Applied Survival Analysis Using R Chapter 12: Additional Topics

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1 Using Piecewise Constant Hazards to Model Survival Data

2 Interval Censoring

3 The Lasso Method for Selecting Predictive Biomarkers



What is "piecswise"?

- The piecewise exponential model is a generalization of the exponential which can offer considerable flexibility for modeling.
- "pieces" means the survival time axis was divided into multiple intervals and the hazard is constant.
- The piecewise exponential is that the likelihood is equivalent to a Poisson likelihood.
- If it is a single piece, that means it's just an ordinary exponential distribution with rate parameter λ.

$$L_{e}(\lambda) = \prod_{i=1}^{n} h(t_{i})^{\delta_{i}} S(t_{i}) = \prod_{i=1}^{n} \lambda^{\delta_{i}} e^{-\lambda t_{i}} = \lambda^{d} e^{-\lambda V}$$
 (1)

where t_i is the failure time of the *i*th subject, $V = \sum t_i$ is the total time at risk.

Poisson Distribution

- In Chapter 2, the m.l.e is given by $\hat{\lambda} = d/V$, now we suppose the d(the number of death) has a *Poisson distribution* with mean μ , so $\mu = V\lambda$.
- The likelihood function for a Poisson distribution is:

$$L_p(\lambda) = (\lambda V)^d e^{-\lambda V} = \lambda^d e^{-\lambda V} \cdot V^d$$
 (2)

Clearly the Poisson likelihood L_p is *proportional* to the exponential likelihood L_e , the constant multiple being V^d

Notations

- Suppose that we divide the time axis into contiguous intervals using cut points 0, c_1 , c_2 ,..., c_k .
- For each subject, say i, we denote the time spent in each interval by t_{i1} , t_{i2} ,..., $t_{ik'}$, where k' denotes the time interval subject i dies, or the *largest* time interval subject i is still known to be alive.
- Define δ_{ij} to be 0 for each interval j in which the ith subject is known to be alive, and 1 for interval k if the subject died in that interval.
- Then for patient i, the survival time of that patient is $t_i = \sum_{j=1}^k t_{ij}$ and the censoring indicator is $\delta_i = \sum_{j=1}^k \delta_{ij}$.

Formulas

Assume a proportional hazards model

$$\lambda_i(t_i,\beta) = \lambda_0(t_i)e^{z_i\beta} \tag{3}$$

where now the *baseline hazard* is a *piecewise* exponential, $\lambda_0(\mu) = \lambda_j$, j being jth interval, the one in which μ falls, so the full likelihood is:

$$L_{pe}(\lambda_1, \lambda_2, ..., \lambda_k, \beta) = \prod_{i=1}^n \prod_{j=1}^{k'(i)} \lambda_{ij}^{\delta_{ij}} e^{-\lambda_{ij} t_{ij}}$$
(4)

where $\lambda_{ij} = \lambda_j e^{z_i \beta}$, which is the proportional hazards assumption for the piecewise exponential

• With the single exponential, treat the censoring indicators δ_{ij} as a Poisson distribution with mean $\lambda_{ij}t_{ij}$

m.l.e

 so the maximum likelihood estimates obtained from the Poisson model with

$$\log(\lambda_{ij}) = \log(\lambda_j) + z_i \beta = \alpha_j + z_i \beta \tag{5}$$

so for the piecewise exponential proportional hazards model

$$\log(\lambda_{ij}t_{ij}) = \log(\lambda_j) + z_i\beta + \log(t_{ij}) = \alpha_j + z_i\beta + \log(t_{ij})$$
 (6)

where the α_j are the logs of the baseline hazard coefficients and β , is the log of the hazard ratio for z_i , The constant log.tij/ is called an *offset*.

Consider a synthetic example in Chapter 4.

```
1 > tt <- c(6, 7, 10, 15, 19, 25)
2 > delta <- c(1, 0, 1, 1, 0, 1)
3 > trt <- c(0, 0, 1, 0, 1, 1)
4 > id <- 1:6
5 > simple <- data.frame(id, tt, delta, trt)
6 > simple
7    id tt delta trt
8    1    1    6    1    0
9    2    2    7    0    0
10    3    3    10    1    1
11    4    4    15    1    0
12    5    5    19    0    1
13    6    6    25    1    1
```

 Define three intervals that we will use to define a piecewise exponential distribution, and partition the data into events and person-weeks per interval:

Fig. 12.1 Survival times of a synthetic data set. The intervals defined by the cut points as are shown using vertical defined lines

Patient 1 (C)

Patient 2 (C)

Patient 4 (C)

Patient 6 (T)

Patient 6 (T)

Patient 6 (T)

```
1 > tau.s <- c(0, 8, 16, 30)
2 > simple.split.s <- survSplit(data=simple, cut=tau.s, end="tt",
3 start="t0", event="delta", episode="diagGrp")
4 > simple.split.s$expo <- simple.split.s$tt - simple.split.s$t0
5 > ord <- order(simple.split.s$id)
6 > simple.split.ord <- simple.split.s[ord,]
7 > simple.split.ord
```

1		id	tt	delta	trt	t0	diagGrp	expo
2	7	1	6	1	0	0	1	6
3	8	2	7	0	0	0	1	7
4	9	3	8	0	1	0	1	8
5	15	3	10	1	1	8	2	2
6	10	4	8	0	0	0	1	8
7	16	4	15	1	0	8	2	7
8	11	5	8	0	1	0	1	8
9	17	5	16	0	1	8	2	8
10	23	5	19	0	1	16	3	3
11	12	6	8	0	1	0	1	8
12	18	6	16	0	1	8	2	8
13	24	6	25	1	1	16	3	9

 To obtain parameter estimates for the piecewise exponential distribution in (6)

```
1 > result.simple.poisson <- glm(delta ~ -1 + factor(diagGrp)+trt+
2 offset(log(expo)), family=poisson, data=simple.split.ord)
3 > summary(result.simple.poisson)
```

```
Coefficients:
                             Estimate
                                       Std. Error
                                                   z value
                                                             Ρr
(> |z|)
factor (diagGrp) 1
                   -3.2942
                            1.0370
                                       -3.177
                                                   0.00149 **
factor (diagGrp) 2
                   -1.7463
                            0.8569
                                       -2.038
                                                  0.04156 *
factor (diagGrp) 3
                  -1.0912
                            1.5949
                                       -0.684
                                                  0.49389
                   -1.3937
                            1.2425
                                       -1.122
                                                   0.26199
t.rt.
Signif. codes: 0***0.001 **0.01 * 0.05 . 0.1
```

 The "-1" in the model definition tells R to fit a separate term for each of the three interval factors (rather than to consider the first level as a reference level), more details see book!

Right Censoring and Interval Censoring

 Right censoring occurs naturally in clinical trials in Chapter 1, and the general likelihood function:

$$L(\beta) = \prod_{i \in D} f(t_i) \cdot \prod_{i \in C} S(t_i)$$
 (7)

where *D* represents the set of all subjects who fail and *R* the set of all who are right-censored.

• Suppose that some of the observations are *interval-censored*, so that, patient k has an event that lies between times L_k and R_k .

$$L(\beta) = \prod_{i \in D} f(t_i) \cdot \prod_{i \in C} [S(t_i) - S(R_i)]$$
 (8)



- If an event is only known to have occurred before a particular time R_i , this is known as *left censoring*, and accommodated by setting $L_i = 0$
- A special optimization technique known as the Expectation -Maximization algorithm to get m.l.e.

Breast Cosmesis Study

Fit a proportional hazards model to interval-censored data, and illustrated it using data from a breast cosmesis study. In this study, 94 breast cancer patients treated with radiation therapy with or without adjuvant chemotherapy were followed to determine the time until cosmetic deterioration (specifically, the appearance breast retraction) of the treated breast. Since patients were evaluated at office visits separated by a number of months, the data were interval-censored.

```
library(interval)
  data (bcos)
 bcos[c(1,33, 47, 62, 90),]
       left.
                right
                           t.reatment
        4.5
                   Tnf
                                 Rad
33
                                 Rad
                    12
                             RadChem
62
        14
                    17
                             RadChem
90
        16
                    60
                             RadChem
```

 Patient 47 received radiation and adjuvant chemotherapy, had not had the event at an office visit at 8 months, but the breast retraction had been observed at the next visit four months later. Thus, the event took place sometime between 8 and 12 months



obtain m.l.e of the survival distributions and plot:

```
1 > icout <- icfit(Surv(left,right,type="interval2")~treatment,
2 data=bcos, conf.int=F)
3 > plot(icout, XLAB="Time in months", YLAB="Survival probability",
4 COL=c("lightblue", "pink"), LEGEND=F, estpar=list(col=
5 c("blue", "red"), lwd=2, lty=1))
6 > legend("bottomleft",legend=c("Radiation alone", "Radiation and
7 chemo"), col=c("blue", "red"), lwd=2)
```

• fit a Weibull proportional hazards model to the interval-censored data, first we must define modified left and right endpoints of the intervals and set a maximum possible time.

```
> bcos <- within(bcos, {</pre>
left.alt <- left
left.alt[left == 0] <- 0.1
right.alt <- right
right.alt[is.infinite(right)] <- 65})
> bcos[c(1,33,47,62,90),]
       left
                right
                         treatment
                                        right.alt
                                                     left.alt
        45
                                               65
                                                          45.0
                  Tnf
                                Rad
33
                                Rad
                                                           0.1
47
                   12
                           RadChem
                                               12
                                                           8.0
62
        14
                   17
                           RadChem
                                               17
                                                          14.0
90
        16
                   60
                           RadChem
                                               60
                                                          16.0
```

• fit the Weibull model and add the fitted survival curves as follows:



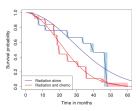


Fig. 12.6 Survival times (times to appearance of breast retractions) for breast cancer patients treated with radiation alone (blue) or radiation with adjuvant chemotherapy (red). The smooth lines are from a proportional bazards (also accelerated failure time) Weibull fit to the interval-censored data, whereas the step functions are non-parametric proportional hazards estimates. The shaded areas represent intervals on which the form of the survival curve estimate is ambiguous

```
bcos.survreq <-survreq(Surv(left.alt, right.alt, type="interval2")
treatment, dist="weibull", data=bcos)
> pct <- 1:999/1000
> ptime <- predict (bcos.survreg, type='quantile',
newdata=data.frame(treatment=c("Rad", "RadChem")),p=pct, se=F)
> ines(ptime[1,], 1-pct, xlab="Hours", ylab="Survival", type='1',
lty=c(1,2,2), lwd=c(2,1,1), xlim=c(0,20), col="blue")
> lines(ptime[2,], 1-pct, xlab="Hours", ylab="Survival", type='1',
lty=c(1,2,2), lwd=c(2,1,1), xlim=c(0,20), col="red")
```

select covariates

- In some applications, by contrast, our interest focuses on the predictive ability of a set of covariates. the biomarker focus on using those measurements to *predict* a patient's survival prospects.
- An important such method is the "lasso" procedure in the R package "penalized"
- This approach *maximizes* the partial likelihood function $\ell(\beta) = \log L(\beta)$

$$L(\beta) = \prod_{j=1}^{D} \frac{h_0(t_j)\psi_j}{\sum_{k \in B_j} h_0(t_j)\psi_k} = \prod_{j=1}^{D} \frac{\psi_j}{\sum_{k \in B_j} \psi_k}$$
(9)

lasso procedure

• The parameter satisfies the *constraint* $\sum_{j=1}^{p} |\beta_j| \le t$ for a constant t, where p is the number of parameters,this may be shown to be equivalent to maximizing the penalized likelihood which, for a pre-specified value of λ .

$$\ell_{pen}(\beta) = \ell(\beta) - \lambda \sum_{j=1}^{p} |\beta_j|$$
 (10)

- A sufficiently large value of λ will result in *no covariates* at all in the model, and smaller λ values will cause non-zero coefficient estimates.
- How do we select λ?



Cross Validation

- The goal of the procedure is accurate prediction, to add the predictive accuracy, we have a procedure called cross validation.
- Cross Validation:
 - Start with an initial value of λ and randomly divide the data set into *five* subsets of approximately equal size.
 - Select one of the subsets to be what we shall call the "validation" set(20%), and combine the remaining subsets into what we shall call the "training" set(80%).
 - Use the training set to construct the lasso model based on (10), and use this model to predict the survivals of patients in the validation set.

Cross Validation

- Use a partial-likelihood-based measure of goodness-of-fit to these data.
- Repeat this four more times, with each of the remaining four subsets in turn playing the role of the 20% validation set and the others being the training set.
- Derive an *average* partial-likelihood goodness of fit, and repeated for a range of values of λ , and we select that value that produces the *optimum* goodness of fit.

Example 2

The data contains 17 clinical and biomarker measurements on 227 patients, as well as overall survival and time to recurrence, both recorded in months. Of the 227 patients, 117 have levels of a variety of chemokines and other markers, some representing levels in the tumor itself and some outside the tumor, use 26 of these measurements (columns 23 to 48) as potential predictors of overall survival using a lasso model

Selecting the 117 patients with complete data:

```
hepatoCellularNoMissing<-hepatoCellular[
complete.cases(hepatoCellular),]
```

• "OS" (overall survival), "Death" (censoring) and a few of the cytokine measurements

```
hepatoCellularNoMissing[c(1,5,12), c(16,17,23:27)]
                           CD4N
                                             CD8N
                                                       CD20T
2
     OS Death
                 CD4T
                                     CD8T
     83
             2.600000 0.000000 190.6000
                                           126.80
                                                   20.950000
 76 20
           1 14.450000 2.758621
                                   2.1500 38.95
                                                   26.100000
 131 35
          1 2.821133
                        8.294828
                                   8.0064
                                            62.64
                                                    2.821133
```

• Then we fit a simple lasso model using the 26 predictors (columns 23 to 48), and we fix the penalty at $\lambda = 10$.

```
1 > attach(hepatoCellularNoMissing)
2 > library(penalized)
3 > hepato.pen <- penalized(Surv(OS, Death),
4 penalized=hepatoCellularNoMissing[,23:48],
5 standardize=T, lambdal=10) # nonzero coefficients: 7</pre>
```

List their values using the "coef" function

 Use cross-validation to select a value that optimizes the predictive ability of the lasso model, as defined by maximizing the cross-validated partial log-likelihood(CVL).

```
1 > set.seed(34)
2 > hepato.prof <- profLl(Surv(OS, Death),
3 penalized=hepatoCellularNoMissing[,23:48],
4 standardize=T, fold=10, minlambda1=2, maxlambda1=12)
5 > plot(hepato.prof$cvl ~ hepato.prof$lambda, type="l", log="x",
6 xlab="lambda", ylab="Cross-validated log partial likelihood")
```

• "set.seed" is to set the random number seed so that we can reproduce this model fit exactly.

 To find the optimal value, we use "OptL1" with the same starting seed.

```
1 > set.seed(34)
2 > hepato.opt <- optLl(Surv(OS, Death),
3 penalized=hepatoCellularNoMissing[,23:48], standardize=T,
4 fold=10)
5 > hepato.opt$lambda
6 [1] 8.242321
7 > abline(v=hepato.opt$lambda, col="gray")
```

• The optimal value is 8.24.

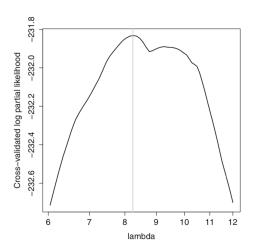


Fig. 12.7 Cross-validated log partial likelihood for a range of values of lambda for the hepatocellular data. The *vertical gray line* shows the global maximum at $\lambda = 8.24$, which was obtained using the "OptL1" function in the "penalized" package

26/26