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**Title:** Collaborative Evaluation of *In silico* Predictions for High Throughput Toxicokinetics

Authors:

|  |  |  |
| --- | --- | --- |
| John Wambaugh | 1 EPA/ORD/CCTE | [Wambaugh.john@epa.gov](mailto:Wambaugh.john@epa.gov) |
| Nisha Sipes | 1 | [Sipes.nisha@epa.gov](mailto:Sipes.nisha@epa.gov) |
| Kamel Mansouri | 2 Innotiv  3 NICEATM | kamel.mansouri@nih.gov |
| Jon Arnot | 4 Arnot Research & Consulting | jon@arnotresearch.com |
| Trevor Brown | 4 | trevor.n.brown@gmail.com |
| Chris Cook | 1 | cookchristopher732@gmail.com |
| Daniel Dawson | 1 | ddawson@integral-corp.com |
| Sarah Davidson Fritz | 1 | DavidsonFritz.Sarah@epa.gov |
| John DiBella | 5 Simulations Plus | john.dibella@simulations-plus.com |
| Stephen Ferguson | 6 NIEHS | stephen.ferguson@nih.gov |
| Rocky Goldsmith | 1 | rgoldsmith@congruencetx.com |
| Richard Judson | 1 | Judson.Richard@epa.gov |
| Michael Lawless | 5 | mlawless@simulations-plus.com |
| Gilberto Padilla Mercado | 1 | PadillaMercado.Gilberto@epa.gov |
| Grace Patlewicz | 1 | Patlewicz.Grace@epa.gov |
| Ester Papa | 7 University of Insubria, Varese | ester.papa@uninsubria.it |
| Prachi Pradeep | 8 German Federal Institute for Risk Assessment (BfR) | prachipradeep.15@gmail.com |
| Alessandro Sangion | 2 | alessandro.sangion@mail.utoronto.ca |
| Risa Sayre | 1 | sayre.risa@epa.gov |
| Russell Thomas | 1 | Thomas.Russell@epa.gov |
| Rogelio Tornero-Velez | 1 | tornero-velez.rogelio@epa.gov |
| Barbara Wetmore | 1 | wetmore.barbara@epa.gov |
| Michael Devito | 1 | Devito.Michael@epa.gov |

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Four bullet point summary:

* Toxicokinetic (TK) information, such as elimination half-life (thalf, plotted below), is critical for understanding chemical risk
* Here we collected *in silico* (quantitative structure-property relationship, QSPR) predictions of key *in vitro* determinants of TK from several different models
* The models were evaluated for ability to reproduce *in vitro* and *In vivo* measurements of TK
* Overall, high throughput physiologically-based TK (PBTK) model performed similarly when using TK QSPRs as when the actual *in vitro* measured data were used

**One sentence description:** This collaborative trial demonstrates that multiple QSPRs exist that make reasonably accurate chemical structure-based predictions for *in vitro* TK parameters

**Chemicals Involved:** 87 ToxCast chemicals with diverse uses for which high-throughput toxicokinetic (HTTK) data are available

Collaborative Evaluation of *In silico* Predictions for High Throughput Toxicokinetics

John Wambaugh1,\*, Nisha Sipes1, Jon Arnot2, Trevor Brown2, Christopher Cook1,3, Daniel Dawson1, Sarah Davidson1, , John DiBella4, Stephen Ferguson5, Rocky Goldsmith1, Chris Grulke1, Richard Judson1, Michael Lawless4, Kamel Mansouri6, Grace Patlewicz1, Ester Papa7, Prachi Pradeep1,8, Alessandro Sangion2, Risa Sayre1, Russell Thomas1, Rogelio Tornero-Velez1, Barbara Wetmore1, and Michael Devito1

1. Center for Computational Toxicology and Exposure, Office of Research and Development, United States Environmental Protection Agency, Research Triangle Park, North Carolina 28311
2. ARC Arnot Research and Consulting Inc.
3. Oak Ridge Institute for Science and Education, Oak Ridge, Tennessee 38331
4. Simulations Plus, Inc.,
5. Division of the National Toxicology Program (NTP), National Institute of Environmental Health Sciences, National Institute of Environmental Health Sciences, Research Triangle Park, North Carolina 28317
6. NTP Interagency Center for the Evaluation of Alternative Toxicological Methods, National Institute of Environmental Health Sciences, Research Triangle Park, North Carolina 28317
7. Department of Theoretical and Applied Sciences, University of Insubria, Varese
8. German Federal Institute for Risk Assessment (BfR)

\*Corresponding Author: wambaugh.john@epa.gov

# Abstract (313 out of 250)

To assess public health risks posed by chemicals we need chemical-specific toxicokinetic (TK) data to understand chemical absorption, distribution, metabolism, and elimination by the body*.* High throughput TK (HTTK) methods have the potential to address data gaps. This collaborative trial uses a database of *in vivo* measured toxicokinetic data to evaluate *in silico* approaches for HTTK. Six different sets of quantitative structure-property-relationship (QSPR) tools were evaluated. The predicted parameter values were used within a high throughput physiologically based TK (PBTK) model to predict *in vivo* measured plasma concentrations for 92 chemicals, mostly in rats. Root mean squared log10 error (RMSLE) was calculated for toxicologically-relevant dosimetry statistics AUC (time-integrated area under the curve) and Cmax (peak concentration). Early time points, including Cmax, are driven by physicochemical properties and are insensitive to *in vitro* measured HTTK data. Cmax could be predicted with RMSLE 0.8-0.9 in contrast to 0.7 for *in vivo* fits. For AUC *in vivo* parameters gave RMSLE 0.5. Chemical-specific *in vitro* data had RMSLE 1.3 and QSPRs ranged from 1.3-1.4. *In vivo* parameters for the full concentration time-course data had RMSLE of 0.62. *In vitro* HTTK predicted gave RMSLE of 1.18. Greater discrimination between QSPRs was observed at later time points, which impacts AUC and is driven by estimated metabolism and elimination. QSPRs predict plasma binding well (RMSLE 0.03 – 0.07) but have difficulty predicting metabolic clearance (RMSLE 0.37 – 1.28). A consensus prediction using the maximum clearance predicted across all QSPRs predicted AUC with RMSLE 1.1 and the full time-course with RMSLE 1.1. The consensus predictions outperformed the *in vitro* measured data for the evaluation chemicals. For novel compounds a consensus QSPR approach may yield reasonable predictions of TK. This evaluation characterizes the accuracy of HTTK approaches for new chemicals based on both *in vitro* measurement and structure-based *in silico* predictions.

# Introduction

Toxicokinetics (TK) describes the absorption, distribution, metabolism, and excretion (ADME) of a chemical compound in the body as a function of time [1]. Since TK allows the prediction of internal tissue concentrations as a function of chemical exposure it provides critical information for assessing risk posed by a chemical to public health [2]. TK allows interpretation of biomonitoring data [3], dosimetric anchoring of animal toxicity studies [4], and quantitative *in vitro*-*in vivo* extrapolation (or IVIVE) from high throughput bioactivity studies [5]. These *in vitro* bioactivity data are available for thousands of chemicals (for example, the ToxCast [6] and Tox21 [7] screening programs). IVIVE of bioactivity data relies upon TK to relate the concentrations found to be active *in vitro* with doses that could cause these concentrations in tissues [8-12]. Unfortunately, chemical-specific information on TK are often unavailable for thousands of chemicals in commerce and the environment [13]. New approach methodologies (NAMs) are being developed throughput the chemical risk assessment process [14] including new methods for assessing TK [15,16].

For non-therapeutic compounds TK data were traditionally developed using animal studies; for reasons of both ethics and resources these studies are no longer desirable nor practical for the thousands of remaining chemicals [17]. An alternative technology developed by the pharmaceutical industry relies on characterizing certain aspects of TK *in vitro* and then extrapolating to *in vivo* conditions to estimate TK parameters such as AUC, often within a three-fold error [18]. For the past decade, government chemical regulatory agencies working with their collaborators and contractors have been collecting chemical-specific *in vitro* data that allow prediction of TK [8-12,19]. These data currently span approximately one thousand chemicals used for industry, pesticides, diet, therapy, and consumer products. However, thousands more remain uncharacterized.

The U.S. National Academies of Science, Engineering, and Mathematics have recognized that *in vitro* TK data “enabled first-tier risk-based rankings of chemicals on the basis of margins of exposure—the ratio of exposures that cause effects (or bioactivity) to measured or estimated human exposures” [20]. To address the remaining chemicals for which even *in vitro* TK data are unavailable, multiple organizations have developed *in silico* quantitative structure-property relationship (QSPR) models for predicting these values [21-25]. Meanwhile, EPA and other organizations are continuing to accumulate chemical-specific TK data, both *in vivo* – that is, curation of data from the scientific literature as well as a targeted animal studies only when needed [26,27] – as well as *in vitro* [11,19,12].

Here we examine the TK predictions from six different modeling teams. Four modeling teams produced QSPR models for two key toxicokinetic parameters that can be measured *in vitro*: intrinsic hepatic clearance (Clint measured with hepatocyte incubations, [28]) and fraction unbound in plasma (fup, typically measured via rapid equilibrium dialysis [29]). The models are initially evaluated for their ability to reproduce *in vitro* measured values. However, we focus on analysis of the predictions for chemical concentration as a function of time (CvT) that can be made when a generic physiologically-based TK (PBTK) model [30] is used with the predictions from each QSPR. For this study *in vivo* plasma and blood concentration vs. time data for rat and human were initially available for 101 chemicals from the CvTdb [27]. Models are evaluated for ability to reproduce the full CvT curve as well as summary statistics (such as peak plasma concentration) and parameters (such as chemical half-life). Two additional models for chemical half-life are also evaluated. The six modeling teams were provided with chemical identities and some physico-chemical descriptors but were not provided with the actual *in vivo* evaluation data.

# Methods

As summarized in Table 1, three levels of evaluation were made. First (Level 1), QSPR predictions of parameters were compared against *in vitro*-measured values for each chemical where data were available. Then (Level 2) the ability of a PBTK model to predict chemical concentration vs. time behavior was evaluated against *in vivo* measurements for the full time-course observed for each combination of QSPR and chemical. Finally (Level 3) the ability of the QSPRs and PBTK model to predict summary statistics (for example Cmax, AUC, half-life) was evaluated. All analyses were performed in the free, open-source statistical analysis language R [31] v4.4.0. Analyses were performed on a Dell Precision 7560 laptop personal computer. Scripts to perform all analyses are available in RMarkdown [32] format as supplemental material at <https://github.com/USEPA/CompTox-ExpoCast-HTTKQSPRs>.

## HTTK

R package “httk” [30] v2.3.2 was used for this analysis. All *in vitro* and *in vivo* data are made available through data tables released as part of this package. “httk” can parameterize a physiologically based toxicokinetic (PBTK) model based on chemical-specific values for fraction unbound in plasma (fup, unitless) and intrinsic hepatic clearance (Clint, µL/min/106 hepatocytes). These experimentally measured values are collected from peer-reviewed literature and provided by “httk”. All the QSPRs analyzed here were trained to human data.

The generic PBTK model used here (model “pbtk”) consists of well-mixed compartments for the gut, kidney, liver, and rest of body. The model is parameterized for a chemical using fup and Clint plus equilibrium tissue:plasma partition coefficients predicted with a modified Schmitt’s method [33,34]. The model simulates both oral and intravenous dosing. Oral dosing is subject to first-pass metabolism by the liver before the compound distributes systemically. Among other species, the model includes physiological information for parameterizing both humans and rats (primarily from [35,33,36]. The model was simulated using command “httk::solve\_pbtk()” with option default.to.human=TRUE – that is, since no rat-specific values are predicted by the models under evaluation, comparisons to data from rats were done using rat physiology but human *in vitro* TK parameters.

The steady-state ratio of the concentration of chemical in blood and plasma (Rb:p) is an important parameter in the PBTK model and is used for converting between plasma predictions and observations in whole blood. The function httk::get\_rblood2plasma() either retrieves measured values of Rb:p from the literature or predicts the ratio by predicting the red blood cell:plasma equilibrium partition coefficient and then using the hematocrit fraction for the relevant species.

R is an interpreted language (primarily operated by a user from the command-line although scripts are common). The user can alter the values within a table which stores the fup and Clint values for all chemicals (httk::chem.phys\_and\_invitro.data). After alteration, the httk function will proceed using the new values in the table. The HTTK data can be returned to their default values via the command “httk::reset\_httk()”. By default, no QSPR values are included in the table. However, predictions can be loaded with the commands “httk::load\_sipes2017(overwrite=TRUE)”, “httk::load\_pradeep2020(overwrite=TRUE)”, or “httk::load\_dawson2021(overwrite=TRUE)” [23-25]. The argument “overwrite=TRUE” is needed so that *in vitro* measured data are overwritten whenever a chemical-specific prediction is available. To facilitate comparisons, a custom function “clear\_httk()” is included in the supplemental material which deletes all Human Clint and fup values. The OPERA predictions are available as Supplemental Table 1.

## QSPR Models

The QSPR models evaluated are summarized in Table 2. Four different modeling teams previously [23-25] produced quantitative structure-property relationship (QSPR) models for two key toxicokinetic parameters that can be measured *in vitro*: intrinsic hepatic clearance (Clint measured with hepatocyte incubations) and fraction unbound in plasma (fup). The consensus QSPR was constructed for fup by taking the inverse logit of the mean of the logit-transformed fup from each QSPR. For Clint the maximum value predicted by any QSPR was used. Two additional models for chemical half-life were also evaluated. Individual model predictions are available in Supplemental Table 1.

Due to the potential presence of the model evaluation data (that is, measured *in vitro* parameters) in the training sets for some or all the models, we attempted to remove predictions that seemed more like a direct retrieval of the chemical-specific values from a training set. Model predictions were removed for a particular model-chemical combination if both the predictions for Clint and fup were within 1% absolute fold error of the measured values, the predictions were omitted from the evaluations. Omitted predictions are listed in Supplemental Table 6. When a QSPR model prediction was missing for a particular chemical the mean prediction of the other models was used for evaluation purposes.

QSAR training set summary data

* number chemicals, max, min, average values
* will use the averages for missing values

## *In vitro* Data

For 61 of the study chemicals *in vitro* measurements were also available for comparison. These data are collected by the R package “httk” but are drawn from the peer reviewed scientific literature (including [9,5,12]). It is likely that these data were in the training set of some of the QSPR models. The *in vitro* measured values are available in Supplemental Table 1.

## *In vivo* Data

EPA has developed a public database of concentration vs. time data for building, calibrating, and evaluating TK models [27]. Curation and development of the database are ongoing, but when this study began there were 101 chemicals with either rat or human *in vivo* blood or plasma concentration vs. time data. The *in vivo* measured concentration vs. time values are available as Supplemental Table 2.

## Compartmental Model Fits

For chemicals with CvT data, parameters were estimated for empirical one- and two-compartment toxicokinetic models using R package “invivoPKfit” (<https://github.com/USEPA/CompTox-ExpoCast-invivoPKfit>). Three models were considered: one- and two-compartment empirical TK models and a flat “null hypothesis” where there was no systematic change in concentration vs. time. The model with the lowest Akaike Information Criterion (AIC) value – indicating model parsimony – was selected [37]. Data sets where the flat model was selected were omitted from further analysis. The empirical model fit was then used as a “best case” prediction scenario for comparison with PBTK parameterize by *in vitro* or QSPR predictions.

For both models a half-life was calculated from the terminal elimination rate as thalf = ln(2)/kelim. For the two-compartment model the volume of distribution at steady-state was used as Vd. For both models, clearance was calculated as Cltot = Vd \* kelim. The estimated TK parameters for both models are provided as Supplemental Table 3.

## Evaluation Metrics

Multiple statistics were used to evaluate predictions (*pred*) relative to observed values (*obs*) as appropriate. Relative Predictive Error (RPE) was calculated as RPE =, where if the observed value was 0 then the error was set to zero. We note that if the predicted value is 0 then RPE = -1. Absolute Average Fold Error (AAFE) was calculated as AAFE = , where if pred=0 and obs=0 we assigned = 0. Root Mean Squared Log Error (RMSLE) was calculated as RMSLE = .

Kolmogorov-Smirnov tests were performed using R function *ks.test*.

# Results

## Evaluation Chemicals and Predictions

There are 101 chemicals present in the public release of CvTdb (Sayre, 2020) that had plasma concentration data following either oral or intravenous doses given to rats or humans. These chemicals included: 57 from the Toxic Substances Control Act (TSCA) active inventory [38], 20 pharmaceuticals, 24 pesticides, 99 that are found in consumer products, 7 per- and poly-fluorinated substances (PFAS) [39], and 64 that are part of the ToxCast screening program. Note that a chemical could be in more than one of these categories.

As illustrated in Figure 1, a typical TK concentration vs. time curve following a single oral dose consists of separate phases: 1) an initial increase driven by absorption of the chemical into the body, 2) a steep decrease driven by distribution of the chemical from the blood into tissues with varying affinities for the chemical, followed by 3) metabolism and excretion of the chemical (elimination) from the blood. Metabolism and excretion are present during the distribution phase, and that phase can be more or less pronounced depending on how much chemical the tissues can sequester. Following an intravenous dose there is no absorption phase, although there maybe a very short initial phase as the chemical is distributed from the site of injection to the systemic blood circulation. The different phases are more or les sensitive to the sorts of HTTK parameters that are predicted here, with the metabolism and elimination being the most relevant.

Data sets were modeled for their suitability to evaluate HTTK predictions by systematically optimizing parameters for one- and two-compartment models [40]. Of the 101 chemicals, four were best described by the “flat” model indicating that the data sets were too noisy to estimate empirical TK parameters:

Perfluorohexanoic acid, Perfluorooctanoic acid. 1,4-Dioxane, and Perfluorooctanesulfonate. Given that these chemicals were poorly described by basic TK models – potentially indicating problems with chemical analysis sensitivity -- these chemicals were withheld from subsequent analysis. Further, the reported limits of quantitation for C.I. Solvent Red 1 and Propylparaben were higher than the measured concentrations, so these data were also withheld. The data for each remaining chemical could be described using either a one- or two-compartment empirical pharmacokinetic models. There were 11 chemicals where we only had oral data, so that we could not estimate the parameters needed to make novel predictions but were well-described by an empirical TK model. Separate parameter estimates were made for each combination of compound and species for which there were data. For each remaining chemical the better of the one or two compartment models was used based on model parsimony.

## QSPR Predictions

The number of chemicals for which predictions could be made (that is, domain of applicability) varied from QSPR model to QSPR model. Throughout this effort we have reported statistics on different subsets of chemicals. There was only a subset of chemicals with predictions from representing an intersection of the models.

Additionally, HTTK *in vitro* data were not available for all the chemicals with CVT data, so evaluations aimed at characterizing the predictivity of *in vitro* HTTK only relied on 64 chemicals. Second Finally, statistics are reported for the maximal number of chemicals for which the QSPR could make predictions. The subset of chemicals with overlap between all models is more rigorous statistically, however the superset of all chemicals with predictions is more rigorous chemically.

For each QSPR we removed predictions where the predicted values for a given chemical were within 1% for both fup and Clint assuming these values reflected the chemical data present in the training set and the model method allowing for recall of the measurements. This affected 21 chemicals as predicted by OPERA (see Supplement Materials Table SupTable-PossibleTrainingChems.txt).

We summarize the chemical-specific properties and predictions in Figure 2. In Figure 2 similar chemicals (rows) and properties/predictions (columns) are clustered together based on Euclidean distance. All properties/predictions were centered (mean changed to zero) and scaled (divided by standard deviation) such that the value reflects the number of standard deviations from the mean. Interestingly, the first division between clusters in Figure 2 places all the Clint measurements and predictions on the one side and all the fup measurements and predictions on the other. The physico-chemical properties are divided between those two clusters, with Octanol:Water (partition coefficient, PC), Octanol:Air PC, Molecular Weight, Boiling Point, and Melting Point all clustering with Clint. Water solubility, vapor pressure, and the Henry’s law constant all clustered with fup.

## Level 1 Analysis

Our first level of evaluation directly compared the predictions of QSPR's with the *in vitro* measured values. Supplemental Table SupTable-QSPRPredCounts.txt gives the per QSPR number of chemicals that were made – the values ranged from a low of 43 for Clint for the Pradeep et al. (2020) QSPR to 51 for IVBP. For fup Pradeep et al. (2020) predicted a low of 38 and Simulations Plus covered a high of 45. Evaluation data were available for Clint for 64 chemicals and fup for 60. Table 3 summarizes the fold errors by parameter and QSPR. Note that there were several potential training chemicals that were removed from OPERA because its nearest-neighbor algorithm predicted the values too accurately.

We evaluate model performance for Clint in Figure 3. Note that the QSPRS models perform reasonably similarly despite that the Dawson (2021) model is categorical (that is, predicting only three values: very slow, slow, and fast) while the other models are continuous. Across the chemicals the RMSLE (root mean squared log10 error) for Clint predictions for the three QSPRs trained in part on chemicals from the ToxCast project (OPERA, Dawson (2021), Pradeep (2020)) were lower. The QSPRs trained more broadly, Simulations Plus ADMET predictor and IVBP had larger RSMLEs. IVBP tends to overestimate the *in vitro* measured clearance when it predicts that there is clearance. In Table 3 we summarize the fold errors for the five QSPRs. QSPRs not trained to ToxCast chemicals tended to overestimate the clearance measured *in vitro*. Also shown in Table 3 is that the predictions of Clint range from 100x lower than experimental values (log10 folder error of -2) to 1000x higher (log10 fold error of 3). Each QSPR’s predictions are compared separately to observations in Supplemental Figure 1.

Figure 4 shows that all four models perform very well for predicting fup. Most models are within ten-fold and predictions are highly correlated with observed. For fup, we note that the errors for OPERA are extremely low (effectively zero for fup), even with the obvious training set chemicals removed.

We examined the distributions of fold errors between the predictions and the measured data using a Kolmogorov-Smirnov test. For both Clint and fup the only QSPRs that differed from the others were OPERA and IVBP. Those two QSPRs which had a significant (p-value < 0.05) difference between the distribution of predicted values and the distributions of the three other QSPRs as well as each other.

## Level 2 Analysis

For the second level of analysis, we compared predictions based on the QSPR predicted values with actual tissue concentration vs. time data. We show all CvT curve fits and predictions on a per chemical, species, and route basis in the Supplemental materials (Supplemental Figures SupFig-ChembyQSPRCvTPlots.pdf).

All predicted values were used with a HTTK PBTK model to make predictions. We bracket QSPR model performance in three ways: First, we use the HTTK PBTK model with the actual *in vitro* measured values. In subsequent figures this is labeled as “HTTK-InVitro”. Next, for a best-case performance, we use empirical (one or two compartment toxicokinetic model) fits to the *in vivo* data, labelled as “*In vivo* Fits”. The one and two compartment models are simpler than the high throughput PBTK model used for all other scenarios, but because they have been optimized to the *in vivo* evaluation data itself, they are expected to outperform the other approaches here. Finally, for worst case performance we use y-randomization so that the measured values across all chemicals in the R package “httk” library are scrambled and assigned to the incorrect chemicals, labelled “HTTK-YRandom”.

Figure 5 shows the full predicted time-courses for each set of model predictions as well as the actual *in vitro* data and empirical model fits. Across the QSPRs the predictions tended to be within a factor of ten (indicated by the dashed lines) with a bias toward over-predicting at low concentrations. We see chemicals where there are vertical bars in Figure 5, indicating that the predicted concentrations were relatively constant (and low) over time while the observed concentrations changed. Typically, these are chemicals where the CvT time course was especially biphasic, with an initial rapid decline and then long tails where low levels of the chemical remained. In the tails the models tend to underestimate concentration, even for the *in vivo* fits.

Prediction error as characterized RMSLE is calculated first on a per chemical and method basis (error for predictions for a single method and a single chemical aggregating over all doses, routes, and time points). For each prediction method the mean RMSLE across all chemicals is reported in Table 4. Optimal performance is given by the empirical fits to the data. Worst case performance is given by the *y*-randomized measured data. Performance of HTTK using the *in vitro* measured parameters is closer to the *y*-randomization than the *in vivo* fits – RMSLE of 1.2 indicates slightly more than a factor of ten on average.

The QSPRs perform roughly equally, with RMSLE generally ranging between 1.1 and 1.3 in Table 4. Notably the QSPRs are close to the performance of the *in vitro* HTTK data. Remarkably, the consensus predictor (using the mean plasma binding and the maximal predicted metabolism) outperforms the *in vitro* HTTK data. Absolute Average Fold Error results are summarized in Supplemental Table SupTable-Level2-Cvt-AAFEstats.txt.

In Table 4 we note that error was roughly consistent regardless of the subset of chemicals used (that is, 1) those chemicals with *in vitro* HTTK data, 2) those without *in vitro* data, 3) those chemicals representing the intersection of all QSPR models, and 4) all chemicals predicted by a given method. For the subsequent analyses we report only the results for those chemicals with *in vitro* HTTK data in the manuscript, although results for the other subsets are available in Supplemental Material.

In Figure 6 we examine the distribution of per chemical RMSLE. In the first panel of Figure 6 all observed time points are valued equally, without consideration of phase (absorption/distribution/metabolism) and measurement accuracy. Here it is clear that the performance of HTPBTK with parameters for a random chemical, while worse, is not a marked departure from the performance HTPBTK with the correct parameters or a QSPR model. At early time points (second panel of Figure 6) all methods are more accurate than for all time points. This is consistent with Figure 1: The early absorption and distribution phases are dominated by the accuracy of volume of distribution (Vd). Prediction of Vd largely depends on physico-chemical properties (which have not been randomized) and weakly on fup. We note that the same absorption rate is used for all QSPR models (and indeed for all chemicals) based on the mean absorption observed across the chemicals profiled in Wambaugh et al. [26].

Also consistent with Figure 1, the most discriminating data for judging HTTK-based Cvt predictions depend on the later time points which characterize metabolism and excretion (the elimination phase of TK). In the third panel of Figure 6 we see that all predictors perform worse in the elimination phase, which is driven by the estimated Clint. For the late time points the specific values of the HTTK *in vitro* parameters (measured or predicted) have greater influence on the accuracy of the predictions – *y*-randomization performs notably worse than the QSPRs. The HTPBTK predictions based upon *in vitro*-measured HTTK data have an RMSLE which is indistinct from the QSPRs. Remarkably, the consensus QSPR predictions, using the most rapid predicted clearance, outperform the *in vitro* data, with an RMSLE of 1.1 across the evaluation chemicals.

In Figure 7 we plot each chemical-specific RMSLE by prediction method with each method as a row and each chemical as a column. These data are also provided in Supplemental Table SupTable-RMSLEbyChem.txt. Chemicals and methods have been clustered based upon Euclidean distance. White gaps mark chemicals that were not in the domain of applicability of different QSPRs (or, in the case of *in vitro* data, no measurements were available). We see that the consensus QSPR also gives the best coverage of chemicals, because the domain of applicability of the different QSPRs are varied.

## Level 3 Analysis

The third level evaluates prediction of TK summary parameters; specifically, peak concentration (Cmax), area under the plasma concentration time course (AUC), volume of distribution (Vd), half-life for elimination from the body (thalf), and whole-body clearance (Cltot). Where available, we compare the predictions to the values estimated from the empirical fits to the CvT data. The values predicted for each method are provided in Supplemental Table SupTable-Level3.txt.

Early time points are dominated by the ability to correctly predict peak plasma concentration (Cmax). In Table 5 we examine each methods accuracy in predicting Cmax as determined from the CvT data. While empirical fits are again best, predictions based on *in vitro* measured HTTK data are not that different from QSPR predictions. Y-randomization shows that Cmax is relatively insensitive to fup and Clint, with RMSLE of 1. That is, we do not see large differences between *in vivo* data, QSPRs, or y-randomization with respect to Cmax. As shown in Supplemental Figure 3, all the models tend to do a good job predicting Cmax greater than 1 mg/L but have a tendency to overestimate when Cmax is less than 1 mg/L. Cmax depends on Vd (for intravenous doses Cmax = dose / Vd). Given that the Cvt data were relatively insensitive to HTTK parameters at early time points (Figure 6), as shown in Table 6, y-randomization was much not worse for predicting Vd when compared with both *in vitro* data and QSPR-based predictions.

The most discriminating data for judging HTTK-based Cvt predictions depend on the later time points which characterize metabolism and elimination and inform metrics such as AUC. In Table 5 we examine the ability to predict time-integrated area under the plasma concentration time course (Area Under the Curve or AUC). Again, the empirical fits give a clear best-case scenario, but here the y-randomization more clearly gives a worst-case scenario. *In vitro* measured HTTK data predict AUC with an RMSLE of 1.3, while the QSPRs range from RMSLE 1.3 to 1.5. The consensus QSPR predictions were the best model, with an RMSLE of 1.1.

In Table 6, we also examine two quantities that inform our ability to predict AUC at late time points -- thalf and CLtot examine predicted vs. observed thalf. Most of the models and the *in vitro* measured data were unsuccessful for predicting thalf, with RMSLE indicating errors larger than 100x. Interestingly the y-randomization outperforms the models. Both the QSARINS and IFS-QAPR models are good, predicting thalf within a factor of 10x. The consensus model is improved over any of the other models, with a RMSLE of 1.7 (50x error). Finally, in Table 6 we examine predictions for CLtot, which depends on both elimination rate (inverse of thalf) and Vd. Both IFS-QSAR and QSARINS-Chem again performed best. The other QSPRs performed about as well as using the *in vitro* measured data.

# Discussion

To understand chemical risk to human health TK information are needed, including half-life (thalf) and toxicological dose metrics like peak concentration (Cmax) and time-integrated plasma concentration (AUC). Unfortunately, TK is chemical-specific, requiring some sort of method tailored on a per chemical basis. Large TK data gaps exist for analytes identified in biological matrices by both targeted (NHANES [41]) and non-targeted analyses (Woodruff [42]). Multiple governments have recognized [43-46] that high throughput (chemical-agnostic) TK models parameterized with chemical-specific *in vitro* data are a powerful tool for facilitating next generation risk assessment based on *in vitro* screening for chemical toxicity. Governments and industry are continuing to accumulate chemical-specific TK data including both *in vivo* concentration vs. time data in key tissues [27,26] and *in vitro* HTTK data [8,9,11,12,10,19]. However, several thousand chemicals remain in need of TK info; the QSPRs for HTTK evaluated here provide options to fill this gap.

Since the *in vitro* data for HTTK are limited in number, here we have conducted a collaborative trial of four QSPRs for *in vitro* TK parameters and two additional predictors of *in vivo* TK half-life. The focus of the analysis was using *in vivo* data from rat or human TK studies collected by CvTdb [27]. We aimed to characterize the accuracy of HTTK approaches for new chemicals based on structure-based *in silico* predictions. For novel compounds a consensus quantitative structure-property-relationship (QSPR) approach may yield a reasonable result. This collaborative trial used a database of *in vivo* measured toxicokinetic data to evaluate *in silico* approaches. *In vivo* data had to be carefully reviewed to include only those data that could be well-described by empirical TK models. Six different sets of QSPR tools for predicting TK were evaluated. Four of the QSPR models made predictions for chemical-specific *in vitro* measurements (HTTK data). An additional consensus prediction was constructed from the various QSPRs. By comparing predictions with observations, the root mean squared log10 error (RMSLE) and other key statistics could be calculated. The RMSLE characterizes the expected accuracy for new predictions. Given sufficient observations for evaluation, the RMSLE can be interpreted as a coefficient of variation for normally distributed errors about the prediction. For normally distributed errors one has 95% confidence that the actual value will occur within +- 2 RMSLE of the prediction.

Throughout this effort we have reported statistics on different subsets of chemicals. First, HTTK *in vitro* data were not available for all the chemicals with CVT data. For the context of decision makers relying either on *in vitro* HTTK or QSPR-based HTTK we report chiefly on the statistics associated with either the *in vitro* only chemicals or the chemicals with no existing *in vitro* data.

QSPRs were first evaluated for their ability to predict the chemical specific HTTK parameters Clint and fup that are expected by high throughput PBTK models. These *in vitro* parameters, of course, are not true TK. Even if QSPR perfectly reproduces an *in vitro* parameter, it is only as good as the *in vitro* assays. However, HTTK assays provide rapid methods for partially characterizing TK. [18,15]. There is an ongoing proliferation of high throughput PBTK models developed to make use of these HTTK *in vitro* parameters to allow HT-PBTK models to make chemical-specific predictions [47,30,15,48,49]. Most QSPRs predict plasma binding well (RMSLE 0.03 – 0.07) but have difficulty predicting metabolic clearance (RMSLE 0.37 – 1.28).

Predictions for the full concentration time-course were then evaluated. Our “level 2” evaluation used the QSPR predicted parameter values within a physiologically based TK (PBTK) model to predict the full TK chemical concentration time course. TK in the absorption and distribution phases are relatively insensitive to the *in vitro* HTTK parameters Clint and fup. For example, distribution depend largely on physico-chemical properties and only somewhat on fup for accurate prediction of equilibrium tissue partition coefficients (and, in turn, volume of distribution) [34]. However, the elimination phase of a TK time course is dominated by metabolism and excretion, which are both characterized to some extent by HTTK *in vitro* parameters.

To frame the level 2 evaluation, statistics were also calculated for empirical TK models using parameter optimized using the *in vivo* data. The *in vivo* fits were intended to approximate a best case given that the data themselves are noisy. As a worst-case scenario HTTK based on randomly selected (incorrect) *in vitro* HTTK parameters, while keeping the physico-chemical parameters correct. The y-randomization approximates helps account for potential correlations within the *in vitro* measured chemical data. During early time points (absorption and distribution) the PBTK models parameterized with QSPRs perform closer to y-randomized predictions than to empirical fits to the data. However, at later time points (elimination phase) the separation between QSPRs and y-randomized predictions grew.

Although nearly 100 chemicals is substantial, the existing annotated TK data do not constitute machine learning “Big Data” which might rely on thousands if not millions of observations [50]. While more than one thousand chemicals have available *in vitro* HTTK measurements for QSPR modeling, it is still a constrained random set of chemicals reflecting the correlation and the property distribution of the set. For example, to be suitable for *in vitro* measurement the volatility and solubility of the chemicals must be somewhat constrained. When chemical space is too narrow the overall statistics are limited; with the CvTdb, like any data set, one can only evaluate and model things that vary across our dataset. Among the evaluation chemicals only two had metabolic clearance (Clint above 103 µL/min/106 hepatocytes and 94% are within two-fold of the median. Four out of 64 chemicals with CvT data have no measured *in vitro* clearance compared with 254 out of 1023 measured Clint values (25 percent). The two parameters fup and Clint further interact in how they influence TK; if a chemical has low metabolic clearance, it may accumulate regardless of how highly the chemical binds; conversely if a chemical is highly bound it may not matter how fast the free chemical clears.

Here we have found the HTTK PBTK model performed similarly when using TK QSPRs for Clint and fup as when the actual *in vitro* measured data were used. Fits to the *in vivo* data had RMSLE of 0.62, while predictions based on *in vitro* HTTK had RMSLE of 1.18 without using any *in vivo* data. RMSLE for individual QSPRs ranged from 1.19 to 1.3. Any one QSPR performed slightly better than using *in vitro* values for random chemicals (RMSLE 1.35). A consensus prediction using the maximum clearance predicted across all QSPRs had RMSLE 1.09 for the full time course – this is better than using the *in vitro* measured data for the evaluation chemicals. For example, the IVBP model tends to over predict *in vitro* clearance, moving predicted clearance rates higher (correctly as it turns out).

QSPR models for metabolism are limited by many factors, including the limitations of the *in vitro* intrinsic clearance model. Since the majority of the *in vitro* Clint assays is based upon hepatocytes suspended in media and losing viability over a four-hour measurement [28,8], it in unsurprising that the *in vitro* assays underestimate clearance. We currently cannot evaluate the impacts of three-dimensional aspects of chemical structures (including chirality) – all the QSPR models are based on two-dimensional structure descriptors because the data do not exist to train the models otherwise (for example chiral pairs). Further, the data used to train the models are based on human biological material, but the data used for evaluation here are largely drawn from rat, again due to the much wider availability of human than rat *in vitro* HTTK data [51,52].

Geci et al. [53] recently evaluated HT-PBTK using a large data set of chemical concentration vs. time course data (including the CvTdb) for which AUC and Cmax were estimated via non-compartmental analysis. They found that the error for HT-PBK parametrized with *in vitro* data was two-fold for Cmax and 1.8-fold for AUC. With *in silico* predictors the best that could be achieved was 2.2-fold for Cmax and 2.4-fold for AUC. Here we observe that empirical one- or two-compartment fits to the Cvt data can predicted Cmax with RMSLE 0.65 (4.5-fold) and AUC with RMSLE 0.58 (3.8-fold) (Table 5, [40]). A key factor considered by Geci et al. [53] that was not considered here was intestinal permeability, however given that the majority of the data analyzed here were for rat and there is little reason to expected concordance between rat and human oral absorption Musther et al. [54] the source of the discrepancy is unclear. Geci et al. [53] reports the median, rather than mean error for their results, and for non-normally distributed results (such as log-normal, with a long “tail” of larger values) the mean can be much larger than the median. We have focused on the RMSLE here because *in vivo* data are often-log-normally distributed and RMSLE provides an estimate of the prediction standard deviation. In model evaluation literature there is significant discussion of the appropriate model evaluation stastistic, with mean absolute error (MAE) often preferred for scaling with the magnitude of the prediction (errors are weighted equally) while root mean squared error (RMSE) being preferred for indicating the standard deviation for normally distributed predictions. {Willmott, 2005; Chai, 2014}. RMSLE provides the advantageous of both, by scaling with the magnitude of prediction (because the data are logarithmic) and allowing statistical interpretation as the coefficient of variation.

Geci et al. [53] is consistent with Wang [18] which found that the SimCYP model, parameterized with *in vitro* data, could predict AUC within 2.3-fold when evaluated across 54 pharmaceuticals. Wambaugh et al. [26] found evaluated HTTK-based IVIVE for TK using just over forty chemicals including pharmaceuticals and non-pharmaceuticals. Wambaugh et al. [26] found Cmax could be predicted with RMSLE 2.2 and that AUC could be predicted with RMSLE 2.0. As summarized in Table 5, here we found that Cmax could be predicted with RMSLE 0.83 – equivalent to 6.8-fold. AUC can be predicted with RMSLE 1.3 – equivalent to 20-fold. We speculate that differences relatively to the literature are due to 1) curation of the CvT data by requiring that they can be fit to empirical models, [53] 2) the inclusion of many non-pharmaceuticals [18], and 3) the inclusion of more chemicals [26]. By design the ADME properties of pharmaceuticals are often constrained to have shorter half-lives than occur for non-pharmaceuticals – this means that the distribution of measured time points may be more sparse for non-pharmaceuticals {Padilla, 2024}.

Among the summary TK statistics (level 3) evaluated, the related quantities AUC, CLtot, and half-life are more challenging to predict than Cmax and Vd. Vd only depends on partitioning (partially characterized by fup) and Cmax only depends on Vd; neither of these two quantities depend upon Clint. For AUC, optimized fits to the *in vivo* data indicated a RMSLE of 0.5 while using *in vitro* values for random (incorrect) chemicals had an AUC RMSLE of 0.89. Using chemical-specific *in vitro* data predicted AUC with RMSLE 0.74, while QSPRs ranged in accuracy from 0.74 to 0.84. The consensus prediction using the maximum clearance predicted across all QSPRs predicted AUC with RMSLE 1.03 – this is better than using the *in vitro* measured data for the evaluation chemicals. For total clearance CLtot the RMSLEis 1.57. For Cmax *in vivo* fits gave RMSLE 0.5 and random chemicals gave 1.7. Chemical-specific *in vitro* data had RMSLE 1.3 and QSPRs ranged from 1.3-1.4. Vd had RMSLE 0.89.

For a novel chemical it may be safer to assume that the prediction errors will be closer to the conservative values estimated here. This is supported by the observed accuracy for the comparing the predicted full concentration vs. time curves against the observations, which are similar accurate to the predictions of summary TK statistics Cmax and AUC that have been analyzed elsewhere. We do not necessarily think that the improved ability of consensus modeling is sole influence of any one model, rather that consensus QSPRs represent the “wisdom of the crowd” [55,56]. By using the maximum predicted clearance, if any one QSPR predicts clearance then chemical may be moe likely to be metabolized. Used in this manner, we hope these QSPRs will enable public health risk-based prioritization of many more chemicals in commerce and the environment than *in vivo* and *in vitro* testing alone.

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# Conflict of Interest

Please declare any COI here

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# Figures

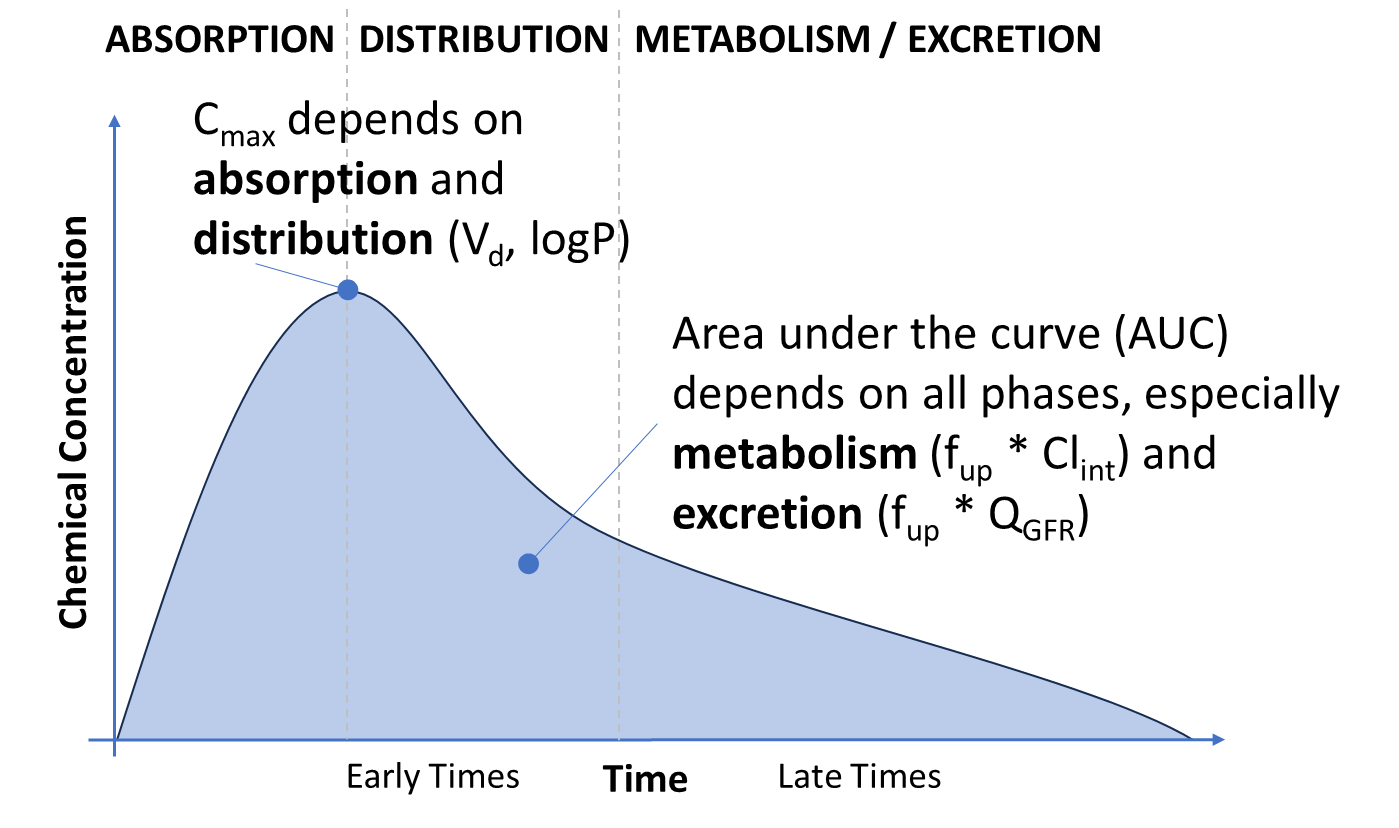


Figure : Conceptual illustration of toxicokinetic concentration vs. time (CvT) data.

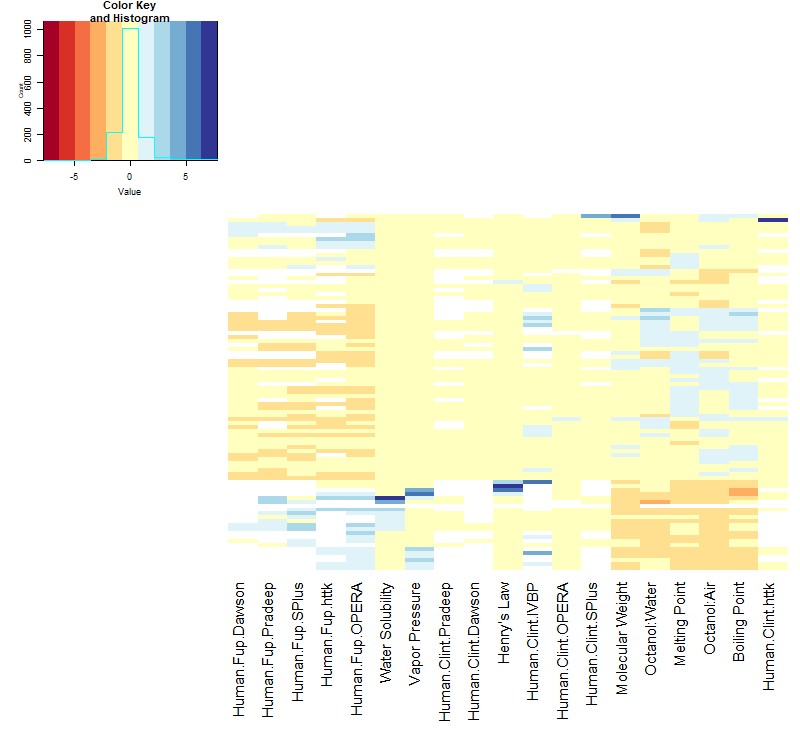


Figure : Columns in this heatmap indicate the physico chemical properties and measured/predicted values for in vitro TK (intrinsic hepatic clearance, Clint, and fraction unbound in plasma, fup). Each row corresponds to one of the 83 chemicals. The in vitro TK measurements (“Human.Clint.InVitro” and “Human.Fup.InvItro”) and predictions for these values from the various QSPRs (Table 2) are indicated by name. Data are normalized on a per column basis by centering (subtracting the mean) and scaling (by standard deviation). Thus, the "Value" of each entry in heatmap indicates the number of standard deviations from the mean. Blank values indicate no prediction.

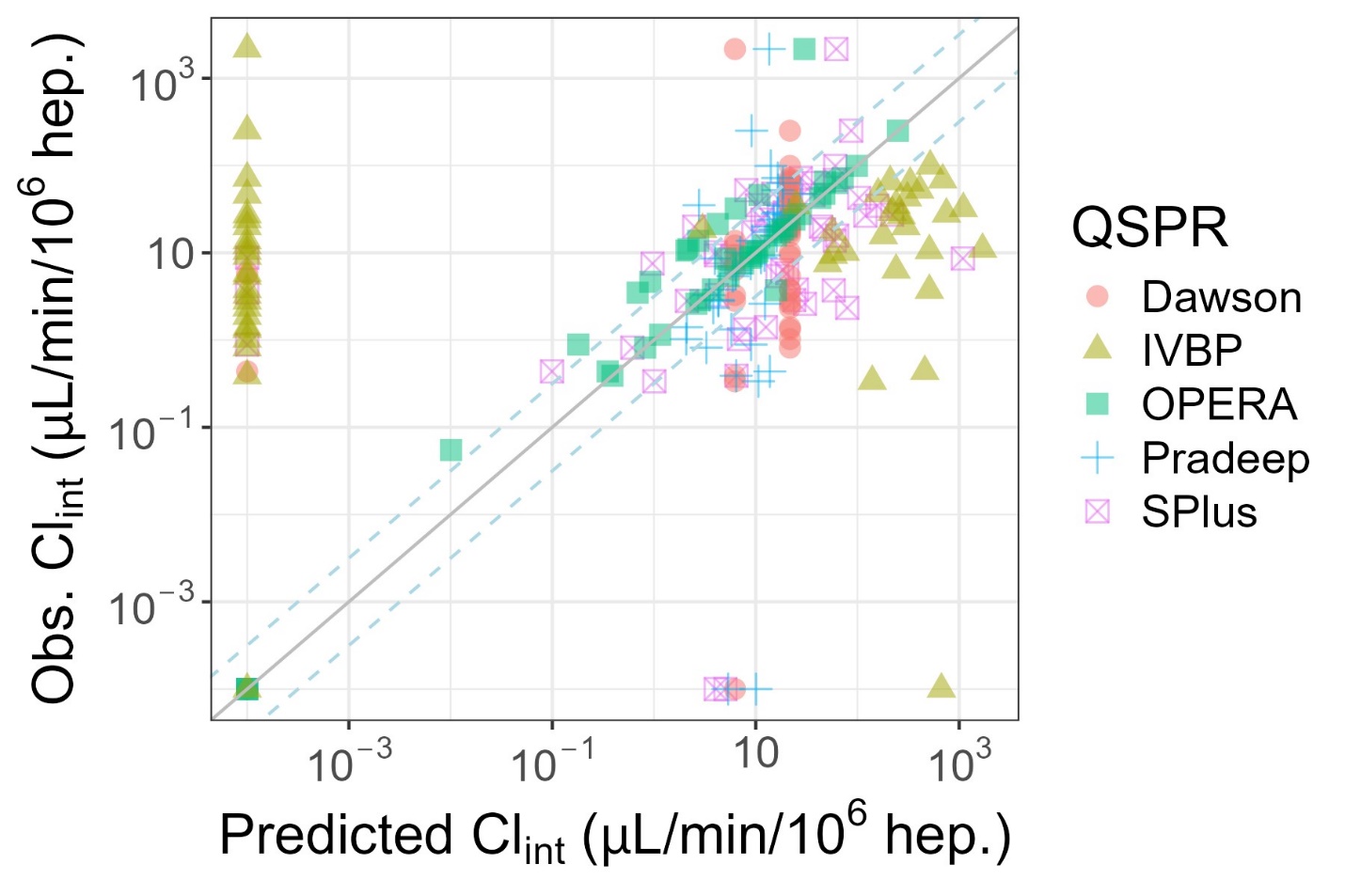
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Figure : Evaluation of Predictions for Intrinsic Hepatic Clearance (Clint). Zero values were plotted at 10-1, the solid line indicates identity (1:1) while the dashed lines indicate 3.2-fold difference.

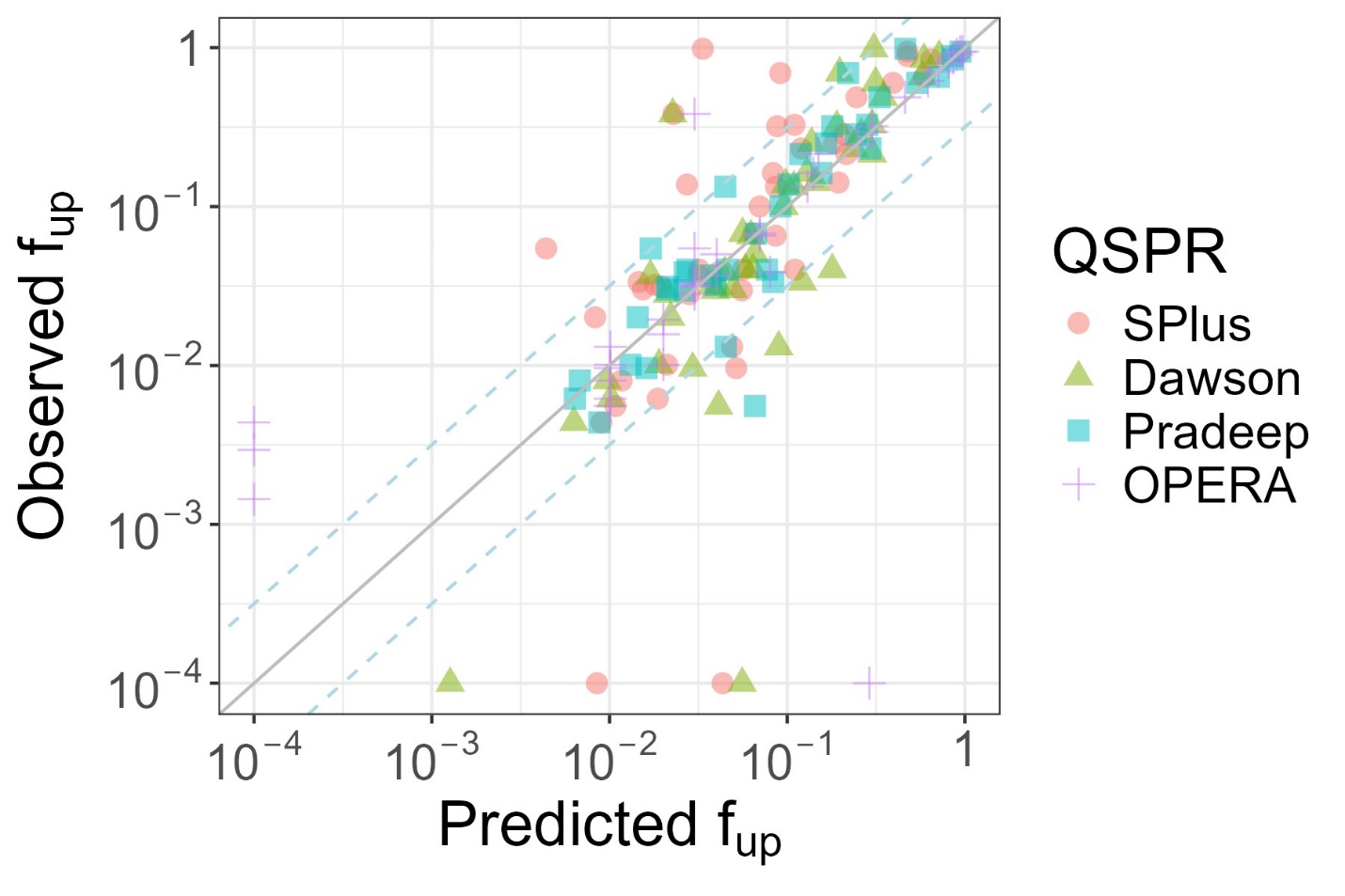


Figure : Evaluation of Predictions for Fraction Unbound in Plasma (fup). Zero values were plotted at 10-4, the solid line indicates identity (1:1) while the dashed lines indicate 3.2-fold difference.

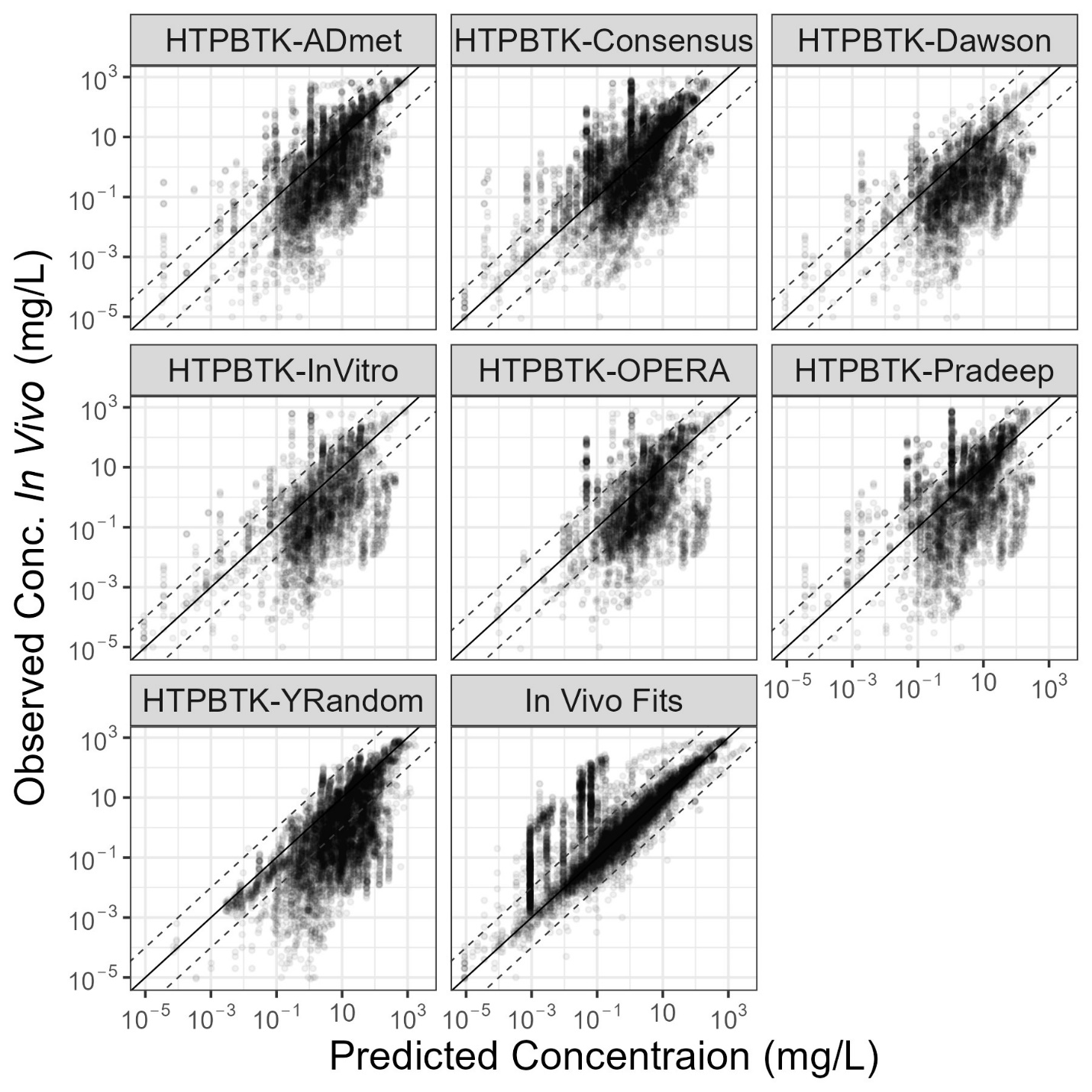


Figure : Comparison of in vivo measured chemical concentration vs. time (CvT) data [27] vs. predictions for empirical models fit to the data (“In vivo Fits”), and predictions for a PBTK model (“HTTK”) parameterized with chemical specific values either measured in vitro (“HTTK-InVitro”) or predicted with various QSPRs. In each sub-plot the y-axis shows the measured data while the x-axis shows the predictions made using chemical-specific parameters from the various sources. The solid line indicates identity (1:1) while the dashed lines indicate ten-fold difference.

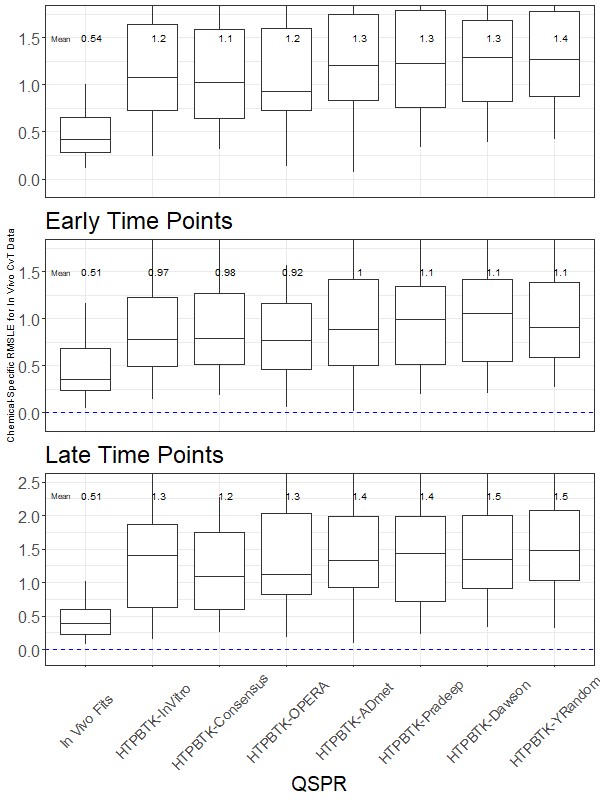


Figure : Chemical-Specific RMSLE for In vivo CvT Data. The upper and lower extent of the box for each model indicates the 25th to 75th quantiles, the mid-line indicates the median (50th quantile) and vertical line indicates 1.5x the range of the box.

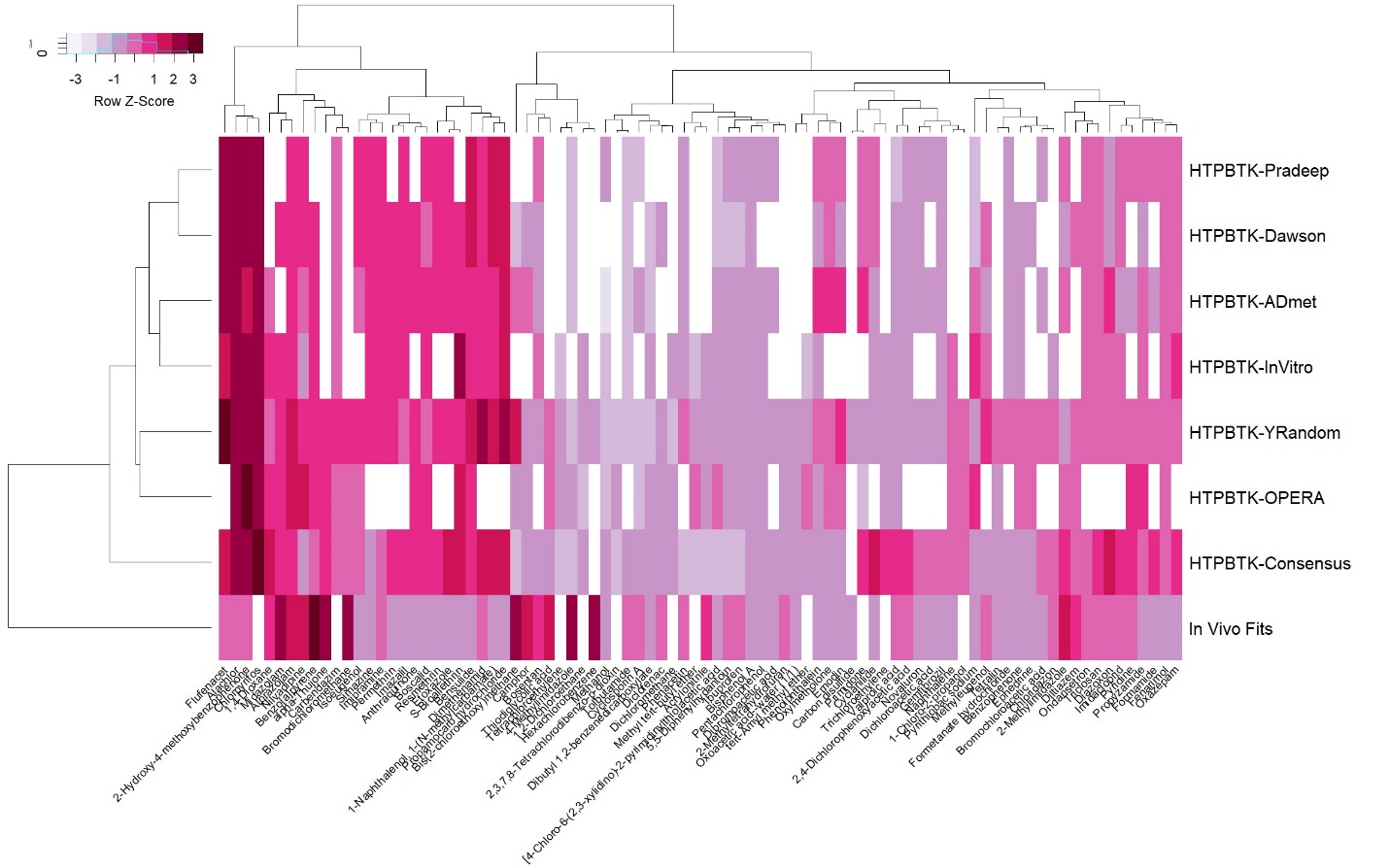


Figure : Values in this heatmap are the root mean squared log10 error (RMSLE) by chemical and predictor (that is, lighter indicates better predictive ability). The columns indicate different evaluation chemicals. The rows give the different prediction methods: The empirical fits to the data are given by “In vivo Fits”. All other values are calculated using the HTTK PBTK model and either measured values “In vitro”, y-randomized measured values (“Y-Random”) or the various QSPRS.

# Tables

Table Three Levels of Evaluation were performed

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Evaluation | TK Quantities | Chemicals for Evaluation | QSPR Models Evaluated | Reference |
| Level 1 | *In vitro* TK Measurements  (fup, Clint) | 64 with Measured *In vitro* Data | 4 (fup),  5 (Clint) | [30] |
| Level 2 | TK Concentration vs. Time  (all points, Cmax, time-integral/AUC) | 88 with predictions across multiple QSPRs and empirical model fits | 5 + Consensus | [27] |
| Level 3 | Summary Statistics  (Vd, thalf, Cltot) | 88 | 6 | [27] |

Table QSPR Models Evaluated

|  |  |  |  |
| --- | --- | --- | --- |
| Model | Predictions | Mechanism | Reference |
| Simulations Plus ADMET Predictor® | Level 1  (*in vitro* parameters) | Sum of CYP-specific Artificial Neural Network (ANN) | [23] |
| Pradeep 2020 | Level 1 | Random forest and support vectors method | [24] |
| Dawson 2021 | Level 1 | Random forest, clearance organized by categories | [25] |
| OPERA | Level 1 | Nearest-neighbors | [57,58] |
| IFS-QSAR | Level 3  (Half-lives) | Fragment-based Multiple Linear Regressors (MRL) | [21] |
| QSARINS-Chem | Level 3 | Ordinary Least Squares MLR | [22] |

Table : Biases of the QSPRs for predicting in vitro measured values in terms of log10 fold error (FE) – no bias would be FE = 0

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
|  | Num  Clint  Compared | Median  Clint  AbsFE | Median  Clint  FE | Num  Fup  Compared | Median  fup  AbsFE | Median  Fup  FE |
| SPlus | 50 | 0.575 | 0.0239 | 45 | 0.287 | -0.116 |
| Dawson | 48 | 0.362 | -0.0245 | 43 | 0.15 | 0.0115 |
| Pradeep | 43 | 0.232 | 0.00985 | 38 | 0.137 | -0.0429 |
| OPERA | 43 | 0.00144 | -0.00014 | 38 | 0.0336 | 1.09E-05 |
| IVBP | 51 | 2.62 | 0.507 | 0 |  |  |

Table Level 2 Analysis of predictions based on empirical model fits and a PBPK model (“HTTK”) parameterized with chemical specific values either measured in vitro (“HTTK-InVitro”) or predicted with various QSPRs. RMSLE was first calculated on a per chemical basis and then averaged across chemicals. The number of chemicals in each calculation is given in parentheses. The four sets of chemicals refer to those with in vitro HTTK parameters (“In vitro”), those without in vitro HTTK parameters (“No In vitro”), those chemicals for which all QSPRs could make predictions, and the maximum number of predictions available for each chemical. “HTTK-YRandom” indicates the HTTK PBPK model parameterized with a random permutation of the in vitro data.

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | HTPBTK-InVitro | HTPBTK-ADmet | HTPBTK-Dawson | HTPBTK-Pradeep | HTPBTK-OPERA | HTPBTK-Consensus | HTPBTK-YRandom | *In vivo* Fits |
| *In vitro* | 1.2 (55) | 1.3 (45) | 1.3 (43) | 1.3 (38) | 1.2 (33) | 1.1 (55) | 1.4 (55) | 0.54 (47) |
| No *In vitro* |  | 1.2 (18) | 1.1 (15) | 1.1 (15) | 0.85 (22) | 0.95 (29) | 1.2 (31) | 0.88 (33) |
| QSPR Intersection | 1.4 (17) | 1.4 (23) | 1.3 (23) | 1.3 (23) | 1.3 (23) | 1.1 (23) | 1.4 (23) | 0.58 (20) |
| Maximal | 1.2 (55) | 1.3 (63) | 1.3 (58) | 1.3 (53) | 1.1 (55) | 1.1 (84) | 1.3 (86) | 0.68 (80) |

Table Level 3 Analysis of predictions based on empirical model fits and a PBPK model (“HTTK”) parameterized with chemical specific values either measured in vitro (“HTTK-InVitro”) or predicted with various QSPRs. Both the coefficient of determination (R2) and root mean squared error (RMSLE) are calculated on a log10 scale. “HTTK-YRandom” indicates the HTTK PBPK model parameterized with a random permutation of the in vitro data.

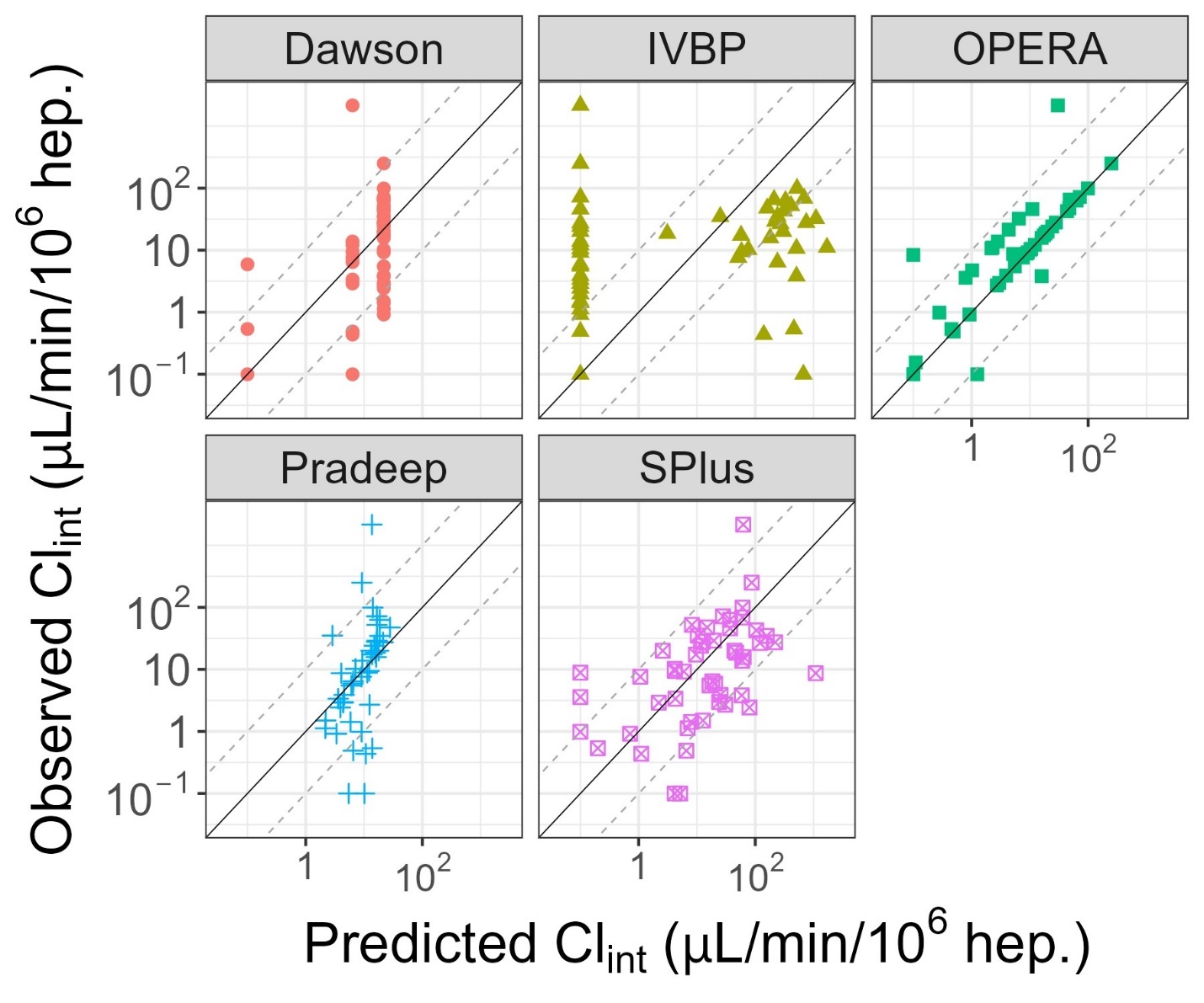
|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | HTPBTK-InVitro | HTPBTK-ADmet | HTPBTK-Dawson | HTPBTK-Pradeep | HTPBTK-OPERA | HTPBTK-Consensus | HTPBTK-YRandom | In vivo Fits |
| Cmax RMSLE | 0.85 (55) | 0.92 (45) | 0.99 (43) | 0.98 (38) | 0.79 (33) | 0.87 (55) | 1 (55) | 0.56 (47) |
| AUC RMSLE | 1.3 (55) | 1.4 (45) | 1.5 (43) | 1.5 (38) | 1.3 (33) | 1.1 (55) | 1.9 (55) | 0.46 (47) |

Table Analysis of predictions based on empirical model fits and a PBPK model (“HTTK”) parameterized with chemical specific values either measured in vitro (“HTTK-InVitro”) or predicted with various QSPRs. Both the coefficient of determination (R2) and root mean squared error (RMSLE) are calculated on a log10 scale. “HTTK-YRandom” indicates the HTTK PBPK model parameterized with a random permutation of the in vitro data.

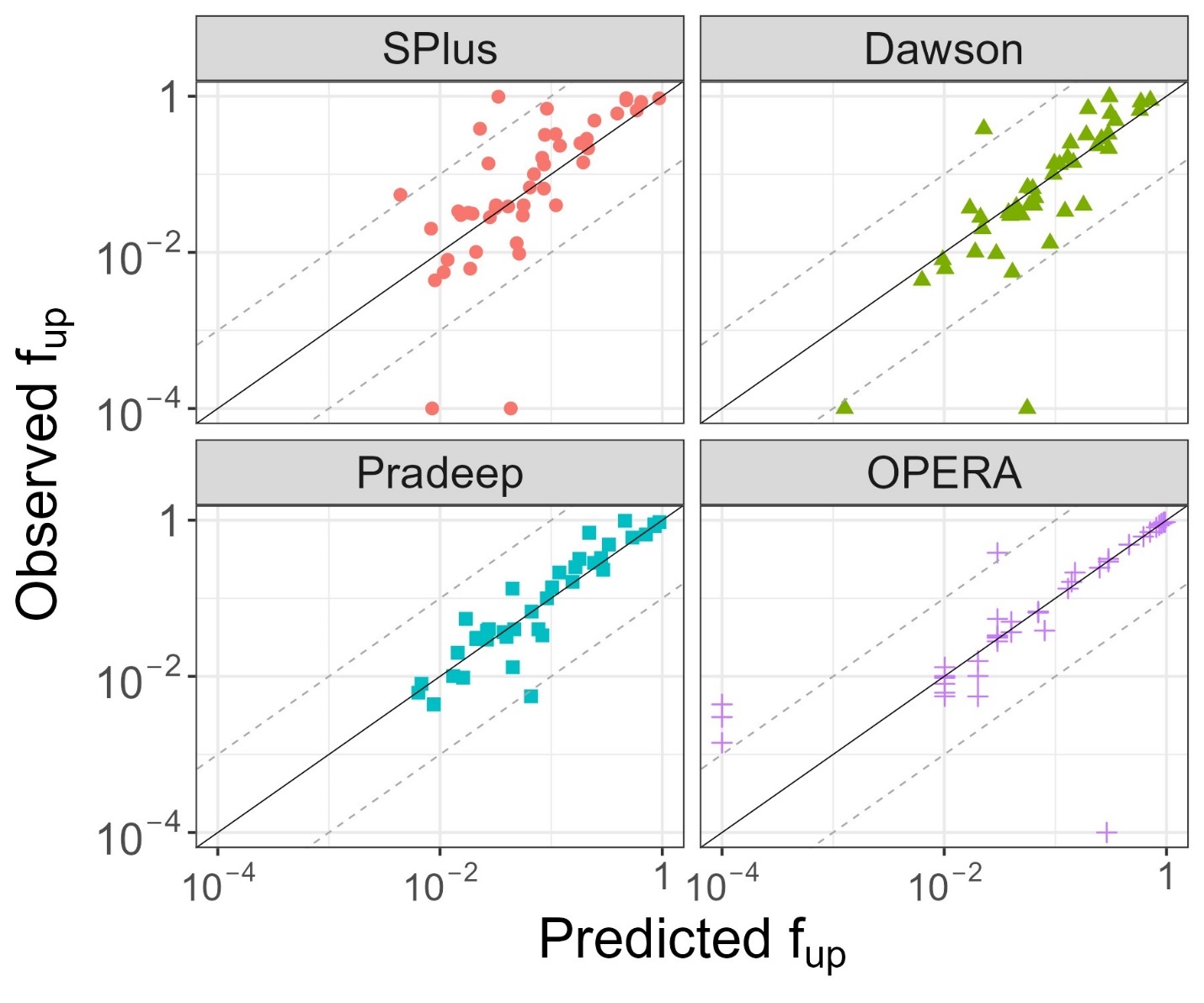
|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | HTPBTK-InVitro | HTPBTK-ADmet | HTPBTK-Dawson | HTPBTK-Pradeep | HTPBTK-OPERA | HTPBTK-Consensus | HTPBTK-YRandom | QSARINS | IFS-QAPR |
| Half-Life RMSLE | 1.6 | 1.6 | 1.6 | 1.6 | 1 | 2.2 | 2.1 | 1.8 | 1.8 |
| Half-Life RSquared | 0.01 | 0.0027 | 0.016 | 0.011 | 0.013 | 0.0047 | 0.0027 | 0.061 | 0.006 |
| Cltot RMSLE | 1.3 | 1.2 | 1.3 | 1.2 | 1.2 | 1.6 | 1.9 | 2.6 | 2.4 |
| CLtot RSquared | 0.033 | 0.17 | 0.0036 | 0.012 | 0.11 | 0.046 | 0.012 | 0.061 | 0.24 |
| Vd RMSLE | 1.4 | 1.4 | 1.4 | 1.2 | 1.3 | 1.4 | 1.1 |  |  |
| Vd RSquared | 0.26 | 0.36 | 0.41 | 0.3 | 0.31 | 0.29 | 0.28 |  |  |

# Supplemental Figures

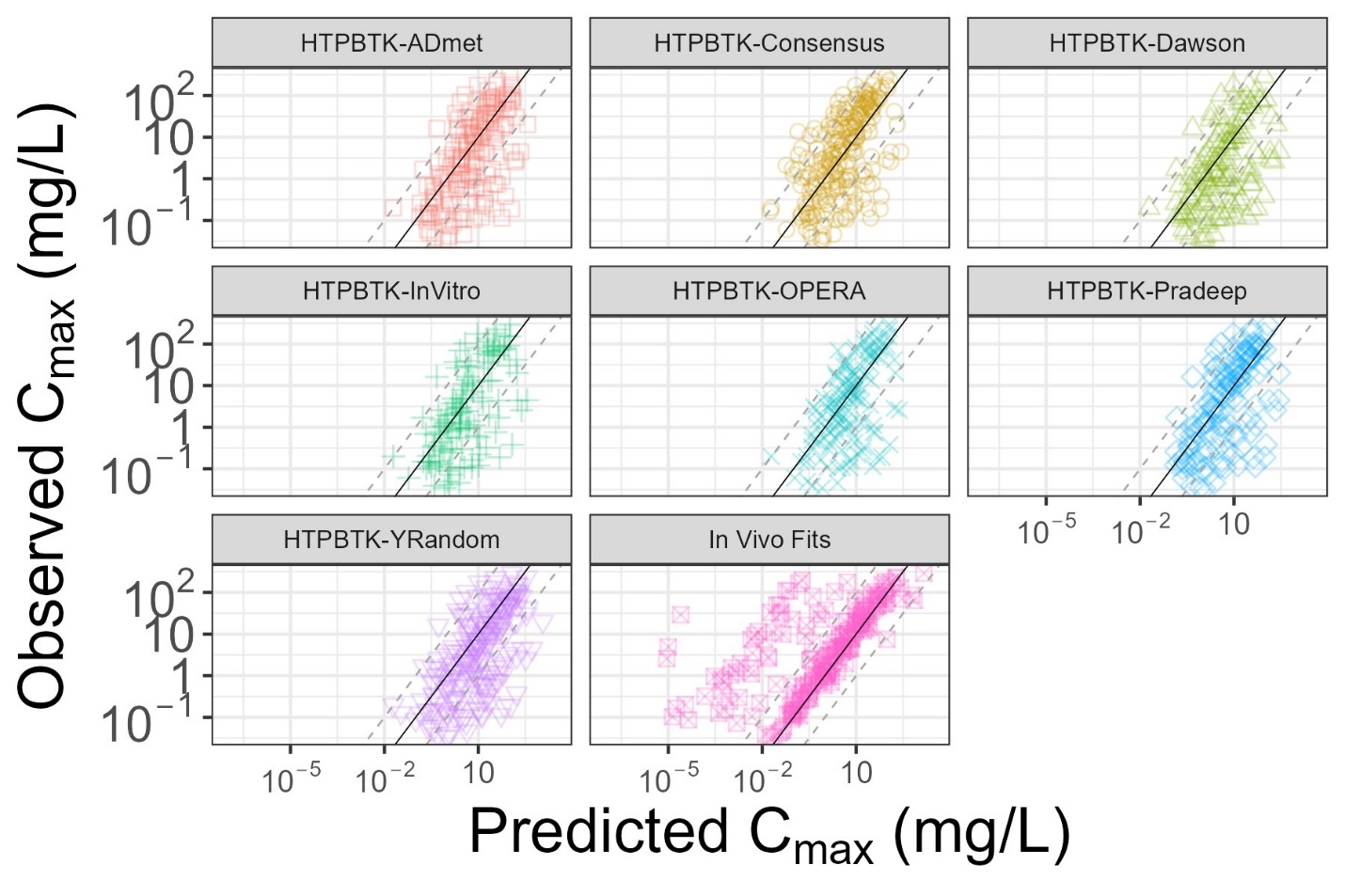
Supplemental Materials are available at: <https://github.com/USEPA/CompTox-ExpoCast-HTTKQSPRs>



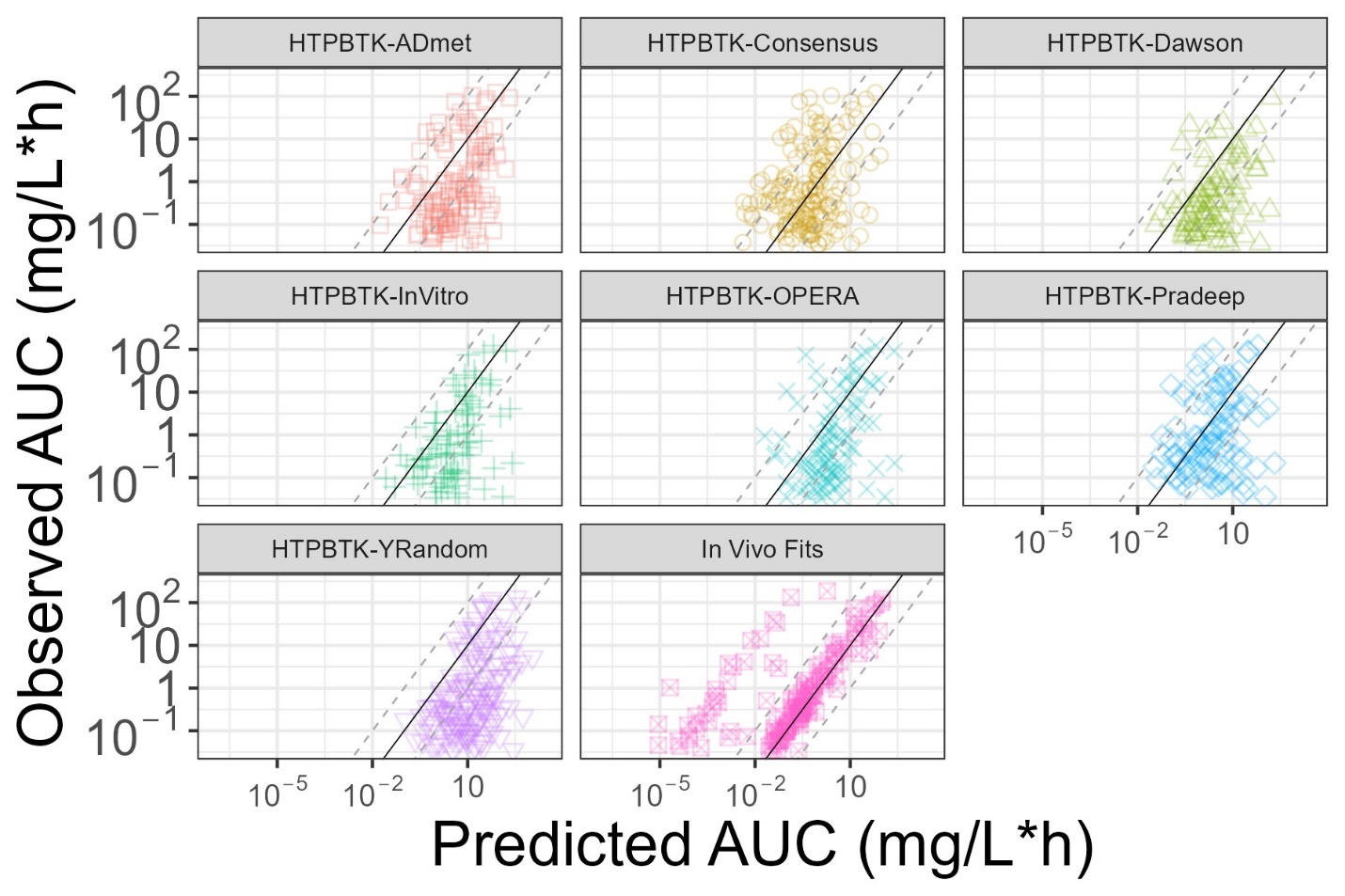
Supplemental Figure 1: Evaluation of Predictions for Intrinsic Hepatic Clearance (Clint). Zero values were plotted at 10-1, the solid line indicates identity (1:1) while the dashed lines indicate 3.2-fold difference.



Supplemental Figure 2: Evaluation of Predictions for Fraction Unbound in Plasma (fup). Zero values were plotted at 10-4, the solid line indicates identity (1:1) while the dashed lines indicate 3.2-fold difference



Supplemental Figure 3: Evaluation of Predictions for Cmax based on empirical model fits (“1CompFits”), and a PBPK model (“HTTK”) parameterized with chemical specific values either measured in vitro (“HTTK-InVitro”) or predicted with various QSPRs.



Supplemental Figure 4: Evaluation of Predictions for AUC based on empirical model fits (“1CompFits”), and a PBPK model (“HTTK”) parameterized with chemical specific values either measured in vitro (“HTTK-InVitro”) or predicted with various QSPRs.

# Supplemental Tables

Supplemental Materials are available at: <https://github.com/USEPA/CompTox-ExpoCast-HTTKQSPRs>

Supplemental Table 1: List of 102 chemicals, physico-chemical descriptors, and *in vitro* measured values, and QSPR predictions

SupTable-QSPRPredsandInVitroData.xlsx

Supplemental Table 2: Concentration vs. time data (Sayre et al., 2020)

SupTable-CvTData.xlsx

Supplemental Table 3: Empirical (one and two compartment model) toxicokinetic parameter estimates *SupTable-TKFits.txt*

Supplemental Table 4: Chemicals whose measured values were potentially retrieved “as is” from model training sets and were therefore removed from the evaluation: *SupTable-PossibleTrainingChems.txt*

Supplemental Table 5: Chemical-specific root mean square log10 errors for the full TK concentration time course data by QSPRs*SupTable-RMSLEbyChem.txt*

Supplemental Table 6: Level 3 Predictions

SupTable-Level3.xlsx