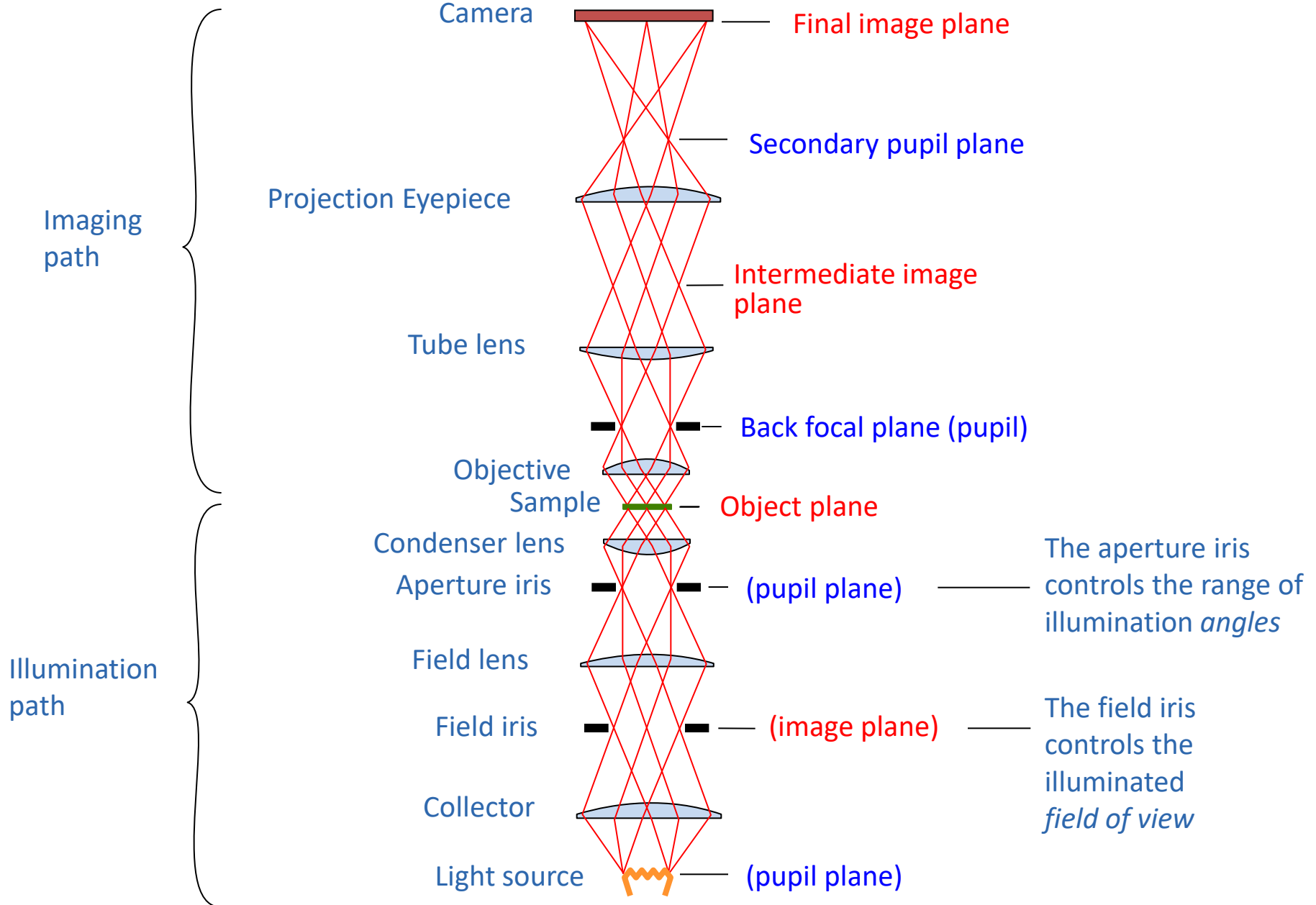


# Principles and Practices of Light Microscopy

II: Brightfield optics, resolution, and aberrations

# Trans-illumination Microscope



By far the most important part:  
*the Objective Lens*



Each major manufacturer sells 20-30 different *categories* of objectives.  
What are the important distinctions?



# Working Distance

## Objective Working and Parfocal Distance



In general, high NA lenses have short working distances

However, extra-long working distance objectives do exist

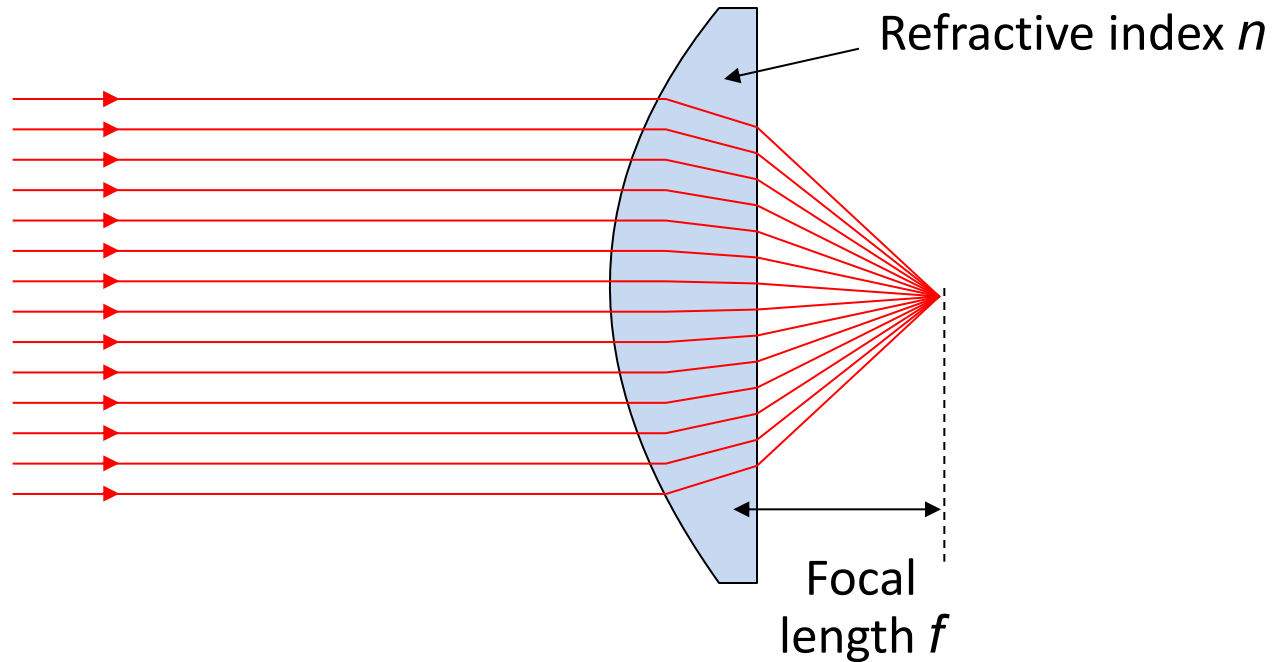
Some examples:

10x/0.3 WD = 15.2mm

20x/0.75 WD = 1.0mm

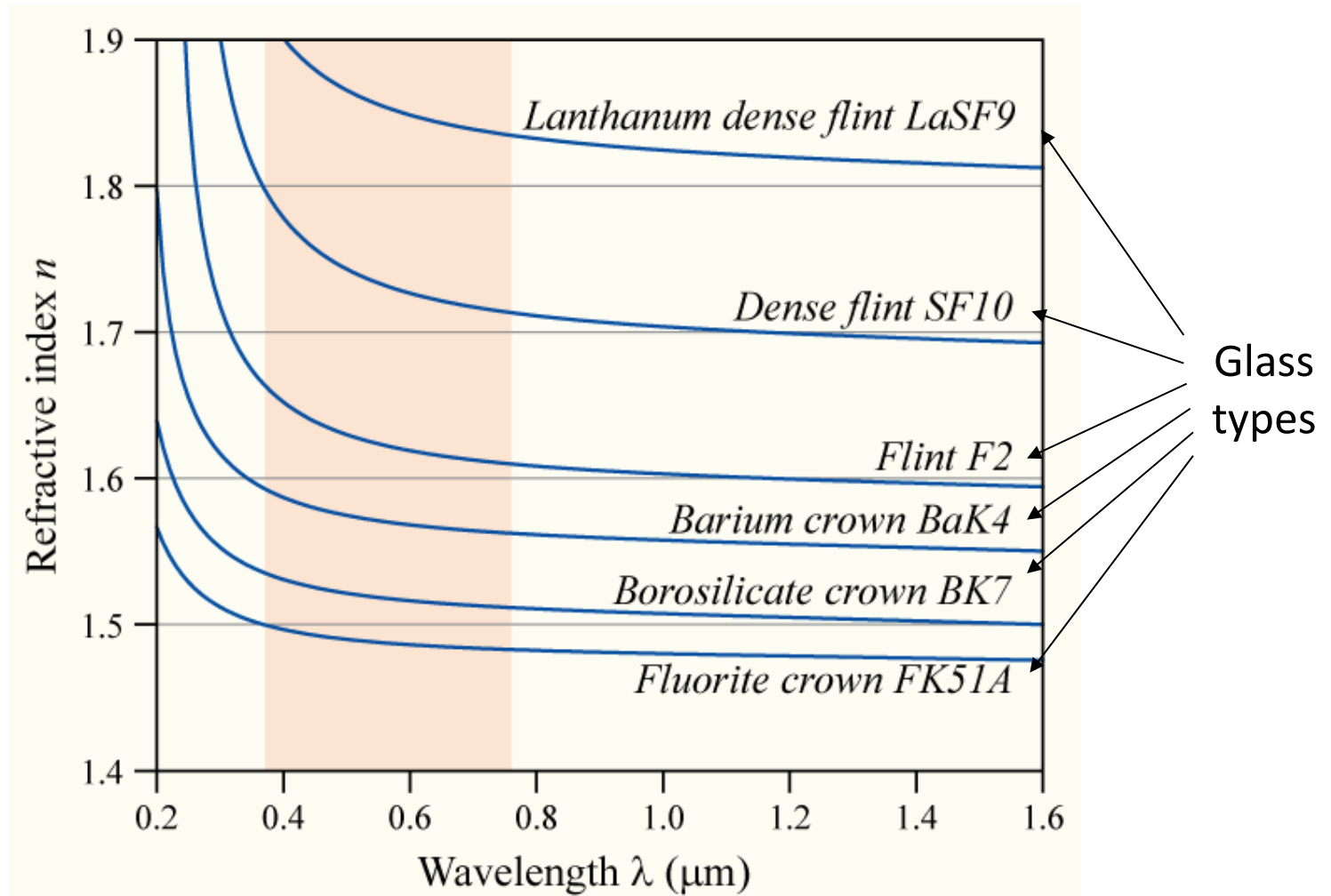
100x/1.4 WD = 0.13mm

The focal length of a lens depends on the refractive index...



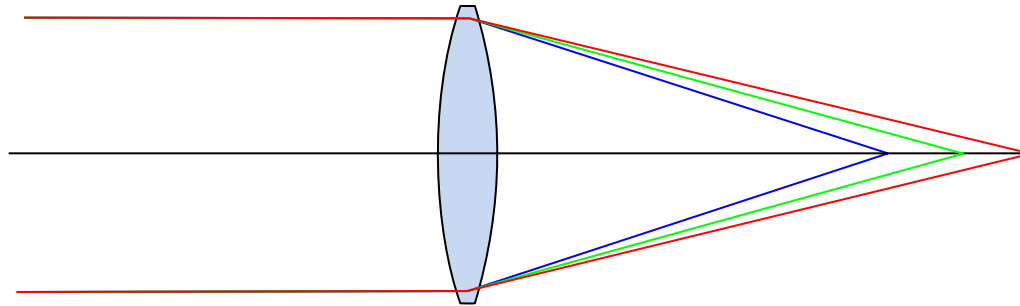
$$f \propto 1/(n-1)$$

... and the refractive index  
depends on the wavelength  
("dispersion")

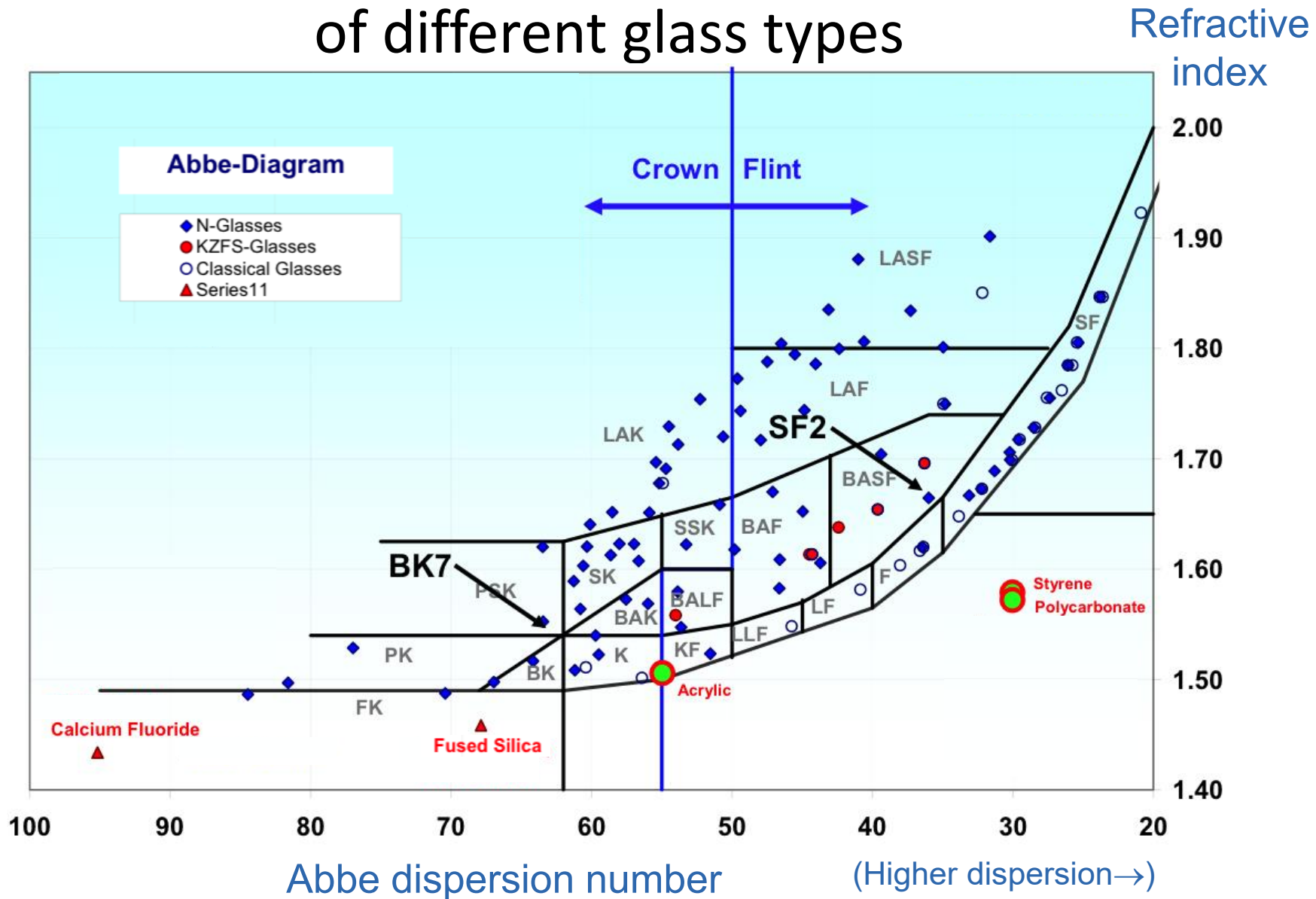


## $\Rightarrow$ Chromatic aberration

- Different colors get focused to different planes
- Not good...



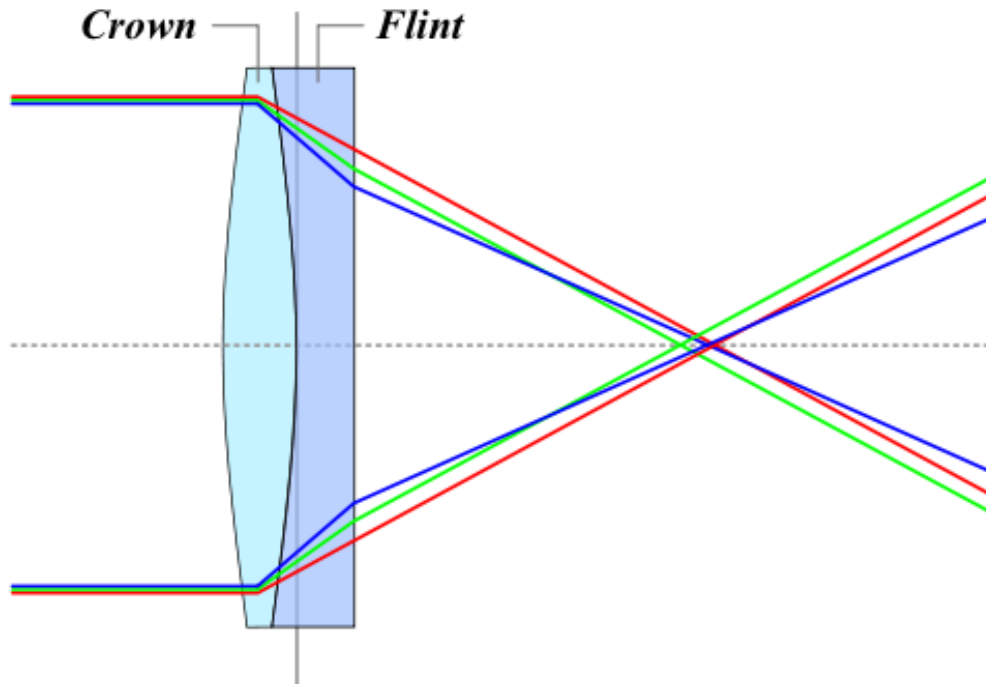
# Dispersion vs. refractive index of different glass types



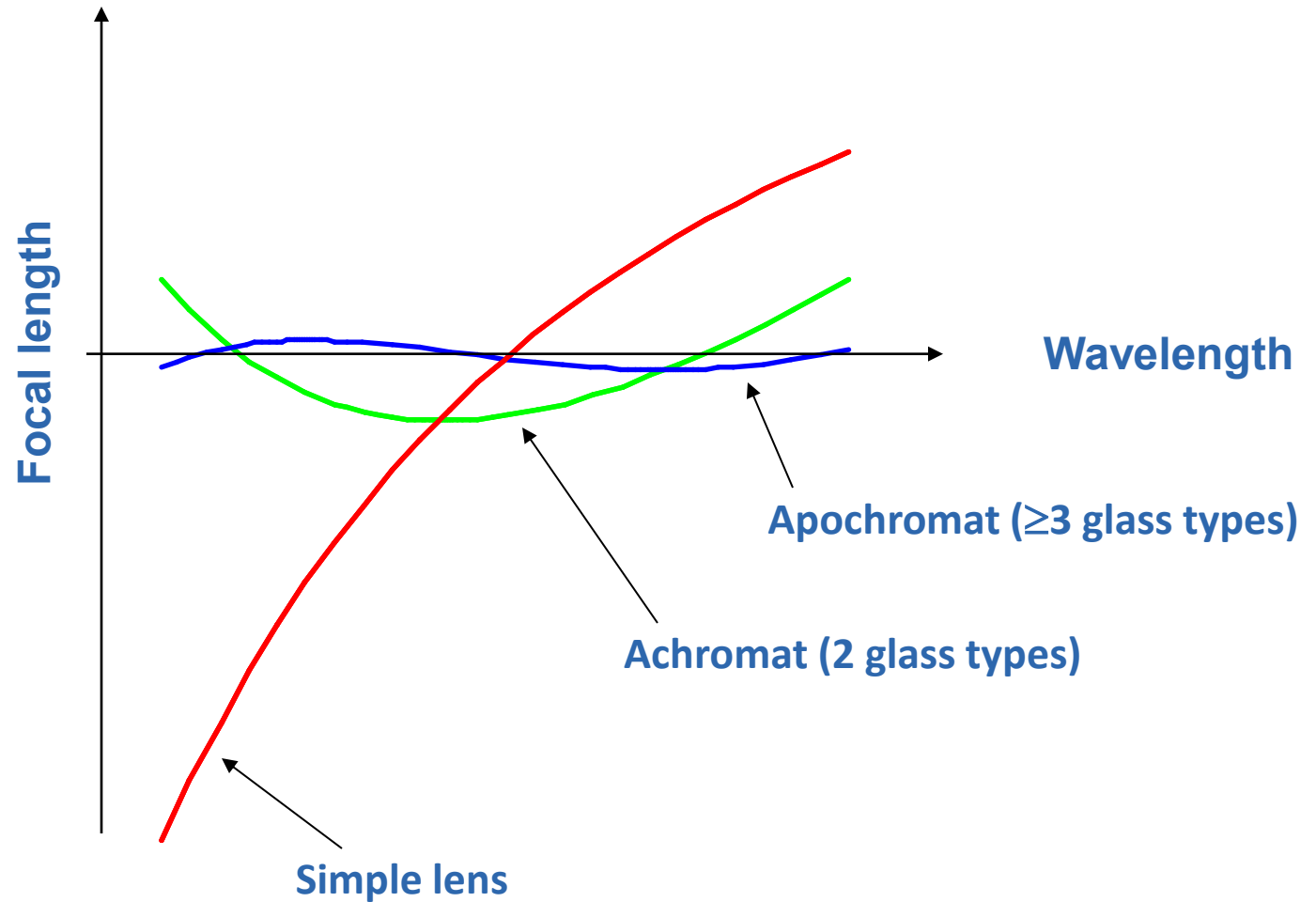


# Achromatic Lenses

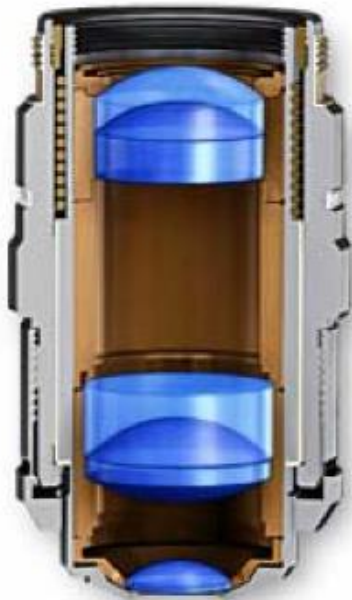
- Use a weak negative flint glass element to compensate the dispersion of a positive crown glass element



# Achromats and Apochromats



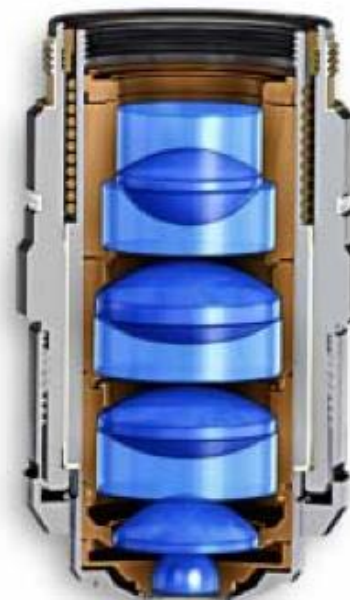
# Correction classes of objectives



**Achromat**  
(cheap)



**Fluor**  
“semi-apo”  
(good correction,  
high UV  
transmission)

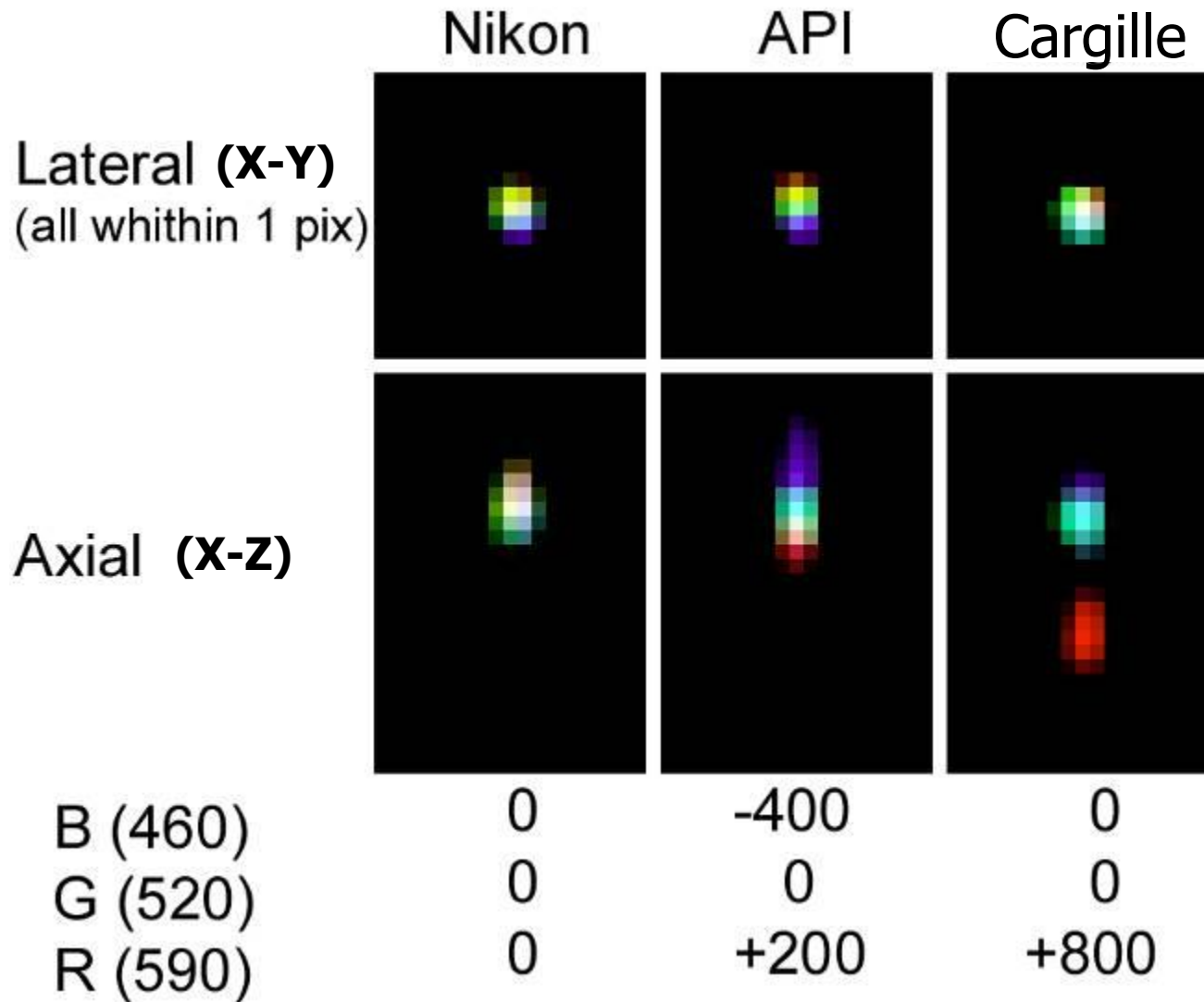


**Apochromat**  
(best correction)

Correction for other (i.e. monochromatic) aberrations  
also improves in the same order



# Using the wrong immersion oil can induce axial chromatic aberration



# Putting one brand of objectives onto another brand of microscope?

## Usually a bad idea:

- May not even fit
- May get different magnification than is printed on the objective
- Incompatible ways of correcting lateral chromatic aberration (LCA)  
⇒ mixing brands can produce severe LCA



### Tube lens focal length

Nikon	200
Leica	200
Olympus	180
Zeiss	165

### LCA correction:

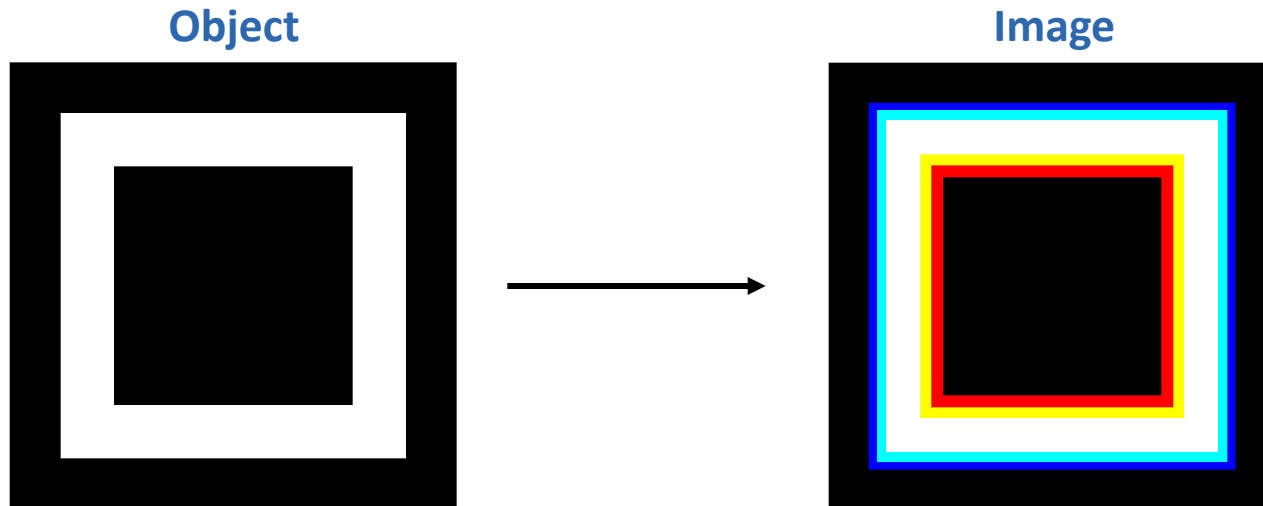
<u>In objective</u>	<u>In tube lens</u>
Nikon	Leica
Olympus	Zeiss



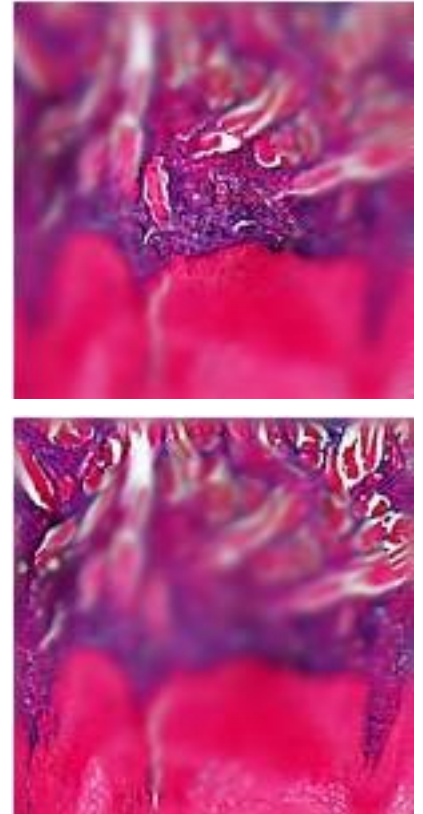
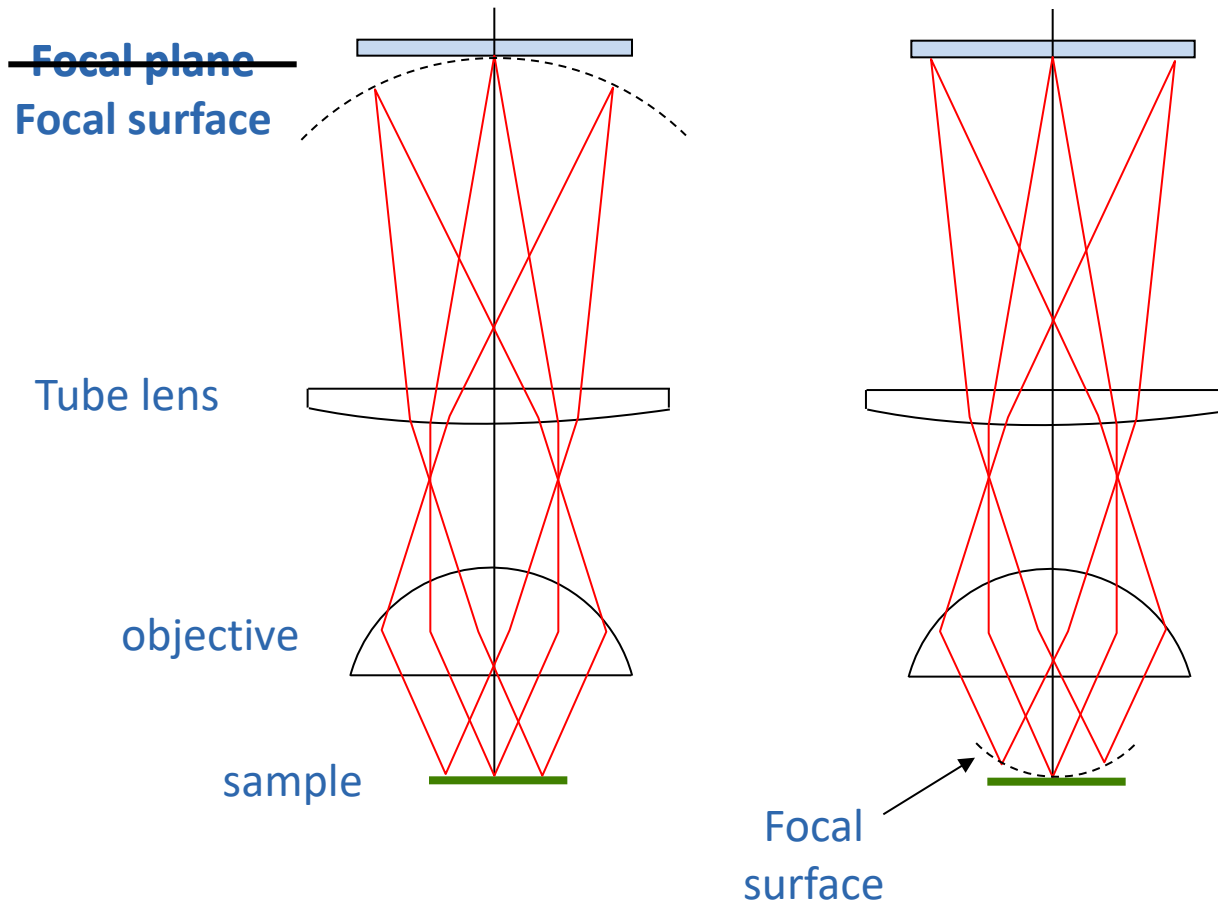
# Lateral chromatic aberration

(= LCA, lateral color, chromatic difference of magnification)

= Different magnification for different colors

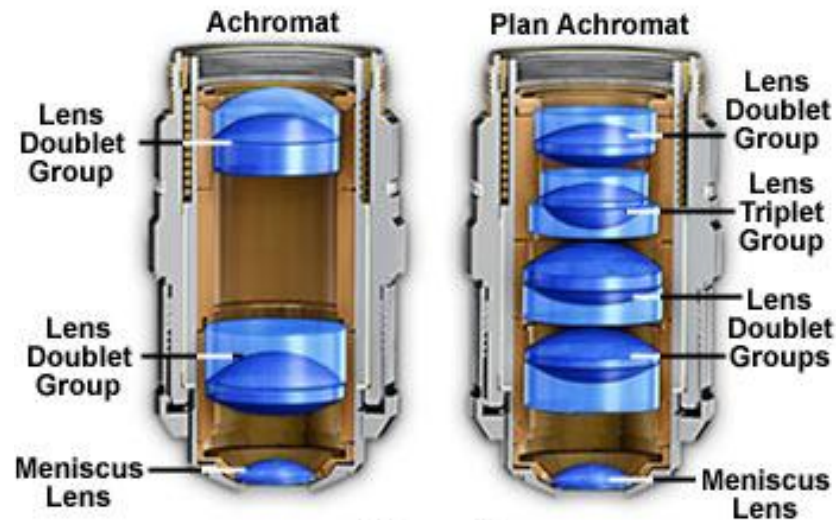


# Curvature of Field



# Plan objectives

- Corrected for field curvature
- More complex design
- Needed for most photomicrography



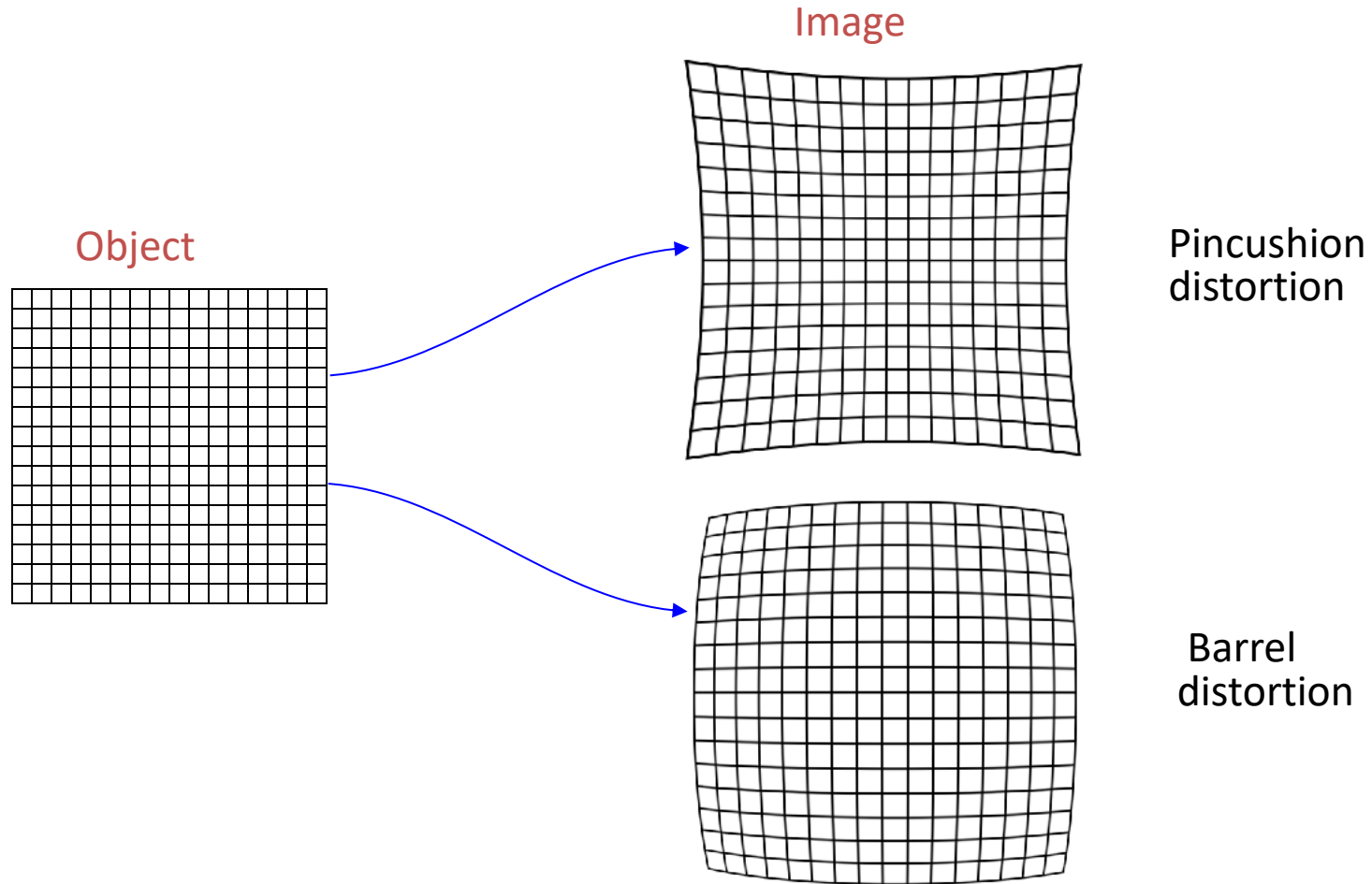
- **Plan-Apochromats** have the highest performance (and highest complexity and price)

# Aberrations

- Chromatic aberrations
  - Longitudinal chr. Ab.
  - Lateral chr. Ab.
- Curvature of field
- Distortion
- Wavefront aberrations
  - Spherical aberration
  - Astigmatism
  - Coma
  - ...

# Geometric Distortion

= Radially varying magnification

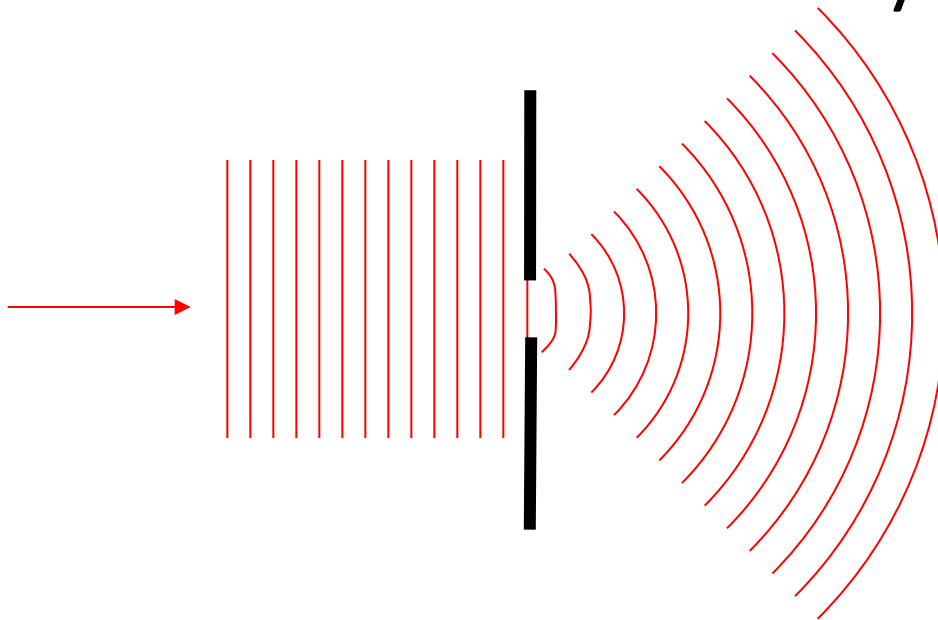


May be introduced by the projection eyepiece

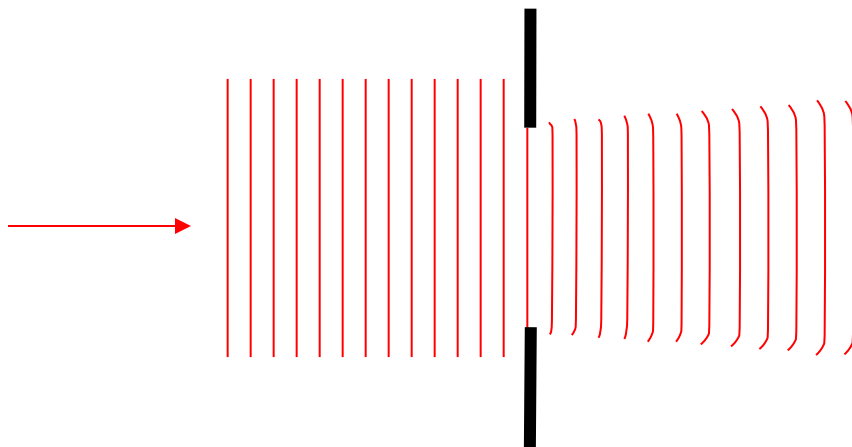


# Diffraction by an aperture

drawn as waves



Light spreads to new angles



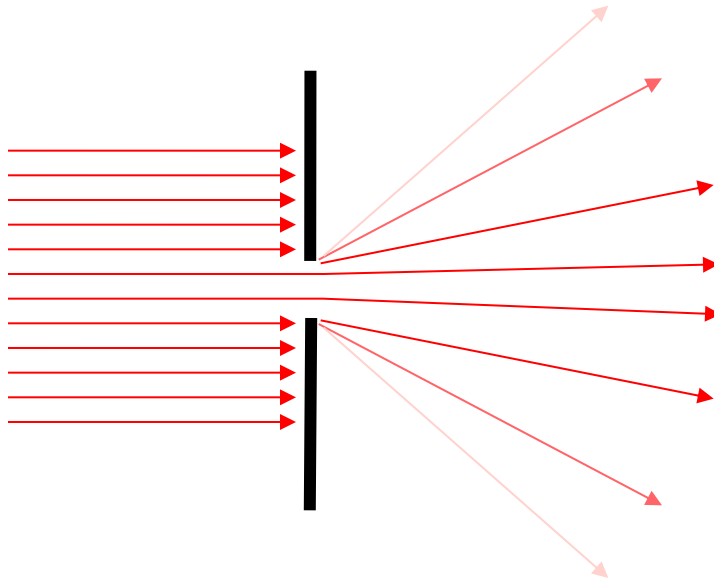
Larger aperture



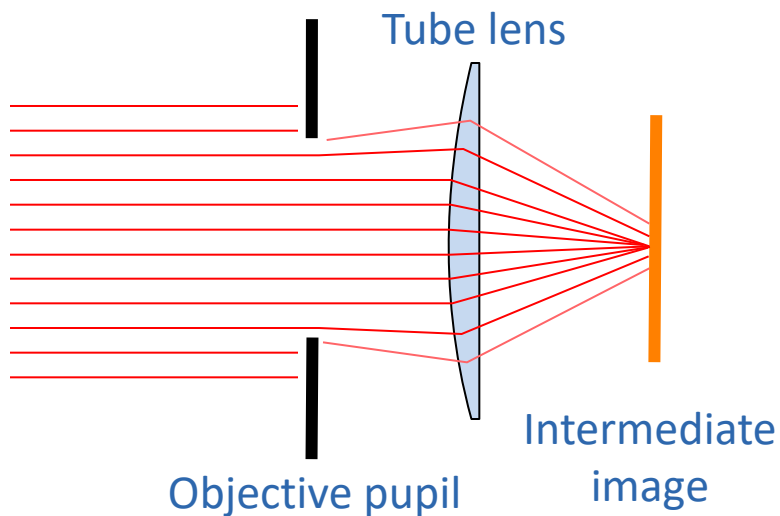
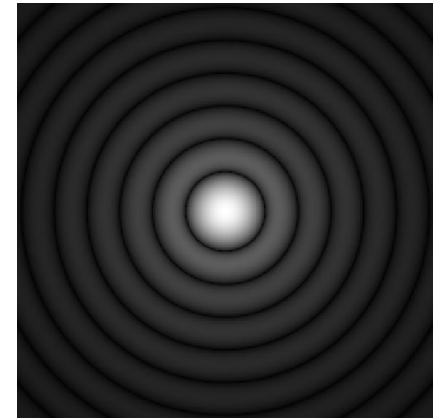
weaker diffraction

# Diffraction by an aperture

drawn as rays



The pure, “far-field”  
diffraction pattern  
is formed at  $\infty$  distance...

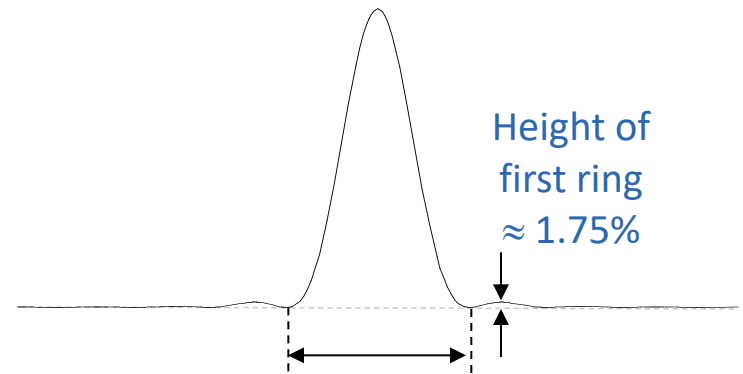
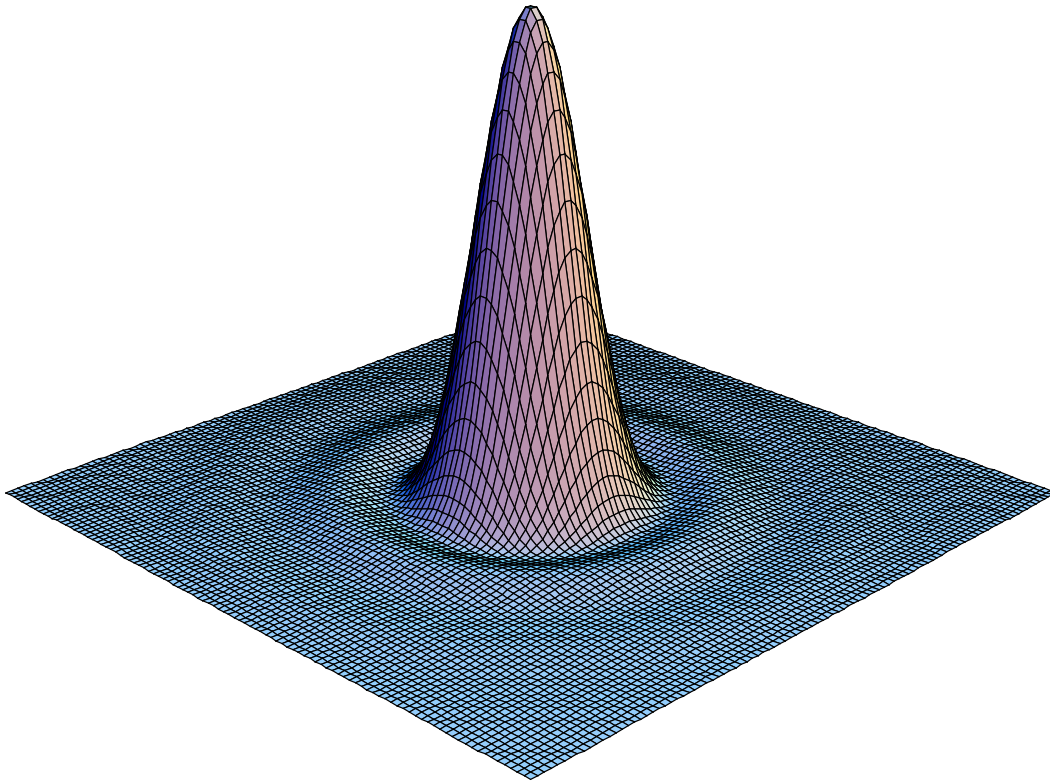


...or can be formed  
at a finite distance  
by a lens...

*...as happens in a microscope*

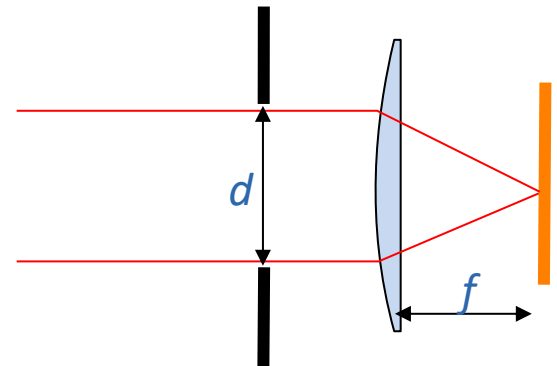
# The Airy Pattern

= the far-field diffraction pattern from a round aperture

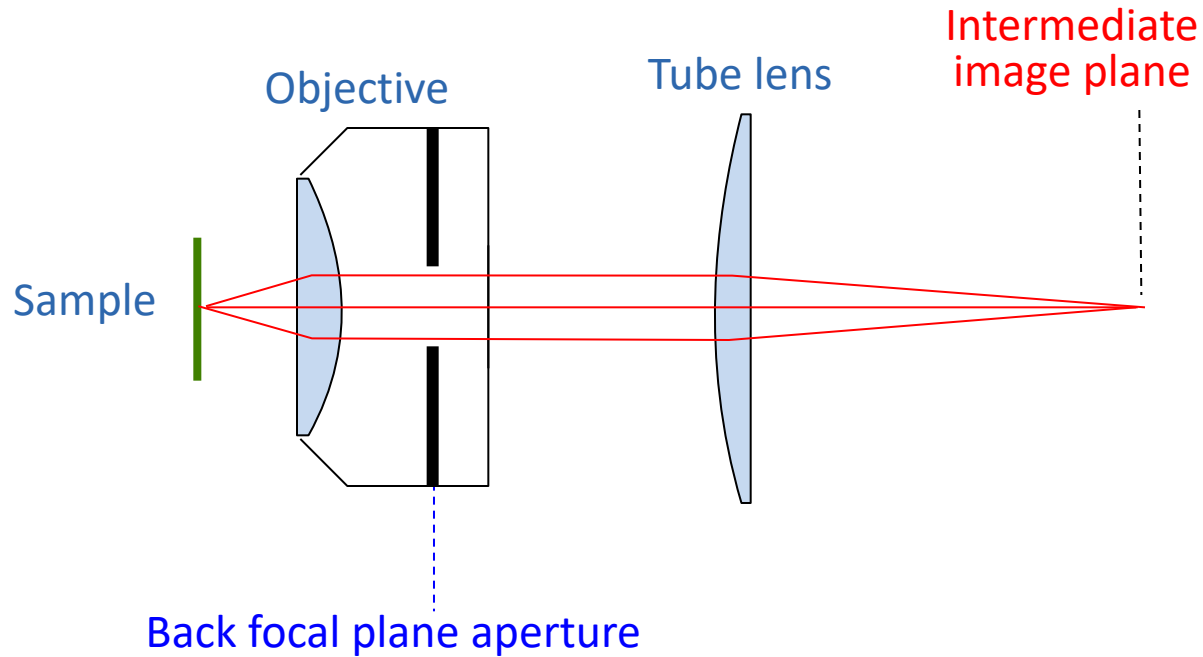


Height of  
first ring  
 $\approx 1.75\%$

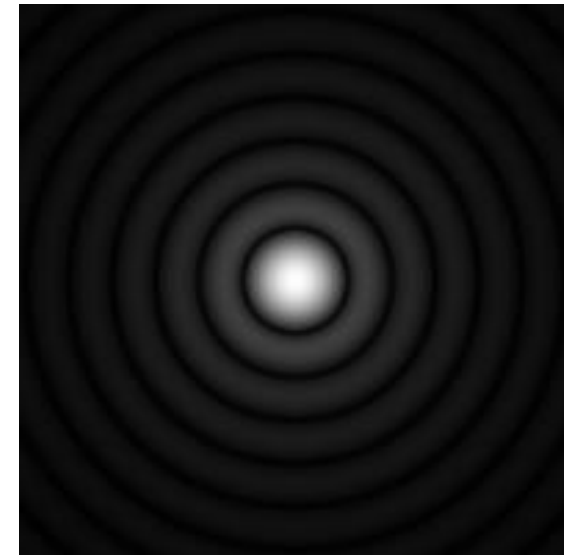
“Airy disk” diameter  
 $d = 2.44 \lambda f/d$   
(for small angles  $d/f$ )



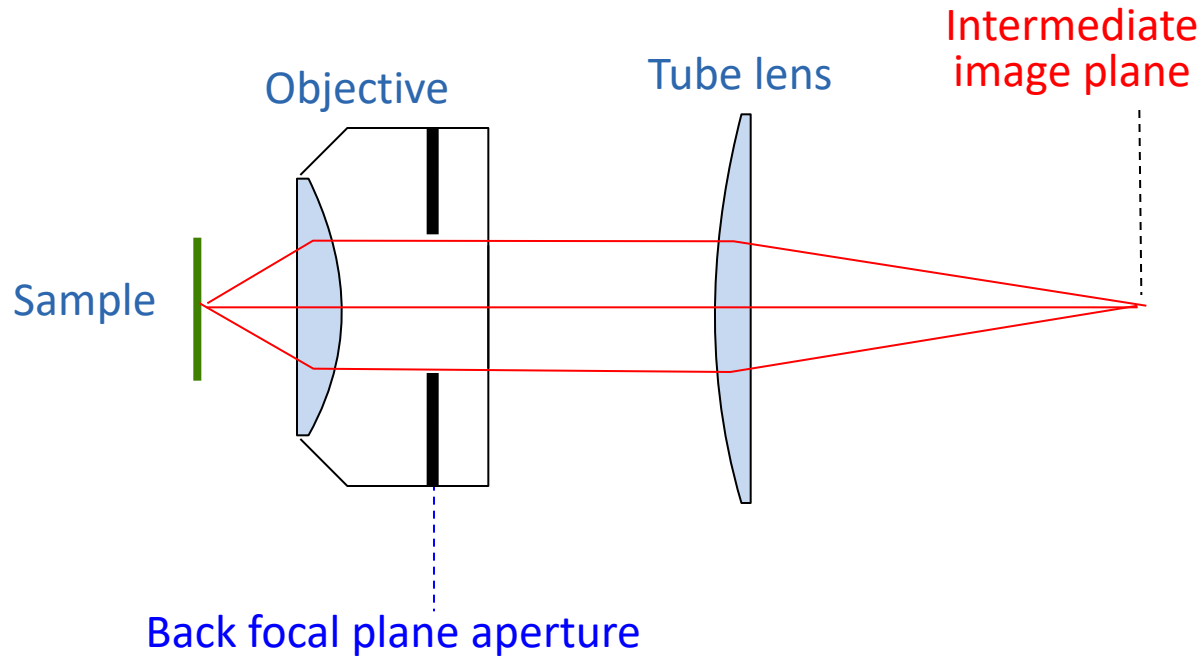
# Aperture and Resolution



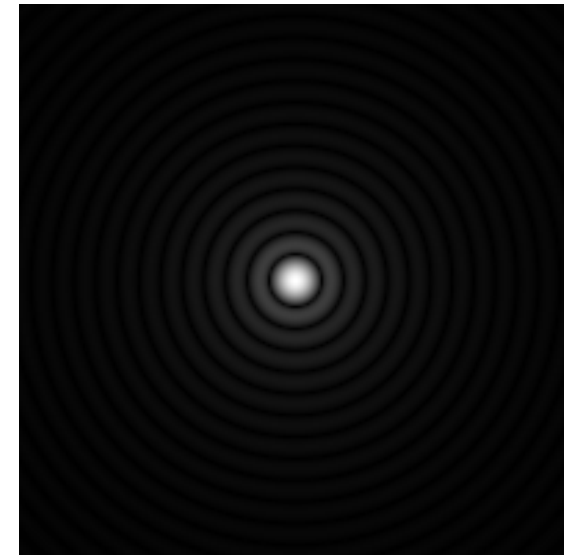
Diffraction spot  
on image plane  
= *Point Spread Function*



# Aperture and Resolution

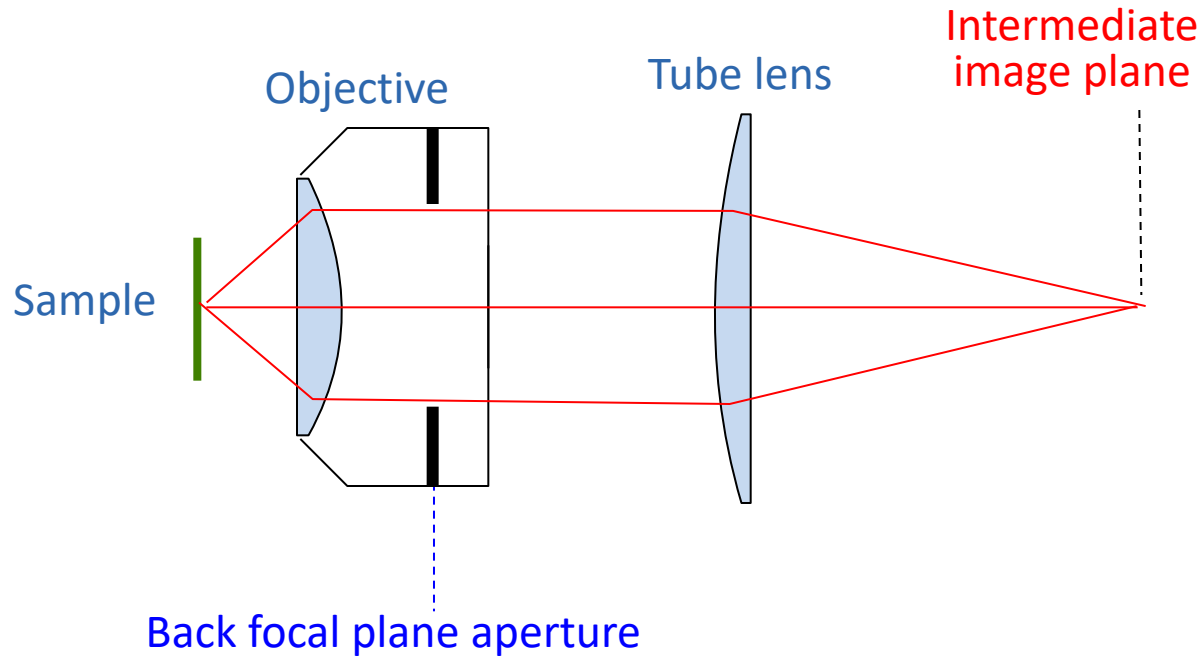


Diffraction spot  
on image plane  
= ***Point Spread Function***

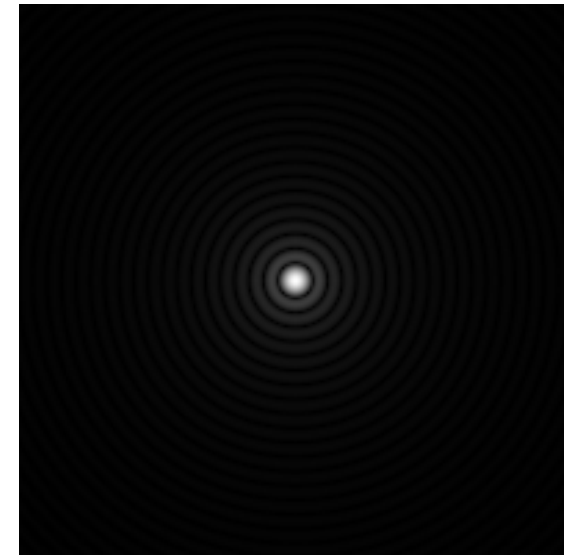




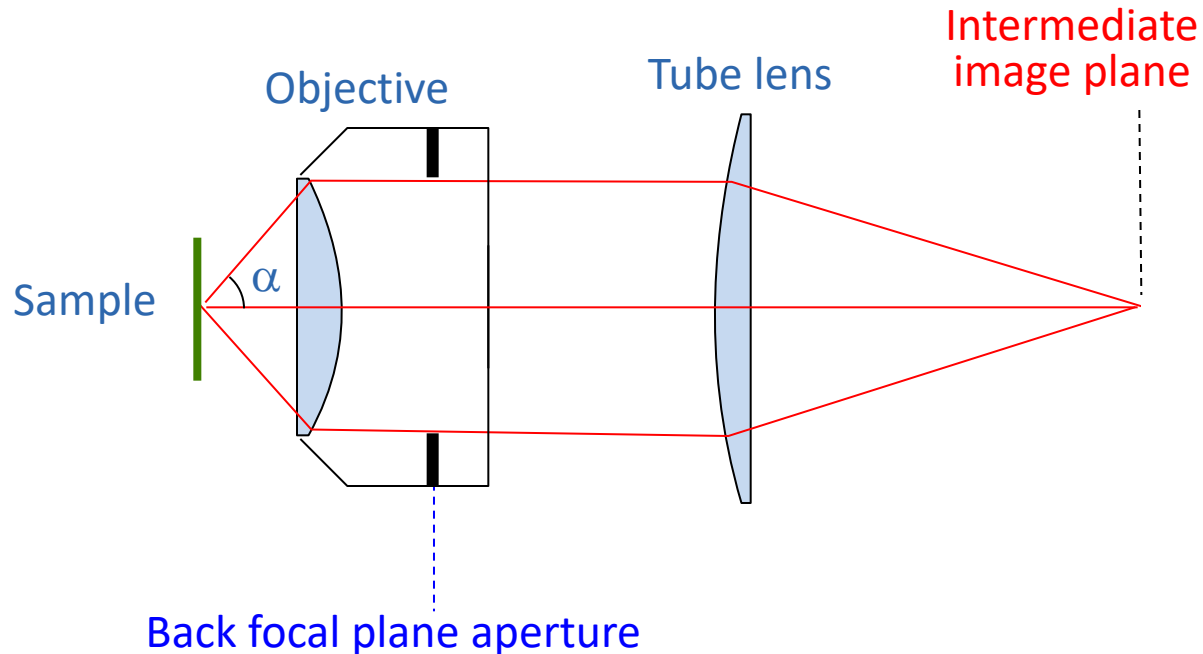
# Aperture and Resolution



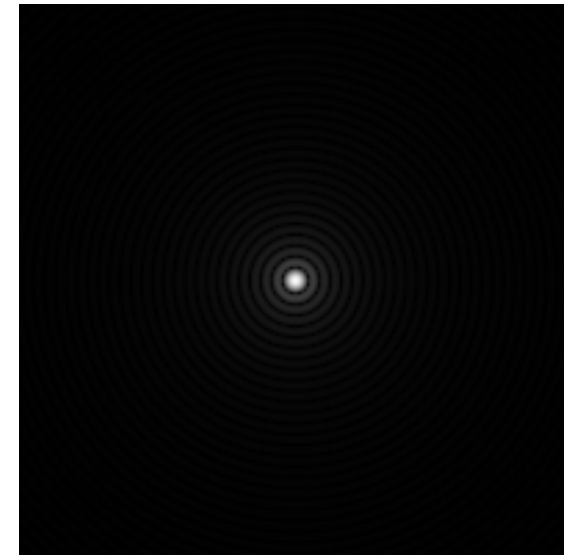
Diffraction spot  
on image plane  
= ***Point Spread Function***



# Aperture and Resolution



Diffraction spot  
on image plane  
(resolution)

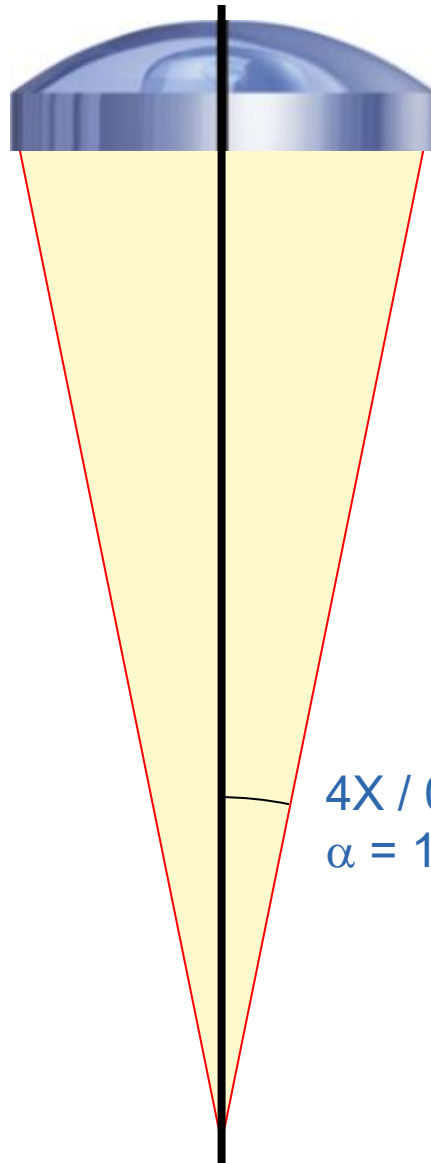


- Image resolution improves with aperture size — Numerical Aperture (NA)

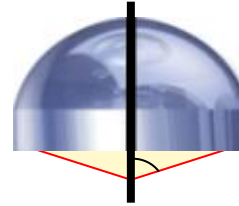
$$NA = n \sin(\alpha)$$

where:  $\alpha$  = light gathering angle  
 $n$  = refractive index of sample

# Numerical Aperture



4X / 0.20 NA  
 $\alpha = 11.5^\circ$



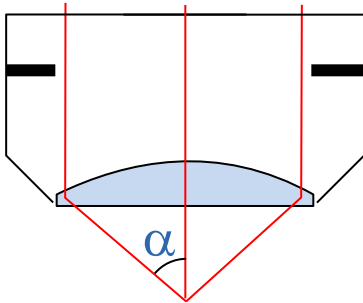
100X / 0.95 NA  
 $\alpha = 71.8^\circ$

# Numerical Aperture

Compare:

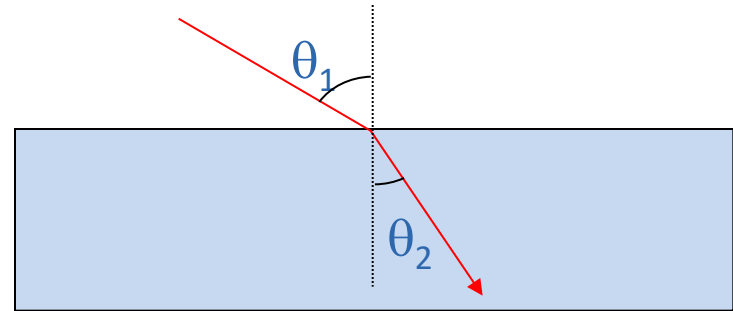
Numerical Aperture:

$$NA = n \sin(\alpha)$$



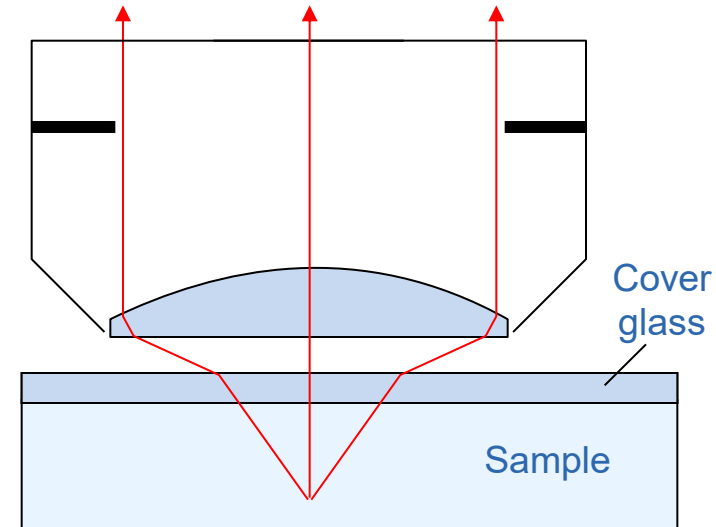
Snell's law:

$$n_1 \sin(\theta_1) = n_2 \sin(\theta_2)$$



- $n \sin(\theta)$  doesn't change at horizontal interfaces
- $\sin(\text{anything}) \leq 1$

⇒ NA cannot exceed the *lowest*  $n$  between the sample and the objective lens

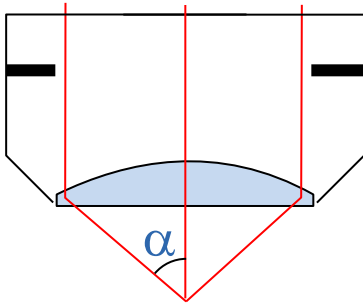


# Numerical Aperture

Compare:

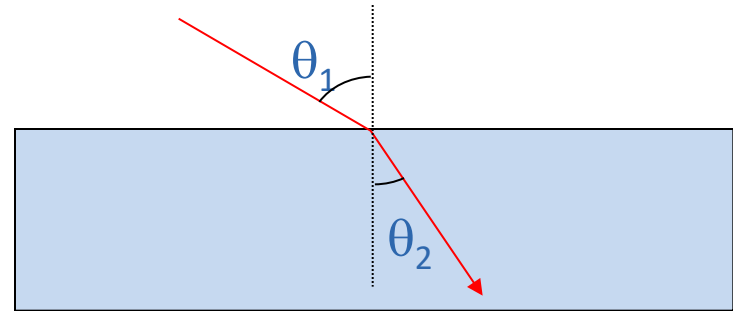
Numerical Aperture:

$$NA = n \sin(\alpha)$$



Snell's law:

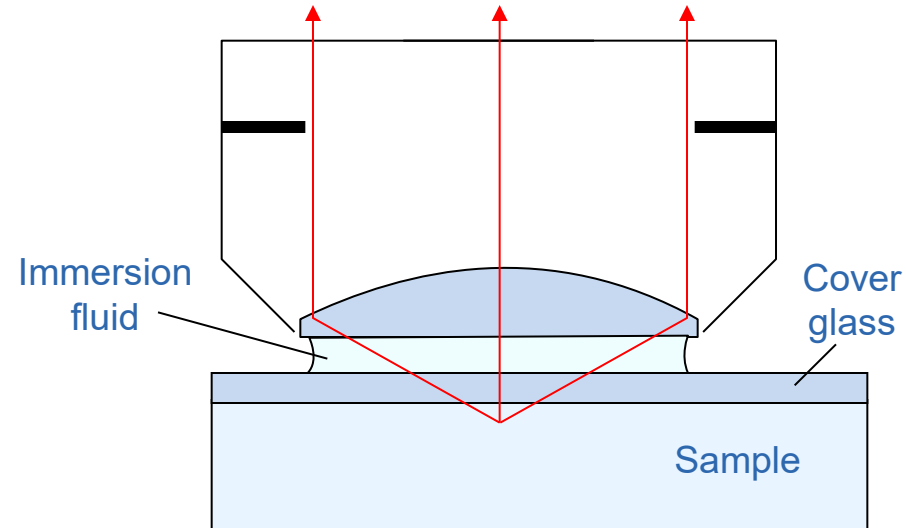
$$n_1 \sin(\theta_1) = n_2 \sin(\theta_2)$$



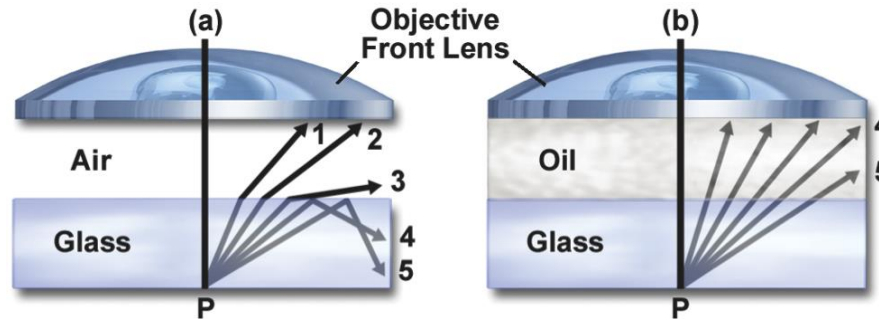
- $n \sin(\theta)$  doesn't change at horizontal interfaces
- $\sin(\text{anything}) \leq 1$

⇒ NA cannot exceed the *lowest*  $n$  between the sample and the objective lens

⇒  $NA > 1$  requires **fluid immersion**



# Immersion Objectives



⇒ NA cannot exceed  
the *lowest*  $n$  between the  
sample and the objective lens

⇒  $NA > 1$  requires **fluid immersion**

NA can approach  
the index of the  
immersion fluid

Oil immersion:

$n \approx 1.515$

max NA  $\approx 1.4$  (1.45–1.49 for TIRF)

Glycerol immersion:

$n \approx 1.45$  (85%)

max NA  $\approx 1.35$  (Leica)

Water immersion:

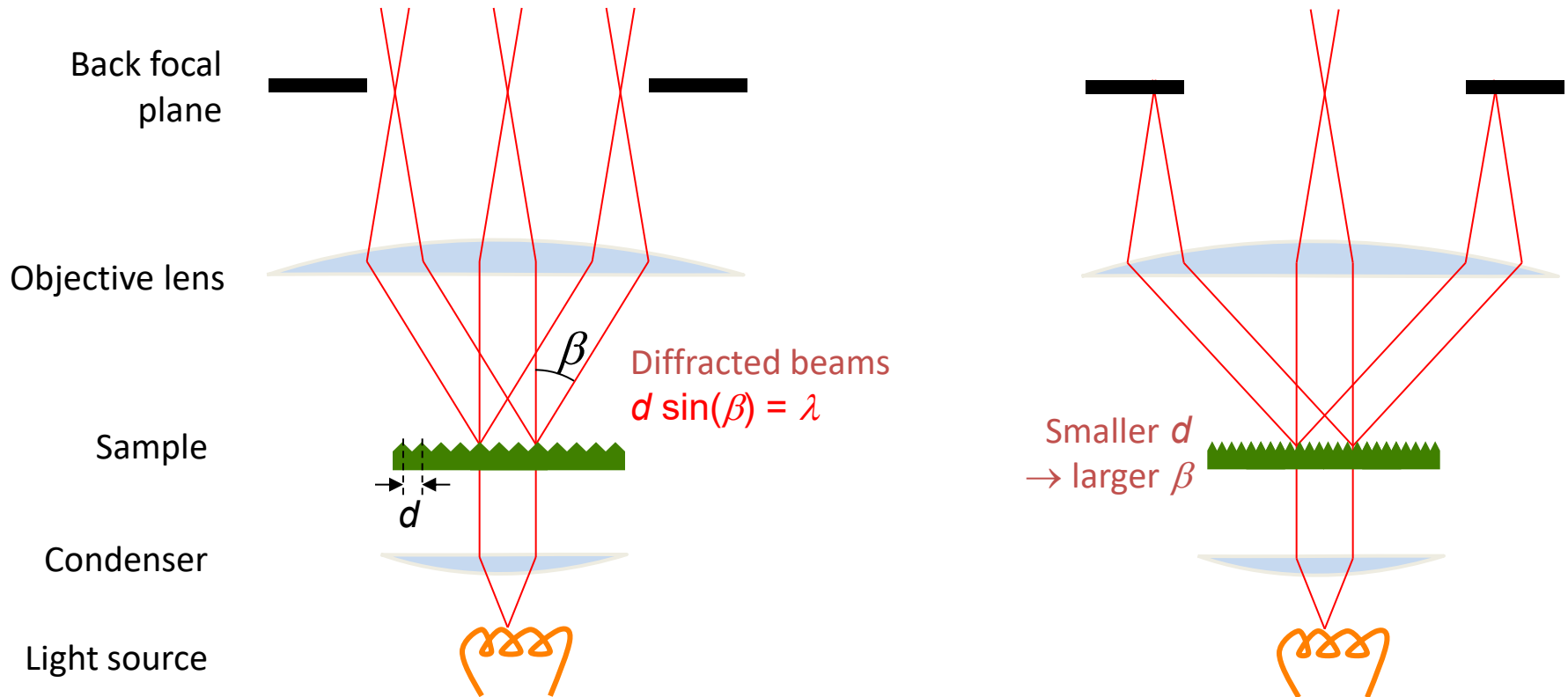
$n \approx 1.33$

max NA  $\approx 1.2$

# Resolution

## Ernst Abbe's argument (1873)

Consider a striped sample  $\approx$  a diffraction grating



Consider first  
a point light source

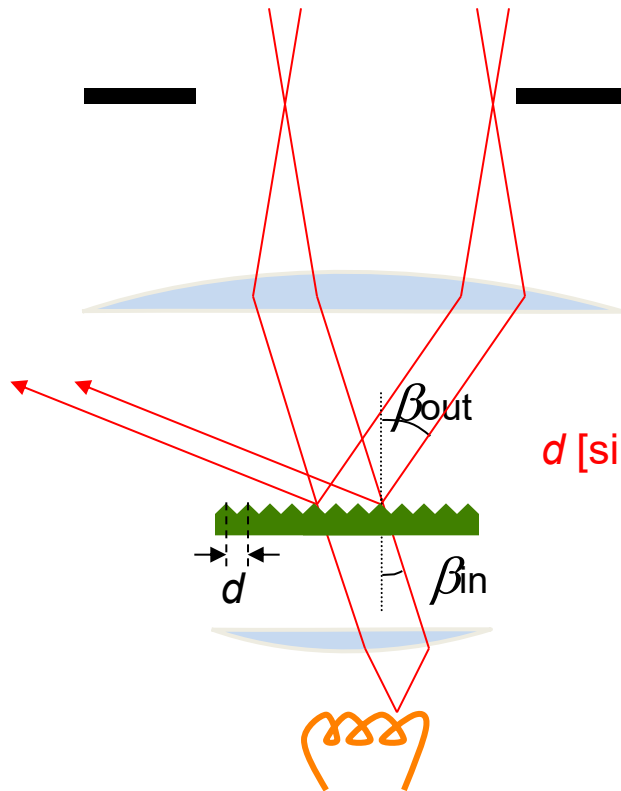
If  $\beta > \alpha$ , only one spot makes it through  
 $\Rightarrow$  no interference  $\Rightarrow$  no image formed

Resolution (smallest resolvable  $d$ ):

$$d_{\min} = \lambda_{\text{sample}} / \sin(\alpha) = \lambda / n \sin(\alpha) = \lambda / NA$$

## (Abbe's argument, continued)

Now consider oblique illumination  
(an off-axis source point):



One spot hopelessly lost,  
but **two** spots get through  
→ interference → image formed!

$$d [\sin(\beta_{in}) + \sin(\beta_{out})] = \lambda$$

Two spots get through if  
 $\beta_{out} < \alpha$  and  $\beta_{in} < \alpha$ .

Resolution (smallest resolvable  $d$ )  
with incoherent illumination (all possible illumination directions):

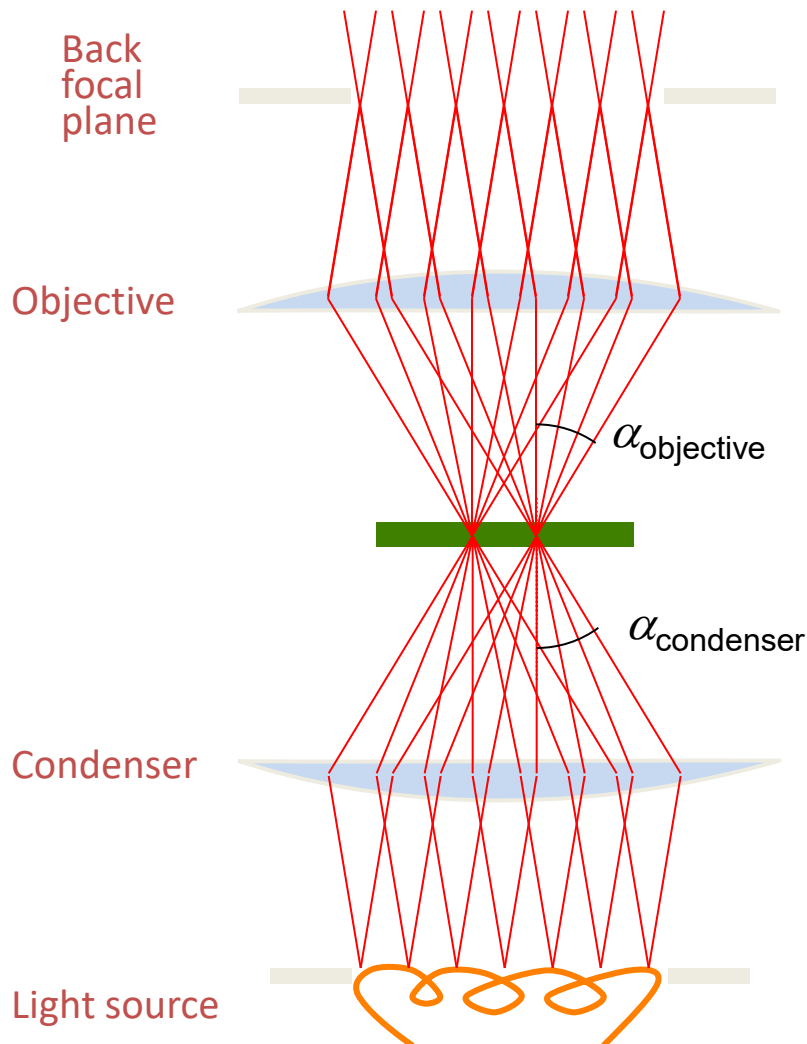
$$d_{min} = \lambda / (NA_{obj} + NA_{condenser})$$

→  $\lambda/2 NA$  if  $NA_{condenser} \geq NA_{obj}$  ("Filling the back focal plane")



# Filling the back focal plane

*In trans-illumination microscopy, to get maximum resolution, the illumination must “fill the back focal plane”*



For the highest resolution,  
we need to have

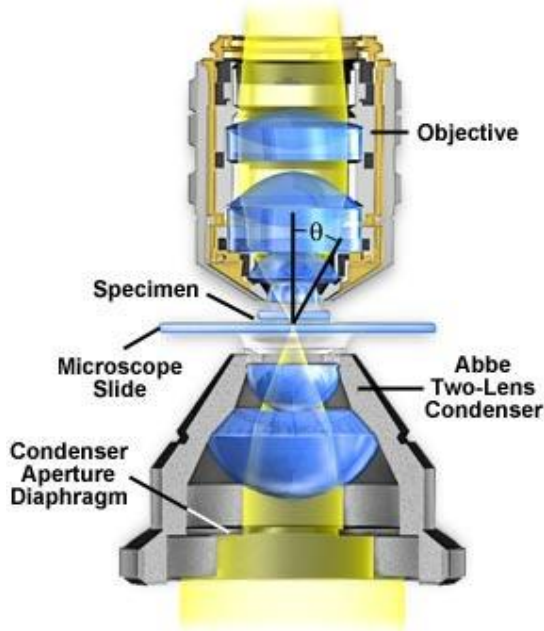
$$\alpha_{\text{condenser}} \geq \alpha_{\text{objective}}$$

$$NA_{\text{condenser}} \geq NA_{\text{objective}}$$

$\Rightarrow$  with oil immersion objectives,  
we need an *oil immersion condenser*!

# The Condenser

Abbe Condenser Optical Pathway



## Tasks:

- Illuminate at all angles  $< \alpha_{\text{objective}}$
- Concentrate light on the field of view for *all* objectives to be used

## Problem:

- Low mag objectives have large FOV,
- High mag objectives have large  $\alpha$   
(With 2X and 100x objectives we need  $(100/2)^2 = 2500$  times more light than any objective uses!)

## Solutions:

- Compromise
- Exchangable condensers, swing-out front lenses,...

Grade of correction

NA

Achromat/Aplanat Condenser (Numerical Aperture = 1.38)

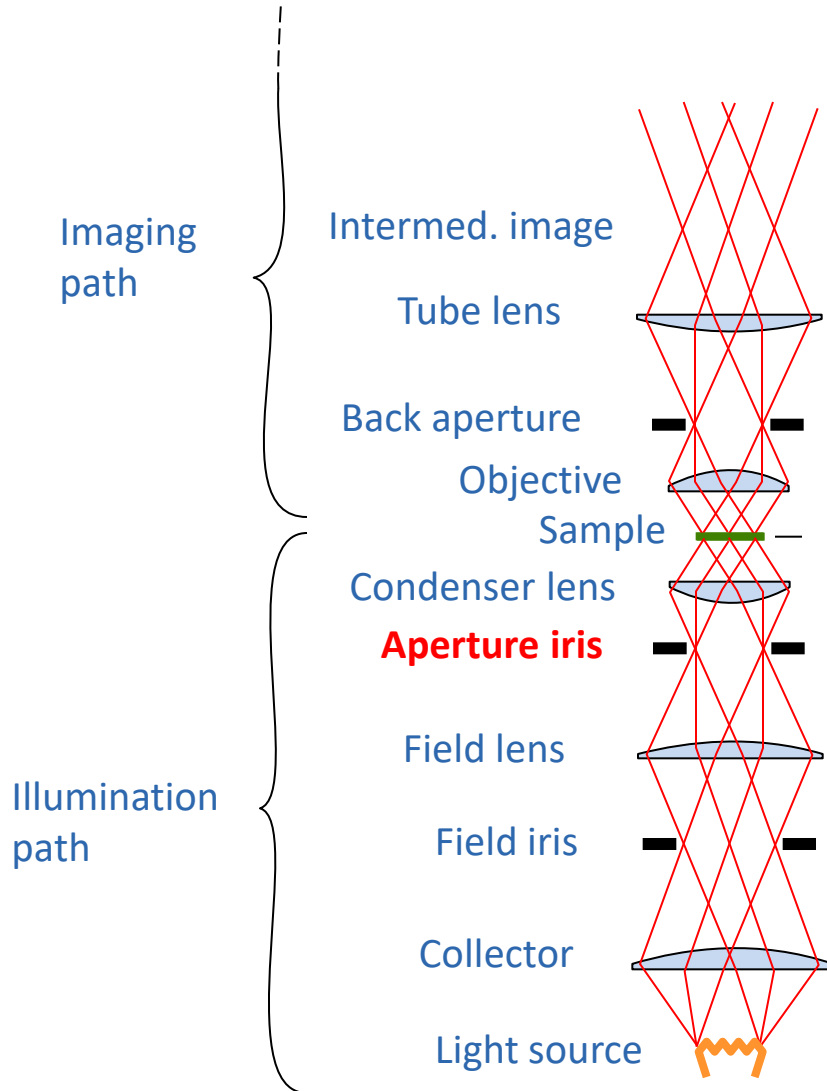


Swing-Out Top Lens Condenser (Numerical Aperture = 1.35)



# Aperture, Resolution & Contrast

Can adjust the condenser NA with the **aperture iris**

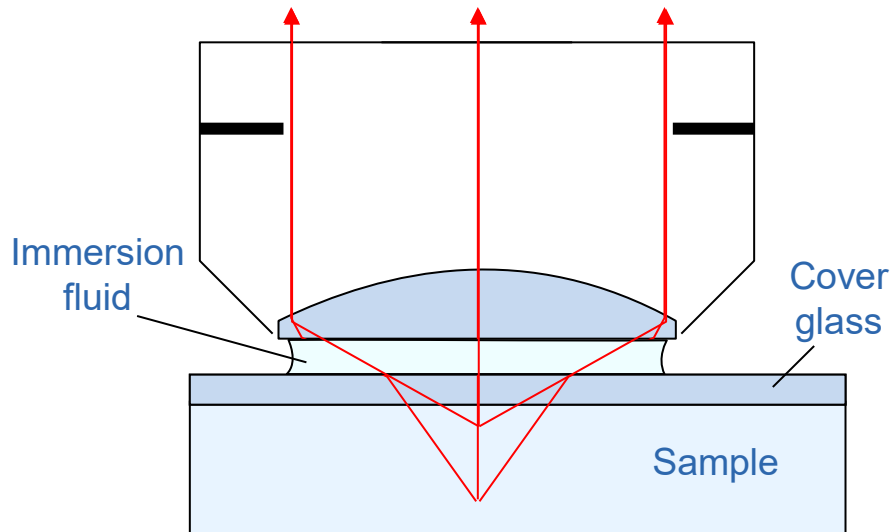


Q: Don't we always want it full open??

A: **No**

Why? Tradeoff:  
resolution vs. **contrast**

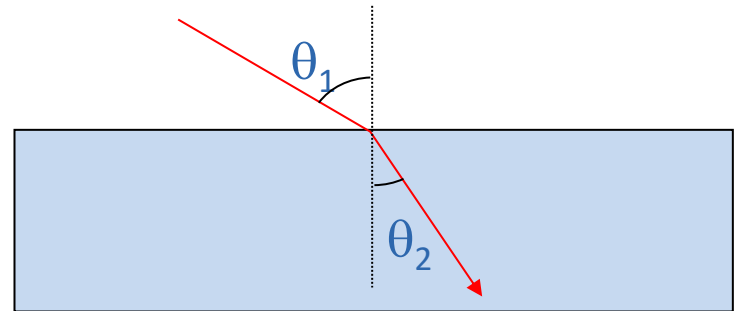
# Numerical Aperture and Resolution



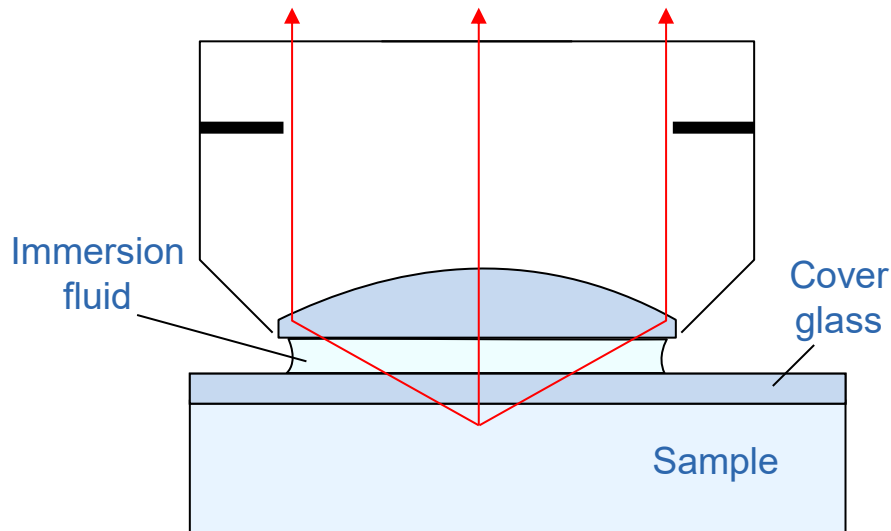
$$NA = n \sin(\theta)$$

$$d_{min} = \frac{\lambda}{NA} = \frac{\lambda}{n \sin(\theta)}$$

Snell's law:  
 $n_1 \sin(\theta_1) = n_2 \sin(\theta_2)$



# Numerical Aperture and Resolution



$$NA = n \sin(\theta)$$

$$d_{min} = \frac{\lambda}{NA} = \frac{\lambda}{n \sin(\theta)}$$

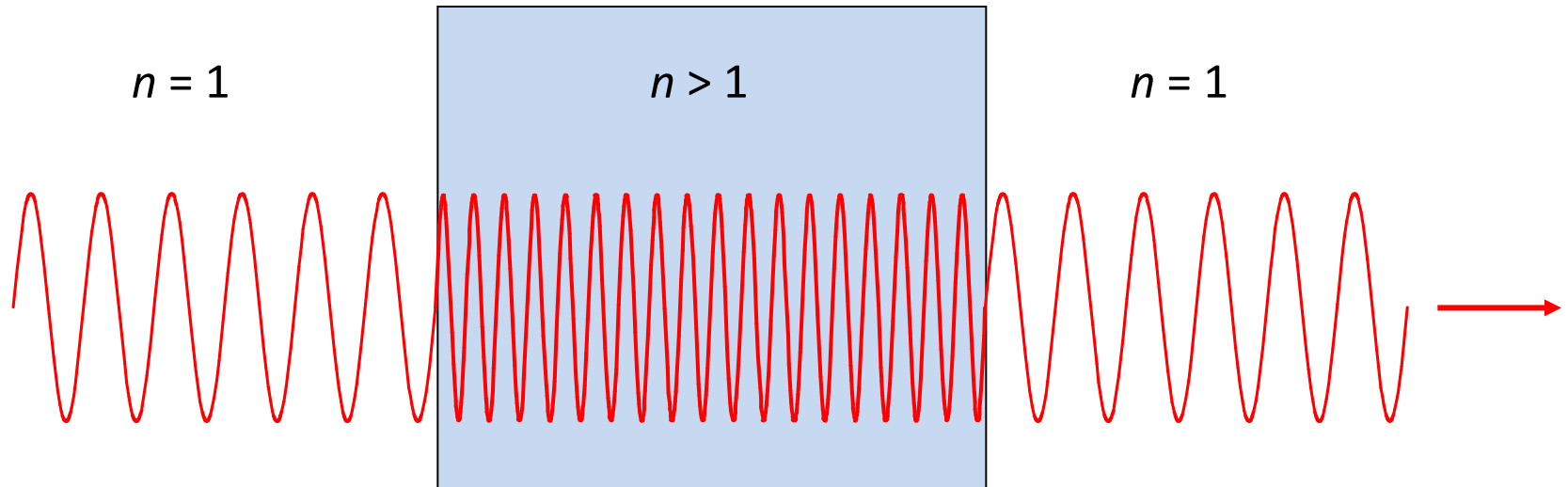
Sample, coverglass, immersion fluid, and top lens of objective all have same refractive index.

What happens if we change that refractive index?

Resolution improves with RI! Why?

# Light travels more slowly in matter

And the wavelength shortens



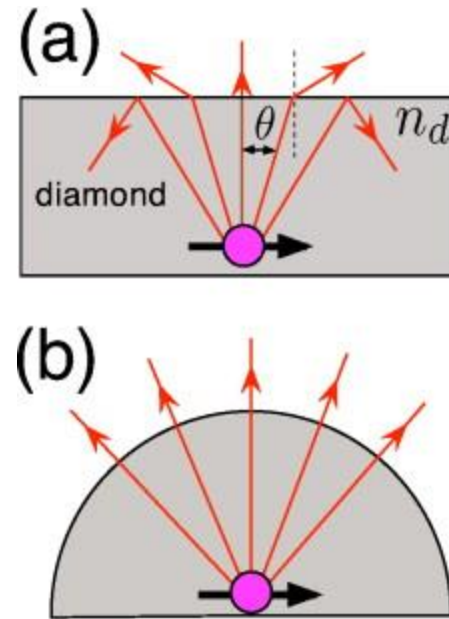
Recall Abbe's experiment:

$$d_{\min} = \lambda_{\text{sample}} / \sin(\alpha) = \lambda / n \sin(\alpha) = \lambda / NA$$

# Modifying RI to change resolution

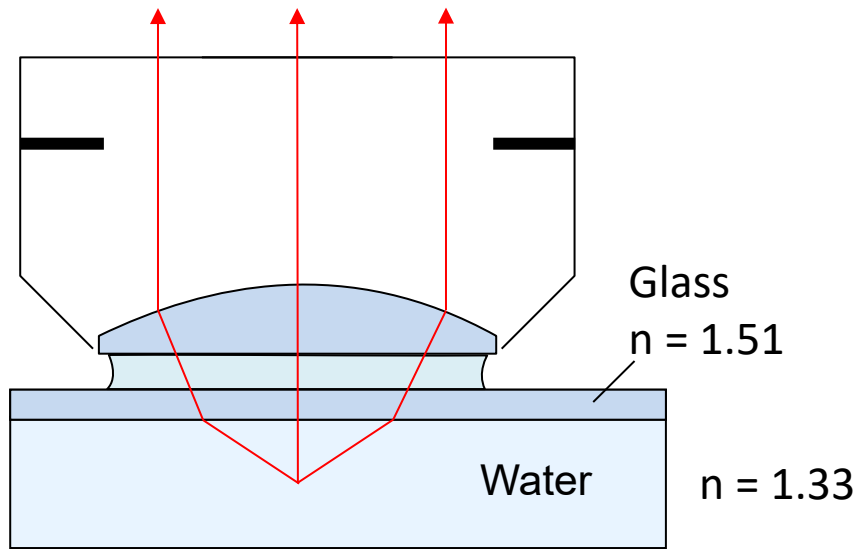


Olympus 1.65 NA TIRF lens:  
Uses sapphire coverslips ( $n=1.76$ ),  
diiodomethane ( $n=1.74$ )

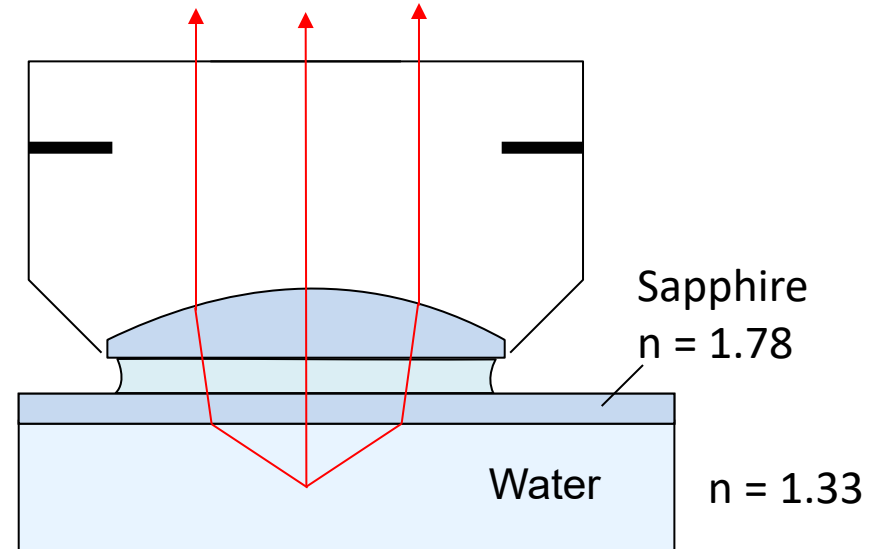


Diamond solid immersion lens  
Appl. Phys. Lett. 97, 241902 (2010)

# Why not use NA 1.65 lenses?



NA 1.4 oil immersion objective



NA 1.65 oil immersion objective

What's the largest angle we can collect?

$$\begin{aligned}n_1 \sin \theta_1 &= n_2 \sin \theta_2 \\1.33 \sin 90 &= n_2 \sin \theta_2 \\1.33/n_2 &= \sin \theta_2\end{aligned}$$

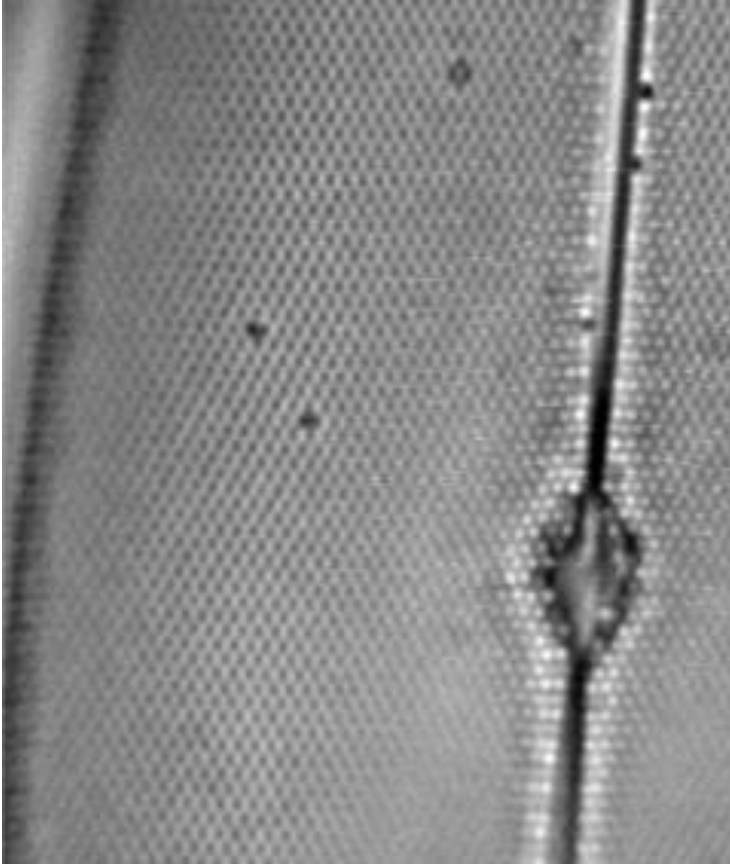


# Effective NA is limited by the sample

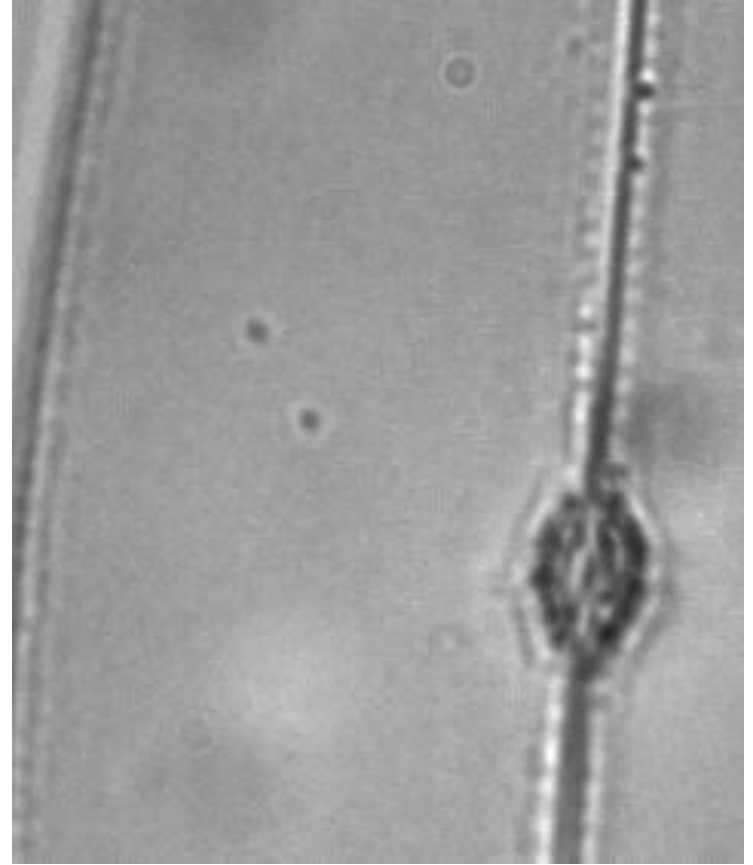
- To use a high NA objective, need a high RI mounting medium

Water	1.33
Glycerol	1.45
Vectashield	1.44
Prolong Gold	1.39 – 1.46
2,2-thiodiethanol	1.52
Methyl Salicylate	1.53
Benzyl benzoate	1.57

# NA and Resolution



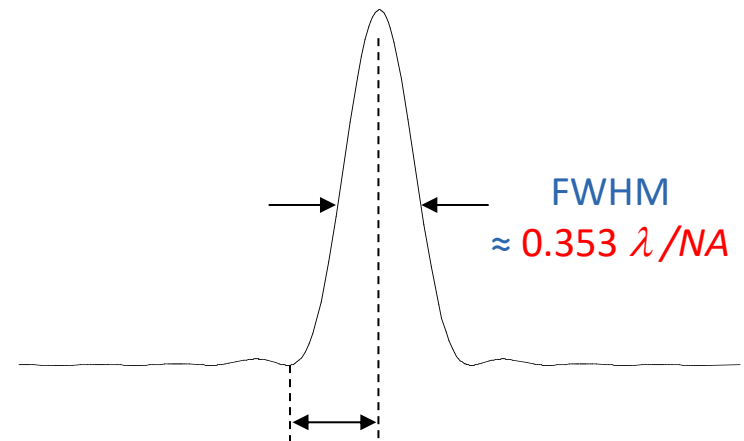
High NA Objective



Low NA Objective

# Alternate Definitions of Resolution

As the Full Width at Half Max  
(FWHM) of the PSF



As the diameter of the Airy disk  
(first dark ring of the PSF)  
= “Rayleigh criterion”

(Probably most common definition)

# Objective Types



## Basic properties

- Magnification
- Numerical Aperture (NA)
- Infinite or finite conjugate
- Cover slip thickness if any
- Immersion fluid if any

## Correction class

- Achromat
- Fluor
- Apochromat

## Field flatness

- Plan or not

## Phase rings for phase contrast

- Positive or negative
- Diameter of ring (number)

## Special Properties

- Strain free for Polarization or DIC

## Features

- Correction collar for spherical aberration
- Iris
- Spring-loaded front end
- Lockable front end

## Further reading

[www.microscopyu.com](http://www.microscopyu.com)

[micro.magnet.fsu.edu](http://micro.magnet.fsu.edu)

Michael W. Davidson and Douglas B. Murphy  
“Fundamentals of Light Microscopy and Electronic Imaging”

James Pawley, Ed. “Handbook of Biological Confocal  
Microscopy, 3rd ed.”

## Acknowledgements

Ron Vale / Mats Gustafsson / Steve Ross