

Interpreting your gene list

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UCLA School of Medicine
Division of Hematology-Oncology

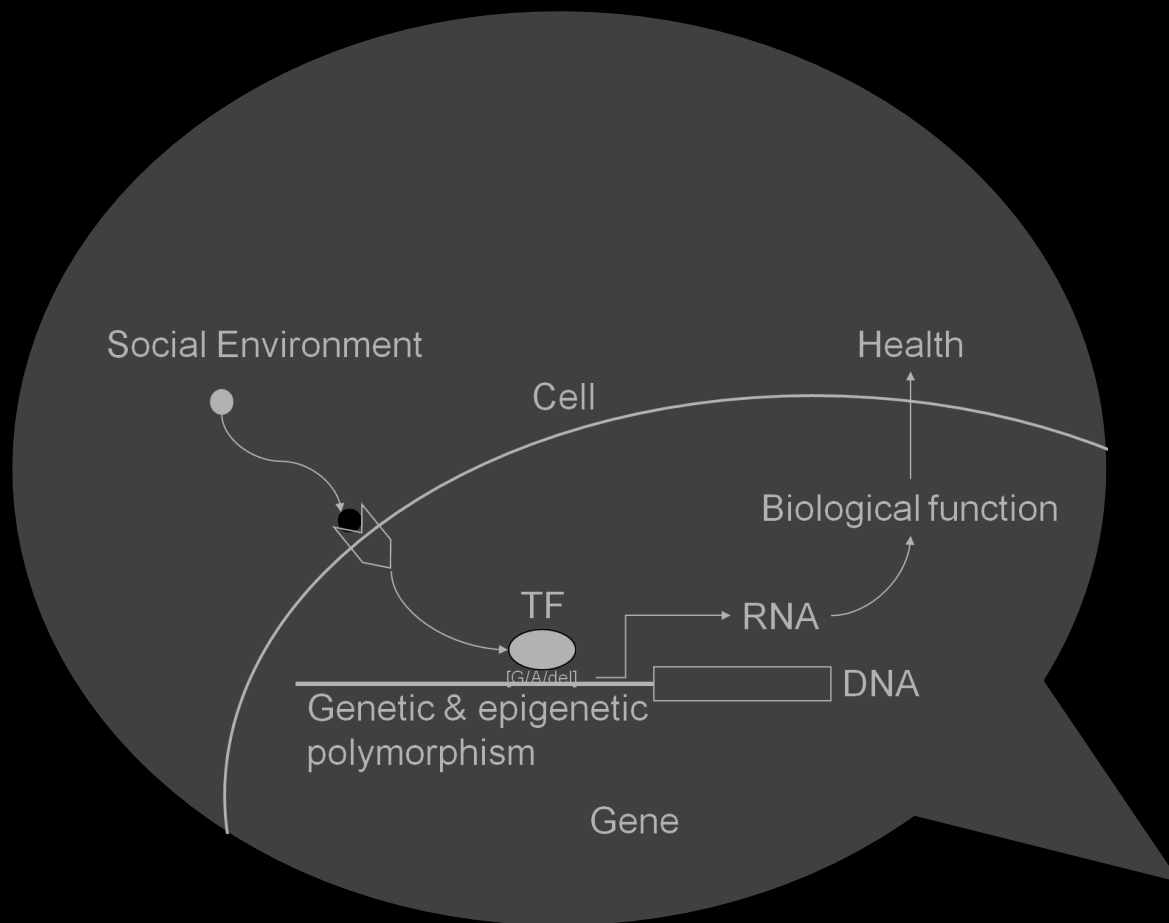
gene 1
gene 2
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gene 5
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gene 12
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gene 15
gene 16
gene 17
gene 18
gene 19
gene 20
gene 21
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gene 24
gene 25
gene 26
gene 27
gene 28
gene 29
gene 30
gene 31
gene 32
gene 33
gene 34
gene 35
gene 36
gene 37
gene 38
gene 39
gene 40

I've heard
of this one!

And this
one...

... and this
one too!

OK done.



gene 1
gene 2
gene 3
gene 4
gene 5
gene 6
gene 7
gene 8
gene 9
gene 10
gene 11
gene 12
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gene 36
gene 37
gene 38
gene 39
gene 40

Environment

Health

Causes

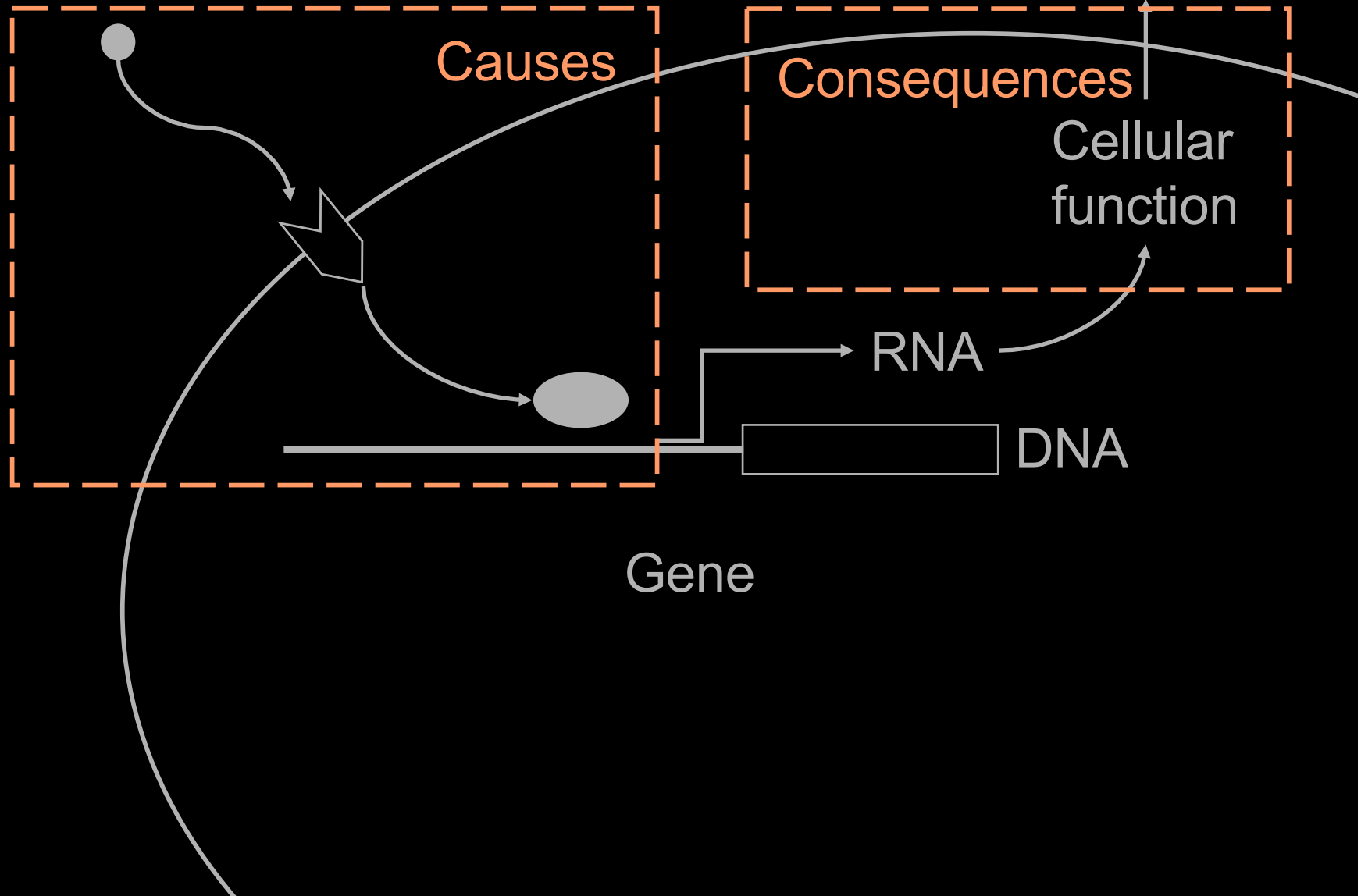
Consequences

Cellular
function

RNA

DNA

Gene



Interpreting results within an unbiased
“expert annotation” reference space

gene 1
gene 2
gene 3
gene 4
gene 5
gene 6
gene 7
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gene 40

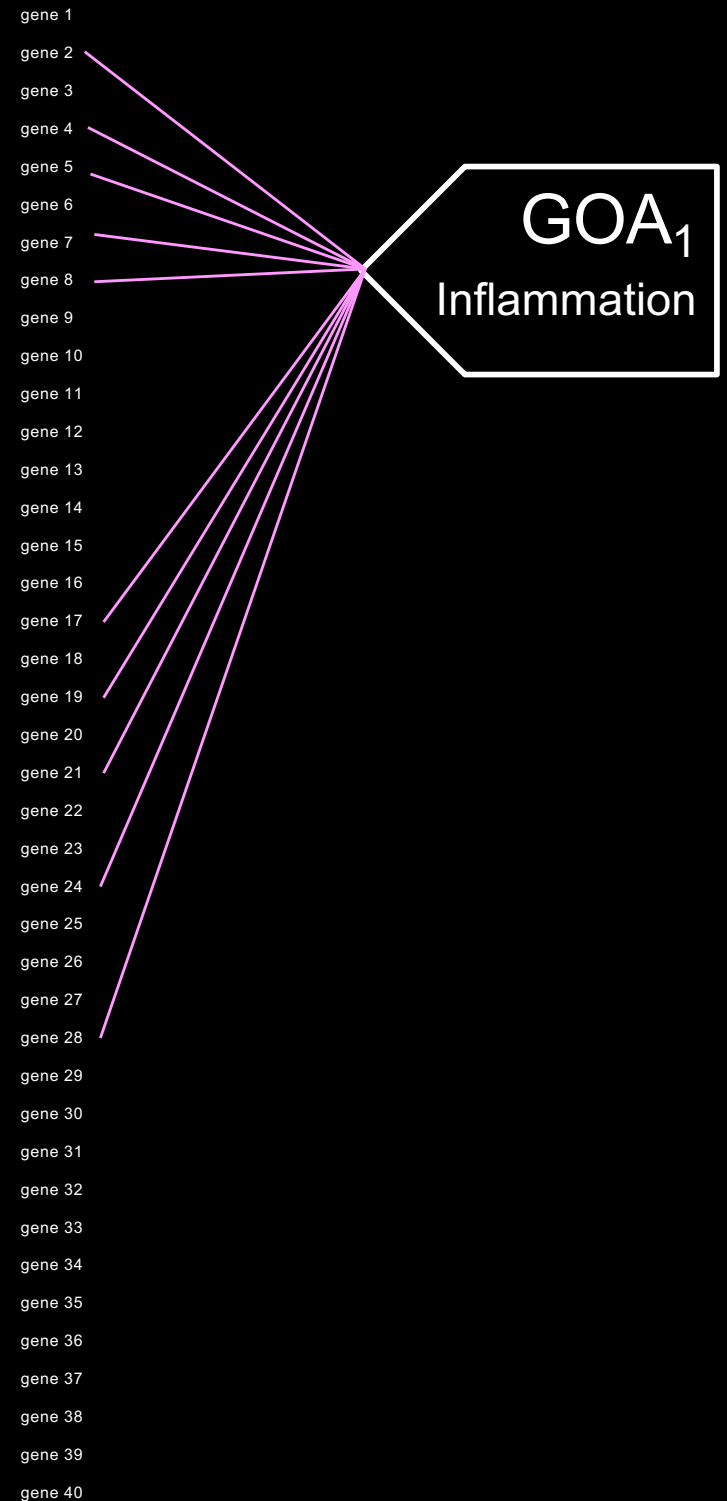


GOA₁
Inflammation

Interpreting results within an unbiased
“expert annotation” reference space

A statistical caveat: what’s the right
reference point for a null hypothesis?

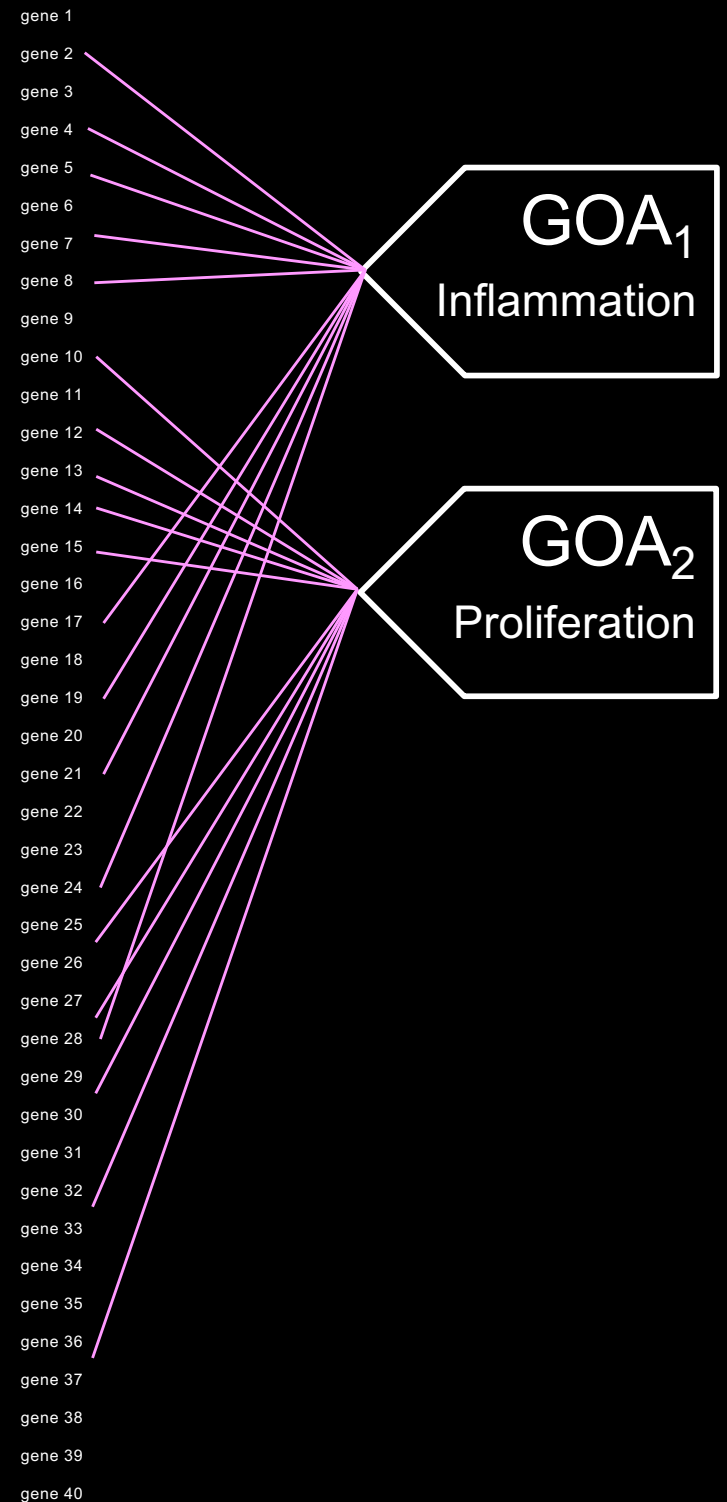
- Genome-wide baseline?
- Expressed transcriptome?
- Up- vs. down-regulated gene sets?



Interpreting results within an unbiased
“expert annotation” reference space

A statistical caveat: what’s the right
reference point for a null hypothesis?

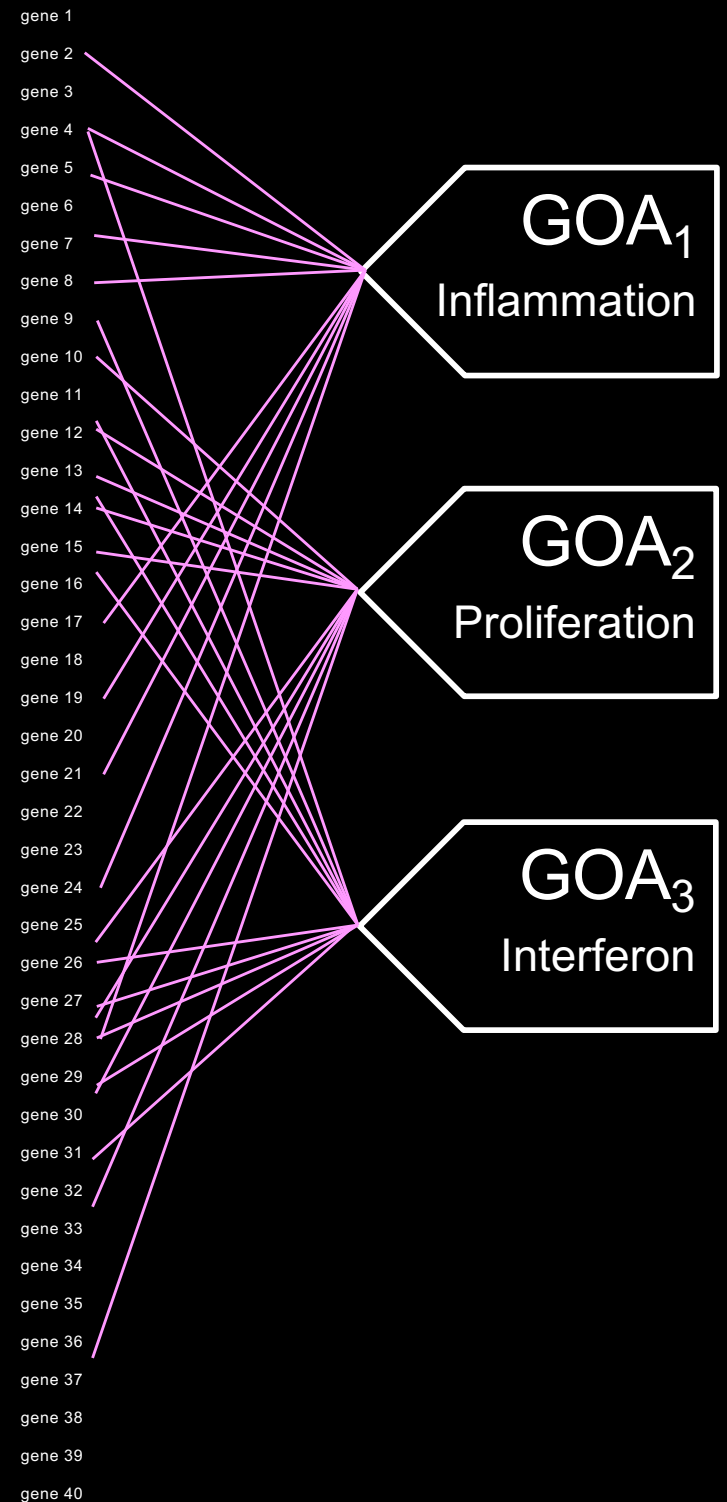
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Interpreting results within an unbiased
“expert annotation” reference space

A statistical caveat: what’s the right
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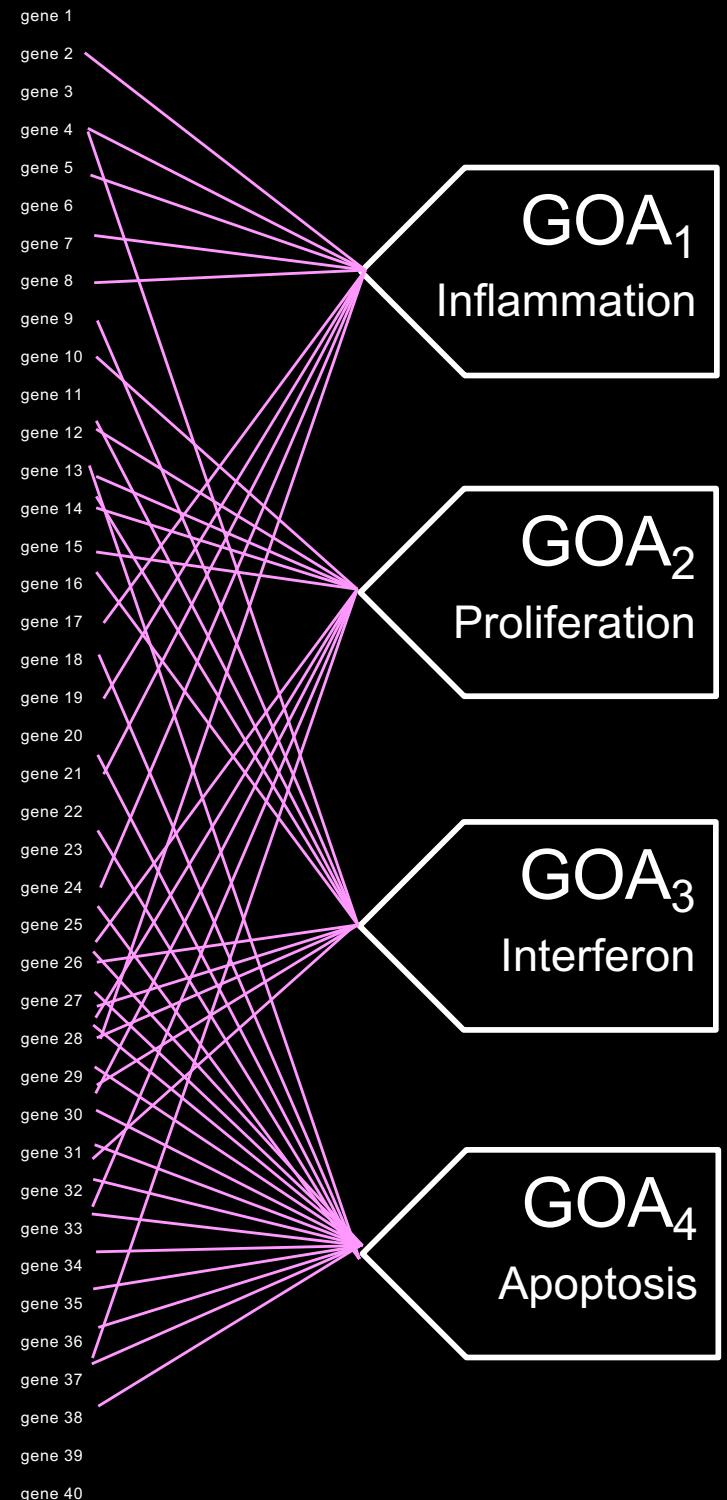
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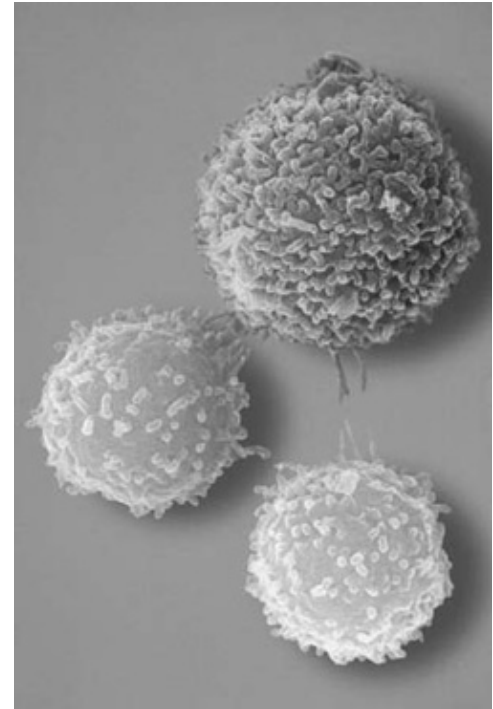
Interpreting results within an unbiased
“expert annotation” reference space

A statistical caveat: what’s the right
reference point for a null hypothesis?

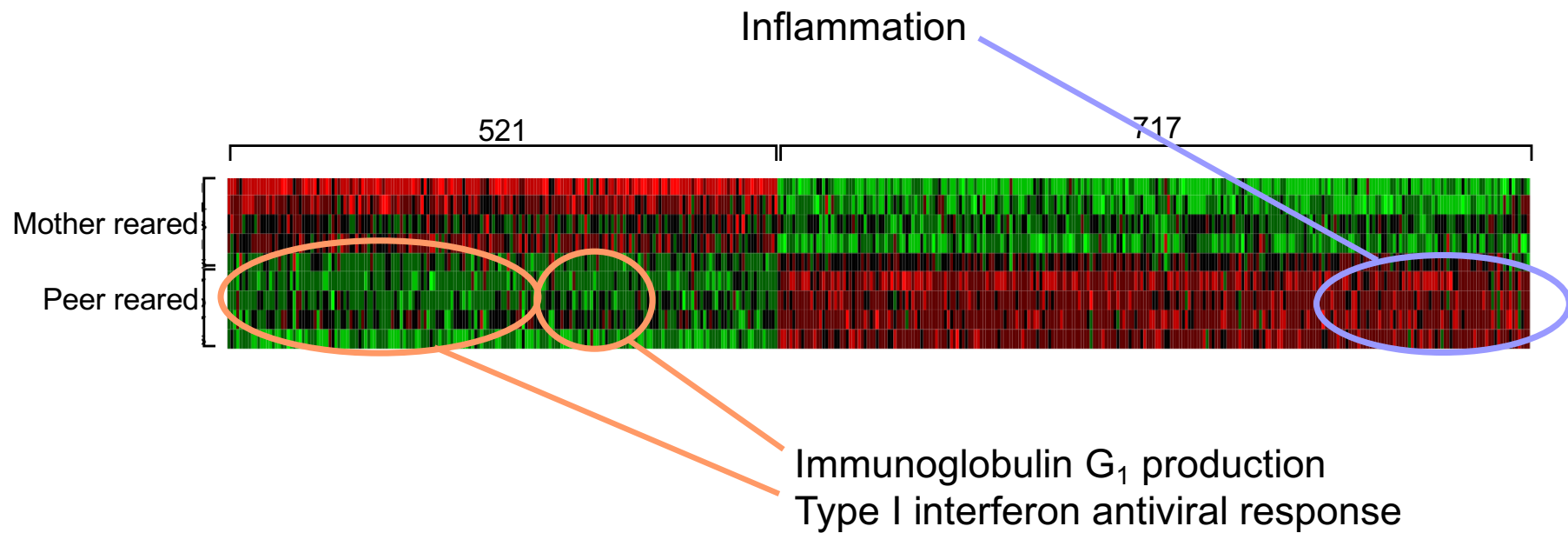
- Genome-wide baseline?
- Expressed transcriptome?
- Up- vs. down-regulated gene sets?



Social instability



CTRA – conserved transcriptional response to adversity



Low SES

Social loss / bereavement

Post-traumatic stress

Cancer diagnosis

Social threat

Loneliness

Social instability

Chronic stress

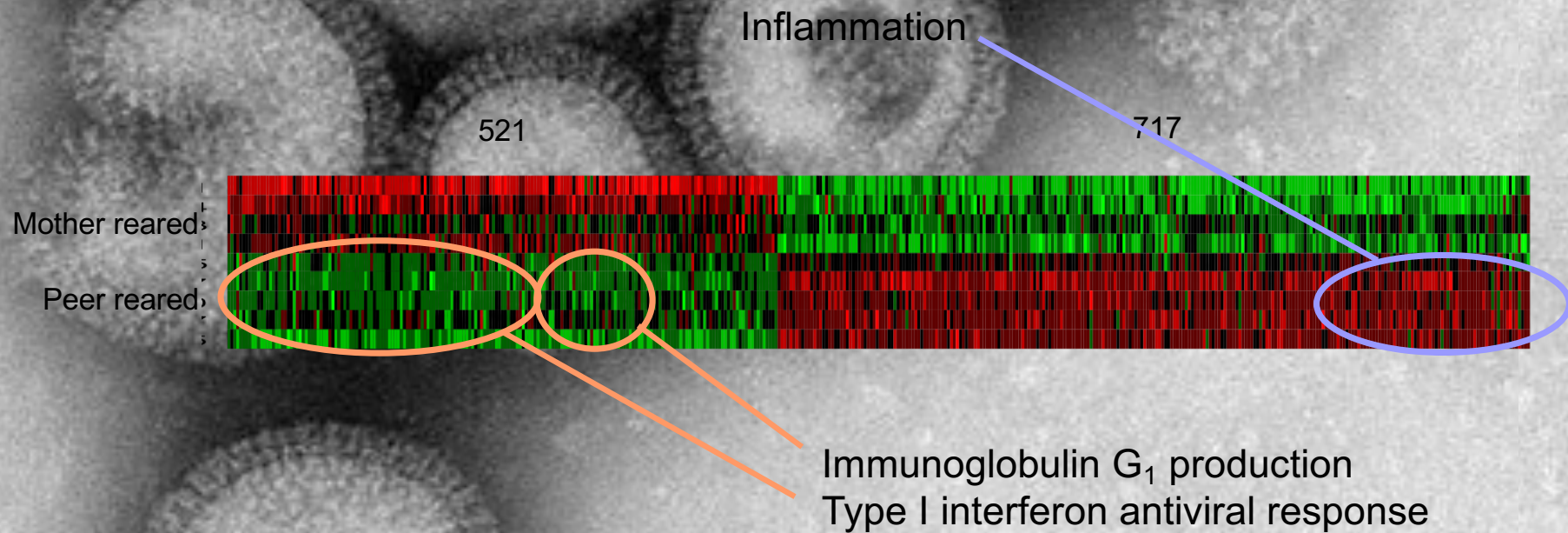
Low social rank

Caregiving for seriously ill

Anxiety

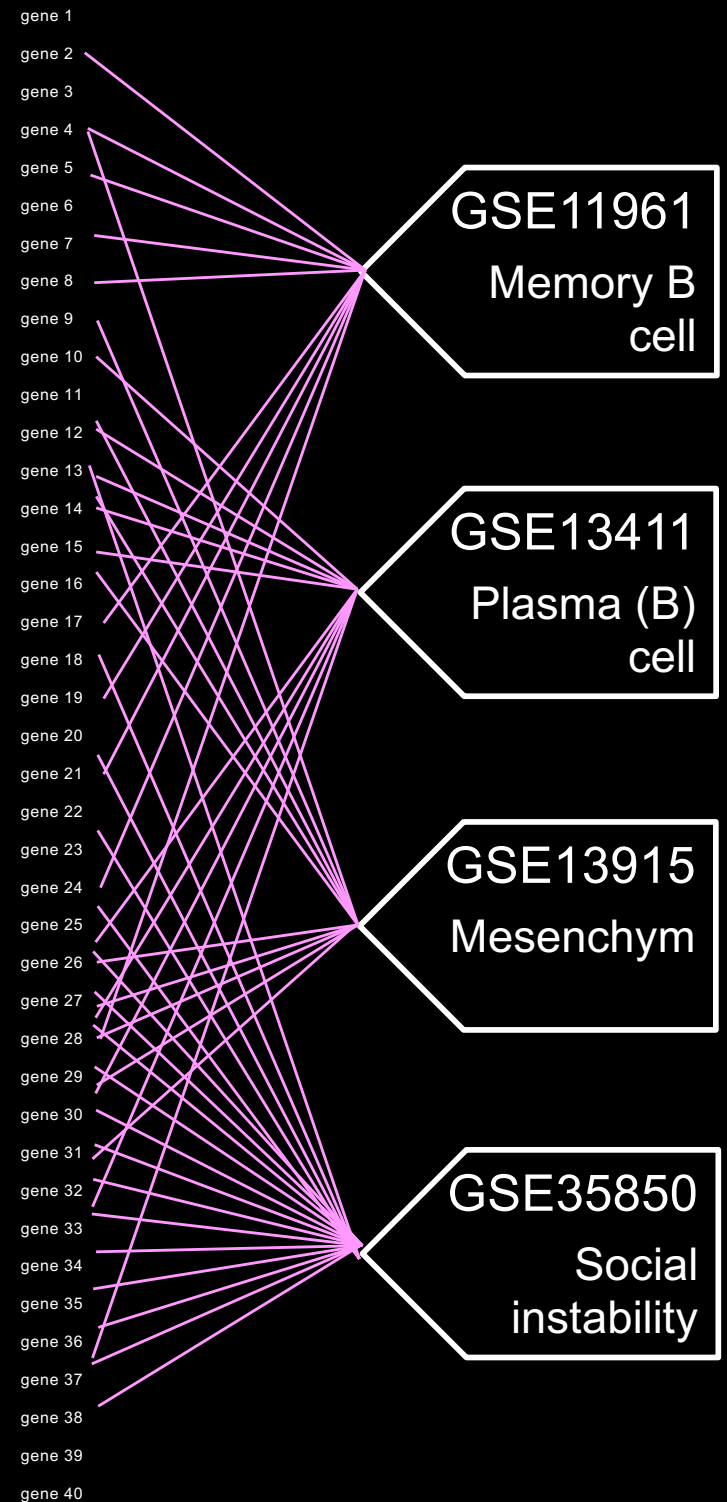
Early life adversity

CTRA – conserved transcriptional response to adversity



Interpreting results within an empirical transcriptome reference space (vs. “expert” GO tags)

1. GEO search
(or ArrayExpress, GSEA, etc.)
2. Grab the data & compute diagnosticity scores for their experimental “poles”
3. Test for gene set differential expression in your data (either them|you or you|them)



Environment

Health

Causes

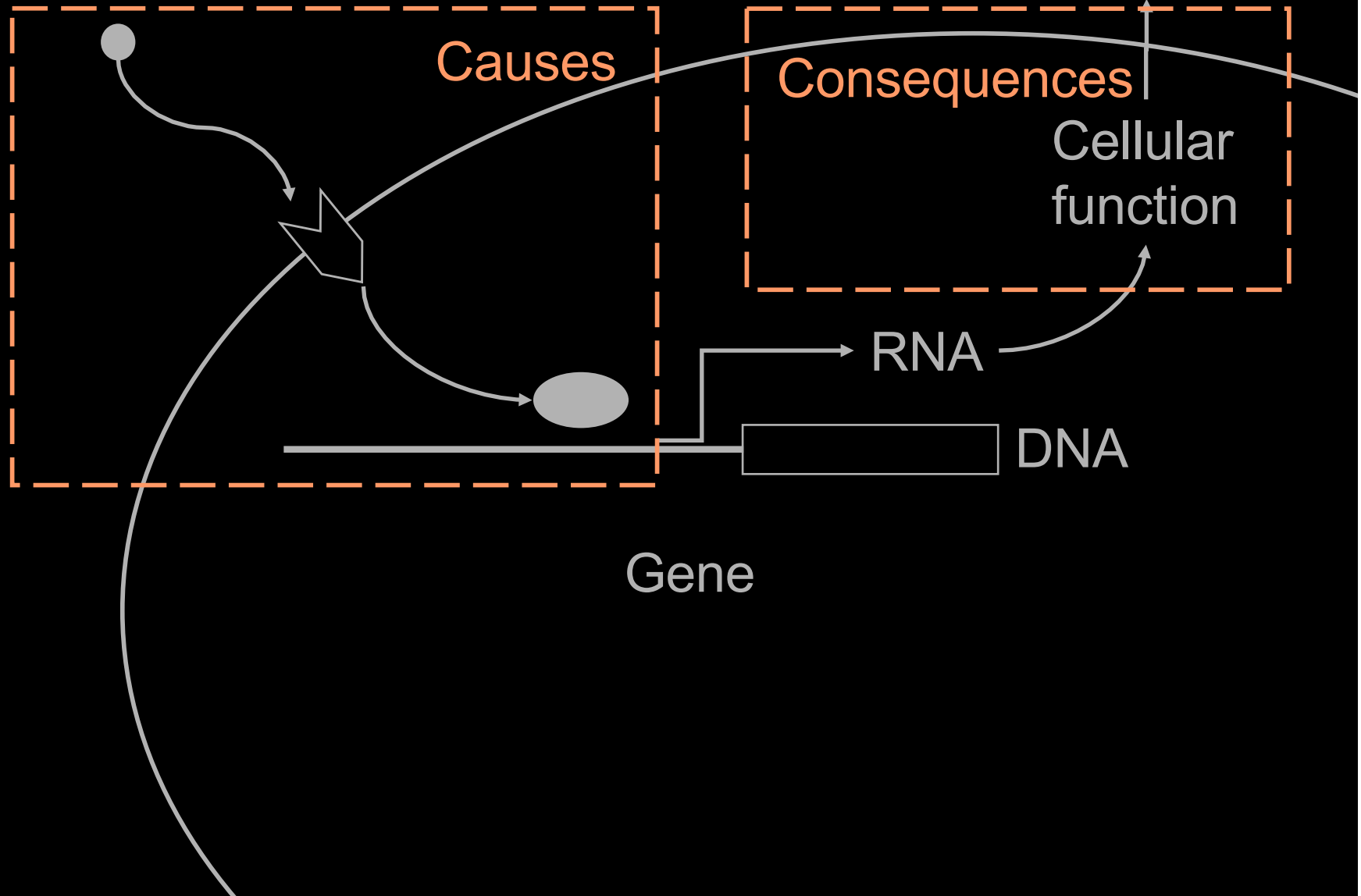
Consequences

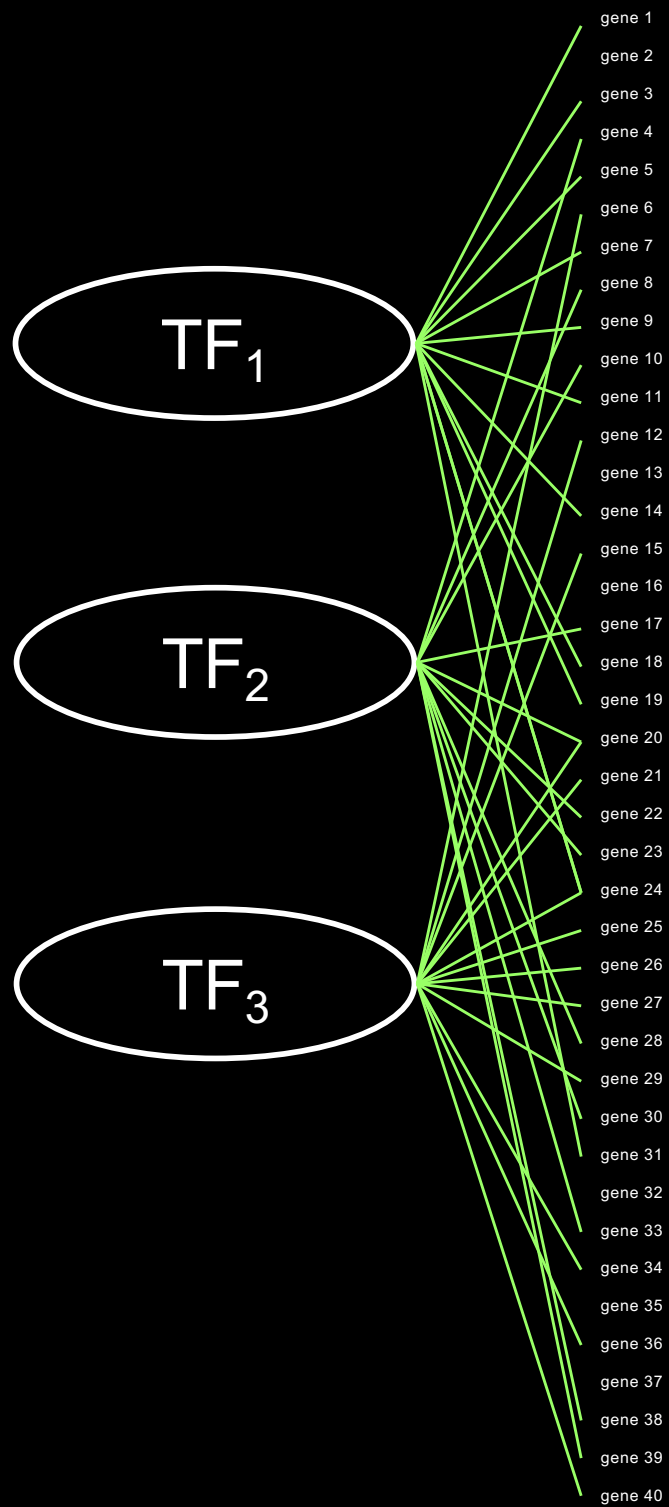
Cellular
function

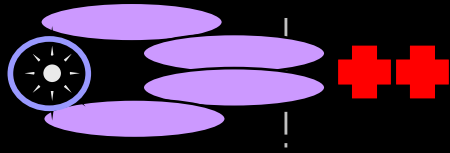
RNA

DNA

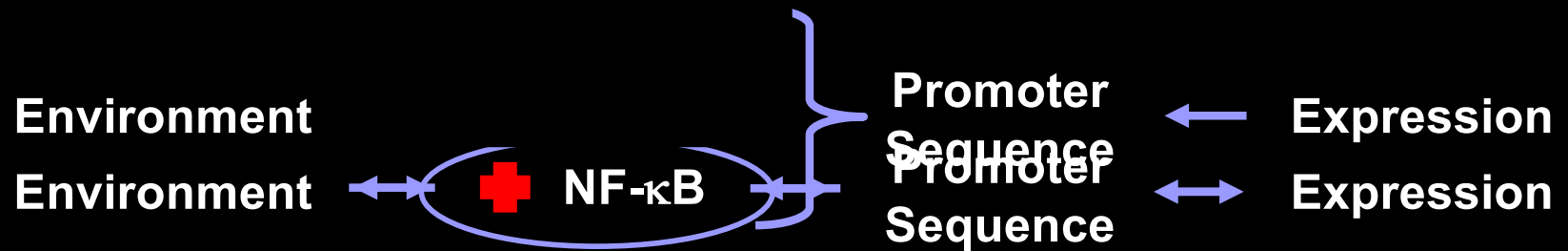
Gene



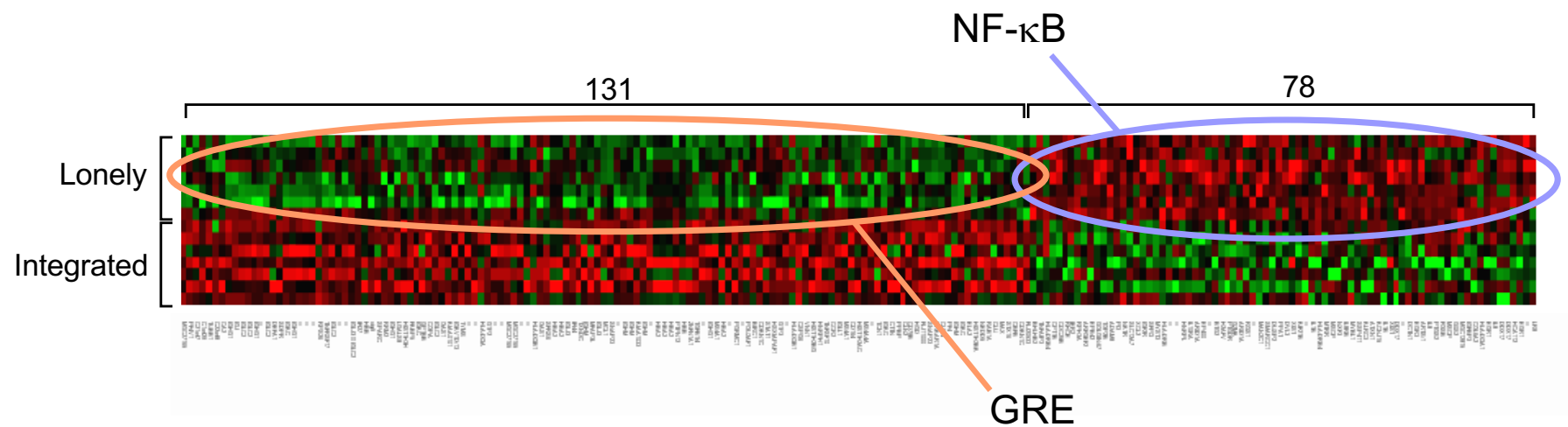




?



CTRA – Conserved Transcriptional Response to Adversity



Environment

Health

Causes

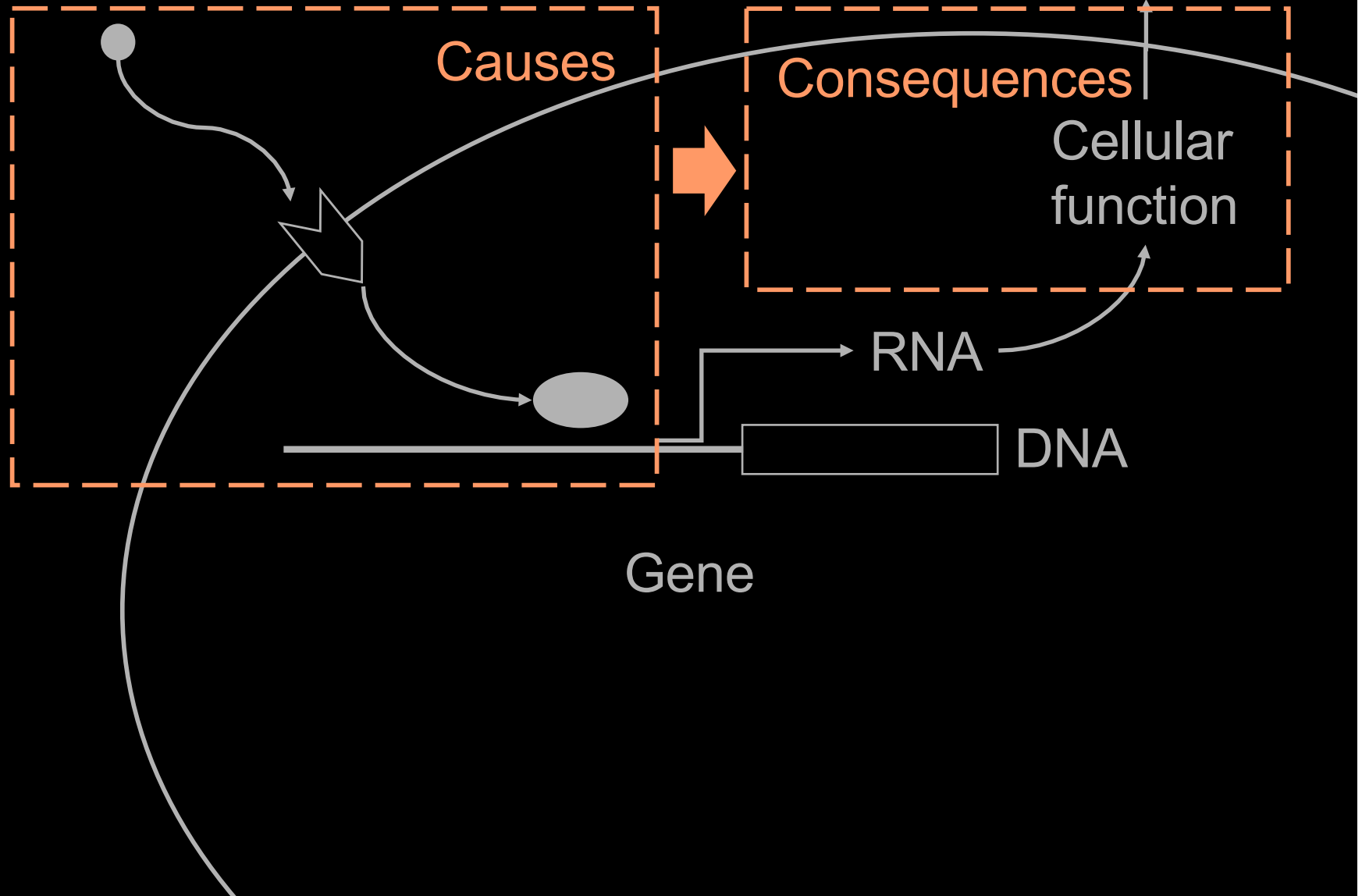
Consequences

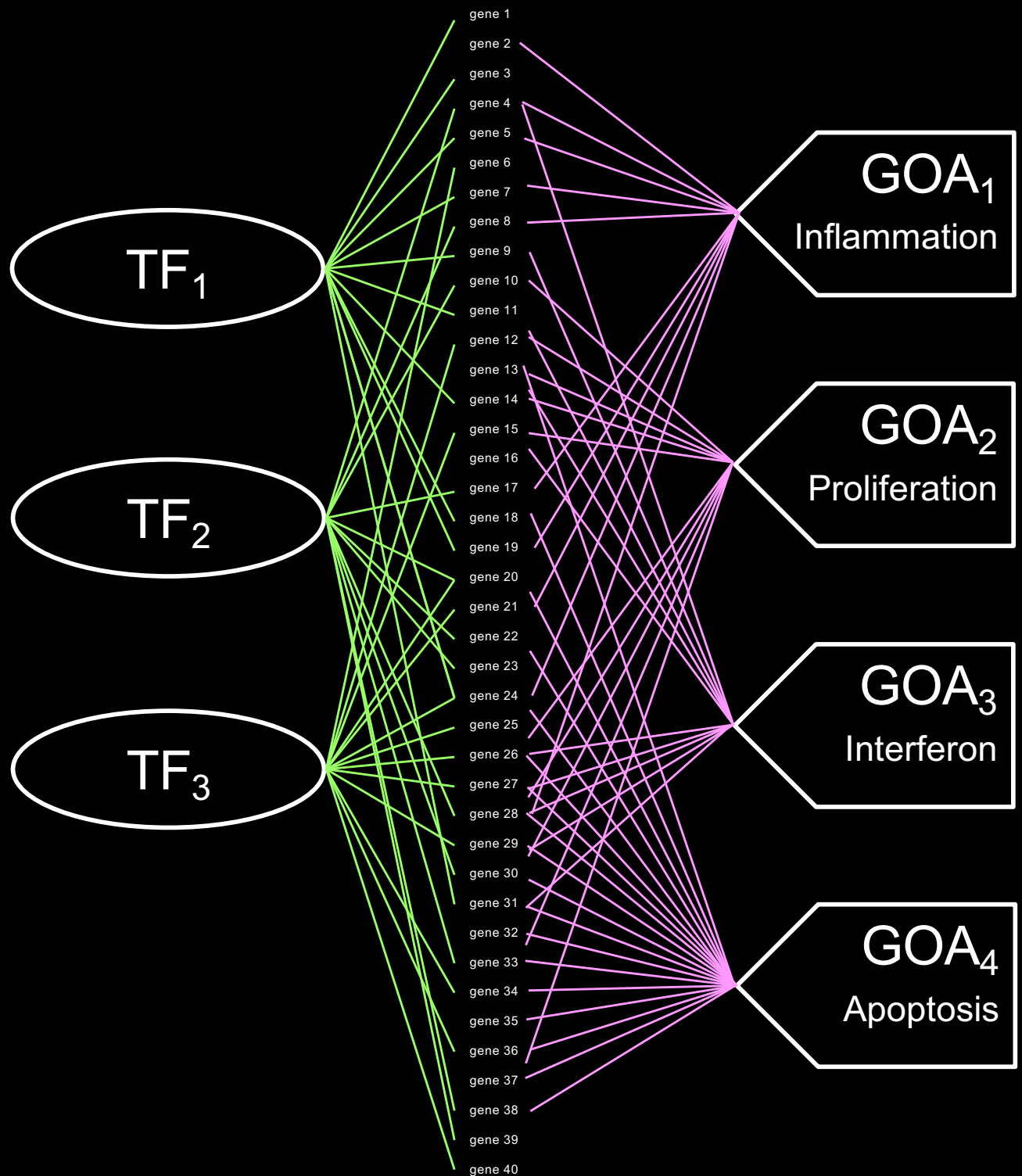
Cellular
function

RNA

DNA

Gene





Environment

Health

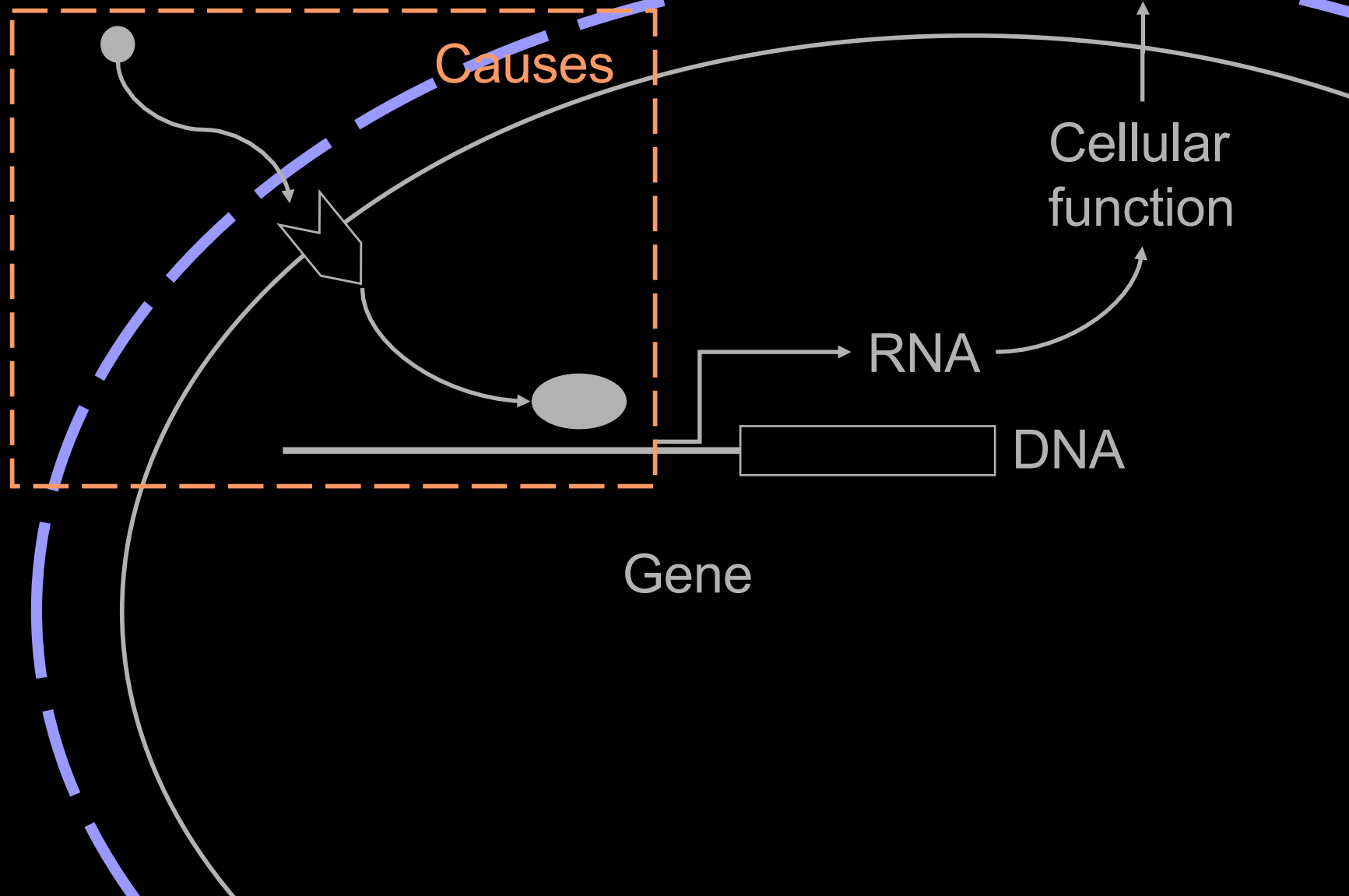
Causes

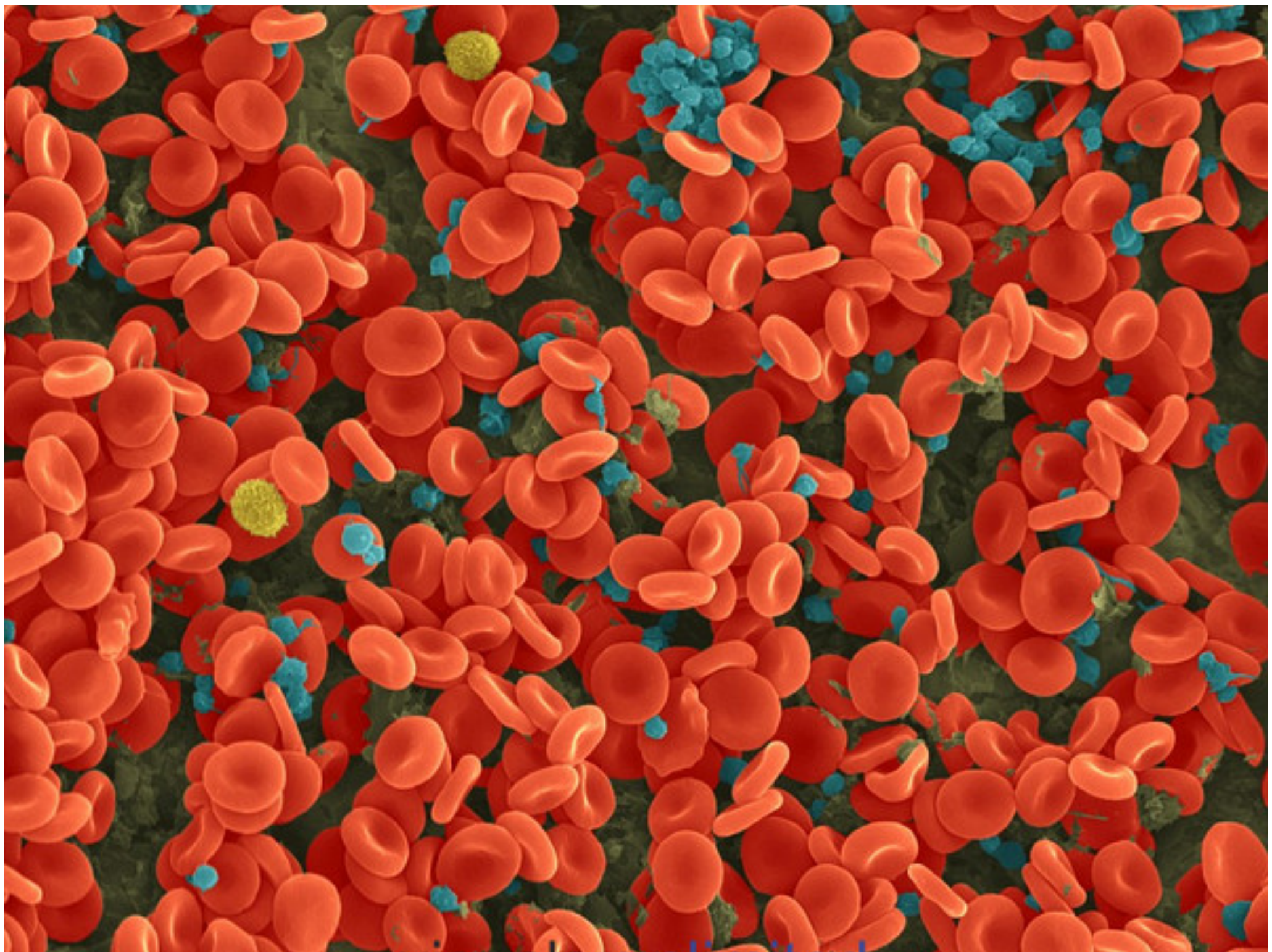
Cellular
function

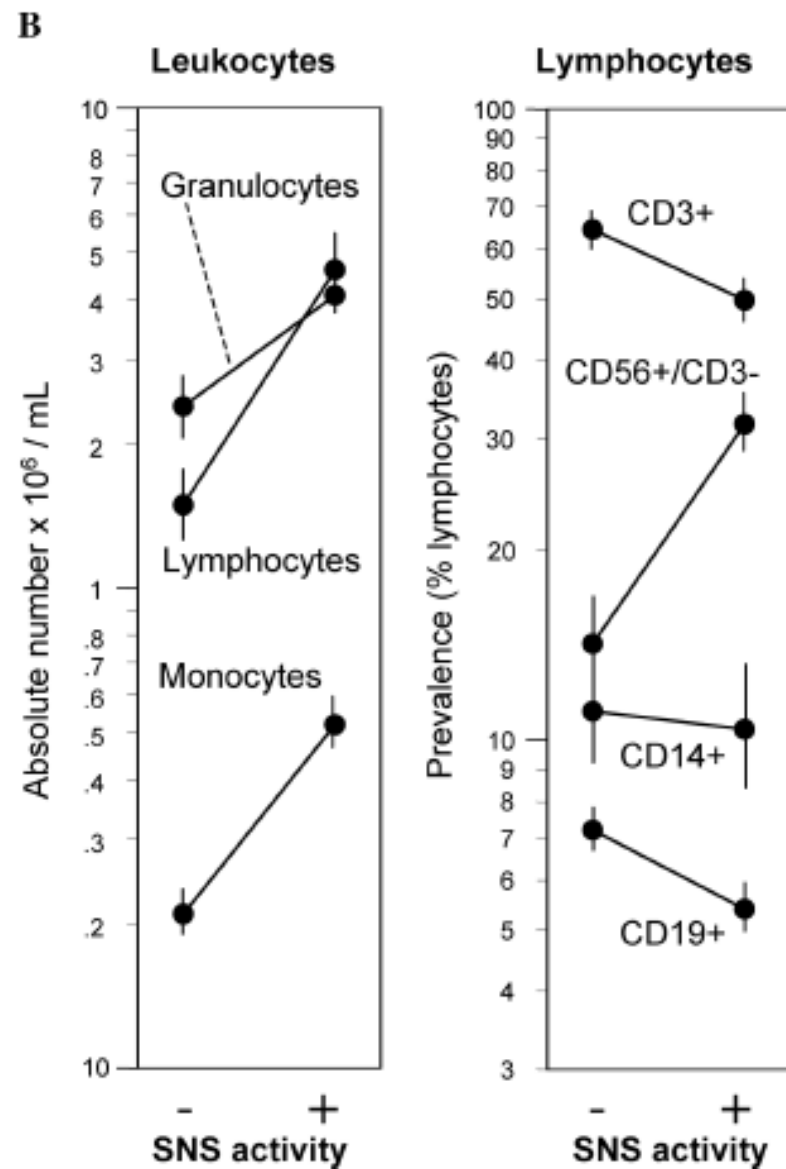
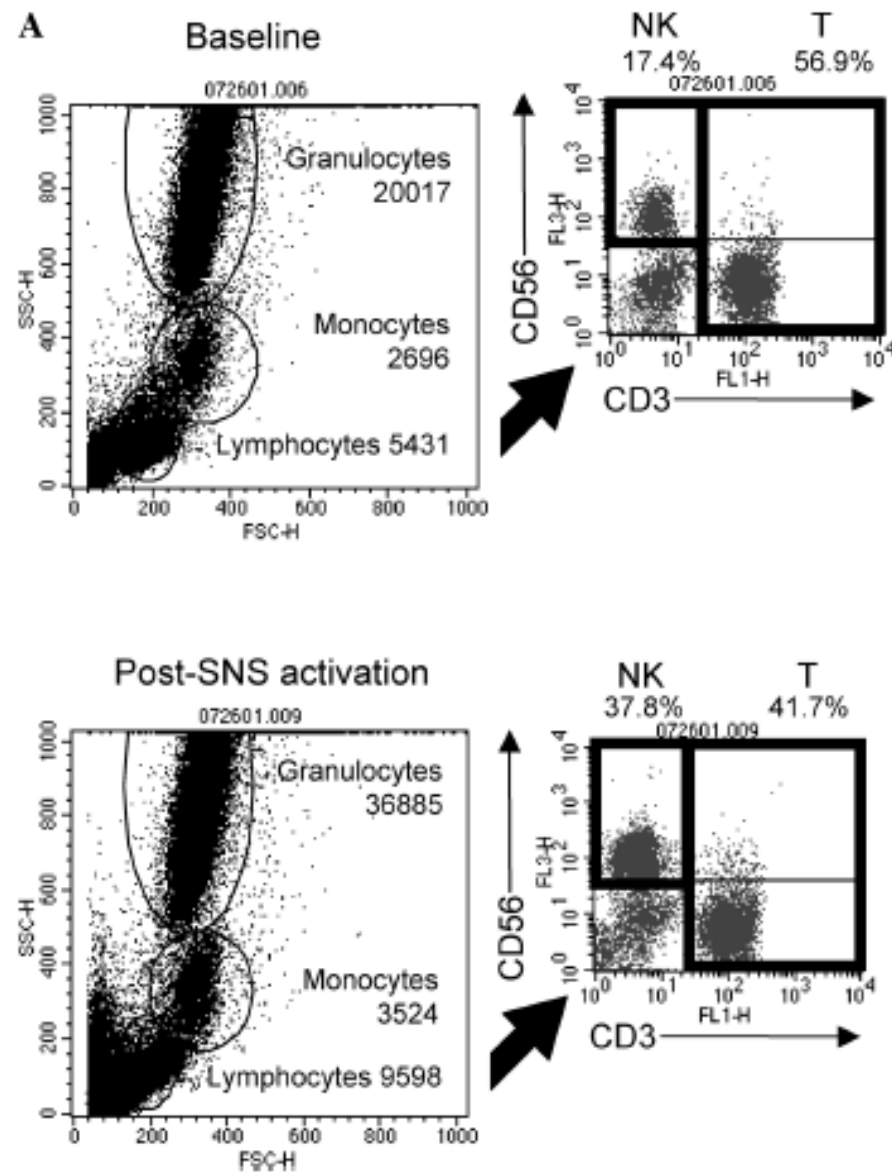
RNA

DNA

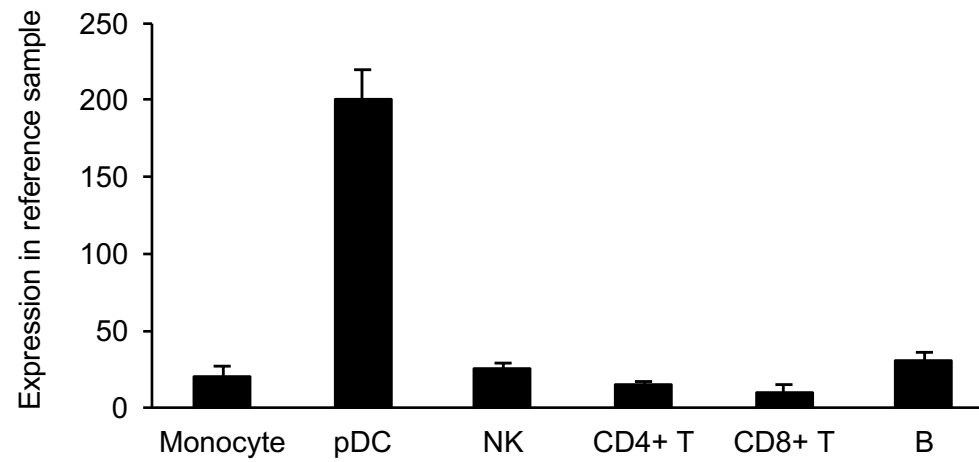
Gene



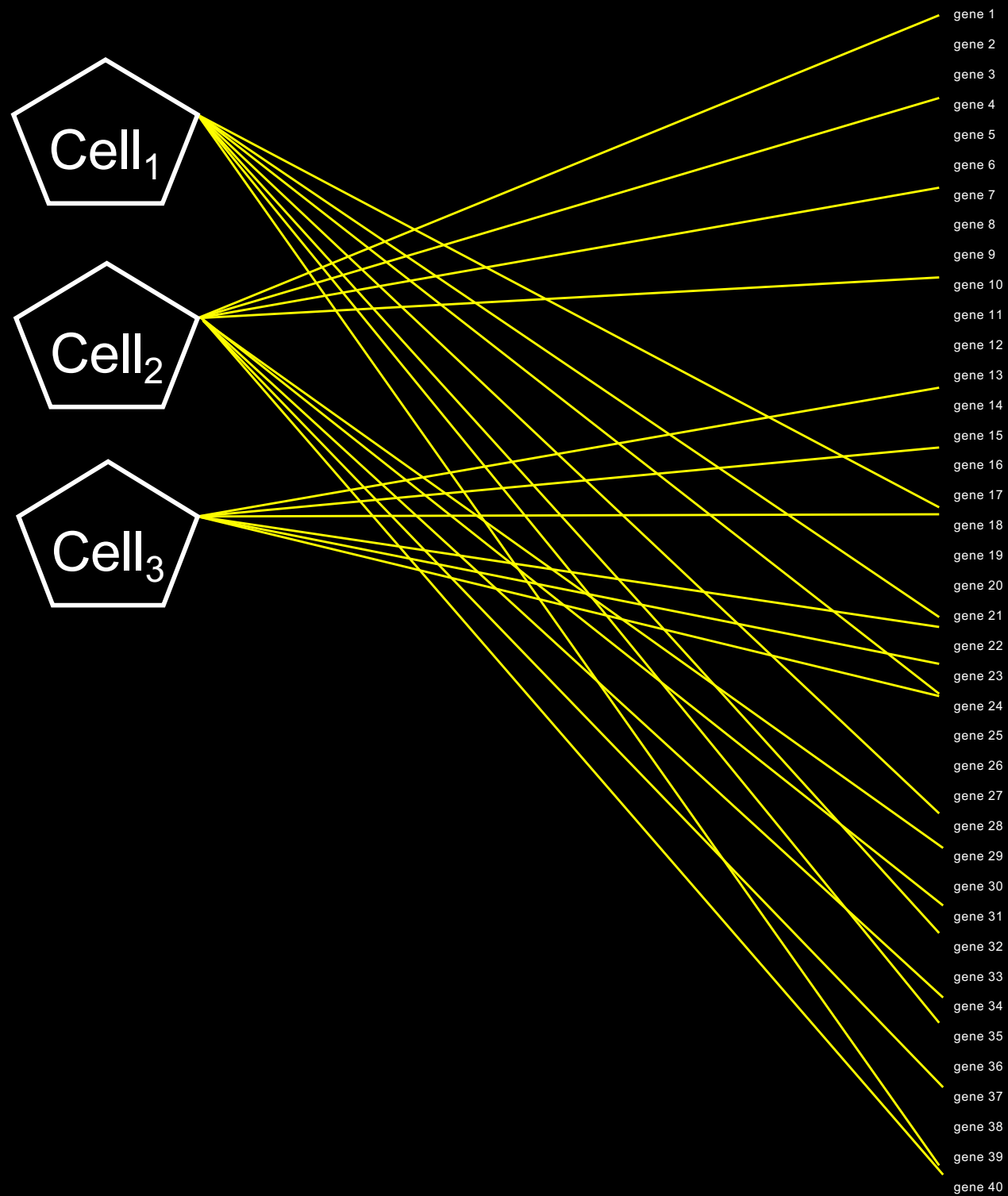




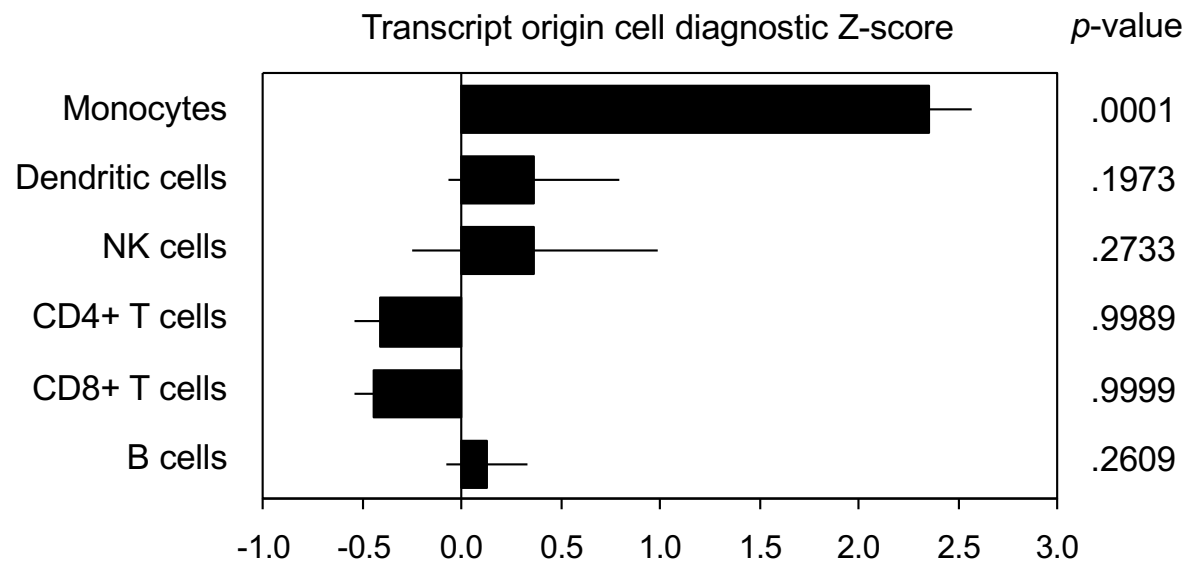
Transcript origin analysis

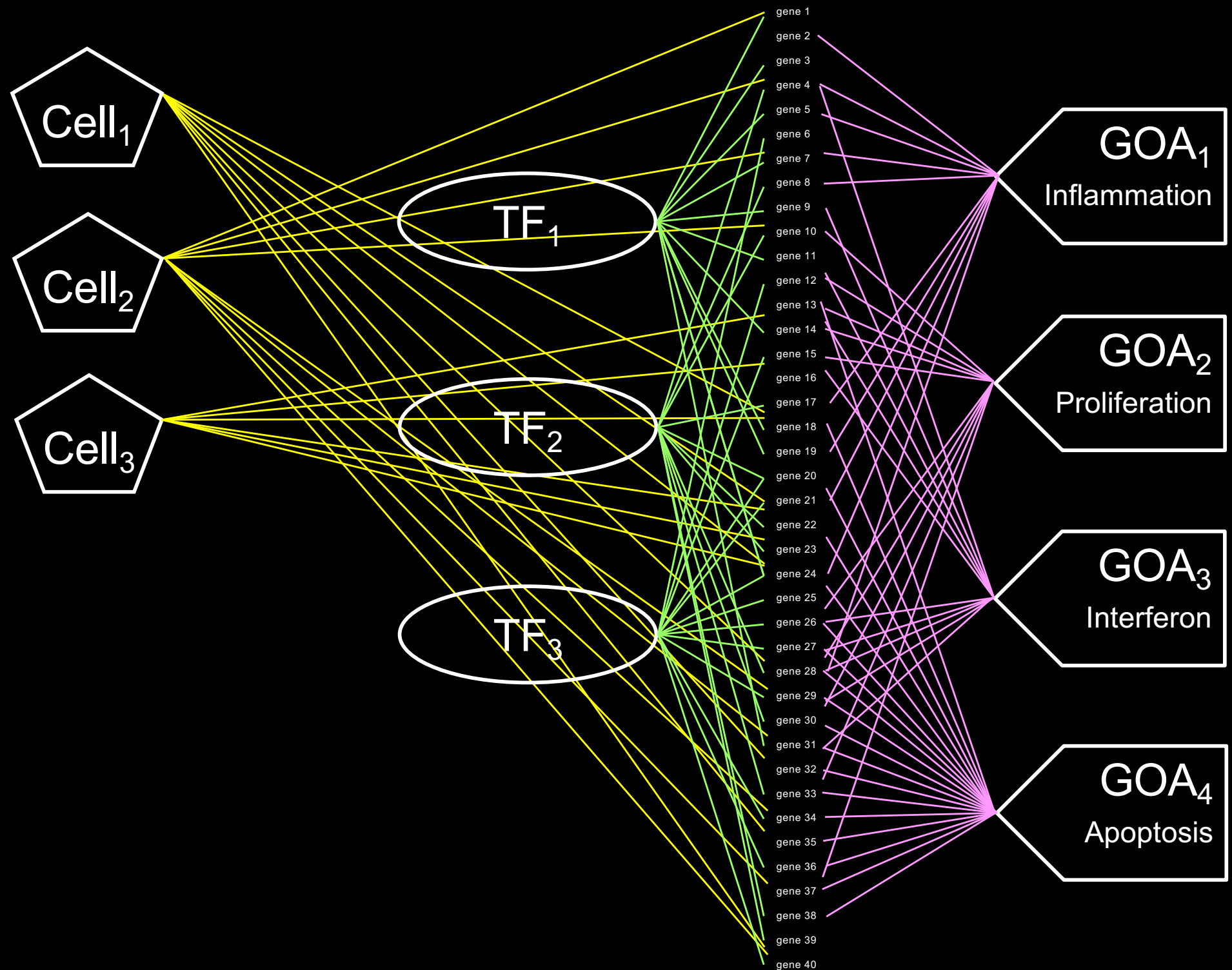


TOA Z-score_{gc} = -0.4 4.9 -0.3 -0.5 -0.6 -0.3



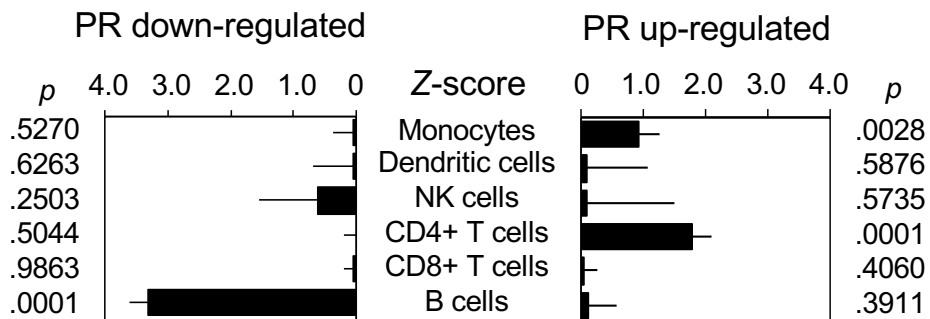
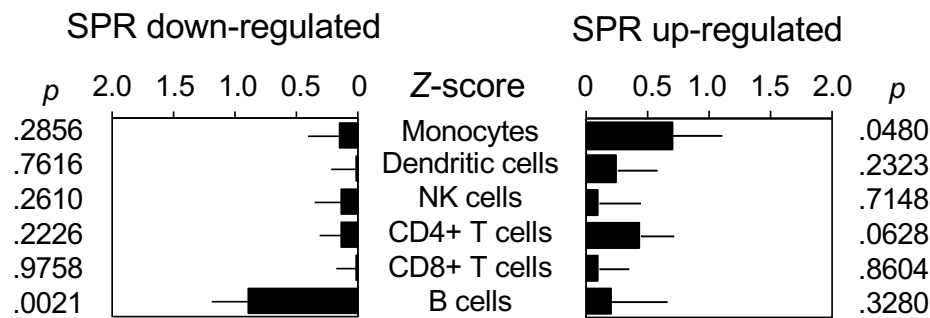
Cellular target of social adversity



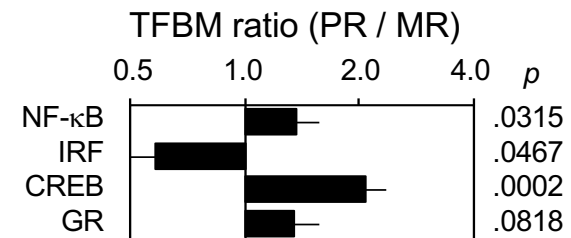
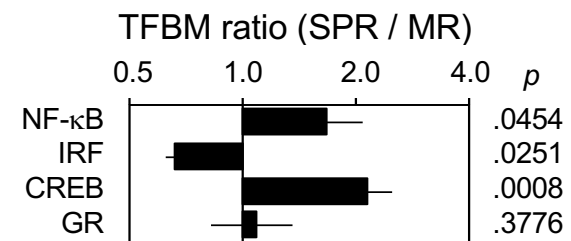


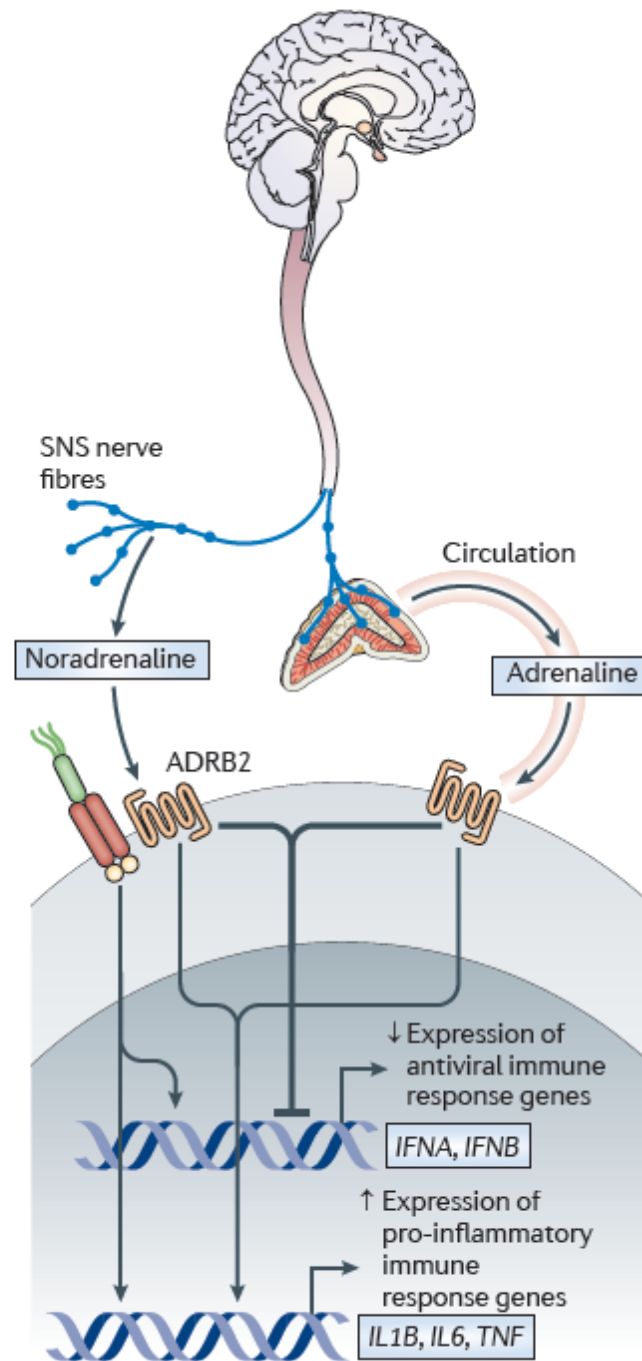
Social instability

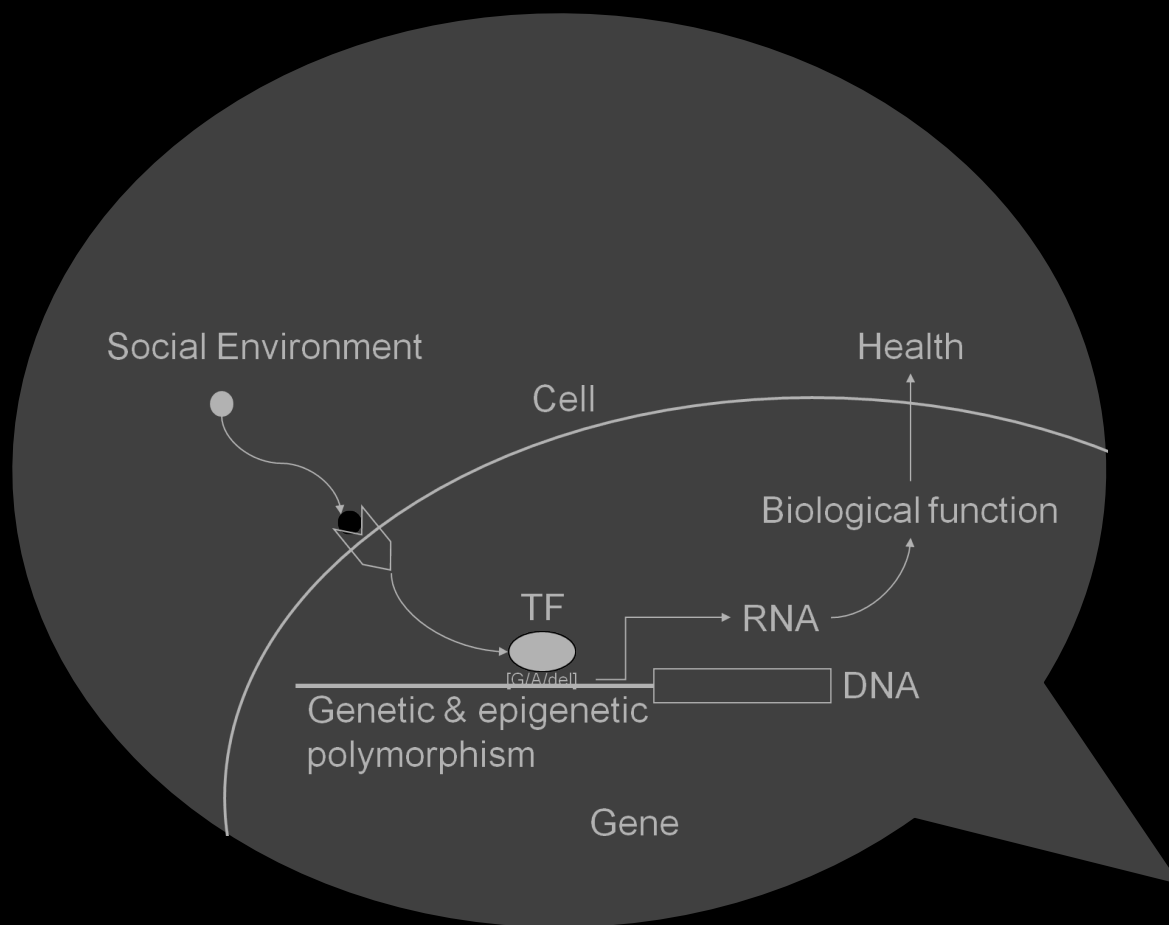
Cellular origin



Transcription factor







- gene 1
- gene 2
- gene 3
- gene 4
- gene 5
- gene 6
- gene 7
- gene 8
- gene 9
- gene 10
- gene 11
- gene 12
- gene 13
- gene 14
- gene 15
- gene 16
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- gene 39
- gene 40

Social Environment

Health

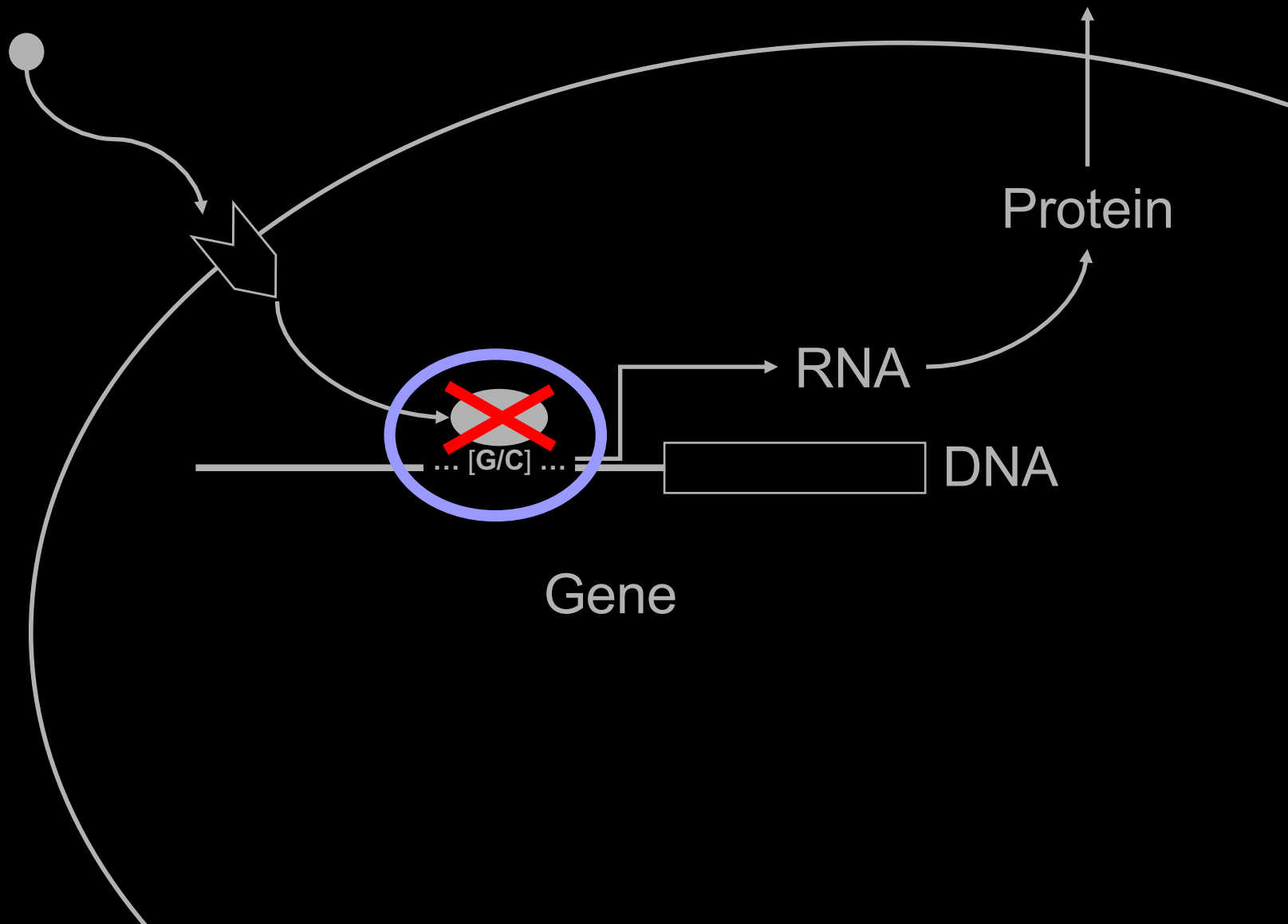
Protein

RNA

DNA

Gene

... [G/C] ...

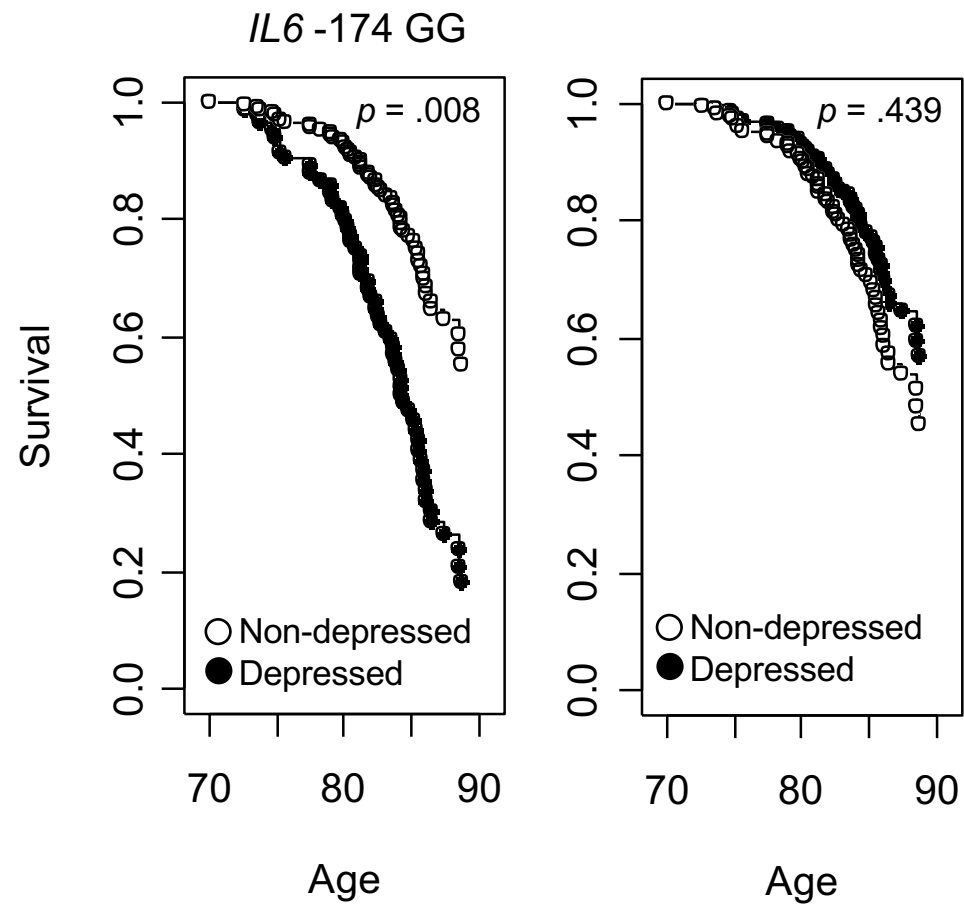


Gene x Environment Interaction

In silico



Gene x Environment Interaction



Adversity (depression)

Social Environment

● **SNS / NE**

β -AR

GATA1



...GATG...

rs1800795

Inflammation

RNA

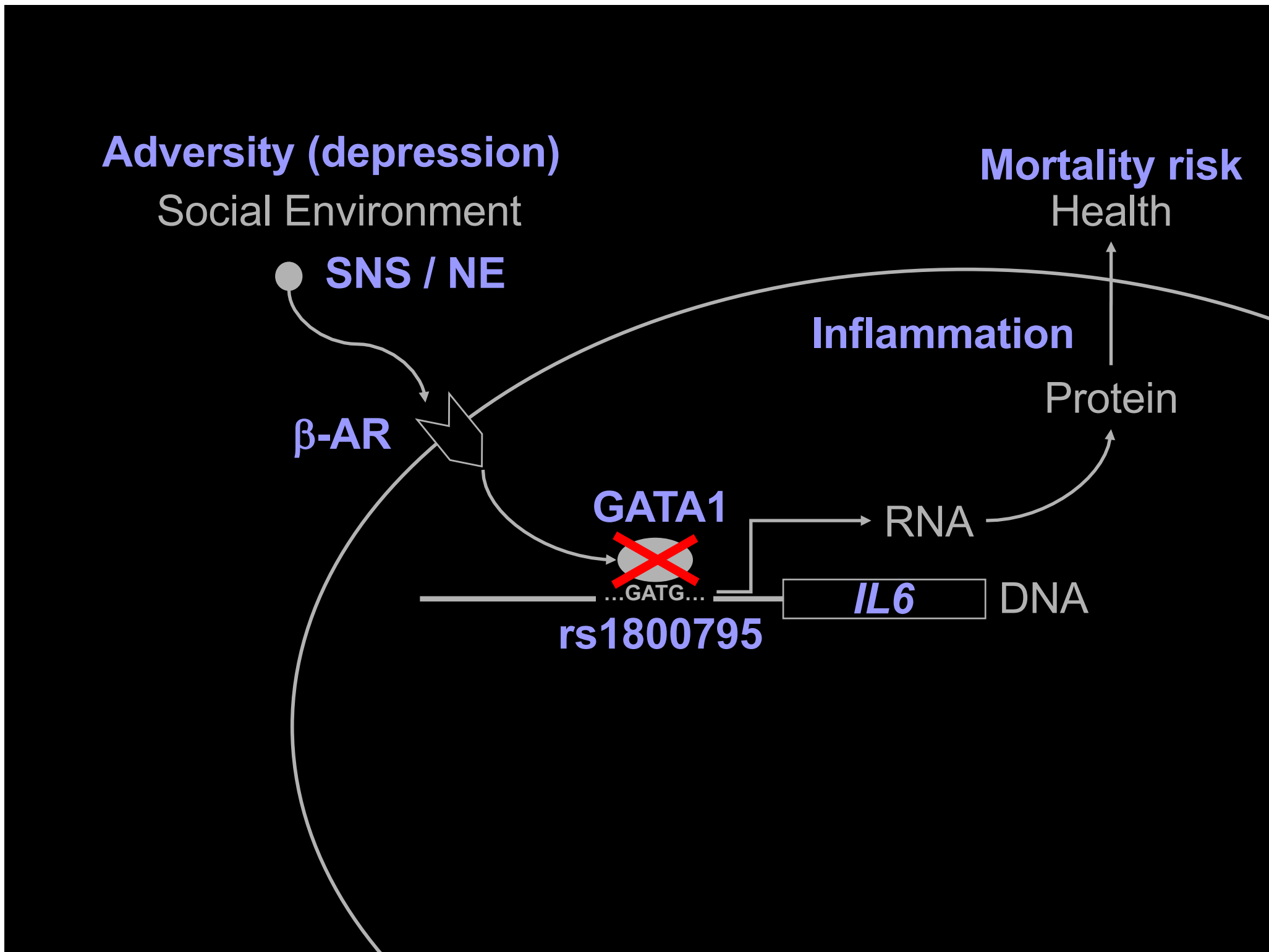
IL6

DNA

Protein

Mortality risk

Health



[illegible]

Environment

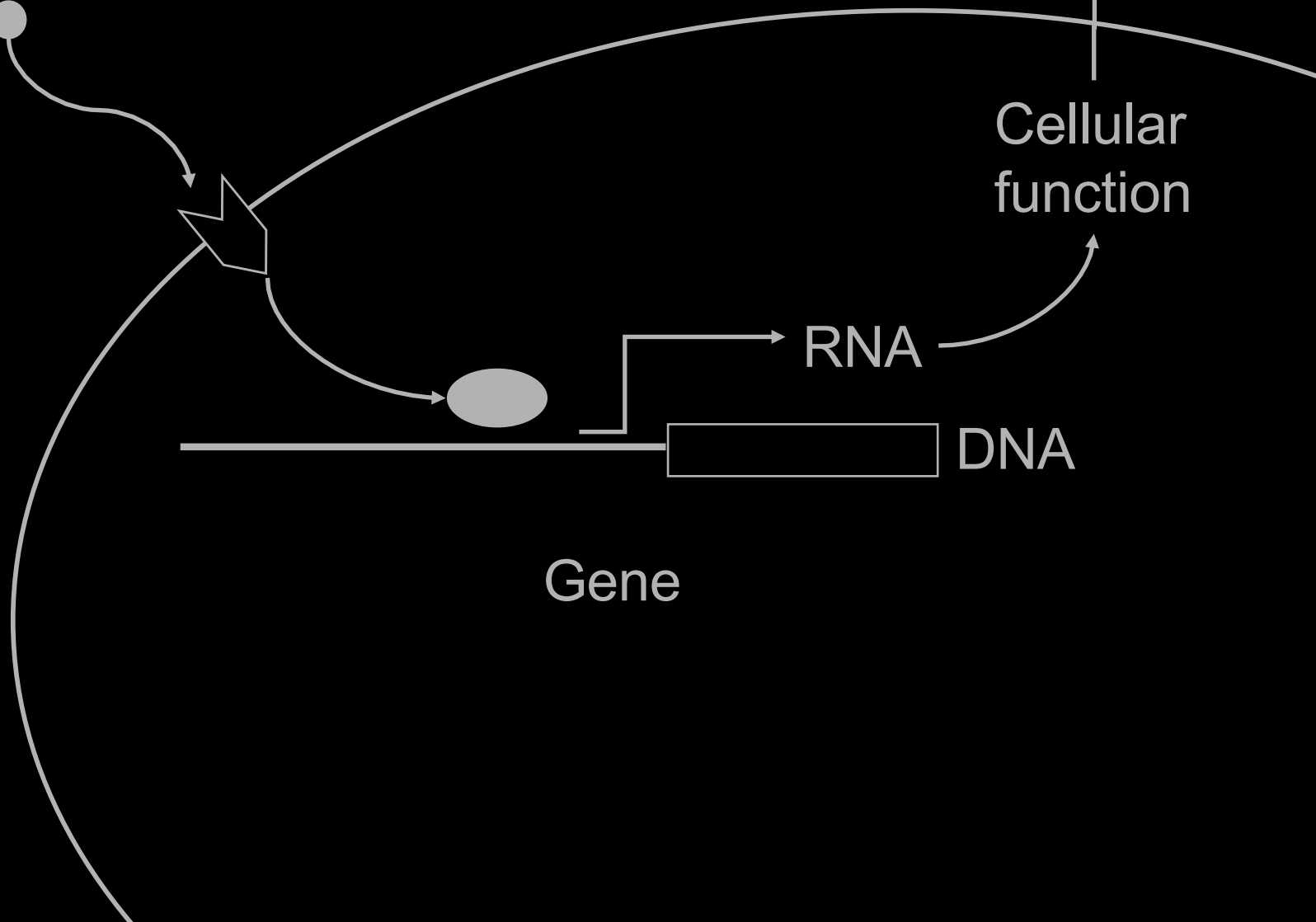
Health

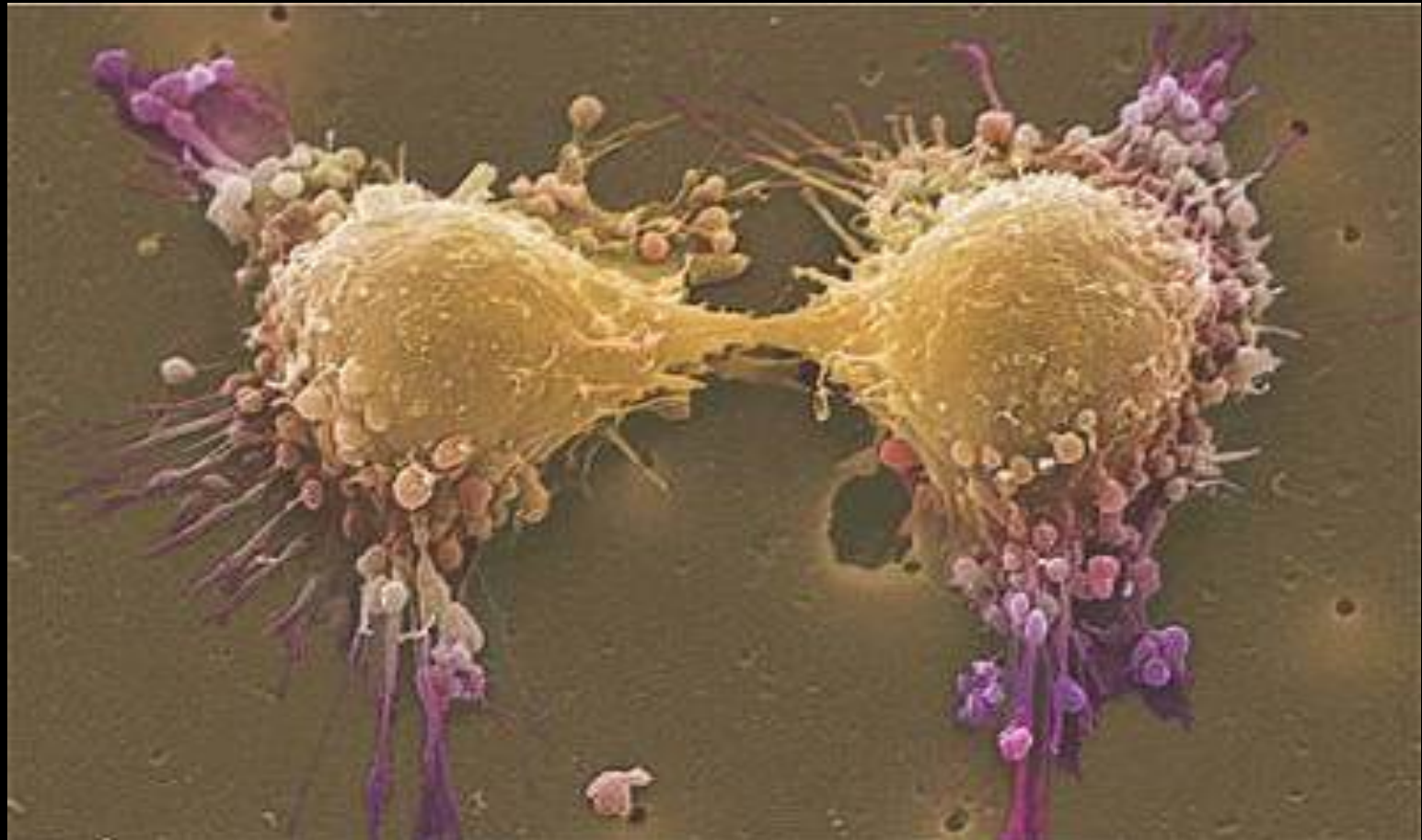
Cellular
function

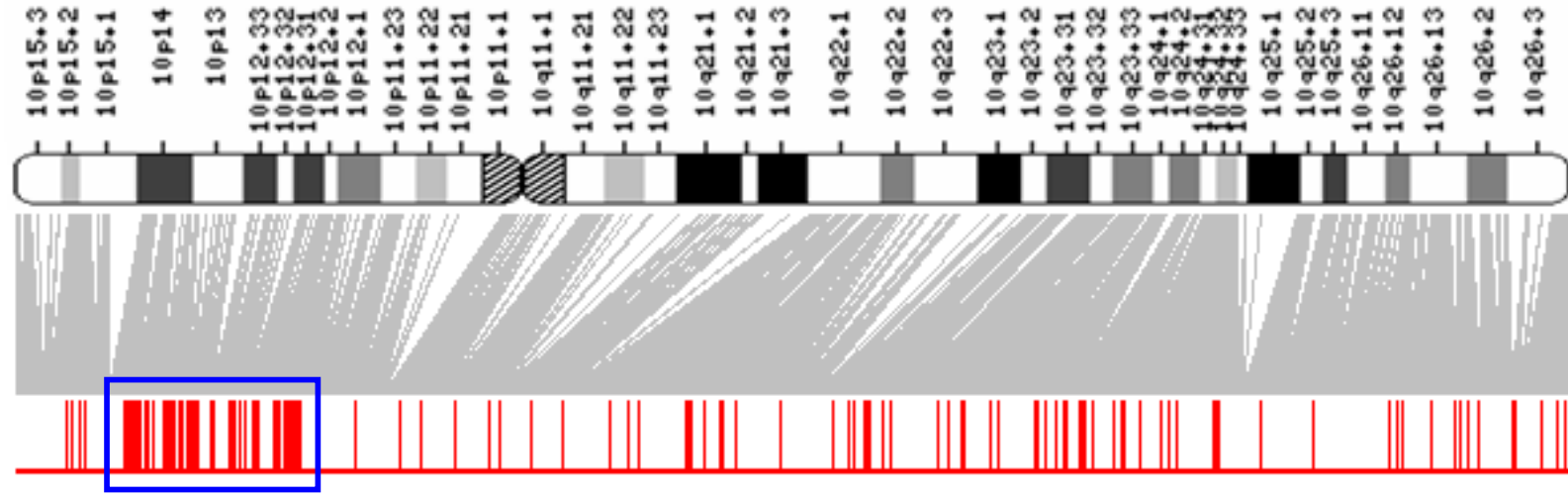
RNA

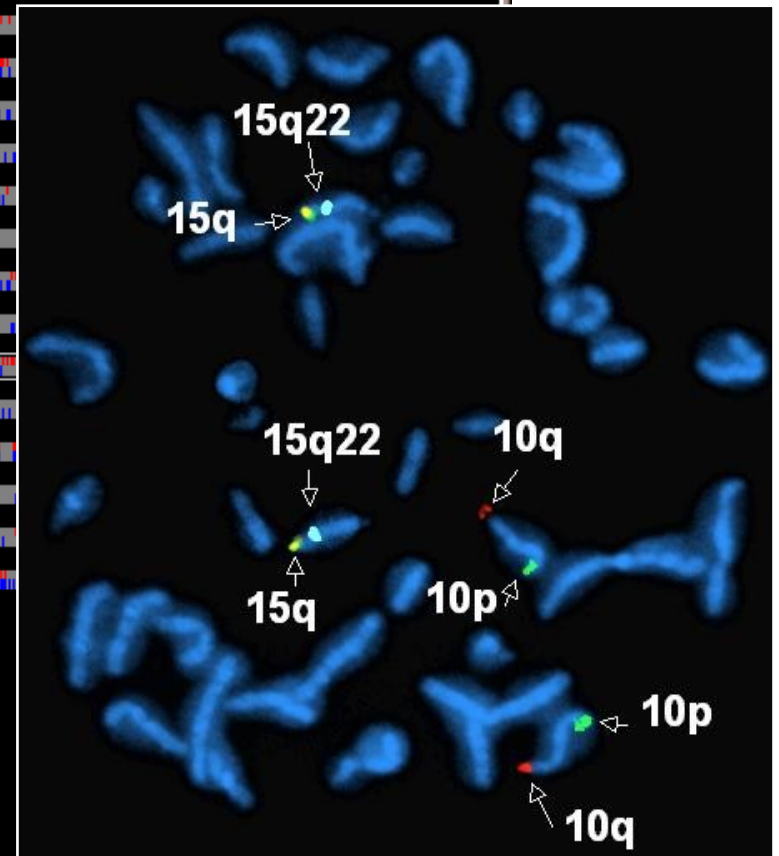
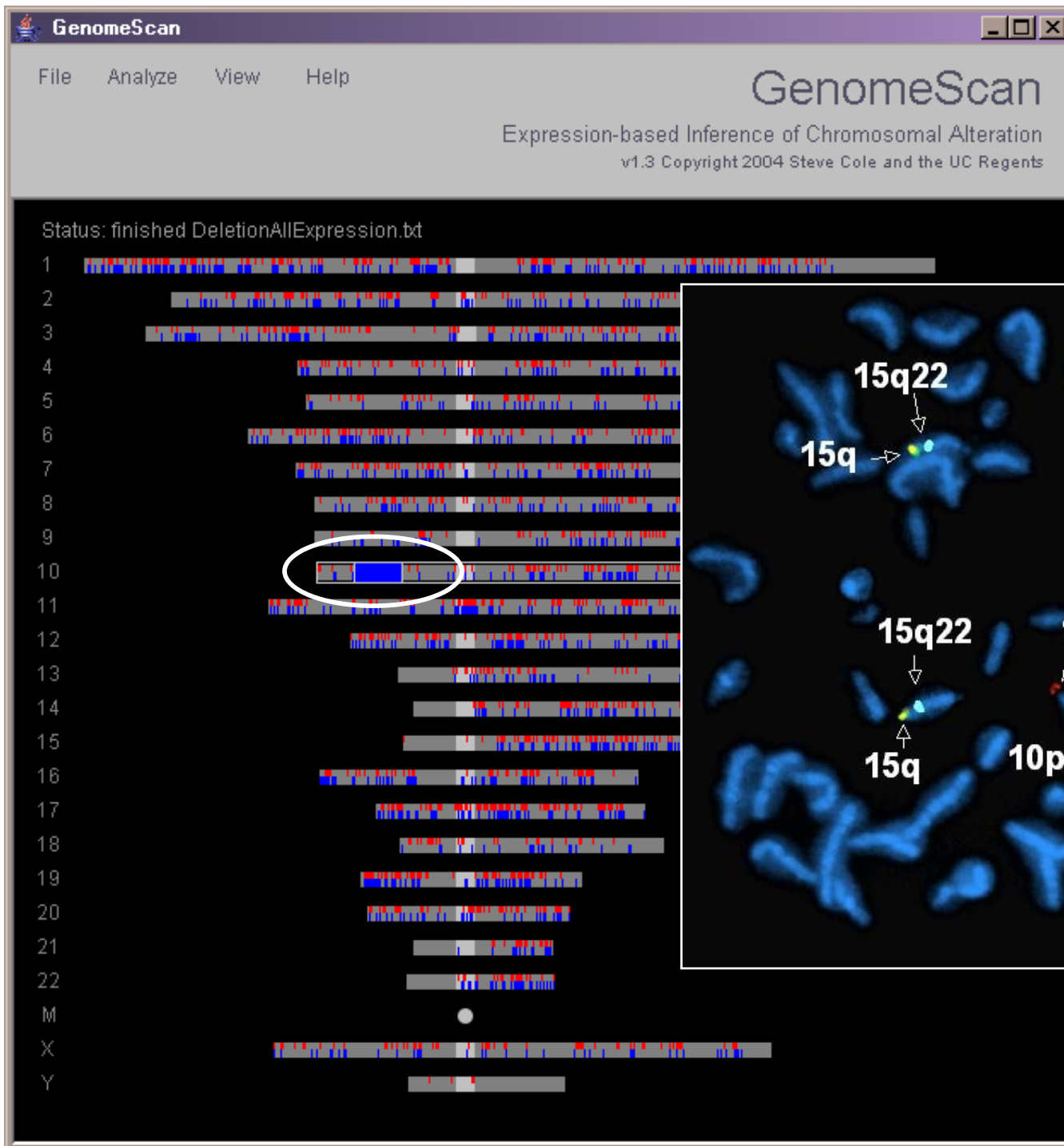
DNA

Gene

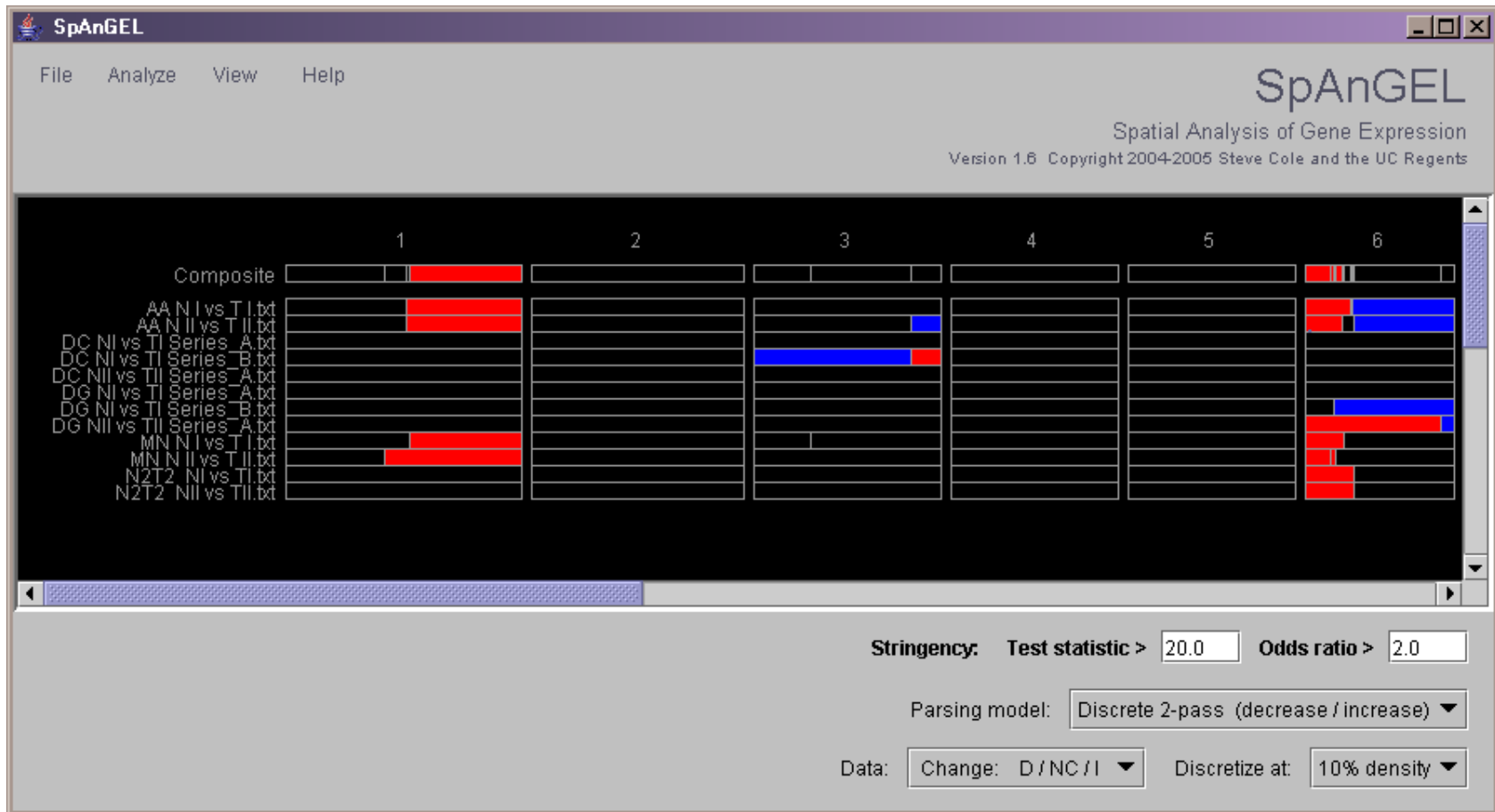




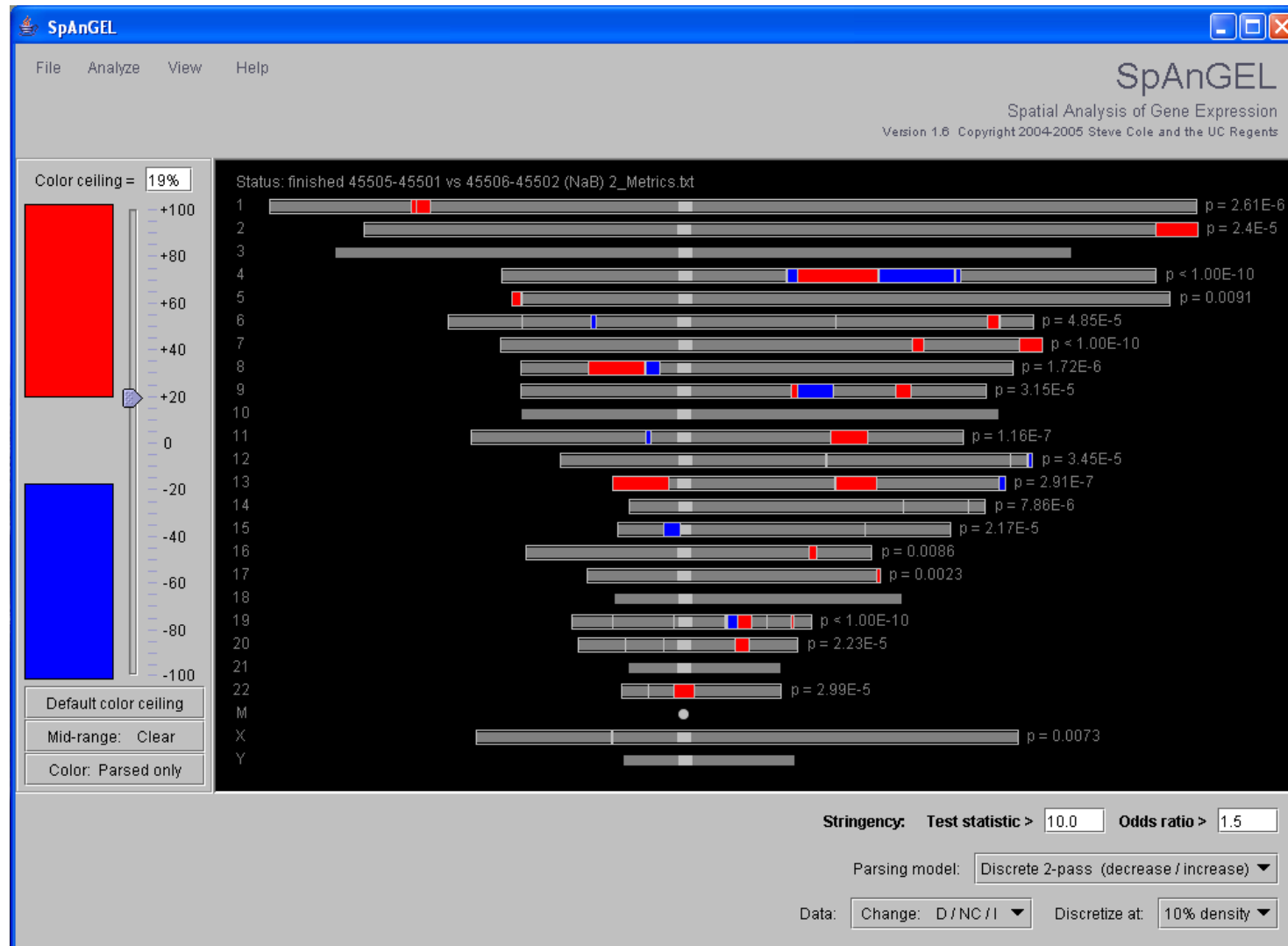


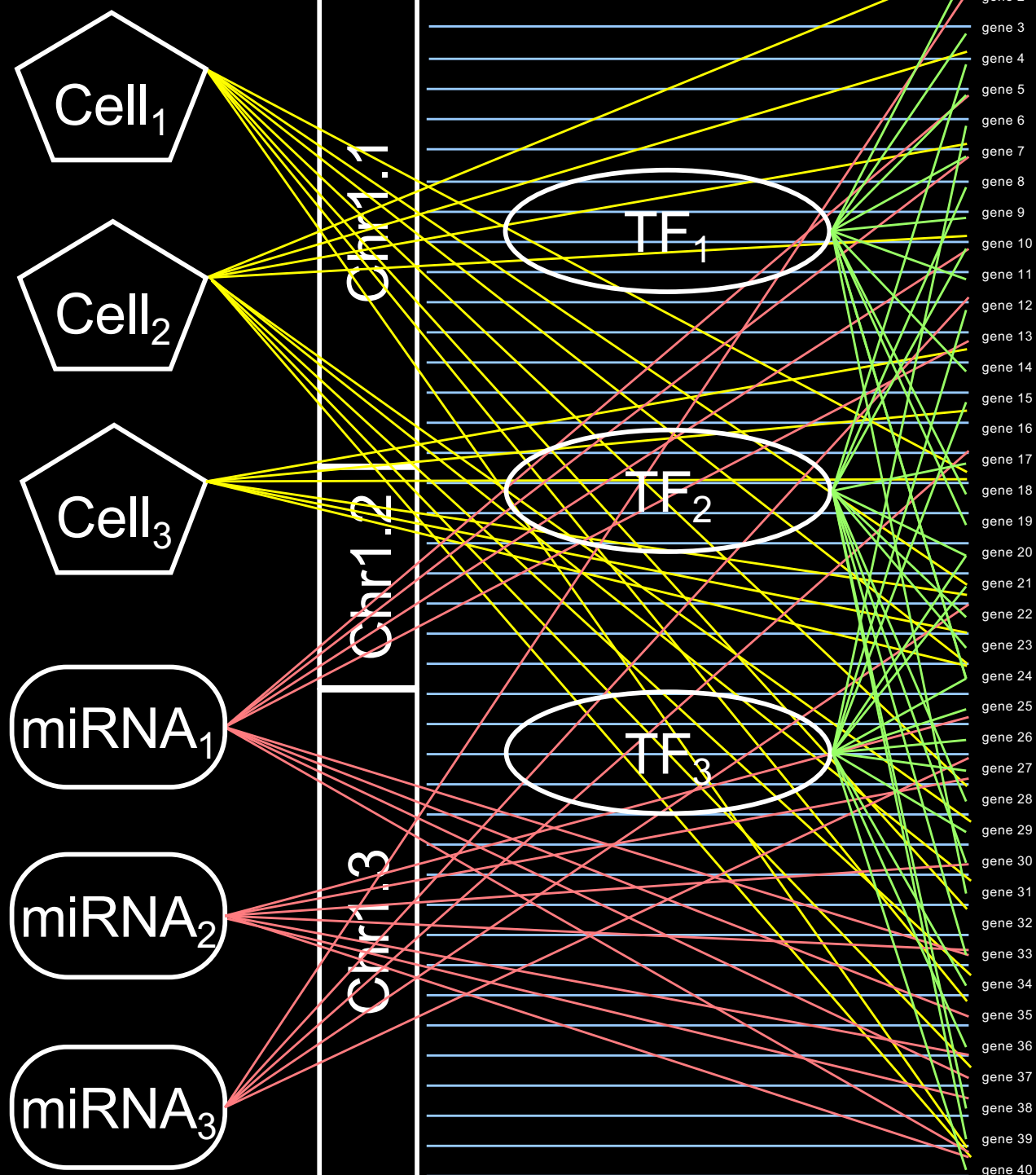


Chromosomal aberration profiles

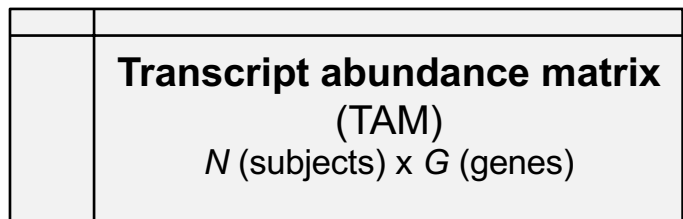


Epigenetics, chromatin accessibility, & RIDGEs

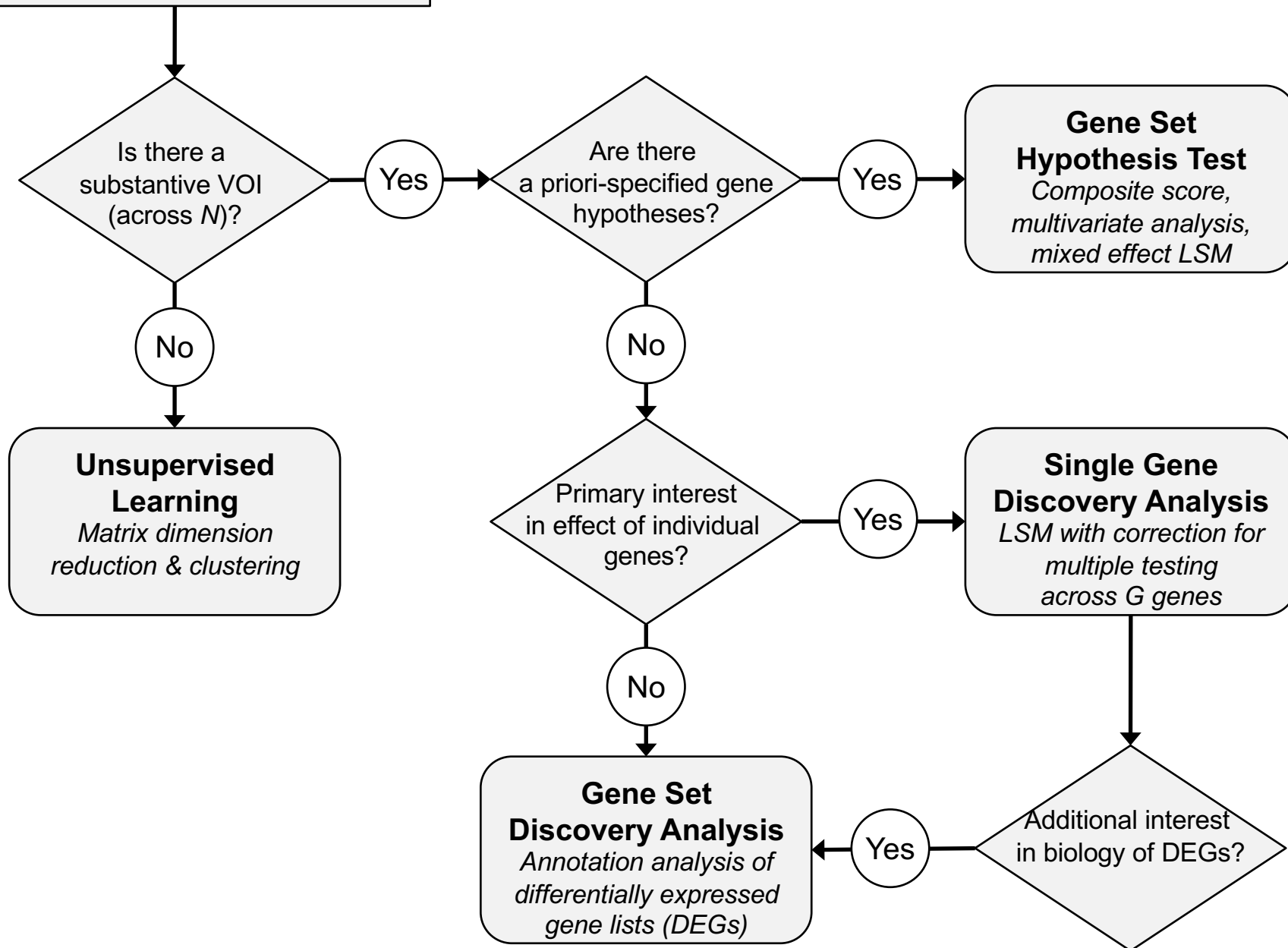




Some notes on statistical testing for gene list analysis

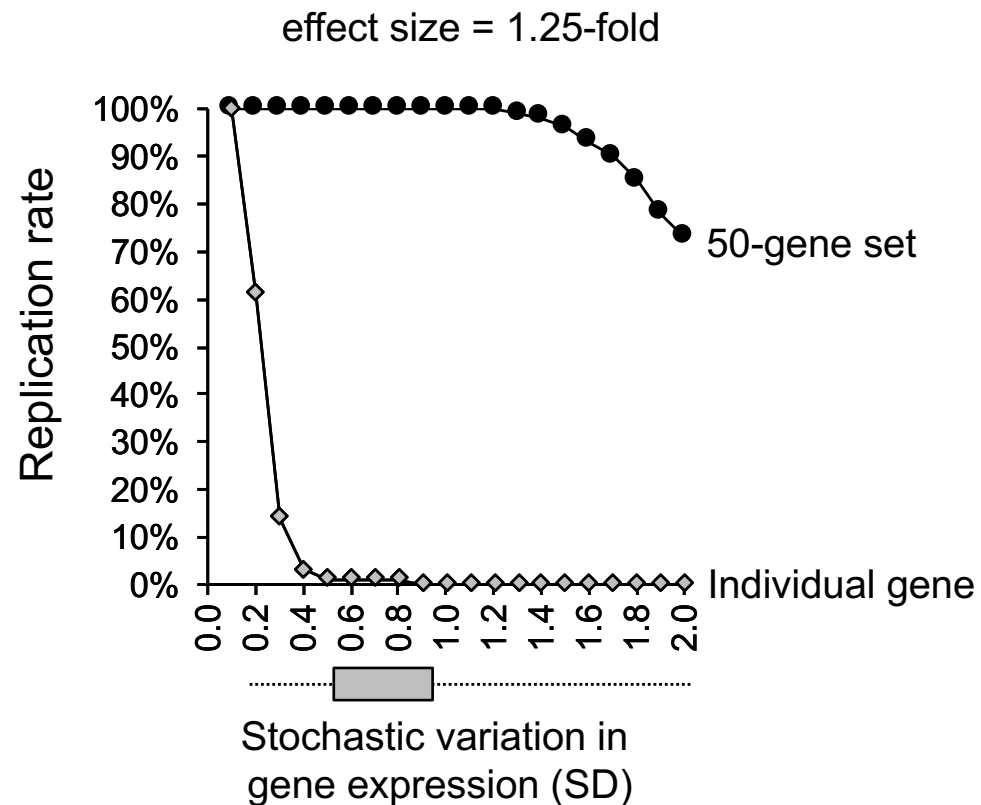


VOI = variable of interest (regressor/phenotype/experimental condition)
LSM = linear statistical model
DEG = differentially expressed gene



Painful lesson #1: If you want replicable results, focus on functionally related **sets** of genes (not individual gene transcripts)

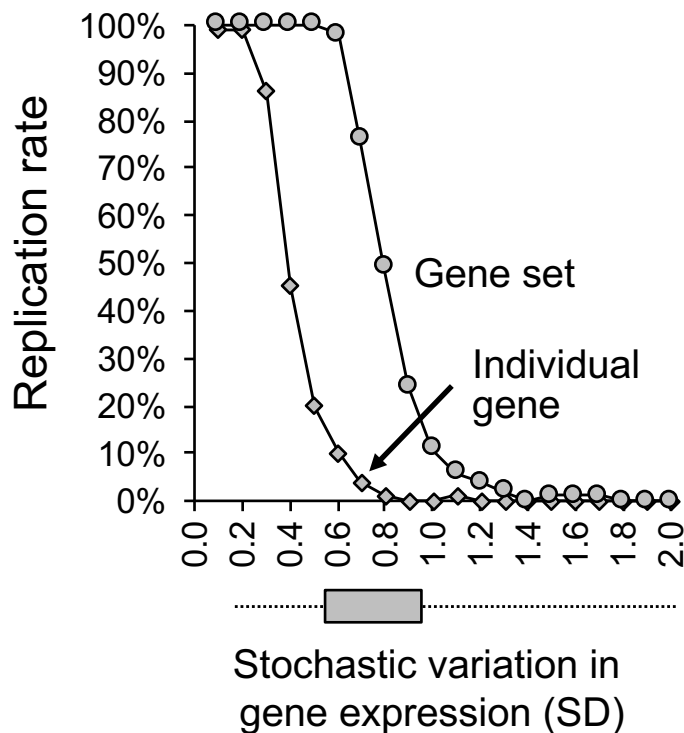
Differential expression of an a priori gene set



Painful lesson #2: If you want replicable results, conduct 2nd stage bioinformatics on gene lists derived from point estimates of **effect size** (not *p/q*-values)

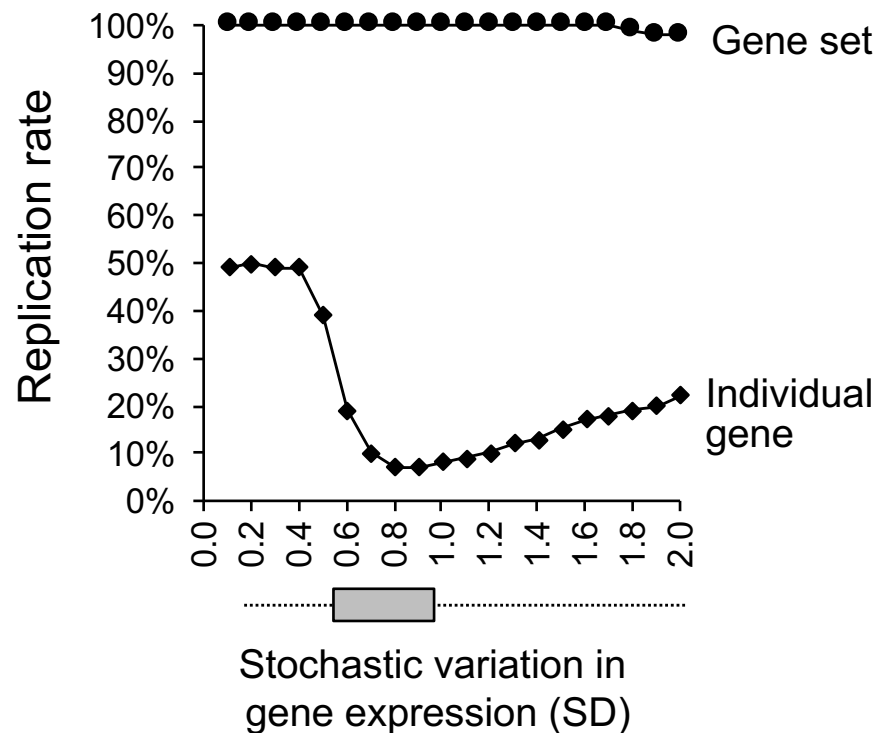
Gene set enrichment analysis

Low level input: *p*-value gene list



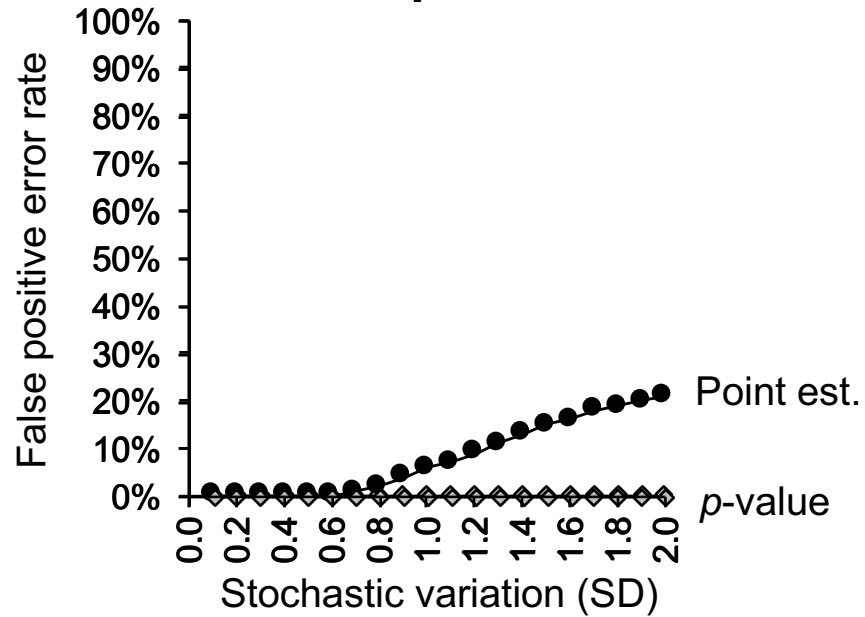
Gene set enrichment analysis

Low level input: point estimate gene list

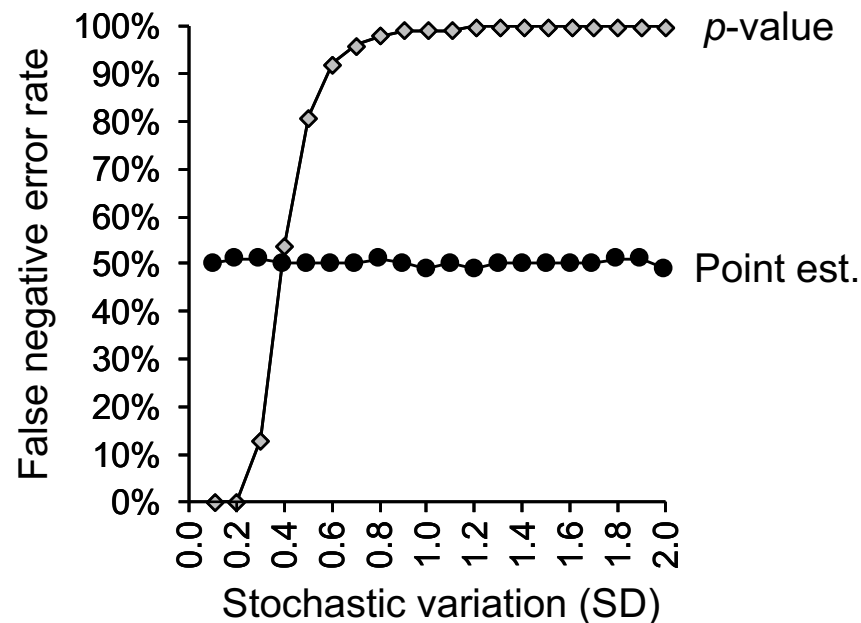


Individual gene association

False positive

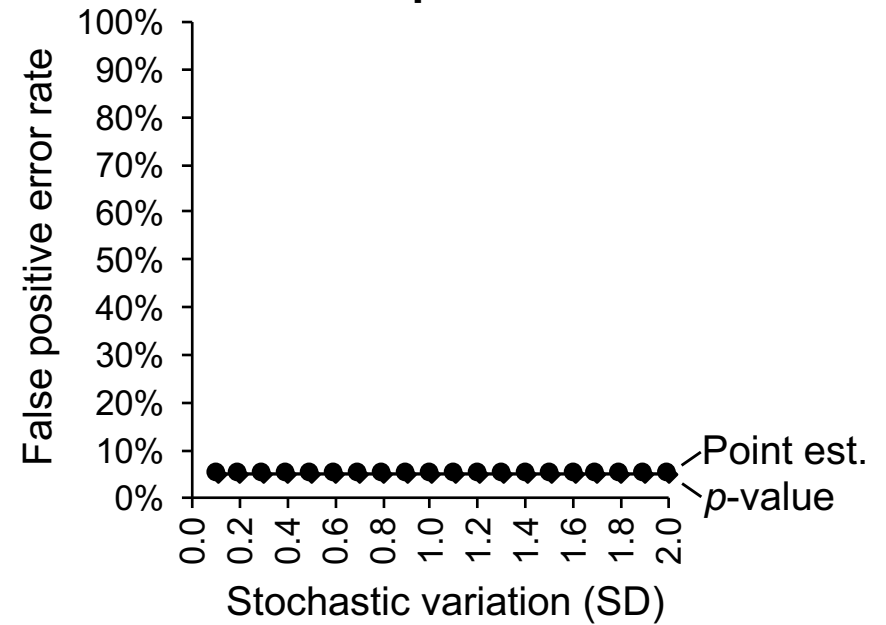


False negative

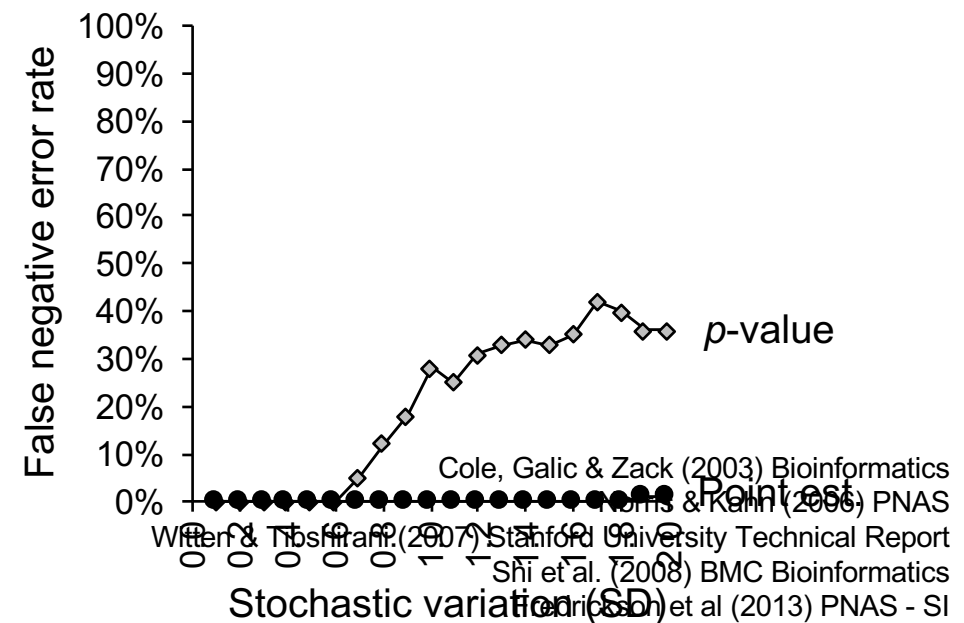


Gene set association

False positive

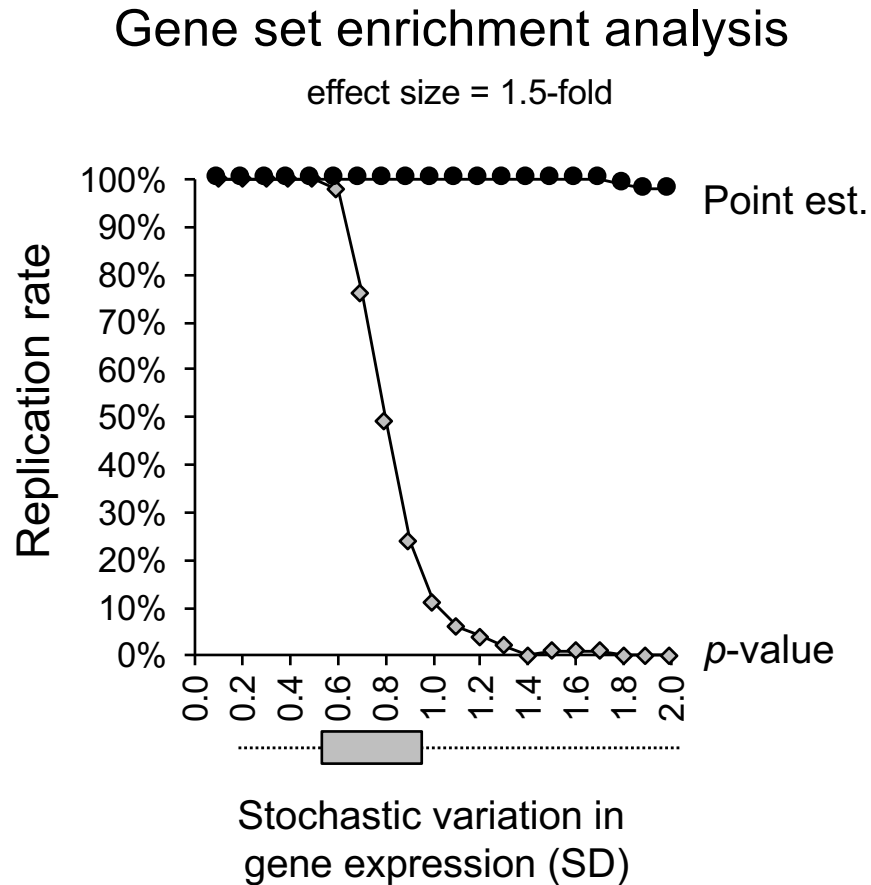


False negative



Cole, Galic & Zack (2003) Bioinformatics
 Witten & Tibshirani (2007) Stanford University Technical Report
 Shi et al. (2008) BMC Bioinformatics
 Morris et al (2013) PNAS - SI

Implication of false negative errors for result replication



Cole, Galic & Zack (2003) Bioinformatics

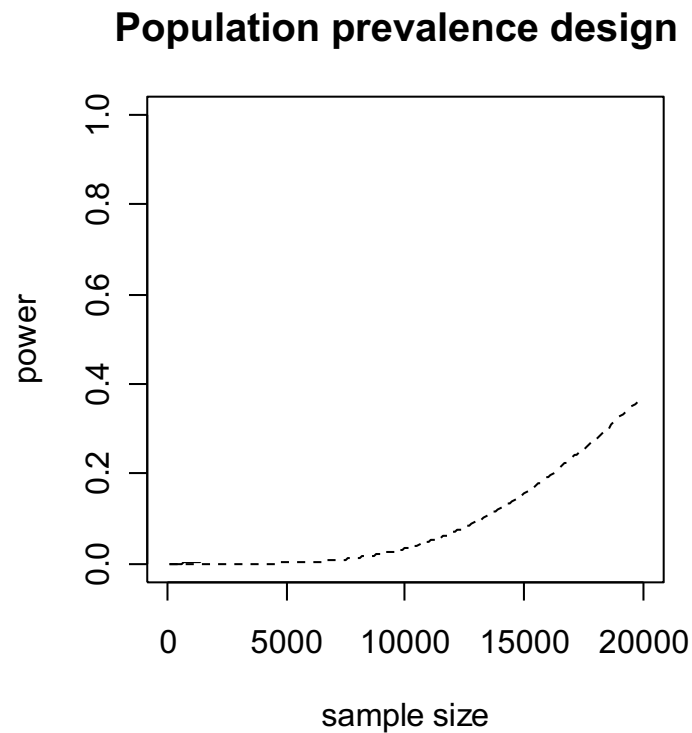
Norris & Kahn (2006) PNAS

Witten & Tibshirani (2007) Stanford University Technical Report

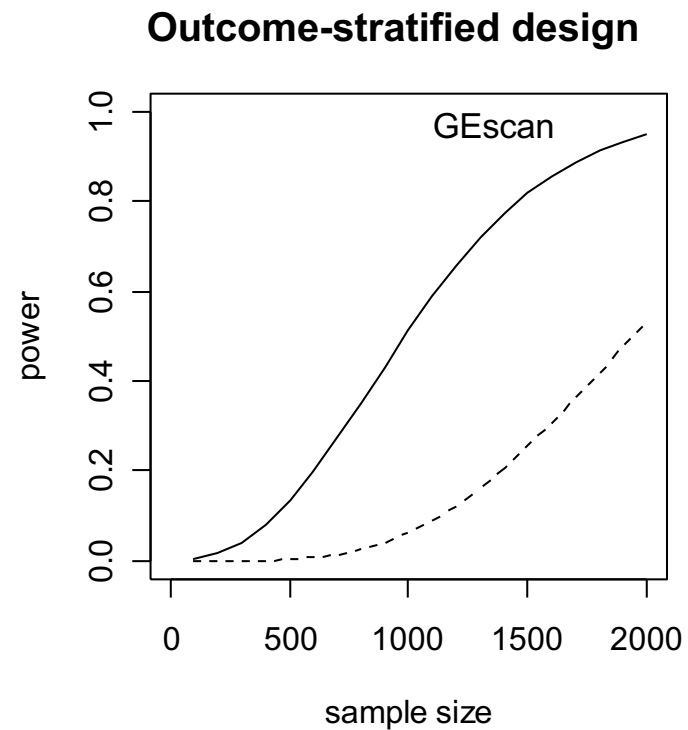
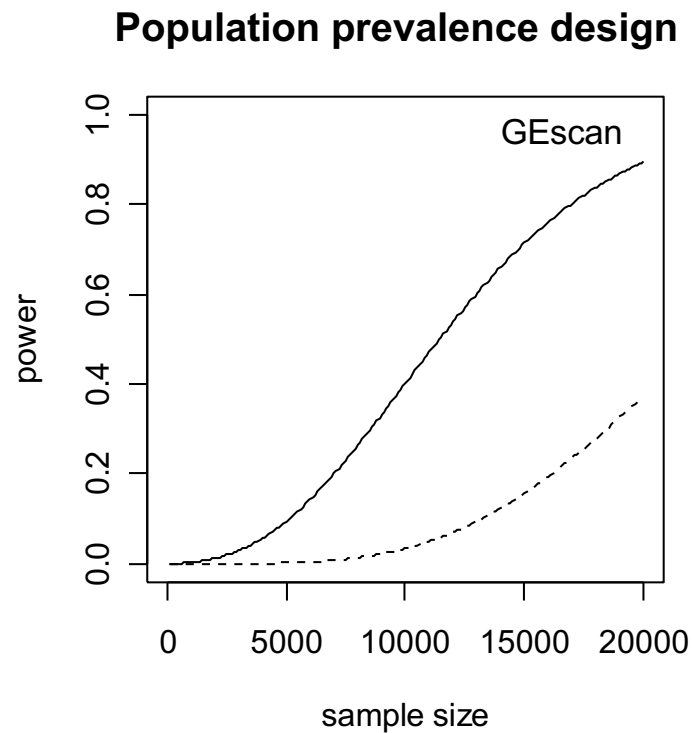
Shi et al (2008) BMC Bioinformatics

Fredrickson et al (2013) PNAS - SI

Painful lesson #3: Hypotheses are ***extremely*** valuable for statistical power in genomics. Don't pretend you know nothing (unless you really do).

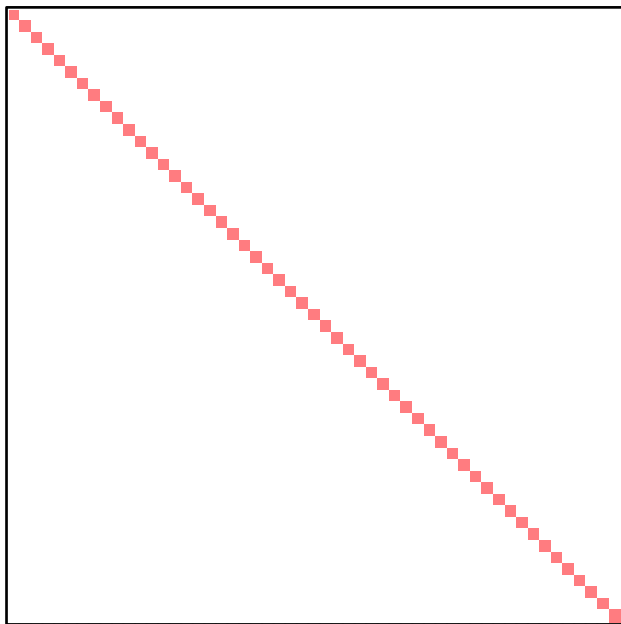


Painful lesson #3: Hypotheses are ***extremely*** valuable for statistical power in genomics. Don't pretend you know nothing (unless you really do).

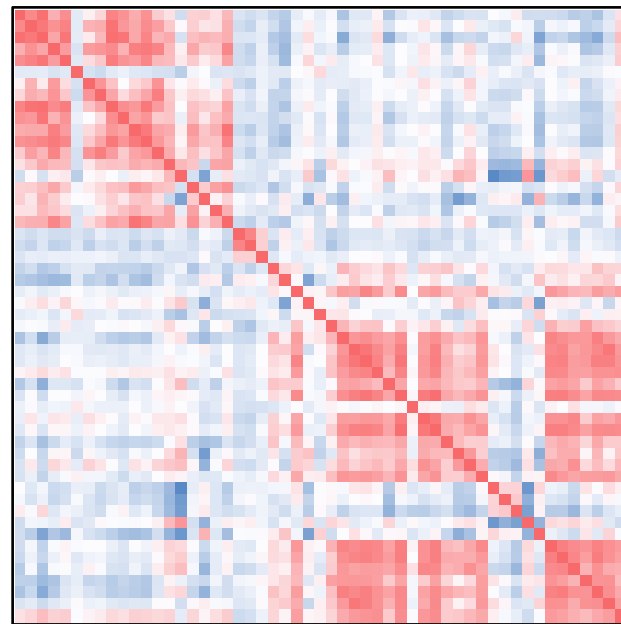


Painful lesson #4: Most p - and q -values are wrong (due to non-independence).
Don't trust them.

**Inter-gene correlation
assuming independence**



**Empirical inter-gene
correlation**



Painful lesson #1: If you want replicable results, analyze functionally related **sets** of genes (not individual gene transcripts)

Sets derive from a biological model of mass action

(i.e., environmental conditions, pathogens, distinct cell types, differentiation states, TFs, chromosomal & epigenetic alterations, genetic polymorphisms, etc.)

Painful lesson #2: If you want replicable results, conduct interpretive bioinformatics on gene lists derived from point estimates of **effect size** (not *p/q*-values)

Effect sizes (not SNRs) reflect mass biological processes

(i.e., environmental conditions, pathogens, cells, differentiation states, TFs, chromosomal & epigenetic alterations, genetic polymorphisms, etc.)

Painful lesson #3: Hypotheses are **extremely** valuable for statistical power, and that is the limiting resource in genomics. Don't pretend you know nothing.

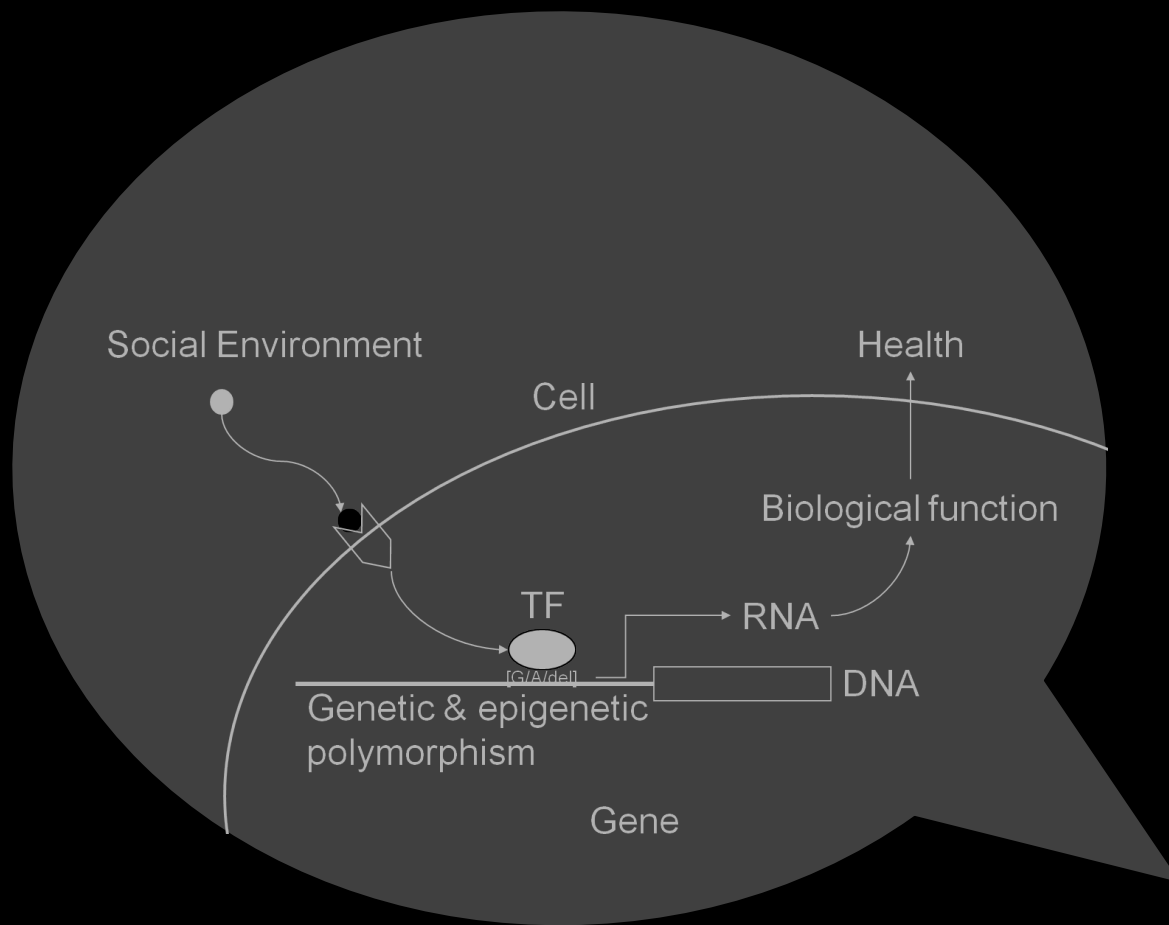
Hypotheses derive from a biological model

(i.e., or perhaps at least a set of ~100 imaginable models \ll 20,000 genes)

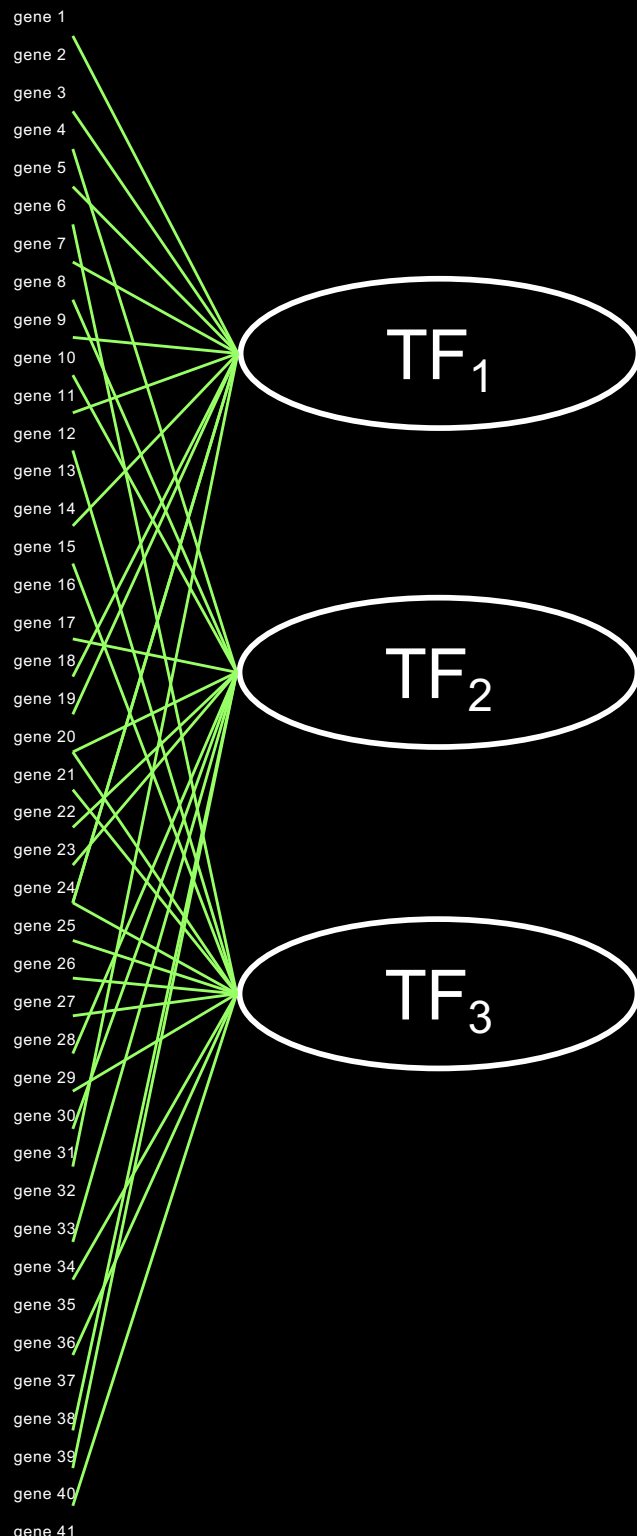
Painful lesson #4: *p/q*-values are inaccurate due to mass action correlation.

Model the biological processes that create the correlation

(work with the gene sets, not against them)



- gene 1
- gene 2
- gene 3
- gene 4
- gene 5
- gene 6
- gene 7
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- gene 39
- gene 40



Statistical power

100x narrowed search space

Increased reliability

100x indicators

Substantive understanding

Move from *what happened* to *why*



