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## Spatial Generalized Linear Models in R Using **spmodel**

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### Abstract

Non-Gaussian data are common in practice and include binary, count, skewed, and proportion data types. Often, non-Gaussian data are modeled using a generalized linear model (GLM). GLMs typically assume that observations are independent of one another. This is an impractical assumption for spatial data, as nearby observations tend to be more similar than distant ones. The **spmodel** package in R provides a suite of tools for fitting spatial generalized linear models (SPGLMs) to non-Gaussian data and making spatial predictions (i.e., Kriging). SPGLMs for point-referenced data (x- and y-coordinates) are fit using the `spglm()` function, while SPGLMs for areal (lattice, polygon) data are fit using the `spgautor()` function. Both `spglm()` and `spgautor()` maximize a novel Laplace likelihood which marginalizes over the model's fixed effects and latent mean while formally incorporating spatial covariance (i.e., spatial correlation). The inputs and outputs of `spglm()` and `spgautor()` closely resemble the `glm()` function from base R, easing the transition from GLMs to SPGLMs. **spmodel** provides and builds upon several commonly used helper functions for model building like `summary()`, `plot()`, `fitted()`, and `tidy()`, among others. Spatial predictions of the latent mean at unobserved locations are obtained using `predict()` or `augment()`. **spmodel** accommodates myriad advanced modeling features like geometric anisotropy, nonspatial random effects, analysis of variance, and more. Throughout, we use **spmodel** to fit SPGLMs to moose presence and counts in Alaska, United States (US), skewed conductivity data in the Southwestern US, harbor seal abundance trends in Alaska, US, and voter turnout rates in Texas, US.

*Keywords:* autoregressive model, geostatistical model, spatial covariance, spatial correlation.

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## 1. Introduction

In practice, non-Gaussian data (e.g., binary, count, skewed, and proportion data) are ubiquitous. Non-Gaussian that belong to an exponential family data can be naturally modeled using a generalized linear model (GLM) regression framework (Nelder and Wedderburn 1972; McCullagh and Nelder 1989). In a GLM, an  $n \times 1$  response variable  $\mathbf{y}$  belongs to a statistical distribution (e.g., Binomial, Poisson) with some mean and variance. Often, the analysis goal is to study the impact of a linear function of several explanatory variables on  $\mathbf{y}$  through a GLM. In this context, the latent (i.e., unobserved) mean of  $\mathbf{y}$ ,  $\boldsymbol{\mu}$ , is linked to these explanatory variables via a link function:

$$f(\boldsymbol{\mu}|\mathbf{X}, \boldsymbol{\beta}) \equiv \mathbf{w} = \mathbf{X}\boldsymbol{\beta}, \quad (1)$$

where for a sample size  $n$ ,  $f(\cdot)$  is a link function that connects  $\boldsymbol{\mu}$  to  $\mathbf{w}$ ,  $\mathbf{X}$  is the  $n \times p$  design matrix of explanatory variables, and  $\boldsymbol{\beta}$  is the  $p \times 1$  vector of fixed effects. While the mean is typically constrained in some way (e.g., if a probability, between zero and one), the link function generally makes  $\mathbf{w}$  unconstrained. Common link functions include the log odds (i.e., logit) link for binary and proportion data and the log link for count and skewed data.

Equation 1 can also be written in terms of the inverse link function,  $f^{-1}(\cdot)$ :

$$\boldsymbol{\mu}|\mathbf{X}, \boldsymbol{\beta} \equiv f^{-1}(\mathbf{w}) = f^{-1}(\mathbf{X}\boldsymbol{\beta}).$$

The GLM fixed effects ( $\boldsymbol{\beta}$ ) are typically estimated via maximum likelihood (Chambers and Hastie 1992). It is often convenient to compute the maximum likelihood estimates using the iteratively reweighted least squares (IRWLS) algorithm (Wood 2017), which is the approach used by the `glm()` function in the R programming language (R Core Team 2024). GLMs add an additional layer of complexity compared to linear regression models, as the left-hand size of Equation 1 is a function of the mean of  $\mathbf{y}$  rather than  $\mathbf{y}$  itself (as in linear regression models).

The standard GLM assumes the elements of  $\mathbf{y}$  are independent. This independence assumption is typically impractical for spatial data. For spatial data, nearby observations tend to be more similar than distant observations (Tobler 1970), which leads to positive spatial covariance. The consequences of ignoring spatial covariance in statistical models for spatial data can be severe and include imprecise parameter estimates as well as misleading standard errors that inflate Type-I error rates and decrease power (Zimmerman and Ver Hoef 2024).

An approach for handling spatial data using a GLM is to assume the elements of  $\mathbf{w}$  share covariance that varies spatially. This is achieved by adding to Equation 1 two random effects,  $\boldsymbol{\tau}$  and  $\boldsymbol{\epsilon}$ . The random effect  $\boldsymbol{\tau}$  is an  $n \times 1$  column vector of spatially dependent random errors. We assume that  $E(\boldsymbol{\tau}) = \mathbf{0}$  and  $Cov(\boldsymbol{\tau}) = \sigma_{\tau}^2 \mathbf{R}$ , where  $E(\cdot)$  and  $Cov(\cdot)$  denote expectation and covariance, respectively. The variance parameter  $\sigma_{\tau}^2$  controls the magnitude of spatial covariance and is often called a partial sill. The matrix  $\mathbf{R}$  is an  $n \times n$  spatial correlation matrix that depends on a range parameter controlling the distance-decay rate of the spatial correlation. One example of a spatial covariance matrix is the “exponential”, which is given by

$$Cov(\boldsymbol{\tau}) = \sigma_{de}^2 \mathbf{R}_{exp} = \sigma_{de}^2 \exp(-\mathbf{H}/\phi), \quad (2)$$

where  $\mathbf{H}$  is a matrix of pairwise distances among the elements of  $\mathbf{y}$  and  $\phi$  is the range parameter. From Equation 2, as the distance between two elements of  $\mathbf{y}$  increases, the spatial

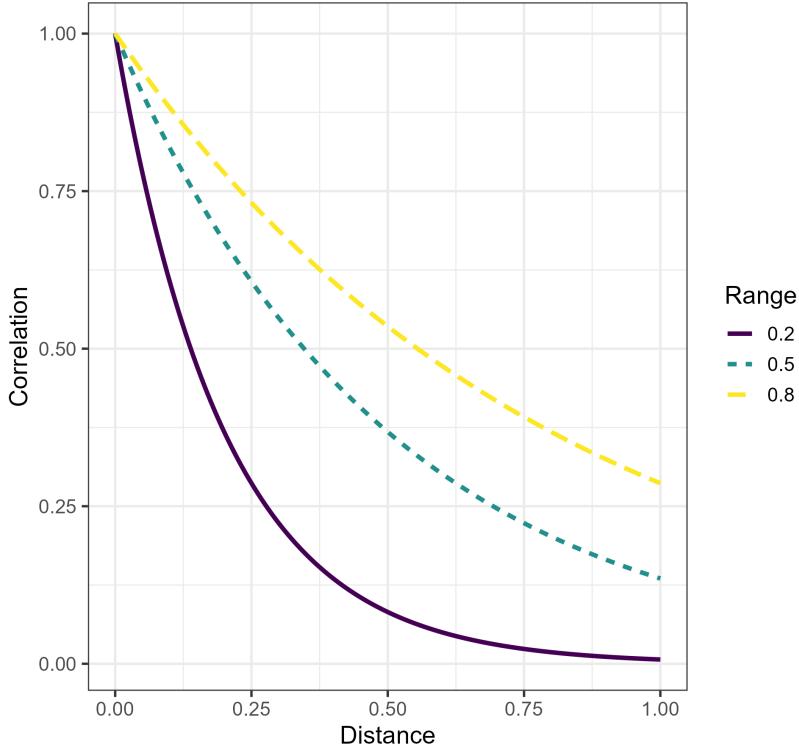


Figure 1: An exponential spatial correlation function with varying range parameters.

39 covariance decreases, which reflects intuition. Moreover, as the range parameter,  $\phi$ , increases,  
 40 the strength of spatial dependence increases (Figure 1). The random effect  $\epsilon$  is an  $n \times 1$   
 41 column vector of independent random errors. We assume that  $E(\epsilon) = \mathbf{0}$  and  $\text{Cov}(\tau) = \sigma_\epsilon^2 \mathbf{I}$ ,  
 42 where  $\mathbf{I}$  is an  $n \times n$  identity matrix. The variance parameter  $\sigma_\epsilon^2$  controls the magnitude  
 43 of nonspatial variability (i.e., fine-scale variation) and is often called a nugget. Often in  
 44 spatial statistics, quantities are explicitly referenced with respect to  $\mathbf{s}$ , a vector of coordinates  
 45 indexing the observation (Cressie 1993). For example,  $\mathbf{y}$  and  $\mathbf{X}$  may instead be written  $\mathbf{y}(\mathbf{s})$   
 46 and  $\mathbf{X}$ , respectively. We acknowledge the utility of this nomenclature but drop the explicit  
 47 dependence on  $\mathbf{s}$  for simplicity of notation.

48 Through inclusion of  $\tau$  and  $\epsilon$ , the spatial GLM (SPGLM) can be written as

$$f(\mu | \mathbf{X}, \beta, \tau, \epsilon) \equiv \mathbf{w} = \mathbf{X}\beta + \tau + \epsilon. \quad (3)$$

49 Assuming independence among  $\tau$  and  $\epsilon$ , it follows that

$$\text{Cov}(\tau + \epsilon) = \text{Cov}(\tau) + \text{Cov}(\epsilon) = \sigma_\tau^2 \mathbf{R} + \sigma_\epsilon^2 \mathbf{I}.$$

50 Henceforth, we refer to  $\sigma_\tau^2$  as  $\sigma_{de}^2$  (for spatially dependent error variance) and  $\sigma_\epsilon^2$  as  $\sigma_{ie}^2$   
 51 (for independent error variance). The parameters  $\sigma_{de}^2$ ,  $\sigma_{ie}^2$ , and  $\phi$ , in addition to any other  
 52 parameters in  $\mathbf{R}$ , compose  $\theta$ , the covariance parameter vector.

53 Fitting and using SPGLMs is challenging both conceptually and computationally (Bolker,  
 54 Brooks, Clark, Geange, Poulsen, Stevens, and White 2009). Recently, however, there have  
 55 been numerous, significant advances in R software that have made these models more acces-  
 56 sible to practitioners. The **brms** (Bürkner 2017), **carBayes** (Lee 2013), **ngspatial** (Hughes and

57 Cui 2020), **R-INLA** (Lindgren and Rue 2015) and **inlabru** (Bachl, Lindgren, Borchers, and  
 58 Illian 2019), **spBayes** (Finley, Banerjee, and Carlin 2007), **spOccupancy** (Doser, Finley, Kéry,  
 59 and Zipkin 2022), **spAbundance** (Doser, Finley, Kéry, and Zipkin 2024), and **spNNGP** (Finley,  
 60 Datta, and Banerjee 2022) packages take a Bayesian approach, either directly sampling from  
 61 posterior distributions of parameters (e.g., using MCMC) or approximating them. A benefit  
 62 of Bayesian approaches is that prior information can be incorporated and uncertainty quan-  
 63 tification of parameter estimates is straightforward. However, Bayesian approaches, especially  
 64 those using MCMC, can be computationally expensive. In order to reduce computation time,  
 65 many of these packages work with the precision matrix instead of the covariance matrix so  
 66 that computationally expensive matrix inversion is not required. For example, **R-INLA** uses  
 67 the precision matrix and tends to be very fast. Working with precision matrices, however,  
 68 can be more restrictive and less intuitive than working directly with the covariance matrix.  
 69 The **FRK** (Sainsbury-Dale, Zammit-Mangion, and Cressie 2024), **glmmTMB** (Brooks, Kris-  
 70 tensen, van Benthem, Magnusson, Berg, Nielsen, Skaug, Maechler, and Bolker 2017), **hglm**  
 71 (Ronnegard, Shen, and Alam 2010), **mgcv** (Wood 2017), and **spaMM** (Rousset and Ferdy  
 72 2014) packages directly use Laplace, quasi-likelihood, or reduced-rank approaches to estimate  
 73 parameters. These direct approaches tend to be computationally efficient, as they don't rely  
 74 on MCMC sampling. In contrast to the Bayesian approach, a drawback of these direct ap-  
 75 proaches is that prior information cannot be formally incorporated and covariance parameter  
 76 uncertainty is more challenging to quantify. The **sdmTMB** (Anderson, Ward, English, Bar-  
 77 nett, and Thorson 2024) package combines elements of **R-INLA**, **glmmTMB**, and Gaussian  
 78 Markov random fields to fit a wide variety of SPGLMs, while **tinyVAST** (Thorson, Ander-  
 79 son, Goddard, and Rooper 2025) extends some of these models to multivariate or (dynamic)  
 80 structural equation models.

81 Building from Evangelou, Zhu, and Smith (2011) and Bonat and Ribeiro Jr (2016), Ver Hoef,  
 82 Blagg, Dumelle, Dixon, Zimmerman, and Conn (2024) proposed a novel approach for fitting  
 83 SPGLMs that leverages the Laplace approximation while marginalizing over both the latent  
 84  $\mathbf{w}$  and the fixed effects ( $\beta$ ) and accommodating spatial covariance. This approach performed  
 85 efficiently in a variety of simulation settings, generally having appropriate confidence interval  
 86 coverage for the fixed effects and prediction interval coverage for  $\mathbf{w}$  at new locations. The  
 87 approach performed similarly to the Bayesian SPGLM approach in **spBayes** and the automatic  
 88 differentiation SPGLM approach in **glmmTMB** but was much faster. At small sample sizes,  
 89 the approach outperformed the approximate Bayesian SPGLM approach in **R-INLA** and had  
 90 similar computational times. For moderate sample sizes, it performed similarly to **R-INLA**,  
 91 though **R-INLA** was faster. The novel Laplace approach is particularly attractive for two  
 92 reasons. First, it is general enough that can be applied to any covariance structure (not just  
 93 spatial). Second, after estimating the covariance parameters, analytical solutions exist for the  
 94 fixed effects (and their standard errors) as well as predictions of the latent  $\mathbf{w}$  at new locations  
 95 (and their standard errors).

96 The **spmodel** R package (Dumelle, Higham, and Ver Hoef 2023) recently provided a full set  
 97 of modeling tools for SPGLMs fit using the novel Laplace approach described by Ver Hoef  
 98 *et al.* (2024). These modeling tools are approachable and mirror the familiar **glm()** syntax  
 99 from base-R, making the transition from GLMs to SPGLMs relatively seamless. The **spglm()**  
 100 function fits SPGLMs for point-referenced data (e.g., x-coordinates and y-coordinates repre-  
 101 senting point locations in a field; models are sometimes called “geostatistical” models), while  
 102 the **spgautor()** function fits SPGLMs for areal data (e.g., polygon boundaries represent-

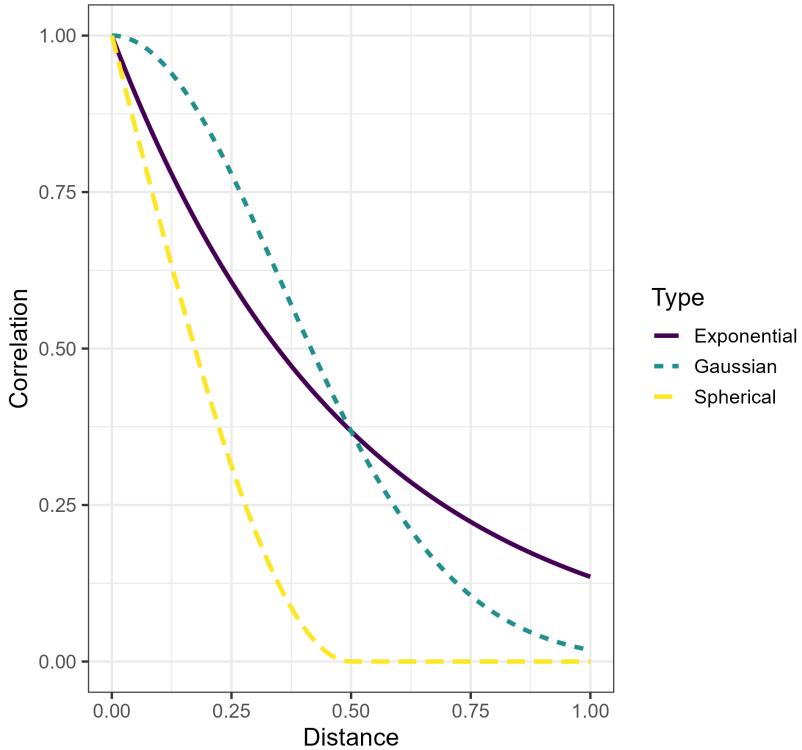


Figure 2: Exponential, Gaussian, and spherical spatial correlation functions all with range parameters equal to 0.5.

ing geographic subsets of a region; models are sometimes called “autoregressive” models). **spmodel** supports the binomial distribution for binary data, Poisson and negative binomial distributions for count data, Gamma and inverse Gaussian distributions for skewed data, and the beta distribution for proportion data. There are 20 different spatial covariance structures available including the exponential, Gaussian, and spherical for point-referenced data (Figure 2) and the conditional autoregressive, and simultaneous autoregressive structures for areal data. **spmodel** provides tools for commonly used model summaries, visualizations, and diagnostics (e.g., Cook’s distance) using standard R helper functions like `summary()`, `plot()`, `fitted()`, and `tidy()`, among others. **spmodel** also provides tools to predict `w` at new locations and quantify uncertainty in those prediction using `predict()` and `augment()`. This core functionality, combined with several advanced features we describe throughout the manuscript, enables **spmodel** to introduce novel, important SPGLM modeling tools previously missing from the existing R ecosystem.

**spmodel** (version 0.11.0) is arguably most similar to **sdmTMB** (version 0.7.4) in terms of scope and feel. Both packages use similar syntax as `glm()`, accommodate flexible `formula` arguments (e.g., offsets, splines), handle spatial covariance that decays at different rates in different directions (i.e., geometric anisotropy), incorporate nonspatial random effects, support other R packages for modeling like **broom** (Robinson, Hayes, and Couch 2021; Kuhn and Silge 2022), **emmeans** (Lenth 2024), and **car** (Fox and Weisberg 2019), and have tools for model summaries, prediction, and simulating data. There are some notable differences between the two packages, however. **sdmTMB** supports several additional GLM distributions like the

<sup>124</sup> Tweedie, supports Hurdle models, and can incorporate prior information through Bayesian  
<sup>125</sup> applications. **sdmTMB** also provides tools for working with temporal data and spatiotem-  
<sup>126</sup> poral data and provides enhanced visualizations of the model's marginal effects. **sdmTMB**  
<sup>127</sup> does require a preprocessing step of constructing a mesh for the stochastic partial differential  
<sup>128</sup> equation approach, and the density of the mesh can affect model results and computational  
<sup>129</sup> complexity. On the other hand, **spmodel** does not require the construction of a mesh prior  
<sup>130</sup> to modeling. **spmodel** also supports 20 different spatial covariances and models them di-  
<sup>131</sup> rectly, rather than using a precision matrix approximation to the Matérn spatial covariance  
<sup>132</sup> as in **sdmTMB**. **spmodel** can also model areal data directly using neighborhood distance and  
<sup>133</sup> autoregressive models rather than relying on the polygon centroid (as in **sdmTMB**), which  
<sup>134</sup> may not be within the polygon's boundaries. **spmodel** also provides experimental design  
<sup>135</sup> tools (e.g., analysis of variance, contrasts), supports **sf** objects in modeling and prediction  
<sup>136</sup> functions (Pebesma 2018), has several specialized model diagnostics like leverage values and  
<sup>137</sup> Cook's distances, and has analytic solutions for fixed effect and prediction standard errors.  
<sup>138</sup> Other similarities and differences do exist between **sdmTMB** and **spmodel**, and both packages  
<sup>139</sup> continue to evolve. Overall, we believe that these packages are complementary and enhance  
<sup>140</sup> the suite of SPGLM tools accessible to practitioners.

<sup>141</sup> The rest of this article is organized as follows. In Section 2, we provide some background for  
<sup>142</sup> the SPGLM fitting and prediction routines in **spmodel**. In Section 3, we provide an overview  
<sup>143</sup> of core SPGLM functionality in **spmodel** by modeling moose presence in Alaska, United  
<sup>144</sup> States (US). In Section 4, we model moose counts in Alaska, US; skewed lake conductivity  
<sup>145</sup> in the Southwestern US; harbor seal abundance trend behavior in Alaska, US; and voter  
<sup>146</sup> turnout rates in Texas, US. And in Section 5, we end with a discussion synthesizing **spmodel**'s  
<sup>147</sup> contributions to the analysis of SPGLMs in R.

## 2. The spatial generalized linear model and marginalization

<sup>148</sup> The novel Laplace approach implemented in **spmodel** formally maximizes a hierarchical GLM  
<sup>149</sup> likelihood (Lee and Nelder 1996; Wood 2017), making likelihood-based statistics for model  
<sup>150</sup> comparison like AIC (Akaike 1974), AICc (Hoeting, Davis, Merton, and Thompson 2006), BIC  
<sup>151</sup> (Schwarz 1978), deviance (McCullagh and Nelder 1989), and likelihood ratio tests available.  
<sup>152</sup> These types of statistics are not available for quasi-likelihood (Wedderburn 1974; Breslow  
<sup>153</sup> and Clayton 1993) or pseudo-likelihood approaches (Wolfinger and O'Connell 1993), which  
<sup>154</sup> only specify the first two moments of a distribution. Next, we describe a brief overview of  
<sup>155</sup> the approach and how it can be used for several primary data analysis tasks (Tredennick,  
<sup>156</sup> Hooker, Ellner, and Adler 2021) like model comparison, parameter estimation, inference,  
<sup>157</sup> model diagnostics, and prediction.

### 158 2.1. Formulating the hierarchical likelihood

<sup>159</sup> We can write the SPGLM likelihood hierarchically as

$$[\mathbf{y}|\mathbf{X}, \varphi, \boldsymbol{\theta}] = \int_{\mathbf{w}} \int_{\boldsymbol{\beta}} [\mathbf{y}|f^{-1}(\mathbf{w}), \varphi] [\mathbf{w}|\mathbf{X}, \boldsymbol{\theta}] d\boldsymbol{\beta} d\mathbf{w}, \quad (4)$$

<sup>160</sup> where  $[\mathbf{y}|f^{-1}(\mathbf{w}), \varphi]$  is the density for the appropriate response distribution of  $\mathbf{y}$  (e.g., bino-  
<sup>161</sup> mial, Poisson) given the latent  $\mathbf{w}$  and dispersion parameter ( $\varphi$ ), and  $[\mathbf{w}|\mathbf{X}, \boldsymbol{\theta}]$  is the multi-  
<sup>162</sup> variate Gaussian density for  $\mathbf{w}$  given the explanatory variables ( $\mathbf{X}$ ), fixed effects ( $\boldsymbol{\beta}$ ), and

<sup>163</sup> spatial covariance parameters ( $\boldsymbol{\theta}$ ). The elements of  $[\mathbf{y}|f^{-1}(\mathbf{w}), \varphi]$  are conditionally independent (given  $\mathbf{w}$ ), but the elements of  $[\mathbf{w}|\mathbf{X}, \boldsymbol{\theta}]$  share spatial covariance. Following Harville (1977), we can integrate  $\boldsymbol{\beta}$  out of Equation 4, which yields

$$[\mathbf{y}|\mathbf{X}, \varphi, \boldsymbol{\theta}] = \int_{\mathbf{w}} [\mathbf{y}|f^{-1}(\mathbf{w}), \varphi] [\mathbf{w}|\mathbf{X}, \boldsymbol{\theta}] d\mathbf{w}, \quad (5)$$

<sup>166</sup> where  $[\mathbf{w}|\mathbf{X}, \boldsymbol{\theta}]$  is the restricted (i.e., residual) multivariate Gaussian density (Patterson and <sup>167</sup> Thompson 1971) for  $\mathbf{w}$  given the explanatory variables and covariance parameters. Equation 5 <sup>168</sup> can synonymously be written after profiling the overall variance out of  $\boldsymbol{\Sigma}$ , which reduces the <sup>169</sup> dimension of  $\boldsymbol{\theta}$  by one for optimization (Wolfinger, Tobias, and Sall 1994). The restricted <sup>170</sup> multivariate Gaussian density is given by

$$[\mathbf{w}|\mathbf{X}, \boldsymbol{\theta}] = \frac{\exp(-\frac{1}{2}(\mathbf{y} - \mathbf{X}\tilde{\boldsymbol{\beta}})\boldsymbol{\Sigma}^{-1}(\mathbf{y} - \mathbf{X}\tilde{\boldsymbol{\beta}})^T)}{(2\pi)^{(n-p)/2}|\boldsymbol{\Sigma}|^{1/2}|\mathbf{X}^T\boldsymbol{\Sigma}^{-1}\mathbf{X}|^{1/2}},$$

<sup>171</sup> where  $\tilde{\boldsymbol{\beta}} = (\mathbf{X}^T\boldsymbol{\Sigma}^{-1}\mathbf{X})^{-1}\mathbf{X}^T\boldsymbol{\Sigma}^{-1}\mathbf{w}$  and  $|\cdot|$  denotes the determinant. Next, let

$$\ell_{\mathbf{w}} = \log([\mathbf{y}|f^{-1}(\mathbf{w}), \varphi][\mathbf{w}|\mathbf{X}, \boldsymbol{\theta}])$$

<sup>172</sup> and rewrite Equation 5 as

$$[\mathbf{y}|\mathbf{X}, \varphi, \boldsymbol{\theta}] = \int_{\mathbf{w}} \exp(\ell_{\mathbf{w}}) d\mathbf{w}.$$

<sup>173</sup> A second-order Taylor series expansion of  $\ell_{\mathbf{w}}$  around a point  $\hat{\mathbf{w}}$  yields

$$[\mathbf{y}|\mathbf{X}, \varphi, \boldsymbol{\theta}] \approx \int_{\mathbf{w}} \exp(\ell_{\hat{\mathbf{w}}} + \mathbf{g}^T(\mathbf{w} - \hat{\mathbf{w}}) + \frac{1}{2}(\mathbf{w} - \hat{\mathbf{w}})^T \mathbf{G}(\mathbf{w} - \hat{\mathbf{w}})) d\mathbf{w},$$

<sup>174</sup> where  $\mathbf{g}$  and  $\mathbf{G}$  are the gradient and Hessian, respectively, of  $\ell_{\mathbf{w}}$  with respect to  $\mathbf{w}$ . If  $\hat{\mathbf{w}}$  is a <sup>175</sup> value for which  $\mathbf{g} = \mathbf{0}$ ,

$$[\mathbf{y}|\mathbf{X}, \varphi, \boldsymbol{\theta}] \approx \exp(\ell_{\hat{\mathbf{w}}}) \int_{\mathbf{w}} \exp(-\frac{1}{2}(\mathbf{w} - \hat{\mathbf{w}})^T (-\mathbf{G})(\mathbf{w} - \hat{\mathbf{w}})) d\mathbf{w}. \quad (6)$$

<sup>176</sup> The integral in Equation 6 can be solved by leveraging properties of the normalizing constant <sup>177</sup> of a multivariate Gaussian distribution. Thus, rewriting  $\exp(\ell_{\hat{\mathbf{w}}})$  yields

$$[\mathbf{y}|\mathbf{X}, \varphi, \boldsymbol{\theta}] \approx [\mathbf{y}|f^{-1}(\hat{\mathbf{w}}), \varphi][\hat{\mathbf{w}}|\mathbf{X}, \boldsymbol{\theta}] (2\pi)^{n/2} |-\mathbf{G}_{\hat{\mathbf{w}}}|^{-1/2}. \quad (7)$$

<sup>178</sup> Maximizing the natural logarithm of Equation 7 requires a doubly iterative process over <sup>179</sup>  $\boldsymbol{\theta}$  and  $\varphi$  as well as  $\mathbf{w}$ , eventually yielding the the marginal restricted maximum likelihood <sup>180</sup> estimators  $\hat{\varphi}$  and  $\hat{\boldsymbol{\theta}}$  and their corresponding values of  $\hat{\mathbf{w}}$ . Maximizing this log likelihood is a <sup>181</sup> computationally expensive operation that involves repeatedly evaluating  $\boldsymbol{\Sigma}^{-1}$ ,  $\mathbf{g}$ , and  $\mathbf{G}$ ; see <sup>182</sup> Ver Hoef *et al.* (2024) for more details and forms of  $\mathbf{g}$  and  $\mathbf{G}$  for various response distributions.

## <sup>183</sup> 2.2. Estimating fixed effects

<sup>184</sup> Though the fixed effects are integrated out of the likelihood, we can still estimate them using <sup>185</sup> generalized least squares (GLS) principles, a common practice for linear models estimated

186 using restricted maximum likelihood methods. Had we observed  $\mathbf{w}$ , a GLS estimator for  $\beta$  is  
 187 given by

$$\hat{\beta} = (\mathbf{X}^\top \Sigma^{-1} \mathbf{X})^{-1} \mathbf{X}^\top \Sigma^{-1} \mathbf{w} = \mathbf{B}\mathbf{w},$$

188 where  $\mathbf{B} = (\mathbf{X}^\top \Sigma^{-1} \mathbf{X})^{-1} \mathbf{X}^\top \Sigma^{-1}$ . However, we only observe  $\hat{\mathbf{w}}$ , so it is reasonable to define  
 189  $\hat{\beta} = \mathbf{B}\hat{\mathbf{w}}$ . Thus, to derive properties of  $\hat{\beta}$  like expectation and variance, we must derive  
 190 these properties for  $\hat{\mathbf{w}}$ . To do so, we must condition on  $\mathbf{w}$  as if it were observed and invoke  
 191 properties of the laws of total expectation and variance. Because  $\hat{\mathbf{w}}$  was optimized via the  
 192 likelihood, we assume that given  $\mathbf{w}$ ,  $\hat{\mathbf{w}}$  has mean  $\mathbf{w}$  and variance approximately equal to  
 193  $-\mathbf{H}^{-1}$  (the inverse Hessian). It follows that  $E(\hat{\mathbf{w}})$  is given by

$$E(\hat{\mathbf{w}}) = E(E(\hat{\mathbf{w}}|\mathbf{w})) = E(\mathbf{w}) = \mathbf{X}\beta$$

194 and  $\text{Var}(\hat{\mathbf{w}})$  is given by

$$\begin{aligned} \text{Var}(\hat{\mathbf{w}}) &= E(\text{Var}(\hat{\mathbf{w}}|\mathbf{w})) + \text{Var}(E(\hat{\mathbf{w}}|\mathbf{w})) \\ &= E(-\mathbf{H}^{-1}) + \text{Var}(\mathbf{w}) \\ &= -\mathbf{H}^{-1} + \Sigma \end{aligned}$$

195 Putting this all together, it follows that

$$E(\hat{\beta}) = E(\mathbf{B}\hat{\mathbf{w}}) = \mathbf{B}E(\hat{\mathbf{w}}) = (\mathbf{X}^\top \Sigma^{-1} \mathbf{X})^{-1} (\mathbf{X}^\top \Sigma^{-1} \mathbf{X}) \beta = \beta$$

196 and

$$\begin{aligned} \text{Var}(\hat{\beta}) &= \text{Var}(\mathbf{B}\hat{\mathbf{w}}) \\ &= \mathbf{B}\text{Var}(\hat{\mathbf{w}})\mathbf{B}^\top \\ &= \mathbf{B}(-\mathbf{H}^{-1} + \Sigma)\mathbf{B}^\top \\ &= \mathbf{B}(-\mathbf{H})^{-1}\mathbf{B}^\top + \mathbf{B}\Sigma\mathbf{B}^\top \\ &= \mathbf{B}(-\mathbf{H})^{-1}\mathbf{B}^\top + (\mathbf{X}^\top \Sigma^{-1} \mathbf{X})^{-1} \end{aligned}$$

197 In practice,  $\text{Var}(\hat{\beta})$  is estimated by evaluating  $\Sigma$  at  $\hat{\theta}$ , the estimated covariance parameter  
 198 vector.

199 These results are important because they justify closed-form solutions for  $\hat{\beta}$  and its associated  
 200 variance. Closed-form solutions are useful because they bypass the need for sampling-based  
 201 strategies to evaluate the mean and variance of  $\hat{\beta}$ , a common technique for other approaches  
 202 to SPGLMs like Bayesian MCMC.

### 203 2.3. Inspecting model diagnostics

204 Inspecting model diagnostics is an important step of the modeling process that can yield  
 205 valuable insights into model behavior and unusual observations. [Montgomery, Peck, and](#)  
 206 [Vining \(2021\)](#) contextualize three components of unusual observations: outliers, leverage,  
 207 and influence. An observation is an outlier if it has an unusual response value relative to  
 208 expectation. The response GLM residuals simply compare the observation to its fitted latent  
 209 mean:

$$\mathbf{r}_r = \mathbf{y} - f^{-1}(\hat{\mathbf{w}})$$

Because observations often have a unique support in a GLM (e.g., only two possible response values for binary data) and the variance of an observation generally depends on its mean, response residuals lack some utility. Deviance residuals are a function of response residuals that are appropriately scaled to behave more like response residuals in a standard linear model. Deviance residuals are given by

$$\mathbf{r}_d = \text{sign}(\mathbf{r}_r) \sqrt{\mathbf{d}},$$

where  $\mathbf{d}$  is a vector of individual deviances. The sum of the squared deviance residuals equals the sum of  $\mathbf{d}$ . The sum of  $\mathbf{d}$  is the deviance of the model fit, which quantifies twice the difference in log likelihoods between the a saturated model that fits every observation perfectly (i.e.,  $\mathbf{y} = f^{-1}(\hat{\mathbf{w}}_i)$  for all  $i$ ) and the fitted model (Myers, Montgomery, Vining, and Robinson 2012). Deviance is often used as a fit statistic; lower values of deviance imply a better model fit. Pearson and standardized residuals are other types of GLM residuals that involve a scaling of the response residuals; the Pearson residuals scale  $\mathbf{r}_r$  by the square root of  $\mathbf{V}$ , while the standardized residuals scale the deviance residuals by  $\frac{1}{\sqrt{(1-\mathbf{L}_{ii})}}$ , where  $\mathbf{L}_{ii}$  is the  $i$ th diagonal element of the leverage matrix, which we discuss next.

An observation has high leverage if its combination of explanatory variables is far away from other observations. In a linear model, the leverage (i.e., hat) values are the diagonal of the leverage (i.e., projection, hat) matrix,  $\mathbf{L} = \mathbf{X}(\mathbf{X}^\top \mathbf{X})^{-1}\mathbf{X}^\top$ . In a GLM, the leverage matrix is given by

$$\mathbf{L} = \mathbf{V}^{1/2} \mathbf{X} (\mathbf{X}^\top \mathbf{V} \mathbf{X})^{-1} \mathbf{X}^\top \mathbf{V}^{1/2},$$

where  $\mathbf{V}$  is a diagonal matrix with  $i$ th diagonal element equal to the variance of the response distribution evaluated at  $f^{-1}(\mathbf{w}_i)$  (Faraway 2016);  $\mathbf{V}$  is sometimes called the GLM weight matrix. The larger the value of  $\mathbf{L}_{ii}$ , the more severe the leverage from the  $i$ th observation.

An observation is influential if it has a sizable impact on model fit. Influence is measured using Cook's distance (Cook 1979; Cook and Weisberg 1982), which is given for a GLM by

$$\mathbf{c} = \frac{\mathbf{r}_s^2}{\text{tr}(\mathbf{L})} \frac{\text{diag}(\mathbf{L})}{(1 - \text{diag}(\mathbf{L}))},$$

where  $\mathbf{r}_s^2$  are the standardized residuals and  $\text{diag}(\mathbf{L})$  indicates the diagonal elements of the leverage matrix. The larger the value of  $\mathbf{c}_i$ , the more severe the influence from the  $i$ th observation. Montgomery *et al.* (2021) provide guidance for interpreting these types of statistics, including cutoffs to consider when identifying unusual residual, leverage, or influence values.

In a linear model, the  $R^2$  (R-squared) statistic quantifies the proportion of variability in the data captured by the explanatory variables. It is calculated as one minus the ratio of the error sum of squares to the total sum of squares (Rencher and Schaalje 2008). In a GLM, there are many ways to define a statistic that emulates the aforementioned meaning of  $R^2$  from the linear model (Smith and McKenna 2013). This statistic is called a pseudo R-squared ( $PR^2$ ). One  $PR^2$  for GLMs simply replaces the sums of squares ratio from the linear model with the deviance ratio:

$$PR^2 = 1 - \frac{\text{deviance}_{\text{fit}}}{\text{deviance}_{\text{null}}},$$

where  $\text{deviance}_{\text{fit}}$  is the deviance of the fitted model (sometimes called the residual deviance) and  $\text{deviance}_{\text{null}}$  is the deviance of the model taking  $\mathbf{X} \equiv \mathbf{1}$ , a column of all ones (i.e., an intercept-only model). In practice,  $\text{deviance}_{\text{null}}$  is derived by computing  $\hat{\mathbf{w}}$  when  $\mathbf{X} \equiv \mathbf{1}$  given  $\hat{\boldsymbol{\theta}}$  and  $\hat{\varphi}$  from the fitted model. Like  $R^2$ ,  $PR^2$  can be adjusted to account for the numbers of parameters estimated in a model. Because the  $\text{deviance}_{\text{null}}$  denominator changes across fitted models (as the values of  $\hat{\boldsymbol{\theta}}$  and  $\hat{\varphi}$  change), this statistic should not be used as a model comparison tool. Rather, it should be used as an informative diagnostic tool that unique to each model fit and describes how much variability from that model is attributable to the explanatory variables.

## 2.4. Predicting at new locations

We may also predict values of the latent mean (on the link scale) at new locations by leveraging the spatial covariance between observed locations and new locations (spatial prediction is also called Kriging; see Cressie (1990)). Again suppose that we observed  $\mathbf{w}$  and we want to make predictions at  $\mathbf{u}$ , a vector of latent means at the new locations that follows the same SPGLM from Equation 3 and having fixed effects design matrix,  $\mathbf{X}_u$ . The vector  $(\mathbf{w}, \mathbf{u})^\top$  has expectation  $(\mathbf{X}\boldsymbol{\beta}, \mathbf{X}_u\boldsymbol{\beta})^\top$  and covariance matrix  $\begin{bmatrix} \boldsymbol{\Sigma} & \boldsymbol{\Sigma}_{wu} \\ \boldsymbol{\Sigma}_{uw} & \boldsymbol{\Sigma}_{uu} \end{bmatrix}$ , where  $\boldsymbol{\Sigma} = \text{Var}(\mathbf{w}, \mathbf{w})$ ,  $\boldsymbol{\Sigma}_{wu} = \text{Var}(\mathbf{w}, \mathbf{u})$ ,  $\boldsymbol{\Sigma}_{uw} = \boldsymbol{\Sigma}_{wu}^\top$  and  $\boldsymbol{\Sigma}_{u,u} = \text{Var}(\mathbf{u}, \mathbf{u})$ . By assumption, we have observed  $\mathbf{w}$ , so we may derive the conditional distribution of  $\mathbf{u}|\mathbf{w}$ , which has the following properties:

$$\begin{aligned} E(\mathbf{u}|\mathbf{w}) &= \mathbf{X}_u\boldsymbol{\beta} + \boldsymbol{\Sigma}_{u,w}\boldsymbol{\Sigma}^{-1}(\mathbf{w} - \mathbf{X}\boldsymbol{\beta}) \\ E(\mathbf{u}|\mathbf{w}) &= \boldsymbol{\Sigma}_{u,u} - \boldsymbol{\Sigma}_{u,w}\boldsymbol{\Sigma}^{-1}\boldsymbol{\Sigma}_{w,u} \end{aligned}$$

Ver Hoef *et al.* (2024) show how these equations are adjusted to reflect uncertainty in both  $\hat{\boldsymbol{\beta}}$  and  $\hat{\mathbf{w}}$  while leveraging the laws of total expectation and variance yet again. They derive the predictor of  $\mathbf{u}$ ,  $\hat{\mathbf{u}}$ , and its associated variance, given by:

$$\begin{aligned} \hat{\mathbf{u}} &= \mathbf{X}_u\hat{\boldsymbol{\beta}} + \boldsymbol{\Sigma}_{u,w}\boldsymbol{\Sigma}^{-1}(\hat{\mathbf{w}} - \mathbf{X}\hat{\boldsymbol{\beta}}) \\ \text{Var}(\hat{\mathbf{u}}) &= \boldsymbol{\Sigma}_{u,u} - \boldsymbol{\Sigma}_{u,w}\boldsymbol{\Sigma}^{-1}\boldsymbol{\Sigma}_{w,u} + \mathbf{K}(\mathbf{X}^\top\boldsymbol{\Sigma}^{-1}\mathbf{X})^{-1}\mathbf{K}^\top + \boldsymbol{\Lambda}(-\mathbf{H})^{-1}\boldsymbol{\Lambda}^\top, \end{aligned}$$

where  $\mathbf{K} = \mathbf{X}_u - \boldsymbol{\Sigma}_{u,w}\boldsymbol{\Sigma}^{-1}\mathbf{X}$  and  $\boldsymbol{\Lambda} = \mathbf{X}_u\mathbf{B} + \boldsymbol{\Sigma}_{u,w}\boldsymbol{\Sigma}^{-1}(\mathbf{1} - \mathbf{X}\mathbf{B})$  for a vector of ones,  $\mathbf{1}$ . As with  $\hat{\boldsymbol{\beta}}$ , in practice these covariance matrices are evaluated at  $\hat{\boldsymbol{\theta}}$ .

## 3. Modeling moose presence in Alaska, USA

The `moose` data in **spmodel** contain information on moose (*Alces Alces*) presence in the Togiak region of Alaska, USA. `moose` is an `sf` object, a special data frame that is supplemented with spatial information using the `sf` package in R (Pebesma 2018). The first few rows of `moose` look like:

```
R> head(moose)
```

```
Simple feature collection with 6 features and 4 fields
Geometry type: POINT
```

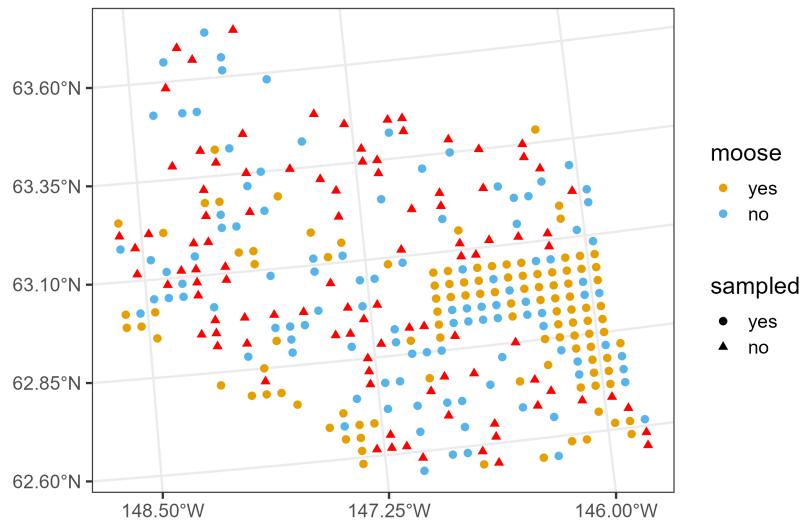


Figure 3: Moose presence in Alaska. Circles represent moose presence or absence (based on color) and triangles represent locations at which moose presence probability predictions are desired.

```
Dimension:      XY
Bounding box:  xmin: 281896.4 ymin: 1518398 xmax: 311325.3 ymax: 1541016
Projected CRS: NAD83 / Alaska Albers
# A tibble: 6 x 5
  elev strat count presence      geometry
  <dbl> <chr> <dbl> <fct>        <POINT [m]>
1 469. L     0 0   yes          (293542.6 1541016)
2 362. L     0 0   yes          (298313.1 1533972)
3 173. M     0 0   yes          (281896.4 1532516)
4 280. L     0 0   yes          (298651.3 1530264)
5 620. L     0 0   yes          (311325.3 1527705)
6 164. M     0 0   yes          (291421.5 1518398)
```

- 271 There are five columns: `elev`, the numeric site elevation (meters); `strat` a stratification  
 272 variable for sampling with two levels, "L" and "M", which are categorized by landscape metrics  
 273 at each site; `count`, the number of moose at each site; `presence`, a factor that indicates  
 274 whether at least one moose was observed at each site (0 implies no moose; 1 implies at least  
 275 one moose); and `geometry`, the NAD83/Alaska Albers (EPSG: 3338) projected coordinate of  
 276 each site (these data are point-referenced because each observation occurs at point coordinates  
 277 and are represented by a `POINT` geometry. Moose are most prevalent in the southwestern and  
 278 eastern parts of the Togiak region (Figure 3).  
 279 The `moose_preds` data in `spmodel` is an `sf` object with point locations at which moose  
 280 presence predictions are desired. Like `moose`, `moose_preds` contains `elev` and `strat` for each  
 281 site:

```
R> head(moose_preds)
```

```
Simple feature collection with 6 features and 2 fields
Geometry type: POINT
Dimension:      XY
Bounding box:   xmin: 291839.8 ymin: 1436192 xmax: 401239.6 ymax: 1512103
Projected CRS: NAD83 / Alaska Albers
# A tibble: 6 x 3
  elev strat      geometry
  <dbl> <chr>    <POINT [m]>
1 143. L     (401239.6 1436192)
2 324. L     (352640.6 1490695)
3 158. L     (360954.9 1491590)
4 221. M     (291839.8 1466091)
5 209. M     (310991.9 1441630)
6 218. L     (304473.8 1512103)
```

### 282 3.1. Model Fitting

283 SPGLMs in **spmodel** are fit using the `spglm()` function. The `spglm()` function requires  
 284 four arguments: `formula`, the relationship between the response and explanatory variables;  
 285 `family`, the response distribution assumed for the repsonse variable; `data`, the data frame  
 286 that contains the variables in `formula`, and `spcov_type`, the type of spatial covariance.  
 287 The `formula`, `family`, and `data` arguments are the three required arguments to `glm()` for  
 288 nonspatial GLMs. So, the transition from `glm()` to `spglm()` simply requires one additional  
 289 argument: `spcov_type`. When `data` is not an `sf` object, `spglm()` also requires the `xcoord`  
 290 and `ycoord` arguments, which indicate the columns in `data` that represent projected the x-  
 291 and y-coordinates, respectively.

292 We use `spglm()` to fit a spatial GLM (i.e., here, a spatial logistic regression) quantifying the  
 293 effect of elevation and strata on moose presence:

```
R> spbin <- spglm(
+   formula = presence ~ elev + strat,
+   family = binomial,
+   data = moose,
+   spcov_type = "exponential"
+ )
```

294 The `summary()` function returns a model summary that returns relevant information like the  
 295 function call, deviance residuals, a coefficients table of fixed effects, the pseudo R-squared,  
 296 spatial covariance parameter coefficient estimates, and the GLM dispersion parameter (fixed  
 297 at one in logistic regression):

```
R> summary(spbin)
```

Call:

```
spglm(formula = presence ~ elev + strat, family = binomial, data = moose,
```

```

spcov_type = "exponential")

Deviance Residuals:
    Min      1Q  Median      3Q     Max
-1.7535 -0.8005  0.3484  0.7893  1.5797

Coefficients (fixed):
            Estimate Std. Error z value Pr(>|z|)
(Intercept) -2.465713   1.486212 -1.659  0.097104 .
elev         0.006036   0.003525  1.712  0.086861 .
stratM       1.439273   0.420591  3.422  0.000622 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Pseudo R-squared: 0.06275

Coefficients (exponential spatial covariance):
      de      ie      range
5.145e+00 1.294e-03 4.199e+04

Coefficients (Dispersion for binomial family):
dispersion
      1

```

298 The model suggests that elevation is positively associated with the log odds of moose presence  
 299 ( $p$ -value  $\approx 0.087$ ), after controlling for strata. The model also suggests that moose have a  
 300 higher log odds of presence in the "M" strata compared to the "L" strata ( $p$ -value  $< 0.001$ ),  
 301 after controlling for elevation.

302 The fixed effects coefficients table from `summary()` is often of primary scientific interest, but  
 303 it is not immediately usable when printed directly to the R console. The `tidy()` function  
 304 tidies this table, turning it into a data frame (i.e., a tibble) with standard column names:

```

R> tidy(spbin, conf.int = TRUE)

# A tibble: 3 x 7
  term      estimate std.error statistic p.value conf.low conf.high
  <chr>        <dbl>     <dbl>      <dbl>    <dbl>     <dbl>      <dbl>
1 (Intercept) -2.47      1.49      -1.66  0.0971   -5.38e+0   0.447
2 elev         0.00604   0.00353     1.71  0.0869   -8.73e-4   0.0129
3 stratM       1.44       0.421      3.42  0.000622  6.15e-1    2.26

```

### 305 3.2. Model Comparison

306 The strength of spatial covariance in the data affects how beneficial a SPGLM is relative to  
 307 a GLM. When the spatial covariance is strong, the SPGLM should notably outperform the  
 308 GLM. When the spatial covariance is weak, the SPGLM and GLM should perform similarly.

309 We can quantify the benefits of incorporating spatial covariance for a particular data set  
310 by comparing the fit of a SPGLM to a GLM. We can fit a GLM in **spmodel** by specifying  
311 `spcov_type = "none"`:

```
R> bin <- spglm(
+   formula = presence ~ elev + strat,
+   family = binomial,
+   data = moose,
+   spcov_type = "none"
+ )
```

312 While the `spglm()` approach evaluates the HGLMM likelihood with  $\sigma_{de}^2 = 0$  and  $\sigma_{ie}^2 \approx 0$   
313 instead of just the GLM likelihood, the parameter estimates and their standard errors are the  
314 same:

```
R> bin_glm <- glm(
+   formula = presence ~ elev + strat,
+   family = binomial,
+   data = moose,
+ )
R> round(coef(bin), digits = 4)

(Intercept)      elev      stratM
-0.4247     -0.0003     0.8070

R> round(coef(bin_glm), digits = 4)

(Intercept)      elev      stratM
-0.4247     -0.0003     0.8070

R> round(sqrt(diag(vcov(bin))), digits = 4)

(Intercept)      elev      stratM
 0.4208      0.0019     0.2906

R> round(sqrt(diag(vcov(bin_glm))), digits = 4)

(Intercept)      elev      stratM
 0.4208      0.0019     0.2906
```

315 However, using `spglm()` instead of `glm()` ensures that **spmodel** helper functions are available  
316 and that each of the `spglm()` models uses the same likelihood:

```
R> glance(spin)
```

```
# A tibble: 1 x 10
  n      p    npar value    AIC   AICc    BIC logLik deviance
<int> <dbl> <int> <dbl> <dbl> <dbl> <dbl> <dbl>    <dbl>
1 218     3      3  676.  682.  683.  693. -338.     176.
# i 1 more variable: pseudo.r.squared <dbl>

R> glance(bin)

# A tibble: 1 x 10
  n      p    npar value    AIC   AICc    BIC logLik deviance
<int> <dbl> <int> <dbl> <dbl> <dbl> <dbl> <dbl>    <dbl>
1 218     3      0  708.  708.  708.  708. -354.     294.
# i 1 more variable: pseudo.r.squared <dbl>
```

317 The likelihood-based statistics AIC, AICc, BIC, and deviance are much lower for the SPGLM,  
 318 indicating a better fit relative to the GLM. We may also perform a likelihood ratio test (LRT)  
 319 between the two models, as the GLM is a special case of the SPGLM (i.e., is nested within  
 320 the SPGLM):

```
R> anova(spbin, bin)

Likelihood Ratio Test

Response: presence
          Df Chi2 Pr(>Chi2)
spbin vs bin  3 31.546 6.525e-07 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

321 The LRT provides strong evidence that the SPGLM is preferred to the GLM ( $p$ -value <  
 322 0.001).  
 323 An alternative approach to model comparison is to use a cross-validation procedure (James,  
 324 Witten, Hastie, and Tibshirani 2013). The `loocv()` function performs leave-one-out cross  
 325 validation, comparing the predicted mean (on the response scale) to the observed response  
 326 variable for each hold-out observation, recomputing estimates of  $\beta$  each iteration. Then, statis-  
 327 tics like bias, mean-squared-prediction error (MSPE), and the square root of MSPE (RMSPE)  
 328 can be used to evaluate models:

```
R> loocv(spbin)

# A tibble: 1 x 3
  bias  MSPE RMSPE
  <dbl> <dbl> <dbl>
1 0.0000206 0.156 0.394
```

```
R> loocv(bin)
```

```
# A tibble: 1 x 3
  bias  MSPE RMSPE
  <dbl> <dbl> <dbl>
1 -1.23e-9 0.240 0.490
```

329 Both models have negligible bias, but the SPGLM has much lower MSPE and RMSPE than  
 330 the GLM, indicating the SPGLM predictions are far more efficient. Three separate metrics  
 331 (likelihood-based statistics, likelihood-ratio test, and leave-one-out cross validation) prefer  
 332 the SPGLM to the GLM.

333 We can compare two SPGLMs with different spatial covariance functions using likelihood-  
 334 based statistics and leave-one-out cross validation, but we can't use the LRT because generally,  
 335 the spatial covariance functions aren't nested:

```
R> spbin2 <- update(spbin, spcov_type = "gaussian")
R> glances(spbin, spbin2)

# A tibble: 2 x 11
  model     n     p   npar value    AIC   AICc    BIC logLik deviance
  <chr> <int> <dbl> <int> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl>
1 spbin2    218     3     3  674.  680.  680.  690. -337.   198.
2 spbin     218     3     3  676.  682.  682.  683. -338.   176.
# i 1 more variable: pseudo.r.squared <dbl>

R> loocv(spbin)

# A tibble: 1 x 3
  bias  MSPE RMSPE
  <dbl> <dbl> <dbl>
1 0.0000206 0.156 0.394

R> loocv(spbin2)

# A tibble: 1 x 3
  bias  MSPE RMSPE
  <dbl> <dbl> <dbl>
1 -0.000261 0.146 0.382
```

336 The "exponential" spatial covariance (**spbin**) has a slightly lower (better) deviance but  
 337 slightly higher (worse) AIC, AICc, and BIC than the "gaussian" spatial covariance (**spbin2**).  
 338 Both spatial covariance functions have similar leave-one-out cross validation metrics, though  
 339 the "gaussian" spatial covariance RMSPE is slightly lower (better). For practical purposes,  
 340 these models fit similarly.

341 Often times in spatial statistics, the difference in model fit between the best spatial model  
 342 and worst spatial model is much smaller than the difference in model fit between the worst  
 343 spatial model and the nonspatial model, implying that accounting for some form of spatial  
 344 covariance is very beneficial. Two spatial covariance functions to consider starting with are the

<sup>345</sup> exponential and Gaussian, which have quite different origin behaviors (Figure 2), something  
<sup>346</sup> Stein (1999) argues is important to characterize accurately.

### <sup>347</sup> 3.3. Model Diagnostics

<sup>348</sup> `spmodel` provides a suite of tools for model diagnostics. One is `augment()`, which augments  
<sup>349</sup> the model data with diagnostics:

```
R> augment(spbin)

Simple feature collection with 218 features and 8 fields
Geometry type: POINT
Dimension:      XY
Bounding box:  xmin: 269085 ymin: 1416151 xmax: 419057.4 ymax: 1541016
Projected CRS: NAD83 / Alaska Albers
# A tibble: 218 x 9
   presence elev strat .fitted .resid     .hat   .cooksdi .std.resid
   * <fct>    <dbl> <chr>   <dbl>  <dbl>   <dbl>    <dbl>
 1 0          469. L      -1.95 -0.516  0.0476  0.00465  -0.528
 2 0          362. L      -2.70 -0.361  0.0123  0.000548  -0.363
 3 0          173. M      -1.96 -0.514  0.00455  0.000405  -0.516
 4 0          280. L      -3.15 -0.290  0.00413  0.000117  -0.291
 5 0          620. L      -1.19 -0.728  0.168   0.0427   -0.798
 6 0          164. M      -1.71 -0.576  0.00534  0.000598  -0.578
 7 0          164. M      -1.60 -0.606  0.00576  0.000714  -0.608
 8 0          186. L      -2.50 -0.397  0.00439  0.000233  -0.398
 9 0          362. L      -1.88 -0.532  0.0239   0.00237  -0.539
10 0         430. L      -1.54 -0.623  0.0497   0.00713  -0.639
# i 208 more rows
# i 1 more variable: geometry <POINT [m]>
```

<sup>350</sup> The fitted values (`.fitted`) can be returned on either the link ( $\hat{w}$ ) or response ( $f^{-1}(\hat{w})$ )  
<sup>351</sup> scale and the residuals (`.resid`) can deviance, Pearson, or response residuals. The default  
<sup>352</sup> fitted values are the link scale and the default residuals are deviance residuals. Also re-  
<sup>353</sup> turned by `augment()` are the leverage (`.hat`), Cook's distance (`.cooksdi`), and standardized  
<sup>354</sup> residuals `.std.resid`. A benefit of using `augment()` when data is an `sf` object is that the  
<sup>355</sup> output is also an `sf` object, which makes it straightforward to create spatial diagnostic plots  
<sup>356</sup> (Figure 4). Standard R helpers (e.g., `fitted()`, `residuals()`) are also available to extract  
<sup>357</sup> model diagnostics from the model object.

<sup>358</sup> The `plot()` function can also be used to return similar diagnostics as from `lm()` and `glm()`  
<sup>359</sup> with additional tools for diagnosing spatial covariance. For example, we can inspect Cook's  
<sup>360</sup> distance values and the empirical spatial covariance as a function of distance with (Figure 5):

```
R> plot(spbin, which = c(4, 7))
```

<sup>361</sup> The `varcomp()` function partitions model variability into several different components, help-  
<sup>362</sup> ing to elucidate the model's structure:

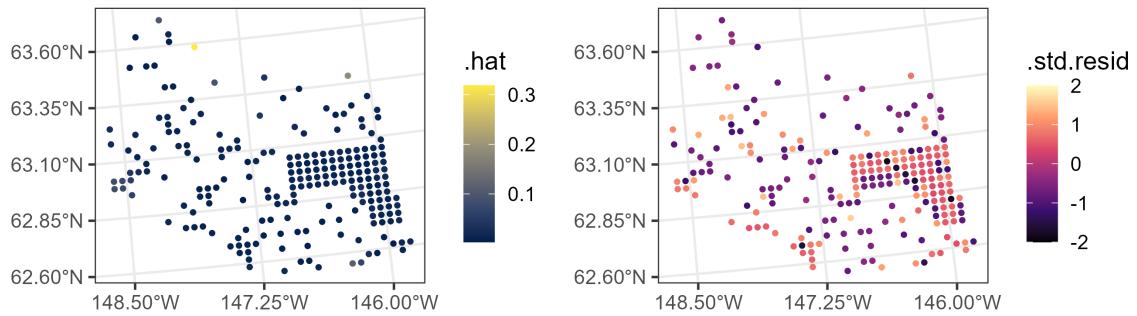


Figure 4: Moose presence model diagnostics, including leverage values (left) and standardized residuals (right).

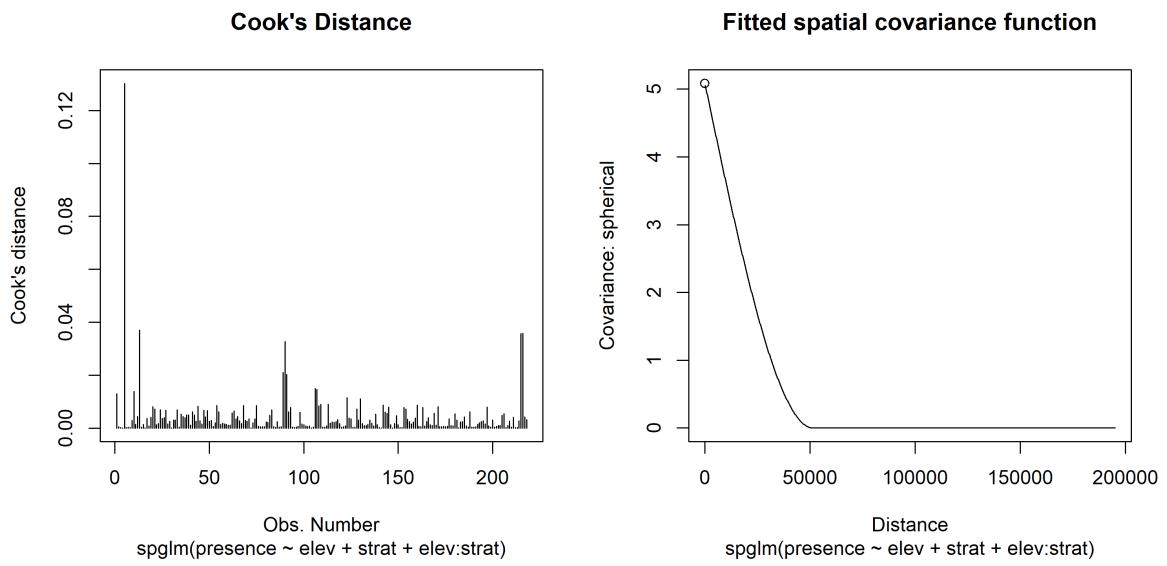


Figure 5: Moose presence model diagnostics, including Cook's distance and the fitted spatial covariance as a function of distance (right).

```
R> varcomp(spbin)

# A tibble: 3 x 2
  varcomp      proportion
  <chr>          <dbl>
1 Covariates (PR-sq) 0.0627
2 de            0.937
3 ie            0.000236
```

363 The pseudo R-squared ( $PR^2$ ) is reported in the first row. The remaining variability ( $1 - PR^2$ )  
 364 is allocated proportionally to `de` and `ie` according to  $\sigma_{de}^2$  and  $\sigma_{ie}^2$ . This variability partitioning  
 365 is a useful that helps quantify how much the explanatory variables, residual spatial variance,  
 366 and residual nonspatial variance contribute to model fit, but as with  $PR^2$ , should not be used  
 367 as a model comparison tool.

### 368 3.4. Prediction

369 We can predict the probability of moose presence using `predict()`:

```
R> predict(spbin, newdata = moose_preds)[1:5]
```

1	2	3	4	5
0.06664165	-0.79069107	-1.60387940	-0.83159357	1.38183928

370 By default, predictions are returned on the link scale, but this can be changed to the response  
 371 scale via `type`:

```
R> predict(spbin, newdata = moose_preds, type = "response")[1:5]
```

1	2	3	4	5
0.5166542	0.3120203	0.1674401	0.3033082	0.7992862

372 Predictions on the response scale are visualized alongside the fitted values ( $f^{-1}(\hat{\mathbf{w}})$ ) in  
 373 Figure 6.

374 Prediction intervals for the probability of moose presence (on the link scale) are returned by  
 375 supplying `interval`:

```
R> predict(spbin, newdata = moose_preds, interval = "prediction")[1:5, ]
```

	fit	lwr	upr
1	0.06664165	-2.0374370	2.1707203
2	-0.79069107	-3.4758514	1.8944692
3	-1.60387940	-4.0953329	0.8875741
4	-0.83159357	-3.0704818	1.4072947
5	1.38183928	-0.7692107	3.5328893

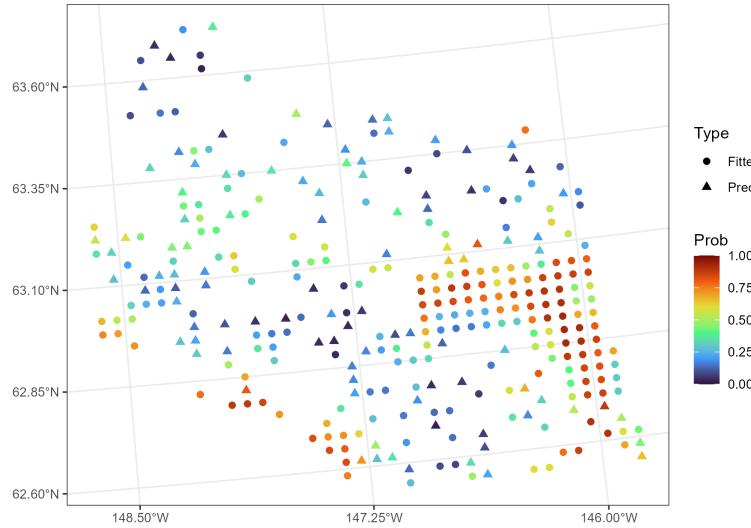


Figure 6: Moose presence probability fitted values and predictions. Fitted values are represented by circles and predictions by triangles.

376 We can alternatively use `augment()` to augment the prediction data with predictions. Arguments  
377 to `predict()` can also be passed to `augment()`:

```
R> augment(spbin, newdata = moose_preds, interval = "prediction")

Simple feature collection with 100 features and 5 fields
Geometry type: POINT
Dimension:      XY
Bounding box:  xmin: 269386.2 ymin: 1418453 xmax: 419976.2 ymax: 1541763
Projected CRS: NAD83 / Alaska Albers
# A tibble: 100 x 6
  elev strat .fitted .lower .upper      geometry
* <dbl> <chr>   <dbl>  <dbl>  <dbl>      <POINT [m]>
 1 143. L     0.0666 -2.04   2.17 (401239.6 1436192)
 2 324. L    -0.791  -3.48   1.89 (352640.6 1490695)
 3 158. L    -1.60   -4.10   0.888 (360954.9 1491590)
 4 221. M    -0.832  -3.07   1.41 (291839.8 1466091)
 5 209. M     1.38   -0.769  3.53 (310991.9 1441630)
 6 218. L    -2.59   -5.20   0.0177 (304473.8 1512103)
 7 127. L    -2.73   -5.24  -0.220 (339011.1 1459318)
 8 122. L    -2.32   -4.74   0.0920 (342827.3 1463452)
 9 191. L    -1.17   -4.01   1.66 (284453.8 1502837)
10 105. L   -0.905  -3.05   1.24 (391343.9 1483791)
# i 90 more rows
```

378 By using `augment()` when `newdata` is an `sf` object, predictions and their corresponding  
379 uncertainties are readily available for spatial mapping (Figure 7).

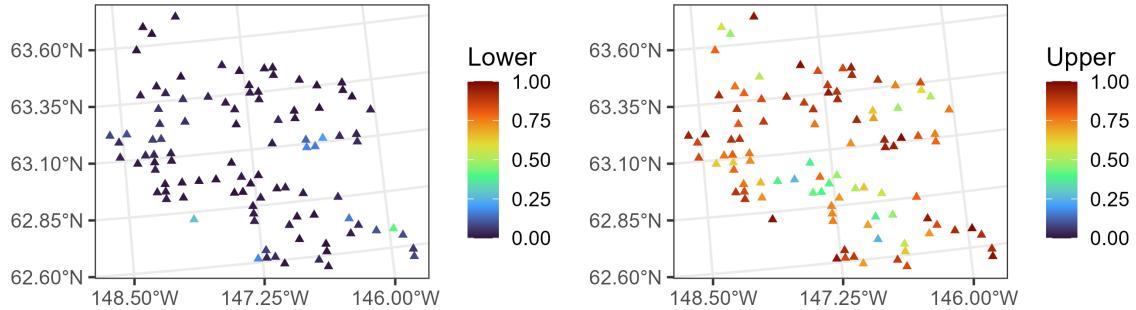


Figure 7: Moose presence probability prediction intervals. 95% prediction interval lower bound (left) and 95% prediction interval upper bound (right).

## 4. Additional applications

Throughout the remainder of this section, we briefly highlight some additional **spmodel** capabilities for SPGLMs. In Section 4.1, we fit Poisson and negative binomial models with and without geometric anisotropy for the point-referenced moose count data. In Section 4.2, we fit a Gamma model to the point-referenced lake conductivity data, showing how to perform a spatial analysis of variance (ANOVA), fit contrasts for models with interactions and use maximum likelihood to compare two models with different explanatory variables. In Section 4.3, we fit a binomial model to the areal harbor seal trend data with a nonspatial random effect. Finally in Section 4.4, we fit beta models to Texas voter turnout data, which can be treated as point-referenced or areal.

### 4.1. Modeling moose counts in Alaska, USA

In addition to moose presence, moose counts are also recorded in **moose** (Figure 8). The Poisson and negative binomial response distributions can be used to model SPGLMs for count data. The Poisson distribution mean is equal to its variance, while the negative binomial has an extra parameter to accommodate overdispersion (where the variance is larger than the mean). Using a spherical spatial covariance function, we may fit both a Poisson and negative binomial SPGLM changing the **family** argument:

```
R> sppois <- spglm(
+   formula = count ~ elev + strat,
+   family = poisson,
+   data = moose,
+   spcov_type = "spherical"
+ )
R> spnb <- update(sppois, family = nbinomial)
```

Because the Poisson and negative binomial distributions have the same support (nonnegative integers), we can compare them using AIC, AICc, or BIC:

```
R> BIC(sppois, spnb)
```

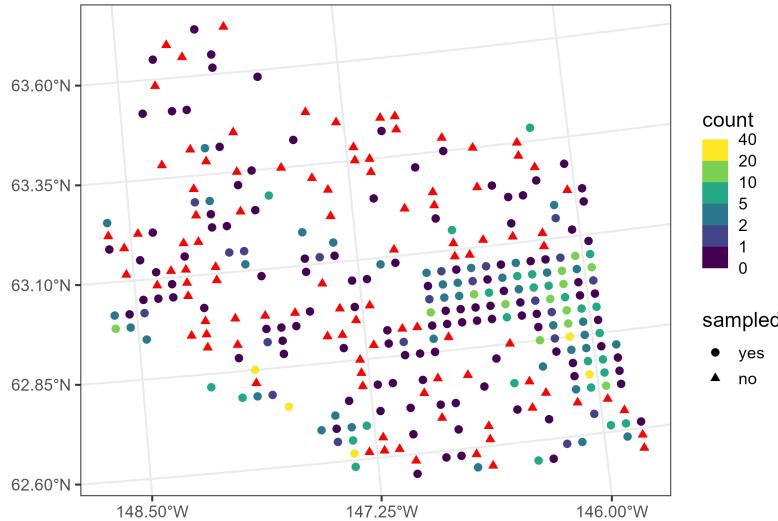


Figure 8: Moose counts in Alaska. Circles represent moose counts (based on color) and triangles represent locations at which mean count predictions are desired.

	df	BIC
sppois	3	1344.574
spnb	4	1343.105

398 Implicit in our spatial covariance functions thus far has been an assumption of geometric  
 399 isotropy. A spatial covariance function is geometrically isotropic if it decays with distance at  
 400 the same rate in all directions (Figure 9; left). A spatial covariance is geometrically isotropic  
 401 if it decays with distance at different rates in different directions (Figure 9; right). Geometric  
 402 anisotropy is formally incorporated by rotating and scaling original coordinates, yielding  
 403 transformed coordinates that are geometrically isotropic:

$$\begin{bmatrix} x^* \\ y^* \end{bmatrix} = \begin{bmatrix} 1 & 0 \\ 0 & 1/\omega \end{bmatrix} \begin{bmatrix} \cos(\alpha) & \sin(\alpha) \\ -\sin(\alpha) & \cos(\alpha) \end{bmatrix} \begin{bmatrix} x \\ y \end{bmatrix}.$$

404 The parameters  $\omega$  and  $\alpha$  controls the scaling and rotation, respectively, of the major and minor  
 405 axes of a level curve of equal correlation (Figure 9). Using these transformed coordinates, the  
 406 partial sill ( $\sigma_{de}^2$ ), nugget ( $\sigma_{ie}^2$ ), and range ( $\phi$ ) parameters are estimated. We accommodate  
 407 geometric anisotropy by supplying `anisotropy`:

```
R> sppois_anis <- update(sppois, anisotropy = TRUE)
R> spnb_anis <- update(spnb, anisotropy = TRUE)
```

408 According to BIC, the spatial negative binomial model with geometric anisotropy performs  
 409 best:

```
R> BIC(sppois, spnb, sppois_anis, spnb_anis)
```

	df	BIC
sppois	3	1344.574

```
spnb           4 1343.105
sppois_anis  5 1341.143
spnb_anis    6 1339.714
```

- <sup>410</sup> The `plot()` function can be used to visualize the anisotropy (Figure 9):

```
R> plot(spnb, which = 8)
R> plot(spnb_anis, which = 8)
```

- <sup>411</sup> The spatial correlation is strongest in a northwest-southeast direction and weakest in the  
<sup>412</sup> northeast-southwest direction (Figure 9), which is intuitive given the similar patterns in moose  
<sup>413</sup> counts from Figure 8.

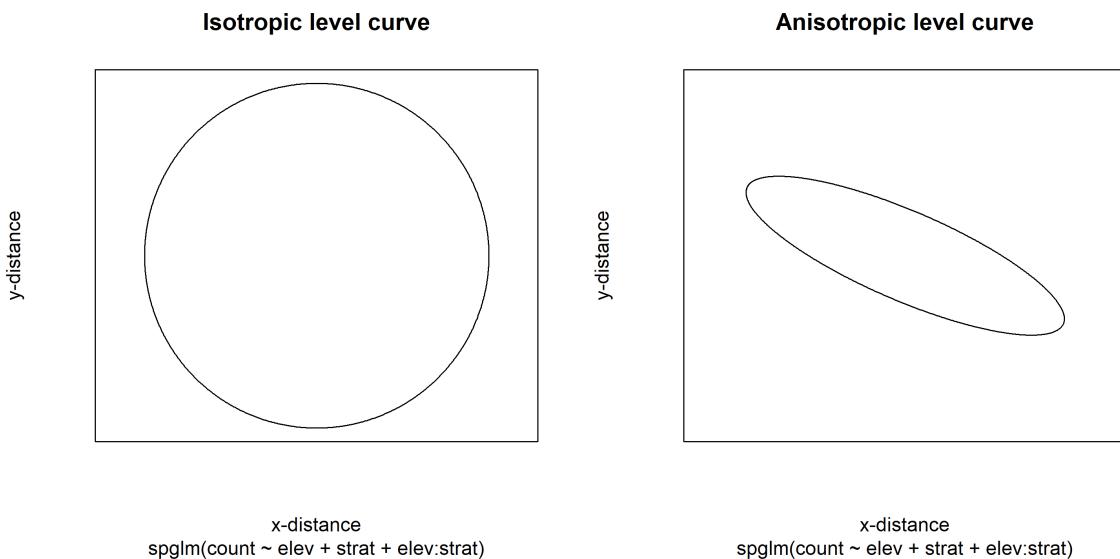


Figure 9: Level curves of equal correlation for the negative binomial moose count models. The ellipse is centered at zero distance in the x-direction and y-direction, and points along the ellipse have equal levels of correlation. In the isotropic level curve (left), spatial covariance decays equally in all directions. In the anisotropic level curve (right), spatial covariance decays fastest in the northeast-southwest direction and slowest in the northwest-southeast direction (this pattern can be seen in the observed counts).

<sup>414</sup> **4.2. Modeling lake conductivity in Southwest, USA**

- <sup>415</sup> The `lake` data in `spmodel` contains climate and chemical data for several lakes in four south-  
<sup>416</sup> western states in the United States: Arizona, Colorado, Nevada, and Utah. We desire an  
<sup>417</sup> SPGLM that characterizes the effect of temperature, state, and lake origin (whether the lake  
<sup>418</sup> is naturally occurring or human made) on lake conductivity. Conductivity is a measure of  
<sup>419</sup> dissolved ions (measured here in water), which is important for various physical, chemical,  
<sup>420</sup> and biological processes. Chemical data is often heavily right-skewed, so we model it using  
<sup>421</sup> an SPGLM assuming a Gamma distribution for the response. The `log_cond` variable in `lake`

<sup>422</sup> is the logarithm of conductivity, which we dynamically exponentiate within `formula` so that  
<sup>423</sup> it is on the original scale:

```
R> spgam <- spglm(
+   formula = exp(log_cond) ~ temp + state + temp:state + origin,
+   family = "Gamma",
+   data = lake,
+   spcov_type = "cauchy",
+   partition_factor = ~ year
+ )
```

<sup>424</sup> We model conductivity as a function of temperature, state, and lake origin, and we allow the  
<sup>425</sup> effect of temperature to vary by state (`temp:state`). The `year` partition factor (specified via  
<sup>426</sup> `partition_factor`) restricts spatial covariance to be nonzero only for observations sampled  
<sup>427</sup> during the same year. Data were collected in 2012 and 2017, so this partition factor assumes  
<sup>428</sup> independence between observations in 2012 and 2017. While we used the partition factor here  
<sup>429</sup> illustratively, more generally, the utility of partition factors can be highly context dependent.  
<sup>430</sup> When categorical variables have more than two levels, the default reference group contrasts  
<sup>431</sup> are not well-suited to assess the variable's overall significance:

```
R> summary(spgam)
```

Call:

```
spglm(formula = exp(log_cond) ~ temp + state + temp:state + origin,
      family = "Gamma", data = lake, spcov_type = "cauchy", partition_factor = ~year)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.35762	-0.20796	-0.03706	0.17869	1.10616

Coefficients (fixed):

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	3.59325	0.50058	7.178	7.06e-13 ***
temp	0.15182	0.03006	5.051	4.39e-07 ***
stateCO	-0.03214	0.56098	-0.057	0.95432
stateNV	0.75664	0.66851	1.132	0.25771
stateUT	-0.19696	0.55916	-0.352	0.72466
originNATURAL	0.08313	0.21988	0.378	0.70538
temp:stateCO	0.13679	0.04808	2.845	0.00444 **
temp:stateNV	0.01882	0.05820	0.323	0.74645
temp:stateUT	0.20015	0.04846	4.131	3.62e-05 ***

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Pseudo R-squared: 0.7061

```

Coefficients (cauchy spatial covariance):
      de      ie      range      extra
2.069e-02 2.952e-01 4.119e+06 5.645e-01

Coefficients (Dispersion for Gamma family):
dispersion
      3.761

```

- 432 A more effective approach is to use an analysis of variance (ANOVA), which is well-suited to  
 433 asses the overall significance of each variable:

```
R> anova(spgam)
```

```
Analysis of Variance Table
```

```

Response: exp(log_cond)
      Df    Chi2 Pr(>Chi2)
(Intercept) 1 51.5270 7.062e-13 ***
temp         1 25.5146 4.390e-07 ***
state        3  3.0747 0.3802528
origin       1  0.1429 0.7053819
temp:state   3 19.7668 0.0001897 ***
---
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

- 434 The main effect for temperature and the temperature by state interaction are highly significant  
 435 ( $p$ -value  $< 0.001$ ), while the main effects for state and lake origin are not significant. The  
 436 ANOVA table can be tidied using `tidy()`.  
 437 Variance inflation factors assess the degree to which standard errors  $\hat{\beta}$  are inflated due to  
 438 covariance among the columns of  $\mathbf{X}$ . Generalized variance inflation factors can capture the  
 439 variance inflation for subsets of  $\mathbf{X}$  that may include categorical variables with more than two  
 440 levels (Fox and Monette 1992):

```
R> library(car)
```

```
R> vif(spgam)
```

	GVIF	Df	GVIF^(1/(2*Df))
temp	4.691914	1	2.166083
state	127.082397	3	2.242234
origin	1.264940	1	1.124695
temp:state	76.387383	3	2.059856

- 441 The  $GVIF^{1/2df}$  values for `temp`, `state`, and `temp:state` are just greater than two, which  
 442 suggests moderate multicollinearity for these terms – unsurprising given the `temp:state`

<sup>443</sup> interaction in the model. The GVIF<sup>1/2df</sup> for **origin** is close to one, which suggests little to  
<sup>444</sup> no multicollinearity for this term.

<sup>445</sup> Because of the interaction between **temp** and **state**, contrasts that assess mean differences  
<sup>446</sup> among states should condition upon a specific temperature value. By default, **emmeans** uses  
<sup>447</sup> the mean temperature value (here, 7.63) to assess contrasts:

```
R> library(emmeans)

R> pairs(emmeans(spgam, ~ state / temp))

temp = 7.63:
contrast estimate   SE  df z.ratio p.value
AZ - CO    -1.012 0.337 Inf  -3.004  0.0142
AZ - NV     -0.900 0.348 Inf  -2.584  0.0480
AZ - UT     -1.331 0.326 Inf  -4.082  0.0003
CO - NV      0.112 0.258 Inf   0.434  0.9727
CO - UT     -0.319 0.223 Inf  -1.427  0.4822
NV - UT     -0.431 0.244 Inf  -1.763  0.2915

Results are averaged over the levels of: origin
Degrees-of-freedom method: asymptotic
Results are given on the log (not the response) scale.
P value adjustment: tukey method for comparing a family of 4 estimates
```

<sup>448</sup> Again, because of the interaction between **temp** and **state**, we should assess temperature  
<sup>449</sup> trends separately for each state:

```
R> emtrends(spgam, ~ state, var = "temp")

state temp.trend     SE  df asymp.LCL asymp.UCL
AZ        0.152 0.0301 Inf    0.0929    0.211
CO        0.289 0.0370 Inf    0.2161    0.361
NV        0.171 0.0504 Inf    0.0718    0.270
UT        0.352 0.0372 Inf    0.2791    0.425

Results are averaged over the levels of: origin
Degrees-of-freedom method: asymptotic
Results are given on the exp (not the response) scale.
Confidence level used: 0.95
```

#### <sup>450</sup> 4.3. Modeling harbor seal trends in Alaska, USA

<sup>451</sup> The **seal** data in **spmodel** contains harbor seal abundance trends for two different harbor  
<sup>452</sup> seal stocks (genetically distinct populations). While the **moose** and **lake** data were point-  
<sup>453</sup> referenced, the **seal** data are areal. Each polygon in the **seal** data represents a distinct

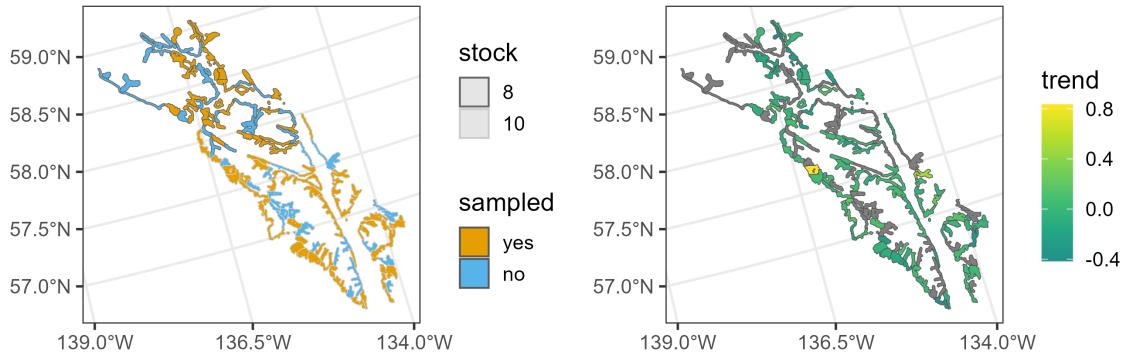


Figure 10: Seal trend distribution in Alaska. Observed and missing seal polygons by stock (left) and observed log seal trends (right).

454 harbor seal haulout location (Figure 10). A haulout location is a spot on coastal rocks that  
 455 harbor seals go to rest, molt, and give birth.  
 456 For each polygon, a Poisson regression was used to quantify the mean trend in abundance  
 457 over approximately 30 years (Ver Hoef, Peterson, Hooten, Hanks, and Fortin 2018). If the  
 458 logarithm of mean abundance trends (`log_trend`) is negative (positive), it means abundance  
 459 is decreasing (increasing). We use a binomial SPGLM to quantify the likelihood that mean  
 460 abundance trends are decreasing:

```
R> is_decreasing <- seal$log_trend < 0
R> spbin <- spgautor(
+   formula = is_decreasing ~ 1,
+   family = binomial,
+   data = seal,
+   spcov_type = "car",
+   random = ~ stock
+ )
```

461 To model spatial dependence, we used a conditional autoregressive function. Conditional  
 462 and simultaneous autoregressive functions characterize spatial distance through neighborhood  
 463 relationships (rather than Euclidean distance) and have `spcov_type` values of "car" and  
 464 "sar", respectively. By default, Queen's distance is used to determine whether two sites are  
 465 neighbors, though custom neighborhood matrices are passed via `W`. Row standardization is  
 466 also assumed by default; this can be changed via `row_st`. Using `random`, we also specified a  
 467 nonspatial random effect for seal stock. The `random` argument uses similar syntax as `lme4`  
 468 (Bates, Mächler, Bolker, and Walker 2015) and `nlme` (Pinheiro and Bates 2006) to specify  
 469 nonspatial random effects.

470 Tidying the model reveals the estimates and confidence intervals on the log odds scale:

```
R> tidy(spbin, conf.int = TRUE)
# A tibble: 1 x 7
  term      estimate std.error statistic p.value conf.low conf.high
```

<chr>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>
1 (Intercept)	0.340	0.673	0.506	0.613	-0.979	1.66

- <sup>471</sup> Back-transforming the confidence interval to the probability scale yields:

```
R> emmeans(spbin, ~ 1, type = "response")

1      prob    SE  df asympt.LCL asympt.UCL
overall 0.584 0.164 Inf     0.273      0.84
```

Degrees-of-freedom method: asymptotic  
 Confidence level used: 0.95  
 Intervals are back-transformed from the logit scale

- <sup>472</sup> The **SE** column is the standard error on the response scale obtained from the delta method  
<sup>473</sup> (Oehlert 1992).

- <sup>474</sup> In contrast to point-referenced data, prediction locations for areal data must be specified  
<sup>475</sup> at the time of model fitting, as they affect the spatial covariance function's neighborhood  
<sup>476</sup> structure. Prediction locations whose response values have an **NA** (i.e., missing) value are  
<sup>477</sup> converted into a **newdata** object that is stored in the model output. For example, rows one  
<sup>478</sup> and nine are locations without seal trends, meaning they are not used in model fitting but  
<sup>479</sup> are desired for prediction:

```
R> seal

Simple feature collection with 149 features and 2 fields
Geometry type: POLYGON
Dimension:      XY
Bounding box:  xmin: 913618.8 ymin: 855730.2 xmax: 1221859 ymax: 1145054
Projected CRS: NAD83 / Alaska Albers
# A tibble: 149 x 3
  log_trend stock                         geometry
* <dbl>   <fct>                         <POLYGON [m]>
 1 NA     8    ((1035002 1054710, 1035002 1054542, 1035002 105354~
 2 -0.282 8    ((1037002 1039492, 1037006 1039490, 1037017 103949~
 3 -0.00121 8   ((1070158 1030216, 1070185 1030207, 1070187 103020~
 4  0.0354 8    ((1054906 1034826, 1054931 1034821, 1054936 103482~
 5 -0.0160 8    ((1025142 1056940, 1025184 1056889, 1025222 105683~
 6  0.0872 8    ((1026035 1044623, 1026037 1044605, 1026072 104461~
 7 -0.266  8    ((1100345 1060709, 1100287 1060706, 1100228 106070~
 8  0.0743 8    ((1030247 1029637, 1030248 1029637, 1030265 102964~
 9  NA     8    ((1043093 1020553, 1043097 1020550, 1043101 102055~
10 -0.00961 8   ((1116002 1024542, 1116002 1023542, 1116002 102254~

# i 139 more rows
```

- <sup>480</sup> Then, **predict()** can be called without having to specify **newdata**:

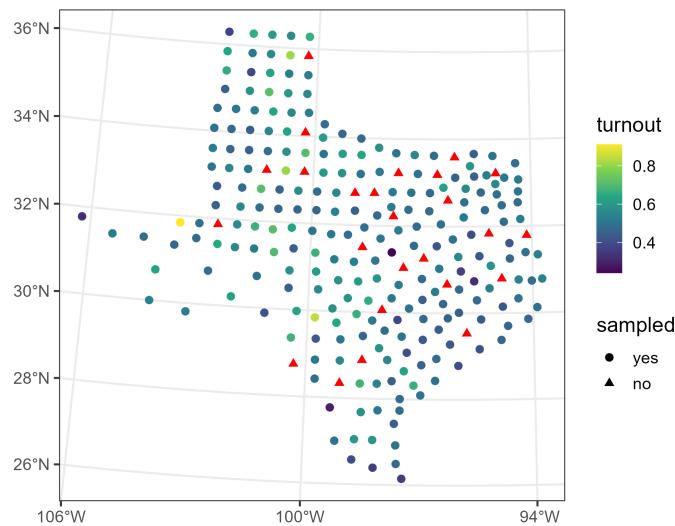


Figure 11: Proportion of voter turnout in Texas for the 1980 presidential election. Circles represent voter turnout (based on color) and triangles represent locations at which voter turnout predictions are desired.

```
R> predict(spbin, type = "response", interval = "prediction")[1:5, ]
```

	fit	lwr	upr
1	0.6807677	0.3863736	0.8783808
9	0.5945680	0.2467634	0.8678078
13	0.6189055	0.2974432	0.8616799
15	0.6040102	0.2921802	0.8493132
18	0.6375700	0.3356282	0.8596641

- 481 We could have alternatively used a (geostatistical) SPGLM via `spglm()`. When areal data are  
 482 used with `spglm()`, the centroids of each polygon are used as the point-referenced coordinates.  
 483 We further explore comparisons between point-referenced and aerial data in the next example.

#### 484 4.4. Modeling voter turnout in Texas, USA

- 485 The `texas` data in `spmodel` contains voter turnout data for Texas counties in the 1980 United  
 486 States Presidential Election (Bivand, Nowosad, and Lovelace 2024). The data are point-  
 487 referenced, with polygon centroids representing the spatial location of each county (Figure 11).  
 488 Beta regression is a GLM used to model rate and proportion data in the (0, 1) interval (Ferrari  
 489 and Cribari-Neto 2004; Cribari-Neto and Zeileis 2010). We model voter turnout rates as a  
 490 function of mean log income of county residents using an SPGLM assuming a beta distributed  
 491 response variable:

```
R> spbeta_geo <- spglm(  
+   formula = turnout ~ log_income,  
+   family = "beta",  
+   data = texas,
```

```
+   spcov_type = "matern"
+ )
```

492 Alternatively, we could use an autoregressive model to fit the model, constructing a neighborhood matrix by assuming centroids within cutoff of one another are neighbors:

```
R> spbeta_auto <- spgautor(
+   formula = turnout ~ log_income,
+   family = "beta",
+   data = texas,
+   spcov_type = "car",
+   cutoff = 1e5
+ )
```

494 According to AIC, the SPGLM for point-referenced data is preferred:

```
R> AIC(spbeta_geo, spbeta_auto)

      df      AIC
spbeta_geo  5 -44.53113
spbeta_auto  3 -22.46104
```

495 The default estimation method in **spmodel** is restricted maximum likelihood (REML). One  
496 drawback of REML is that likelihood-based statistics are only valid for model comparison  
497 when the models have the same explanatory variable and fixed effect structure. This is  
498 because the error contrasts used to construct the REML likelihood change based on **X** and  
499  $\beta$ . An alternative is to use maximum likelihood, which can use likelihood-based statistics  
500 to compare models with different explanatory variable and fixed effect structures. While we  
501 could assess the significance of log turnout with the **spbeta\_geo** model fit with REML (e.g.,  
502 via **tidy()**), an alternative approach is to use a likelihood ratio test:

```
R> spbeta_full_ml <- update(spbeta_geo, estmethod = "ml")
R> spbeta_red_ml <- update(spbeta_geo, estmethod = "ml", formula = turnout ~ 1)
R> anova(spbeta_full_ml, spbeta_red_ml)
```

#### Likelihood Ratio Test

Response: turnout	Df Chi2 Pr(>Chi2)
spbeta_red_ml vs spbeta_full_ml	1 23.155 1.494e-06 ***
---	
Signif. codes:	0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

503 The likelihood ratio test suggests that log income is significantly related to voter turnout  
504 (*p*-value < 0.001). We could instead use a different likelihood-based statistic like AIC:

```
R> AIC(spbeta_full_ml, spbeta_red_ml)
```

	df	AIC
spbeta_full_ml	7	-31.25900
spbeta_red_ml	6	-10.10354

505 The AIC prefers the full model, which suggests that log income is important for predicting  
 506 voter turnout.

## 5. Discussion

507 SPGLMs are fit in **spmodel** using a novel application of the Laplace approximation that  
 508 simultaneously marginalizes over the latent (i.e., unobserved) random effects and the fixed  
 509 effects,  $\beta$ . The approach is very flexible and accommodates general response distributions and  
 510 covariance structures, though here we focus on spatial applications. **spmodel**'s `spglm()` and  
 511 `spgautor()` fit SPGLMs that are similar in structure and syntax as base R's `glm()` function,  
 512 easing the transition from GLMs to SPGLMs for practitioners. The `spglm()` and `spgautor()`  
 513 functions support six response distributions for binary, count, and skewed data and 20 spatial  
 514 covariance functions. **spmodel** also provides a suite of tools for data visualization, inference,  
 515 model diagnostics, and prediction, providing a framework that can be used for all stages of a  
 516 data analysis. There are many additional **spmodel** features that are not covered here, includ-  
 517 ing fitting multiple models simultaneously, fixing spatial covariance and dispersion parameters  
 518 at known values, fitting models to large non-Gaussian data having thousands of observations  
 519 via spatial indexing (Ver Hoef, Dumelle, Higham, Peterson, and Isaak 2023), incorporating  
 520 spatial dependence in machine learning (e.g., random forests; Breiman (2001)), simulating  
 521 spatially dependent data (e.g., `spbinom()`, `sprpois()`, etc.), and more. Further details are  
 522 provided by <https://CRAN.R-project.org/package=spmodel> and links therein.

## Computational details

523 The results in this paper were obtained using R 4.4.0 with the **spmodel** 0.11.0 package. Figures  
 524 were created using the `ggplot2` 3.5.1 package (Wickham 2016) and base R.

## Data and code availability

525 All writing and code associated with this manuscript is available for viewing and download on  
 526 GitHub at <https://github.com/USEPA/spmodel.glm.manuscript>. All data used are part  
 527 of the **spmodel** R package available for download from CRAN at <https://CRAN.R-project.org/package=spmodel>.

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 531 The views expressed in this manuscript are those of the authors and do not necessarily  
 532 represent the views or policies of the U.S. Environmental Protection Agency or the National

533 Oceanic and Atmospheric Administration. Any mention of trade names, products, or services  
 534 does not imply an endorsement by the U.S. government, the U.S. Environmental Protection  
 535 Agency, or the National Oceanic and Atmospheric Administration. The U.S. Environmental  
 536 Protection Agency and the National Oceanic and Atmospheric Administration do not endorse  
 537 any commercial products, services or enterprises.

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