# Fish-Tracking-Visualization

Project: Developing Exploration Behavior

Holds the scripts to visualize the molly's trajectory. The folder tex/trajectory contains the resulting pdfs. To make use of the links to the mp4 and csv-files – connect to the server loopbio\_data. Currently the links work for MacOX systems and best with Adobe Reader.

## Requirements

- python3, gcc, latex
- install dependencies:
  - pip3 install -r requirements.txt

## Build

• To compile the *Cython* code and creating you own script/env.sh, run:
- python3 setup.py build\_ext --inplace

## Trajectory Visualization PDFs

scripts/env.sh contains the paths to the trajectory data. One can configure these to point to the correct location of the data. Reading the data directly from the server  $loopbio_data$  results in long running times. It is recommended to use a external hard drive. If you path uses spaces, for example the name of the hard drive, rename it to underscores - \_.

Accessing the data from the server is very slow.

### Generate the trajectory visualizations, run:

- python3 main.py program={trajectory, feeding}
- optional arguments: fish\_id=<<cam\_pos>>, test={0,1}

### Then run the bash-script:

- bash scripts/build-trajectories.sh
- optional argument:
  - --feeding or -f for the feeding trajectories.
  - --test, -t is used to test the script, to generate only the fist pdf.
  - --local, -l to use the paths of the local hard drive to link the csv file in the pdf.
  - --cam-id cameraID, -cam cameraID\_position, to create only the pdf for the given camera.

**Remark:** For the bash-script you can not build feeding and non feeding trajectories in parallel as they use the same files.

## Data File and Path Validation

The python script path\_validation.py is used to validate the filenames and paths of the data files. It logs all error messages into log-path-validation.txt.

### Run the script:

• python path\_validation.py — optional arguments <<delete=1, n\_files=15>> to delete duplicated filenames and change the expected number of files in a folder for a day.

## Trajectory Analysis

- run: python3 main.py program={metric} <<optional>> time\_interval=<<default>>100 fish\_id=<<cam\_pos>> For the metric argument use one out of {activity, angle, tortuosity, entropy, abs\_angle, wall\_distance, all}.
- run python3 main.py program={metric} time\_interval=hour to record mean and standard derivation per fish per hour in one csv-file.
- run: bash scripts/build\_analytics.sh to generate the pdfs.

#### **Metrics:**

- step length is the length of the vector drawn between to consecutive data frames
- the mean and standard derivation illustrated in the visualization is computed from filtered data frames, removing obvious error point and normed by the distance between data frame when erroneous data point where removed.
- The number of spikes is defined by the threshold of '>  $\mu + 3\sigma$ '
- For the sum of angles we take each angle between consecutive steps anticlockwise ' $\alpha \in [-\pi, \pi]$ '.
- For the average angle each angle ' $\alpha > 0$ '

Compute: function(fish\_id, time\_interval in sec) In methods.py there are four function to calculate the metrics and store mean and standard derivation in /results/<<time\_interval>>\_<<metrics\_name>>.csv.

- activity\_per\_interval
- turning\_angle\_per\_interval
- tortuosity\_per\_interval
- entropy\_per\_interval

### Start on the GPU

- srun -pty -partition=ex scioi gpu -gres=gpu:1 -time=0-02:00 bash -i
- conda activate rapids-22.04

- Type ifconfig and get the inet entry for eth0, i.e. the IP address of the node
- ssh -L localhost:5000:localhost:5000 luka.staerk@[IP address]
- jupyter-lab --no-browser --port=5000

## Open TODOs

- pdfs for windows adapted the root to C:\data\.... (Not needed for now)
- \href in windows not working.
- start BioTracker from link. Install BioTracker on Mac?
- How to start the BioTracker .\BioTracker.lnk --video="X:\path-to-file.mp4"

Further documentation will follow here...