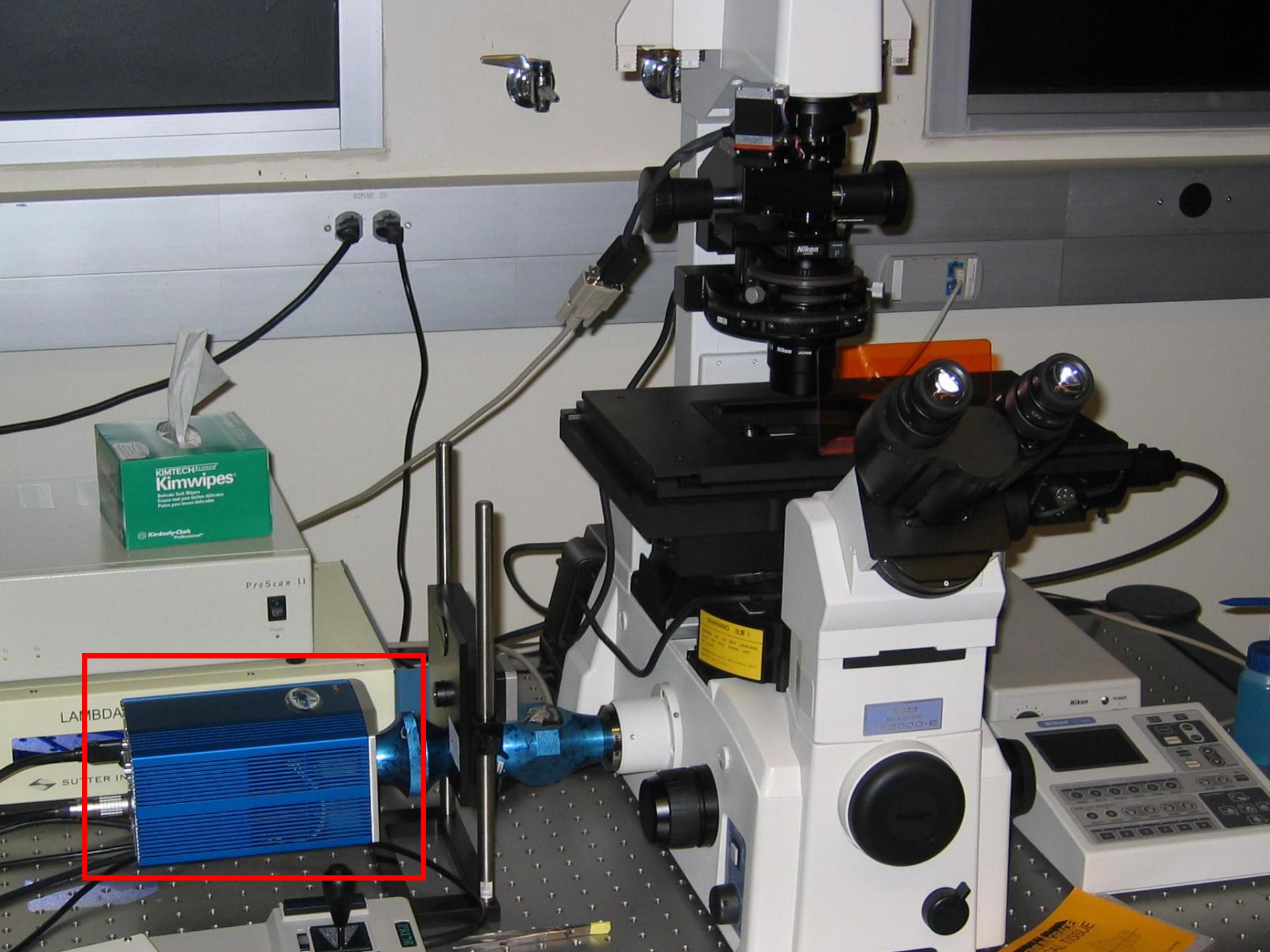


# Digital Cameras in Microscopy

Kurt Thorn

Nikon Imaging Center @ QB3/UCSF



# What does a camera need to do?

- Convert light into an electrical signal
- Accurately measure this signal
- Do this in a spatially resolved way

# CCD architecture

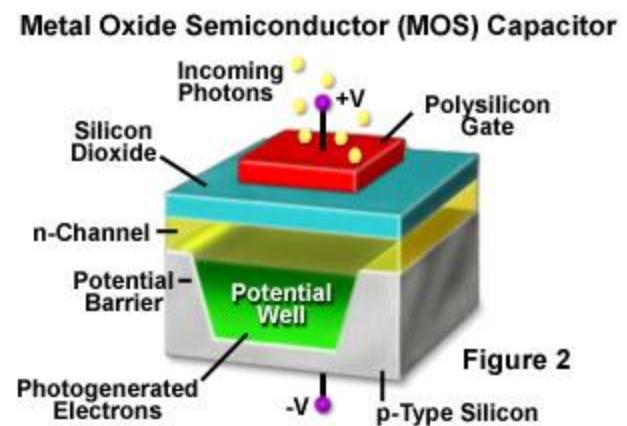
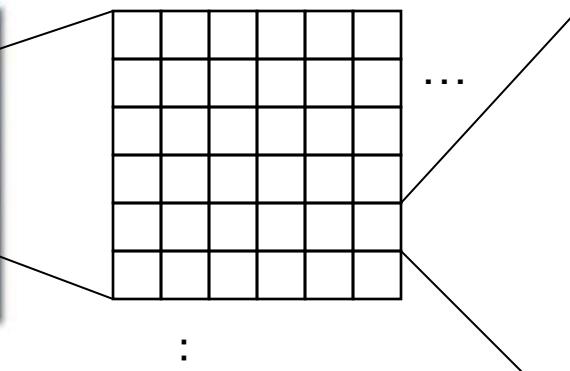
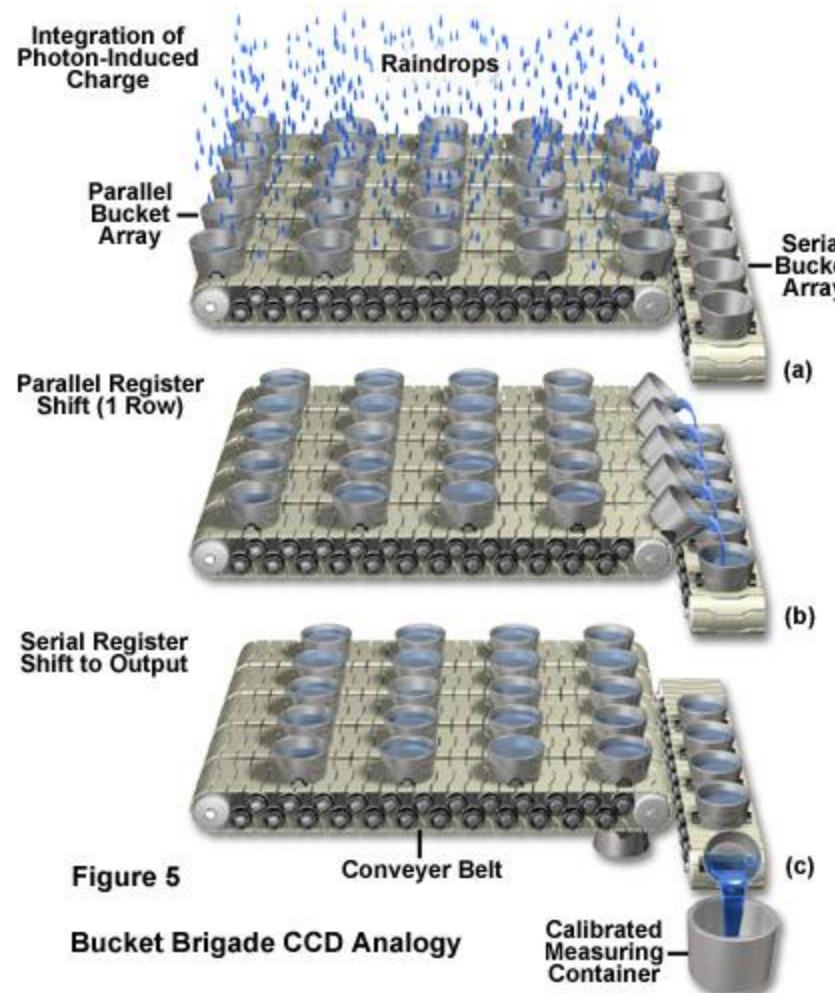


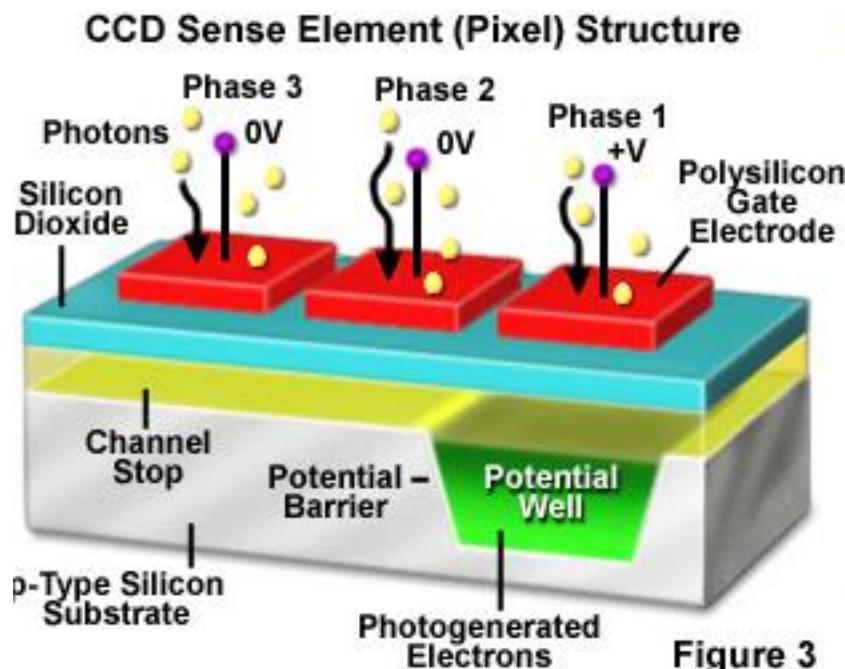
Figure 2

# CCD readout “bucket-brigade” analogy

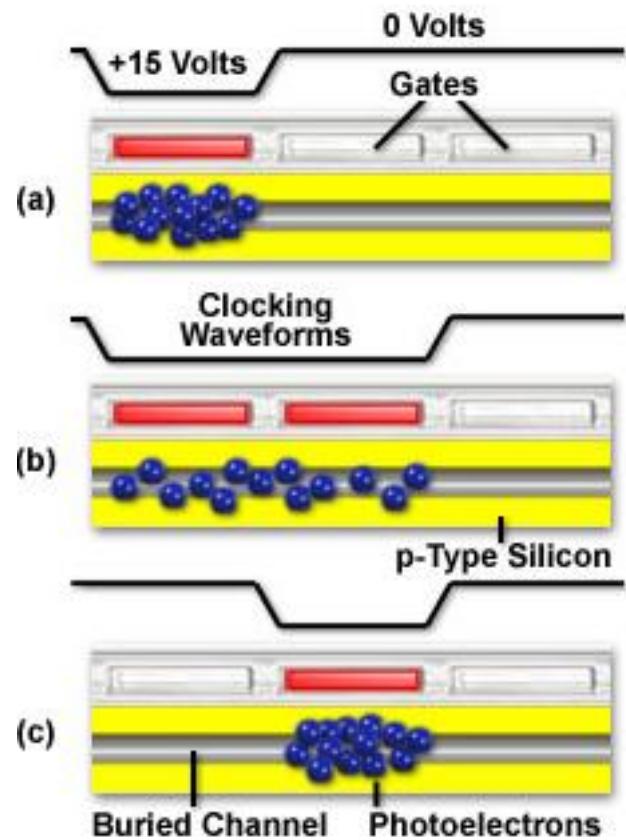


# A little more realistic....

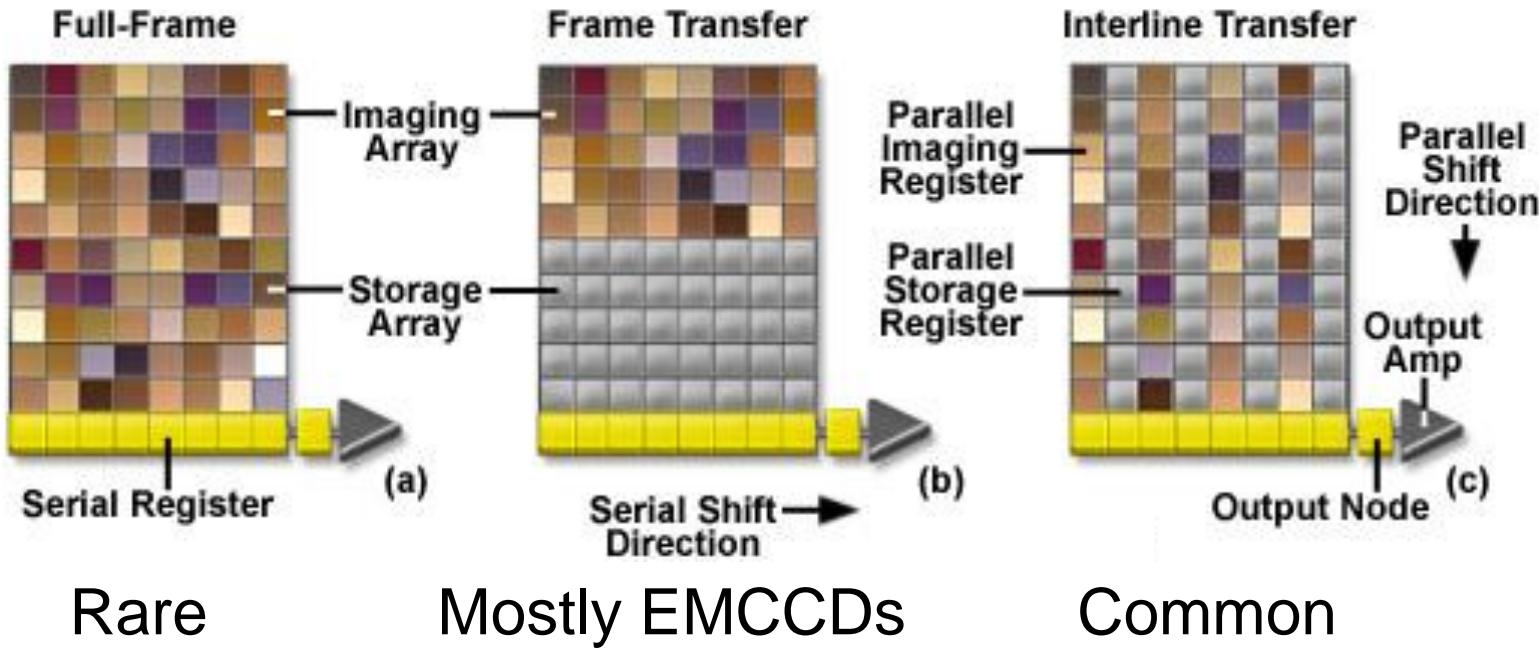
Each pixel is subdivided into three phases



**Three Phase CCD Clocking Scheme**



# CCD Architectures



Full frame CCDs cannot acquire while being read out;  
They also require a mechanical shutter to prevent smearing  
during readout.

# Interline CCDs and microlenses

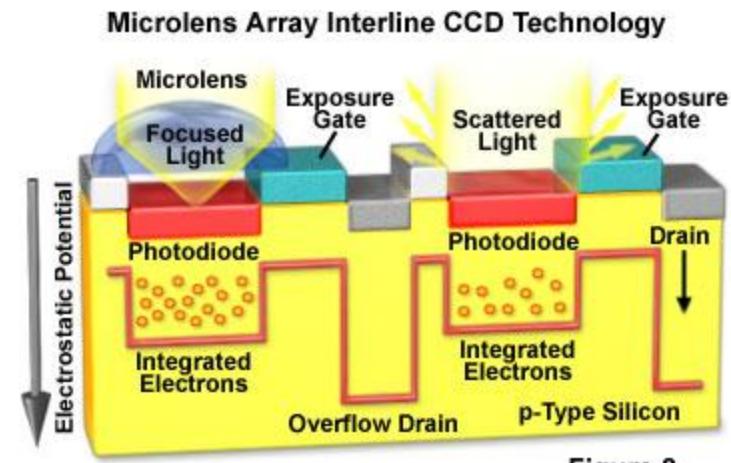
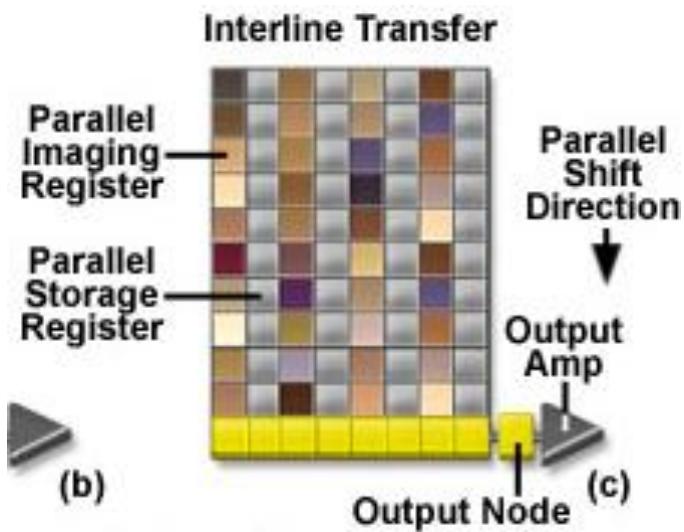
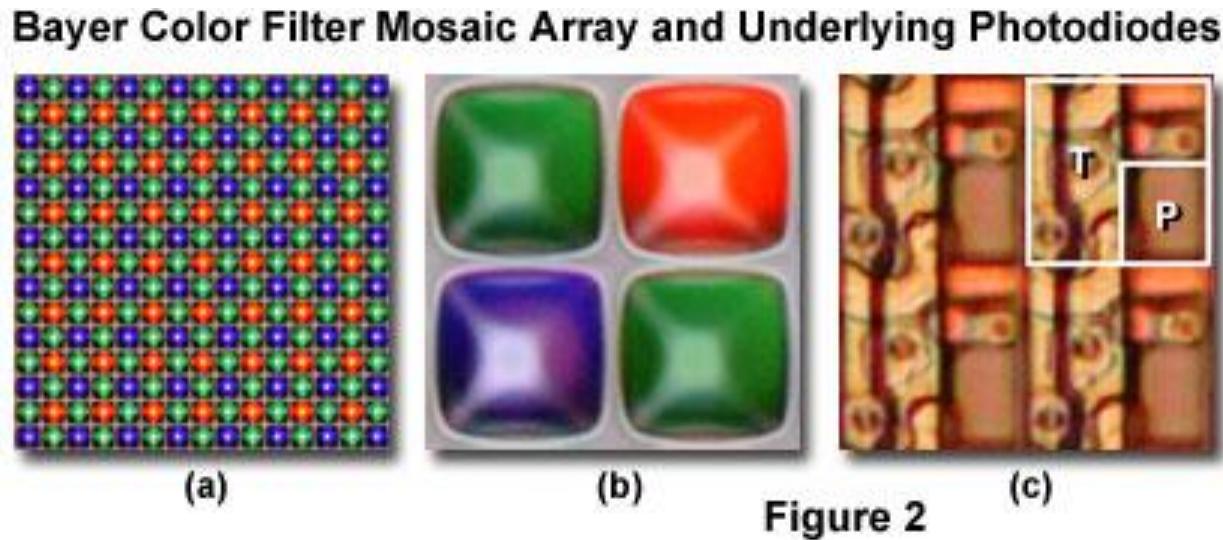


Figure 8

Interline storage registers take up half the light gathering area on the CCD

Solution: use microlenses to focus light onto the light-gathering areas

# Why don't we use color CCDs?

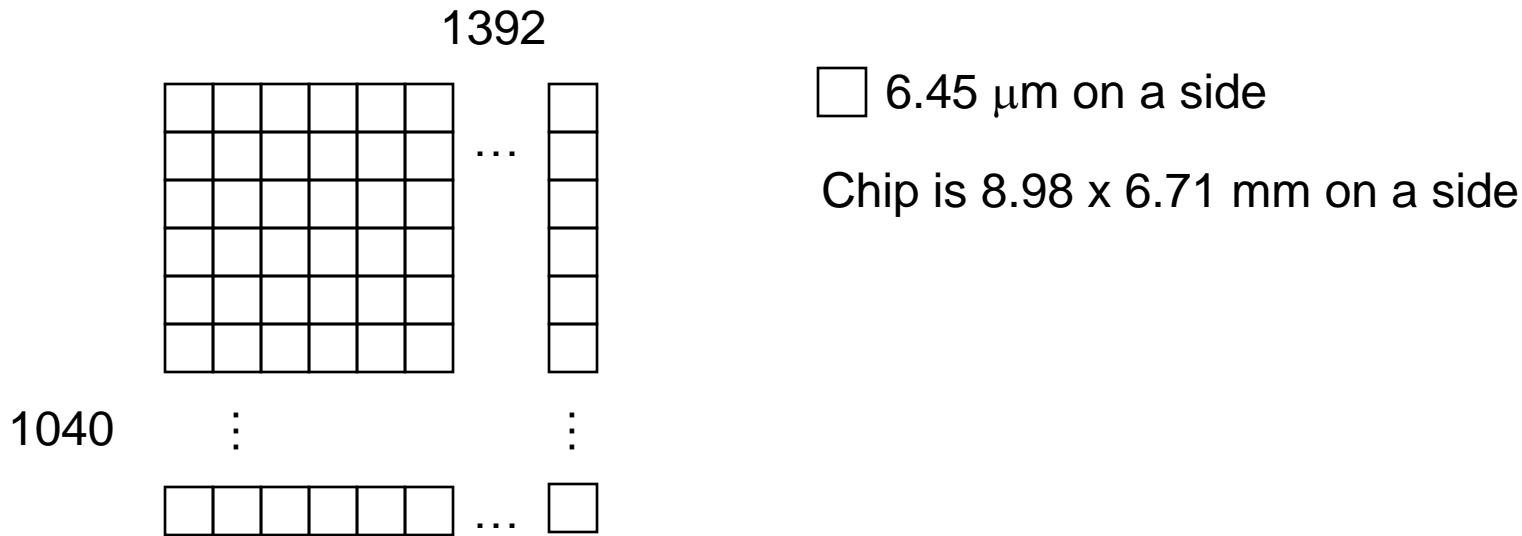


- Four monochrome pixels are required to measure one color pixel
- Your 5MP digital camera really acquires a 1.25 MP red and blue image and a 2.5 MP green image and uses image processing to reconstruct the true color image at 5 MP

# Vital Statistics for CCDs

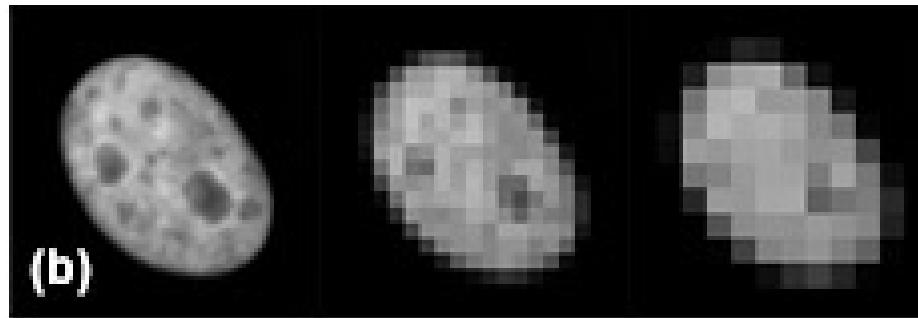
- Pixel size and number
- Quantum efficiency: the fraction of photons hitting the CCD that are converted to electrons
- Full well depth: total number of electrons that can be recorded per pixel
- Read noise
- Dark current (negligible for most biological applications)
- Readout time

# Magnification and CCDs



Typical magnification from sample to camera is roughly objective magnification

# Resolution and magnification



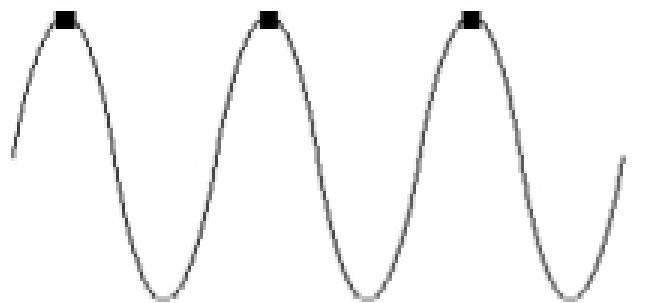
More pixels / resolution element

Where is optimum?

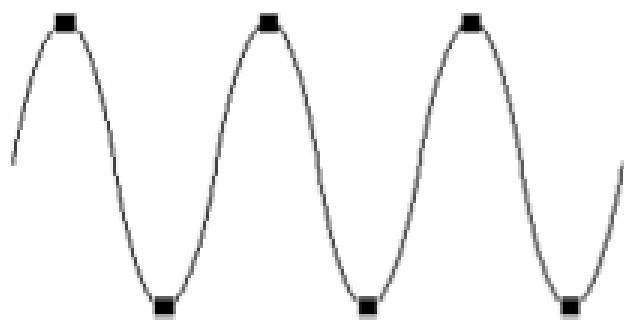
# Nyquist-Shannon Sampling

- How many CCD pixels are needed to accurately reproduce the smallest object that can be resolved by the scope?
- Nyquist-Shannon Sampling theorem:  
Must have at least two pixels per resolvable element
  - 2.5 – 3 is preferable

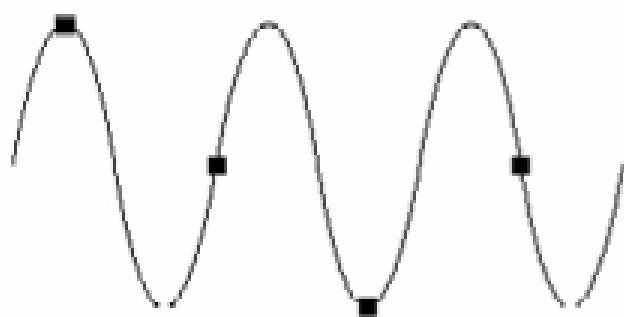
# Nyquist-Shannon Sampling



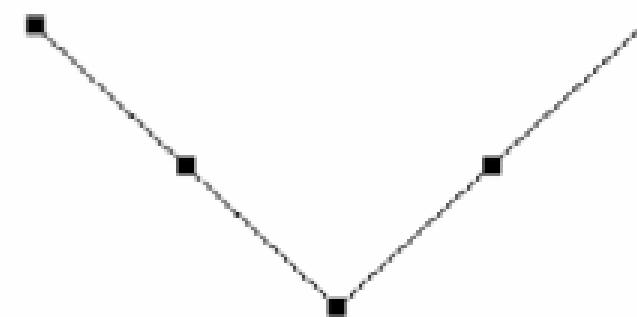
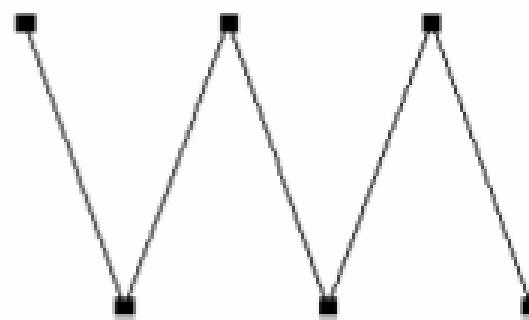
A  
Sampled at  $f$



B  
Sampled at  $2f$



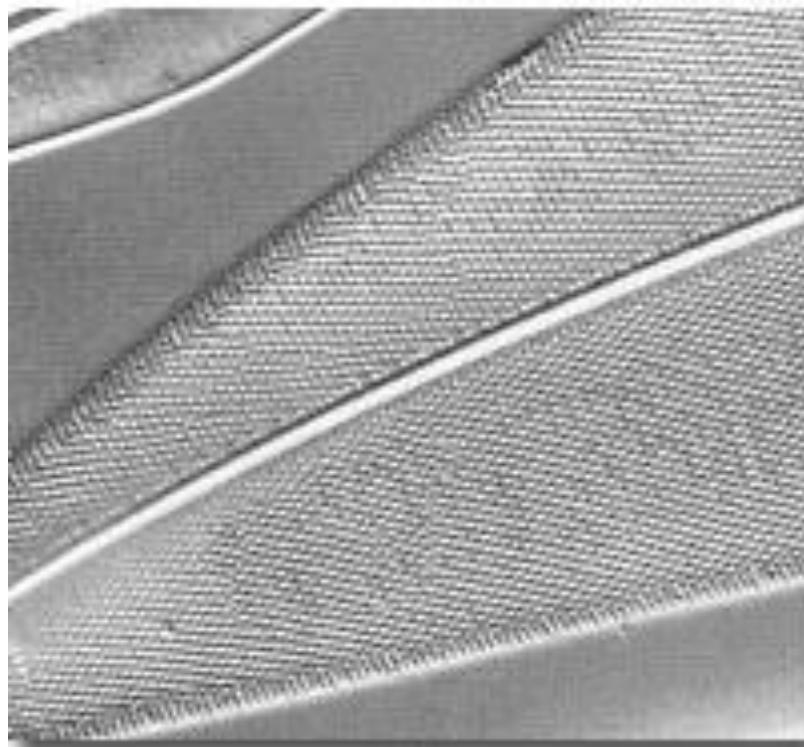
C  
Sampled at  $4f/3$



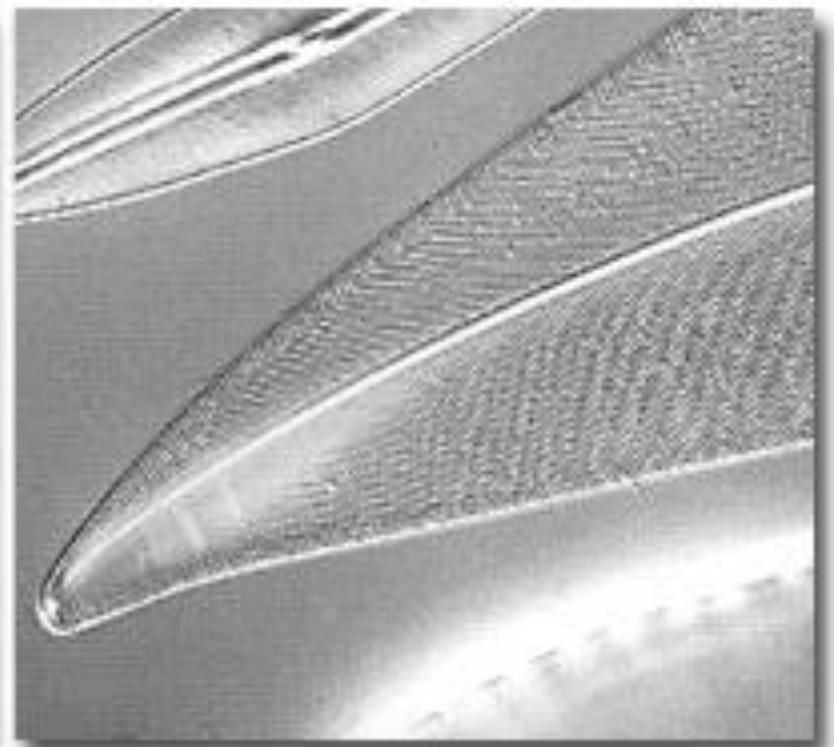
# Resolution and CCDs

- Nyquist-Shannon Sampling theorem:  
Must have at least two pixels per resolvable element
- E.g: if your resolution is 300 nm, your image should be magnified to so that 150 nm in the sample corresponds to at least one pixel on the camera
- If you fail to do this, you will miss features smaller than twice your sampling size
- You can also run into aliasing problems

# Aliasing



Nyquist sampled

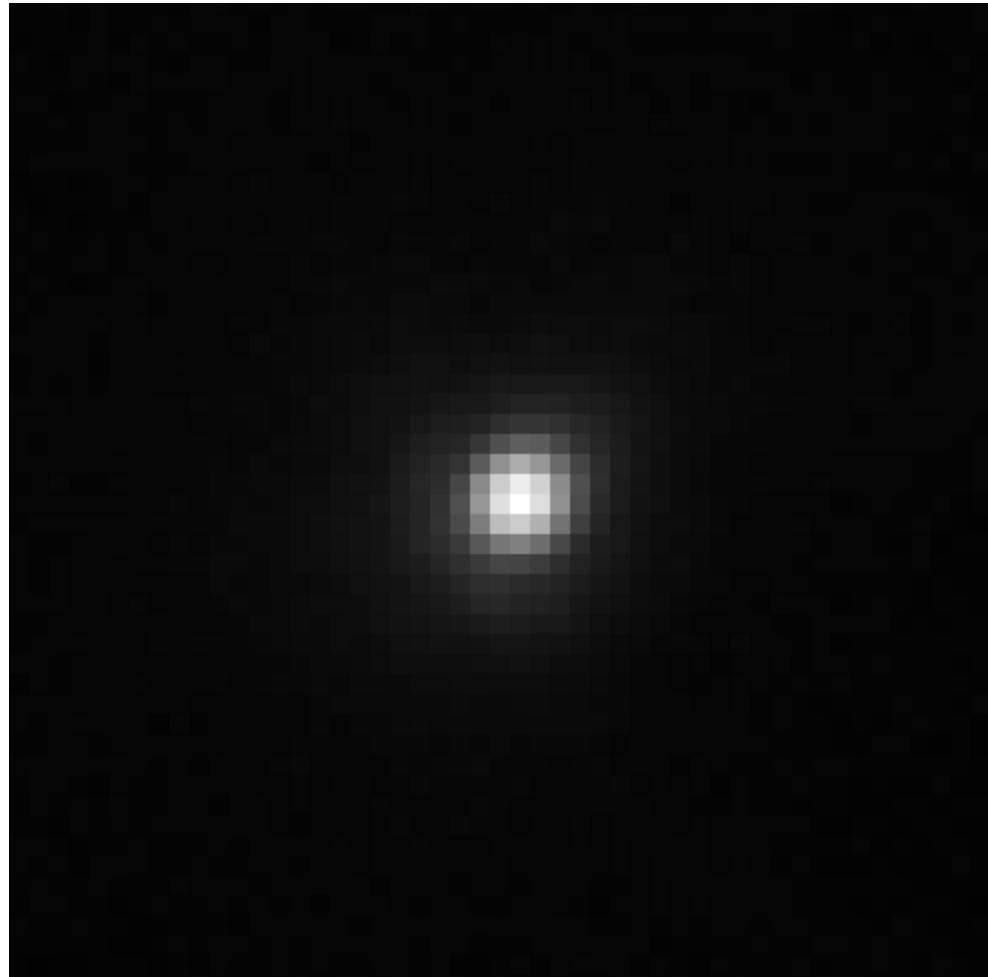


Undersampled

# A resolution-centric view of imaging

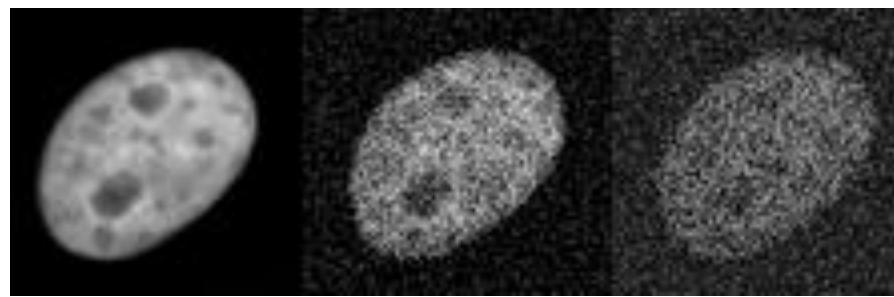
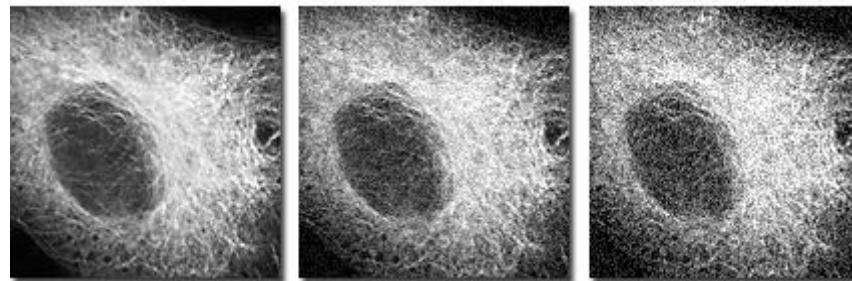
- The objective NA sets the highest resolution you can measure ( $1.4 \text{ NA} \sim 220 \text{ nm}$ )
- To achieve this resolution, 220 nm in your image must cover 2 pixels
- Choose your magnification to achieve this
- For  $6.45 \mu\text{m}$  pixels, we need a total magnification of  $6450/110 = 58.6$
- So for 1.4 NA, a 40x lens would be undersampled, a 60x would be just at the Nyquist limit, and a 100x lens would oversample

# Actual PSF



# Noise

- Longer exposure times are better – why?



←

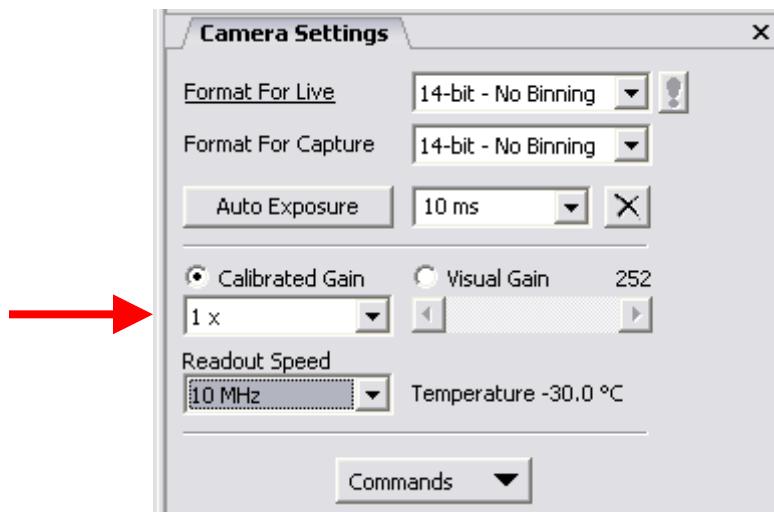
Increasing exposure time

# Noise

- Read noise – inherent in reading out CCD
  - Scales as the square root of readout speed (faster = noisier)
  - For CoolSNAP HQ2:  $4.5 \text{ e}^- / \text{pixel}$  @ 10MHz (180 ms readout)
  - $5.5 \text{ e}^- / \text{pixel}$  @ 20MHz (90ms readout)
- Dark current – thermal accumulation of electrons
  - Cooling helps, so negligible for most applications
  - CoolSNAP HQ2:  $0.001 \text{ e}^- / \text{pixel} / \text{s}$  (@ -30°C)

# Noise

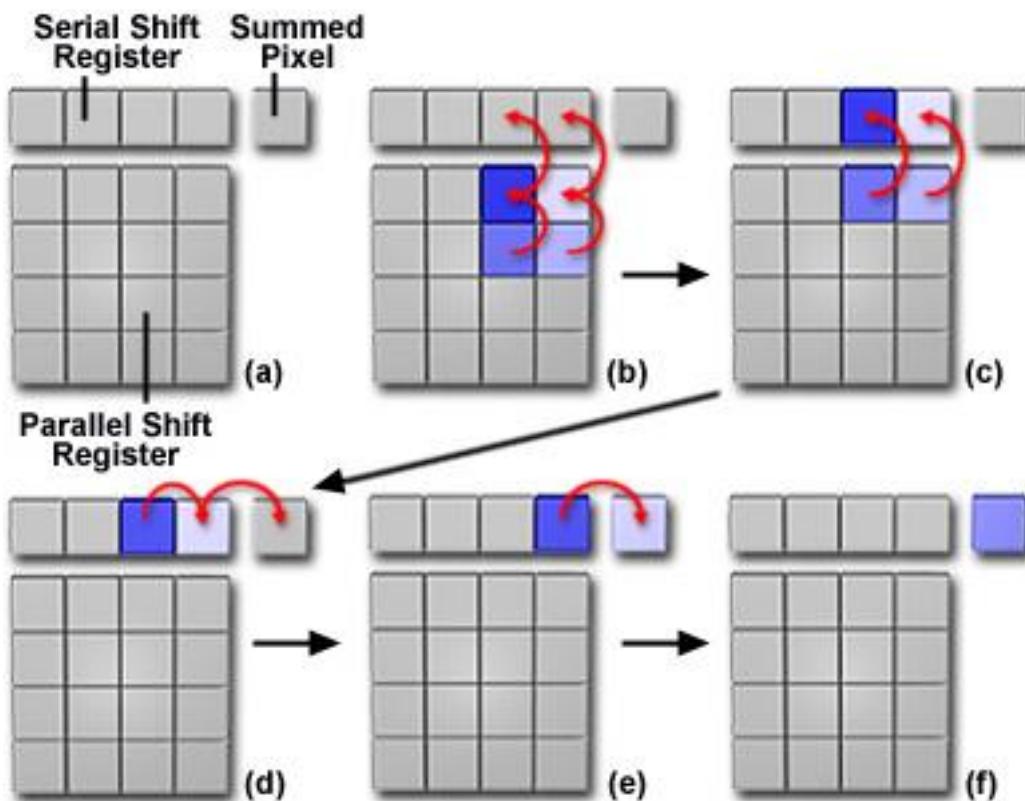
- Photon Shot Noise: Due to the fact that photons are particles and collected in integer numbers
  - Square root of the number of photons
- 1 photon  $\neq$  1 count in your image – depends on the camera (A/D) gain
- Zero photons collected doesn't result in zero being measured on the camera – it has an offset



# Signal/Noise Ratio (SNR)

- Signal = # of photons
- Noise =  $\sqrt{(\text{read noise}^2 + (\# \text{ of photons}))}$
- At low photon numbers, read noise dominates
- At high photon numbers,  
$$\begin{aligned}\text{SNR} &= (\# \text{ of photons}) / \sqrt{(\# \text{ of photons})} \\ &= \sqrt{(\# \text{ of photons})}\end{aligned}$$
- So, to double your SNR, you need to acquire four times as long (or 2x2 bin)

# Binning



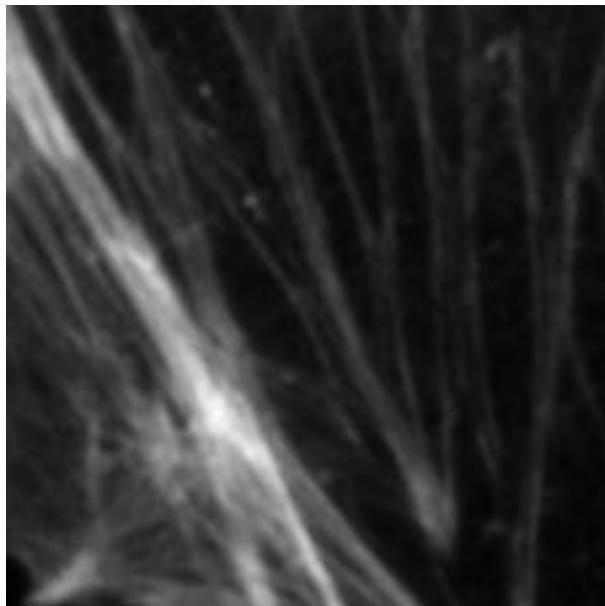
- Read out 4 pixels as one
- Increases SNR by 2x
- Decreases read time by 2 or 4x
- Decreases resolution by 2x

# Signal/Noise Ratio (SNR)

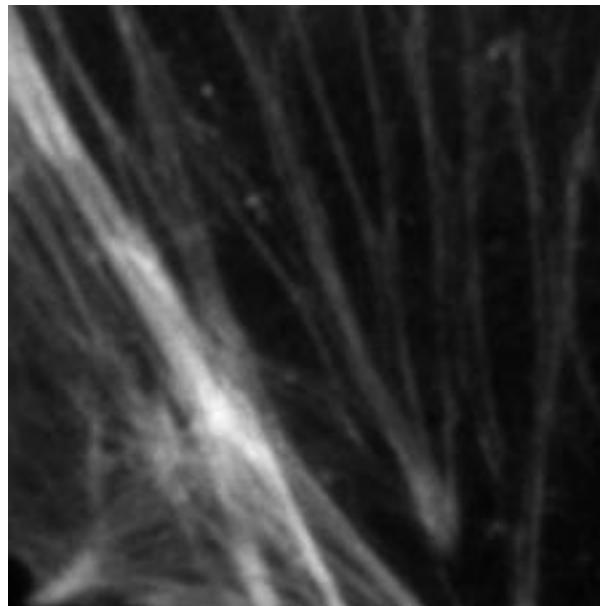
- Read noise dominates whenever  
 $\text{read noise}^2 = \# \text{ of photons}$
- 8 e- read noise → 64 photons
- 16 e- read noise → 256 photons
- 50 e- read noise → 2500 photons
- Full range on Coolsnap HQ2 with 4x gain: 4095 photons

# What does this look like?

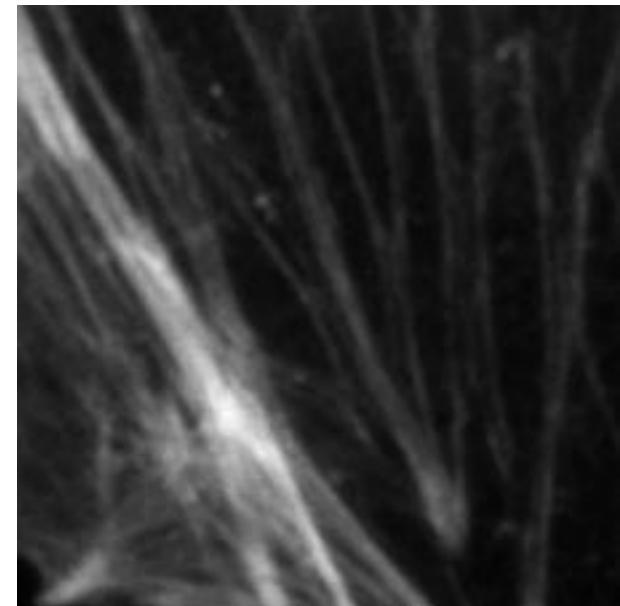
1000 photons / pixel on average; ~5000 in brightest areas



Test image



no read noise

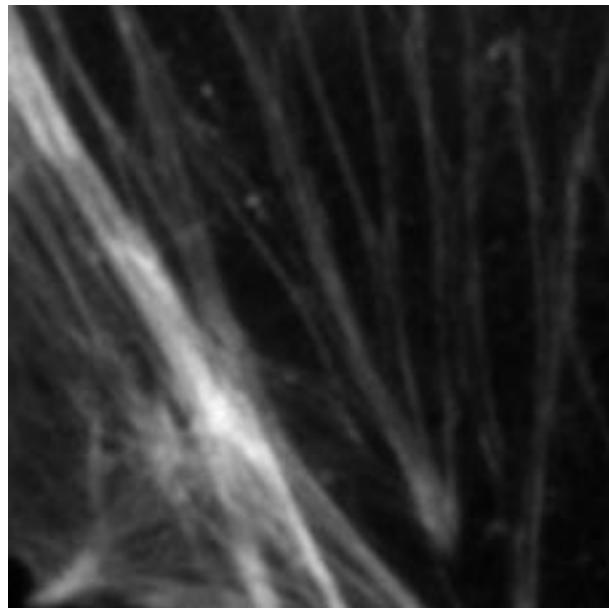


5 e<sup>-</sup> read noise

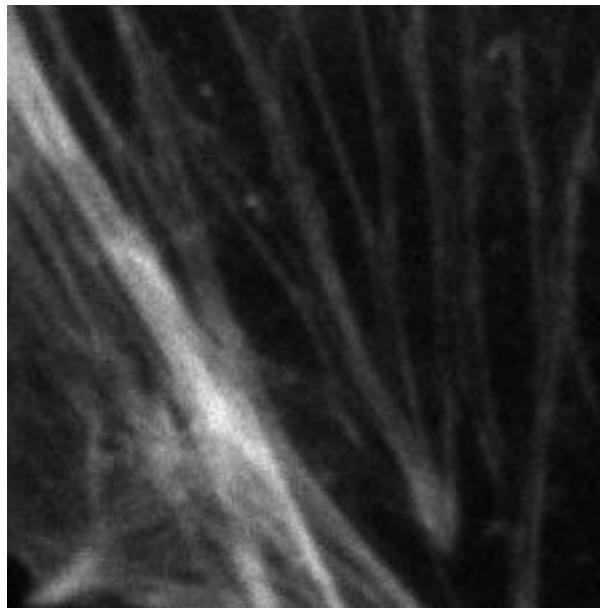
Photon shot noise ~ 6x read noise

# What does this look like?

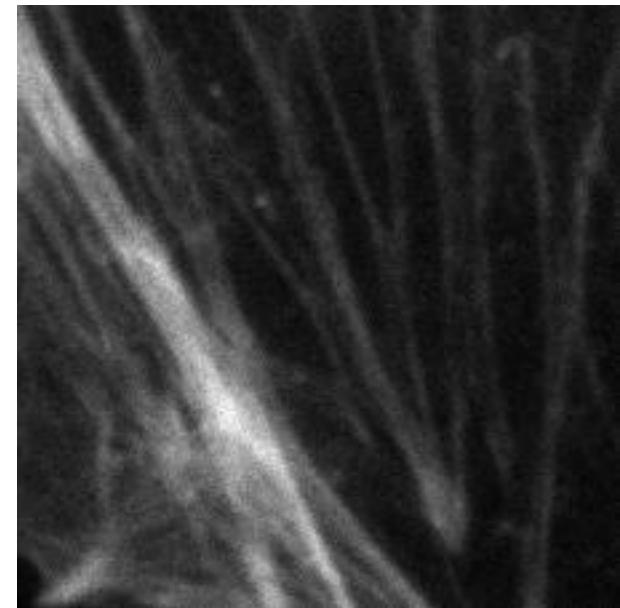
100 photons / pixel on average; ~500 in brightest areas



Test image



no read noise

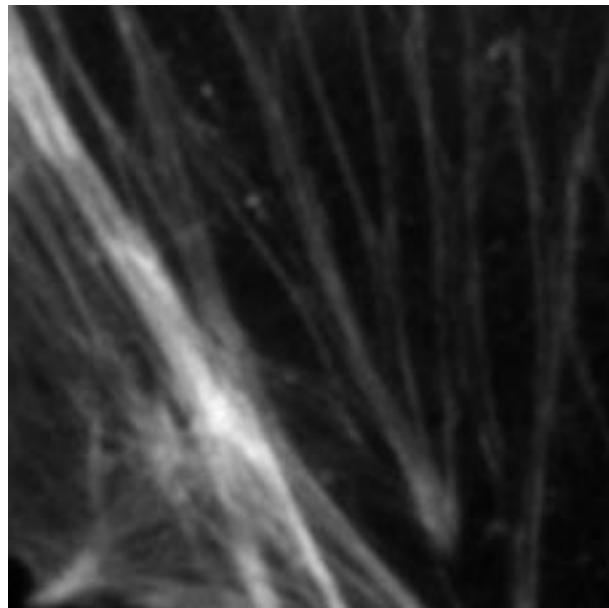


5 e<sup>-</sup> read noise

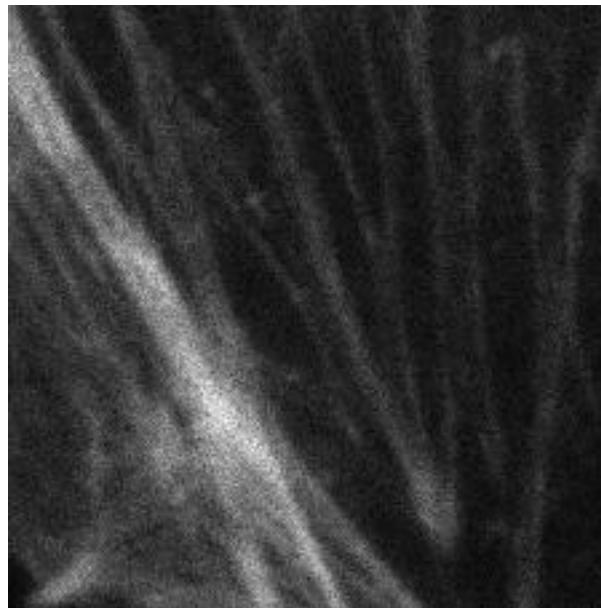
Photon shot noise = 2x read noise

# What does this look like?

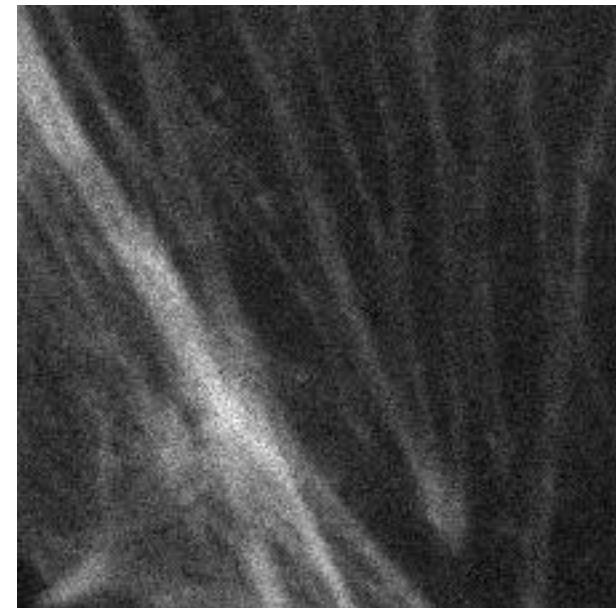
25 photons / pixel on average; ~125 in brightest areas



Test image



no read noise

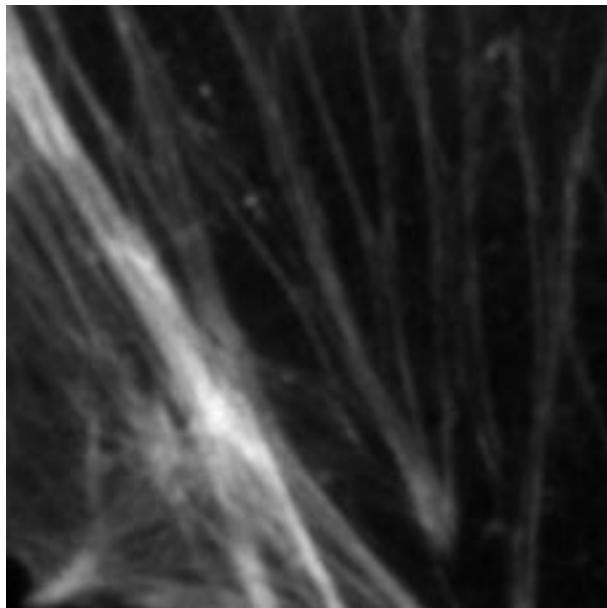


5 e<sup>-</sup> read noise

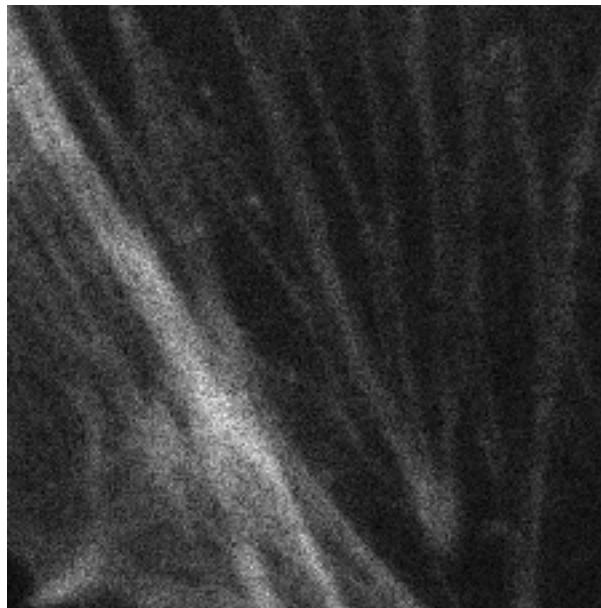
Photon shot noise = read noise

# What does this look like?

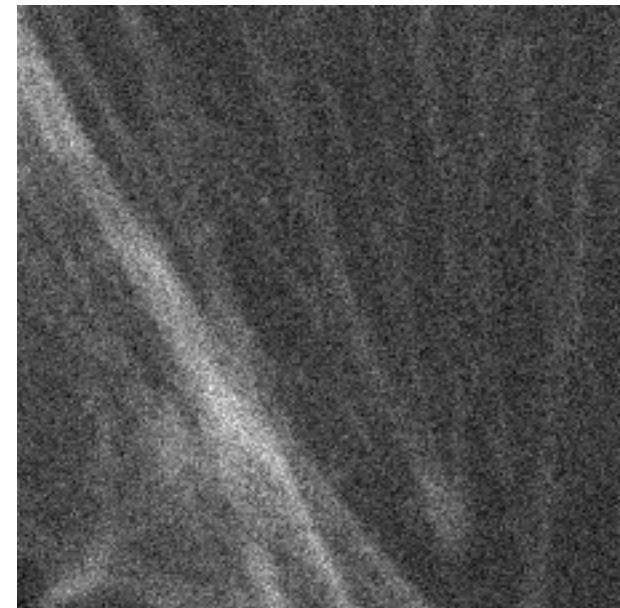
10 photons / pixel on average; ~50 in brightest areas



Test image



no read noise

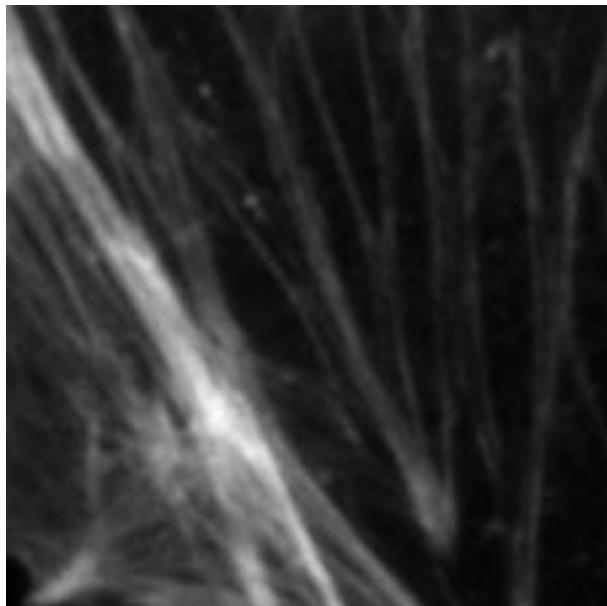


5 e<sup>-</sup> read noise

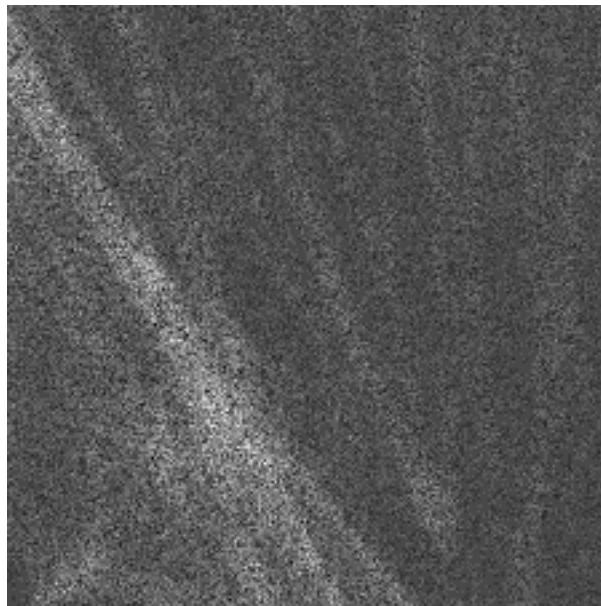
Photon shot noise ~ 2/3 read noise

# What does this look like?

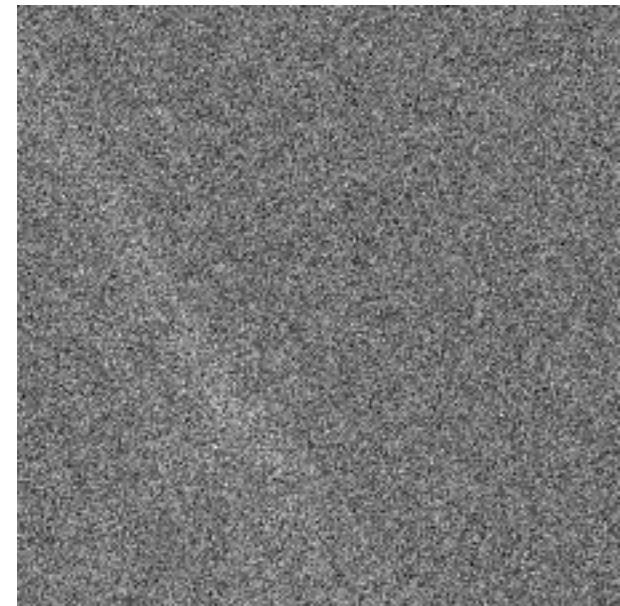
1 photon / pixel on average; ~5 in brightest areas



Test image



no read noise



5 e<sup>-</sup> read noise

Photon shot noise ~ 1/5 read noise

# Beating the read-out noise EMCCD

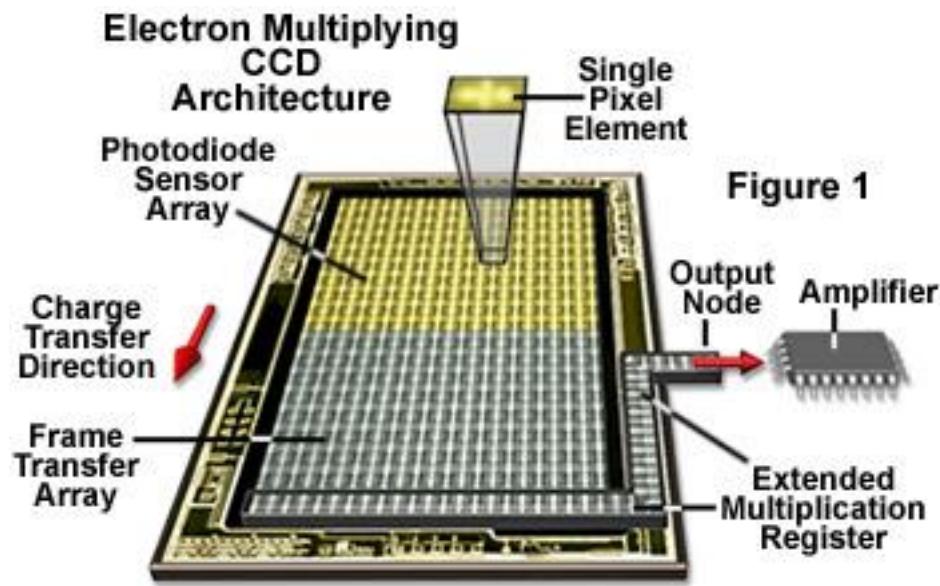


Figure 1

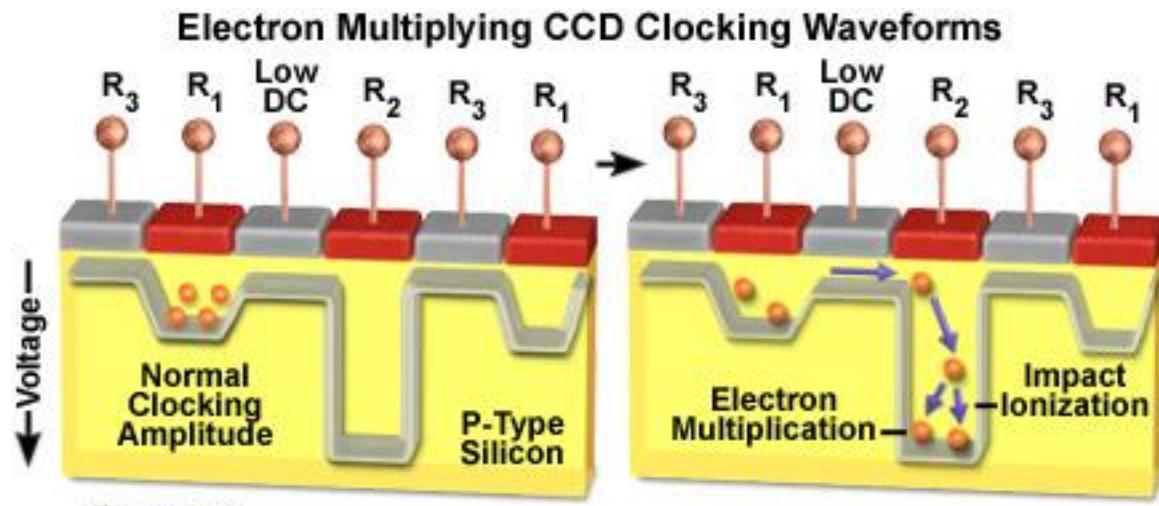


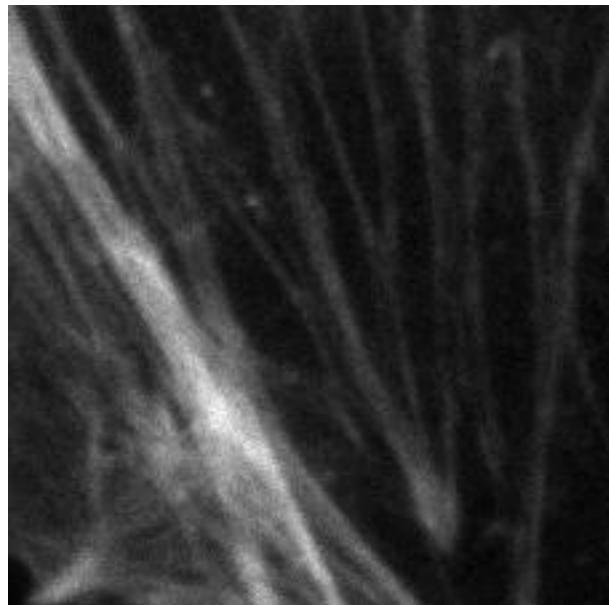
Figure 4

# EMCCD result

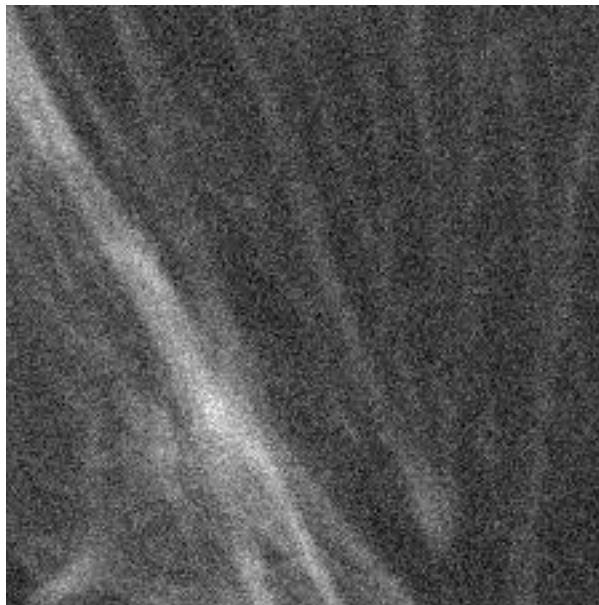
- Fast noisy CCD – runs at 30 fps, but 50 e<sup>-</sup> read noise
- Multiply signal by 100-fold – now read noise looks like 0.5 e<sup>-</sup>
- Downside – multiplication process adds additional Poisson noise, so your QE looks like it's halved
- Upside – you get to image fast without worrying about read noise

# Hypothetical CCD/EMCCD comparison

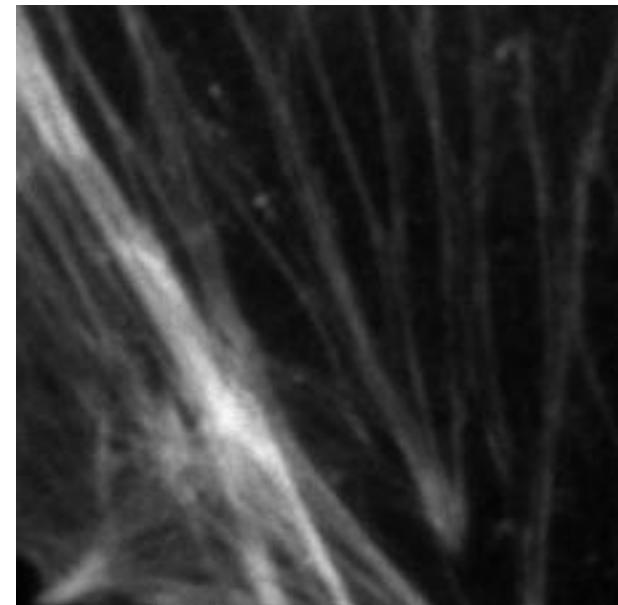
100 photon / pixel on average; ~500 in brightest areas



Slow scan CCD  
 $4e^-$  read noise  
(1 sec read time)



Video rate CCD,  
 $50e^-$  read noise



Video rate EMCCD  
 $50e^-$  read noise  
200x gain

# CCDs vs. CMOS

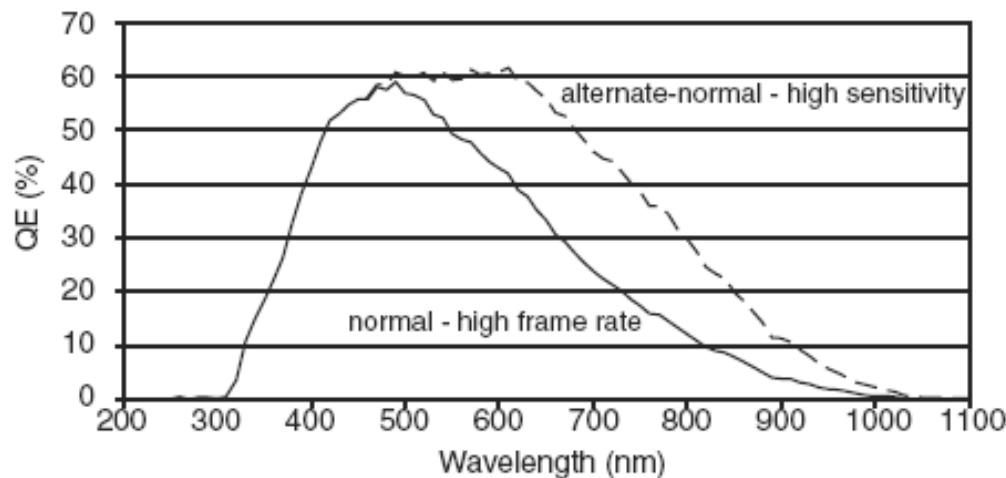
- CCDs:
  - Output electrons
  - Off chip amplifier and A/D converter generate output signal
  - Slow, low noise
- CMOS
  - Outputs digital signal
  - Each pixel has its own amplifier; each row has its own A/D converter
  - Fast, noisy

# New: sCMOS

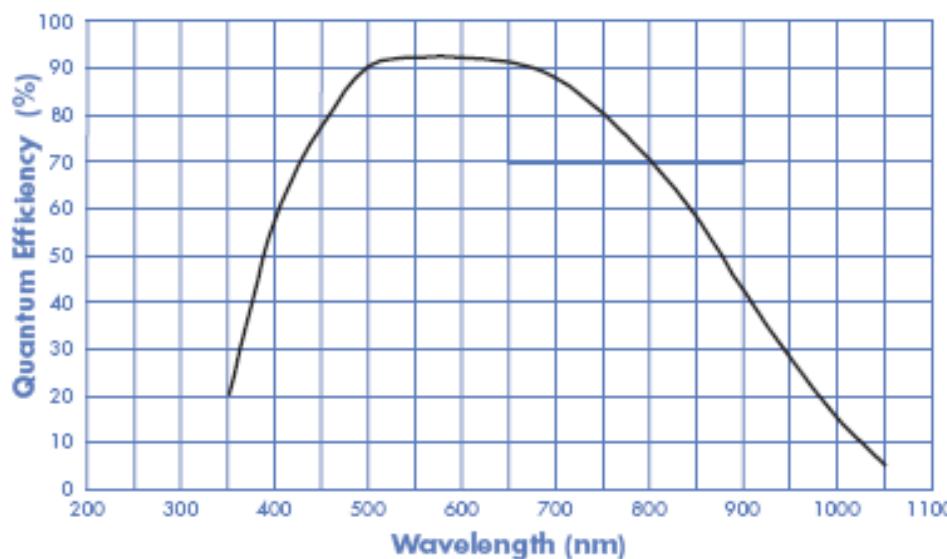
## Scientific CMOS

- Like CMOS, but better
  - Differences are proprietary
  - Vendors: Hamamatsu, Andor, PCO
- Specs (Andor Neo):
  - 5.5 megapixels (2560 x 2160)
  - 6.5  $\mu\text{m}$  pixels
  - 100 fps readout
  - 1.5 e<sup>-</sup> read noise

# Quantum efficiency



HQ2

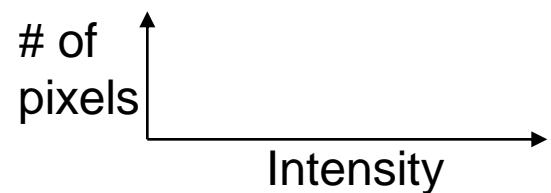
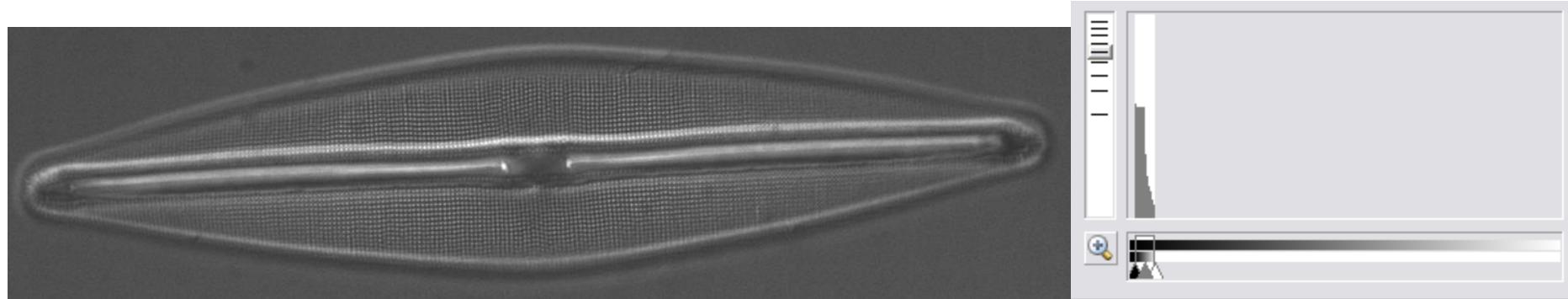
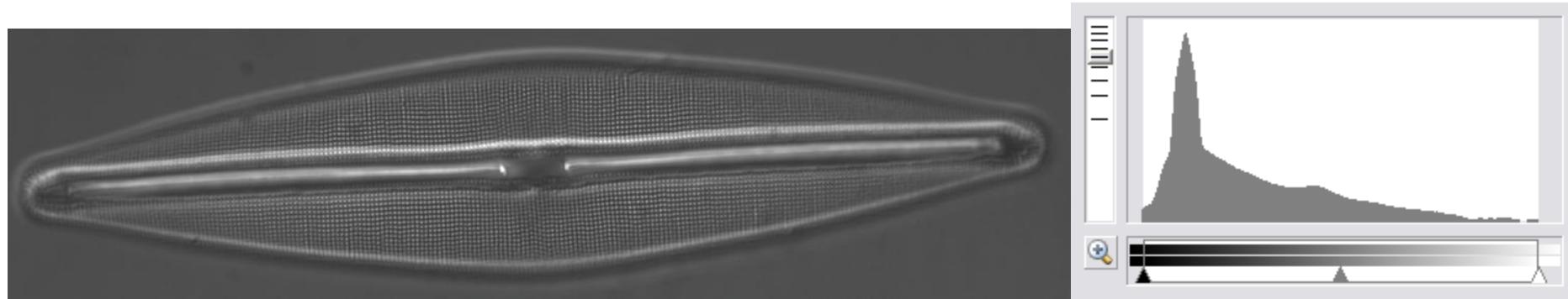


Cascade II

# How many intensity levels can you distinguish?

- Full well capacity (16 000 e<sup>-</sup>)
- Readout noise: 5e-
- Dynamic range:
  - FWC/readout noise: 3200
  - $0.9 * \text{FWC} / (3 * \text{readout noise}) = 960$
- (Human eye ~ 100)

# Check your histogram



# Improve Signal/noise

- Use bright, non-bleaching fluorophores
- Best possible optics (high NA lenses, high QE camera, high transmission filters, reduce spherical aberration, no phase!)
- Minimize optical elements between your sample and the camera (use bottom port!)
- Work in the dark, use clean cover slips, reagents, etc..
- Increase exposure or use frame averaging
- Binning (at the expense of spatial resolution)

# Acknowledgements

- [www.microscopyu.com](http://www.microscopyu.com)
- Nico Stuurman
- James Pawley, Ed. “Handbook of Biological Confocal Microscopy, 3<sup>rd</sup> ed.), especially appendix 3: “More than you ever really wanted to know about charge-coupled devices”
- James Janesick, “Scientific Charge Coupled Devices” (if you really, really, want to know about CCDs)