

# MGMA2024

Lecture 11. Serotyping, genotyping and phenotyping microbes

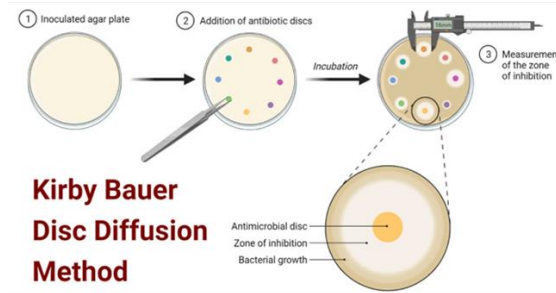
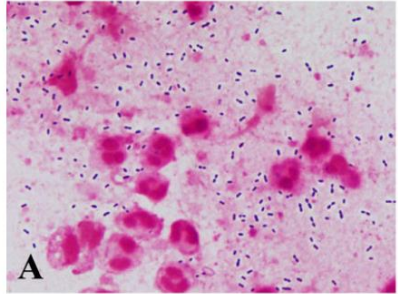
## Section 2. Genotype and molecular subtyping methods

# Content

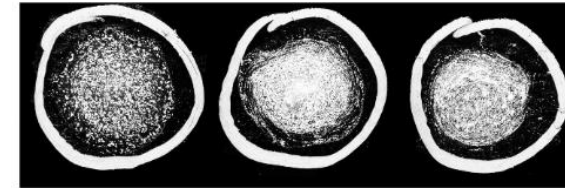
1. Genotype and subtyping methods
2. Molecular subtyping methods
  - Band–base methods
  - Sequence-Based methods
3. Summary of methods

# Subtyping methods

*S. pneumoniae*



A. Negative with *E. coli* K12



B. Positive with *E. coli* O26

FIGURE 1. Typical latex agglutination results with latex anti-O26 STEC.



Results (+/-)	-	-	+	-	+	-	-	+	+	-	+	-	+	+	-	+	-	+	-	+
Scores for results	1	2	4	1	2	4	1	2	4	1	2	4	1	2	4	1	2	4	1	2
	DNPG	ADH	LDC	ODC	CIT	H2S	URE	TDA	IND	VP	IGL	GLU	MAN	INO	SOR	RHA	SAC	MEL	AMY	ARA
Total value of + Samples score	4					2			6		4		5			6			2	

*Escherichia coli* 613

Culture-**positive** samples



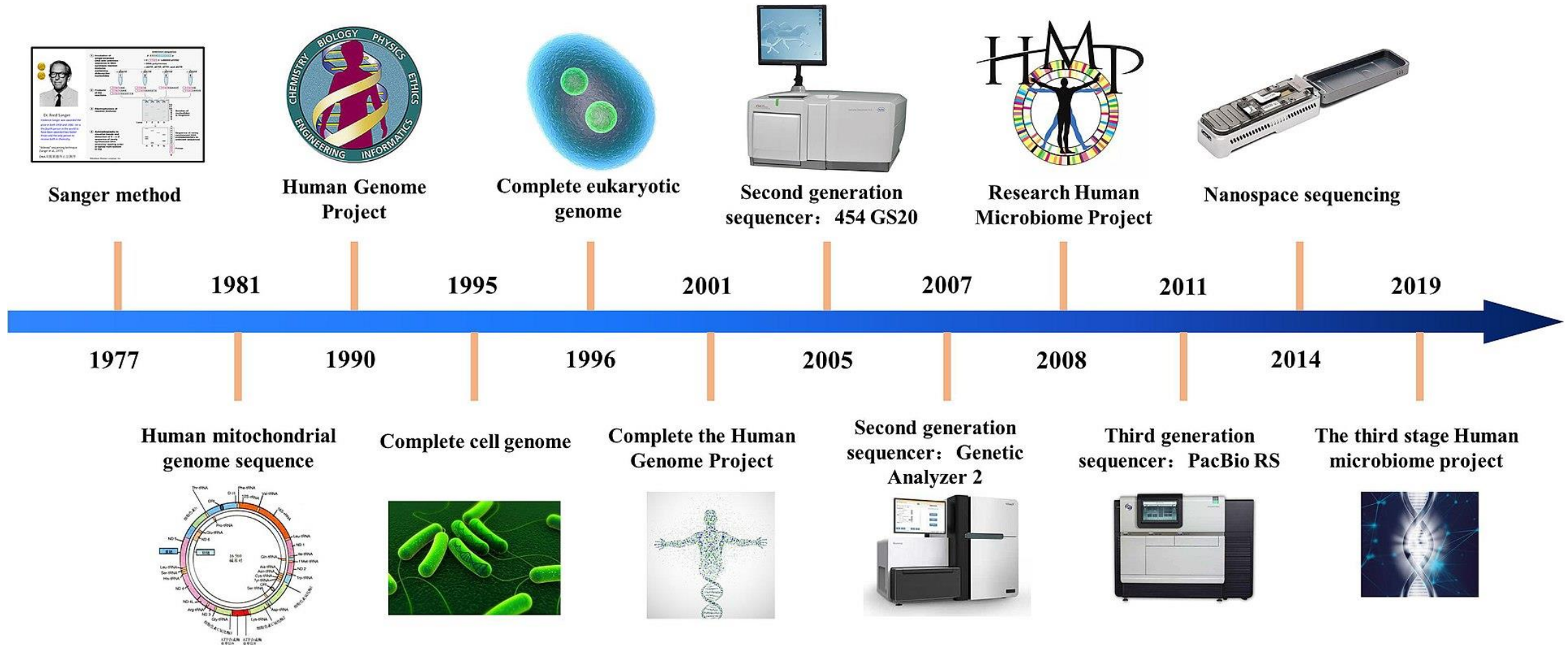
Phenotyping and Serotyping

Culture-**negative** samples

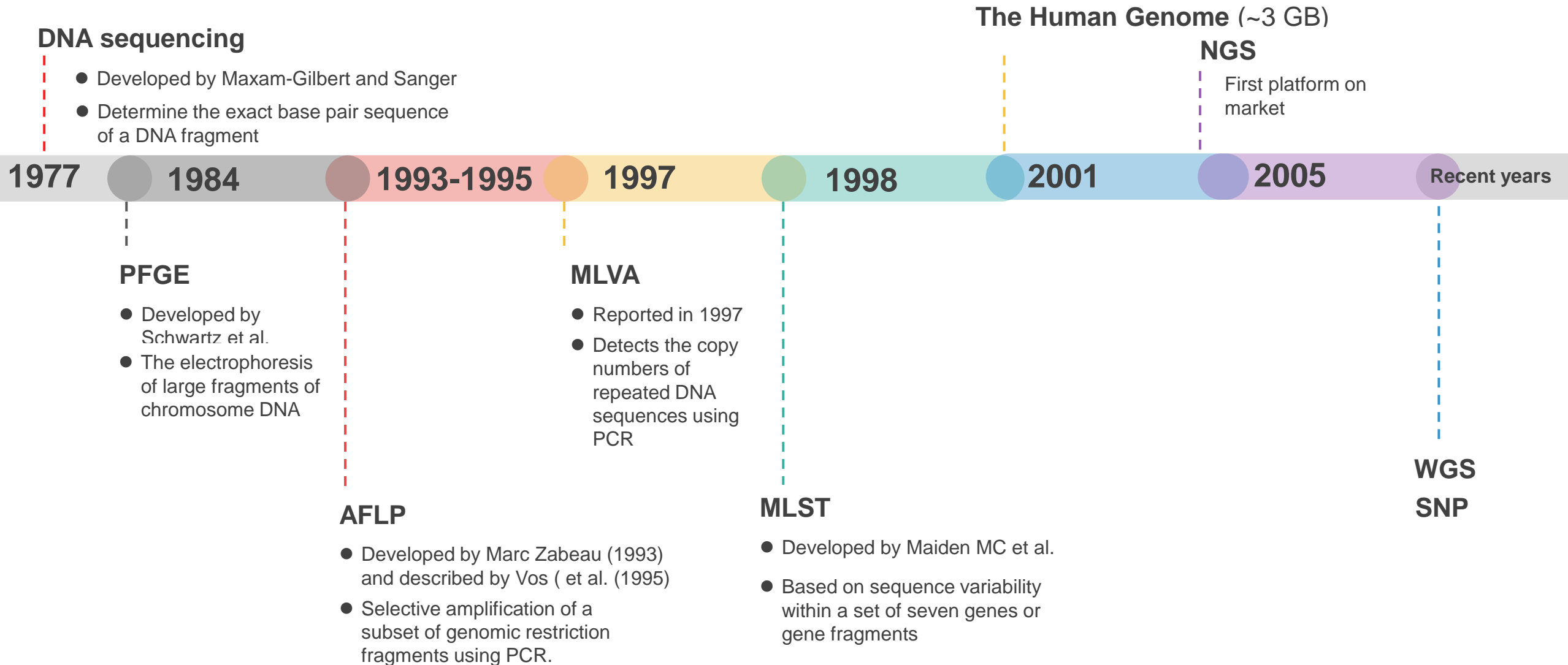


Methods?

# A brief history of Next Generation Sequencing (NGS)



# Timeline of the development of molecular subtyping methods



# Molecular subtyping methods

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graph TD; A[Molecular subtyping methods] --> B[Band –base methods]; A --> C[Sequence-Based methods];
```

## Band –base methods

(Based on comparing variation in DNA banding pattern on gels)

### ❖ Restriction digestion-based method

- ❑ Pulsed Field Gel Electrophoresis (PFGE)

### ❖ Amplification-based method

- ❑ Amplified Fragment Length Polymorphisms (AFLP)
- ❑ Multiple- Locus Variable number tandem repeat Analysis (MLVA)

## Sequence-Based methods

(Based on polymorphism of DNA sequences)

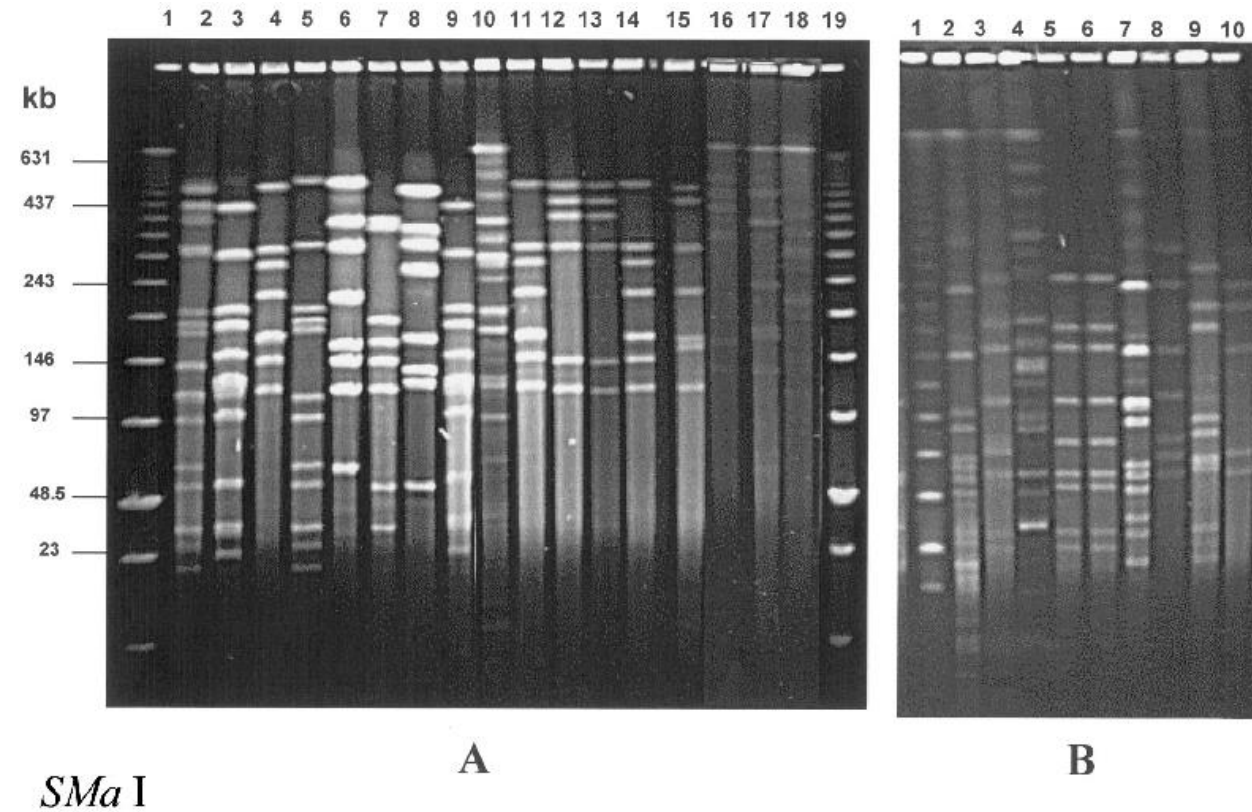
### ❖ Multi Locus Sequence Typing (MLST)

- ❑ Traditional MLST
- ❑ cg MLST
- ❑ wg MLST

### ❖ Whole Genome Sequence (WGS)

### ❖ Single-Nucleotide Polymorphism (SNP)

# BAND – BASE METHODS

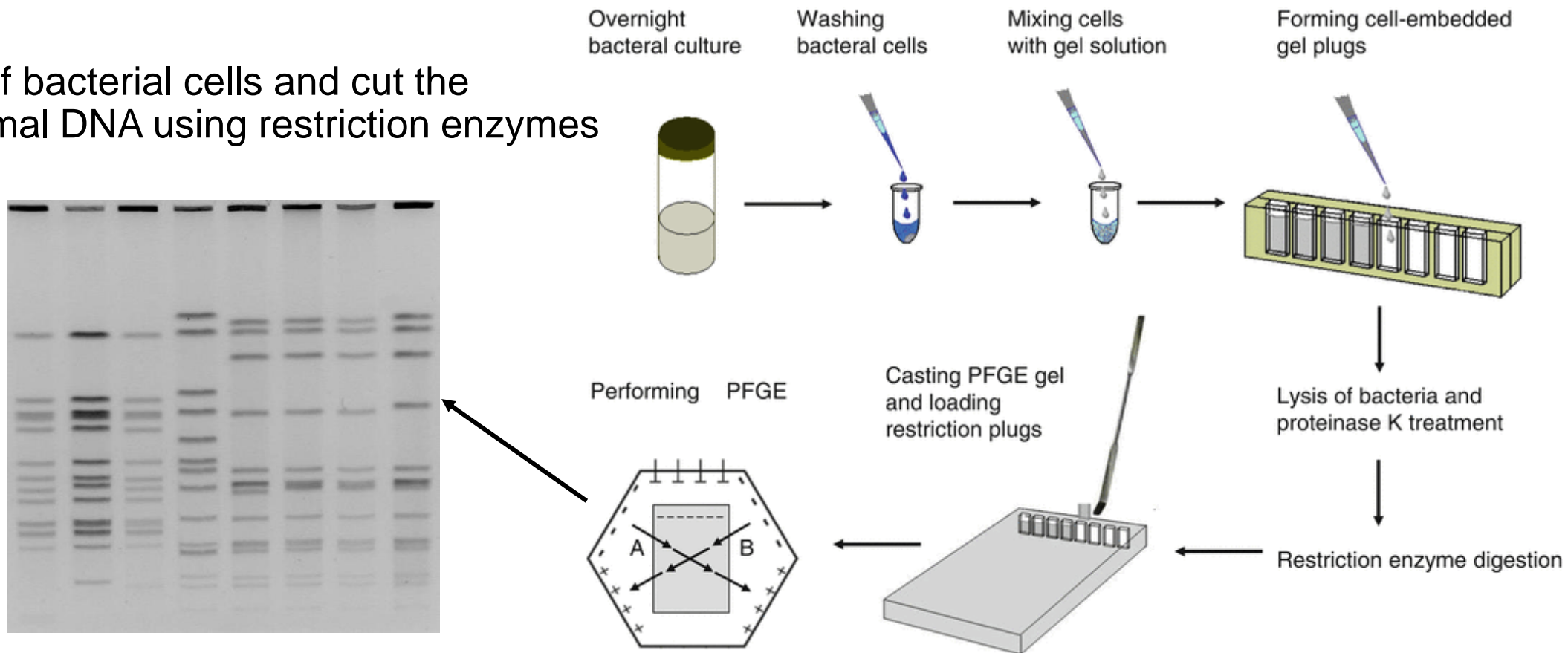




# Band –base methods

## ❖ Pulse Field Gel Electrophoresis (PFGE)

- The electrophoresis of large fragments (10 Mb) of chromosome DNA
- The direction of the electric field is constantly changed
- The lysis of bacterial cells and cut the chromosomal DNA using restriction enzymes





# Band –base methods

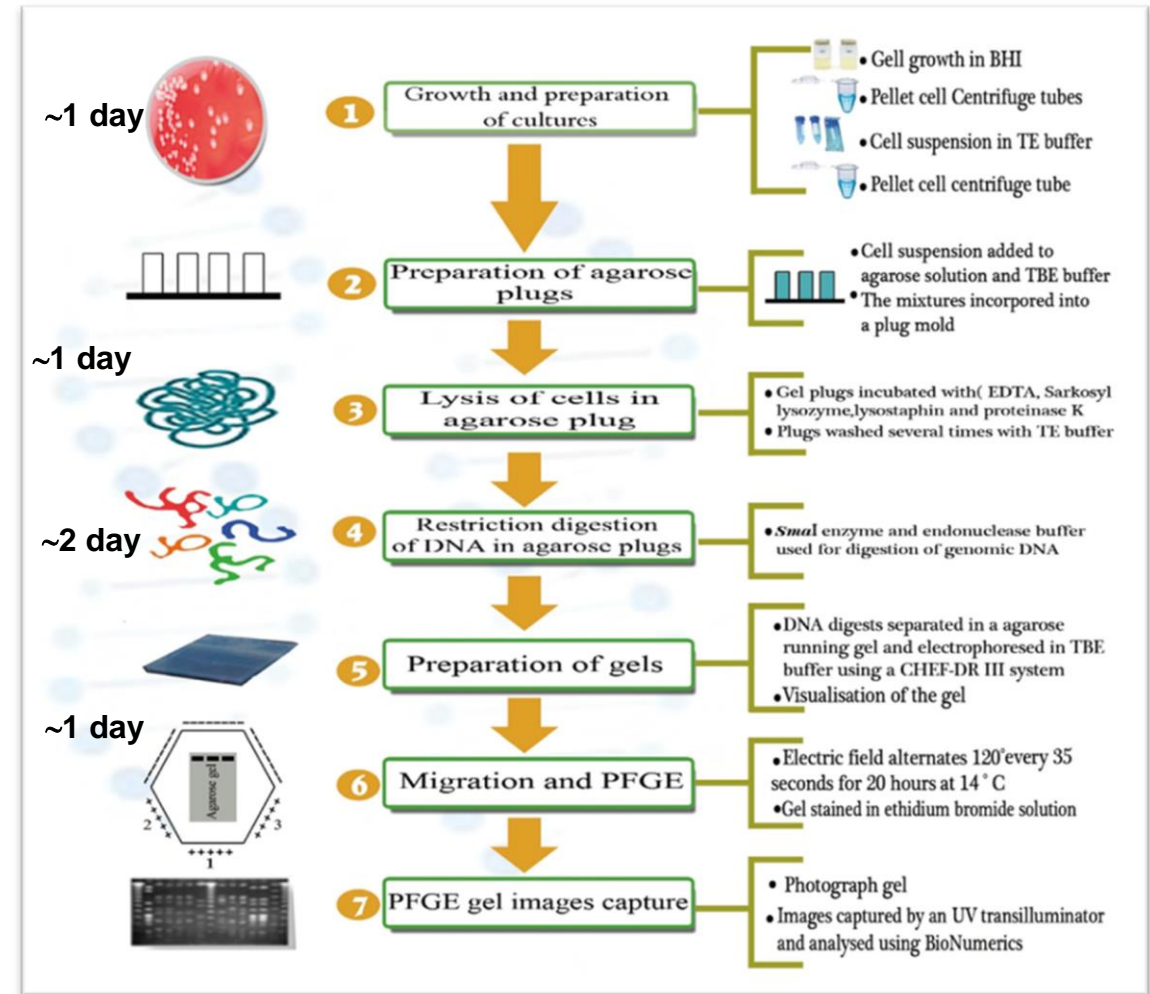
## ❖ Pulse Field Gel Electrophoresis (PFGE)

### □Application

- Standard genetic typing method
- Epidemiological studies (e.g. *S. aureus*, *N. meningitidis*, *V. cholerae*, *M. tuberculosis*)

### □Requirement

- Instruments (CHEF/FIGE...)
- The lysing methods
- Selection restriction enzymes
- Protocol for electrophoresis



<https://doi.org/10.7860%2FJCDR%2F2016%2F15718.7043>

<https://doi.org/10.3390/pathogens12070966>

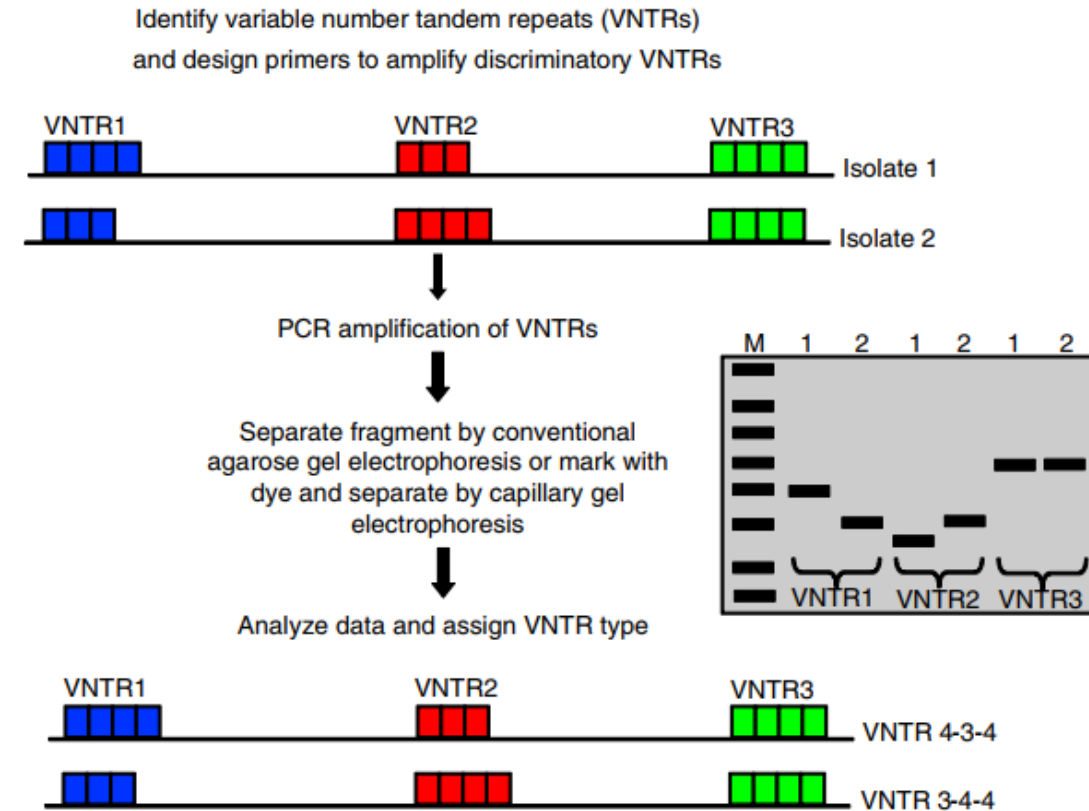
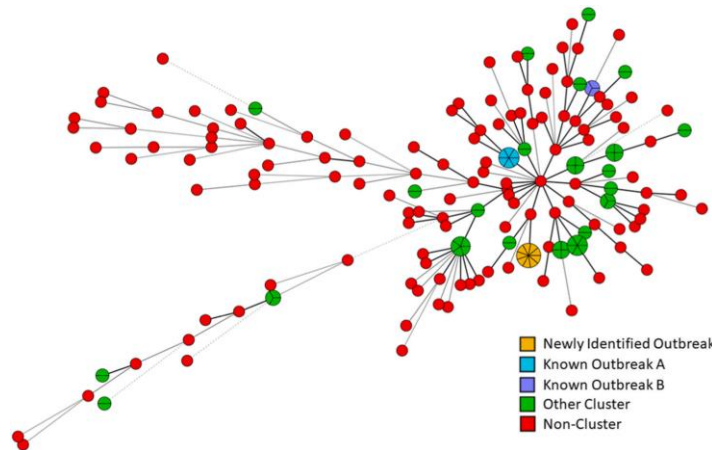
6/25/2024

Figure 1. PFGE process for the subtyping of *S. aureus* strains\*\*

# Band –base methods

## ❖ Multiple-Locus Variable number tandem repeat Analysis (MLVA)

- ❑ From data genomic sequencing
- ❑ Individual strains often carry the same elements with different copy numbers
- ❑ Display all MLVA profiles, MLVA clusters and Minimum spanning tree



**Figure 3** Multiple-locus variable-number tandem repeat analysis (MLVA).

# Band –base methods

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## ❖ Multiple-Locus Variable number tandem repeat Analysis (MLVA)

### ❑ Application

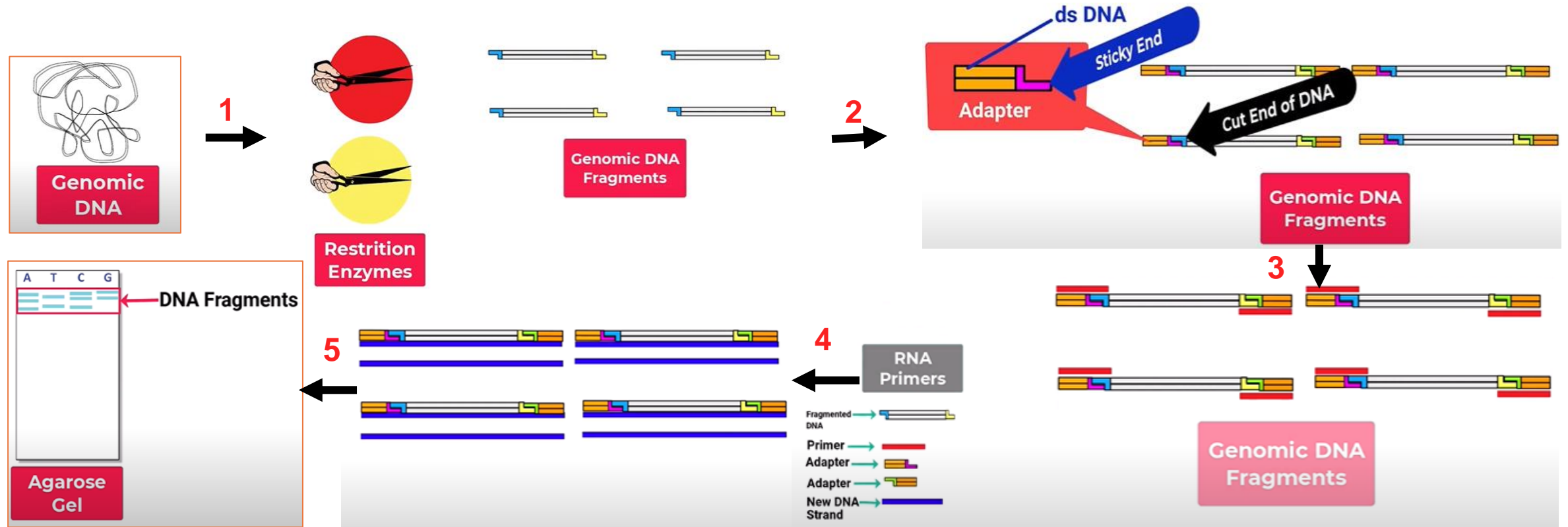
- The most recent analysis of multiple prokaryotic  
*e.g. E. coli O157, Mycoplasma pneumoniae\*, Cryptosporidium parvum*
- The epidemiological surveillance, inferring linkage and the investigation of outbreaks

### ❑ Requirement

- Data from genomic sequencing projects
- Design multiplex PCR primers (loci containing tandem repeats)
- Assessed on agarose gel/capillary electrophoresis
- BioNumerics software

# Band –base methods

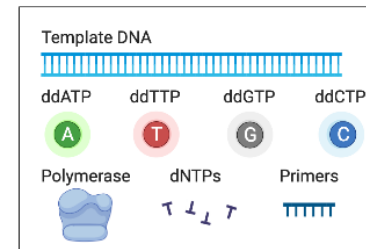
## ❖ Amplified Fragment Length Polymorphism (AFLP)



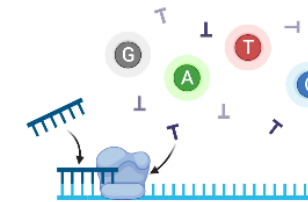
<https://www.youtube.com/watch?app=desktop&v=kuRuY25z9TY>

# SEQUENCE-BASED METHODS

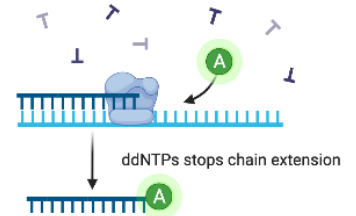
## Reagents



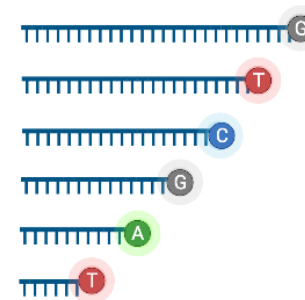
## ① Primer annealing and chain extension



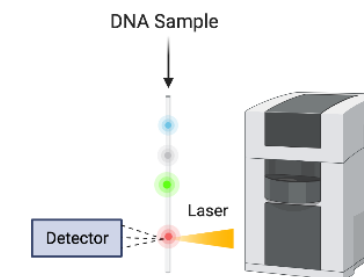
## ② ddNTP binding and chain termination



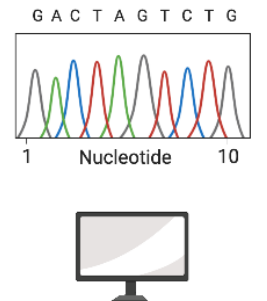
## ③ Fluorescently labelled DNA sample



## ④ Capillary gel electrophoresis and fluorescence detection



## ⑤ Sequence analysis and reconstruction

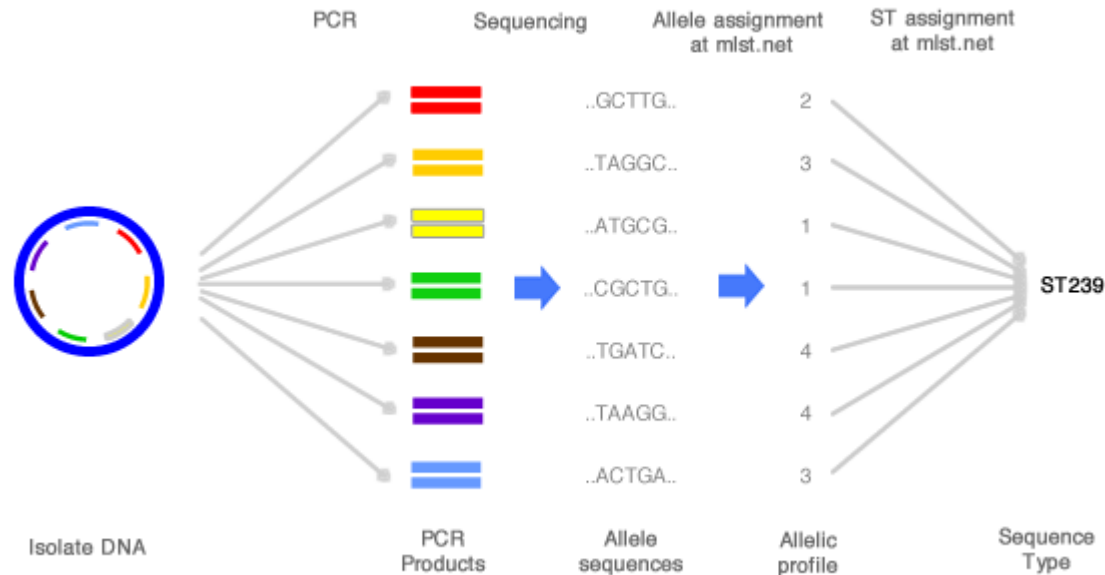


# Sequence-based methods

## ❖ Multi-Locus Sequence Typing (MLST)

### Traditional MLST

- Using fragments of 7 house-keeping genes (loci)
- 450-500 bp each gene
- Define the allelic profile or sequence types
- Comparisons result with a centralized database (PubMLST; [pubmlst.org](http://pubmlst.org))



### Current MLST Workflow

Isolate bacterial DNA samples from specimens

PCR amplification of housekeeping genes using MLST primers

Sequencing with PCR primers and Data Generation

Inspect sequence data manually, including alignment, reference trimming, and exporting sequences

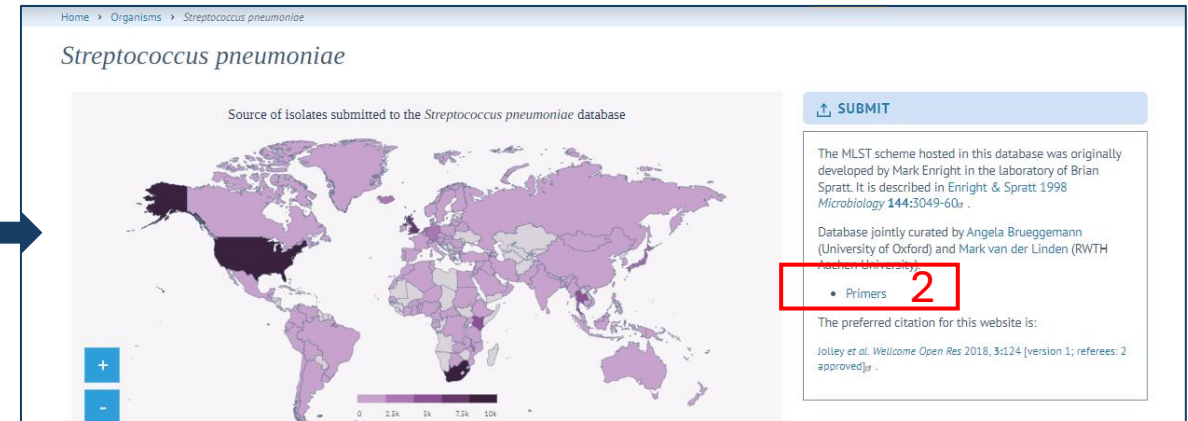
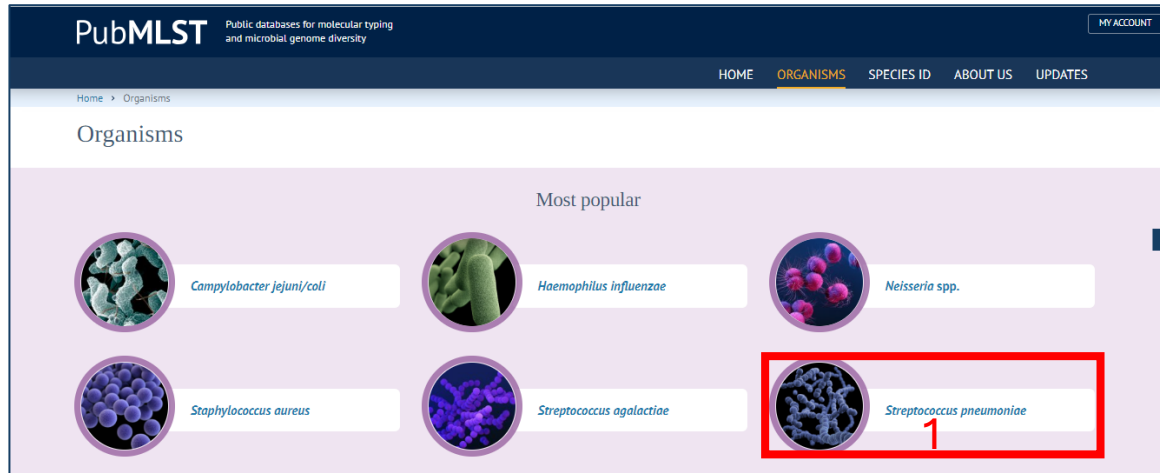
Match sequences to allelic library in the MLST website to determine the allelic profile

Use the MLST website to identify sequence type from the allelic profile.

**Analysis time:**  
4-5 hours per sample



# Sequence-based methods



## Primers and PCR conditions for MLST of *S. pneumoniae*

The pneumococcal MLST scheme uses internal fragments of the following seven house-keeping genes:

- aroE (shikimate dehydrogenase)
- gdh (glucose-6-phosphate dehydrogenase)
- gki (glucose kinase)
- recP (transketolase)
- spi (signal peptidase I)
- xpt (xanthine phosphoribosyltransferase)
- ddl (D-alanine-D-alanine ligase)

The primer pairs used for the PCR amplification of internal fragments of these genes are:

aroE-up, 5'-GCC TTT GAG GCG ACA GC  
aroE-dn, 5'-TGC AGT TCA (G/A)AA ACA T(A/T)T TCT AA

gdh-up, 5'-ATG GAC AAA CCA GC(G/A/T/C) AG(C/T) TT  
gdh-dn, 5'-GCT TGA GGT CCC AT(G/A) CT(G/A/T/C) CC

gki-up, 5'-GGC ATT GGA ATG GGA TCA CC  
gki-dn, 5'-TCT CCC GCA GCT GAC AC

recP-up, 5'-GCC AAC TCA GGT CAT CCA GG  
recP-dn, 5'-TGC AAC CGT AGC ATT GTA AC

spi-up, 5'-TTA TTC CTC CTG ATT CTG TC  
spi-dn, 5'-GTG ATT GGC CAG AAG CGG AA

xpt-up, 5'-TTA TTA GAA GAG CGC ATC CT  
xpt-dn, 5'-AGA TCT GCC TCC TTA AAT AC.

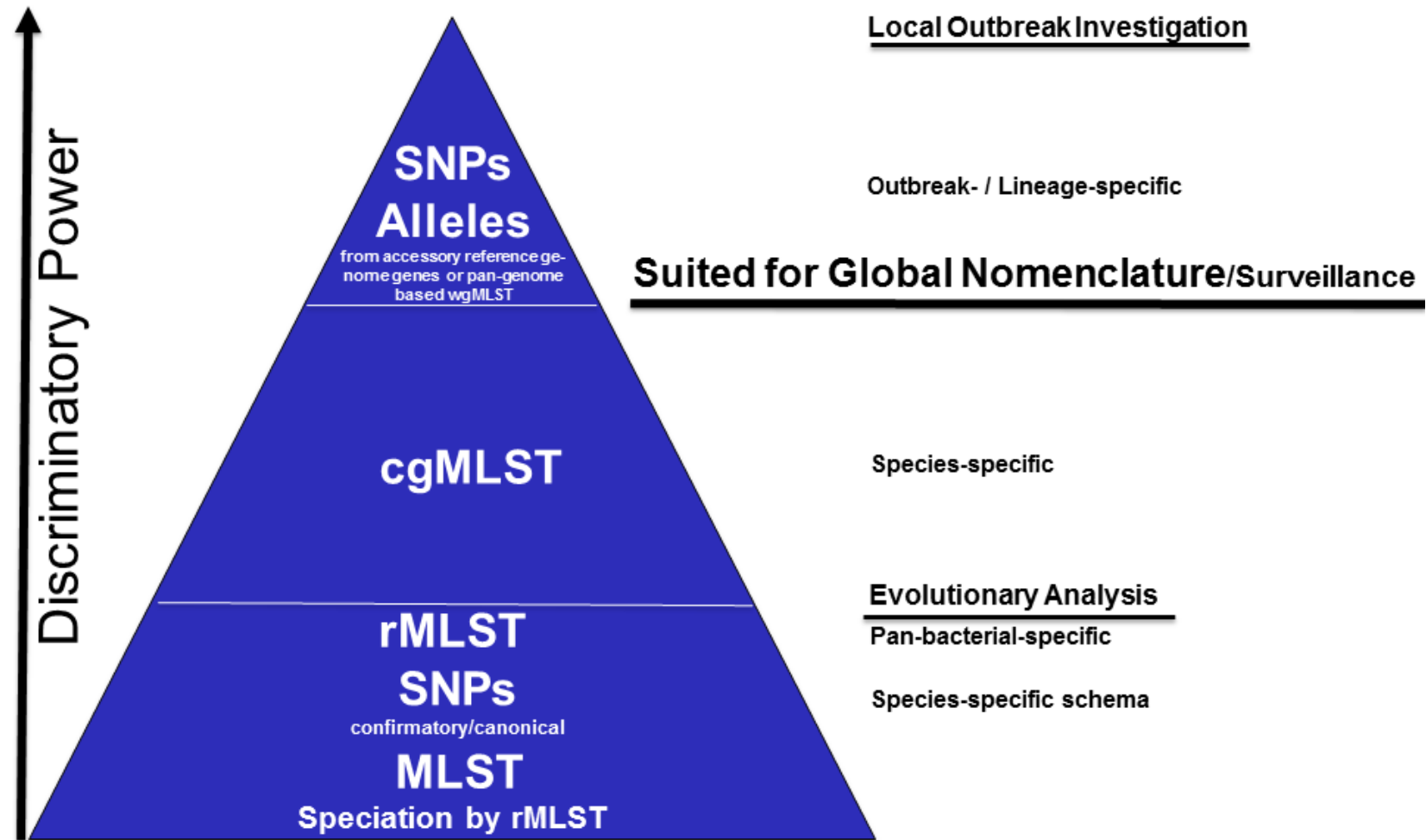
ddl-up, 5'-TGC (C/T)CA AGT TCC TTA TGT GG  
ddl-dn, 5'-CAC TGG GT(G/A) AAA CC(A/T) GGC AT

PCR amplification is carried out on chromosomal DNA using an extension time of 30 seconds, and an annealing temperature of 50 °C, with Taq polymerase. As the same primers are used for amplification and sequencing, it is important that only a single DNA fragment is amplified in the initial PCR. This may involve some optimisation of the annealing temperature.

The following housekeeping genes and primers for MLST

<https://pubmlst.org/organisms>

# Sequence-based methods



# Molecular Typing Methods

Typing Method	Basis Features	Public Database Link
<b>PFGE</b>	<ul style="list-style-type: none"> <li>Commonly used for epidemiological investigations of foodborne diseases and others</li> </ul>	<ul style="list-style-type: none"> <li><a href="http://www.pulsenetinternational.org">www.pulsenetinternational.org</a></li> </ul>
<b>AFLP</b>	<ul style="list-style-type: none"> <li>Analyses a subset of DNA regions of restriction enzyme-digested bacterial genome</li> </ul>	
<b>MLVA</b>	<ul style="list-style-type: none"> <li>Determines the number of tandem repeats in multiple loci; capable of detecting genetic differences between strains of highly homogeneous species</li> </ul>	<ul style="list-style-type: none"> <li><a href="http://www.pasteur.fr/mlva">http://www.pasteur.fr/mlva</a></li> <li><a href="http://minisatellites.upsud.fr/MLVAnet">http://minisatellites.upsud.fr/MLVAnet</a></li> <li><a href="http://www.pulsenetinternational.org/protocols/Pages/mlva.aspx">http://www.pulsenetinternational.org/protocols/Pages/mlva.aspx</a></li> </ul>
<b>MLST</b>	<ul style="list-style-type: none"> <li>Standard strain typing method including six to eight loci; typing scheme has been developed for many bacterial species</li> </ul>	<ul style="list-style-type: none"> <li><a href="http://pubmlst.org">http://pubmlst.org</a></li> <li><a href="http://www.mlst.net">http://www.mlst.net</a></li> </ul>
<b>WGS</b>	<ul style="list-style-type: none"> <li>Detects genetic variations at the genome level and provides higher resolution than other common typing methods</li> </ul>	<ul style="list-style-type: none"> <li><a href="http://www.ncbi.nlm.nih.gov/genome/">http://www.ncbi.nlm.nih.gov/genome/</a></li> <li><a href="http://img.jgi.doe.gov">http://img.jgi.doe.gov</a></li> </ul>
<b>SNP</b>	<ul style="list-style-type: none"> <li>Targeting polymorphic sequences of the whole or partial genome sequences; valuable for tracking the spread of monomorphic pathogens</li> </ul>	

# Molecular Typing Methods

Features of molecular strain typing methods for bacterial organisms

Method	Type of markers used for differentiation	Discriminatory power	Reproducibility	Bioinformatic knowledge needed	Cost
Pulsed-field gel electrophoresis (PFGE)	Number of bands depending on restriction enzyme	•	••	•	••
Multilocus sequence typing (MLST)	7–8 housekeeping genes	••	•••• <sup>a</sup>	••	••
Core genome MLST (cgMLST)	Hundreds to thousands of core genes	•••	•••• <sup>a</sup>	•••	•••
Whole genome MLST (wgMLST)	Hundreds to thousands of core plus accessory genes	•••	••••	•••	•••
Reference-based single nucleotide polymorphism (SNP) calling	Depends on organism of interest plus reference choice	••••	•••	••••	••••
Reference-agnostic/k-mer based SNP calling	Depends on organism of interest	••••	••••	••••	••••

• low, •• medium, ••• high, •••• very high.

<sup>a</sup> Generally high, but depends on organism of interest and chosen reference.

*Curr Opin Infect Dis.* 2021 August 01; 34(4): 339–345. doi:10.1097/QCO.0000000000000743.

**Thank you for your attention!**