

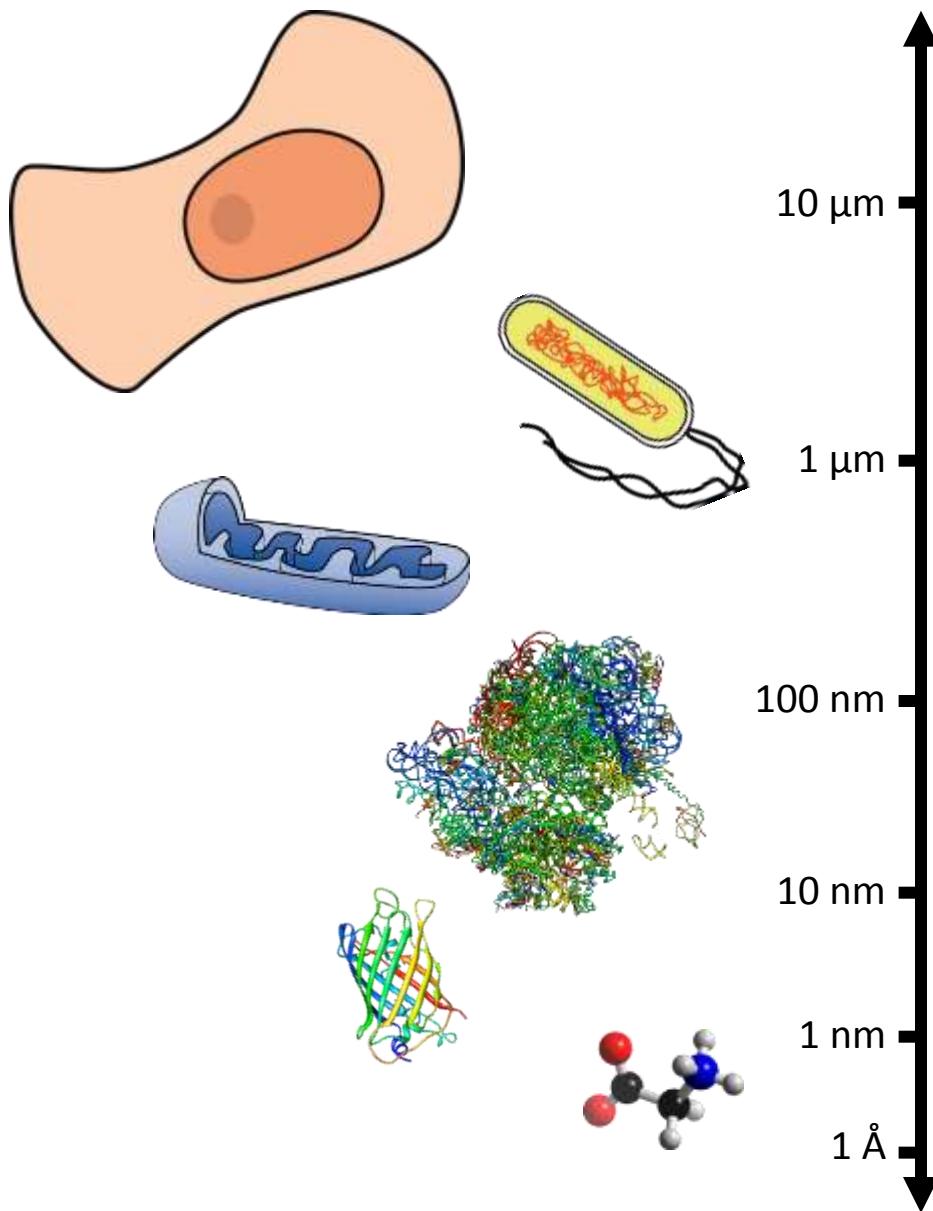
Super-Resolution Microscopy

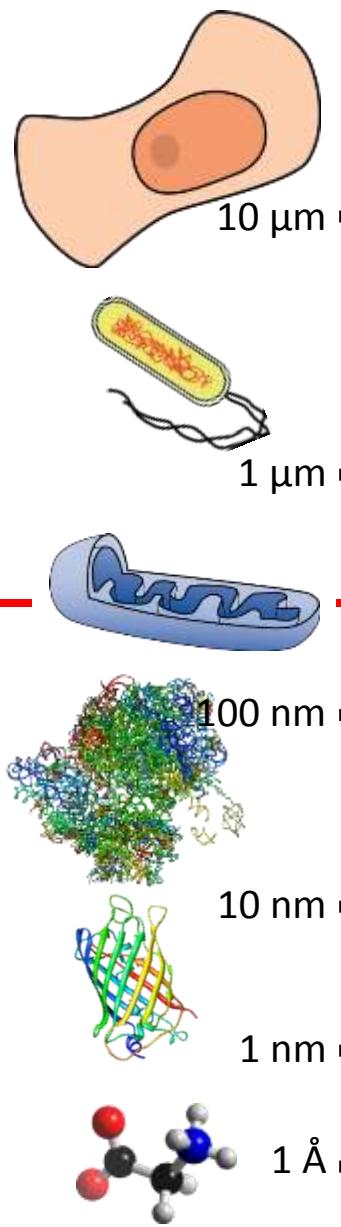
Structured Illumination



Bo Huang

Looking into microscopic world of life...

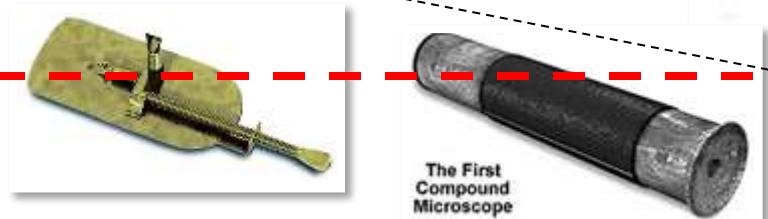




Naked eye: $\sim 50\text{-}100 \mu\text{m}$

★ 1595, Zaccharias and Hans Janssen
First microscope, 9x magnification

★ Antonie van Leeuwenhoek
(1632-1723), 200x



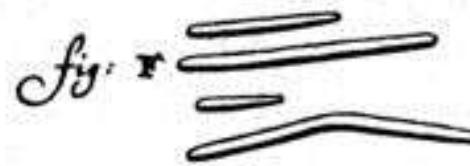
Compound microscope
 $>1000\times$

PLATE XXIV

fig. A

fig. B

Ernst Abbe (1840-1905)
The “physical” diffraction limit

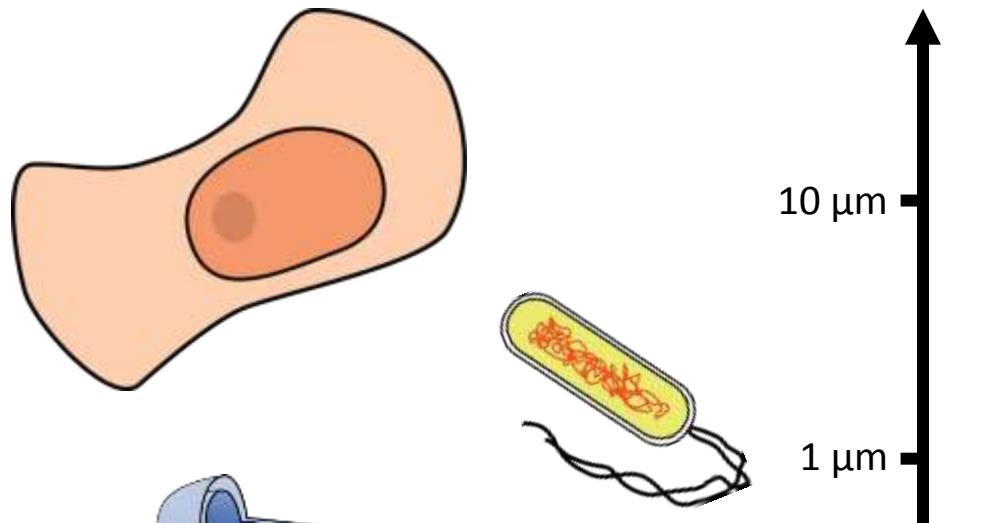


Timeline (horizontal axis):

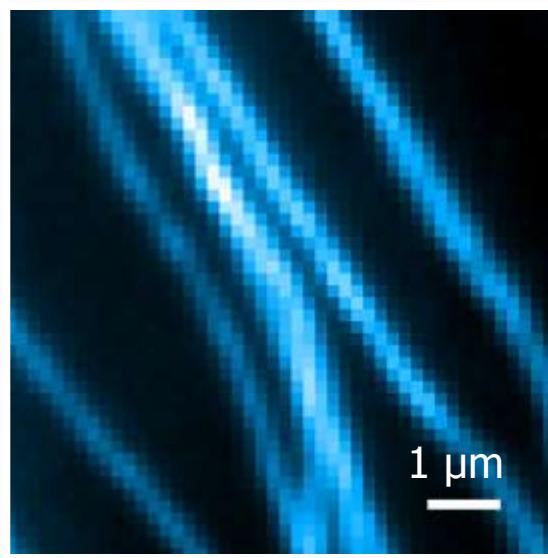
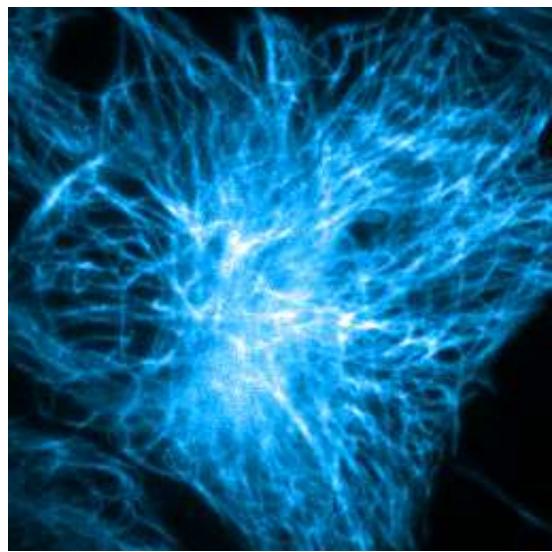
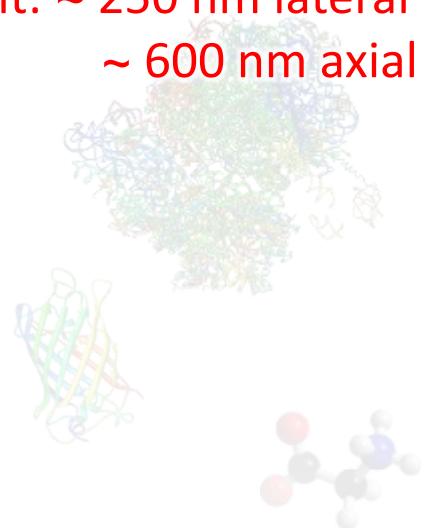
1600 1700 1800 1900 2000

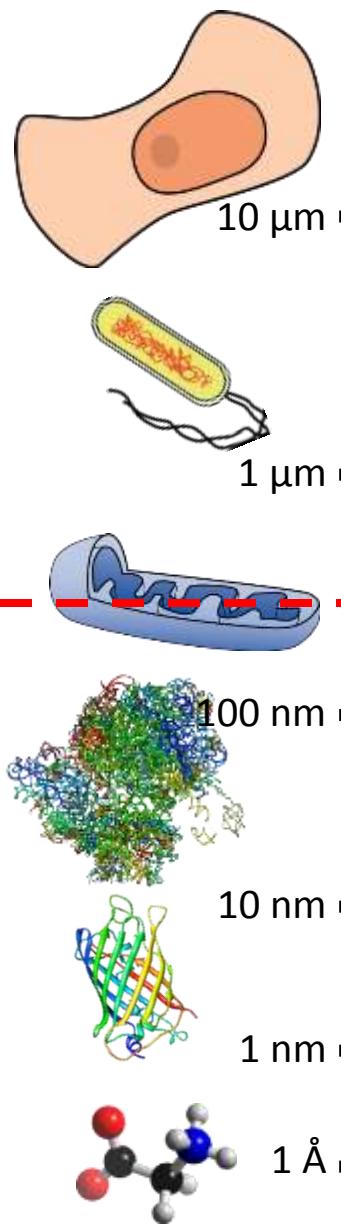


The diffraction barrier



Diffraction limit: ~ 250 nm lateral
~ 600 nm axial

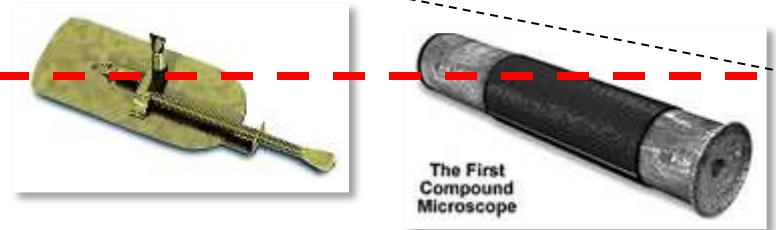




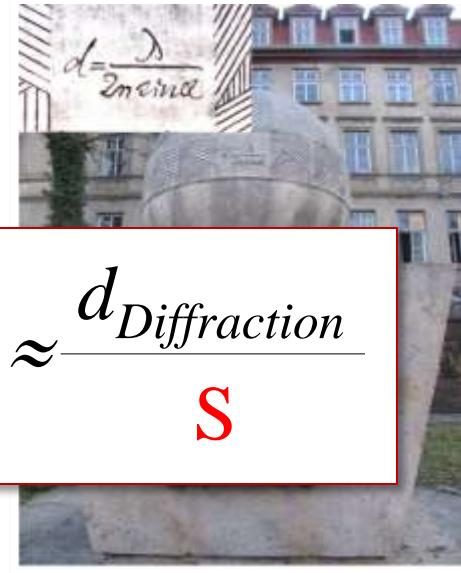
Naked eye: $\sim 50\text{-}100 \mu\text{m}$

★ 1595, Zaccharias and Hans Janssen
First microscope, 9x magnification

★ Antony Van Leeuwenhoek
(1632-1723), 200x



$$D \approx \frac{d_{\text{Diffraction}}}{S}$$



Ernst Abbe (1840-1905)
The “physical” diffraction limit

Super-resolution
Deconvolution
Optical Microscopy

4-PI microscopy

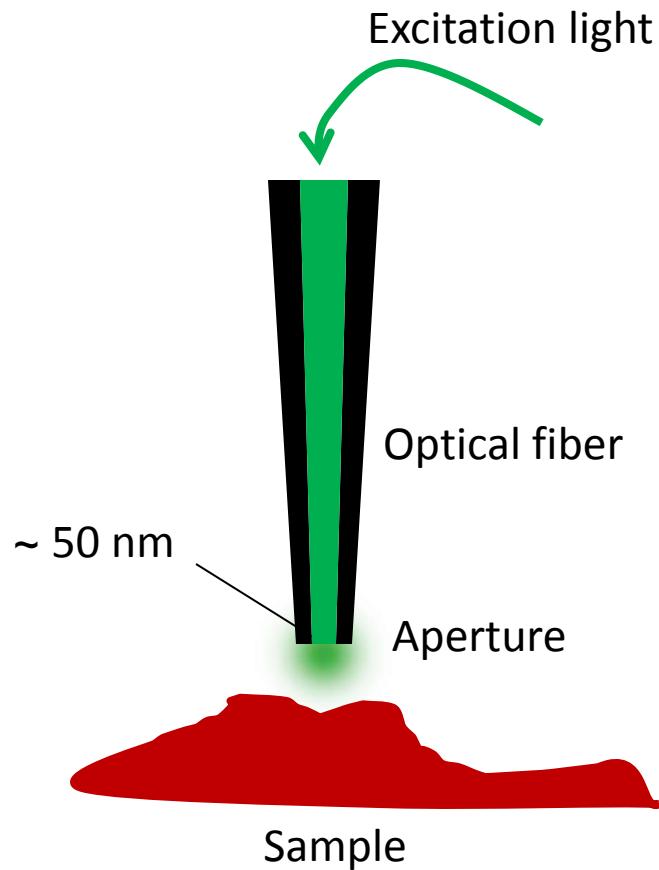
...

1600 1700 1800 1900 2000

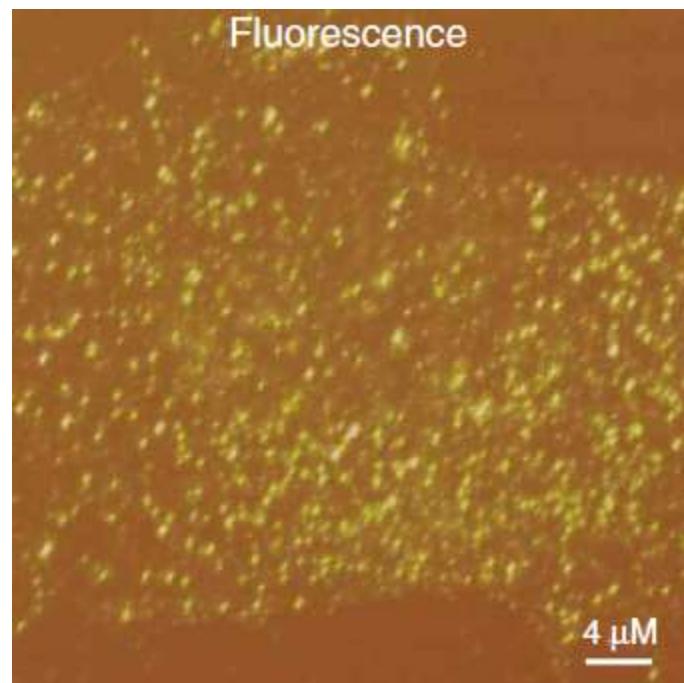
50 years to extend the resolution

- Confocal microscopy (1957)
- Near-field scanning optical microscopy (1972/1984)
- Multiphoton microscopy (1990)
- 4-Pi microscopy / I^5M (1991-1995)
- Structured illumination microscopy (2000)
- Negative refractive index (2006)

Near-field scanning optical microscopy

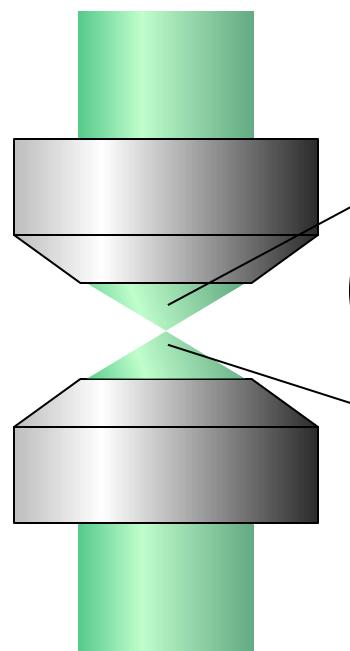


β_2 adrenergic receptor clusters
on the plasma membrane



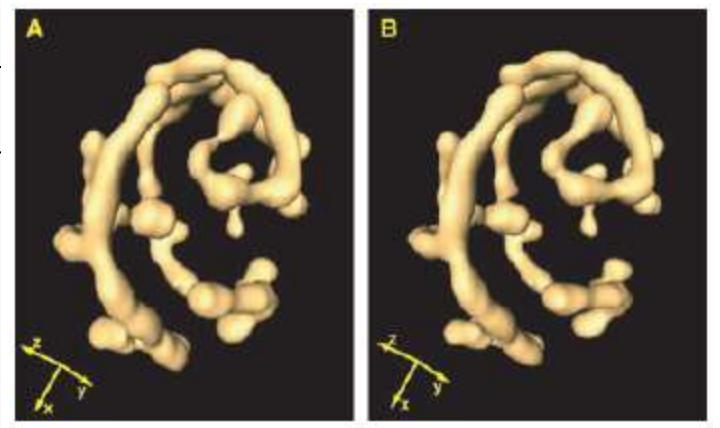
Ianoul et al., 2005

4-Pi / I^5M



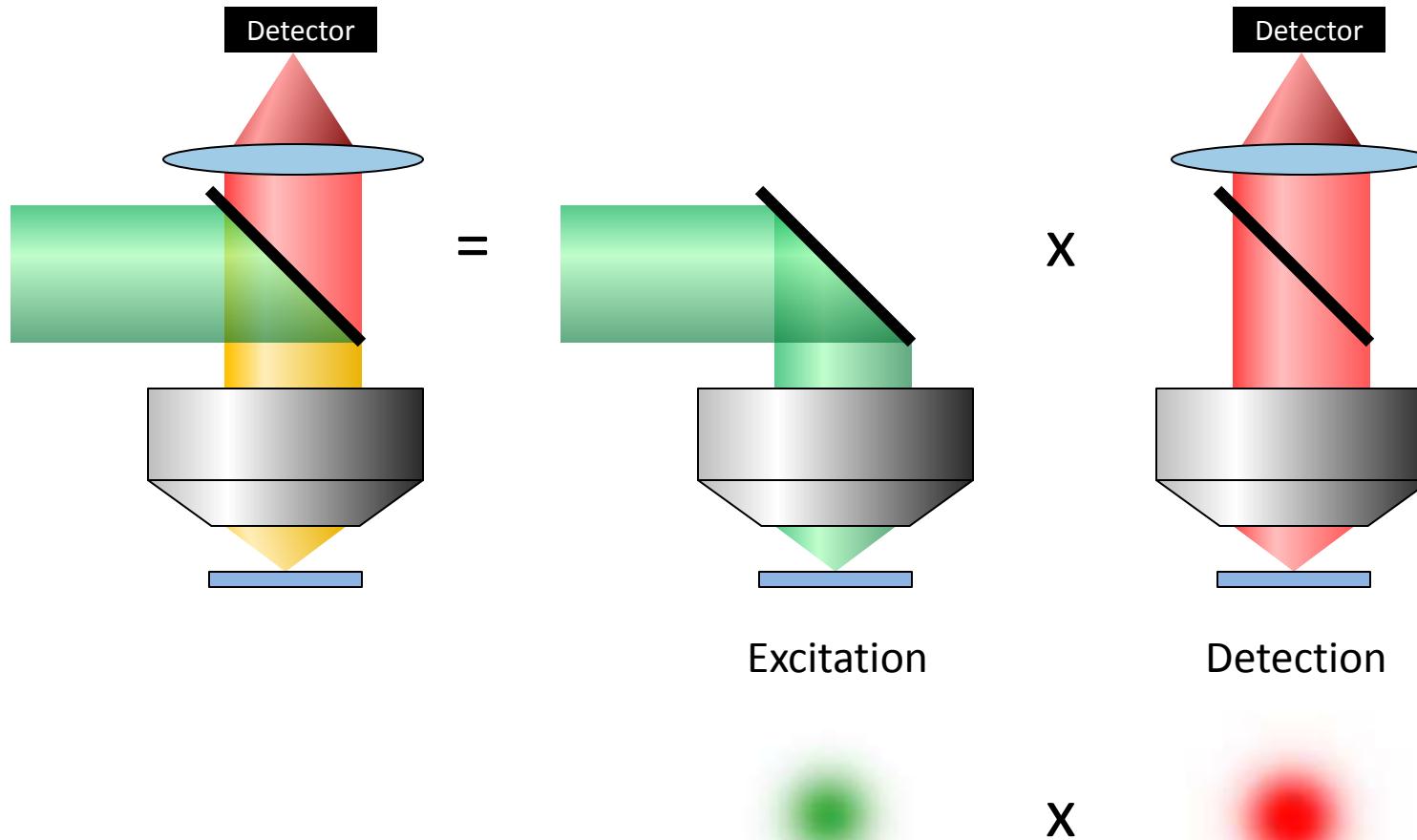
$$NA = n \sin \alpha$$

$$d \approx \frac{\lambda}{2 NA}$$

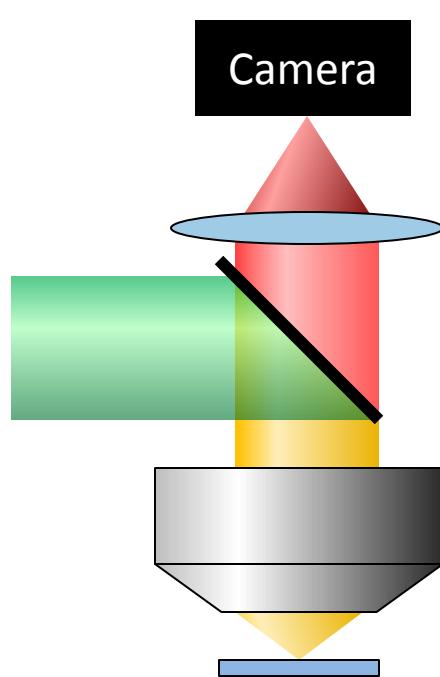


Major advantage:
Similar z resolution as x-y resolution

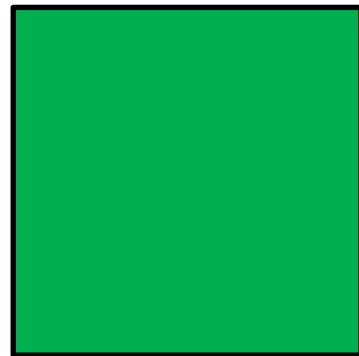
Patterned illumination



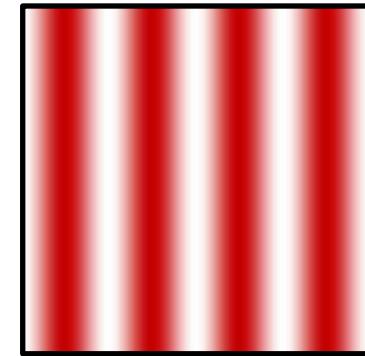
Structured Illumination Microscopy (SIM)



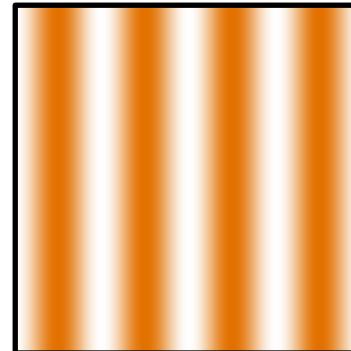
Wide field illumination



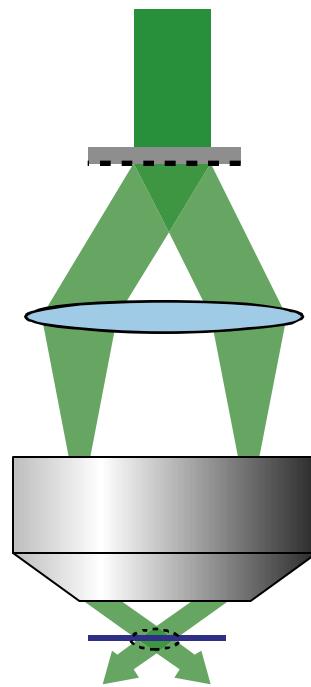
Diffraction-limited detection



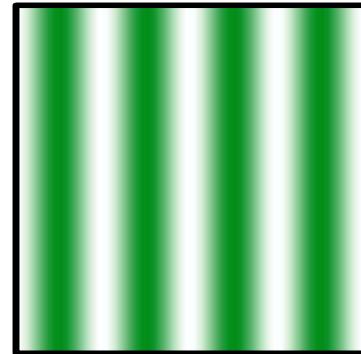
Diffraction-limited image



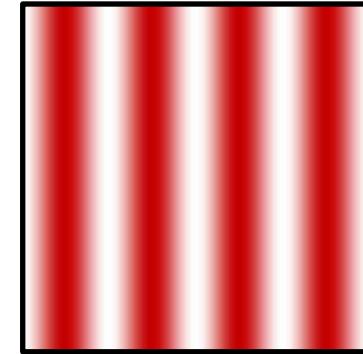
Structured Illumination Microscopy (SIM)



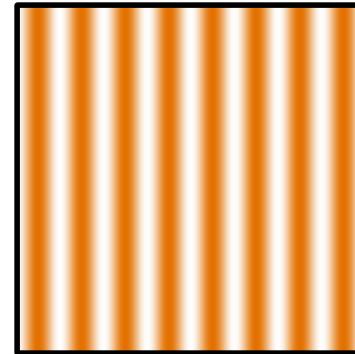
Structured field illumination



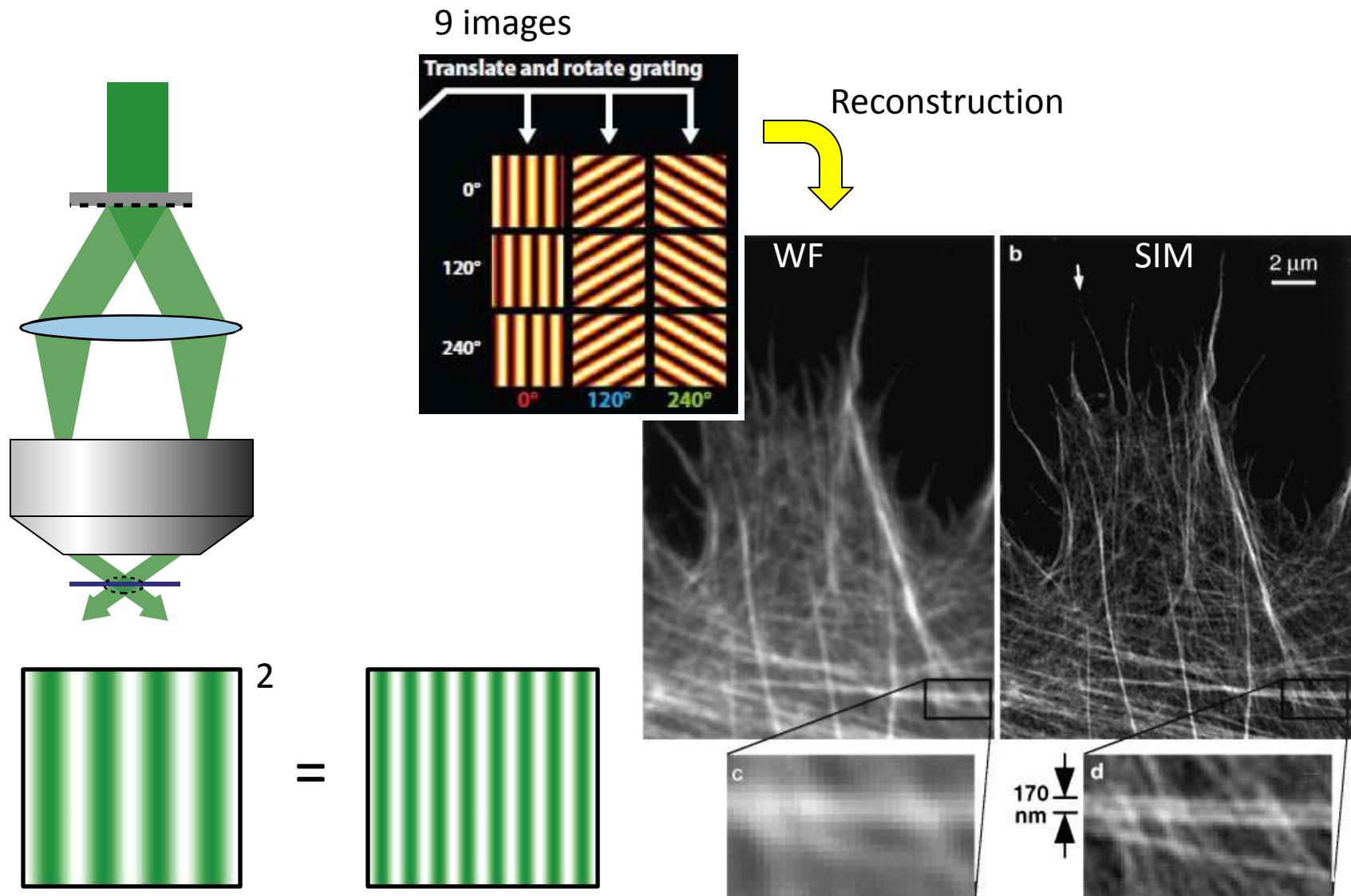
Diffraction-limited detection



Diffraction-limited image



Structured Illumination Microscopy (SIM)

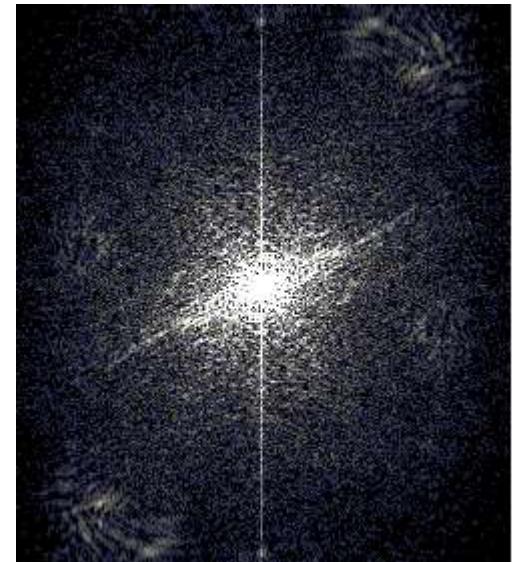


Being (slightly) more rigorous about SIM

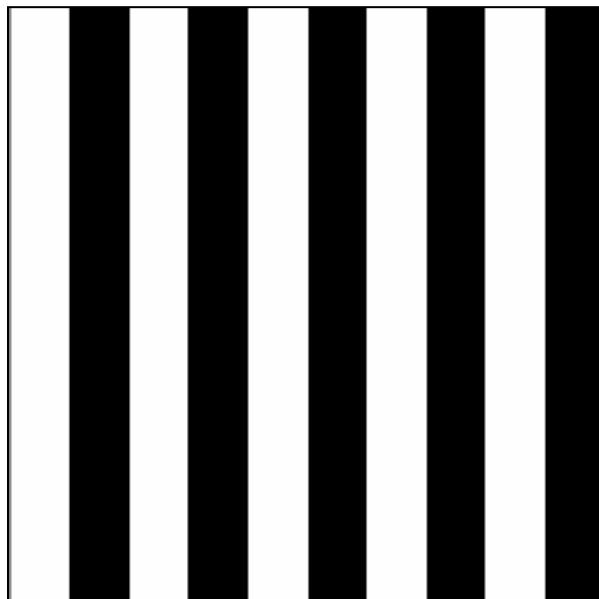


Fourier transform: Examples

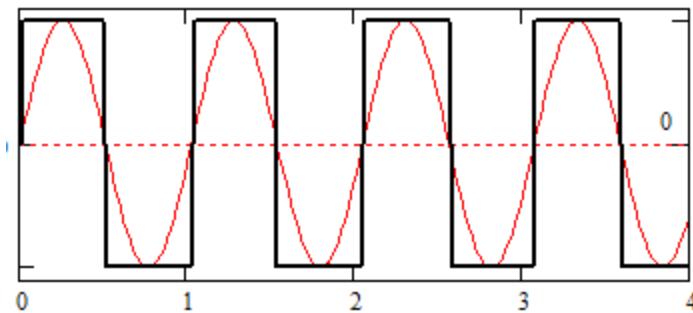
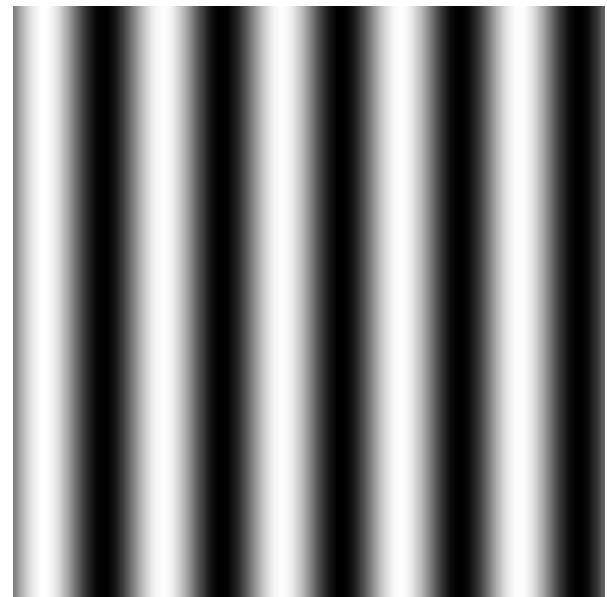
- $f(x) = \delta(x) \quad \Rightarrow \quad \tilde{f}(k) = 1$
- $f(x) = 1 \quad \Rightarrow \quad \tilde{f}(k) = 2\pi \delta(k)$
- $f(x) = e^{-ax|x|} \quad \Rightarrow \quad \tilde{f}(k) = \frac{2a}{k^2 + a^2}$
- $f(x) = \frac{1}{x^2 + a^2} \quad \Rightarrow \quad \tilde{f}(k) = \frac{\pi}{a} e^{-ak/k}$
- $f(x) = e^{-a\frac{|x|}{2}} \quad \Rightarrow \quad \tilde{f}(k) = \sqrt{\frac{2\pi}{a}} e^{-\frac{1}{a} \cdot \frac{k^2}{2}}$
- $$f(x) = \begin{cases} 1 & -a \leq x < 0 \\ 0 & 0 \leq x \leq a \end{cases} \quad \Rightarrow \quad \tilde{f}(k) = 2 \frac{\sin(ak)}{k}$$
A graph showing a rectangular pulse function $f(x)$ plotted against x . The function is 1 for x between - a and a , and 0 elsewhere. The origin is marked as 0. The x-axis is labeled with - a , 0, and a .



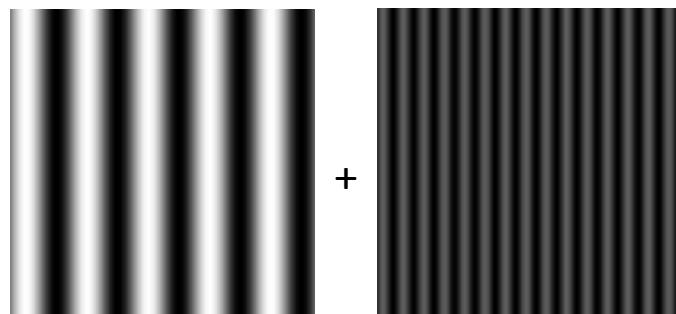
Fourier transform and spatial frequencies



?
=

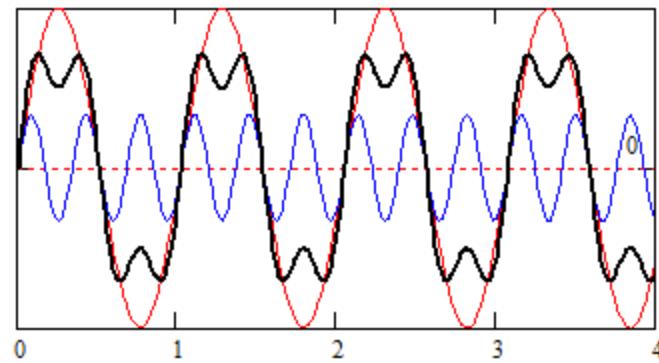
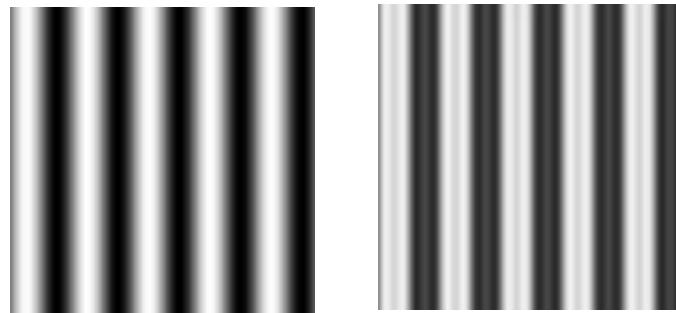


Fourier transform and spatial frequencies



+

Summed image

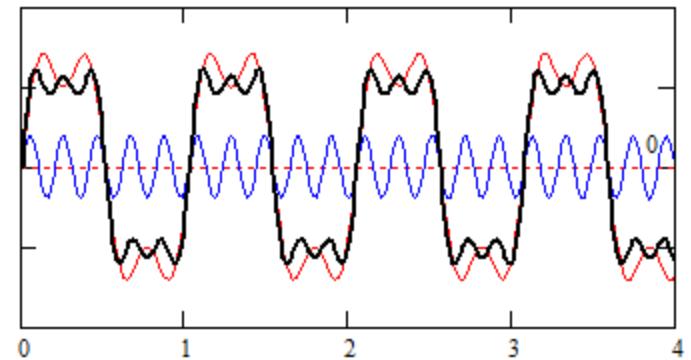
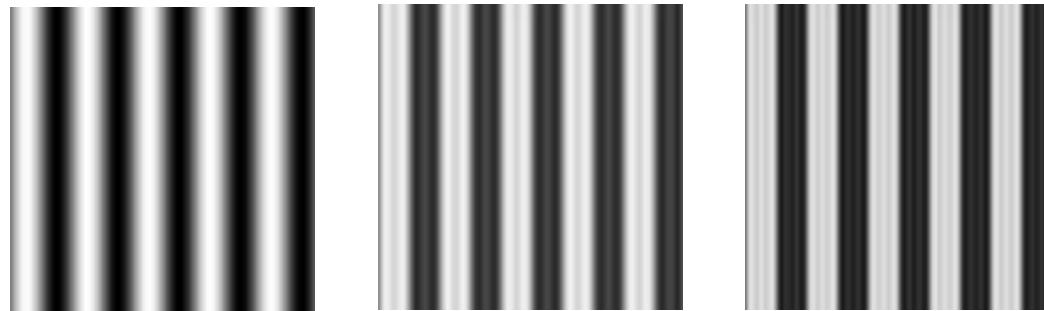


Fourier transform and spatial frequencies



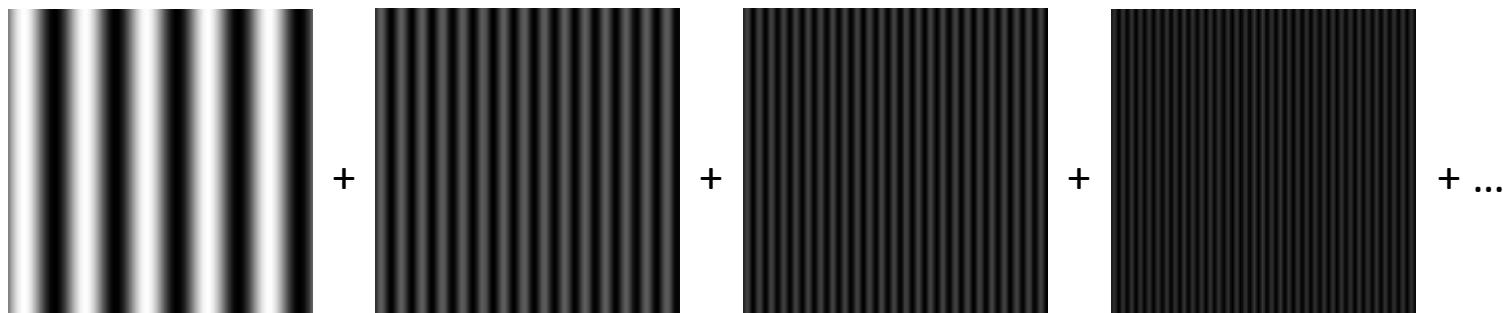
+ +

Summed image



Fourier transform and spatial frequencies

Discrete spatial frequencies



Summed image

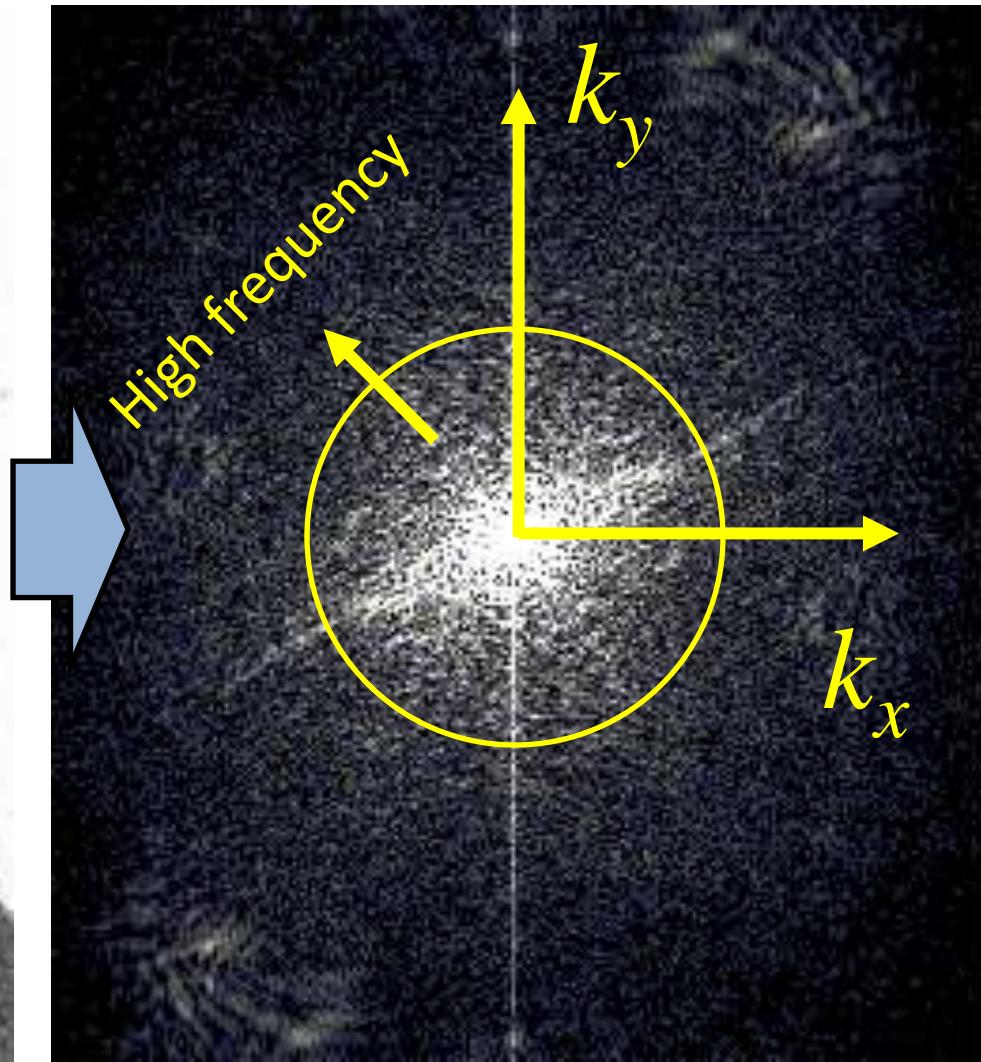


$$G(x) = \sum F(k) \sin(k x)$$

Fourier transform and spatial frequencies

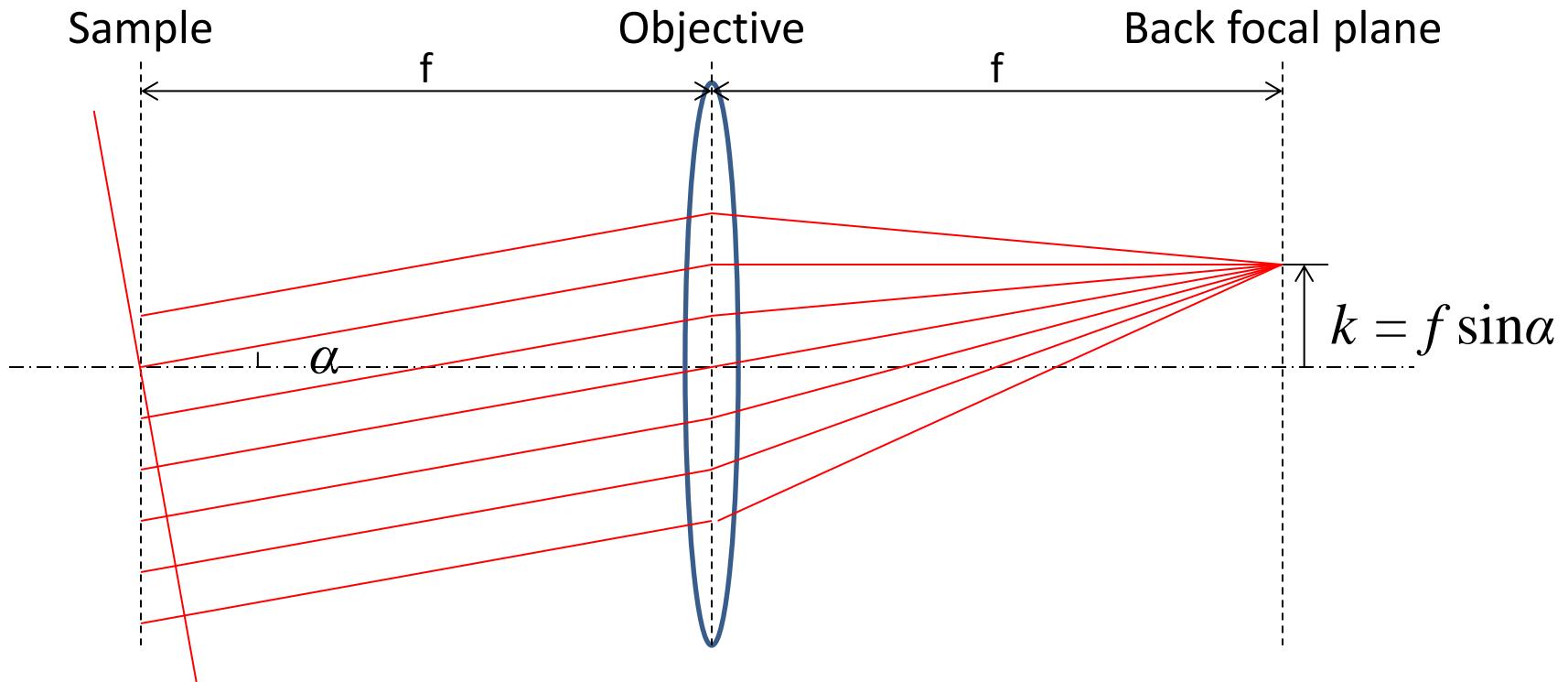


Original Image (real space)

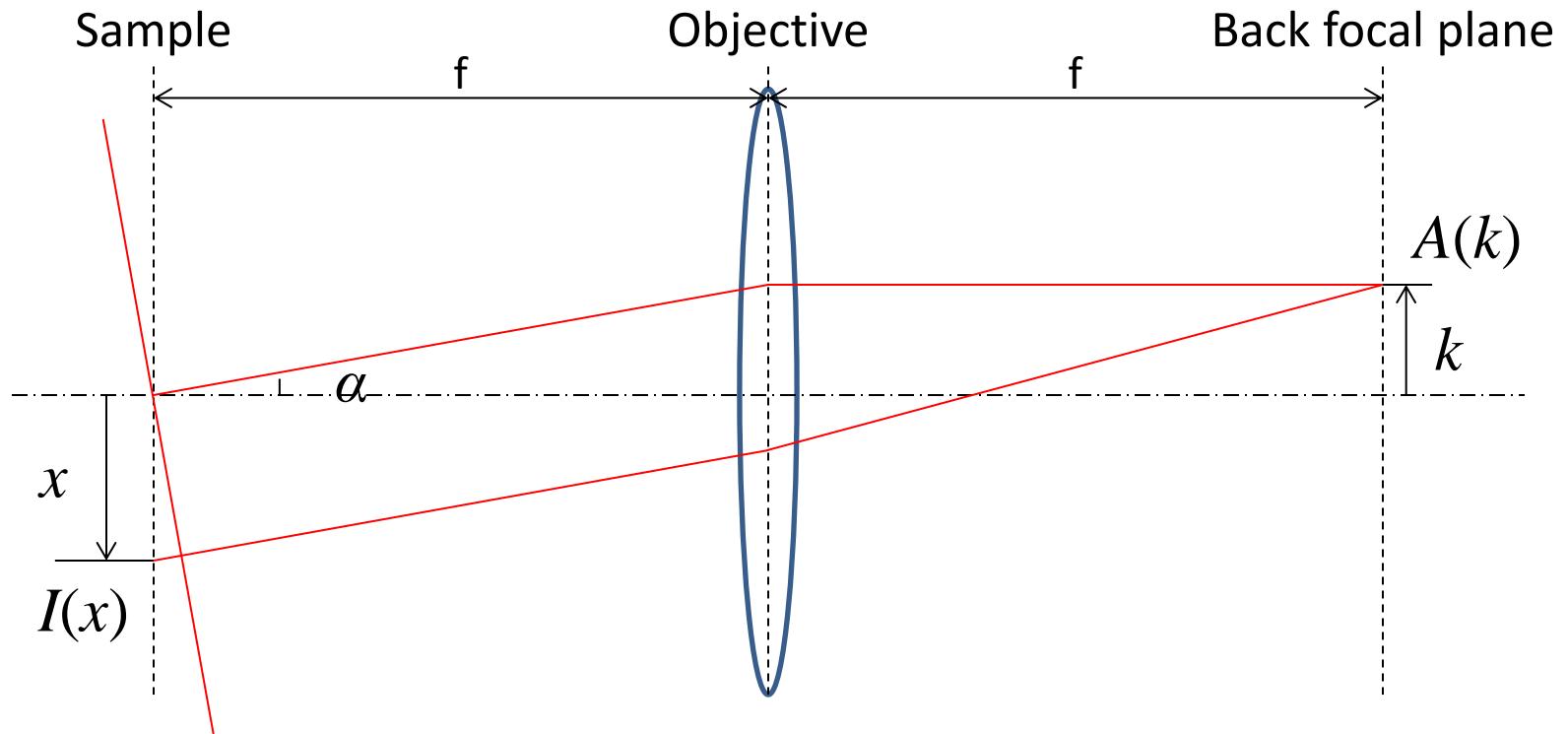


Fourier transform (frequency space)

Fourier optics and microscope resolution



Fourier optics and microscope resolution



Phase delay from the mid-point

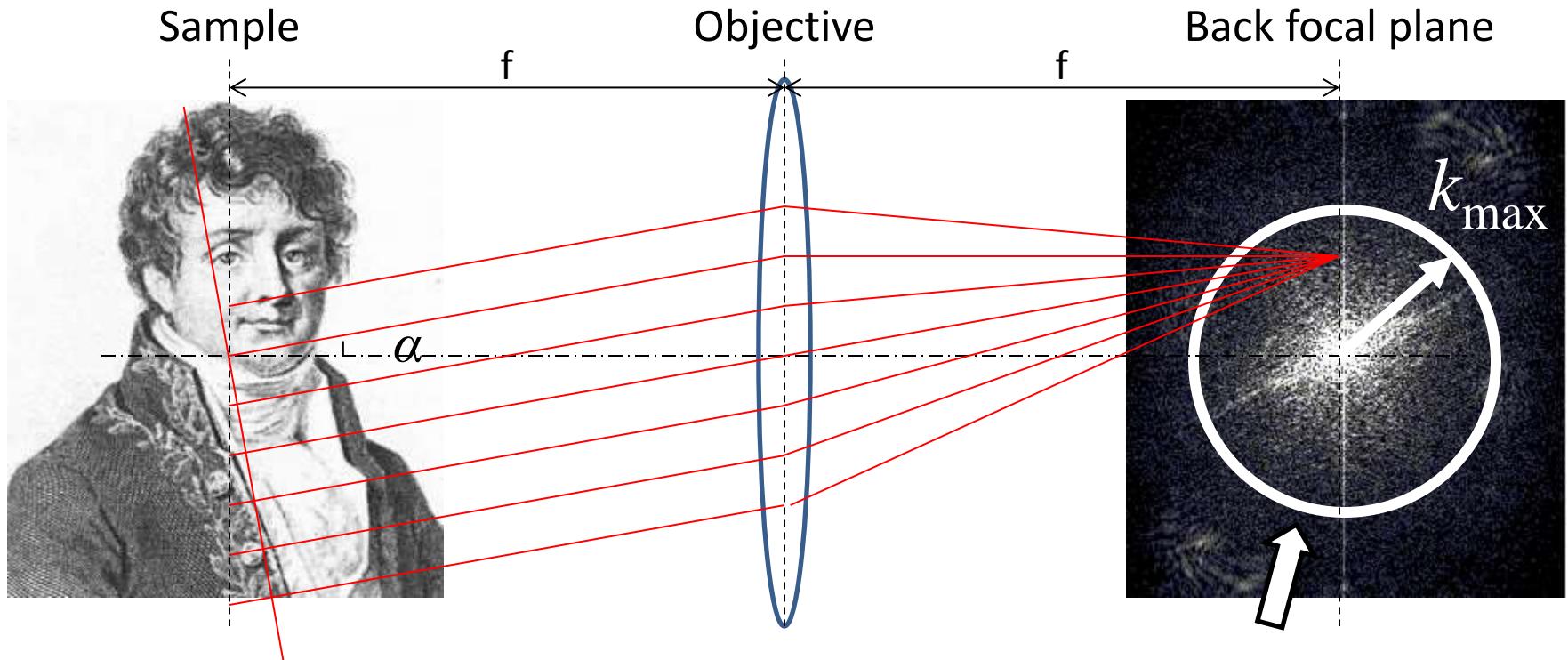
$$\Delta\varphi = x \sin\alpha (2\pi/\lambda) = x (k/f) (2\pi/\lambda) \text{ assuming refractive index} = 1$$

Light intensity at the sample plane

Fourier Transform!

$$I(x) = \sum A(k) \sin(\Delta\varphi) = \boxed{\sum A(k) \sin(x k 2\pi/\lambda f)}$$

Fourier optics and microscope resolution



$$\text{Spatial frequency} = k \cdot 2\pi/\lambda f$$

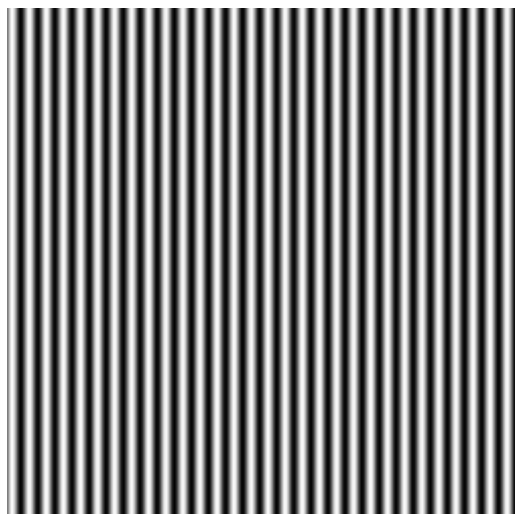
$$k_{\max} = f \sin \alpha_{\max} = f \cdot NA$$

$$\text{Resolution} = \lambda / 2NA$$

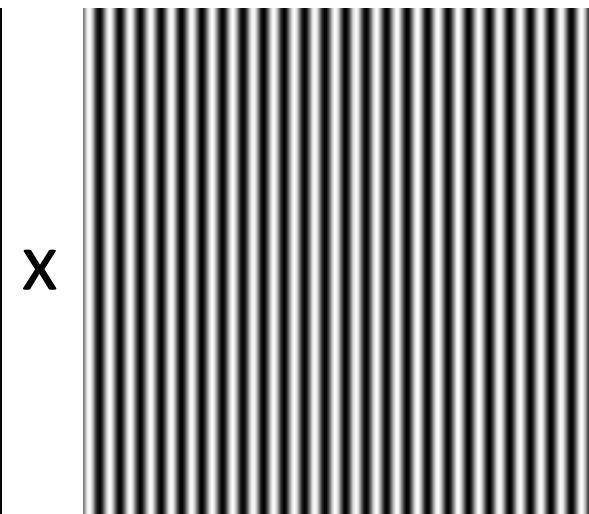
Size of the back focal plane

Extending the measurable freq. range

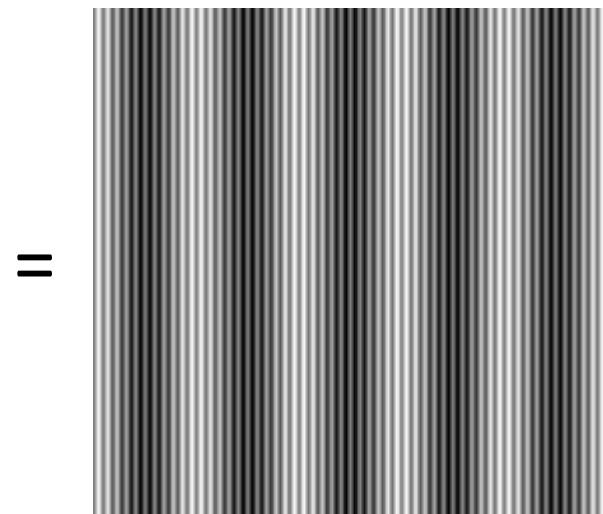
Excitation(x) \times Sample(x) = Observed Signal(x)



Freq = 30



Freq = 25



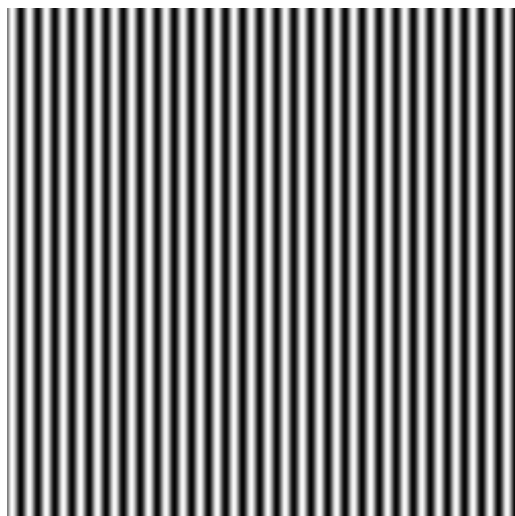
=

Freq = 55 & 5

$$\sin A \cdot \sin B = (\cos(A - B) - \cos(A + B)) / 2$$

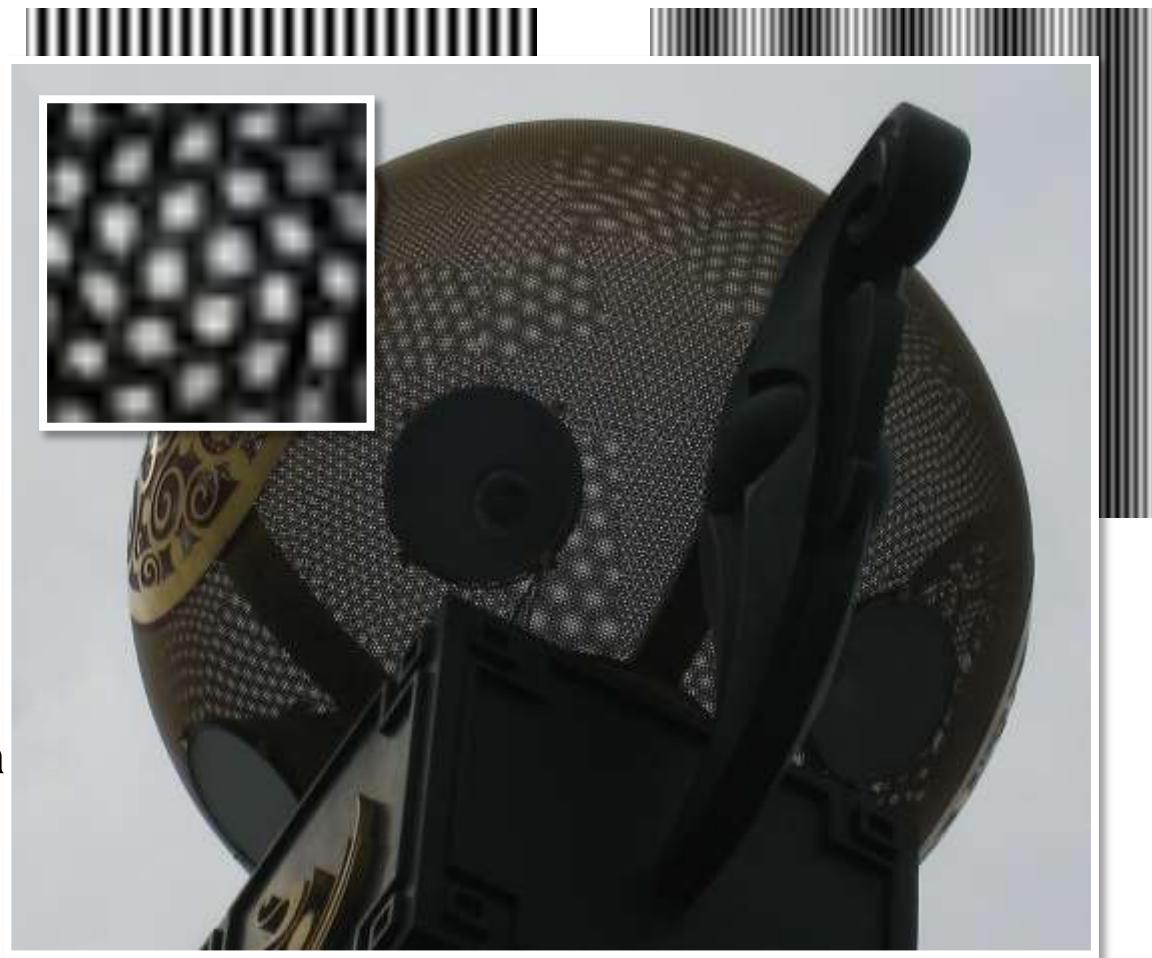
Extending the measurable freq. range

$$\text{Excitation}(x) \times \text{Sample}(x) = \text{Observed Signal}(x)$$



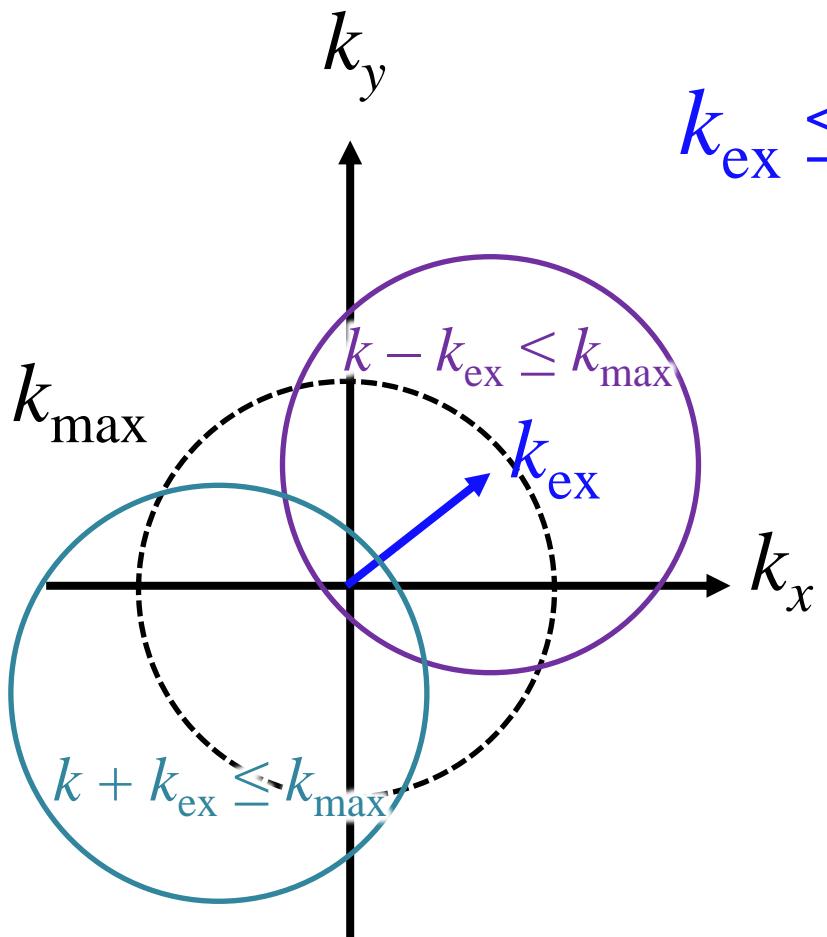
Freq = 30

x

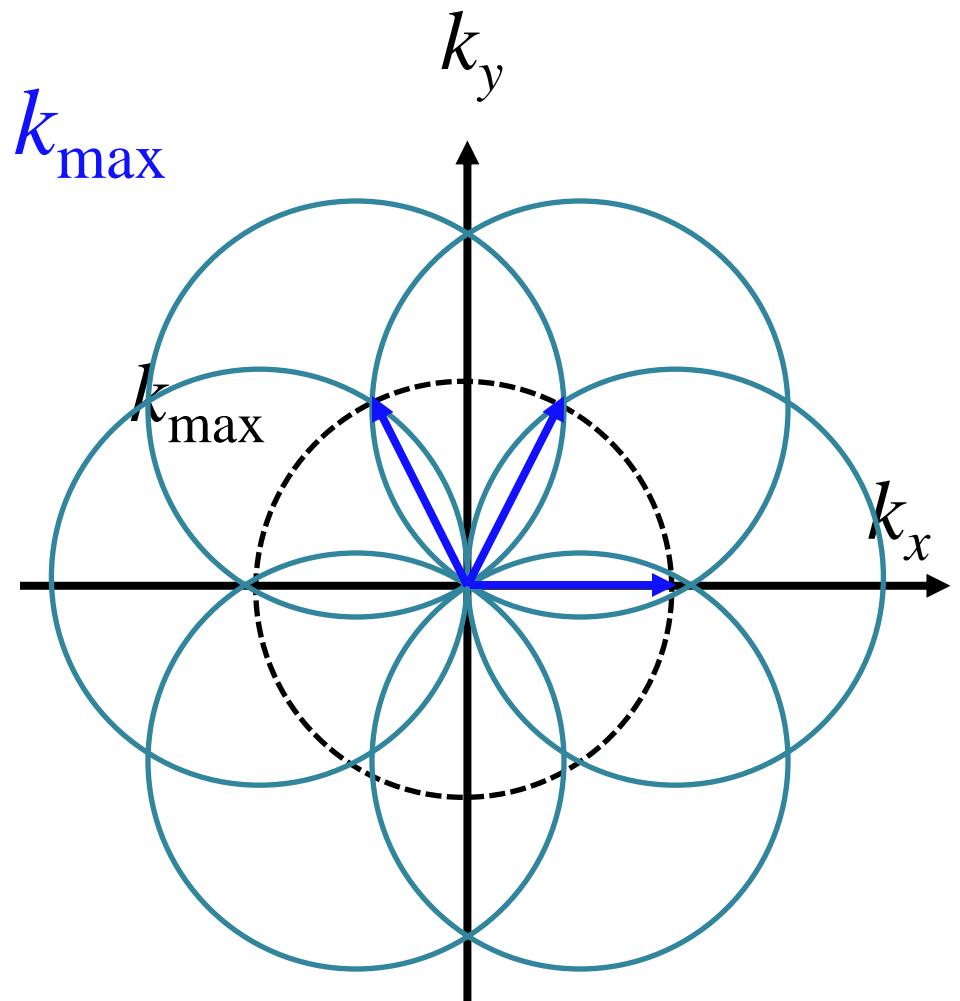


$\sin A \cdot \sin$

Extending the measurable freq. range

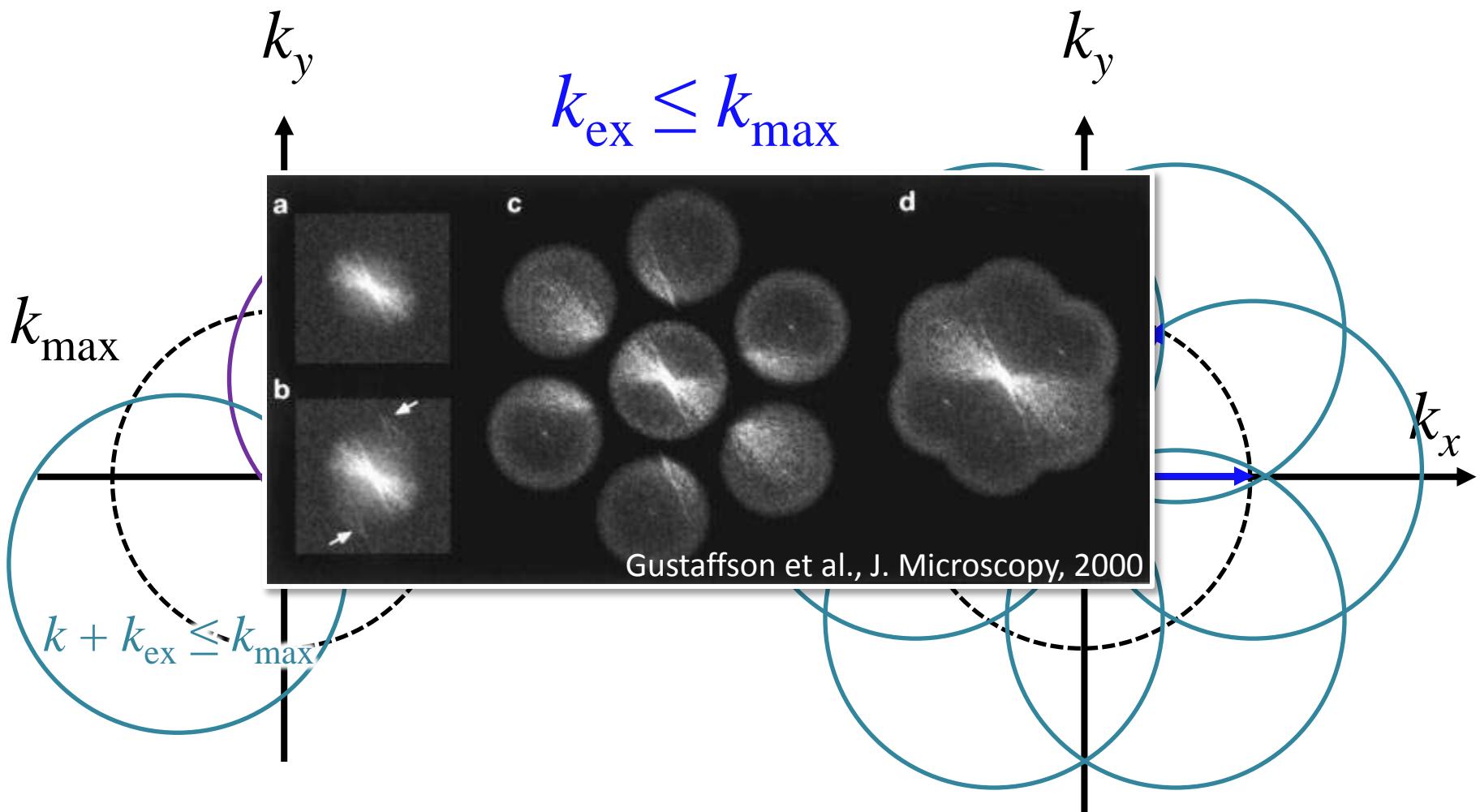


$$k_{\text{ex}} \leq k_{\max}$$

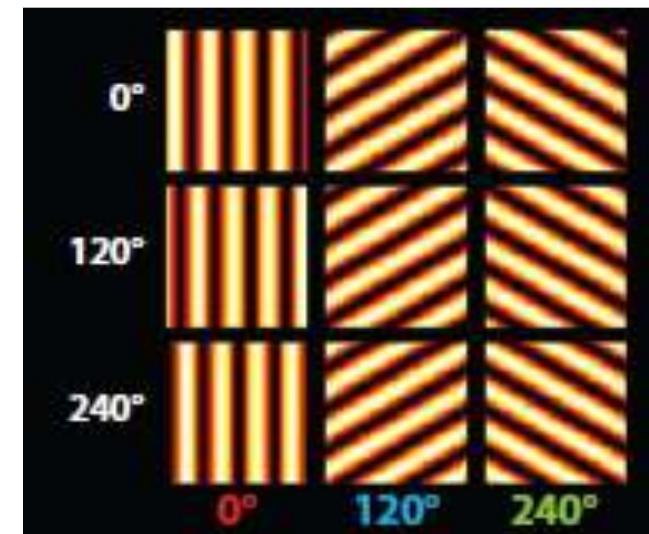
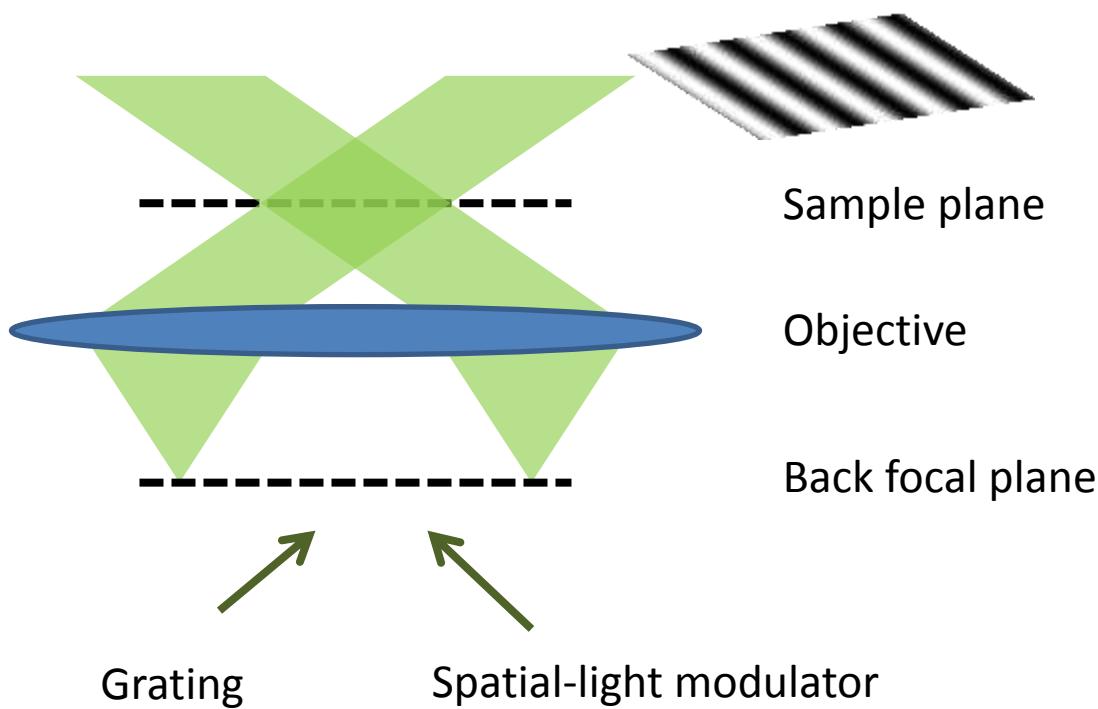


$$k_{\max}$$

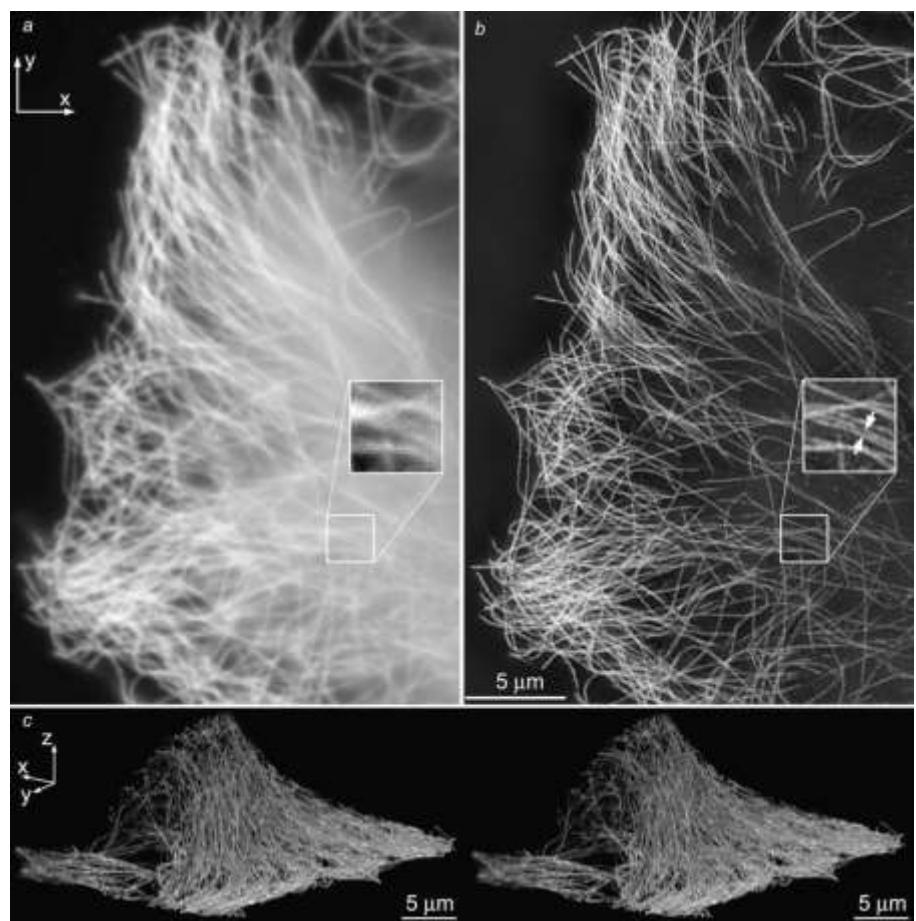
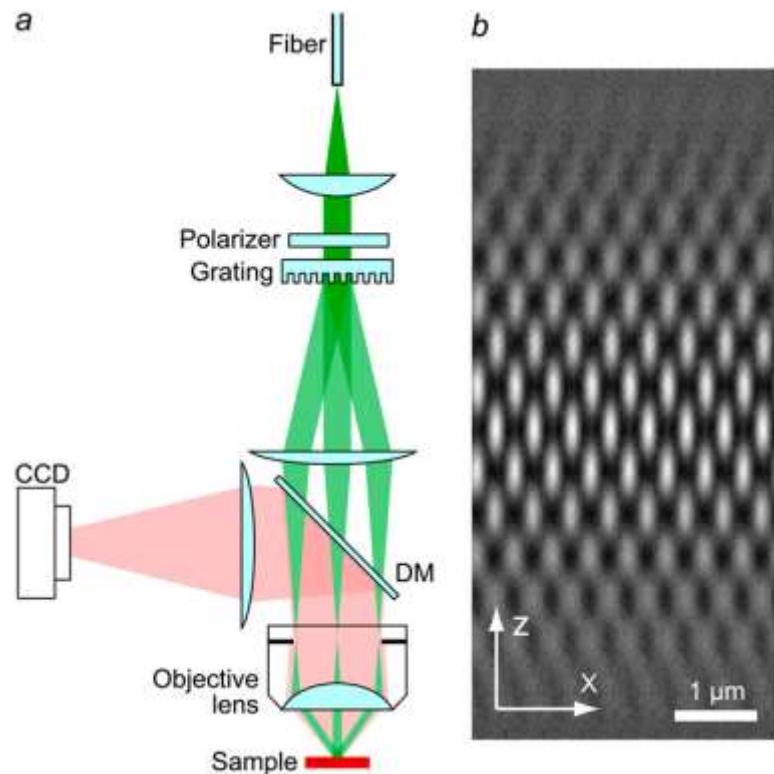
Extending the measurable freq. range



Generating the illumination pattern

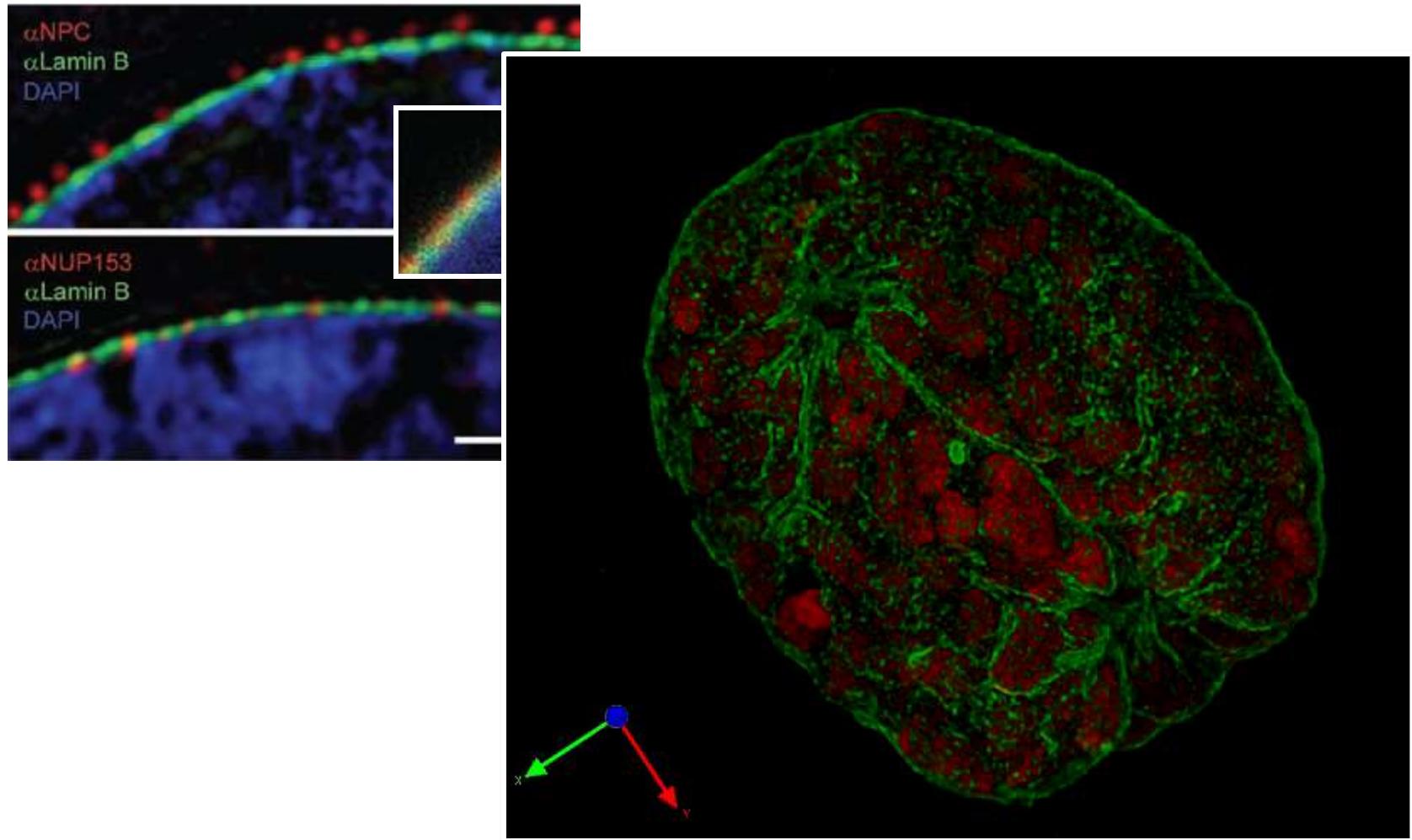


3D SIM: better resolution + optical sectioning

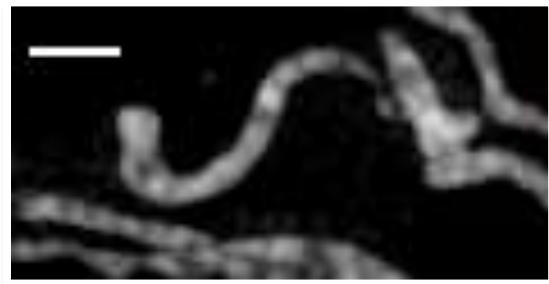
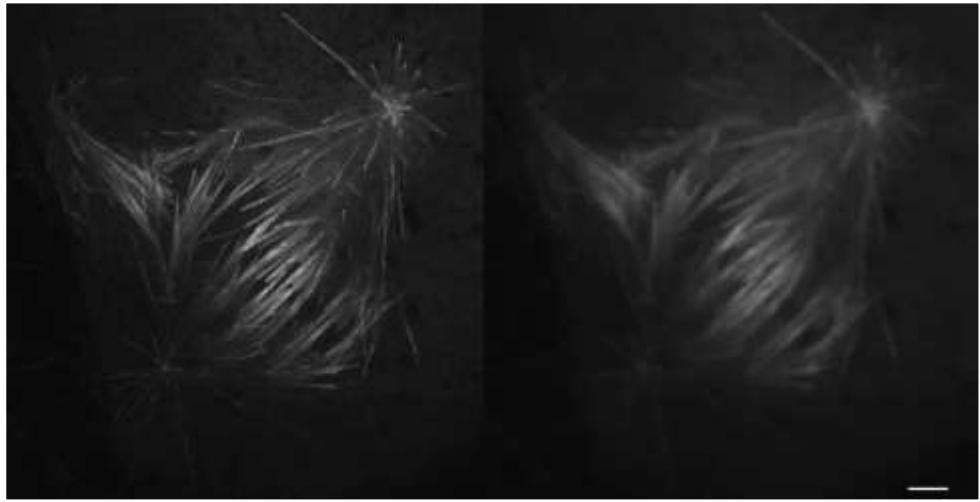


Multicolor SIM

Same as conventional fluorescence microscopy!



Live imaging with SIM

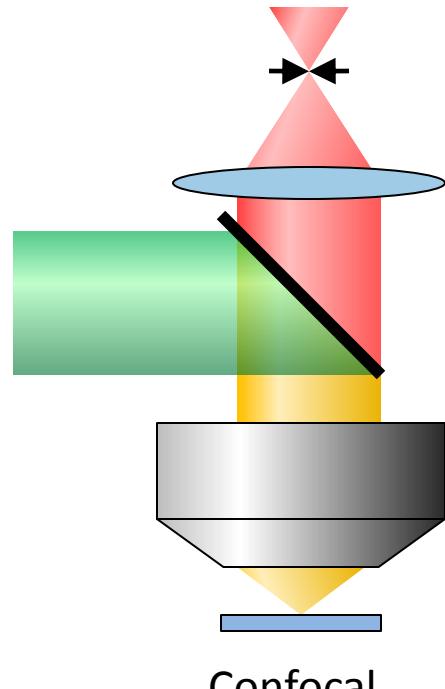


Kner, Chhun et al., Nat Methods,

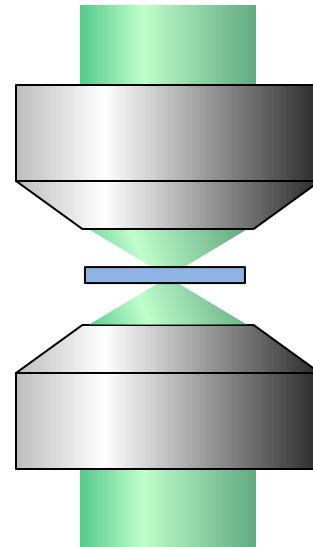


Shao et al., Nat Methods, 2011

The diffraction limit still exists

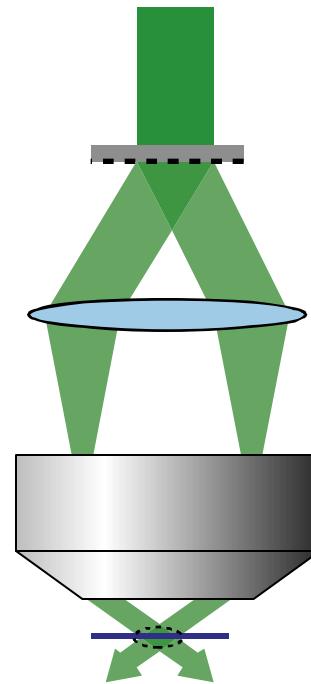


Confocal



$4\text{Pi} / \text{l}^5\text{M}$

$$d \geq \frac{1}{2} \cdot \frac{\lambda}{2NA}$$

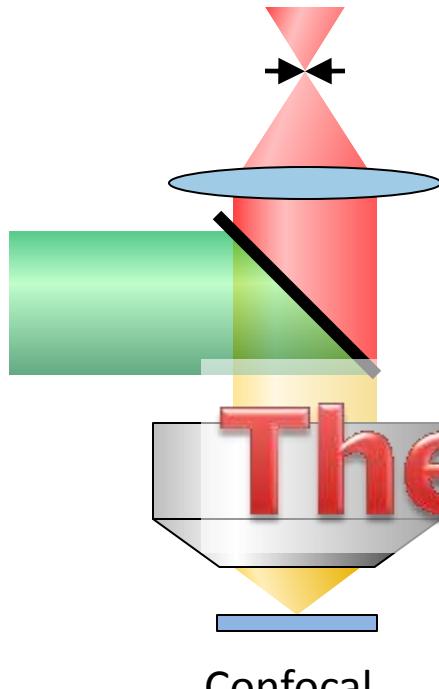


SIM

Breaking the diffraction barrier



Breaking the diffraction barrier

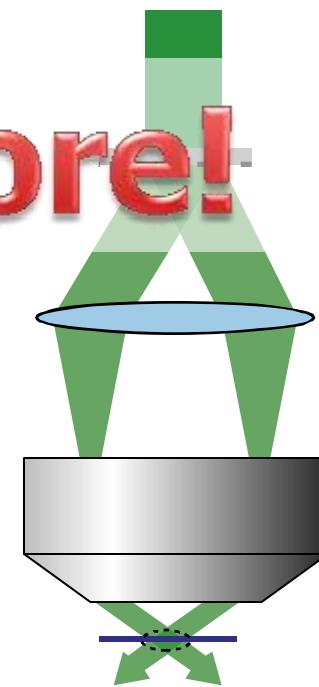


Confocal

The Fluorophore!

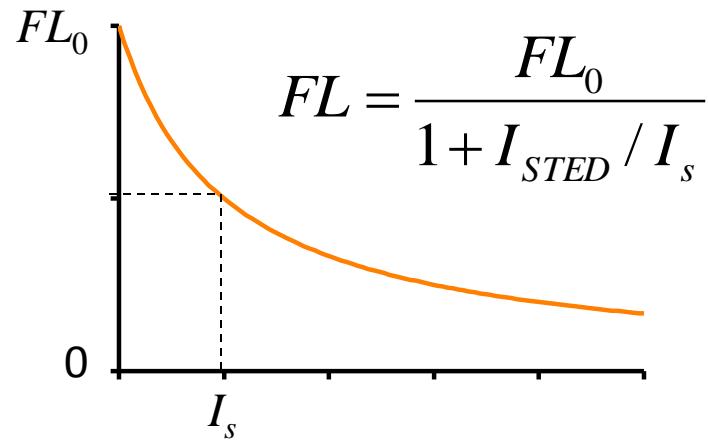
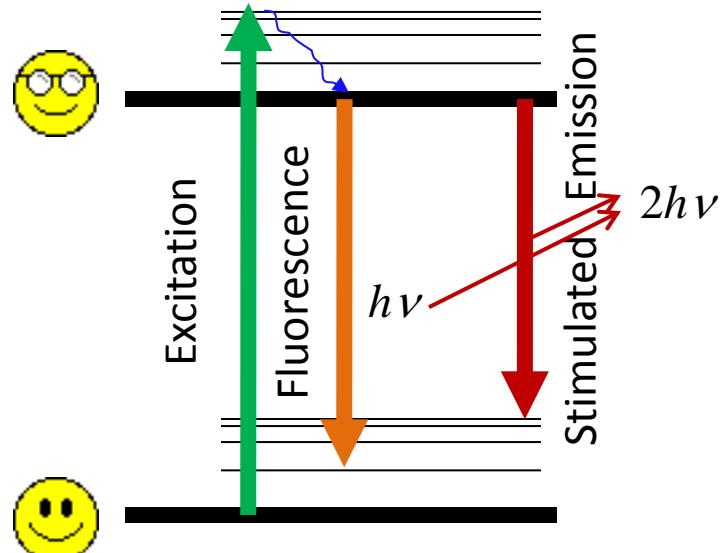
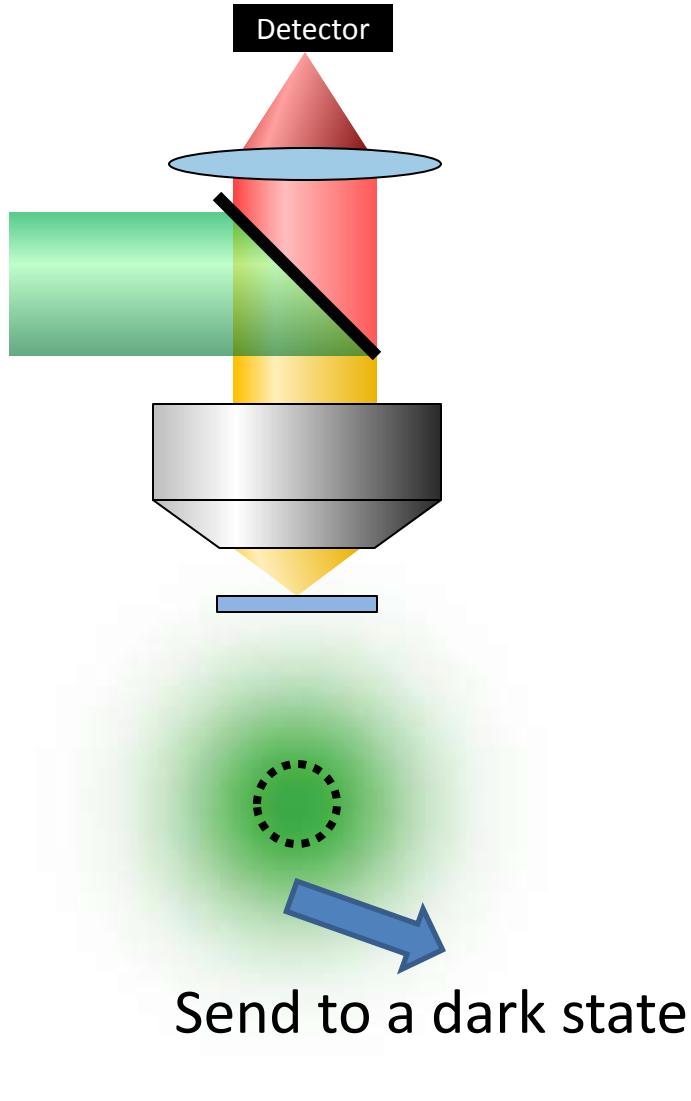


$4\text{Pi} / \text{I}^5\text{M}$

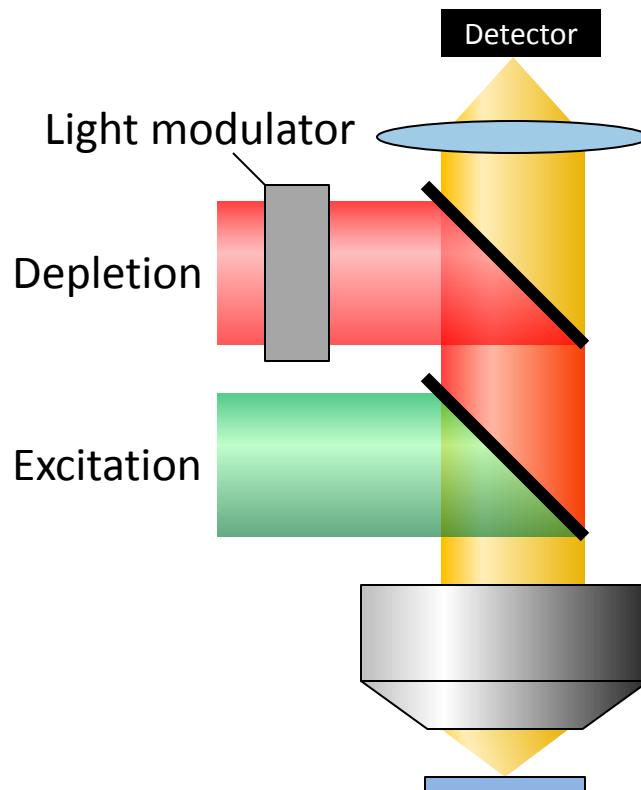
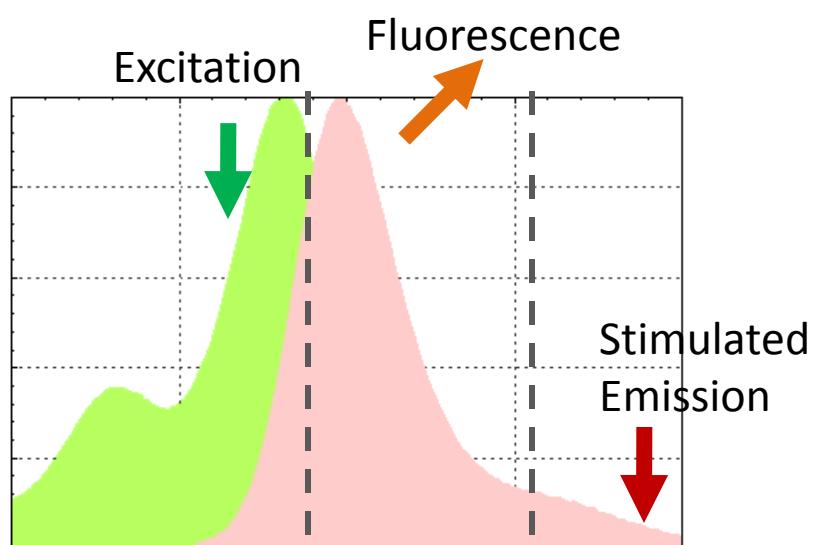


SIM

Stimulated Emission Depletion (STED)



STED microscopy



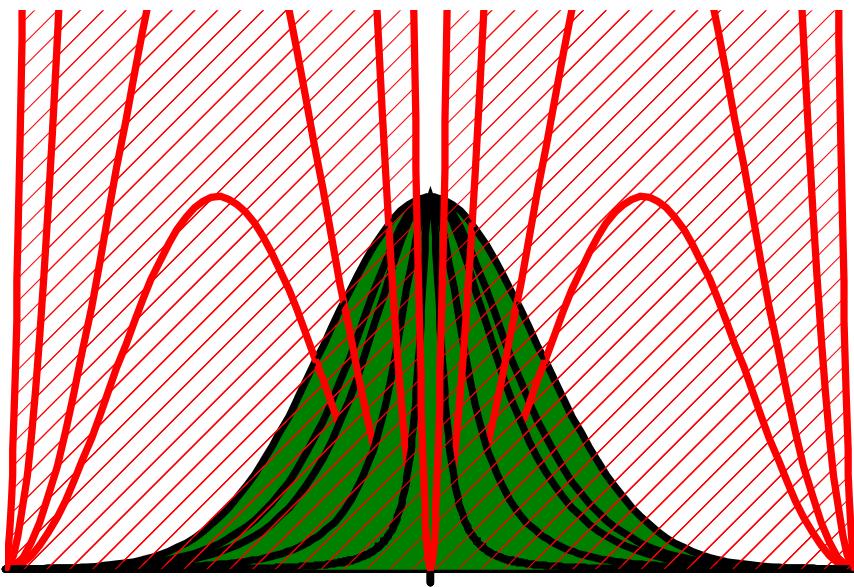
Excitation

STED
pattern

Effective
PSF

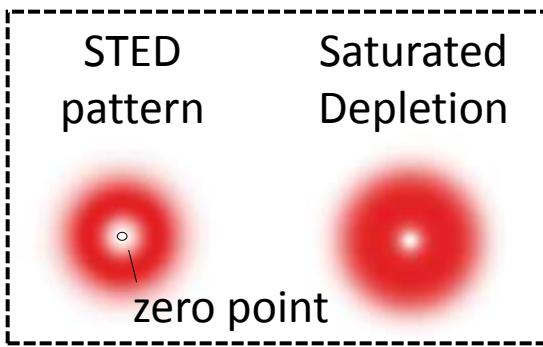
$$\text{Excitation} \quad \div \quad \text{STED pattern} = \text{Effective PSF} \quad ?$$

Saturated depletion

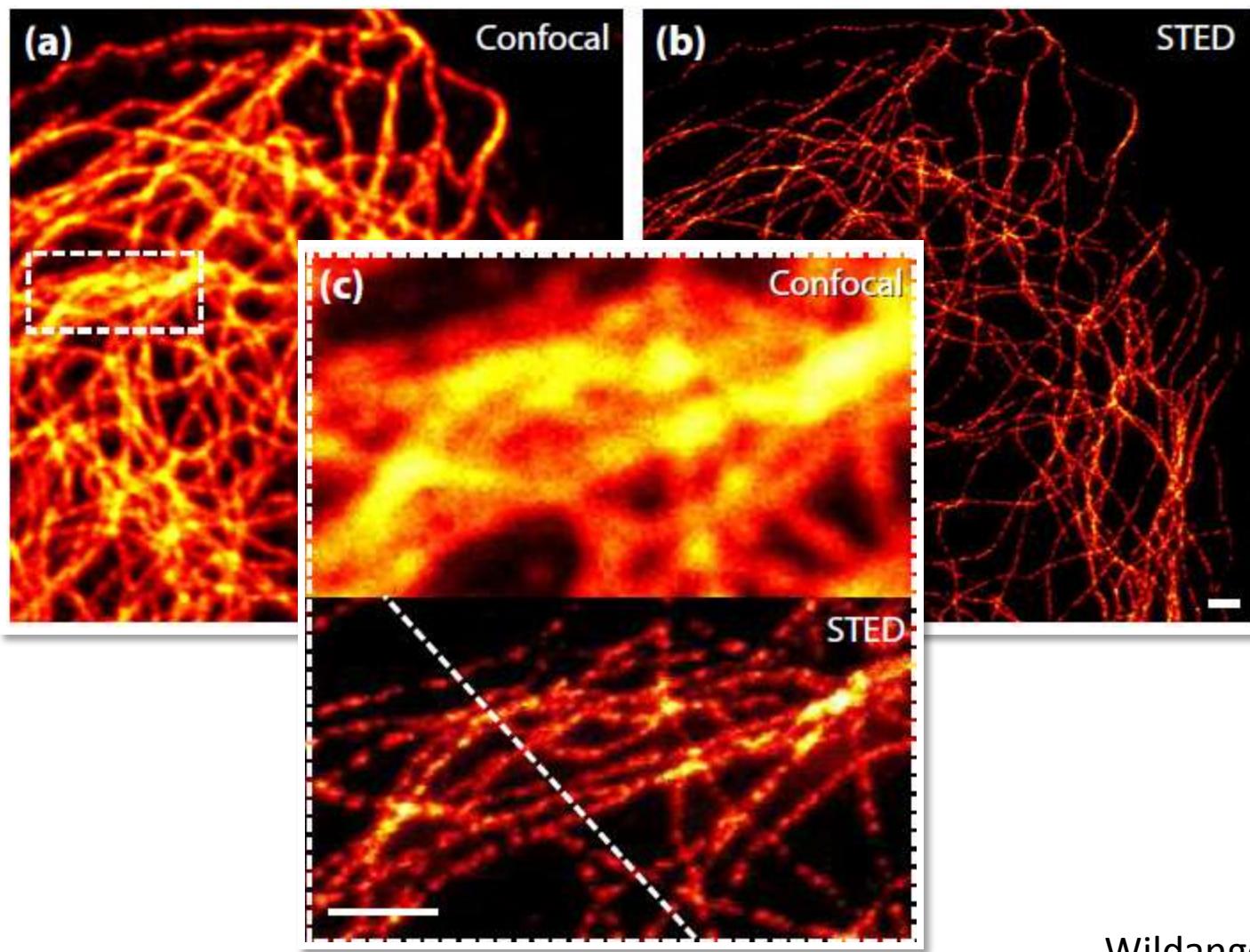


$$I_{\text{STED}} = I_{\text{S}} \Omega Q J_S$$

$$D = \frac{1}{\sqrt{1 + I/I_s}} \cdot \frac{\lambda}{2NA}$$

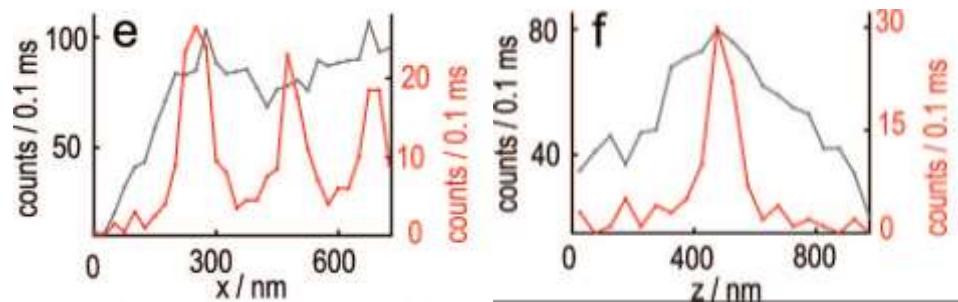
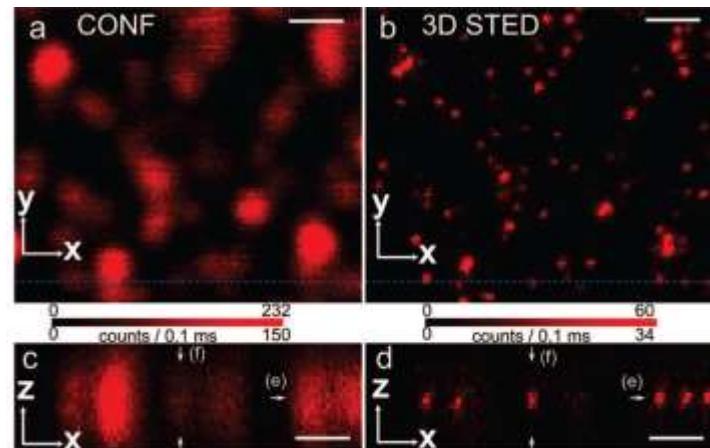
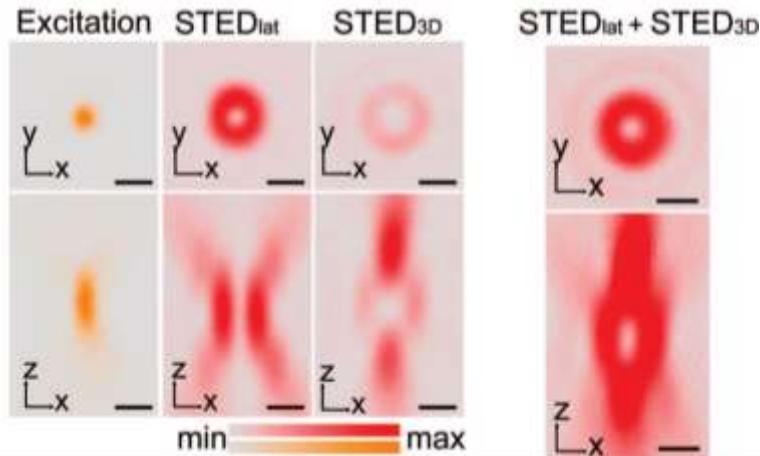


STED images of microtubules

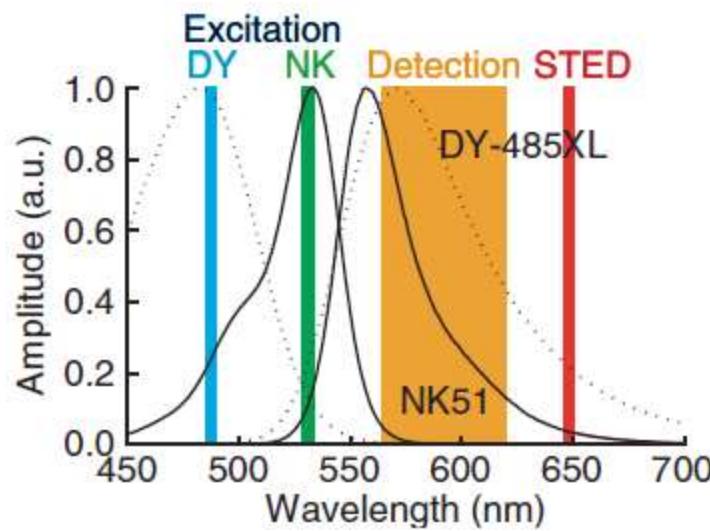
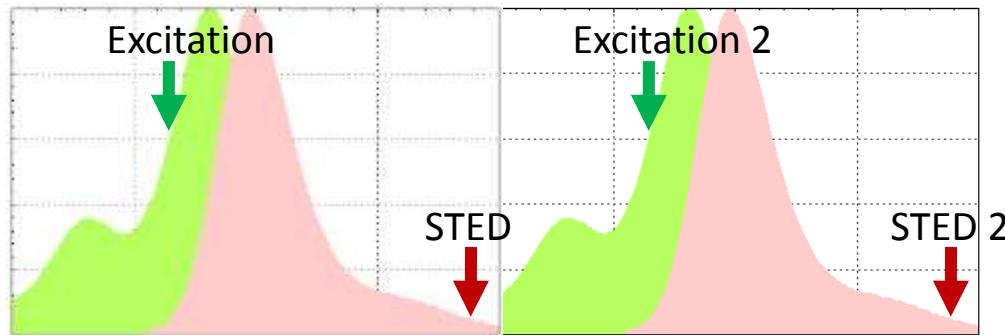


Wildanger et al., 2009

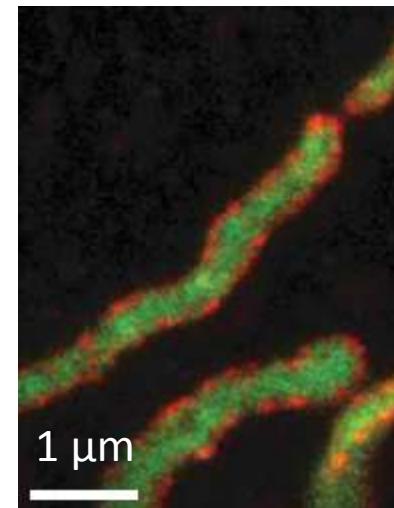
3D STED



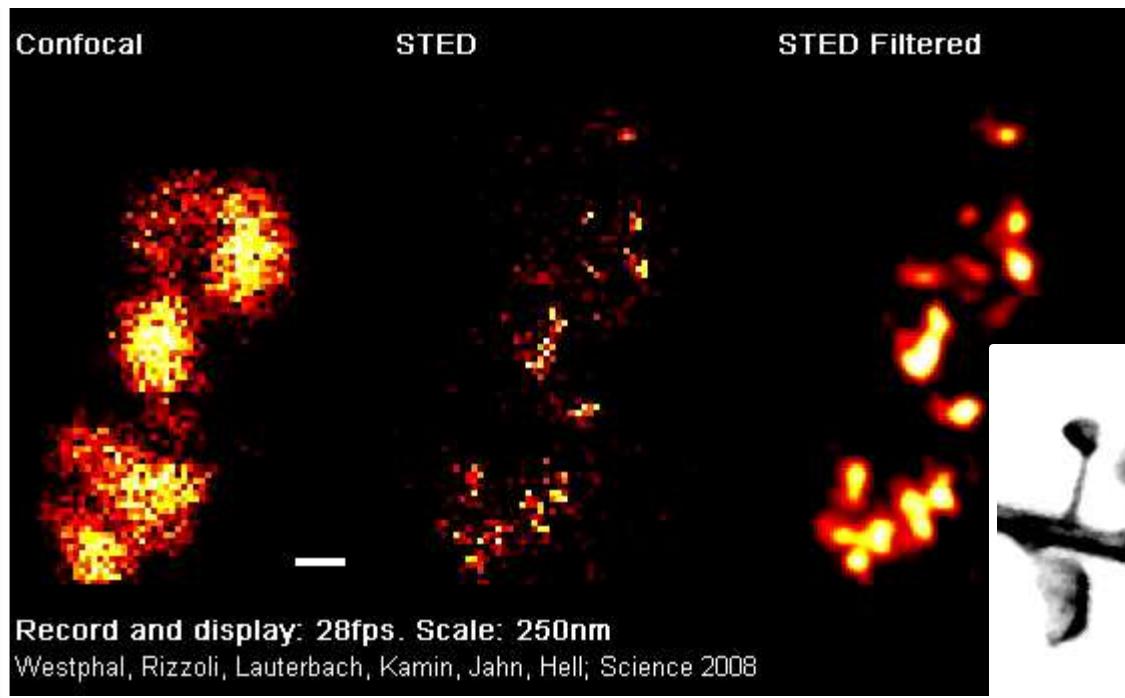
Muticolor STED



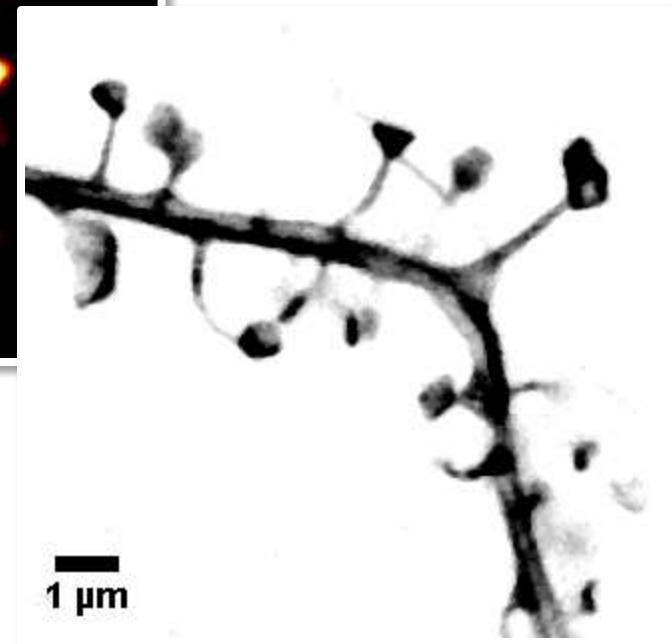
2 color isoSTED resolving
the inner and outer membrane
of mitochondria



Live STED



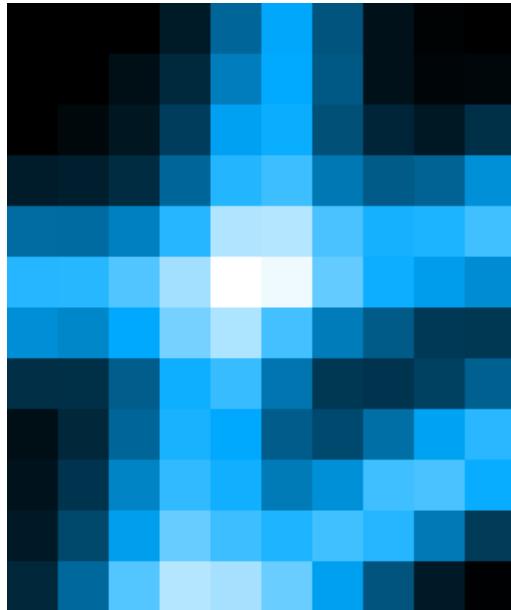
Westaphl et al., Science, 2008



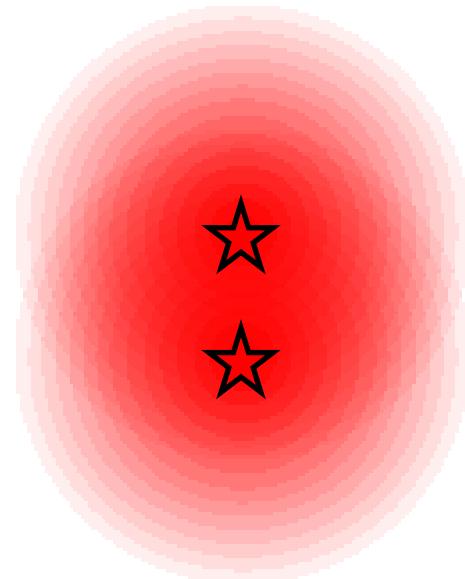
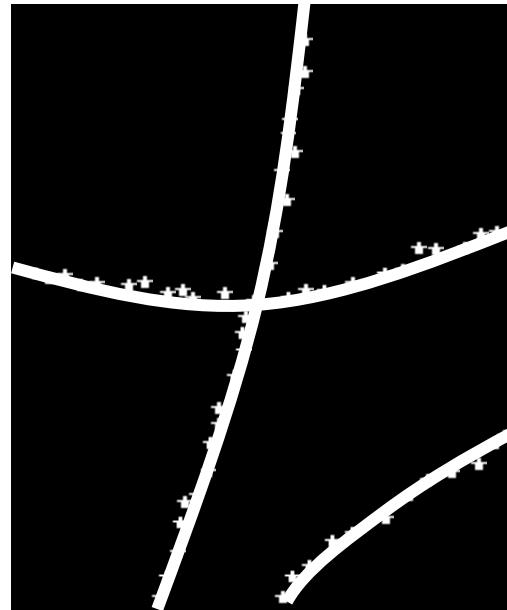
Nagerl et al., PNAS, 2008

Super-resolution by...

Fluorescence image

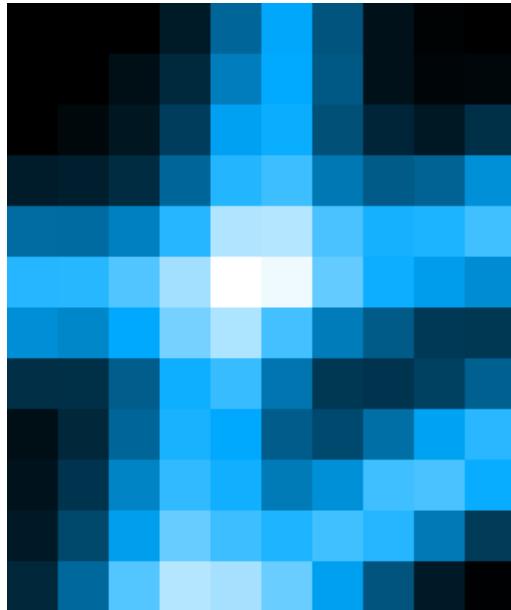


Underlying structure

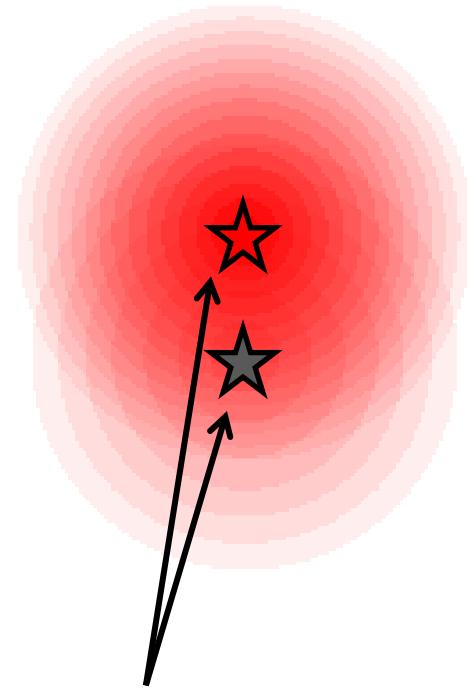
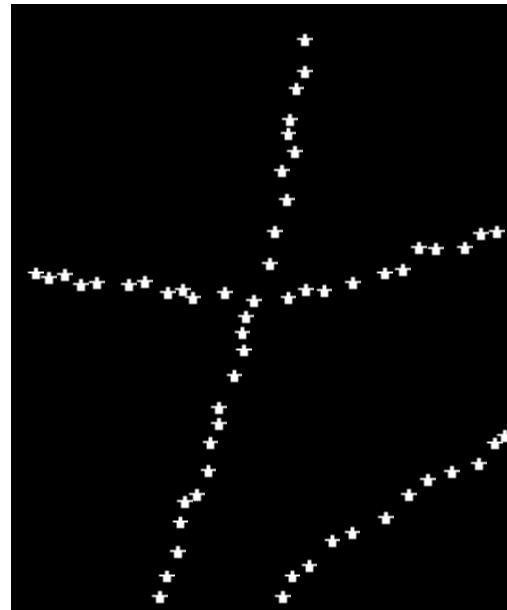


Super-resolution by spatial modulation

Fluorescence image



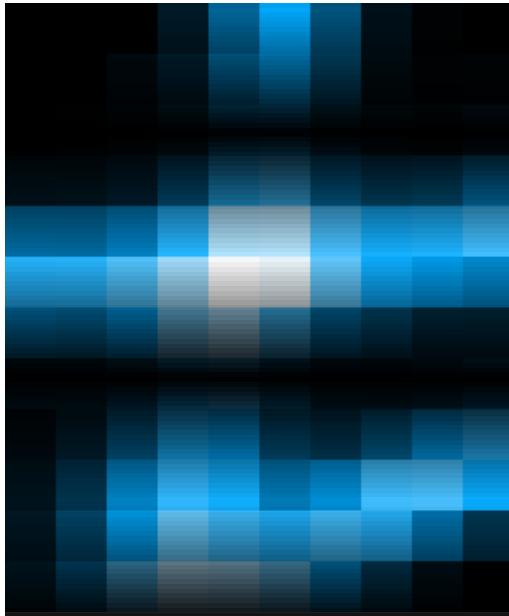
Underlying structure



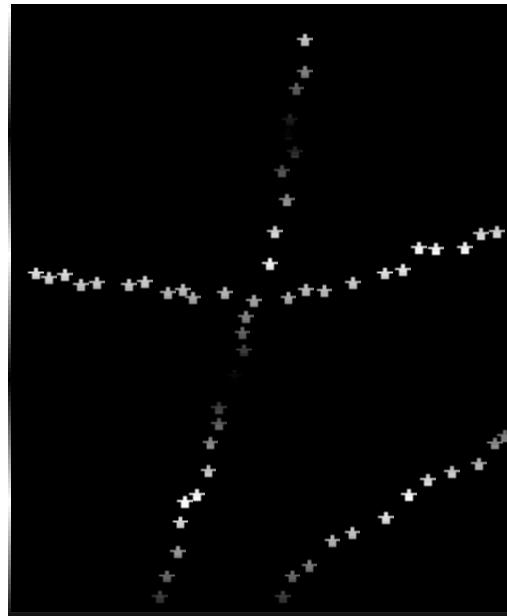
Differential modulation
of the fluorescence response

Super-resolution by differential excitation

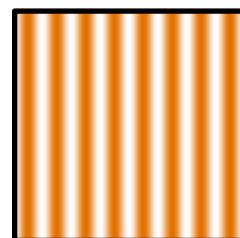
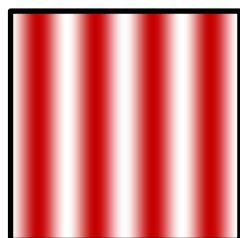
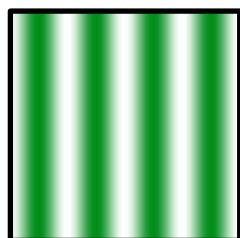
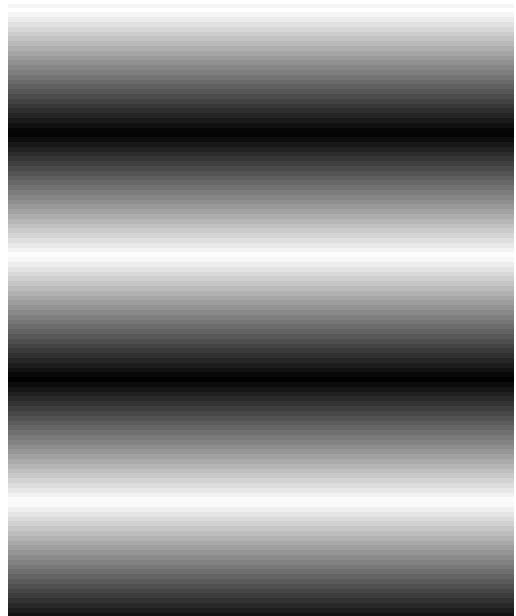
Fluorescence image



Underlying structure



Excitation pattern



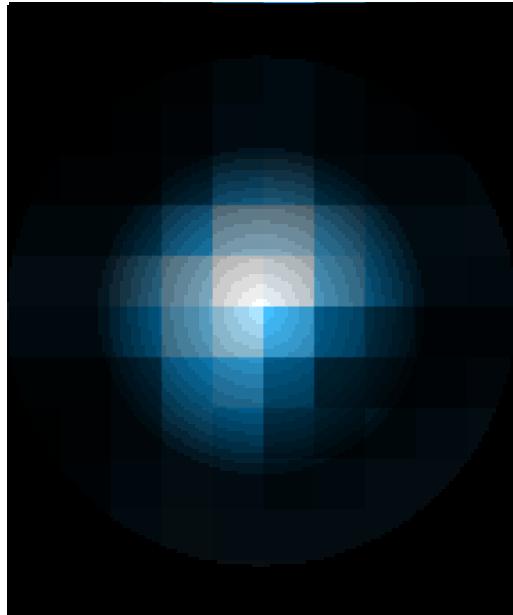
SIM (Gustafsson / Heintzmann)
SSIM (Gustafsson 2005)

Diffraction limited excitation and emission

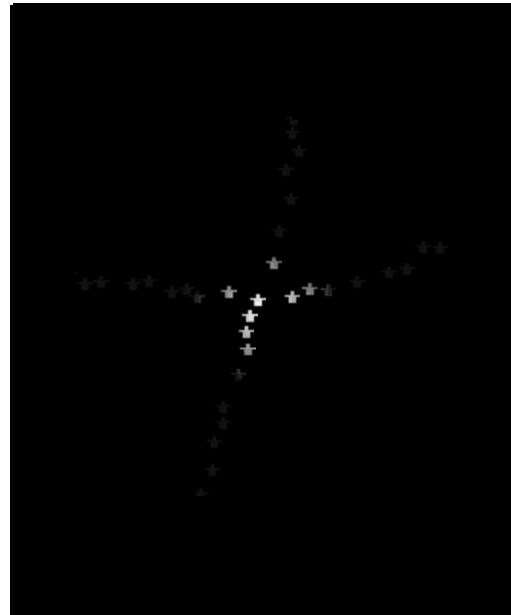
Doubled resolution

Super-resolution by differential depletion

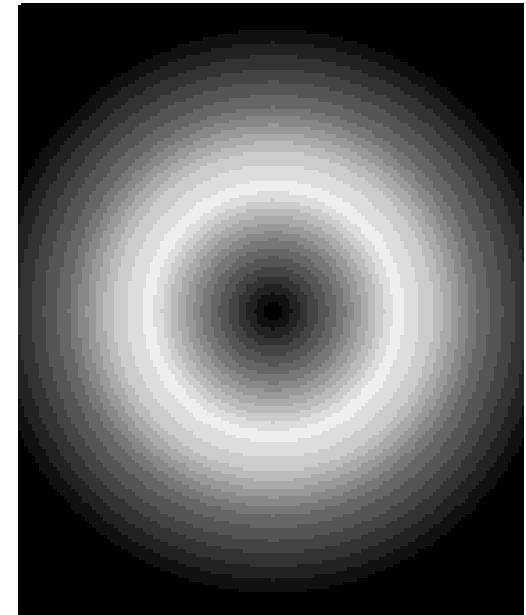
Fluorescence image



Underlying structure



Depletion pattern



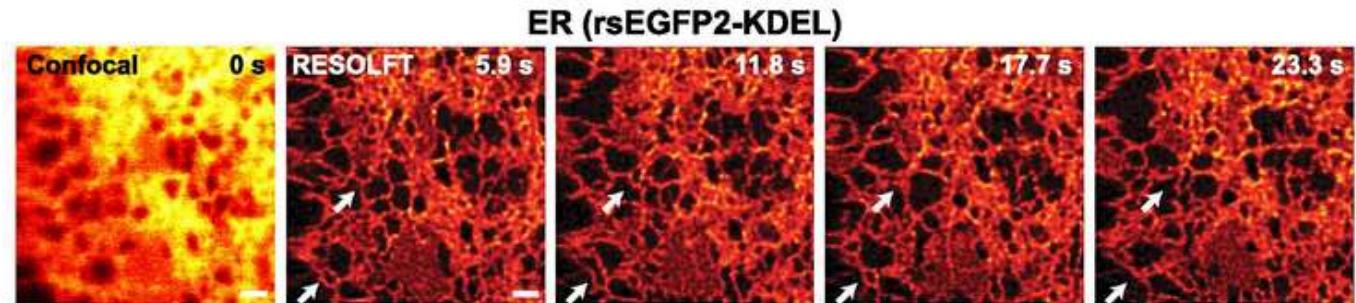
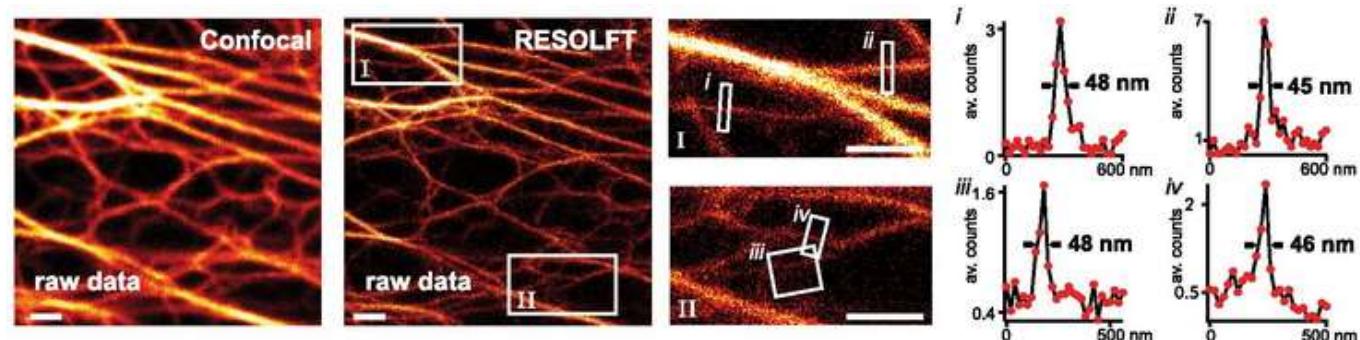
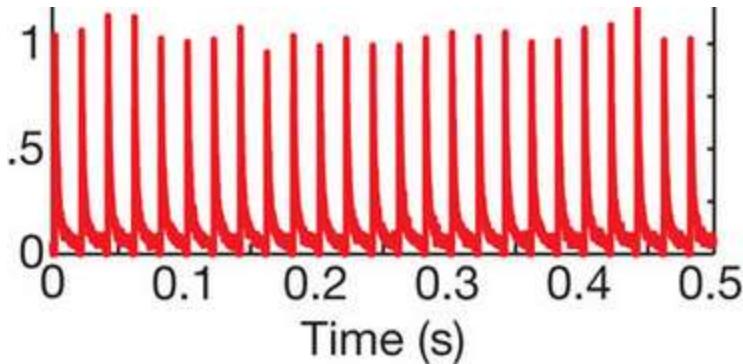
Diffraction limited PSF Saturated depletion = Smaller effective PSF

STED (Hell 1994, Hell 1999)

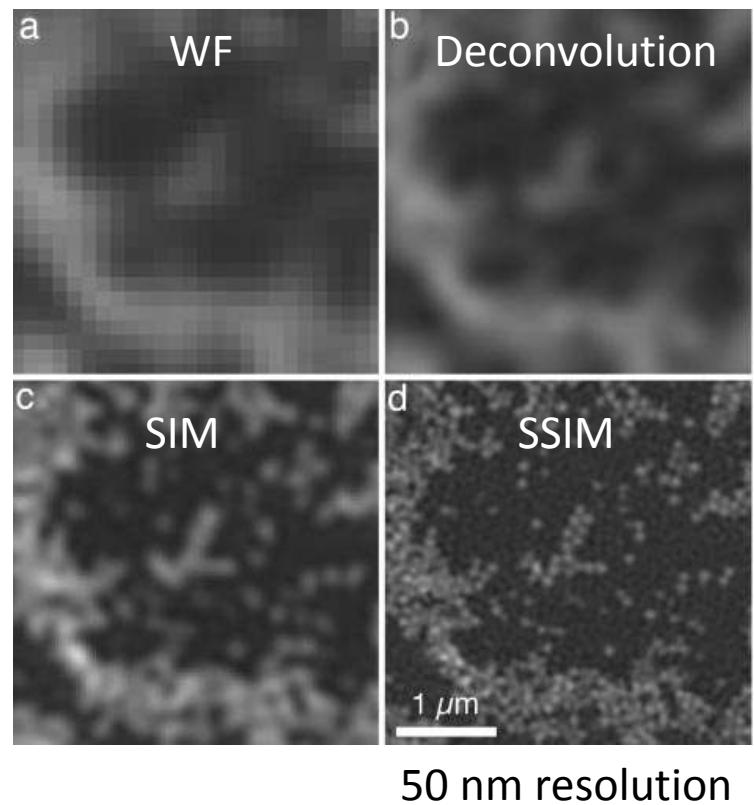
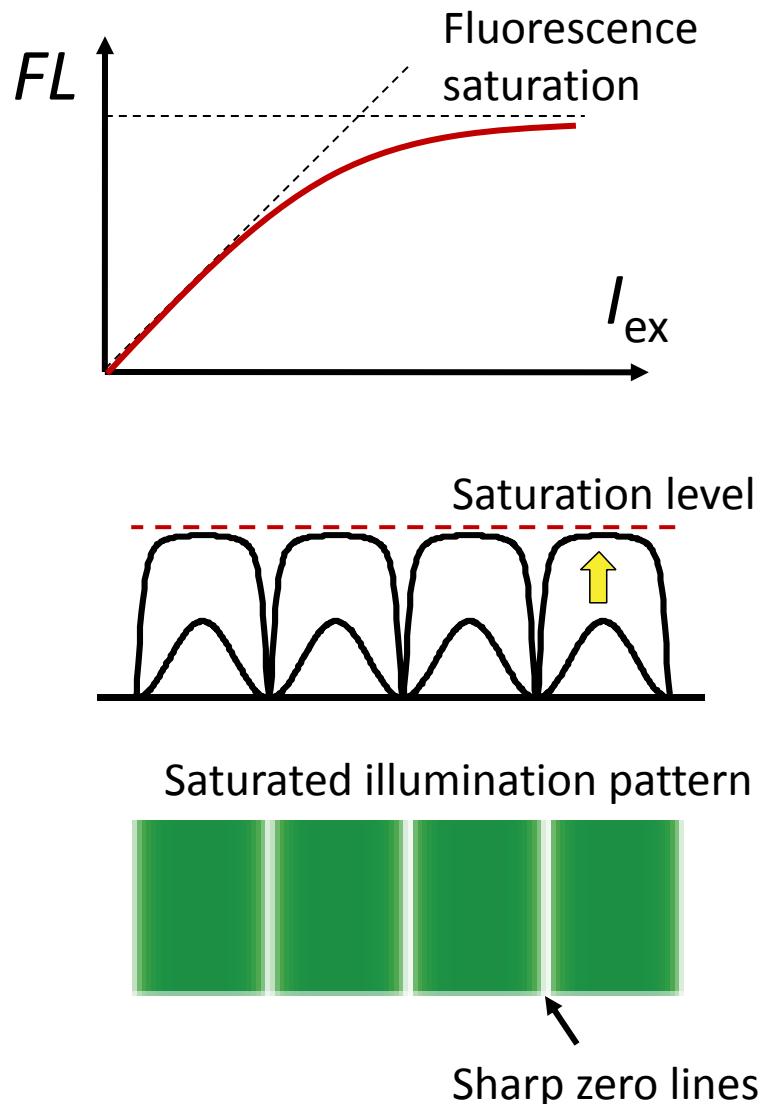
GSD (Hell 1995, Hell 2007)

RESOLFT (Hell 2003, Hell 2011)

RESOLFT by rsEGFP and rsEGFP2



Saturated SIM



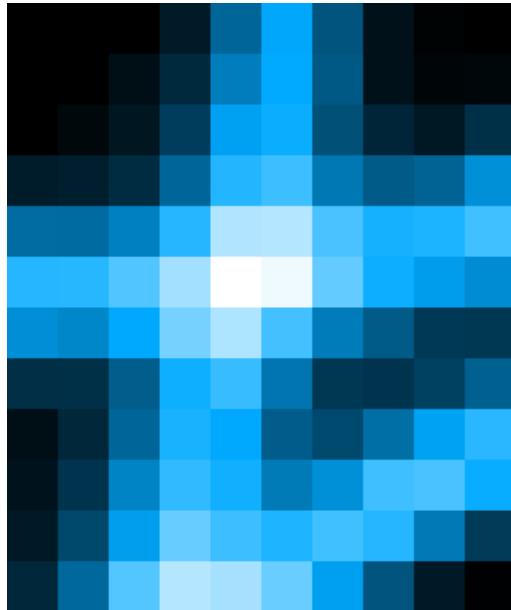
Suffers from fast photobleaching
under saturated excitation condition

Super-resolution by single-molecule switching

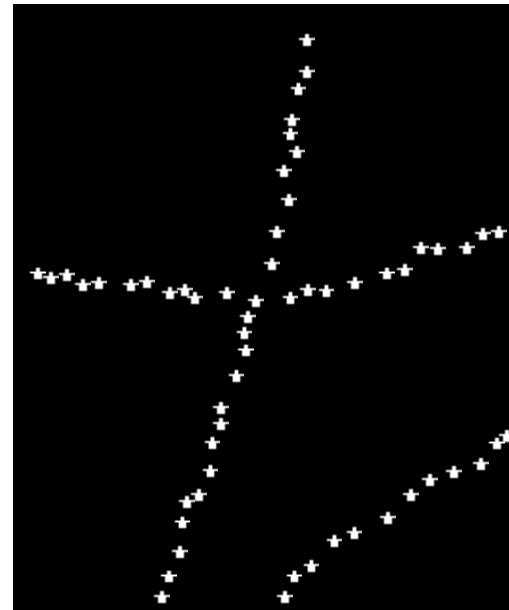


Super-resolution by single-molecule switching

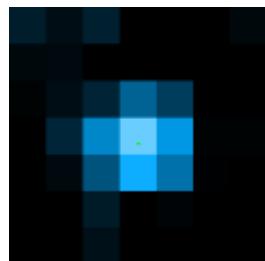
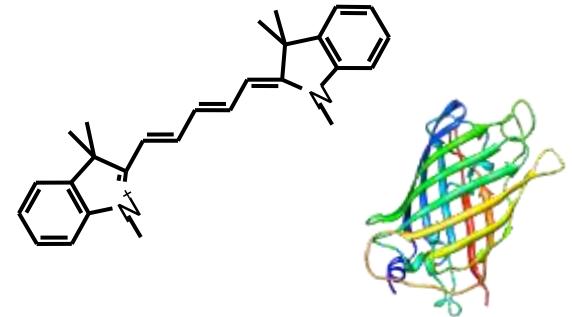
Fluorescence image



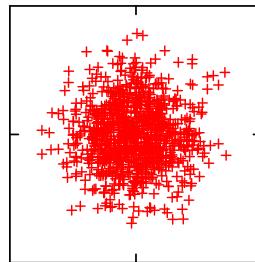
Underlying structure



Photoswitchable molecules



=



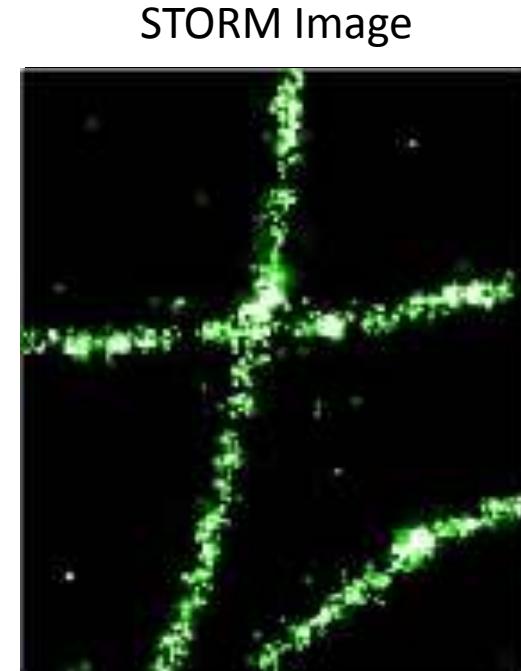
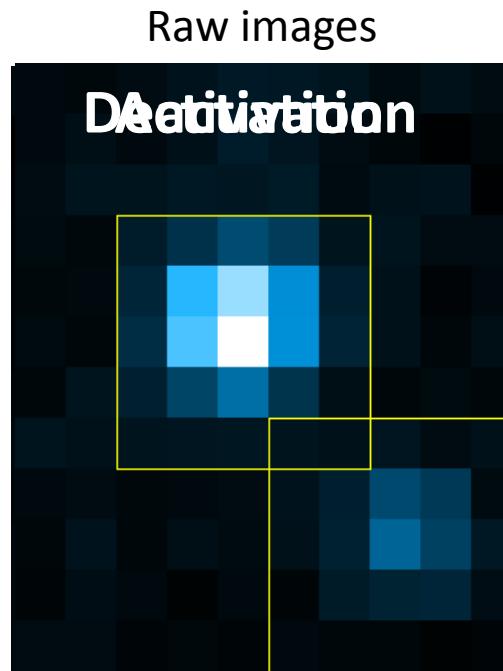
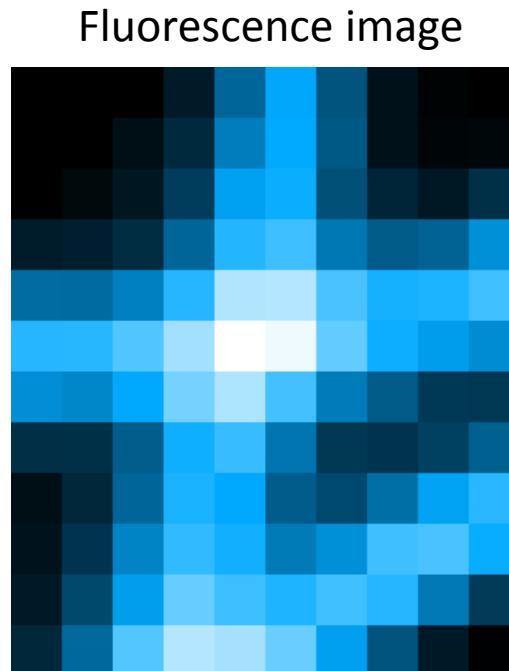
$$D \approx d / \sqrt{N}$$

Single molecule image

N photons

Single-molecule localization

Super-resolution by single-molecule switching



2x real time

STORM = Stochastic Optical Reconstruction Microscopy (Zhuang 2006)

PALM = Photoactivation Localization Microscopy (Betzig & Hess 2006)

FPALM = Fluorescence Photoactivation Localization Microscopy (Hess 2006)

PALMIRA (Hell 2007), **GSDIM** (Hell 2008), **dSTORM** (Sauer 2008), **SMACM** (Moerner 2008)

PAINT (Hochstrasser 2006), **SPRAYPAINT** (Moerner 2011), **SOFI** (Weiss 2009)

Drosophila motoneuron dendrites

