experiment 1

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This experiment supercedes all previous ones. It is a factorial manipulation of diversity of the three groups. It takes about 50 hours to run while using 12 cores.

Setup

\mathbf{R}

```
rm(list = ls())
knitr::opts_knit$set(progress = TRUE, verbose = FALSE, cache = TRUE)
microxanox_release <- "0.2"
#tmplib <- tempfile()</pre>
#dir.create(tmplib)
### From '?remotes::install_github`:
# auth_token
   To install from a private repo, generate a personal access token (PAT) in
   "https://github.com/settings/tokens" and supply to this argument. This is
  safer than using a password because you can easily delete a PAT without
   affecting any others. Defaults to the GITHUB_PAT environment variable.
# remotes::install_qithub(
  "opetchey/microxanox",
# ref = microxanox_release,
  # auth_token = "ENTER YOUR TOKEN or PROVED AS ENVIRONMENT VARIABLE",
  build_vignettes = FALSE,
   force = TRUE,
   upgrade = FALSE,
    lib = tmplib
#library(microxanox, lib.loc = tmplib)
library(microxanox)
library(tidyverse)
```

Version of microxanox package used: 0.2

General simulation conditions

```
default_dynamic_model <- bushplus_dynamic_model
default_event_definition <- event_definition_1
default_event_interval <- 100
default_noise_sigma <- 0
default_minimum_abundances <- rep(1, 3)
names(default_minimum_abundances) <- c("CB", "PB", "SB")
default_sim_duration <- 80000
default_sim_sample_interval <- 100
initial_pars_from <- "bush_ssfig3"
## note that next line (log10a_series is over-ridden with getting stable states)
#default_log10a_series <- c(-2, -2, -2, -10, -10, -10, -10, -10)</pre>
```

Define diversity

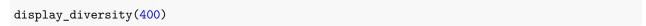
```
num_CB_strains <- 9
num_SB_strains <- 9
num_PB_strains <- 9
CB_gmax_div <- 0.015789474
CB_h_div <- -0.08
SB_gmax_div <- 0.015789474
SB_h_div <- -0.323
PB_gmax_div <- 0.015789474</pre>
```

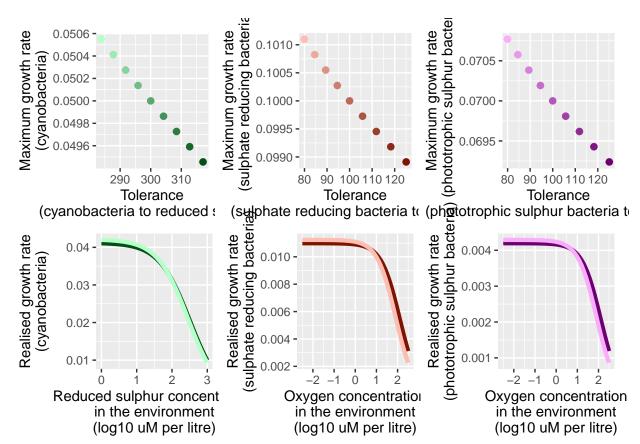
```
PB_h_div <- -0.323
num_div_treatment_levels <- 20
```

Create diversity

```
var_expt <- create_diversity_factorial()</pre>
```

Display diversity



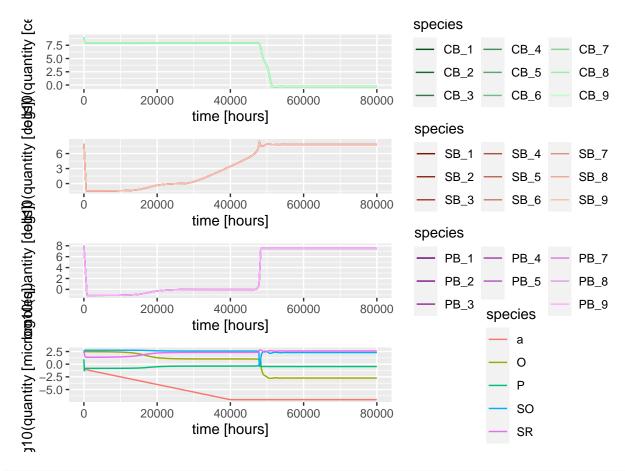


Temporal switching

```
var_expt_levels <- var_expt[,1:6]
no_diversity <- which(rowSums(abs(var_expt_levels))==0)
max_diversty_all <- which(max(rowSums(abs(var_expt_levels))) ==</pre>
```

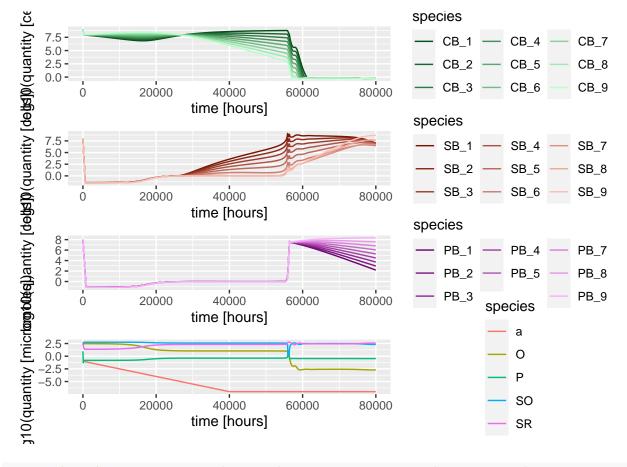
Oxic to anoxic

No diversity



#ggsave(here("simulations/expt2/figures/switching_novar.pdf"), width = 10)

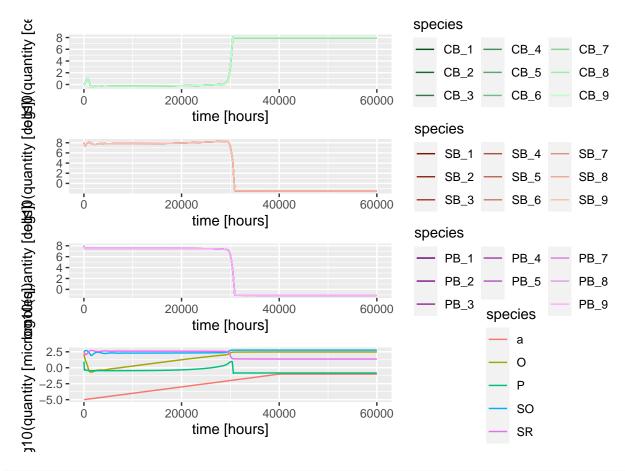
Maximum diversity



#ggsave(here("simulationsexpt2/figures/switching_highvar.pdf"), width = 10)

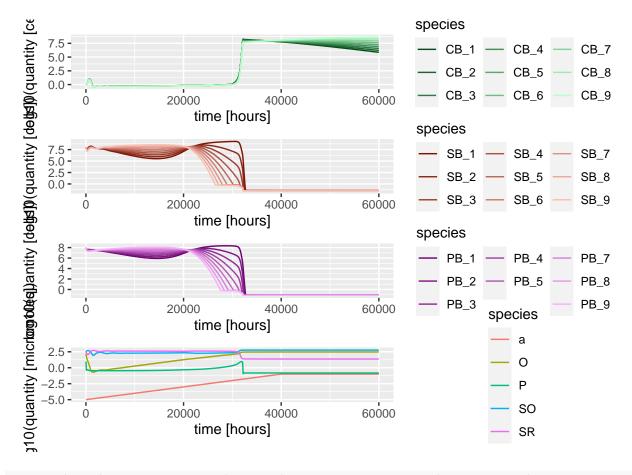
Anoxic to oxic

No diversity



#ggsave(here("simulations/expt2/figures/switching_novar.pdf"), width = 10)

Maximum diversity

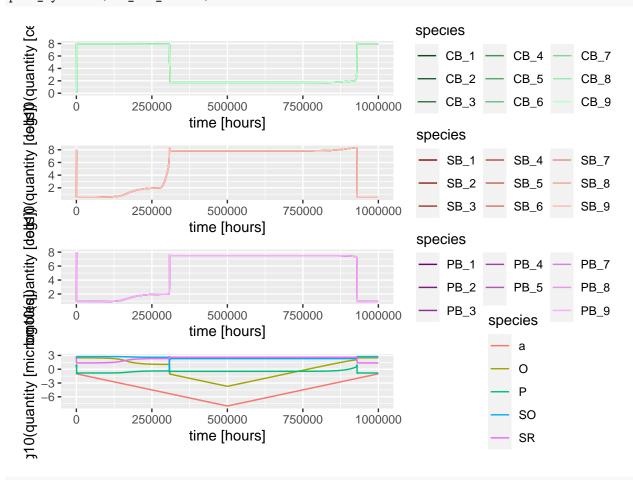


#ggsave(here("simulationsexpt2/figures/switching_highvar.pdf"), width = 10)

Anoxic to oxic to anoxic

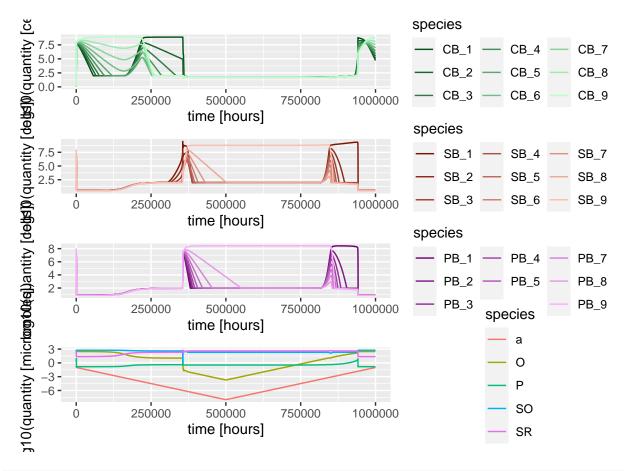
No diversity

sim_res_novar3 <- readRDS(here("experiments/experiment 1/data/sim_res_novar3.RDS"))
plot_dynamics(sim_res_novar3)</pre>



#ggsave(here("simulations/expt2/figures/switching_novar.pdf"), width = 10)

Maximum diversity



#qqsave(here("simulationsexpt2/figures/switching_highvar.pdf"), width = 10)

Visualise

```
visualise_temporal_env_eco()
```

Stable state finding

Setup

```
options(mc.cores = 12)

ssfind_minimum_abundances <- rep(0, 3)
names(ssfind_minimum_abundances) <- c("CB", "PB", "SB")
ssfind_simulation_duration <- default_sim_duration
ssfind_simulation_sampling_interval <- ssfind_simulation_duration
ssfind_event_interval <- ssfind_simulation_duration
grid_num_a <- 1000 #usually 1000 ## number of a_0 values
a_0s <- 10^seq(-7, -1, length=grid_num_a) ## sequence of a_0 values
grid_num_N <- 2 ## number of N values</pre>
```

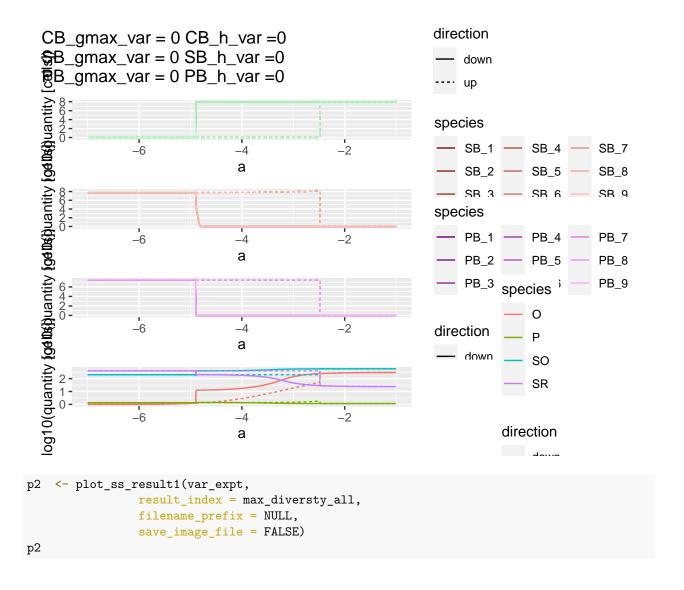
Run stable state finding

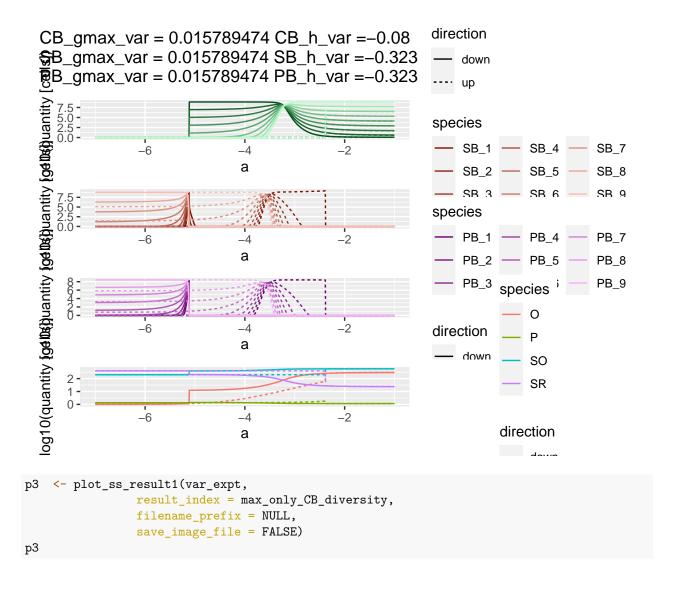
*Careful, this simulation takes about 600 hours on a single core

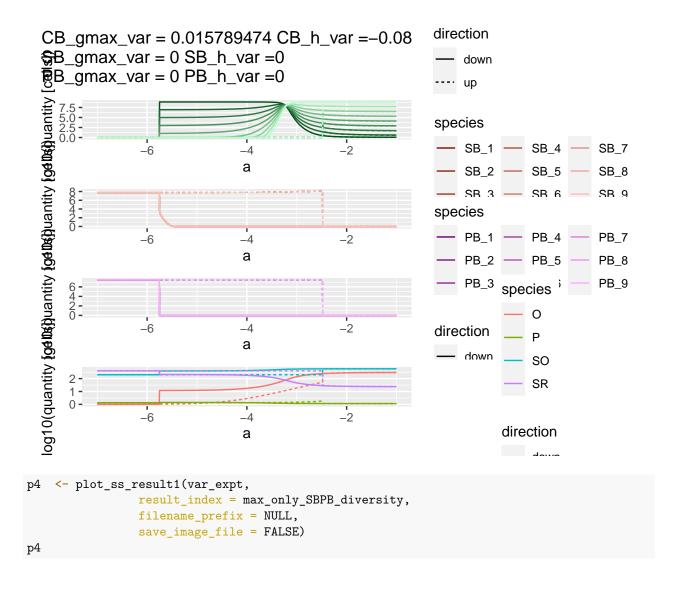
```
#var_expt <- run_ss_var_experiment()
#saveRDS(var_expt, here("experiments/experiment 1/data/ss_data.RDS"))</pre>
```

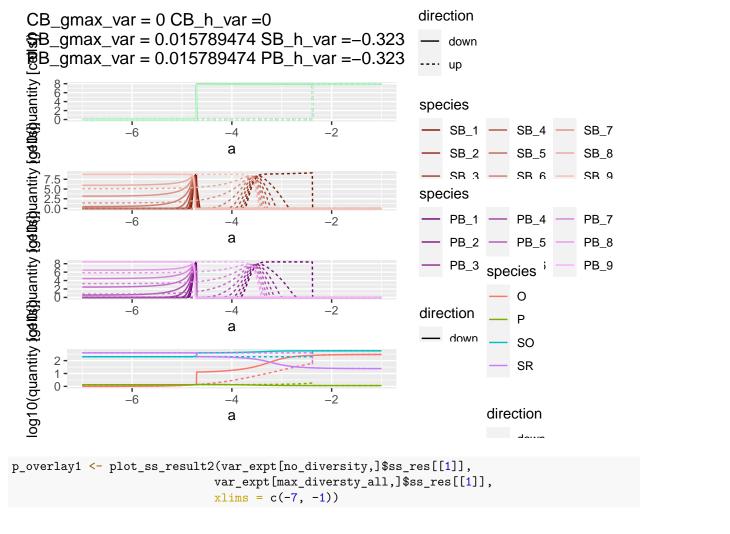
Process the stable state data

various graphs



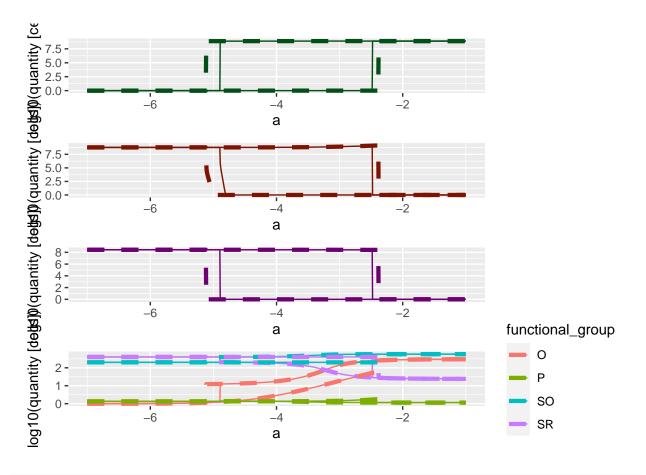






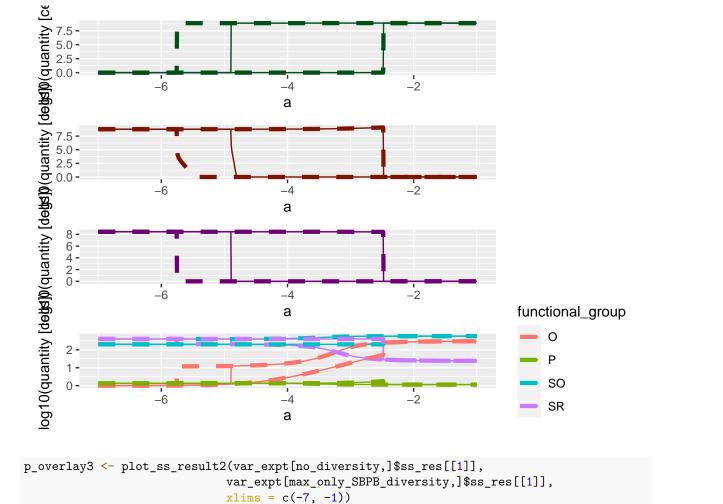
```
## 'summarise()' has grouped output by 'a', 'direction', 'var_type'. You can override using the '.group
## 'summarise()' has grouped output by 'a', 'direction', 'var_type'. You can override using the '.group
```

p_overlay1



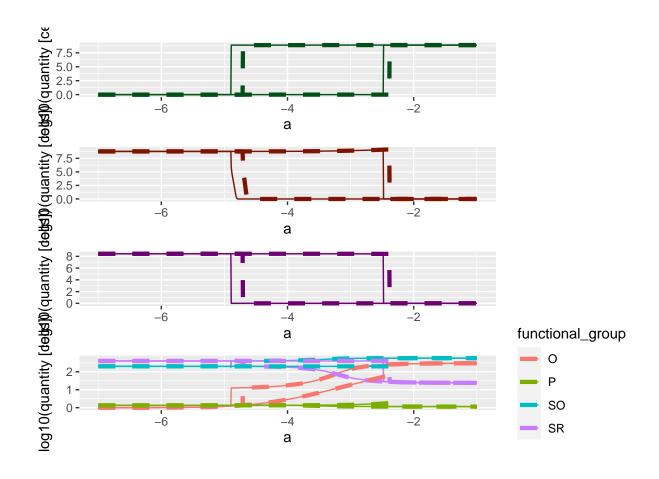
```
## 'summarise()' has grouped output by 'a', 'direction', 'var_type'. You can override using the '.group
## 'summarise()' has grouped output by 'a', 'direction', 'var_type'. You can override using the '.group
```

p_overlay2



```
## 'summarise()' has grouped output by 'a', 'direction', 'var_type'. You can override using the '.group
## 'summarise()' has grouped output by 'a', 'direction', 'var_type'. You can override using the '.group
```

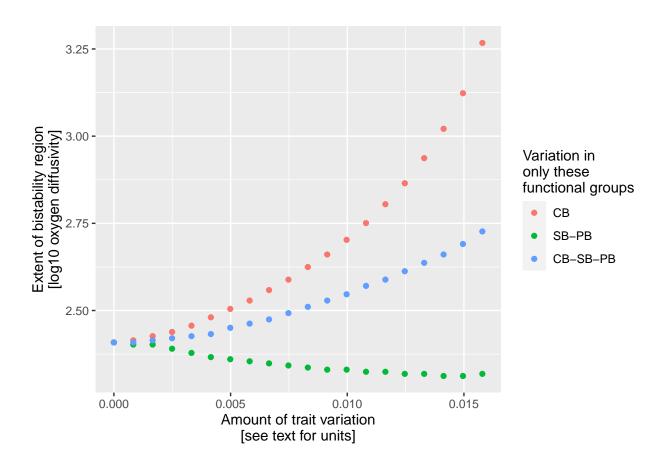
p_overlay3



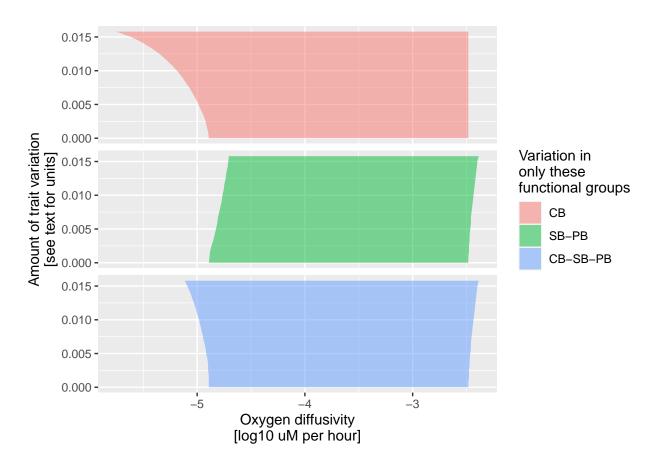
Calculate stability measures

Process and visualise stability measures

```
SBPB_stab_data <- stab_data %>%
  filter(CB_var_gmax_s == 0) %>%
  mutate(var_treat = "SB-PB",
         var_gmax = SB_var_gmax_s)
for_join <- tibble(CB_var_gmax_s = CB_vars,</pre>
                     SB_var_gmax_s = SB_vars)
CBSBPB stab data <- stab data %>%
 right_join(for_join) %>%
 mutate(var_treat = "CB-SB-PB",
        var_gmax = CB_var_gmax_s)
## Joining, by = c("CB_var_gmax_s", "SB_var_gmax_s")
all_stab_results <- CB_stab_data %>%
 bind_rows(SBPB_stab_data) %>%
# bind_rows(results3) %>%
# bind_rows(results4) %>%
 bind_rows(CBSBPB_stab_data)
all_stab_results<- all_stab_results %>%
   mutate(var_treat = forcats::fct_relevel(var_treat, levels = c("CB", "SB-PB", "CB-SB-PB")))
## Warning: Outer names are only allowed for unnamed scalar atomic inputs
#saveRDS(all stab results, here("experiments/experiment summary/all stab.RDS"))
#all_stab_results <- readRDS(here("experiments/experiment summary/all_stab.RDS"))
all_stab_results %>%
  filter(Species == "0") %>%
  ggplot(aes(x = var_gmax, y = hyst_range, col=var_treat)) +
  geom_point() +
 xlab("Amount of trait variation\n[see text for units]") +
  ylab("Extent of bistability region\n[log10 oxygen diffusivity]") +
  labs(col = "Variation in\nonly these\nfunctional groups")
```



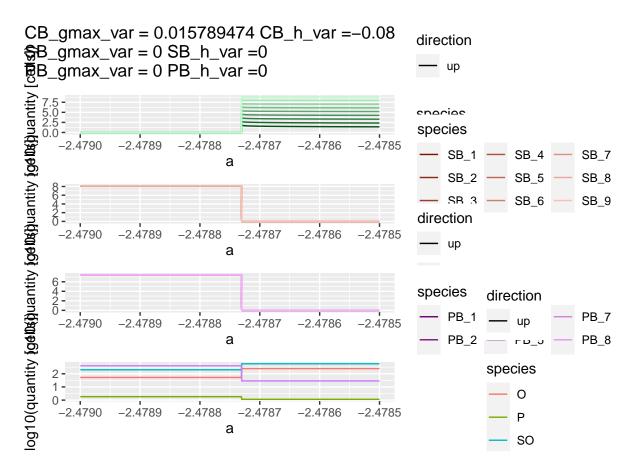
```
##ggsave("manuscript/figures/extent_of_bistab1.pdf", height = 4)
all_stab_results %>%
  #filter(var_treat == "CB") %>%
  filter(Species == "0") %>%
  ggplot(aes(x = var_gmax,
             ymin = hyst_min,
             ymax = hyst_max,
             fill=var_treat)) +
  geom_ribbon(alpha = 0.5) +
  facet_wrap( ~ var_treat, nrow = 3) +
  xlab("Amount of trait variation\n[see text for units]") +
  ylab("Oxygen diffusivity\n[log10 uM per hour]") +
  labs(fill = "Variation in\nonly these\nfunctional groups") +
  coord_flip() +
  theme(
   strip.background = element_blank(),
   strip.text.x = element_blank()
```



##ggsave("manuscript/figures/extent_of_bistab2.pdf", height = 4)

Zoom in on SS

```
result_index = 1,
filename_prefix = NULL,
save_image_file = FALSE)
p1
```



Negative abundance investigation

I (Owen) found that the sampling interval had an effect on the stability of the simulation. If the sampling interval was long, then in some rare cases (see below) the odesolver failed, with negative abundances occurring. I think this is due to abundances becoming very small, and then the computer having trouble with precision. I guess that when a sample is taken, the abundance is somehow altered if it is very low, probably by some rounding.

```
var_expt$pars[[1]]
dd <- var_expt$ss_res[[1]]
dd1 <- filter(dd, PB_1<(-0.0001))
dd1$a_0

ss_expt_master <- ss_expt
ss_expt <- ss_expt_master[abs(ss_expt_master$a_0 - 1.336984e-05)<1e-10,]

var_expt_master <- var_expt
#var_expt <- var_expt[1,]</pre>
```

```
var_expt_test <- run_ss_var_experiment()</pre>
res <- var_expt_test$ss_res[[1]]
test1 <- ss_by_a_N(ss_expt, var_expt$pars[[1]])</pre>
x \leftarrow ss_expt[2,]
param <- var_expt$pars[[1]]</pre>
get_final_states_a_N(x, param)
ssfind parameters <- param
ssfind_simulation_sampling_interval <- 1000
## now run inside the function "get_final_states_a_N"
simres1 <- simres</pre>
ssfind_simulation_sampling_interval <- 5000
## now run inside the function "get_final_states_a_N"
simres2 <- simres # this fails
## now run inside the function "get_final_states_a_N"
plot_dynamics(simres2)
ggplot() +
  geom_line(data = simres1$result,
              mapping = aes(x = time, y = log10(PB_1))) +
  geom_point(data = simres2$result,
              mapping = aes(x = time, y = log10(PB_1))) +
  xlim(c(0, 250000))
ccc <- simres2$result</pre>
simres2$result$PB 1
simres2$result$time
log10_a <- log10(ss_expt$a_0[1]) ## very slowly goes anoxic</pre>
\#log10_a \leftarrow log10(a_0s[354]) \# very slowly goes anoxic
\#loq10_a \leftarrow loq10(a_0s[356]) \# very very very slowly goes anoxic
\#log10_a \leftarrow log10(a_0s[357]) \#\# does not go anoxic
default_dynamic_model <- bushplus_dynamic_model</pre>
default_event_definition <- event_definition_1</pre>
default_event_interval <- ssfind_simulation_duration</pre>
default_noise_sigma <- 0</pre>
default_minimum_abundances <- ssfind_minimum_abundances</pre>
default_sim_duration <- ssfind_simulation_duration</pre>
default_sim_sample_interval <- ssfind_simulation_duration</pre>
#initial_pars_from <- "bush_ssfig3"</pre>
default_log10a_series <- c(log10_a, log10_a)</pre>
initial_state <- new_initial_state(num_CB_strains,</pre>
                                      num_PB_strains,
                                      num_SB_strains,
                                      values = "bush_ssfig3")
```