



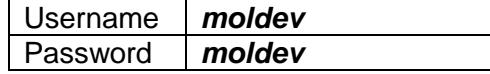
ImageXpress® Micro Confocal & MetaXpress® 6.5

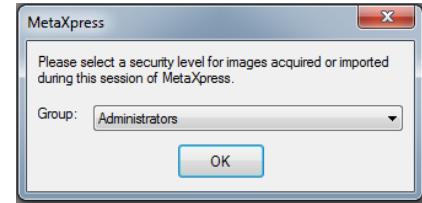
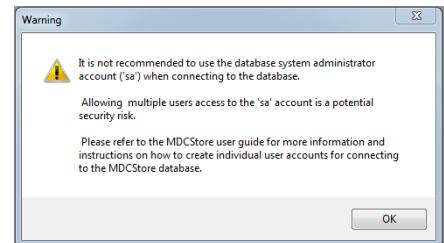


The purpose of this guide is to briefly describe:

- I. Turn on system and acquire plate with saved settings (p. 1)
- II. Test acquisition settings (p. 4)
- III. Define new acquisition settings (p. 6)
- IV. View images and run an analysis (p. 14)

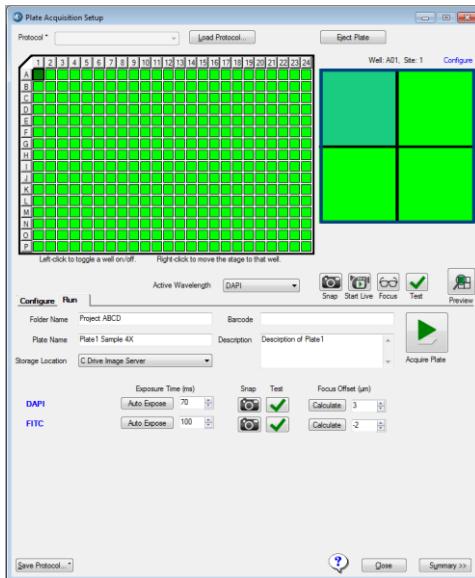
I. Turn on System and Acquire Plate with Saved Settings

1.	Turn on the system: <ul style="list-style-type: none">• IXM power supply controller box• Computer and Monitor
2.	Go to the MetaXpress folder and double-click on the appropriate hardware profile shortcut
3.	Login to MDCStore database with username and password  *NOTE* Your database, username, and password may be different. Refer to your administrator for this information
4.	If you log in as system administrator (sa), the next window is a warning regarding security risks; click OK
5.	Group (security level) and click OK



In the main toolbar, click  Acquisition Setup or in the main menu select **Screening > Acquisition Setup**

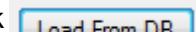
6.

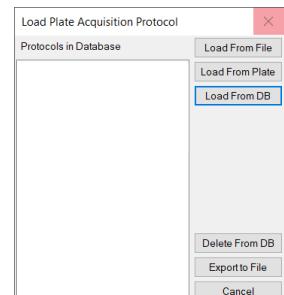


7.

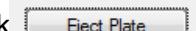
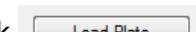
To load a previous saved protocol, click on  in **Plate Acquisition Setup**

8.

- Click  to search windows for the appropriate .hts file.
- If the settings file is saved to the database, highlight the protocol and click 
- If no settings have been saved, protocols can be loaded from an existing plate by clicking 

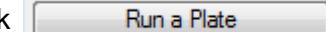
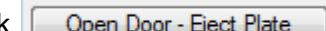
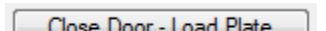


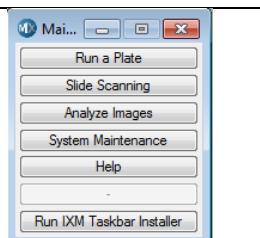
9.

- Click  to open the door and place the plate in the system
- Click  to close the door

10.

Alternatively, you can use the **Main Taskbar** to open and close the door.

- Click 
- Click  or 



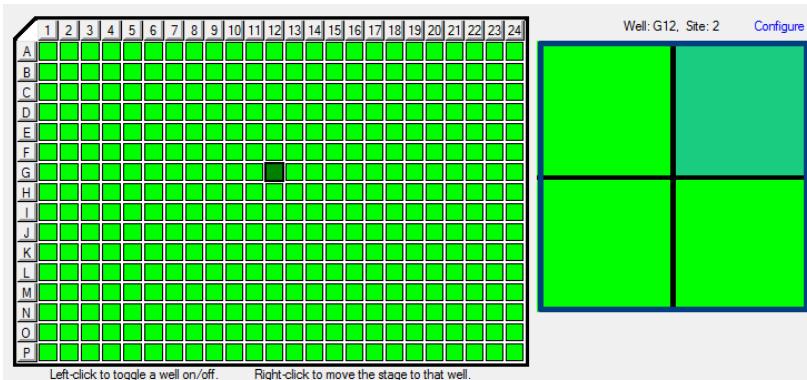
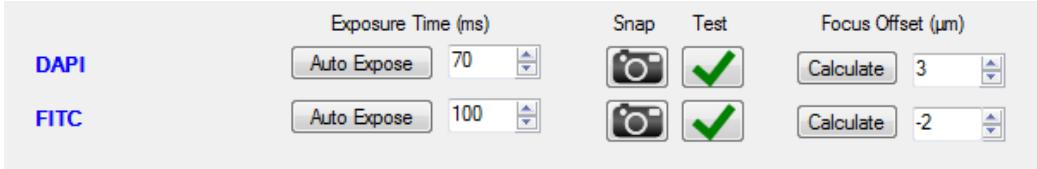
On the **Run** tab, update the folder name, plate name and description as desired

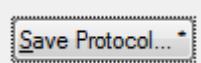
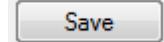
11.

Folder Name	Plate 1 Sample	Barcode
Plate Name	Plate 1 Sample MMDDYY	Description
Storage Location	C Drive Image Server	Spheroids stained with DAPI-Hoechst and FITC-Actin

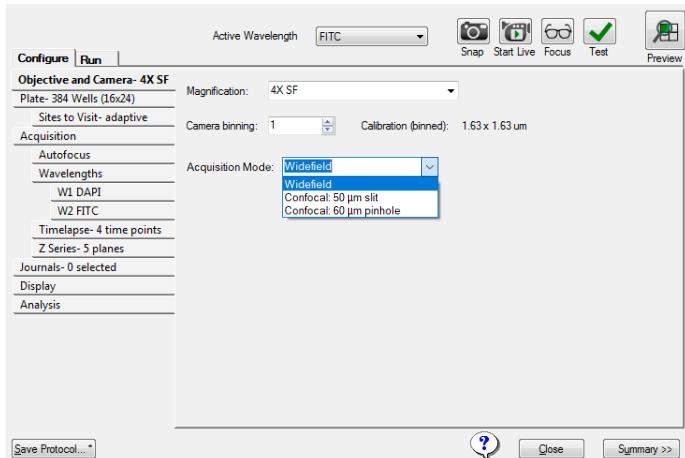
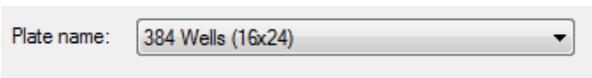
12. Click  to begin acquiring the plate

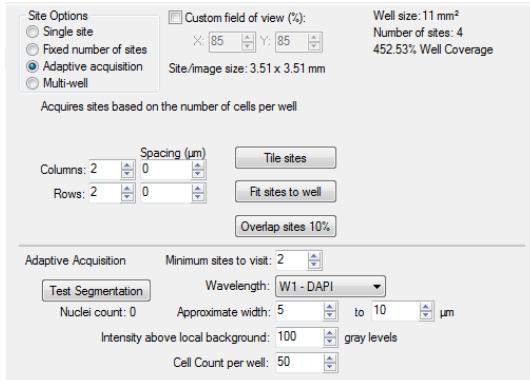
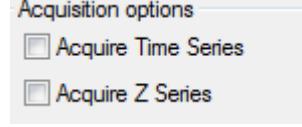
II. Test Acquisition Settings

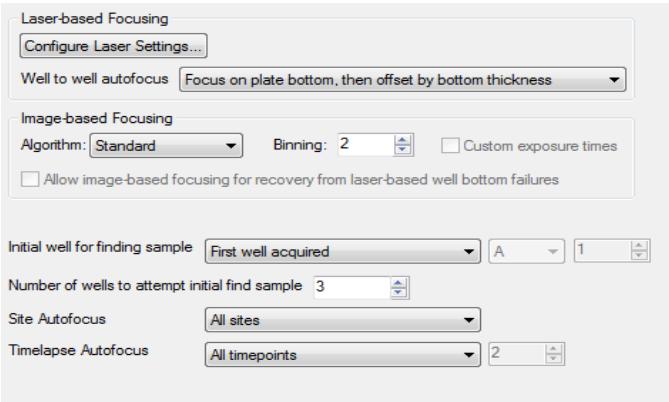
1.	Open Plate Acquisition Setup
2.	In the plate and site section of Plate Acquisition Setup , right-click on the desired well and/or site to move the plate to that position (indicated by a dark green color)  <p>Left-click to toggle a well on/off. Right-click to move the stage to that well.</p>
3.	Test the acquisition settings by clicking: <ul style="list-style-type: none"> to perform a large-range autofocus and snap image routine to perform a focus and snap image routine (if Z series has been activated, all planes will be acquired) to perform an autofocus and snap image routine all for all wavelengths (if Z series has been activated, all planes will be acquired)
4.	Adjust the acquisition settings, if necessary, within the Run tab: <ul style="list-style-type: none">Adjust the focus offset by clicking Calculate or adjust the number manuallyAdjust the exposure time by clicking Auto Expose or change the number manually  <p>*NOTE* Click on the wavelength name to open the corresponding wavelength tab for advanced options</p>

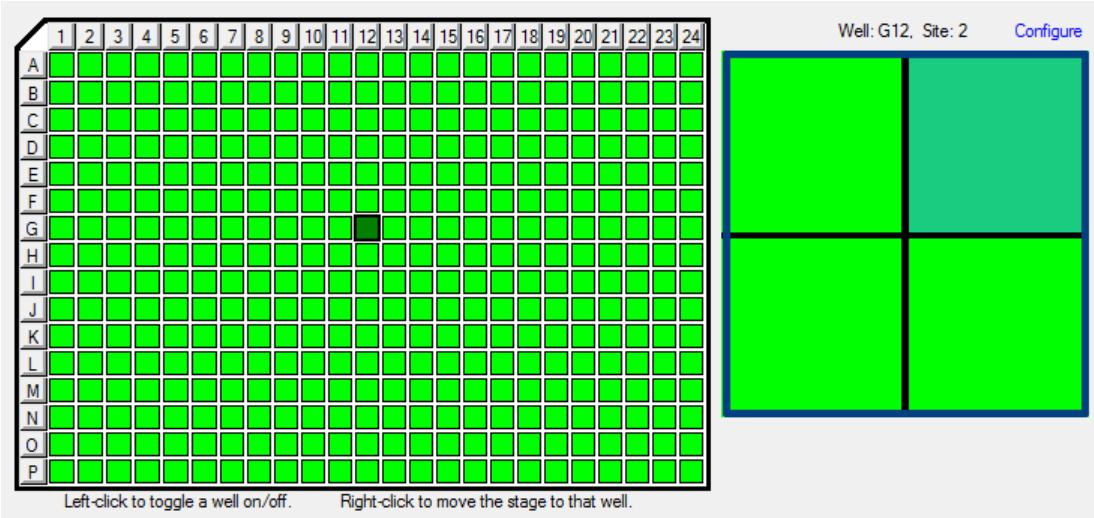
	When you have optimized settings, click 
5.	<ul style="list-style-type: none">Molecular Devices recommends enabling <input type="checkbox"/> Save to file rather than databaseClick  to search for a location on the hard drive.
6.	Click  to begin acquiring the plate

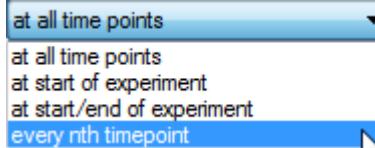
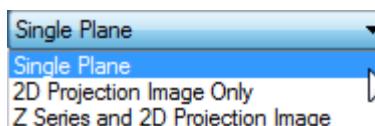
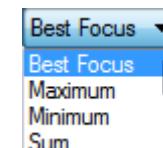
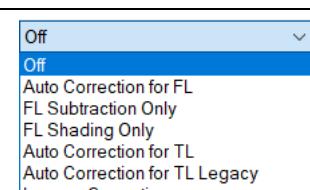
III. Define New Acquisition Settings

1.	Open Plate Acquisition Setup
2.	Select the Configure tab
3.	<p>Select the Objective and Camera tab</p> <p>i. Select the appropriate magnification from the drop-down menu</p> <p>ii. Set binning (2 for cell counting and cell scoring; 1 for fine sub-cellular detail)</p> <p>iii. Select Acquisition Mode: Widefield or Confocal</p> <p>3 confocal options depending on system configuration: 50 µm slit, 60 µm pinhole, 42 µm pinhole</p> 
4.	<p>Adjust the objective correction collar, if necessary, (setting on objective should match physical plate bottom thickness in mm).</p> <p>On the Run a Plate Taskbar, click on Adjust Correction Collar to step through the process.</p>
5.	<p>Select the Plate tab and select the appropriate plate type from the drop-down list</p> 

	<p>Select the Sites to Visit tab and select the appropriate number of sites</p> <ul style="list-style-type: none"> Single Site: image one site per well in the center Fixed number of sites: image the number of selected sites for every well. Adjust number and spacing of sites. Left-click on sites to select (green) and deselect (grey). Right-click on any site to move the plate to that site position (dark green) Adaptive acquisition: collect the minimum number of sites to image at least the cell count indicated by the user. The Adaptive Acquisition section will appear allowing the user to choose wavelength, size and threshold settings, and desired minimum count for cells Multi-well: collect multiple wells within one image which is then cropped to define single wells automatically Custom field of view (%): reduce the size of each image by the percentage entered. This is useful when the field of view covers more than the site/well area desired
6.	 <p>The screenshot shows the 'Site Options' dialog box. Under 'Site Options', 'Adaptive acquisition' is selected. Other options include 'Single site', 'Fixed number of sites', and 'Multi-well'. A 'Custom field of view (%)' checkbox is checked, with X: 85 and Y: 85. 'Well size: 11 mm²' and 'Number of sites: 4' are displayed. Below this, 'Site./Image size: 3.51 x 3.51 mm' and '452.53% Well Coverage' are shown. Under 'Acquires sites based on the number of cells per well', 'Columns: 2' and 'Rows: 2' are set to 0. Buttons for 'Tile sites' and 'Fit sites to well' are present. An 'Overlap sites 10%' button is also visible. In the 'Adaptive Acquisition' section, 'Minimum sites to visit: 2' is set. Under 'Test Segmentation', 'Wavelength: W1 - DAPI' is selected. 'Nuclei count: 0', 'Approximate width: 5 to 10 μm', 'Intensity above local background: 100 gray levels', and 'Cell Count per well: 50' are also specified.</p>
7.	Select the Acquisition tab to select Autofocus and Acquisition options
8.	<p>Autofocus options:</p> <ul style="list-style-type: none"> Always select Enable laser-based focusing Enable image-based focusing for thick samples or those with different focal planes from site-to-site or well-to-well  <p>The screenshot shows the 'Autofocus options' dialog box. It contains two checkboxes: 'Enable laser-based focusing' (checked) and 'Enable image-based focusing (for acquisition or laser recovery)' (unchecked).</p>
9.	<p>Acquisition options:</p> <ul style="list-style-type: none"> Enable Acquire Time series for timelapse experiments Enable Acquire Z series for Z step acquisition  <p>The screenshot shows the 'Acquisition options' dialog box. It contains two checkboxes: 'Acquire Time Series' (unchecked) and 'Acquire Z Series' (unchecked).</p>

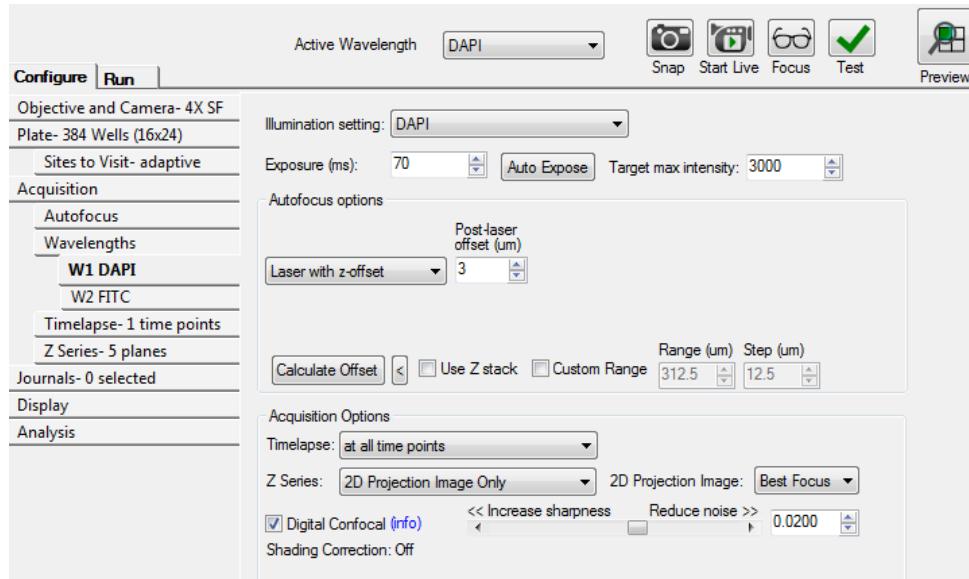
	<p>Other options:</p> <ul style="list-style-type: none"> • If running a journal during acquisition, enable this option to activate the Journals tab • If an analysis has already been setup, enable Analyze Images After Acquisition 	<input type="checkbox"/> Run Journals During Acquisition <input type="checkbox"/> Analyze Images After Acquisition <input type="checkbox"/> Allow Appending to Existing Plate Directory for Stored Correction Images... C:_
10.	<p>*NOTE* this requires an offline computer to be in Auto-run mode or running PowerCore software</p> <ul style="list-style-type: none"> • To enable appending time points, enable Allow Appending to Existing Plate • If using the Legacy Correction shading correction option for any wavelengths, click Directory for Stored Correction Images and select the appropriate directory where shading correction images are saved 	
	<p>Select the Autofocus tab:</p> <ol style="list-style-type: none"> i. Set Well to well autofocus to Focus on well bottom This is the default acquisition setup, however when imaging thin-bottom plates with low magnification objectives (4x and below) or microscope slides, select Focus on plate bottom, then offset by bottom thickness ii. For Image-based Focusing refer to corresponding MetaXpress 6 Software Guide modules for suggested settings iii. Set Initial well for finding sample to First well acquired iv. Set Number of wells to attempt initial find sample to 3 v. If more than one site is acquired, set Site Autofocus to All sites vi. If timelapse is enabled, set Timelapse Autofocus to All timepoints for long term timelapse, and First timepoint only for fast kinetic experiments 	

12.	<p>Select the Wavelengths tab and select the number of wavelengths (colors) including transmitted light that you would like to acquire</p>	<p>Number of wavelengths: <input type="text" value="2"/></p>
13.	<p>In the plate and site section of Plate Acquisition Setup, right-click on the desired well (typically a control well) and/or site to move the plate to that position (indicated by a dark green color)</p>  <p>The image shows a 96-well plate grid with wells labeled A through P on the left and 1 through 24 at the top. A dark green square in the 12th column of the G row indicates it is selected. To the right is a zoomed-in view of Well G12, Site 2, which is also dark green. Below the grid are two instructions: "Left-click to toggle a well on/off." and "Right-click to move the stage to that well."</p>	
14.	<p>Select the W1 (wavelength) tab</p> <ol style="list-style-type: none"> i. Select the desired filter set from the drop-down menu under Illumination setting ii. Click  Focus iii. Examine the image <ul style="list-style-type: none"> a. If the image appears to be dim or saturated, first adjust the image scaling, then adjust exposure time if necessary b. If a blank or snowy image appears, this can indicate that a plate is not in the system or laser autofocus settings are incorrect iv. Click the Calculate Offset to perform an automatic focus determination <ul style="list-style-type: none"> a. For more control, enable <input type="checkbox"/> Use Z stack and follow the prompts b. If necessary, enable <input type="checkbox"/> Custom Range <input type="text" value="312.5"/> <input type="text" value="12.5"/> v. Click  again to test the new post-laser offset. Image should now be in focus. vi. Examine the image for brightness <ul style="list-style-type: none"> a. If necessary, click Auto Expose with Target max intensity: 45000 set to 33000 – 45000 b. You can also increase or decrease exposure manually 	

	If acquiring a Timelapse, select how often to acquire this image from the drop-down menu
15.	
	If acquiring a Z Stack, select the appropriate setting for image collection
16.	 <p>*NOTE* Z Series and 2D Projection Image is not available when acquiring a Timelapse</p> <p>If saving the 2D Projection Image, select the appropriate projection method (press F1 for more information)</p>  <p>*NOTE* Best Focus is not recommended for comparison of intensity measurements</p>
17.	If the option is available, you can enable Digital confocal and select the appropriate K value using the slider bar (press F1 for more information)
18.	<p>Apply a shading correction option for your wavelength, if needed.</p> <p>For Fluorescent wavelengths:</p> <p>FL Shading Only generally works well for most assays.</p> <p>Other options include Auto Correction for FL, FL Subtraction Only, Legacy Correction (requires the use of preset reference images), or Off (no shading correction)</p> <p>For Brightfield or Phase Contrast wavelengths:</p> <p>Auto Correction for TL generally works well for most assays.</p> <p>Other options include Auto Correction for TL Legacy, Legacy Correction (requires the use of preset reference images), or Off (no shading correction)</p> 

Repeat for each subsequent wavelength

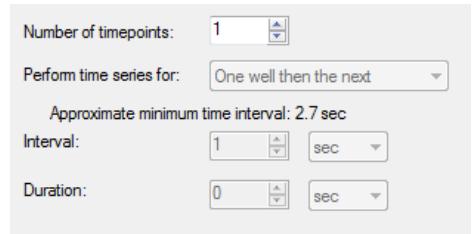
19.



If acquiring with Timelapse, select the **Timelapse** tab

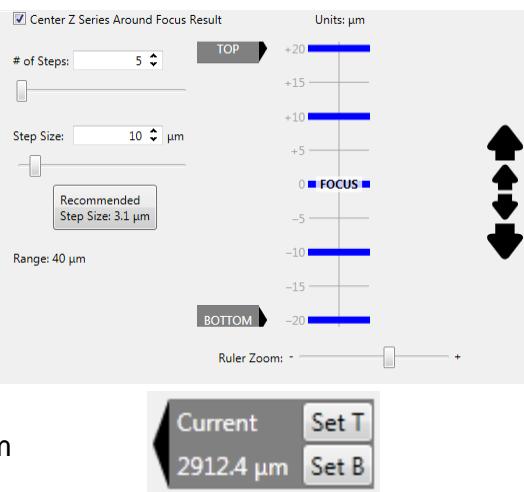
20.

- Enter the number of **Time points** desired
- Set **Interval** as the time between each time point
- Set **Duration** as the total time of the experiment
- Set **Perform time series** for:
 - One well then the next:** entire timelapse is run for one well before acquiring next well
 - One column then the next:** entire timelapse is run for one column before the next
 - One row then the next:** entire timelapse is run for one row before acquiring next row
 - All selected wells:** all wells are imaged before continuing with next time point



If acquiring a Z Series, select the **Z Series** tab

- i. Deselect Center Z Series Around Focus Result
- ii. Adjust Step size for spacing between each Z plane
- iii. Click  to determine the Z start position
- iv. Click  to start **Live Mode**
- v. Use the large and small arrows to move to the top of the focus range for the sample and click 
- vi. Use the large and small arrows to move to the bottom of the focus range for the sample and click 
- vii. Click  to stop **Live Mode**
- viii. Click  to perform focus and snap image routine to acquire all Z steps for the active wavelength. The last image in the stack will be the selected 2D projection image.
- ix. Click  to perform focus and snap image routine to acquire all Z steps for all wavelengths. The last image in the stack will be the selected 2D projection image.



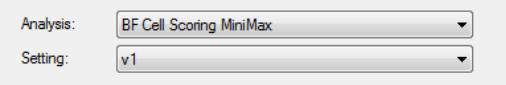
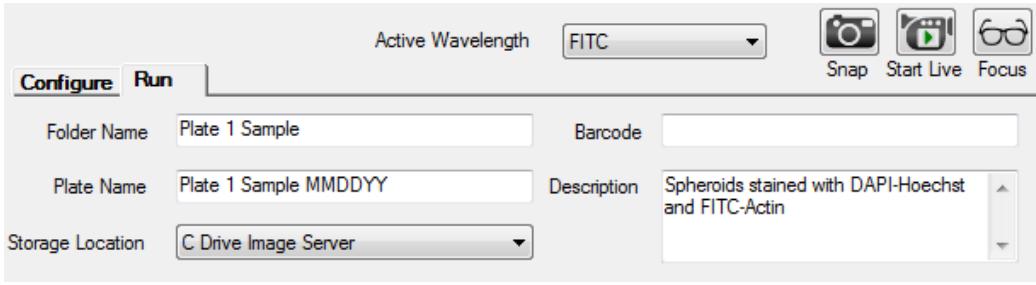
Select the **Journal** tab (enabled on acquisition tab) and activate journals where appropriate.

It might also be necessary to enable

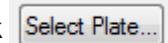
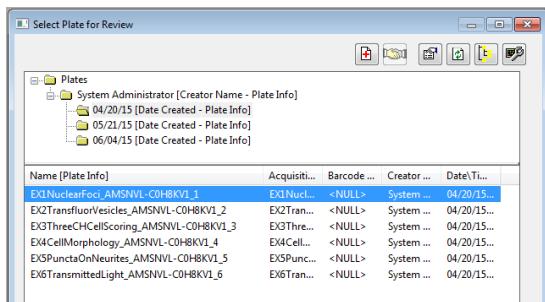
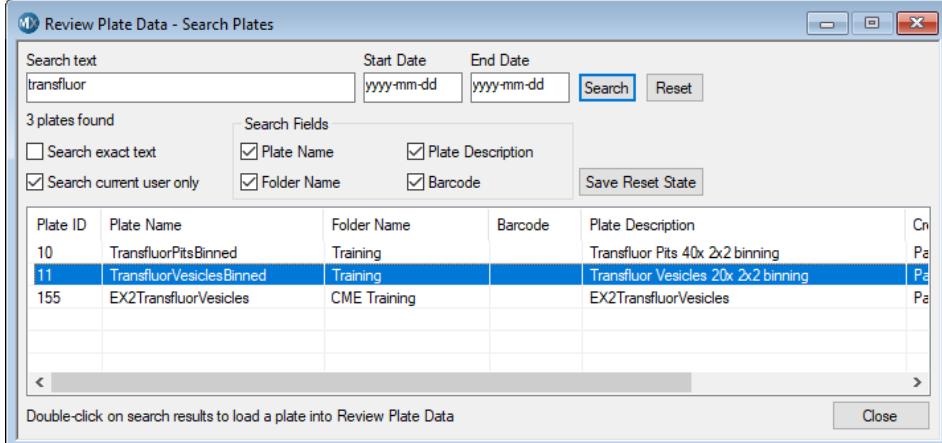
- Prevent asynchronous hardware moves
(recommended if any journals are dependent on hardware positioning).

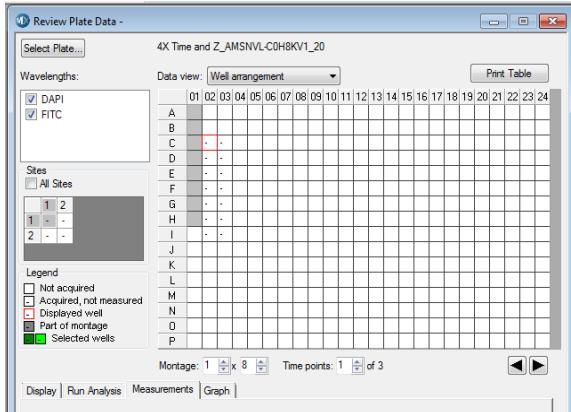
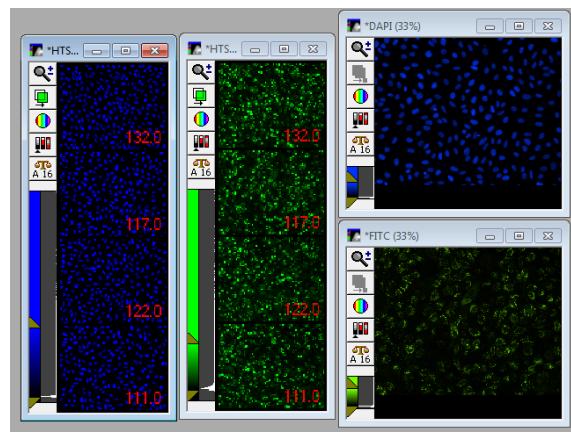
for certain journals (refer to documentation accompanying journals for details)

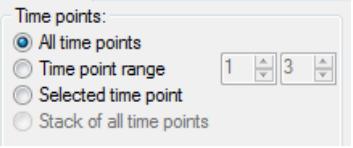
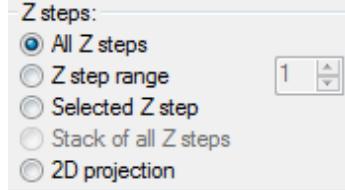
Acquisition Step	Journal
<input type="checkbox"/> Before each image	[None]
<input type="checkbox"/> After each image	[None]
<input type="checkbox"/> Before focusing	[None]
<input type="checkbox"/> Start of z	[None]
<input type="checkbox"/> End of z	[None]
<input type="checkbox"/> Start of site	[None]
<input type="checkbox"/> End of site	[None]
<input type="checkbox"/> Start of well	[None]
<input type="checkbox"/> End of well	[None]
<input type="checkbox"/> Start of time point	[None]
<input type="checkbox"/> End of time point	[None]
<input type="checkbox"/> Start of plate	[None]
<input type="checkbox"/> End of plate	[None]
 <input type="checkbox"/> Prevent asynchronous hardware moves (recommended if any journals are dependent on hardware positioning).	

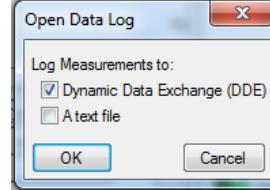
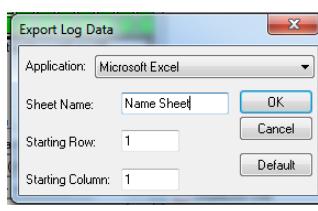
	Select the Display tab to:
23.	<ul style="list-style-type: none"> • <input type="checkbox"/> Auto Arrange Images will use default settings to arrange displayed images • <input type="checkbox"/> Display Acquisition Layout manually adjust image display prior to acquisition <ul style="list-style-type: none"> • Enable <input checked="" type="checkbox"/> Display images during autofocus • Enable <input checked="" type="checkbox"/> Display images during acquisition • (Optional) Enable <input checked="" type="checkbox"/> Display a color overlay of wavelength images during acquisition
24.	<p>Select the Analysis tab (enabled on the acquisition tab) to specify the appropriate optimized Analysis routine and Settings from the drop down-menus</p> <p>*NOTE* This requires an offline computer set in Auto-run mode or running PowerCore software</p> 
25.	<p>Under the Run tab, enter:</p> <ul style="list-style-type: none"> • Folder Name: Project name, your name, PI, etc. All your plates will go under this name. • Plate Name: Name of this experiment • Storage location: Select appropriate server for image storage. *NOTE* There may only be one choice. • Barcode: Enter a barcode if desired • Description: Any text regarding the experiment 
26.	<p>When you have optimized settings, click Save Protocol...</p> <ul style="list-style-type: none"> • Molecular Devices recommends enabling <input type="checkbox"/> Save to file rather than database • Click Save for a location on the hard drive.
27.	<p>Click on  to begin acquiring the plate</p>

IV. Review Images and Run an Analysis

1.	In the main toolbar, click  or in the main menu select Screening > Review Plate
2.	On the Review Plate Data dialog, click 
3.	<p>Navigate through the folders to find the plate of interest.</p> <p>Highlight the plate and click </p> 
4.	<p>If you cannot find your plate, on the Review Plate Data dialog, click </p> <ul style="list-style-type: none"> Enter the Search text and optional date range Enable or disable search options Click Search. Highlight the plate and double-click it to open. <p>*NOTE* Search results can remain open, in case multiple plates need to be checked.</p> 

	In the Plate View section, you will see a ‘-’ in each well that was imaged. Left-click and drag across the wells you want to view. A thumbnail montage of these wells will open for each wavelength.
5.	
6.	If there are multiple sites per well, select an appropriate site to view, or enable All Sites . The image montages will automatically adjust. 
7.	To view all Timelapse or Z Series images at once, change the Data view to Data view: Time Point vs Well or Data view: Z Step vs Well, respectively.
8.	Left-click on a single thumbnail to view full resolution images (all wavelengths) 
9.	To run or set up an analysis, select the Run Analysis tab 

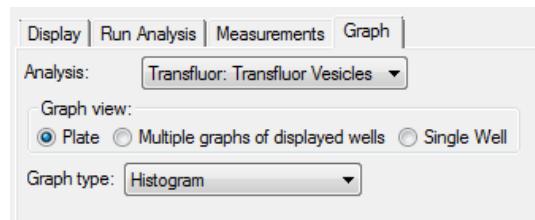
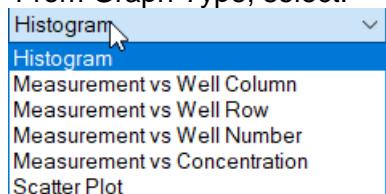
10.	<p>If analysis settings have already been optimized, select the analysis routine (application module, custom module, or journal) and settings from the drop-down menus</p>	<p>Analysis: <Angiogenesis Tube Formation> Settings: No Settings Configured Setting description: Measures angiogenesis tube formation.</p>
11.	<p>Under the Run Analysis tab, select the appropriate button to run the analysis:</p> <ul style="list-style-type: none"> • Run on all wells analysis will be run on all acquired images • Run on selection analysis will be run on selected wells (selected wells are indicated in green; to select wells, right click well(s) in the plate section or image montage) • Run on displayed site analysis will be run only on the currently displayed site 	<p>Legend</p> <ul style="list-style-type: none"> □ Not acquired - Acquired, not measured - Displayed well ■ Part of montage ■ Selected wells
12.	<p>For a Timelapse data set, select the appropriate option for analysis under the Time points section</p> <ul style="list-style-type: none"> • All time points: run analysis on all time points in the data set • Time point range: run analysis on a consecutive range of time points • Selected time point: run analysis on only one time point that is selected in the Time point section below the plate layout <p>Time points: 1 of 3</p> <ul style="list-style-type: none"> • Stack of all time points: use if, in the Analysis field, you select a timelapse journal which analyzes the planes in a stack 	
13.	<p>For a Z Series data set where all Z planes were saved, select the appropriate option in the Z steps section</p> <ul style="list-style-type: none"> • All Z Steps: run analysis on all Z planes • Z Step range: run analysis on consecutive range of Z planes • Selected Z step: run analysis on only one Z plane that is selected in the Z step section below the plate layout <p>Z steps: 3 of 5</p> <ul style="list-style-type: none"> • Stack of all Z steps: run an analysis with a journal that requires a stack of images • 2D projection: only run analysis on the saved 2D projection image 	

14.	<p>If the selected analysis has already been run on the plate, a warning will appear asking to overwrite the data. If you are not sure, save the analysis settings with a new name before analyzing your plate.</p>
15.	<p>To view analysis results, select the Measurements tab</p> <ol style="list-style-type: none"> Select the Analysis (module and settings name) from the drop-down menu Select a measurement from the drop-down menu. The values will be shown in the plate layout. <ul style="list-style-type: none"> Measurements starting with a “Cell” are cell-by-cell data and will give the average of all cells in the displayed site(s) for the well Activate the heat map by enabling <input checked="" type="checkbox"/> Show Heat Map Configure the heat map by clicking on Heat Map... <p>*NOTE* In the plate view, summary measurements, such as counts, are displayed as an average of all sites in the well, rather than a sum. To obtain sum values, the data can be exported via Plate Data Utilities.</p> 
16.	<p>To view the cell-by-cell data, click Cellular Results... at the bottom of the Review Plate Data dialog. Data will be automatically updated based on the well and site selected in the montage view</p>
17.	<p>To export data to Excel:</p> <ol style="list-style-type: none"> On the Measurements tab, click on Open Log Select only Dynamic Data Exchange Select Microsoft Excel and name worksheet as desired. This opens an empty worksheet. Click Log Data. Currently viewed data will be logged into the Excel sheet.  

18.

To create simple graphs in MetaXpress:

- i. Go to **Graph** tab
- ii. From Graph Type, select:



- iii. Select measurements to plot from the drop-down menu
- iv. Click **Show Graph**
- v. Right-click on the graph for more options

NOTE For Measurement vs Concentration, the plate must first be annotated.