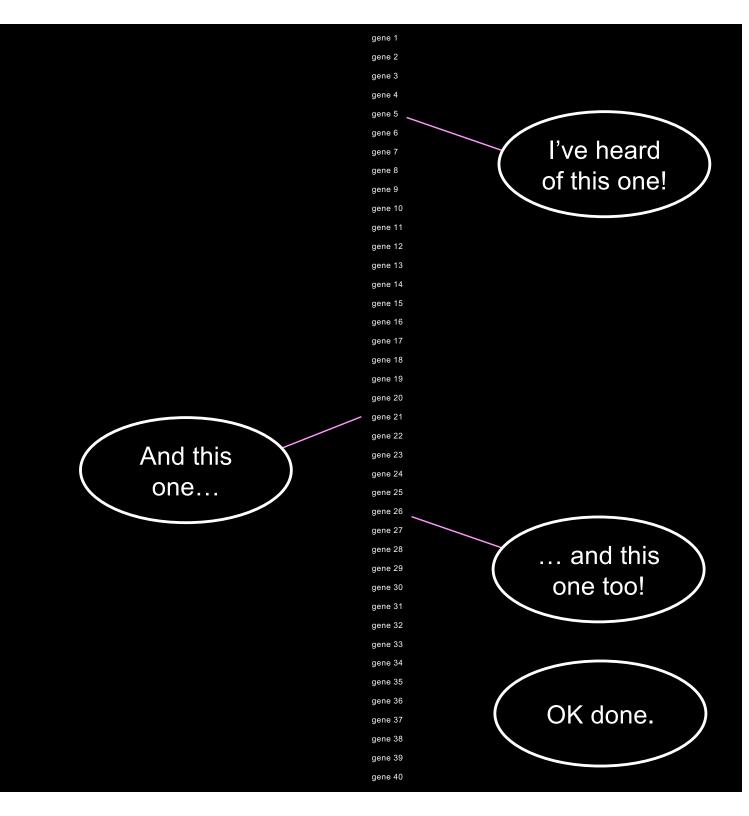
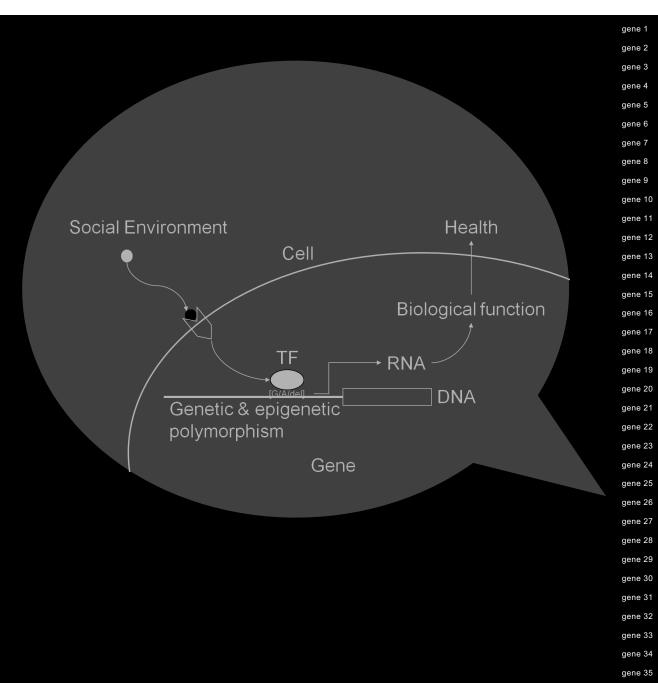
Interpreting your gene list

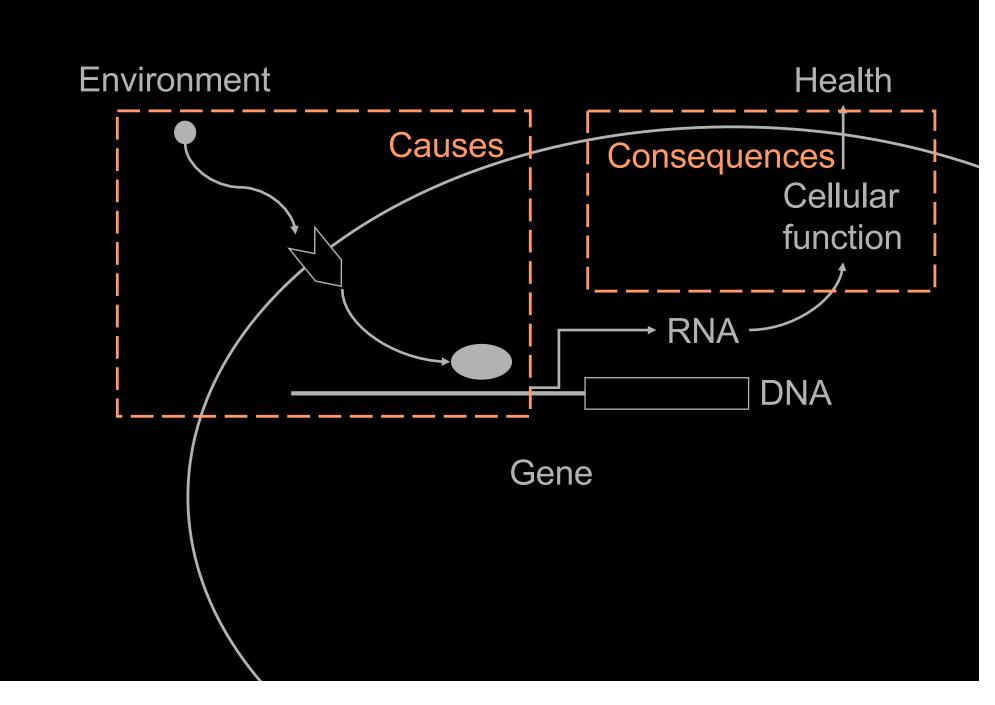
Steve W. Cole, Ph.D.

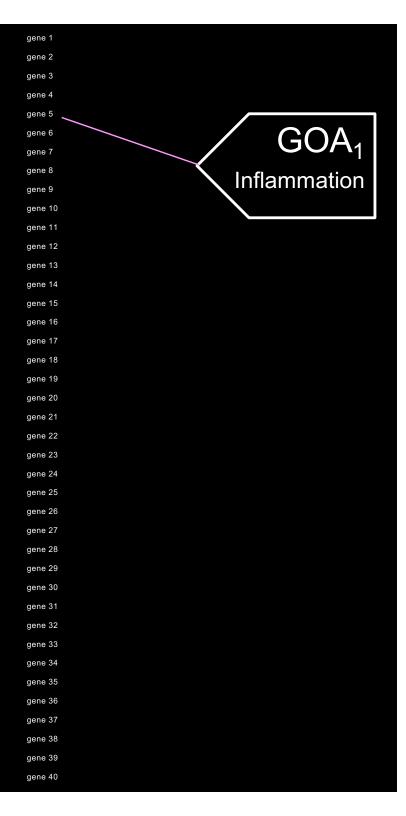
UCLA School of Medicine
Division of Hematology-Oncology



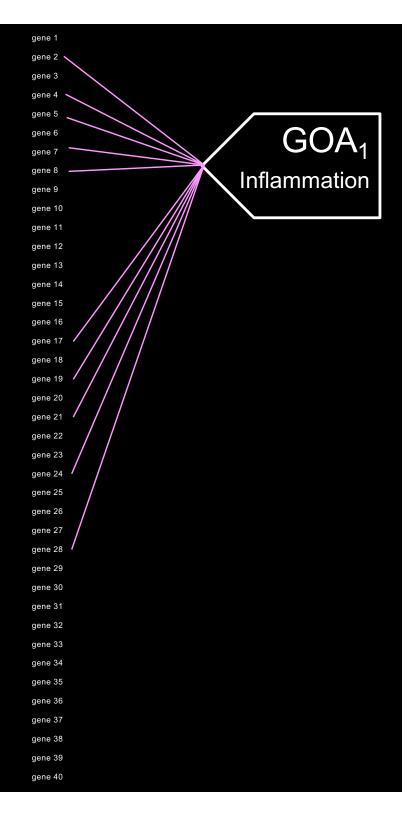


gene 36 gene 37 gene 38 gene 39 gene 40

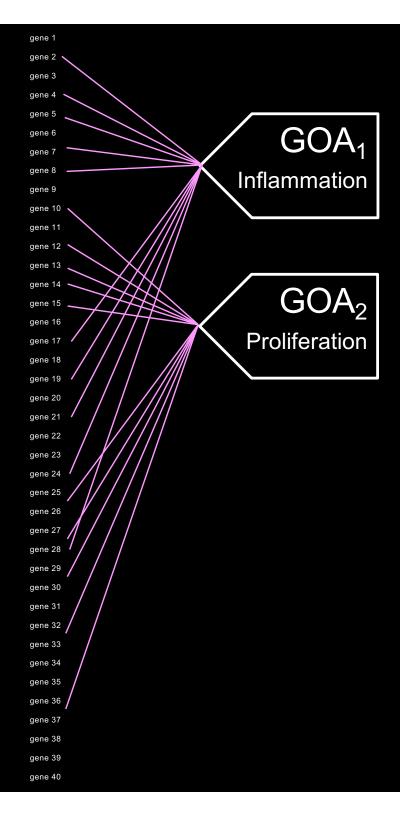




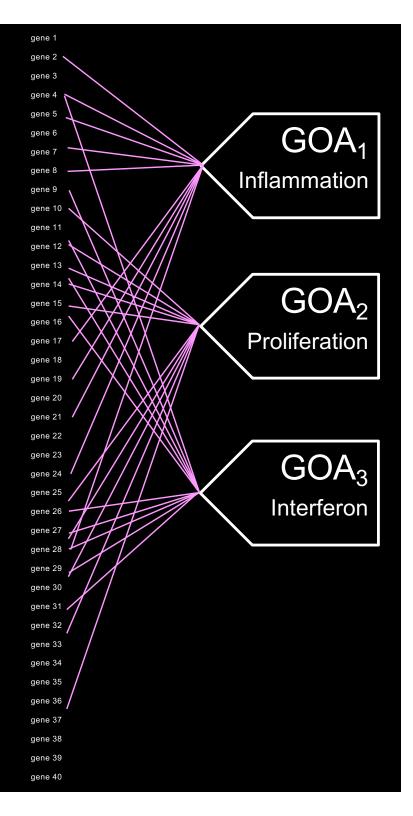
- Genome-wide baseline?
- Expressed transcriptome?
- Up- vs. down-regulated gene sets?



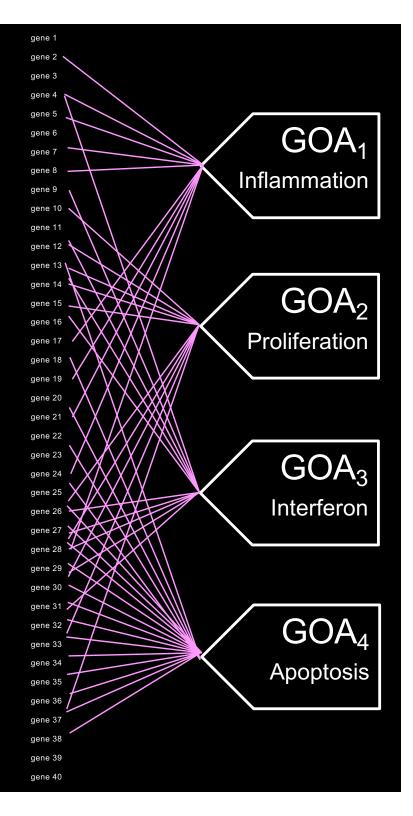
- Genome-wide baseline?
- Expressed transcriptome?
- Up- vs. down-regulated gene sets?



- Genome-wide baseline?
- Expressed transcriptome?
- Up- vs. down-regulated gene sets?

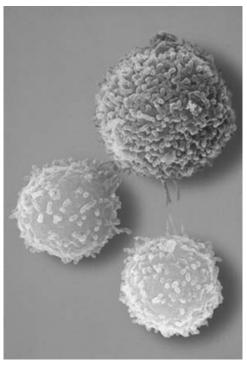


- Genome-wide baseline?
- Expressed transcriptome?
- Up- vs. down-regulated gene sets?

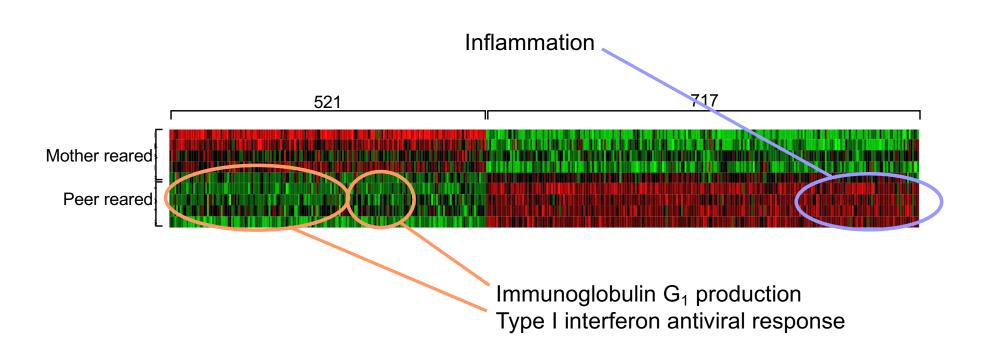


Social instability





CTRA – conserved taristatility sponse to adversity



Low SES

Social loss / bereavement

Post-traumatic stress

Cancer diagnosis

Social threat

Loneliness

Social instability

Chronic stress

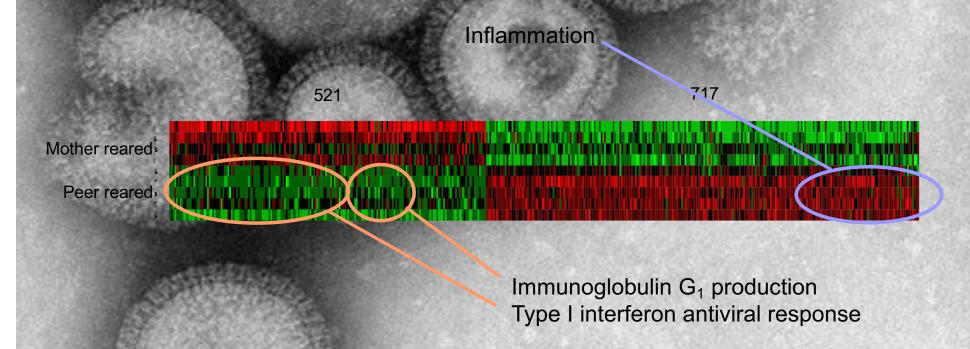
Low social rank

Caregiving for seriously ill

Anxiety

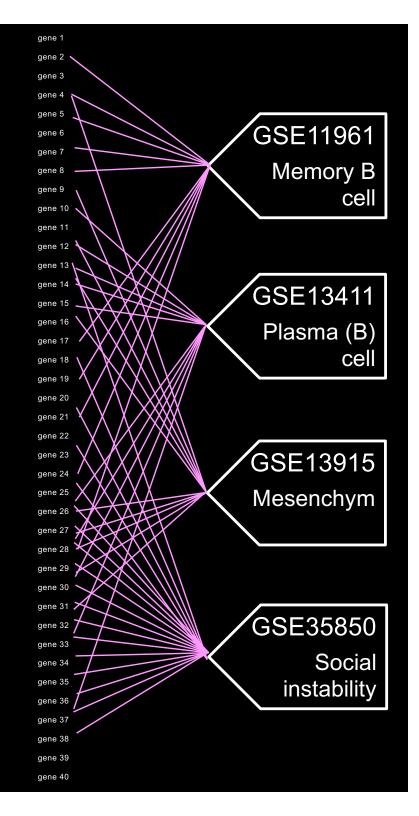
Early life adversity

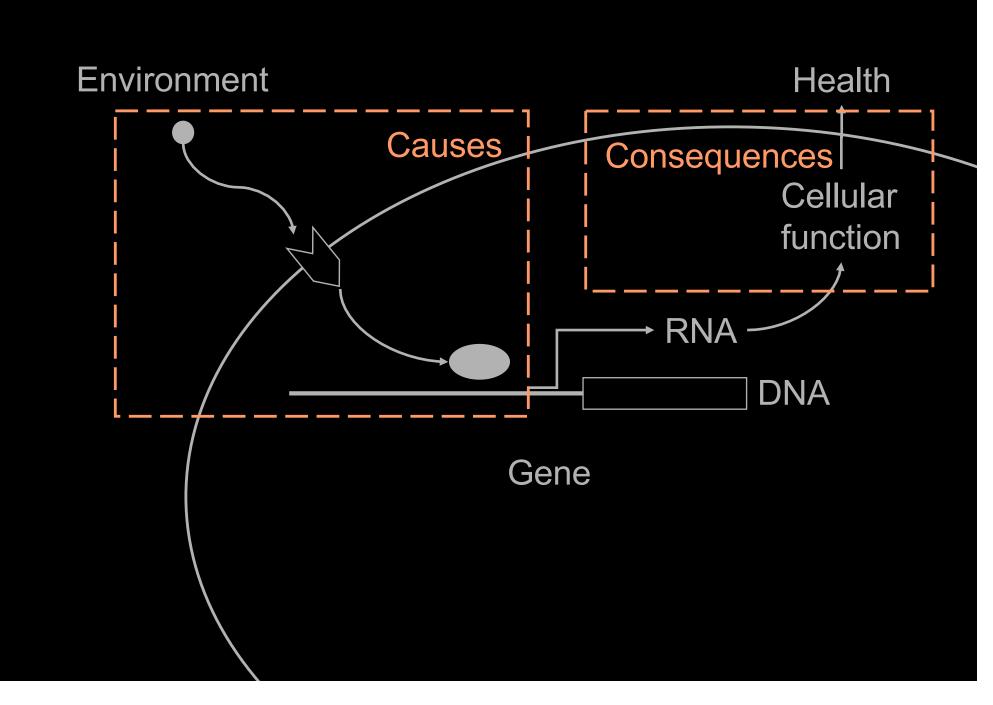


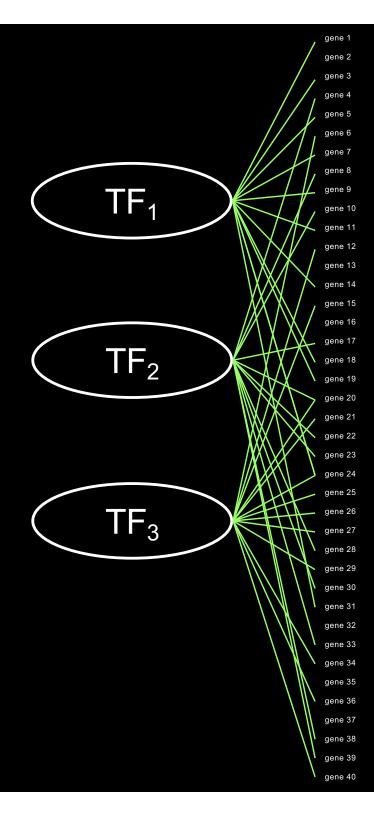


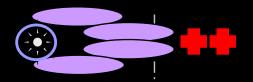
Interpreting results within an empirical transcriptome reference space (vs. "expert" GO tags)

- 1. GEO search (or ArrayExpress, GSEA, etc.)
- 2. Grab the data & compute diagnosticity scores for their experimental "poles"
- 3. Test for gene set differential expression in your data (either them|you or you|them)

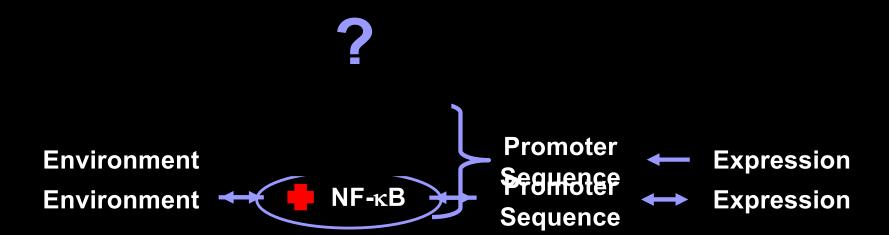




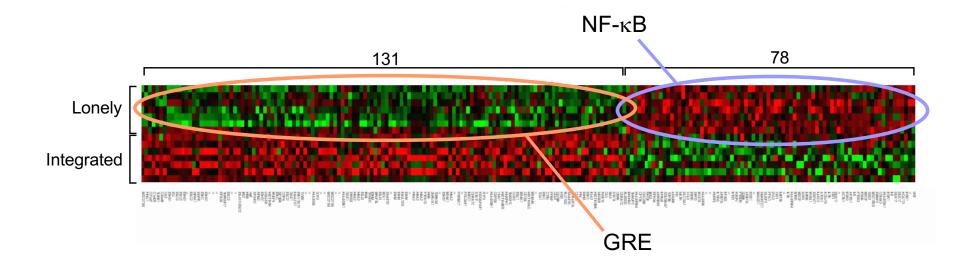


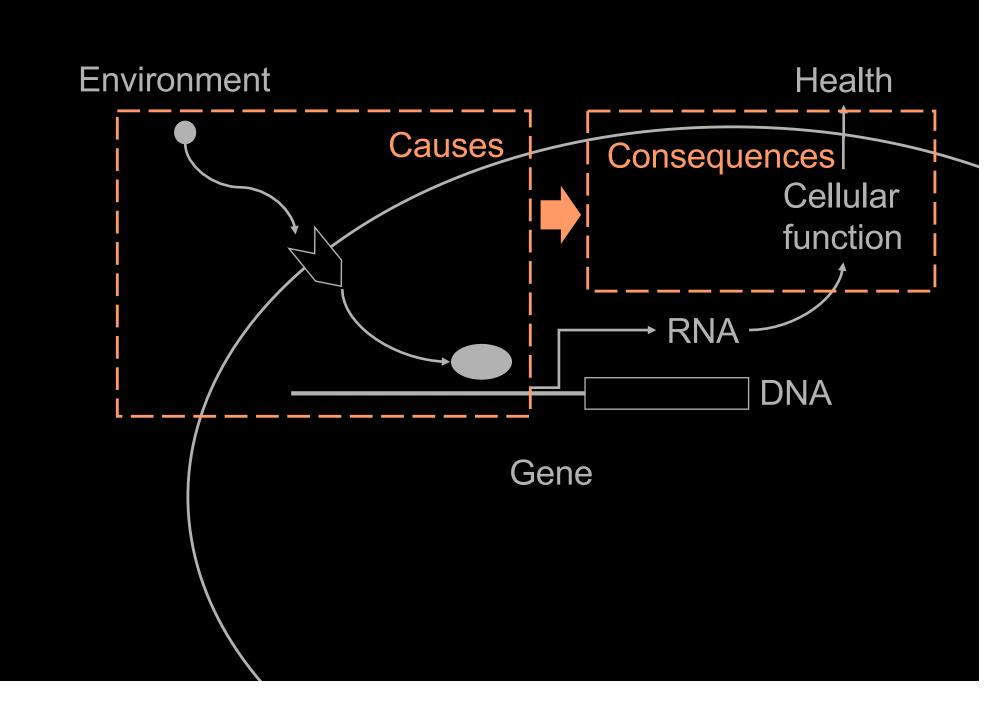


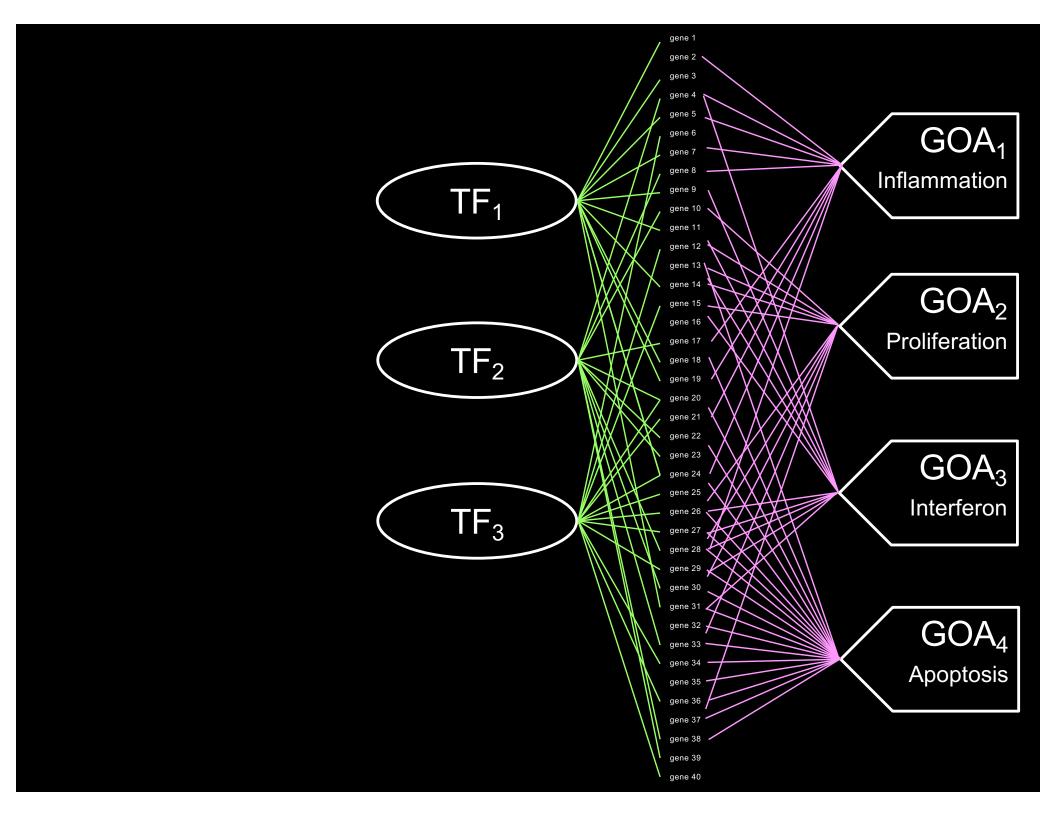


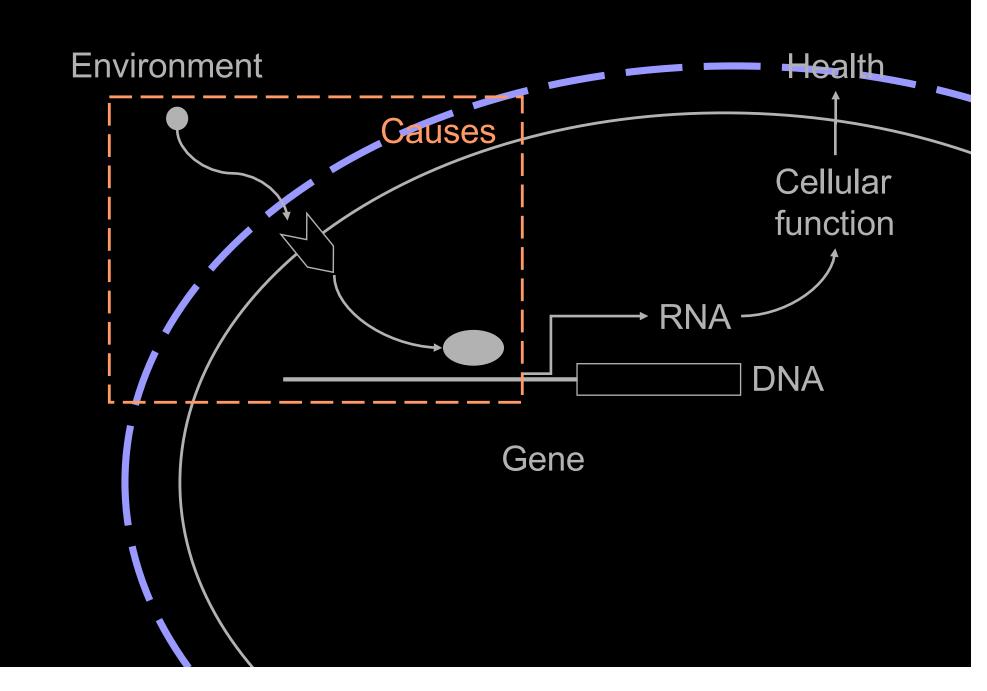


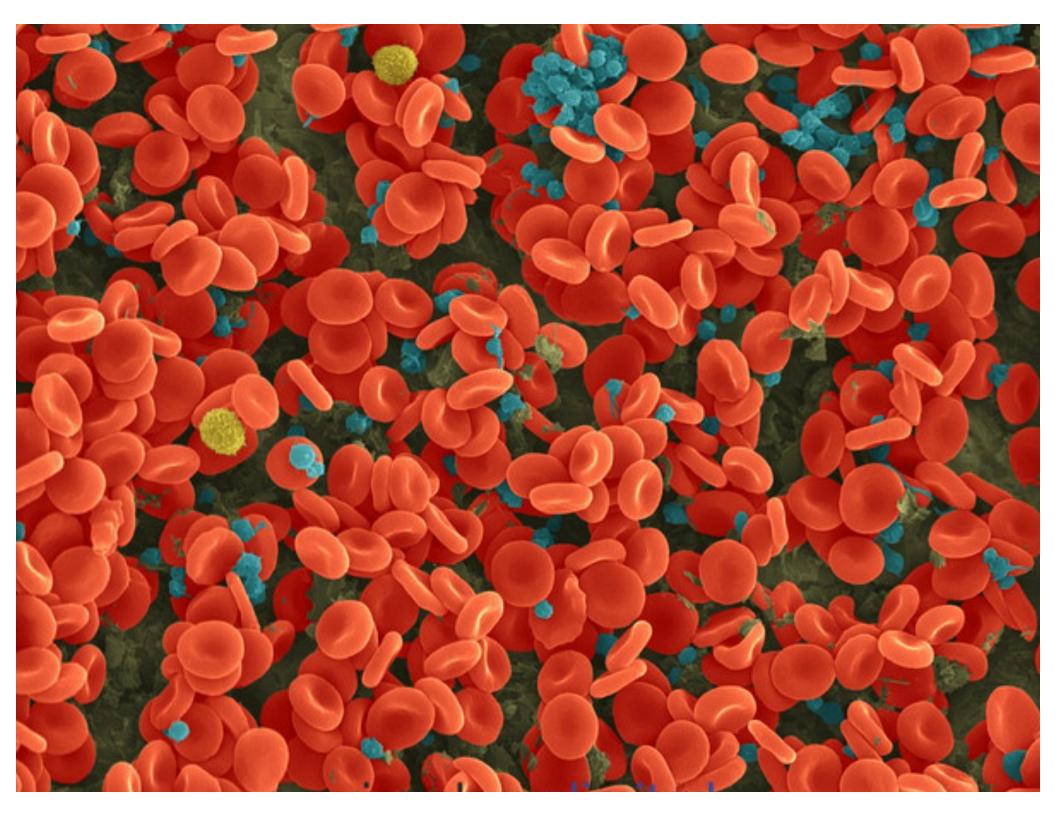
CTRA – Conserved Transcriptional Response to Adversity

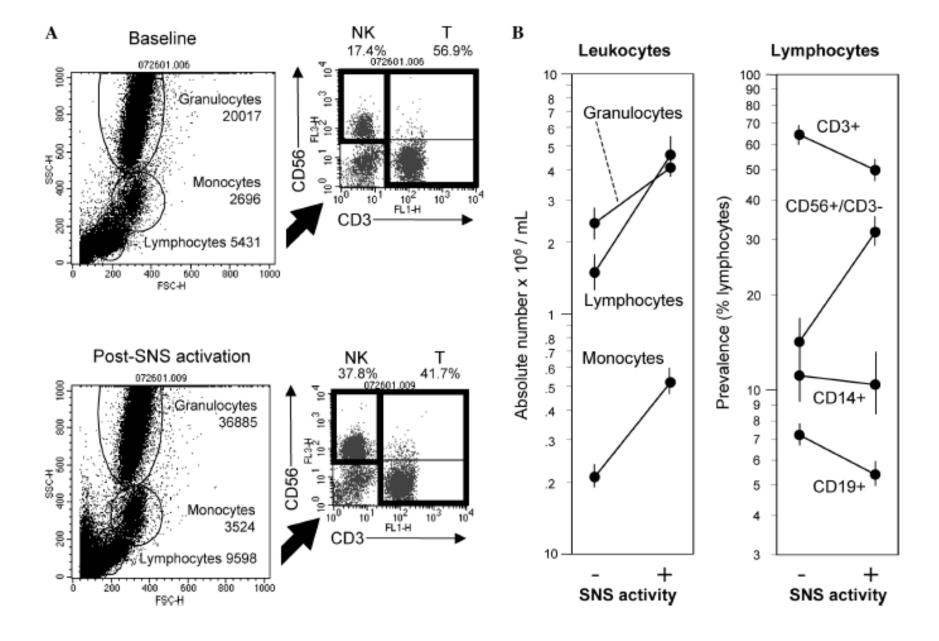




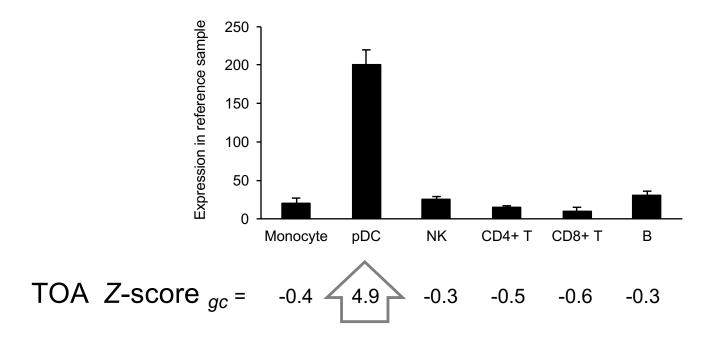


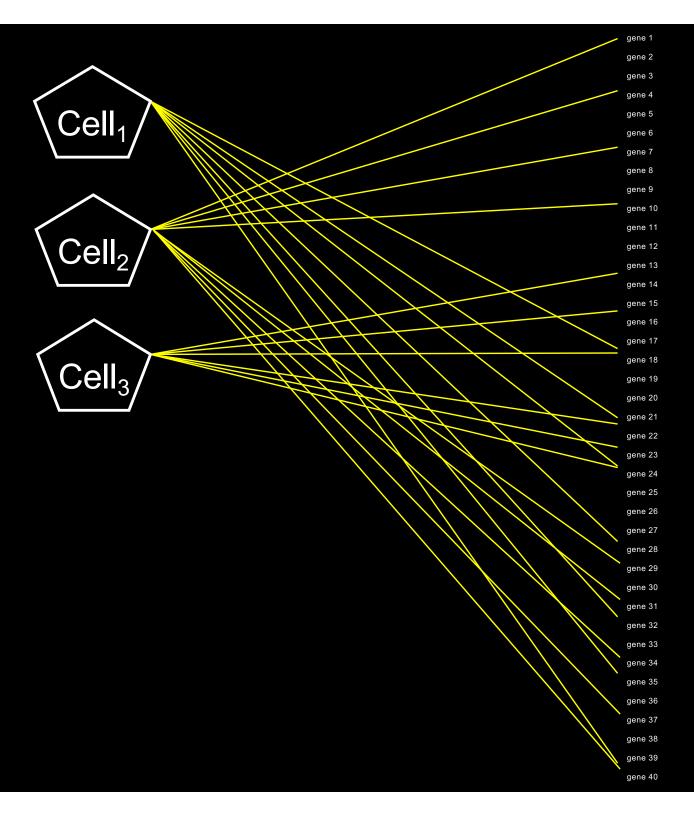




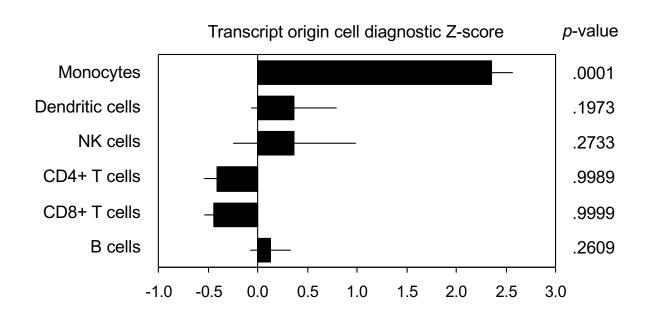


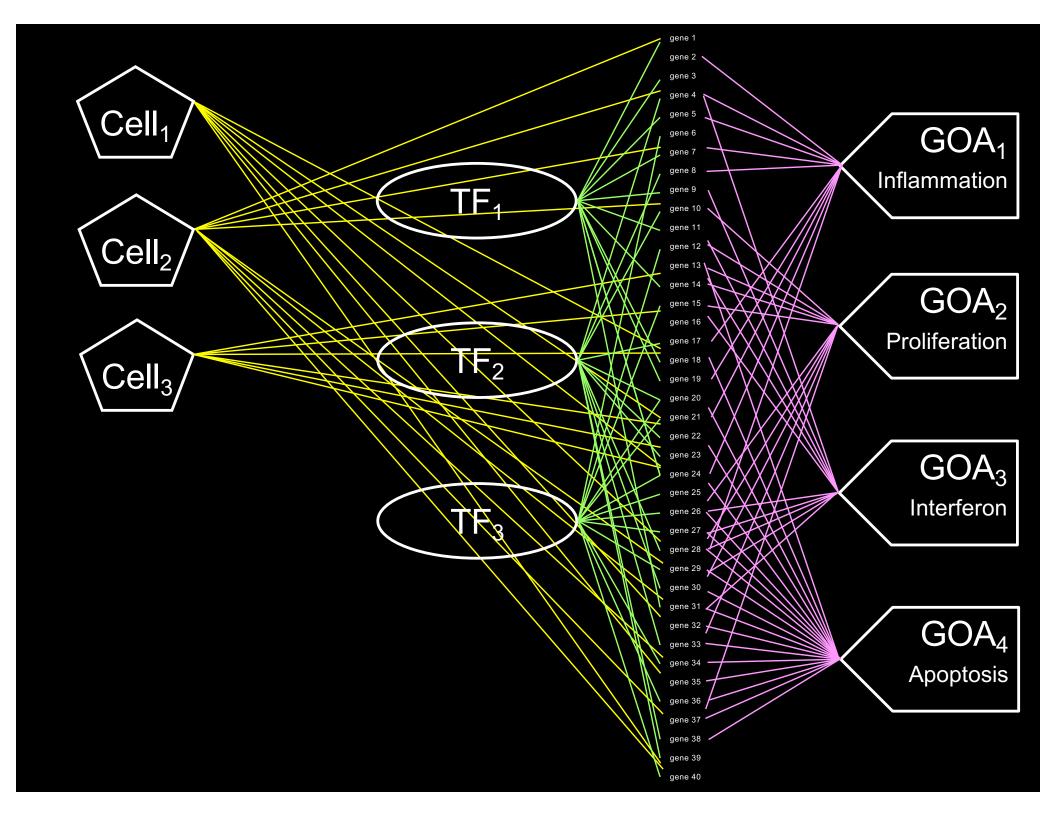
Transcript origin analysis





Cellular target of social adversity

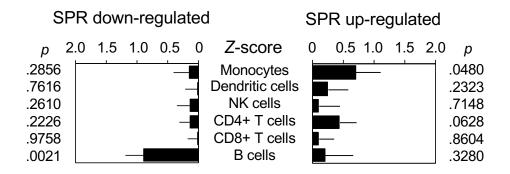


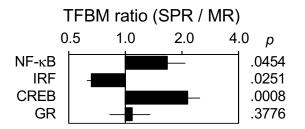


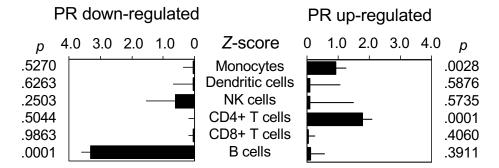
Social instability

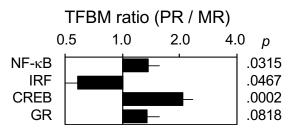
Cellular origin

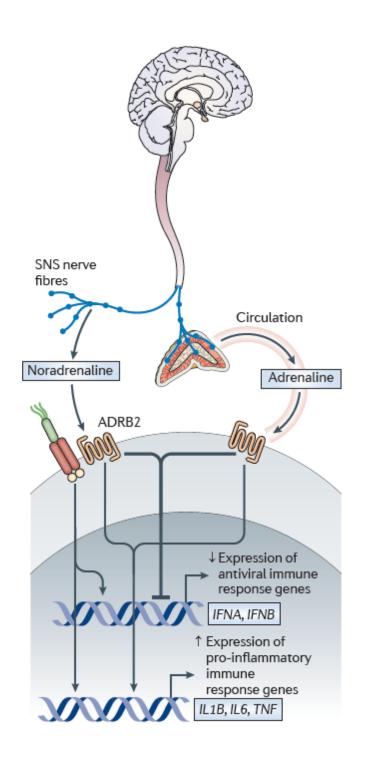
Transcription factor

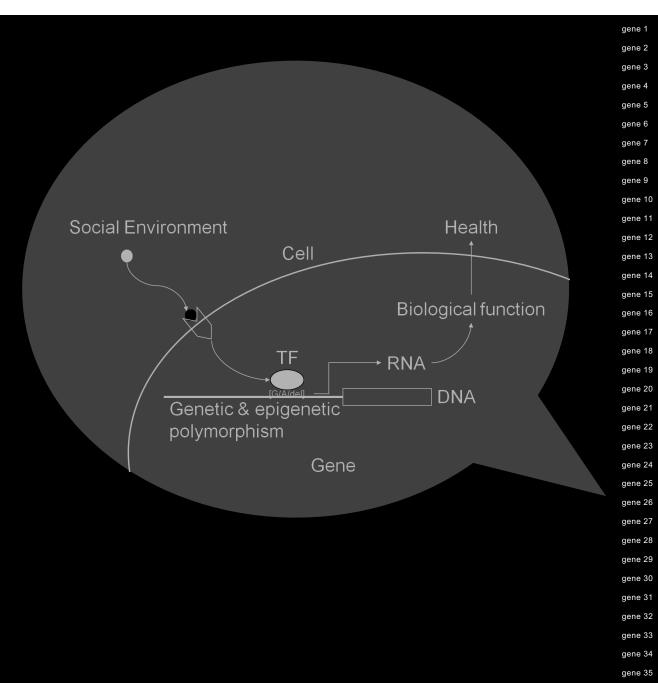




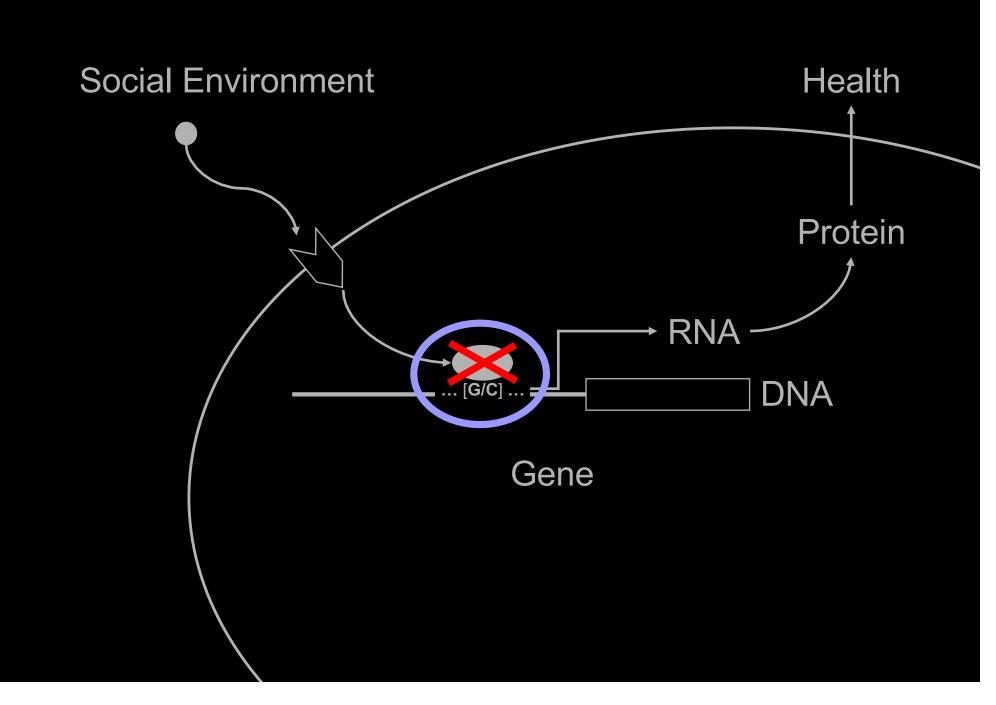








gene 36 gene 37 gene 38 gene 39 gene 40

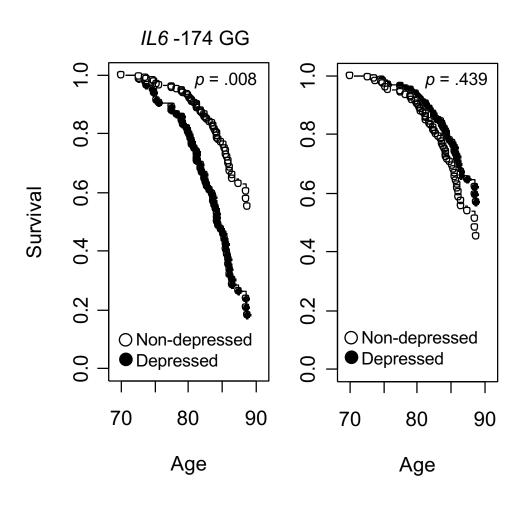


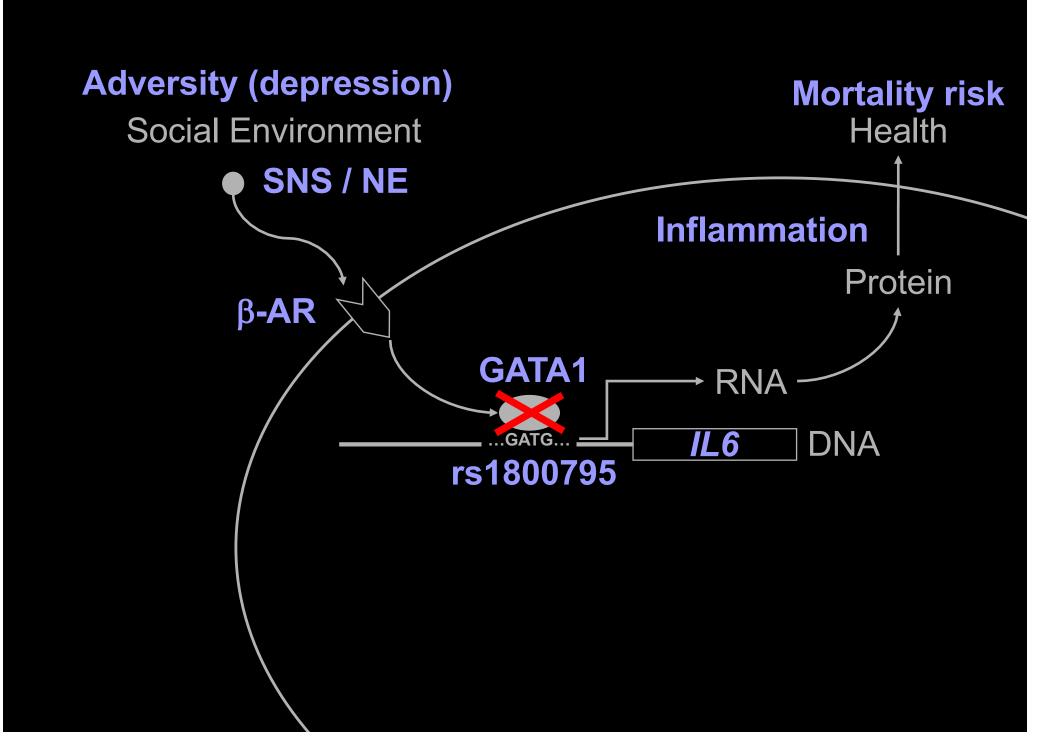
Gene x Environment Interaction

In silico

— TCT TGCGATGCTA AAG — IL6

Gene x Environment Interaction





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SOC -26	FLJ22582 -372 DIA1 -749	LOC401164 -978 LOC391727 -934	LOC222699 -719
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ADAMTS20 -965

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LOC400144 -807

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CDKL1 -902

RTN1 -974

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LOC400320 -443

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LOC390488 -457

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PSMA6 -219

KTN1 -222

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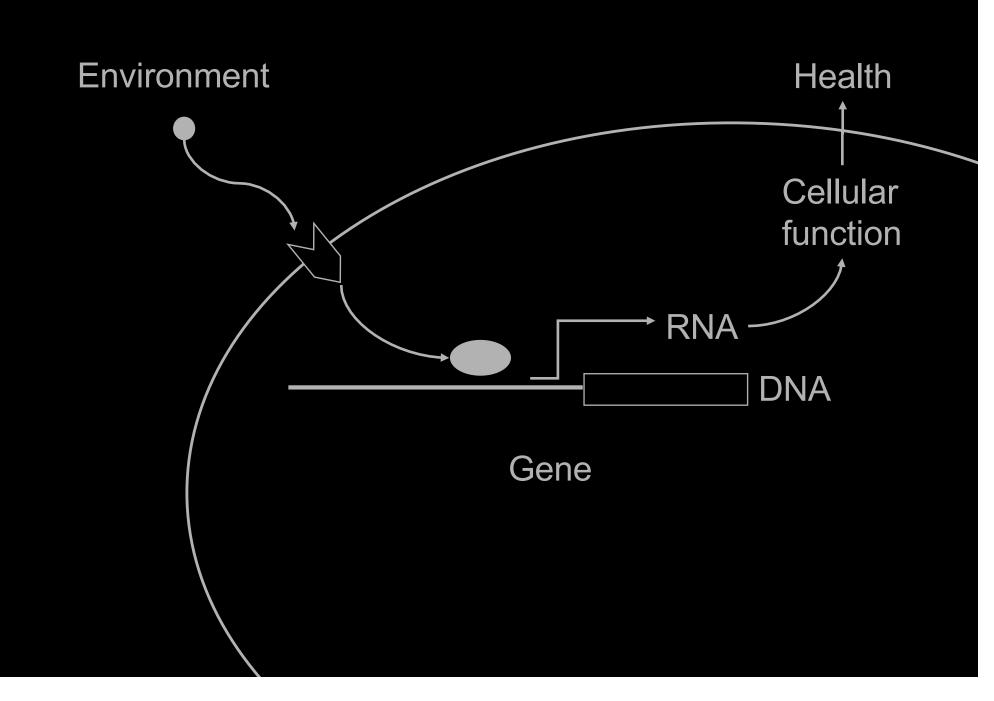
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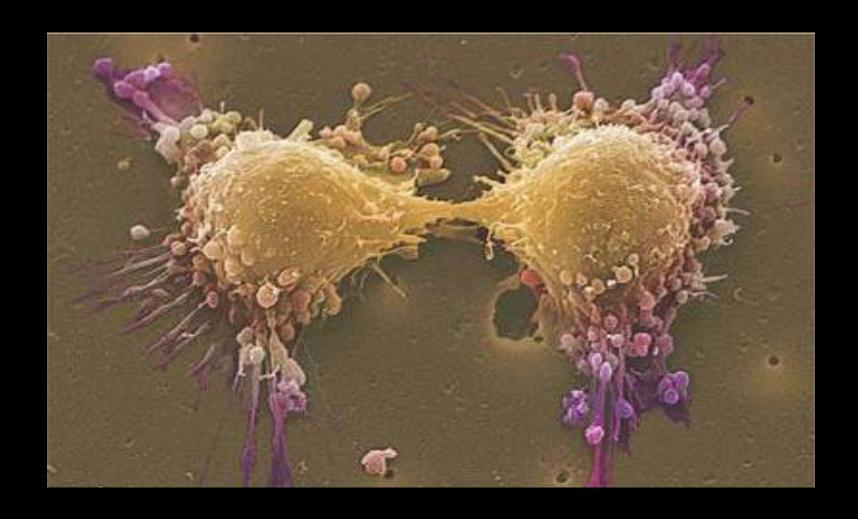
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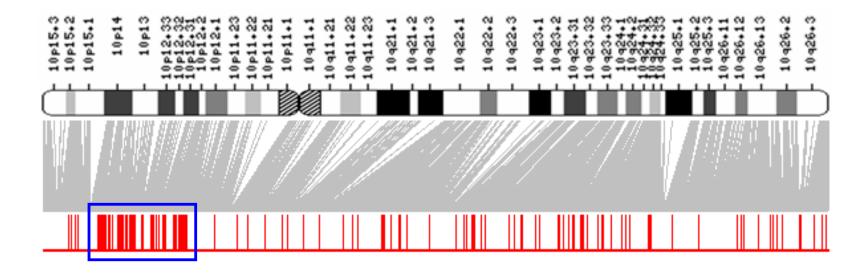
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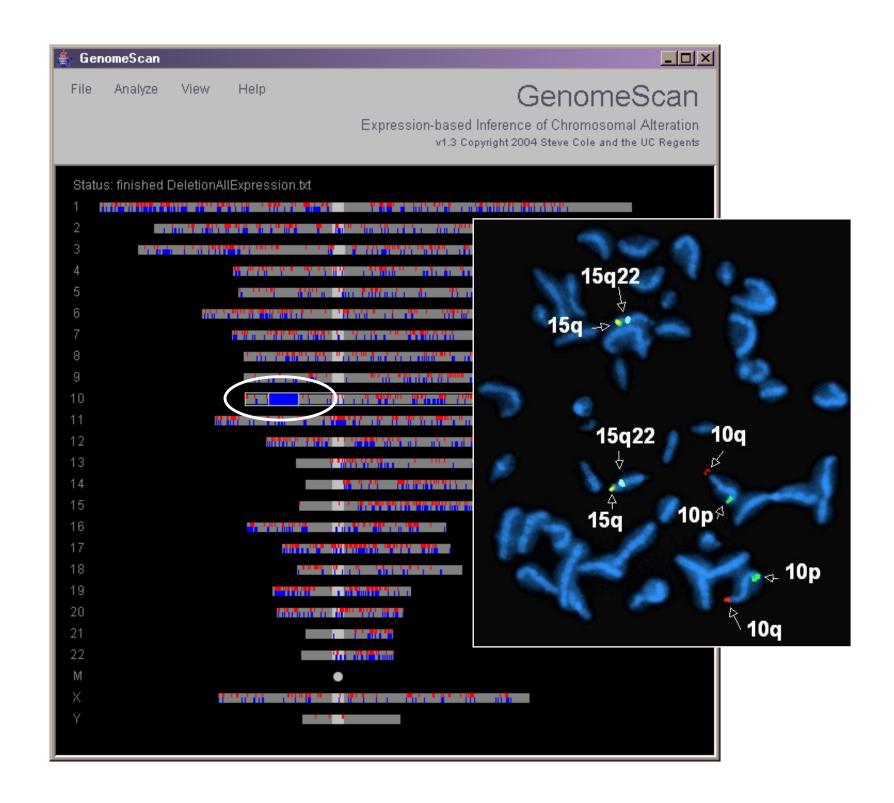
PSMC4 -215 EGLN2 -452 LOC388549 -412 SYNGR4 -825 RPL13A -816 LOC402665 -925 FLJ46385 -176 LOC91661 -13 LAIR2 -705 LAIR2 -705 KIR2DL1 -763 KIR3DL2 3 ZNF583 -867 ZNF71 -861 MGC4728 -490 ZNF211 -76 ZNF211 -76 LOC401895 -957 APBA3 -13 FUT5 -174 TNFSF7 8 SH2D3A -273 8D6A -950 FIF3S4 -547 RAB3D -852 MGC20983 -338 MGC20983 -338 MGC20983 -338 NDUFB7 -741 LOC339377 -660 IL12RB1 -56 IL12RB1 -56 IL12RB1 -56 IL12RB1 -56 LOC148198 -361 CEBPA -564 UNQ467 -521 FLJ22573 -941 CLC -823 DYRK1B -849 DYRK1B -849 DYRK1B -849 PSG11 -297 PSG11 -297 PSG4 -299 PSG4 -299 PSG9 -435 FL J34222 -415 ERCC2 -123 DMPK -988 PGLYRP1 -212 LIG1 -806 FLJ32926 -288 CGB8 -202 TFAD2 -546 FLJ20643 -895 LOC400712 -236 SIGLEC6 -972 SIGLEC6 -972 SIGLEC6 -972 ZNF577 -582 ZNF611 -148 ZNF600 -716 ZNE600 -37 NALP9 -489 PRDM2 -762 PRDM2 -762 LOC400743 -400 PADI1 -598 FLJ44952 -494 PPP1R8 5 PPP1R8 5 PPP1R8 5 ATPIF1 -766 ATPIF1 -766 ATPIF1 -766 LOC440581 -793 CGI-94 -384 FLJ14351 -753 UROD -715 LOC441885 -810 IL23R -322 CTH -6 CTH -6 AK5 -966 DNAJB4 -987 CDC7 -604 LOC388649 -426 DCLRE1B -406 LOC440610 -739 LOC440610 -584 LOC440610 -652 LOC440673 -482 BNIPL -420 BNIPI -419 SPRR1B -826 SPRR1B -826 IL6R -110 II 6R -110 CKS1B -983 SYT11 -785 PMF1 -223 LOC164118 -75 FY -397 NCSTN -809 HSPA6 -839 HSPA6 -611

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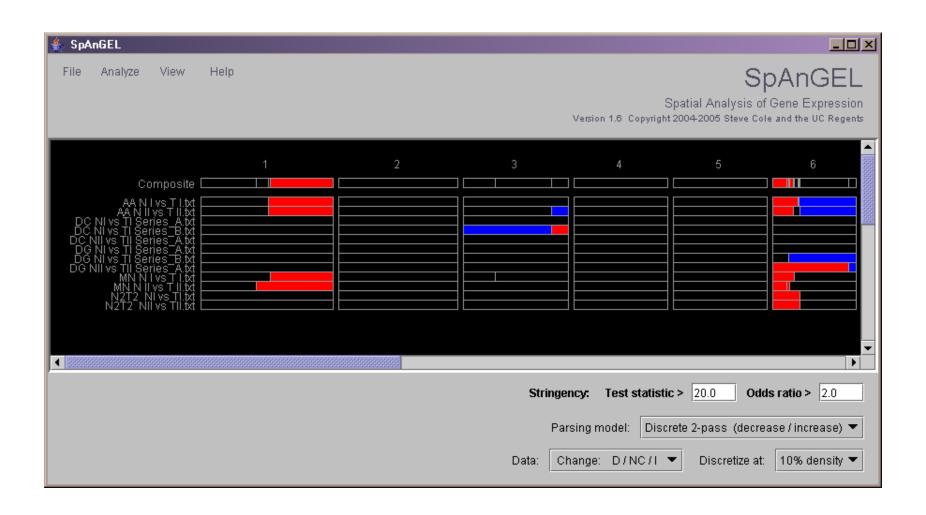




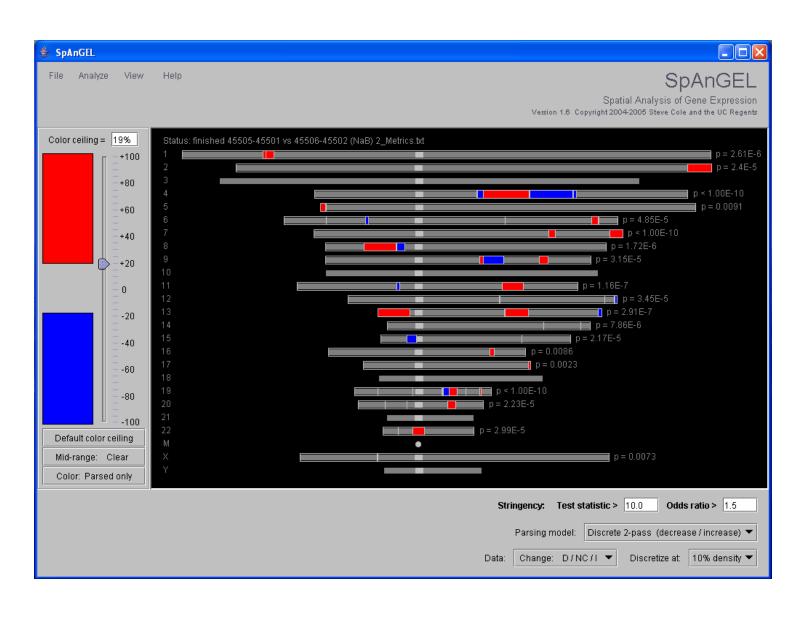


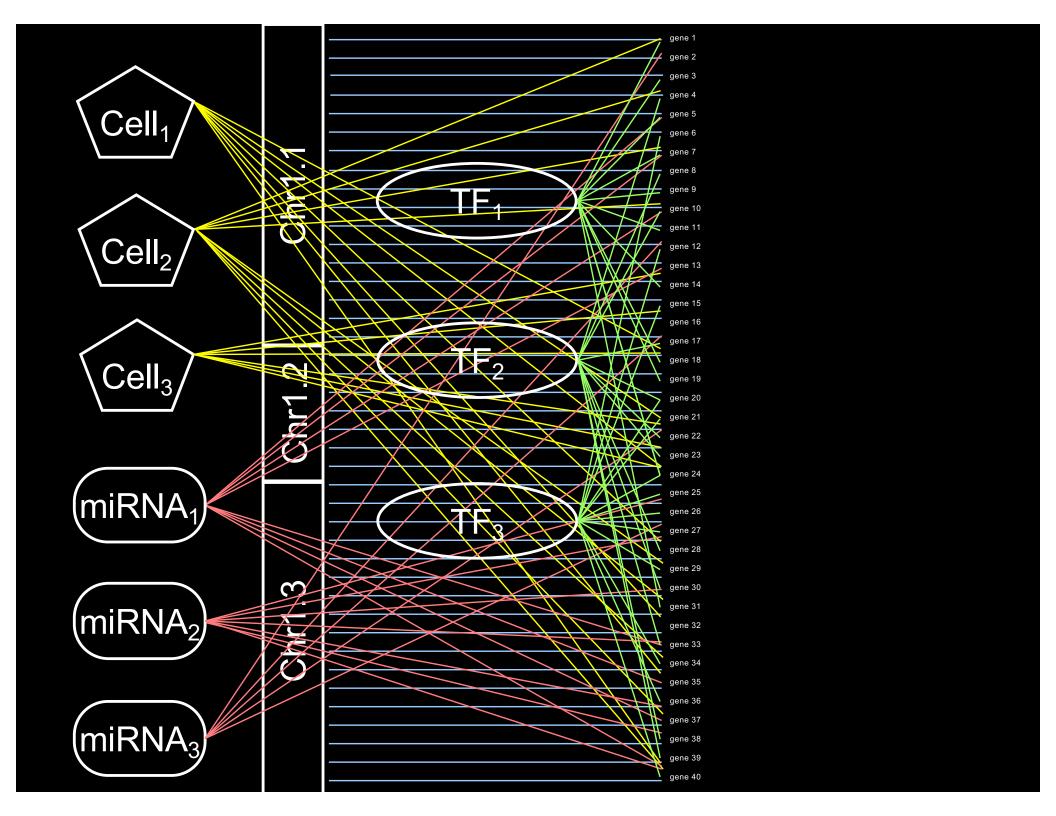


Chromosomal aberration profiles

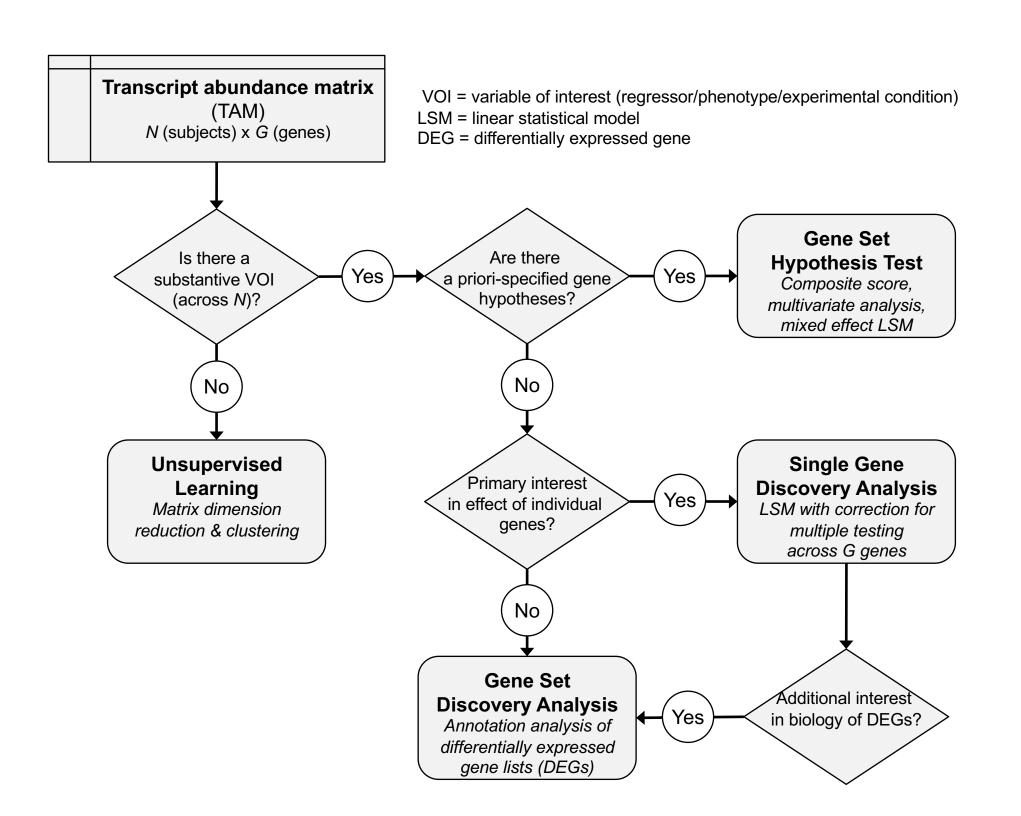


Epigenetics, chromatin accessibility, & RIDGEs



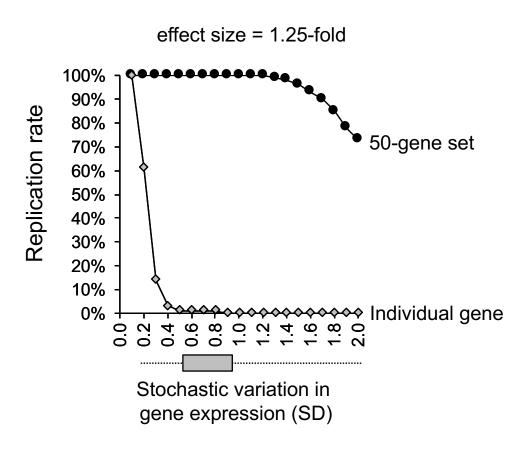






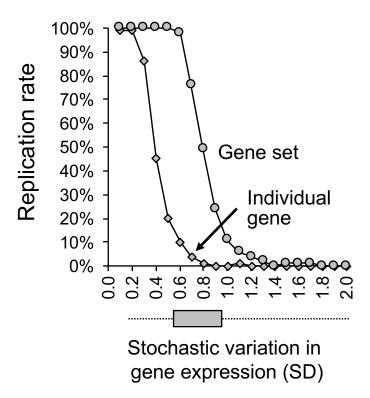
Painful lesson #1: If you want replicable results, focus on functionally related **sets** of genes (not individual gene transcripts)

Differential expression of an a priori gene set

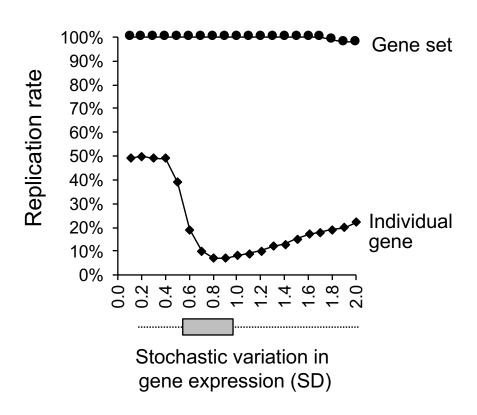


Painful lesson #2: If you want replicable results, conduct 2^{nd} stage bioinformatics on gene lists derived from point estimates of **effect size** (not p/q-values)

Gene set enrichment analysis Low level input: *p*-value gene list

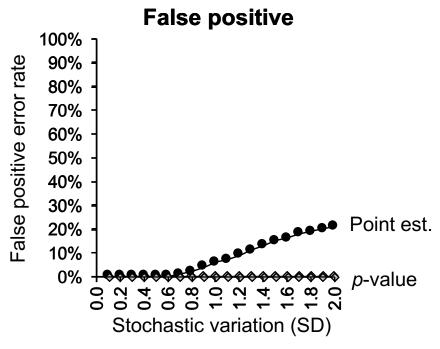


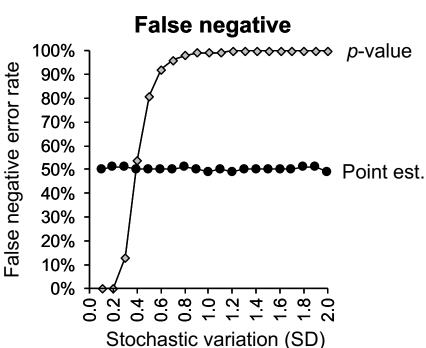
Gene set enrichment analysis Low level input: point estimate gene list

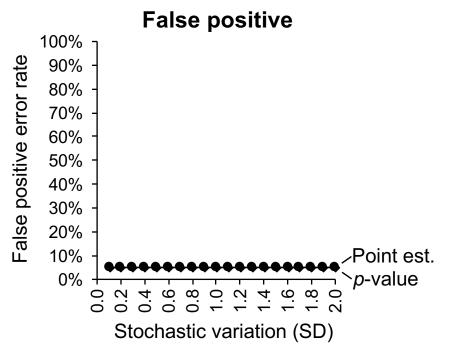


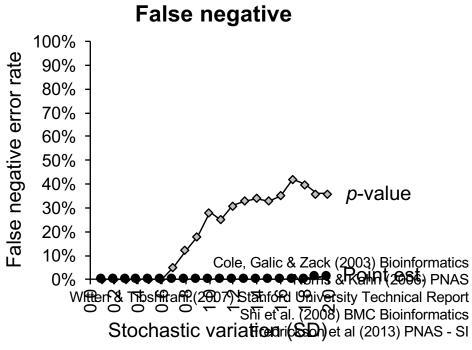
Individual gene association

Gene set association





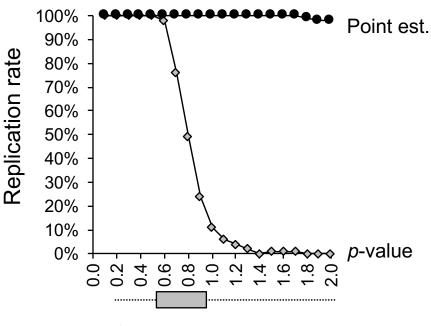




Implication of false negative errors for result replication

Gene set enrichment analysis

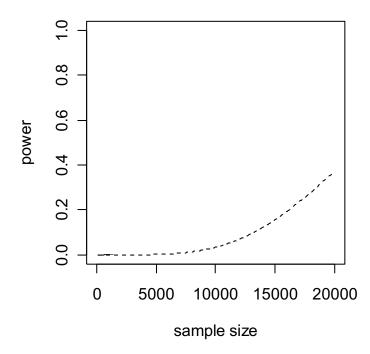
effect size = 1.5-fold



Stochastic variation in gene expression (SD)

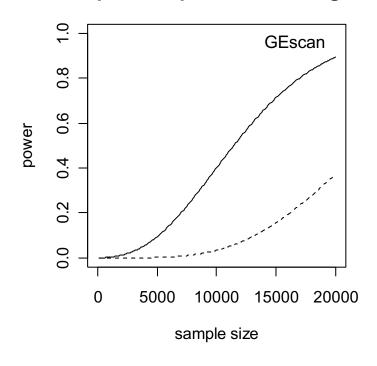
Cole, Galic & Zack (2003) Bioinformatics Norris & Kahn (2006) PNAS Witten & Tibshirani (2007) Stanford University Technical Report Shi et al (2008) BMC Bioinformatics Fredrickson et al (2013) PNAS - SI Painful lesson #3: Hypotheses are *extremely* valuable for statistical power in genomics. Don't pretend you know nothing (unless you really do).

Population prevalence design

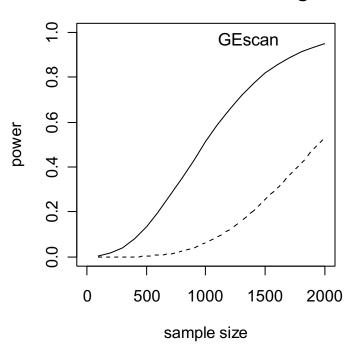


Painful lesson #3: Hypotheses are *extremely* valuable for statistical power in genomics. Don't pretend you know nothing (unless you really do).

Population prevalence design

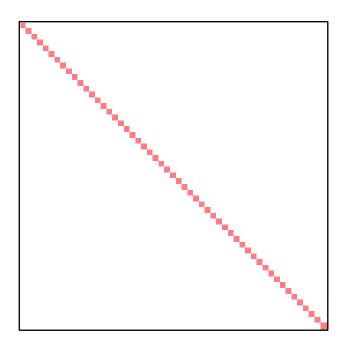


Outcome-stratified design

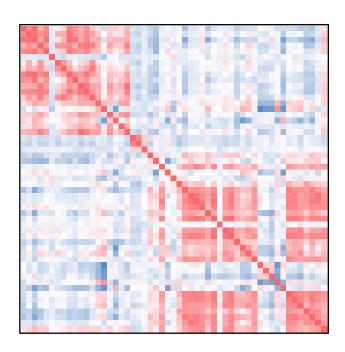


Painful lesson #4: Most *p*- and *q*-values are wrong (due to non-independence). Don't trust them.

Inter-gene correlation assuming independence



Empirical inter-gene correlation



Painful lesson #1: If you want replicable results, analyze functionally related **sets** of genes (not individual gene transcripts)

Sets derive from a biological model of mass action

(i.e., environmental conditions, pathogens, distinct cell types, differentiation states, TFs, chromosomal & epigenetic alterations, genetic polymorphisms, etc.)

Painful lesson #2: If you want replicable results, conduct interpretive bioinformatics on gene lists derived from point estimates of **effect size** (not p/q-values)

Effect sizes (not SNRs) reflect mass biological processes

(i.e., environmental conditions, pathogens, cells, differentiation states, TFs, chromosomal & epigenetic alterations, genetic polymorphisms, etc.)

Painful lesson #3: Hypotheses are *extremely* valuable for statistical power, and that is the limiting resource in genomics. Don't pretend you know nothing.

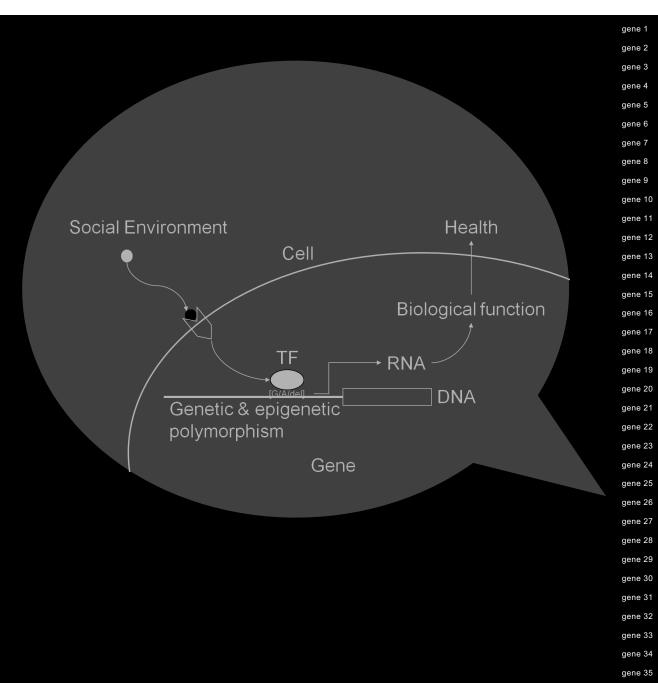
Hypotheses derive from a biological model

(i.e., or perhaps at least a set of ~100 imaginable models << 20,000 genes)

Painful lesson #4: p/q-values are inaccurate due to mass action correlation.

Model the biological processes that create the correlation

(work with the gene sets, not against them)



gene 36 gene 37 gene 38 gene 39 gene 40

