Gene Sequence Analysis

Lecture 2: Pairwise Alignment

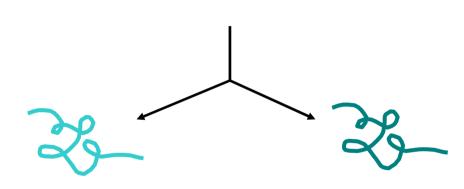
30/05/2024 Phuc-Loi Luu, PhD

Adapted from Dr. Morgan Langille's Lecture

Importance of Similarity

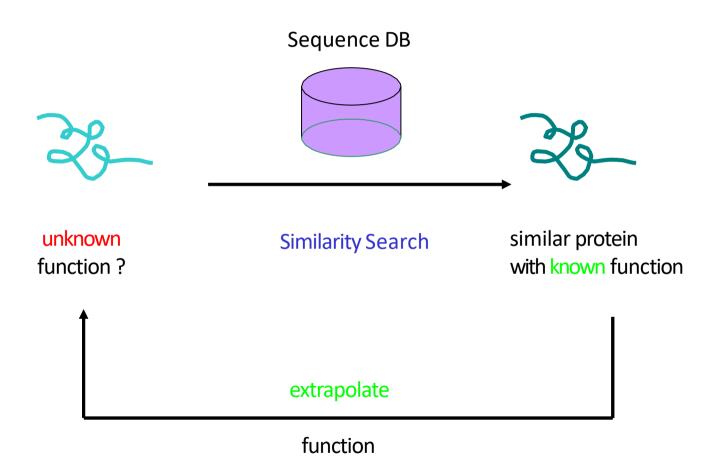
Ancestral protein/gene sequence

Similar (homologous) protein/gene sequences



Similar sequences: probably have the same ancestor, share the same structure, and have a similar biological function

Importance of Similarity



Importance of Similarity

Rule-of-thumb:

If your sequences are more than **100 amino acids** long (or 100 nucleotides long) you can considered them as homologues if **25%** of the **aa** are identical (**70%** of **nucleotide** for DNA). Below this value you enter the twilight zone.

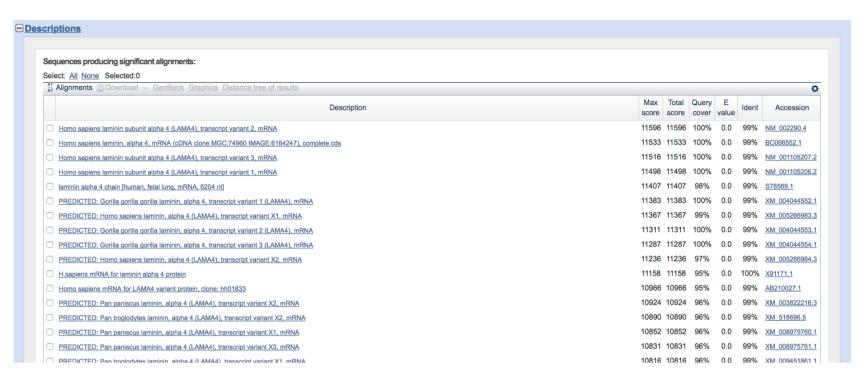
Twilight zone = protein sequence similarity between ~0-20% identity: is not statistically significant, i.e. could have arisen by chance.

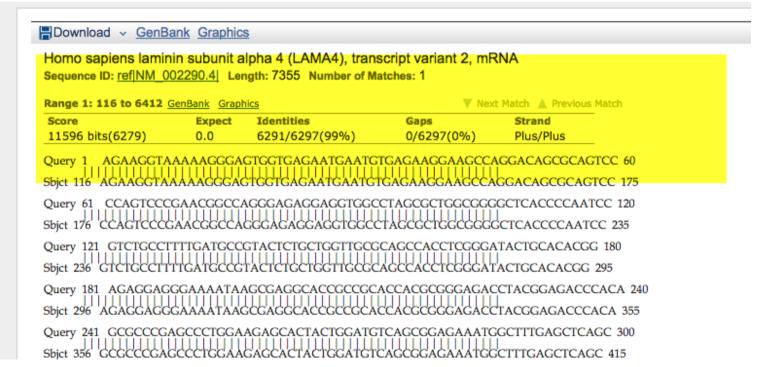
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Beware:

- E-value (Expectation value)
- length of the segments similar between the two sequences
- The number of insertions/deletions

BLAST, FASTA, SSEARCH, and other commonly used similarity searching programs produce accurate statistical estimates that can be used to reliably infer homology. Searches with protein sequences (BLASTP, FASTP, SSEARCH,) or translated DNA sequences (BLASTX, FASTX) are preferred because they are 5- to 10-fold more sensitive than DNA:DNA sequence comparison. The 30% identity rule-of-thumb is too conservative; statistically significant $[E() < 10^{-6} - 10^{-3}]$ protein homologs can share less than 20% identity. E()-values and bit scores (bits >50) are far more sensitive and reliable than percent identity for inferring homology.





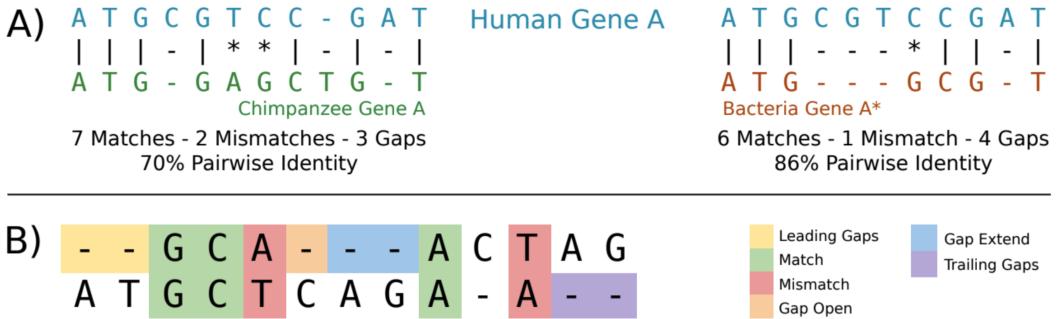


FIGURE 1.2: **Similarity versus homology and alignment scoring:** *A)* Two sequence alignments to Human Gene A highlighting how true homologs (Chimpanzee Gene A) can have similar similarity to non-homologous sequences (Bacteria Gene A*). *B)* A sequence alignment with different reward or penalties highlighted to illustrate how homology search algorithms score alignments based on sequence similarity.

Doolittle's twilight zone is between 15-25% and is a 'rule of thumb' for sequences of 100 amino acid length or greater (29). The 100 amino acids caveat is due to the greater probability of short sequences having the same amino acid sequence by chance and therefore the level of confidence in short alignments is not as easy to give general advice for (19, 29). No quantitative experiment was done to determine this

As a part of a study of structural ncRNAs that were used to benchmark multiple sequence alignment programs the twilight zone for nucleotide alignment accuracy was calculated (11). It was between 50-60% which is much higher then the 10-20% calculated by Thompson *et al.* 1999 for protein alignments (32). This zone represents

billion years ago (e.g. humans to bacteria). Moreover, DNA:DNA alignment statistics are less accurate than protein:protein statistics; while protein:protein alignments with expectation values < 0.001 can reliably be used to infer homology, DNA:DNA expectation values $< 10^{-6}$ often occur by chance, and 10^{-10} is a more widely accepted threshold for homology based on DNA:DNA searches. The most effective way to improve search sensitivity with DNA sequences is to use translated-DNA:protein alignments, such as those produced by BLASTX and FASTX, rather than DNA:DNA alignments.

Outline

- PSSMs/PSI-BLAST
- HMMs/HMMer RNA Alignments
- Genome Alignments
- Assemblers Mappers

Different tools for homology searching

- Searching for protein families
- Aligning genomes
- Looking for RNA genes
- Combining overlapping sequences (assemblers)
- Finding the position of a sequence in a genome

One tool does not do it all

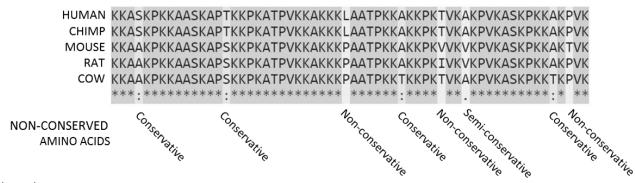
Blast may give you an answer

 BUT you could find the answer much quicker or with more precision by using the right tool!

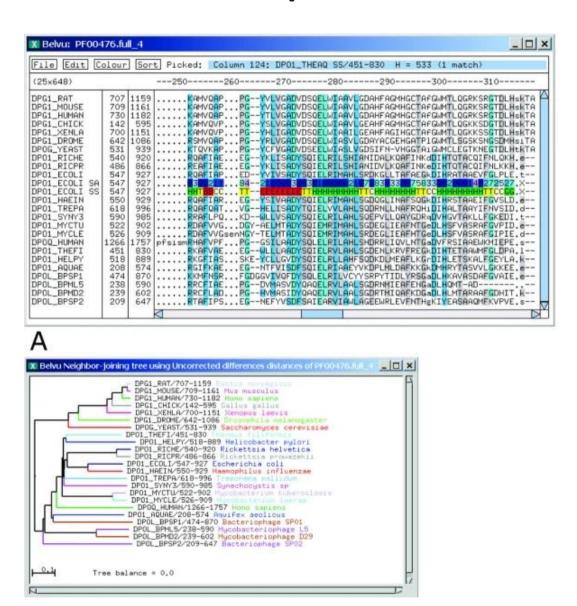
Typical BLAST output



Histone H1 (residues 120-180)



Distance related proteins



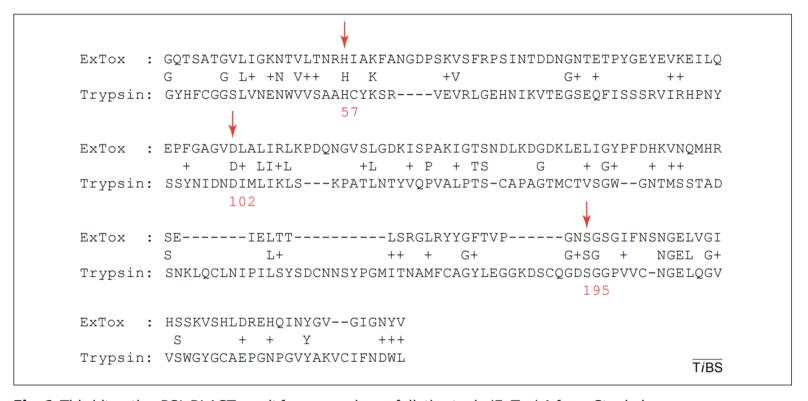


Fig. 1. Third-iteration PSI-BLAST result from querying exfoliative toxin (ExTox) A from *Staphylococcus aureus* (BAA97652.1) against the non-redundant database. His, Asp and Ser residues are indicated with arrows and numbered as for trypsin [anionic; complexed with the inhibitor benzamidine (1bit)]. The alignment has 15% identity (32/206) and the *E*-value = 6×10^{-21} . The threshold *E*-value for inclusion in the profile was 0.005 and the effective search space was 22 926 875 677.

```
Gi|2622094 (AE000872) conserved protein [Methanobacterium thermoautotrophicum]

Length = 143

Score = 84.7 bits (206), Expect = 4e-16

Identities = 56/156 (35%), Positives = 81/156 (51%), Gaps = 16/156 (10%)

Query: 4 MYKKLLYPTDFSETAEIALKHVKAFKTLKAEEVILLHVIDEREIKKRDIFSLLLGVAGLN 63
MY KIL PTD S+ & & +H E+I L V++ S L+G+

Sbjct: 1 MYSKILLPTDGSKQANKAAEHAIWIARESGAEIIALTVMET-----SSLVGLPA-- 49

Query: 64 KSVEEFENELKNKLTEEAKNKMENIKKELEDVGFKVKDIIVV--GIPHEEIVKIAEDEGV 121

++ L+ L EEA +E +KK +E+ G +K + G P E I++ E EGV

Sbjct: 50 ---DDLIIRLREMLEEEASRSLEAVKKLVEESGADIKLTVRTDEGSPAEAILRTVEKEGV 106

Query: 122 DIIIMGSHGKTNLKEILLGSVTENVIKKSNKPVLVV 157

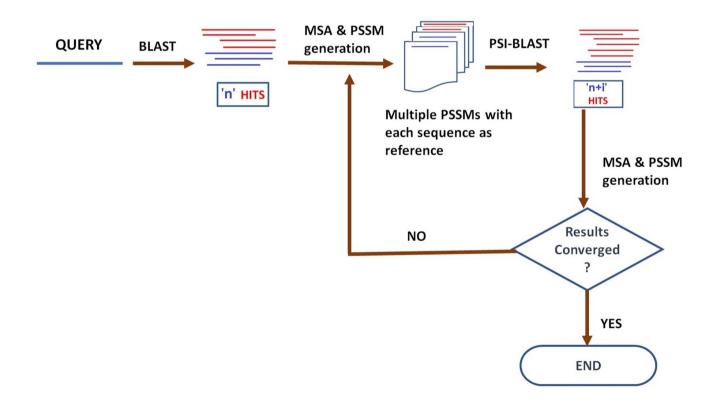
D+++MG+ GK L LLGSV E V++ + PVLVV

Sbjct: 107 DLVVMGTSGKHGLDRFLLGSVAEKVVRSAGCPVLVV 142
```

An example of high-scoring segment pairs (HSP) found by PSI-BLAST. The first peptide pairs as marked by the box are similar, and we assign the secondary structure element of each amino acid in MYKKILY to its counterpart in MYSKILL.

- Position Specific Iterated BLAST A
- cycling/iterative method
 - Gives increased sensitivity for detecting distantly
 - related proteins
 - Can give insight into functional relationships
 - Very refined statistical methods
- Fast still based on BLAST methods
- Simple to use

• Essentially we are using intermediate sequences to infer similarity between two sequences that are too dissimilar to link directly.



Profiles & PSSMs Need Multi-sequence Alignment

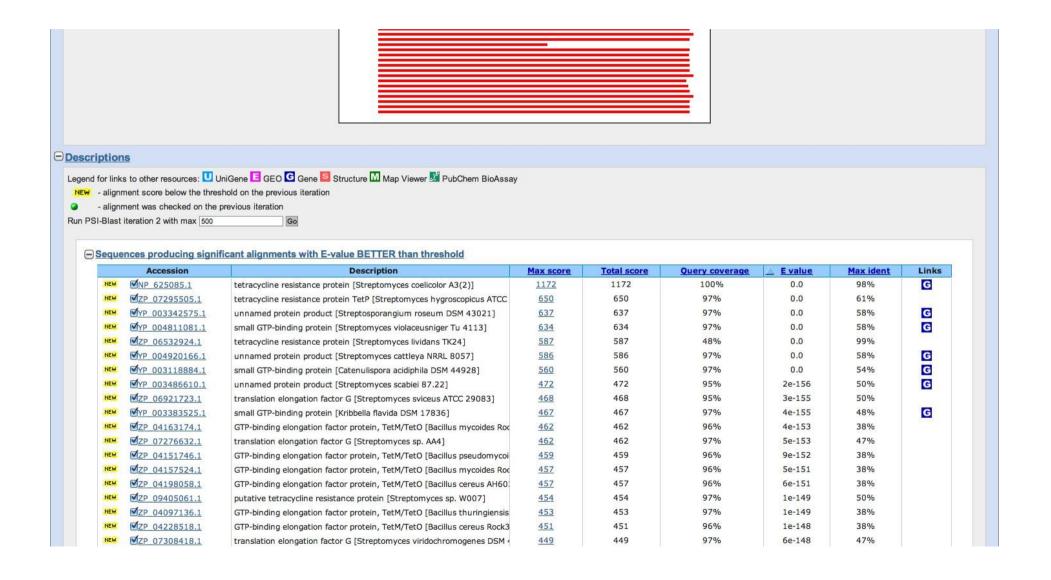
	1				50
P43871-1		IKKLDSN	SIHAIIS <mark>DIP</mark>	YGIDYDDWDI	LHSNTNSALG
S18997-1	LMSKIYQMDA	VDWLKTLENC	SVDLFIT <mark>DPP</mark>	YESL.EKYRQ	IGTTTRLKES
P23192-1	EINKIHQMNC	FDFLDQVENK	SVQLAVI <mark>DPP</mark>	YNL	
P29538-1	MDQRLICSNA	IKALKNLEEN	SIDLIIT <mark>DPP</mark>	YNLG.KDY	
P14751-1	TRHVYDVCDC	LDTLAKLPDD	SVQLIIC <mark>DPP</mark>	YNI	
P34721-1	KNFNIYQGNC	IDFMSHFQDN	SIDMIFA <mark>DPP</mark>	YFLS.NDG.L	TFKNSIIQ
P50178-1	ENAILVHADS	FKLLEKIKPE	SMDMIFA <mark>DPP</mark>	YFLS.NGG.M	SNSGGQIV
P20590-1	FLNTILKGDC	IEKLKTIPNE	SIDLIFA <mark>DPP</mark>	YFMQ.TEGKL	LRTNGDEF
S43876-1	GPETIIHGDC	IEQMNALPEK	SVDLIFA <mark>DPP</mark>	YNLQ.LGGDL	LRPDNSKV
P28638-1	EAKTIIHGDA	LAELKKIPAE	SVDLIFA <mark>DPP</mark>	YNIG.KNF	
P23941-1	DLGKLYNGDC	LELFKQVPDE	NVDTIFA <mark>DPP</mark>	FNLD.KEY	
P14230-1	RSCKIIVGDA	REAVQGLDSE	IFDCVVT <mark>SPP</mark>	YWGL.RDY	
P14243-1	NGATLFEGDA	LSVLRRLPSG	SVRCIVT <mark>SPP</mark>	YWGL.RDY	
Q04845-1	LNNMLLQGNC	AETLKKLPDE	SVNLVFT <mark>SPP</mark>	<u>Y</u> Y	
S53866-1	WVNDIHEGDA	EEVLAELPES	SVHMVMT <mark>SPP</mark>	YFGL.RDY	
P29568-1		MNELKDK	SINLVVT <mark>SPP</mark>	YPMV.EIWDR	LFSELNPKIE

Signature Sequence:

How does PSI-BLAST work?

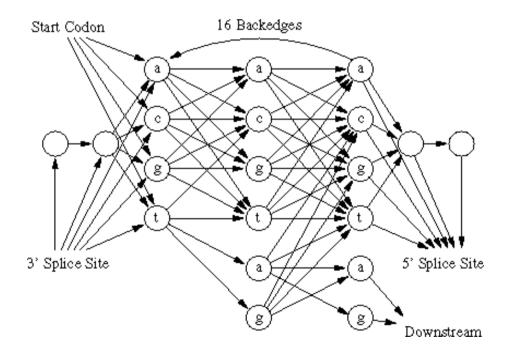
- 1) First, a standard blastp is performed
- 2) The highest scoring hits are used to generate a multiple alignment
- 3) A Position Specific Scoring Matrix (PSSM) is generated from the multiple alignment.
 - Highly conserved residues get high scores
 - Less conserved residues get lower scores
 - The PSSM describes the sequence similarity between your query and all significant blastp hits
- 4) Another similarity search is performed, this time using the new PSSM as the query sequence.
 - This PSSM (scoring matrix) is now customized to find sequences that are related to your original query
 - Steps 2-4 can be repeated until convergence
 - Convergence occurs when no new sequences appear after iteration

PSI-BLAST Example



HMMs & HMMer

The more powerful way to search for protein families than PSSMs



Hidden Markov Models

Hidden Markov Models in Bioinformatics

Used extensively in gene prediction

 Used to create Sequence Profiles and to classify sequences into families

Used in Multiple Sequence Alignment

HMMER 3

 Suite of sequence analysis programs based on HMMs

Used to build the Pfam database

- Available for free download at
 - http://hmmer.org/

HMMER 3

- HMMER 2 was used for many years
 - Biggest draw back was always speed

- HMMER 3 released in 2011
 - Very fast with comparable speeds to BLAST
 - 100X faster than v2

HMMER Programs

- hmmbuild build a HMM from multiple sequence alignment
- hmmscan searches a query sequence(s) against a database of HMMs (used by PFAM)
- hmmsearch—searches a query HMM against a database of sequences (e.g. like psi-blast)
- phmmer search a protein sequence vs a sequence database (e.g. like blastp)

HMMER Search & Software

http://hmmer.janelia.org/search

- PFAM
 - http://pfam.sanger.ac.uk/

RNA Alignments

- RNA alignments are "special"
- RNA genes often have secondary structures that allow improved searching
- Improved searching is needed since
 - Must search in DNA space (less complex then protein sequences)
 - Often shorter length then proteins

Infernal (RNA Search)

- Infernal is like HMMER
 - Includes use of secondary structure information
 - Uses profile "stochastic context-free grammar"
 - SCFGs vs HMMs
 - "consensus RNA secondary structure profiles"
- Infernal is slow!
- Infernal can be used to search RFAM

RNAseq alignment with Magic-BLAST

Software | Open Access | Published: 25 July 2019

Magic-BLAST, an accurate RNA-seq aligner for long and short reads

Grzegorz M. Boratyn, Jean Thierry-Mieg, Danielle Thierry-Mieg, Ben Busby & Thomas L. Madden

<u>BMC Bioinformatics</u> **20**, Article number: 405 (2019) | <u>Cite this article</u> **10k** Accesses | **52** Citations | **51** Altmetric | Metrics

Abstract

Background

Next-generation sequencing technologies can produce tens of millions of reads, often pairedend, from transcripts or genomes. But few programs can align RNA on the genome and accurately discover introns, especially with long reads. We introduce Magic-BLAST, a new aligner based on ideas from the Magic pipeline.

Results

Magic-BLAST uses innovative techniques that include the optimization of a spliced alignment score and selective masking during seed selection. We evaluate the performance of Magic-BLAST to accurately map short or long sequences and its ability to discover introns on real RNA-seq data sets from PacBio, Roche and Illumina runs, and on six benchmarks, and compare it to other popular aligners. Additionally, we look at alignments of human idealized RefSeq mRNA sequences perfectly matching the genome.

RNAseq alignment with Magic-BLAST

NCBI Magic-BLAST RNA-seq mapping tool

NCBI Magic-BLAST Documentation

FASTQ

Paired reads

RNA vs DNA

Multi-threading

Magic-BLAST is a tool for mapping large next-generation RNA or DNA sequencing runs against a whole genome or transcriptome. Each alignment optimizes a composite score, taking into account simultaneously the two reads of a pair, and in case of RNA-seg, locating the candidate introns and adding up the score of all exons. This is very different from other versions of BLAST, where each exon is scored as a separate hit and read-pairing is ignored.

Magic-BLAST incorporates within the NCBI BLAST code framework ideas developed in the NCBI Magic pipeline, in particular hit extensions by local walk and jump (http://www.ncbi.nlm.nih.gov/pubmed/26109056), and recursive clipping of mismatches near the edges of the reads, which avoids accumulating artefactual mismatches near splice sites and is needed to distinguish short indels from substitutions near the edges.

More details about the algorithm and comparison with other similar tools are published here

Boratyn GM, Thierry-Mieg J, Thierry-Mieg D, Busby B, Madden TL. (2019) Magic-BLAST, an accurate RNA-seq aligner for long and short reads. BMC

We call the whole next generation run (from Illumina, Roche-454, ABI, or another sequencing platform excluding SOLiD), a query. The input reads may be provided as SRA accession or a file in a SRA, FASTA, and FASTQ format. Read pairs can be presented as parallel files, or as successive reads in a single

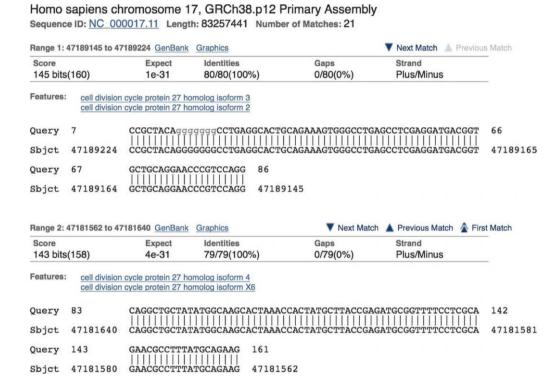
The reference genome or transcriptome can be given as a BLAST database or a FASTA file. It is preferable to use BLAST database for large genomes, such as human, or transcript collections, such as all of RefSeq, Ensembl, or AceView, See here on how to create a BLAST databa-

The full list of options is listed when you use -help option.

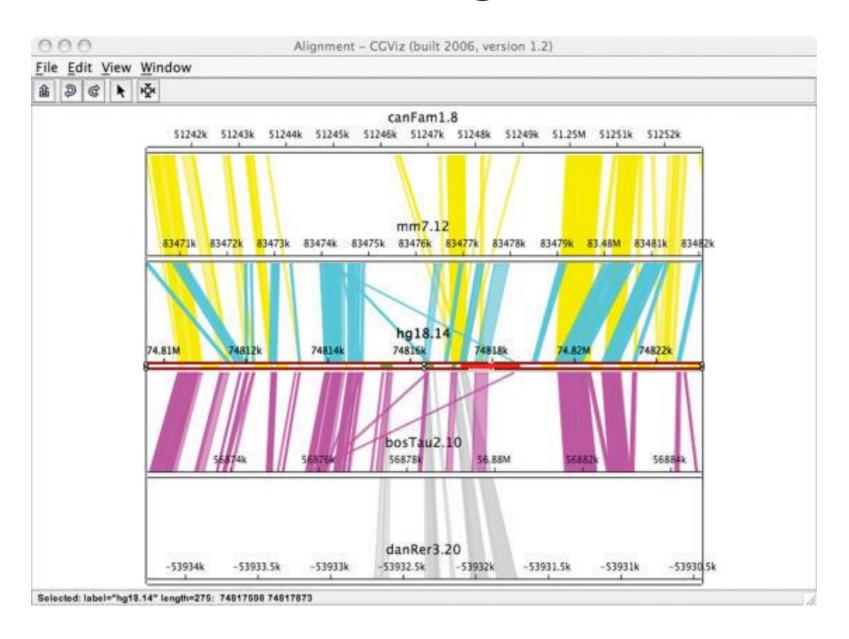
Thank you for trying this tool and providing us with feedback. Please, let us know of any desired enhancement, problem or difficulty

E-mail blast-help@ncbi.nlm.nih.gov with questions or comments

Download NCBI Magic-BLAST



Genome Alignment



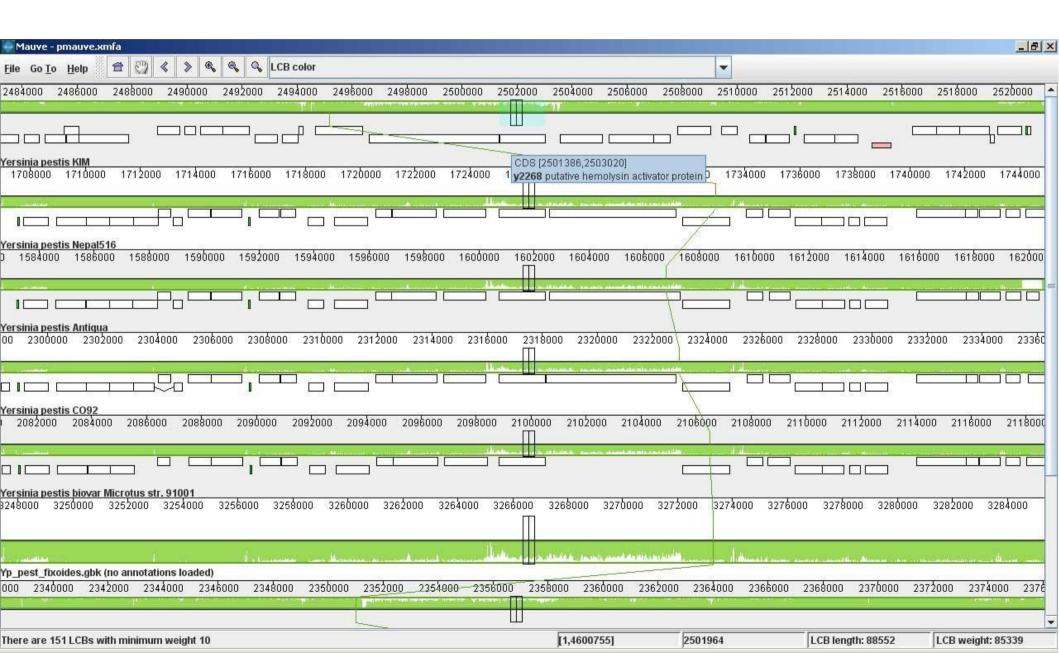
Genome Alignment

- Genome alignment useful for
 - Visualizing genome
 - Rearrangements
 - Insertions/deletions
 - Inversions
 - Annotating genomes
 - Comparing gene annotations across species

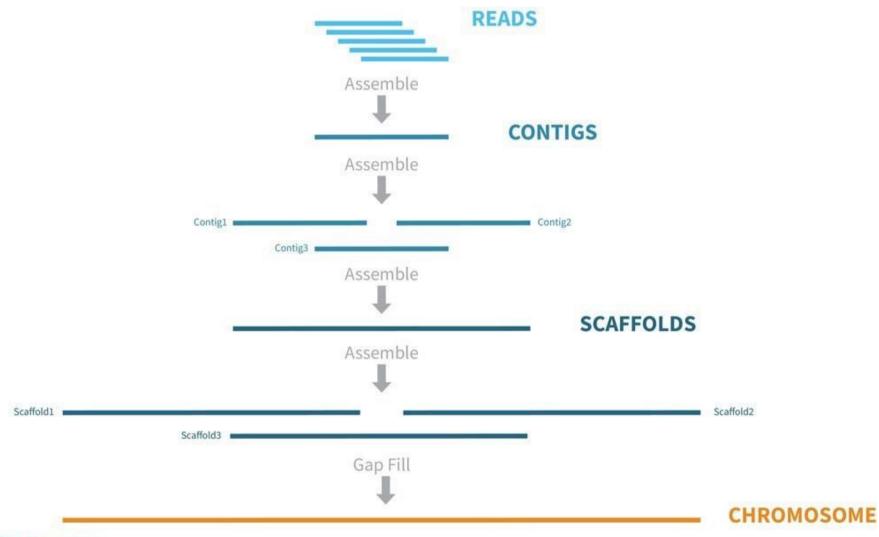
Mauve



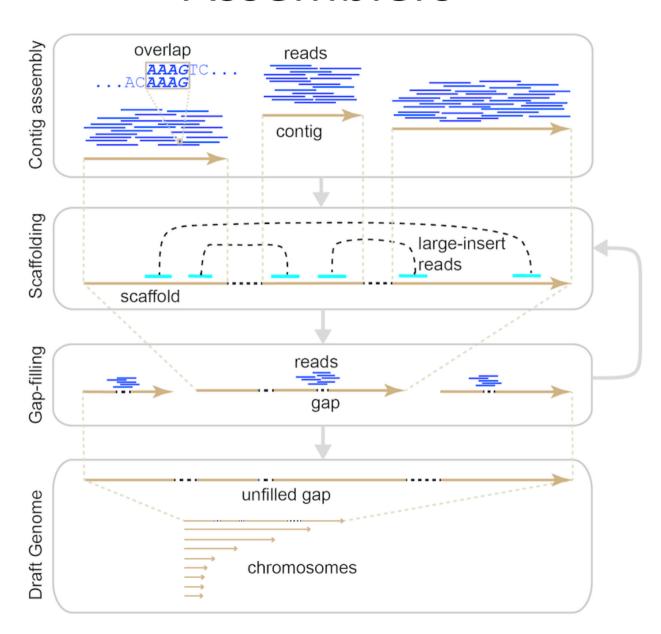
Mauve (zoomed in)



Assemblers



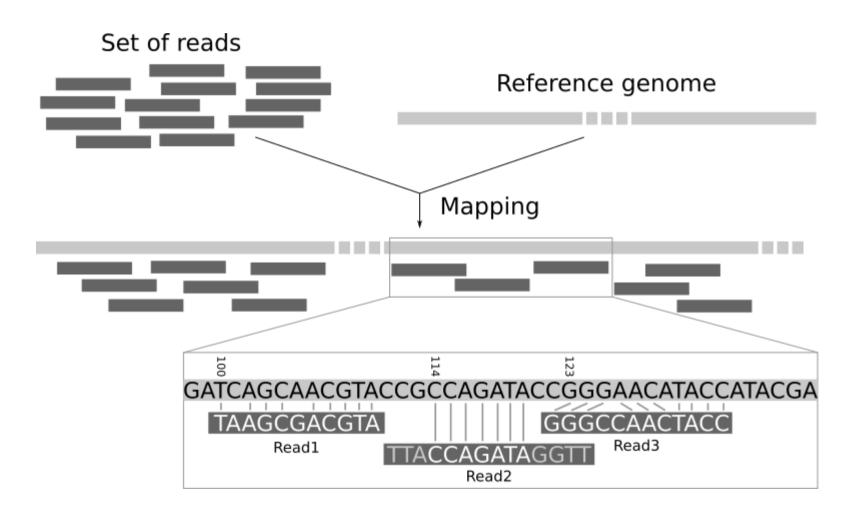
Assemblers



Assemblers

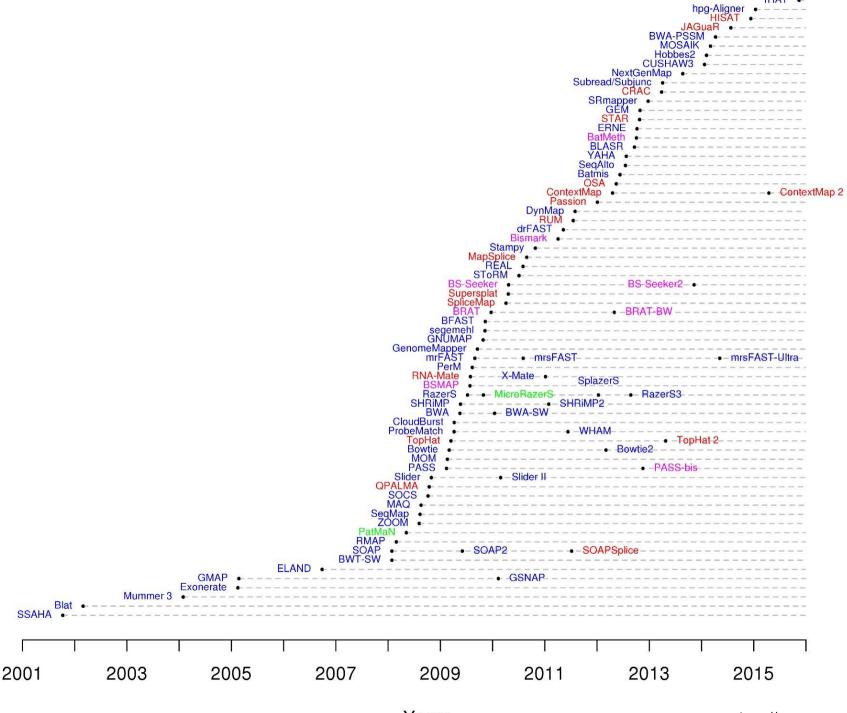
- Assemblers job is to make longer sequences from shorter ones.
- Nothing like homology searching
- Must efficiently compare and join billions of sequences
- Soap-Denovo: http://soap.genomics.org.cn/soapdenovo.html
- Amos: http://sourceforge.net/apps/mediawiki/amos/index.php?title=AMOS
- Many, many, more

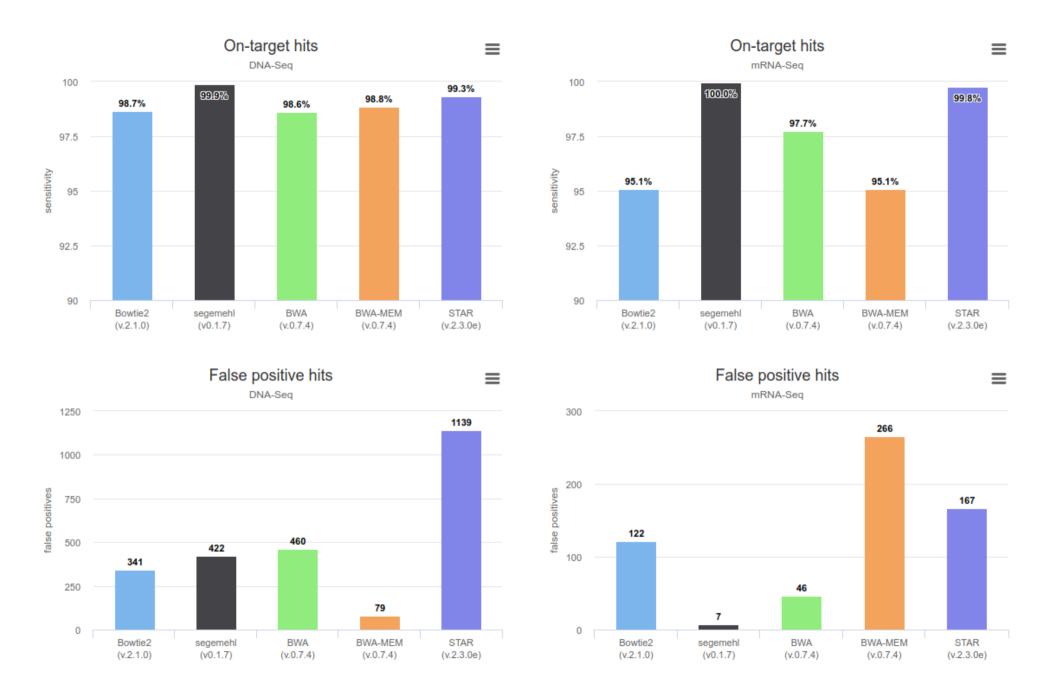
Mappers (Aligners)

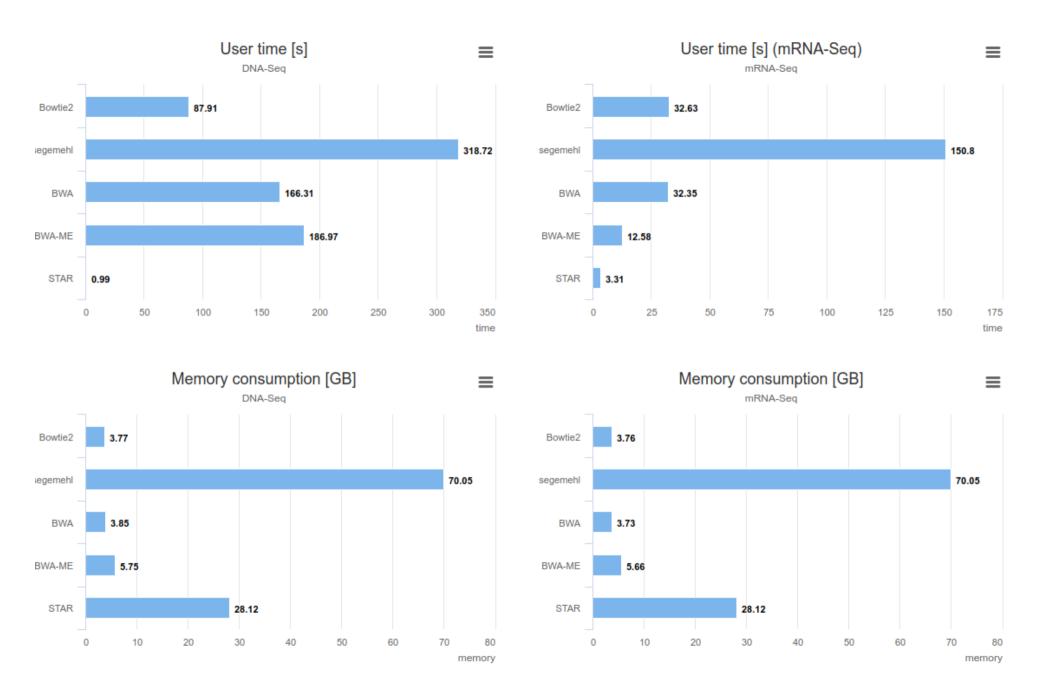


Mappers

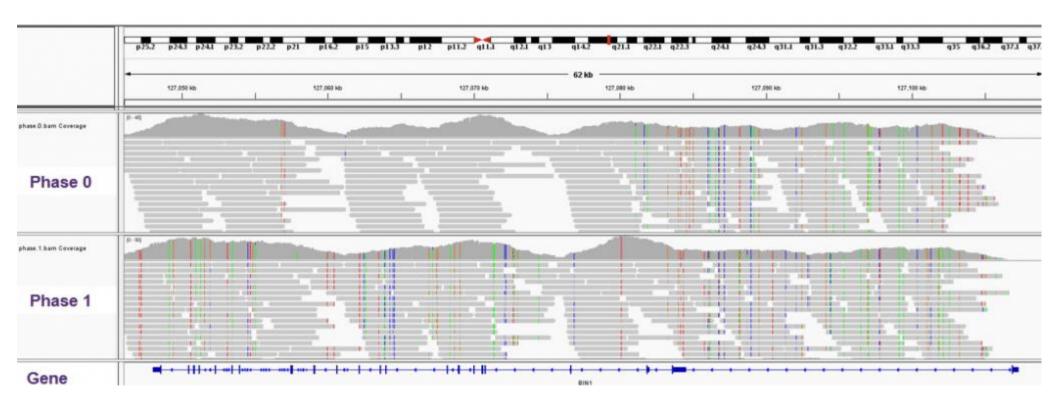
- These map a read to a reference genome
- Useful for assembly when a reference genome is already known
 - (think assembly of personal human genomes)
- Identifying SNPs within the same species Very Fast!
- BWA: https://github.com/lh3/bwa
- Bowtie: http://bowtie-bio.sourceforge.net/index.shtml
- Stampy: http://www.well.ox.ac.uk/project-stampy
- Many Others







Mappers



IGV snapshot with reads in bam file