## Figure-1A.R

## sokole

## 2024-07-15

```
# This Script Generates Figure 1A
# Script By: Eishani Kumar Sokolowski

#Empty the environment & suppress warnings
rm(list = ls())
options(warn=-1)

# Loading the required libraries
library(tidyverse)
```

```
## — Attaching core tidyverse packages —
                                                               —— tidyverse 2.0.0 —
## ✓ dplyr
              1.1.4
                          ✓ readr
                                       2.1.5
## ✓ forcats 1.0.0
                                      1.5.1

✓ stringr

## ✓ ggplot2 3.5.1
                                      3.2.1

✓ tibble

## 🗸 lubridate 1.9.3

✓ tidyr

                                      1.3.1
## ✓ purrr
               1.0.2
## — Conflicts —
                                                         —— tidyverse_conflicts() —
## * dplyr::filter() masks stats::filter()
## x dplyr::lag()
                     masks stats::lag()
## i Use the conflicted package (<a href="http://conflicted.r-lib.org/">http://conflicted.r-lib.org/</a>) to force all conflicts
to become errors
```

```
library(pheatmap)
library(Vennerable)
library(gplots)
```

```
##
## Attaching package: 'gplots'
##
## The following object is masked from 'package:stats':
##
## lowess
```

```
# Load metadata
metadata <- read.csv("./Metadata_Heatmap.csv")</pre>
rownames(metadata) <- metadata$Islet_Number</pre>
# Load normalized expression matrices
my.data.1 <- read.csv("./normalized.with.ERCC.counts.for.heatmap.ER.Stress.csv")</pre>
my.data.2 <- read.csv("./normalized.with.ERCC.counts.for.heatmap.Inflammation.csv")</pre>
my.data <- merge(my.data.1,my.data.2, by="X")</pre>
rownames(my.data) <- my.data[,1]</pre>
my.data <- my.data[-c(1)]</pre>
# Load DEGs
ERS.specific <- read.csv("./Upregulated_only_in_ER_Stress_DEGs.csv")</pre>
ERS.specific <- ERS.specific[-c(2:7)]</pre>
ERS.specific$Regulation <- c("Upregulated_ERS_specific")</pre>
INF.specific <- read.csv("./Upregulated only in Inflammation DEGs.csv")</pre>
INF.specific <- INF.specific[-c(2:7)]</pre>
INF.specific$Regulation <- c("Upregulated INF specific")</pre>
Shared <- read.csv("./Upregulated_common_between_ER_Stress_Inflammation_DEGs.csv")</pre>
Shared \leftarrow Shared [-c(2:7)]
Shared$Regulation <- c("Upregulated ERS and INF Shared")
# Making annotation dataframes
metadata.annotation <- metadata</pre>
metadata.annotation <- metadata.annotation[order(metadata.annotation$Treatment),]</pre>
metadata.annotation <- metadata.annotation[-c(1:7)]</pre>
metadata.annotation.names <- rownames(metadata.annotation)</pre>
matrix.annotation <- rbind(ERS.specific,INF.specific,Shared)</pre>
rownames(matrix.annotation) <- matrix.annotation$X
matrix.annotation <- matrix.annotation[-c(1)]</pre>
# Making the heatmap matrix
genes.list <- rownames(matrix.annotation)</pre>
matrix <- my.data[(genes.list),]</pre>
matrix.ordered <- matrix %>% select(metadata.annotation.names)
matrix.ordered <- as.matrix.data.frame(matrix.ordered)</pre>
matrix.ordered <- na.omit(matrix.ordered)</pre>
matrix.ordered[is.na(matrix.ordered)] <- 0</pre>
# Specify colors
ann_colors = list(
  Treatment = c(A_DMS0="gray", B_Thapsigargin="forestgreen",C_Untreated="grey50",D_Cytok
ines="darkorange"),
  Regulation = c(Upregulated_ERS_specific = "forestgreen",
                  Upregulated_INF_specific="darkorange",
                  Upregulated ERS and INF Shared = "coral4")
)
# Make heatmap
pheatmap(matrix.ordered,
```

```
scale = "row",
clustering_distance_rows = "correlation",
treeheight_row = 0,
treeheight_col = 0,
annotation_row = matrix.annotation,
annotation_col = metadata.annotation,
cluster_row = F,
cluster_cols = F,
fontsize_row = 0.01,
fontsize_col = 0.01,
annotation_colors = ann_colors,
color=bluered(1000))
```

