

# pyWrairLib Documentation

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May 29, 2013

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## 1 Install

It is highly recommended to install utilizing a virtual environment. It will be assumed from here on that you are doing so.

## 1.1 Virtual Env Setup

If you already have a virtual env setup you do not need to continue with this section. That is, the output of the following command is not empty or you know what a virtualenv is:

```
echo $VIRTUAL_ENV
```

Otherwise, simply pick where you want your virtual environment to be and issue the following command:

```
virtualenv --distribute <name of virtualenv>
```

Then issue the following to activate the environment

```
source <name of virtualenv>/bin/activate
```

## 1.2 Install the dependencies

```
pip install -r requirements.txt
```

## 1.3 Install pyWrairLib

1. Edit the config/settings.cfg file inside of the pyWrairLib folder to suit your environment. More on settings.cfg below
2. Execute the installer

```
cd pyWrairLib
python setup.py install
```

# 2 The Pipeline

## 2.1 Introduction

The concept of The Pipeline is quite simple. Execute one script after another performing one task after another on the data until it is processed. Because NGS Data is so large, it is best to run most of these scripts on a Multiprocessor/Multicore system. If you want to know how many CPUs and Cores you have in linux just issue the following command

```
cat /proc/cpuinfo
```

You will have to manually look through this specifically looking for the physical id and cpu cores

## 2.2 Pipeline Pieces

The Pipeline is composed of two parts:

1. Data Setup
2. Analysis

## 2.3 Available Scripts

These are the available scripts that drive the pipeline. The full documentation for each of the scripts is detailed at the bottom of the document.

### 2.3.1 Analysis

- genSummary.sh
  - Runs summary scripts to generate compiled summary output for GsMapper projects inside a directory
- mapSamples.py
  - Creates and runs gsmapper projects based on a RunFile
- mapSummary.py

- Creates AllRefStatus.xls file which gives mapping depth and coverage percentage for each reference that was used in the reference mappings
- genallcontigs.py
  - Creates a directory containing all GsMapper project’s 454AllContigs.fna+.qual consolidated into a single directory. Each AllContigs fasta+qual file is merged into a single .fastq file named after the GsMapper project directory from which it originated.
- allcontig\_to\_allsample.py
  - Merges a GsMapper project’s 454AllContigs.fna and 454AllContigs.qual file into a single .fastq
- genallrefstatus.sh
  - Legacy shell script that merges together all found 454RefStatus.txt files into a single output stream/file
- variant\_lookup
  - Script to aid in the lookup of variant information

### 2.3.2 Data

- demultiplex
  - Demultiplexes all found sff files inside of a given sff directory. Also renames the generated files utilizing a given runfile.
- link\_reads
  - Links all valid reads found in a given directory into the NGSDData’s ReadsBySample directory
- sanger\_to\_fastq
  - Helper script to convert sanger .ab1 sequence files to .fastq files. Useful as Newbler only accepts .sff or .fastq as input sequence format.

### 2.3.3 Misc/Undocumented/InDevelopment

- bestblast
- entrez\_helper
- reads\_for\_contig
- refrename.py
- splitsff

## 3 Running the Pipeline

*Anything between a greater than and less than symbol should be replaced using information for your setup*

### 3.1 Data Setup

The data setup requires that you follow the directory structure in the settings.cfg file which should be located in your PYTHONHOME under the config directory. Anytime you see <ngsdir> in the following section replace it with the path that is in your settings.cfg file set for NGSDATA\_DIR Same goes for <readsbysampledir>, <rawdatadir> and <readdatadir>

### 3.1.1 All Platforms

- Copy the Sequence data from the instrument into its appropriate location under the <rawdatadir> directory

```
rsync -av --progress <path_to_data>/<sequence_run_date> \  
<rawreadsdir>/<sequencer_type>/
```

- Change directory to the copied directory

```
cd <rawreadsdir>/<sequencer_type>/<sequence_run_date>
```

- Proceed with the directions for the platform you are working on below and then return to the next step
- Link into the NGSDData/ReadsBySample directory

```
link_reads
```

After this is completed all the data for each sample will be located under the <readsbysampledir> directory under that sample's name This is important because mapSamples.py looks there for read data

### 3.1.2 Sanger

Sanger data needs to be converted to \*.fastq files using the sanger\_to\_fastq script

- In theory you should be able to link the RawReads directly into the ReadData folder and then simply run sanger\_to\_fastq on them. That being said, you will likely have to rename the files correctly before doing so which will not be detailed here.

```
ln -s <rawdatadir>/Sanger/<sequence_date> <readdatadir>/Sanger/
```

- Change directory to the linked directory and create fastq files

```
cd <readdatadir>/Sanger/<sequence_date>  
sanger_to_fastq -d $(pwd) -t fastq
```

### 3.1.3 Roche454 and IonTorrent

Roche454 and IonTorrent both produce sff files which first need to be demultiplexed before linking

- Create a meta directory for that run

```
mkdir meta
```

- Create Primer and Ref directories and copy RunFile into meta directory. You may have to search a bit for the RunFile

```
mkdir Primer  
mkdir Ref  
cp <RunFile.txt> meta/  
cp <path_to_primer_fasta_files> meta/Primer/
```

- Change directory into the R\_ directory that is not the prewash directory

```
cd R_<run_name>
```

- Link the meta directory into the Signal Processing directory

```
ln -s ../meta D_*signalProcessing*/
```

- Link the Signal processing directory from the RawReads sequencer directory into NGSDData/ReadData

```
ln -s D_*signalProcessing* <readdatadir>/<sequencer_type>/
```

- Change directory to the Signal Processing Directory for the run you want to demultiplex. The following command only works if you issue it directly after the above command.

```
cd !$
```

- Run the demultiplex script to demultiplex the raw sff files and rename them according to the runfile

```
demultiplex -s sff/ -r meta/<RunFile.txt>
```

## 3.2 Analysis

- Change directory to the base of your Analysis data

```
cd <path to Analysis>
```

- Create a new analysis directory and change directory to it. Typically named after the NGS Sequencer run you wish to analyze

```
mkdir <YYYY_MM_DD>  
cd <YYYY_MM_DD>
```

- Link in the meta directory for the NGS Run

```
ln -s <path to NGSDData>RawReads/<sequencer type>/<run date of sequencer>/meta .
```

- Copy any needed references into the meta/Ref folder
- Create a directory for the analysis with whatever name you want with a workfile directory to place summaries and RunFile

```
mkdir myanalysis && mkdir myanalysis/workfile
```

- Copy Runfile from meta directory to workfile directory

```
cp meta/Runfile.txt myanalysis/workfile
```

- Edit the copied runfile inside of the workfile directory and set references for each of the samples. You may specify a directory of references instead of individual reference files to include all \*.fasta files inside of that directory.
- Once the runfile is copied you can run the mapping for all samples inside of the RunFile that are not commented out(See more about RunFiles below). Make sure you have changed directory to the analysis directory you created.
  - Be patient if there are a lot of samples to map.

```
cd myanalysis  
mapSamples.py workfile/Runfile.txt
```

- When mapSamples.py is finished you can generate summaries with the genSummary.sh script

```
genSummary.sh
```

## 4 Config Files

### 4.1 settings.cfg

This is the main configuration file that controls how all of the libraries and scripts interact in the various pyWrairLib modules. When pyWrairLib is installed this file is copied into a directory called config inside of the PYTHONHOME environmental variable.

*You can see where this is by issuing*

```
echo $PYTHONHOME
```

**If nothing is displayed then you need to locate your system's default python home**

### 4.2 MidParse.conf

This file is used by the roche software to demultiplex sff files. The contents of the file contain a mapping between a name and a barcode sequence. The structure of the file is as follows

```
MID  
{  
  mid = "MIDNAME1", "BARCODESEQUENCE", MISMATCHTOLERANCE, "OPTIONAL 3' Trim Sequence";  
  mid = "MIDNAME2", "BARCODESEQUENCE", MISMATCHTOLERANCE, "OPTIONAL 3' Trim Sequence";  
}
```

*More information on this file can be found in the Roche documentation Part C under MIDConfig.parse. An example file that can be used is included in the config directory*

## 4.3 RunFile

Run files are simply a file that links a samplename to the following information about that sample

- Region number
- samplename
- virus name
  - Should conform to a valid wrair virus name(More details later on that)
- barcode/midkey name
  - Has to be a valid midkey name inside of the Midparse.conf file(for sffile to demultiplex with using -mcf option)
- mismatch tolerance
- reference directory/file to use to map to
  - Can be a directory of reference files or a single reference
- unique sampleid
  - Deprecated, just use the same as you put in the samplename column
- primerfile location
  - Fasta file of primers to trim off

The following RunFile templates utilize Python Template Strings You can read more about them at:

<http://docs.python.org/2/library/string.html#template-strings>

Basically replace anything that has a dollar sign in front of it with a value

### 4.3.1 RunFile Header

```
# $platform sample list
# $numregions Region $type
# Run File ID: $date.$id
!Region Sample_name Genotype MIDKey_name Mismatch_tolerance Reference_genome_location Unique_sample_id Primers
```

- The header line (starts with a !) is tab separated
- \$platform would be replaced by Roche454 or IonTorrent
- \$date needs to be a valid date string that is in one of the following formats
  - DDMMYYYY
  - DD\_MM\_YYYY
  - YYYY\_MM\_DD
- \$id is anything that does not contain a space character

### 4.3.2 RunFile Sample Line

This line is also tab separated

```
$region $sample $virus $midkey $mismatch $reference $sample $primeri
```

### 4.3.3 Sample RunFile

Here is an example RunFile with 2 samples

Sample D1\_FST2432 is in region 1 and has barcode IX001 from the MidParse.conf file

Sample D1\_FST2410 is in region 2 and has barcode IX002 from the MidParse.conf file

```
# IonExpress sample list
# 2 Region PTP
# Run File ID: 04192013.PGM.CPTLin
!Region Sample_name Genotype MIDKey_name Mismatch_tolerance Reference_genome_location Unique_sample_id Primers
1 D1_FST2432 Den1 IX001 0 Analysis/PipelineRuns/2013_04_19/Ref/Den1 D1_FST2432 NGSData/RawData/IonTorrent/2013_04_19/meta/Primer/Den1.fna
2 D1_FST2410 Den1 IX002 0 Analysis/PipelineRuns/2013_04_19/Ref/Den1 D1_FST2410 NGSData/RawData/IonTorrent/2013_04_19/meta/Primer/Den1.fna
```

## 5 Libraries

This section lists all of the libraries that make up pyWrairLib

Each library may contain scripts as well which are listed with their usage

### 5.1 wrairdata

#### 5.1.1 sanger\_to\_fastq

Convert .ab1 files into various formats(by default fastq) This script is under development and should be considered as such That being said, it seems to work fine giving it the -d and -t options together

##### Usage

```
usage: sanger_to_fastq [-h] [-s SANGERFILE] [-d SANGERDIR] [-o OUTPUTFILE]
                        [-t OUTPUTTYPE]
```

Convert sanger .ab1 file to another type

##### optional arguments:

```
-h, --help                show this help message and exit
-s SANGERFILE, --sanger-file SANGERFILE
                        Sanger file to convert
-d SANGERDIR, --sanger-dir SANGERDIR
                        Directory containing sanger files
-o OUTPUTFILE, --output-file OUTPUTFILE
                        The output filename. Omit an extension if you use the
                        -t fasta+qual option.
-t OUTPUTTYPE, --output-type OUTPUTTYPE
                        The output file type. Check out
                        http://biopython.org/wiki/SeqIO for options. Use
                        fasta+qual to get both fasta and qual files
```

#### 5.1.2 demultiplex

While there are many options for this command, only the -r and -s options are typically used.

- -r Specifies the location of the runfile so it knows how to map sample names with what barcode/region they are from
- -s Specifies the location of the sff directory that contains the multiplexed sff files to demultiplex

##### Usage

```
usage: demultiplex [-h] [-d PROCDIR] [-r RUNFILE] [-s SFFDIR] [-o OUTPUTDIR]
                  [--mcf MIDPARSEFILE] [--sfffilecmd SFFFILECMD] [--rename]
                  [--demultiplex]
```

##### optional arguments:

```
-h, --help                show this help message and exit
-d PROCDIR, --image-processing-dir PROCDIR
                        Image processing directory path
-r RUNFILE, --runfile RUNFILE
                        Path to the Runfile
-s SFFDIR, --sff-dir SFFDIR
                        Path to directory containing sff files
-o OUTPUTDIR, --output-dir OUTPUTDIR
                        Output directory path[Default: demultiplexed/]
--mcf MIDPARSEFILE        Midkey config parse file[Default:
                        NGSDData/MidParse.conf]
--sfffilecmd SFFFILECMD   Path to sfffile command[Default:
                        bin/sfffile]
--rename                  Only rename already demultiplexed sff files
--demultiplex              Only demultiplex. Don't rename
```

- At this time the -d option is in testing and unstable

#### 5.1.3 link\_reads

link\_reads is a simple helper script to link NGS reads to the NGSDData's ReadsBySample directory link\_reads by default looks to see if there is a demultiplexed directory in the current directory you are in otherwise defaults to just the current directory you are in or you can manually set the directory by using the -d option

Typically link\_reads is run right after running demultiplex



## Usage

```
usage: link_reads [-h] [-d INPUTDIR] [-c CONFIGPATH]
```

### optional arguments:

```
-h, --help            show this help message and exit
-d INPUTDIR, --demultiplexed-dir INPUTDIR
                        Directory containing read data [Default:
                        demultiplexed]
-c CONFIGPATH, --config CONFIGPATH
                        Config file to use
```

## 5.2 wrairlib

## 5.3 wrairnaming

## 5.4 wrairanalysis

### 5.4.1 mapSamples.py

mapSamples.py is used to map samples to a reference genome using the Roche Newbler mapper. It derives all of its parameters from a RunFile. It will create GsMapper project directories inside of the directory that you execute it from. It is usually best to create a directory to run this command from within.

## Usage

```
usage: mapSamples.py [-h] [-c CONFIGPATH] runfile
```

### positional arguments:

```
runfile                Runfile path to use
```

### optional arguments:

```
-h, --help            show this help message and exit
-c CONFIGPATH, --config CONFIGPATH
                        Config file to use
```

### 5.4.2 genSummary.sh

This is a wrapper shell script that runs post mapping summary scripts. It doesn't generate any output or output files itself, but runs scripts that do generate output. It runs all scripts on all GsMapper directories found in the current directory it is executed from. It is easy to run after mapSamples.py is run since mapSamples.py creates GsMapper projects in a directory and then you can just run genSummary.sh after that to get a more compiled look at all of the projects status **Currently runs:**

- Gap analysis
  - Compiles the output of NGSCoverage's gapsformids.py into images that are separated by each virus's segment inside of Gaps/Segments
- genallcontigs.py
- mapSummary.py
- SequenceExtraction's Summary -table

## Usage

```
genSummary.sh
```

*You must run this from within the same directory that mapSamples.py was run*

### 5.4.3 mapSummary.py

Given a directory containing gsMapper projects, this script will find all 454RefStatus.txt files and compiles a single Excel spreadsheet from them for all references used across all projects. You can also specify a reference file to 'target' so that only the references inside that file will be listed in the output excel file.

## Usage

```
mapSummary.py -d <projectdir> [-r <reference>] [-o <outputfile>]
```

- projectdir can be any directory that directly contains GsMapper projects. Usually this directory is the same directory that mapSamples.py was executed from.
- reference is optional and should be one of the reference files that was used in the mapping. This basically filters out all references so that only the reference you specify will be listed in the output
- outputfile is optional and specifies where to write the resulting Excel file. The default is in the current directory with the name AllRefStatus.xls

### 5.4.4 genallcontigs.py

Searches inside a given directory for any gsMapper directories. For every gsMapper directory found, it gathers all contigs from the 454AllContigs.fna and writes them to a file named after the project directory inside of the given output directory. The output is essentially a single directory containing Merged 454AllContigs.fna and .qual files into a .fastq file for each gsMapper directory

## Usage

```
genallcontigs.py -d <projectdir> [-o <outputdir>]
```

- projectdir can be any directory that directly contains GsMapper projects. Usually this directory is the same directory that mapSamples.py was executed from.
- outputdir is optional and defines what directory to place the results in. By default it is FastaContigs inside the current directory.

### 5.4.5 allcontig\_to\_allsample.py

Given a single gsMapper project directory, merges the 454AllContigs.fna and 454AllContigs.qual files into a single .fastq file. Output is by default sent to STDOUT(screen) but can be set to a file by using the -o option

## Usage

```
allcontig_to_allsample.py -p <gsproject> [-o <outputfile>]
```

- gsproject is the path to a single Gs Project
- outputfile is optional and should be a file to write the resulting fastq to. By default it is written to the terminal

### 5.4.6 variant\_lookup

Variant lookup looks up a position in a GsMapper alignment and displays the information about that variant. You just have to provide the nucleotide position and optionally a unique portion of the reference you are looking for.

If you do not provide the reference it will display all references for that alignment

## Usage

```
usage: variant_lookup [-h] [-d PDPATH] variant_pos [ref_name]
```

### positional arguments:

variant_pos	The variant position to display info for
ref_name	The identifier to limit the info for. Default is to show all identifiers

### optional arguments:

-h, --help	show this help message and exit
-d PDPATH, --project_directory PDPATH	GsProject directory path [Default: Current working directory]

- variant\_pos is the Nucleotide position of the alignment to look for the information for
- ref\_name only has to be a portion of the reference you are looking for. This is simply a filter for all reference names in the alignment.

Example:

If the alignment has 3 references

Reference\_ABCD

Reference\_ACDE

Reference\_1

- Specifying Reference as the ref\_name argument would yield all 3 references
  - Specifying ABCD would only yield Reference\_ABCD
  - Specifying Reference\_A would yield Reference\_ABCD and Reference\_ACDE
- project\_directory is only used if your current working directory is not a GsMapper project. It is easiest to invoke variant\_lookup by first changing directory to the GsMapper project you are wanting to work on.