# MGMA2024:

# A Review of R for Microbial Data Analysis

Explore the atlas1006 microbiome dataset

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#### CONTENT

- 1. Working with sample metadata
- 2. The atlas1006 dataset exploration
- 3. Data manipulating with R
- 4. Statistical analysis with R

# **Working with metadata**

### Performing a microbiome study

#### Study design

(power, randomization)
(metadata--standard + specific to your study)

#### Sample collection

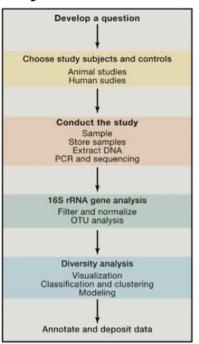
(method, shipping/storage)
lab protocols
(include neg/pos controls)

#### Data processing

#### Data analysis

#### For more on these topics, see:

Conducting a Microbiome Study, by Goodrich et al. 2014
Reagent Contamination, Salter et al. 2014
Storage effects, by Song et al. 2016
Microbiome Quality Control (MBQC), by Sinha et al. 2017
MIMARKS, by Yilmaz et al. 2011
KatharoSeg low biomass workflow, by Minich et al. 2017



#### Sample Information

Sequence Data

**Feature Table** 

**Summary Statistics** 

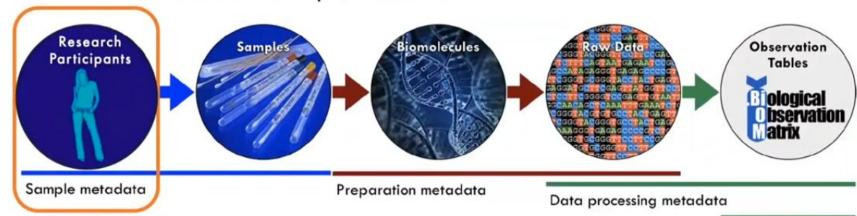
#### Other resources:

Earth Microbiome Project website Human Microbiome Project website American Gut Project website

## What are Metadata?



#### Microbiome Metadata - Sample metadata



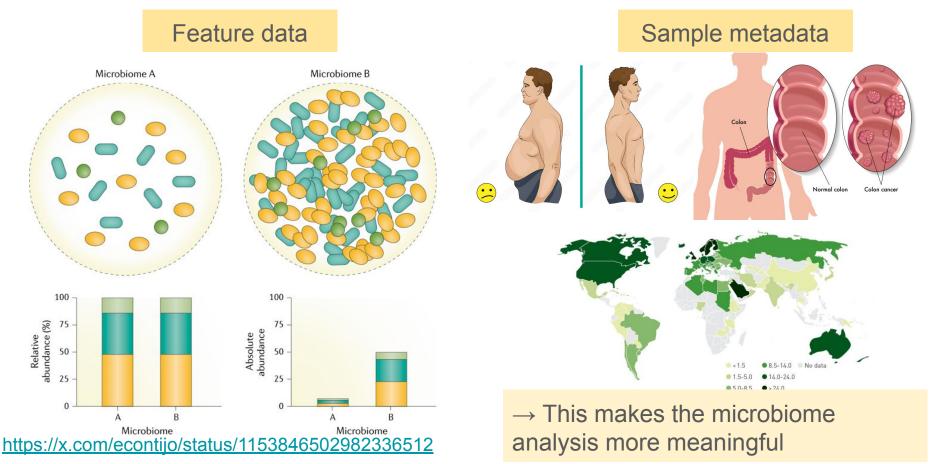
Information about the person providing the sample

Feature metadata

Adapted from: Introduction to Metadata and Ontologies: Everything You Always Wanted to Know About Metadata and Ontologies (But Were Afraid to Ask) DOI: 10.25979/1607365

#### Microbiome feature data & sample metadata

Combine between...



# The atlas1006 dataset

## Data from: Tipping elements in the human intestinal ecosystem

Lahti, Leo <sup>1</sup>; Salojärvi, Jarkko <sup>1</sup>; Salonen, Anne <sup>1</sup>; Scheffer, Marten <sup>2</sup>; de Vos, Willem M. <sup>1</sup>

Author affiliations >

Published May 19, 2015 on Dryad. https://doi.org/10.5061/dryad.pk75d



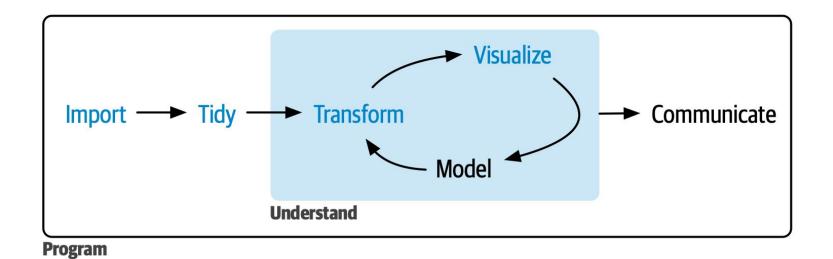
#### Cite this dataset



Lahti, Leo et al. (2015). Data from: Tipping elements in the human intestinal ecosystem [Dataset]. Dryad. https://doi.org/10.5061/dryad.pk75d

1006 individuals

#### Data analysis workflow



## Import packages

```
# Install package
install.packages()

# Get the list of installed packages
installed.packages()

# Import package
library()
```

## Import data

```
# Read CSV file
read.csv("file.csv")

# Reads a file in table format more
flexibility in specifying the delimiter
read.table("file.txt")
```

#### First look of our data

```
> atlas1006
phyloseq-class experiment-level object
otu_table() OTU Table: [ 130 taxa and 1151 samples ]
sample_data() Sample Data: [ 1151 samples by 10 sample variables ]
tax_table() Taxonomy Table: [ 130 taxa by 3 taxonomic ranks ]
```

	age	sex	nationality	DNA_extraction _method	project	project diversity		subject	time	sample
Sample-1	28	male	US	NA	1	5.76	severeobese	1	0	Sample-1
Sample-2	24	female	US	NA	1	6.06	obese	2	0	Sample-2
Sample-3	52	male	US	NA	1	5.5	lean	3	0	Sample-3
Sample-4	22	female	US	NA	1	5.87	underweight	4	0	Sample-4
Sample-5	25	female	US	NA	1	5.89	lean	5	0	Sample-5

Explain the fields of atlas 1006

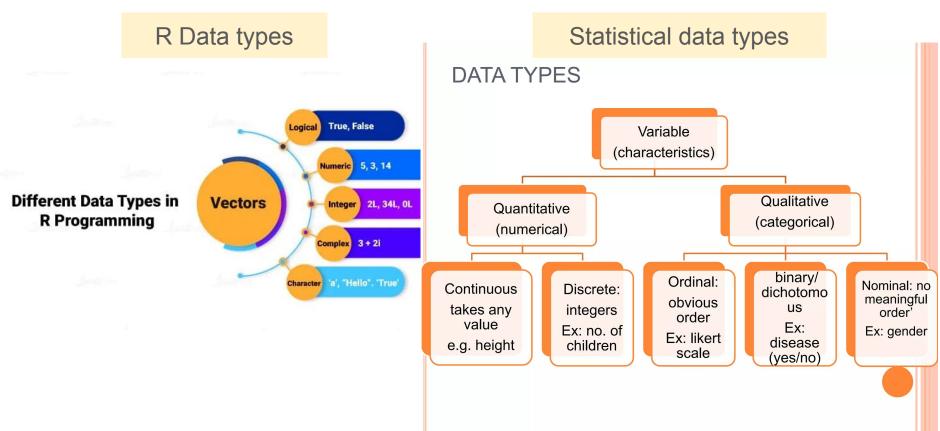
```
# First look of some first fields and values
head()
View()

# Explore the structure and variable type
str()
glimpse()
```

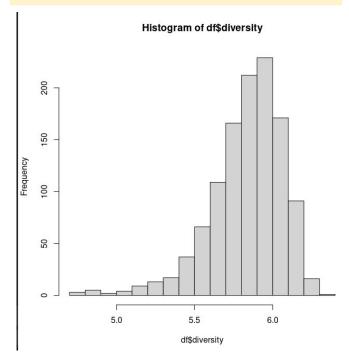


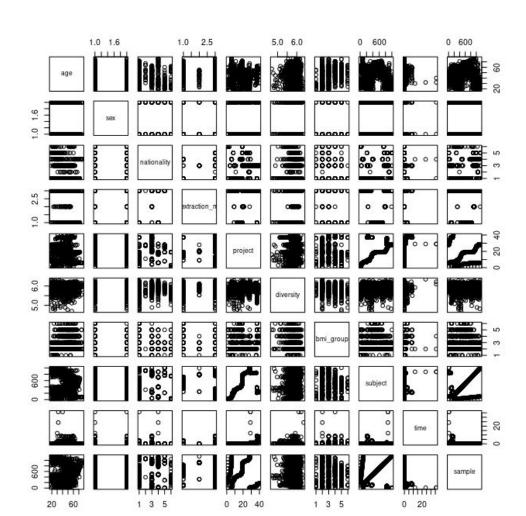
```
> str(df)
'data.frame': 1151 obs. of 10 variables:
                    : int 28 24 52 22 25 42 25 27 21 25 ...
$ age
                    : Factor w/ 2 levels "female", "male": 2 1 2 1 1 2 1 1 1 1 ...
 $ sex
                    : Factor w/ 6 levels "CentralEurope",..: 6 6 6 6 6 6 6 6 6 ...
$ nationality
 : Factor w/ 40 levels "1","2","3","4",..: 1 1 1 1 1 1 1 1 1 ...
 $ project
 $ diversity
                    : num 5.76 6.06 5.5 5.87 5.89 5.53 5.49 5.38 5.34 5.64 ...
 $ bmi group
                    : Factor w/ 6 levels "underweight",..: 5 4 2 1 2 2 1 2 2 2 ...
 $ subject
                    : Factor w/ 1006 levels "1","2","3","4",..: 1 2 3 4 5 6 7 8 9 10 ...
 $ time
                    : num 0 0 0 0 0 0 0 0 0 0 ...
                    : chr "Sample-1" "Sample-2" "Sample-3" "Sample-4" ...
 $ sample
```

A short mention about data type...

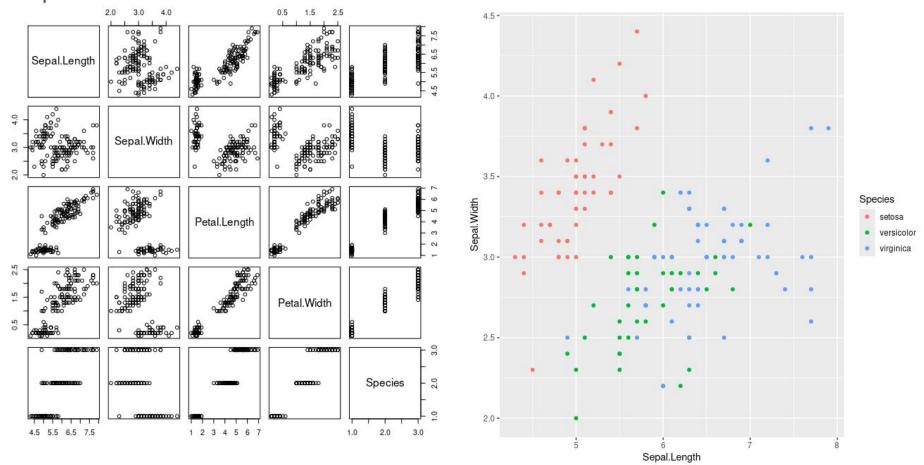


```
# Explore by visualizing
plot()
hist()
```





#### Explore iris



```
# More advance
skim()
```

DataExplorer::create\_report(df)

```
    Variable type: character -

 skim_variable n_missing complete_rate min max empty n_unique whitespace
sample
                                     1 8 11
                                                         1151
 - Variable type: factor
 skim variable
                       n missing complete rate ordered n unique
 sex
                              37
                                         0.968 FALSE
 nationality
                                         0.972 FALSE
                                                               6
 DNA extraction method
                                         0.818 FALSE
                             209
1 project
                                               FALSE
                                                             40
                             106
                                         0.908 FALSE
 bmi aroup
                                                              6
6 subject
                                               FALSE
                                                           1006
 top_counts
fem: 666. mal: 448
2 Cen: 650, Sca: 271, Sou: 89, UKI: 50
3 г: 510, o: 396, p: 36
 19: 303, 7: 107, 28: 90, 9: 84
5 lea: 484, obe: 222, ove: 197, sev: 99
6 831: 5, 832: 5, 833: 5, 834: 5

    Variable type: numeric

 skim_variable n_missing complete_rate mean
                                                  sd p0
                                                            p25 p50 p75 p100
                      56
                                 0.951 45.0
 age
 diversity
                                                                        6 6.35
3 time
                                                                        0 36
 hist
```

#### Data Profiling Report

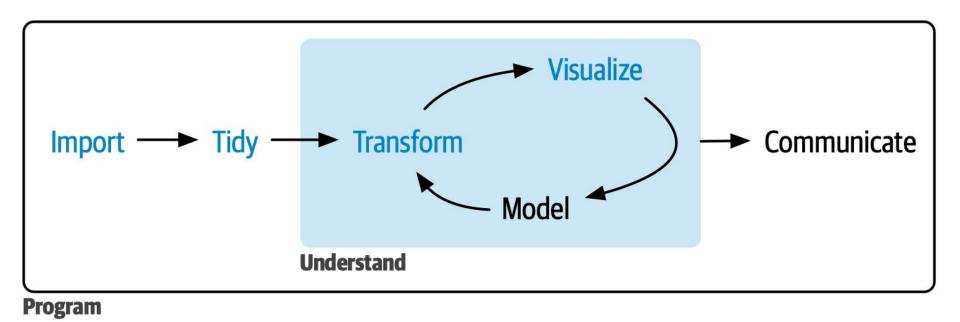
- Basic Statistics
  - Raw Counts
  - Percentages
- Data Structure
- · Missing Data Profile
- · Univariate Distribution
  - Histogram
  - Bar Chart (with frequency)
  - QQ Plot
- Correlation Analysis
- Principal Component Analysis

#### **Basic Statistics**

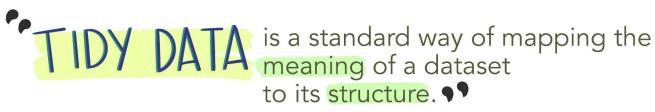
#### Raw Counts

Name	Value
Rows	1,151
Columns	10
Discrete columns	7
Continuous columns	3
All missing columns	0
Missing observations	440
Complete Rows	927
Total observations	11,510
	222.11

#### Data manipulation - The workflow



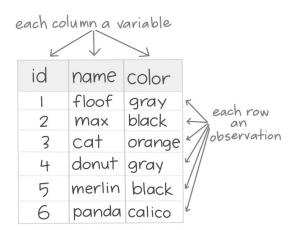
Introduction to data tidying



-HADLEY WICKHAM

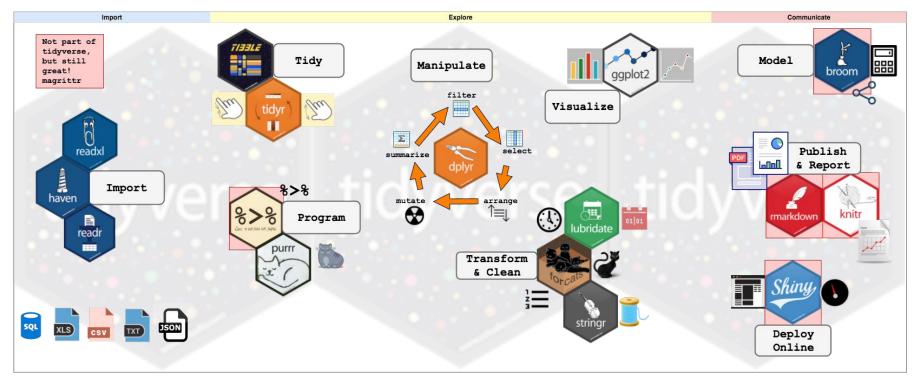
# In tidy data:

- each variable forms a column
- each observation forms a row
- each cell is a single measurement



Wickham, H. (2014). Tidy Data. Journal of Statistical Software 59 (10). DOI: 10.18637/jss.v059.i10

#### tidyverse package

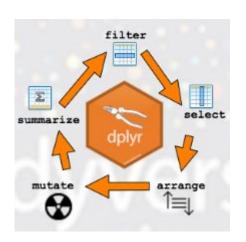


Data tidy - Cleaning and Filtering

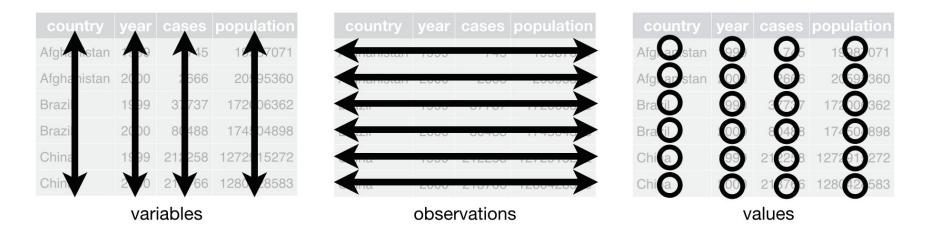
```
# Select interested columns
select()

# Filter the row matched our interested
filter()

# Remove NAs
!is.na()
```



#### Reshaping



The following three rules make a dataset tidy: variables are columns, observations are rows, and values are cells.

## Reshaping

```
# Reshape to a longer format
pivot_longer()

# Reshape to a wider format
pivot_wider()
```

id	bp1	bp2		
Α	100	120		
В	140	115		
С	120	125		



id	measurement	value
Α	bp1	100
Α	bp2	120
В	bp1	140
В	bp2	115
С	bp1	120
С	bp2	125

Wide

Long

#### From a wide table...

	Species	Sample-1	Sample-2	Sample-3	Sample-4	Sample-5	Sample-6	Sample-7	Sample-8	Sample-9	Sample-10
1	Actinomycetaceae	0	0	0	0	0	0	0	0	0	0
2	Aerococcus	0	0	0	0	0	0	0	0	0	0
3	Aeromonas	0	0	0	0	0	0	0	0	0	0
4	Akkermansia	21	36	475	61	34	14	27	21	78	169
5	Alcaligenes faecalis et rel.	1	1	1	2	1	1	1	1	1	1
6	Allistipes et rel.	72	127	34	344	50	57	21	9	9	23
7	Anaerobiospirillum	0	0	0	0	0	0	0	0	0	0
8	Anaerofustis	0	0	0	0	0	0	0	0	5	0
9	Anaerostipes caccae et rel.	176	108	27	203	232	166	32	1032	35	1309
10	Anaerotruncus colihominis et rel.	10	48	38	8	21	15	7	11	15	10

> dim(otu)
[1] 130 1151

#### ...to a long table

	Species	sample	count
1	Actinomycetaceae	Sample-1	0
2	Actinomycetaceae	Sample-2	0
3	Actinomycetaceae	Sample-3	0
4	Actinomycetaceae	Sample-4	0
5	Actinomycetaceae	Sample-5	0
6	Actinomycetaceae	Sample-6	0
7	Actinomycetaceae	Sample-7	0
8	Actinomycetaceae	Sample-8	0
9	Actinomycetaceae	Sample-9	0
10	Actinomycetaceae	Sample-10	0
11	Actinomycetaceae	Sample-11	0

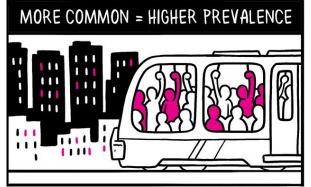
```
# Reshape to a longer format
pivot_longer()
```

```
> dim(otu)
[1] 149630 3
```

Examine the prevalence of microoranisms in atlas1006 cohort

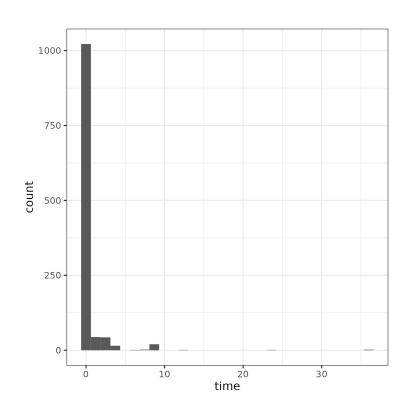
"The number of cases of a disease, number of infected people, or number of people with some other attribute present during a particular interval of time. It is often expressed as a rate (for example, the prevalence of diabetes per 1,000 people during a year)." - **CDC** 

$$\mathbf{P} = \frac{\text{Total number of cases}}{\text{Total population}} = \frac{C}{P}$$





Our population has 1006 people, but why we have 1151 columns?



Based on time distribution,

- → There are some people whose samples collected more than 1 time
- → Filter it. Keep only the first time.

#### From count table...

	Species	Sample-1	Sample-2	Sample-3	Sample-4	Sample-5	Sample-6	Sample-7	Sample-8	Sample-9	Sample-10
1	Actinomycetaceae	0	0	0	0	0	0	0	0	0	0
2	Aerococcus	0	0	0	0	0	0	0	0	0	0
3	Aeromonas	0	0	0	0	0	0	0	0	0	0
4	Akkermansia	21	36	475	61	34	14	27	21	78	169
5	Alcaligenes faecalis et rel.	1	1	1	2	1	1	1	1	1	1
6	Allistipes et rel.	72	127	34	344	50	57	21	9	9	23
7	Anaerobiospirillum	0	0	0	0	0	0	0	0	0	0
8	Anaerofustis	0	0	0	0	0	0	0	0	5	0
9	Anaerostipes caccae et rel.	176	108	27	203	232	166	32	1032	35	1309
10	Anaerotruncus colihominis et rel.	10	48	38	8	21	15	7	11	15	10

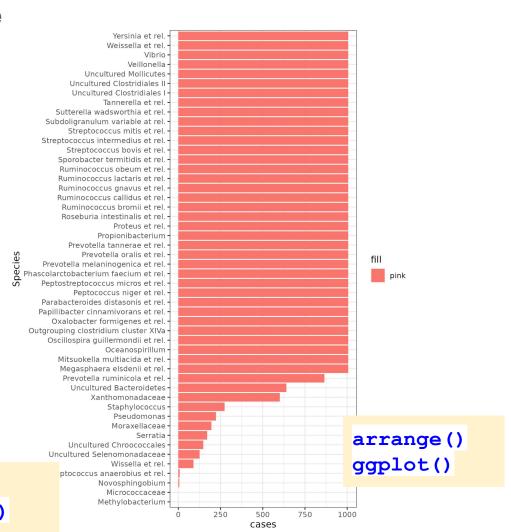
#### ...to case table

	Species	Sample.1	Sample.2	Sample.3	Sample.4	Sample.5	Sample.6	Sample.7	Sample.8	Sample.9	Sample.10
1	Actinomycetaceae	0	0	0	0	0	0	0	0	0	0
2	Aerococcus	0	0	0	0	0	0	0	0	0	0
3	Aeromonas	0	0	0	0	0	0	0	0	0	0
4	Akkermansia	1	1	1	1	1	1	1	1	1	1
5	Alcaligenes faecalis et rel.	1	1	1	1	1	1	1	1	1	1
6	Allistipes et rel.	1	1	1	1	1	1	1	1	1	1
7	Anaerobiospirillum	0	0	0	0	0	0	0	0	0	0
8	Anaerofustis	0	0	0	0	0	0	0	0	1	0
	Anaerostipes caccae et rel.	1	1	1	1	1	1	1	1	1	1
10	Anaerotruncus colihominis et rel.	1	1	1	1	1	1	1	1	1	1

```
mutate()
if_else()
```

#### ...to prevalence table

	Species	cases	total			
1	Actinomycetaceae	292	1006			
2	Aerococcus	3	1006			
3	Aeromonas	0	1006			
4	Akkermansia	1006	1006			
5	Alcaligenes faecalis et rel.	1006	1006			
6	Allistipes et rel.	1006				
7	Anaerobiospirillum	2	1006			
8	Anaerofustis	91	1006			
9	Anaerostipes caccae et rel.	1006	1006			
10	Anaerotruncus colihominis et rel.	1006	1006			
11	Anaerovorax odorimutans et rel.	1006	1006			
12	Aneurinibacillus	11	1006			
13	Aquabacterium	93	sum()			
14	Asteroleplasma et rel.	0 across				

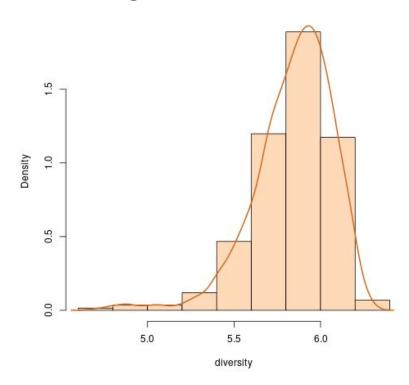


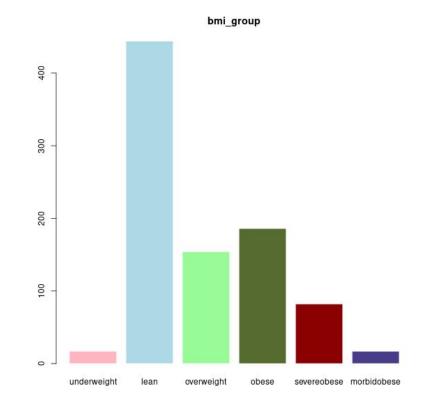
# Statistical Analysis with R

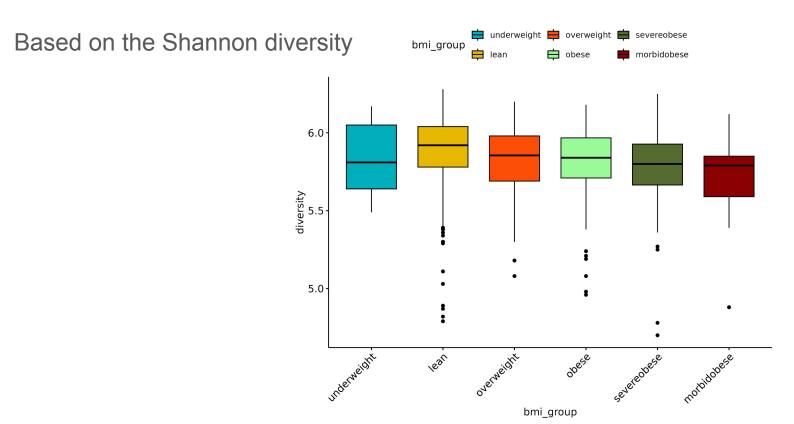
The question can be conducted before study design or after EDA

```
> str(df)
'data.frame': 1151 obs. of 10 variables:
 $ age
                       : int 28 24 52 22 25 42 25 27 21 25 ...
                       : Factor w/ 2 levels "female", "male": 2 1 2 1 1 2 1 1 1 1 ...
 $ sex
 $ nationality
                      : Factor w/ 6 levels "CentralEurope",..: 6 6 6 6 6 6 6 6 6 ...
 $ DNA extraction method: Factor w/ 3 levels "o","p","r": NA ...
 $ project
                      : Factor w/ 40 levels "1","2","3","4",..: 1 1 1 1 1 1 1 1 1 ...
 $ diversity
                       : num 5.76 6.06 5.5 5.87 5.89 5.53 5.49 5.38 5.34 5.64 ...
 $ bmi group
                       : Factor w/ 6 levels "underweight",..: 5 4 2 1 2 2 1 2 2 2 ...
 $ subject
                      : Factor w/ 1006 levels "1","2","3","4",..: 1 2 3 4 5 6 7 8 9 10 ...
 $ time
                             0 0 0 0 0 0 0 0 0 0 ...
                       : num
 $ sample
                       : chr "Sample-1" "Sample-2" "Sample-3" "Sample-4" ...
```

After doing EDA...







Do lean people have more diversity in interstitial microbiota?

Scientific question

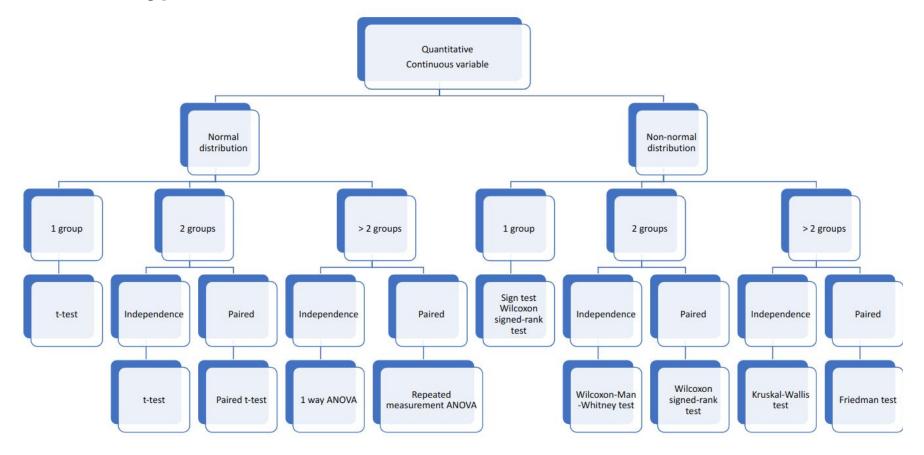
Is there any significant difference in **microbiome diversity** between those who have a **lean** body than those who are **overweight** or **obese**?

#### **Hypothesis**

**Ho:** No significant difference in microbiome diversity between individuals who have a lean body and those who are overweight or obese.

**Ha:** There is a significant difference in microbiome diversity between individuals who have a lean body and those who are overweight or obese.

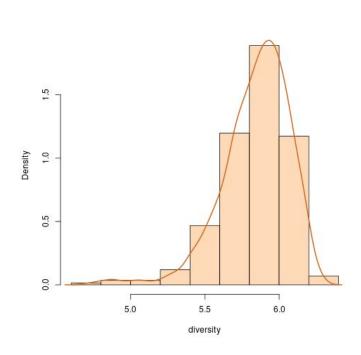
#### **Conduct a hypothesis test**

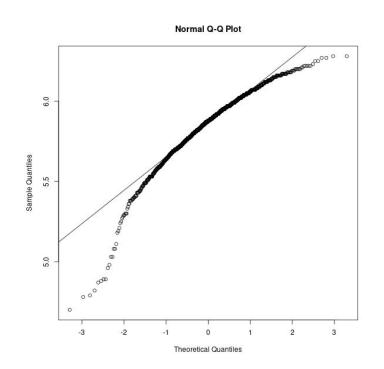


(\*) This beautififul chart is from Tran Ba Thien

## **Conduct a hypothesis test**

#### Normality check





Normaly distributed?

#### **Choose hypothesis test**

Normality check - Shapiro-Wilk test

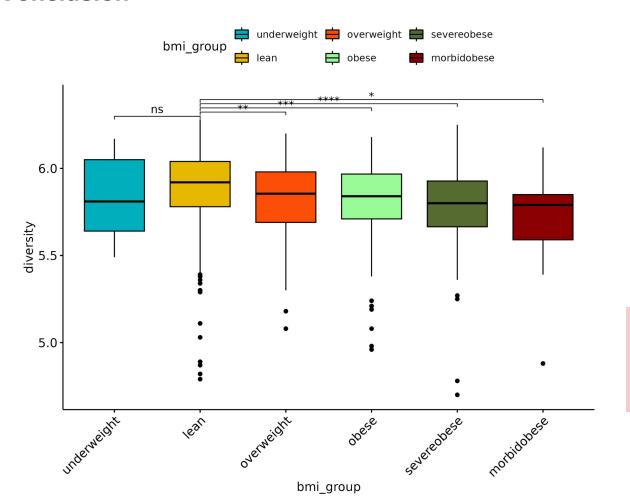
```
> meta |> pull(diversity) |> shapiro.test()

Shapiro-Wilk normality test

data: pull(meta, diversity)
W = 0.93439, p-value < 2.2e-16</pre>
```

- → Not a normal distribution
- → Non-parametric test (Wilcoxon test)

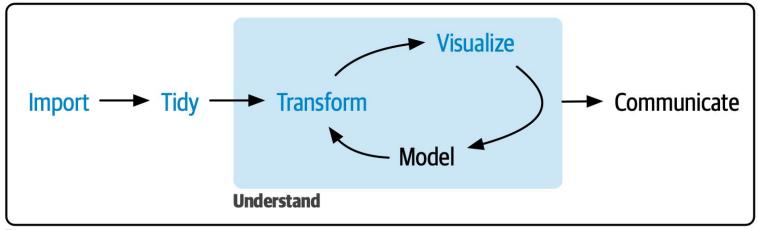
#### Conclusion



Reject Ho

 → Lean people have more diversity in intestinal microbiome

#### Conclusion



**Program** 

#### **Further readings**

Introduction to Urobiome Metadata Standards <a href="https://www.youtube.com/watch?v=nCxZ8m7bcwY">https://www.youtube.com/watch?v=nCxZ8m7bcwY</a>

Sample Metadata

https://www.youtube.com/watch?v=hh6pqmzJWds&t=22s

# THANK YOU