Introduction to R for Microbiome Data (part 1)

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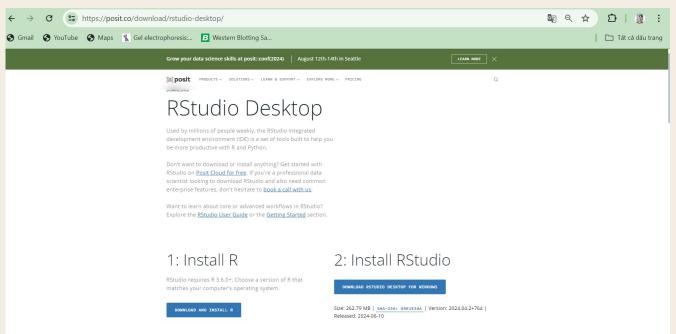
30/06/2024

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https://posit.co/download/rstudio-desktop/

https://www.youtube.com/watch?v=dsjF57fjL40&list=PLMIaO-u3S5-jO2rMt8r8HD5ifiZv_Sd9Ohttps://www.youtube.com/watch?v=ngpB_00Jiyk&t=825s

. . .

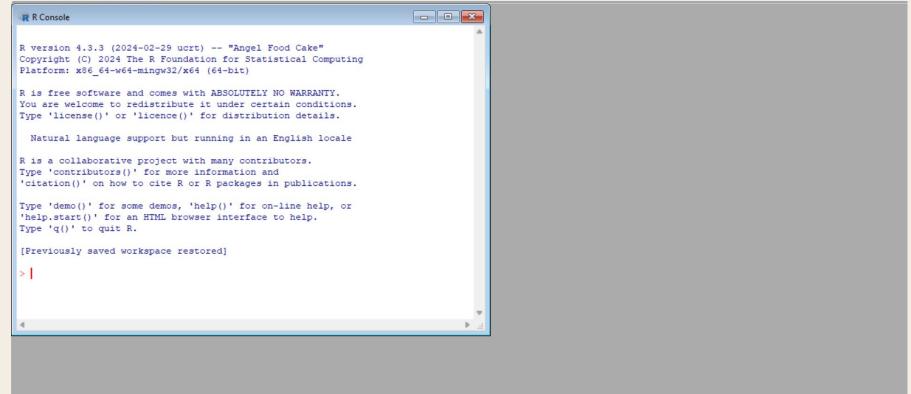
. . .

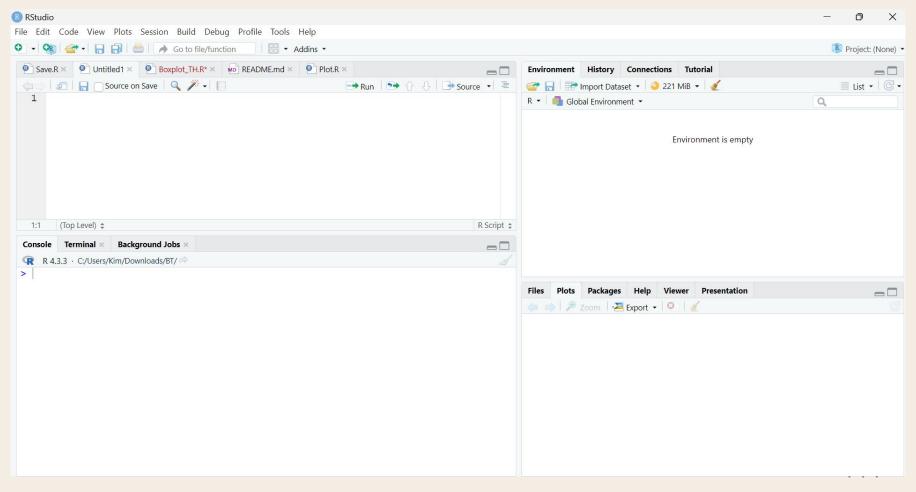
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RGui (64-bit)

File Edit View Misc Packages Windows Help







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1. Some Useful R Functions

1.1. Use SaveRDS() to Save Single R Object

```
# Save an object to a file
saveRDS(object, file = "my_data.rds")
# Restore the object
readRDS(file = "my_data.rds")
```

- object: An R object to save
- file: the name of the file where the R object is saved to or read from

Example:

```
# Saving data as RDS
saveRDS(microbiome_data,"microbiome_data.rds")
# Reading data from RDS file
readRDS("microbiome_data.rds")
# Restore it under a different name
loaded_data <- readRDS("microbiome_data.rds")
head(loaded_data)</pre>
```



1.2. Use Save() to Save Multiple R Objects

```
# Saving on object in RData format
save(data1, file = "data.RData")
# Save multiple objects
save(data1, data2, file = "data.RData")
# To load the data again
load("data.RData")
```

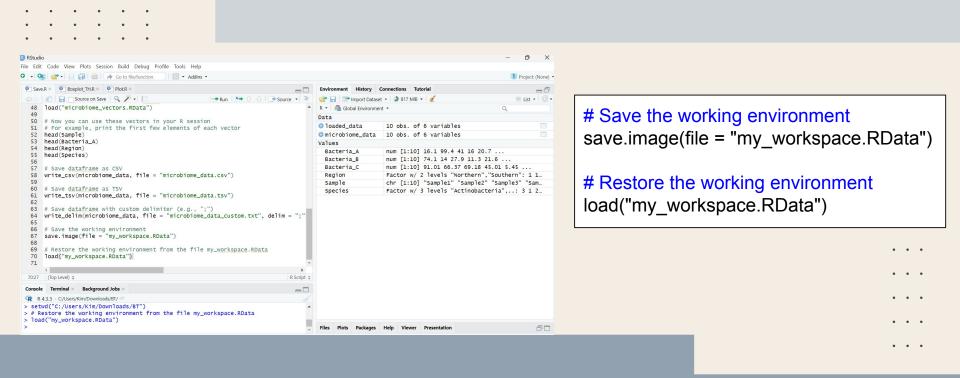
Example:

>	print(microbiome_data)							
	Sample	Bacteria_A	Bacteria_B	Bacteria_C	Region	Species		
1	Sample1	16.14	74.12	91.01	Northern	Firmicutes		
2	Sample2	99.43	14.00	66.37	Northern	Actinobacteria		
3	Sample3	40.97	27.95	69.18	Northern	Bacteroides		
4	Sample4	16.02	11.35	45.01	Northern	Bacteroides		
5	Sample5	20.66	21.61	5.45	Northern	Bacteroides		
6	Sample6	22.11	99.94	35.16	Southern	Bacteroides		
7	Sample7	61.11	63.28	30.94	Southern	Actinobacteria		
8	Sample8	60.13	15.23	48.72	Southern	Firmicutes		
9	Sample9	46.08	86.54	54.43	Southern	Actinobacteria		
10	Sample10	39.31	59.18	49.98	Southern	Firmicutes		

```
# Saving each column of microbiome data as individual vectors
Sample <- microbiome data$Sample
Bacteria A <- microbiome data$Bacteria A
Bacteria B <- microbiome data$Bacteria B
Bacteria C <- microbiome data$Bacteria C
Region <- microbiome data$Region
Species <- microbiome data$Species
# Checking the data types of each vector
class(Sample) # "character"
class(Bacteria A) # "numeric"
class(Bacteria B) # "numeric"
class(Bacteria C) # "numeric"
class(Region) # "factor"
class(Species) # "factor"
# Save to .RData file
save(Sample, Bacteria A, Bacteria B, Bacteria C, Region, Species, file =
"microbiome vectors.RData")
# Load the vectors from the .RData file
load("microbiome vectors.RData")
# For example, print the first few elements of each vector
head(Sample)
head(Bacteria A)
head(Region)
head(Species)
```



1.3. Use Save.Image() to Save Workspace Image

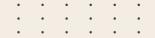




1.4. Use Download.File() to Download File from Website



2. Some Useful R Packages for Microbiome Data

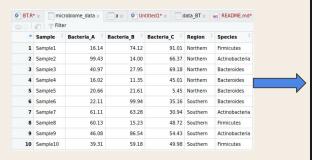


2.1. Readr

```
install.packages("readr")
library(readr)
# General function of writing data from R
write_delim(df, file, delim = " ")
# Write comma (",") separated value files
write csv(df, file)
# Write tab ("\t") separated value files
write tsv(df, file)
where df is a data frame to be written; file is used to specify the file
name to be saved; while delim is the delimiter that is used to separate
values. It must be single character.
```

2.1. Readr

Example



Save dataframe as CSV

write csv(microbiome data, file = "microbiome data.csv")

Save dataframe as TSV

write tsv(microbiome data, file = "microbiome data.tsv")

Save dataframe with custom delimiter (e.g., ";") write delim(microbiome data, file =

"microbiome data custom.txt", delim = ";")

microbiome data.csv

1 Sample, Bacteria A, Bacteria B, Bacteria C, Region, Species 2 Sample1, 16.14, 74.12, 91.01, Northern, Firmicutes 3 Sample2,99,43,14,66,37,Northern,Actinobacteria 4 Sample3, 40.97, 27.95, 69.18, Northern, Bacteroides 5 Sample4, 16.02, 11.35, 45.01, Northern, Bacteroides 6 Sample5, 20.66, 21.61, 5.45, Northern, Bacteroides 7 Sample6, 22, 11, 99, 94, 35, 16, Southern, Bacteroides 8 Sample7,61.11,63.28,30.94,Southern,Actinobacteria 9 Sample8, 60.13, 15.23, 48.72, Southern, Firmicutes 10 Sample9,46.08,86.54,54.43,Southern,Actinobacteria 11 Sample10,39.31,59.18,49.98, Southern, Firmicutes

			1	microbiom	e_data.tsv		×		
1	Sample	Bacter	ia_A	Bacter	ia_B	Bacteria	_C	Region	Species
2	Sample1	16.14	74.12	91.01	Norther	n	Firmicut	es	
3	Sample2	99.43	14	66.37	Norther	n	Actinoba	cteria	
4	Sample3	40.97	27.95	69.18	Norther	n	Bacteroi	des	
5	Sample4	16.02	11.35	45.01	Norther	n	Bacteroi	des	
6	Sample5	20.66	21.61	5.45	Norther	n	Bacteroi	des	
7	Sample6	22.11	99.94	35.16	Souther	n	Bacteroi	des	
8	Sample7	61.11	63.28	30.94	Souther	n	Actinoba	cteria	
9	Sample8	60.13	15.23	48.72	Souther	n	Firmicut	es	
10	Sample9	46.08	86.54	54.43	Souther	n	Actinoba	cteria	
11	Sample10	9	39.31	59.18	49.98	Southern	1	Firmicu	tes

1 Sample; Bacteria A; Bacteria B; Bacteria C; Region; Species 2 Sample1;16.14;74.12;91.01;Northern;Firmicutes 3 Sample2:99.43:14:66.37:Northern:Actinobacteria 4 Sample3;40.97;27.95;69.18;Northern;Bacteroides 5 Sample4:16.02:11.35:45.01:Northern:Bacteroides 6 Sample5; 20.66; 21.61; 5.45; Northern; Bacteroides 7 Sample6;22.11;99.94;35.16; Southern; Bacteroides 8 Sample7:61.11:63.28:30.94:Southern:Actinobacteria 9 Sample8; 60.13; 15.23; 48.72; Southern; Firmicutes 10 Sample9:46.08:86.54:54.43:Southern:Actinobacteria

11 Sample10:39.31:59.18:49.98:Southern:Firmicutes

*microbiome data custom.txt



2.2 Ggpubr

Type: Package

Title: 'ggplot2' Based Publication Ready Plots

- Using CRAN (Comprehensive R Archive Network): install.packages("ggpubr")
- Using GitHub: install.packages("devtools") devtools::install_github("kassambara/ggpubr")

2.2 Ggpubr

Download dataset:

https://datadryad.org/stash/dataset/doi:10.5061/dryad.x95x69pqf

install.packages("readxl")

library(readxl)

data_PT <- read_excel("C:/Users/Kim/Downloads/BT/plateassay_growthrates.xlsx")

View(data PT)

	₽ PF	iiter					Q	
^	index	replicate	species	temperature	photoperiod	bacteria [‡]	mumax [‡]	
1	1	1	T_rotula	4	24	xenic	0.08829063	
2	2	2	T_rotula	4	24	xenic	0.13004346	
3	3	3	T_rotula	4	24	xenic	0.11478548	
4	4	4	T_rotula	4	24	xenic	0.10251854	
5	5	5	T_rotula	4	24	xenic	0.12222029	
6	6	6	T_rotula	4	24	xenic	0.09568441	
7	7	7	T_rotula	4	24	xenic	0.11578493	
8	8	8	T_rotula	4	24	xenic	0.11919964	
9	9	9	T_rotula	4	24	xenic	0.08955854	
10	10	10	T_rotula	4	24	xenic	0.12102892	
11	11	11	T_rotula	4	24	xenic	0.08787527	
12	12	12	T_rotula	4	24	xenic	0.11457817	
13	13	13	T_rotula	4	24	xenic	0.12561646	
14	14	14	T_rotula	4	24	xenic	0.10937829	
15	15	15	T_rotula	4	24	xenic	0.10288063	
16	16	16	T_rotula	4	24	xenic	0.09750211	
17	17	17	T_rotula	4	24	xenic	0.10131356	
18	18	18	T_rotula	4	24	xenic	0.06079321	
19	19	19	T_rotula	4	24	xenic	0.08617507	
20	20	20	T_rotula	4	24	xenic	0.08294450	

index: A unique identifier for each observation in the dataset, typically used to distinguish and identify individual rows of data.

replicate: The biological replicate number of the respective experimental treatment.

species: Defines the species studied in each observation. In this case, species can be "T. rotula" or "T. gravida", denoting specific marine diatom species under investigation.

temperature: The applied temperature in the experimental condition.

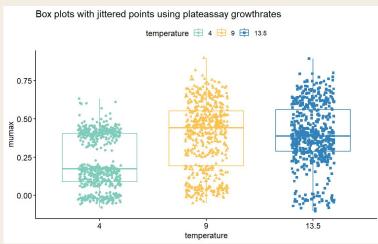
photoperiod: The duration of light applied in the experimental condition.

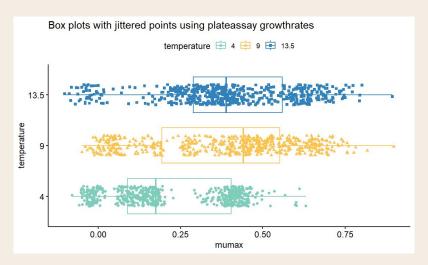
bacteria: Specifies if the native microbiome is present (xenic) or removed (axenic) for the respective observation in the experimental condition.

mumax: Maximum growth rate for each experimental unit.

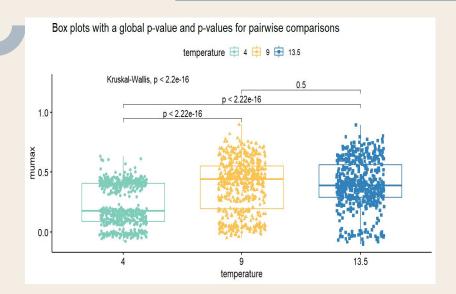
2.2. Ggpubr

2.2.1. Box Plots

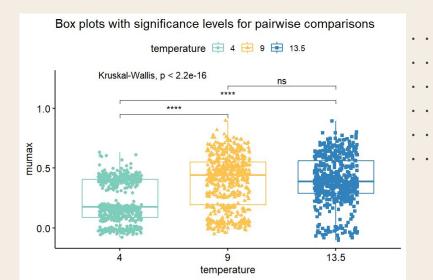




2.2.1. Box Plots

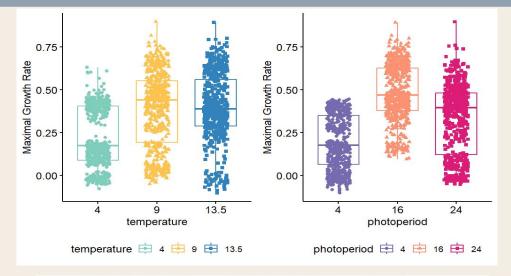


```
# Specify the pairwise group comparisons
comps <- list(c("4", "9"), c("4", "13.5"), c("9", "13.5"))
p_1 <- p_1 +
    stat_compare_means(comparisons = comps) +
    stat_compare_means(label.y = 1.25) +
    labs(title =
    "Box plots with a global p-value and p-values for pairwise comparisons")</pre>
```

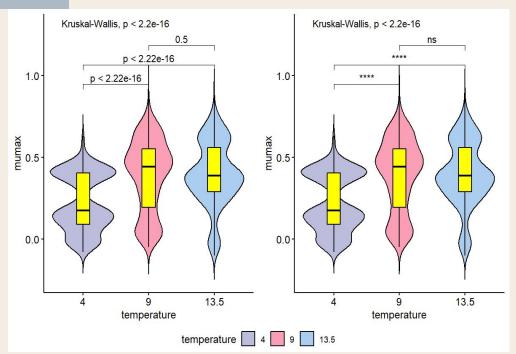


```
# Specify the comparisons of interest
comps <- list(c("4", "9"), c("4", "13.5"), c("9", "13.5"))
p_1 <- p_1 +
    stat_compare_means(comparisons = comps, label = "p.signif") +
    stat_compare_means(label.y = 1.25) +
    labs(title =
    "Box plots with significance levels for pairwise comparisons")</pre>
```

2.2.1. Box Plots



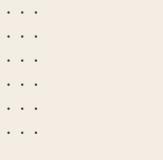
2.2.2 Violin Plots



```
library(ggpubr)
# Specify the pairwise group comparisons
comps <- list(c("4", "9"), c("4", "13.5"), c("9", "13.5"))
# Plot 1: Violin plot with boxplot inside, standard p-value labels
plot1 <- ggviolin(data_PT, x = "temperature", y = "mumax", fill = "temperature",</pre>
                  palette = c("#bcbddc", "#fa9fb5", "#abcdef"),
                  add = "boxplot", add.params = list(fill = "vellow")) +
  stat_compare_means(comparisons = comps) +
  stat_compare_means(label.y = 1.30)
# Plot 2: Violin plot with boxplot inside, customized p-value labels
plot2 <- ggviolin(data_PT, x = "temperature", y = "mumax", fill = "temperature".</pre>
                  palette = c("#bcbddc", "#fa9fb5", "#abcdef"),
                  add = "boxplot", add.params = list(fill = "vellow")) +
  stat_compare_means(comparisons = comps, label = "p.signif") +
  stat_compare_means(label.y = 1.30)
# Combine the plots into a single frame
ggarrange(plot1, plot2, ncol = 2, nrow = 1,
          common.legend = TRUE, legend = "bottom")
```

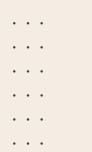


2.2.3. Density Plots



```
temperature 4 4 9 113.5
density
                                  0.25
                                                                          0.75
              0.00
                                                      0.50
                                             mumax
```

2.2.4. Histogram Plots



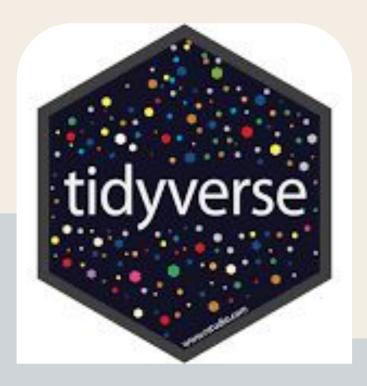
```
temperature 4 9 13.5
  120-
   80
count
   40
                                  0.25
                 0.00
                                                   0.50
                                           mumax
```





R packages for data science
The tidyverse is an opinionated collection
of R packages designed for data science.
All packages share an underlying design
philosophy, grammar, and data structures.

Install the complete tidyverse with: install.packages("tidyverse")



2.3. Tidyverse





- ggplot2, for data visualisation.
- **dplyr** for data manipulation.
- **tidyr**, for data tidying.
- readr, for data import.

```
> library(tidyverse)
                                                       tidyverse 2.0.0 —

    Attaching core tidyverse packages

√ dplyr
           1.1.4
                    √ readr
                               2.1.5

√ forcats 1.0.0 √ stringr
                               1.5.1

√ ggplot2 3.5.0 
√ tibble

                               3.2.1

√ lubridate 1.9.3

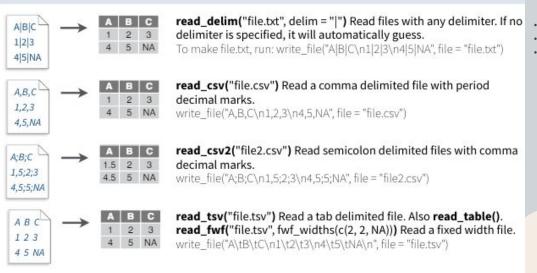
√ tidyr

                               1.3.1
√ purrr
        1.0.2
```



Read Tabular Data with readr

read_*(file, col_names = TRUE, col_types = NULL, col_select = NULL, id = NULL, locale, n_max = Inf,
 skip = 0, na = c("", "NA"), guess_max = min(1000, n_max), show_col_types = TRUE) See ?read_delim

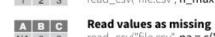


USEFUL READ ARGUMENTS

Α	В	C	No header	1	2	
1	2	3	read_csv("file.csv", col_names = FALSE)	4	5	N
4	5	NA				
	-			A	В	0
X	У	Z	Provide header	1	2	3
A	В	C	read_csv("file.csv",			
1	2	3	col_names = c("x", "y", "z"))	Δ	В	i
4	5	NA		NA	2	3
			Read multiple files into a single table	4	5	N
	>		read_csv(c("f1.csv", "f2.csv", "f3.csv"), id = "origin_file")	A;I	B;C	

Read a subset of lines read csv("file.csv", n max = 1)

read_csv("file.csv", skip = 1)



Skip lines

NA 2 3 read_csv("file.csv", na = c("1"))

Specify decimal marks read_delim("file2.csv", locale = locale(decimal_mark = ","))

tidyr

Data tidying with tidyr:: cheat sheet

Tidy data is a way to organize tabular data in a consistent data structure across packages. A table is tidy if:







its own column

Each observation, or case, is in its own row





Access variables as vectors

Preserve cases in vectorized operations

 $A \cdot B \rightarrow C$

Tibbles

AN ENHANCED DATA FRAME

Tibbles are a table format provided by the tibble package. They inherit the data frame class, but have improved behaviors:

- Subset a new tibble with 1, a vector with [[and \$.
- No partial matching when subsetting columns.
- Display concise views of the data on one screen. options(tibble.print max = n, tibble.print min = m, tibble.width = Inf) Control default display settings.

View() or glimpse() View the entire data set.

CONSTRUCT A TIBBLE

tibble(...) Construct by columns. tibble(x = 1:3, y = c("a", "b", "c"))

tribble(...) Construct by rows. tribble(~x, ~y,



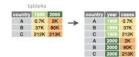


Both make

as_tibble(x, ...) Convert a data frame to a tibble. enframe(x, name = "name", value = "value") Convert a named vector to a tibble. Also deframe(). is_tibble(x) Test whether x is a tibble.

Studio

Reshape Data - Pivot data to reorganize values into a new layout.



pivot_longer(data, cols, names_to = "name", values_to = "value", values_drop_na = FALSE)

"Lengthen" data by collapsing several columns into two. Column names move to a new names to column and values to a new values to column.

pivot longer(table4a, cols = 2:3, names to ="vear", values to = "cases")



pivot wider(data, names from = "name". values from = "value")

The inverse of pivot_longer(). "Widen" data by expanding two columns into several. One column provides the new column names, the other the

pivot wider(table2, names from = type, values from = count)

Expand Tables

Create new combinations of variables or identify implicit missing values (combinations of variables not present in the data).



expand(data, ...) Create a new tibble with all possible combinations of the values of the variables listed in ... Drop other variables. expand(mtcars, cyl, gear,



A 1 3 A 1 3 Isst()) Add missing possible B 1 4 combinations of values of variables listed in ... Fill remaining variables with NA. complete(mtcars, cyl, gear,

- Use these functions to split or combine cells into individual, isolated values.



A 1999 0.7K

B 1999 37K

country year rate

В 1999

A 1999 0.7K

B 2000 80K

B 2000 1741

1999 19M

2000 2K

1999 37K

2000 2K

C 2000 pop 1T

table3

A 1999 0.7K/

B 1999 37K/

B 2000 80K/1/

untry year rate

A 1999 0.7K/

A 2000 2K/

B 1999 37K/

B 2000 80K/

A 2000 2K/

unite(data, col, ..., sep = "_", remove = TRUE, na.rm = FALSE) Collapse cells across several columns into a single column.

unite(table5, century, year, col = "year", sep = "")

separate(data, col, into, sep = "[^[:alnum:]]+", remove = TRUE, convert = FALSE, extra = "warn" fill = "warn", ...) Separate each cell in a column into several columns, Also extract().

separate(table3, rate, sep = "/". into = c("cases", "pop"))

separate_rows(data, ..., sep = "[^[:alnum:].]+", convert = FALSE) Separate each cell in a column into several rows.

separate_rows(table3, rate, sep = "/")

Handle Missing Values

Drop or replace explicit missing values (NA).



drop_na(data, ...) Drop rows containing NA's in ... columns.

drop_na(x, x2)



fill(data, ..., .direction = "down") Fill in NA's in ... columns using the next or previous value. fill(x, x2)



replace_na(data, replace) Specify a value to replace NA in selected columns.

replace na(x, list(x2 = 2))

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dplyr

Data transformation with dplyr:: **CHEAT SHEET**

dplyr functions work with pipes and expect tidy data. In tidy data:







its own column

Each variable is in Each observation, or case, is in its own row

becomes f(x, y)

Summarise Cases

Apply summary functions to columns to create a new table of summary statistics. Summary functions take vectors as input and return one value (see back).

summary function



summarise(.data, ...) Compute table of summaries. summarise(mtcars, avg = mean(mpg))



count(.data, ..., wt = NULL, sort = FALSE, name = NULL) Count number of rows in each group defined by the variables in ... Also tally().

count(mtcars, cvl)

Group Cases

Use group_by(.data, ..., .add = FALSE, .drop = TRUE) to create a "grouped" copy of a table grouped by columns in ... dplyr functions will manipulate each "group" separately and combine the results.



mtcars %>% group_by(cyl) 96>96 summarise(avg = mean(mpg))

Use rowwise(.data, ...) to group data into individual rows. dplyr functions will compute results for each row. Also apply functions to list-columns. See tidyr cheat sheet for list-column workflow.



starwars %>% rousuico/) 06×06

Manipulate Cases

EXTRACT CASES

Row functions return a subset of rows as a new table.



filter(.data, ..., .preserve = FALSE) Extract rows that meet logical criteria. filter(mtcars, mpg > 20)



distinct(.data, ..., .keep_all = FALSE) Remove rows with duplicate values. distinct(mtcars, gear)

slice(.data, ..., .preserve = FALSE) Select rows by position.





slice_sample(.data, ..., n, prop, weight_by = NULL, replace = FALSE) Randomly select rows. Use n to select a number of rows and prop to select a fraction of rows. slice_sample(mtcars, n = 5, replace = TRUE)

slice_min(.data, order_by, ..., n, prop, with_ties = TRUE) and slice_max() Select rows

with the lowest and highest values. slice_min(mtcars, mpg, prop = 0.25)

slice_head(.data, ..., n, prop) and slice_tail() Select the first or last rows. slice head(mtcars, n = 5)

Logical and boolean operators to use with filter()

=	<	<=	is.na()	%in%	1	xor()
!=	>	>=	!is.na()	!	&	

See ?base::Logic and ?Comparison for help.

ARRANGE CASES



arrange(.data, ..., .by_group = FALSE) Order rows by values of a column or columns (low to high), use with desc() to order from high to low. arrange(mtcars, mpg) arrange(mtcars, desc(mpg))

Manipulate Variables

EXTRACT VARIABLES

Column functions return a set of columns as a new vector or table.



pull(.data, var = -1, name = NULL, ...) Extract column values as a vector, by name or index. pull(mtcars, wt)



select(.data, ...) Extract columns as a table. select(mtcars, mpg, wt)



relocate(.data, ..., .before = NULL, .after = NULL) Move columns to new position. relocate(mtcars, mpg, cyl, .after = last_col())

Use these helpers with select() and across()

e.g. select(mtcars, mpg:cvl)

contains(match) starts with(match) matches(match)

num_range(prefix, range) :, e.g. mpg:cyl ends with(match) all of(x)/any of(x, ..., vars) -, e.g., -gear

MANIPULATE MULTIPLE VARIABLES AT ONCE



across(.cols, .funs, ..., .names = NULL) Summarise or mutate multiple columns in the same way. summarise(mtcars, across(everything(), mean))



c_across(.cols) Compute across columns in row-wise data.

transmute(rowwise(UKgas), total = sum(c_across(1:2)))

MAKE NEW VARIABLES

Apply vectorized functions to columns. Vectorized functions take vectors as input and return vectors of the same length as output (see back). vectorized function



mutate(.data, ..., .keep = "all", .before = NULL, .after = NULL) Compute new column(s). Also add_column(), add_count(), and add_tally(). mutate(mtcars, gpm = 1 / mpg)

ggplot2

Data visualization with ggplot2:: CHEAT SHEET

Basics

ggplot2 is based on the grammar of graphics, the idea that you can build every graph from the same components: a data set, a coordinate system, and geoms-visual marks that represent data points.



To display values, map variables in the data to visual properties of the geom (aesthetics) like size, color, and x and y locations.



Complete the template below to build a graph.



ggplot(data = mpg, aes(x = cty, y = hwy)) Begins a plot that you finish by adding layers to. Add one geom function per layer.

last plot() Returns the last plot.

ggsave("plot.png", width = 5, height = 5) Saves last plot as 5' x 5' file named "plot.png" in working directory. Matches file type to file extension.

Aes Common aesthetic values. color and fill - string ("red", "#RRGGBB")

linetype - integer or string (0 = "blank", 1 = "solid", 2 = "dashed", 3 = "dotted", 4 = "dotdash", 5 = "longdash", 6 = "twodash")

lineend - string ("round", "butt", or "square") linejoin - string ("round", "mitre", or "bevel") size - integer (line width in mm) 0 1 2 1 4 5 6 7 8 9 10 11 12

shape - integer/shape name or BH 51617 H 920 21 22 23 23 25 a single character ("a") ⊠⊠□□△◊○○□□◆△▽

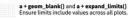


Geoms

Use a geom function to represent data points, use the geom's aesthetic properties to represent variables, Each function returns a layer.

GRAPHICAL PRIMITIVES

a <- ggplot(economics, aes(date, unemploy)) b <- ggplot(seals, aes(x = long, y = lat))



b + geom_curve(aes(yend = lat + 1, xend = long + 1), curvature = 1) - x, xend, y, yend, alpha, angle, color, curvature, linetype, size

a + geom_path(lineend = "butt", linejoin = "round", linemitre = 1) x, y, alpha, color, group, linetype, size

> a + geom_polygon(aes(alpha = 50)) - x, y, alpha, color, fill, group, subgroup, linetype, size b + geom rect(aes(xmin = long, ymin = lat, xmax = long + 1, ymax = lat + 1)) - xmax, xmin.

ymax, ymin, alpha, color, fill, linetype, size a + geom ribbon(aes(vmin = unemploy - 900, vmax = unemplov + 900)) - x, vmax, vmin, alpha, color, fill, group, linetype, size

LINE SEGMENTS

common aesthetics: x, y, alpha, color, linetype, size

b + geom_abline(aes(intercept = 0, slope = 1)) b + geom_hline(aes(yintercept = lat)) b + geom_vline(aes(xintercept = long))

b + geom_segment(aes(yend = lat + 1, xend = long + 1)) b + geom spoke(aes(angle = 1:1155, radius = 1))

ONE VARIABLE continuous

c <- ggplot(mpg, aes(hwy)); c2 <- ggplot(mpg)

c + geom_area(stat = "bin") x, y, alpha, color, fill, linetype, size

c + geom_densitv(kernel = "gaussian") x, y, alpha, color, fill, group, linetype, size, weight

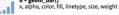
c + geom_dotplot() x, y, alpha, color, fill

> c + geom freqpoly() x, y, alpha, color, group, linetype, size

c + geom_histogram(binwidth = 5) x, y, alpha, color, fill, linetype, size, weight

c2 + geom_qq(aes(sample = hwy)) x, y, alpha, color, fill, linetype, size, weight

d <- ggplot(mpg, aes(fl))



TWO VARIABLES

both continuous e <- ggplot(mpg, aes(cty, hwy))

e + geom_label(aes(label = cty), nudge_x = 1, nudge_y = 1) - x, y, label, alpha, angle, color,

family, fontface, hjust, lineheight, size, vjust e + geom point()

x, y, alpha, color, fill, shape, size, stroke e + geom quantile()

x, y, alpha, color, group, linetype, size, weight

e + geom_rug(sides = "bl") x, y, alpha, color, linetype, size e + geom_smooth(method = lm)

x, y, alpha, color, fill, group, linetype, size, weight

e + geom_text(aes(label = cty), nudge_x = 1, nudge_y = 1) - x, y, label, alpha, angle, color, family, fontface, hjust, lineheight, size, vjust

one discrete, one continuous f <- ggplot(mpg, aes(class, hwy))

f + geom_col() x, y, alpha, color, fill, group, linetype, size

> x, y, lower, middle, upper, ymax, ymin, alpha, color, fill, group, linetype, shape, size, weight

f + geom_dotplot(binaxis = "y", stackdir = "center"; x, y, alpha, color, fill, group

f + geom_violin(scale = "area") x, y, alpha, color, fill, group, linetype, size, weight

both discrete

g <- ggplot(diamonds, aes(cut, color))

g + geom_count() x, y, alpha, color, fill, shape, size, stroke

> e + geom_jitter(height = 2, width = 2) x, y, alpha, color, fill, shape, size

continuous bivariate distribution h <- ggplot(diamonds, aes(carat, price))

h + geom bin2d(binwidth = c(0.25, 500))x, y, alpha, color, fill, linetype, size, weight

ggplot2



h + geom_density_2d() x, v, alpha, color, group, linetype, size



h + geom_hex() x, y, alpha, color, fill, size



continuous function i <- ggplot(economics, aes(date, unemploy))

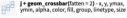


x, y, alpha, color, group, linetype, size

i + geom_step(direction = "hv") x, y, alpha, color, group, linetype, size

visualizing error

df <- data.frame(grp = c("A", "B"), fit = 4:5, se = 1:2) j <- ggplot(df, aes(grp, fit, ymin = fit - se, ymax = fit + se))



j + geom_errorbar() - x, ymax, ymin, alpha, color, group, linetype, size, width

Also geom_errorbarh(). j + geom_linerange()

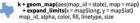
x, ymin, ymax, alpha, color, group, linetype, size

j + geom_pointrange() - x, y, ymin, ymax, alpha, color, fill, group, linetype, shape, size

mans

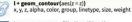
data <- data.frame(murder = USArrests\$Murder, state = tolower(rownames(USArrests))) map <- map data("state")

k <- ggplot(data, aes(fill = murder))



THREE VARIABLES

sealsSz <- with(seals, sgrt(delta_long^2 + delta_lat^2)); I <- ggplot(seals, aes(long, lat))



I + geom contour filled(aes(fill = z)) x, y, alpha, color, fill, group, linetype, size, subgroup



x, y, alpha, fill l + geom_tile(aes(fill = z))

x, v, alpha, color, fill, linetype, size, width



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thank Lyon

ANNALEIGH + BENJAMIN | 06.10.20XX