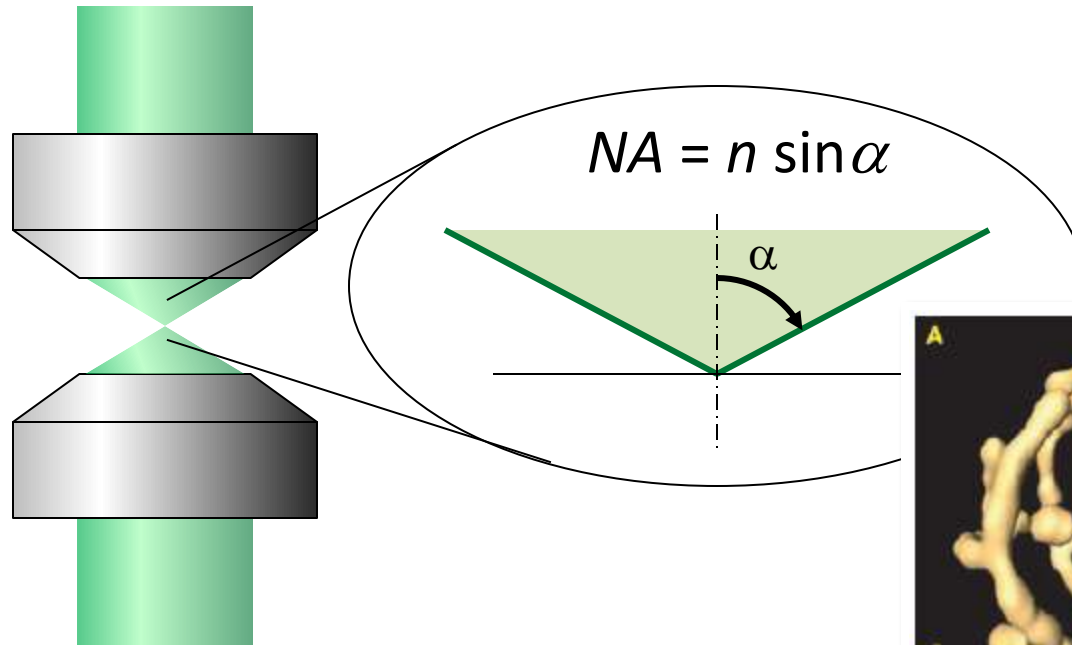


$\beta_2$  adrenergic receptor clusters  
on the plasma membrane

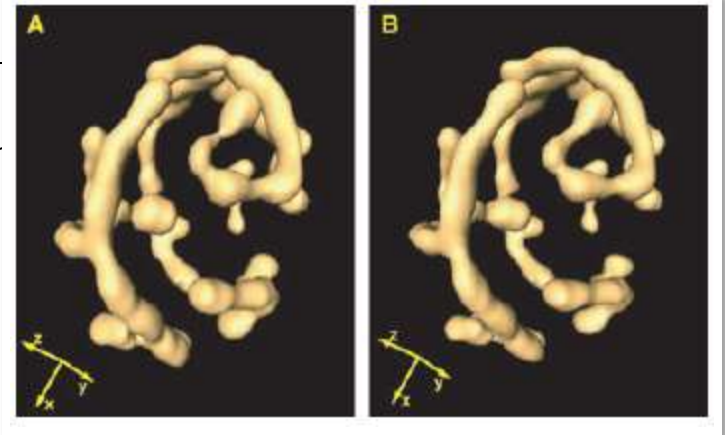


Ianoul et al., 2005

# 4-Pi / I<sup>5</sup>M



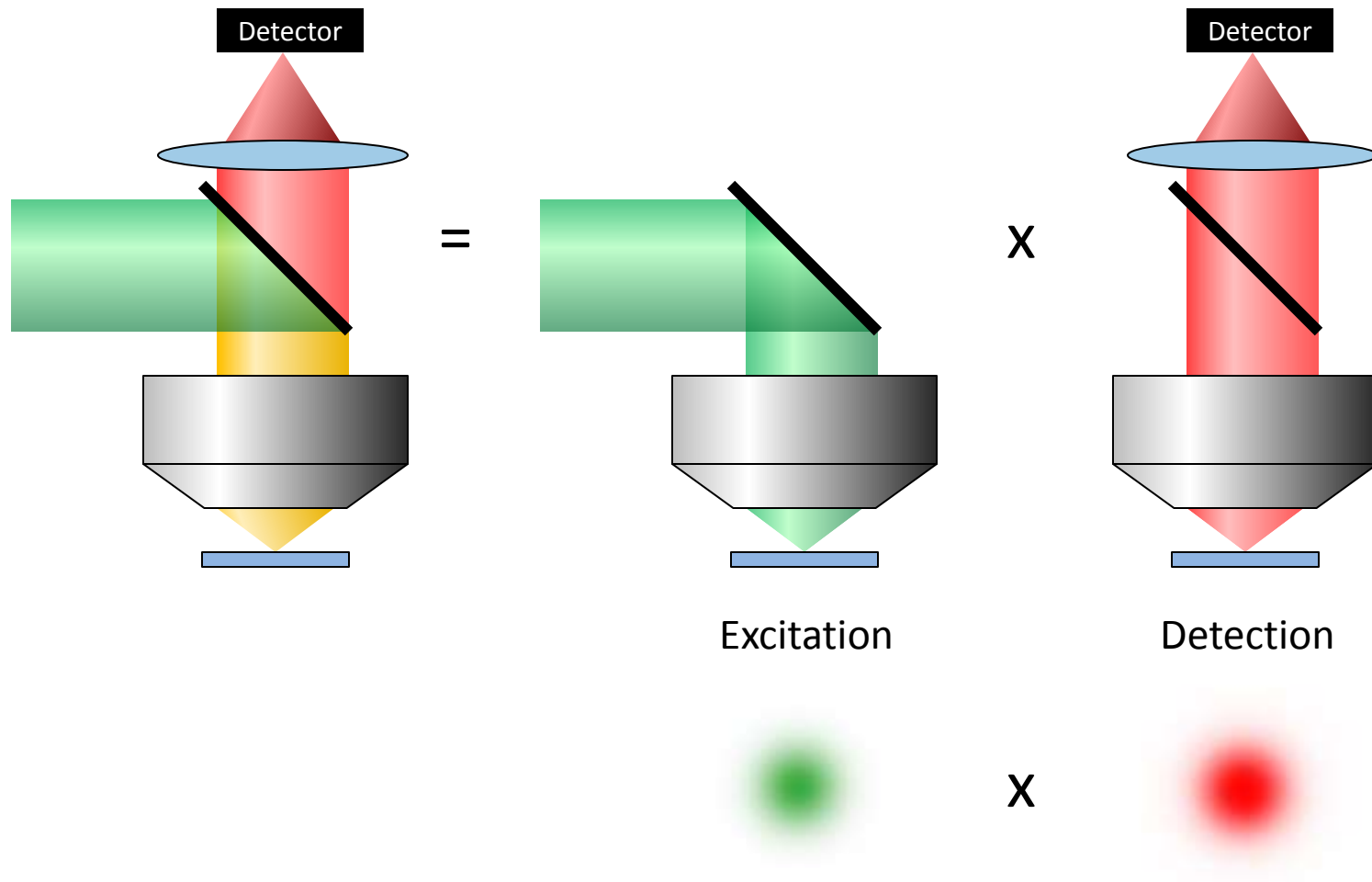
$$d \approx \frac{\lambda}{2 NA}$$



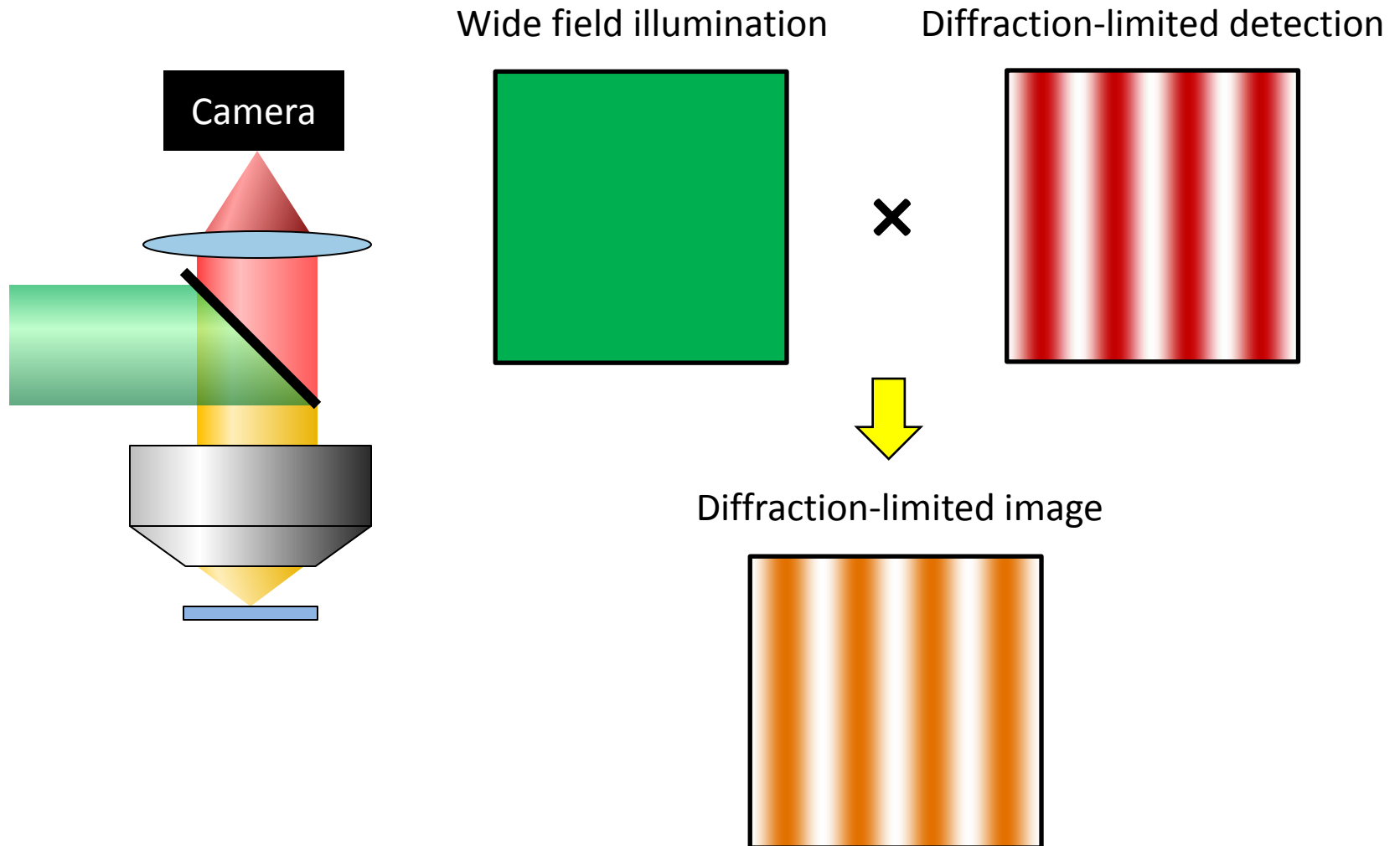
Major advantage:  
Similar z resolution as x-y resolution



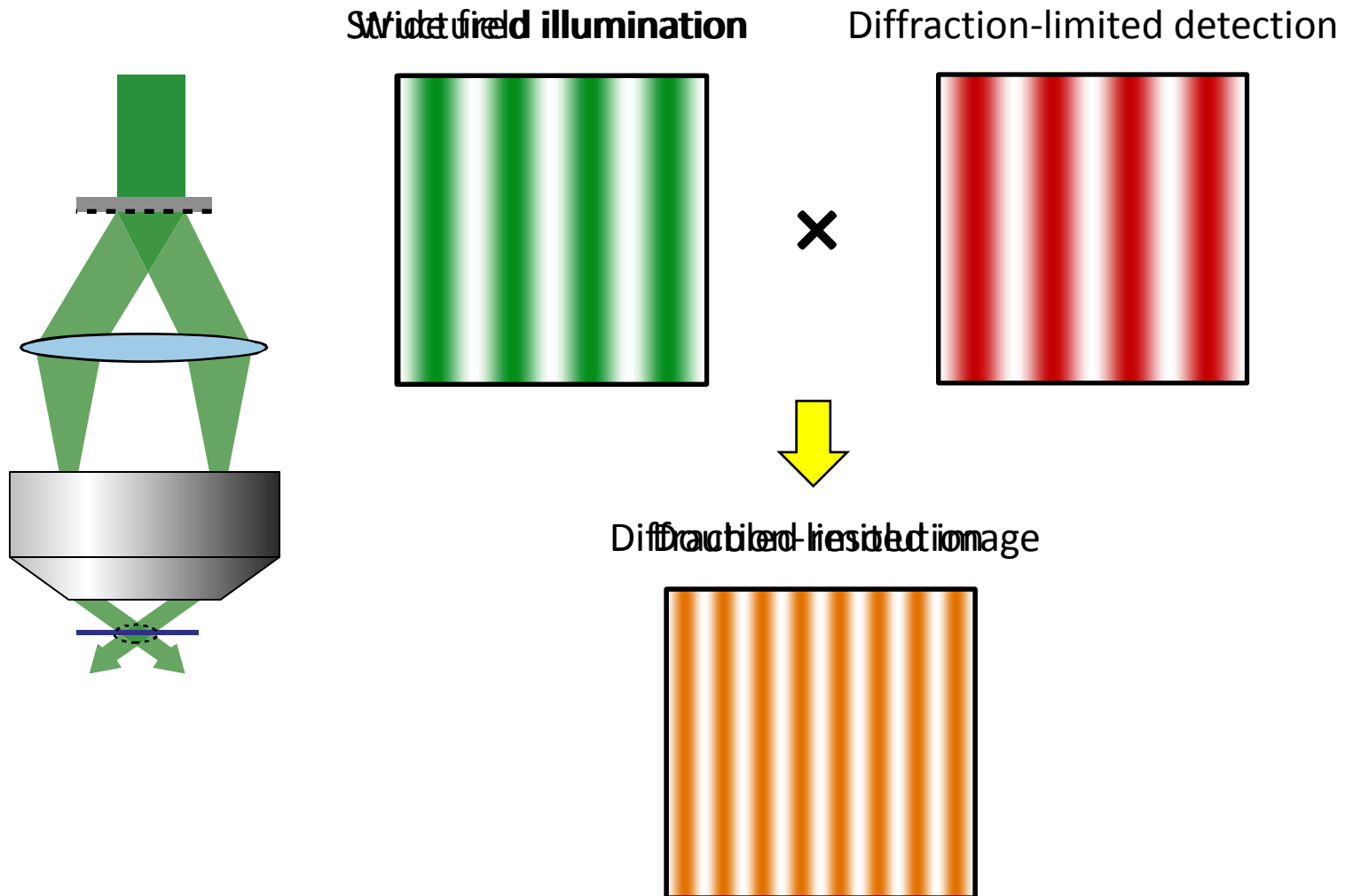
# Patterned illumination



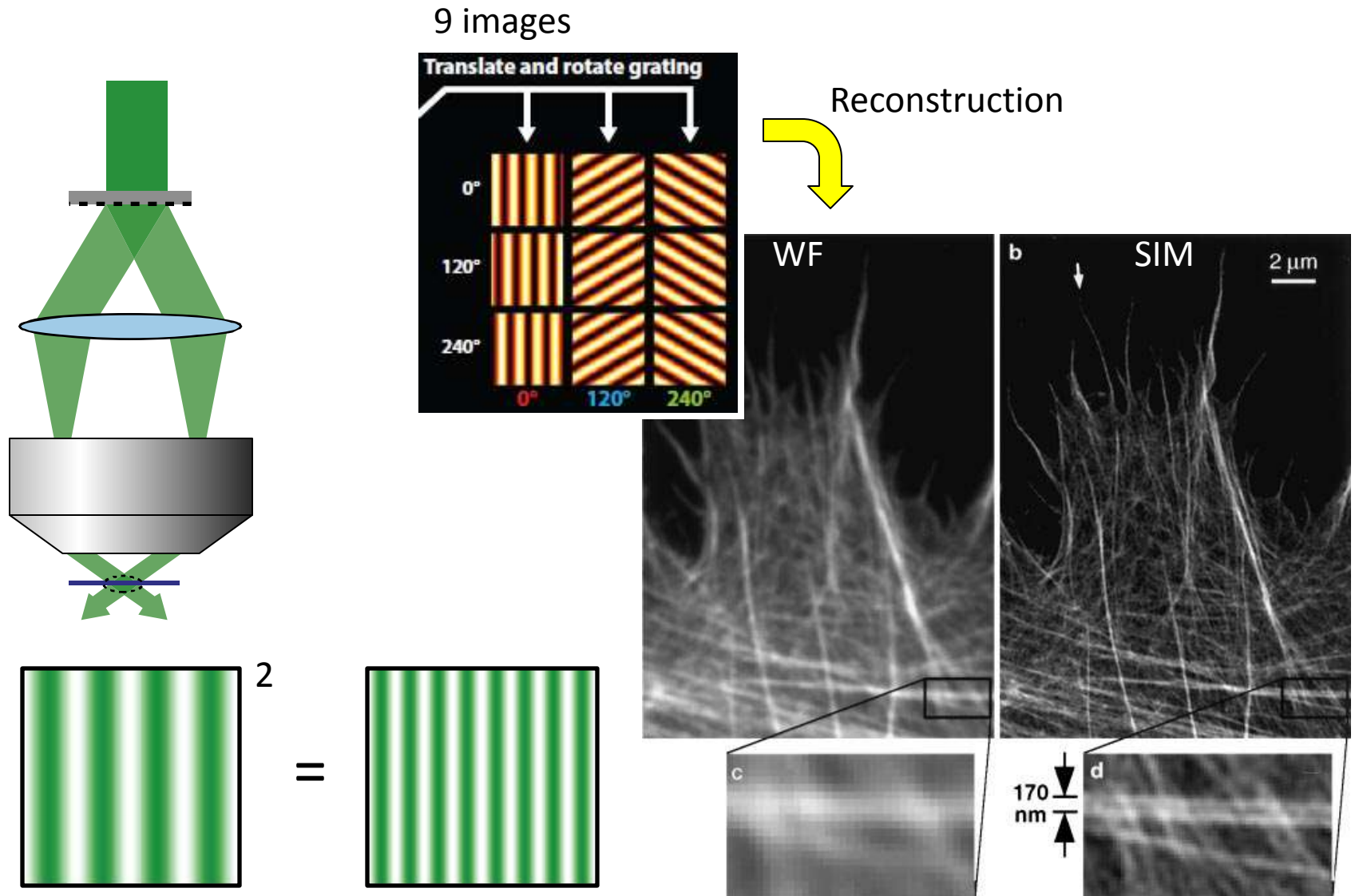
# Structured Illumination Microscopy (SIM)



# Structured Illumination Microscopy (SIM)



# Structured Illumination Microscopy (SIM)

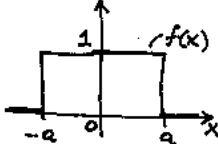


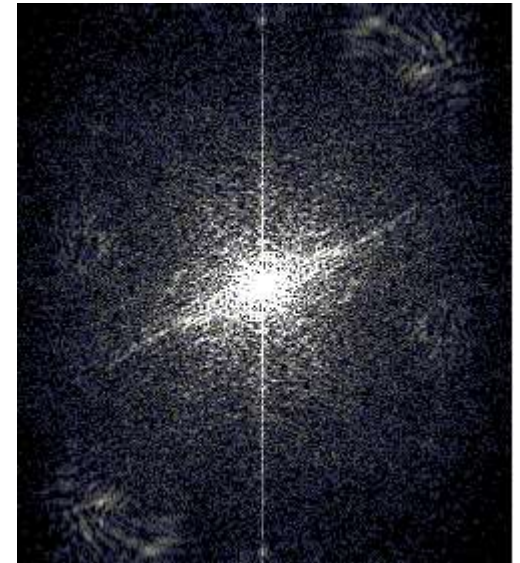
# Being (slightly) more rigorous about SIM



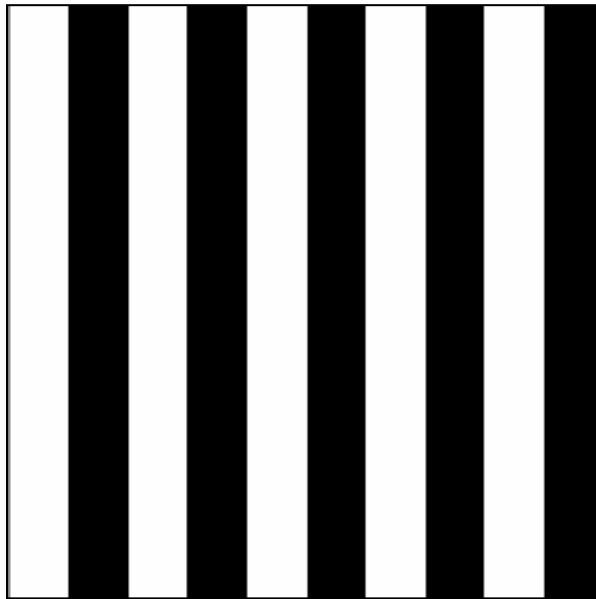
## Fourier transforms: Examples

- $f(x) = \delta(x) \quad \rightarrow \quad \tilde{f}(k) = 1$
- $f(x) = 1 \quad \rightarrow \quad \tilde{f}(k) = 2\pi \delta(k)$
- $f(x) = e^{-a|x|} \quad \rightarrow \quad \tilde{f}(k) = \frac{2a}{k^2 + a^2}$
- $f(x) = \frac{1}{x^2 + a^2} \quad \rightarrow \quad \tilde{f}(k) = \frac{\pi}{a} e^{-a|k|}$
- $f(x) = e^{-a\frac{x^2}{2}} \quad \rightarrow \quad \tilde{f}(k) = \sqrt{\frac{2\pi}{a}} e^{-\frac{1}{a}\frac{k^2}{2}}$

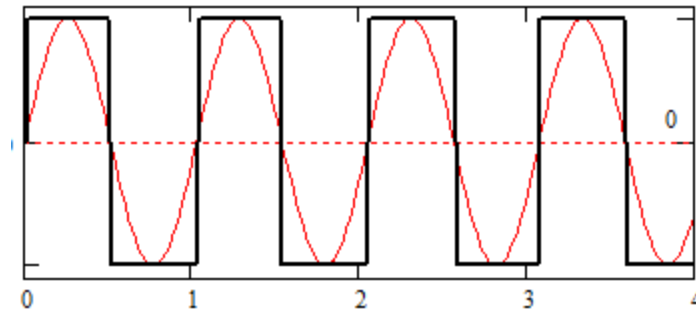
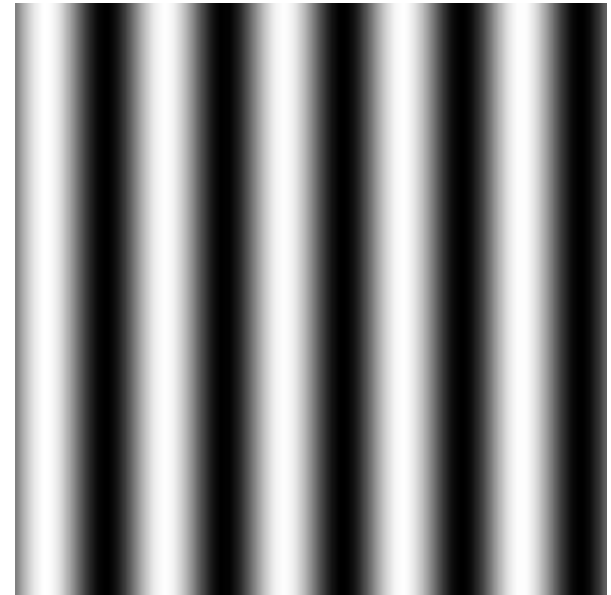
-   $\rightarrow \quad \tilde{f}(k) = 2 \frac{\sin(ak)}{k}$



# Fourier transform and spatial frequencies

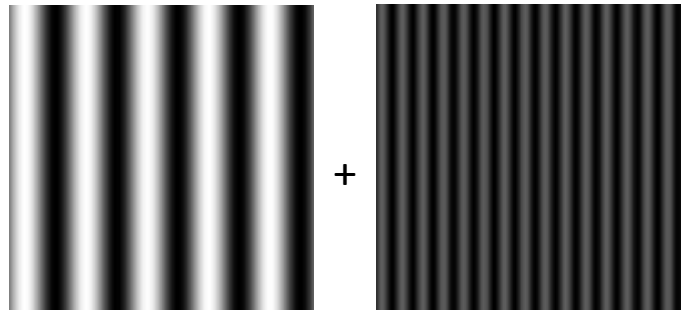


$\hat{=}$   
?

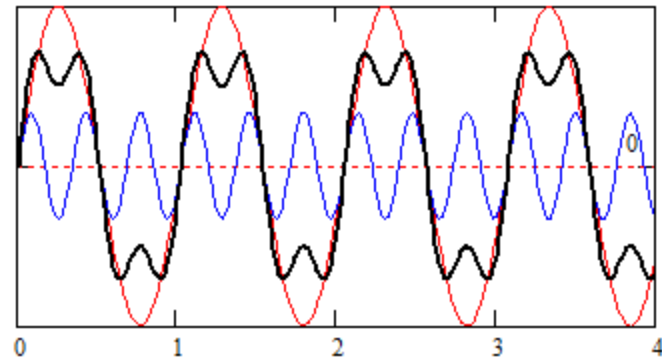
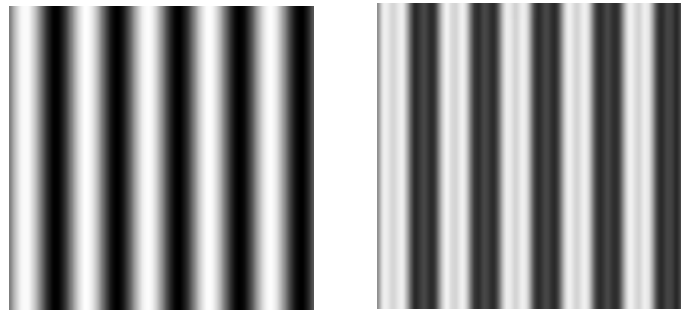




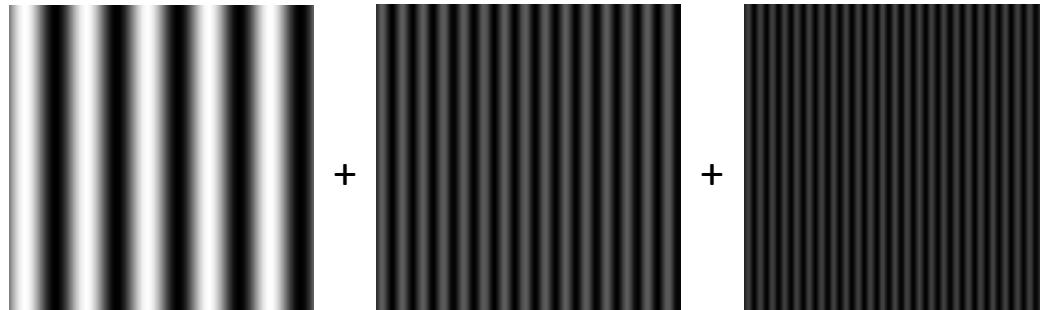
# Fourier transform and spatial frequencies



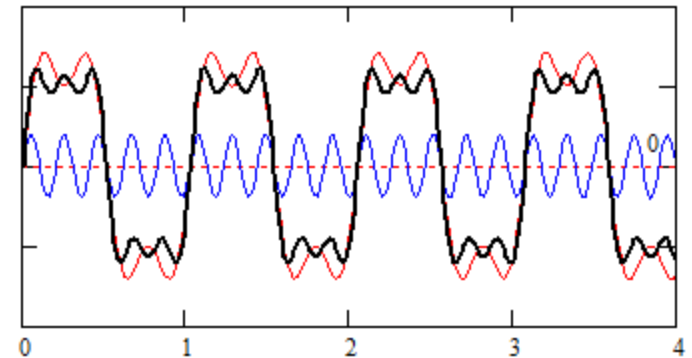
Summed image



# Fourier transform and spatial frequencies

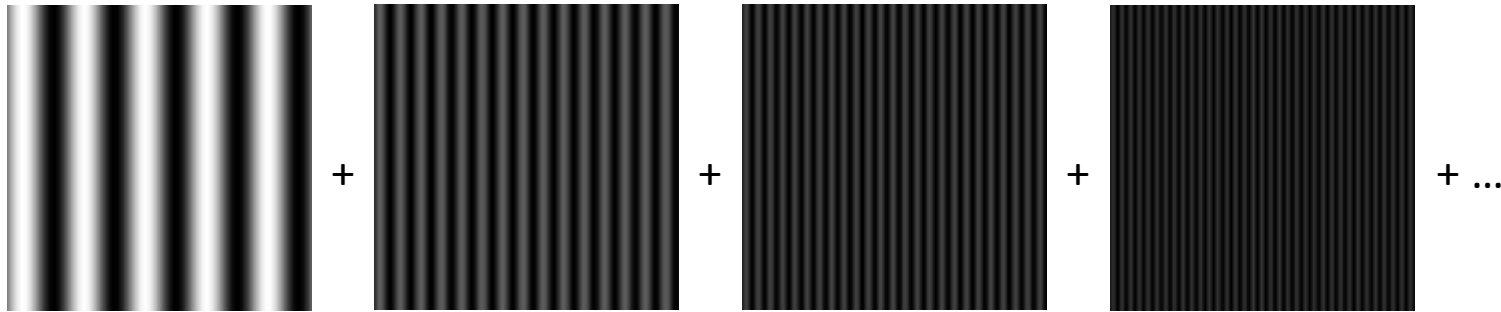


Summed image



# Fourier transform and spatial frequencies

Discrete spatial frequencies



Summed image

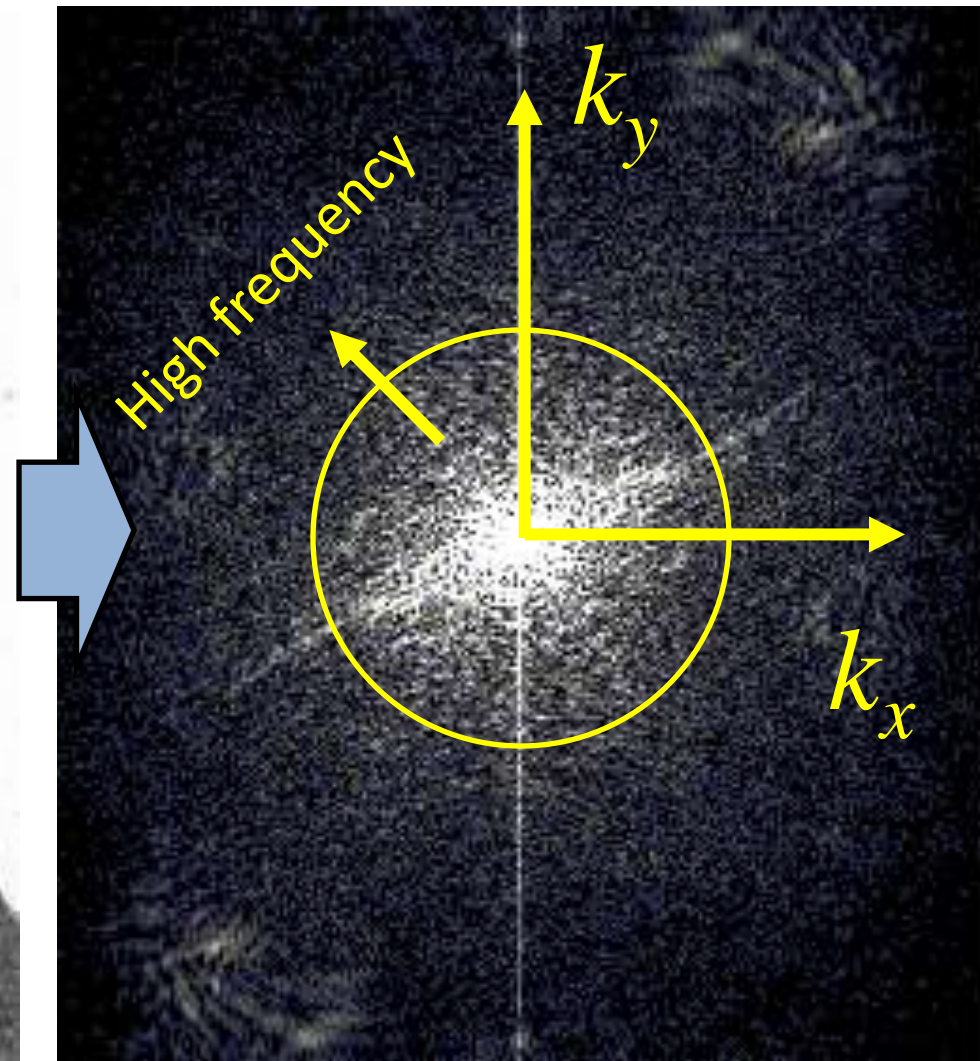


$$G(x) = \sum F(k) \sin(k x)$$

# Fourier transform and spatial frequencies

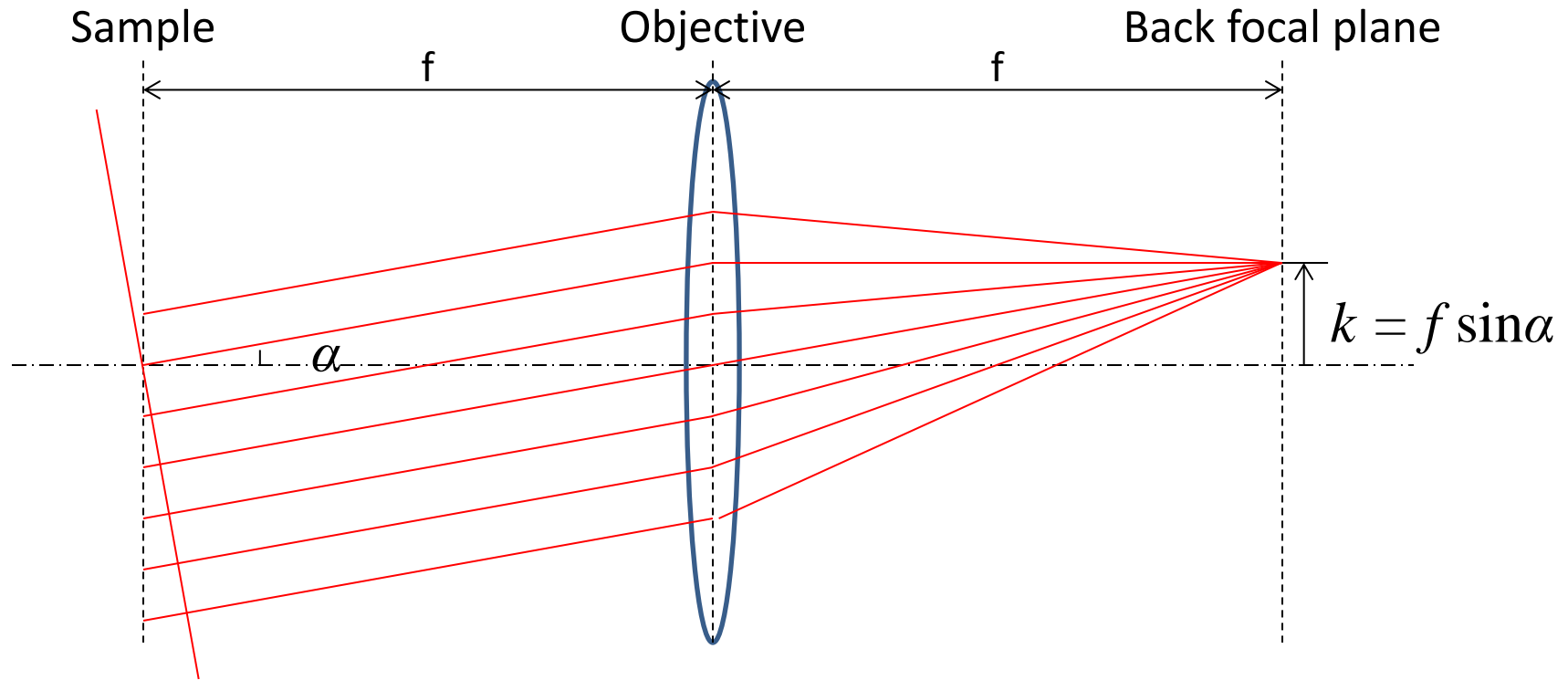


Original Image (real space)

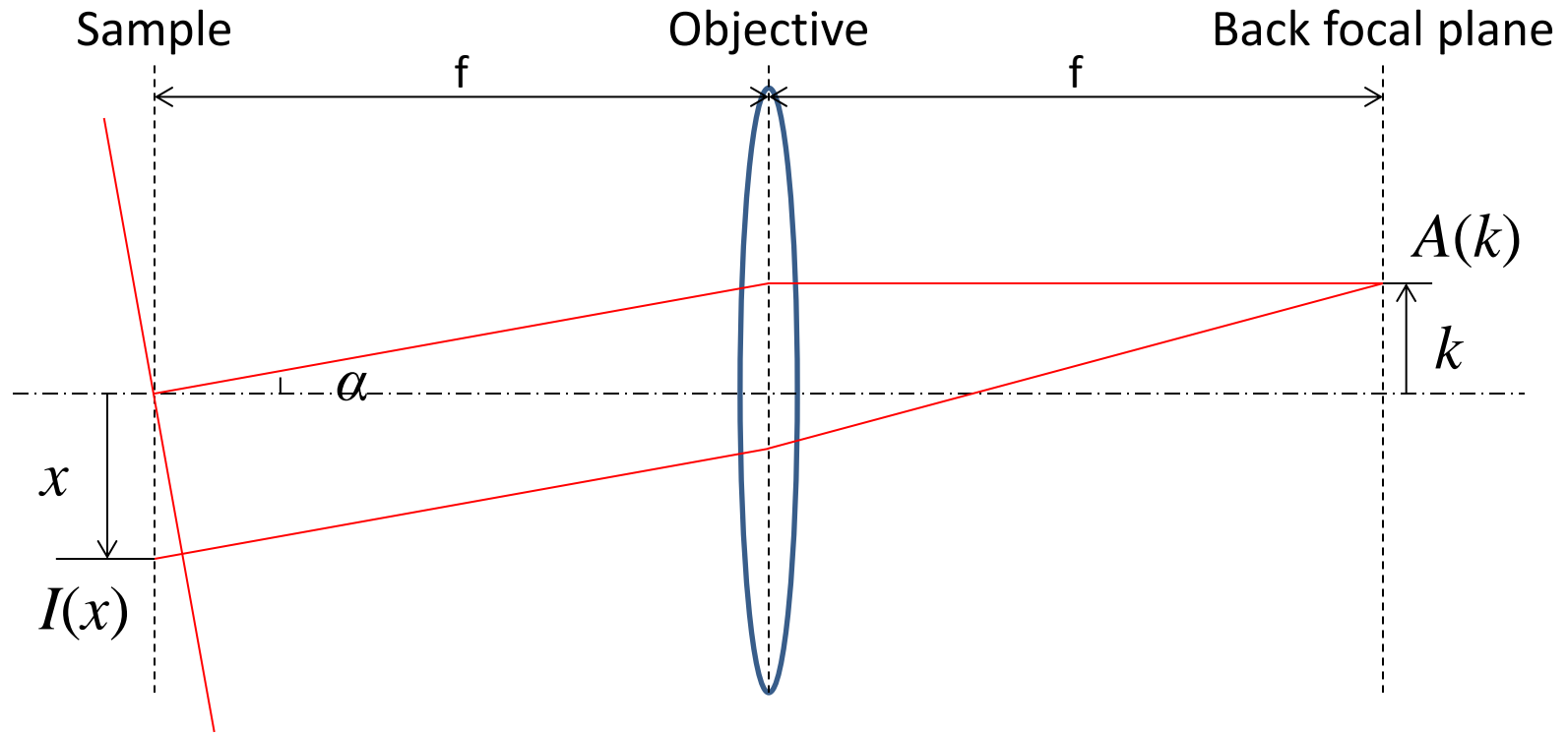


Fourier transform (frequency space)

# Fourier optics and microscope resolution



# Fourier optics and microscope resolution



Phase delay from the mid-point

$$\Delta\varphi = x \sin\alpha (2\pi/\lambda) = x (k / f) (2\pi/\lambda) \text{ assuming refractive index} = 1$$

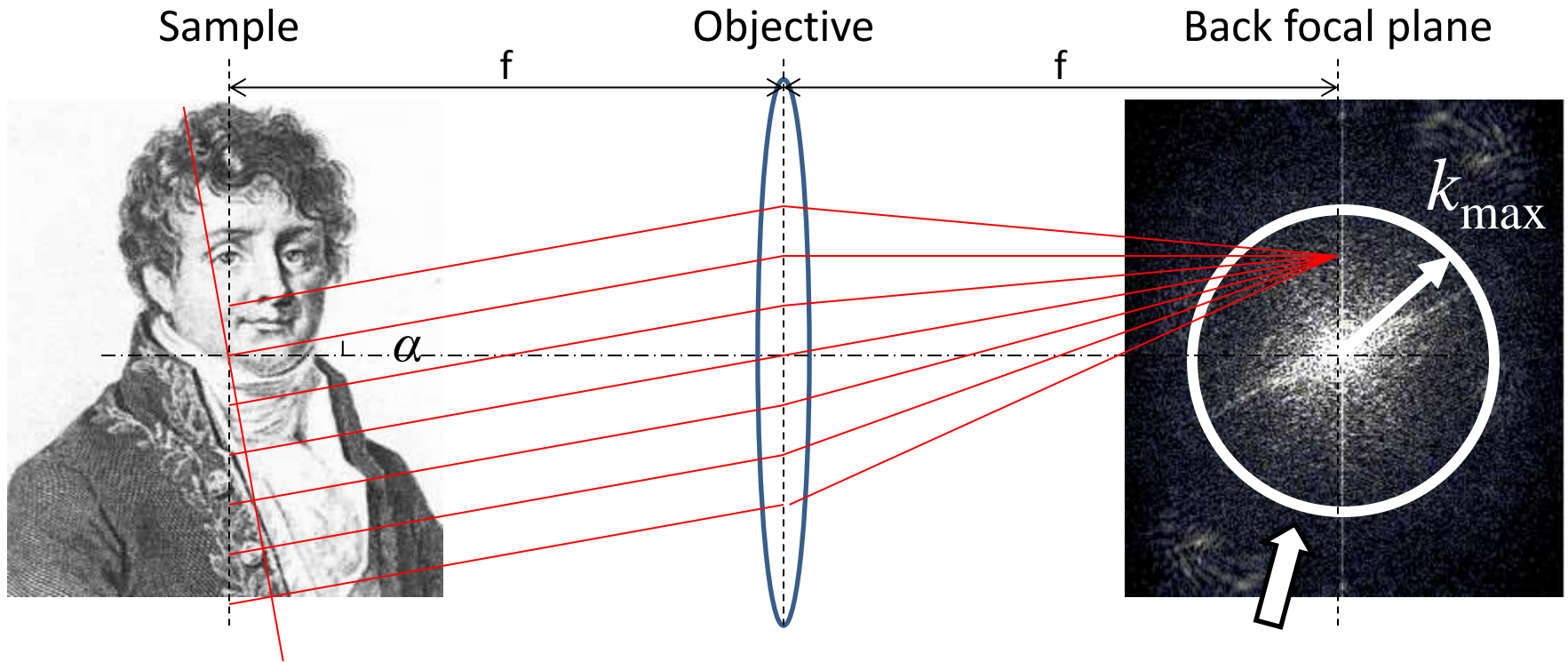
Light intensity at the sample plane

**Fourier Transform!**

$$I(x) = \sum A(k) \sin(\Delta\varphi) = \sum A(k) \sin(x k 2\pi/\lambda f)$$



# Fourier optics and microscope resolution



$$\text{Spatial frequency} = k \cdot 2\pi/\lambda f$$

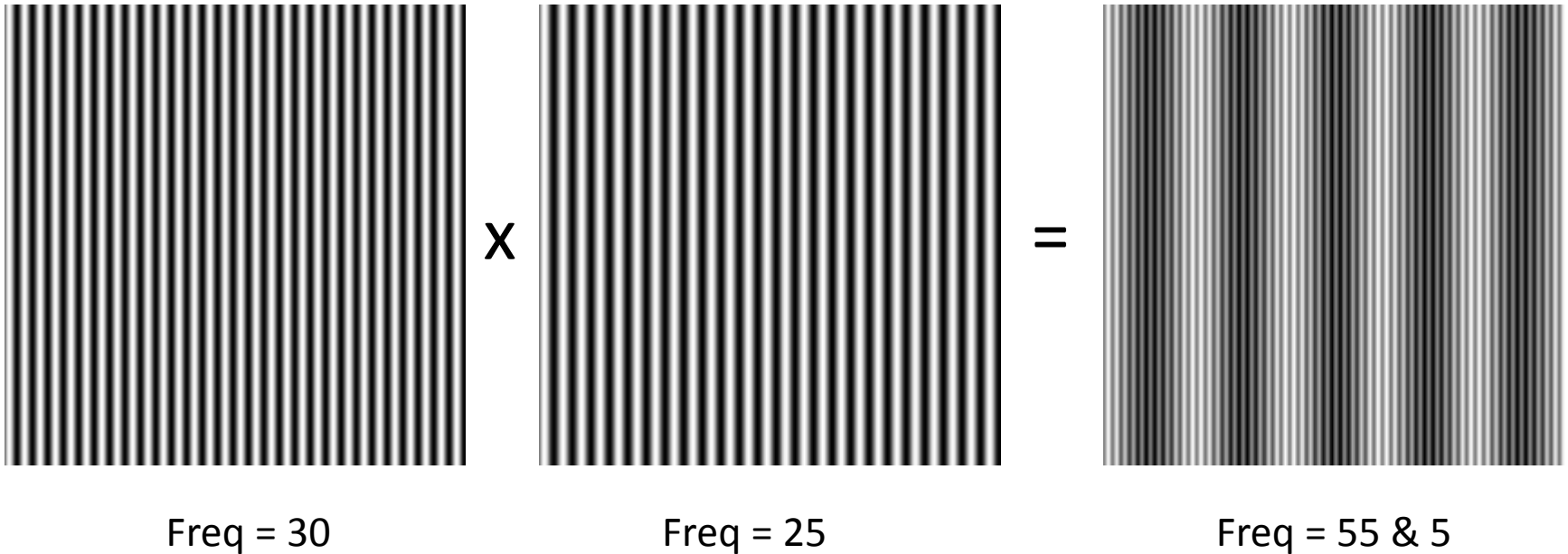
$$k_{\text{max}} = f \sin \alpha_{\text{max}} = f \cdot NA$$

$$\text{Resolution} = \lambda / 2NA$$

Size of the back focal plane

# Extending the measurable freq. range

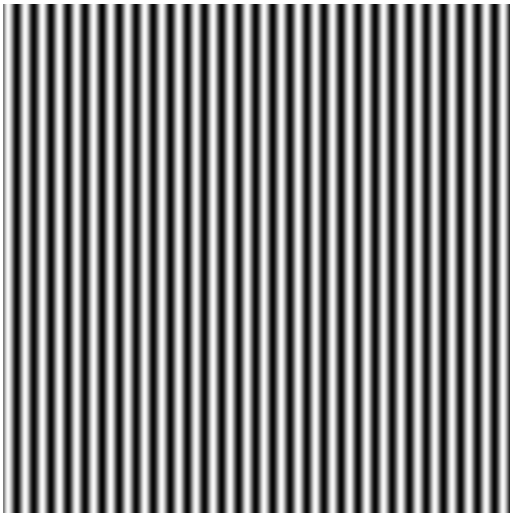
$$\text{Excitation}(x) \times \text{Sample}(x) = \text{Observed Signal}(x)$$



$$\sin A \cdot \sin B = (\cos(A - B) - \cos(A + B)) / 2$$

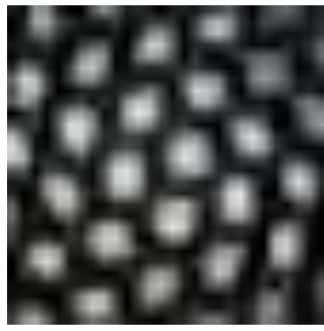
# Extending the measurable freq. range

$$\text{Excitation}(x) \times \text{Sample}(x) = \text{Observed Signal}(x)$$



Freq = 30

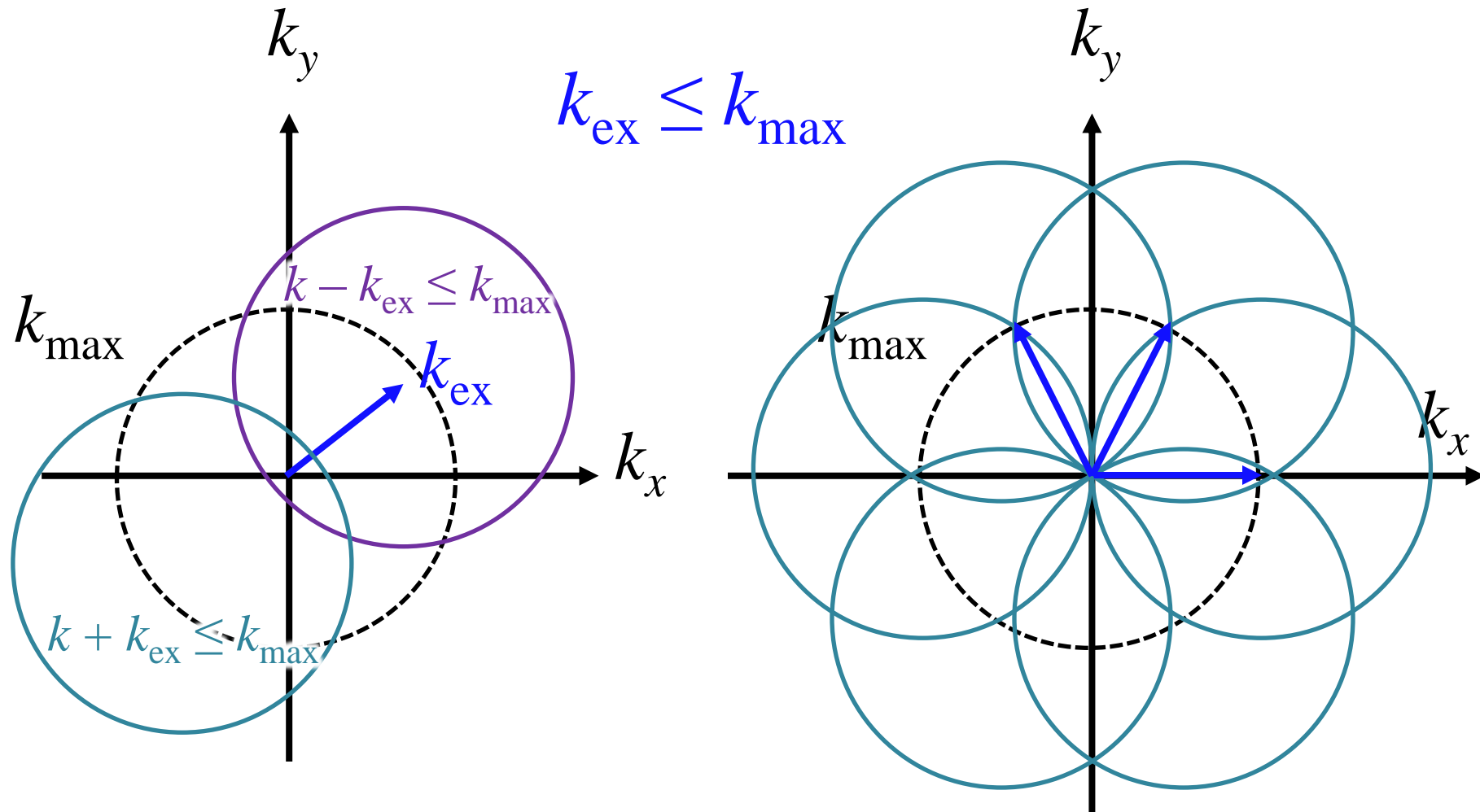
x



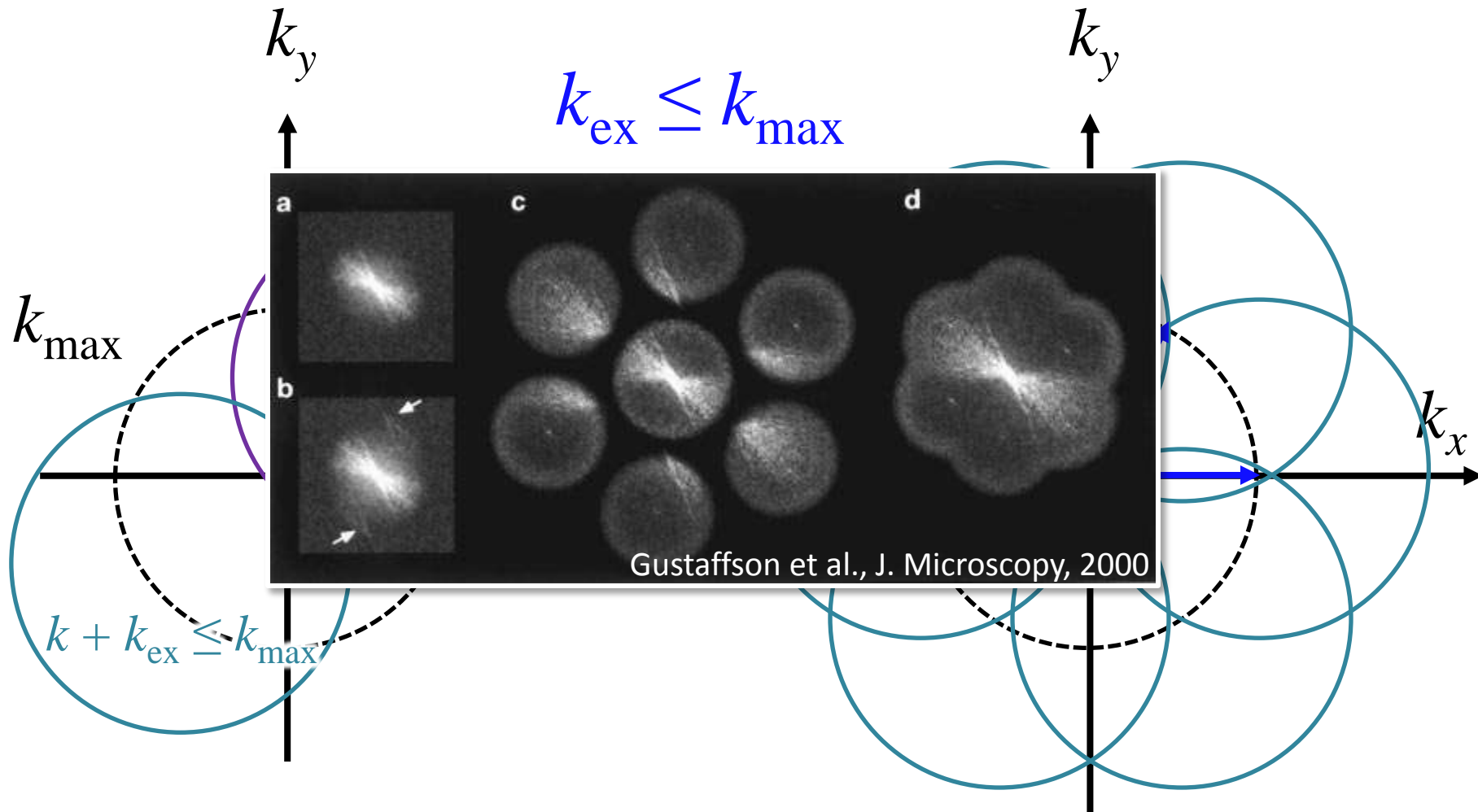
$\sin A \cdot \sin$



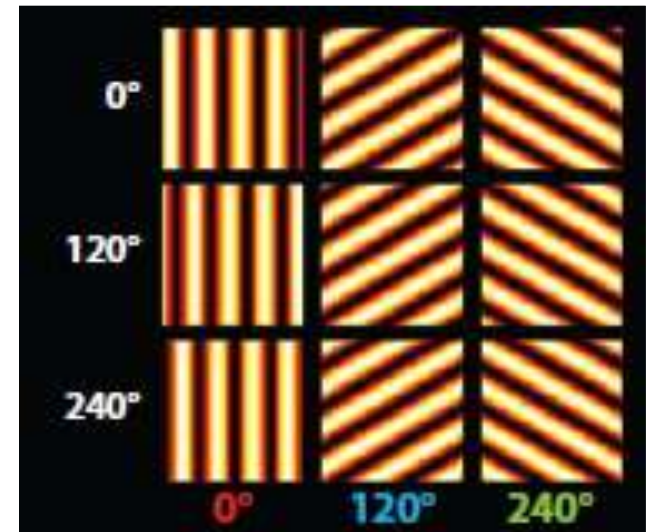
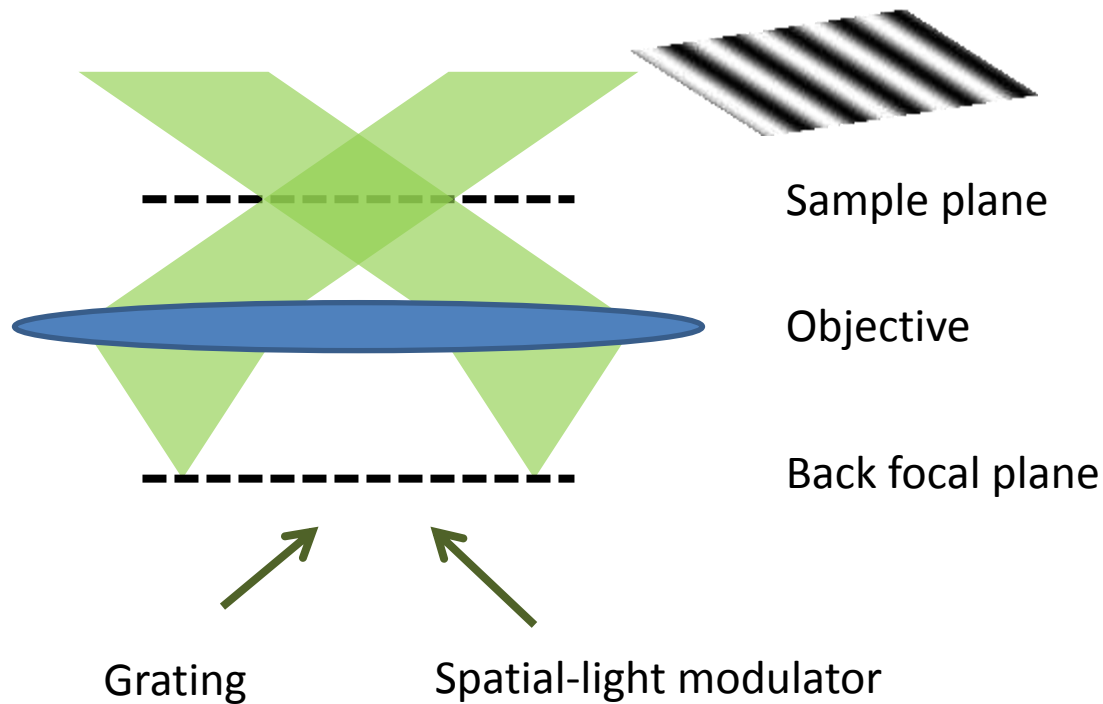
# Extending the measurable freq. range



# Extending the measurable freq. range

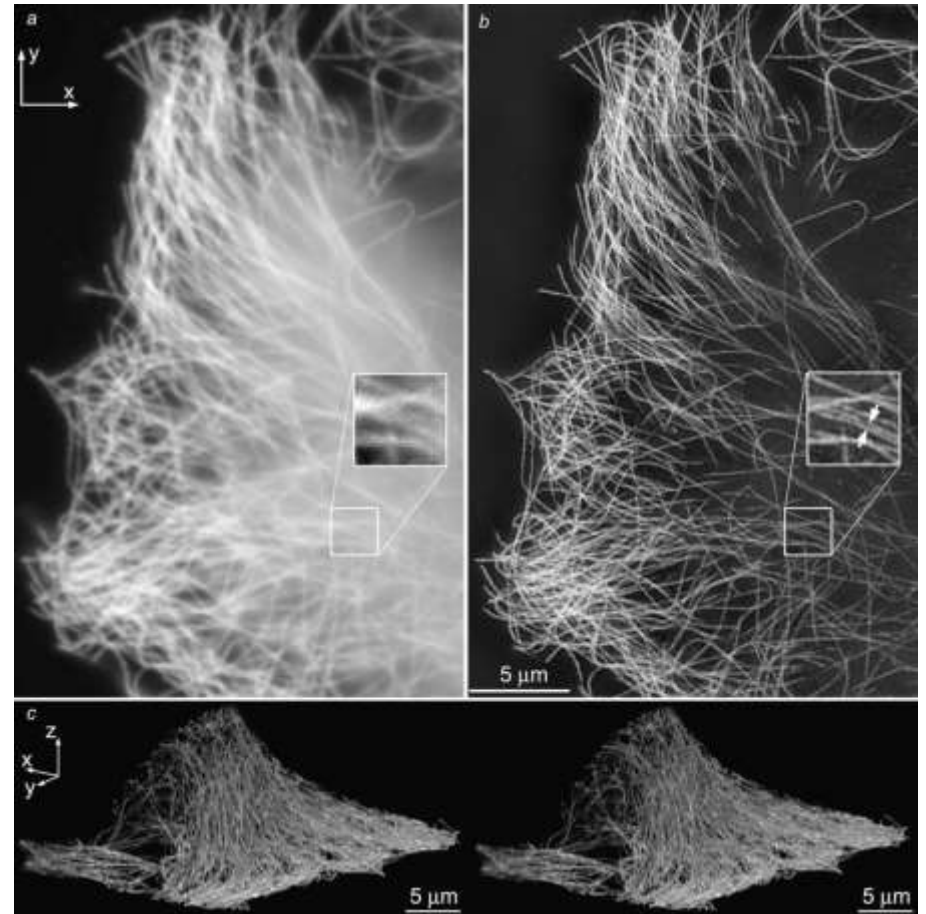
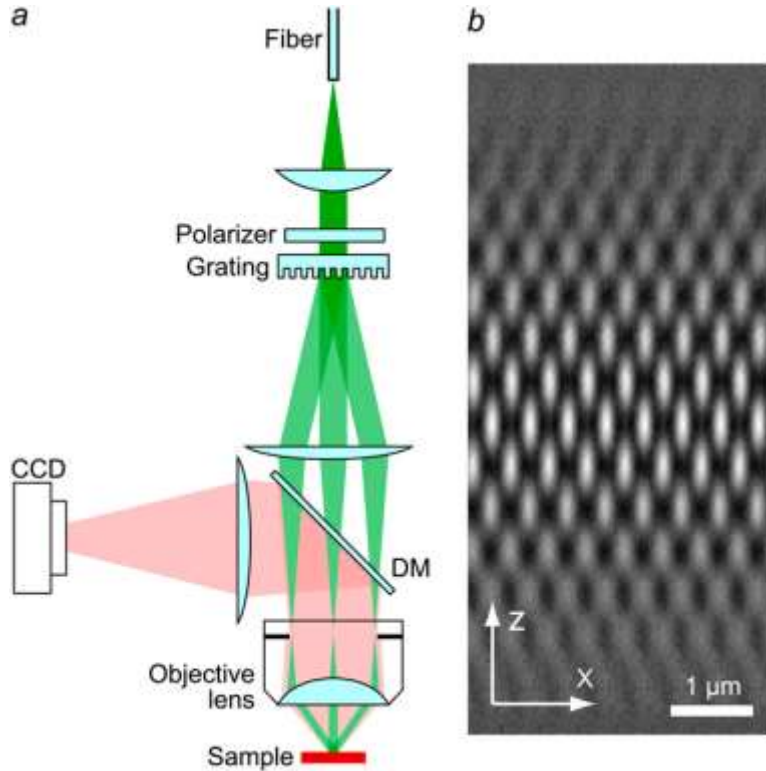


# Generating the illumination pattern



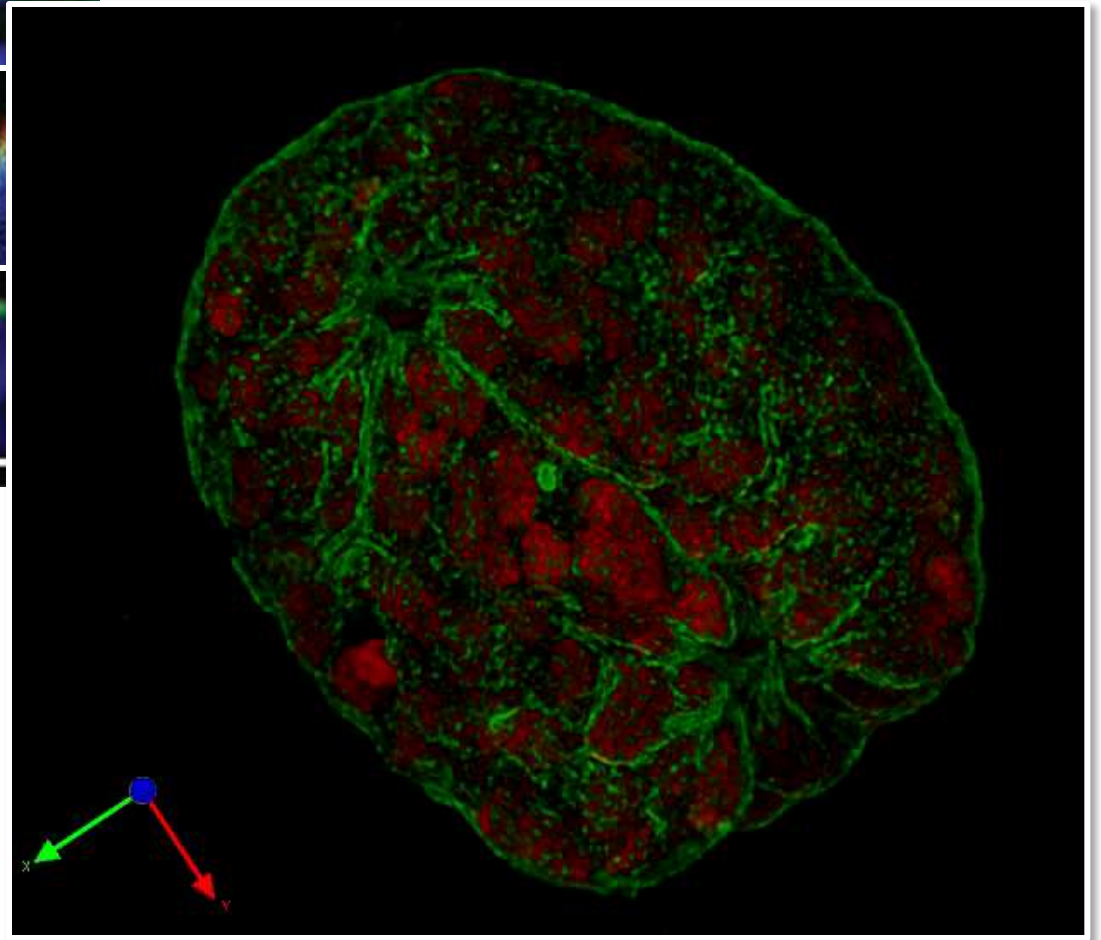
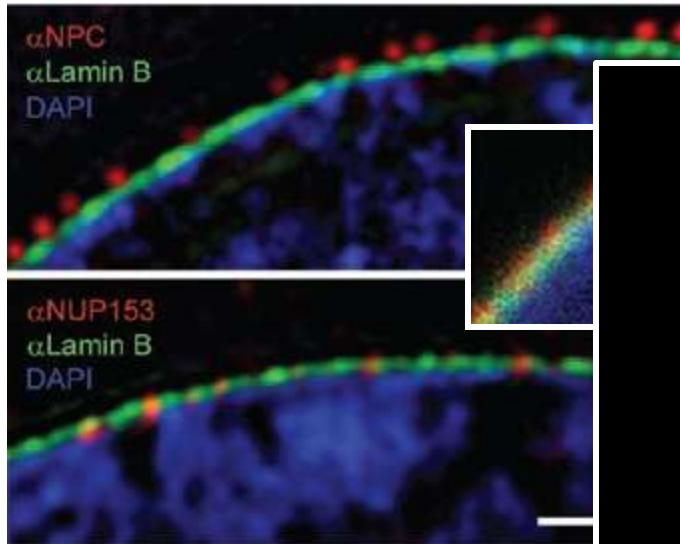


# 3D SIM: better resolution + optical sectioning

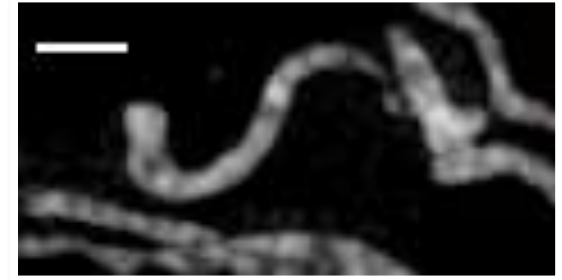
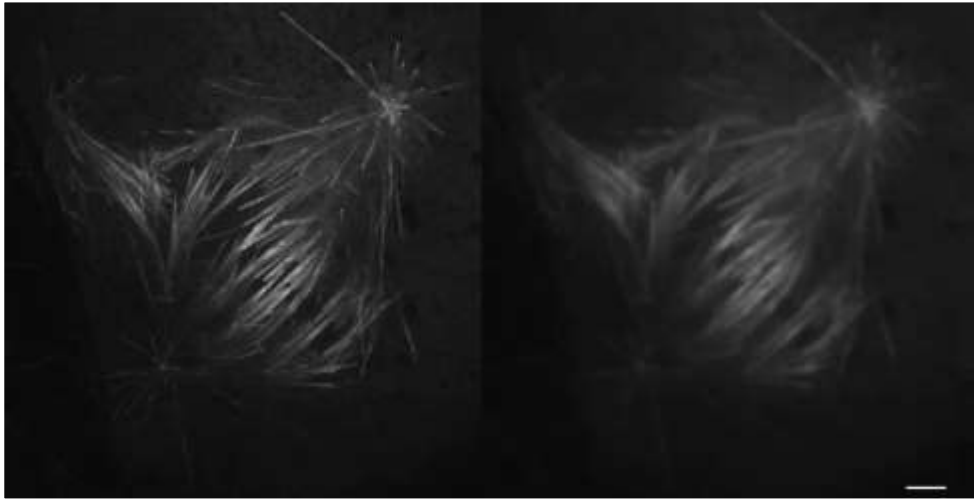


# Multicolor SIM

Same as conventional fluorescence microscopy!



# Live imaging with SIM

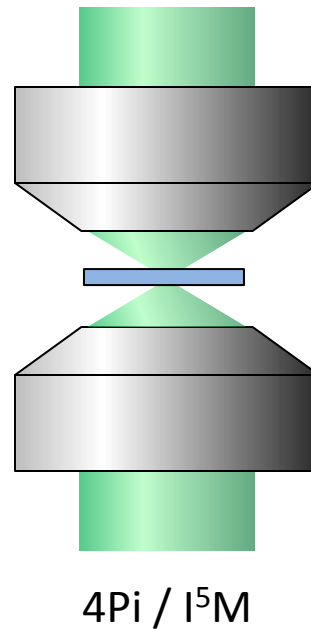
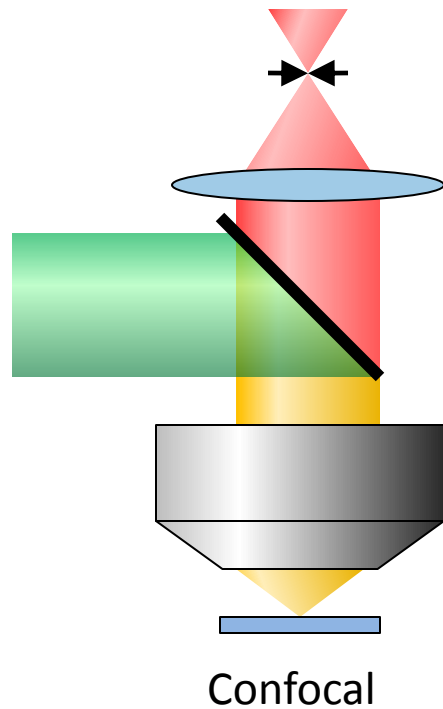


Kner, Chhun et al., Nat Methods,

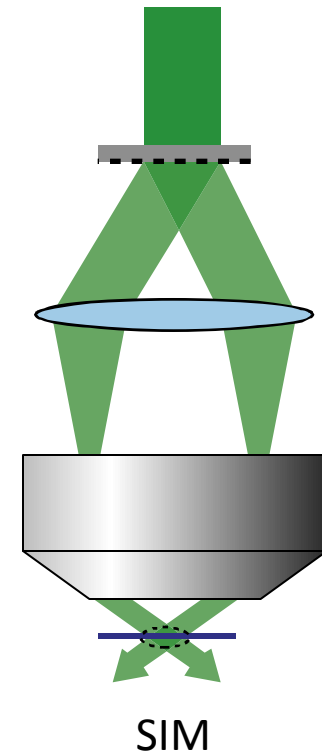
Shao et al., Nat Methods, 2011



# The diffraction limit still exists



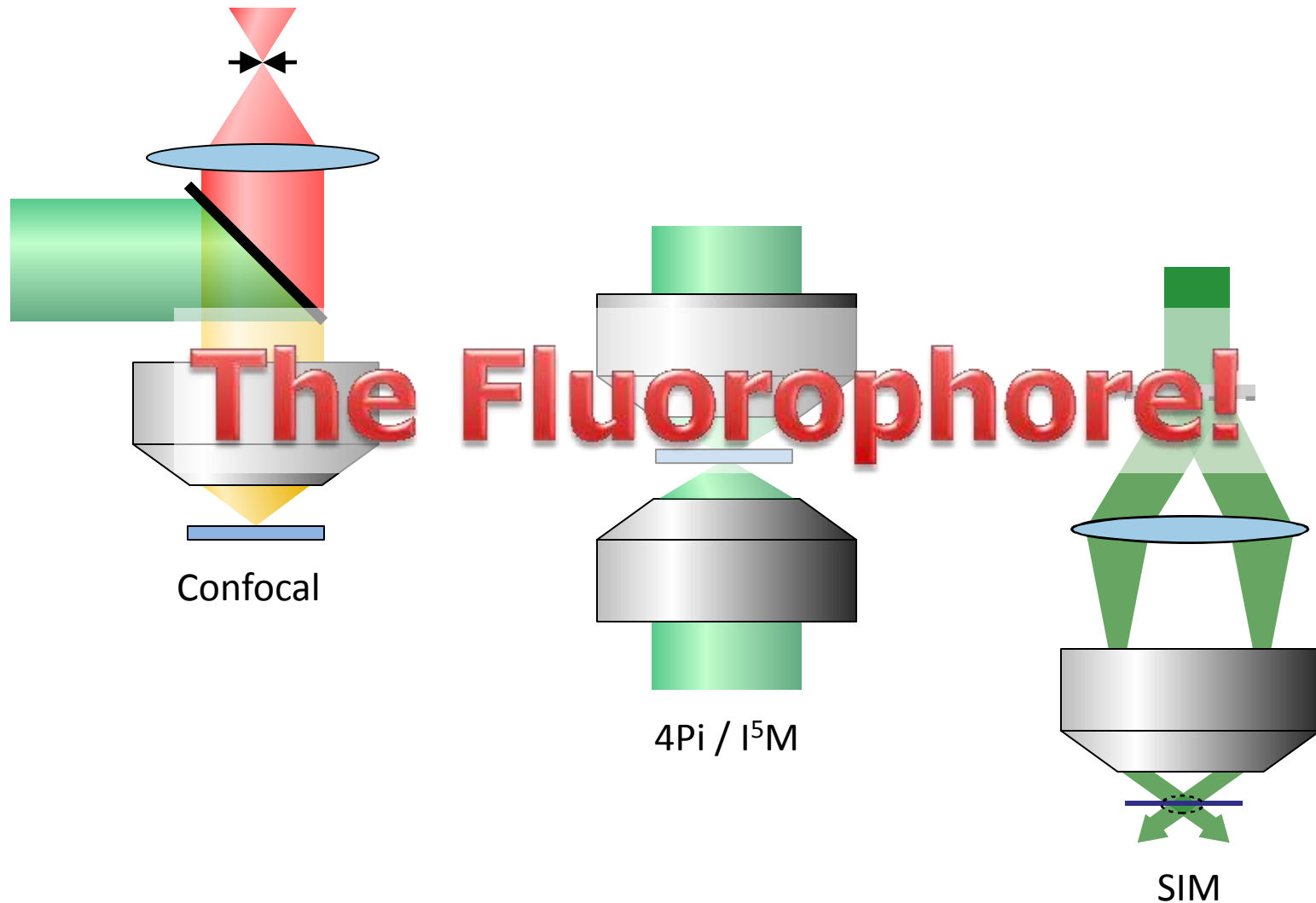
$$d \geq \frac{1}{2} \cdot \frac{\lambda}{2NA}$$



# Breaking the diffraction barrier

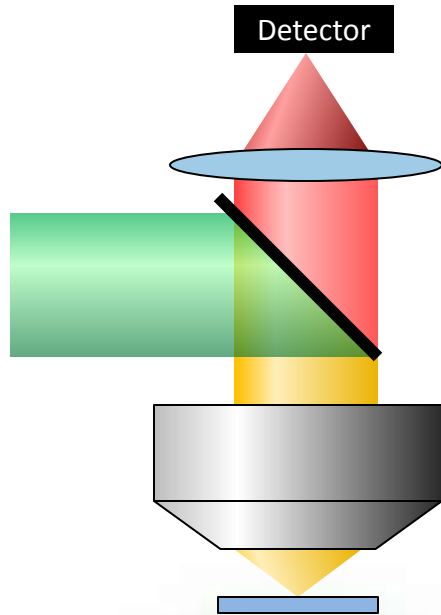


# Breaking the diffraction barrier

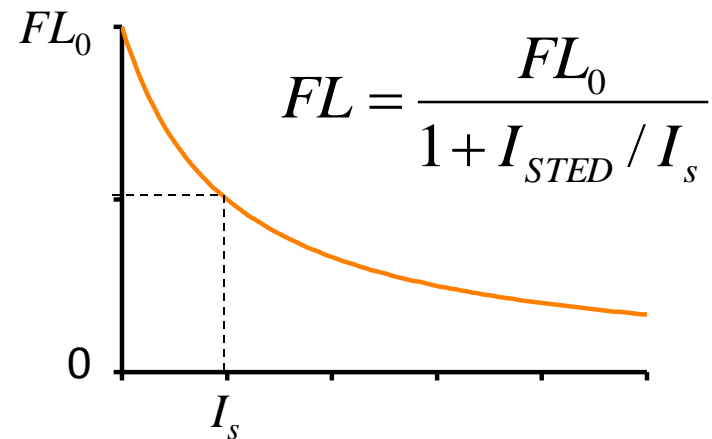
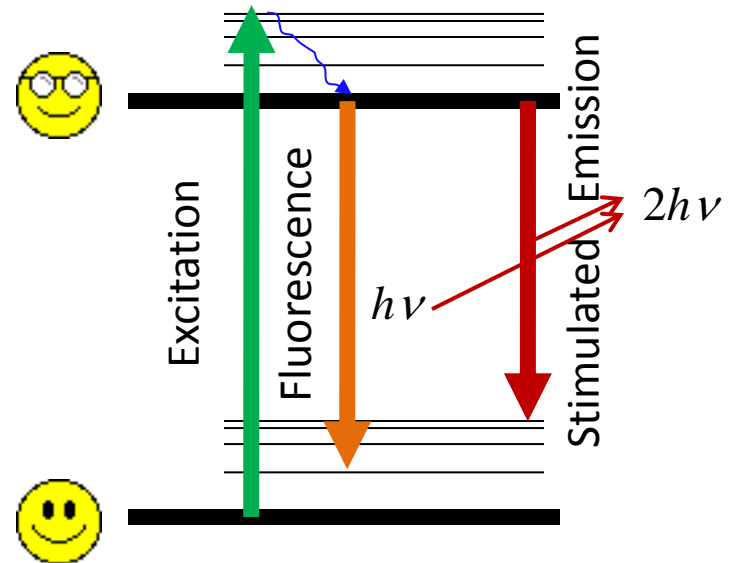




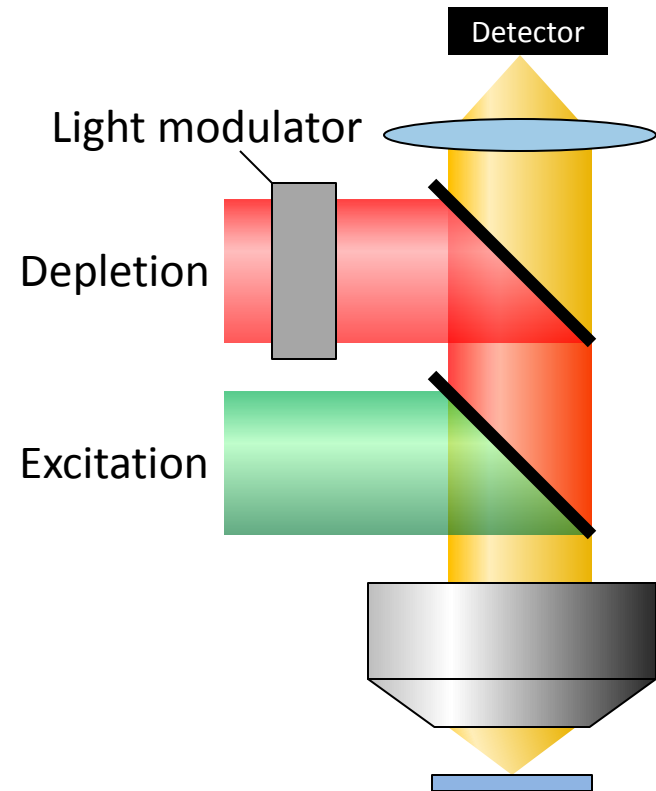
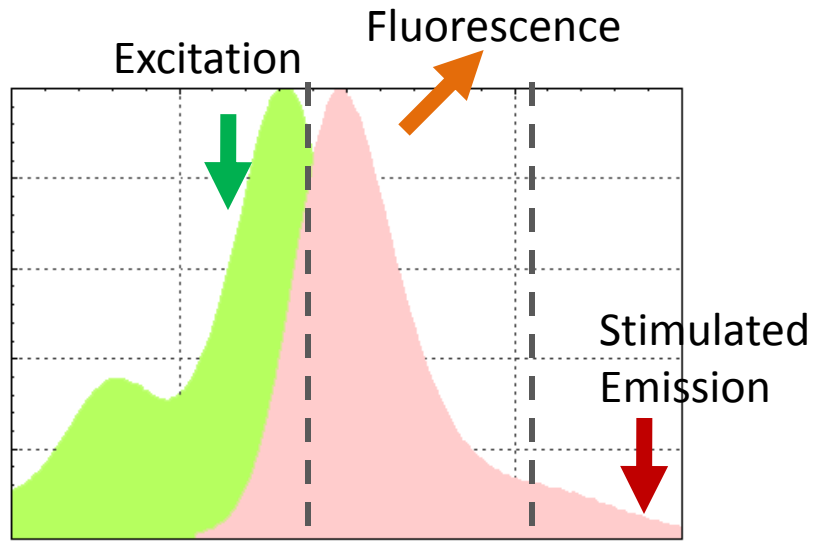
# Stimulated Emission Depletion (STED)



Send to a dark state



# STED microscopy

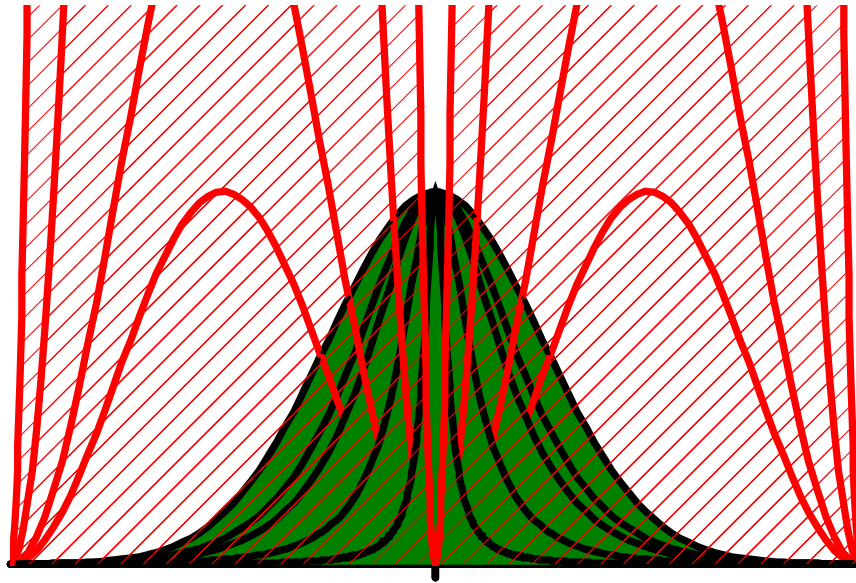


Excitation      STED pattern      Effective PSF

$$\text{Excitation} \div \text{STED pattern} = \text{Effective PSF} \quad ?$$

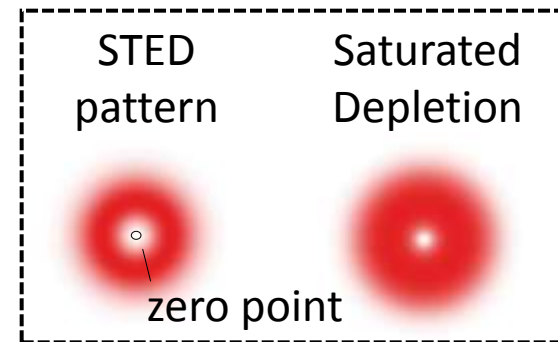
The diagram illustrates the mathematical relationship between the excitation and STED beams to form the effective point spread function (PSF). It shows a green circular spot (Excitation) divided by a red circular spot with a central white spot (STED pattern), resulting in a smaller green circular spot (Effective PSF) followed by a question mark.

# Saturated depletion

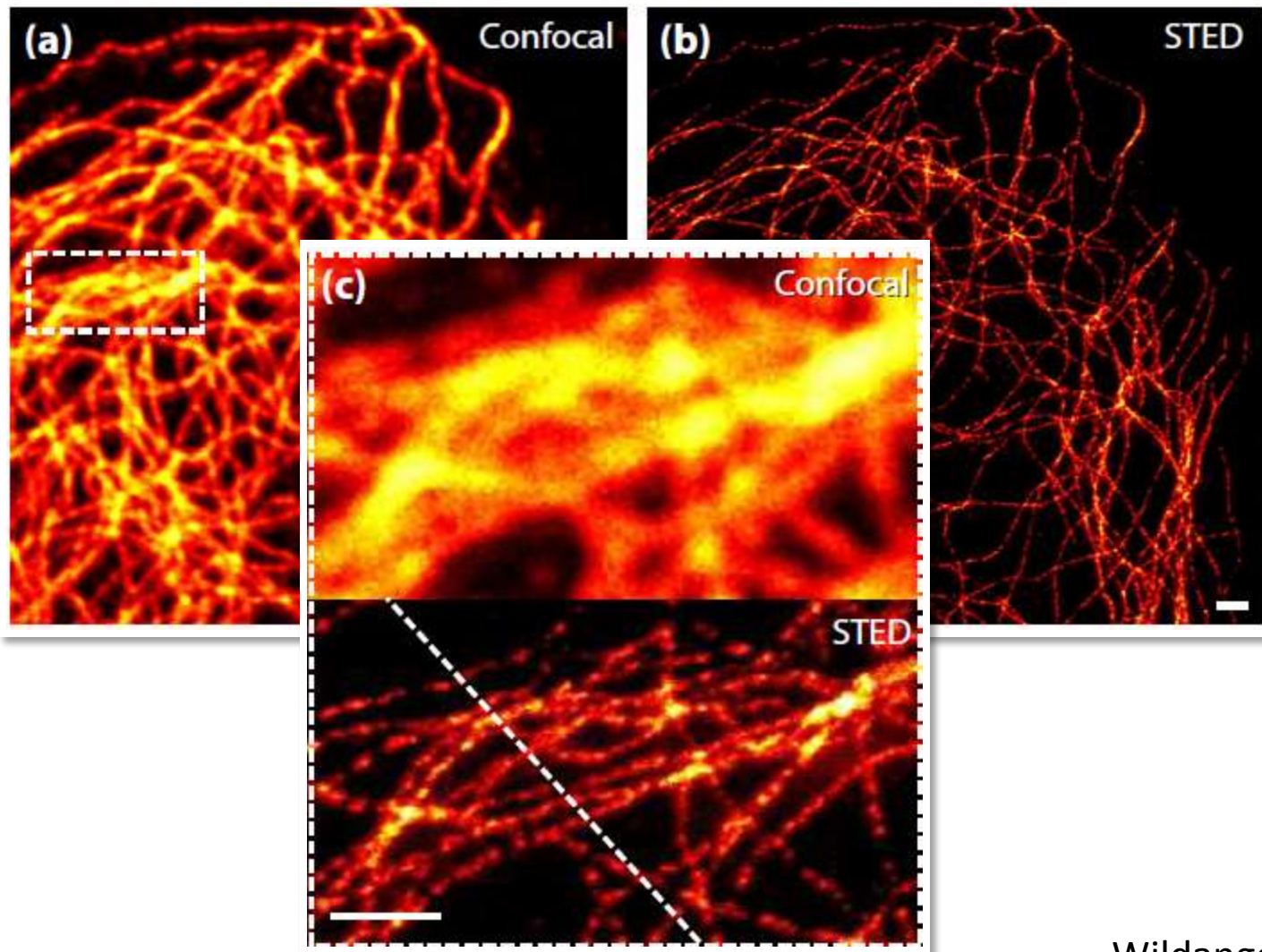


$$I_{\text{STED}} = 200 I_s$$

$$D = \frac{1}{\sqrt{1 + I/I_s}} \cdot \frac{\lambda}{2NA}$$

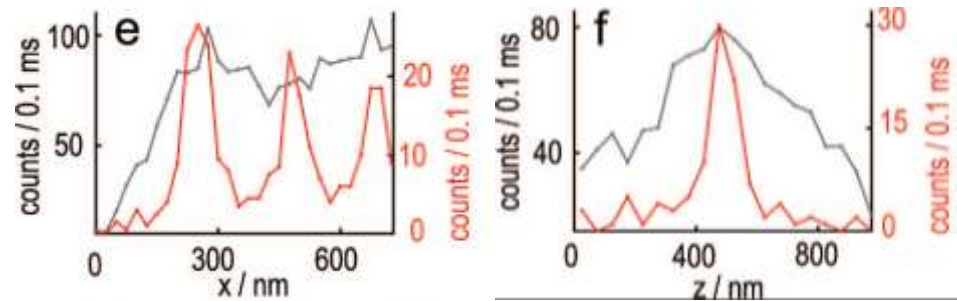
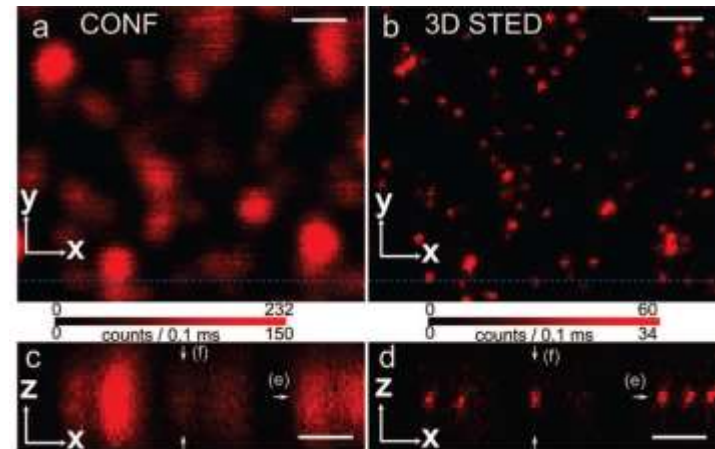
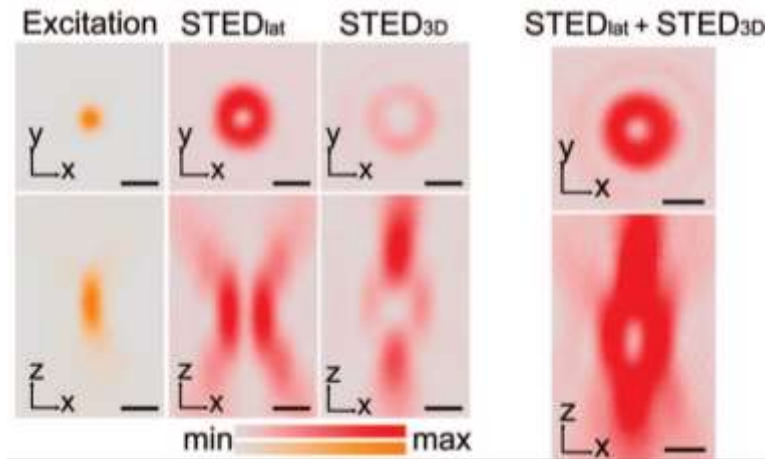


# STED images of microtubules

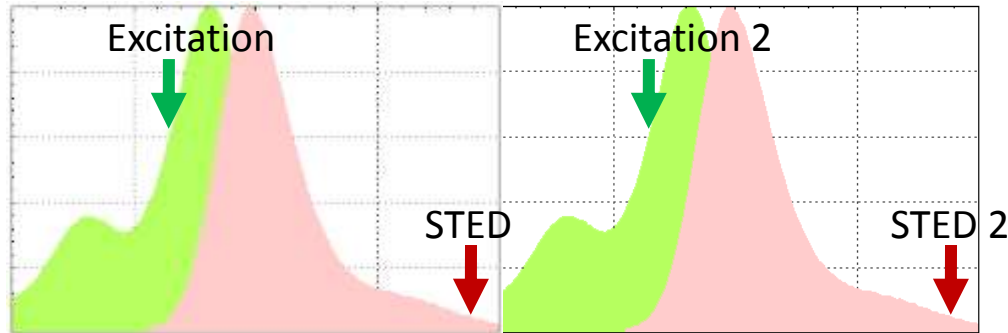


Wildanger et al., 2009

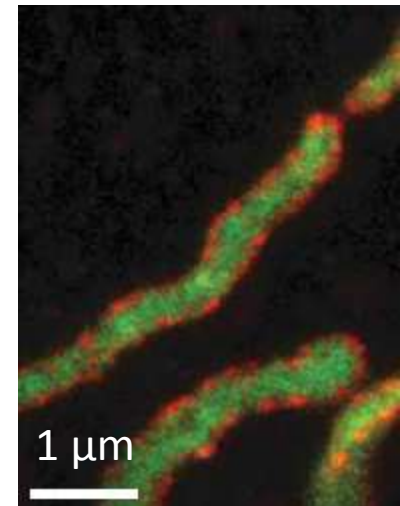
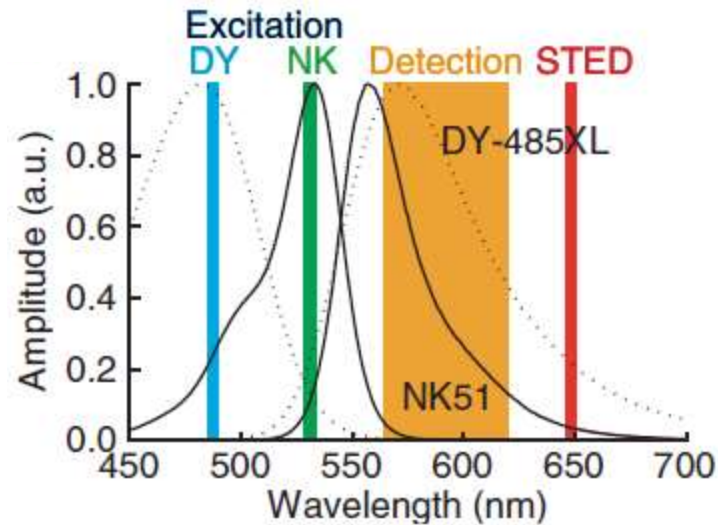
# 3D STED



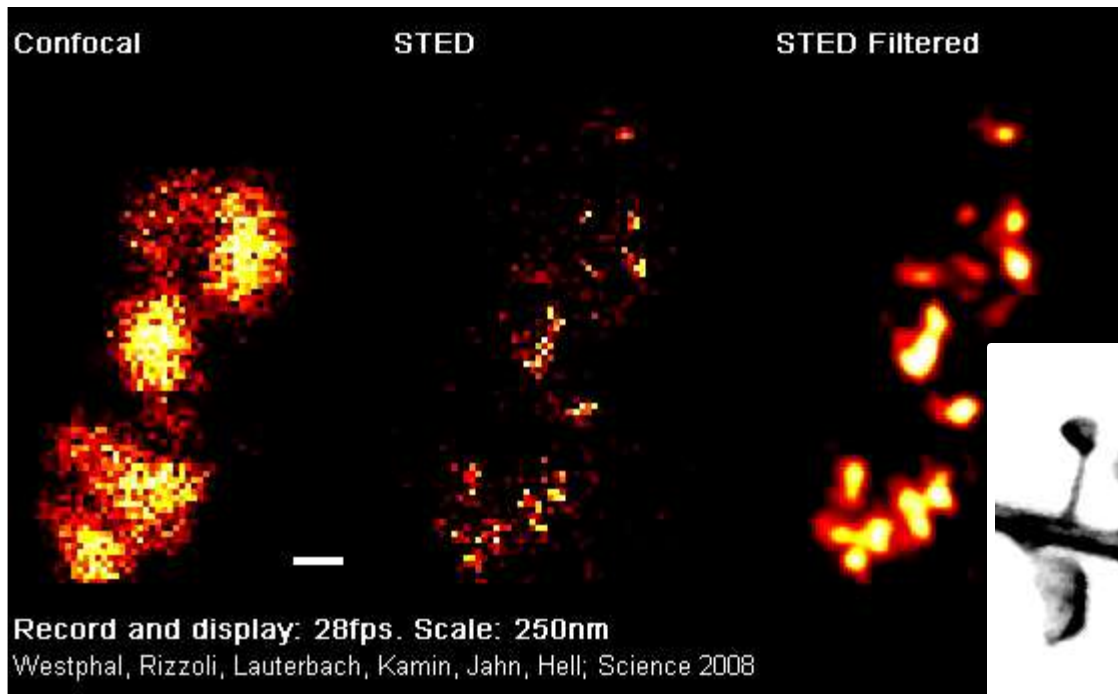
# Multicolor STED



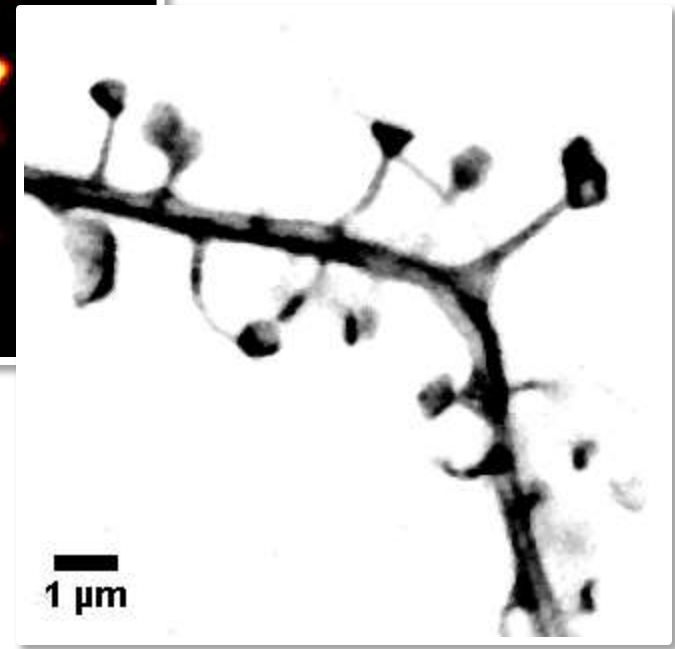
2 color isoSTED resolving  
the inner and outer membrane  
of mitochondria



# Live STED



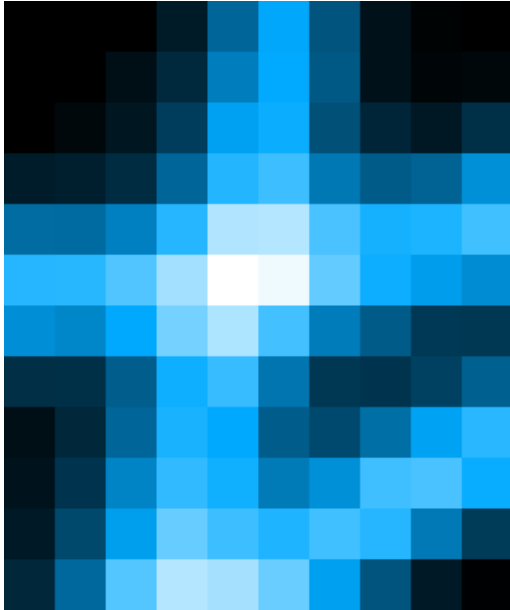
Westphal et al., Science, 2008



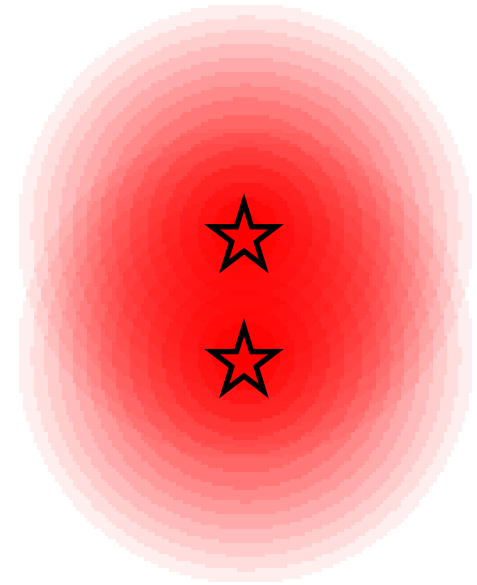
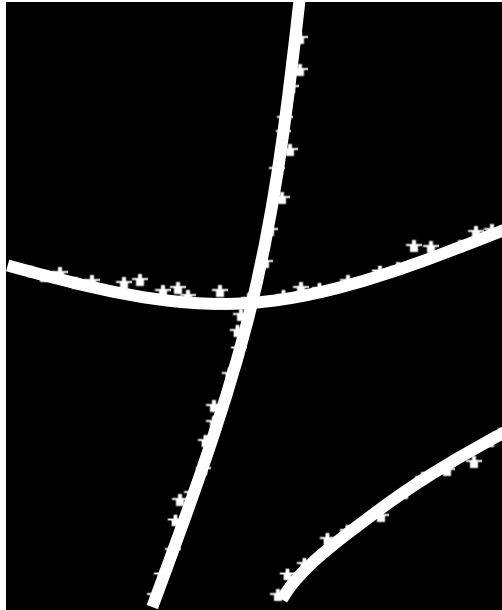
Nagerl et al., PNAS, 2008

# Super-resolution by...

Fluorescence image



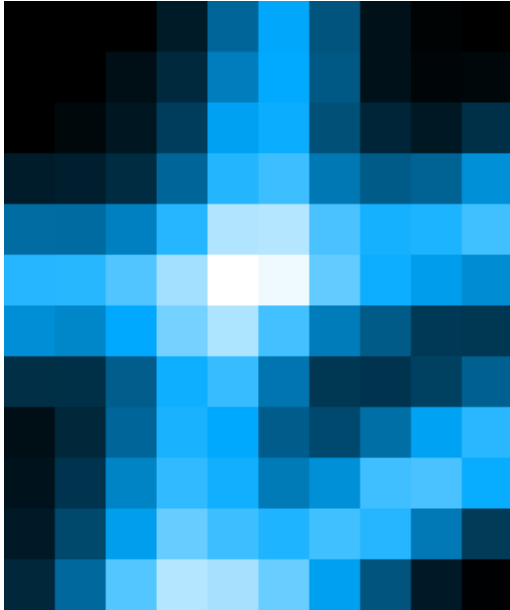
Underlying structure



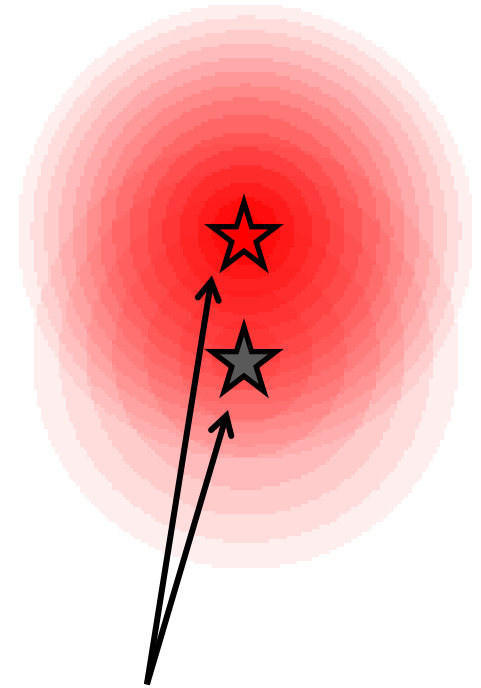
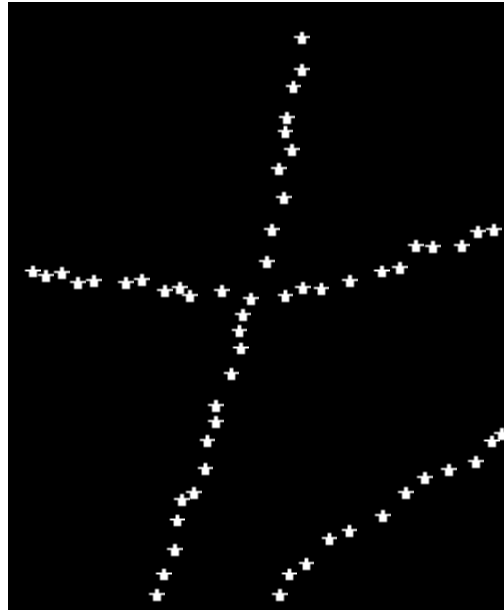


# Super-resolution by spatial modulation

Fluorescence image



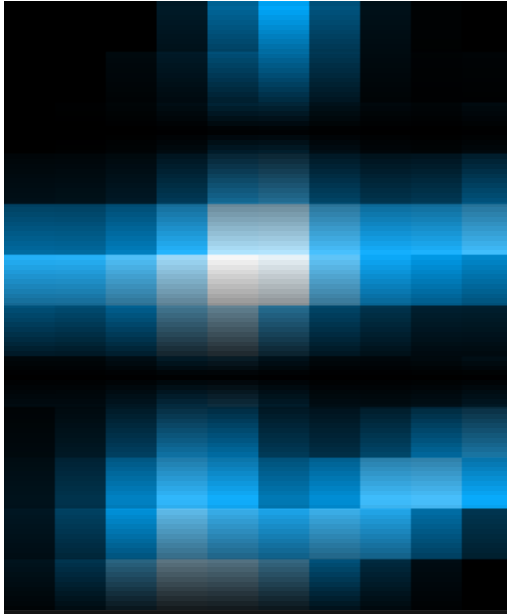
Underlying structure



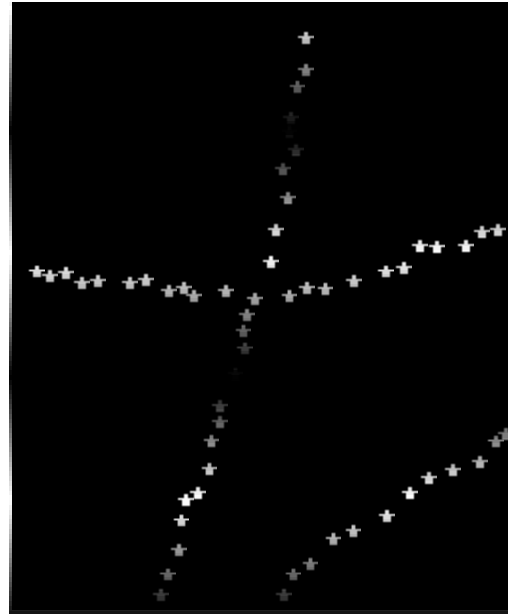
Differential modulation  
of the fluorescence response

# Super-resolution by differential excitation

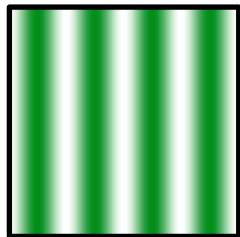
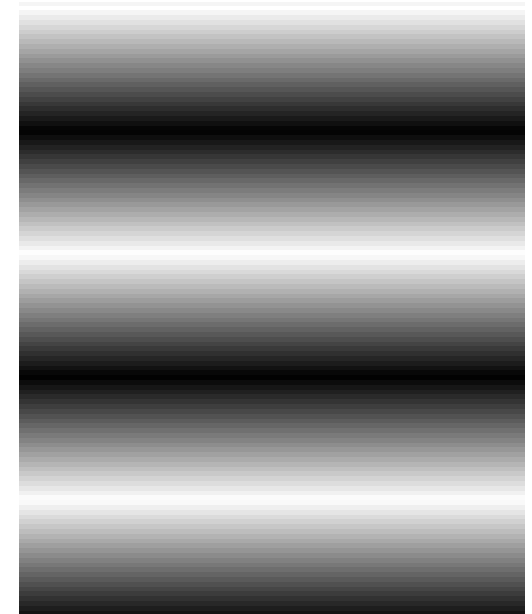
Fluorescence image



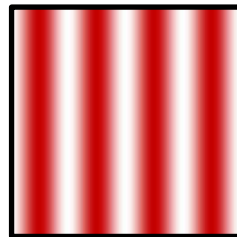
Underlying structure



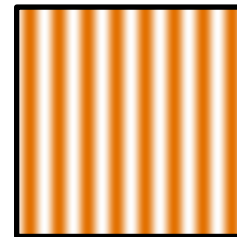
Excitation pattern



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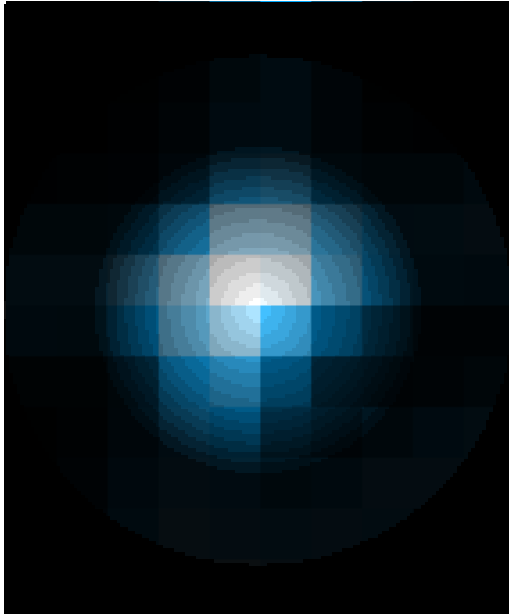
**SIM** (Gustafsson / Heintzmann)  
**SSIM** (Gustafsson 2005)

Diffraction limited excitation and emission

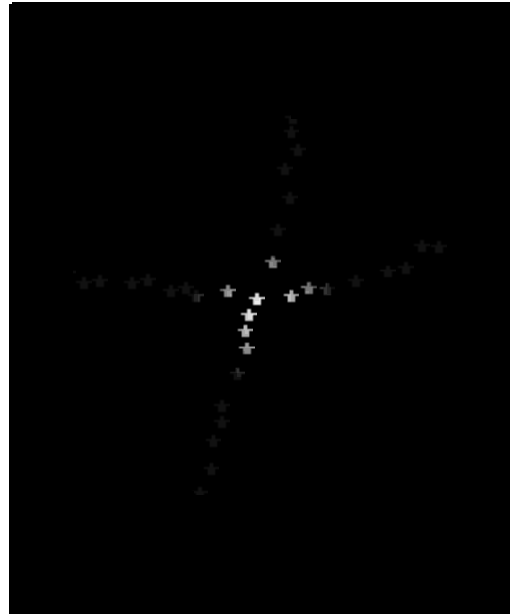
Doubled resolution

# Super-resolution by differential depletion

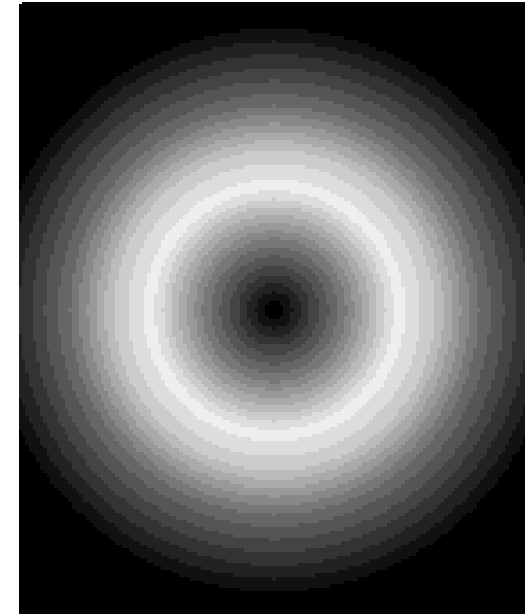
Fluorescence image



Underlying structure



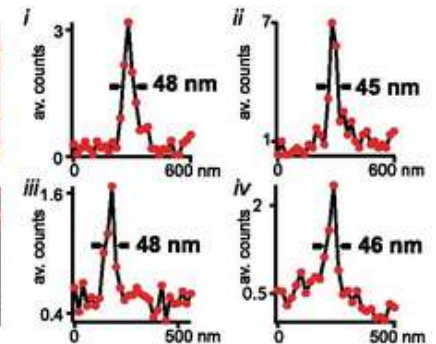
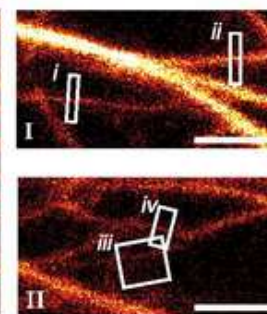
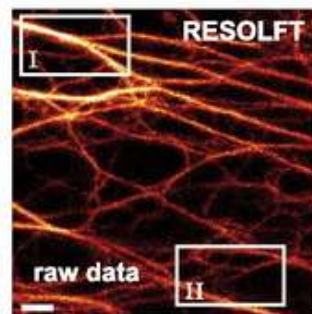
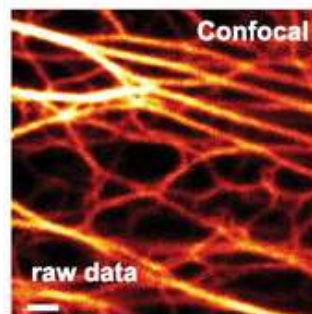
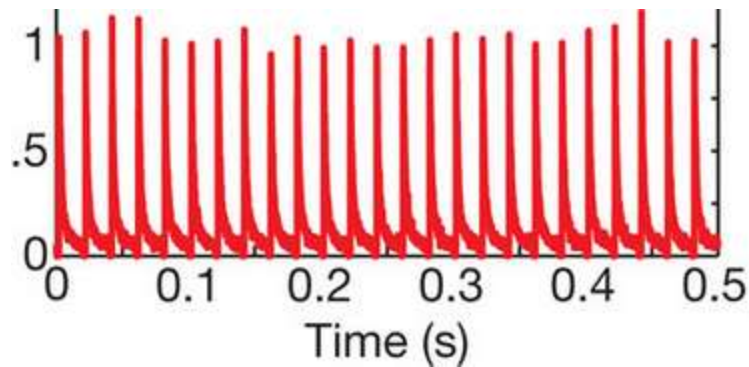
Depletion pattern



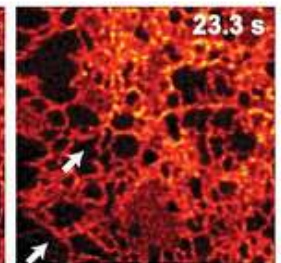
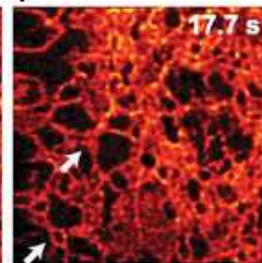
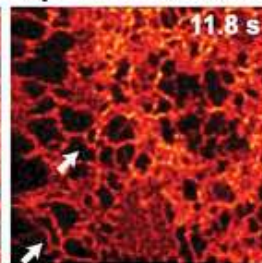
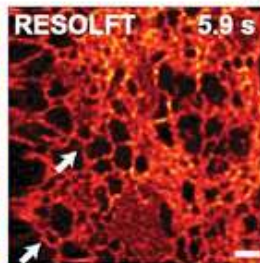
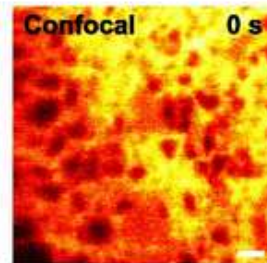
**STED** (Hell 1994, Hell 1999)  
**GSD** (Hell 1995, Hell 2007)  
**RESOLFT** (Hell 2003, Hell 2011)

Diffraction limited PSF   Saturated depletion   Smaller effective PSF

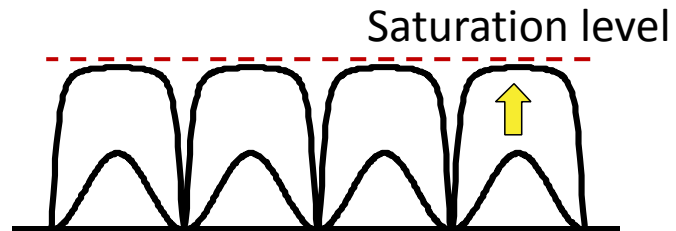
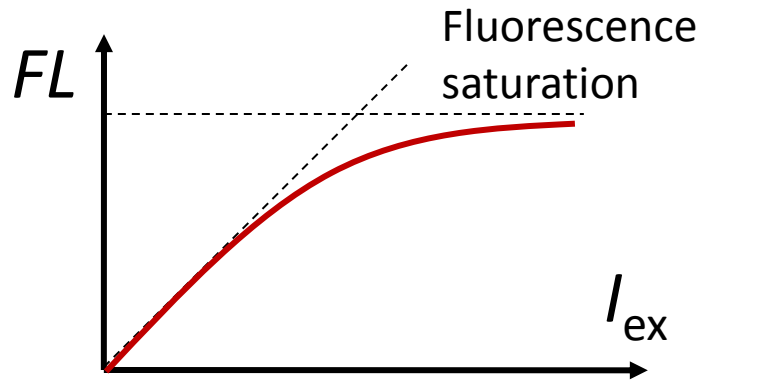
# RESOLFT by rsEGFP and rsEGFP2



## ER (rsEGFP2-KDEL)



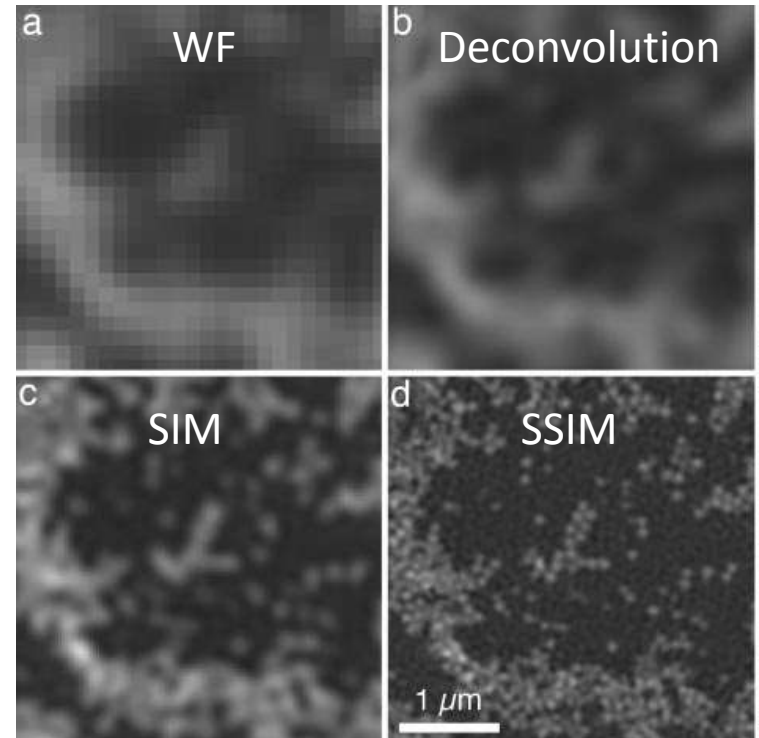
# Saturated SIM



Saturated illumination pattern



Sharp zero lines



50 nm resolution

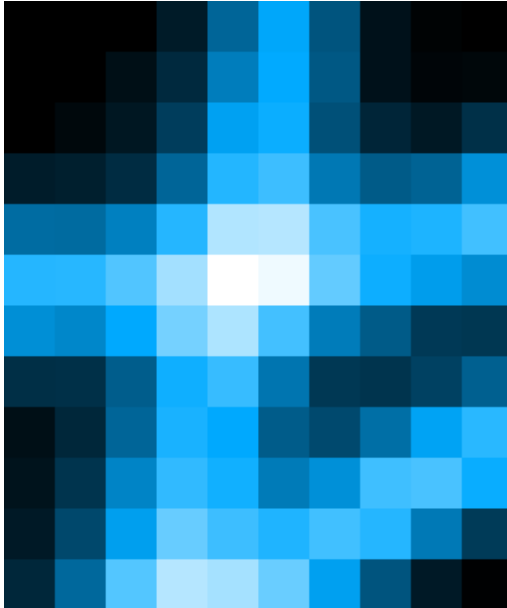
Suffers from fast photobleaching  
under saturated excitation condition

# Super-resolution by single-molecule switching

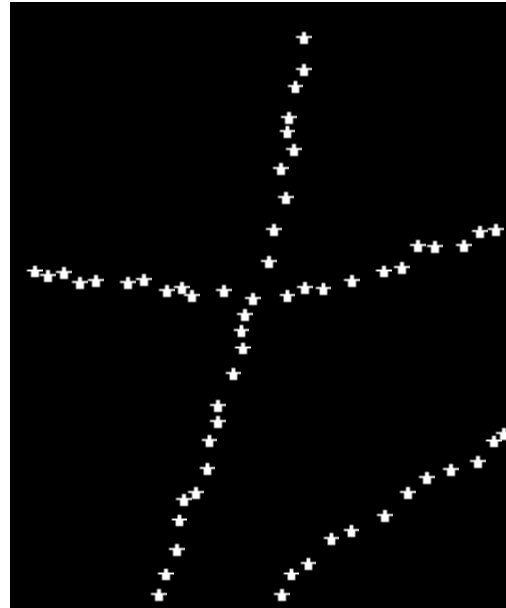


# Super-resolution by single-molecule switching

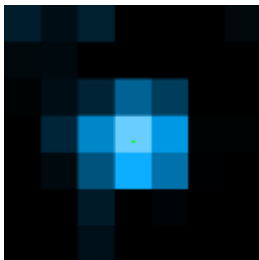
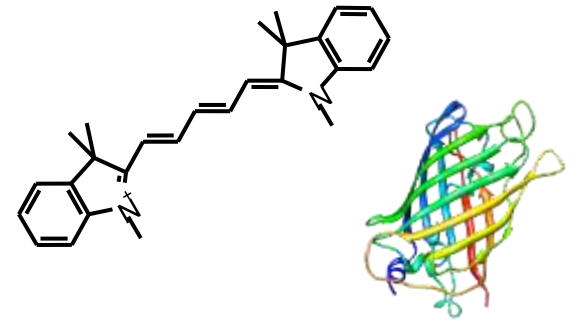
Fluorescence image



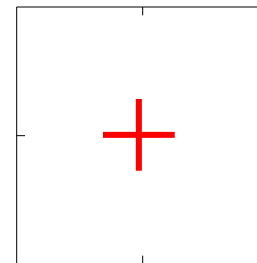
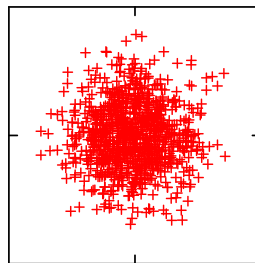
Underlying structure



Photoswitchable molecules



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$$D \approx d / \sqrt{N}$$

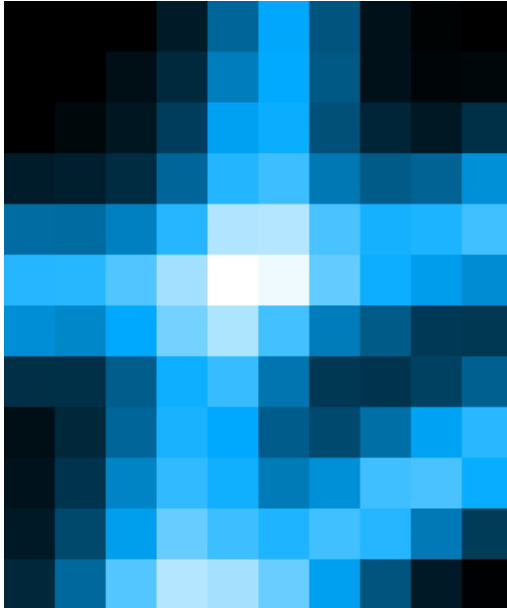
Single molecule image

N photons

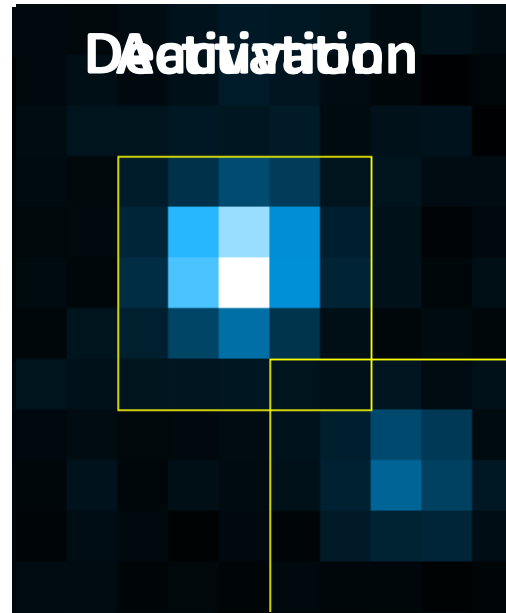
Single-molecule localization

# Super-resolution by single-molecule switching

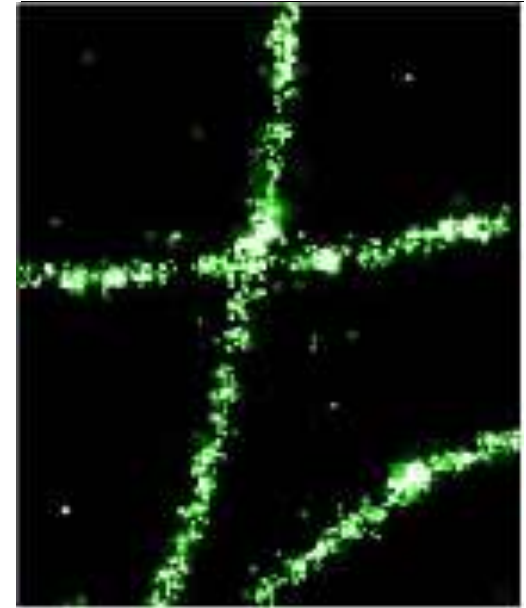
Fluorescence image



Raw images



STORM Image



2x real time

**STORM** = Stochastic Optical Reconstruction Microscopy (Zhuang 2006)

**PALM** = Photoactivated Localization Microscopy (Betzig & Hess 2006)

**FPALM** = Fluorescence Photoactivation Localization Microscopy (Hess 2006)

**PALMIRA** (Hell 2007), **GSDIM** (Hell 2008), **dSTORM** (Sauer 2008), **SMACM** (Moerner 2008)

**PAINT** (Hochstrasser 2006), **SPRAYPAINT** (Moerner 2011), **SOFI** (Weiss 2009)



# *Drosophila* motoneuron dendrites

