Package 'Amplicomsat'

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Type Package

Title Processes microsatellite genotypes from Amplicon generated fastq
read files

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Description A series of functions that can all be wrapped sequentially in a single script to extract
microsatellite genotypes from fastq files with Ampicon sequencing reads. It can extract
mutiple alleles per sample loci combinations and identify sequence variant alleles.

Imports stringr, readr, Biostrings

License GPL (>=2)

NeedsCompilation no

R topics documented:

AlleleCum   *Plots cummulative distributions of scored alleles*

Description

Plots cummulative distributions of scored alleles to help diagnose any possible errors

Usage

```
AlleleCum(GenotypeA.DF="./GenotypesOut/Genotypes allele table.txt",locus,locusInfo="primers",
         ymin=NULL,ymax=NULL)
```

Arguments

GenotypeA.DF   A data.frame as produced by alleleFinder

locus   A string with the name of the locus to plot. The name needs to be written as in
               the file provided to locusInfo argument to other functions, e.g., read.size.freqs

locusInfo   A tab delimited text file containing the information for the microsatellite loci
               used. Should contain four columns with the following header names:

               Locus: The loci names

               Fprimer: The sequence of the FWD primer in upper case

               Rprimer: The sequence of the RVS primer in upper case

               Motif: The sequence for the microsatellite motif (e.g., CTTG for tetranucleotide).
               Note that for some loci you might need to use the reverse complementar se-
               quence of what your reference value is. Thus, inspect your reads first or if you
               are not getting data for some loci try this out. For example, for a motif CAGG,
               you might need to use CCTG.

               nmotif: Reads are retained if they have at least these many repetitions of the
               repeat motif.

ymin   The y lower lim for the y axis. Usefull for loci that are very polymorphic and
               need to be zoomed in.

ymax   The y upper lim for the y axis. Usefull for loci that are very polymorphic and
               need to be zoomed in.

Details

Plots the cumulative distribution of alleles scored for the selected locus stored in the data.frame
supplied to argument GenotypeA.DF. The sample names for specific observations can be obtained
by interacting with the plot using function getpoints.

Value

Plots the cumulative allele distribution for a selected locus

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

getpoints

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alleleFinder	*Scores the diploid genotypes into an output table*

Description

Estimates the diploid genotypes based on read size (primer to primer) in bp. The results are written to two files: one with two alleles per column and the other with the two alleles collapsed in a single column. The files are written to a folder named GenotypesOut created by the function.

Usage

```
alleleFinder(sampleDir="./samplesF",locusInfo="primers",calibrationFiles="CalibrationSamples.txt")
```

Arguments

sampleDir   The directory where the filtered reads are stored. This directory should only contain the read files and nothing else. By default this is a directory named samplesF written in the session working directory, as produced by read.quality.filter

locusInfo   A tab delimited text file containing the information for the microsatellite loci used. Should contain four columns with the following header names:

Locus: The loci names

Fprimer: The sequence of the FWD primer in upper case

Rprimer: The sequence of the RVS primer in upper case

Motif: The sequence for the microsatellite motif (e.g., CTTG for tetranucleotide). Note that for some loci you might need to use the reverse complementar sequence of what your reference value is. Thus, inspect your reads first or if you are not getting data for some loci try this out. For example, for a motif CAGG, you might need to use CCTG.

nmotif: Reads are retained if they have at least these many repetitions of the
repeat motif.

calibrationFiles

The file described below is the same as used for function `GenotypeModel`, ear-
lier in the previous pipeline step. Here the only information used is the last row
containing the sequencing coverage threshold for each marker (number of reads
of similar sequence length) that will applied to filter alleles scored. Inspect the
plots produced by function `reads.plots` carefully to select values for cover-
age filtering. Essentialy use a value below which you don't trust reads to be
true alleles. A tab delimited file with as many columns as loci amplified in the
samples. The order of loci is required to match the order in the locusInfo file.
The header names must start with the locus name, written exactly has in the file
passed to locusInfo. The first row contains the range of alleles for each locus,
the format should be 100-250, for an example of an allelic range between 100
and 250 bp. Users should inspect the plots created by `reads.plots` to deter-
mine the ranges. Be conservative and add a buffer of about 10 bp on each side
but know that the larger the range the slower the function will be. The following
rows contain samplenames (one per row) for a homozygote samples that will
be used to generate the expected relative frequencies under different combina-
tions of alleles. Users can select these homozygote representative samples by
inspecting the plots saved in the Plots directory (generated by `reads.plots`).
Use the exact sample names as the file names stored in directory samplesF. For
convenience, sample names are written to the plots and can be copied from there
when selecting homozygote samples.

Details

Uses the tables created by `GenotypeModel`, containing expected relative frequencies of read cover
for different combinations (2 out of N) of alleles, to contrast with observed data in each sample and
find the diploid genotypes minimizing a sum of squares. For each observed sample, the row table
that minimizes the sum of squares: $sum((ORF(a)-ERF(a))^2)$, with ORF and ERF the Observed
and Expected relative frequencies at each read size, is the estimated genotype.

Value

A data frame with a concatenated genotype (the two alleles in the same cell) is returned. This table
is also written to file, as well as a version with the two alleles in one locus in different columns (two
columns per locus). The output files are written to a folder named GenotypesOut that is created by
the function in the R session working directory.

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

GenotypeModel, AlleleCum

Edit.Primer.File   *Checks and corrects for mismatches in primer sequences.*

Description

Sequencing reads might show primer sequences that diverge from published sequences for various reasons. This function will search the reads for primer sequences and correct those that diverge from the supplied parameter file.

Usage

```
Edit.Primer.File(sampleDir="./samplesF",locusInfo="primers",calibN=2,mismatch=1)
```

Arguments

sampleDir	The directory where the filtered reads are stored. This directory should only contain the read files and nothing else. By default this is a directory named samplesF written in the session working directory, as produced by read.quality.filter
locusInfo	A tab delimited text file containing the information for the microsatellite loci used. Should contain four columns with the following header names:
	Locus: The loci names
	Fprimer: The sequence of the FWD primer in upper case
	Rprimer: The sequence of the RVS primer in upper case
	Motif: The sequence for the microsatellite motif (e.g., CTTG for tetranucleotide). Note that for some loci you might need to use the reverse complementar sequence of what your reference value is. Thus, inspect your reads first or if you are not getting data for some loci try this out. For example, for a motif CAGG, you might need to use CCTG.
	nmotif: Reads are retained if they have at least these many repetitions of the repeat motif.
calibN	A numeric value (defaults to 2) or a character vector with sample names. The function will search primers in these many random samples or specific samples supplied in a vector of sample names. The default is two random samples. If using specific samples, we recommend using two or three samples to avoid slowing down the code too much.
mismatch	A numeric value for the maximum mismatch allowed between supplied primer sequences and those searched in sequenced reads. It defaults to 1.

Details

All forward and reverse primers supplied in locusInfo will be searched in the sequenced reads, allowing for mismatches of up to one base pair (default). If differences are found, an edited file with primerInfo is written to the R session working directory. A log file reporting the changes made to the file is also written. The function uses matchPattern from package Biostrings to perform the search with mismatches. Using this function at the start of the pipeline allows for faster read filtering in subsequent steps by searching exact sequences without mismatches.

Value

The function returns a table with the edited primer information in a similar format to argument locusInfo (see above). The table is also written in the R session working directory, where it can be conveniently parsed to the other functions in the Amplicomsat pipeline. A log reporting the changes made to the primer sequences is also written into a file.

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

read.size.freqs

GenotypeModel   *Creates tables with expected relative frequencies of read sizes*

Description

Creates tables with expected relative frequencies of read sizes for each possible diploid genotype within an allelic range set by the user.

Usage

```
GenotypeModel(sampleDir="./samplesF",locusInfo="primers",drop.out.b=0.015,
              calibrationFiles="CalibrationSamples.txt")
```

Arguments

sampleDir   The directory where the filtered reads are stored. This directory should only con-
                   tain the read files and nothing else. By default this is a directory named samplesF
                   written in the session working directory, as produced by read.quality.filter

locusInfo   A tab delimited text file containing the information for the microsatellite loci used. Should contain four columns with the following header names:

Locus: The loci names

Fprimer: The sequence of the FWD primer in upper case

Rprimer: The sequence of the RVS primer in upper case

Motif: The sequence for the microsatellite motif (e.g., CTTG for tetranucleotide). Note that for some loci you might need to use the reverse complementar sequence of what your reference value is. Thus, inspect your reads first or if you are not getting data for some loci try this out. For example, for a motif CAGG, you might need to use CCTG.

nmotif: Reads are retained if they have at least these many repetitions of the repeat motif.

drop.out.b   The proportion of decline in coverage per base pair distance between small and large allele on a heterozygous loci. It defauts to 0.015. It could be estimated from the data from each locus, in a future version, if the user supplies a list of heterozygous samples (upon inspecting the plots produced).

calibrationFiles

A tab delimited file with as many columns as loci amplified in the samples. The order of loci is required to match the order in the locusInfo file. The header names must start with the locus name, written exactly has in the file passed to locusInfo. The first row contains the range of alleles for each locus, the format should be 100-250, for an example of an allelic range between 100 and 250 bp. Users should inspect the plots created by `reads.plots` to determine the ranges. Be conservative and add a buffer of about 10 bp on each side but know that the larger the range the slower the function will be. The following rows contain samplenames (one per row) for a homozygote samples that will be used to generate the expected relative frequencies under different combinations of alleles. Users can select these homozygote representative samples by inspecting the plots saved in the Plots directory (generated by `reads.plots`). Use the exact sample names as the file names stored in directory samplesF. For convenience, sample names are written to the plots and can be copied from there when selecting homozygote samples. The final row contains the coverage threshold (number of reads of similar sequence length) that will applied to filter alleles scored. This function will not use this information but to keep the format of the calibration file similar across this and other functions the information is required here.

Details

Using a set of representative homozygote samples and an allelic range provided by the user, the function produces a large table with as many rows as combinations of diploid genotypes, i.e., all combinations of 2 in N, N being defined by seq(lower.range, upper.range, 1). To these combination of 2 in N the homozygote cases, for all alleles in the defined range, are added as additional possible

genotypes. Then a pattern of relative frequencies from a single allele, generated from the analysis of reads in the homozygote reference samples, is shifted along the allelic range to match the bp position of each different combination of alleles. This procedure generates all the possible overlaps of read relative frequencies for the heterozygote cases. For heterozygotes the relative frequencies are then divided by two so that the row sum adds to 1. The table rows (one for each possible genotype) are then compared against the observed data for each sample by function alleleFinder to find the row table that minimizes a sum of squares relative to the observed relative frequencies for each read in each sample: sum((ORF(a)-ERF(a))^2), with ORF and ERF the Observed and Expected relative frequencies at each read size a. The row with the minimum sum is the estimated genotype.

Value

One model table for each locus is written in a directory, named ModelTables, created by the function in the working directory.

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

alleleFinder

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getpoints   *Interacts with an open plot to extract sample names*

Description

A tool to extract sample name information

Usage

```
getpoints(GenotypeA.DF,locus)
```

Arguments

GenotypeA.DF   A data.frame as produced by alleleFinder

locus   A string with the name of the locus to plot. The name needs to be written as in
                the file provided to locusInfo argument to other functions, e.g., read.size.freqs

Details

Interacts with the plots produced by [AlleleCum](#) to extract sample name information for specific allele sizes. Upon running the command the R locator function is called for plot interaction. The user needs to perform two clicks on the plot, first by moving the cursor below the targeted allele points, the second click above them. The list of samples in GenotypeA.DF that contain alleles sizes within the y range (size in bp) of the two clicks is printed to the screen.

Value

The sample names containing the selected allele sizes are printed to screen.

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

[AlleleCum](#)

M.allele.Finder	*Identifies multiple alleles (possibly more than two) for each loci in each sample.*

Description

The function searches samples for multiple alleles of the same loci. Examples of such samples come from forensic analysis, where DNA from multiple individuals might be present, or eDNA samples containing alleles from several individuals. Only alleles present in the reference population can be detected.

Usage

```
M.allele.Finder(sampleDir="./samplesF.MA", locusInfo="primers",
             calibrationFiles="CalibrationSamples.txt",
             ReadSizesRepeatNumbers="./ReadSizesRepeatNumbers/",
             RefGenotypes="./GenotypesOut/Genotypes allele table.txt")
```

Arguments

sampleDir   The directory where the filtered reads are stored. This directory should only con-
                   tain the read files and nothing else. By default this is a directory named samplesF
                   written in the session working directory, as produced by `read.quality.filter`

locusInfo   A tab delimited text file containing the information for the microsatellite loci
                   used. Should contain four columns with the following header names:

                   Locus: The loci names

                   Fprimer: The sequence of the FWD primer in upper case

                   Rprimer: The sequence of the RVS primer in upper case

                   Motif: The sequence for the microsatellite motif (e.g., CTTG for tetranucleotide).
                   Note that for some loci you might need to use the reverse complementar se-
                   quence of what your reference value is. Thus, inspect your reads first or if you
                   are not getting data for some loci try this out. For example, for a motif CAGG,
                   you might need to use CCTG.

                   nmotif: Reads are retained if they have at least these many repetitions of the
                   repeat motif.

calibrationFiles
                   A tab delimited file with as many columns as loci amplified in the samples.
                   The header names must start with the locus name, written exactly has in the file
                   passed to locusInfo. The first row contains the range of alleles for each locus,
                   the format should be 100-250, for an example of an allelic range between 100
                   and 250 bp. Users should inspect the plots created by `reads.plots` to deter-
                   mine the ranges. Be conservative and add a buffer of about 10 bp on each side
                   but know that the larger the range the slower the function will be. The following
                   rows contain sample names (one per row) for a homozygote samples that will be
                   used to generate the expected relative frequencies under different combinations
                   of alleles. Users can select these homozygote representative samples by inspect-
                   ing the plots saved in the Plots directory (generated by `reads.plots`). Use the
                   exact sample names as the file names stored in directory samplesF. For conve-
                   nience, sample names are written to the plots and can be copied from there when
                   selecting homozygote samples. I used six samples per locus in my test (thus my
                   calibration table had a total of seven rows).

ReadSizesRepeatNumbers
                   The path to the folder with the same name created by function `read.size.freqs`.

RefGenotypes   A string with the path for the reference file genotype file in the 2 columns per
                   locus format. The format is the same as produced by function `alleleFinder`
                   and thus the default path is the folder where that file is written by this function
                   "./GenotypesOut/Genotypes allele table.txt". Only alleles present in the refer-
                   ence population can be found.

Details

The function searches for the presence of alleles, from those present in the reference population, in samples that may have up to n alleles. Examples of such samples come from forensic analysis where DNA from multiple multiple individuals might be present or eDNA samples containing alleles from several individuals.

Value

The function returns to the working environment a list with the alleles found for each locus and individual. The function also prints to the working directory a pdf file for each sample containing plots of read (allele) distribution, with separate plots per page showing the different loci, and red vertical lines indicating the alleles scored.

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

[GenotypeModel](), [alleleFinder]()

| M.allele.seq.variant | *Identifies multiple sequence based alleles in eDNA or forensic samples.* |

Description

The function searches samples for multiple sequenced based alleles, i.e., alleles of the same size with sequence variation. Examples of such samples come from forensic analysis, where DNA from multiple individuals might be present, or eDNA samples containing alleles from several individuals. Sequenced based AllelesSample are searched from a list containing the multiple size-based alleles found by `M.allele.Finder`.

Usage

```
M.allele.seq.variant(sampleDir="./samplesF.MA",AlleleList,locusInfo,alleleDB,
                HeteroThreshold=0.4,calibrationFiles="CalibrationSamples.txt")
```

Arguments

sampleDir	The directory where the filtered reads are stored. This directory should only contain the read files and nothing else. By default this is a directory named samplesF written in the session working directory, as produced by read.quality.filter
AlleleList	A list with the alleles found for each loci and sample as created by M.allele.Finder.
locusInfo	A tab delimited text file containing the information for the microsatellite loci used. Should contain four columns with the following header names:

Locus: The loci names

Fprimer: The sequence of the FWD primer in upper case

Rprimer: The sequence of the RVS primer in upper case

Motif: The sequence for the microsatellite motif (e.g., CTTG for tetranucleotide). Note that for some loci you might need to use the reverse complementar sequence of what your reference value is. Thus, inspect your reads first or if you are not getting data for some loci try this out. For example, for a motif CAGG, you might need to use CCTG.

nmotif: Reads are retained if they have at least these many repetitions of the repeat motif.

alleleDB	A data.frame containing the database of allele size, sequence variation, new allele code, and nucleotide sequence, as created by score.seq.variants. If necessary, the function will update the database with new allele sizes or sequence variants or both, in which case the updated database is written to the working directory. The format is a tab-delimited table with different sequence variants (rows) and the following required columns:

Locus: The locus name for the sequence-based allele. Needs to match the loci names supplied in argument locusInfo.

SIZE: The size in bp for the sequence-based allele.

VARIANT: A low case letter code to distinguish the different sequence-based alleles that share the same sequence size.

SEQ: The nucleotide sequence for each sequence-based allele

AlleleCode: A numeric code used to write alleles in format required by most population genetics software.

HeteroThreshold	

When sequence variation exists within reads of the same size for the same loci, two or more sequences will dominate the read frequencies, with many more in low frequency due to sequencing errors. This argument refers to the proportion of reads in the second and following most frequent variants compared to the first.

The value sets the minimum allowed proportion required to score these reads as sequence-based variant alleles, i.e., not sequencing errors. It defaults to 0.4.

calibrationFiles

The file described below is the same as used for function `GenotypeModel` and `alleleFinder`, in earlier pipeline steps. Here the only information used is the last row containing the sequencing coverage threshold for each marker (number of reads of similar sequence length) that will be applied to filter alleles scored. Inspect the plots produced by function `reads.plots` carefully to select values for coverage filtering. Essentialy, use a value below which the plots become too ambiguous to confidently call alleles. A tab delimited file with as many columns as loci amplified in the samples. The order of loci is required to match the order in the locusInfo file. The header names must start with the locus name, written exactly has in the file passed to locusInfo. The first row contains the range of alleles for each locus, the format should be 100-250, for an example of an allelic range between 100 and 250 bp. Users should inspect the plots created by `reads.plots` to determine these ranges. Be conservative and add a buffer of about 10 bp on each side but know that the larger the range the slower the function will be. The following rows contain sample names (one per row) for examples of homozygote samples at that column locus. These will be used to generate the expected relative frequencies under different combinations of alleles. Users can select these homozygote representative samples by inspecting the plots saved in the Plots directory (generated by `reads.plots`). Use the exact sample names as the file names stored in directory samplesF. For convenience, sample names are written to the plots and can be copied from there when selecting homozygote samples.

Details

The function searches for the presence of sequenced-based alleles in list containing multiple size-based alleles supplied by the user (as produced by `M.allele.Finder`). The function requires a data base of allele variants, as produced by `score.seq.variants`. If new variants are found, the allele database will be updated and written to working environment (Allele DataBase UPDATED.txt). A log file (DataBase new entries.txt) is also written to the working environment with the new alleles added to the database. The fifth column in this file informs if the new allele represents a new size or a new sequence variant.

Value

A list with the sequence-based alleles found for each locus and individual.

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

[score.seq.variants](#), [alleleFinder](#)

---

read.quality.filter   *Filters the fastq reads for sequencing quality*

Description

Filters the reads so that if a single nucleotide is below a phred score of 30 the read is removed. Future versions will allow user defined phred scores.

Usage

```
read.quality.filter(sampleDir="./samples",filterDir="samplesF")
```

Arguments

sampleDir	A string for the path of the directory containing the samples. This should contain only the fastq files and nothing else. The default is a folder named samples
filterDir	A string with the directory path where the filtered fastq files will be stored. The default is a folder named samplesF. This folder is created in the session working directory when running the function. When running multiple times, users might want to rename this folder.

Details

A read is filtered out if a single nucleotide is below a phred score 30. The input fastq files should be stored in a directory named samples, and the output filtered files will be written in a folder named samplesF created when the function is run. Users can change these names.

Value

Fstq read files filtered by sequence quality. A Filterlog file is written to the working directory with the number and percentage of reads that were removed for each sample.

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

[read.size.freqs](#)

---

read.size.freqs	*Finds reads that contain the microsatellite primers and four or more repeats of the microsatellite motif*

Description

Searches sequenced reads for those containing the microsatellite FWD and RVS primers and a user-defined number of microsatellite motif repeats. Writes files for each sample and loci combination containing the read size and the number of msat motif repeats for each read detected.

Usage

```
read.size.freqs(locusInfo="primers",sampleDir="./samplesF")
```

Arguments

locusInfo   A tab delimited text file containing the information for the microsatellite loci used. Should contain four columns with the following header names:

Locus: The loci names

Fprimer: The sequence of the FWD primer in upper case

Rprimer: The sequence of the RVS primer in upper case

Motif: The sequence for the microsatellite motif (e.g., CTTG for tetranucleotide). Note that for some loci you might need to use the reverse complementar sequence of what your reference value is. Thus, inspect your reads first or if you are not getting data for some loci try this out. For example, for a motif CAGG, you might need to use CCTG.

nmotif: Reads are retained if they have at least these many repetitions of the repeat motif.

sampleDir   The directory where the filtered reads are stored. This directory should only contain the read files and nothing else. By default this is a directory named samplesF written in the session working directory, as produced by read.quality.filter

Details

Searches sequenced reads for those containing the microsatellite FWD and RVS primers and a user-defined number of microsatellite motif repeats. Writes files for each sample and loci combination containing the read size and the number of msat motif repeats for each read detected. Running this function is the slowest step in the pipeline, but the files only need to be created once per set of new samples.

Value

Output files are written to folder ReadSizesRepeatNumbers, created by the function. Output files for each sample and locus combinations contain the following two columns for each read detected:

ReadSize: The length in bp of each read

RepeatNumber: The number of microsatellite motifs for each read

These files are written to a folder named ReadSizesRepeatNumbers that is created by the function in the working environment and will be used down the pipeline by the functions reads.plots, GenotypeModel, and alleleFinder.

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

reads.plots, alleleFinder

---

reads.plots   *Creates frequency plots for the reads of specfic locus for each sample*

Description

Creates plots to visualize the read frequencies and help discerning the possible microsatellite alleles.

Usage

```
reads.plots(sampleDir="./samplesF",locusInfo="primers",x.range=c(90,350),
            x.label.cex=0.8,RepNumberPlot=FALSE )
```

Arguments

sampleDir   A string with the path for the directory where the filtered reads are stored. This directory should only contain the read files and nothing else. This should be the samplesF directory written in the session working directory, as produced by read.quality.filter.

locusInfo   A tab delimited text file containing the information for the microsatellite loci used. Should contain four columns with the following header names:

               Locus: The loci names

        Fprimer: The sequence of the FWD primer in upper case

        Rprimer: The sequence of the RVS primer in upper case

        Motif: The sequence for the microsatellite motif (e.g., CTTG for tetranucleotide). Note that for some loci you might need to use the reverse complementar sequence of what your reference value is. Thus, inspect your reads first or if you are not getting data for some loci try this out. For example, for a motif CAGG, you might need to use CCTG.

        nmotif: Reads are retained if they have at least these many repetitions of the repeat motif.

x.range	The x-axis limits plot limits. If some sample has alleles outside this range the function will adjust the x-axis limit for that sample. The defautl range is 90-350 bp.
x.label.cex	The plot size for allele labels (sizes in bp) along the x-axis. The default size is 0.8.
RepNumberPlot	A logical to decide if the second plot per sample should be plotted. It defaults to FALSE.

Details

The function provides a visualization tool that separates samples by loci, i.e. one plot per loci, with all samples and two pages per sample. The first sample plot is the classical barplot with x-axis for read sizes and y-axis for their frequency. The second plot shows read size vs number of motif repeats for reads. The plots are based on information from reads that contain the locus primers and a certain number of repeats for the msat motif in each locus. The function reads the files created by function read.size.freqs written to the folder ReadSizesRepeatNumbers.

Value

Plots for each loci, each containing two pages per sample. The first plot is the classical barplot with x-axis for read sizes and y-axis for their frequency. The second plot shows read size vs number of motif repeats for reads containing both primers and a minimum number of motif repeats (as set by function read.size.freqs).

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

read.size.freqs

---

sampleDist	*Plots barplots with filtered read frequency (coverage) for a selected sample and given locus.*

Description

Plots coverage barplots of a selected sample for a given locus

Usage

```
sampleDist(sample,xlim=c(75,400))
```

Arguments

sample	The name of the sample to plot including the name of the loci. The format is exactly as shown in the main tittle of the plots saved in the Plots directory, thus copy them from here.
xlim	A vector with two values for lower and upper limits of the x-axis

Details

A plotting function to zoom in to specific sample locus filtered read plots. Produces a plot similar to those printed to pdfs in the Plots directory for a single sample.

Value

A barplot with the read size coverage of a selected sample for a given locus.

Author(s)

F. Alberto

References

Alberto F (in prep)...

See Also

reads.plots

score.seq.variants   *Scores alleles based on sequence polymorphism*

Description

The function searches for sequence based polymorphism in a data set of size-only scored alleles.

Usage

```
score.seq.variants(locusInfo="primers",RefGenotypes,sampleDir="./samplesF",
        HeteroThreshold=0.4,alleleDB=NULL,calibrationFiles="CalibrationSamples.txt")
```

Arguments

locusInfo	A tab delimited text file containing the information for the microsatellite loci used. Should contain four columns with the following header names:
	Locus: The loci names
	Fprimer: The sequence of the FWD primer in upper case
	Rprimer: The sequence of the RVS primer in upper case
	Motif: The sequence for the microsatellite motif (e.g., CTTG for tetranucleotide). Note that for some loci you might need to use the reverse complementar sequence of what your reference value is. Thus, inspect your reads first or if you are not getting data for some loci try this out. For example, for a motif CAGG, you might need to use CCTG.
	nmotif: Reads are retained if they have at least these many repetitions of the repeat motif.
RefGenotypes	A tab delimited file with two columns per locus containing the size only based alleles to be inspected for sequence based variation. The format is of file "/GenotypesOut/Genotypes allele table.txt" created by function alleleFinder.
sampleDir	The directory where the filtered reads are stored. This directory should only contain the read files and nothing else. By default this is a directory named samplesF written in the session working directory, as produced by read.quality.filter
HeteroThreshold	
	When sequence variation exists within reads of the same size for the same loci, two variants will dominate the read frequencies, with many more in low frequency due to sequencing errors. This argument refers to the proportion of reads in the second most frequent variant compared to the first. The value sets the minimum allowed proportion required to score the second allele as a sequence-based variant allele, i.e., not a sequencing error. It defaults to 0.4.

alleleDB   An allele database with allele size, sequence variation, new allele code, and nu-
                    cleotide sequence. The default is NULL, in which case the function will produce
                    the database from the data supplied. When a database is supplied, the function
                    will update it with any new variants that might be identified. The format is a
                    tab-delimited table with different sequence variants (rows) and the following re-
                    quired columns:

                    Locus: The locus name for the sequence-based allele. Needs to match the loci
                    names supplied in argument locusInfo.

                    SIZE: The size in bp for the sequence-based allele.

                    VARIANT: A low case letter code to distinguish the different sequence-based
                    alleles that share the same sequence size.

                    SEQ: The nucleotide sequence for each sequence-based allele

                    AlleleCode: A numeric code used to write alleles in format required by most
                    population genetics software.

calibrationFiles

                    The file described below is the same as used for function [GenotypeModel](#) and
                    [alleleFinder](#), in earlier pipeline steps. Here the only information used is the
                    last row containing the sequencing coverage threshold for each marker (number
                    of reads of identical sequence length) that will be applied to filter alleles scored.
                    Inspect the plots produced by function [reads.plots](#) carefully to select values
                    for coverage filtering. Essentialy, use a value below which plots become too am-
                    biguous to confidently call alleles. A tab delimited file with as many columns
                    as loci amplified in the samples. The order of loci is required to match the order
                    in the locusInfo file. The header names must start with the locus name, writ-
                    ten exactly has in the file passed to locusInfo. The first row contains the range
                    of alleles for each locus, the format should be 100-250, for an example of an
                    allelic range between 100 and 250 bp. Users should inspect the plots created
                    by [reads.plots](#) to determine these ranges. Be conservative and add a buffer
                    of about 10 bp on each side but know that the larger the range the slower the
                    function will be. The following rows contain sample names (one per row) for
                    examples of homozygote samples at that column locus. These will be used to
                    generate the expected relative frequencies under different combinations of alle-
                    les. Users can select these homozygote representative samples by inspecting the
                    plots saved in the Plots directory (generated by [reads.plots](#)). Use the exact
                    sample names as the file names stored in directory samplesF. For convenience,
                    sample names are written to the plots and can be copied from there when select-
                    ing homozygote samples.

Details

The function searches for sequence-based polymorphism within a size-only genotype table supplied
by the user (refGenotypes argument to function). The filtered reads are reanalyzed and searched for
sequence-based polymorphism. The analysis of sequence-based variation within loci reads of the

same size reveals many variants in low frequency that represent sequencing errors. Two high-frequency reads, which would otherwise be cryptic (by homoplasy) alleles, are often revealed. The function is intended for diploid data, and therefore, if three or more high-frequency sequence-based alleles are identified, the genotype is scored as NA, and a message is logged to a WarningLog file written to the working directory.

Value

Outputs three files written in a folder created in the working directory named AlleleBySeq. The three files are Allele DataBase.txt, containing the reference database of alllele sequence variants found; Genotypes.by.Sequence.txt, a population genetics file with two columns per locus and alleles represented by the concatenation of allele size and variant letter code; and Genotype Codes.txt contains new numeric allele codes, one column per locus, that are required for most population genetics software. The allele database file contains the correspondence between the different allele code formats. A warningLog file is also written directly to the working directory reporting the cases where more than two alleles were found, in which case NAs were written to the output file.

Author(s)

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References

Alberto F (in prep)...

See Also

GenotypeModel, alleleFinder

Index