Applied Survival Analysis Using R Chapter 7: Model Diagnostics

Qi Guo

Department of Mathematical Sciences The University of Texas at Dallas

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1 Assessing Goodness of Fit Using Residuals

2 Case Deletion Residuals

3 Checking the Proportion Hazards Assumption

4 Schoenfeld Residuals



The use of residuals

- The residuals are plotted versus some quantity(STAT 6337:
 Advanced Statistical Method I, the first two chapters), such as a covariate value, and the observed pattern is used to diagnose possible problems with the fitted model.
- The <u>residuals</u> have the additional property of not only indicating problems but also suggesting <u>remedies</u>, that means suggest an alternative model that fits the data better.
- Many of these residuals have been generalized to survival analysis. And survival data evolves over time, and requires special assumptions such as proportional hazard.

Martingale and Deviance Residuals

- An important tool for assessing the goodness of fit of a model is to compare the censoring indicator (0 for censored, 1 for death) for each subject to the expected value of that indicator under the proportional hazards Cox model.
- If there are no time dependent covariates (Chapter 8) and if the survival times are right-censored. Martingale Residuals

$$m_{i} = \delta_{i} - \hat{H}_{0}(T_{i}) \exp(z_{i}'\hat{\beta})$$
 (1)

- The residual is essentially the <u>difference</u> between the observed value (1 or 0) of the censoring indicator and its expected value under a particular Cox model.
- But the sum of squares of Martingale residuals cannot be used as a measure of goodness of fit, so "deviance" residual is an alternative way does have this property.

$$d_i = sign(m_i) \cdot \{-2 \cdot [m_i + \delta_i \log(\delta_i - m_i)]\}^{1/2}$$
 (2)

Difference

- 1 Deviance Residuals:
 - These residuals are symmetrically distributed with expected value 0 (if the fitted model is correct)
 - The sum of squares of these residuals is the value of the likelihood ratio test.
- 2 Martingale Residuals:
 - These residuals have the property of showing us the *functional form of a covariate*.
- Conclusion: For this reason, in practice, the *martingale residuals* are more useful.

Example 7.1

Consider the "pharmacoSmoking" dataset, and a fit of the "null" Cox proportional hazards model. A null model is one with no fitted covariates. plot Martingale Residuals against continuous predictors to get a preliminary assessment of which of these predictors should be in the model, and what form they should take

 Read in the data and truncate the variable "priorAttempts" at 20.

```
1 > pharmacoSmoking <- read.csv("PharmacoSmoking.csv")
2 > priorAttemptsT <- priorAttempts
3 > priorAttemptsT[priorAttempts > 20] <- 20</pre>
```

• Fit the null model and obtain these residuals as follows:

```
1 > library(survival)
2 > result.0.coxph <- coxph(Surv(ttr, relapse) ~ 1)
3 > rr.0 <- residuals(result.0.coxph, type="martingale")</pre>
```

- Plot martingale residuals versus age, prior attempts at quitting, and longest prior period without smoking.
- Plot these null model residuals versus each of these variables and also versus log transformations of these variables.

In this figure, the three plots on the *left* are for *untransformed*variables, and the three on the *right* are for *log transformations* of
these variables.

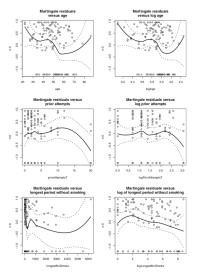


Fig. 7.1 Martingale residuals from a null model fit to the pharmacoSmoking data, plotted versus three continuous predictors and one log-transformed predictor



Case Deletion Residuals

- Some subjects may have an especially large influence on the parameter estimates and may cause problems, so we have to identify them.
- Case deletion residuals (also called "jackknife residuals") serve this purpose.

Case Deletion Residuals

For each subject, a case deletion residual is the *difference* in the value of the coefficient using all of the data and its value when that *subject is deleted* from the data set.

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Delete

- For each subject in turn ("n.obs" subjects in all), we delete the ith
 subject from the survival time "tt", censoring indicator
 "relapse", and covariates "grp", employment", and "age"
- Fit a Cox model to this reduced data set. The results for the ith subject go into "result.coxph.i"
- Extract the age coefficient(4th) into "coef.i" and compute the
 jackknife residual as the difference of "coef.i" and "coef.all":

```
1 > n.obs <- length(ttr)
2 > jkbeta.vec <- rep(NA, n.obs)
3 > for (i in 1:n.obs) {
4 tt.i <- ttr[-i]
5 delta.i <- relapse[-i]
6 grp.i <- grp[-i]
7 employment.i <- employment[-i]
8 age.i <- age[-i]
9 result.coxph.i <- coxph(Surv(tt.i, delta.i) ~ grp.i +
10 employment.i + age.i)
11 coef.i <- result.coxph.i$coef[4]
12 jkbeta.vec[i] <- (coef.all - coef.i)}</pre>
```

Plot

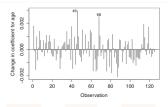
Plot these residuals versus the patient id's, which we place in the vector "index.obs", "type=' h' " causes the residuals to be plotted as spikes. "abline (h=0)" plots a horizontal line through 0.

```
1 > index.obs <- 1:n.obs
2 > plot(jkbeta.vec ~ index.obs, type="h",xlab="Observation",
3 ylab="Change in coefficient for age", cex.axis=1.3, cex.lab=1.3)
4 > abline(h=0)
```

 The "identify" function allows us to identify the index numbers of select patients by manually selecting them with a mouse

```
1 > identify(jkbeta.vec ~ index.obs)
```

Conclusion



- Conclusion: No single patient changes the estimate of the "age" coefficient by more than 0.003, which is less than 10% of the value of that coefficient. and patients 46 and 68 have the most influence over the parameter estimate for age.
- A more convenient way to obtain case deletion residuals is using the "residuals" function with "type = 'dfbeta'".



debeta residuals

```
1 > resid.dfbeta <- residuals(result.coxph, type="dfbeta")
2 > n.obs <- length(ttr)
3 > index.obs <- l:n.obs
4 > plot(resid.dfbeta[,4] ~ index.obs, type="h",xlab="Observation",
5 ylab="Change in coefficient")
6 > abline(h=0)
7 > identify(resid.dfbeta[,4] ~ index.obs)
```

The resulting dfbeta residuals plot (not shown) is nearly
identical, it has the advantage that it is slightly easier to use it to
produce multiple plots for all of the coefficients.

Log Cumulative Hazard Plots

- log-log transformation:
 - Under the assumption, we have

$$S_1(t) = [S_0(t)]^{exp(\beta)}$$
(3)

where $exp(\beta)$ is the proportional hazards constant.

Take logs both sides

$$\log[S_1(t)] = \exp(\beta) \cdot \log[S_0(t)] \tag{4}$$

The survival functions are less than 1, so log will cause negative.

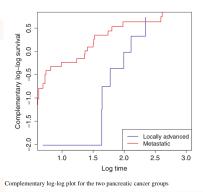
$$\log\{-\log[S_1(t)]\} = \beta + \log\{-\log[S_0(t)]\}$$
 (5)

where the function $g(\mu) = \log\{-\log(\mu)\}$ is called a *complementary log-log transformation*.

• Plot $g[S_1(t)]$ and $g[S_0(t)]$ versus t or $\log(t)$ will yield two parallel curves separated by β if the proportional hazards assumption is correct.

Illustrate this with the pancreatic cancer data from Chapter 3.

```
# Plot the LA(Locally advanced)
  > result.surv.LA <- survfit (Surv (pfs.month) ~ stage,
  subset=stage == "LA")
  > time.LA <- result.surv.LA$time
  > surv.LA <- result.surv.LA$surv
  > cloglog.LA <- log(-log(surv.LA))</pre>
  > logtime.LA <- log(time.LA)
  # Plot the M(Metastatic)
10 > result.surv.M <- survfit(Surv(pfs.month) ~ stage,
  subset=stage == "M")
 > time.M <- result.surv.M$time
12
13 > surv.M <- result.surv.M$surv
14 > cloglog.M <- log(-log(surv.M))
  > loatime.M <- loa(time.M)
15
16 > plot(cloglog.LA ~ logtime.LA, type="s", col="blue", lwd=2)
17 > lines(cloglog.M ~ logtime.M, col="red", lwd=2, type="s")
18 > legend("bottomright", legend=c("Locally advanced",
19 Metastatic"), col=c("blue", "red"), lwd=2)
```



Conclusion: The curves are clearly not parallel. However, one
problem with this approach is that we don't have a clear way to
assess statistical significance. This is a critical issue here due to
the small sample size, particularly in the locally advanced group.

Recall

- The Schoenfeld residuals are the individual terms of the score function, and each term is the observed value of the covariate for patient i minus the expected value $E(Z_i) = \bar{z}(t_i)$, which is a weighted sum, with weights given by $p_k(\beta)$, of the covariate values for subjects at risk at that time.
- Recall:
 - The partial log-likelihood function

$$\ell(\beta) = \sum_{i \in D} \left\{ \log(\psi_i) - \log\left(\sum_{k \in R_i} \psi_k\right) \right\} = \sum_{i \in D} \left\{ z_i \beta - \log\left(\sum_{k \in R_i} e^{z_k \beta}\right) \right\}$$
(6)

The score function:

$$\ell'(\beta) = \sum_{i \in D} \left\{ z_i - \sum_{k \in R_i} z_k \cdot p(\beta, z_k) \right\} \text{where } p(\beta, z_k) = \frac{e^{z_k \beta}}{\sum_{i \in R_k} e^{z_j \beta}} \quad (7)$$

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• For an estimate $\hat{\beta}$ the residual for the *i*th failure time is:

$$\hat{r}_i = z_i - \sum_{k \in R_i} z_k \cdot p(\hat{\beta}, z_k) = z_i - \bar{z}(t_i)$$
 (8)

Example:

```
1 > tt <- c(6, 7, 10, 15, 19, 25)
2 > delta <- c(1, 0, 1, 1, 0, 1)
3 > trt <- c(0, 0, 1, 0, 1, 1)
4 > result.coxph <- coxph(Surv(tt, delta) ~ trt) > result.coxph$coef
5 [1] trt -1.326
```

• We see that $\hat{\beta} = 1.326$. To compute the Schoenfeld residuals, we first compute the weights as follows:

t_i	n_{0i}	n_{1i}	$p(\beta, z_k = 0)$	$p(\beta, z_k = 1)$
6	3	3	$\frac{1}{3+3e^{-1.326}}$	$\frac{e^{-1.326}}{3+3e^{-1.326}}$
10	1	3	$\frac{1}{1+3e^{-1.326}}$	$\frac{e^{-1.326}}{1+3e^{-1.326}}$
15	1	2	$\frac{1}{1+2e^{-1.326}}$	$\frac{e^{-1.326}}{1+2e^{-1.326}}$
25	0	1	$\frac{1}{e^{-1.326}}$	$\frac{e^{-1.326}}{e^{-1.326}} = 1$



Compute the expected values, and then the residuals:

t _i	$E(Z_i) = \sum_{k \in R_i} z_k \cdot p_k(\hat{\beta})$	zį	$\widehat{r_i} = z_i - E(Z_i)$
6	$3 \times 0 \times \frac{1}{3+3e^{-1.326}} + 3 \times 1 \times \frac{e^{-1.326}}{3+3e^{-1.326}} = 0.2098$	0	-0.2098
10	$1 \times 0 \times \frac{1}{1+3e^{-1.326}} + 3 \times 1 \times \frac{e^{-1.326}}{1+3e^{-1.326}} = 0.4434$	1	0.5566
15	$1 \times 0 \times \frac{1}{1 + 2e^{-1.326}} + 2 \times 1 \times \frac{e^{-1.326}}{1 + 2e^{-1.326}} = 0.3468$	0	-0.3468
25	1	1	0

• In R:

The scaled residual

 Gramsch and Therneau proposed scaling each residual by an estimate of its variance. This scaled residual conveniently approximated as follows:

$$r_i^* = r_i \cdot d \cdot var(\hat{\beta}) \tag{9}$$

where d is the total number of deaths, and $var(\hat{\beta})$ is the variance of the parameter estimate.

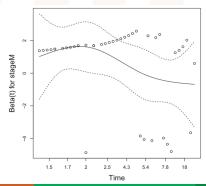
```
> resid.unscaled <- residuals(result.coxph, type= \'schoenfeld'')
> resid.scaled <- resid.unscaled*result.coxph$var*sum(delta)
> resid.unscaled
[1] 6 10 15 25
-0.2098004 0.5566351 -0.3468347 0.0000000
> resid.scaled
[1] -1.313064 3.483776 -2.170712 0.000000
```

The scaled residuals

• An approximate estimate of $\beta(t)$ may be obtained by adding the estimate $\hat{\beta}$ from the Cox proportional hazards model to the standardized residuals.

 Using this residuals to the pancreatic data, compute the scaled Schoenfeld residuals and plot

```
1 > result.coxph <- coxph(Surv(pfs.month) ~ stage)
2 #The '`transform'' option specifies that the time axis is scaled to
3 > result.sch.resid <- cox.zph(result.coxph, transform="km")]
4 > plot(result.sch.resid)
```





- The shape of the smoothed(loess) curve is an estimate of the difference parameter as a function of time.
- The hypothesis test for a constant β

- Alternatively, variations of this test may be obtained by plotting $\beta(t)$ versus time or versus other transformations of time, and the "cox.zph" function offers:
 - "rank" option: the time axis is ordered by the ranks of the times.
 - "identity" option: the time variable is untransformed.

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