

# ADAMTS-13 Activity Assay

for Research Use Only in US & Canada

Tips for Training & Validation

# Assay & Reagent Prep

- Allow all reagents to reach Room

- **Dilute** wash buffer by mixing  
1 part

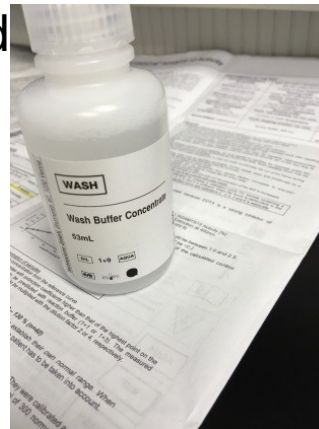
- wash buffer with 9 parts  
diH<sub>2</sub>O

- **Reconstitute** vWF substrate  
solution with 6mL diH<sub>2</sub>O.

- 15 min after  
reconstitution mix for 10  
sec using vortex mixer

- **Reconstitute** calibrators and  
controls with 500μL diH<sub>2</sub>O

- 15 min after  
reconstitution mix for 10  
sec using vortex mixer



# Reagent Info & Storage



Material/ Reagent	State	Storage	Expiration
ELISA test strips	after opening	2-8°C in storage bag with desiccant	expiration date on bag
GST-vWF73 Substrate	after reconstitution	-20°C	6 weeks
Calibrators, Controls	after reconstitution	-20°C	6 months
Reaction Buffer	after opening	2-8°C	6 months
Conjugate	after opening	2-8°C	4 months
TMB Color Reagent	after opening	2-8°C	expiration date on bottle
Wash Buffer Concentrate (10x)	after opening	2-8°C	6 months
Wash Buffer	after reconstitution	2-8°C	3 weeks

Expiration date on labels refers to storage of unopened vial at 2-8°C.  
 Tech tip: Aliquot reconstituted calibrators, controls & substrate in no less than 250 µl.  
 Storage in Eppendorf vials is acceptable. Thaw at room temperature for best results.

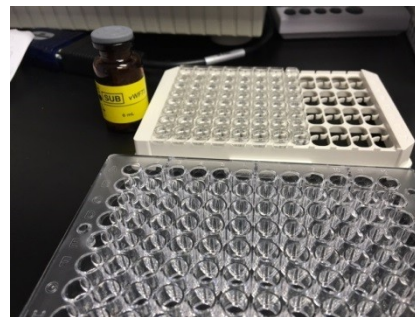
# Sample Dilution & Preparation



te samples and reconstituted calibrators & controls 31-fold:

Using the sample dilution microplate, pipette **5 $\mu$ L** of **sample**, reconstituted **calibrator** or **control**. Then add **150 $\mu$ L** of **reaction buffer** and mix well.

**Note:** Samples, cals & controls should be tested in duplicate so this dilution will need to be done 2x for each sample.



For higher precision, use dilution tubes and add **20 $\mu$ L** of **sample**, reconstituted **calibrator** or **control** to the tube. Then add **600 $\mu$ L** of **reaction buffer** and mix well.

**Note:** Samples, cals & controls should be tested in duplicate so pipette duplicate wells from this 1 dilution preparation.

**Reverse pipetting** is recommended.

**Incubation times** begin after pipetting the last sample. *Pipetting time for all samples should not exceed 60 seconds (cals/controls/samples/conjugate).*

**Samples > highest cal** should be re-tested by pre-diluting 1:2 or 1:4 with **reaction buffer**. The measured concentration is then multiplied by the dilution factor.

# Assay Tips

- **Reagents from kits with different lot numbers should not be used together.**

- exclusions: wash buffer concentrate, stop solution, sample dilution microplate & plate sealers

- **Avoid using thawed calibrators, controls or substrate with fresh calibrators, controls or substrate**

- i.e. If using thawed calibrators, all calibrators should be tested

- **Precision & Performance depend on:**

- ELISA reader: verify appropriate maintenance
  - Appropriate incubation temperature (20-25°C)
  - Appropriate incubation times – should not vary by more than 10%
    - GST-vWF73 substrate incubation: 57-63 min
    - Sample incubation: 28.5-31.5 min
    - Conjugate reaction: 57-63 min
    - TMB Color Reagent reaction: 28.5-31.5 min
    - *Pipetting time during substrate reactions and at stopping should not exceed 10 sec per test strip.*

Tech Tip: Incubating at the highest allowable temp (25°C) & for longest allowable time (63 or 31.5 min) will increase assay OD values.

- **Use of a multichannel pipette is encouraged.** *Remember to change tips when pipetting from row to row.*



# Limitations & Interferences

TA is a strong inhibitor of ADAMTS-13 function. **Do not use samples with EDTA**

**hemolysis:** No interference is observed with samples containing up to 200mg/dL hemoglobin, which corresponds with a moderate haemolysis /dL.

**emia:** No interference is observed with samples containing up to 300 mg/dL Int which corresponds with a moderate to severe concentration.

**erus:** No interference was observed with samples containing up to 15mg/dL bili (conjugated as well as unconjugated), which corresponds with a moderate to severe concentration.

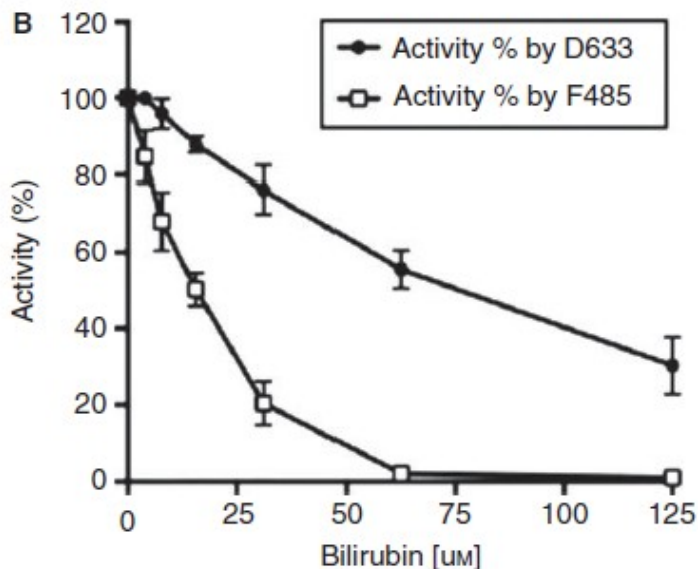
**umatoid factor:** No interference was observed up to 30U/mL RF, with corresponding a 2-fold concentration of normal.

**ti CD20 antibodies:** No interference was observed up to a level of 200µg/mL, which corresponds to the upper level of serum concentrations found after rituximab administration.

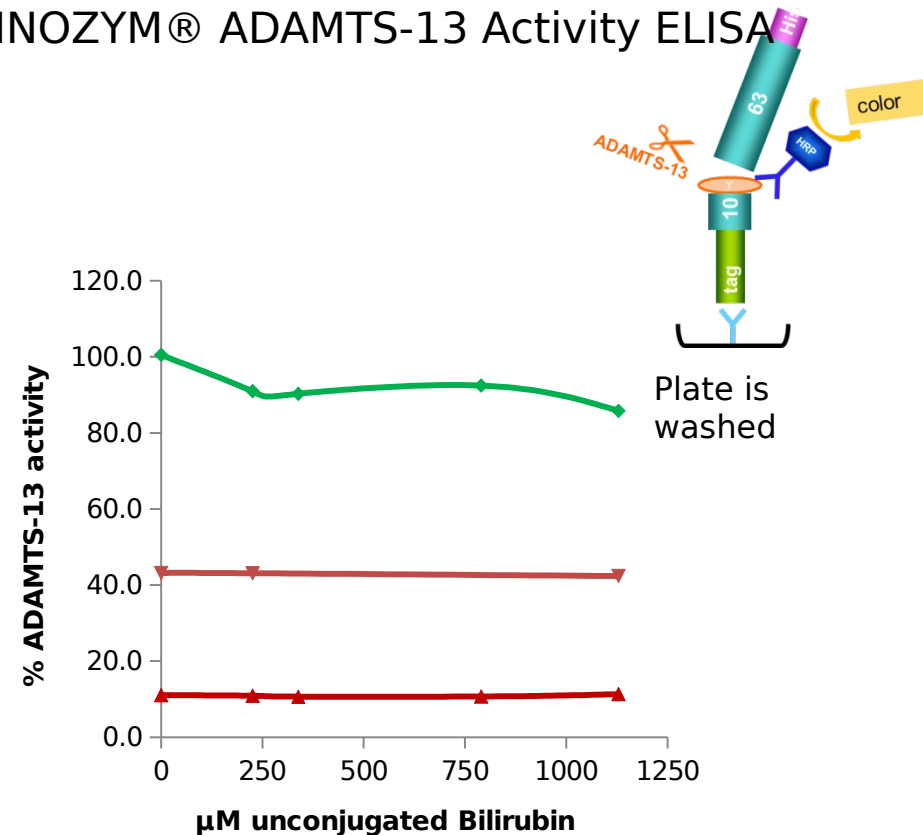
# Interferences - ADAMTS13 Assays

## Fluorogenic Assays

**Unconjugated bilirubin interferes with fluorogenic assays:**

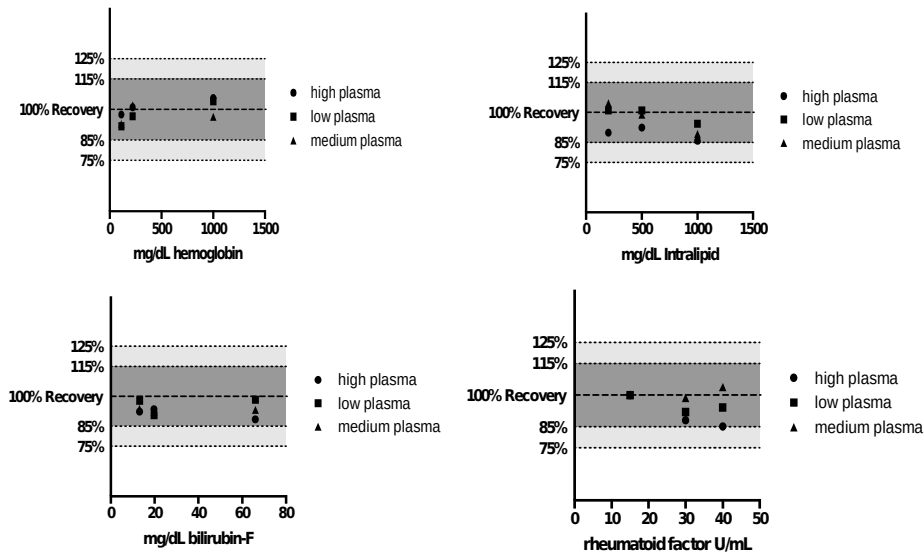


## TECHNOZYM® ADAMTS-13 Activity ELISA



**No interference of unconjugated bilirubin during measurement independent of ADAMTS-13 concentration range**

# Interferences - TECHNOZYM® ADAMTS-13 Activity ELISA



**No interference of hemolysis, lipemia, conjugated bilirubin and rheumatoid factor during measurement.**



# Analysis of Results



Reader settings should be established with the aid of your instrument's technical support &/or internal IT department.

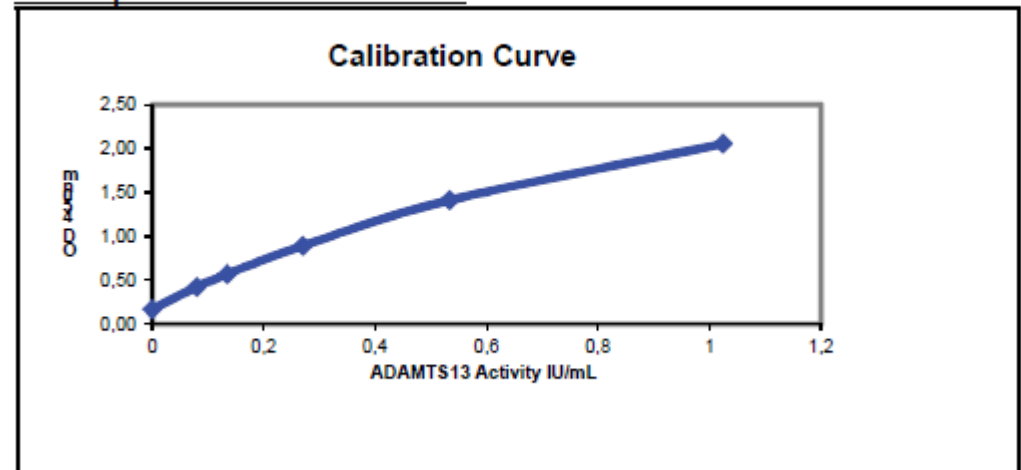
- Evaluation software (Excel spreadsheet with calculations) is available from Technoclone to convert raw OD values to IU/mL, though most readers can do this as well.

Set up a reference curve:

- X-axis = ADAMTS-13 activity (IU/mL)
- Y-axis = extinction at 450nm
- Graph a linear-linear plot with a best fit curve.

The validity of the test is assessed using the calculated control values.

Example of standard curve



# Lot-to-Lot Qualification Suggestions

**Lot qualification procedures should always be established in conjunction with your institution's QA policies & procedures.**

Perform 20 assay runs of each control to establish your laboratory control range.

- Test using a previously qualified lot; and
- Use control range on the Certificate of Analysis from the new lot of control to accept/reject the “test” control.

Take the mean of these 20 values to establish the mean for your laboratory.

- Use Westgard rules & a Levey-Jennings chart to accept/reject runs based on control values with the new lot.

Test previously assayed samples using the new lot. Verify sample interpretation is the same between lots.

# Method to Prepare Low-Activity Samples

- Heat-treat normal human plasma at **56°C** for **30 minutes** (inactivated plasma)
  - *This inactivates endogenous ADAMTS13 protease activity.*
- Heat-treated plasma can then be diluted in different concentrations (1:1, 1:2, 1:4, 1:8, 1:16) with normal plasma (pooled 5-10 individuals).
- Run the normal plasma straight to calculate expected values of diluted heat-inactivated samples & then run those as low-activity samples.

# Training Video

- [www.diapharma.com/a13](http://www.diapharma.com/a13)