

# **VEuPathDB User Documentation**

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Date	Version /release	Description/Change log
September 08, 2021	Release 54	<p>Changes relative to last submission</p> <ul style="list-style-type: none"><li>The Apollo sandbox link has been updated to remove broken links (page 11).</li><li>A new page of public datasets planned for integration has been added and will be regularly updated. This is noted on page 14 and the page is available here <a href="https://veupathdb.org/veupathdb/app/static-content/datalnprogress.html">https://veupathdb.org/veupathdb/app/static-content/datalnprogress.html</a></li><li>The 'Frequently Asked Questions' section has been reorganized into categories (page 21).</li><li>A new section, 'What can I do if a gene model is incorrect or missing?' has been added to Frequently Asked Questions (page 29).</li></ul>

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# Introduction

This material is made available to our users and NIH administrators to help them use VEuPathDB resources and understand the underlying tools, experiments and analyses provided by this Bioinformatics Resource Center funded in part by the US National Institute of Allergy and Infectious Diseases (Contract HHSN75N93019C00077). Note that the content of this document is also provided through the website, often in a context dependent manner. The web links are provided in the appropriate places throughout.

This report summarizes the categories of user documentation and provides a link to the landing page for each. The documentation can be divided into four broad categories: Site usage; Analysis methods; Dataset descriptors and technical infrastructure and software documentation.

This report is available from all VEuPathDB sites, e.g. <https://veupathdb.org/>, from the Help menu.

## About VEuPathDB.org

The Eukaryotic Pathogen, Vector and Host Informatics Resource (VEuPathDB) is one of two [Bioinformatics Resource Centers \(BRCs\)](#) funded by the US National Institute of Allergy and Infectious Diseases (NIAID), with additional support from the Wellcome Trust (UK).

These resources stem from support initially provided for the Plasmodium Genome Database by the Burroughs Wellcome Fund (2000-2) and a research grant from NIAID (2002-6). The BRC program was initiated in 2004 to provide public access to computational platforms and analysis tools enabling collection, management, integration and mining of genomic information and other large-scale datasets relevant to infectious disease pathogens including their interaction with mammalian hosts and invertebrate vectors of disease. Two BRCs are currently funded:

- VEuPathDB focuses on eukaryotic pathogens and invertebrate vectors of infectious diseases, encompassing data from prior BRCs devoted to parasitic species (EuPathDB), fungi (FungiDB) and vector species (VectorBase).
- BV-BRC - [PATRIC](#) & [VIPR](#) focus on bacterial and viral pathogens.

VEuPathDB has also received funding from the Wellcome Trust (UK), Bill & Melinda Gates Foundation, and US Department of Agriculture to support informatics efforts focusing on kinetoplastida and fungal organisms with special emphasis on improving functional annotation for select genome sequences and families of genes.

VEuPathDB provides access to diverse genomic and other large scale datasets related to eukaryotic pathogens and invertebrate vectors of disease (see [Genome Info and Stats](#)). Organisms supported by this resource include (but are not limited to) the NIAID list of [emerging and re-emerging infectious diseases](#).

Component web sites are constructed using a common infrastructure and standard data analysis and loading procedures, allowing the use of [VEuPathDB](#) as a single point of entry for each (or all) of these community resources, and the opportunity to leverage orthology for searches across taxa.

- [Organisms - Genome Info and Stats](#) provides a list of all organisms available in this website.
- [Data Sets](#) provides a list of all information in this website integrated into VEuPathDB, with relevant references.

#### Current Funding

The Eukaryotic Pathogen, Vector and Host Informatics Resources (VEuPathDB) is funded by the National Institute of Allergy and Infectious Diseases (NIH/DHHS) under Contract No. NIH HHS 75N93019C00077.

VEuPathDB also receives funding from the Wellcome Trust (UK) to support informatics efforts focusing on kinetoplastida and fungal organisms with special emphasis on improving functional annotation of genomes. Grant numbers: 212929/Z/18/Z and 218288/Z/19/Z.

## About OrthoMCL

### Current Release

See [news about the Current Release](#).

To see the current set of organisms as well as their proteome sources and orthology statistics, go to [Proteome Sources and Statistics](#).

**Downloads:** Go to the [download site](#) to obtain the protein sequences and ortholog groups used in this release.

### Method for Forming and Expanding Ortholog Groups

Proteins are placed into Ortholog Groups by the following steps:

1. The OrthoMCL algorithm (see below) is employed on proteins from a set of 150 Core species to form Core ortholog groups. These species were carefully chosen based on proteome quality and widespread placement across the tree of life. Each Core protein is placed by the algorithm into a Core ortholog group consisting of one or more proteins. Core group names have the format OG6\_xxxxxx (e.g., OG6\_101327). OG6 refers to OrthoMCL release 6; for each sub-release (e.g., 6.1, 6.2, etc), the Core species and the Core ortholog group names will remain constant.
2. The proteins from hundreds of additional organisms, termed Peripheral organisms, are mapped into the Core groups. To do this, NCBI BLASTP is used to compare each Peripheral protein to each Core protein in the Core groups. (Note that Peripheral proteins that were previously added to the Core group are NOT used in the BLASTP.) Then, each Peripheral protein is assigned to the Core group containing the Core protein with the best BLAST score, but only if the E-Value is <1e-5 and the percent match length is >=50%.
3. All Peripheral proteins that fail to map to a Core group are collected and subjected to independent OrthoMCL analysis, forming Residual groups consisting of one or more proteins. Residual group names have the format OG6r1\_xxxxxx (e.g., OG6r1\_101327), where OG6 refers to release 6 and r1 refers to sub-release 1.
4. For each subsequent sub-release (which will occur every ~3 months along with other VEuPathDB sites), proteomes from additional Peripheral organisms will be processed as in steps 2 and 3 above. However, step 3 will differ slightly because the previous set of

Residual groups will be disassembled, leaving the previous unmapped Peripheral proteins to be combined with the new unmapped Peripheral proteins. All of these proteins will be used to form new Residual groups (e.g., OG6r2\_xxxxx).

5. During a sub-release, the proteomes of some species will be updated to the latest version. This can be easily done for a Peripheral species: the old set of proteins are removed from ortholog groups and then the new set is mapped into groups as above. However, this is not possible for Core species because these proteins are used to define Core groups. Thus, the Core species with the older proteome remains on the site but is superficially retired by appending its abbreviation with -old (e.g., aaeg becomes aaeg-old). Then, the latest version of the proteome is mapped in as a peripheral species and obtains the original species abbreviation (e.g., aaeg is a peripheral with a more recent proteome than aaeg-old). These retired species will be eliminated fully when a new set of Core species is defined, as described in the next point.
6. On occasion, the set of Core species will be re-defined, as more appropriate proteomes become available and/or when a large number of Core species are retired. In this case, new Core groups (e.g., OG7\_xxxxx) and Residual groups (e.g., OG7r1\_xxxxx) will be formed from the latest version of proteomes from a carefully-chosen set of core species.

This design allows for the addition of proteomes at every sub-release (e.g., 6.1, 6.2, etc). Note that Core groups (e.g., OG6\_101327) will remain between sub-releases, though these groups will expand as Peripheral proteins are mapped in. In contrast, Residual groups will exist only for that sub-release; thus, Residual groups are useful in allowing the user to find proteins related to their protein(s) of interest, but are not stable groups.

## The OrthoMCL Algorithm

See the [OrthoMCL Algorithm Document](#) for a detailed description of the OrthoMCL algorithm.

In overview:

- All-v-all BLASTP of the proteins.
- Compute *percent match length*
  - Select whichever is shorter, the query or subject sequence. Call that sequence S.
  - Count all amino acids in S that participate in any HSP.
  - Divide that count by the length of S and multiply by 100
- Apply thresholds to blast result. Keep matches with E-Value < 1e-5 percent match length >= 50%
- Find potential inparalog, ortholog and co-ortholog *pairs* using the Orthomcl Pairs program. (These are the pairs that are counted to form the *Average % Connectivity* statistic per group.)
- Use the [MCL](#) program to cluster the pairs into groups

## Background on Orthology and Prediction

Orthologs are homologs separated by speciation events. Paralogs are homologs separated by duplication events. Detection of orthologs is becoming much more important with the rapid progress in genome sequencing ([Glover et al. 2019](#)).

OrthoMCL is a genome-scale algorithm for grouping orthologous protein sequences. It provides not only groups shared by two or more species/genomes, but also groups representing species-specific gene expansion families. Thus, it serves as an important utility for automated eukaryotic

genome annotation. OrthoMCL starts with reciprocal best hits within each genome as potential in-paralog/recent paralog pairs and reciprocal best hits across any two genomes as potential ortholog pairs. Related proteins are interlinked in a similarity graph. Then, MCL (Markov Clustering algorithm; [Dongen 2000](#); [www.micans.org/mcl](#)) is invoked to split mega-clusters. This process is analogous to the manual review in COG construction. MCL clustering is based on weights between each pair of proteins, so to correct for differences in evolutionary distance the weights are normalized before running MCL.

OrthoMCL is similar to the INPARANOID algorithm ([Remm et al. 2001](#)) but is extended to cluster orthologs from multiple species. OrthoMCL clusters are coherent with groups identified by EGO ([Lee et al. 2002](#)), and an analysis using EC number suggests a high degree of reliability ([Li et al. 2003](#)).

We evaluated the performance of seven widely-used orthology detection algorithms that use three general prediction strategies: phylogeny-based, evolutionary distance-based and BLAST-based ([Chen, et al. 2007](#)). Specifically, we used Latent Class Analysis (LCA), a statistical technique appropriate for testing large data sets when no gold standard is available. Our results show an overall trade-off between sensitivity and specificity among these algorithms, with INPARANOID and OrthoMCL performing best with False Positive (FP) and False Negative (FN) error rates lower than 20%.

## Software

OrthoMCL was originally implemented by Li Li. The software was not available for download.

[Version 1.4](#) was developed as publicly available software by Feng Chen (This version is now not supported).

[Version 2.0](#) was re-engineered to handle large-scale datasets (hundreds of genomes) by Steve Fischer, Mark Heiges, John Iodice, and Ryan Thibodeau.

## Data Access Policy

All data on these websites are provided freely for public use, through the contributions of many researchers involved in generating genome sequences, functional genomics datasets, and additional information. These data often derive from ongoing research and are not guaranteed to be accurate. When using data obtained from VEuPathDB, it is important to cite the original publications and contributors. Please see [Citing VEuPathDB](#).

## Citing VEuPathDB in Publications and Presentations

If you use a VEuPathDB resource, we invite you to please cite the most relevant publication. This [PubMed filter](#) provides a list of the most recent VEuPathDB publications.

Please note that much of the data in VEuPathDB is provided by independent researchers, please cite them if you use their data. See [Data Sets](#) for a list of all information integrated into VEuPathDB, and related publications.

For acknowledgements in presentations, you may wish to use one or more of the following logos:



Additional resources leveraging the same infrastructure: [MicrobiomeDB](#) and [ClinEpiDB](#).

## Community Interactions and Data Submission Policies

VEuPathDB serves a global scientific community that demands direct active support and community involvement. VEuPathDB outreach activities include:

- Organizing and running hands on training workshops and webinars ([Google Map](#)).
- Developing educational material in the form of [exercises](#) and [online tutorials](#).
- Responding to support emails for users who contact us directly by clicking the "Contact Us" links in the header or footer of any VEuPathDB webpage (average response time is 48 hours).
- Holding open community meetings/forums with our diverse user base. These meetings are held in person at scientific conferences or using an online conferencing platform.
- Attending national and international meetings with active participation in the form of posters, presentations or help desks.
- Authoring [peer reviewed manuscripts](#).
- Maintaining active social media presence in the form of a [FaceBook page](#) and [Twitter feed](#).
- Providing a clear [data handling and release policy](#) to investigators to encourage submission of data prepublication.

## How to Submit Data to VEuPathDB

The Eukaryotic Pathogen, Vector & Host Informatics Resources (<https://VEuPathDB.org>) is a Bioinformatics Resource Center (BRC) operated under contract from the US National Institute of Allergy and Infectious Diseases (NIAID) and the Wellcome Trust. VEuPathDB is charged with ensuring that genomic (and other large-scale) datasets pertaining to supported pathogens are conveniently accessible to the worldwide community of biomedical researchers. This document summarizes policies associated with releasing datasets on VEuPathDB. Our goal is to help the communities that we serve ensure that their data are FAIR, Findable, Accessible, Interoperable and Reusable.

VEuPathDB welcomes submissions of genomic-scale data concerning eukaryotic microbes, fungi, vectors of human disease, and host-pathogen interactions. The VEuPathDB contract from NIAID provides support for biosecurity pathogens, including *Babesia*, *Cryptosporidium*, *Entamoeba*, *Giardia*, *Microsporidia* (various genera), *Toxoplasma*, *Plasmodium*, and related taxa (*Acanthamoeba*, *Gregarina*, *Neospora*, *Theileria*) and also arthropod vectors (ticks, mites, mosquitoes, kissing bugs, tsetse flies, sand flies, lice, etc.) of human disease, as well as a snail that serves as an intermediate host, and comparator species. Support for kinetoplastid parasites (*Crithidia*, *Endotrypanum*, *Leishmania*, *Trypanosoma*) is provided by The Wellcome Trust. The FungiDB project encompasses a large (and growing) number of species supported by both NIAID and the Wellcome Trust. Please [contact us](#) if you have data from other species that should be incorporated into VEuPathDB! Please review our [Data Submission Policy](#).

Our most common data types include transcriptomics, proteomics, metabolomics, epigenomics, population-level and isolate information. In one form or another, VEuPathDB currently represents datasets in the following categories:

- Sequence (genomic [nuclear and organellar])
- ESTs and RNA-seq, generated on various platforms)
- Host-response data
- Comparative genomic information
- DNA polymorphism and population genetics data
- Sequences and metadata pertaining to field and clinical isolates and collections (with geo-spatiotemporal and other metadata)
- Chromatin modification data (ChIP-chip and ChIP-seq)
- Manually curated and automatically generated gene models and other annotation (GO terms, InterPro domains, etc.)
- Transcript and proteomic profiling
- Host response data sets (multiple platforms)
- Interactome data
- Protein structural information
- Metabolic pathways and metabolomics data
- Phenotype information, reagents (clones, antibodies, etc.)
- Publication references
- Image data, etc.

We also accept other genomic-scale data and are open to suggestions. Use the [Contact Us](#) link to make suggestions. We look forward to working with you!

## How to submit data for integration in VEuPathDB

To submit your data for integration, fill out the appropriate VEuPathDB Dataset nomination form listed below. If your data cannot be submitted via our forms, use the [Contact Us](#) link to send a brief description (two or three sentences) of your data.

Genomes & high throughput sequencing data (e.g. RNA-Seq, ChIP-Seq, isolates typed by Whole Genome Sequencing or by sequencing limited genetic loci) must be available in The International Nucleotide Sequence Database Collaboration (INSDC) such as NCBI GenBank, EMBL-EBI ENA or DDBJ.

Once the dataset is prioritized for loading, we will export the data directly from INSDC. Note: while genome sequences must be available in INSDC, functional annotation (e.g. gene names, GO terms, etc.) can be submitted directly to VEuPathDB.

Tell us about your data as early as possible, to allow ample time for scheduling into VEuPathDB release cycles. Depending on the dataset type, we can provide instructions on how to transfer your data to us (e.g., formats of proteomics datasets may differ depending on the nature and scale of the data to be transferred), or we may be able to facilitate data submission to a repository (e.g. GenBank, GEO/ArrayExpress, etc.).

For the gene manual annotations, the change of exon-intron boundaries and creation of new genes can be submitted via Apollo for all VectorBase genomes and some of the other VEuPathDB organisms too. The gene metadata (gene names/symbols and functional description) can be submitted by sending a spreadsheet file, follow this link for information about the 12 columns heading that are required. Before you submit a manuscript for publication, please inquire if we can generate new GeneIDs that can be linked out to within the publication; this process can take a few months, we strongly encourage you to please contact us as early as possible to allow ample time for us to process your data. Gene deletions are not handled via Apollo, please Contact Us with supporting evidence.

### Genome Sequence and/or Annotation

Genomes must be available in The International Nucleotide Sequence Database Collaboration (INSDC) such as NCBI GenBank, EMBL-EBI ENA or DDBJ.

- If your genome is uploaded to a repository, complete the [Genome Sequence and/or Annotation Description Form](#) making sure to include the accession numbers of your data when prompted. We will download your data from the repository.
- If the annotation file **is not** uploaded to a repository, use the [Contact Us](#) form to send us the genome annotation file only (e.g. gff file format).

### Structural and Functional Community Curation in Apollo

Welcome to the VEuPathDB Apollo service ([Dunn et al. 2019](#)), a real time collaborative genome annotation and curation platform. Use Apollo to integrate new or update current structural and functional data for gene models in the organisms available in VEuPathDB. Organisms in AmoebaDB, CryptoDB, FungiDB, GiardiaDB, MicrosporidiaDB, PiroplasmaDB, PlasmoDB, ToxoDB, TriTrypDB, & VectorBase are available for community curation.

We are currently in the process of a long term project to update our genome annotation which will mean some organisms are currently unavailable in Apollo. However, you can request these organisms to be available for curation by emailing our team at [help@veupathdb.org](mailto:help@veupathdb.org) for more information.

Apollo help and documentation:

- A “play” Apollo site, e.g., a [sandbox \(Login required\)](#) is available for you to experiment and get familiar with all Apollo menus, tools, and tracks before you decide to use it for your real gene manual annotations. These changes will not affect any of the organism's official gene set, neither will be preserved.
- A comprehensive webinar recording to learn [how to use Apollo](#) (57:40 min)
- A description of [Quick commands](#) (Apollo now has the BLAT tool!)
- [Functional annotation tutorial](#)
- [Structural annotation tutorial](#)
- [About Apollo \(Login required\)](#)
- [User Guide](#)
- [Request feature/Report a bug \(Login required\)](#)
- [Powered by JBrowse](#)
- [Web Service API \(Login required\)](#)

## High Throughput or Next Generation Sequencing – RNA, DNA or ChIP Sequencing

We prefer to download the raw read data in FASTQ format from a sequence read archive. We integrate the data into the database using the raw reads and use the raw reads during future database releases to remap or update our analyses when necessary.

How to transfer a copy of your data to VEuPathDB:

- Upload your data to a sequence read archive such as the European Nucleotide Archive or NCBI's Sequence Read Archive and provide us the accession numbers. We will export the data directly from INSDC
- Complete the appropriate data description form making sure to enter your data archive accession numbers when prompted

[RNA-Seq Data Description Form](#)

[DNA-Seq Data Description Form](#)

[ChIP-Sequencing Data Description Form](#)

## Microarray

Transfer a copy of your data to VEuPathDB using one of these options:

- Upload your data to a repository such as GeneExpression Omnibus. Complete the data description form linked below making sure to enter your data archive accession numbers when prompted. We will export the data directly from a repository.

- Send your data as an attachment to an email. Use the [Contact Us](#) form to send us an email. Files (e.g. CEL, CSV) must include expression levels and probe set information. Pay special attention to clearly indicate the identity of columns in the data files you transferred to VEuPathDB.

### [Microarray Data Description Form](#)

## Proteomics

Excel or tab delimited text files are preferred. We can accommodate xml file format. Required columns include gene IDs, peptide sequences, peptide counts and scores.

How to transfer a copy of your data to VEuPathDB:

- Send your data as an attachment to an email. Use the [Contact Us](#) form to send us an email.
- Complete the [Proteomics Data Description Form](#) making sure to clearly indicate the content of each column in your file.

## Quantitative Proteomics

Excel or tab delimited files are preferred. We can accommodate xml file format. Required columns include gene IDs and scores.

How to transfer a copy of your data to VEuPathDB:

- Send your data as an attachment to an email. Use the [Contact Us](#) form to send us an email.
- Complete the [Quantitative Proteomics Data Description Form](#) making sure to include a description of data columns, for example, time course units and arrangement if not apparent from column headers.

## ChIP-chip

Your data files should include expression levels and probe set information.

Transfer a copy of your data to VEuPathDB using one of these options:

- Upload your data to a repository such as Gene Expression Omnibus and complete the [ChIP-chip Data Description Form](#) making sure to enter the archive accession numbers (if any) for your data when prompted.
- Send your data as an attachment to an email. Use the [Contact Us](#) form to send us an email.

## Isolates typed by sequencing limited genetic loci

If your data **ARE already** uploaded to the GenBank, use the [Contact Us](#) form to tell us about your data. Note: GenBank Isolate records and the associated metadata are automatically

updated with each VEuPathDB release. There is no need to complete our Isolate Submission Form.

If your data **ARE NOT** already uploaded to the GenBank, we can facilitate this upload. Complete the Isolate Submission Form linked below and we will use the information to generate a GenBank submission for your isolates. The new isolate records will be downloaded to VEuPathDB when the records are released by the GenBank. Use the [Contact Us](#) form to send us instructions for retrieving your data.

[Isolate Submission Form](#)

[Help for submitting Isolate Data](#)

## Isolates or Strains typed by High-Throughput Sequencing

We prefer to receive the raw read data in FASTQ or FASTA file format. We integrate your data into the database using the raw reads. We also use the raw reads during future database releases to remap your data when the reference genome is reloaded and to update our analyses when needed.

How to transfer a copy of your data to VEuPathDB:

- Upload your data to a sequence read archive such as DNA Data Bank of Japan, the European Nucleotide Archive or NCBI's Sequence Read Archive. We will retrieve your data using the read archive's accession numbers for your data set.
- Complete our [DNA Seq Data Description Form](#) making sure to enter the read archive accession numbers for your data when prompted. We also ask that you complete an abbreviated [Abbreviated Isolate Submission Form](#) to describe meta data associated with your isolates.

## Bulk user comments form

- Instructions: download the form, fill out required fields and provide any additional information, if available, and send it back to the email listed in the form
- Download the [Bulk user comments form](#)

## Phenotype collection form

- Instructions: download the form, fill out required fields and provide any additional information, if available, and send it back to the email listed in the form
- Download the [Phenotype collection form](#)
- Download the [Parasite phenotype collection form](#)

**General Data Submission** – Use the [Contact Us](#) form to tell us about data that does not fit any of the above categories

## Datasets we are working on loading

- Below is a list of datasets that VEuPathDB is working on integrating into future releases. Note that this list is fluid and will be updated frequently.
- Once a dataset is released it will be removed from this list. To view all loaded datasets please explore the [datasets in VEuPathDB page](#).
- Let us know about missing datasets by using the [Contact Us](#) form or through the “[How to submit data](#)” section of VEuPathDB resources.

**\*Note - the list of planned datasets is not included in this report but can be accessed on veupathdb.org under the ‘About’ menu and at this direct link:**

<https://veupathdb.org/veupathdb/app/static-content/dataInprogress.html>

## Data submission and release on VEuPathDB databases

issued February 2010, most recent revision 02 April 2020



The Eukaryotic Pathogen, Vector & Host Informatics Resources (<http://VEuPathDB.org>) is a Bioinformatics Resource Center (BRC) operated under contract from the US National Institute of Allergy and Infectious Diseases (NIAID) and the Wellcome Trust. VEuPathDB is charged with ensuring that genomic (and other large-scale) datasets pertaining to supported pathogens are conveniently accessible to the worldwide community of biomedical researchers. This document summarizes policies associated with releasing datasets on VEuPathDB and affiliated knowledgebases. Our goal is to help the communities that we serve ensure that their data are FAIR, Findable, Accessible, Interoperable and Reusable.

### General principles:

- **Data providers define the schedule for data release (in consultation with funders, publishers, etc).** While there is no point in providing VEuPathDB with data that will never become public, deposition does not in itself authorize immediate release. Data become accessible to the public only when the data providers and VEuPathDB staff agree that it is accurately represented and ready to go live. Note that knowledgebase staff are not active research scientists; they are distinct from researchers in the groups responsible for VEuPathDB, who see new data only when it becomes accessible to the general public.
- **Data providers know their data best.** We expect to work with those who generated the underlying data to determine how best to analyze and represent new data types. This typically means taking in relatively raw data – often earlier, and in a more unprocessed form than the published dataset – and building an in-house analysis pipeline to ensure that all comparable datasets are handled similarly.
- **The earlier we learn about new datasets, the easier it is to schedule timely release.** The nature of our knowledgebase production, and competing demands from the many communities we support, means that several months' notice are often required to prepare for release. Note that it is often possible to use a preliminary dataset for planning, which can be swapped for the final version before public release.

- ***Experience has shown that data not deposited prior to publication often fails to emerge at all!*** After publication, it may be difficult to focus on tracking down the raw data, associated metadata, analysis methods, etc. It is never too early to discuss planned datasets with the VEuPathDB team!
- ***While not required, pre-publication data release often results in favorable attention from scientific colleagues (including journal editors and grant reviewers).*** Note that all major scientific journals now agree that early release of genomic-scale datasets does not compromise publication.

## Why submit my data to VEuPathDB?

- Inclusion in VEuPathDB facilitates your own analysis of the data, in the context of other genomic-scale experiments already available from researchers around the world.
- Electronic access permits others to analyze your data in greater depth than possible in print (even in advance of publication, if you wish to allow this).
- Availability within VEuPathDB keeps your data alive on a highly visible genomics knowledgebase resource: VEuPathDB is accessed by ~13,000 unique users each month.

## How do I submit data to VEuPathDB?

- Fill out the appropriate form to indicate the data availability
- Contact the VEuPathDB by clicking the 'Contact Us' link on any VEuPathDB page or emailing us at [help@VEuPathDB.org](mailto:help@VEuPathDB.org).
- Tell us about your data as early as possible, to allow ample time for scheduling into VEuPathDB release cycles.
- Once you tell us about your data, we will provide instructions on how to transfer your data to us (formats may differ depending on the nature and scale of the data to be transferred).
- In order to avoid any confusion and ensure accuracy, we adhere to strict Standard Operating Procedures (SOPs), as outlined below.

## What data types are supported by VEuPathDB?

In one form or another, VEuPathDB currently represents sequence (genomic [nuclear and organellar], ESTs and RNA-seq, generated on various platforms), host-response data, comparative genomic information, DNA polymorphism and population genomics data, sequences and metadata pertaining to field and clinical isolates and collections (with geo-spatiotemporal and other metadata), chromatin modification data (ChIP-chip and ChIP-seq), manually curated and automatically generated gene models and other annotation (GO terms, InterPro domains, etc.), transcript and proteomic profiling, host response data sets (multiple platforms), interactome data, protein structural information, metabolic pathways and metabolomics data, phenotype information, reagents (clones, antibodies, etc.), publication references, image data, etc. We can support additional data types as needed.

## Please let us know if you have data to provide that is not currently supported! What species are supported by VEuPathDB?

The VEuPathDB contract from NIAID provides support for biosecurity pathogens, including *Babesia*, *Cryptosporidium*, *Entamoeba*, *Giardia*, *Microsporidia* (various genera), *Toxoplasma*, *Plasmodium*, and related taxa (*Acanthamoeba*, *Gregarina*, *Neospora*, *Theileria*) and also

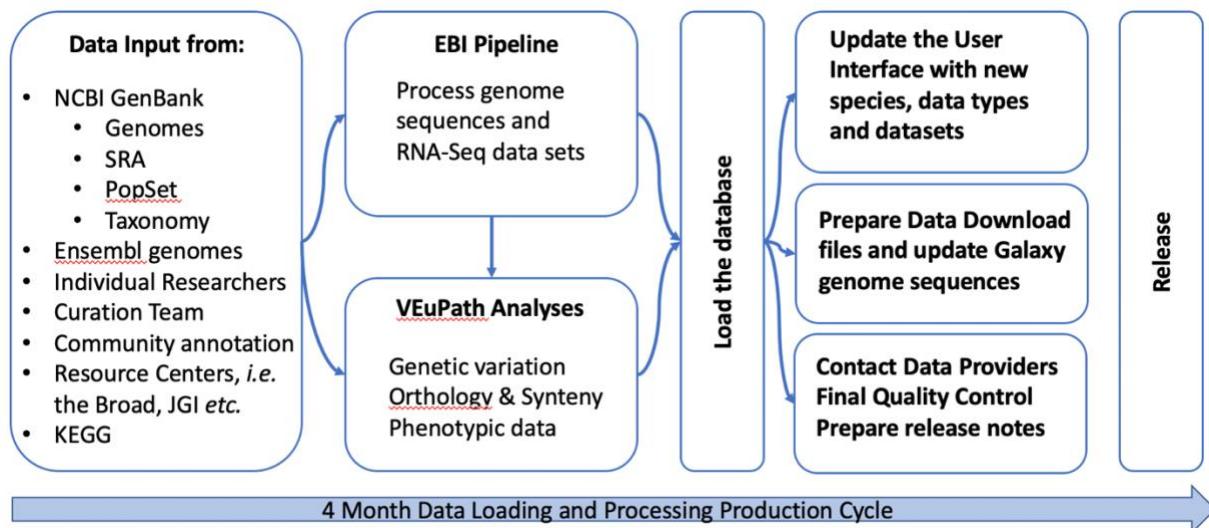
arthropod vectors (ticks, mites, mosquitoes, kissing bugs, tsetse flies, sand flies, lice, etc.) of human disease, as well as a sail that serves as an intermediate host, and comparator species. Support for kinetoplastid parasites (*Crithidia*, *Endotrypanum*, *Leishmania*, *Trypanosoma*) is provided by The Wellcome Trust. The FungiDB project encompasses a large (and growing) number of species supported by both NIAID and the Wellcome Trust. **Please contact us if you have data from other species that should be incorporated into VEuPathDB!**

## The Data Production Cycle

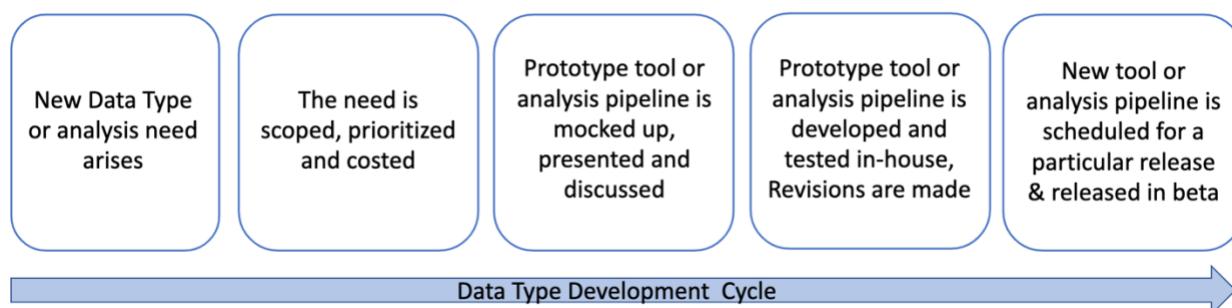
### Guiding Principle:

- We balance the need to process as many datasets as possible from known data types in our production pipeline with the need to scale in capacity and containerize analyses for these same data types and the need to continually build new tools and infrastructure in our data type development to accept new data types and facilitate emerging community analysis needs as they emerge.

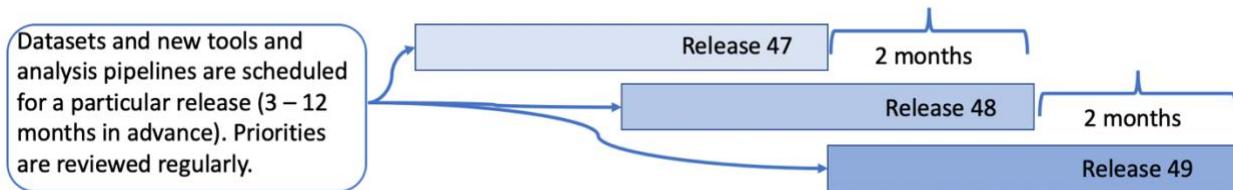
(A)



(B)



(C)



**High level views** of the VEuPathDB Production Cycle (A), Datatype Development Cycle (B), and Release Timeline (C; overlapping 4 month cycles of production and data type development with bimonthly releases).

## Data Management SOPs (Standard Operating Procedures) for VEuPathDB databases

VEuPathDB routinely handles datasets provided prior to publication, in addition to those already in the public domain. In order to ensure timely and accurate data integration we strictly adhere to the following Standard Operating Procedures (SOPs):

1. Datasets come to our attention in several ways, including
    - Direct contact from researchers generating the data (during the earliest stages of project design, as
      - data is being produced, in the course of data analysis, or in the context of manuscript preparation).
      - Information provided by members of the VEuPathDB or larger pathogen community.
      - Information obtained by VEuPathDB staff at meetings and conferences.
      - Publicly available information from the scientific literature, genomic dataset repositories, etc.
- \* Note that VEuPathDB can often facilitate data deposition in the appropriate archival repositories (GenBank, SRA, GEO/ArrayExpress, etc.)
2. **Decisions to include a dataset in VEuPathDB are based on value to the research community.** When prioritizing data for integration, we rely heavily on discussions with active researchers, including the scientific advisory committees established for each of the taxonomic groups supported by VEuPathDB and the objectives and priorities of our funders. **Please contact us if you are interested in participating in these discussions.**
  3. **Regardless of how we first learn about a given dataset, communication is established with the original data producer** through email, teleconference, and/or face-to-face meetings to discuss the desirability and feasibility of integration into VEuPathDB. In the course of these discussions, we consider what data are likely to be available, data formats and transfer protocols, questions the community may wish to ask of this data, and ways to represent or display such information. We also collect appropriate metadata (regarding samples, experimental protocols, etc.), and information on data sources, data providers, appropriate citation, etc.
  4. **Data provided to VEuPathDB is housed on secure servers and never shared outside of VEuPathDB staff without prior consent of the data provider.** Note that database staff are not active researchers; they are distinct from students and postdocs in the groups responsible for VEuPathDB, who see new datasets only when they become accessible to

the general public.

5. Datasets are assigned a provisional release date, in consultation with the data provider. ***Scheduling a dataset does not mean that it will be released without final examination and approval by the data provider!*** We operate on the assumption that those who generate the data are best placed to evaluate its proper integration and representation in the knowledgebase. Note that this ‘golden rule’ applies to both published and unpublished data.
6. Three to four months before the scheduled release date, the data provider is contacted by the ***Outreach*** team, to ensure that we have the most up-to-date version of the data, along with appropriate metadata and information on data sources and citations. The ***Data Loading*** team then processes and integrates this data into our internal knowledgebases.
7. After data loading is complete, the ***Data Development*** team begins to analyze and develop searches against the data. At this point we will likely communicate with the data provider, if questions arise.
8. ***Once data development is underway, the data provider is given access to a password-protected version of the VEuPathDB website containing their data.*** This development site is similar to the current production knowledgebase, except that it also includes new data from the provider. We also provide instruction on how to search and view these new data, including sample searches integrating new data with relevant information already available in the knowledgebase. Important questions to consider include:
  - Does the knowledgebase accurately represent your data?
  - Are the values and/or graphical displays provided appropriate?
  - Are the questions that one can ask of your data appropriate?
  - Are there additional questions that you would like to see implemented?
  - Are the data appropriately described, including relevant metadata and reference / citation details?
9. ***A series of exchanges typically ensues,*** in which we work iteratively with data providers to address any concerns, with changes reviewed on the password protected site so that providers can view and interrogate their data in the context of the rest of the database.
10. ***Public release is only considered after everyone is satisfied with how the data is represented.*** If the provider is not yet ready to authorize data public release, data is rescheduled for a future release, and removed from the development site before it goes live.
11. Once data are approved for public release, a description is included in the ‘News’ accompanying the next release, ***highlighting new datasets and functionality, and acknowledging all data providers.***
12. ***Post-release quality assurance*** provides the opportunity to modify displays and develop new queries if/as appropriate.

## How to Use Our Sites

We provide our users with a variety of mechanisms to learn about how the VEuPathDB site works and can facilitate their research. In addition to the instructions provided here, users

should know that they can learn about VEuPathDB in person, via recorded instructional material or via help located throughout the website, detailed in the list below.

## Online instructional material

- a. Users can request that a member of the VEuPathDB outreach staff attend a lab meeting or institutional or regional workshop to ask questions and have training in a specific topic
- b. User can participate in or view previously recorded webinars (online recordings of training sessions conducted by VEuPathDB staff) that are provided as YouTube recordings.
- c. Users can browse and download “quick start” materials and tutorials to follow at their own pace, each with color pictures and well explained prompts to help users explore the VEuPathDB site or one of its projects, e.g. VectorBase, FungiDB or PlasmoDB. All project use the same underlying architecture and software, only the local environment is customized so user can learn from a tutorial on any organism.
- d. VEuPathDB host several workshops annual and all training materials (step by step tutorials with exercises and answers) are provided for all to use and can be easily downloaded.
- e. Context sensitive help is available via the many help icons located on the web site, particularly on search pages.
- f. Textual descriptions are associated with search pages to provide information about the data type and how to use the search.

To quickly access these online resources, please visit:

<https://beta.veupathdb.org/veupathdb.beta/app/static-content/landing.html>

## BLAST<sup>beta</sup> (an update to our BLAST service to provide multi-sequence BLAST)

BLAST finds regions of similarity between sequences. The program compares nucleotide or protein sequences to sequence databases and calculates the statistical significance of matches. BLAST<sup>beta</sup> is VEuPathDB's updated BLAST tool that accommodates input as a single sequence or several FASTA-formatted sequences at once, e.g. multi-BLAST.

### Video tutorial

Our tool uses the [NCBI BLAST program](#) to find genes, genomic sequences, ESTs, or PopSet Sequences that have BLAST similarity to your input sequence. The tool accommodates up to 100 sequences of up to 100,000 amino acids or 1 million nucleotides. Each 'job' you submit is logged in your 'My Jobs' page and remains available indefinitely. Jobs run on multiple sequences include results in two formats:

- Combined Result - a nonredundant list of all sequences with similarity to your input sequence
- Individual Result - categorized lists of sequences with similarity to your input sequence. One list per input sequence.

All results can be downloaded in your choice of several formats, and individual results can be exported as a search strategy for further data mining in VEuPathDB.

## Frequently Asked Questions

### How to cite a reference

How to cite a genome assembly and/or gene set?

If you use any of the genomes from VEuPathDB genomic sites, when citing them in your work please provide their complete scientific name, strain, assembly, and gene set version, together with the most recent publication that corresponds, as shown in the FAQ “How to cite a VEuPathDB site?”

### Template

VectorBase, [vectorbase.org](http://vectorbase.org) (VectorBase most recent paper),  
<Species\_name><isolate/strain> <Assembly, Gene set>.

### Worked example

For example if you want to reference your usage of the *Anopheles gambiae* AgamP4.12 set of predictions you would use:

VectorBase, [www.vectorbase.org](http://www.vectorbase.org) (Giraldo-Calderón et al 2015), *Anopheles gambiae* PEST, AgamP4.13.

## How to cite a VEuPathDB site?

If you use VEuPathDB data, tools or resources, we invite you to please cite our most recent publication, for your database of interest:

Aurrecoechea, C., Barreto, A., Basenko, E. Y., Brestelli, J., Brunk, B. P., Cade, S., Crouch, K., Doherty, R., Falke, D., Fischer, S., Gajria, B., Harb, O. S., Heiges, M., Hertz-Fowler, C., Hu, S., Iodice, J., Kissinger, J. C., Lawrence, C., Li, W., Pinney, D. F., ... Zheng, J. (2017). **EuPathDB**: the eukaryotic pathogen genomics database resource. Nucleic acids research, 45(D1), D581–D591. <https://doi.org/10.1093/nar/gkw1105>

Basenko, E. Y., Pulman, J. A., Shanmugasundram, A., Harb, O. S., Crouch, K., Starns, D., Warrenfeltz, S., Aurrecoechea, C., Stoeckert, C. J., Jr, Kissinger, J. C., Roos, D. S., & Hertz-Fowler, C. (2018). **FungiDB**: An Integrated Bioinformatic Resource for Fungi and Oomycetes. Journal of fungi (Basel, Switzerland), 4(1), 39. <https://doi.org/10.3390/jof4010039>

Fischer, S., Brunk, B. P., Chen, F., Gao, X., Harb, O. S., Iodice, J. B., Shanmugam, D., Roos, D. S., & Stoeckert, C. J., Jr (2011). Using OrthoMCL to assign proteins to **OrthoMCL-DB** groups or to cluster proteomes into new ortholog groups. Current protocols in bioinformatics, Chapter 6, Unit–6.12.19. <https://doi.org/10.1002/0471250953.bi0612s35>

Giraldo-Calderón, G. I., Emrich, S. J., MacCallum, R. M., Maslen, G., Dialynas, E., Topalis, P., Ho, N., Gesing, S., VectorBase Consortium, Madey, G., Collins, F. H., & Lawson, D. (2015). **VectorBase**: an updated bioinformatics resource for invertebrate vectors and other organisms related with human diseases. Nucleic acids research, 43(Database issue), D707–D713. <https://doi.org/10.1093/nar/gku1117>

Oliveira, F. S., Brestelli, J., Cade, S., Zheng, J., Iodice, J., Fischer, S., Aurrecoechea, C., Kissinger, J. C., Brunk, B. P., Stoeckert, C. J., Jr, Fernandes, G. R., Roos, D. S., & Beiting, D. P. (2018). **MicrobiomeDB**: a systems biology platform for integrating, mining and analyzing microbiome experiments. Nucleic acids research, 46(D1), D684–D691. <https://doi.org/10.1093/nar/gkx1027>

Ruhamyankaka, E., Brunk, B. P., Dorsey, G., Harb, O. S., Helb, D. A., Judkins, J., Kissinger, J. C., Lindsay, B., Roos, D. S., San, E. J., Stoeckert, C. J., Zheng, J., & Tomko, S. S. (2020). **ClinEpiDB**: an open-access clinical epidemiology database resource encouraging online exploration of complex studies. Gates open research, 3, 1661. <https://doi.org/10.12688/gatesopenres.13087.2>

Older publications for each VEuPathDB site can be found [here](#).

How to reference a MapVEu (formerly PopBio) project ID, VBP\*?

Here is an example from one of our users:

**Data availability.**

Microsatellite and SNP genotypes were deposited in VectorBase<sup>48,49</sup> PopBio projects: VBP0000201 (new data), and VBP0000138, VBP0000176-177 (previously published data). Sequencing data were deposited in NCBI under accession numbers MF371160–MF371174 and MG241351–MG241354.

Gloria-Soria, A., Lima, A., Lovin, D. D., Cunningham, J. M., Severson, D. W., & Powell, J. R. (2018). Origin of a High-Latitude Population of *Aedes aegypti* in Washington, DC. *The American journal of tropical medicine and hygiene*, 98(2), 445–452. <https://doi.org/10.4269/ajtmh.17-0676>

## In silico experiments, data mining and hypothesis testing

How do I populate a strategy with my existing list of genes of interest?

1. From the home page click on “Genes” > “Annotation, curation, and identifiers” > “Gene ID(s)”
2. Paste in your existing list of genes or upload a text file
3. Click “Get Answer”

The screenshot shows the 'Search for...' interface. At the top, there's a search bar with 'expand all | collapse all' and a filter dropdown. Below it, a sidebar has 'Genes' expanded, showing options like 'Find a gene by its ID.', 'Function prediction', and 'Gene models'. A circled '1' is next to 'Find a gene by its ID.'. To the right, the main area is titled 'Identify Genes based on Gene ID(s)'. It has a 'Gene ID input set' section with a text input field containing 'AGAP004730, AGAP004731, AGAP004732'. A circled '2' is next to this input field. Below it are sections for 'Upload a text file' (with a note about file size), 'Copy from My Basket' (with a note about records copied), and 'Copy from My Strategy' (with a note about records in a strategy). A circled '3' is next to the 'Get Answer' button at the bottom.

How do I populate a strategy with all the genes in an organism?

First start a new strategy from the home page

1. From the home page click on “Genes” > “Taxonomy” > “Organism”
2. Filter for your organism of interest
3. Click “Get Answer”

The screenshot shows two main panels. The left panel is titled "Search for..." and lists categories like Genes, Taxonomy, and Organisms. A callout bubble labeled "1" highlights the "Organism" section, which contains a note: "Find all genes from one or more species/organism." The right panel is titled "Identify Genes based on Organism" and shows a taxonomy tree under the heading "Organism". A callout bubble labeled "2" highlights the "Diptera" node, specifically "Culex quinquefasciatus Johannesburg". A callout bubble labeled "3" highlights the "Get Answer" button at the bottom right of the taxonomy panel.

**Search for...**

expand all | collapse all

Filter the searches below...

- ▼ Genes
  - ▶ Annotation, curation and identifiers
  - ▶ Function prediction
  - ▶ Gene models
  - ▶ Genomic Location
  - ▶ Immunology
  - ▶ Orthology and synteny
  - ▶ Pathways and interactions
  - ▶ Protein features and properties
  - ▶ Protein targeting and localization
  - ▶ Sequence analysis
  - ▶ Structure analysis
- ▼ Taxonomy
- Organism**
- Organisms

**Identify Genes based on Organism**

Organism

Note: You must select at least 1 values for this parameter.  
1 selected, out of 43

select all | clear all | expand all | collapse all

Filter list below...

- ▼ Arthropoda
  - ▶ Arachnida
  - ▼ Insecta
    - ▼ Diptera
      - ▼ Culicidae
        - ▶ Aedes
        - ▶ Anopheles
        - ▶ Culex** (selected)
        - ▶ Glossinidae
        - ▶ Muscidae
        - ▶ Psychodidae
      - ▶ Hemiptera
      - ▼ Phthiraptera
        - ▶ Pediculus humanus USDA
    - ▶ Mollusca

select all | clear all | expand all | collapse all

**3**

Get Answer

How do I do an enrichment analysis on a list of genes by GO term or metabolic pathway?

Start with a list of genes using an existing strategy, a list of genes you submit (see above), or all the genes in a given organism (see above)

1. click “Analyze results” This will give you the option to run a variety of analyses on your list of genes See the following worksheet to learn more: [GO enrichment](#)

The screenshot shows the VEuPathDB Gene Results interface. At the top, it displays "1,029 Genes (970 ortholog groups)" and a "Revise this search" button. Below this is an "Organism Filter" section with options like "select all", "clear all", "expand all", "collapse all", and a search bar for "Search organisms...". A blue circle with the number "1" is drawn around the "Analyze Results" button, which is highlighted in blue. To the right of the button are navigation links for "Gene Results", "Genome View", and "New Analysis". There are also buttons for "Download", "Add to Basket", and "Add Columns". The main content area is titled "Analyze your Gene results with a tool below." It features three boxes: "Gene Ontology Enrichment" (with a large "GO" logo), "Metabolic Pathway Enrichment" (with a network diagram), and "Word Enrichment" (with a list of terms: kinase, phosphatase, exported, membrane). On the far left, there is a vertical sidebar with a "Hide Organism Filter" button.

How do I transform a list of genes into homologs in a different organism?

Start with a list of genes using an existing strategy, a list of genes you submit (see above), or all the genes in a given organism (see above)

1. Click “Add a step”
2. Choose “Transform”
3. Click “Orthologs”
4. Locate the species/strain that you want to transform your list of genes, into and tick it
5. Click “Run Step” You will now have a list of genes in the new species (A) and corresponding list of homologue(s) in the input species (B).

**Step 1**

Step 1  
Unnamed Search Strategy \*

Organism 19,793 Genes + Add a step

19,793 Genes (14,063 ortholog groups) Revise this search

Organism Filter select all | clear all | expand all | collapse all

**Step 2**

Transform into related records

Organism

**Step 3**

Transform 19,793 Genes into...

- Orthologs
- Metabolic Pathways
- Compounds

**Step 4**

Add a step to your search strategy

Your Genes from Step 1 will be converted into:

Organism Note: You must select at least 1 values for this parameter. 1 selected out of 43

select all | clear all | expand all | collapse all

Filter list below... ④

- Arthropoda
- Arachnida
- Insecta
- Diptera
  - Culicidae
    - Aedes
      - Aedes aegypti LVP\_AGGW
      - Aedes albopictus Foshan
      - Anopheles
      - Culex
  - Glossinidae
  - Muscidae
  - Pipunculidae
  - Hemiptera
  - Phlebotomidae
  - Pediculidae
  - Pedicularis humanus USDA
  - Biomphalaria glabrata BB02
- Mollusca

select all | clear all | expand all | collapse all

**Step 5**

⑤ Run Step

**A**

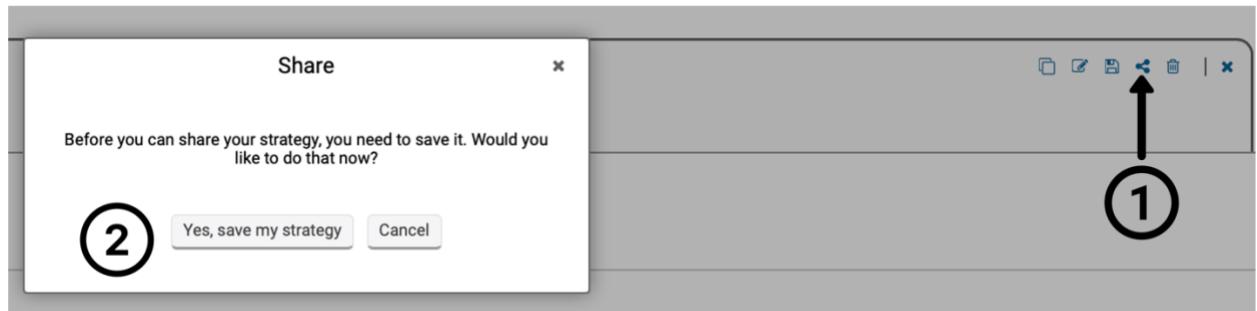
Gene ID	Transcript ID	Organism	Genomic Location (Gene)	Product Description	Input Ortholog(s)	Ortholog Group
AAEL013858	AAEL013858- Aedes aegypti LVP_AGGW	Aaegl_5_3:275,464,569..275,506,423(+) [Source: VB Community Annotation]	Aaegl_5_3:275,464,569..275,506,423(+) [Source: VB Community Annotation]	rat ion channel, nompC [Source: VB Community Annotation]	CPIJ004499	Og6_100006
AAEL008117	AAEL008117- Aedes aegypti LVP_AGGW	Aaegl_5_3:14,211,977..14,216,346(+) [Source: VB Community Annotation]	Aaegl_5_3:14,211,977..14,216,346(+) [Source: VB Community Annotation]	ion channel, nompC [Source: VB Community Annotation]	CPIJ001846,CPIJ002399,CPIJ010503,CPIJ018694	Og6_100012
AAEL011563	AAEL011563- Aedes aegypti LVP_AGGW	Aaegl_5_3:106,964,924..107,226,182(+) [Source: VB Community Annotation]	Aaegl_5_3:106,964,924..107,226,182(+) [Source: VB Community Annotation]	unspecified product	CPIJ001846,CPIJ002399,CPIJ010503,CPIJ018694	Og6_100012

**B**

How do I share my strategy /list of genes with someone else?

Ensure you are logged in first

1. Click on the share icon
2. If necessary, you will be prompted to save your strategy before you can share. Click the share icon again. A link will be provided that you can share.



## Manual gene annotation using Apollo

What can I do if a gene model is incorrect or missing?

1. Go to the gene record page of your gene of interest. In case of a missing gene, choose the gene record page of a neighboring gene.
2. There are two options. Option 1 is to add a user comment. Select “Add the first user comment”. To add a user comment you need to be logged in.
3. In the user comment window provide details about the incorrect/missing gene. The comment will be displayed on the gene record page after you’ve submitted it.
4. The second option is to correct the gene model in Apollo. To do so, click on the “View and update community annotations in Apollo” link on the gene record page.
5. You need to be logged in to VEuPathDB to use Apollo. If you want to correct a gene model in Apollo follow the steps in the tutorials and FAQs on the [Apollo page](#). Some organisms are currently unavailable in Apollo. You can request these organisms to be available for curation, [click here to contact us](#).

**1**

**2**

**3**

**4**

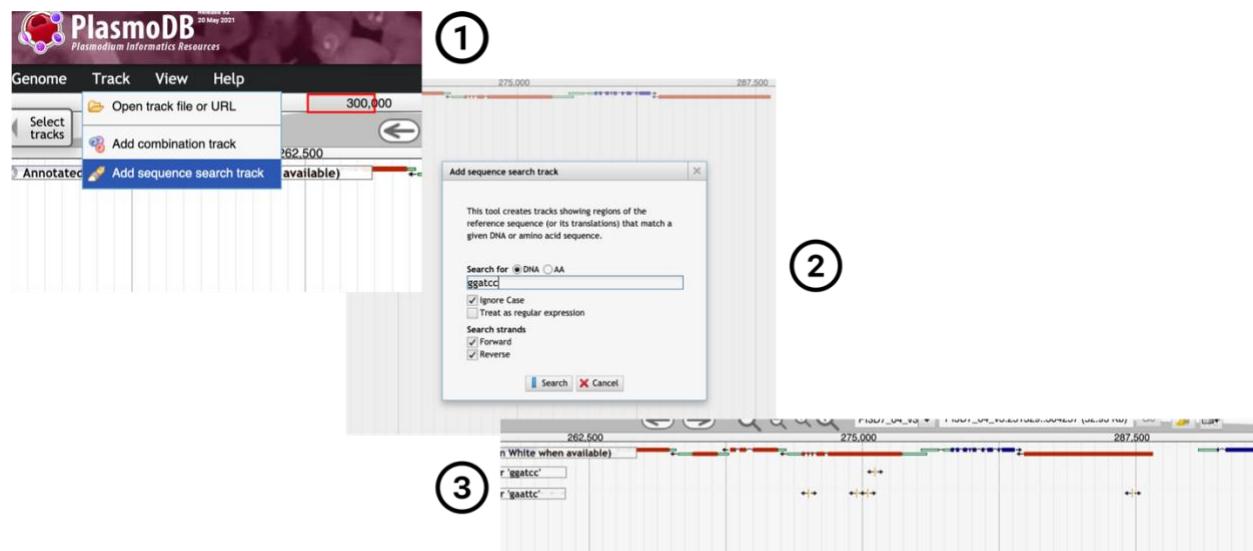
**5**

## Other website tools

How do I add a restriction site to JBrowse?

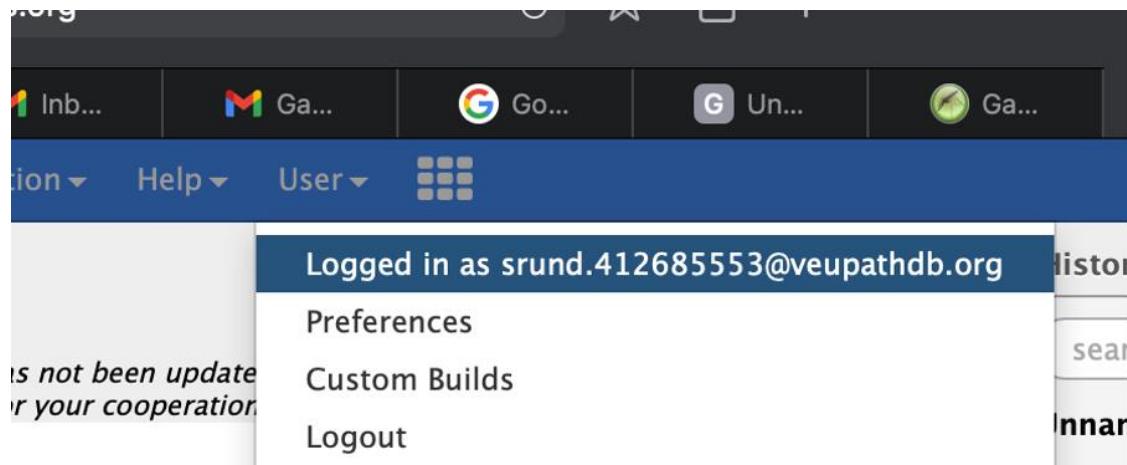
In JBrowse

1. Select “Add sequence search track” from the Track menu
2. Enter the enzyme sequence in the popup
3. The location of the sequence will now show up in the browser. The cool thing is that you can add multiple tracks with sequences that you define



Exporting from Galaxy/Globus to VEuPathDB returns a user email not valid error. How can I fix this?

This is most likely caused by you having a Galaxy account created outside VEuPathDB. To check this, click on “User” from the home page. If the email address listed does not end in “@veupathdb.org”, then you are using a non-VEuPathDB account. This account could have been created through the Globus system. If you do not have a “@veupathdb.org” address listed here, our systems will not be able to link your Galaxy account to your VEuPathDB account.



To fix this, go here: <https://app.globus.org/account/identities> You will likely have a primary identity consisting of your non-VEuPathDB email address:

A screenshot of the Globus account identities page. The page shows a list of identities linked to the user's account. The first identity, 'ryan2@globusid.org', is highlighted with a red circle and an orange arrow pointing down to it. This identity is listed as 'PRIMARY'. The second identity, 'rdoherty.12401223@veupathdb.org', is listed as 'IDENTITY'. Both identities show details such as Identity Provider, Name, Email, Organization, and 'Use It To' options. A note at the bottom states: 'This id is managed by Globus ID, where you can change your password, update your profile, and manage SSH and X.509 keys.'

If so, you need to unlink your VEuPathDB account from your other account by selecting ‘Manage Identities’ and pressing the Trash icon next to your VEuPathDB user name. This will not delete the VEuPathDB account, but will remove it from this particular Globus identity. Log out from this Globus identity.

Then, you must log out of Galaxy. Then log back into Galaxy using your VEuPathDB credentials -- this will likely create a new Galaxy account. Also, this will create a new Globus identity in which your VEuPathDB is the only account and also the primary identity. To verify that this worked, press the User menu in Galaxy and you should see your VEuPathDB user name: for example, mark123456789@veupathdb.org.

To transfer your work from your old non-VEuPathDB account to your new VEuPathDB account, share your history from the non-VEuPathDB account to the VEuPathDB account. Import the history into your VEuPathDB account.

Now you can export to our VEuPathDB websites. You should use this particular Galaxy account to do your future VEuPathDB work, so that you can export to VEuPathDB and so that we can contact you with important information about VEuPathDB Galaxy.

## Data Download

Download genomic or amino acid sequences (one gene)?

Start with a list of genes using an existing strategy, a list of genes you submit (see above), or all the genes in a given organism (see above)

1. From the gene page of your gene of interest, select “Download Gene”
2. Select “FASTA”
3. Select what kind of sequence you want
4. Select the region of sequence you want to download including sequences upstream and downstream of the coding region. Instructions at the bottom of the download page will guide you.
5. Click “Get Sequence”

The screenshot shows the VectorBase gene download interface. At the top, it displays "Release 50 beta" and "17 Dec 2020". Below the header, there are links for "Add to basket" and "Add to favorites". The main content area shows a gene entry for "CPIJ040096 unspecified product". The gene details are: Name: tRNA-His, Type: ncRNA gene, Chromosome: mitochondrial\_chromosome. To the right, there are "Shortcuts" for "Custom" and "All genes". Step 1 is highlighted with a circle around the "Download Gene" link. Step 2 is highlighted with a circle around the "Choose a Report" section, which includes options for "Text - choose from columns and/or tables" and "FASTA - sequence retrieval, configurable". Step 3 is highlighted with a circle around the "Choose the type of sequence:" section, which includes options for Genomic, Protein, CDS, and Transcript. Step 4 is highlighted with a circle around the "Choose the region of the sequence(s):" section, which includes fields for "Begin at" (Transcription Start\*\*\*) and "End at" (Transcription Stop\*\*\*), both with "+/-" buttons and "nucleotides" input fields. Step 5 is highlighted with a circle around the "Get Sequences" button at the bottom right.

Download genomic or amino acid sequences (lots of genes)?

Start with a list of genes using an existing strategy, a list of genes you submit (see above), or all the genes in a given organism (see above)

1. From the gene page of your gene of interest, select “Download Gene”
2. Select “FASTA”
3. Select what kind of sequence you want
4. Select the region of sequences you want to download including sequences upstream and downstream of the coding region. Instructions at the bottom of the download page will guide you.
5. Click “Get Sequences”

The screenshot shows the 'Gene Results' tab of the VEuPathDB interface. At the top, it displays 'Genes: 13,649' and 'Transcripts: 14,415'. A checkbox for 'Show Only One Transcript Per Gene' is checked. Below this, there's a pagination control with pages 1 through 721 and a 'Rows per page' dropdown set to 20. To the right of the table, there's a large 'Download' button with a circled '1' over it, and an 'Add to Basket' button. The main table has columns for Gene ID, Transcript ID, Organism, Genomic Location (Gene), and Computed GO Component IDs. A row is selected for 'AARA007236' from 'Anopheles arabiensis' with location 'KB704125:4,085..4,785(-)' and 'N/A' for GO components. To the right of the table, there's a 'Download Genes' section with a 'Choose a Report' dropdown (circled '2') containing options like 'Tab- or comma-delimited (openable in Excel)', 'FASTA - sequence retrieval (configurable)', 'GFF3 - gene models and optional sequences', and 'Standard JSON'. Below this, there's a 'Choose the type of sequence:' dropdown with options for 'Genomic', 'Protein', 'CDS', and 'Transcript' (circled '3'). Further down, there's a 'Choose the region of the sequence(s):' section with 'Begin at' and 'End at' fields for transcription start and stop, both set to 0 nucleotides. There are also sections for 'Download Type' (Text File, Show in Browser), 'Fasta define' (Only Gene ID, Full Fasta Header), and 'Sequence format' (Single line, Default (60 chars on a line)). A 'Get Sequences' button is at the bottom. A note at the bottom right explains the behavior for genomic and protein sequences.

**Note:**  
For "genomic" sequence: If UTRs have not been annotated for a gene, then choosing "transcription start" may have the same effect as "coding start".  
For "protein" sequence: you can only retrieve sequence contained within the ID(s) listed, i.e. from downstream of amino acid sequence.

How do I download a table with a list of genes including any field(s) from our gene pages (e.g., gene names and symbols)?

Start with a list of genes using an existing strategy, a list of genes you submit (see above), or all the genes in a given organism (see above)

1. Click “Add Columns”
2. Add any relevant columns of data you wish
3. Click “Update Columns”
4. Click “Download”

The screenshot shows the VEuPathDB interface with a 'Select Columns' dialog open over a table of gene data.

**1** (top right): A circular callout pointing to the 'Download' button in the top right corner of the main interface.

**2** (middle left): A circular callout pointing to the 'Product Description' checkbox in the 'Annotation, curation and identifiers' section of the 'Select Columns' dialog.

**3** (bottom center): A circular callout pointing to the 'Update Columns' button at the bottom of the 'Select Columns' dialog.

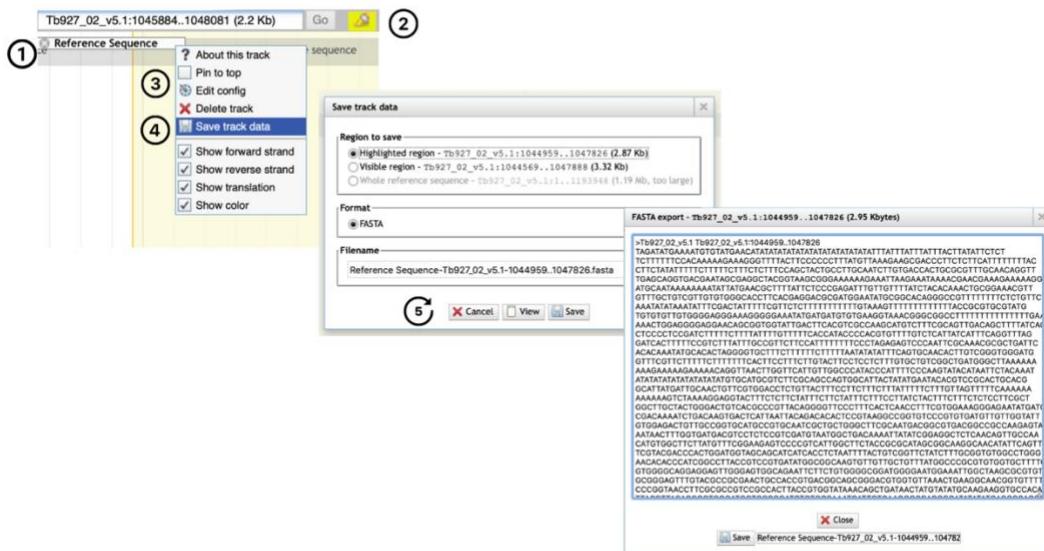
**4** (top right): A circular callout pointing to the 'Download' button in the 'Select Columns' dialog.

**Main Interface (Table View):**

	Gene ID	Gene Name	Product Description
PBANKA_00001	PBANKA_00001	Plasmodium berghei ANKA	PIR protein
PBANKA_00002	PBANKA_00002	Plasmodium berghei ANKA	fam-c protein, pseudogene
PBANKA_00003	PBANKA_00003	Plasmodium berghei ANKA	fam-b protein
PBANKA_00004	PBANKA_00004	Plasmodium berghei ANKA	tryptophan-rich antigen, putative, pseudogene
PBANKA_00006	PBANKA_00006	Plasmodium berghei ANKA	PIR protein
PBANKA_00007	PBANKA_00007	Plasmodium berghei ANKA	PIR protein
PBANKA_00008	PBANKA_00008	Plasmodium berghei ANKA	PIR protein
PBANKA_00009	PBANKA_00009	Plasmodium berghei ANKA	fam-a protein
PBANKA_00010	PBANKA_00010	Plasmodium berghei ANKA	PIR protein
PBANKA_00011	PBANKA_00011	Plasmodium berghei ANKA	PIR protein
PBANKA_00012	PBANKA_00012	Plasmodium berghei ANKA	PIR protein
PBANKA_00013	PBANKA_00013	Plasmodium berghei ANKA	PIR protein
PBANKA_0001401.1	PBANKA_0001401.1	Plasmodium berghei ANKA	PIR protein
PBANKA_0001501.1	PBANKA_0001501.1	Plasmodium berghei ANKA	PIR protein
PBANKA_0001601.1	PBANKA_0001601.1	Plasmodium berghei ANKA	fam-a protein
PBANKA_0006300.1	PBANKA_0006300.1	Plasmodium berghei ANKA	PIR protein
PBANKA_0006401.1	PBANKA_0006401.1	Plasmodium berghei ANKA	PIR protein, pseudogene

How do I download a sequence in FASTA format from a region of interest in JBrowse?

1. Make sure the “annotated transcripts” and the “reference sequence” tracks are turned on
  2. Click on the “highlight a region” button in the navigation bar. It should turn yellow when activated
  3. Click and drag in the genome view region and select the area you would like to highlight (tip: highlight region from within annotation track)
  4. Click on the down arrow on the reference sequence track and select “Save track data”
  5. In the next popup window you can keep everything as the default and either save or view the sequence



## Data submission

How do I submit data?

Please visit: [data submission](#)

How do I add a comment to a gene pages?

Our user comment system links your expert knowledge to both gene and other records. Your comments are immediately integrated into the database and become searchable. Your comments will also guide our curators' efforts.



Add to basket Add to favorites Download Gene

## EDI\_244000 Phosphatidylchol

**Type:** protein coding gene

**Shortc**

**Chromosome:** Not Assigned

**Location:** DS548829:7,885..9,171(-)

**Synten**

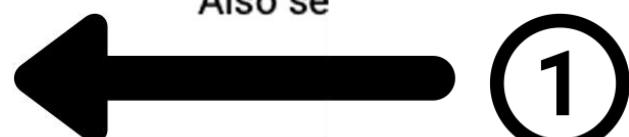
**Species:** Entamoeba dispar

**Also se**

**Strain:** SAW760

**Status:** Reference Strain

**Add the first user comment**



## Public Strategies

Users can share their strategies publicly so that others may use them. The public strategies link is located under the **About** menu followed by **Community** and **Public Strategy**. A table of available public strategies will appear and there is a filter box located at the top of the public strategies so that you can search for the author or subject of the strategy among other items. The public strategies for PlasmoDB, as an example, are located at:

<https://plasmodb.org/plasmo/app/workspace/strategies/public>

## Print-based instructional material

The following tutorials are included in [Appendix A](#) and for download (<https://veupathdb.org/veupathdb/app/static-content/tutorials.html> )

- OrthoMCL Tutorial
- Site Search
- Search Strategies
- Advanced Search Strategies
- Exploring the Gene Page
- JBrowse Basics
- Interpreting RNA-seq data (Browser Exercise II)
- Gene Ontology (GO) Enrichment
- Regular Expressions & Genomic Colocation
- Variant calling in VEuPathDB galaxy (Part 1)
- Variant calling in VEuPathDB Galaxy (Part 2)
- Genome Annotation with Companion (Part 1)
- Genome Annotation with Companion (Part 2)
- Genetic Variation Exercises
- Host Response
- Metabolic Pathways - Exploring pathways and compounds
- Orthology and Phyletic Patterns
- Exploring Transcriptomic data
- MapVEu: part I - An Introduction
- MapVEu: part II - Additional features
- Functional Annotation in Apollo
- Structural Annotation in Apollo

## Related sites of interest to our communities

- [Previous EuPathDB Workshops](#)
- [Companion](#)
- [OrthoMCL](#)
- [GeneDB](#)
- [ModBase at UCSF](#)
- [Tetrahymena Genome](#)
- [The Arabidopsis Information Resource](#)
- [NAR Database Summary Paper Categories](#)

# VEuPathDB Publications and Citations

View [publications that cite us](#) in Google Scholar.

This [PubMed filter](#) provides a current list of the most recent publications about VEuPathDB resources

Recent publications include:

(Book Chapter)

Omar Harb, Jessica C. Kissinger and David S. Roos on behalf of the EuPathDB group

**ToxoDB: the functional genomic resource for *Toxoplasma***

*Toxoplasma gondii* 3rd edition

Edited by Louis Weiss and Kami Kim (2020)

<https://doi.org/10.1016/B978-0-12-815041-2.00023-2>

(Book Chapter)

Susanne Warrenfeltz and Jessica Kissinger on behalf of the EuPathDB Consortium

[\*\*Accessing \*Cryptosporidium\* omic & isolate data via CryptoDB.org\*\*](#)

Methods in Molecular Biology

Editor, Jan Mead (2020)

[https://doi.org/10.1007/978-1-4939-9748-0\\_22](https://doi.org/10.1007/978-1-4939-9748-0_22)

(Book Chapter)

Omar S. Harb and David S. Roos on behalf of the EuPathDB Consortium

**ToxoDB: Functional Genomics Resource for *Toxoplasma* and Related Organisms**

Methods in Molecular Biology

Editor, Christopher J. Tonkin (2020)

[https://doi.org/10.1007/978-1-4939-9857-9\\_2](https://doi.org/10.1007/978-1-4939-9857-9_2)

(Book Chapter)

Susanne Warrenfeltz, Evelina Y Basenko, Kathryn Crouch, Omar S. Harb, Jessica C. Kissinger, Achchuthan Shanmugasundram and Fatima Silva-Franco

**EuPathDB: the eukaryotic pathogen genomics database resource**

Methods in Molecular Biology

Editor, Martin Kollmar (2018)

[https://doi.org/10.1007/978-1-4939-7737-6\\_5](https://doi.org/10.1007/978-1-4939-7737-6_5)

Evelina Y. Basenko, Jane A. Pulman, Achchuthan Shanmugasundram, Omar S. Harb, Kathryn Crouch, David Starns, Susanne Warrenfeltz, Cristina Aurrecoechea, Christian J. Stoeckert, Jr., Jessica C. Kissinger, David S. Roos and Christiane Hertz-Fowler

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J. Fungi 2018, 4(1), 39

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Francislon S Oliveira, John Brestelli, Shon Cade, Jie Zheng, John Iodice, Steve Fischer, Cristina Aurrecoechea, Jessica C Kissinger, Brian P Brunk, Christian J Stoeckert, Jr, Gabriel R Fernandes, David S Roos, Daniel P Beiting

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[Nucleic Acids Research. 2017 doi.org/10.1093/nar/gkx1027](https://doi.org/10.1093/nar/gkx1027)

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**EuPathDB: the eukaryotic pathogen genomics database resource**

[Nucleic Acids Research 2017 doi: 10.1093/nar/gkw1105](https://doi.org/10.1093/nar/gkw1105)

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Nucleic Acids Res. 2013 Jan;41(Database issue):D684-91

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Nucleic Acids Res. 2013 Jan;41(Database issue):D728-31

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## Publications that use our resource

Our project resources are cited in >20,000 publications. You can view the [publications that cite us](#) in Google Scholar.

## Release Notes

Release notes are generated for each project site within VEuPathDB with each new release. They are located in the “New” tab on the right-hand expandable panel of each project site. An example for VectorBase is shown below.

### Example release notes - VectorBase 48 Released

(27 Aug 2020)

**We are pleased to announce the release of VectorBase 48.**

Beginning with VectorBase 48, the URL <https://VectorBase.org> directs you to the newest version of our interface. The site contains previous VectorBase.org data plus some new tools and functions in an updated and streamlined interface that emphasizes easy access to help information. Navigate to <https://legacy.VectorBase.org> for the previous version of the interface. The legacy interface will remain active for at least one release.

#### Release 48 Webinar

Join us on September 3, 2020 at 10am EDT for our Release 48 Highlights webinar where we will demonstrate and discuss new data and features in VEuPathDB 48. [Register here](#)

## New features in VectorBase 48

- Gene pages: The sequence section now contains links to copy the sequence to your clipboard. Sequences are copied in FASTA format with the gene ID in the def line.
- Gene pages: It is now possible to scroll and zoom within the JBrowse views that are present in the 'Gene models' and 'Protein features and properties' sections.
- Contact Us form: A new option on the Contact Us form makes it possible to paste a screenshot into the form when sending us questions or suggestions.
- Search result page: The Download and Add to Basket options from a search result tab now appear as buttons for better visibility.
- Error messages: The text in our error messages was reviewed and edited to improve clarity. And we've made it easier to report errors by adding links in error messages to the Contact Us form.

## Omics data sets

- Fifteen additional microarray datasets are available in the merged VectorBase site. These are marked as 'New' in the [Identify Genes based on Microarray Evidence](#) search page.
- The results of nine comparative genomic study analyses (VCF files) are available in [JBrowse](#). The data are categorized under Genetic Variation in the Select Tracks tool.

## Population Biology

This release represents our second largest abundance data release ever – over 219,000 new non-zero abundance records. It is also our largest increase in pathogen status assays, doubling our records to over 174,000 assays.

45 projects have been added. You can use any of the Project IDs from the list below, in the Search box of the MapVEu (formerly PopBio map) tool: <https://vectorbase.org/popbio-map/web/>

- Indoor Human landing catches from Western Cameroon (VBP0000626: [map](#), 52 collections, 166 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in Uganda by PRISM ICEMR group, 2018 (VBP0000627: [map](#), 1113 collections, 7791 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in Marion County, Indiana, USA. 2019. (VBP0000628: [map](#), 1758 collections, 7539 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in Manatee County, Florida, USA. 2017 (VBP0000629: [map](#), 1845 collections, 10390 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in Manatee County, Florida, USA. 2018 (VBP0000630: [map](#), 1217 collections, 8462 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in Manatee County, Florida, USA. 2019 (VBP0000631: [map](#), 1859 collections, 11223 samples, 0 phenotypes, 0 genotypes)
- Maryland Dept. of Agriculture mosquito surveillance, Maryland, USA, 2016 (VBP0000632: [map](#), 558 collections, 4373 samples, 0 phenotypes, 0 genotypes)
- Maryland Dept. of Agriculture mosquito surveillance, Maryland, USA, 2019 (VBP0000634: [map](#), 757 collections, 7016 samples, 0 phenotypes, 0 genotypes)

- Mosquito surveillance in Lucas County, Ohio, USA. 2018 (VBP0000635: [map](#), 1438 collections, 6626 samples, 0 phenotypes, 0 genotypes)
- Supplemental Maryland Dept. of Agriculture mosquito surveillance, Maryland, USA, 2016 (VBP0000636: [map](#), 45 collections, 259 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in Lucas County, Ohio, USA. 2019 (VBP0000637: [map](#), 1554 collections, 7260 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in Hernando County, Florida, USA. 2018 (VBP0000638: [map](#), 361 collections, 2167 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in Hernando County, Florida, USA. 2019 (VBP0000639: [map](#), 513 collections, 2656 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2016 (VBP0000664: [map](#), 1750 collections, 4745 samples, 0 phenotypes, 0 genotypes)
- Dynamic of resistance alleles of two major insecticide targets in *Anopheles gambiae* (s.l.) populations from Benin, West Africa (VBP0000640: [map](#), 28 collections, 28 samples, 54 phenotypes, 0 genotypes)
- Contrasting resistance patterns to type I and II pyrethroids in two major arbovirus vectors *Aedes aegypti* and *Aedes albopictus* in the Republic of the Congo, Central Africa (VBP0000641: [map](#), 7 collections, 17 samples, 45 phenotypes, 12 genotypes)
- Maryland Dept. of Agriculture mosquito surveillance, Maryland, USA, 2017 (VBP0000633: [map](#), 614 collections, 5500 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Norfolk County Mosquito Control, in Massachusetts, USA, 2019 (VBP0000644: [map](#), 934 collections, 4297 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in Desplaines Valley Mosquito Abatement, Illinois, USA, 2019 (VBP0000645: [map](#), 3006 collections, 25974 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in Salt Lake County, Utah, USA. 2018 (VBP0000646: [map](#), 1585 collections, 6134 samples, 0 phenotypes, 0 genotypes)
- Mosquito infection assays for *Plasmodium falciparum* in Uganda by PRISM ICEMR group, 2015-2017 (VBP0000647: [map](#), 7621 collections, 92524 samples, 92524 phenotypes, 0 genotypes)
- Mosquito infection assays for *Plasmodium falciparum* in Uganda by PRISM ICEMR group, 2018 (VBP0000648: [map](#), 379 collections, 1497 samples, 1497 phenotypes, 0 genotypes)
- Mosquito surveillance in Uganda by PRISM ICEMR group, 2017 - 2019 (VBP0000649: [map](#), 2706 collections, 6791 samples, 0 phenotypes, 0 genotypes)
- Susceptibility of *An. gambiae* s.l. from Côte d'Ivoire to Insecticides used on Insecticide-Treated Nets: Evaluating the Additional Entomological Impact of Piperonyl Butoxide and Chlорfenapyr (VBP0000650: [map](#), 15 collections, 165 samples, 165 phenotypes, 0 genotypes)
- Mosquito abundance in Central Mozambique (VBP0000651: [map](#), 28 collections, 55 samples, 0 phenotypes, 0 genotypes)
- Spatial and temporal distribution of *Anopheles arabiensis* larvae (Sudan) (VBP0000652: [map](#), 3293 collections, 3296 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2017 (VBP0000653: [map](#), 1623 collections, 5221 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2019 (VBP0000654: [map](#), 1376 collections, 4778 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2007 (VBP0000655: [map](#), 602 collections, 1408 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2008 (VBP0000656: [map](#), 692 collections, 1627 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2009 (VBP0000657: [map](#), 831 collections, 2389 samples, 0 phenotypes, 0 genotypes)

- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2011 (VBP0000659: [map](#), 1064 collections, 3050 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2012 (VBP0000660: [map](#), 1482 collections, 3972 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2013 (VBP0000661: [map](#), 1819 collections, 5103 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2014 (VBP0000662: [map](#), 1488 collections, 5151 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2015 (VBP0000663: [map](#), 1625 collections, 4856 samples, 0 phenotypes, 0 genotypes)
- Montana State University mosquito data for 2015 (VBP0000666: [map](#), 223 collections, 1458 samples, 0 phenotypes, 0 genotypes)
- Montana State University mosquito data for 2018 (VBP0000667: [map](#), 239 collections, 1640 samples, 0 phenotypes, 0 genotypes)
- Montana State University mosquito data for 2016 (VBP0000668: [map](#), 283 collections, 1790 samples, 0 phenotypes, 0 genotypes)
- Montana State University mosquito data for 2019 (VBP0000669: [map](#), 80 collections, 639 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance in South Walton County, Florida, USA. 2019 (VBP0000670: [map](#), 856 collections, 5264 samples, 0 phenotypes, 0 genotypes)
- Mosquito surveillance, Ada County Weed, Pest, and Mosquito, in Idaho, USA, 2010 (VBP0000658: [map](#), 798 collections, 1976 samples, 0 phenotypes, 0 genotypes)
- Larval habitats seasonality and species distribution (VBP0000671: [map](#), 4278 collections, 8866 samples, 0 phenotypes, 0 genotypes)
- Comparative toxicity of larvicides and growth inhibitors on Aedes aegypti from select areas in Jamaica (VBP0000642: [map](#), 6 collections, 238 samples, 238 phenotypes, 0 genotypes)
- Variation in Malaria Transmission Dynamics in Three Different Sites in Western Kenya (VBP0000665: [map](#), 133 collections, 1701 samples, 0 phenotypes, 0 genotypes)

Note: Our bimonthly database releases incorporate new data and correct errors in old data when necessary. Changes in annotation and new experimental data may slightly alter your search results by increasing or decreasing the number of hits. When search parameters change with a new release, we invalidate ( $\emptyset$ ) the search and ask you to rerun it. When IDs are updated or removed, we map the old IDs to the new ones, remove the old IDs from your Basket, and leave your Favorites page alone.

## Analyses methods

VEuPathDB draws data from many sources. To facilitate comparisons across data sets, we analyze all data with standardized, data type-specific analyses. All data of one type are analyzed with the same workflow. Although our results may show some differences from an author's publication, our re-analysis of the data makes it feasible to compare data sets from very different sources and to update the data analysis with contemporary methods. For transparency, the methods we use to analyze data are presented below. They are also located online at: *Link:* <https://veupathdb.org/veupathdb/app/static-content/methods.html>

## Genome analyses

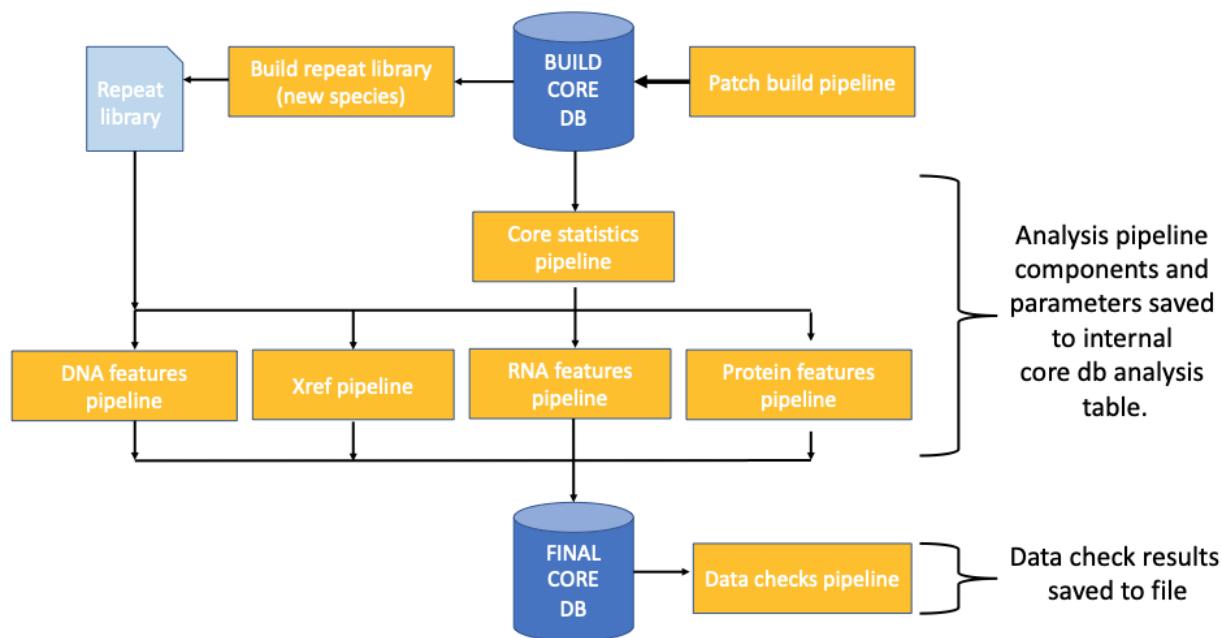
VEuPathDB employs the [Ensembl genome analysis](#) pipelines for analyzing genomic sequence to enhance annotations. While most of the genomic sequence (FASTA) are integrated into VEuPathDB from an INSDC repository, genome annotation (GFF3) may come from either the INSDC repository or a community submission.

[Core database pipelines](#) (next figure)- Primary genomic sequence and structural annotation data are loaded into a [core database](#) and run through 6 pipelines: core statistics, DNA feature annotation, [external cross reference](#) annotation, [RNA gene](#) annotation, [repeat feature](#) annotation, and protein feature annotation. The main pipelines applied to the core database and their components are listed in table 1.

Table 1 - Core database analysis pipelines and hive components.

Pipeline	Hive pipeline component	Git repository link
Core db build	Bio::EnsEMBL::EGPipeline::LoadGFF3 ::LoadGFF3	Under code review, details coming soon
Protein features	InterProScan	<a href="https://github.com/Ensembl/ensembl-production">https://github.com/Ensembl/ensembl-production</a>
DNA features	Bio::EnsEMBL::EGPipeline::DNA Features::*	Under code review, details coming soon
RNA features	Bio::EnsEMBL::EGPipeline::RNA Features::*	Under code review, details coming soon
Xrefs	Bio::EnsEMBL::EGPipeline::Xref::*	Under code review, details coming soon
Core Statistics	shortnoncodingdensity	<a href="https://github.com/Ensembl/ensembl-production">https://github.com/Ensembl/ensembl-production</a>
Core Statistics	snpdensity	<a href="https://github.com/Ensembl/ensembl-production">https://github.com/Ensembl/ensembl-production</a>
Core statistics	codingdensity	<a href="https://github.com/Ensembl/ensembl-production">https://github.com/Ensembl/ensembl-production</a>
Core statistics	percentgc	<a href="https://github.com/Ensembl/ensembl-production">https://github.com/Ensembl/ensembl-production</a>
Core statistics	percentagerepeat	<a href="https://github.com/Ensembl/ensembl-production">https://github.com/Ensembl/ensembl-production</a>

Core statistics	pseudogenedensity	<a href="https://github.com/Ensembl/ensembl-production">https://github.com/Ensembl/ensembl-production</a>
Core statistics	longnoncodingdensity	<a href="https://github.com/Ensembl/ensembl-production">https://github.com/Ensembl/ensembl-production</a>



### Core database analysis pipelines and hive components

### Example ehive pipelines, modules, programs and parameter data from coredb analysis table

Table 2 - Example ehive pipelines, modules, programs and parameter data from coredb analysis table.

Pipeline	Program	Program version	Parameters	Ehive module (Bio::EnsEMBL::)	Database	Database version
DNA features	dustmasker	NULL	NULL	Analysis::Runnable::DustMasker		

DNA features	trf	4	25 7 80 10 40 500 -d -h	Analysis::Runnable::TRF		
DNA features	RepeatMasker	4.0.5	-nolow -gccalc -species "Aedes aegypti" -engine crossmatch -q	Analysis::Runnable::RepeatMasker		
DNA features	RepeatMasker	4.0.5	-nolow -gccalc -lib "location1" -engine crossmatch -q	Analysis::Runnable::RepeatMasker		
DNA features	RepeatMasker	4.0.5	-nolow -gccalc -lib "location2" -engine crossmatch -q	Analysis::Runnable::RepeatMasker		
	NULL	NULL	NULL	EGPipeline::LoadGFF3::LoadGFF3		
RNA features	Infernal	1.1		Analysis::Runnable::CMScan		
RNA features	tRNAscan-SE	1.23		Analysis::Runnable::tRNAscan		
RNA features	Infernal	1.1		Analysis::Runnable::CMScan		
RNA features	rfam_12.2_gene	NULL	NULL	EGPipeline::RNAFeatures::CreateCmscanGenes		
RNA features	mirbase_gene	NULL	NULL	EGPipeline::RNAFeatures::CreateMirbaseGenes		
RNA features	trnascan_gene	NULL	NULL	EGPipeline::RNAFeatures::CreateTrnascanGenes		
Xref	xrefchecksum	NULL	NULL	EGPipeline::Xref::LoadUniParc		
Xref	xrefuniparc	NULL	NULL	EGPipeline::Xref::LoadUniProt		
Xref	gouniprot	NULL	NULL	EGPipeline::Xref::LoadUniProtGO		
Xref	xrefuniprot	NULL	NULL	EGPipeline::Xref::LoadUniProtXrefs		

DNA features	blastp	NULL	-word_size 3 -num_alignments 100000 -num_descriptions 100000 -lcase_masking -seg yes -num_threads 3	Analysis::Runnable::Bla stEG		
DNA features	blastp	NULL	-word_size 3 -num_alignments 100000 -num_descriptions 100000 -lcase_masking -seg yes -num_threads 3	Analysis::Runnable::Bla stEG		
Protein Features	InterProScan	5.37-76.0	NULL		Prosite patterns	2019_01
Protein Features	InterProScan	5.37-76.0	NULL		SFLD	4
Protein Features	InterProScan	5.37-76.0	NULL		CDD	3.17
Protein Features	InterProScan	5.37-76.0	NULL		Gene3D	4.2.0
Protein Features	InterProScan	5.37-76.0	NULL		HAMAP	2019_01
Protein Features	InterProScan	5.37-76.0	NULL		PANTHER	14.1
Protein Features	InterProScan	5.37-76.0	NULL		ncoils	2.2.1
Protein Features	InterProScan	5.37-76.0	NULL		Prosite profiles	2019_01
Protein Features	InterProScan	5.37-76.0	NULL		Pfam	32
Protein Features	InterProScan	5.37-76.0	NULL		PRINTS	42
Protein Features	InterProScan	5.37-76.0	NULL		Smart	7.1
Protein Features	InterProScan	5.37-76.0	NULL		SuperFamily	1.75
Protein Features	InterProScan	5.37-76.0	NULL		TIGRfam	15

Protein Features	InterProScan	5.37-76.0	NULL		InterPro- 2GO	NULL
Protein Features	InterProScan	5.37-76.0	NULL		PIRSF	3.02
Protein Features	InterProScan	5.37-76.0	NULL		SignalP	4.1
Protein Features	InterProScan	5.37-76.0	NULL		TMHMM	2.0c
Protein Features	InterProScan	5.37-76.0	NULL		Seg	NULL
Protein Features	InterProScan	5.37-76.0	NULL		MobiDBLite	2
Protein Features	InterProScan	5.37-76.0	NULL		InterPro- 2Pathway	NULL

\*Location 1

"/homes/jallen/scratch/vb/recent\_assemblies/aedes\_aegypti/RepeatModeler/aedes\_aegypti.rm.lib"

\*Location 2 "/nfs/panda/ensemblgenomes/vectorbase/data/tefam/aegypti\_tefam.lib"

## Supplements to the EBI Pipelines

VEuPathDB supplements the EBI pipeline with workflows that produce data for EST alignments, Open reading frames, and synteny (Table).

EST alignments: BLAT is applied to EST sequences that have been blocked using RepeatMasker.

Open reading frame generation: Open reading frames are generated from genomic DNA or EST sequences. The analysis produces a gff file containing the ORFs (50 or more amino acid translations of the input nucleic acid) for the 6 reading frames. The translations in all 6 reading frames do not necessarily begin with MET but always end at a stop codon. ORF names are in the form template-frame-start-end, e.g. AAEL01000396-5-5847-4366.

Synteny: VEuPathDB uses an in-house script called runMercator to run pair-wise alignments that employs Mercator and MAVID for comparative genome analysis. Mercator generates orthology maps using genomes and exon coordinates to create exon translations for protein BLAT alignments. The orthology maps are used as a guide for MAVID which also uses a phylogenetic newick tree to generate gene alignments.

### Details for the supplements to the EBI pipelines

Table 3: Details for supplements to the EBI Pipeline for genomes

Data	Program	Parameters or VEuPathDB GitHub repository	Version
------	---------	---	---------

EST alignments	BLAT	-nohead -maxIntron=1000 -t=dna -q=dna -dots=10	35
Open Reading Frames	orfFinder	--minPepLength 50 <a href="https://github.com/VEuPathDB/.../orfFinder">https://github.com/VEuPathDB/.../orfFinder</a>	
Synteny	Mercator and MAVID	-p <PATH TO MERCATOR DIRECTORY> -t <tree string> -m <MAVID_EXE> -c <CNDSRC_DIR> -d draftGenome1... -d draftGemoneN -n nonDraftGeome1... -n nonDraftGenomeN -r referenceGenome <a href="https://github.com/VEuPathDB/.../runMercator">https://github.com/VEuPathDB/.../runMercator</a>	Mercator mapmaker 0.4 (2016-01-21)  Mavid Version 2.0.4

## In-house genome analyses in lieu of the EBI Pipelines

On rare occasions the EBI pipeline cannot be applied to a genome. For example, genomes that are not housed at an INSDC repository cannot be analyzed by the EBI pipeline. VEuPathDB uses the following in-house analyses in lieu of the EBI pipeline.

[BLAT against NRDB](#): For every genome, VEuPathDB runs BLAT alignments of the annotated proteins against the GenBank Non-Redundant Protein Sequence Database (NRDB) to identify possible relationships and alignments outside the scope of VEuPathDB-supported organisms.

[Compute open reading frames](#): Open reading frames are generated from genomic DNA or EST sequences. The analysis produces a gff file containing the ORFs (50 or more amino acid translations of the input nucleic acid) for the 6 reading frames. The translations in all 6 reading frames do not necessarily begin with MET but always end at a stop codon. ORF names are in the form template-frame-start-end, e.g. AAEL01000396-5-5847-4366.

[DNA repeats](#): The Tandem Repeats Finder program locates and displays tandem repeats in genomic sequences.

[EST alignments](#): BLAT is applied to EST sequences that have been blocked using RepeatMasker.

[Protein domain annotations](#): InterProScan scans protein sequences against the protein signatures of the InterPro member databases and generates a file containing the domain matched, description of the InterPro entry, GO descriptions and E-values.

[Signal peptide prediction](#): Signal P is used to identify signal peptides and their likely cleavage sites. A signal peptide is a short peptide present at the N-terminus of most newly synthesized proteins that are destined towards the secretory pathway.

Syntenic sequences: VEuPathDB uses an in-house script called runMercator to run pair-wise alignments that employs Mercator and MAVID for comparative genome analysis. Mercator generates orthology maps using genomes and exon coordinates to create exon translations for protein BLAT alignments. The orthology maps are used as a guide for MAVID which also uses a phylogenetic newick tree to generate gene alignments.

Transmembrane domain prediction: TMHMM is used to predict transmembrane domain presence and topology from protein sequences.

tRNA gene prediction: tRNAScan identifies transfer RNA genes in transcript or genome sequences.

#### Details for the VEuPathDB in-house pipelines

Table 4: Details for genome analyses in lieu of EBI Pipeline

Data	Program	Parameters or VEuPathDB GitHub repository	Version
BLAT against NRDB	BLAT	-nohead -maxIntron=1000 -t=dna -q=prot -dots=10 -minScore=25 -minIdentity=20	35
Computed Open Reading Frames	orfFinder	--minPepLength 50 <a href="https://github.com/VEuPathDB/.../orfFinder">https://github.com/VEuPathDB/.../orfFinder</a>	
DNA Repeat regions	Tandem Repeats Finder	2 7 7 80 20 50 500	4.04
EST alignments	BLAT	-nohead -maxIntron=1000 -t=dna -q=dna -dots=10	35
Signal peptide predictions	SignalP	-t euk -f short -m nn+hmm -q -trunc 70	3.0

Synteny	Mercator and MAVID	-p <PATH TO MERCATOR DIRECTORY> -t <tree string> -m <MAVID_EXE> -c <CNDSRC_DIR> -d draftGenome1... -d draftGemoneN -n nonDraftGeome1... -n nonDraftGenomeN -r referenceGenome <a href="https://github.com/VEuPathDB/.../runMercator">https://github.com/VEuPathDB/.../runMercator</a>	Mercator mapmaker 0.4 (2016-01-21)  Mavid Version 2.0.4
Transmembrane domain prediction	TMHMM	Nice -short	2.0c

## Orthology

The identification of orthologs is an important concept in gene evolution, commonly used to infer likely gene function(s) in newly-sequenced genomes. Orthologs are proteins in different species that have diverged solely through speciation, and are therefore relatively likely to retain similar function ([Gabaldón & Koonin 2013](#)). Paralogs are proteins that arose through gene duplication; while initially identical, duplication provides the opportunity for functional divergence and evolutionary selection. “In-paralogs” refer to recent duplications that are likely to retain similar function; “out-paralogs” derive from more ancient duplications predating divergence of the species of interest, and are more likely to have diverged in function. See [Chen 2007](#) and the [Orthology Benchmarking Service](#) ([Altenhoff 2020](#)) for descriptions and assessments of widely-used ortholog detection algorithms

### OrthoMCL

VEuPathDB websites use the OrthoMCL algorithm ([Li 2003](#)) to infer orthology and paralogy, employing protein sequence similarity (BLASTP) to relate proteins across the tree of life, several normalization steps to improve performance on complex eukaryotic genomes, and Markov clustering to group these proteins into ortholog groups. Each protein in every species is assigned to precisely one OrthoMCL ortholog group (e.g., OG6\_135465). The [OrthoMCL website](#) allows users to explore ortholog groups and the proteins they contain, including sequences, similarity metrics, alignments, domain architecture, and phyletic pattern profiles. Users can also assign putative ortholog groups to proteins from additional genomes of interest ([Fisher 2011](#)). Because of the large and rapidly growing number of available genomes, the most recent implementation of OrthoMCL has been modified to improve scalability and usability, as described [here](#).

### Orthology on the Gene Page

All Gene Pages in VEuPathDB databases include an ‘Orthology and Synteny’ section providing: a link to the ortholog group on the OrthoMCL database, a list of orthologs within the same VEuPathDB database component (e.g., PlasmoDB; see the OrthoMCL database for orthologs in other species), and a JBrowse genome browser display of syntenic chromosomes in other strains and species (shaded to indicate orthology).

## Orthology-based functional prediction

Many genes have yet to be functionally characterized, but function can often be inferred using orthology and protein feature information. VEuPathDB assigns putative Enzyme Commission (EC) numbers (e.g., 2.7.4.3) based on a heuristic scoring algorithm that considers the length, sequence similarity, and domain architecture of other proteins in the same ortholog group that have previously been annotated with this EC number. Scoring details and other statistics are provided in a table in the 'Functional Prediction' section of the Gene Page; further details on EC prediction are available [here](#). The inferred EC number is also represented in metabolic pathways graphs and tables accessible via the 'Pathways and Interactions' section of the Gene Page.

## Searches for genes based on orthology

Several searches employ orthology to identify genes of possible interest. The [Orthology Phylogenetic Profile Search](#), identifies genes based on the taxonomic groups in which orthologs are or are **not** found (i.e. genes displaying a specific pattern of conservation across the tree of life). For example, users seeking a pathogen-specific therapeutic target may wish to search for genes present in the pathogen(s) of interest, but not in non-pathogenic species or host species (e.g., mammals). The [Paralog Count Search](#) allows users to identify genes based on the number of paralogs identified, e.g. genes that have undergone a recent expansion through one or more duplications. Finally, any list of genes identified through previous searches or [Search Strategies \(tutorial\)](#) may be converted into a list of orthologs in other species. This is particularly useful when seeking to identify genes in an understudied species based on functional information in more intensively characterized species (e.g., essentiality data derived from mutational screens conducted in model organisms).

## Proteomics

VEuPathDB integrates the results of proteomics experiments as peptides aligned to a reference genome or as abundance data assigned to a gene. We do not reanalyze the raw mass spec data but instead use an in-house plugin that loads found peptides or abundance data from tab delimited input files of a specific format.

[Details for the VEuPathDB in-house proteomics pipeline](#)

## RNA-Sequence

VEuPathDB integrates RNA-Seq data from many different experiments and analyzes all data with the same EBI RNA-Seq analysis pipeline. The RNA sequence data that we integrate is processed at EBI.

The following is a general outline of the analysis process.

- Trim poor quality data (Trimmomatic)
- HiSAT2 alignment to a reference genome
- HT-Seq-count to tally aligned reads per gene
- Convert to transcripts per kilobase million (TPM)

- DESeq2 to determine differential expression

[EBI RNA-Seq pipeline details](#)

## ChIP-Sequence

VEuPathDB integrates ChIP-Seq data from many different experiments and sources. DNA seq data are aligned to the reference genome using Bowtie2. Alignment results are converted to bigwig and displayed in JBrowse.

## Copy Number Variation

VEuPathDB uses coverage from whole genome sequencing data to estimate gene and chromosome copy numbers in sequenced strains. The bowtie2 alignments generated during SNP analysis are used as a starting point. HTseq-count is used to count the number of reads that align to each gene and the values are converted to transcripts per million (TPM). Assuming that the median TPM value represents a single copy gene on a chromosome of constitutive ploidy, we can infer gene or chromosome duplications by comparing the TPM values for individual genes or the median TPM for individual chromosomes to the whole genome median using custom scripts based on the method described in [PMID: 22038252](#). Additionally, coverage is calculated in 1 kb bins across the genome, normalised to the constitutive ploidy and converted to bigwig format for visualisation in JBrowse.

Haploid number and gene dose are metrics used to define copy number in VEuPathDB. Haploid number is the number of genes on an individual chromosome. Gene dose is the total number of genes in an organism, accounting for copy number of the chromosome. For example, a single-copy gene in a diploid organism has a haploid number of 1 and a gene dose of 2.

### Searches for genes based on Copy Number Variation

There are two searches that query copy number data in VEuPathDB. The first, [Identify Genes based on Copy Number \(CNV\)](#) returns genes that are present at copy numbers within a range that you specify. The search can be configured to return genes based on the haploid number or gene dose. The second search, [Identify Genes based on Copy Number Comparison \(CNV\)](#), returns genes for which the copy number varies between the reference and your chosen isolates. This search compares the estimated copy number of a gene in the resequenced strain(s) with the copy number in the reference annotation. The copy number in the reference annotation is calculated as the number of genes that are both on the same chromosome and in the same ortholog group as the gene of interest. We infer that these genes have arisen as a result of tandem duplication of a common ancestor. In this search, the metric for copy number is the haploid number, which is the number of copies of a gene on a single chromosome.

## Genetic Variation and SNP calling

VEuPathDB analyzes whole genome resequencing data to call single nucleotide polymorphisms of isolates. The method employed by VEuPathDB to call SNPs from short read sequencing like Illumina reads, follows these steps:

- Reads are aligned to the reference genome using bowtie2
- The resulting BAM file from bowtie2 is sorted and a pileup file using samtools is generated
- Reads around indels are realigned using GATK
- SNPs and indels are called consensus sequence using VarScan is generated:
  - P value  $\leq 0.01$
  - minimum aligned reads  $\geq 5$
  - minimum read frequency  $\geq 0.8$
- SNP calls where coverage is  $>2.5x$  the median coverage are removed to limit erroneous calls in repeat regions
- At each SNP position “like reference” calls are generated for each strain that is identical to the reference to give the full picture of each SNP

## Microarray data

VEuPathDB integrates microarray data from high density oligonucleotide as well as spotted arrays. In general, the data comes to us as intensities associated with probes. VEuPathDB does not reanalyze the original fluorescence data. We process the data we receive according to the following outline:

- Map the array probes to the reference genome's transcriptome
- Filter the data to remove outliers.
- Normalize
  - For one channel data we perform a robust multi-array average (RMA) normalizations.
  - For two channel data we perform a Loess normalization
- Compute the average probe intensity per gene.
- Compute the expression average per gene.
  - First, average the technical replicates.
  - Second, average the biological replicates (if any).
- Optional: perform differential expression analysis if there is a sufficient number of biological replicates.

## Protein Array data

VEuPathDB integrates protein array data from serum antibody microarray experiments. In general, the data comes to us as intensities associated with probes. VEuPathDB does not

reanalyze the original fluorescence data. Although each experiment and data set can have special considerations, we process the data according to the following outline:

- Map the array probes to the reference genome's transcriptome
- Filter the data to remove outliers.
- Normalize
  - For one channel data we perform a robust multi-array average (RMA) normalizations.
  - For two channel data we perform a Loess normalization
- Compute the average probe intensity per gene.
- Compute the expression average per gene.
  - First, average the technical replicates.
  - Second, average the biological replicates (if any).
- Optional: perform differential expression analysis if there is a sufficient number of biological replicates.

## Metabolic Pathways

VEuPathDB integrates metabolic pathways from [KEGG](#) and [MetaCyc](#). For TriTrypDB, pathways are also integrated from [LeishCyc](#) and [TrypanoCyc](#). Metabolic pathways are associated with genes via Enzyme Commission annotations.

## Dataset descriptions

This document provides a high-level overview of the software infrastructure utilized by the VEuPathDB BRC to load, integrate and provide data to users. Please check [a list of all the data sets](#) loaded in our VEuPathDB sites utilizing this infrastructure.

*Link: <https://veupathdb.org/veupathdb/app/search/dataset/AllDatasets/result>*

All loaded datasets are described appropriately with links to publications and repositories as appropriate.

- Data Set (Name)
- Organism(s) (source or reference)
- VEuPathDB Project
- Release # /Date
- Category
- Publications
- Summary
- Contact

## Organisms - Genome info and stats

This document provides a comprehensive overview of the organisms and genome sequence versions and annotation versions within the VEuPathDB BRC.

*Link:*

<https://veupathdb.org/veupathdb/app/search/organism/GenomeDataTypes/result>

All genome sequences and annotations are searchable and described appropriately with links to VEuPathDB organism pages.

- VEuPathDB project
- Organism
- NCBI taxon ID
- Species NCBI taxon ID
- Genome source
- Genome version
- Genome size (Mbp)
- Annotation version
- Total gene count
- Is reference strain indicator?
- Contig count
- Supercontig count
- Chromosome count
- Protein coding gene count
- Non protein coding gene count
- Pseudogene count
- User comment count
- Ortholog count
- Go Terms count

## Technical infrastructure and software documentation:

*Link:* <https://beta.veupathdb.org/veupathdb.beta/app/static-content/infrastructure.html>

The above document describes the overall infrastructure of our data loading system and websites with links to appropriate GitHub repositories for users who want to explore using VEuPathDB infrastructure to build their own genomics database and website.

## Browser Compatibility Statement

We recognize that our users access VEuPathDB using various Internet Browsers and Operating Systems. Our goal is to ensure that you have the best possible experience on VEuPathDB, but

it is impossible to develop applications that work identically, efficiently and effectively on all web browsers.

Based on our site usage statistics we support the following browsers used by greater than 95% of our visitors:

- Firefox
- Safari
- Chrome

Feel free to [contact us](#) about any browsing issues you might come across.

## Data Loading and Database Schema

We use the [Genomics Unified Schema \(GUS\) database schema](#) and data loading infrastructure and its framework available at [GusAppFramework](#). This includes not only a comprehensive database schema for integrating and representing genomic and functional (or post) genomic data but also tools for loading said data into that system. We have made some extensions to the schema and tools for VEuPathDB specific purposes primarily to generate de-normalized views of the data for query optimization purposes.

Our data are all stored in Oracle12c databases. Our software infrastructure also supports PostgreSQL but we have some Oracle specific SQL constructs in our model that would need to be changed in order to run successfully in PostgreSQL.

We load all data using an in house engineered workflow system called [ReFlow](#). Briefly, ReFlow is engineered to be an efficient graph-based workflow system. In it each step (node in the graph) has the ability to be undone and subsequently rerun with updated data. This was a significant requirement as it enables us to undo entire genomes when the annotation or underlying sequence changes. This results in automated removal of all data dependent on that genome. When the step is re-run with the new annotation, all dependent data are recomputed and reloaded automatically, thus greatly improving our ability to keep these complex databases up-to-date.

The ReFlow workflow system utilizes another piece of software developed at the University of Pennsylvania to schedule, manage and monitor running tasks called [DistribJob](#). DistribJob distributes tasks generated from a large input dataset such as a set of sequences to compute nodes in a cluster for analysis and retrieves and collates the results in an efficient manner. We automate the running of large compute tasks on compute clusters located at the University of Pennsylvania and the University of Georgia.

## Code Availability

To facilitate greater transparency and tool reuse, our codebase has been migrated to the GIT repository (<https://github.com/VEuPathDB>).

## Web Presentation System and User Interfaces

Our websites are based on code that we developed and have released to the community called the Strategies-WDK (Strategies Web Development Kit) which enables the graphical strategies search system. You can download the software and see documentation for this toolkit at [Strategies-WDK](#). This toolkit enables us to represent our data as an XML model which is then turned into the web interfaces that are presented to users using these tools.

## Software Code Repository

To facilitate greater transparency and tool reuse, our codebase has been migrated to the GIT repository (<https://github.com/VEuPathDB>).

## System Hardware and Third-Party Software

VEuPathDB maintains redundant database and content web servers at the University of Pennsylvania and the University of Georgia to minimize interruptions for our users during maintenance periods. Additionally VEuPathDB compute and data loading servers are located at the University of Pennsylvania.

Server configurations are coordinated and deployed through Puppet automation software (<http://puppetlabs.com/>). Custom infrastructure software is versioned and deployed through standard RPM/YUM mechanisms. When appropriate, software builds are automated using Jenkins Continuous Integration Server (<http://jenkins-ci.org/>)

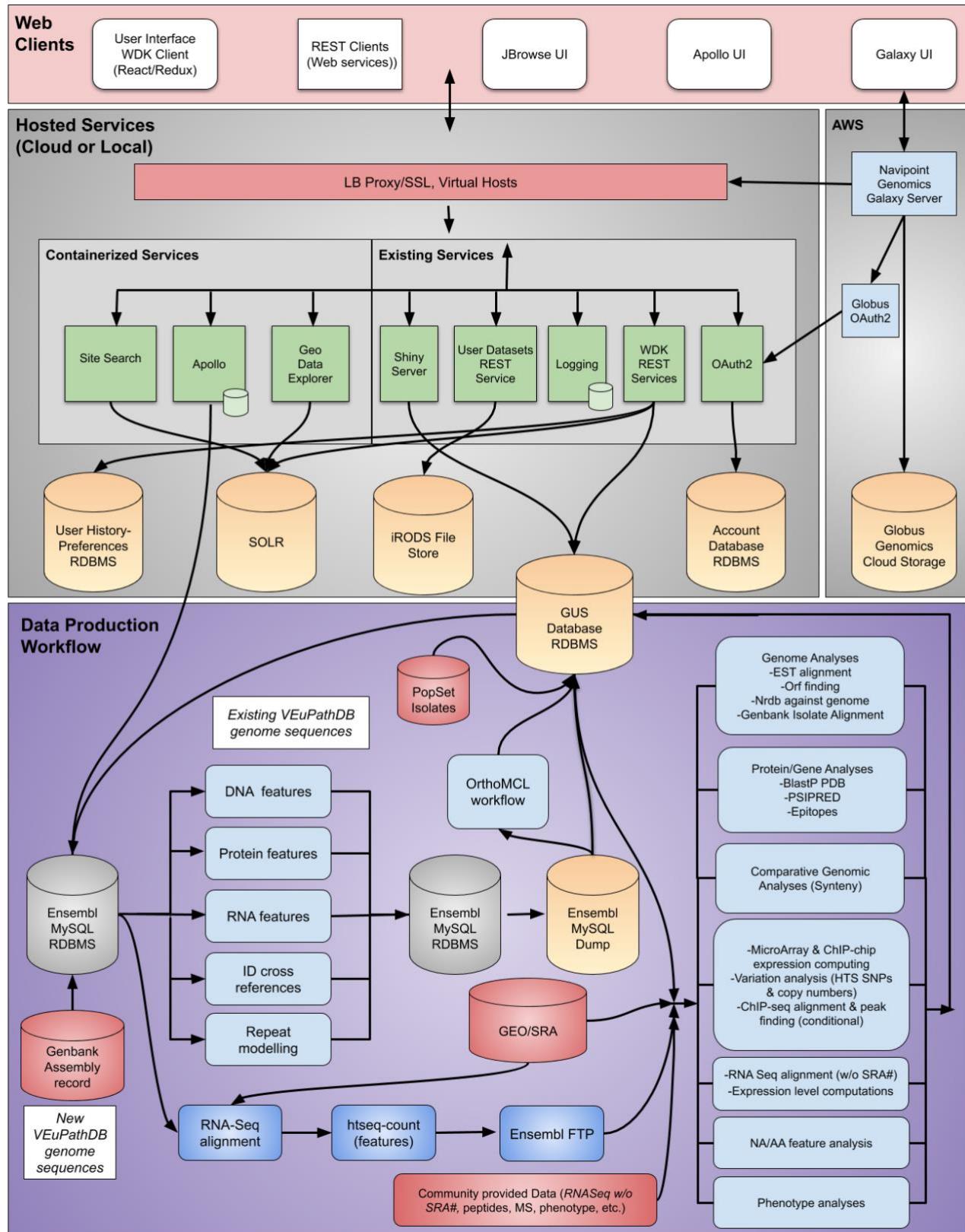
System infrastructure statistics (CPU load, I/O, etc) are gathered with collectd (<http://collectd.org/>) and in-house applications and feed to Graphite (<http://graphite.wikidot.com/>) for human review. Nagios (<http://www.nagios.org/>) provides notifications of system degradations.

Both Universities also maintain large compute clusters that are heavily utilized by VEuPathDB in order to analyze and load incoming data in a timely fashion. The linked document below describes our actual hardware and includes a list of third-party software required in order to analyze, load and present data via our websites.

## Overview of the VEuPathDB Data Production Workflow and Architecture

The complete pathway from data acquisition to web presentation and utilization by users is detailed in the next figure. The data production workflow (bottom, purple rectangle) begins with data acquired from numerous sources (indicated in red) including direct deposition by the community. These data are processed through two main pipelines (indicated in blue), one targeting genome features and RNA-Seq mapping using Ensembl pipelines and the second focused on comparative genomics and analysis of other types of omics-scale data including proteomics and high-throughput phenotypic screens among others. Quality-controlled analysed data are loaded into the GUS database (indicated in gold spanning the purple data production and grey services rectangles). The GUS database links the data to the servers and services (green) and resources that allow the user community to securely access, interrogate, visualize,

download and further analyse data, including their own data via the provided web clients (indicated in the pink rectangle).



# VEuPathDB Website Privacy Policy

We do not use or share any of your personal information for any purpose unrelated to the functionality of the websites; however, we do collect some information to help us understand how our sites are being used and to improve community support.

UPDATED: April 12, 2020

## Introduction

VEuPathDB (also referred to as ‘we’ throughout this document) is committed to protecting its users’ data and privacy. The purpose of this page is to provide you with information about how the data we collect from users of VEuPathDB websites is used or shared. We may update this Privacy Notice from time to time. We encourage you to visit this page frequently and take note of the date updated field above.

We do not use or share any of your personal information for any purpose unrelated to the functionality of the websites; however, we do collect some information to help us understand how our sites are being used in order to improve community support and to enhance the VEuPathDB community’s experience when visiting our sites.

### Information Automatically Collected

When you browse VEuPathDB sites, certain information about your visit will be collected. We automatically collect and store the following type of information about your visit:

- The IP address of the client making the request. Often the IP address is that of your personal computer or smart phone; however, it might be that of a firewall or proxy your internet provider manages.
- The operating system and information about the browser used when visiting the site.
- The date and time of each visit.
- Pages visited.
- The address of a referring page. If you click a link on a website that directs you to a VEuPathDB page, the address of that originating web page will be collected. This “referrer” information is transmitted as part of the browser and server communications; it is not based on any marketing or partnering agreements with the referring site.

This automatically collected information does not identify you personally unless you include personally identifying information in a support form request; see the “Contact Us” policy below for details. We use this information to measure the number of visitors to our site. The aggregate data may be included in prospectuses and reports to funding agencies.

## Information You Directly Provide

The Basket, Favorites, Public Strategies, Gene/Sequence Comment and GBrowse Track features of the VEuPathDB websites require that you register for an account. A valid email address is required so we can send you your temporary account password. An anonymous email service can be used if you do not want to provide personally identifying information.

Your email address will be used to send you infrequent alerts if you subscribe to receive them. We do not sell or distribute email addresses to third parties.

We also ask for your name and institution during account registration. If you add a comment to a Gene or a Sequence, your name and institution will be displayed with the comment. If you make one of your strategies public, your name and institution will be displayed with it. We do not routinely verify the validity of names and institutions associated with comments or public strategies; however, we will delete accounts or comments if we believe them to be fraudulent based on inappropriate activity or posted content. We will not sell or distribute your name or institution to third parties.

When you log in, the client IP address is recorded. This IP address can be correlated with the address automatically collected as noted above. If your user profile personally identifies you, then it may be possible to associate you with your detailed activity on VEuPathDB web sites.

## “Contact Us” Form

The header on each web page includes a “Contact Us” link to a form where users can submit questions, error reports, feature requests, and dataset proposals. Submissions through this form are emailed to VEuPathDB staff and recorded in a project management application accessible only by VEuPathDB staff.

The form includes a field for an email address. If the email address identifies you personally, say if you use your institutional email, then your correspondence with us will likewise be linked to you. A valid email is not strictly required, although we cannot reply to you without one.

When you submit the form, your IP address and browser version will be recorded for internal use. In the case of reported bugs or other site errors, this information may be used by technical staff to help locate your session in the server logs to aid in troubleshooting the issue. This does have the side effect of making it possible to associate an IP address with an email address which may, in turn, personally identify you. However, VEuPathDB does not publicly release this information.

## How VEuPathDB Uses Cookies

VEuPathDB uses cookies to associate multiple requests by your web browser into a stateful session. Cookies are essential to track the state of query strategies, gene baskets and authentication.

Some cookies persist only for a single session. The information is recorded temporarily and is erased when the user quits the session or closes the browser. Others may be persistently stored on the hard drive of your computer until you manually delete them from a browser folder or until they expire, which can be months after they were last used.

Cookies can be disabled in your browser (refer to your browser's documentation for instructions); however, the majority of the website functionality will be unavailable if cookies are disabled.

## Google Analytics

Google Analytics provides aggregate measurements of website traffic including counts of page hits and unique users along with statistics on countries of origin.

The raw measurements and statistics are only available to approved VEuPathDB staff. Aggregated data may be included in prospectuses and reports to funding agencies.

## Third-Party Websites and Applications

Third-party websites and applications are not exclusively operated or controlled by VEuPathDB. By using these third-party websites, individuals may be providing nongovernmental third-parties with access to personally identifying information.

### Twitter

VEuPathDB maintains a presence on Twitter in the form of a [VEuPathDB branded page](#). This page allows for a direct connection with end users to promote information related VEuPathDB services and to disseminate educational information on research publications, news and events related to the biology of eukaryotic pathogens. Postings may also include information about planned service maintenance and outages.

Twitter collects profile information such as name and email address about users who register to use this third-party website. Depending on the user's privacy settings, this information may be displayed on the user's profile page or in the user's tweets which may be retweeted on VEuPathDB's page. The VEuPathDB Twitter account may post the authors and institutions of publicly published scientific papers and news articles. VEuPathDB does not actively collect or maintain personally identifying information through its use of Twitter. VEuPathDB will redact or refrain from retweeting a posting that contains obviously identifiable personal information. A Twitter account is not required to read VEuPathDB postings on Twitter. VEuPathDB does not collect or use personal information outside of Twitter's site.

Twitter is hosted and maintained by a third party which may use browser tracking and related technologies to collect information about visitors to twitter.com and its affiliates. Refer to Twitter's privacy statement, <https://twitter.com/en/privacy>, for more information.

### Facebook

VEuPathDB maintains a presence on Facebook in the form of a [VEuPathDB branded page](#). This page allows for a direct connection with end users to promote information related VEuPathDB services and to disseminate educational information on research publications, news and events related to the biology of eukaryotic pathogens. Postings may also include information about planned service maintenance and outages.

Like Twitter, Facebook collects profile information, including name and email address, from its users. Depending on the user's privacy settings, this information may be displayed on the user's

profile page along with any activity such as comments or “likes” on the VEuPathDB Facebook page or in posts that VEuPathDB shares on Facebook. VEuPathDB does not collect or use any personally identifying information outside of our Facebook page. To understand how Facebook collects and uses personal information, refer to their data policy page, <https://www.facebook.com/policy.php>.

## YouTube

VEuPathDB maintains a presence on YouTube in the form of a [VEuPathDB branded page](#). This page provides tutorials on the use of our websites.

YouTube also requires some information when users create an account, including an email address, and users may choose to provide a name and other identifying information in their public profile. Depending on their individual privacy settings, some personally identifiable information may be available to other users, including VEuPathDB. However, VEuPathDB does not collect or use any of that information outside of its YouTube interactions. You can view videos without signing in to an account, but you must be a registered user in order to comment. As YouTube is a Google service, the VEuPathDB youtube channel is subject to [Google's privacy policy](#).

## Globus Genomics

The VEuPathDB Galaxy Data Analysis Service is a workspace for large-scale data analyses. Developed in partnership with [Globus Genomics](#), workspaces offer a private analysis platform with published workflows and pre-loaded annotated genomes. The workspace is accessed through the “Analyze My Experiment” tab on the home page of any VEuPathDB resource and can be used to upload your own data, compose and run custom workflows, retrieve results and share workflows and data analyses with colleagues.

The VEuPathDB Galaxy Data Analysis Service is hosted by Globus Genomics, an affiliate of Globus. The first time you visit VEuPathDB Galaxy you will be asked to sign up with Globus in order to set up your private Galaxy workspace. Linking your Globus account with your VEuPathDB account is necessary so that input data and analysis results can be transferred between the two systems. We encrypt data transfers and storage, but ultimately, we cannot guarantee the security of data transmissions among VEuPathDB, Globus and affiliates, Amazon Cloud Services, and the user. It is your responsibility to back up your data and obtain any required permissions from your study and/or institution prior to uploading data for analyses on the VEuPathDB Galaxy platform. Do not use, transmit, upload or share any human identifiable information in the files you analyze. VEuPathDB, Globus and affiliates, the University of Georgia, the University of Pennsylvania, the University of Liverpool, and Amazon Cloud Services do not take any responsibility and are not liable for the loss and/or release of any data you analyze via the VEuPathDB Galaxy platform. We encourage you to review the [Globus' privacy policy](#).

## Your Rights based on the General Data Protection Regulation (GDPR)

To read more about GDPR please check the [GDPR website](#).

1. The right of transparency and modalities. The privacy policy should be clear and easy to follow in explaining what data we collect and how we use it.

2. The right to be informed about when data is gathered. This is described in the privacy policy, during the registration process (if you choose to register), site banner and an email sent out to all registered users on May 25, 2018.
3. The right of access. You can ask for what specific data we have about you and how we use it.
4. The right to rectification. We will correct any errors in your personal data that you point out to us.
5. The right to be forgotten. We are happy to delete your account and info when you make such a request.
6. The right to restrict processing. You have the right to request that we restrict the use of your data.
7. The right for notification obligation regarding rectification/erasure/restriction.
8. The right to data portability.
9. The right to object to the processing of your personal data at any time.
10. The right in relation to automated decision making and profiling. Basically, you have the right not to be subject to decisions based solely on automated processing which significantly affect you.

To make any of the above stated requests or if you have any questions please email us at [help@VEuPathDB.org](mailto:help@VEuPathDB.org)

## VEuPathDB Accessibility Conformance

We strive to make the VEuPathDB website accessible. Wherever possible, our sites have alternative text for images, the pages can be magnified and read by screen readers. The results of some searches are less accessible as they are dynamic and user-specific, thus alternative text describing images cannot be generated. We update our Voluntary Product Accessibility Template, VPAT, 508 accessibility report annually. The more recent version is located at: [https://veupathdb.org/documents/VEuPathDB\\_Section\\_508\\_BRC4.pdf](https://veupathdb.org/documents/VEuPathDB_Section_508_BRC4.pdf)

## VEuPathDB personnel

### VEuPathDB Management

Beatrice Amos, Annotation Manager  
Cristina Aurrecoechea, User Interface and Portal Manager  
Bob Belnap, Systems and Databases Manager  
John Brestelli, Data Development Manager  
Brian Brunk, VEuPathDB Senior Manager  
George Christophides, Co-I, NIAID BRC Contract  
Kathryn Crouch, Co-I, Wellcome Trust  
Jeremy DeBarry, VEuPathDB Project Manager  
Steve Fischer, Software and Infrastructure Manager  
Paul Flückeck, Co-I, NIAID BRC Contract  
Omar Harb, Director of Scientific Outreach & Education  
Andy Jones, Wellcome Trust PI; Co-I NIAID BRC Contract  
Jessica C Kissinger, Joint-PI, NIAID BRC Contract; WT Co-PI  
Wei Li, Data Loading Manager

Gareth Maslen, EBI Data Processing Manager  
Mary Ann McDowell, Joint-PI, NIAID BRC Contract  
David S Roos, Joint-PI, NIAID BRC Contract; WT Co-PI  
Chris J Stoeckert, Co-I, NIAID BRC Contract

To contact any one of us please use the [contact us form](#).

## Current VEuPathDB Team members

Beatrice Amos<sup>4</sup>, Rachel Ankirskiy<sup>1</sup>, Cristina Aurrecoechea<sup>1</sup>, Matthieu Barba<sup>9</sup>, Ana Barreto<sup>3</sup>, Evelina Basenko<sup>4</sup>, Wojtek Bazant<sup>2</sup>, Dan Beiting<sup>2</sup>, Bob Belnap<sup>1</sup>, Ann Blevins<sup>2</sup>, Ulrike Böhme<sup>4</sup>, John Brestelli<sup>3</sup>, Brian Brunk<sup>2</sup>, Danielle Callan<sup>2</sup>, Mikkel Christensen<sup>9</sup>, George Christophides<sup>8</sup>, Kathryn Crouch<sup>6</sup>, Katie Cybulski<sup>7</sup>, Jeremy DeBarry<sup>1</sup>, Ryan Doherty<sup>3</sup>, Yikun Duan<sup>2</sup>, Dave Falke<sup>1</sup>, Steve Fischer<sup>3</sup>, Paul Flicek<sup>9</sup>, Bindu Gajria<sup>2</sup>, Gloria I. Giraldo-Calderón<sup>7</sup>, Omar S. Harb<sup>2</sup>, Elizabeth Harper<sup>2</sup>, Danica Helb<sup>2</sup>, Mark Hickman<sup>2</sup>, Connor Howington<sup>7</sup>, Sufen Hu<sup>2</sup>, Jay Humphrey<sup>1</sup>, John Iodice<sup>3</sup>, Andy Jones<sup>4</sup>, John Judkins<sup>2</sup>, Sarah Kelly<sup>8</sup>, Jessica C. Kissinger<sup>1</sup>, Dae Kun Kwon<sup>7</sup>, Kris Lamoureux<sup>1</sup>, Wei Li<sup>2</sup>, Disha Lodha<sup>9</sup>, Jamie Long<sup>2</sup>, Bob MacCallum<sup>8</sup>, Gareth Maslen<sup>9</sup>, Mary Ann McDowell<sup>7</sup>, Greg Milewski<sup>2</sup>, Jarek Nabrzyski<sup>7</sup>, David S. Roos<sup>2</sup>, Samuel Rund<sup>7</sup>, Steph Wever Schulman<sup>2</sup>, Achchuthan Shanmugasundram<sup>4</sup>, Vasili Sitnik<sup>9</sup>, Drew Spruill<sup>1</sup>, David Starns<sup>4</sup>, Christian J. Stoeckert Jr.<sup>3</sup>, Sheena Shah Tomko<sup>2</sup>, Haiming Wang<sup>1</sup>, Susanne Warrenfeltz<sup>1</sup>, Robert Wieck<sup>7</sup>, Paul Wilkinson<sup>4</sup>, Mariann Winkelmann<sup>2</sup>, Lin Xu<sup>2</sup>, Jie Zheng<sup>3</sup>.

## Previous VEuPathDB Team members, 2004-2020

Antelmo Aguilar<sup>7</sup>, James Allen<sup>9</sup>, Alexis Allot<sup>9</sup>, Nora Besansky<sup>7</sup>, Austin Billings<sup>2</sup>, Sanjay Boddu<sup>9</sup>, Steve Bogol<sup>7</sup>, Ewan Birney<sup>9</sup>, Andrew Brockman<sup>8</sup>, Robert Bruggner<sup>7</sup>, Ja'Shon Cade<sup>3</sup>, Mark Caddick (Principal Investigator, WT 2019-2020)<sup>4</sup>, David Campbell<sup>7</sup>, Cristian Cocos<sup>2</sup>, Frank Collins (VectorBase Principal Investigator, 2004-2018)<sup>7</sup>, Kathy Couch<sup>1</sup>, Greg Davies<sup>7</sup>, Elaine Daugan<sup>4</sup>, Ale Diaz Miranda<sup>2</sup>, Emmanuel Dialynas, Jennifer Dommer<sup>3</sup>, Vicky Dritsou, Scott Emrich<sup>10</sup>, Xin Gao<sup>2</sup>, William Gelbart<sup>12</sup>, Sandra Gesing<sup>7</sup>, Alan Gingle<sup>1</sup>, Greg Grant<sup>3</sup>, Matt Guidry<sup>1</sup>, Martin Hammond<sup>9</sup>, Mark Heiges<sup>1</sup>, Christiane Hertz-Fowler (Principal Investigator, WT 2008-2019)<sup>4</sup>, Nicholas Ho<sup>8</sup>, Daniel Hughes<sup>9</sup>, Frank Innamorato<sup>3</sup>, San James<sup>14</sup>, Amie Jaye<sup>8</sup>, Fotis Kafatos<sup>8</sup>, Paul Kersey<sup>9</sup>, Ioannis Kimitzoglou<sup>8</sup>, Nathan Konopinski<sup>7</sup>, Carolyn Knoll<sup>2</sup>, Eileen T. Kraemer<sup>1</sup>, Nick Langridge<sup>9</sup>, Cris Lawrence<sup>2</sup>, Daniel Lawson<sup>8</sup>, Brianna Lindsay<sup>2</sup>, Neil Lobo<sup>7</sup>, Christos (Kitsos) Louis<sup>11</sup>, Ross Madden<sup>6</sup>, Greg Madey<sup>7</sup>, Elisabetta Manduchi<sup>3</sup>, Karine Megy<sup>9</sup>, John A. Miller<sup>6</sup>, Elvira Mitraka<sup>11</sup>, Vishal Nayak<sup>3</sup>, Cary Pennington<sup>1</sup>, Deborah F. Pinney<sup>3</sup>, Brian Pitts<sup>1</sup>, Jane A. Pulman<sup>4</sup>, Caleb Reinking<sup>7</sup>, Seth Redmon, Chris Ross<sup>1</sup>, Andrew Sheehan<sup>7</sup>, Fatima Silva<sup>4</sup>, Ganesh Srinivasamoorthy<sup>1</sup>, Scott Szakonyi<sup>7</sup>, Pantelis Toplais<sup>11</sup>, Ryan Thibodeau<sup>1</sup>, Charles Treatman<sup>2</sup>, Betsy Wenthe<sup>1</sup>, Matt Vander Werf<sup>7</sup>, Maggie Werner-Washburne<sup>13</sup>, Patricia L. Whetzel<sup>3</sup>, Derek Wilson<sup>9</sup>, Andrew Yates<sup>9</sup>

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- All the community members who have contributed data (often pre-publication), entered user comments or sent us their suggestions.
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**AmoebaDB:**

William Petri

Serge Ankri

Craig Roberts

Fiona Henriquez

Hugo Aguilar-Díaz

**CryptoDB:**

Boris Striepen

**FungiDB:****ZyGoLife**

Jason Stajich

Zachary Lewis

**GiardiaDB:**

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Tineke Lauwaet

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Scott Dawson

**MicrosporidiaDB:**

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Pattana Jaroenlak (Michael)

Damian Ekiert

Michael Cammer

**PiroplasmaDB:**

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Lowell Kappemeyer

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Brendan Elsworth

Caroline Keroack

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**PlasmoDB:**

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Pedro Moura

**ToxoDB:**

David Roos

**TrichDB:**

Antonio Pereira-Neves

Marlene Benchimol

**TriTrypDB:**

Rick Tarleton

Richard Wheeler

Leandro Lemgruber Soares

Margaret Mullin

Camila Silva Gonçalves

Wanderley de Souza

Maria Cristina Machado Motta

## VEuPathDB Community Representatives

VEuPathDB encourages community members to provide feedback about our resources. We get feedback from many community members including those listed below who have been active in our open community meetings. We encourage you to get involved. Feel free to [contact us](#) any time.

**Amoeba:** Open community call and Carol Gilchrist, Upi Singh

**Cryptosporidium:** Gregory Buck, Guy Robinson, Karin Troell, Sumiti Vinayak, Jonathon Wastling, Giovanni Widmer, Lihua Xiao, Guan Zhu

**Fungi:** Bridget Barker, Elaine Bignell, Katherine Borkovich, Michael Bromley, Christina Cuomo, Tamara Doering, Jay Dunlap, Michael Freitag, Louise Glass, Kim Hammond-Kosack, Guilhem Janbon, Seogchan Kang, Theo Kirkland, Corby Kistler, Jennifer Lodge, Robin May, Jessie Uehling, Sinem Beyhan, Douglas Lake, Natalie Mitchell, Maureen Donlin, Vera Meyer, Marc Orbach, Nadia Potts, Antonis Rokas, Jason Stajich, Matt Sachs, George R. Thompson, Martin Urban, Nathan P. Wiederhold

**Giardia:** Scott Dawson, Fran Gillin, Adrian Hehl, Aaron Jex, Hilary Morrison, John Samuelson, Cornelia Spycher, Staffan Svard

**Microsporidia:** James Becnel, Nicolas Corradi, Elizabeth Didier, Patrick Keeling, Emily Troemel, Louis Weiss

**Piroplasma:** Open community call and Choukri Mamoun

**Plasmodium:** John Adams, Chris Janse, Rays H.Y. Jiang, Shahid Khan, Stuart Ralph, Akhil Vaidya, Andy Waters

**Toxoplasma:** John Boothroyd, Jon P. Boyle, Vern B. Carruthers, Marc-Jan Gubbels, Kami Kim, Markus Meissner, Jeroen Saeij, Lilach Sheiner, Ross Waller, Michael White

**Trichomonas:** Jane Carlton, Patricia Johnson, Steven Sullivan, Jan Tachazy

**Trypanosoma/Kinetoplastids:** Fernan Aguero, Vivian Bellofatto, Richard Burchmore, George Cross, Angela Cruz, Antonio Estevez, Mark Field, Catarina Gadelha, Eva Gluenz, Keith Gull, John Kelly, Annette MacLeod, Jeremy Mottram, Torsten Ochsenreiter, Marc Ouellette, Barbara Papadopoulou, Laurie Read, Sergio Schenkman, Rick Tarleton, Brent Weatherly, Bill Wickstead, Michael (Mick) Urbaniak

**Vectors:** Gregory Dasch, Jeff Grabowski, María de Lourdes Muñoz, Monika Gulia-Nuss, Sukanya Narasimhan, Kristin Michel, Michael Povelones, Igor Sharakhov, Ronald van Rij, Rob Waterhouse

## Previous Scientific Working Group

VEuPathDB wishes to acknowledge previous scientific working group members. They provided regular feedback oversight and guidance.

Lyric Bartholomay	Michael Gottlieb	Malcolm McConville	John Taylor
Matt Berriman	Keith Gull	Nicola Mulder	Jake Tu
Bill Black	Matthew Hahn	Ulli Munderloh	Brett Tyler
John Boothroyd	Adrian Hehl	Daniel Neafsey	Kenneth Vernick
Greg Buck	Steve Higgs	Kenneth Olson	Sarah Volkman
Geraldine Butler	Catherine Hill	Bill Petri	Jonathan Wastling

Angela Cruz	Marcelo Jacobs-Lorena	Barry Pittendrigh	Scott Weaver
George Dimopoulos	Anthony (Tony) James	Jeffrey Powell	Louis Weiss
Martin Donnelly	Pedro Lagerblad de Oliveira	Hillary Ranson	Dyann Wirth
Patrick Duffy	Greg Lanzaro	Alexander Raikhel	Jennifer Wortman
Pascale Gaudet	Daniel Masiga	Lincoln Stein	Guilyan Yan

## Website usage statistics

We collect project-specific website usage statistics using the program awstats. The links to all statistics (they are continually updated) are listed below. A sample website usage statistics report, as prepared by awstats is shown for an example of the types of data that are available. Users can visit the site and select custom reporting periods. A monthly usage statistics report is available online at all VEuPathDB sites in the About-> Technical Information menu.

### Website usage links:

- <https://amoebadb.org/awstats/awstats.pl>
- <https://cryptodb.org/awstats/awstats.pl>
- <https://fungidb.org/awstats/awstats.pl>
- <https://giardiadb.org/awstats/awstats.pl>
- <https://hostdb.org/awstats/awstats.pl>
- <https://microsporidiadb.org/awstats/awstats.pl>
- <https://piroplasmadb.org/awstats/awstats.pl>
- <https://plasmodb.org/awstats/awstats.pl>
- <https://toxodb.org/awstats/awstats.pl>
- <https://trichdb.org/awstats/awstats.pl>
- <https://tritrypdb.org/awstats/awstats.pl>
- <https://orthomcl.org/awstats/awstats.pl>
- <https://veupathdb.org/awstats/awstats.pl>
- <https://vectorbase.org/awstats/awstats.pl>

### Sample awstats report from FungiDB.org

**Statistics for:** fungidb.org

Last Update: 17 Sep 2020 - 01:00  
Reported period: Sep 2020

**Summary**

**When:**  
Monthly history  
Days of month  
Days of week  
Hours  
**Who:**  
Countries  
Full list  
**Navigation:**  
File type  
Downloads  
Full list  
Viewed  
Full list  
Entry  
Exit  
**Operating Systems:**  
Versions  
Unknown  
**Browsers:**  
Versions  
Unknown  
Screen sizes  
**Others:**  
Miscellaneous

Reported period	Month Sep 2020
First visit	01 Sep 2020 - 00:01
Last visit	17 Sep 2020 - 00:59
Unique visitors	5,080
Number of visits	743,449
Pages	(146.34 Pages/Visit)
Hits	1,034,297
Bandwidth	(203.6 Hits/Visit) (4882.72 KB/Visit)
Viewed traffic *	2,423
(2.09 visits/visitor)	
Not viewed traffic *	291,199
	334,657
	9.01 GB

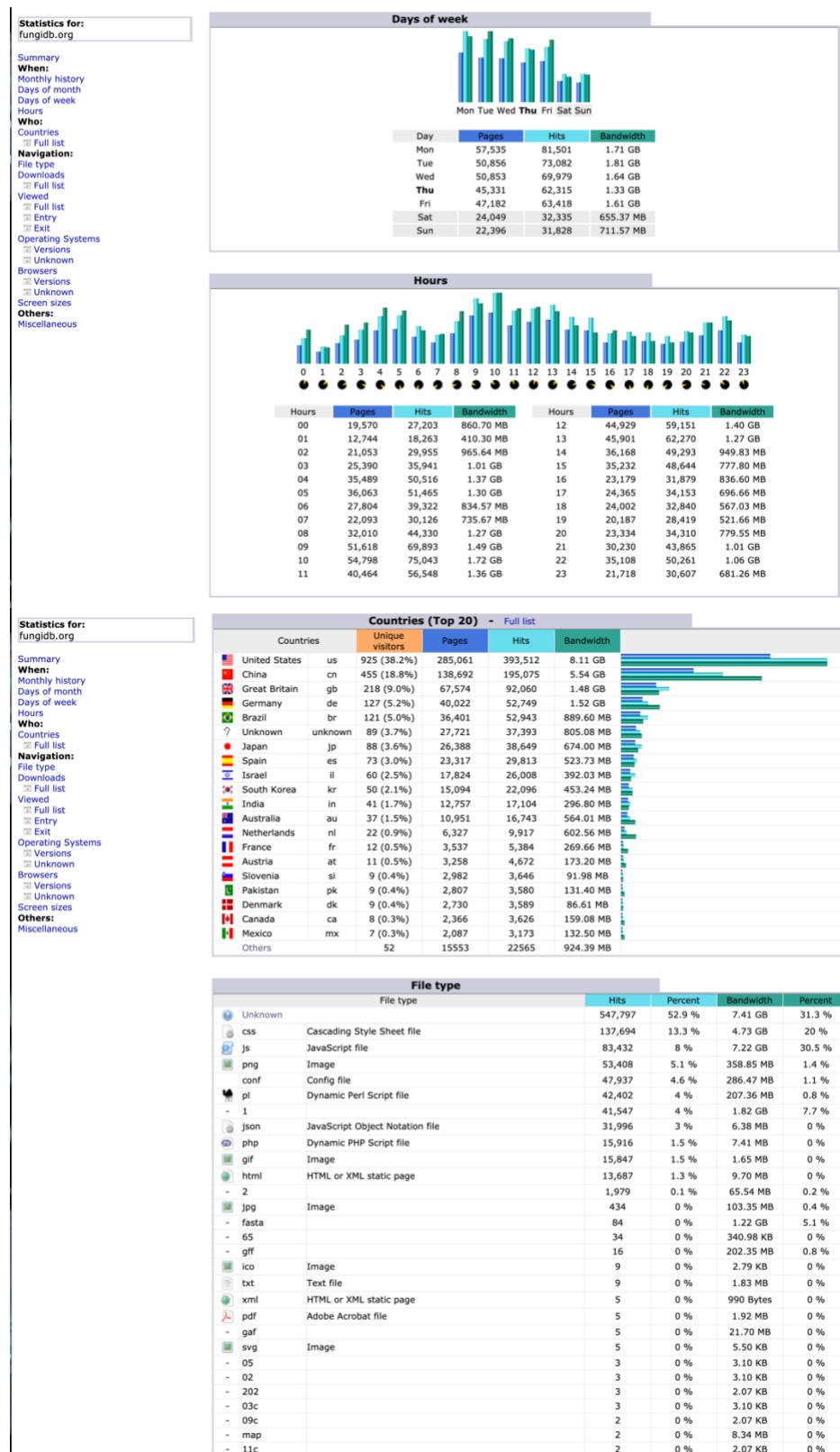
\* Not viewed traffic includes traffic generated by robots, worms, or replies with special HTTP status codes.

**Monthly history**

Month	Unique visitors	Number of visits	Pages	Hits	Bandwidth
Jan 2020	3,644	7,822	1,143,872	1,387,925	29.51 GB
Feb 2020	4,429	9,010	847,864	1,044,521	23.63 GB
Mar 2020	3,965	7,925	1,052,667	1,237,807	47.52 GB
Apr 2020	3,807	8,657	1,260,543	1,476,644	27.14 GB
May 2020	4,626	10,120	1,474,243	1,739,026	34.97 GB
Jun 2020	4,296	9,220	1,245,047	1,488,054	30.23 GB
Jul 2020	4,189	9,495	1,417,204	1,679,054	38.86 GB
Aug 2020	3,826	8,711	1,213,322	1,467,604	31.25 GB
<b>Sep 2020</b>	<b>2,423</b>	<b>5,080</b>	<b>743,449</b>	<b>1,034,297</b>	<b>23.66 GB</b>
Oct 2020	0	0	0	0	0
Nov 2020	0	0	0	0	0
Dec 2020	0	0	0	0	0
Total	35,205	76,040	10,398,211	12,554,932	286.77 GB

**Days of month**

Day	Number of visits	Pages	Hits	Bandwidth
01 Sep 2020	304	56,276	79,716	1.68 GB
02 Sep 2020	333	47,162	65,008	1.68 GB
03 Sep 2020	370	63,508	89,315	2.08 GB
04 Sep 2020	356	43,424	57,688	2.02 GB
05 Sep 2020	181	17,145	22,268	540.29 MB
06 Sep 2020	222	24,581	34,378	798.19 MB
07 Sep 2020	334	43,870	59,084	1.62 GB
08 Sep 2020	336	47,544	65,825	1.99 GB
09 Sep 2020	437	52,328	72,512	1.96 GB
10 Sep 2020	420	71,755	96,542	1.87 GB
11 Sep 2020	344	50,941	69,148	1.20 GB
12 Sep 2020	204	30,954	42,403	770.46 MB
13 Sep 2020	194	20,212	29,278	624.94 MB
14 Sep 2020	379	71,201	103,919	1.80 GB
15 Sep 2020	344	48,749	73,707	1.77 GB
16 Sep 2020	312	53,069	72,418	1.26 GB
<b>17 Sep 2020</b>	<b>10</b>	<b>732</b>	<b>1,088</b>	<b>47.24 MB</b>
18 Sep 2020	0	0	0	0
19 Sep 2020	0	0	0	0
20 Sep 2020	0	0	0	0
21 Sep 2020	0	0	0	0
22 Sep 2020	0	0	0	0
23 Sep 2020	0	0	0	0
24 Sep 2020	0	0	0	0
25 Sep 2020	0	0	0	0
26 Sep 2020	0	0	0	0
27 Sep 2020	0	0	0	0
28 Sep 2020	0	0	0	0
29 Sep 2020	0	0	0	0
30 Sep 2020	0	0	0	0
Average	298	43,732	60,841	1.39 GB
Total	5,080	743,449	1,034,297	23.66 GB



**Statistics for:** fungidb.org

**When:**  
 Summary  
 Monthly history  
 Days of month  
 Days of week  
 Hours  
**Who:**  
 Countries  
 IP addresses  
**Navigation:**  
 Full list  
 Downloads  
 Full list  
 Viewed  
 Full list  
 Entry  
 Exit  
 Operating Systems  
 Versions  
 Unknown  
 Browsers  
 Versions  
 Unknown  
 Screen sizes  
**Others:**  
 Miscellaneous

**Downloads (Top 10) - Full list**

Downloads	Hits	206 Hits	Bandwidth	Average size
/common/downloads/release-41/CalbicansSC5314/txt/FungiDB-41_Calb...	2	0	820.91 KB	410.45 KB
/common/downloads/release-39/AcarbanusarITEM5010/txt/FungiDB-39...	2	0	1.33 MB	679.82 KB
/common/downloads/release-39/SpunktatusDAOMBR117/txt/FungiDB-39...	2	0	1.31 MB	672.60 KB
/common/downloads/release-39/CoposadasicC735deltSOWgp/txt/FungiDB...	2	0	979.30 KB	489.65 KB
/common/downloads/release-39/CgatticCA1873/txt/FungiDB-39_Cgatt...	2	0	818.65 KB	409.32 KB
/common/downloads/release-39/Alumigatusf293/txt/FungiDB-39_Alum...	2	0	1.21 MB	621.91 KB
/common/downloads/release-39/CneoformansK99/txt/FungiDB-39_Cneo...	2	0	923.01 KB	461.50 KB
/common/downloads/release-36/CposadasiiRMSCC3488/txt/FungiDB-36...	2	0	987.31 KB	493.66 KB
/common/downloads/release-39/AculeatusATCC16872/txt/FungiDB-39_...	2	0	1.38 MB	707.43 KB
/common/downloads/release-41/Ferticillioides7600/txt/FungiDB-41...	2	0	1.83 MB	937.20 KB

**Pages-URL (Top 10) - Full list - Entry - Exit**

Viewed	Average size	Entry	Exit
20,154 different pages-url			
/cgi-bin/dataPlotter.pl	42,360	5.01 KB	6
/a/service/browse/stats/global	36,358	67 Bytes	4
/fungidb/service/record-types/gene/records	20,349	15.11 KB	43
/a/browse/browse.conf	15,483	539 Bytes	7
/a/browse/browse_conf.json	15,478	230 Bytes	11
/a/browse/functions.conf	15,425	16.22 KB	3
/a/browse/	13,016	737 Bytes	8
/site-search	12,923	6.54 KB	219
/fungidb/services/WsfService	10,877	38.12 KB	56
/fungidb/service/	9,366	495 Bytes	171
Others	551,814	19.02 KB	4,552
			4,314

**Operating Systems (Top 10) - Full list/Versions - Unknown**

Operating Systems	Pages	Percent	Hits	Percent
Windows	458,583	61.6 %	632,903	61.1 %
Macintosh	238,971	32.1 %	344,467	33.3 %
Linux	31,325	4.2 %	40,875	3.9 %
Unknown	10,944	1.4 %	10,952	1 %
Unknown Unix system	2,333	0.3 %	3,313	0.3 %
iOS	1,285	0.1 %	1,777	0.1 %
BSD	6	0 %	10	0 %
BlackBerry	1	0 %	1	0 %
Sun Solaris	1	0 %	1	0 %

**Browsers (Top 10) - Full list/Versions - Unknown**

Browsers	Grabber	Pages	Percent	Hits	Percent
Google Chrome	No	489,742	65.8 %	688,488	66.5 %
Firefox	No	134,227	18 %	178,981	17.3 %
Safari	No	79,061	10.6 %	117,729	11.3 %
Edge	No	26,909	3.6 %	34,104	3.2 %
Unknown	?	10,935	1.4 %	10,941	1 %
Opera	No	1,676	0.2 %	2,857	0.2 %
MS Internet Explorer	No	562	0 %	836	0 %
Mozilla	No	290	0 %	311	0 %
Android browser (PDA/Phone browser)	No	29	0 %	31	0 %
Netscape	No	15	0 %	18	0 %
Others		3	0 %	3	0 %

**Screen sizes (Top 10)**

Screen sizes	Percent

**Miscellaneous**

Miscellaneous	
Javascript disabled	-
Browsers with Java support	-
Browsers with Macromedia Director Support	-
Browsers with Flash Support	-
Browsers with Real audio playing support	-
Browsers with Quicktime audio playing support	-
Browsers with Windows Media audio playing support	-
Browsers with PDF support	-

Advanced Web Statistics 7.7 (build 20180105) - Created by awstats (plugins: geolip)

## VEuPathDB Glossary

Please also check the [NCBI glossary](#)

- **3-Frame translation (forward)**

[Translation](#) of a nucleotide sequence in all three possible reading frames in one direction, usually "on the top [strand](#)" of DNA.

- **3-Frame translation (reverse)**

[Translation](#) of a nucleotide sequence in all three possible reading frames in the reverse direction, usually "on the bottom [strand](#)" of DNA.

- **AA sequence**

Amino acid sequence.

- **Affymetrix genotyped SNP probes**

Probes on Affymetrix [SNP](#) (single nucleotide polymorphism) arrays, which are used for [SNP genotyping](#). See [Affymetrix microarray technology](#) and [www.affymetrix.com](#)

- **Affymetrix microarray technology**

[Microarray](#) manufacturing technology developed by Affymetrix. Combines semiconductor fabrication techniques, solid phase chemistry, combinatorial chemistry, molecular biology, and robotics to generate a photolithographic manufacturing process in which oligonucleotides are synthesized directly on a chip. See [www.affymetrix.com](#)

- **Affymetrix probes**

Probe on an Affymetrix [microarray](#) designed to determine whether or not the complementary sequence of RNA or DNA is present in a sample. Generally 25 nucleotides in length (25-mers), their short length provides higher specificity than longer probes. See [Affymetrix microarray technology](#) and [www.affymetrix.com](#)

- **Amitochondriate**

Eukaryotic organism that lacks a [mitochondrion](#). Examples include Giardia and other parasites such as Trichipleistophora and Entamoeba. However, most of these organisms contain what appear to be mitochondrial remnants as well as mitochondrial [genes](#) in their nuclear genomes.

- **Annotation**

Identified feature within a sequence, such as a known or predicted [gene](#), domain, motif, post-translational modification, etc.

- **Annotation density**

Level to which a nucleotide or protein sequence has been annotated. See [Annotation](#).

- **ApiCyc**

Database/utility on EuPathDB used for searching and visualizing metabolic pathway information for organisms in EuPathDB; derivative database generated by analyzing various genomes (for example from Plasmodium, Cryptosporidium, and Toxoplasma) with SRI International's pathway tools.

- **ApiDots alignments**

Consensus sequences found in the ApiDots database and generated by clustering and assembling Apicomplexan mRNA and [EST](#) sequences. These consensus sequences were subjected to database searches against protein and protein domain sequences.

- **Apicoplast**

Nonphotosynthetic plastid found in almost all protozoan parasites belonging to the phylum Apicomplexa that have been examined. The apicoplast is surrounded by four membranes, giving rise to the theory that its presence in the Apicomplexa is the result of a secondary endosymbiosis (acquired by the engulfment of an ancestral alga and retention of the algal plastid). Similar to other endosymbiotic organelles (mitochondria, chloroplasts), the apicoplast contains its own genome as well as proteins that are encoded in the nucleus and post-translationally imported. The apicoplast is a vital organelle to the parasite's long-term survival.

- **Attribute**

Inherent characteristic or feature; in a database, a data item related to a database object. For example, attributes of [genes](#) can include features such as introns and untranslated regions (UTRs).

- **BLAST**

Basic local alignment search tool, a [sequence similarity](#) search tool used to quickly find local alignments between a [query](#) sequence and sequences in nucleotide or protein databases. Different versions of this search tool are available to match the types of query sequence and database used. See [blastn](#), [blastp](#), [blastx](#), [tblastn](#), and [tblastx](#).

- **Boolean**

System of logical thought developed by George Boole (1815-1864). In Boolean searching, an "and" operator between two words or values (for example, "apple AND orange") generates a search for items in a database containing both of the words or values. Similarly, an "or" operator between two words or values (for example, "apple OR orange") generates a search for items containing either word.

- **CDS**

Coding sequence. Region of nucleotides that corresponds to the sequence of amino acids in a predicted protein and that includes start and stop codons. Unexpressed sequences (for example, the 5'-UTR, the 3'-UTR, and introns) are not included within a CDS. The CDS usually does not correspond to the actual mRNA sequence.

- **Centromere**

Region of the [chromosome](#) or chromosomal structure essential for division and retention of the chromosome within the cell; point of a chromosome where the spindle fibers attach to pull the chromosome apart during cell division.

- **Chromosome**

Macromolecule of DNA constituting the physical organization of DNA in a cell.

- **Coil**

Three-dimensional spiral structure in protein macromolecules.

- **Contig**

Contiguous genomic sequence assembled from overlapping primary sequences representing overlapping regions of a particular [chromosome](#).

- **CryptoCyc**

Database/utility built by analyzing the Cryptosporidium genome with SRI International's pathway tools; used for searching and visualizing Cryptosporidium metabolic pathway information.

- **Curated annotation**

[Annotation](#) made under the supervision of a curator as opposed to a purely computational prediction. Curated predictions often contain combinations of different types of evidence to support the annotation.

- **DNA/GC content**

Content of guanine (G) and cytosine (C) in a fragment of DNA or a genome. Because GC pairs are more thermostable compared to the AT pairs, it was commonly believed that GC content played a vital part in adaptation to high temperatures, a hypothesis that has been refuted. In the genome browser, the DNA/GC content track displays a GC content graph of the reference sequence at low magnifications and the DNA sequence itself at higher magnifications.

- **Dalton**

Unit of mass abbreviated Da and used to express atomic and molecular masses.

- **Deprecated gene**

[Genes](#) with little or no evidence (similarities / expression) that overlapped (or were subsumed by) larger genes for which there was evidence such as protein similarities, expression evidence from [EST alignments](#), SAGE or [proteomics](#) data were marked as deprecated. These genes will likely be removed in a subsequent release (and in GenBank) unless additional evidence is provided indicating they should be moved into the real gene category.

- **EC numbers**

Enzyme Commission numbers. EC numbers constitute the numerical classification scheme for enzymes based on the chemical reactions they catalyze. EC numbers do not refer to the enzymes, but to the reactions they catalyze.

- **EST**

Expressed sequence tag. Short (typically 100-500 base pairs) partial [cDNA](#) produced by single-shot sequencing of a cloned mRNA (cDNA) and often used to identify [gene](#) transcripts.

- **EST alignments**

Alignments of expressed sequence tags (ESTs) with a corresponding genomic region. For example, in ToxoDB you can visualize [EST](#) alignments by clicking on the "View this sequence in the genome browser" link and turning on the EST Alignments track. Useful for identifying intron boundaries.

- **EST clusters**

Groups of homologous, overlapping [EST](#) sequences created to reduce redundancy of the EST database.

- **Expression level**

Level at which an mRNA or protein is present in a sample. Value can be absolute or relative to other mRNA or protein species in the sample.

- **Expression profile**

Pattern of expression of one or more [genes](#) or proteins over time or over a set of experimental conditions (for example, during development or treatment, or as a result of a genetic mutation such as a knockout).

- **Expression profile correlation**

Method for correlation of [gene expression profiles](#) with gene ontology (GO) [annotations](#) developed for the purpose of identifying groups of genes, pathways, and processes reacting in concert to experimental perturbations.

- **Expression timing**

Timing of [gene](#) expression during a developmental, metabolic, regulatory, or other biological process or response.

- **GBrowse**

Interactive genome browser developed by the Generic Model Organism Database (GMOD) project ([www.gmod.org](http://www.gmod.org)) that can be customized to show selected chromosomal features as well as display user-provided [annotations](#).

- **GBrowse track**

In the [GBrowse](#) viewer, a line of data that corresponds to a particular type of genomic information or feature and that is distinguished by a particular shape or color.

- **Gametocyte**

Eukaryotic germ cell that divides by mitosis to generate other gametocytes or by meiosis to generate gametes. Male gametocytes are called spermatocytes, and female gametocytes are called oocytes. Term often used to describe gametocytes of Plasmodium.

- **GenBank protein record**

Protein sequence file in the GenBank database generally derived by [translation](#) of a related nucleotide record.

- **GenPept protein**

Protein record from the GenPept database at the [NCBI](#) GenBank, which contains inferred [translations](#) of [protein-coding](#) sequences.

- **Gene**

Fundamental physical and functional unit of heredity. Ordered sequence of nucleotides located in a particular position on a particular [chromosome](#) that encodes a specific functional product, such as a protein or RNA molecule. A gene may have a number of parts, including the [promoter](#) region, untranslated regions (5' and 3' [UTRs](#)), introns, and exons.

- **GeneDB**

Project developed by the Sanger Institute Pathogen Sequencing Unit (PSU) and aimed at developing and maintaining curated database resources for all projects handled by the PSU. The database is accessible at [www.genedb.org](http://www.genedb.org)

- **Genetic markers**

Known DNA sequences that can be identified by a simple assay. Generally genetic variations caused by mutation or alterations in loci that can be observed, examples include restriction length polymorphisms (RFLPs), short tandem repeats (STRs), variable number tandem repeats (VNTRs), short DNA sequences surrounding single base-pair changes (single nucleotide polymorphisms, or [SNPs](#)), or longer [microsatellite](#) sequences.

- **Genomic context**

Location of a [gene](#) in the genome, which can influence the expression of the gene and functional interactions of the gene expression products. In this database, genes are depicted on individual gene pages with their surrounding genomic region and [annotations](#).

- **Genome Sequence**

The inherited nucleic acid component of the genome. In eukaryotes, organized into linear [chromosomes](#) that are capped with [telomeres](#) and located in the nucleus.

- **Genotyped SNPs**

Single nucleotide polymorphisms (SNPs) identified during [genotyping](#) of individual organism strains. See [SNP genotyping](#).

- **Genotyping**

Process of determining the genotype of an individual with a biological assay using PCR, DNA sequencing, or hybridization to DNA [microarrays](#) or beads. Provides a measurement of the genetic variation between members of a species.

- **GLEAN gene**

Predicted [gene](#) sequence generated by GLEAN, an algorithm that integrates different sources of gene structure evidence (for example, gene model predictions, [EST](#) and protein sequence alignments to the genome, and SAGE or peptide tags) to produce a consensus gene prediction in the absence of known genes.

- **GO**

[Gene](#) Ontology project. Collaborative project that has developed three structured, controlled vocabularies (ontologies) to describe gene products in terms of their associated biological processes, cellular components, and molecular functions in a species-independent manner. The use of a consistent vocabulary allows genes from different species to be compared based on their GO [annotations](#).

- **GO component**

[Gene](#) Ontology term used to describe a cellular component, or the location where a gene product may act, rather than physical features of proteins or RNAs. For example, membrane (GO:0016020), extrinsic to membrane (GO:0019898), and integral to membrane (GO:0016021).

- **GO function**

[Gene](#) Ontology term used to describe the molecular function of a gene product, the jobs that it performs, or the "abilities" that it has (for example, transporting compounds, binding to things, holding things together, and changing one thing into another). This is different from the biological processes the gene product is involved in, which involve more than one activity.

- **GO process**

[Gene](#) Ontology term used to describe a biological process, a recognized series of events, or molecular functions associated with a gene product. A biological process is not equivalent to a pathway, though some [GO terms](#) do describe pathways.

- **GO term**

[Gene](#) Ontology term. The building blocks of the Gene Ontology, each term is assigned to one of the three ontologies: molecular function, cellular component, or biological process. Each [GO](#) term consists of a unique alphanumerical identifier, a common name, synonyms (if applicable), and a definition. When a term has multiple meanings depending on species, the

GO uses a "sensu" tag to differentiate among them. For example, the enzyme fumarase has the GO term GO:0004333, fumarate hydratase activity (fumarase activity), catalysis of the reaction: (S)-malate = fumarate + H<sub>2</sub>O.

- **GPI anchor**

C-terminal post-translational modification of many eukaryotic proteins. The two fatty acids within the glycoprophatidylinositol (GPI) moiety anchor the protein to the outer leaflet of the plasma membrane. GPI-anchored proteins are believed to be involved in signal transduction and immune responses, as well as the pathobiology of many parasites.

- **HMM**

Hidden Markov model. Statistical model in which the system being modeled is assumed to be a Markov process with unknown parameters, and the challenge is to determine the hidden parameters from the observable parameters. The extracted model parameters can then be used to perform further analysis, for example for pattern recognition applications.

- **Homolog**

Related by evolutionary descent, either between species (ortholog) or within a species (paralog).

- **Hydropathy**

Hydropathicity. Degree to which a peptide or protein is likely to be soluble in water. Protein hydropathy plots can be useful in predicting [transmembrane domains](#), potential antigenic sites, and regions that are likely to be exposed on the protein's surface.

- **Interactome**

The complete set of physical interactions between proteins within a specific biological context such as an organelle, cell, or organism. Can also refer to the complete set of genetic interactions between genes. Usually determined using genome-wide techniques, although the complete set of interactions is difficult to capture due to experimental noise and limits of detection.

- **Intergenic region**

Stretch of DNA located between [genes](#) that may act to regulate gene expression.

- **Intersect**

Similar to the [Boolean](#) operator "AND", an action used in the [query](#) history to find items that are common to two query result sets. For example, to search for items that appear in both result set X and result Y, type, "X INTERSECT Y".

- **Join**

Similar to the [Boolean](#) operator "UNION", an action used in the [query](#) history to combine query sets. For example, to combine result sets X and Y, type, "X JOIN Y".

- **JBrowse**

Interactive genome browser ([jbrowse.org](#)) developed by the Generic Model Organism Database (GMOD) project ([www.gmod.org](#)) that can be customized to show selected chromosomal features as well as display user-provided [annotations](#).

- **KEGG map**

Metabolic or regulatory interaction pathway generated by the Kyoto Encyclopedia of [Genes](#) and Genomes (KEGG) or by the use of their tools ([www.genome.jp/kegg](#)).

- **Locus**

Position on a [chromosome](#) of a [gene](#), feature (such as a [telomere](#)), or other chromosomal marker; also, the DNA at that position. Use of this term is sometimes restricted to mean expressed DNA regions.

- **Low complexity**

Pertaining to sequence regions that have an unusually repetitive nature (for example, a protein sequence of low complexity might look like PPTDPPPKDGGPPL, and a low-complexity nucleotide sequence might be AAATAAAAAAAAAAATTTAA). Low-

complexity regions can create problems in [sequence similarity](#) searching by causing artifactual hits. For this reason, filters are often used to remove low-complexity sequences. Low-complexity regions also contribute to antigenic variation in apicomplexan parasites.

- **Mass spec data**

Mass spectrometry data. Mass spectrometry is an analytical technique used to measure the mass-to-charge ratios of small molecules in several applications, including identification of proteins or peptides. In our databases, the "Identify [Genes](#) by Mass Spec Evidence" [query](#) is used to identify genes that have evidence of protein expression based on mass spec data.

- **Metabolic pathways**

Series of chemical reactions occurring within a cell and often catalyzed by enzymes. In a pathway, a molecule is often changed or modified into another product, which can be stored by the cell, used as a metabolic product, or used to initiate another pathway.

- **Microarray**

Microscopic array of biological molecules (for example, DNA or protein) used to determine the presence and/or amount (referred to as quantitation) of other biomolecules (other proteins, transcripts, etc.) in biological samples.

- **Microsatellite**

Polymorphic [locus](#) in nuclear and organellar DNA that consists of repeating units of 1-4 base pairs in length. Mostly neutral and codominant, microsatellites are used as molecular markers and to study [gene](#) dosage (looking for duplications or deletions of a particular genetic region). Also known as simple sequence repeats (SSRs).

- **Microsatellite map**

Map of [microsatellite](#) locations and linkages on a genome.

- **Mitochondrion**

Organelle responsible for respiration in a eukaryotic cell. Proteins required for mitochondrial function are encoded both in the nucleus and within the smaller mitochondrial genome.

- **Motif search**

Tool used to identify and locate sequence patterns (motifs) in protein and nucleic acid sequences. In our databases, this flexible search can be based on the general characteristics of the pattern and not solely on specific sequences (for example, Cys-[9-11 amino acids]-Cys or Leu-Leu-[basic residue]-Val). This allows the user to [query](#) using previously undescribed motifs.

- **NCBI**

National Center for Biotechnology Information. Established in 1988 as a national resource for molecular biology information, NCBI creates public databases, conducts research in computational biology, develops software tools for analyzing genome data, and disseminates biomedical information - all for the better understanding of molecular processes affecting human health and disease.

- **Nonredundant protein DB (NRDB)**

Peptide sequence database containing all nonidentical protein sequences from the same species extracted from GenPept, [NCBI](#) RefSeq, Swiss-Prot, and PRF databases. Used for [BLAST](#) protein database searches because its smaller size results in shorter search times and more meaningful statistics.

- **Nucmer**

NUCleotide MUMmer. Part of the MUMmer alignment package, an alignment tool used for the rapid alignment of very large DNA and amino acid sequences.

- **ORF**

Amino acid sequences computed by translating the six frames of raw genomic sequence using the standard genetic code. We save the translated sequences having at least 50 amino acids. The sequences are not annotated nor human reviewed. ORF's do not necessarily begin with methionine residues, but they all terminate with a stop codons.

- **Oligo**

Oligonucleotide. Short sequence of DNA or RNA, typically of 20-70 nucleotides.

- **Oligonucleotide microarray**

Collection of microscopic oligonucleotide spots arrayed on a solid surface by covalent attachment to chemically suitable matrices. Used for expression profiling, the monitoring of the [expression levels](#) of hundreds or thousands of [genes](#) simultaneously. Probes for oligonucleotide [microarrays](#) are designed to match parts of known or predicted mRNAs.

- **Open Reading Frame**

See [ORF](#)

- **Organellar genome sequence**

The genome sequence contained in an organelle *i.e.* the [mitochondria](#) and/or [apicoplast](#) organelles. These sequences may be circular or linear and exist in addition to the much larger nuclear genome sequence.

- **OrthoMCL**

Genome-scale algorithm for grouping orthologous protein sequences. It provides [genes](#) shared by two or more species/genomes and also genes representing species-specific gene expansion families. Therefore, it serves as a utility for automated eukaryotic genome [annotation](#) and phylogenetic profiling. Available at [orthomcl.org](#)

- **Ortholog**

Same [gene](#) in different organisms; having evolved from the same ancestral [locus](#).

- **Ortholog group**

Orthologous [genes](#) shared by group of organisms; the group of genes can also contain [paralogs](#).

- **Orthology-based phylogenetic profile**

Tool used to find [genes](#) that are present or not present in a desired group of organisms on the tree of life (currently computed for 81 complete genomes). The user has control over the profile and over whether or not genes must be found in any particular group of organisms (for example, in Apicomplexa but not in mammals). Taxa can also be marked as indifferent (for example, it does not matter if the gene is also found in plants). [Ortholog](#) and [paralog](#) relationships are determined using the [OrthoMCL](#) algorithm.

- **PATS**

Neural network analysis tool that identifies amino acid sequences within a [query](#) sequence that are potentially targeted to the [apicoplast](#) matrix of Plasmodium falciparum.

- **PDB 3D structure**

Three-dimensional macromolecular structure in the Protein Data Bank (PDB) ([www.pdb.org](#)) obtained by one of three methods: X-ray crystallography (over 80%), solution nuclear magnetic resonance (NMR) (about 16%), or theoretical modeling (2%). A few structures were determined by other methods.

- **PROSITE motif**

Protein sequence pattern or profile derived from multiple alignments of homologous sequences and stored in the PROSITE database ([prosite.expasy.org](#)), an annotated collection of motif descriptors dedicated to the identification of protein families and domains.

- **Paralog**

Related by [gene](#) duplication within a genome; originated by duplication and then diverged from the parent copy by mutation and selection or drift.

- **Pearson correlation**

Pearson Product Moment Correlation, the most common measure of the correlation between two variables. Reflects the degree of linear relationship between two variables and ranges from +1 to -1, with a correlation of +1 indicating a perfect positive linear relationship between variables.

- **Peptide mass fingerprinting**

Analytical technique for protein identification wherein an unknown protein of interest is cleaved into peptides by a protease such as trypsin, and the peptides resulting from this cleavage are analyzed using a mass spectrometric method such as MALDI-TOF or ESI-TOF. The masses derived for the peptides are then compared to a database containing known protein sequences or even to the genome. Computer programs theoretically cut the protein sequences in the database into peptides with the same protease (for example trypsin), and calculate the absolute masses of the peptides from each protein. They then compare the masses of the peptides of the unknown protein to the theoretical peptide masses of each protein encoded in the genome. The results are statistically analyzed to find the best match.

- **Pfam domain**

Conserved protein region in the Pfam database ([Pfam.org](#)), a collection of multiple sequence alignments and hidden Markov models covering many common protein families. The alignments may represent evolutionarily conserved structures that may shed light on protein function. Profile hidden Markov models (profile [HMMs](#)) built from the Pfam alignments can be useful for associating a new protein to a known protein family, even if the homology is weak. Unlike standard pairwise alignment methods (for example, [BLAST](#) and [FASTA](#)), Pfam HMMs deal sensibly with multidomain proteins.

- **Phylogeny**

Historical relationships among lineages of organisms or their parts, including their [genes](#).

- **PlasmoAP**

Algorithm/tool that predicts the likelihood that a protein sequence is targeted to the [apicoplast](#). It provides the position of [signal peptide](#) cleavage sites in amino acid sequences if targeting is predicted.

- **PlasmoCyc**

Database/utility built by analyzing the genomes of the Plasmodium species in EuPathDB with SRI International's pathway tools; used for searching and visualizing Plasmodium metabolic pathway information.

- **ProDom**

Database of protein domain families generated from the global comparison of all available protein sequences ([prodom.prabi.fr](#)).

- **Promoter**

Regulatory region of DNA located upstream (towards the 5' region) of a [gene](#) and providing a control point for regulated gene transcription.

- **Protein-coding**

Capable of encoding a protein sequence; generally refers to a sequence of DNA.

- **Proteomics**

The large-scale study of proteins, particularly of the full set of proteins encoded by a genome.

- **Pseudogene**

Defunct relatives of known [genes](#) that have lost their [protein-coding](#) ability or are otherwise no longer expressed in the cell. Although they may have some gene-like features (such as [promoters](#), CpG islands, and splice sites), they are nonetheless considered nonfunctional due to their lack of protein-coding ability resulting from various genetic disablements (stop codons, frameshifts, or a lack of transcription) or their inability to function as an RNA (such as with [rRNA](#) pseudogenes).

- **PubCrawler**

Free service that scans daily updates to the [NCBI](#) Medline (PubMed) and GenBank databases and alerts users to any relevant updates. Available at [pubcrawler.gen.tcd.ie](#)

- **Query**

Sequence or term used in a database search. For example, the sequence submitted for a [BLAST](#) search is the query sequence.

- **RNA predictions**

Predictions of [genes](#) that encode nonprotein-encoding RNA's such as [tRNA](#), snoRNA, [rRNA](#), etc.

- **Reference genome sequence**

The community gold standard genome sequence. Usually the most complete assembly including an annotation. Reference genome sequences do change as superior assemblies and annotation become available.

- **RefSeq mRNA**

Nonredundant mRNA sequence in the RefSeq database. RefSeq mRNA sequences with an NM\_XXXXXX accession are curated sequences and are, therefore, considered more reliable than those with XM\_XXXXXX accessions (predicted mRNA sequences).

- **RefSeq noncoding RNA**

Nonredundant noncoding RNA (ncRNA) sequence in the RefSeq database.

RefSeq [ncRNA](#) sequences with an NR\_XXXXXX accession are curated sequences and are, therefore, considered more reliable than those with XR\_XXXXXX accessions (predicted ncRNA sequences).

- **RefSeq protein**

Nonredundant protein sequence in the RefSeq database. RefSeq protein sequences with NP\_XXXXXX accessions are curated sequences and are, therefore, considered more reliable than those with XP\_XXXXXX accessions (predicted protein sequences).

- **RefSeq**

[NCBI](#) reference sequences. A curated nonredundant collection of sequences representing genomes, transcripts, and proteins as annotated by NCBI (available at [www.ncbi.nlm.nih.gov/refseq](http://www.ncbi.nlm.nih.gov/refseq)). The [annotation](#) in these records is often different from the original GenBank submission, which may not be updated every time new information is obtained.

- **Repeat regions**

Sequences present in many identical or highly similar copies in the genome.

- **SAGE tags**

Serial analysis of [gene](#) expression (SAGE) tags. Short (14-nucleotide) sequences found within mRNA, the relative abundance of which indicates the level of expression of the mRNA containing that tag.

- **SNP**

Single nucleotide polymorphism. Small genetic changes or variations that can occur within a DNA sequence, for example when a single nucleotide, such as an A replacing one of the other three nucleotide letters C, G, or T. Most SNPs are found outside of coding sequences, but SNPs found within a coding sequence are more likely to alter the biological function of a protein. SNPs may be synonymous (generating a conservative change not altering the amino acid sequence) or they can be nonsynonymous and change the amino acid that is encoded.

- **SNP density**

Amount or number of single nucleotide polymorphisms (SNPs) in a region of the genome.

- **SNP genotyping**

Identifying and mapping single nucleotide polymorphisms (SNPs) in an effort to determine the genotype members of a species. [SNPs](#) usually consist of two alleles (where the rare allele frequency is less than 1%), are evolutionarily conserved, and are the most common type of genetic variation. See [Genotyping](#).

- **Scaffolds**

In genomic mapping, a series of [contigs](#) that are in the right order and orientation, but not necessarily connected in one continuous stretch of sequence.

- **Sequence similarity**

Degree of similarity between two or more protein or nucleotide sequences.

- **Signal peptide**

Short (3-60 amino acids) peptide sequence that directs the co-translational import of a protein to certain organelles or for secretion.

- **SignalP**

Program that predicts the presence and location of [signal peptide](#) cleavage sites in amino acid sequences from Gram-positive and -negative prokaryotes and eukaryotes. The method incorporates a prediction of cleavage sites and a signal peptide/nonsignal peptide prediction based on a combination of several artificial neural networks and hidden Markov models (HMMs).

- **Strand**

One half of the DNA helix; string or stretch of covalently linked nucleotides.

- **Subtract**

Similar to the [Boolean](#) operator "NOT", an action used in the [query](#) history to remove items from one result set that occur in another result set. For example, to remove items that exist in set Y from those in set X, type, "X SUBTRACT Y".

- **Syntenic**

Loci located on the same [chromosome](#) but not necessarily linked. For example, [genes](#) that are part of a syntenic group share a common chromosomal location. Also used to refer to conservation of gene order across species.

- **TIGR**

The Institute for Genomic Research, a nonprofit center dedicated to deciphering and analyzing genomes.

- **TM domains**

See [transmembrane domain](#).

- **TWINSCAN gene models**

[Gene](#) models generated using TWINSCAN, a tool that integrates traditional probability models (such as those underlying GENSCAN and FGENESH) with information from alignments between two genomes. TWINSCAN is based on the idea that functional sequences show different patterns of evolutionary conservation than sequences under little selective pressure, such as the central regions of introns. TWINSCAN is designed for the analysis of high-throughput genomic sequences containing an unknown number of genes.

- **Telomere**

Nucleoprotein complexes that constitute the physical ends of linear eukaryotic [chromosomes](#) and that have important functions, primarily in the protection, replication, and stabilization of the chromosome ends. Telomeres often contain lengthy stretches of tandemly repeated simple DNA sequences composed of a G-rich [strand](#) and a C-rich strand (called terminal repeats). These terminal repeats are highly conserved.

Sequences adjacent to the telomeric repeats are often highly polymorphic and rich in repetitive elements (termed subtelomeric repeats); in some cases, [genes](#) have been found in the proterminal regions of chromosomes.

- **TigrScan gene**

[Gene](#) model generated using TigrScan, a gene-finding tool based on the generalized hidden Markov model (HMM) framework, similar to GENSCAN and Genie. It is highly reconfigurable and includes software for retraining.

- **ToxoCyc**

Database/utility built by analyzing the *Toxoplasma gondii* genome with SRI International's pathway tools; used for searching and visualizing *Toxoplasma* metabolic pathway information.

- **Translation**

Synthesis of protein from an mRNA template.

- **Transmembrane domain**

Three-dimensional protein structure that is thermodynamically stable in a membrane. This may be a single alpha helix, a stable complex of several transmembrane alpha helices, a transmembrane beta barrel, a beta-helix of gramicidin A, or any other structure.

Transmembrane domains average 20 amino acid residues in length, though they may be much smaller or much longer.

- **Transmembrane protein**

Protein that spans an entire biological membrane.

- **UTR**

Untranslated region. Section of messenger RNA (mRNA) that either precedes (5' UTR) or follows (3' UTR) the coding region and is not itself translated. The UTR contains several regulatory regions, including the polyadenylation (polyA) site in the 3' UTR, sequences involved in the initiation of [translation](#) (in the 5' UTR), and binding regions for proteins and other regulatory molecules in both the 3' and 5' UTR.

- **UniGene**

Project and database at [NCBI](#) aimed at defining [gene](#)-oriented clusters of expressed sequence tags (ESTs). Sets of [ESTs](#) are clustered based on strong sequence homology in an attempt to define a specific, nonredundant cluster for each transcript in a tissue or genome. Each UniGene cluster contains sequences that represent a unique gene in addition to information about the tissue types in which the gene has been expressed and map location.

- **Wildcard character**

Character used to substitute for any other character(s) in a string.

- **Xenolog**

[Gene](#) found in an unrelated species and that is related by gene transfer rather than common vertical descent.

- **blastn**

Version of the basic local alignment search tool (BLAST) used to compare a nucleotide [query](#) sequence against a nucleotide sequence database.

- **blastp**

Version of the basic local alignment search tool (BLAST) used to compare a protein [query](#) sequence against a protein sequence database.

- **blastx**

Version of the basic local alignment search tool (BLAST) used to compare a nucleotide [query](#) sequence translated in all reading frames against a protein sequence database.

- **cDNA**

Complementary DNA. DNA molecule synthesized by the enzyme reverse transcriptase using an mRNA as template.

- **cDNA microarray**

Collection of microscopic [cDNA](#) spots commonly representing single [genes](#) and arrayed on a solid surface (commonly glass slides) by covalent attachment to chemically suitable matrices. Used for expression profiling, the monitoring of the [expression levels](#) of hundreds or thousands of genes simultaneously.

- **ePCR**

Electronic PCR (polymerase chain reaction). Computational procedure used to check for uniqueness in spacing and number of primer binding sites within DNA sequences. Searches for subsequences that closely match a set of PCR primers and that have the correct order, orientation, and spacing to make a PCR product. Used to check the expected length of a PCR product, which can provide information regarding unexpected repetitive sequences.

- **ncRNA**

Noncoding RNA. Any RNA that is not translated into a protein. Includes transfer RNA (tRNA), ribosomal RNA (rRNA), small RNAs such as snoRNAs, microRNAs, siRNAs and piRNAs, as well as long ncRNAs.

- **rRNA**

Ribosomal RNA. Component of the ribosomes, which function in protein synthesis.

- **snRNA**

Small nuclear RNA. Class of small RNA molecules found within the nucleus, transcribed by RNA polymerase II or III, and involved in a variety of important processes such as RNA splicing (removal of introns from hnRNA), regulation of transcription factors (7SK RNA) or RNA polymerase II (B2 RNA), and maintaining [telomeres](#). They are always associated with specific proteins, and the complexes are referred to as small nuclear ribonucleoproteins (snRNP) or snurps. These elements are rich in uridine. A large group of snRNAs known as small nucleolar RNAs (snoRNAs) are small RNA molecules that play an essential role in RNA biogenesis and chemical modification of ribosomal RNAs (rRNAs) and other RNA [genes](#) (tRNA and snRNAs). They are located in the nucleus and the Cajal bodies of eukaryotic cells (the major sites of RNA synthesis).

- **tRNA**

Transfer RNA. Small RNA chain (73-93 nucleotides) that transfers a specific amino acid to a growing polypeptide chain at the ribosomal site of protein synthesis during [translation](#). A three-base region, the anticodon, pairs to the corresponding three-base codon region on the template mRNA. Each type of tRNA molecule can be attached to only one type of amino acid, but because the genetic code contains multiple codons that specify the same amino acid, tRNA molecules bearing different anticodons may also carry the same amino acid.

- **tblastn**

Version of the basic local alignment search tool (BLAST) used to compare a protein [query](#) sequence against a translated nucleotide sequence database.

- **tblastx**

Version of the basic local alignment search tool (BLAST) used to compare the six-frame [translations](#) of a nucleotide [query](#) sequence against the six-frame translations of a nucleotide sequence database.

## **Appendix A: Print-based instructional materials**

## Basic OrthoMCL Functionality

<http://orthomcl.org/>

### Learning objectives:

- Run searches in OrthoMCL
- Run phyletic pattern searches using check boxes or an expression
- Combine searches using the strategy system
- Explore individual ortholog group pages
- Explore the group cluster graphs

### 1. Using the Phyletic Pattern search in OrthoMCL

The “Phyletic Pattern” search is an ortholog group search – look under the ortholog groups category and explore the available searches. Can you find the one called “Phyletic Pattern”? There are two ways to specify a phyletic pattern:

The screenshot shows the OrthoMCL DB homepage with a sidebar on the left containing search filters for Ortholog Groups and Proteins. A large central window displays the "Identify Ortholog Groups based on Phyletic Pattern" tool. This tool includes a graphical tree display, a search bar for taxonomic names, and an expression input field with a key for constraint types. A blue callout box highlights the expression input field and its key.

**Key:** ● = no constraints | ✓ = must be in group | □ = at least one subtaxon must be in group | ✗ = must not be in group | \* = mixture of constraints

**Ortholog Groups**

- Q % Pairs w/ Similarity
- Q All Groups
- Q Avg % Homology
- Q Avg % Identity
- Q Avg % Match Length
- Q Avg E-Value
- Q EC Number (specific)
- Q Group ID(s)
- Q Group or Sequence ID
- Q Number of Sequences
- Q Number of Taxa
- Q PFam ID or Keyword
- Q Phyletic Pattern
- Q Text Terms

**Proteins**

**Overview of Resources and Tools**

**Identify Ortholog Groups based on Phyletic Pattern**

Find Ortholog Groups that have a particular phyletic pattern, i.e., that include or exclude taxa or species that you specify.

The search is controlled by the Phyletic Pattern Expression (PPE) shown in the text box. Use either the text box or the graphical tree display, or both, to specify your pattern. The graphical tree display is a friendly way to generate a pattern expression. You can always edit the expression directly. For PPE help see the instructions at the bottom of this page.

In the graphical tree display:

- Click on the + icon to expand a node or hide subnodes and species.
- Click on the ✗ icon to specify which taxa or species to include or exclude in the profile.
- Refer to the legend below to understand other icons.

Expression: BACT+OT AND ARCH+OT

**Key:** ● = no constraints | ✓ = must be in group | □ = at least one subtaxon must be in group | ✗ = must not be in group | \* = mixture of constraints

**Get Answer**

**Community Chat**

1. Using the expression box. Type the expression using hints available at the bottom of the search page.

2. Using the selectable tree menu. Click on the circle next to the taxon you want to include or exclude.
- a. How many protein groups do not contain orthologs from bacteria and archaea?
  - b. Find all groups that contain orthologs from at least one species of *Cryptosporidium* and *Giardia* but not from bacteria or archaea. If you are getting frustrated trying to figure this one out, you have a right to be! You cannot answer this question by using the check boxes. However, OrthoMCL has an added feature that allows you to enter an expression to define the phyletic pattern. This option provides additional flexibility. Can you figure out what expression to use to answer this question? (hint: scroll down to the bottom of the search page to find additional information about expression parameters.)

Before looking at the answer below, try this on your own to see if you can figure it out.

Find Ortholog Groups that have a particular phyletic pattern, i.e., that include or exclude taxa or species that you specify.

The search is controlled by the Phyletic Pattern Expression (PPE) shown in the text box. Use either the text box or the graphical tree display, or both, to specify your pattern. The graphical tree display is a friendly way to generate a pattern expression. You can always edit the expression directly. For PPE help see the [instructions at the bottom of this page](#).

In the graphical tree display:

- Click on -/+ to show or hide subtaxa and species.
- Click on the  icon to specify which taxa or species to include or exclude in the profile.
- Refer to the legend below to understand other icons.

Expression:

BACT=0T AND ARCH=0T AND cpar+cand+choi+chot+chom+chod+cmei+cmur+cpia+ctyz+cubi>=1T AND gass+gadh+gasb+gabb+gase+gmur>=1T

All VEuPathDB sites also have a phyletic pattern search that uses OrthoMCL data under *Genes -> Orthology and Synteny -> Orthology Phylogenetic Profile*. This search is very useful to identify genes in your organism of interest that are restricted in their profile. For example, you frequently want to identify genes that are conserved among organisms in your genus of interest but not present in the host as these genes may make good drug targets or vaccine candidates.

## 2. Combining searches in OrthoMCL

Find all plant proteins that are likely phosphatases that do not have orthologs outside of plants.

- a. Use the site search box at the top of the page in the header **to find OrthoMCL groups** that contain the word “\*phosphatase\*” (note that the search should be run without the quotation marks but with the asterisks).



The screenshot shows the OrthoMCL DB homepage. At the top, there's a logo with a network of colored dots and the text "OrthoMCL DB" and "Ortholog Groups of Protein Sequences". Below the logo, it says "Release 6.2 beta" and "17 Dec 2020". A search bar contains the query "\*phosphatase\*". Below the search bar, there are links for "My Strategies", "Searches", "Tools", "My Workspace", "Data", "About", "Help", and "Contact Us". On the right side, there are social media icons for Twitter, Facebook, YouTube, and a "Guest" link.

- How many proteins did you identify?
- How many groups did you identify?

**Note: that numbers in screen shots will likely be different on the site due to frequent updates.**

## All results matching \*phosphatase\*

Export as a Search Strategy  
to download or mine your results ➔

1 - 20 of 54,032

◀ 1 2 3 ... 2,702 ▶

### Filter results

Hide zero counts

Genome	49,228
Protein Sequences	

Orthology	4,804
Ortholog Groups	

### Filter fields

Select a result filter above

Protein Sequence - aaeg-old|AAEL000109 Enolase-phosphatase E1 (EC 3.1.3.77)(2,3-diketo-5-methylthio-1-phosphopentane phosphatase)

Current group: OG6\_104759  
Taxon Name: Aedes aegypti LVP\_AGWG

▶ Fields matched: Product

Protein Sequence - aaeg-old|AAEL000372 myo inositol monophosphatase

Current group: OG6\_100401  
Pfam Domains: PF00459

Taxon Name: Aedes aegypti LVP\_AGWG

▶ Fields matched: EC Numbers; Product

Protein Sequence - aaeg-old|AAEL000840 skeletal muscle/kidney enriched inositol 5-phosphatase

Current group: OG6\_108700

- b. Click on ortholog groups to filter the results to only show the ortholog groups containing the word phosphatase. Notice that you can filter the group results even further by the group fields. Also, notice that the export as a Search Strategy button is now active. (Note: this is because all the results are now only one type of record: groups.)
- c. Export the ortholog group results as a search strategy by clicking on the blue “Export as a Search Strategy” button at the top right of the page.

## Ortholog Groups matching \*phosphatase\*

Export as a Search Strategy  
to download or mine your results ➔

1 - 20 of 4,804

◀ 1 2 3 ... 241 ▶

### Filter results

Hide zero counts

Orthology	
Ortholog Groups	4,804

**Clear filter**

### Filter Ortholog Group fields

[select all](#) | [clear all](#)

<input type="checkbox"/> Domains	72
<input type="checkbox"/> EC Number	1,526
<input type="checkbox"/> Keywords	1,394
<input type="checkbox"/> List of All Sequences	3,203
<input type="checkbox"/> Pfam Domains	2,476

Ortholog Group - OG6\_100000

▶ Fields matched: EC Number; Pfam Domains

Ortholog Group - OG6\_100006

▶ Fields matched: Pfam Domains

Ortholog Group - OG6\_100011

▶ Fields matched: List of All Sequences; Pfam Domains

Ortholog Group - OG6\_100012

▶ Fields matched: List of All Sequences

Ortholog Group - OG6\_100013

## My Search Strategies

Opened (1) All (25) Public (7) Help

Unnamed Search Strategy \*

Step 1

**Text** **4,804 Ortholog Groups** **+ Add a step**

4,804 Ortholog Groups Revise this search

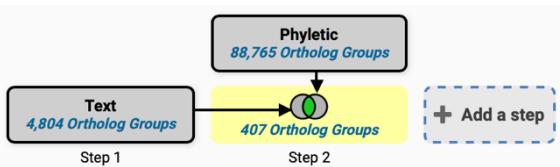
Ortholog Group Results

Rows per page: 50

Ortholog Group	Score	Total Number Proteins	Keywords	PFam Domains	Archaea	Bacteria	Alveolata	Amoeba	Euglenozoa	Fungi
OG6_100000	1	12129	N/A	N/A	0 / 27 (0%)	12 / 47 (26%)	62 / 108 (57%)	4 / 13 (31%)	22 / 52 (42%)	133 / 171 (78%)

**Download** **Add to Basket** **Add Columns**

- d. Add a step and run a phyletic pattern search for groups that contain any plant protein but do not contain any other organism outside plants. (hint: make sure everything has a red x on it except for plants (Viridiplantae (VIRI)), which should be a grey circle).
- e. How many groups did you return?



- \* Root (ALL)
  - ✖ Bacteria (BACT)
  - ✖ Firmicutes (FIRM)
  - ✖ Proteobacteria (PROT)
  - ✖ Other Bacteria (OBAC)
  - ✖ Archaea (ARCH)
    - ✖ Nitrosopumilus maritimus (strain SCM1) (nmar)
    - ✖ Euryarchaeota (EURY)
    - ✖ Crenarchaeota (CREN)
    - ✖ Nanoarchaeota (NANO)
    - ✖ Korarchaeota (KORA)
  - \* Eukaryota (EUKA)
    - ✖ Alveolates (ALVE)
    - ✖ Amoebozoa (AMOE)
    - ✖ Euglenozoa (EUGL)
    - ✖ Viridiplantae (VIRI)
    - ✖ Fungi (FUNG)
    - ✖ Metazoa (META)
    - ✖ Other Eukaryota (OEUK)

### 3. Exploring a specific OrthoMCL group - examining the cluster graph.

- a. Visit the OrthoMCL group OG6\_131670. You can quickly get to this group by first running a site search with the group ID. Click on the group ID to get to its page.

OrthoMCL DB Release 6.2 beta 17 Dec 2020 OG6\_131670 **My Strategies Searches Tools My Workspace Data About Help Contact Us**

All results matching OG6\_131670

1 - 20 of 94

**Ortholog Group - OG6\_131670**

Fields matched: Current group

Filter results

Genome Protein Sequences 93

Orthology Ortholog Groups 1

Filter fields

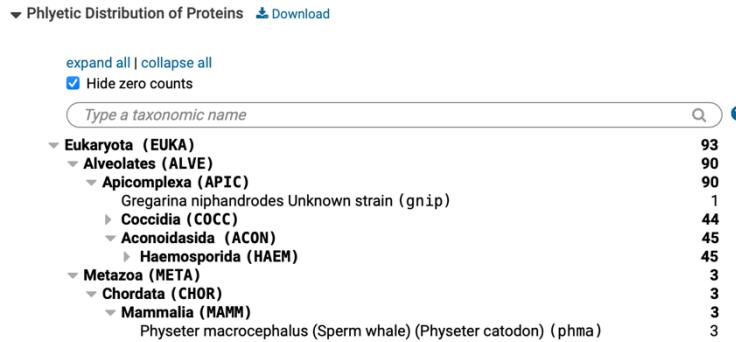
Select a result filter above

Protein Sequence - hhsDRFSR R10R3N alveolin domain containing intermediate filament IMC10

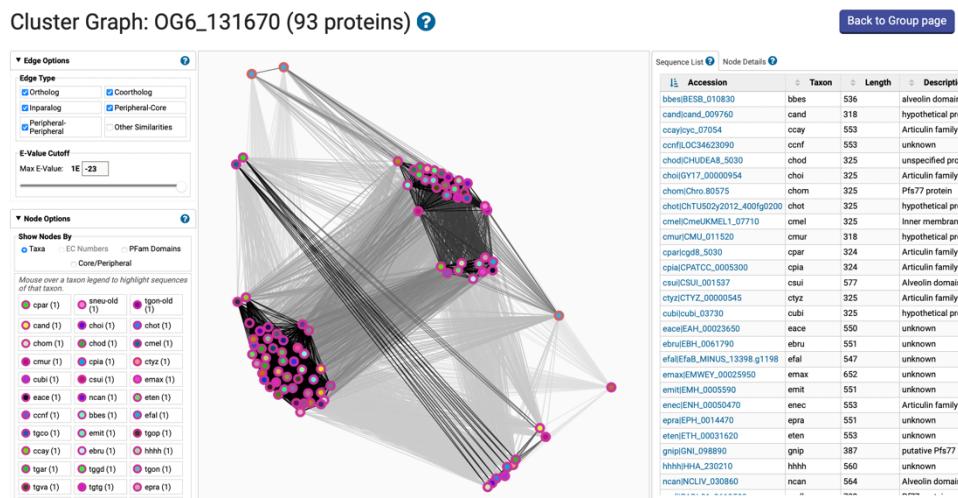
- b. The group page is divided into 5 sections:
- Phyletic distribution
  - Group summary
  - List of proteins
  - PFam domains
  - Cluster graph

Examine each of the above sections - is it clear what each section contains?

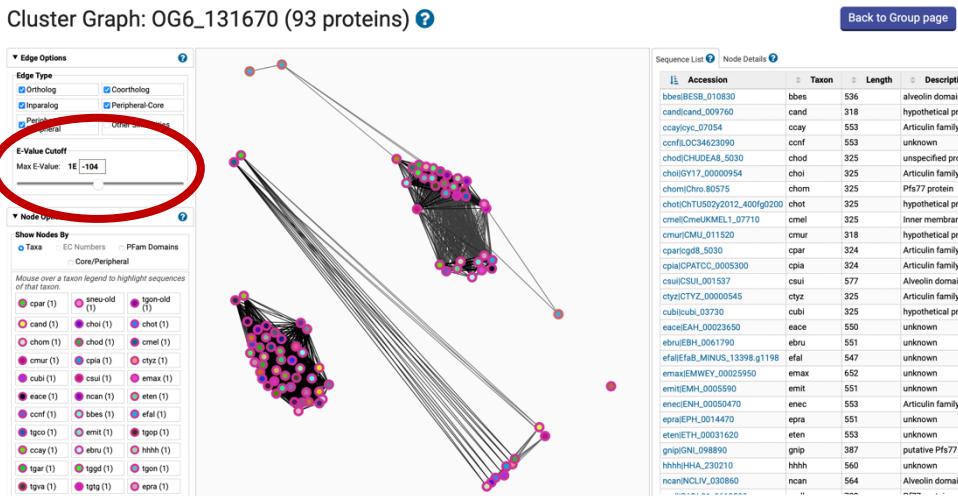
c. Examine the phyletic distribution tree. What taxa does this group contain?



d. Examine the cluster graph for this group (*hint: go to the cluster graph section of the page and then click on the “Click to open the Cluster graph in a new tab”*)



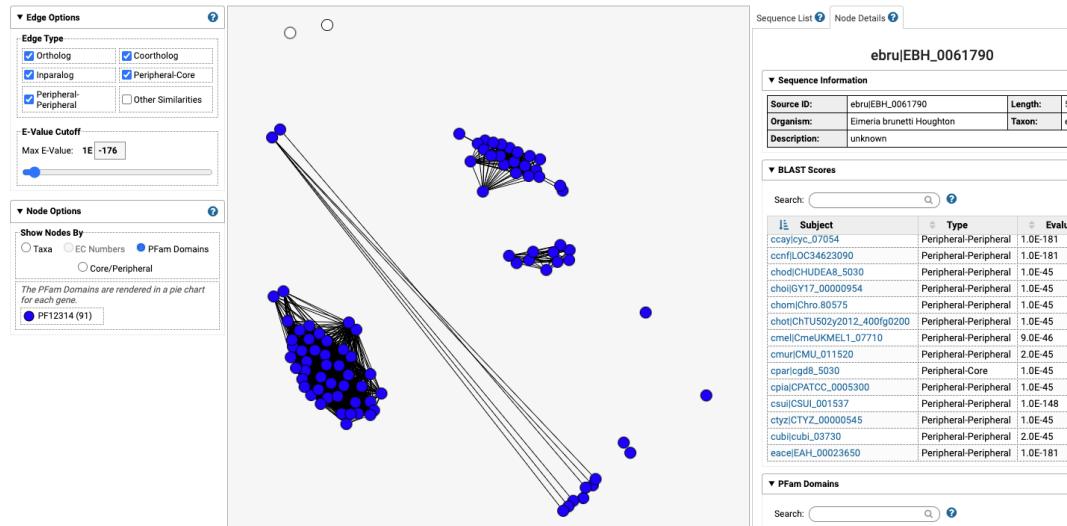
You can interact with the cluster graph. For example, move the slide to increase the E-value cutoff stringency (e.g., to a more negative number). Can you identify subclusters? Click on the nodes in the graph – notice how the organism is updated on the right.



On the left of the page in the Node Options panel, click on PFam Domains to see which proteins have the various PFam domains.

### Cluster Graph: OG6\_131670 (93 proteins) ?

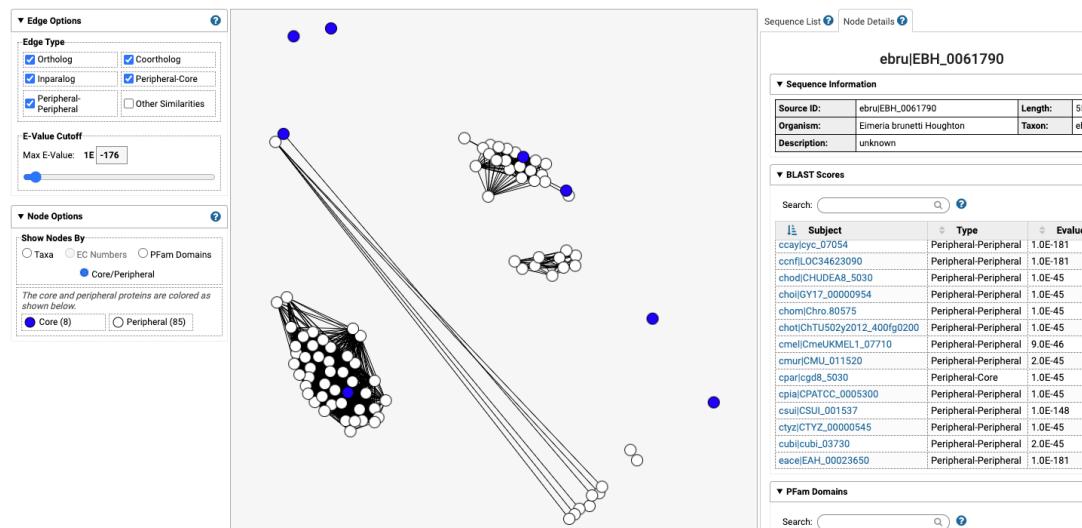
[Back to Group page](#)



And in the same panel, click on Core/Peripheral to observe which proteins were derived from Core species and which proteins were derived from Peripheral species. Proteins from Core species were used in the initial OrthoMCL algorithm to form Core ortholog groups. Proteins from Peripheral species were mapped into these Core groups by sequence similarity (determined by BLAST score).

### Cluster Graph: OG6\_131670 (93 proteins) ?

[Back to Group page](#)



# Site Search

**Note:** this exercise uses PlasmoDB.org as an example database, but the same functionality is available on all VEuPathDB resources.

## Learning objectives:

- Use keywords in site search
- Explore site search results
- Filter site search results by categories
- Filter site search results by organisms
- Filter site search results by category fields
- Export results to a search strategy
- Find a specific gene using its ID in site search

1. Enter the word *kinase* in the site search window (top center of the page, arrow in the image below). Then click enter on your keyboard or click on the search icon (square in the image below).

The screenshot shows the PlasmoDB beta homepage. At the top, there is a search bar with the word "kinase" typed into it. To the right of the search bar is a blue square button with a white magnifying glass icon. The page has a dark purple header with the PlasmoDB logo and navigation links like "My Strategies", "Searches", "Tools", etc. Below the header, there's a main content area titled "Overview of Resources and Tools". On the left, there's a sidebar with a "Search for..." section containing a dropdown menu and a list of categories: Genes, Organisms, Popset Isolate Sequences, Genomic Sequences, Genomic Segments, SNPs, SNPs (from Array), ESTs, and Metabolic Pathways. The "Genes" option is currently selected. The main content area shows various tools and resources with small icons and brief descriptions.

2. How many results with the word kinase did you get? Are all the results genes? Explore the filter panel on the left side of the webpage. Filter the results so that you only view gene results (hint: click on the word *genes* in the *Filter results* section; arrow in image below).

The screenshot shows the search results page for "kinase". The title at the top says "All results matching kinase". Below the title, it says "1 - 20 of 17,367". There are two filter panels on the left: "Filter results" and "Filter organisms". In the "Filter results" panel, there is a checkbox for "Hide zero counts" and a list of categories with their counts: Genes (16,684), Population biology (249), Metabolism (345), and others. A red arrow points to the "Genes" link. Below these filters, there is a "Select a result filter above" section and a "Filter organisms" section with a dropdown menu. The main content area lists several search results, each with a gene name, organism, and a "Fields matched" section. The first result is "Gene - PCYB\_132500 kinase" from Plasmodium cynomolgi strain B. The last few results listed are Gene - AK88\_00505 pantethenate kinase, Gene - AK88\_01656 phosphoglycerate kinase, and Gene - AK88\_02186 pyridoxal kinase.

3. How many of the genes included the word kinase in their product descriptions? Notice that once you filter the result by genes (click on the *Genes* filter), the fields section expands to reveal additional filtering options. Once you select the *Product descriptions* field you are provided the option to *apply* this filter or cancel it (box middle panel below). Once a filter is applied it can be cleared by clicking on *Clear filter* (box left panel below).

**Filter results**

Hide zero counts

**Genome Genes**

**Clear filter** 16,684

**Filter Gene fields**

Alternate product descriptions 7  
 EC descriptions and numbers 13,510  
 GO terms 5,432  
 InterPro domains 6,955  
 Orthologs 7,136  
 PDB chains 4,814  
 Product descriptions 6,007  
 PubMed 689  
 Rodent malaria phenotype 87  
 User comments 255

**Filter organisms**

select all |  clear all |  expand all |  collapse all

Type a taxonomic name

- ▶  Plasmodium adleri 322
- ▶  Plasmodium berghei 370
- ▶  Plasmodium hilli*linnei* 375

**Filter results**

Hide zero counts

**Genome Genes**

**Clear filter** 16,684

**Filter Gene fields**

**Apply**  **X**

Alternate product descriptions 7  
 EC descriptions and numbers 13,510  
 GO terms 5,432  
 InterPro domains 6,955  
 Orthologs 7,136  
 PDB chains 4,814  
 Product descriptions 6,007  
 PubMed 689  
 Rodent malaria phenotype 87  
 User comments 255

**Filter organisms**

select all |  clear all |  expand all |  collapse all

Type a taxonomic name

- ▶  Plasmodium adleri 322
- ▶  Plasmodium berghei 370
- ▶  Plasmodium hilli*linnei* 375

**Filter results**

Hide zero counts

**Genome Genes**

**Clear filter** 6,007

**Filter Gene fields**

**Clear filter**

Alternate product descriptions 7  
 EC descriptions and numbers 13,510  
 GO terms 5,432  
 InterPro domains 6,955  
 Orthologs 7,136  
 PDB chains 4,814  
 Product descriptions 6,007  
 PubMed 689  
 Rodent malaria phenotype 87  
 User comments 255

**Filter organisms**

select all |  clear all |  expand all |  collapse all

Type a taxonomic name

- ▶  Plasmodium adleri 157
- ▶  Plasmodium berghei 121
- ▶  Plasmodium billcollinsi 147
- ▶  Plasmodium blacklocki 144
- ▶  Plasmodium chabaudi 122
- ▶  Plasmodium coatneyi 117
- ▶  Plasmodium cynomolgi 244
- ▶  Plasmodium falciparum 2,353
  - Plasmodium falciparum 3D7 145
  - Plasmodium falciparum 7G8 147
  - Plasmodium falciparum CD01 146
  - Plasmodium falciparum Dd2 146
  - Plasmodium falciparum GA01 147
  - Plasmodium falciparum GB4 149
  - Plasmodium falciparum GN01 146
  - Plasmodium falciparum HB3 145
  - Plasmodium falciparum IT 147
  - Plasmodium falciparum KE01 146
  - Plasmodium falciparum KH01 146
  - Plasmodium falciparum KH02 148
  - Plasmodium falciparum ML01 151
  - Plasmodium falciparum SD01 146
  - Plasmodium falciparum SN01 147
  - Plasmodium falciparum TG01 151
  - ▶  Plasmodium fragile 94
  - ▶  Plasmodium gaboni 302
  - ▶  Plasmodium gallinaceum 129
  - ▶  Plasmodium iui 111
  - ▶  Plasmodium knowlesi 241
  - ▶  Plasmodium malariae 125
  - ▶  Plasmodium ovale curtisi 125
  - ▶  Plasmodium praefalciparum 144
  - ▶  Plasmodium reichenowi 291
  - ▶  Plasmodium relictum 128
  - ▶  Plasmodium vinckei 213
  - ▶  Plasmodium vivax 247

4. How many of the above genes are found in *Plasmodium falciparum* 3D7? How did you find this number? (hint: explore the *Filter organisms* section of the results filter). Select the correct organism and apply the filter.

**Filter organisms**

add these |  clear these |  select only these  
 select all |  clear all

Type a taxonomic name  Plasmodium falciparum 3D7

- Plasmodium falciparum 2,238
- Plasmodium falciparum 3D7 138

**Filter organisms**

select all |  clear all |  expand all |  collapse all

Type a taxonomic name

- ▶  Plasmodium adleri 157
- ▶  Plasmodium berghei 121
- ▶  Plasmodium billcollinsi 147
- ▶  Plasmodium blacklocki 144
- ▶  Plasmodium chabaudi 122
- ▶  Plasmodium coatneyi 117
- ▶  Plasmodium cynomolgi 244
- ▶  Plasmodium falciparum 2,353
  - Plasmodium falciparum 3D7 145
  - Plasmodium falciparum 7G8 147
  - Plasmodium falciparum CD01 146
  - Plasmodium falciparum Dd2 146
  - Plasmodium falciparum GA01 147
  - Plasmodium falciparum GB4 149
  - Plasmodium falciparum GN01 146
  - Plasmodium falciparum HB3 145
  - Plasmodium falciparum IT 147
  - Plasmodium falciparum KE01 146
  - Plasmodium falciparum KH01 146
  - Plasmodium falciparum KH02 148
  - Plasmodium falciparum ML01 151
  - Plasmodium falciparum SD01 146
  - Plasmodium falciparum SN01 147
  - Plasmodium falciparum TG01 151
  - ▶  Plasmodium fragile 94
  - ▶  Plasmodium gaboni 302
  - ▶  Plasmodium gallinaceum 129
  - ▶  Plasmodium iui 111
  - ▶  Plasmodium knowlesi 241
  - ▶  Plasmodium malariae 125
  - ▶  Plasmodium ovale curtisi 125
  - ▶  Plasmodium praefalciparum 144
  - ▶  Plasmodium reichenowi 291
  - ▶  Plasmodium relictum 128
  - ▶  Plasmodium vinckei 213
  - ▶  Plasmodium vivax 247

5. Export the results to a search strategy. (hint: to achieve this click on the blue *Export as a search strategy* button at the top right-hand side of the results).

The screenshot shows the 'My Search Strategies' page. At the top, there is a blue button labeled 'Export as a Search Strategy' with a white arrow pointing right. Below this, a red arrow points down to the search strategy interface. The interface includes a title 'Unnamed Search Strategy \*' and a 'Step 1' section containing a 'Text' box with '145 Genes'. The main results table shows 145 genes from Plasmodium falciparum, with columns for Gene ID, Transcript ID, Organism, Genomic Location (Gene), and Product Description. The results are paginated at the top.

Gene ID	Transcript ID	Organism	Genomic Location (Gene)	Product Description
PF3D7_0102600	PF3D7_0102600.1	Plasmodium falciparum 3D7	Pf3D7_01_v3:119,041..121,249(-)	serine/threonine kinase, FIK
PF3D7_0103700	PF3D7_0103700.1	Plasmodium falciparum 3D7	Pf3D7_01_v3:166,513..168,687(+)	L-seryl-RN putative
PF3D7_0107600	PF3D7_0107600.1	Plasmodium falciparum 3D7	Pf3D7_01_v3:314,618..319,405(+)	eukaryotic factor 2-alp

6. Return to the site search results page. How did you do this? (hint: you can achieve this in two ways: 1. Click on your browser's back arrow. 2. Click on the back to results arrow in the site search window. Notice that your previous results and filter settings were preserved.



7. Clear all filters. How did you do this? (hint: you can achieve this in two ways: 1. You can click on each of the clear filter options in the filter results panel on the left (boxes below). 2. You can click on the single *clear filters* option in the site search window.

The screenshot shows the BioMart interface. On the left, the 'Filter results' panel is open, displaying various filtering options. A red box labeled '1' highlights the 'Clear filter' button under the 'Filter Gene fields' section. Another red box highlights the 'Clear filter' button under the 'Filter organisms' section. At the top right, a search bar contains the word 'kinase'. A red box labeled '2' highlights the 'Clear filters' button in the search bar's toolbar.

**Filter results**

Hide zero counts

**Filter Gene fields**

[select all](#) | [clear all](#)

<input type="checkbox"/> Alternate product descriptions	3
<input type="checkbox"/> EC descriptions and numbers	320
<input type="checkbox"/> GO terms	156
<input type="checkbox"/> InterPro domains	165
<input type="checkbox"/> Orthologs	164
<input type="checkbox"/> PDB chains	112
<input checked="" type="checkbox"/> Product descriptions	145
<input type="checkbox"/> PubMed	126
<input type="checkbox"/> Rodent malaria phenotype	42
<input type="checkbox"/> User comments	50

**Filter organisms**

[select all](#) | [clear all](#) | [expand all](#) | [collapse all](#)

Type a taxonomic name  [?](#)

<input type="checkbox"/> Plasmodium adleri	157
<input type="checkbox"/> Plasmodium berghei	121
<input type="checkbox"/> Plasmodium bilcollinsi	147
<input type="checkbox"/> Plasmodium blacklocki	144
<input type="checkbox"/> Plasmodium chabaudi	122
<input type="checkbox"/> Plasmodium coatneyi	117
<input type="checkbox"/> Plasmodium cynomolgi	244
<input checked="" type="checkbox"/> Plasmodium falciparum	2,353
<input type="checkbox"/> Plasmodium fragile	0

8. Try the *Hide zero counts* check box in the *Filter results* panel. What does this do?

The screenshot shows the 'Filter results' panel with two states. On the left, the 'Hide zero counts' checkbox is checked (indicated by a red arrow pointing from the previous screenshot). This results in a list of items with counts: Genome (Genes: 16,684), Population biology (Popset isolate sequences: 249), Metabolism (Metabolic pathways: 345, Compounds: 85), Data access (Data sets: 1, Searches: 3). On the right, the 'Hide zero counts' checkbox is unchecked. This results in a list where all items have a count of 0: Genome (Genes: 16,684, Genomic sequences: 0), Organism (Organisms: 0), Transcriptomics (ESTs: 0), Population biology (Popset isolate sequences: 249, Field samples: 0), Metabolism (Metabolic pathways: 345, Compounds: 85), Data access (Data sets: 1, Searches: 3), Instructional (Tutorials: 0, Workshop exercises: 0), About (News: 0, General info pages: 0).

9. Try running a search with a wild card. The wild card is denoted by an asterisk \*. The wild card can be used alone to retrieve all results available to the site search or combined with a word such as \*kinase to retrieve compound words ending with the word kinase like phosphofructokinase. As usual results can then be explored using the filters in the *Results filter* on the left side of the website.

### All results matching \*

1 - 20 of 516,501

Compound - CHEBI:10000	Vismione D	
Compound - CHEBI:10001	Visnadin	
Compound - CHEBI:10002	Visnagin	
Compound - CHEBI:10003	ribostamycin sulfate	
<b>Definition:</b> An aminoglycoside sulfate salt resulting from the reaction of ribostamycin with sulfuric acid.		
Compound - CHEBI:100147	naldixic acid	
<b>Definition:</b> A monocarboxylic acid comprising 1,8-naphthyridin-4-one substituted by carboxylic acid, ethyl and methyl groups at positions 3, 1, and 7, respectively.		
Compound - CHEBI:10014	Voacamine	
Compound - CHEBI:10015	vobasine	
<b>Definition:</b> An indole alkaloid that is vobasine in which the bridgehead methyl group is substituted by a methoxycarbonyl group and an additional oxo substituent is present in the 3-position.		
Compound - CHEBI:10016	vobtusine	
Compound - CHEBI:10017	volementol	
<b>Definition:</b> A heptitol that is heptane-1,2,3,4,5,6,7-heptol that has R-configuration at positions 2, 3, 5 and 6.		
Compound - CHEBI:10018	volkenin	
<b>Definition:</b> A cyanogenic glycoside that is (4R)-4-hydroxycyclopent-2-ene-1-carbonitrile attached to a beta-D-glucopyranosyloxy at position 1.		
Compound - CHEBI:10019	Vomicine	

### All results matching \*kinase

1 - 20 of 18,073

Gene - AK88_00104	CK1/CK1/CK1-D protein kinase	
<b>Organism:</b> Plasmodium fragile strain nilgiri		
▶ Fields matched: EC descriptions and numbers; GO terms; InterPro domains; Orthologs; PDB chains; Product descriptions		
Gene - AK88_00479	CAMK protein kinase	
<b>Organism:</b> Plasmodium fragile strain nilgiri		
▶ Fields matched: EC descriptions and numbers; GO terms; InterPro domains; Orthologs; PDB chains; Product descriptions		
Gene - AK88_00505	pantothenate kinase	
<b>Organism:</b> Plasmodium fragile strain nilgiri		
▶ Fields matched: EC descriptions and numbers; GO terms; Orthologs; PDB chains; Product descriptions		
Gene - AK88_00565	Atypical/ABC1 protein kinase	
<b>Organism:</b> Plasmodium fragile strain nilgiri		
▶ Fields matched: InterPro domains; Orthologs; Product descriptions		

10. Try searching for a specific gene ID. Enter the gene ID below in the site search window:

*PF3D7\_0310100*

The screenshot shows the PlasmoDB search results for the gene ID PF3D7\_0310100. The search bar at the top contains the query. Below the search bar, there are links for "My Strategies", "Searches", "Tools", "My Workspace", "Data", "About", "Help", and "Contact Us". On the right side, there are social media icons for Twitter, Facebook, and YouTube, along with a user profile icon for "Omar". A blue button on the right says "Export as a Search Strategy" with a link to "download or mine your results". The main content area displays "Genes matching PF3D7\_0310100" and "1 - 2 of 2". On the left, a sidebar titled "Filter results" shows a count of 2 for "Genome Genes". Below it, "Filter Gene fields" lists "External links" (1), "Gene ID" (1), and "Notes from annotators" (1). Further down, "Filter organisms" lists "Plasmodium falciparum" (1) and "Plasmodium gaboni" (1). The main results area shows two entries: "Gene - PF3D7\_0310100 calcium-dependent protein kinase 3" and "Gene - PGSY75\_0310100 calcium-dependent protein kinase 3". Both entries show "Gene name or symbol: CDPK3" and "Organism: Plasmodium falciparum 3D7". Below each entry is a note about matched fields: "Fields matched: External links; Gene ID" for the first and "Fields matched: Notes from annotators" for the second.

Notice that the gene of interest appears at the top for easy access. You can click on the Gene ID to go the gene page.

# Search Strategies

**Note:** this exercise uses *PlasmoDB* (<https://PlasmoDB.org>) as an example database, but the same functionality is available on all *VEuPathDB* resources.

## Learning objectives:

- Running a search to start a search strategy
- Adding steps in a search strategy
- Adding and sorting results
- Revising steps

There are three options to start a Search Strategy. 1) From the “Site Search” box ---> Export as Search Strategy, 2) In the site header from the “Searches” menu and 3) In the home page (left hand side) from the “Search for ...” section.

The screenshot shows the PlasmoDB beta homepage. A red box labeled '1' highlights the top navigation bar's 'Searches' menu. A red box labeled '2' highlights the 'Search for...' dropdown menu on the left sidebar. A red box labeled '3' highlights the 'Search for...' input field on the left sidebar. The page features a purple header with the PlasmoDB logo and a 'Release 48 beta 27 Aug 2020' notice. The main content area includes sections for 'Overview', 'Getting Started', and links to 'Transcriptomic Resources', 'Phenotypic Data', and 'Analyze My Data'.

1. Go to the home page and in the “Search for ...” section on the left, filter the searches by typing the word *transmembrane* to find the ***Transmembrane Domain Count*** search in the filtered results.

**Search for...**

expand all | collapse all

Filter the searches below...

- ▶ Genes
- ▶ Organisms
- ▶ Popset Isolate Sequences
- ▶ Genomic Sequences
- ▶ Genomic Segments
- ▶ SNPs
- ▶ SNPs (from Array)
- ▶ ESTs
- ▶ Metabolic Pathways
- ▶ Compounds

**Search for...**

transm

Genes

Protein targeting and localization

Transmembrane Domain Count

2. Click on the transmembrane (TM) domain count search to get to the search page. Configure this search to find all genes from *Plasmodium vivax* P01 that have at least 6 TM domains and at most 8 TM domains. See image below if you need help with the configuration.

## Identify Genes based on Transmembrane Domain Count

1 Selected, OUT OF 45

add these | clear these | select only these  
select all | clear all

vivax  ←

Plasmodium vivax  Plasmodium vivax P01 ←  
 Plasmodium vivax Sal-1  
 Plasmodium vivax-like sp.  
 Plasmodium vivax-like PvI01

add these | clear these | select only these  
select all | clear all

Minimum Number of Transmembrane Domains  ←

Maximum Number of Transmembrane Domains  ←

Get Answer ←

3. How many genes did you obtain? (hint: look at the number results in the strategy step in yellow, or the number right above the results and below the search strategy).

## My Search Strategies

Opened (1) All (404) Public (42) Help

Unnamed Search Strategy \*

Transmb Dom  
101 Genes

+ Add a step

Step 1

101 Genes (97 ortholog groups) Revise this search

Gene Results Genome View Analyze Results

Organism Filter select all | clear all | expand all | collapse all Hide zero counts

Search organisms...

Gene ID	Transcript ID	Organism	Genomic Location (Transcript)	# TM Domains
PVP01_0104300	PVP01_0104300.1	Plasmodium vivax P01	PvP01_01_v1:213970..224298(+)	6
PVP01_0113600	PVP01_0113600.1	Plasmodium vivax P01	PvP01_01_v1:607892..610837(+)	6
PVP01_0114900	PVP01_0114900.1	Plasmodium vivax P01	PvP01_01_v1:664719..665566(-)	6
PVP01_0317200	PVP01_0317200.1	Plasmodium vivax P01	PvP01_03_v1:744775..747000(+)	6
PVP01_0606500	PVP01_0606500.1	Plasmodium vivax P01	PvP01_06_v1:261479..262988(-)	6
PVP01_0703300	PVP01_0703300.1	Plasmodium vivax P01	PvP01_07_v1:187292..193820(-)	6
PVP01_0706600	PVP01_0706600.1	Plasmodium vivax P01	PvP01_07_v1:352530..354459(-)	6

4. Explore the results table. Try the following things:

- Sort the #TM domain column to show genes with 8 TM domains first.
- Add a column for transcript length (Click on add columns and find the transcript length column, then click on update columns).

Gene Results Genome View Analyze Results

Rows per page: 50

Download Add to Basket Add Columns

Gene ID	Transcript ID	Organism	Genomic Location (Transcript)	# TM Domains
PVP01_0208500	PVP01_0208500.1	Plasmodium vivax P01	PvP01_02_v1:3252862..356879(+)	8
PVP01_0412900	PVP01_0412900.1	Plasmodium vivax P01	PvP01_04_v1:530187..531415(-)	8
PVP01_0509600	PVP01_0509600.1	Plasmodium vivax P01	PvP01_05_v1:429961..434135(-)	8
PVP01_0702700	PVP01_0702700.1	Plasmodium vivax P01	PvP01_07_v1:158919..167549(-)	8
PVP01_0817900	PVP01_0817900.1	Plasmodium vivax P01	PvP01_08_v1:778581..780290(+)	8
PVP01_0914100	PVP01_0914100.1	Plasmodium vivax P01	PvP01_09_v1:554670..655704(+)	8
PVP01_0936100	PVP01_0936100.1	Plasmodium vivax P01	PvP01_09_v1:1551091..1552320(-)	8
PVP01_1011300	PVP01_1011300.1	Plasmodium vivax P01	PvP01_10_v1:501278..502747(-)	8
PVP01_1028700	PVP01_1028700.1	Plasmodium vivax P01	PvP01_10_v1:1224465..1226132(-)	8

Select Columns

Update Columns

select all | clear all | expand all | collapse all | current | reset to default |

Search Columns

Gene models

- # Exons in Gene
- # Transcripts
- Annotated 5' UTR length
- Annotated 3' UTR length
- Gene Strand
- Gene Type
- Is Pseudo
- JBrowse
- Transcript Length

Annotation, curation and identifiers

- Link outs
- Genomic Location
- Chromosome
- Genomic location (Gene)
- Genomic Location (Transcript)
- Genomic Sequence ID

Taxonomy

- Organism
- Phylogeny and synteny
- Phenotype
- Genetic variation
- Transcription
- Protein features and properties
- Protein targeting and localization
- SignalPeptide

Add Columns

5. Add a step to your strategy. Click on the add step button then find the search for genes with GO Terms. When you find it select it and configure the search to find all genes with the GO term “Transporter activity”. See screen shots below if you need help.

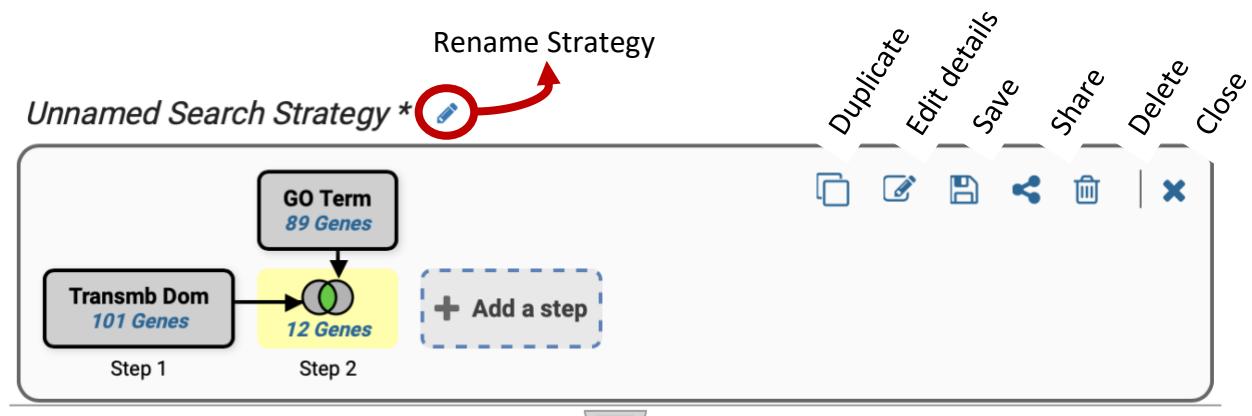
The top screenshot shows the main search strategy interface. A red arrow points from the 'Add a step' button in the 'Step 1' section to the 'Add a step to your search strategy' dialog box. Inside the dialog, another red arrow points to the '1 INTERSECT 2' radio button under 'Choose how to combine with other Genes'. The search bar contains 'go|'.

The bottom screenshot shows the 'Add a step to your search strategy' dialog box expanded. A red arrow points to the 'vivax' entry in the 'Organism' dropdown. Another red arrow points to the 'GO Term or GO ID' input field containing 'GO:0005215 : transporter activity'.

Notice that you have different options on how to combine results from searches in your strategy. What do you each of the operations do?

Operator	Combined Result will contain
<input checked="" type="radio"/>  2 INTERSECT 3	IDs in common between the two lists
<input type="radio"/>  2 UNION 3	IDs from list 2 and list 3
<input type="radio"/>  2 MINUS 3	IDs unique to 2
<input type="radio"/>  3 MINUS 2	IDs unique to 3
	IDs whose features are near each other (collocated) in the genome

6. You can rename, duplicate, delete, save and share strategies (saving and sharing strategies requires creating an account and logging in). You can also rename each



individual step in a strategy.

7. Revising a step in a strategy. You can revise any step in a strategy by moving your mouse over the step you want to revise until you see the edit button appear on the step.
8. Revise the first step in your strategy and change the TM domain parameter to include genes with a minimum of 5 TMs and a maximum of 12 TMs. How does this change your final results?

Opened (1) All (404) Public (42) Help

Unnamed Search Strategy \*

Step 1 Step 2

**Details for step Transmb Dom**  
101 Genes

Organism Plasmodium vivax P01

Minimum Number of Transmembrane Domains 6

Maximum Number of Transmembrane Domains 8

► Give this search a weight

12 Genes (12 ortholog groups)

Organism Filter select all | clear all | expand all | collapse all

Rows per page: 50

Download Ad

Transmb Dom 101 Genes

GO Term 89 Genes

Minimum Number of Transmembrane Domains  
5

Maximum Number of Transmembrane Domains  
12

Revise

## Advanced Search Strategies

**Note:** this exercise uses PlasmoDB.org as an example database, but the same functionality is available on all VEuPathDB resources.

### Learning objectives:

- Integrate diverse datatypes in a search strategy
- Leverage orthology and phylogenetic profile searches

This exercise walks you through the process of building a multi-step strategy, integrating different datatypes. The final search strategy identifies plasmodium genes that are likely secreted, or membrane bound, highly polymorphic, “essential” for parasite survival, not conserved in mammals and expressed in liver stages of the Plasmodium life cycle. There are many ways to build these strategies and order the steps to reach a similar answer.

1. Identify all genes in PlasmoDB that are predicted to have a secretory signal peptide as defined by SignalP. An easy way to identify a search type is to filter the searches on the left of the home page. Start typing a word to identify the search type. For example, start typing the word "secreted", you should see the searches being filtered even before you finish typing the complete word.

The screenshot shows the PlasmoDB beta homepage. On the left, there is a sidebar with a 'Search for...' dropdown menu containing options like 'Genes', 'Organisms', 'Popset Isolate Sequences', etc. A red arrow points from this menu to a search bar on the main page. The main page has a large search bar at the top with the placeholder 'Site search, e.g. PF3D7\_1133400 or \*reductase or \*binding protein\*'. Below it, a 'Search for...' box is highlighted with a blue border. Inside this box, the word 'secr' is typed into a search input field. Another red arrow points from this input field to a sub-section titled 'Predicted Signal Peptide' which is also highlighted with a blue border. This section contains a brief description of SignalP and a 'Read More' link. The top of the page features the PlasmoDB logo, a release date of 'Release 48 beta 27 Aug 2020', and various navigation links like 'My Strategies', 'Searches', 'Tools', 'My Workspace', 'Data', 'About', 'Help', and 'Contact Us'. The right side of the page includes social media icons and a user profile for 'Omar'.

2. Click on the search for genes by predicted signal peptide. On the next page select all organisms and click on the get answer button at the bottom of the page.

## Identify Genes based on Predicted Signal Peptide

### Organism

*Note: You must select at least 1 values for this parameter.*  
45 selected, out of 45

[select all](#) | [clear all](#) | [expand all](#) | [collapse all](#)

(Filter list below...) ?

- ▶  Plasmodium adleri
- ▶  Plasmodium berghei
- ▶  Plasmodium bilcollinsi
- ▶  Plasmodium blacklocki
- ▶  Plasmodium chabaudi
- ▶  Plasmodium coatneyi
- ▶  Plasmodium cynomolgi
- ▶  Plasmodium falciparum
- ▶  Plasmodium fragile
- ▶  Plasmodium gaboni
- ▶  Plasmodium gallinaceum
- ▶  Plasmodium inui
- ▶  Plasmodium knowlesi
- ▶  Plasmodium malariae
- ▶  Plasmodium ovale curtisi
- ▶  Plasmodium praefalciparum
- ▶  Plasmodium reichenowi
- ▶  Plasmodium relictum
- ▶  Plasmodium vinckeii
- ▶  Plasmodium vivax
- ▶  Plasmodium vivax-like sp.
- ▶  Plasmodium yoelii

[select all](#) | [clear all](#) | [expand all](#) | [collapse all](#)

### Advanced Parameters

↓ Get Answer

3. The next step is to combine the signal peptide results with results of genes that are predicted to have at least one transmembrane domain (TM). Click on the add step button in the search strategy panel.

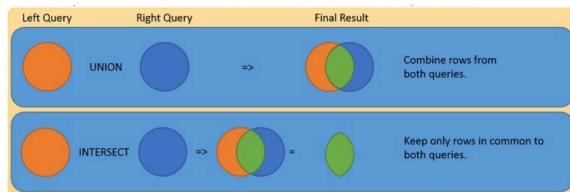
## My Search Strategies

Opened (1) All (415) Public (42) Help

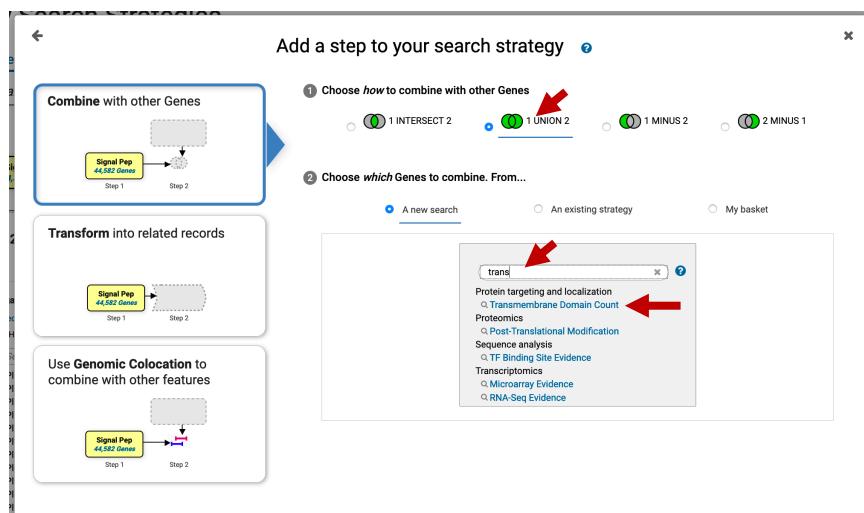
Unnamed Search Strategy \* (edit)

The screenshot shows the 'Search Strategy' interface. At the top, there's a header with 'Opened (1)', 'All (415)', 'Public (42)', and 'Help'. Below it is a section titled 'Unnamed Search Strategy \*' with a blue '(edit)' link. The main area contains a single step: 'Signal Pep 44,582 Genes'. To the right of this step is a blue dashed box containing a white plus sign and the text '+ Add a step'. A red arrow points from the text 'Add a step' towards this button. At the bottom right of the interface are several small icons: a square, a magnifying glass, a document, a person, a gear, a trash can, and an 'X'.

The popup window offers you option to add additional steps and ways to combine the searches (intersect, union, minus). For this exercise we are interested in finding genes that have a signal peptide or a TM domain or both. What operation will you use to combine the searches – Union or Intersect?

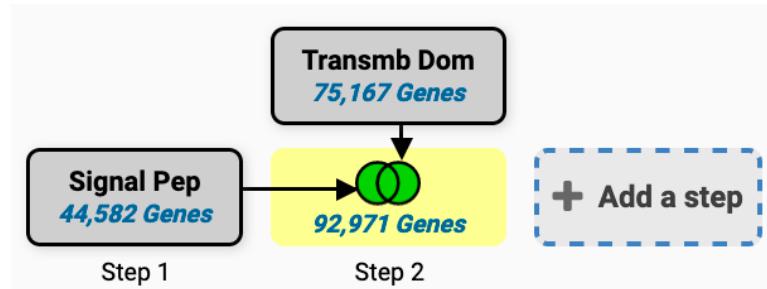


Once you select the option for combining the searches, find the search for transmembrane domain count. Notice that you can use the same query filtering mechanism as before. Start typing transmembrane to find this search. Once you find it click on to open the search parameters.



- For the TM search, again select all organisms, use the default parameters and click on the get answer button.

5. How many genes did you get? Since you used a union the number of results should be more than each of the individual steps that were combined.



6. Next, identify genes from step 2 that contain at least 5 non-synonymous SNPs (non-synonymous SNPs are single nucleotide polymorphisms that result in an amino acid change). Were you able to find the SNP search by clicking on add step and filtering the searches with a keyword? Which operation will you select to combine the searches?

7. On the Genes by SNP characteristics search popup, select Plasmodium falciparum from the drop down and select all available isolates by selecting the checkbox at the top of the filter panel (See image below).

8. Next scroll down and select the following parameters. SNP class = Non-synonymous. Number of SNPs of above class  $\geq 5$ . After you select these parameters, scroll down to the bottom and click on Run Step.

[←](#) Add a step to your search strategy [?](#)

[expand all](#) | [collapse all](#)

**② Read frequency threshold**

[▼](#)

**② Minor allele frequency  $\geq$**

**② Percent isolates with a base call  $\geq$**

**② SNP Class**

[▼](#) 

**② Number of SNPs of above class  $\geq$**



**② Number of SNPs of above class  $\leq$**

What do the results look like? What species are represented in the results? Is this surprising? Remember that your last search only queried *P. falciparum* data.

## My Search Strategies

Opened (1) All (415) Public (42) Help

Unnamed Search Strategy \* 



1,578 Genes (6,987 ortholog groups)

Some Genes in your combined result have Transcripts that were not returned by one or both of the two input searches. [Explore](#)

Gene Results		Genome View		Analyze Results	
Organism Filter	select all   clear all   expand all   collapse all	Rows per page:	50	Download	Add to Basket
<input type="text" value="Search organisms..."/> <span>?</span>				<span>Gene ID</span> <span>Transcript ID</span> <span>Genomic Location (Gene)</span> <span>Product Description</span> <span>Ortholog Group</span>	
<input type="checkbox"/> Plasmodium adleri <input type="checkbox"/> Plasmodium berghei <input type="checkbox"/> Plasmodium billrothi <input type="checkbox"/> Plasmodium blacklocki <input type="checkbox"/> Plasmodium chabaudi <input type="checkbox"/> Plasmodium coatneyi <input type="checkbox"/> Plasmodium cynomolgi <input type="checkbox"/> Plasmodium falciparum		Genes: 1,578   Transcripts: 1,597 <input type="checkbox"/> Show Only One Transcript Per Gene		rifin rifin erythrocyte membrane protein 1 (PfEMP1), exon 1, pseudogene rifin	
1,578		1 2 3 ... 32		<a href="#">Ortholog Group</a> <span>?</span> <span>x</span> 	
Ortholog Group Filter		Download		<a href="#">Add to Basket</a>	
Ortholog Group Filter		Download		<a href="#">Add to Basket</a>	

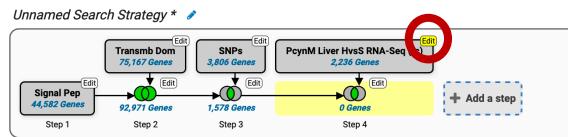
9. Determine how many of these genes are also differentially expressed in liver stages. Click on add step then search for the RNA-seq search. Type RNA in the search filter in the popup.

10. On the next page find data that queries liver stages. You can filter the data by typing the word liver in the filter box at the top of the page. This should yield two datasets from *P. cynomolgi* and *P. vivax*. For this exercise, select the fold change query for the *P. cynomolgi* dataset: Liver stage hypnozoite vs schizont transcriptomes (primary culture) (Voorverg-van der Wel et al.).

11. Configure the RNA-Seq search to identify genes that are differentially regulated by at least 2-fold between all the hypozoite stages and the sporozoite stages. For example, select the hypozoite stages in the reference selection box and the sporozoite samples in the comparator selection box, then click on run step.

12. How many results did you get? Why did you get 0 results? How can you change this? Remember that the previous search was a list of *P. falciparum* genes and this RNA-Seq was from *P. cynomolgy*. What you would like to do is convert the *P. cynomolgy* genes into *P. falciparum* genes. To do this follow these steps:

- hover your mouse over the RNA-seq step then click on the edit option on that step.



- In the popup window, click on the **orthologs** link.

- c. In the next window select which organism(s) you would like to transform to. For this exercise select *P. falciparum* 3D7 and click on run step.

**Organism**

Note: You must select at least 1 values for this parameter.  
1 selected, out of 45

[add these](#) | [clear these](#) | [select only these](#)  
[select all](#) | [clear all](#)

3d7

Plasmodium falciparum

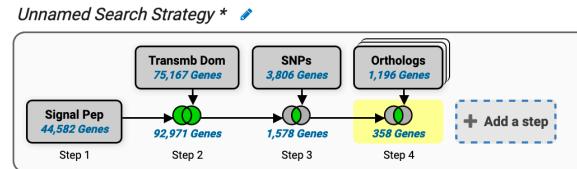
[add these](#) | [clear these](#) | [select only these](#)  
[select all](#) | [clear all](#)

**Syntenic Orthologs Only?**

no

**Run Step**

- d. Did you get results now?



13. Next identify how many of these genes do not have orthologs in mammals. To do this add a step for genes based on orthology phylogenetic profile. Again you can filter the searches by typing the word “phylogenetic”.

Add a step to your search strategy

① Choose how to combine with other Genes

4 INTERSECT 5  4 UNION 5  4 MINUS 5  5 MINUS 4

② Choose which Genes to combine. From...

A new search  An existing strategy  My basket

phy

Orthology and synteny  
↳ Orthology Phylogenetic Profile

Combine with other Genes  
Transform into related records  
Use Genomic Colocation to combine with other features

On the next page select *P. falciparum* 3D7 the configure the phylogenetic profile by finding Mammalia under Chordata which are under Metazoa. Click twice on the circle next to Mammalia – it should become a red x (See image below).

Add a step to your search strategy x

add these |  clear these |  select only these  
 select all |  clear all

3d7 ←

Plasmodium falciparum  
 Plasmodium falciparum 3D7

add these |  clear these |  select only these  
 select all |  clear all

**Select orthology profile**

Click on + to determine which organisms to include or exclude in the orthology profile.  
( ) = no constraints / \* = must be in group / \* = must not be in group / ? = mixture of constraints

All Organisms + expand all | collapse all

- Bacteria (BACT)
- Firmicutes (FIRM)
- Proteobacteria (PROT)
- Other Bacteria (OBAC)
- Archaea (ARCB)
  - Nitrosopumilus maritimus SCMT1 (nmar)
  - Euryarchaeota (EURY)
  - Crenarchaeota (CREN)
  - Nanoarchaeota (NANO)
  - Korarchaeota (KORA)
- Eukaryota (EUKA)
  - Alveolates (ALVE)
  - Amoebozoa (AMOE)
  - Euglenozoia (EUGL)
  - Viridiplantae (VIRI)
  - Fungi (FUNG)
  - Metazoa (META)
    - Nemata (NEMA)
    - Arthropoda (ARTH)
    - Chordata (CHOR)
      - Branchiostoma floridae (bflo)
      - Xenopus (Silurana) tropicalis (xtro)
      - Actinopterygii (ACTI)
      - Aves (AVES)
      - Mammalia (MAMM)
      - Tunicates (TUNI)
      - Other Metazoa (OMET)
  - Other Eukaryota (OEUK)

←

14. Determine if a mutation in any of these genes affects fitness. Click on add step and find the search for phenotype evidence.

Add a step to your search strategy x

**Combine with other Genes**

OrthoPh Pro 3,306 Genes + Step 5 Step 6

**Transform into related records**

OrthoPh Pro 3,306 Genes + Step 5 Step 6

**Use Genomic Colocation to combine with other features**

OrthoPh Pro 3,306 Genes + Step 5 Step 6

**Choose how to combine with other Genes**

① 5 INTERSECT 6  ② 5 UNION 6  ③ 5 MINUS 6  ④ 6 MINUS 5

**Choose which Genes to combine. From...**

A new search  An existing strategy  My basket

phen Phenotype  Phenotype Evidence

15. Select the P. falciparum piggyBac insertion mutagenesis (John Adams) experiment.

Add a step to your search strategy x

**Search for Genes by Phenotype Evidence**

The results will be + intersected with  the results of Step 5.

Legend: + AS Association to Genomic Segments CP Curated Phenotype S Similarity SA Similarity of Association

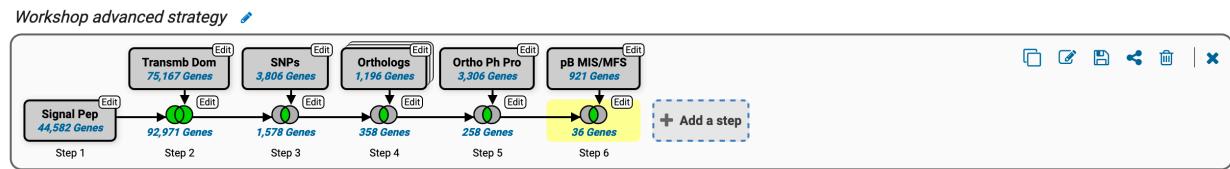
Filter Data Sets:  + AS CP S SA

Organism	Data Set	Choose a Search
Plasmodium berghei ANKA	P. berghei knockout (PlasmoGEM) growth phenotypes (Bushell, Gomes and Sanderson et al.)	<span style="color: blue;">CP</span>
Plasmodium berghei ANKA Plasmodium falciparum 3D7 Plasmodium yoelii yoelii 17XLN	RMgM08 - Rodent Malaria genetically modified Parasites (Chris J. Janse)	<span style="color: blue;">CP</span>
Plasmodium falciparum 3D7	eQTL for HB3, Dd2 and 34 progeny (Gonzales et al.)	<span style="color: blue;">AS</span> <span style="color: blue;">S</span> <span style="color: grey;">SA</span>
Plasmodium falciparum 3D7	piggyBac insertion mutagenesis (John Adams)	<span style="color: blue;">CP</span> <span style="color: red; border: 2px solid red; border-radius: 50%; padding: 2px;">CP</span>

16. On the next page select the Mutan Fitness Score (MFS) option and choose any score range – generally the more negative the bigger the effect is on fitness. For this example a score range of -4.078 to -3.07 was chosen.



Explore your final results. Do they make sense/plausible? Note that you can revise any of the steps in the strategy to explore the data further. You can also save your strategy and share it with others or make it public. Here is a link to this search stragey:



<https://plasmodb.org/plasmo/app/workspace/strategies/import/fd387e8d3acda856>

## Exploring the Gene Page

**Note:** this exercise uses *TriTrypDB* (<https://TriTrypDB.org>) as an example database, but the same functionality is available on all *VEuPathDB* resources.

### Learning objectives

#### Gene pages:

- Become familiar with the information in gene pages
- Navigate to and from the gene pages

#### 1. Navigation to the Gene pages

For this exercise visit the gene page for Tb927.10.13780 (Glycogen synthase kinase 3). How did you get to this gene? (hint: copy and paste the ID in the site search, then click on the gene ID in the results.

The screenshot shows the TriTrypDB gene search results for the query "Tb927.10.13780". The results section displays a single entry:

**Gene - Tb927.10.13780 Glycogen synthase kinase 3 short**

Gene name or symbol: GSK3s  
Organism: Trypanosoma brucei brucei TREU927

Fields matched: Cellular localization; External links; Gene ID; GO terms; Transcripts

**Filter results**

Genome	1
Genes	1

**Filter Gene fields**

select all   clear all	
<input type="checkbox"/> Cellular localization	1
<input type="checkbox"/> External links	1
<input type="checkbox"/> Gene ID	1
<input type="checkbox"/> GO terms	1
<input type="checkbox"/> Transcripts	1

**Filter organisms**

select all   clear all   expand all   collapse all	
Type a taxonomic name	<input type="text"/>
<input type="checkbox"/> Trypanosomatidae	1
<input type="checkbox"/> Trypanosoma	1

Export as a Search Strategy ►

#### 2. Explore the top section of the gene page

What information is in this section? Can you easily find which chromosome this gene is located on? Does this gene have any user comments?

Add to basket Add to favorites Download Gene

# Tb927.10.13780 Glycogen synthase kinase 3 short

**Name:** GSK3s  
**Type:** protein coding  
**Chromosome:** 10  
**Location:** Tb927\_10\_v5.1:3,361,774..3,366,257(-)

**Species:** Trypanosoma brucei  
**Strain:** brucei TREU927  
**Status:** Curated Reference Strain

This genome is actively curated at GeneDB. User comments added to this gene will be reviewed and incorporated into the official annotation if appropriate.

**Shortcuts**

Synteny BLAT Alignments Phenotype SNPs Transcriptomics Protein Features Proteomics

View this gene at GeneDB   
View 3 user comments, or add a comment 

Also see Tb927.10.13780 in the [Genome Browser](#) or [Protein Browser](#)

### **3. Explore the gene model section.**

Scroll down to the gene model section of the gene page. What direction is the transcript relative to the chromosome? Does the gene have UTRs?

**1 Gene models**

# Exons in Gene 1

# Transcripts 1

Gene Models

View in JBrowse genome browser

Scroll and zoom ?

Annotated Transcripts (UTRs in White when available)

3,350,000 3,362,500 3,375,000 3,387,500

Tb927.10.13730 mRNA  
Tb927.10.13730  
Tb927.10.13740 mRNA  
Tb927.10.13740  
Tb927.10.13750 mRNA  
Tb927.10.13750  
Tb927.10.13770 mRNA  
Tb927.10.13770  
Tb927.10.13790 mRNA  
Tb927.10.13790  
Tb927.10.13800 mRNA  
Tb927.10.13800  
Tb927.10.13820 mRNA  
Tb927.10.13820  
Tb927.10.13840 mRNA  
Tb927.10.13840  
Tb927.10.13850 mRNA  
Tb927.10.13850  
Tb927.10.13860 mRNA  
Tb927.10.13860  
Tb927.10.13870 mRNA  
Tb927.10.13870  
Tb927.10.13890 mRNA  
Tb927.10.13890  
Tb10.NT.146 mRNA  
Tb10.NT.146  
Tb10.NT.148 mRNA  
Tb10.NT.148  
13700.rRNAc mRNA  
13700.rRNAc  
7.10.13700 mRNA  
7.10.13700  
927.10.13710 mRNA  
927.10.13710  
Tb927.10.13710

View in JBrowse genome browser

How long is the transcript? You can determine transcript length by expanding the Transcripts section.

View in SBrowse <a href="#">Genomic Browser</a>			
▼ Transcripts	<a href="#"> Download</a>	<a href="#"> Data sets</a>	
Tb927.10.13780:mRNA	1	4484	352

## 4. Content navigation.

How do you find/navigate to the different sections of the page? Use the “Contents” menu on the left side, type a keyword and click on the menu, click on the work to

navigate to it on the page. In the example below the word “synteny” is used. You can also click on the images in the Shortcuts section in the top of the page.

## 5. Running an alignment of selected sequences

- Expand the “Orthologs and Paralogs in TriTrypDB” section.
- Select a few genes from the table using the checkbox.
- Scroll to the bottom of the table and click on the Run Clustal Omega button.

<input checked="" type="checkbox"/>	TcYC6_0115420	Trypanosoma cruzi Y C6	protein kinase
<input type="checkbox"/>	Tc_MARK_4866	Trypanosoma cruzi marinellei strain B7	glycogen synt alpha, putative
<input type="checkbox"/>	TevSTIB805.10.14480	Trypanosoma evansi strain STIB 805	glycogen synt
<input type="checkbox"/>	DQ04_000191000	Trypanosoma grayi ANR4	putative glyco kinase-3 alpha
<input checked="" type="checkbox"/>	TM35_000033680	Trypanosoma theileri isolate Edinburgh	putative glyco kinase-3 alpha
<input type="checkbox"/>	TvY486_1013940	Trypanosoma vivax Y486	protein kinase

Select sequence type for Clustal Omega multiple sequence alignment:

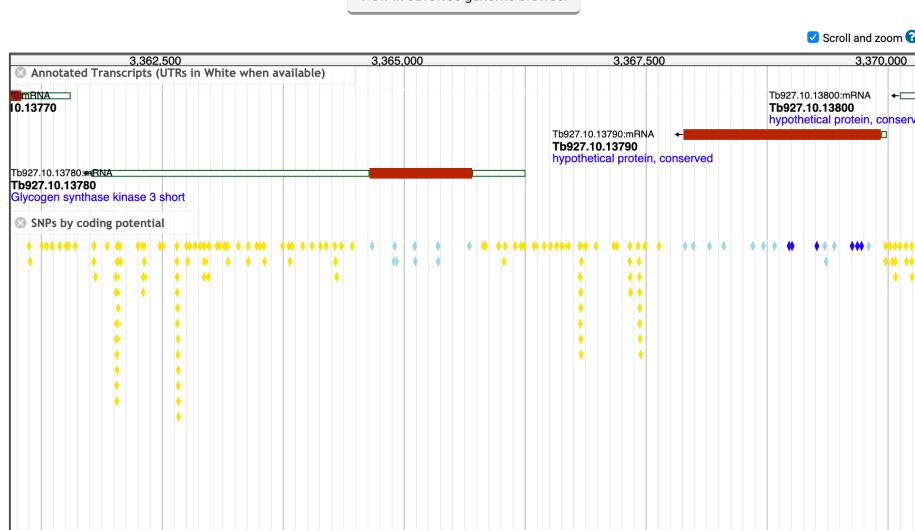
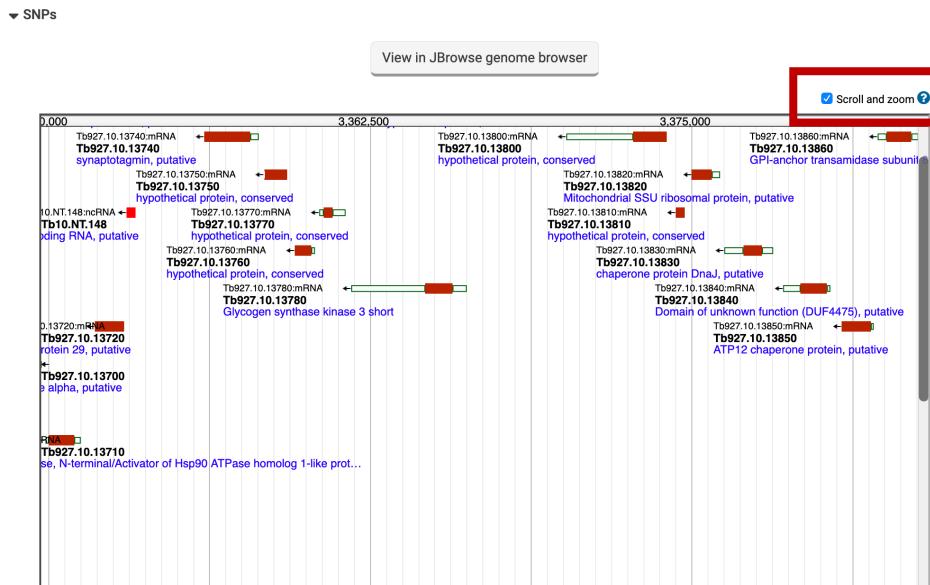
Please note: selecting a large flanking region or a large number of sequences will take several minutes.

Protein  CDS (spliced)  Genomic

Output format:

## 6. Exploring the genetic variation section

Go to the Genetic variation section of the gene page and expand the SNP section. Notice that by default you cannot scroll within the embedded browser window. To enable scrolling, select the “Scroll and Zoom” check box in the upper right-hand side of the browser window. To scroll down within the browser window, you click and drag or use two-finger scrolling. You can also double click in an area to zoom in.



## 7. Explore other sections of the gene page.

Feel free to scroll around the gene page and ask questions for clarification. Here are some questions you may want to ask about this gene:

- a. Is there evidence that this protein is phosphorylated? (hint: go to the proteomics section and expand the Post Translational Modification section).
- b. Where is the protein localized? (hint: go to the Protein Targeting and Localization section and expand the cellular localization section).
- c. Is there any phenotypic data available for this gene? (hint: go to the Phenotype section and expand its subsections).
- d. Is there any RNA-Seq data available for this gene? (hint: go to the Transcriptomics section and expand the subsections called RNA-Seq transcription summary and Transcript Expression).

## JBrowse Basics

**Note:** this exercise uses *TriTrypDB* as an example database, but the same functionality is available on all VEuPathDB resources.

### Learning objectives:

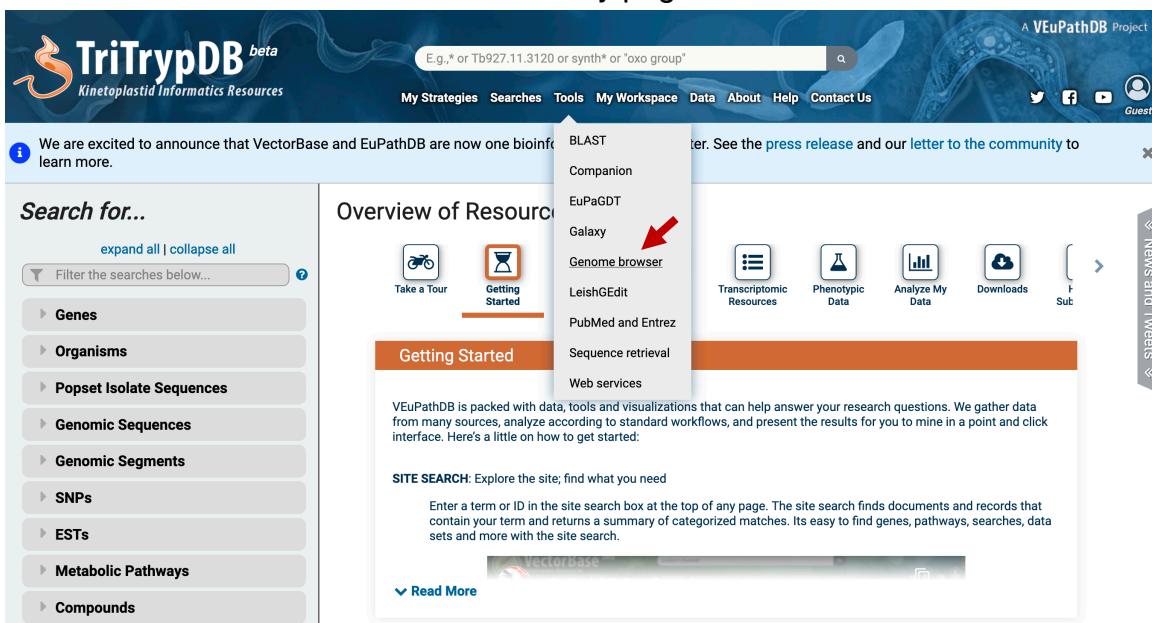
- Navigate to the genome browser
- Use the menu and navigation bars
- Run searches
- Add pre-loaded data tracks
- Upload your own data tracks
- Configure tracks
- Download track data

### 1. Navigating to the Genome Browser (JBrowse)

JBrowse is a fast and full-featured genome browser built with JavaScript and HTML5. You can read more about JBrowse and its features here:  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4830012/>

Links to the genome browser are available from multiple locations:

- a. The tools menu in the banner of any page.



The screenshot shows the TriTrypDB homepage. At the top, there's a banner with a message about VectorBase and EuPathDB being merged. Below the banner, the header includes the TriTrypDB logo, a search bar, and links for My Strategies, Searches, Tools, My Workspace, Data, About, Help, and Contact Us. On the far right are social media icons and a 'Guest' link. The main content area has a blue header 'Overview of Resources'. On the left, there's a sidebar titled 'Search for...' with categories like Genes, Organisms, Popset Isolate Sequences, etc. In the center, under 'Overview of Resources', there's a grid of icons for various tools: Take a Tour, Getting Started (which is highlighted with a red arrow), EuPaGDT, Galaxy, Genome browser (also highlighted with a red arrow), LeishGEedit, PubMed and Entrez, Sequence retrieval, and Web services. To the right of the grid, there are links for Transcriptomic Resources, Phenotypic Data, Analyze My Data, Downloads, and a 'Sub' link. At the bottom of the central column, there's a 'Read More' button. A vertical sidebar on the right is titled 'News and Tweets'.

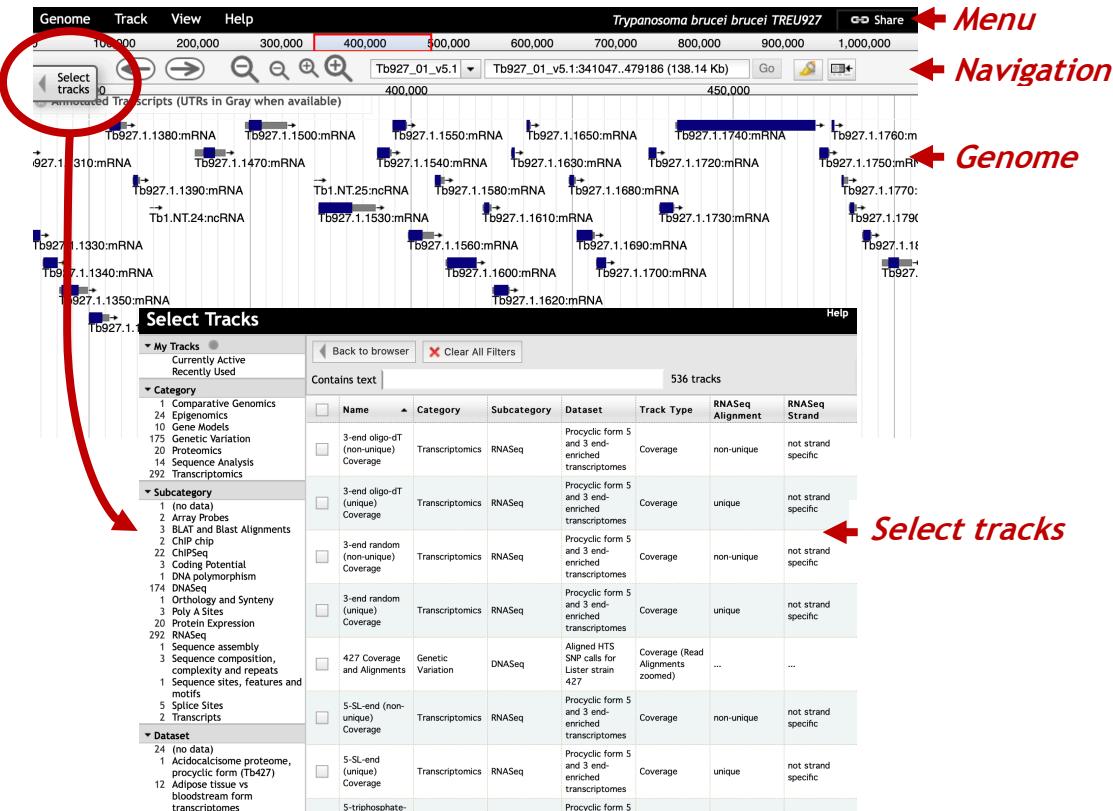
- b. From record pages such as gene, SNP or genomic sequence pages – these links are usually to a specific JBrowse configuration that includes data relevant to the section on that record page. For example, a JBrowse link from an RNAseq dataset on the gene page would display the gene of interest

along with the RNAseq data as density or coverage plots. These links are usually indicated by “View in JBrowse genome browser” button.

## View in JBrowse genome browser

### 2. Getting around JBrowse.

- Use any of the above described JBrowse linking strategies to get to the genome browser.
- Once in JBrowse examine the following features:
  - The **menu bar**: located at the top of the JBrowse frame. This includes the Genome menu, Track menu, View menu, Help menu and the Sharing link. What do each of these do/provide?
  - The **navigation bar**: located below the menu bar. This contains zooming (magnifying glass icons), panning (left/right arrows) and highlighting (yellow highlighter) buttons, reference sequence selector (drop down with sequences from the selected genome sorted by length), a text box to search for features such as gene IDs and overview bar which shows the location of the region in view.
  - The **genome view**: this is where the data tracks are displayed.



- c. Selecting tracks: click on the “select track” button (top left). You can search/filter for tracks and then select them for display by checking the check box next to the track name.

### 3. Navigating to a specific gene in JBrowse.

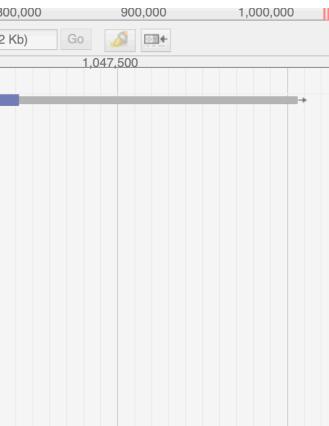
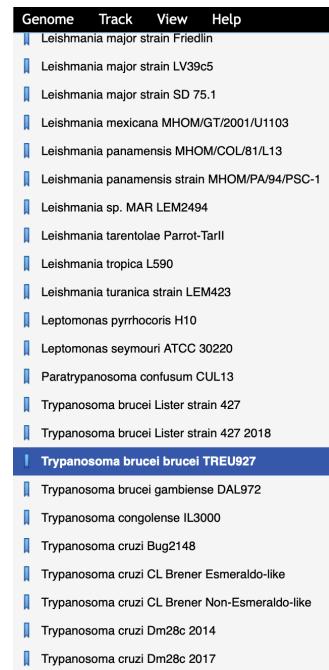
The goal of this step is to navigate to the sedoheptulose-1,7-bisphosphatase (SBPase) gene of *T. brucei* 927.

- Make sure the *Trypanosoma brucei* brucei TREU927 genome is selected from the genome menu.
- Start typing the word sedoheptulose in the search box. After a few seconds you should see the result of the search (do not hit enter). Select the gene from the search dropdown. This will take you to Tb927.2.5800.

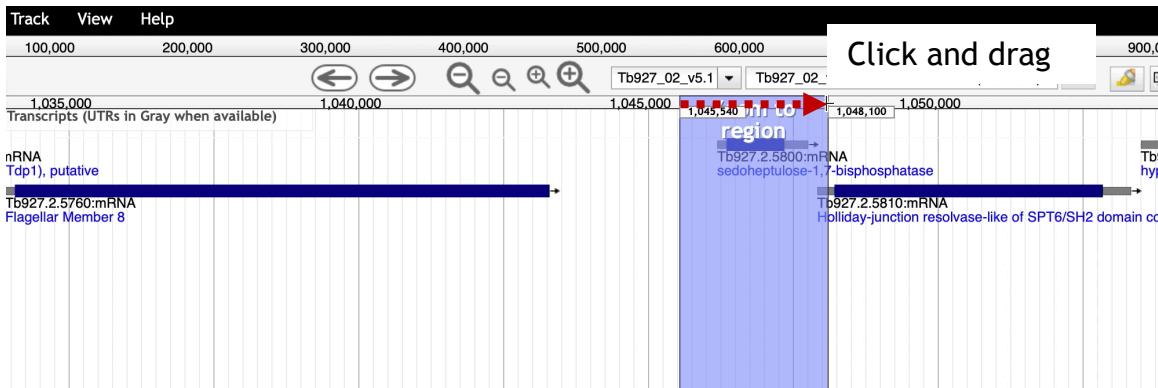
The screenshot shows the JBrowse interface. At the top, a search bar contains the query "sedoheptulose". Below it, a dropdown menu lists "Tb927\_01\_v5.1" and "Tb927.2.5800 - sedoheptulose-1,7-bisphosphatase (SBPase)". A "Go" button is to the right. The main view shows a genomic track with a blue bar representing the gene. A tooltip indicates the gene's ID is Tb927.2.5800. The genomic scale ranges from 400,000 to 800,000. Below the main track, a secondary track shows coordinates 1,046,500 to 1,047,500. A small pop-up window titled "Tb927.2.5800:mRNA details" displays the following information:

Species:	Trypanosoma brucei brucei TREU927
ID:	Tb927.2.5800:mRNA
Gene ID:	Tb927.2.5800
Gene Type:	Protein Coding
Description:	sedoheptulose-1,7-bisphosphatase
5' UTR:	1046195..1046356
CDS:	1046357..1047355
3' UTR:	1047356..1047765
Download:	<a href="#">CDS</a>   <a href="#">protein</a>
OrthoMCL:	<a href="#">OG5_134853</a>
Links:	<a href="#">JBrowse</a>   <a href="#">Gene Page</a>

- You can get information about any feature in the genome view window by clicking on it. Click on the gene feature. What information is available in the popup?
- You can also right click (or control click) on a feature to display the context menu which provides quick links to highlight a feature, go to the feature page (like the gene page) or get the info popup (the same one you get when you click on the feature).
- What genes are immediately upstream and downstream of SBP? (Hint: use the zoom out button in the navigation bar). What is the difference between the small and large zoom buttons? (*Tip 1*: another way to zoom in and out is by clicking on shift and the up or down arrows. What happens if you click shift



and left or right arrows? *Tip2*: you can also zoom in by clicking and dragging your cursor in the location ruler in the navigation bar).



#### 4. Exploring transcription start sites.

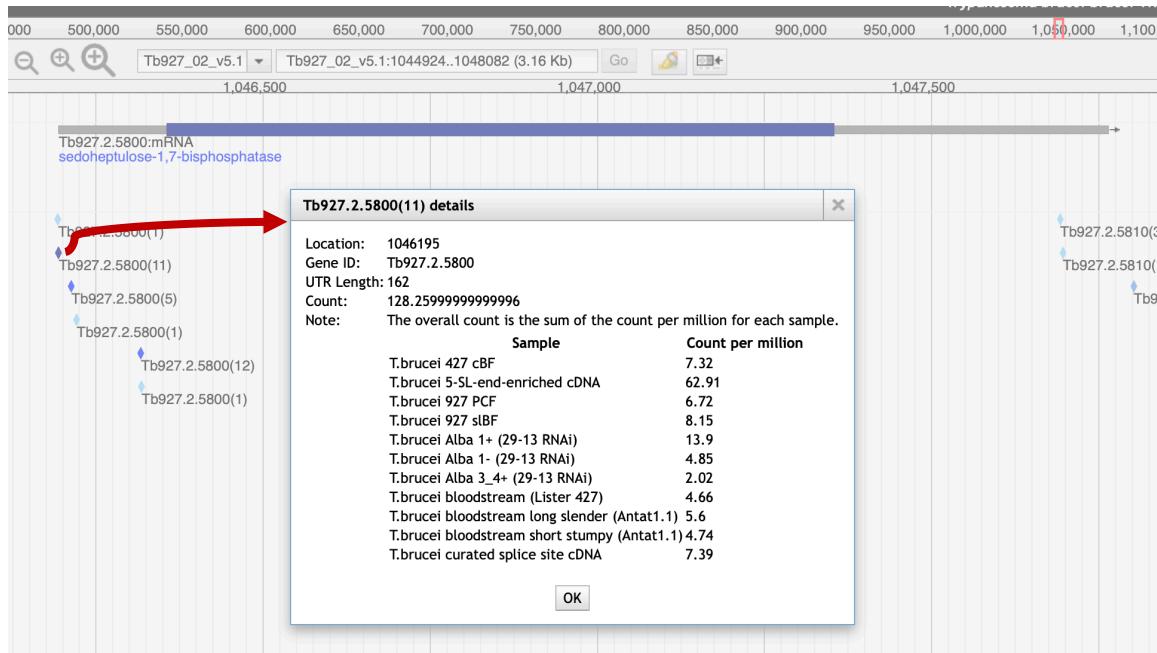
Are you confident about the gene transcription start? (Note: gene features are in blue (left to right) or red (right to left) with untranslated regions (UTRs) in grey).

**Select Tracks**

Back to browser  Contains text  7 matching tracks

Name	Category	Subcategory	Dataset	Track Type	RNASeq Alignment	RNASeq Strand
Bloodstream and Procyclic form spliced leader transcriptomes (927, 427)(2014) - Splice Sites	Gene Models	Splice Sites	...	Segments	...	...
Bloodstream and procyclic forms spliced leader transcriptomes (427, Antat) (2010) - Splice Sites	Gene Models	Splice Sites	...	Segments	...	...
Curated Poly A Sites from bloodstream and procyclic forms	Gene Models	Poly A Sites	...	Segments	...	...
Procyclic form spliced leader transcriptome - Poly A Sites	Gene Models	Poly A Sites	...	Segments	...	...
Procyclic form spliced leader transcriptome - Splice Sites	Gene Models	Splice Sites	...	Segments	...	...
Spliced Leader and Poly A Sites from bloodstream and procyclic forms - Splice Sites	Gene Models	Splice Sites	...	Segments	...	...
Unified Spliced Leader Addition Sites	Gene Models	Splice Sites	...	Segments	...	...

Unified Spliced Leader Addition Sites



What additional data track would be useful for you to assess this? (hint: Click on the “Select Tracks” button to reveal all available tracks. Now type the word “splice” in the “contains text” box. This will filter all tracks that contain the word splice. Find the one called “Unified Splice Leader Addition Sites” and select it. Click on the “Back to browser” button). What do the different diamond colors mean? Click on them and see if you can figure this out from the popups? Which color provides the most evidence for a splice junction?

## 5. Exploring synteny between genomes.

Synteny helps define conservation of homologous genes and gene order between genomes.

- Go to the “Select Tracks” tab on the left of the page and turn on the track called “Syntenic Sequences and Genes”. How did you find this track? One option is to click on the “Comparative Genomics” category on the left side to filter the tracks.

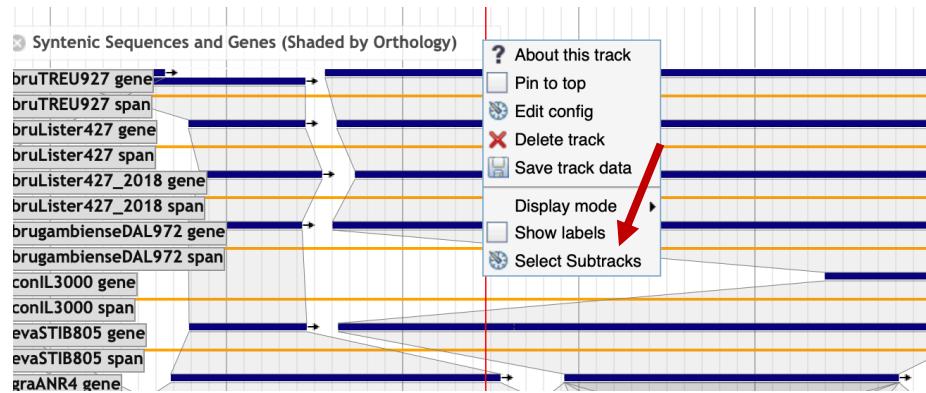
Name	Category	Subcategory	Dataset	Track Type	RNASeq Alignment	RNASeq Strand
Syntenic Sequences and Genes (Shaded by Orthology)	Comparative Genomics	Orthology and Synteny	...	Segments	...	...

- Return to the browser by clicking “Back to Browser” and zoom out so you can see a couple of genes on either side of SBP (does not have to be exact)
- Configure the synteny track to include the following species subtracks:  
*Trypanosoma brucei* 927, *T. brucei* 427, *T. brucei gambiense*, *T. congolense*, *T. evansi*, *T. grayi*, *T. theileri* and *T. vivax*.

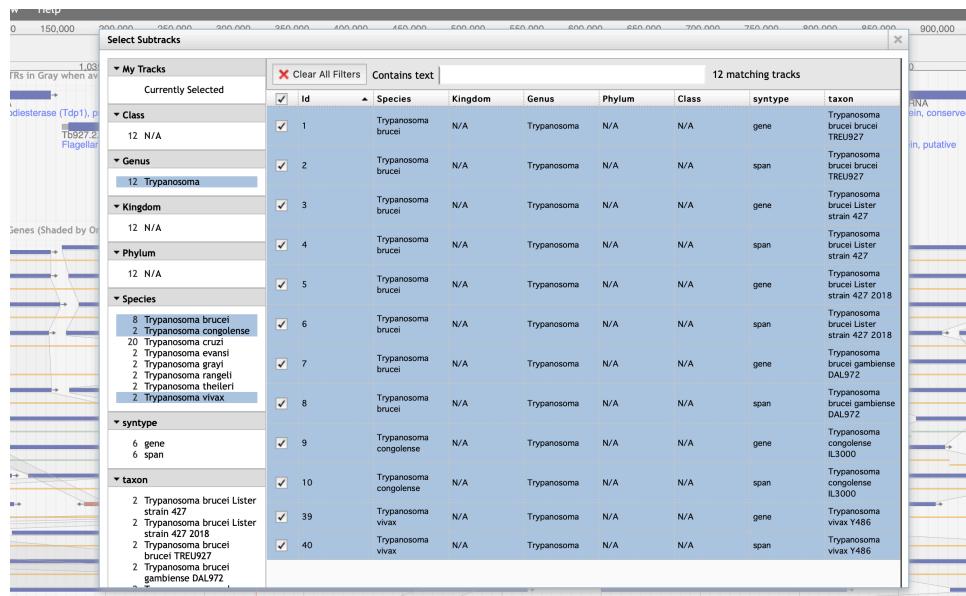
- To configure the subtracks:
  - Click on the down arrow in the track name



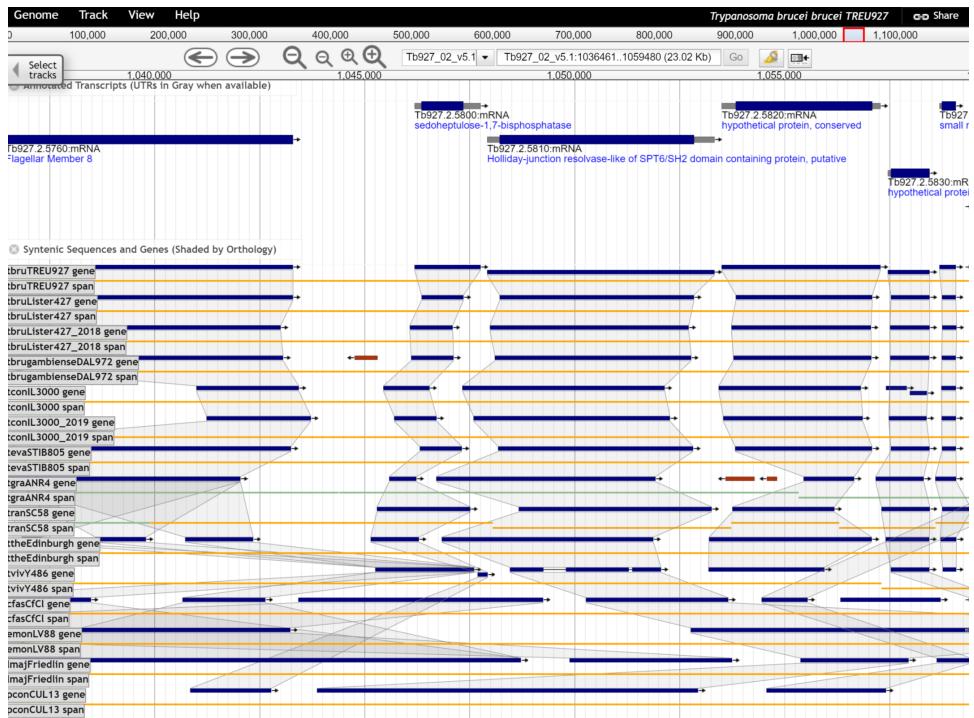
- Select the option called “Select Subtracks” from the menu



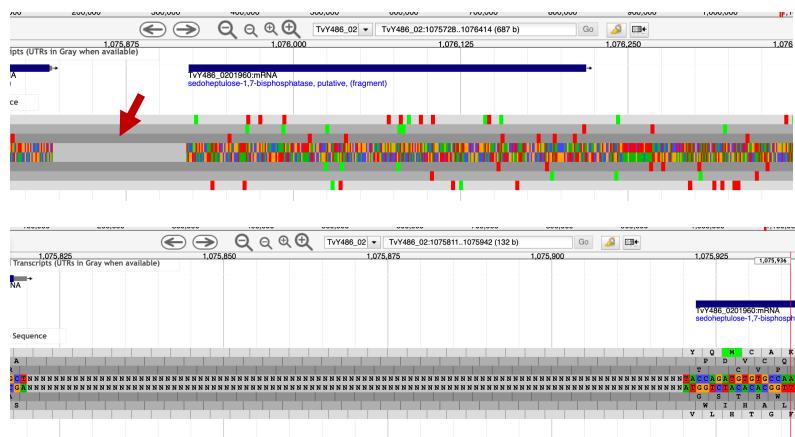
- In the next popup first uncheck all organisms, second use the filters on the left to select Trypanosoma, third select the species of interest (note that you should select both the gene and span subtracks for each species), fourth click on the save button at the bottom of the popup.



- What does the synteny track in this region look like? Feel free to zoom out some more. Are genes (in general) similarly organized between these species? What does the shading between genes mean?
- What direction is the SBPase gene relative to the chromosome?
- What genes are upstream and downstream of the SBPase? Are these genes syntenic?
- What does synteny look like if you add more distantly related species? Does SBPase appear to have orthologs in *Leishmania*? *Endotrypanum*? *Critidilia*?



- Examine the gene corresponding to the *T. vivax* SBPase in the synteny track. Hover over the gene image to find the gene name in the popup. Does this gene appear to be a fragment? What could be some possible reasons for this?
- Do you think all the genomes in the database are fully sequenced? Is it possible that gaps in sequence exist in the available genomes? Let's find out if there is a gap next to the SBPase gene in *T. vivax*:
  - Select *T. vivax* from the list of genomes in the menu bar.
  - Turn on the **annotated transcripts** and the **Reference sequence** tracks.
  - Search for the SBPase gene by typing “sedoheptulose” in the search box then select the gene.
  - Zoom to about 600bps. Do you see something missing on the left side of the gene?
  - Zoom in to this area (click and drag). What do you see? What do all of these Ns mean?



## 6. Exploring other data tracks in Jbrowse.

For this example, we will view *T. brucei* data, so the data tracks you turn on will display data only if the data is aligned to the *T. brucei* genome. Return to the SBPase gene in *T. brucei* by searching for the gene ID in the (Tb927.2.5800) in ‘Landmark or Region’ to redirect the browser. Then zoom to the area between 0.7M and the end of the chromosome.

Turn on the ChIP-seq coverage plots and turn off the syntenic gene and region tracks. The data tracks are from an experiment called: **ChIP-Seq - Four histone Variants ChIP-Seq Coverage aligned to T brucei TREU927 (Cross) (linear plot)**.

For this experiment, chromatin was immunoprecipitated using several different histone antibodies. The DNA that precipitated with the histone was sequenced and aligned to the *T. brucei* TREU927 genome. Peaks in the sequence coverage plots represent areas of histone binding. Different histone variants can be associated with start and termination sites for transcription (

<http://www.ncbi.nlm.nih.gov/pubmed/19369410>)

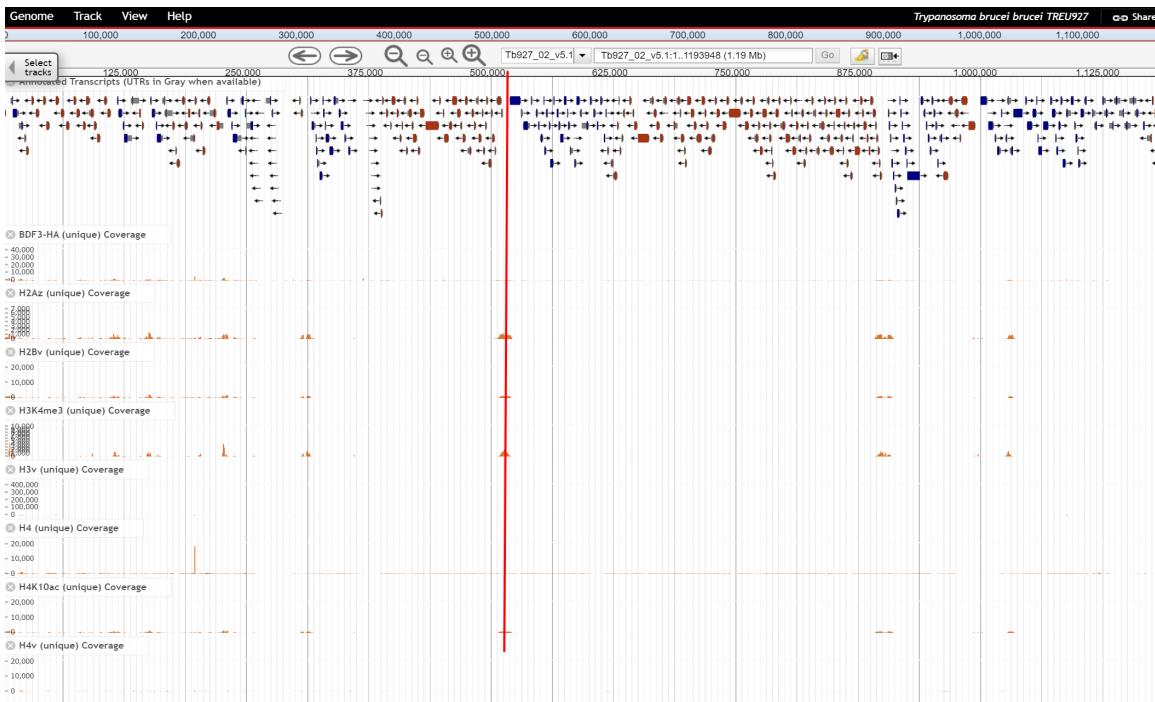
- You may need to adjust the y-axis scaling to bring the tracks into proper view (try

The screenshot shows the 'Select Tracks' interface in JBrowse. On the left, there's a sidebar with categories like 'My Tracks', 'Category', 'Dataset', 'Track Type', 'RNASeq Alignment', and 'RNASeq Strand'. A search bar at the top has a 'Contains text' field with 'Four histone variants' entered. Below the search bar is a table titled 'matching tracks' with columns for Name, Category, Subcategory, Dataset, Track Type, RNASeq Alignment, and RNASeq Strand. The table lists various ChIP-Seq tracks for different histone variants (H3K4me3, H3V, H4, H4K10ac, H2A, H2Bv, H2Az, H3K4me3, H3V, H4, H4K10ac, H4V) across different datasets and categories.

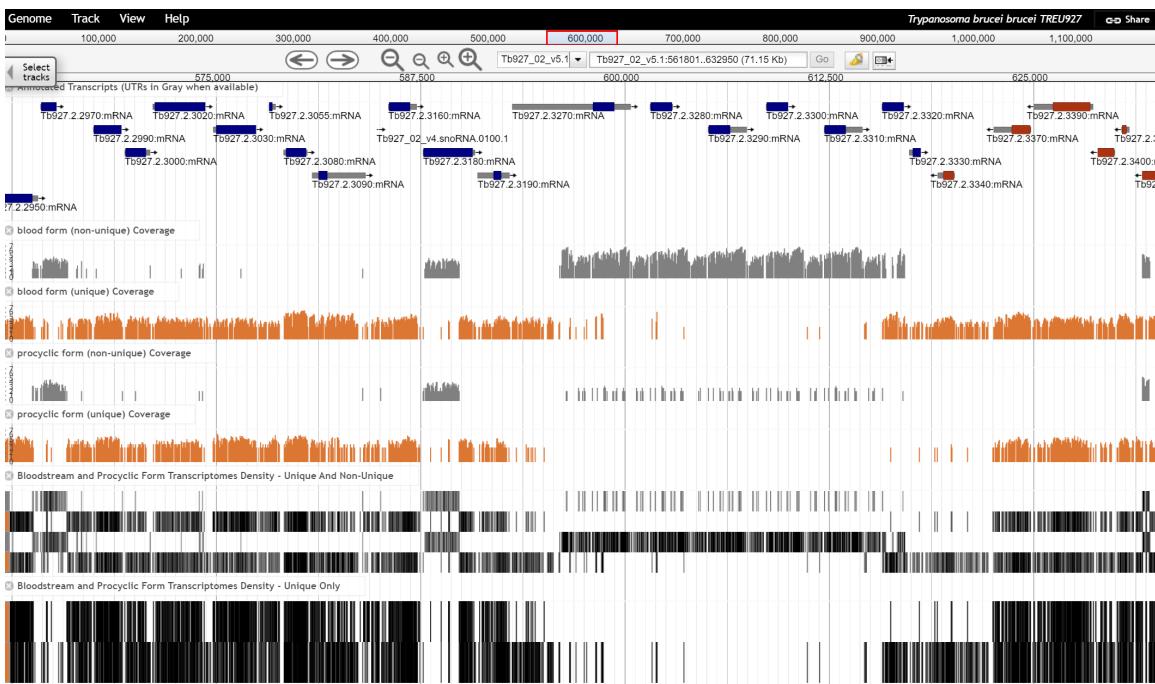
Name	Category	Subcategory	Dataset	Track Type	RNASeq Alignment	RNASeq Strand
BDF3-HA (unique) Coverage	Epigenomics	ChIPSeq	ChIP-Seq - Four histone Variants	Coverage	...	...
ChIP-Seq - Four histone Variants	Epigenomics	ChIPSeq	ChIP-Seq - Four histone Variants	Multi-Density	...	...
Density - Unique And Non-Unique	Epigenomics	ChIPSeq	ChIP-Seq - Four histone Variants	Coverage	...	...
H2Az (unique) Coverage	Epigenomics	ChIPSeq	ChIP-Seq - Four histone Variants	Coverage	...	...
H2Bv (unique) Coverage	Epigenomics	ChIPSeq	ChIP-Seq - Four histone Variants	Coverage	...	...
H3K4me3 (unique) Coverage	Epigenomics	ChIPSeq	ChIP-Seq - Four histone Variants	Coverage	...	...
H3V (unique) Coverage	Epigenomics	ChIPSeq	ChIP-Seq - Four histone Variants	Coverage	...	...
H4 (unique) Coverage	Epigenomics	ChIPSeq	ChIP-Seq - Four histone Variants	Coverage	...	...
H4K10ac (unique) Coverage	Epigenomics	ChIPSeq	ChIP-Seq - Four histone Variants	Coverage	...	...
H4V (unique) Coverage	Epigenomics	ChIPSeq	ChIP-Seq - Four histone Variants	Coverage	...	...

setting the score range to “global” by mousing over the track name, clicking the dropdown arrow and selecting “Change Score Range”).

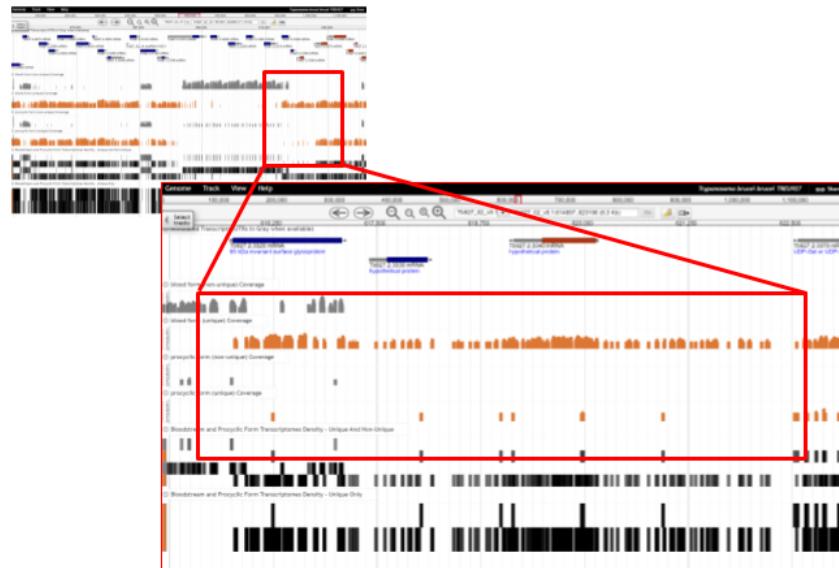
- What does this data show you?
- Roughly how many polycistronic units does this chromosome have? Zoom out to the entire chromosome.



- Do the ChIP-seq peaks correlate with the direction of gene transcription (blue vs. red)?
- Now zoom back to around 50Kb. Turn off the ChIP-Seq tracks and turn on the RNASeq Coverage track called: **Bloodstream and Procyclic Form Transcriptomes mRNASeq Coverage aligned to T brucei TREU927**.



- Move to the **region around 0.6Mbs of the chromosome** (you should be on chromosome 2) and turn on all four subtracks. Take note of the orange and grey bars in the coverage plots. What do you think the grey bars indicate?
- Now zoom out to 100Kb – do you see a difference between the blood and procyclic forms?

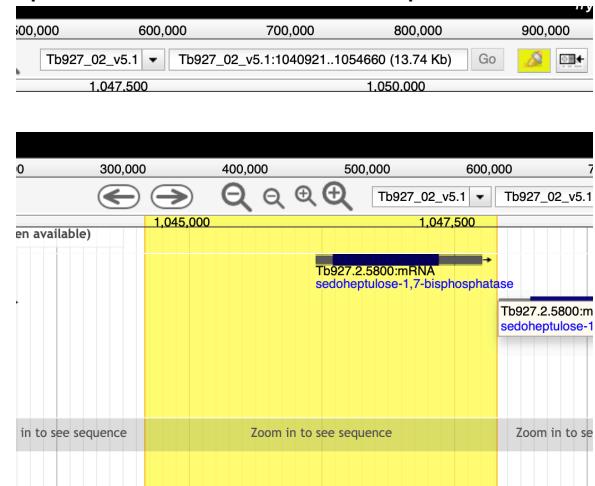


- Zoom in to a gene that looks like it is differentially expressed. What are your conclusions? Are the reads supported by unique or non-unique reads?
- Can you turn on additional tracks that may give some more support to your conclusions?  
Hint: turn on the EST and *T. brucei* protein expression evidence tracks.
  - Is there any proteomics evidence for this region?
  - How about EST evidence? Click on an EST graphic (glyph) to get additional information.
- Turn off the RNA-seq graphs and make sure the *T. brucei* protein expression evidence tracks are on. **Zoom out to 500Kb**. Explore the evidence for gene expression based on mapped peptides from proteomics experiments – which gene in this view has the highest number of peptide hits? Try looking at the “All MS/MS peptides (feature density)” track for an overview.

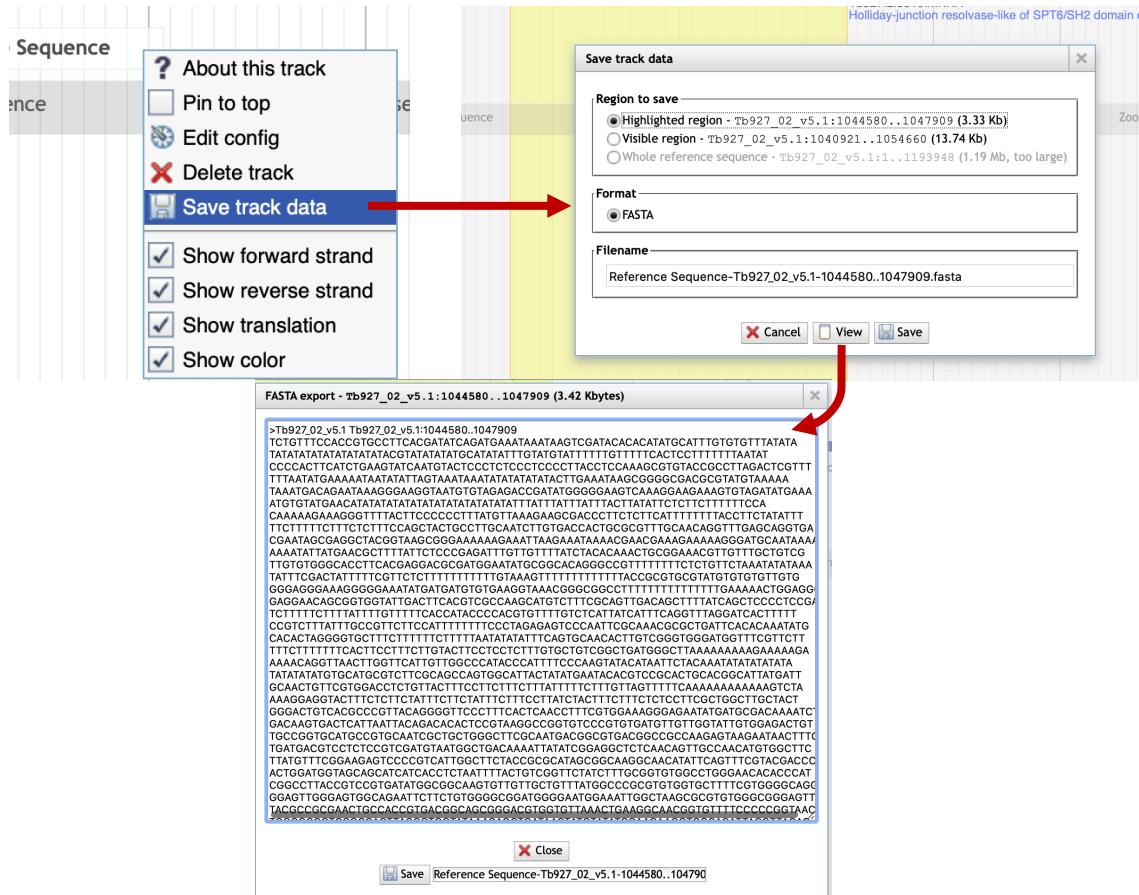


## 7. Retrieving data from and uploading your own tracks to JBrowse

- Downloading sequence in FASTA format from a region of interest:
  - Make sure the “annotated transcripts” and the “reference sequence” tracks are turned on.
  - Click on the “highlight a region” button in the navigation bar. It should turn yellow when activated.
  - Click and drag in the genome view region and select the area you would like to highlight.
  - Click on the down arrow on the reference sequence track and select “Save track data”.



- v. In the next popup window you can keep everything as the default and either save or view the sequence.



b. Uploading data to JBrowse:

JBrowse can accept several standard-format data files by direct upload or through a URL if the data is stored remotely. Some file formats like BAM and VCF require indexing before uploading. In this exercise we will download a bigwig file from GEO and then upload it to JBrowse:

- i. Go to this GEO sample record:

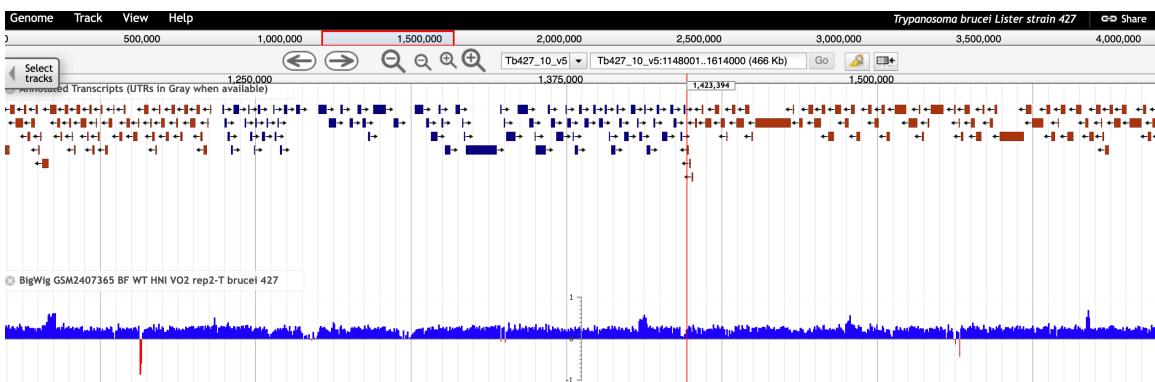
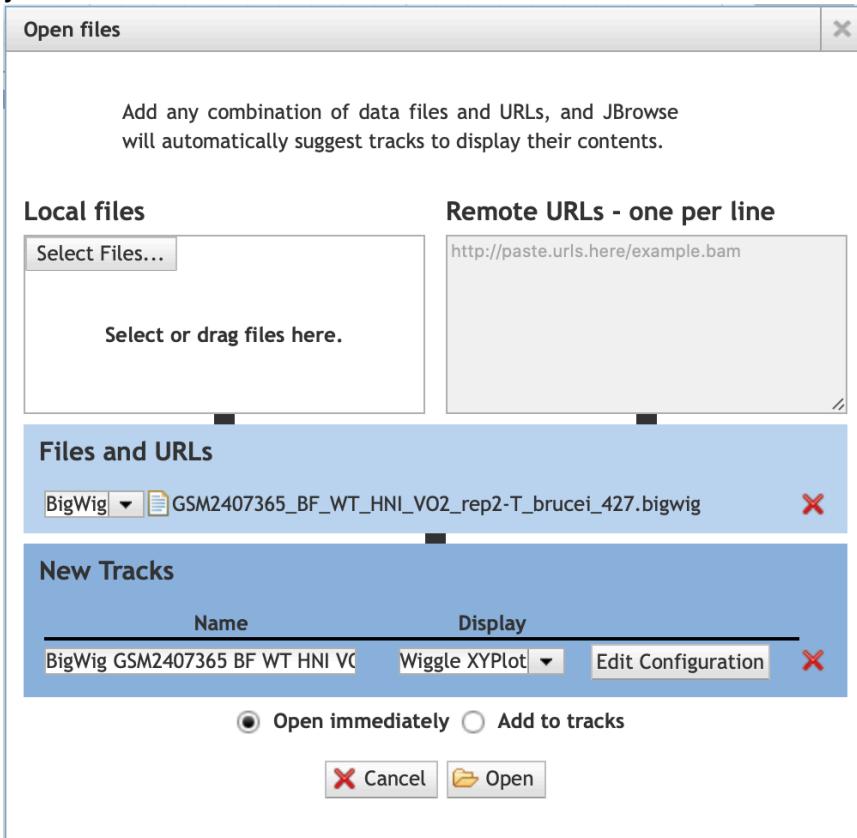
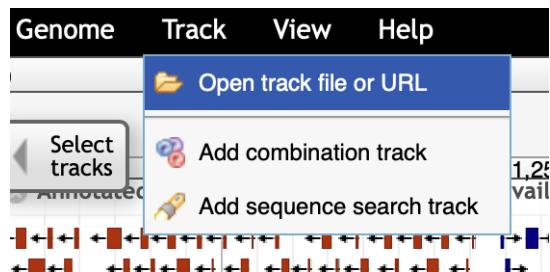
<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSM2407365>

- ii. Scroll down to the bottom of the page and download the bigwig file with the http link.

Supplementary file	Size	Download	File type/resource
GSM2407365_BF_WT_HNI_VO2_rep2-T_brucei_427.bigwig	12.4 Mb	(ftp)(http)	BIGWIG

- iii. Once the file is downloaded go to JBrowse and select *Trypanosoma brucei brucei* Lister 427 as the reference genome (hint: use the Genome link in the menu panel, top left).
- iv. Turn on the track for annotated transcripts if it is not on already.

- v. Click on the Tracks menu item and select “Open track file or URL”.
- vi. In the popup click on select file then select the file you just downloaded. JBrowse should automatically recognize that the file is in bigwig format.
- vii. Click on the Open button. The bigwig output should appear very quickly in your browser.



## Interpreting RNA-seq data (Browser Exercise II)

In previous exercises, you spent some time learning about gene pages and examining genes in the context of the JBrowse genome browser. It is important to recognize that gene models (structural annotation) are often open to interpretation, however, especially with respect to:

- transcript initiation and termination sites (5' and 3' untranslated regions, or UTRs)
- alternative processing events ... if you sequence deep enough, virtually *all* genes (in organisms that process transcripts) display alternative splicing, even for single exon genes
- the potential significance of non-coding RNAs

Even heavily curated genomes (*Plasmodium falciparum*, *Trypanosoma brucei*, *Saccharomyces cerevisiae*) do not fully reflect all available knowledge about stage-specific splicing, as new information is emerging all the time! In addition, many gene models were computationally derived using methods that may have not relied on experimental evidence supporting intron/exon boundaries (e.g. RNAseq data).

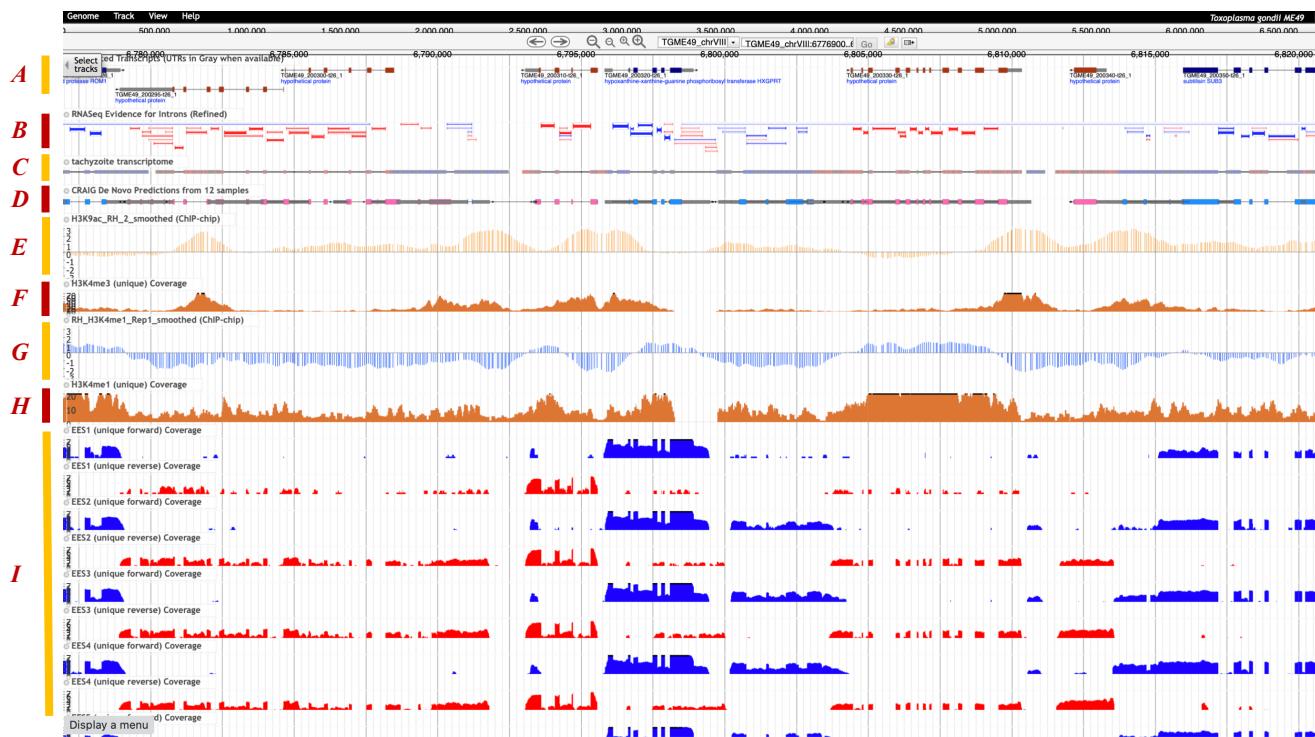
*In this exercise, we will explore genome browser track configuration options in greater detail, focusing on the interpretation of RNA-seq datasets, and using this information to examine the differentially-spliced HXGPRT gene of *T. gondii*. You will then apply your newfound skills to examine other genes that may be alternatively spliced ... and report your findings back to the group as a whole.*

The screen shot below (Fig. 1) shows a sample of data tracks that can be turned on and configured in JBrowse. There are a number of tracks that are worth examining which help in determining the accuracy of annotated gene models and that help in defining possible alternative splice variants of a gene. The link below will display the JBrowse view from figure 1, except for any special configurations with are not stored in the URL. For example, tracks 1c and 1d are collapsed in figure one but will appear expanded in the JBrowse view after clicking on the link:

<https://tinyurl.com/y5lhcvcf>

- What evidence do each of the tracks provide?
- Are the ChiP-ChIP and Chip-seq tracks similar in what they show?
- How many alternative splice variants of HXGPRT would you be willing to annotate based on the evidence?

- Are there other data tracks that might be useful to examine?



*Figure 1: Screen shot from ToxoDB JBrowse. A. Official gene models. B. Splice junction evidence based on available RNAseq data. C. Nanopore long-read transcriptomic data (collapse view). D. Alternative gene models using RNAseq evidence from 12 experiments (collapsed view). E. Chip-ChIP H3K9ac. F. Chip-Seq H3K4me3. G. Chip-ChIP H3K4me1. H. Chip-Seq H3K4me1. I. RNAseq coverage from *Toxoplasma gondii* strain CZ clone H3 in feline enteroeithelial stage (strand specific).*

**Working in groups of four, please select at least two genes from this list to evaluate, based on RNA-seq data and any other available evidence. See if you can discover which exon(s) were represented ... and determine whether these genes are actually alternatively spliced (constitutively or stage-specifically). We will then reconvene to hear a brief report from each group.**

TgME49_200320 (HXGPRT)	TGME49_211420	TGME49_281440
TGME49_246490	TGME49_214440	TGME49_279390
TGME49_256650	TGME49_250115	TGME49_202770
TGME49_283540	TGME49_261720	TGME49_217490
TGME49_226410	TGME49_268610	TGME49_292150
TGME49_225730	TGME49_270520	TGME49_276170
TGME49_213610	TGME49_280380	TGME49_266640
TGME49_213660	TGME49_293720	TGME49_266920
TGME49_297160	TGME49_248445	TGME49_299010
TGME49_211250	TGME49_230180	

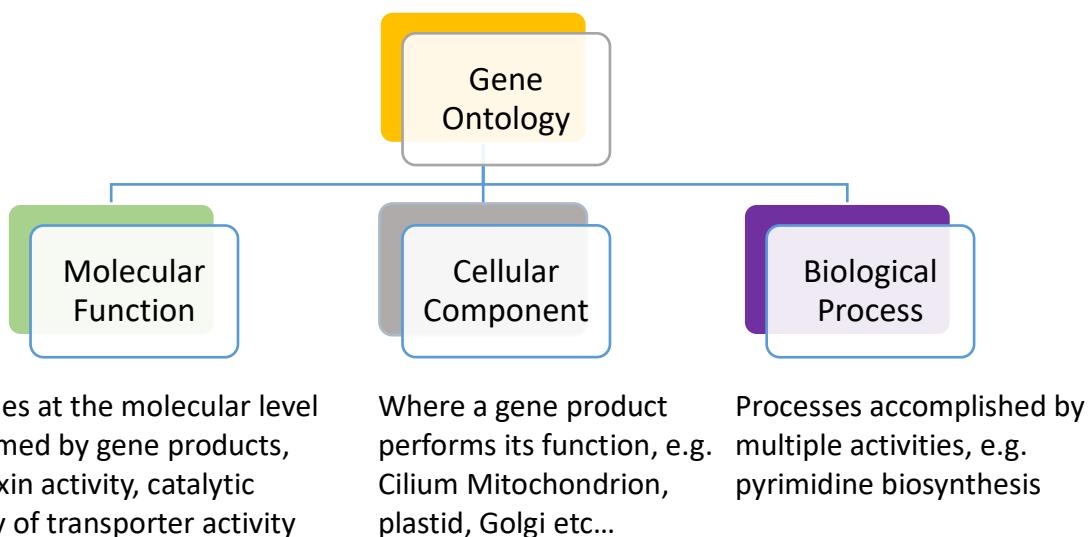
# Gene Ontology (GO) Enrichment

## Learning objectives:

- Run a GO enrichment analysis
- Explore GO enrichment results

## Background:

The gene ontology describes the knowledge of biological sciences and divides this knowledge into three broad categories: Molecular function, cellular component and biological process.



To learn more about Gene Ontology, please visit:  
<http://geneontology.org/docs/ontology-documentation/>

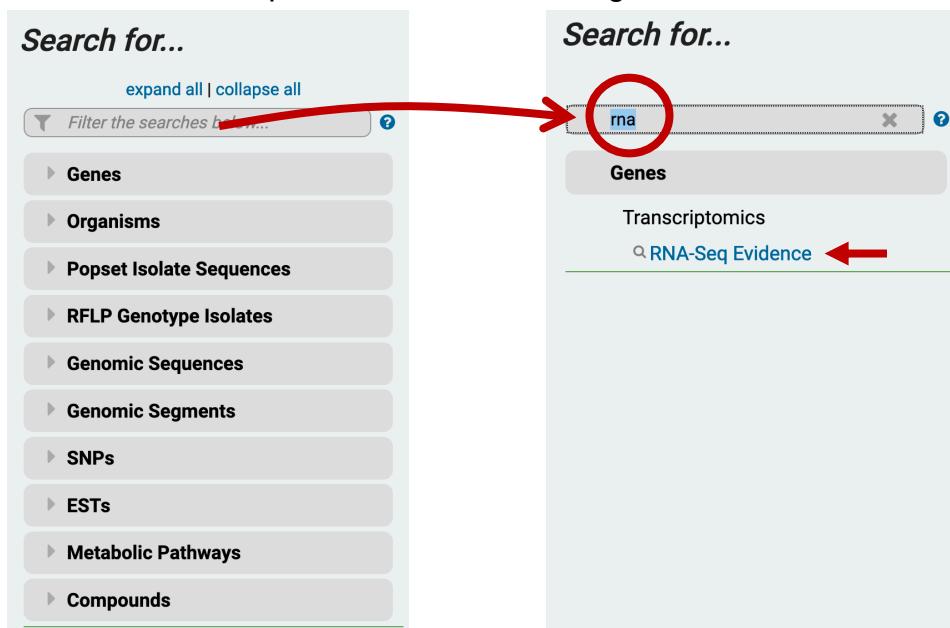
A gene can be assigned a GO term either manually (by an annotator evaluating experimental evidence) or computationally (based on the GO terms of genes that share sequence or functional domains). These GO terms can be used to test whether your set of genes are enriched for a molecular function, cellular component, or biological process.

**For example:** Does my list of genes have an over-representation of specific GO terms compared to the rest of the genome?

A standard enrichment method employs Fisher's exact test, a statistical test that evaluates a 2x2 contingency table (in this case, number of genes in my set *versus* number of genes from genome not in my set, and number of genes with GO term Z

*versus* number of genes without term Z). This test produces a p-value between 0 and 1, where  $p \leq 0.05$  is considered significant (that is, less than 5% probability that the enrichment is due to chance). However, the test is performed for each of the 100s of GO terms, increasing the chances that a GO term will be incorrectly considered enriched (a false positive, or type I, error). Thus, the original p-value must be adjusted for so-called multiple hypothesis testing, resulting in an adjusted p-value such as the Benjamini-Hochberg false discovery rate (FDR) or Bonferroni adjusted p-value.

1. In order to run a GO enrichment analysis, you need a list of genes to test. This can be a list of gene IDs from your experimental results that you can upload using the ID search or a gene list resulting from a search you conducted on a VEuPathDB website. For this example, in ToxoDB, we will identify genes that are differentially regulated over time.
  - a. Navigate to the RNA-Seq searches and find the data set called “**Oocyst Time Series (M4)**” from Fritz *et al.* A quick way of getting to the RNA-Seq searches is to type ‘rna’ in the filter box on the left of the home page and click on the RNA-Seq Evidence link. See image below.



- b. The RNA-Seq evidence page includes a list of all data sets that are loaded in the website. To quickly find a dataset, you can start typing key words in the “Filter Data Sets” box. For example, start typing the word “oocyst”.

### Identify Genes based on RNA-Seq Evidence

- c. Once you find the data set of interest, click on the fold-change (FC) option. This will open a search page that contains the parameters that you can manipulate to search this data set. For this exercise, identify genes that are upregulated by 20-fold in the day 4 and day 10 time points compared to the day 0 time point. Parameters to set:
1. Up-regulated
  2. 20-fold
  3. Maximum
  4. Day 0
  5. Minimum
  6. Day 4 and 10

#### Identify Genes based on T. gondii ME49 Oocyst Time Series (M4) RNA-Seq (fold change)

**For the Experiment**

Oocyst Time Series (M4) - Sense

return protein coding  genes

that are up-regulated

with a Fold change >= 20

between each gene's maximum  expression value

(or a Floor of 10 reads )

in the following Reference Samples

day 0  
 day 4  
 day 10

[select all](#) | [clear all](#)

and its minimum  expression value

(or the Floor selected above)

in the following Comparison Samples

day 0  
 day 4  
 day 10

[select all](#) | [clear all](#)

**Example showing one gene that would meet search criteria**

(Dots represent this gene's expression values for selected samples)

Up-regulated

Expression

Reference Comparison Samples

Minimum Expression Value Comparison

Expression Value Reference

20 fold

For each gene, the search calculates:

$$\text{fold change} = \frac{\text{minimum expression value in comparison}}{\text{reference expression value}}$$

and returns genes when fold change  $\geq 20$ .

You are searching for genes that are up-regulated between one reference sample and at least two comparison samples.

This calculation creates the narrowest window of expression values in which to look for genes that meet your fold change cutoff. To broaden the window, use the average or maximum comparison value.

[Get Answer](#)

d. Once you have set the parameters, click the “Get Answer” button at the bottom of the search. This will return a one-step search strategy. How many genes did you get?

2. To run a GO enrichment analysis on these results, do the following:

142

- a. Click on the Analyze Results tab just above the list of genes (arrow in image below).

## My Search Strategies

Opened (1) All (1) Public (17) Help

Unnamed Search Strategy \*

Step 1

**1,029 Genes (970 ortholog groups)** [Revise this search](#)

Analyze Results

Organism Filter  
select all | clear all | expand all | collapse all  
 Hide zero counts  
Search organisms...   
 Eimeriidae 0

Gene Results Genome View Analyze Results

1 2 3 ... 52 Rows per page: 20

- b. Click on the “Analyze Results” tab to reveal the different analyses that you can run on your results. Besides GO enrichment, what other analyses are available?

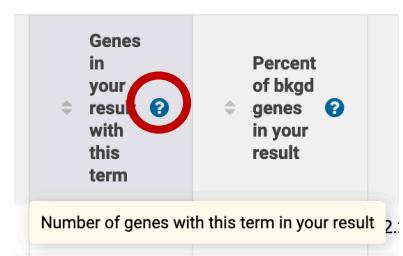
Analyze your Gene results with a tool below.

Gene Results Genome View New Analysis

Gene Ontology Enrichment Metabolic Pathway Enrichment Word Enrichment

Hide Organism Filter

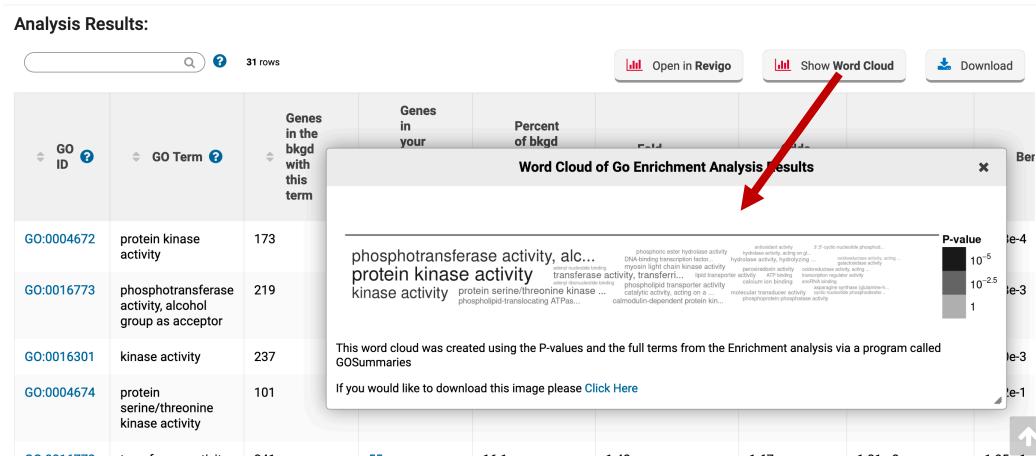
- c. Click on the GO enrichment option. This will reveal the parameters that you can modify. For the purpose of this exercise, keep all the defaults and click on “Submit”.
- d. What is the top enriched GO term from this analysis?
- e. What do each of the columns in the analysis table represent? (Hint: move your mouse over the question mark next to each column header to get more information.)



- f. Try rerunning the GO enrichment analysis, but this time select the Molecular Function ontology. What is the top enriched GO term?

The screenshot shows the 'Gene Results' tab selected at the top. Below it, the 'Gene Ontology Enrichment' tab is active. A red arrow points to the 'Molecular Function' radio button under the 'Ontology' section. Other settings include 'Organism: Toxoplasma gondii ME49', 'Evidence: Computed, Curated', 'P-Value cutoff: 0.05', and 'Limit to GO Slim terms: No'.

- g. Click on the “Word Cloud” button above the analysis results. What does this do? (See image below).



**Additional resources:**

Gene Ontology:

<http://geneontology.org/docs/ontology-documentation/>

Enzyme Commission numbers:

<https://www.qmul.ac.uk/sbcs/iubmb/enzyme/>

More info on Fischer's exact test:

<http://www.biostathandbook.com/fishers.html>

Fisher's Exact Test and the Hypergeometric Distribution (the M&M example):

<https://youtu.be/udyAvvaMjfM>

Some more info about Odds ratios:

<http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2938757/>

False discovery rates and P value correction:

<http://brainder.org/2011/09/05/fdr-corrected-fdr-adjusted-p-values/>

GO Slim:

<http://www-legacy.geneontology.org/GO.slims.shtml>

REVIGO:

<https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0021800>

## Regular Expressions & Genomic Colocation

**Note:** this exercise uses different VEuPathDB resources as example database, but the same functionality is available on all VEuPathDB resources.

### Learning objectives:

- Run a regular expression search on amino acid sequences
- Run a regular expression search on nucleotide sequences
- Use the genomic colocation search

Protein or nucleotide sequences can be identified using the regular expression searches in VEuPathDB. This search is very useful to identify patterns of sequences.

Searches can be accessed from categorized menus in the left search for panel (A) or from the searches menu in the header (B).

The screenshot shows the VEuPathDB website interface. At the top, there's a search bar with placeholder text "E.g., \* or NCU06658 or synth\* or oxo group\*". Below the search bar is a navigation bar with links: My Strategies, Searches (highlighted with a red box), Tools, My Workspace, Data, About, Help, Contact Us, and a Guest link. To the right of the navigation bar is a "News and Tweets" sidebar. The main content area has a heading "Overview of Resources" and a "Getting Started" section. On the left, there's a sidebar with a "Search for..." dropdown (labeled A) containing categories like Genes, Organisms, Popset Isolate Sequences, etc. To the right of the dropdown is a "Filters" section with "expand all | collapse all" and a "Filter the searches below..." button. Below the filters is another list of categories (labeled B) including Genes, Organisms, Popset Isolate Sequences, etc. At the bottom of the sidebar, there's a "Read More" link.

#### Accessing the protein motif pattern search:

- Click on the *Genes* category then click on the sequence analysis category

This screenshot shows the "Genes" category expanded in the sidebar. Under "Sequence analysis", there are two options: "BLAST" and "Protein Motif Pattern". A red arrow points to the "Protein Motif Pattern" option.

#### Accessing the DNA motif pattern search:

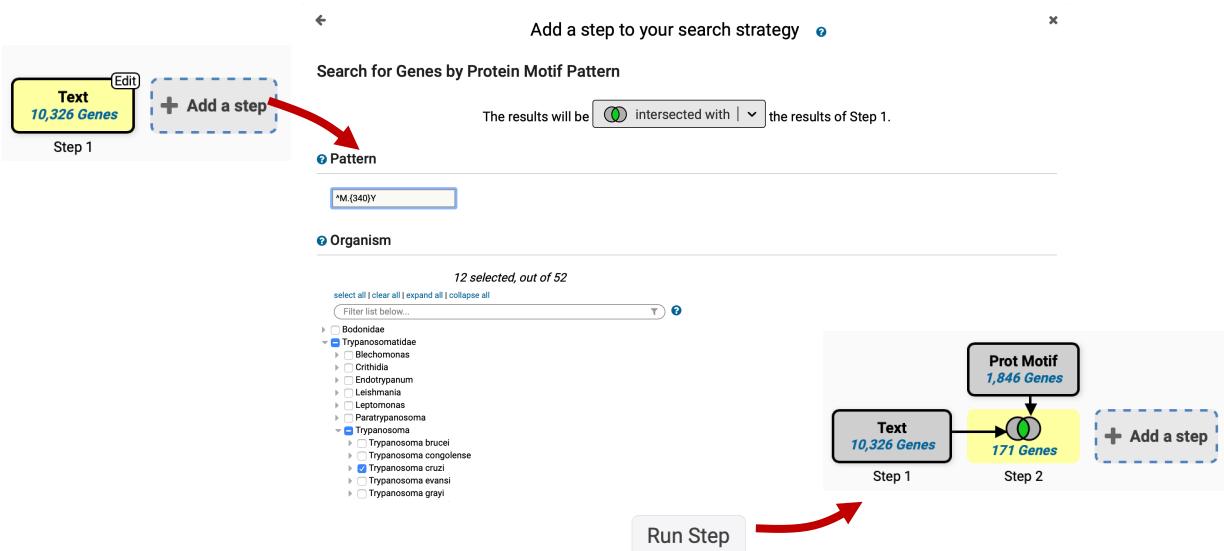
- Click on the *Genomic segments* category

This screenshot shows the "Genomic segments" category expanded in the sidebar. Under "DNA Motif Pattern", there are two options: "DNA Motif Pattern" and "Genomic Location". A red arrow points to the "DNA Motif Pattern" option.

**Note:** the appendix at the end of this document includes additional regular expression help.

## 1. Using regular expressions to find motifs in TriTrypDB: finding active trans-sialidases in *T. cruzi* (TriTrypDB).

- T. cruzi* has an expanded family of trans-sialidases. In fact, if you run a text search for any gene with the word “trans-sialidase” in its *product description*, you return over 10000 genes among the strains in the database!!! Try this and see what you get.
- Not all of the genes returned in (a) are predicted to be active. It is known that active trans-sialidases have a signature tyrosine (Y) at position 342 in their amino acid sequence. Add a motif search step to the text search in ‘a’ to identify only the active trans-sialidases.
- Write a regular expression that defines a protein sequence that starts with a methionine, and is followed by 340 of any amino acids, followed by a tyrosine ‘Y’.



## 2. Find Cryptosporidium genes with the YXXΦ receptor signal motif (CryptoDB)

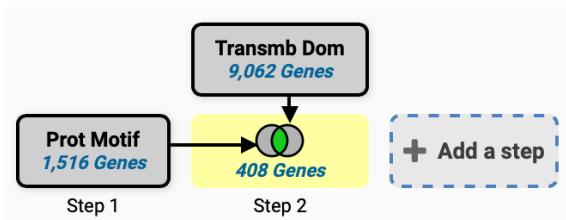
The YXXΦ (Y=tyrosine, X=any amino acid, Φ=bulky hydrophobic [phenylalanine, tyrosine, threonine]) motif is conserved in many eukaryotic membrane proteins that are recognized by adaptor proteins for sorting in the endosomal/lysosomal pathway. This motif is typically located in the c-terminal end of the protein. \*\*\*Note: do not look for the Φ symbol on your keyboard – this will not work. Rather you should use the amino acid symbols.

- a. Use the “protein motif pattern” search to find all *Cryptosporidium* proteins that contain this motif anywhere in the terminal 10 amino acids of proteins. (hint: for your regular expression, remember that you want the first amino acid to be a tyrosine, followed by any two amino acids, followed by any bulky hydrophobic amino acid (phenylalanine, tyrosine, threonine)).
- b. How many of these proteins also contain at least one transmembrane domain.

Identify Genes based on Protein Motif Pattern

Pattern: [ ]

Organism: 11 selected, out of 14  
 select all | clear all | expand all | collapse all  
 Filter list below...   
 Apicomplexa  
 Coccidia  
 Euoccidiorida  
 Cryptosporidiidae  
 Cryptosporidium  
 Gregarinina  
 Chromerida  
 select all | clear all | expand all | collapse all



- c. What would happen if you revise the first step (the motif pattern step) to include genes with the sorting motif in the C-terminal 20 amino acids? (hint: edit the first step and modify your regular expression).

Note: if you need help with the regular expression the answers are in appendix B.

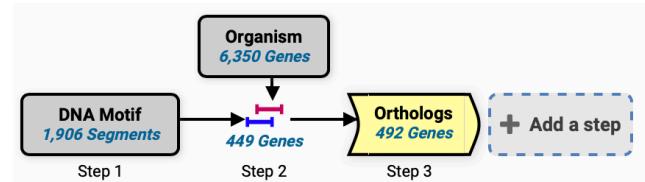
### 3. Find fungal genes downstream of a regulatory DNA motif (FungiDB).

Transcriptional start sites are often located within a certain distance upstream of the genes or gene clusters that they regulate. In fungi, DNA motifs are also important for regulation of processes linked to host cell invasion or production of secondary metabolites. Readily available genomic data facilitate the discovery of regulatory motifs via examination of orthologous sequences.

The goal of this exercise is to identify all genes harboring upstream CACGTG motif, known for its role in transcriptional regulation. We will start our search in an extensively studied model organism *Saccharomyces cerevisiae* and expand our search to *Fusarium graminearum*.

Here is a summary of the search strategy:

- a. Find the CACGTG DNA motif in the *Saccharomyces cerevisiae* genome.
- Select the “Search for genomic segments (DNA motif)” menu from the Search menu and look for CACGTG in *S. cerevisiae*.



2. Your search returns over 1900 DNA segments containing GACGTG motif. Next, let's look for putative regulatory targets of this motif by searching for genes that are located 600bp downstream of this sequence.

**Identify Genomic Segments based on DNA Motif Pattern**

**Search for...**

expand all | collapse all

Filter the searches below... [?](#)

- ▶ Genes
- ▶ Organisms
- ▶ Popset Isolate Sequences
- ▶ Genomic Sequences
- ▶ **Genomic Segments**
- 🔍 DNA Motif Pattern
- 🔍 Genomic Location
- ▶ SNPs
- ▶ ESTs
- ▶ Metabolic Pathways
- ▶ Compounds

**Organism**

1 selected, out of 164

add these | clear these | select only these  
select all | clear all

cer

Fungi
 

- Ascromycota
  - Eurotiomycetes
    - Orygenesales
    - Orygenaceae
      - Byssomyces
      - Byssomyces ceratinophila
      - Byssomyces ceratinophila isolate UAMH 5669
  - Saccharomycetes
    - Saccharomycetales
      - Saccharomycetaceae
        - Saccharomyces
          - Saccharomyces cerevisiae**
          - Saccharomyces cerevisiae S288c**

**Pattern**

Type sequence pattern **CACGTG**

[Get Answer](#)

- b. Identify genes with the CACGTG motif located 600bp upstream of an open reading frame.

EuPathDB offers a colocation function to identify genomic features within a specified distance of each other. Run a search for all genes in *Saccharomyces cerevisiae* and use the colocation tool to identify genes that contain the CACGTG motif in their upstream regions. Follow these steps:

1. Click “Add Step”. Choose the option on the left called “Use **Genomic Colocation** to combine with other features” then select the *organism* gene search which can be found under the *Taxonomy* category.

Add a step to your search strategy [?](#)

Combine with other Genomic Segments

Use Genomic Colocation to combine with other features

Choose which features to colocate. From...

A new search    An existing strategy    My basket

tax

Genes  
Taxonomy  
Organism

2. On the next page select *Saccharomyces cerevisiae* from the taxonomy browser and click on continue.

The screenshot shows a search interface for organisms. A search bar at the top contains the text 'cere'. Below the search bar, a tree view shows the taxonomic hierarchy: Fungi > Ascomycota > Saccharomycetidae > Saccharomyces > Saccharomyces cerevisiae S288c. The 'Saccharomyces cerevisiae S288c' node is selected, indicated by a blue checkmark. At the bottom of the search interface, there are buttons for 'Continue...' and 'Run Step'.

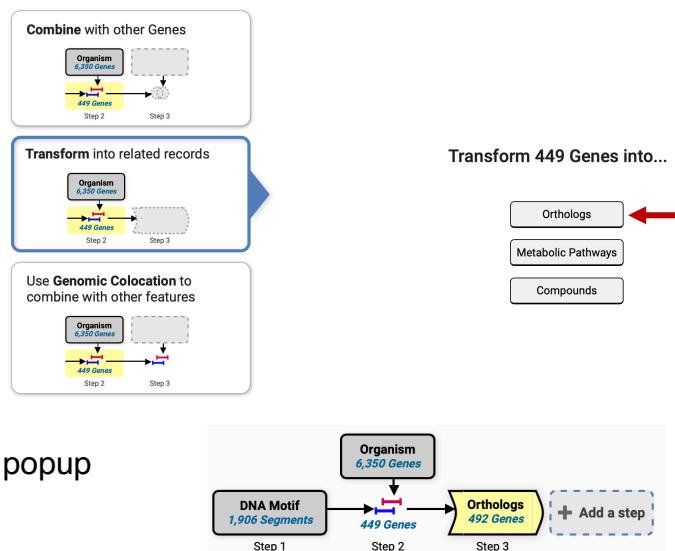
3. Configure the parameters on the next page to return each gene from step 2 whose upstream region (600bp) overlaps the exact region of a Genomic Segment in Step1 (CACGTG) and is on either strand.

The screenshot shows a configuration interface for a search step. It includes two main panels: one for selecting genes based on their upstream region and another for selecting genes based on their genomic segment. The left panel has a dropdown menu 'Return each Gene from the new step' with the condition 'whose upstream region overlaps the exact region of a Genomic Segment from the current step and is on either strand'. The right panel has a similar dropdown menu. Both panels have sections for 'Exact', 'Upstream: 600 bp', and 'Downstream: 1000 bp'. The 'Upstream: 600 bp' section is highlighted with a red box. At the bottom, there is a 'Run Step' button.

### c. Identify orthologs *S. cerevisiae* genes in *Fusarium graminearum*.

All VEuPathDB sites offer tools to transform results between record types. The "Transform by Orthology" tool uses orthology clusters assigned by the OrthoMCL algorithm to enable transformation of a list of genes from one or more species to another (or more) species.

- click on add step then select the **Transform into related records** option from the left side of the popup. Next click on the *Orthologs* option.
- Select *F. graminearum* in the next popup window and click on *Run Step*.



## Appendix A

### **Regular expression help**

The following codes can be used to represent classes of amino acids.

AA property	Amino acids	Code
Acidic	DE	0
Alcohol	ST	1
Aliphatic	ILV	2
Aromatic	FHWY	3
Basic	KRH	4
Charged	DEHKR	5
Hydrophobic	AVILMFYW	6
Hydrophilic	KRHDENQ	7
Polar	CDEHKKNQRST	8
Small	ACDGNPSTV	9
Tiny	AGS	B
Turnlike	ACDEGHKNQRST	Z
Any	ACDEFGHIKLM NPQRSTVWY	X

**The following is a simple explanation of regular expressions.**

Perl regular expressions are terms used for pattern matching in text strings, e.g. '*aadgt*', '*'aa+dgt'*', '*'aldc'*', '*[mac]a*'.

Because nucleotide and amino acid sequences are text strings, regular expressions are very useful for finding motifs within sequences.

Motifs often include repetitive or ambiguous assignments at some locations. The rules and special characters used in regular expressions help define the full set of strings that match the motif pattern.

The following is a description of some of these characters and examples of how they are used.

Although regular expressions seem complicated at first, they are very useful and easy to understand after going through some examples.

### **Special Characters**

- Match any character.

- +** Matches "one or more of the preceding characters".
- \*** Matches "any number of occurrences of the preceding character", including 0.
- ?** Matches "zero or one occurrences of the preceding character".
- [ ]** Matches any character contained in the brackets.
- [^ ]** Match any character *except* those in the brackets.
- {n}** Matches when the preceding character, or character range, occurs exactly n times.
- {n,}** Matches when the preceding character occurs at least n times.
- Matches when the preceding character occurs at least n times, but no more than m  
**{n,m}** times.

### Here are some examples of searches.

**ad+f** (1 or more occurrences of 'd') would match any of the following:

adf  
addf  
adddf  
addddf  
.....

**ad\*f** (0 or more occurrences of 'd') would match:

af  
adf  
addf  
adddf  
.....

**ad?f** (0 or 1 occurrence of 'd') would match:

af  
adf

**a[yst]c** would match:

atc  
asc  
ayc

### Specify the number of occurrences of a residue.

**P{1,5}** would match P from 1 to 5 times.

**.{1,30}** would match any amino acid 1 to 30 times so you could find a motif within 30 amino acids of something like the beginning.

## Pattern Anchors

- ^ Match only at the beginning of the string.
- \$ Match only at the end of the string.

Here are examples of expressions using pattern anchors.

***^mdef*** (e.g. a protein sequence **starting with** 'mdef') would match:

- mdef
- mdefab
- mdefaredfadfk

**but not match:**

- edefa
- emdefa
- eeeemdef

***kdel\$*** (searches for proteins **ending with** 'kdel', a standard ER retention signal) would match:

- eeeekdel
- kdel

**but not match :**

- edefkdell
- akdeleef

## Appendix B

Answers to exercise 2:

A: YXXΦ in the terminal 10 amino acids → ReEX = Y..[FTY].{0,6}\$

B: YXXΦ in the terminal 20 amino acids → ReEX = Y..[FTY].{0,16}\$

# Variant calling in VEuPathDB galaxy (Part 1)

Learning objectives:

1. Retrieve DNA sequence data from the sequence repository EBI and upload data to VEuPathDB Galaxy using Globus Data Transfer;
2. Name a new project/history;
3. Deploy a Variant calling workflow in the VEuPathDB Galaxy.

Galaxy is an open, web-based platform for data-intensive biomedical research. Galaxy allows you to perform, reproduce, and share complete analyses without the use of command-line scripting. VEuPathDB developed its Galaxy instance in collaboration with Globus Genomics (VEuPathDB Galaxy). To learn how to use Galaxy, follow this link to access tutorials prepared by the Galaxy Training Network: [https://wiki.galaxyproject.org/Learn#Galaxy\\_101](https://wiki.galaxyproject.org/Learn#Galaxy_101)

There are different ways to get data into Galaxy. In this exercise we will use Globus Data Transfer to get data from the EBI server using a unique project ID.

## 1. Retrieve DNA sequence data from the sequence repository and upload data to VEuPathDB Galaxy using Globus Data Transfer option.

- a. Click on the “Globus Data Transfer” menu on the left to expand the Data Transfer section.
- b. Click on the “Get Data via Globus from the EBI server” link.

The screenshot shows the VEuPathDB Galaxy Site interface. On the left, there is a sidebar with a 'Tools' section containing several options under 'VEUPATHDB APPLICATIONS' and a 'DATA TRANSFER' section. The 'Globus Data Transfer' link is highlighted with a red box and an arrow pointing to it. Below it, another link 'Get Data via Globus from the EBI server using your unique file identifier' is also highlighted with a red box and an arrow pointing to it. The main content area features a 'Welcome to the VEuPathDB Galaxy Site' header and a brief description: 'A free, interactive, web-based platform for large-scale data analysis'. It then lists 'With VEuPathDB Galaxy you can:' followed by a numbered list of six items. At the bottom, there are sections for 'Get started with VEuPathDB pre-configured workflows:' and 'OrthoMCL'.

**Welcome to the VEuPathDB Galaxy Site**  
A free, interactive, web-based platform for large-scale data analysis

**With VEuPathDB Galaxy you can:**

1. Start analyzing your data now with pre-configured workflows. All VEuPathDB genomes are pre-loaded.
2. Perform large-scale data analysis with no prior programming or bioinformatics experience.
3. Create custom workflows using an interactive workflow editor. [Learn how](#)
4. Export your results to VEuPathDB, so that you can explore your data with our tools, such as JBrowse and search strategies. See [this tutorial](#).
5. View your results on Galaxy or download results to your computer.
6. Keep data private, or share data with colleagues or the community.

*To learn more about Galaxy, visit the [public Galaxy resources](#).*

**Get started with VEuPathDB pre-configured workflows:**

**OrthoMCL**

This workflow uses BLASTP and the OrthoMCL algorithm to assign your set of proteins to OrthoMCL groups. Version OG6r1 is the latest set of groups (as of April 2020), but you can also select the previous set (OG5). [Explore this tutorial to learn more.](#)

- Workflow to map your proteins to OrthoMCL groups

- c. Enter ENA sample ID and define the dataset type to be transferred into the VEuPathDB Galaxy workspace.

The ENA ID should start with the letters ‘SAM’. For this exercise, we will use SAMN01815907, which is a paired-ended dataset. Take care to specify

whether a dataset is a single or paired-ended as incorrect selection will cause the upload to fail.

- d. Once the form is properly filled, click on the “Execute” button to start the data transfer process.

- e. When the job has been successfully deployed and added to the queue, the screen will refresh, and the added job will appear in the history on the right.

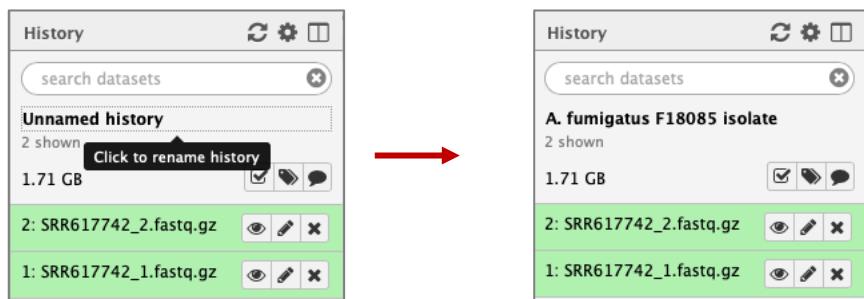
Note: new jobs are highlighted in grey, in progress – yellow, and those completed are in green.

Notice that there are two files appearing in the history on the right. This is because the uploaded data is paired-ended.

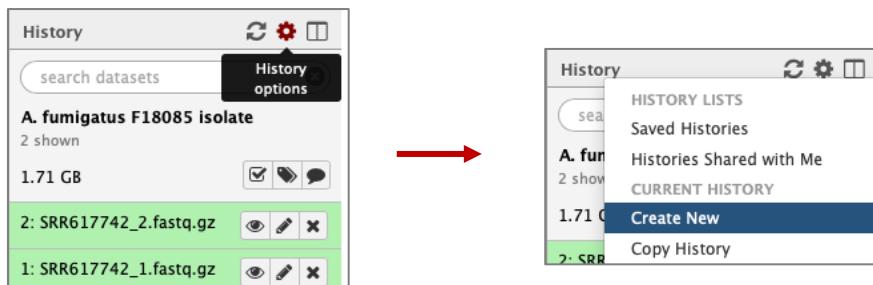
## 2. Rename your history.

By default, all new jobs will be added to the current history on the right. Unless renamed, the history will show up in your history as “Unnamed history”. Let’s rename the history to help us track this project in the future.

- Click on the “Unnamed history” and type “A. fumigatus F18085 isolate”, and then press “enter” to rename this history.



Note: if you would like to start a new project/history, click on the wheel button at the top of the history section and select “Create a new history”.



## 3. Deploy a Variant calling workflow.

VEuPathDB Galaxy main landing page has several workflows for variant calling.

- To navigate to the main page, click on the “Analyze Data”, which is located in the main menu at the top.

The image shows a screenshot of the VEuPathDB Galaxy Analysis home view. The top navigation bar includes "Analyze Data", "Workflow", "Shared Data", "Visualization", "Admin", "Help", "User", and a grid icon. A green success message box is displayed, stating: "Analysis home view" and "1 job has been successfully added to the queue – resulting in the following datasets:". It lists "1: SRR617742\_1.fastq.gz" and "2: SRR617742\_2.fastq.gz". Below this, a note says: "You can check the status of queued jobs and view the resulting data by refreshing the History pane. When the job has been run the status will change from 'running' to 'finished' if completed successfully or 'error' if problems were encountered."

- b. Scroll down to the Variant calling section and choose the workflow for paired-end reads.

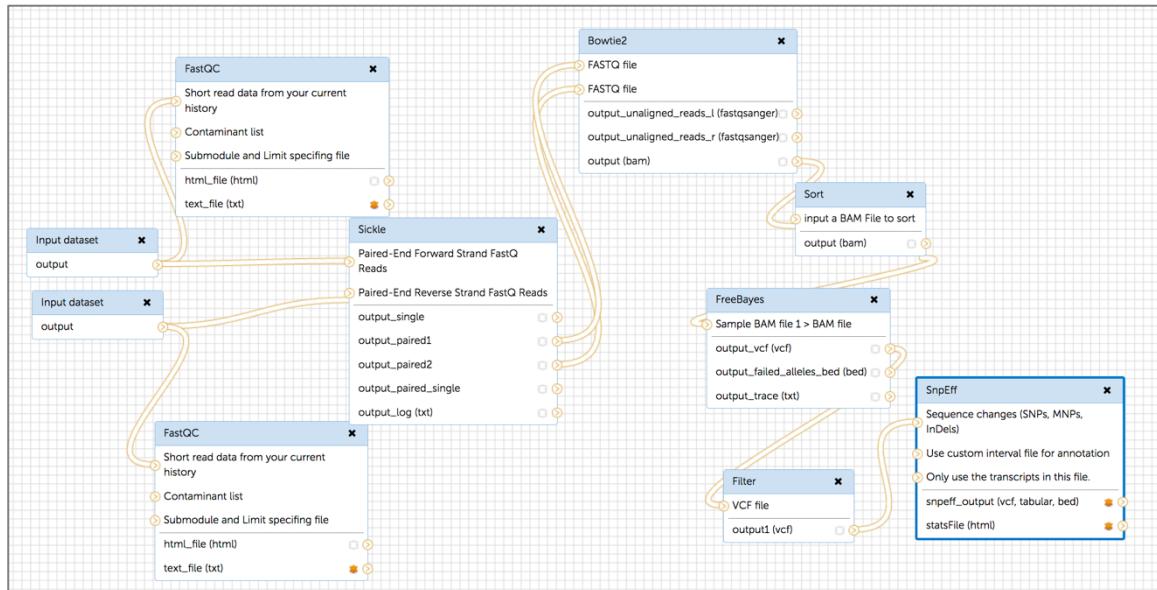
### Variant calling

Use the following workflows to analyze your FASTQ files. The workflows use Sickle for preparation of reads, Bowtie2 for mapping reads to a VEuPathDB reference genome, Freebayes for variant detection, SnpEff to evaluate the effect of variants, and SnpSift for filtering types of variants. Choose the appropriate workflow based on your input data. A VCF file is generated that can be analyzed in Galaxy or downloaded to your computer. NOTE: Export of VCF files to VEuPathDB will be available soon.

- Workflow for single-end reads
- Workflow for paired-end reads

The pre-configured variant calling workflows include the following steps:

- Determine quality of the reads and generate reports (FastQC);
- Trim reads based on their quality scores (Sickle);
- Align reads to a reference genome using Bowtie2 and generate coverage plots ;
- Sort alignments with respect to their chromosomal positions (Sort);
- Detect variants (FreeBayes);
- Filter SNP candidates (Filter);
- Analyze and annotate variants, and calculate the effects of SNPs via SnpEff.



- c. Click on the workflow for paired-end reads and set workflow parameters.

- Make sure that the input steps for paired-end data are set to the xxxx\_1.fastq.gz and xxxx\_2.fastq.gz file (by default the same file will be selected in both fields).

Workflow: imported: EuPathDB\_Workshop\_VariantCalling\_PairedEnd

History Options  
Send results to a new history  
Yes No

1: Input dataset - 1  
1: SRR617742\_1.fastq.gz

2: Input dataset - 2  
2: SRR617742\_2.fastq.gz

History  
A. fumigatus F18085 isolate  
2 shown  
1.71 GB  
1: SRR617742\_1.fastq.gz  
2: SRR617742\_2.fastq.gz

- Select the correct reference genome.
  - Select *Aspergillus fumigatus* Af293 as a reference genome (steps: Bowtie2, FreeBayes, SnpEff).

FreeBayes – Bayesian genetic variant detector (Galaxy Version FREEBAYES: v0.9.21-19-gc003c1e; SAMTOOLS: 0.1.18)

Choose the source for the reference list  
Locally cached

Sample BAM file  
1: Sample BAM file  
BAM file  
Output dataset 'output' from step 7

Using reference genome  
FungiDB-29\_AfumigatusAf293\_Genome

- Choose to deploy the analysis within the same history and click on the Run workflow button.

Workflow: imported: EuPathDB\_Workshop\_VariantCalling\_PairedEnd

History Options  
Send results to a new history  
Yes No

Note: You can use the same workflow to analyze multiple samples in batches. The Upload steps remain the same, however, when setting up the workflow, click on multiple dataset button within the input dataset section.

1: Input dataset - 1  
Multiple datasets

2: Input dataset - 2

This is a batch mode input field. Separate jobs will be triggered for each dataset selection.  
Batch options:

# Variant calling in VEuPathDB galaxy (Part 2)

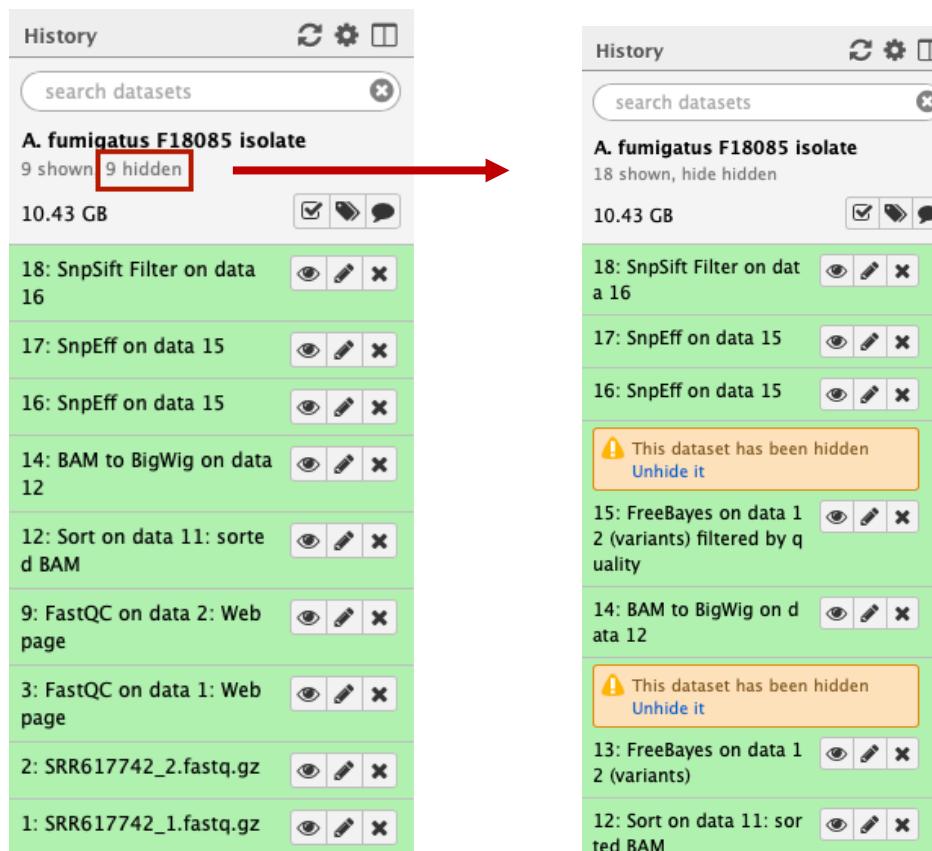
## Learning objectives:

1. [Examine your results and the outputs of the workflow](#)
2. [View VCF file results in the JBrowse genome browser](#)
3. [Filter data in VCF files](#)
4. [Examine the filtered VCF file, extract Gene IDs, and create a Venny diagram](#)
5. [Share or Publish history](#)

## 1. Examine your results and the outputs of the workflow

### a. Examine your results:

Click on the hidden files link in the history panel to reveal all workflow output files.



- The output of Sickle is used by a program called Bowtie2.
- Bowtie generates a file called a BAM file. Whenever dealing with sequence alignment files you will likely hear of file formats called SAM or BAM. SAM stands for Sequence Alignment/Map format, and BAM is the binary version of a SAM file. Many of the downstream analysis programs that use BAM files require a sorted BAM file. This allows access to reads to be done more efficiently.
- The sorted BAM file is the input for a program called FreeBayes. This program is a Bayesian genetic variant detector designed to find small

polymorphisms, specifically SNPs (single-nucleotide polymorphisms), indels (insertions and deletions), MNPs (multi-nucleotide polymorphisms), and complex events (composite insertion and substitution events) smaller than the length of a short-read sequencing alignment.

- The output for many variant callers is a file called a VCF file. VCF stands for variant interchange format.
  - SnpEff is a variant annotation and effect prediction tool. It annotates and predicts the effects of variants on genes (such as amino acid changes).
- b. Examine the output files.
- The output of Sickle is used by a program called Bowtie2.
  - Bowtie generates a file called a BAM file. Whenever dealing with sequence alignment files you will likely hear of file formats called SAM or BAM. SAM stands for Sequence Alignment/Map format, and BAM is the binary version of a SAM file. Many of the downstream analysis programs that use BAM files require a sorted BAM file. This allows access to reads to be done more efficiently.
  - The sorted BAM file is the input for a program called FreeBayes. This program is a Bayesian genetic variant detector designed to find small polymorphisms, specifically SNPs (single-nucleotide polymorphisms), indels (insertions and deletions), MNPs (multi-nucleotide polymorphisms), and complex events (composite insertion and substitution events) smaller than the length of a short-read sequencing alignment.
  - The output for many variant callers is a file called a VCF file. VCF stands for variant interchange format.
  - SnpEff is a variant annotation and effect prediction tool. It annotates and predicts the effects of variants on genes (such as amino acid changes).
- c. Examine the VCF file in your results (click on the eye icon to view its contents). Detailed information about VCF file content is available here: <https://samtools.github.io/hts-specs/VCFv4.2.pdf>

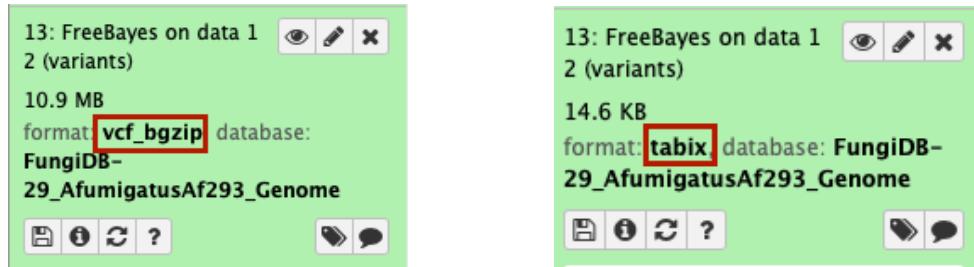
## 2. View VCF file results in the JBrowse genome browser:

*In order to view a VCF file in JBrowse, it first has to be indexed and compressed. We will use a trick in galaxy to produce the required files:*

- a. Find the VCF file you would like to view in JBrowse. This can be the FreeBayes output or the quality filtered VCF. Click on this file to expose the available options.
- b. Click on “display with IGV local” (red circle).
- c. A new window will open up (you are not going to use this window).
- d. Go back to the galaxy window, you will notice that the number of hidden files has increased by 2 files.
- e. Show the hidden files by clicking on the word hidden.

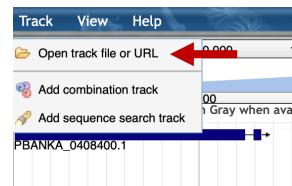


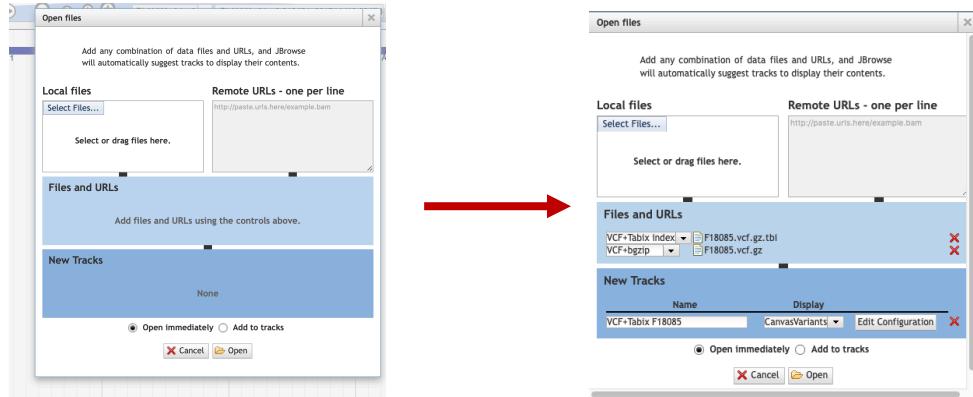
- f. The two new files you want are in the **vcf\_bgzip** and **tabix** format:



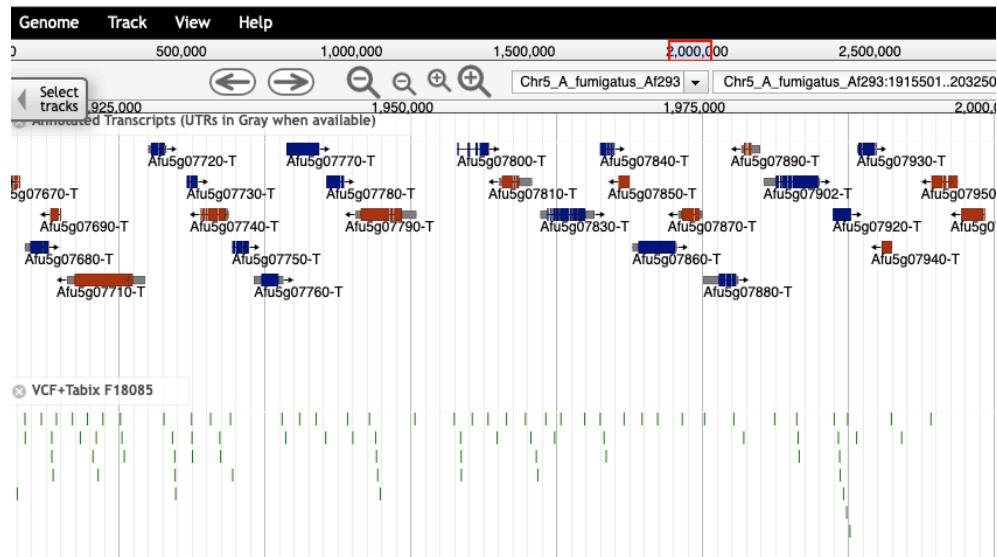
- g. Download both files by clicking on the download icon. *You will need both files.*
- h. After the files are downloaded, After the files are downloaded, rename them as follows:
- Rename the *Galaxy13-[FreeBayes\_on\_data\_12\_(variants)].vcf\_bgzip* file to *sample.vcf.gz* (i.e. F18085.vcf.gz)
  - Rename the *Galaxy13-[FreeBayes\_on\_data\_12\_(variants)].tabix* file to *sample.vcf.gz.tbi* (i.e. F18085.vcf.gz.tbi)
- i. Next, navigate to the Tool section in FungiDB, click on Genome Browser link and select *Aspergillus fumigatus Af293* from the Genome drop down list:

- j. Click on the Track menu, select “Open track file or URL”
- k. Drag and drop your files in the window that appears.  
Notice that the file formats are autodetected.





- I. Click on “Open”. You should see SNP positions displayed in a new track. You can zoom in and click on the SNPs to get more info.

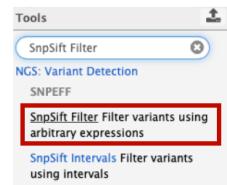


### 3. Filter data in VCF files:

VCF files contain a lot of data about variants and their positions.

SnpEff generates various analyses/summaries of VCF files (including GenIDs that overlap variant positions). However, it is often necessary to filter VCF files further to obtain useful information for your specific question. For example, you may want to filter out SNP positions that have an impact on the coding sequence. One tool that can be used is called SnpSift Filter. This tool allows you to write complex expressions to filter a VCF file.

- a. Navigate to the Tools section and find SnpSift Filter.
  - b. Click on the tool.
  - c. Enter Expression argument



We will run the filter analysis on SNPs with high and moderate impact. Copy and paste the following expression into the SnpSift Filter tool:

((((ANN[\*].IMPACT has 'HIGH') | (ANN[\*].IMPACT has 'MODERATE')) & ((na FILTER) | (FILTER = 'PASS'))))

SnpSift Filter Filter variants using arbitrary expressions (Galaxy Version latest) ▾ Options

VCF input

18: SnpSift Filter on data 16 ▾

Expression

((((ANN[\*]).IMPACT has 'HIGH') | (ANN[\*]).IMPACT has 'MODERATE')) & (((na FILTER) | (FILTER = 'PASS'))))

Execute

#### **4. Examine the filtered VCF file, extract Gene IDs, and create a Venny diagram**

i. 19408\_RPR:2 RUN=1;SAF=2;SAP=12.6832;SAR=9;SRF=6;SRP=26.4622;SRR=23;TYPE=snp;ANN=A|missense\_variant|MODERATE[Afu1g00140|Afu1g00140];  
i;RPR=2;RUN=1;SAF=2;SAP=10.8287;SAR=8;SRF=15;SRP=29.1058;SRR=14;TYPE=snp;ANN=A|missense\_variant|splice\_region\_variant|MODERATE[Afu1g00140];  
i;9253\_RPR:2 RUN=1;SAF=0;SAP=7.3532;SAR=2;SRF=2;SRP=24.5973;SRR=15;TYPE=complex;ANN=TAGTA|splice\_donor\_splice|missense\_variant|splice\_region\_variant|MODERATE[Afu1g00140];  
i;RPR=15;RUN=1;SAF=1;SAP=8;SRF=9;SRP=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=C|missense\_variant|MODERATE[Afu1g00230|Afu1g00230];  
i;0;RPR=16;RUN=1;SAF=1;SAP=18;SRF=4.76149;SAR=13;SRF=0;SRP=0;SRP=0;TYPE=complex;ANN=GGC|missense\_variant|MODERATE[Afu1g00230|Afu1g00230];  
i;RPR=17;RUN=1;SAF=1;SAP=9.45532;SAR=13;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=T|missense\_variant|MODERATE[Afu1g00230|Afu1g00230];  
i;RPR=19;RUN=1;SAF=20;SAP=3.23888;SAR=18;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=A|missense\_variant|MODERATE[Afu1g00230|Afu1g00230];  
i;RPR=17;RUN=1;SAF=1;SAP=20;SAP=3.23888;SAR=18;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=G|missense\_variant|MODERATE[Afu1g00230|Afu1g00230];  
i;RUN=1;SAF=1;SAP=14.8328;SAR=25;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=G|missense\_variant|MODERATE[Afu1g00230|Afu1g00230];  
i;RUN=1;SAF=1;SAP=9.16302;SAR=25;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=G|missense\_variant|MODERATE[Afu1g00230|Afu1g00230];  
i;RPR=0;RPR=18;RUN=1;SAF=1;SAP=10.73113;SAR=13;SRF=0;SRP=0;SRP=0;TYPE=complex;ANN=GATGCCGA|missense\_variant|MODERATE[Afu1g00230|Afu1g00230];  
i;PR=28;RUN=1;SAF=1;SAP=29.44;SAR=21.2375;SAR=35;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=G|missense\_variant|MODERATE[Afu1g00300|Afu1g00300];  
i;RPR=33;RUN=1;SAF=38;SAP=3.25768;SAR=41;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=G|c splice\_acceptor|variant|intron|variant|HIGH|Afu1g00300;  
i;7;RPR=19;RUN=1;SAF=21;SAP=4.16534;SAR=26;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=C|missense\_variant|MODERATE[Afu1g00300|Afu1g00300];  
i;=18;RUN=1;SAF=21;SAP=2.30771;SAR=23;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=G|missense\_variant|MODERATE[Afu1g00300|Afu1g00300];  
i;8777;RPR=0;RUN=1;SAF=1;SAP=3.7342;SAR=2;SRF=0;SRP=5.18177;SRR=1;TYPE=mnp;ANN=AG|missense\_variant|MODERATE[Afu1g00300|Afu1g00300];  
i;=23;RUN=1;SAF=18;SAP=4.09604;SAR=14;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=A|stop\_gained|HIGH|Afu1g00550|Afu1g00550;transcript[Afu1g00550];  
i;7;RPR=26;RUN=1;SAF=25;SAP=3.42611;SAR=22;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=T|missense\_variant|MODERATE[Afu1g00550|Afu1g00550];  
i;7;RPR=11;RUN=1;SAF=1;SAP=6.05036;SAR=21;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=T|missense\_variant|MODERATE[Afu1g00550|Afu1g00550];  
i;=11;RUN=1;SAF=8;SAP=4.77478;SAR=12;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=A|missense\_variant|MODERATE[Afu1g00550|Afu1g00550];transcript[Afu1g00550];  
i;RPR=17;RUN=1;SAF=10;SAP=3.12459;SAR=9;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=T|missense\_variant|MODERATE[Afu1g00570|Afu1g00570];transcript[Afu1g00570];  
i;=1;SAP=11;SAP=3.44459;SAR=9;SRF=0;SRP=0;SRP=0;TYPE=complex;ANN=T|missense\_variant|MODERATE[Afu1g00570|Afu1g00570];transcript[Afu1g00570];  
i;RPR=4;RUN=1;SAF=10;SAP=3.49285;SAR=8;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=T|missense\_variant|MODERATE[Afu1g00570|Afu1g00570];transcript[Afu1g00570];  
i;RPR=25;RUN=1;SAF=14;SAP=3.09072;SAR=13;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=T|missense\_variant|MODERATE[Afu1g00880|Afu1g00880];transcript[Afu1g00880];  
i;=9;RUN=1;SAF=16;SAP=3.08035;SAR=15;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=T|missense\_variant|MODERATE[Afu1g00880|Afu1g00880];transcript[Afu1g00880];  
i;JN=1;SAF=18;SAP=3.51143;SAR=21;SRF=0;SRP=0;SRP=0;TYPE=complex;ANN=CC|missense\_variant|MODERATE[Afu1g00880|Afu1g00880];transcript[Afu1g00880];  
i;RPR=26;RUN=1;SAF=19;SAP=12.2286;SAR=34;SRF=0;SRP=0;SRP=0;TYPE=complex;ANN=TAAGA|missense\_variant|MODERATE[Afu1g00880|Afu1g00880];  
i;RPR=10;RUN=1;SAF=1;SAP=8.72747;SAR=24;SRF=0;SRP=5.18177;SRR=0;TYPE=snp;ANN=T|missense\_variant|MODERATE[Afu1g00880|Afu1g00880];  
i;

```
search datasets
A. fumigatus F18085 Isolate
1 shown
0.45 GB
9: SnpSift Filter on dat
18
18: SnpSift Filter on da
ta 16
10,300 lines, 55 comments
format:vcf, database: FungiDB-
29_AfumigatusFA293_1_Genome
View data

Command to execute: java -
Xmx4g -jar
[mm/gatk/GATKTools/tools/snpeff/sn-
perf filter -f
[scratch/galaxy/files/000/185/dat
e
[scratch/galaxy/job_working_direc


display with IGV local
1. Chrom
#INFO<=ID=DP_Number=1,Type=Integer
#INFO<=ID=DPB_Number=1,Type=Float,
#INFO<=ID=CN_Number=1,Type=Integer
#INFO<=ID=CNR_Number=1,Type=Integer
#INFO<=ID=AF_Number=4,Type=Float,[
```

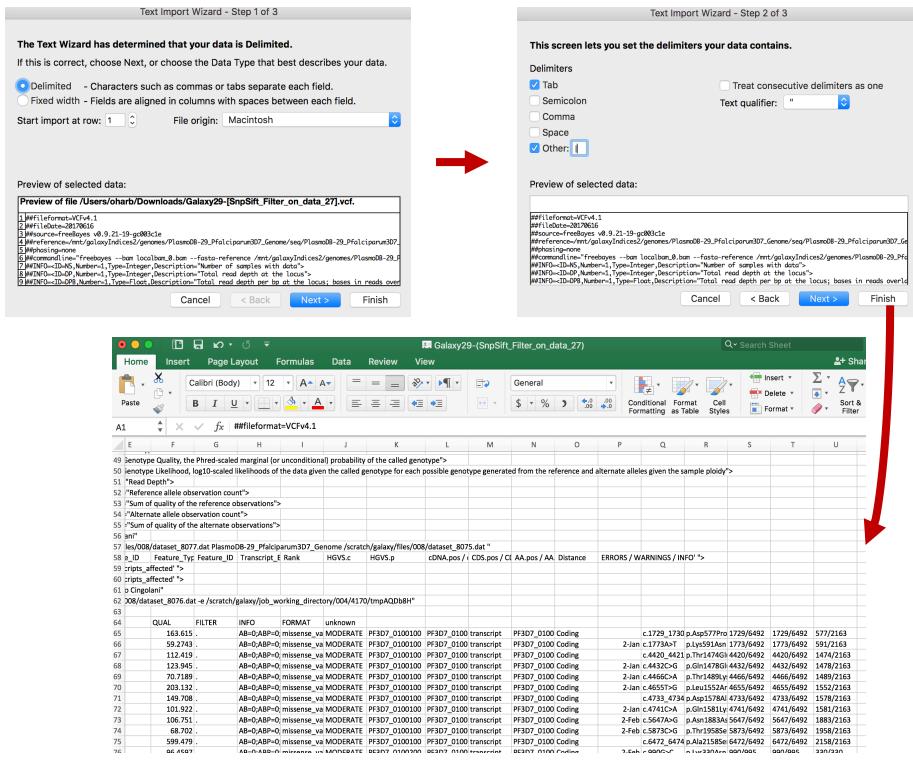
Notice that the GeneIDs are buried in the file but the file has some structure which means you can extract them either programmatically or using a program like Excel.

Here are some steps you can take to extract Gene IDs from two VCF files then compare them to identify genes that are in common or that distinguish the two files.

- a. Download the SnpSift Filter output by clicking on the save icon

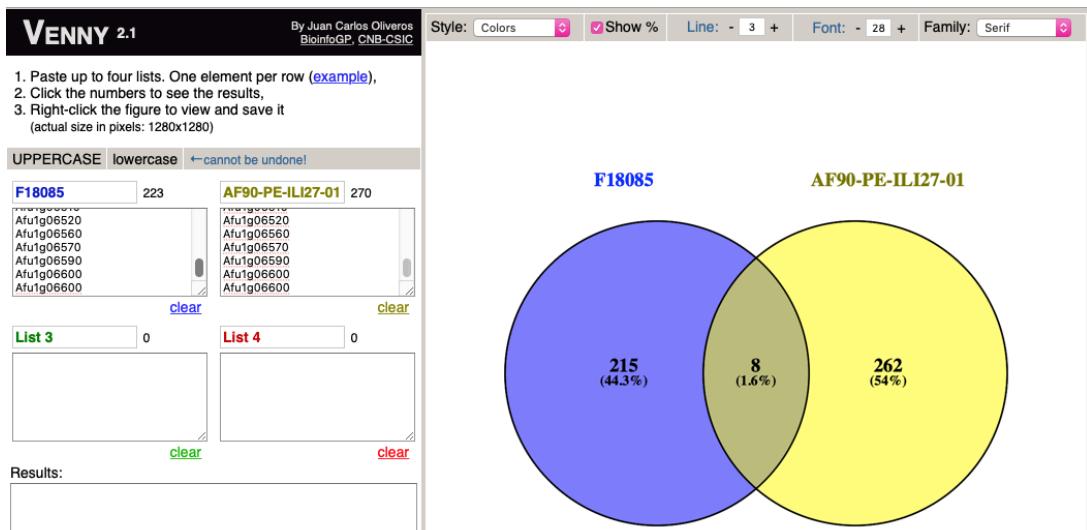
```
18: SnpSift Filter on da  
 16  
10,300 lines, 55 comments  
format: vcf, database: FungiDB-  
29_AfumigatusAf293_Genome  
  
Command to execute: java -  
Xmx4g -jar  
/mnt/galaxyTools/tools/snpeff/snij  
filter filter -f  
/scratch/galaxy/files/000/185/dat  
-e  
/scratch/galaxy/job_working_direc
```

- b. Open this file using Excel. Select a column to split up, choose Data>Tab to Column and make sure you select tabs and | as column delimiter



- c. Now you can look for Gene IDs of interest in the excel file. For example, if this is a known drug resistant line you can find the gene(s) that might be responsible for the resistance and see what kinds of SNPs are present.

If you are comparing a mutant and a wild type or two different strains you can extract gene IDs from both VCF files and use Venny website (<https://bioinfogp.cnb.csic.es/tools/venny/>) to visualize your data.



\*Note that in the above steps you are ultimately comparing gene IDs – do you think you might be missing some important polymorphisms using this method? Of course, the answer is yes😊

It is quite possible that a gene with a SNP in the WT and a SNP in the mutant that will be in the intersection of the two gene lists, contains different SNPs – you will miss this by doing the above steps. Below is a description of steps you can take to create a list of unique IDs for SNPs. This list of unique IDs can then be used in Venny.

1. Start with the same excel files that you opened in the above section.
2. To create a unique ID for SNPs we will combine information from multiple columns to create something that looks like this:  
chromosome:position:genelID
3. To do this you will use the concatenate function in Excel:  
`=concatenate(cell#1,":",cell#2,":",cell#3)`  
Cell#1 = cell with chromosome number  
Cell#2 = cell with position  
Cell#3 = cell with GenelD

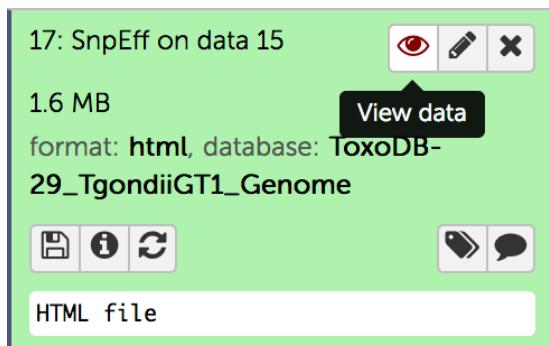
A	B	C	D	E	F	G	H	I	J	K	BS	BT
50 ##SnpEffVersion= 4.21 (build 2015-10-03), by Pablo Cingolani												
51 ##SnpEffCmd="SnpEff -vcf -vcf-stats /scratch/galaxy/files/008/dataset_8107.dat ToxoDB-29_TgondiiGT1_Genome /scratch/galaxy/files/008/dataset_B105.dat "												
52 ##INFO<ID=ANN,Num=Annotation Annotation_Gene_Name Gene_ID Feature_Type Feature_ID Transcript_E_Rank HGVS_c HGVS_p												
53 ##INFO<ID=LOF,Numb_Gene_ID Number_of_Percent_of_transcripts_affected>												
54 ##INFO<ID=NMD,Numb_Gene_ID Number_of_Percent_of_transcripts_affected>												
55 ##SnpSiftVersion="SnpSift 4.1 (build 2015-10-03), by Pablo Cingolani"												
56 ##SnpSiftCmd="SnpSift filter -f /scratch/galaxy/files/008/dataset_8106.dat -e /scratch/galaxy/job_working_directory/004/4169/tmpBopqfU"												
57 ##FILTER=<ID=SnpSift_D (ANN*)>.IMF (FILTER = "PASS")">												
64 #CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	unknown			
65 TGGT1_chrla	227230		A	C	1156.55	AB=0;ABP=0; missense Va MODERATE	TGGT1_293300					
66 TGGT1_chrla	1340271		G	C	2387.77	AB=0;ABP=0; missense Va MODERATE	TGGT1_295040					
67 TGGT1_chrla	1396177		A	C	387.162	AB=0;ABP=0; missense Va MODERATE	TGGT1_295125					
68 TGGT1_chrlb	78769		A	G	1780.8	AB=0;ABP=0; missense Va MODERATE	TGGT1_207440					
69 TGGT1_chrlb	153771		T	G	1414.57	AB=0;ABP=0; missense Va MODERATE	TGGT1_207480					
70 TGGT1_chrlb	276348		T	G	2066.14	AB=0;ABP=0; missense Va MODERATE	TGGT1_207750					
71 TGGT1_chrlb	622140		G	C	2335.06	AB=0;ABP=0; missense Va MODERATE	TGGT1_208310					
72 TGGT1_chrlb	1446003		C	T	60.6579	AB=0;ABP=0; missense Va MODERATE	TGGT1_209755B					
73 TGGT1_chrlb	1446022		G	T	82.4046	AB=0;ABP=0; missense Va MODERATE	TGGT1_209755B					

A	B	C	D	E	F	G	H	I	J	K	BS	BT
50 ##SnpEffVersion= 4.21 (build 2015-10-03), by Pablo Cingolani												
51 ##SnpEffCmd="SnpEff -vcf -vcf-stats /scratch/galaxy/files/008/dataset_8107.dat ToxoDB-29_TgondiiGT1_Genome /scratch/galaxy/files/008/dataset_B105.dat "												
52 ##INFO<ID=ANN,Num=Annotation Annotation_Gene_Name Gene_ID Feature_Type Feature_ID Transcript_E_Rank HGVS_c HGVS_p												
53 ##INFO<ID=LOF,Numb_Gene_ID Number_of_Percent_of_transcripts_affected>												
54 ##INFO<ID=NMD,Numb_Gene_ID Number_of_Percent_of_transcripts_affected>												
55 ##SnpSiftVersion="SnpSift 4.1 (build 2015-10-03), by Pablo Cingolani"												
56 ##SnpSiftCmd="SnpSift filter -f /scratch/galaxy/files/008/dataset_8106.dat -e /scratch/galaxy/job_working_directory/004/4169/tmpBopqfU"												
57 ##FILTER=<ID=SnpSift_D (ANN*)>.IMF (FILTER = "PASS")">												
64 #CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	unknown			
65 TGGT1_chrla	227230		A	C	1156.55	AB=0;ABP=0; missense Va MODERATE	TGGT1_293300					
66 TGGT1_chrla	1340271		G	C	2387.77	AB=0;ABP=0; missense Va MODERATE	TGGT1_295040					
67 TGGT1_chrla	1396177		A	C	387.162	AB=0;ABP=0; missense Va MODERATE	TGGT1_295125					
68 TGGT1_chrlb	78769		A	G	1780.8	AB=0;ABP=0; missense Va MODERATE	TGGT1_207440					
69 TGGT1_chrlb	153771		T	G	1414.57	AB=0;ABP=0; missense Va MODERATE	TGGT1_207480					
70 TGGT1_chrlb	276348		T	G	2066.14	AB=0;ABP=0; missense Va MODERATE	TGGT1_207750					
71 TGGT1_chrlb	622140		G	C	2335.06	AB=0;ABP=0; missense Va MODERATE	TGGT1_208310					
72 TGGT1_chrlb	1446003		C	T	60.6579	AB=0;ABP=0; missense Va MODERATE	TGGT1_209755B					
73 TGGT1_chrlb	1446022		G	T	82.4046	AB=0;ABP=0; missense Va MODERATE	TGGT1_209755B					

4. You should get unique SNP IDs that look like this (for example):  
TGGT1\_chrlb:1446003:TGGT1\_209755B
5. Copy this function to the rest of the column to replicate the concatenate function.
6. Copy the these newly generated unique IDs into Venny and compare the mutant and wild type.

## Supporting information - Examining SnpEff summary:

- Click on the view icon (eye) in the SnpEff output file that has the html format.



- This will open the html file right in galaxy where you can view it.
- The header contains a short summary and information about the run and it has several major components:
  1. Summary table that warns about possible genomic annotation errors or inconsistencies identified in the reference genome. If there are many, use caution interpreting results and examine associated gff files for any issues (ex. missing feature values in gff files, incomplete gene sequences, more than one stop codon per gene, etc.).
  2. Summary statistics for variant types

Number variants by type	
Type	Total
SNP	114,034
MNP	12,864
INS	6,907
DEL	7,304
MIXED	2,180
INTERVAL	0
Total	143,289

Here is an example of variant calls and what they mean in terms of nucleotide changes:

Type	What it means	Example
SNP	Single-Nucleotide Polymorphism	Reference = 'A', Sample = 'C'
Ins	Insertion	Reference = 'A', Sample = 'AGT'
Del	Deletion	Reference = 'AC', Sample = 'C'
MNP	Multiple-nucleotide polymorphism	Reference = 'ATA', Sample = 'GTC'
MIXED	Multiple-nucleotide and an InDel	Reference = 'ATA', Sample = 'GTCAGT'

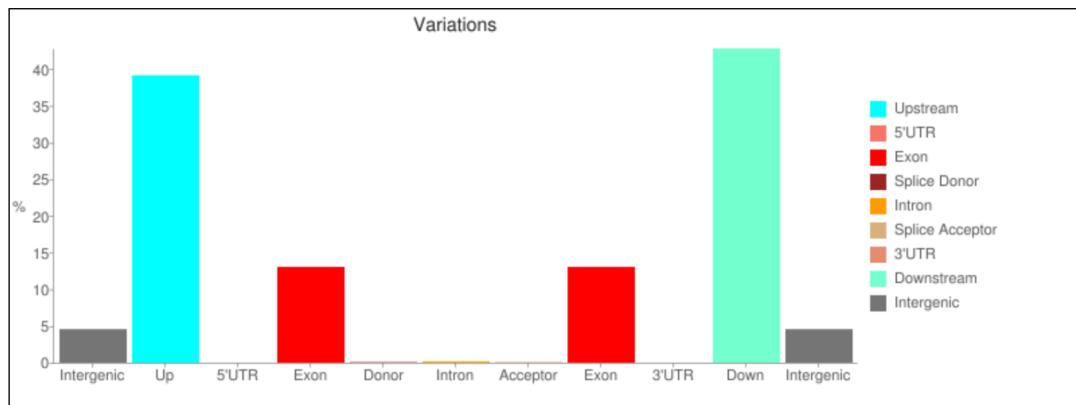
### 3. Statistics for the variant effects and impacts:

**Number of effects by functional class**

Type (alphabetical order)	Count	Percent
MISSENSE	21,588	35.949%
NONSENSE	131	0.218%
SILENT	38,332	63.832%

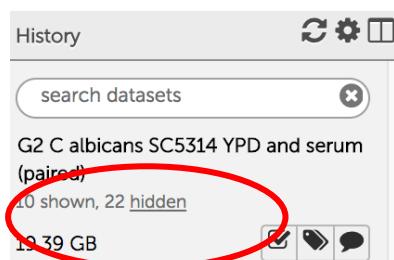
Type (alphabetical order)	Count	Percent
DOWNSTREAM	321,858	40.292%
EXON	67,505	8.451%
INTERGENIC	74,749	9.358%
INTRON	1,064	0.133%
NONE	1	0%
SPlice_SITE_ACCEPTOR	5	0.001%
SPlice_SITE_DONOR	4	0.001%
SPlice_SITE_REGION	176	0.022%
TRANSCRIPT	12	0.002%
UPSTREAM	333,432	41.741%

Base changes summary. SnpEff html files provides a breakdown of SNPs across gene features:



The SNP workflow you are using is set up to generate certain files that will provide you with the information you can export and use further in your analysis (yellow stars).

If you select certain options they will be shown in your history. If you do not select to display these files, you can view the output by clicking on displaying the hidden files from the history menu:



Now, lets take a look at the files generated by the workflow and steps that you can take to further evaluate them.

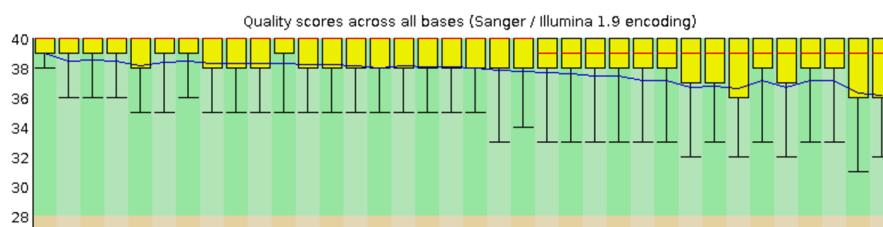
## 1. Examine sequence quality based on FastQC quality scores.

FastQC provides an easy-to-navigate visual representation sequencing data quality and distribution of nucleotides per read position.

### Basic Statistics

Measure	Value
Filename	SRR298691.fastq
File type	Conventional base calls
Encoding	Sanger / Illumina 1.9
Total Sequences	4887868
Sequences flagged as poor quality	0
Sequence length	36
%GC	58

### Per base sequence quality



## 2. Download vcf files and evaluate workflow results.

The vcf file generated by SnpEff contains information about SNPs and the genomic location.

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	unknown			
CM001231	189057	.	AG	CT	787.449	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:143:0:0:143:5341:-207.887,-43.0473,0					
CM001231	483825	.	G	A	64.8756	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:4:0:0:4:146:-10.0999,-1.20412,0					
CM001231	518226	.	G	C	51.7908	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:8:0:7:276:-11.5007,-2.10721,0					
CM001231	574021	.	C	G	237.265	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:32:17:0:17:583:-39.079,-5.11751,0					
CM001231	609879	.	GAA	CAG	55.2785	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:32:8:277:22:861:-18.1711,-0.694735,0					
CM001231	1090073	.	G	T	79.4156	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:8:2:75:6:238:-11.5539,-1.36362,0					
CM001231	1090104	.	A	T	70.961	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:6:0:0:6:220:-12.5146,-1.80618,0					
CM001231	1153611	.	CCTC	GCTG	111.123	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:8:5:188:3:97:-9.30616,-6.1461,0					
CM001231	1159150	.	CT	GC	126.126	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:31:0:0:19:741:-29.7713,-5.71957,0					
CM001231	1159438	.	C	G	82.3312	.	AB=0;ABP=0;GT:DP:RO:QF 0/0:47:30:17:640:0,-9.53002,-3.50705					
CM001231	1159465	.	G	C	249.656	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:126:47:1770:79:3013:-53.8644,-25.2134,0					
CM001231	1159499	.	T	C	124.95	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:143:32:1167:111:4248:-76.1575,-33.4865,0					
CM001231	1181576	.	CC	TG	191.675	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:27:0:0:25:924:-41.7448,-7.52575,0					
CM001231	1293309	.	C	G	51.22	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:2:0:0:2:78:-6.92763,-0.60206,0					
CM001231	1323058	.	TT	GC	71.3001	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:6:0:0:6:223:-12.5485,-1.80618,0					
CM001231	1485397	.	A	G	3558.42	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:499:0:0:497:18671:-804.678,-149.612,0					
CM001231	1485429	.	G	A	3783.33	.	AB=0;ABP=0;GT:DP:RO:QF 1/1:517:1:38:516:20010:-843.425,-151.978,0					

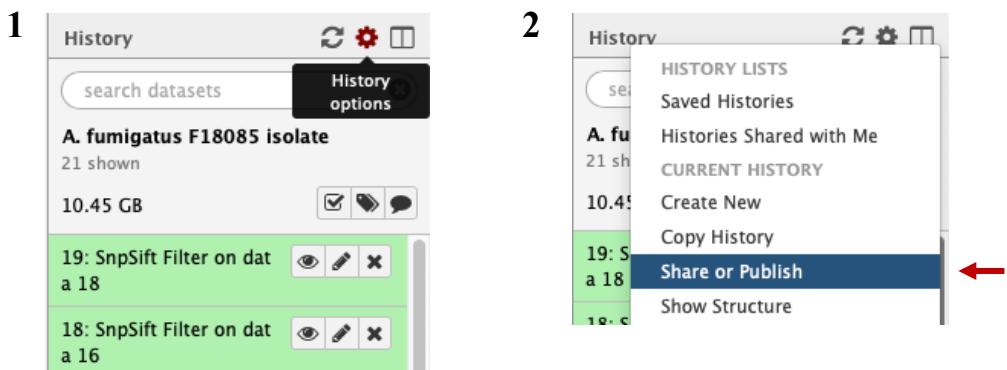
Post-processing of SNP data is normally required to make sense of thousands of SNPs and to decide which ones have biological and functional importance. Data processing can help you to extract SNP distribution and parse associated data including Gene IDs, protein-coding annotations, and effects in sequence ontology terms such as missense or synonymous variants, stop codon gain, etc. and also link changes to the genome model.

Summary	
<b>Genome</b>	ToxoDB-29_TgondiiGT1_Genome
<b>Date</b>	2017-06-17 05:56
<b>SnpEff version</b>	SnpEff 4.11 (build 2015-10-03), by Pablo Cingolani
<b>Command line arguments</b>	SnpEff -i vcf -o vcf -stats /scratch/galaxy/files/008/dataset_8107.dat ToxoDB-29_TgondiiGT1_Genome /scratch/galaxy/files/008/dataset_8105.dat
<b>Warnings</b>	3,941
<b>Errors</b>	0
<b>Number of lines (input file)</b>	8,411
<b>Number of variants (before filter)</b>	8,483
<b>Number of not variants (i.e. reference equals alternative)</b>	0
<b>Number of variants processed (i.e. after filter and non-variants)</b>	8,483
<b>Number of known variants (i.e. non-empty ID)</b>	0 ( 0 % )
<b>Number of multi-allelic VCF entries (i.e. more than two alleles)</b>	72
<b>Number of effects</b>	14,149
<b>Genome total length</b>	63,945,332
<b>Genome effective</b>	.....

## 4. Share or publish workflow results (history).

There are three ways to share your history. Click on the on the “History options”, “Share or Publish” to access these options.

- Select “Make History Accessible via Link” if you would like to share a link with your colleague who also has an account with VEuPathDB.
- Select “Make History Accessible and Publish” if you would like to make the history visible in the Shared section of the VEuPathDB Galaxy.
- Click “Share with a user” to share the history directly with one or several users.



### 3 Share or Publish History 'A. fumigatus F18085 isolate'

#### Make History Accessible via Link and Publish It

This history is currently restricted so that only you and the users listed below can access it. You can:

[Make History Accessible via Link](#)

Generates a web link that you can share with other people so that they can view and import the history.

[Make History Accessible and Publish](#)

Makes the history accessible via link (see above) and publishes the history to Galaxy's Published Histories section, where it is publicly listed and searchable.

#### Share History with Individual Users

You have not shared this history with any users.

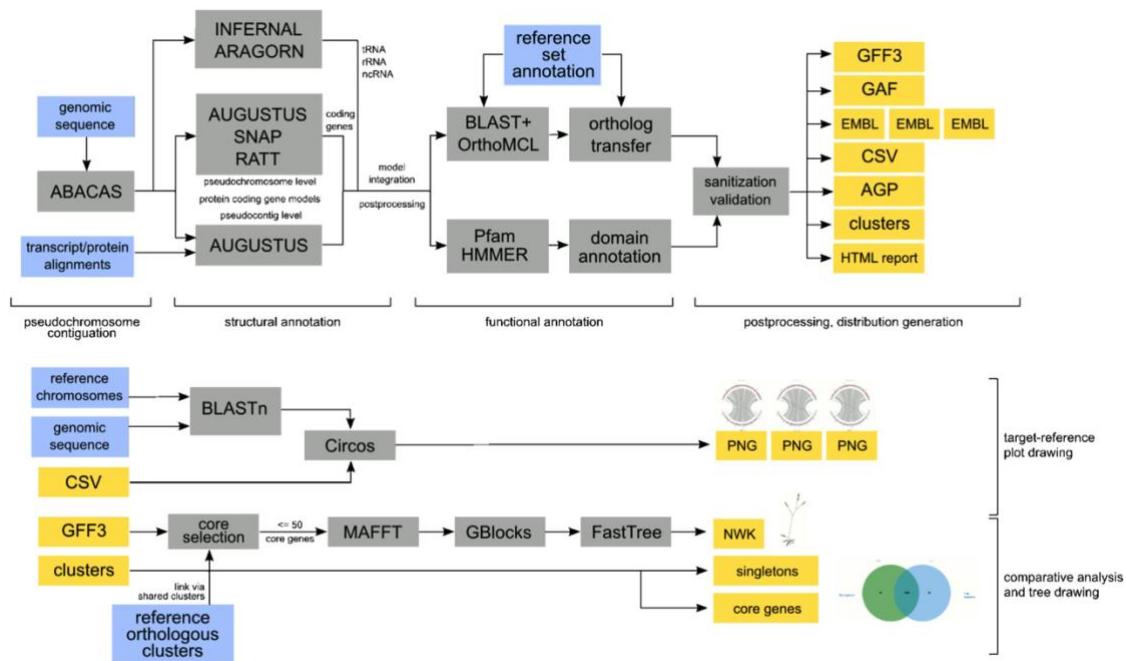
[Share with a user](#)

## Genome Annotation with Companion (Part 1)

### Learning objectives:

- Download genomes and chromosomes from VEuPathDB
- Annotate a genome with Companion
- Interpreting the Companion result
- Download the Companion output and load it into Artemis
- Learn how to use ACT to compare genomes and look at the Companion output

Companion, is an online pipeline that employs different software to annotate and compare an assembled sequence to a reference-annotated genome. The figure below illustrates the Companion pipeline, the software used and the expected output.



For this exercise, we will start with an unannotated genome. We will obtain the assembled FASTA files from VEuPathDB sites. Companion can be accessed here: <http://companion.gla.ac.uk/>

Each group will download one of the following genomes (the tinyURL links will initiate the download) and will use Companion to compare with the specified genome as reference.

Group 1 – *Plasmodium coatneyi* Hackeri using *Plasmodium knowlesi* as reference  
<https://tinyurl.com/y47vvsoj>

Group 2 - *Plasmodium coatneyi* Hackeri using *Plasmodium falciparum* as reference  
<https://tinyurl.com/y47vvsoj>

Group 3 – *Cryptosporidium meleagridis* using *Cryptosporidium parvum* as reference  
<https://tinyurl.com/y4fgc3p5>

Group 4 *Cryptosporidium baileyi* using *Cryptosporidium parvum* as reference  
<https://tinyurl.com/y44ucs5t>

Group 5 *Trypanosoma congolense* using *Trypanosoma brucei* 927 as reference.  
<https://tinyurl.com/yxausbhg>

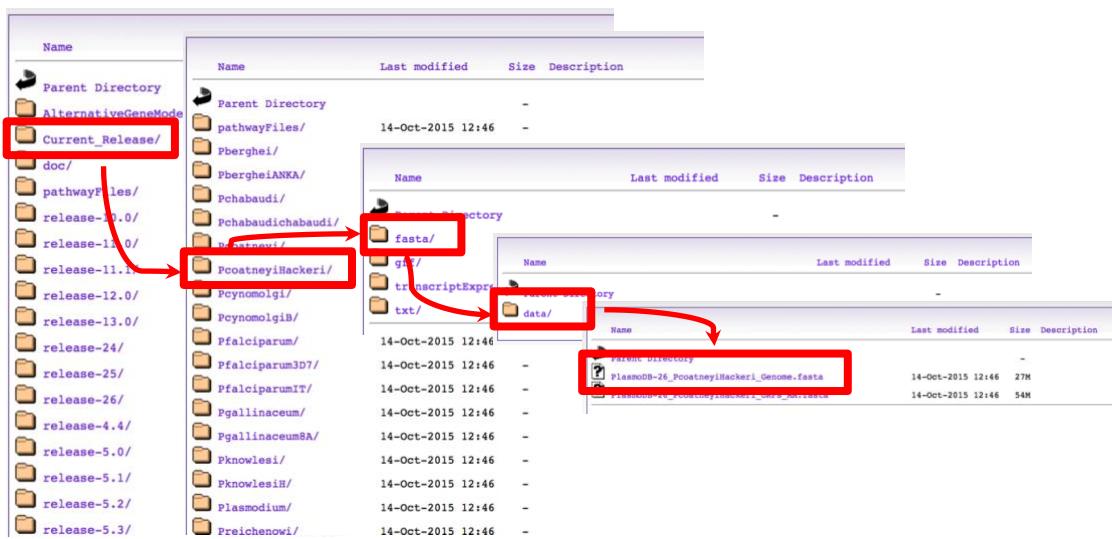
Group 6 *Trypanosoma congolense* 2019 using *Trypanosoma brucei* 927 as reference.  
<https://tinyurl.com/y4pqscrm>

### A word about downloads:

TinyUrls above are direct links to our genome FASTA files in the corresponding VEuPathDB site downloads section. All genomes in VEuPathDB sites are available for download from the “Data File” download section, which you can access from the top menu by clicking on “Data”.



Selecting the option “Download data files” takes you to the download directories where you can navigate to the genome and data type you are looking for.



To download specific contigs/scaffolds/chromosomes instead of entire genomes, use a genomic sequence search and place the desired sequences into your basket.

**Search for...**

expand all | collapse all

Filter the searches below... 

- ▶ Genes
- ▶ Organisms
- ▶ Popset Isolate Sequences
- ▶ **Genomic Sequences**
- 🔍 BLAST
- 🔍 Copy Number/Ploidy
- 🔍 Genomic Sequence ID(s)
- 🔍 **Organism** 
- ▶ Genomic Segments
- ▶ SNPs
- ▶ SNPs (from Array)
- ▶ ESTs
- ▶ Metabolic Pathways
- ▶ Compounds

## Identify Genomic Sequences based on Organism

 **Organism**

Note: You must select at least 1 values for this parameter.  
1 selected, out of 45

select all | clear all | expand all | collapse all

Filter list below... 

- Plasmodium adleri
- Plasmodium berghei
- Plasmodium biliocelluli
- Plasmodium blacklocki
- Plasmodium chabaudi
- Plasmodium coatneyi 
- Plasmodium coatneyi Hacketti
- Plasmodium cynomolgi
- Plasmodium falciparum
- Plasmodium fragile
- Plasmodium gaboni
- Plasmodium gallinaceum
- Plasmodium inui
- Plasmodium knowlesi
- Plasmodium malariae
- Plasmodium ovale curtisi
- Plasmodium praefalciparum
- Plasmodium reichenowi
- Plasmodium relictum
- Plasmodium vinckei
- Plasmodium vivax
- Plasmodium vivax-like sp.
- Plasmodium yoelli

select all | clear all | expand all | collapse all

**My Search Strategies**

Opened (1) All (27) Public (42) Help

Unnamed Search Strategy \*

Organism 14 Sequences + Add a step Step 1

14 Genomic Sequences Revise this search

Genomic Sequence Results

Rows per page: 20

Sequence ID CP016250 CP016249 CP016247 CP016246 CP016251

Genomic Sequence Results

Rows per page: 20

Sequence ID	Organism	Length
CP016250	Plasmodium coatneyi Hackeri	4,930,710
CP016249	Plasmodium coatneyi Hackeri	2,627,280
CP016247	Plasmodium coatneyi Hackeri	2,583,428
CP016246	Plasmodium coatneyi Hackeri	2,003,671

Download Add to Basket Add Columns

You can access the basket from the top menu.

PlasmoDB Release 48 beta 27 Aug 2020 A VEuPathDB Project. My Strategies Searches Tools My Workspace Data About Help Contact Us

My Search Strategies

Opened (1) All (27) Public (42) Help

Unnamed Search Strategy \*

My Baskets

Genomic Sequences (2)

Empty Basket Save Basket to Strategy

In case of Error, please Contact Us or empty your basket. On new releases IDs sometimes change or are retired.

2 Genomic Sequences

Genomic Sequence Results

Rows per page: 20

Sequence ID	Organism	GenbankRecord	Genome Browser	JbrowseUrl
CP016247	Plasmodium coatneyi Hackeri	N/A	Browser	/plasmo/app/browse?loc=CP016247:1..2583428&data=/plasmo/service/jbrowse/tracks/pcoa-hackeri&track...
CP016250	Plasmodium coatneyi Hackeri	N/A	Browser	/plasmo/app/browse?loc=CP016250:1..4930710&data=/plasmo/service/jbrowse/tracks/pcoa-hackeri&track...

Download Add Columns Retrieve Sequence

Nucleotide positions: 1 to 2583428 Reverse & Complement Get Sequence

Nucleotide positions: 1 to 4930710 Reverse & Complement Get Sequence

**Download Genomic Sequence Basket**

Choose a Report:

- Tab- or comma-delimited (openable in Excel) - choose columns to make a custom table [?](#)
- Tab- or comma-delimited (openable in Excel) - choose a pre-configured table [?](#)
- FASTA - sequence retrieval, configurable [?](#) ←
- Standard JSON [?](#)

---

Choose the region of the sequence(s):

Reverse & Complement ←

Nucleotide positions:  to  (0 = end)

Download Type:

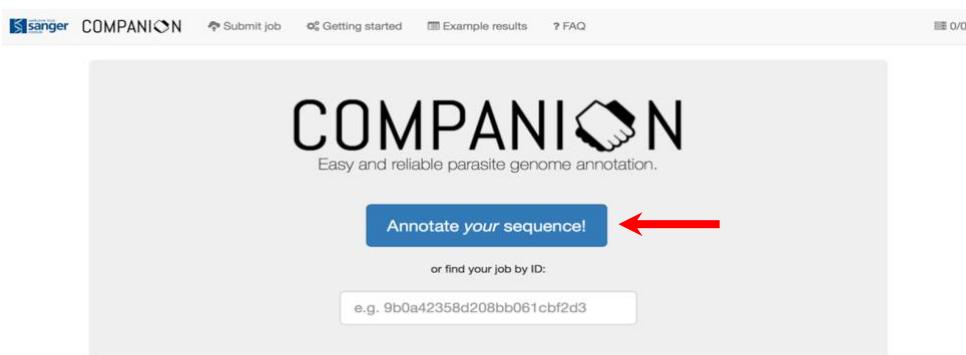
Text File ←

Show in Browser

**Get Sequences**  

-**Back to the Annotation:** Once you have downloaded your sequence file, go to the Companion site:  
<http://companion.gla.ac.uk/>

- Click on the “Annotate your sequence” link.



-Follow the instructions as described on the Companion website:  
1. Provide basic information about the job you are about to submit. This includes a job name, species prefix (usually the first letter of the genus and the first three letters of the species: *Cryptosporidium parvum* = CPAR).

## Submit a new annotation job

Step 1: Basic job properties			
<p>First of all, please specify a free-text <b>name</b> for your new job. It should reflect the purpose of your job, and should probably include the organism you are annotating.</p> <p>Example: <i>My new species annotation</i></p> <table border="1"><tr><td>Job name</td><td></td></tr></table>		Job name	
Job name			
<p>Please also give a short <b>species prefix</b> that will be used to name entities (such as genes, pseudochromosomes, etc.) generated during the annotation run. It should not contain spaces or special characters.</p> <p>Example: <i>LDON</i></p> <table border="1"><tr><td>Species prefix</td><td>LFOO</td></tr></table>		Species prefix	LFOO
Species prefix	LFOO		
<p>Finally, please provide a <b>species name</b> that describes the target species you are annotating.</p> <p>Example: <i>Leishmania donovani</i></p> <table border="1"><tr><td>Species name</td><td>Leishmania donovani</td></tr></table>		Species name	Leishmania donovani
Species name	Leishmania donovani		

2. In step 2, choose the assembly file that you downloaded.
3. In step 3, indicate if you will be using RNAseq evidence to guide the annotation – in this exercise we will not use any RNAseq data.
4. In step 4, select the reference sequence you would like to use to transfer the annotation and to compare your sequence to. Typically, you would like to use a reference that is closely related, so a phylogenetic tree might be useful to look at. Here are examples of phylogenies for *Plasmodium* and *Cryptosporidium*.

<http://tolweb.org/Cryptosporidium/124803>

<http://tolweb.org/Plasmodium/68071>

*Trypanosoma* phylogenetic tree

[https://projects.exeter.ac.uk/meeg/sites/default/files/pictures/tryp\\_tree.jpg](https://projects.exeter.ac.uk/meeg/sites/default/files/pictures/tryp_tree.jpg)

## Step 2: Target sequence

Please upload a **target sequence file** to be annotated from your local filesystem using the button below. The file (FASTA, EMBL or GenBank format) can be gzip- or bzip2-compressed. In this case it must have a .gz or .bz2 suffix.

Note: The maximal size of your uploaded file is **64 MB**, and the maximum number of individual sequences in it is **3000**.

Choose File no file selected

[Here](#) is an example sequence input file for a *Plasmodium falciparum* IT chromosome 5 sequence that can be used with the *Plasmodium falciparum* 3D7 example reference set (choose below in step 4) for a quick example run. To use it, please download it to your local machine and upload it using the button above.

## Step 3: Transcript evidence

The Companion pipeline can optionally make use of assembled transcripts in the GTF format as created by Cufflinks.

- Yes, use transcript evidence.  
 No, do not use transcript evidence.

## Step 4: Reference organism

Please pick a (if possible closely related) **reference organism** for this annotation run. This organism will be used to specify the models for gene finding, functional annotation transfer and pseudochromosome contiguation.

Please select a reference species

5. In step 5, there are a few more parameters you may want to examine. For the purpose of our exercise we will keep these at the default values.

## Step 5: Pseudochromosome contiguation

The contiguation step will try to orientate the sequences in your input file to align with the chromosomal sequences of the reference organism to build pseudochromosomes, which will then be used as the target sequences for gene annotation. This step is optional; if it is not desired then no modifications will be made to the input sequences.

- Yes, contiguate pseudochromosomes.  
 No, do not modify my input sequences.

Select minimum required match length for contig placement: 500 bp

200  20000

Select minimum required match similarity for contig placement: 85 %

30  100

6. Enter your email address to get an update when your job starts running and when it is complete. Next, click on the “I’m not a robot” captcha (Completely Automated Public Turing test to tell Computers and Humans Apart). Finally, click on the “Submit Job” link.

**Step 6: Advanced settings (click chevron to the right to show/hide)**



**Your contact information (optional)**

You can leave your email address if you want to be notified when your job starts and finishes. This is absolutely optional, if you choose not to share your email address, you can always manually check the status of your job using a private link provided by us after submission.

Email

To protect the service from automated bots, please prove that you are a human by ticking the box below.

I'm not a robot



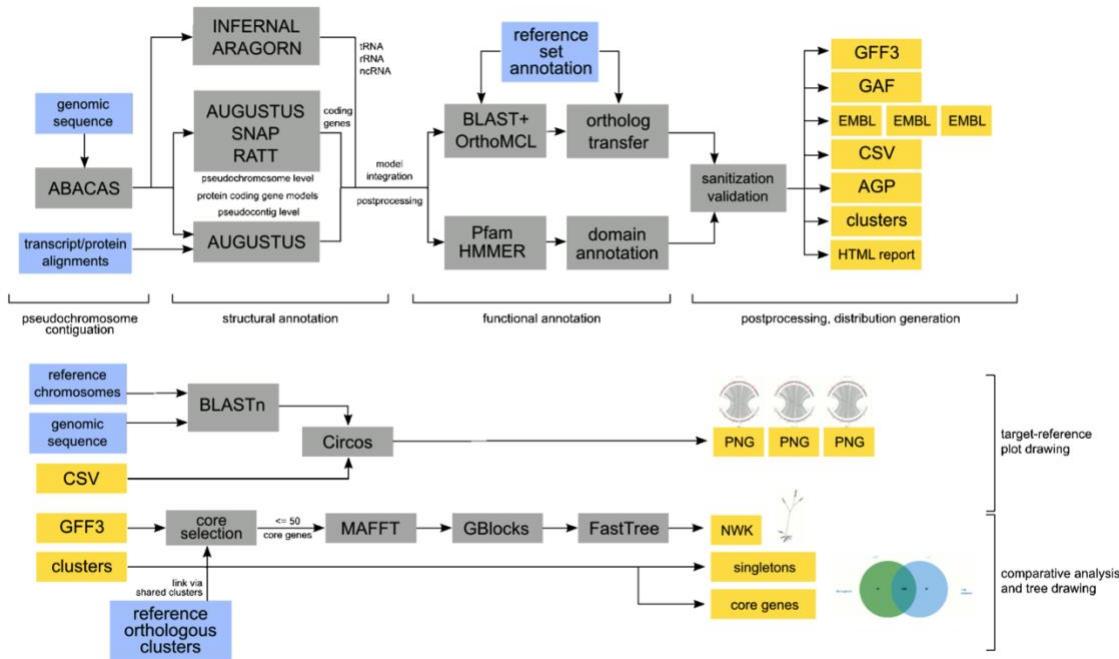
reCAPTCHA

[Privacy](#) · [Terms](#)

Submit job

## Genome Annotation with Companion (Part 2)

You should have gotten an email indicating the status of your annotation (ie. job started and job complete). The email contains a link to the annotation output.



- Explore your results with your group and discuss the annotation findings:
  - What does the genome statics tab tell you about your annotation? Are the results surprising? You can explore the reference genome you used in VEuPathDB to help you assess the results. (For example, are you getting a reasonable number of genes? What about the GC content? Number of non-coding genes?)

## Pcoa-Pkno (PCOA)

Completed

This job was submitted **6 days ago** and ran for **about 3 hours**, finally finishing at **2020-09-09 15:33:07 UTC**.

Genome statistics	Result files	Orthology	Phylogeny	Synteny	Job parameters	Pipeline logs	Validator report	Value
Number of annotated regions/sequences								14
Number of genes								5011
Gene density (genes/megabase)								178.54
Number of coding genes								4943
Number of pseudogenes								533
Number of genes with function								4793
Number of pseudogenes with function								528
Number of non-coding genes								68
Number of genes with multiple CDSs								2693
Overall GC%								39.65
Coding GC%								41.89

- What does the “Result files” tab contain? What is an AGP file? What is a GFF3 file?

Genome statistics	Result files	Orthology	Phylogeny	Synteny	Job parameters	Pipeline logs	Validator report	Format	MD5	Size
	<a href="#">Pseudochromosome level genomic sequence</a>							FASTA		7.79 MB
	<a href="#">Pseudochromosome level gene annotations</a>							GFF3		5.44 MB
	<a href="#">Pseudochromosome layout</a>							AGP		618 Bytes
	<a href="#">Scaffold level genomic sequence</a>							FASTA		7.79 MB
	<a href="#">Scaffold level gene annotations</a>							GFF3		5.47 MB
	<a href="#">Scaffold layout</a>							AGP		825 Bytes
	<a href="#">Pseudochromosome level sequence and annotation</a>							EMBL		13.4 MB
	<a href="#">Gene Ontology function assignments</a>							GAF1		1.65 MB
	<a href="#">Protein sequences</a>							FASTA		3.99 MB

- What does the “orthology” tab display? How many predicted proteins from your new genome are in common with ones from the reference genome? How many are unique to yours? What do singletons represent (click on the singleton number to see what these genes are?)

- What does the phylogeny tab represent? Does it make sense?

## Pcoa-Pkno (PCOA)

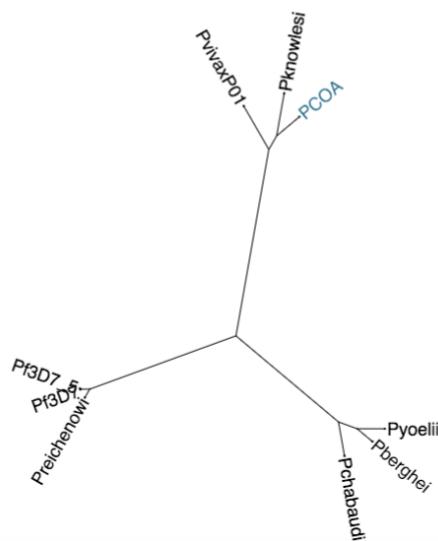
Completed

This job was submitted 6 days ago and ran for about 3 hours, finally finishing at 2020-09-09 15:33:07 UTC.

Genome statistics Result files Orthology Phylogeny Synteny Job parameters Pipeline logs Validator report

Click and drag in the diagram below to pan around. Use the mouse wheel to zoom in and out. The newly annotated genome in this job is highlighted: PCOA.

Rectangular Circular Radial Diagonal



- Examine the Synteny tab – are these genomes syntenic?

## Pcoa-Pkno (PCOA)

Completed

This job was submitted 6 days ago and ran for about 3 hours, finally finishing at 2020-09-09 15:33:07 UTC.

Genome statistics Result files Orthology Phylogeny Synteny Job parameters Pipeline logs Validator report

Each circle below represents a single target-reference pseudochromosome alignment. Click on the thumbnail to zoom in.

[Download all 14 images \(ZIP\)](#)



## Genetic Variation Exercises

### SNPs and CNVs

**Single Nucleotide Polymorphisms (SNPs):** single nucleotide changes between isolates or strains. SNPs have different functional effects with most having no consequential effect on gene function. SNPs may directly affect protein function when they are non-synonymous (results in a change in the amino acid; missense) or when they are cause a premature stop codon (nonsense). SNPs that do not fall within genes are non-coding (between genes or intronic). These types of SNPs may still affect splicing, mRNA stability, transcription, etc.

**Copy number variation (CNV):** variation in copy number of genes or regions of a genome. CNVs may be result of deletions or duplications.

#### Learning Objective:

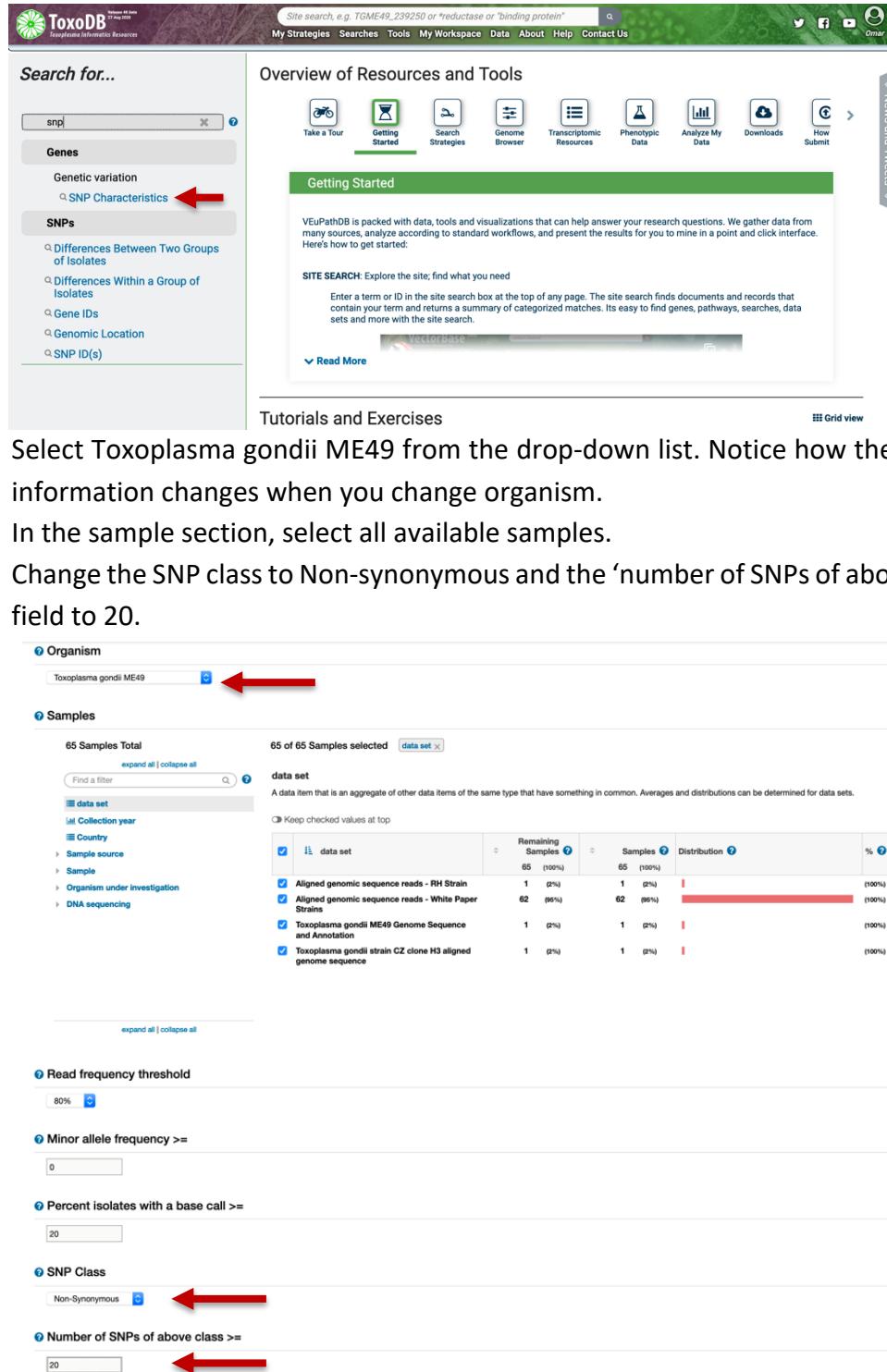
- Run SNP searches in VEuPathDB
- Explore SNP search parameters and their effect on search results
- Use SNP searches to identify genes that are under diversifying or stabilizing selection
- Run CNV searches in VEuPathDB
- Explore CNV search parameters
- Use CNV searches to identify regions of a genome that exhibit duplications or deletions.

### SNP Searches

In VEuPathDB SNPs can be used to characterize similarities and differences within a group of isolates or that distinguish between two groups of isolates. They can also be utilized to identify genes that may be under evolutionary pressure, either to stay the same (purifying selection) or to change (diversifying or balancing selection). Isolates are assayed for SNPs in VEuPathDB by two basic methods; re-sequencing and then alignment of sequence reads to a reference genome or DNA hybridization to a SNP-chip array (available in PlasmoDB only). In these exercises we'll explore both of these methods and ask a variety of questions to identify SNPs or genes of interest. If you do not understand the purpose of a parameter, please remember to mouse over the "?" icon and/or read the more detailed description at the bottom of the question page.

**1. Identify *T. gondii* genes that contain at least 20 nonsynonymous SNPs.**

- a.** Start by running a search for genes based on SNP characteristics – this search can be found under the ‘Genetic Variation’ category.



The screenshot shows the ToxoDB website interface. The top navigation bar includes links for Site search, My Strategies, Searches, Tools, My Workspace, Data, About, Help, Contact Us, and a user profile for 'Omar'. Below the navigation is a search bar with the placeholder 'Site search, e.g. TGME49\_239250 or "reductase or binding protein"'. On the left, a sidebar titled 'Search for...' contains a text input with 'snp' and a dropdown menu with options like 'Genes', 'Genetic variation' (which is selected), and 'SNPs'. A red arrow points to the 'SNP Characteristics' link under 'Genetic variation'. The main content area is titled 'Overview of Resources and Tools' and features a 'Getting Started' section with a green header. This section includes a 'SITE SEARCH' input field and a 'Read More' link. Below it is a 'Tutorials and Exercises' section with a 'Grid view' link. The bottom half of the page shows a detailed search results table for samples. The table has columns for 'Remaining Samples', 'Samples', and 'Distribution'. It lists several entries, including 'Aligned genomic sequence reads - RH Strain' (1 sample, 2%), 'Aligned genomic sequence reads - White Paper Strains' (62 samples, 96%), 'Toxoplasma gondii ME49 Genome Sequence and Annotation' (1 sample, 2%), and 'Toxoplasma gondii strain CZ clone H3 aligned genome sequence' (1 sample, 2%). The table also includes a 'data set' link and a note about averages and distributions. Below the table are several filter sections: 'Read frequency threshold' (set to 80%), 'Minor allele frequency >=' (set to 0), 'Percent isolates with a base call >=' (set to 20), 'SNP Class' (set to Non-Synonymous), and 'Number of SNPs of above class >=' (set to 20). Red arrows point to the 'Non-Synonymous' checkbox, the '20' input field for the SNP class count, and the '20' input field for the number of SNPs.

	Remaining Samples	Samples	Distribution
<input checked="" type="checkbox"/> <a href="#">data set</a>	65 (100%)	65 (100%)	(100%)
<input checked="" type="checkbox"/> <a href="#">Aligned genomic sequence reads - RH Strain</a>	1 (2%)	1 (2%)	(100%)
<input checked="" type="checkbox"/> <a href="#">Aligned genomic sequence reads - White Paper Strains</a>	62 (96%)	62 (96%)	(100%)
<input checked="" type="checkbox"/> <a href="#">Toxoplasma gondii ME49 Genome Sequence and Annotation</a>	1 (2%)	1 (2%)	(100%)
<input checked="" type="checkbox"/> <a href="#">Toxoplasma gondii strain CZ clone H3 aligned genome sequence</a>	1 (2%)	1 (2%)	(100%)

- e. How many genes did you return? Which gene has the highest number of non-synonymous SNPs? (hint: sort the non-synonymous SNP columns).

Unnamed Search Strategy \*

**Step 1:** SNPs 3,184 Genes + Add a step

**Step 2:** 3,184 Genes (2,934 ortholog groups) Revise this search

Gene Results | Genome View | Analyze Results

Organism Filter: select all | clear all | expand all | collapse all | Hide zero counts | Search organisms...

Gene ID	Transcript ID	Product Description	Chromosome	Total SNPs	Nonsynonymous SNPs	
TGME49_280660	TGME49_280660- t26_1	HECT-domain (ubiquitin-transferase) domain-containing protein	VIIa	2878	1417	846
TGME49_248510	TGME49_248510- t26_1	hypothetical protein	XII	1984	1054	628
TGME49_313630	TGME49_313630- t26_1	hypothetical protein	XI	1837	981	571

Download | Add to Basket | Add Columns | **Add a step**

- f. What happens if you revise this search and change the “Percent isolates with a base call >=” field to 100?
- g. How many of these genes have a predicted secretory signal peptide? (hint: add a step that identifies all genes with a signal peptide).
- h. What kinds of genes are in this result list? One way to determine if you have anything enriched in your results is to run an enrichment analysis. Click on the “Analyze Results” tab then compare the results you get from the GO enrichment and from the Word enrichment, we will discuss these results.

Unnamed Search Strategy \*

**Step 1:** SNPs 2,674 Genes + Add a step

**Step 2:** Signal Pep 564 Genes + Add a step

**Step 3:** SNPs 663 Genes + Add a step

**Step 4:** 663 Genes (591 ortholog groups) Revise this search

Gene Results | Genome View | New Analysis X

Organism Filter: select all | clear all | expand all | collapse all | Hide zero counts | Search organisms...

Analyze your Gene results with a tool below.

Gene Ontology Enrichment (GO)

Metabolic Pathway Enrichment

Word Enrichment: kinase, phosphatase, exported, membrane

**2. Identify SNPs that distinguish parasites with rapid clearance times following treatment with the anti-malarial drug Artesunate vs. those that have delayed clearance times.**

We have a published study in PlasmoDB (Takala-Harrison et. al.) with sufficient metadata about the samples to ask this interesting question.

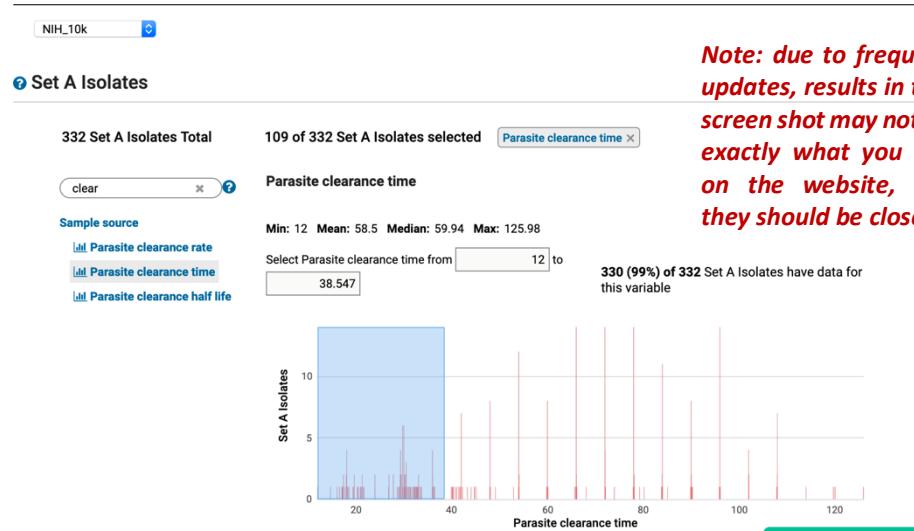
For this exercise use <http://PlasmoDB.org>

Navigate to the “Differences between two groups of isolates” search under “Search for SNPs (from Array).

- Unlike re-sequencing experiments that can identify any SNPs in the sequence, SNP-Chips have a pre-determined set of SNPs that are assayed and there are multiple different Chips on which these assays can be run. For this study, the authors used the NIH\_10K Chip, an array with approximately 10,000 SNPs of which ~8000 can be assayed. Choose this in the Isolate assay type parameter.
- Once this is done, an interesting set of characteristics are seen in the parameters to choose isolates. In addition to geographic location, there are clinical parameters like Clearance Time, Parasitemia levels, etc. In this exercise we want to identify SNPs that distinguish parasites with rapid clearance times from those with delayed clearance times but you could try other possibilities once you are finished. In Set A Isolates, click on some of the characteristics to explore the data. Then choose

The screenshot shows the PlasmoDB search interface. At the top, there is a search bar labeled "SNP". Below it is a sidebar with several search filters: "Differences Between Two Groups of Isolates" (selected), "Differences Within a Group of Isolates", "Gene IDs", "Genomic Location", and "SNP ID(s)". On the right, there is a section titled "SNPs (from Array)" containing the same filter options. A red arrow points to the "Differences Between Two Groups of Isolates" option in this section.

### Identify SNPs (from Array) based on Differences Between Two Groups of Isolates



*Note: due to frequent updates, results in this screen shot may not be exactly what you see on the website, but they should be close.*

Clearance Time and select 0 – 38 or 39 minutes. Do these rapid clearance samples appear to be evenly distributed geographically? Hint: click on Geographic Location to view the distribution of these selected samples (pink section of histogram).

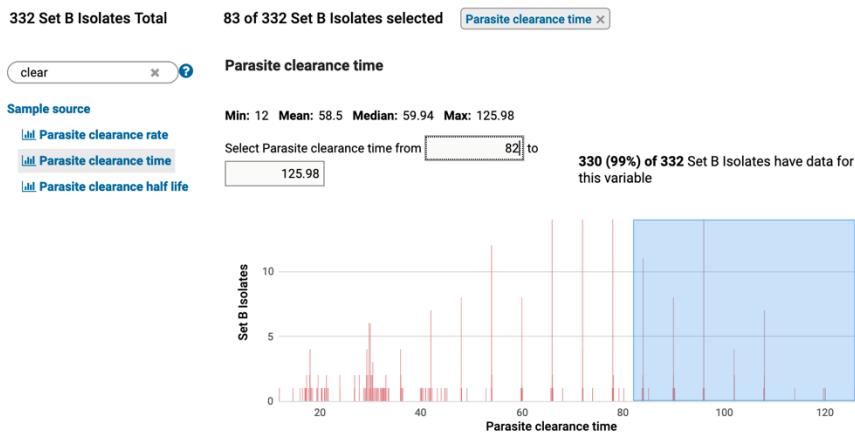
#### Country

Check items below to apply this filter      331 (>99%) of 332 Set A Isolates have data for this variable

	Country	Remaining Set A Isolates <small>?</small>	Set A Isolates <small>?</small>	Distribution <small>?</small>	% <small>?</small>
		109 (100%)	331 (100%)		
<input type="checkbox"/>	Bangladesh	85 (78%)	101 (31%)	<div style="width: 84%; background-color: #e67e22;"></div>	(84%)
<input type="checkbox"/>	Cambodia	15 (14%)	200 (60%)	<div style="width: 8%; background-color: #808080;"></div>	(8%)
<input type="checkbox"/>	Thailand	9 (8%)	30 (9%)	<div style="width: 30%; background-color: #804040;"></div>	(30%)

- c. We'll keep the defaults of 80 for both Major Allele Frequency and Percent Isolates with Call for this exercise.
- d. Now select Clearance times of 82 – end for Set B Isolates. Are these isolates geographically biased?

#### Set B Isolates



- e. Keep defaults for Major Allele and Percent with call and run the search. How many SNPs did you find?

A gene (Kelch13) has been identified that is involved in Artemesinin resistance in South East Asia. Is one or more of your SNPs in the region (+/- 10 KB) of the kelch13 gene? Note that we are not expecting that the SNP would be within the gene as this is a Chip experiment where the SNPs were pre-determined and there may not be a SNP on the array within a particular gene that we care about. However, if there is a haplotype that is being selected for in the presence of artemesinin, any SNPs within that haplotype (region of the genome) should likewise be selected.

*Hint: add a step to search for genes by text and search for kelch13. This will require you to use the genomic co-location operation as outlined in exercise 3. Set it up the same way except choose custom and start – 10000, stop + 10000 to define the region.*

- 3. Find SNPs that distinguish *Toxoplasma gondii* strains isolated from chickens as compared to those isolated from cats. NOTE: This exercise in ToxoDB explores the hypothesis that we can identify SNPs/genes involved in *T. gondii* host preference.**

Navigate to “Identify SNPs based on Differences Between Two Groups of Isolates”.

- a. Click select set A isolates and select hosts from the left column. Check the chicken (*Gallus gallus*) box to select the 11 chicken isolates.

**Set A Isolates**

65 Set A Isolates Total		11 of 65 Set A Isolates selected		Host organism
<input type="checkbox"/> expand all	<input type="checkbox"/> collapse all	<input type="checkbox"/> Host organism	<input type="checkbox"/> Find a variable	<input type="checkbox"/>
		<input type="checkbox"/> Keep checked values at top		
		59 (91%) of 65 Set A Isolates have data for this variable		
	Host organism	Remaining Set A Isolates	Set A Isolates	Distribution
	<input type="checkbox"/> Host organism	59 (100%)	59 (100%)	(100%)
	<input type="checkbox"/> Canis lupus familiaris	1 (2%)	1 (2%)	(100%)
	<input type="checkbox"/> Capra hircus	1 (2%)	1 (2%)	(100%)
	<input type="checkbox"/> Felis catus	12 (20%)	12 (20%)	(100%)
	<input checked="" type="checkbox"/> Gallus gallus	11 (19%)	11 (19%)	(100%)
	<input type="checkbox"/> Homo sapiens	22 (37%)	22 (37%)	(100%)
	<input type="checkbox"/> Ovis aries	4 (7%)	4 (7%)	(100%)
	<input type="checkbox"/> Panthera onca	1 (2%)	1 (2%)	(100%)
	<input type="checkbox"/> Panthera tigris altaica	1 (2%)	1 (2%)	(100%)
	<input type="checkbox"/> Puma concolor couguar	1 (2%)	1 (2%)	(100%)
	<input type="checkbox"/> Ramphastidae	1 (2%)	1 (2%)	(100%)
	<input type="checkbox"/> Sus scrofa	2 (3%)	2 (3%)	(100%)

- b. Click select set B isolates and select hosts from the left column. Check the cat (*Felis catus*) box to select the 12 cat isolates.
- c. Let's run a very stringent search and change the “major allele frequency” parameters for both sets to 90. (What does that mean?). Also, set the isolates with base call parameter to 100 for both sets A and B.
- How many SNPs did your search return? Does this large number that distinguish these two fairly large groups of isolates surprise you?

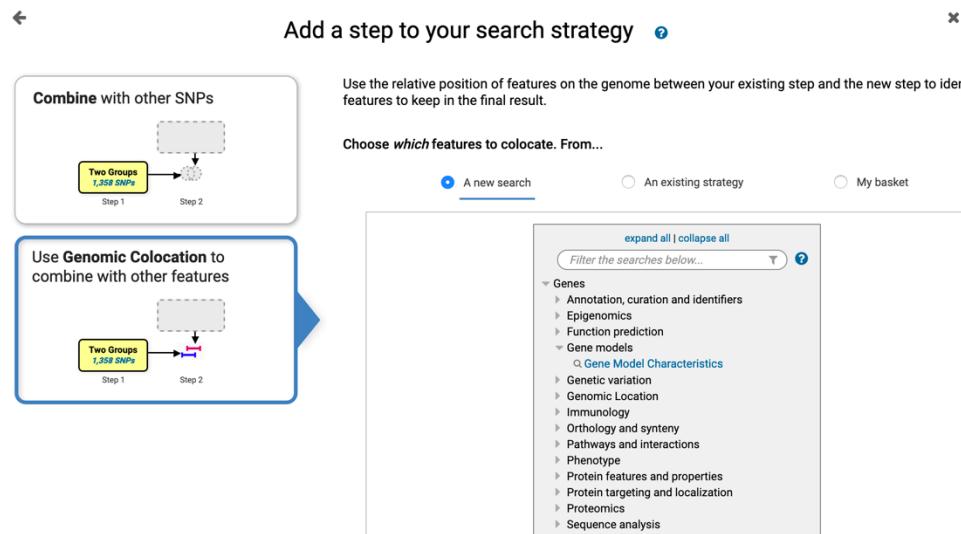
**Set B read frequency threshold >=**  
80%

**Set B major allele frequency >=**  
90

**Set B percent isolates with base call >=**  
100

You want to identify genes that could potentially be involved in host preference in *Toxoplasma gondii* and you expect that the SNPs from this search you just ran may be in protein coding regions of genes involved in this preference. How might you identify genes containing these SNPs?

- d. Add a step to identify *protein-coding genes* in *Toxoplasma gondii* ME49. Select the “Use Genomic Colocation...” option. Then select the gene search called “Gene Model Characteristics”.



- e. Configure the gene model characteristics search to find protein coding genes only.

Add a step to your search strategy ?

## ② Genes or Transcripts

Transcripts 

## ③ Gene Model Characteristics

255,437 Genes/Transcripts Total  
[expand all](#) | [collapse all](#)

Find a variable  

249,971 of 255,437 Genes/Transcripts selected  

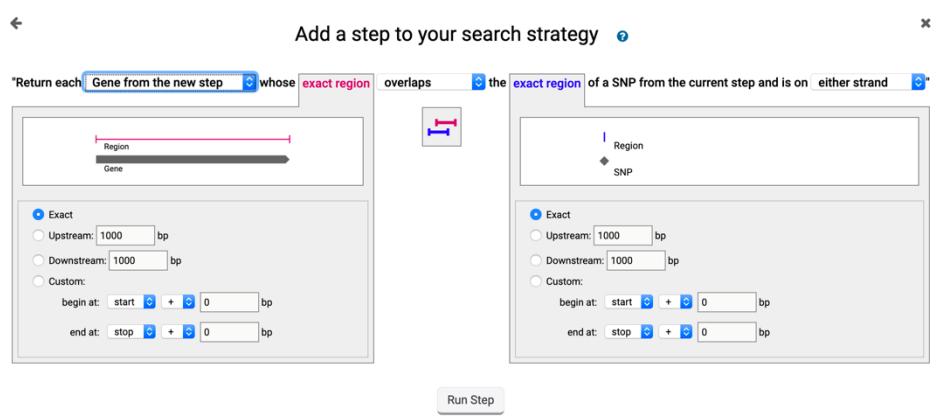
### Gene Type

Select gene type

Keep checked values at top 255,437 (100%) of 255,437 Genes/Transcripts have data for this variable

Gene Type	Remaining Genes/Transcripts	Genes/Transcripts	Distribution	%
<input checked="" type="checkbox"/> protein coding	249,971 (98%)	249,971 (98%)	<div style="width: 100%; background-color: #e6f2ff;"></div>	(100%)
<input type="checkbox"/> rRNA encoding	578 (< 1%)	578 (< 1%)	<div style="width: 10%; background-color: #ffccbc;"></div>	(100%)
<input type="checkbox"/> snoRNA encoding	2 (< 1%)	2 (< 1%)	<div style="width: 10%; background-color: #ffccbc;"></div>	(100%)
<input type="checkbox"/> snRNA encoding	18 (< 1%)	18 (< 1%)	<div style="width: 10%; background-color: #ffccbc;"></div>	(100%)
<input type="checkbox"/> tRNA encoding	4,868 (2%)	4,868 (2%)	<div style="width: 10%; background-color: #ffccbc;"></div>	(100%)

- f. Configure the genome colocation page to return “Gene from Step 2 whose exact region overlaps the exact region of a SNP in Step 1 and is on either strand”



- How many genes are returned?
- What is the gene that contains the most SNPs on your list? *Hint: sort the list high to low by match count.*
- Does this gene have orthologs in other species from ToxoDB? *Hint: go to the gene page and look at the genomic context and orthologs/paralogs in ToxoDB table.*
- Does it have orthology in any other species? *Hint: click on the link under the orthologs table and look at in OrthoMCL.*
- What does this say about this gene? How can you follow up on what role this gene may be playing for the organism? *Hint: you are a biologist and will need to look at the data on the gene record page and interpret it based on your experience and intuition.*
- Do these genes appear to be randomly distributed along the genome? *Hint: click the “Genome View” tab to view the distribution.* If you are a *Toxoplasma* biologist, do you have any hypotheses why the distribution may be skewed?

As a last resort: <https://toxodb.org/toxo/im.do?s=4fe2f7409d4ba4d6>

#### 4. Identifying SNPs within a group of isolates

For this exercise use <http://TriTrypDB.org>

##### a. Go to the “Differences Within a Group of Isolates” search.

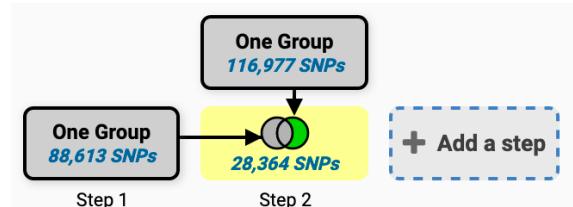
*Hint:* you can find this under the “SNPs” category (remember you can filter the searches by typing a key word like “snps” in the filter box.

The screenshot shows the search interface for identifying SNPs within a group of isolates. On the left, there's a sidebar with a search bar containing 'snps' and a 'Samples' section. A red arrow points from the sidebar to the 'Sample source' dropdown in the main search results area. The main area displays '252 Samples Total' and '208 of 252 Samples selected'. Under 'Host organism', it shows 'Homo sapiens' selected. Below this, there are sections for 'Read frequency threshold' (set to 40%), 'Minor allele frequency >=' (set to 0), and 'Percent isolates with a base call >=' (set to 20%).

##### b. What does this search do? Choose *Leishmania donovani* for the organism and select isolates from the human host. Use default parameters for the rest of the parameters.

Run the query and look at your results.

- How many SNPs were returned?
  - Are any of these heterozygous SNPs?
  - How would you identify heterozygous SNPs?  
Add a step to your strategy to identify SNPs from these isolates that may be heterozygous.
- Hint: choose a read frequency threshold of 40% and select the 2 minus 1 operation.*
- How many SNPs did you identify?



- Click on the second step results to view them. What do you notice about the %minor alleles? (*many are quite low ... i.e. in one or two of the isolates*). How can you remove these from your search results? *Hint: revise this search and increase the minor allele frequency threshold (try 20 and 40 and compare results).*

The screenshot shows the search interface with revised parameters: 'Read frequency threshold' set to 40%, 'Minor allele frequency >=' set to 40, and 'Percent isolates with a base call >=' set to 20. A 'Revise' button is located at the bottom right.

- Why might you want to increase the minor allele threshold when you run SNP searches?
- Try increasing / decreasing the “Percent isolates with base call”. How does this impact your results? Why might you want to change this parameter?
- Go to a record page for a SNP with a high minor allele frequency. What do you see in the Strains table? Why are many of the strains repeated?

## CNV Searches

### 5. Using resequencing data to identify regions of copy number variation (CNV)

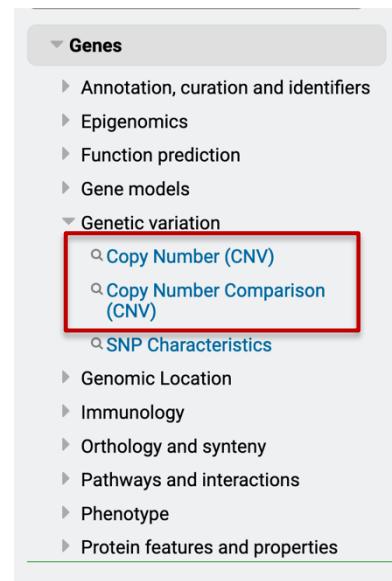
In addition to being useful for variant calling, high throughput sequencing data can be used for determining regions of copy number variation (CNV). All reads in ToxoDB are mapped to the same reference strain ME49, as a result we can estimate a gene’s copy number in each of the aligned strains.

The goal of this exercise is to identify

Gene searches taking advantage of sequence alignment data can be found under the under the “Genetic Variation” category. Two available searches that define regions of CNV are:

**Copy number:** This search returns genes that are present at copy numbers (haploid number or gene dose) within a range that you specify.

**Copy number comparison:** This search compares the estimated copy number of a gene in the re-sequenced strain with the copy number in the reference annotation. The copy number in the reference annotation is calculated as the number of genes that are in the same ortholog group as the gene of interest. We infer that these genes have arisen as a result of tandem duplication of a common ancestor.



You have the choice between two different metrics for defining copy number: **haploid number or gene dose**. Haploid number is the number of genes on an individual chromosome. Gene dose is the total number of genes in an organism, accounting for copy number of the chromosome. For example, a single-copy gene in a diploid organism has a haploid number of 1 and a gene dose of 2. You can choose to search for genes where at least one of your selected isolates

meets your cutoff criteria for the chosen metric (By Strain/Sample), or where the median of the chosen metric across all the selected isolates meets the cutoff (Median of Selected Strains/Samples)

Begin by choosing an Organism (reference genome) and one or more re-sequenced isolates. Choose whether you want to apply your search criteria to individual samples or to the median of your chosen samples. Then choose your Metric, Operator and Copy Number, and initiate the search by clicking the GET ANSWER button. Genes returned by the search will have a copy number based on your chosen metric within the range that you specified. For example, searching with the haploid number equal to 4 will return genes with 4 copies on a chromosome.

- Use the copy number search to identify genes that are present at a copy number greater than 5. Set up the copy number search to include all available isolates/strains, select the median of selected strains/samples, use Gene Dose for copy number metric and set the copy number to 5.

#### Identify Genes based on Copy Number (CNV)

**Organism**

Toxoplasma gondii ME49

**Strain/Sample**

64 Strain/Sample Total      64 of 64 Strain/Sample selected

Keep checked values at top      64 (100%) of 64 Strain/Sample have data for this variable

	Remaining Strain/Sam...	Strain/Sam...	Distribution	%
<input checked="" type="checkbox"/> Aligned genomic sequence reads - RH Strain	64 (100%)	64 (100%)		(100%)
<input checked="" type="checkbox"/> Aligned genomic sequence reads - White Paper Strains	62 (97%)	62 (97%)		(100%)
<input checked="" type="checkbox"/> Toxoplasma gondii strain CZ clone H3 aligned genome sequence	1 (2%)	1 (2%)		(100%)

**Median Or By Strain/Sample?**

Median of Selected Strains/Samples

**Copy Number Metric**

Gene dose

**Operator**

Greater than or equal to

**Copy Number**

4

How many genes did you get? Are any of these genes clustered in the same location? (*hint*: click on the “Genome view” tab and examine the red and blue lines in the gene location column – wider lines indicate more than one gene in that location, click on the line to view what is there).

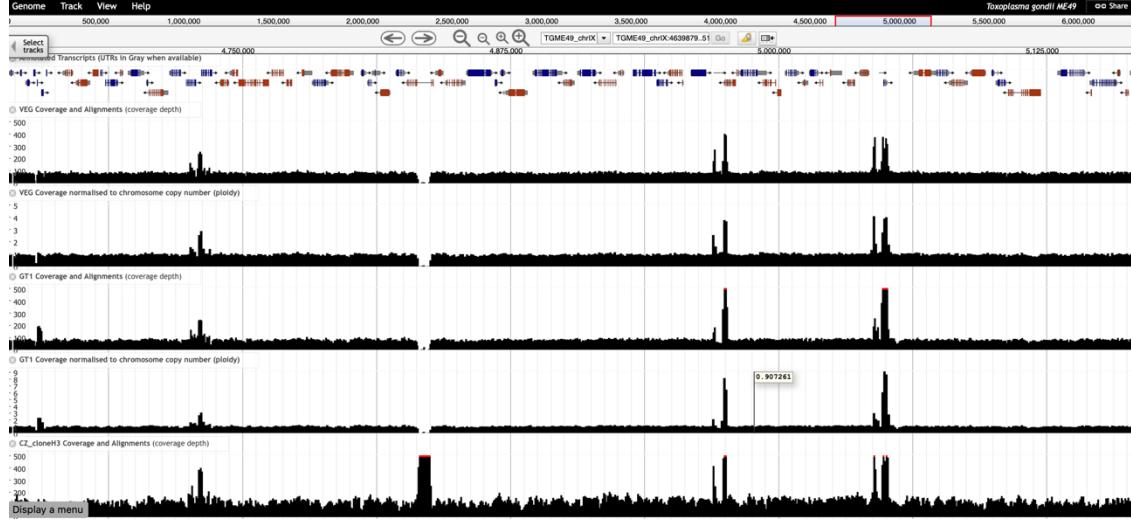
The screenshot shows the 'Gene Results' section of a search strategy. At the top, a yellow box highlights 'CopyNumber 164 Genes'. Below this, a table lists 164 genes across five chromosomes:

Sequence	Organism	Chromosome	#Genes	Length	Gene Locations
TGME49_chrVI	Toxoplasma gondii ME49	VI	32	3656745	
TGME49_chrXII	Toxoplasma gondii ME49	XII	24	7094428	
TGME49_chrX	Toxoplasma gondii ME49	X	20	7486190	
TGME49_chrIX	Toxoplasma gondii ME49	IX	17	6327655	
TGME49_chrV	Toxoplasma gondii ME49	V	16	3331915	

Below the table is a genome browser view for chromosome IX, showing gene locations (blue and red bars) and genomic tracks (black bars representing coverage depth). A legend indicates: Genes on forward strand (blue), Genes on reverse strand (red), Show empty chromosomes (unchecked), Rows per page: 20, and an up arrow icon.

What happens if you edit this step and change the “Median Or By Strain/Sample” parameter to “By Strain/Sample (at least one selected strain/sample meets criteria)?” Do you get more or less genes? Which genes have the highest CNV? (*hint*: sort the median gene dose column from highest to lowest). Is this what you expected? Does the coverage of reads from resequenced strains aligned to the reference support this conclusion? Here is a link to a JBrowse view with some of the resequenced strain coverage data turned on:

<https://tinyurl.com/y3mc53zm>



# Host Response

## Learning objectives:

- Exploring host responses by running a search strategy in HostDB
- Add steps in a search strategy
- Performing a GO enrichment analysis
- Revising steps in a search strategy

### 1. Find host genes that are upregulated in infected mouse cells compared to uninfected ones. For this exercise use <http://hostdb.org>

- a. HostDB has data from a published study that performed a comparative transcriptome analysis of 29 different strains of *Toxoplasma gondii* and the murine macrophages infected with them. We loaded the parasite component of the data in ToxoDB and the host component in HostDB. Go to HostDB.org and navigate to the “Transcriptomics” section then select “RNA Seq Evidence”. Select the fold change query for the “Mouse transcriptomes during infection with 29 strains of T gondii (Minot et al.)” experiment.

The screenshot shows the HostDB search interface. A red arrow points from the 'RNA-Seq Evidence' link in the sidebar of the first window down to the 'Fold Change' button in the second window.

**Search for...**

expand all | collapse all

Filter the searches below...

Genes

- Annotation, curation and identifiers
- Function prediction
- Gene models
- Genomic Location
- Orthology and synteny
- Pathways and interactions
- Protein features and properties
- Protein targeting and localization
- Proteomics
- Sequence analysis
- Structure analysis
- Taxonomy
- Text

Transcriptomics

- RNA-Seq Evidence

Organisms

Genomic Sequences

**Identify Genes based on RNA-Seq Evidence**

Filter Data Sets:

Legend: S Similarity DE Differential Expression FC Fold Change P Percentile SA SenseAntisense

Choose a Search

Organism	Data Set	Search Options
Bos taurus breed Hereford	Host cell transcriptome in bovine cells infected with Cryptosporidium parvum (Widmer et al.)	DE, FC, P
Bos taurus breed Hereford	Transcriptome of Bos taurus during infection with virulent and avirulent N. caninum strains (Horcajо et al.)	DE, FC, P, SA
Homo sapiens REF	HFF transcriptional response to virulent and avirulent T. cruzi (Blew et al. 2017)	DE, FC, P
Homo sapiens REF	Transcriptomes of 46 malaria-infected Gambian children (Lee et al.)	FC, P, SA
Homo sapiens REF	H. sapiens Transcriptome during infection with T. cruzi (Li et al.)	DE, FC, P
Homo sapiens REF	Leishmania major and Leishmania amazonensis RNA-Seq during human macrophage infection (Fernandes et al.)	DE, FC, P
Macaca mulatta isolate 17573	M. mulatta infected with P. cynomolgi over 100 days (Joyner et al.)	FC, P
Mus musculus C57BL/6J	Mouse transcriptome during infection with 29 strains of T. gondii (Minot et al.)	S, FC, P
Mus musculus C57BL/6J	Transcriptomes of 4 M. musculus cell types during infection with T. gondii (Swierzy et al.)	FC, P
Mus musculus C57BL/6J	Mouse transcriptome during early and late chronic infection with T. gondii (Garfoot et al.)	DE, FC, P
Mus musculus C57BL/6J	Transcriptional analysis of sorted subpopulations of mouse macrophages infected with C. albicans (Munoz et al.)	DE, FC, P
Mus musculus C57BL/6J	Transcriptome of mouse bone marrow derived macrophages infected by Wild-Type and grx18 mutant strains of T. gondii (He et al.)	FC, P
Mus musculus C57BL/6J	Transcriptomes of mouse macrophages infected with Leishmania mexicana (Fiebig et al.)	DE, FC, P

- b. Configure the search to return genes that are up-regulated at least 10-fold across all strains in the experiment compared to the uninfected control. Make sure to select upregulated. In the example below a fold change of 10 was selected and the “**average**” operation was applied on the comparison samples.

Identify Genes based on *M. musculus* C57BL6J Mouse transcriptome during infection with 29 strains of *T. gondii* RNA-Seq (fold change)

For the Experiment  
Mouse transcriptome during infection with 29 strains of *T. gondii* unstranded

return protein coding Genes  
that are up-regulated  
with a Fold change >= 10  
between each gene's average expression value  
(or a Floor of 10 reads)

in the following Reference Samples  
 TgCATB9 infected  
 TgCATB9 infected  
 VAND infected  
 VEG infected  
 WTD3 infected  
 Un-infected  
[select all](#) | [clear all](#)

and its average expression value  
(or the Floor selected above)  
in the following Comparison Samples  
 TgCATB9 infected  
 TgCATB9 infected  
 VAND infected  
 VEG infected  
 WTD3 infected  
 Un-infected  
[select all](#) | [clear all](#)

**Example showing one gene that would meet search criteria**  
(Dots represent this gene's expression values for selected samples)

Up-regulated

Average Expression Value Comparison  
Expression Value Reference

A maximum of four samples are shown when more than four are selected.

For each gene, the search calculates:  

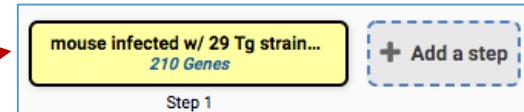
$$\text{fold change} = \frac{\text{average expression value in comparison}}{\text{reference expression value}}$$

and returns genes when fold change  $\geq 10$ .

You are searching for genes that are up-regulated between one reference sample and at least two comparison samples.

To narrow the window, use the minimum comparison value. To broaden the window, use the maximum comparison value.

[Get Answer](#)



- c. What are the functional characteristics of the genes in this result? What kinds of GO terms are enriched? Does the host immune response appear to be turned on? Is there a particular cellular location that is common in this group of genes?  
*Hint:* click on the “Analyze Results” tab and perform a GO enrichment analysis for the biological process ontology.

Gene Ontology Enrichment																																																																																
Find Gene Ontology terms that are enriched in your gene result. <a href="#">Read More</a>																																																																																
Parameters																																																																																
Analysis Results:																																																																																
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- d. Expand the result set to include human orthologs/paralogs of these genes. Hint: add a “Transform by Orthology” step choosing Homo sapiens.



- e. Does this set of human genes also show enriched GO terms? What, if any, are the enriched GO terms?
- f. Do any of these human genes also have peptide evidence for their expression during infection? Hint: add a step and explore the proteomics data “Human Proteome During T. gondii infection”

### Experiments and Samples

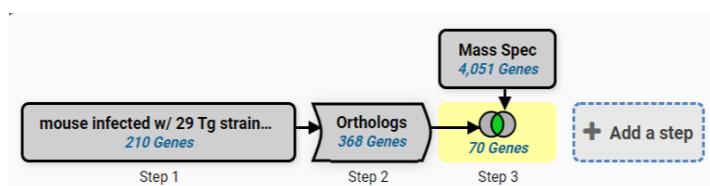
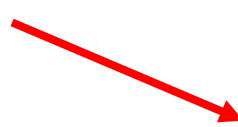
Note: You must select at least 1 values for this parameter.  
7 selected, out of 14

[select all](#) | [clear all](#) | [expand all](#) | [collapse all](#)

[Filter list below...](#) [?](#)

- [Homo sapiens](#)
- [Homo sapiens REF](#)
  - [Human Proteome During Infection with 4 strains of T. gondii and one strain of N. caninum \(Wastling\)](#)
    - 16 hour infection of H.sapiens cells (GT1)
    - 16 hour infection of H.sapiens cells (ME49)
    - 16 hour infection of H.sapiens cells (VEG)
    - 36 hour infection of H.sapiens cells (RH)
    - 36 hour infection of H.sapiens cells (ncLIV)
    - 44 hour infection of H.sapiens cells (ME49)
    - 44 hour infection of H.sapiens cells (VEG)
  - [Giardia secretome IEC Infection \(Maayeh et al.\)](#)
  - [Human Erythrocyte Phosphoproteome during infection with P. falciparum 3D7 schizonts \(2012\) \(Lasonder et al.\)](#)
    - Enriched schizont phospho-proteins (2012)

[select all](#) | [clear all](#) | [expand all](#) | [collapse all](#)



## 2. Identify host genes that are differentially regulated in multiple infection models (e.g. different hosts and parasites).

- Go to the RNAseq searches in HostDB. How many different pathogen infections are available? How many host organisms?
- Start by running a search using the “M. musculus C57BL6J Transcriptomes of 4 M. musculus cell types during infection with T. gondii (Swierzy et al.)”. Identify all genes that are differentially regulated (up or down) in all infected cell types compared to uninfected cell types.

Identify Genes based on M. musculus C57BL6J Transcriptomes of 4 M. musculus cell types during infection with T. gondii RNA-Seq (fold change)

For the Experiment  
Transcriptomes of 4 M. musculus cell types during infection with T. gondii unstranded

return protein coding Genes  
that are Up or down regulated  
with a Fold change >= 2  
between each gene's average expression value  
(or a Floor of 10 reads)  
in the following Reference Samples  
 SKBR3 Uninfected  
 SKMG3 Infected  
 Astrocytes Uninfected  
 Astrocytes Infected  
 Fibroblasts Uninfected  
 Fibroblasts Infected  
select all | clear all

and its average expression value  
(or the Floor selected above)  
in the following Comparison Samples  
 SKBR3 Uninfected  
 SKMG3 Infected  
 Astrocytes Uninfected  
 Astrocytes Infected  
 Fibroblasts Uninfected  
 Fibroblasts Infected  
select all | clear all

Example showing one gene that would meet search criteria  
(Dots represent this gene's expression values for selected samples)

Up or down regulated

For each gene, the search calculates:

$$fold\ change_{up} = \frac{average\ expression\ value\ in\ comparison}{average\ expression\ value\ in\ reference}$$

$$fold\ change_{down} = \frac{average\ expression\ value\ in\ reference}{average\ expression\ value\ in\ comparison}$$

and returns genes when  $fold\ change_{up} \geq 2$  or  $fold\ change_{down} \geq 2$ .

You are searching for genes that are up or down regulated between at least two reference samples and at least two comparison samples.

Get Answer

- How many of these results are also differentially regulated in a *Leishmania* infection model? Try adding a step and running a search for genes that are differentially **regulated** in “M. musculus C57BL6J Transcriptomes of mouse macrophages infected with Leishmania mexicana (Fiebig et al.)”.

Add a step to your search strategy

The results will be intersected with the results of Step 1.

Filter Data Sets:  Legend: S Similarity DE Differential Expression FC Fold Change P Percentile SA SenseAntisense

Organism: Mus musculus C57BL6J Data Set: Transcriptomes of mouse macrophages infected with Leishmania mexicana (Fiebig et al.) Choose a Search: DE FC P

Differential Expression Fold Change Percentile

For the Experiment  
Transcriptomes of mouse macrophages infected with Leishmania mexicana unstranded

return protein coding Genes  
that are Up or down regulated  
with a Fold change >= 2  
between each gene's average expression value  
(or a Floor of 10 reads)  
in the following Reference Samples  
 uninfected  
 infected  
select all | clear all

and its average expression value  
(or the Floor selected above)  
in the following Comparison Samples  
 uninfected  
 infected  
select all | clear all

Example showing one gene that would meet search criteria  
(Dots represent this gene's expression values for selected samples)

Up or down regulated

For each gene, the search calculates:

$$fold\ change_{up} = \frac{comparison\ expression\ value}{reference\ expression\ value}$$

$$fold\ change_{down} = \frac{reference\ expression\ value}{comparison\ expression\ value}$$

and returns genes when  $fold\ change_{up} \geq 2$  or  $fold\ change_{down} \geq 2$ .

You are searching for genes that are up or down regulated between one reference sample and one comparison sample.

Run Step

- Continue adding steps from other pathogen infections. For example, try the “Host cell transcriptome in bovine cells infected with Cryptosporidium parvum”. How many results in common did you get? If your answer is zero, did you remember to transform these results from bovine to mouse?

Here is an example search strategy to explore:  
<https://hostdb.org/hostdb/im.do?s=4d0a7299510641cf>

Step	Search Query	Number of Genes
Step 1	M mus Infect w/ Tgmn RNA-Seq	47 Genes
Step 2	Minus Infected w/ Lmex RNA-S...	1,390 Genes
Step 3	Orthologs	850 Genes
Step 4	Orthologs	871 Genes

**Gene Results**

Gene ID	Transcript ID	Genomic Location (Gene)	Product Description	# Transcripts
ENSMUSG00000029380.11	ENSMUST0000031327	MmusC5BL6J.chr5:90,891,241..90,893,115(+)	chemokine (C-X-C motif) ligand 1	2
ENSMUSG00000029380.11	ENSMUST00000201245	MmusC5BL6J.chr5:90,891,241..90,893,115(+)	chemokine (C-X-C motif) ligand 1	2

### 3. Identify host genes from a Plasmodium infection that are phosphorylated, secreted and have similarity to a 3D structure in the PDB database.

- To do this you can start by looking at the available proteomics data. Can you find an experiment that identifies phosphoproteins?

### Identify Genes based on Mass Spec. Evidence

**Experiments and Samples**

Note: You must select at least 1 values for this parameter.  
1 selected, out of 14

select all | clear all | expand all | collapse all

Homo sapiens  
 Homo sapiens REF

- >  Human Proteome During Infection with 4 strains of *T. gondii* and one strain of *N. caninum* (Wastling)
- >  Giardia secretome IEC infection (Maayeh et al.)
- >  Human Erythrocyte Phosphoproteome during Infection with *P. falciparum* 3D7 schizonts (2012) (Lasonder et al.)
- >  Enriched schizont phospho-proteins (2012)

select all | clear all | expand all | collapse all

**Minimum Number of Unique Peptide Sequences**

1

**Apply min # peptide sequences / sample OR across samples**

Per Sample

- How many of these genes are also predicted to be secreted? To figure this out add a step and search for genes that have a secretory signal peptide.
- Using the same logic as above, add another step to identify any gene with similarity to any structure in the PDB database.

Add a step to your search strategy

**Combine with other Genes**

① Choose how to combine with other Genes

2 INTERSECT 3    2 UNION 3    2 MINUS 3    3 MINUS 2

② Choose which Genes to combine. From...

A new search    An existing strategy    My basket

**Transform into related records**

**Use Genomic Colocation to combine with other features**

**PDB 3D Structures**

Add a step to your search strategy

**Search for Genes by PDB 3D Structures**

The results will be  intersected with |  the results of Step 2.

**Organism**

Note: You must select at least 1 values for this parameter.  
4 selected, out of 4

select all |  clear all |  expand all |  collapse all  
 Filter list below...

Bovidae  
 Primates  
 Rodentia  
 select all |  clear all |  expand all |  collapse all

**With similarity to PDB Proteins from**

Archaea  
 Bacteria  
 Eukaryota - all  
 Eukaryota - only Metazoa  
 select all |  clear all

**BLAST P-value less than 10 to the**

**Signal Pep 30,833 Genes**

**PDB 3D Struc 60,617 Genes**

**Mass Spec 187 Genes**

**39 Genes**

**25 Genes**

**+ Add a step**

Step 1   Step 2   Step 3

# Metabolic Pathways

## Exploring pathways and compounds

**Note:** this exercise uses *PlasmoDB.org* as an example database, but the same functionality is available on all *VEuPathDB* resources.

### Learning objectives:

- Explore the metabolic pathways searches and visualization tools
- Search for a pathway using the name or pathway identifier
- Paint data onto pathway maps to explore:
  - a. Which enzymes in a pathway are present in different genera
  - b. How transcriptional abundance of enzymes in a pathway differs under experimental conditions
- Explore the compound search options

### 1. Find and explore the metabolic pathway for glycolysis.

For this exercise use <http://plasmodb.org>

Navigate to the search page for Identify Metabolic Pathways based on Pathway Name/ID.

- Find the metabolic pathway searches on the home page. You can look under “Metabolic Pathways” or use the search filter. You can find metabolic pathways based on the pathway name or identifier, or using genes or compounds involved in the pathway. Search for the **glycolysis** pathway using the Pathway Name/ID option.
- This search is equipped with a type-ahead function for finding the metabolic pathway name. Begin typing glycolysis and then choose the pathway name from the list that appears.

**Search for...**

expand all | collapse all

Filter the searches below... ?

▶ Genes  
▶ Organisms  
▶ Popset Isolate Sequences  
▶ Genomic Sequences  
▶ Genomic Segments  
▶ SNPs  
▶ SNPs (from Array)  
▶ ESTs  
▼ Metabolic Pathways  
     Compounds  
     Genes  
     Identifier (pathway, gene, compound, etc.)  
     Pathway Name/ID

→

**a. Examine the Glycolysis / Gluconeogenesis pathway.**

Identify Metabolic Pathways based on Pathway Name/ID

Pathway Source: Any

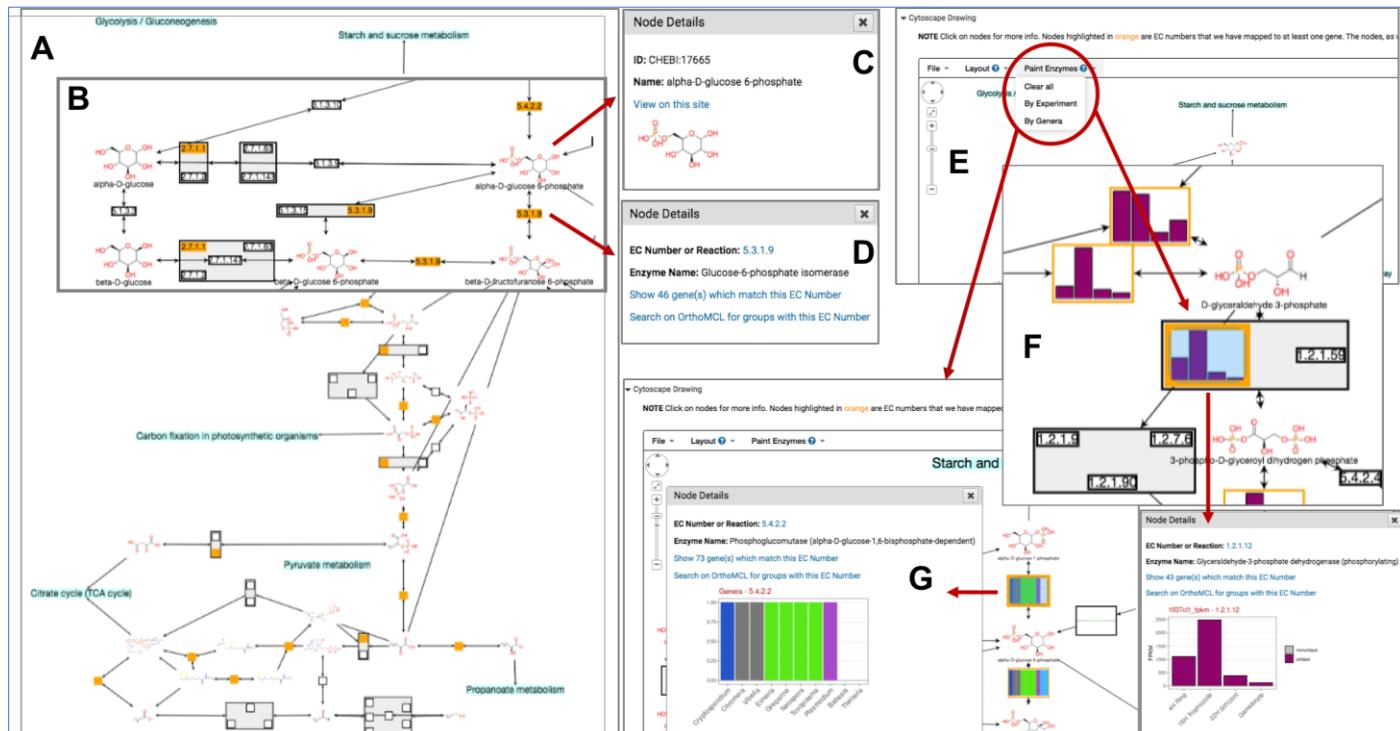
Pathway Name or ID: Glyco

Results:

- C-glycosylflavone biosynthesis I (PWY-6602) (MetaCyc)
- C-glycosylflavone biosynthesis II (PWY-7188) (MetaCyc)
- C-glycosylflavone biosynthesis III (PWY-7189) (MetaCyc)
- CMP-N-glycoloylneuraminate biosynthesis (PWY-6144) (MetaCyc)
- Glycolysis / Gluconeogenesis (ec00010) (KEGG)
- Glycosaminoglycan biosynthesis - chondroitin sulfate / dermatan sulfate (ec00532) (KEGG)
- Glycosaminoglycan biosynthesis - heparan sulfate / heparin (ec00534) (KEGG)
- Glycosaminoglycan degradation (ec00531) (KEGG)
- Glycosphingolipid biosynthesis - ganglio series (ec00604) (KEGG)
- Glycosphingolipid biosynthesis - globo and isoglobio series (ec00603) (KEGG)

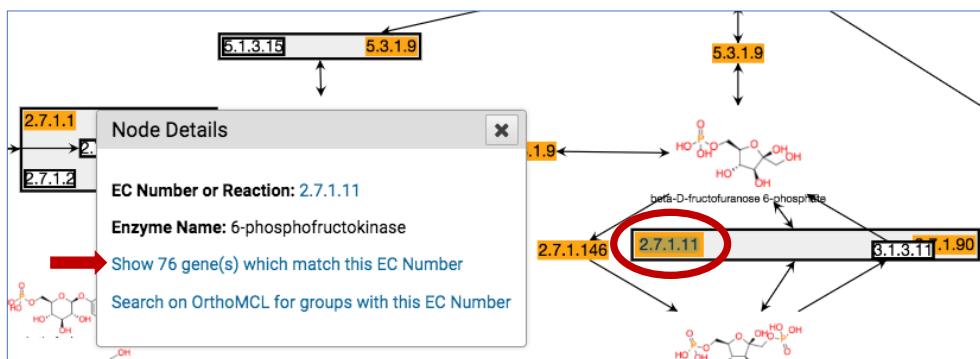
- The search takes you straight to the record page for the Glycolysis / Gluconeogenesis (ec00010) metabolic pathway from KEGG. The overview section of the record page contains an interactive graphical representation of the pathway. The pathway map and the legend can be repositioned.

- Initial pathway view is zoomed out.
- Zoom in to see more details including EC numbers and metabolite structures.
- Click on a metabolite structure to get additional information.
- Click on the EC number to get more info about the enzyme including links to retrieve all genes in the database assigned to this EC number.



- E. The drop-down menu under the heading “Paint Enzymes” allows you paint the pathway based on experimental data or phyletic pattern.
- F. Painting pathway by experiment provides a graphical representation of experimental results. Click on the graph to see more details.
- G. Painting pathway based on phyletic pattern provides a graphical representation of phyletic distribution. Clicking on the phyletic pattern graphic provides additional information.

- Use the Tool Box to move (drag) the map and individual nodes. Zoom in and out to help explore the map.
- What do the rectangles with numbers like 2.7.1.11 represent?
- What is the difference between the rectangular nodes that are orange and those that are not?
- Why are some enzymes grouped?
- Find the node representing 6-phosphofructokinase (EC number = 2.7.1.11). You may need to zoom and reposition the map to find the node.
- Click on the 2.7.1.11 node to open a popup with information about this enzyme.



- How many genes in the database matched this EC number?
- Try the link ‘Search for Gene(s) by EC Number’. Where did you end up? What do the 76 genes in the result list represent? Is 6-phosphofructokinase unique to *P. falciparum*? Notice the two columns called “EC numbers” and “EC numbers from OrthoMCL”. What do these columns represent?

EC Number 76 Genes + Add a step Step 1

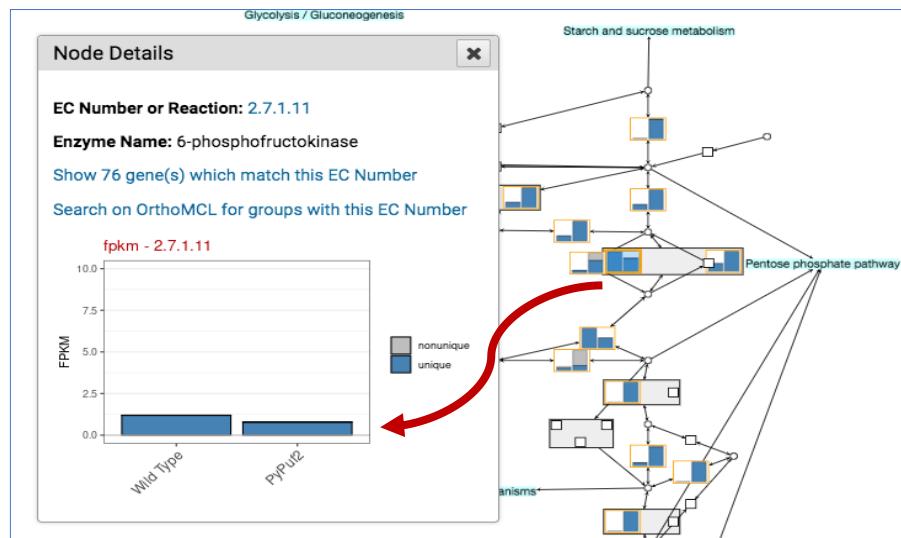
76 Genes (3 ortholog groups) Revise this search

Organism Filter select all | clear all | expand all | collapse all Hide zero counts Search organisms... ?

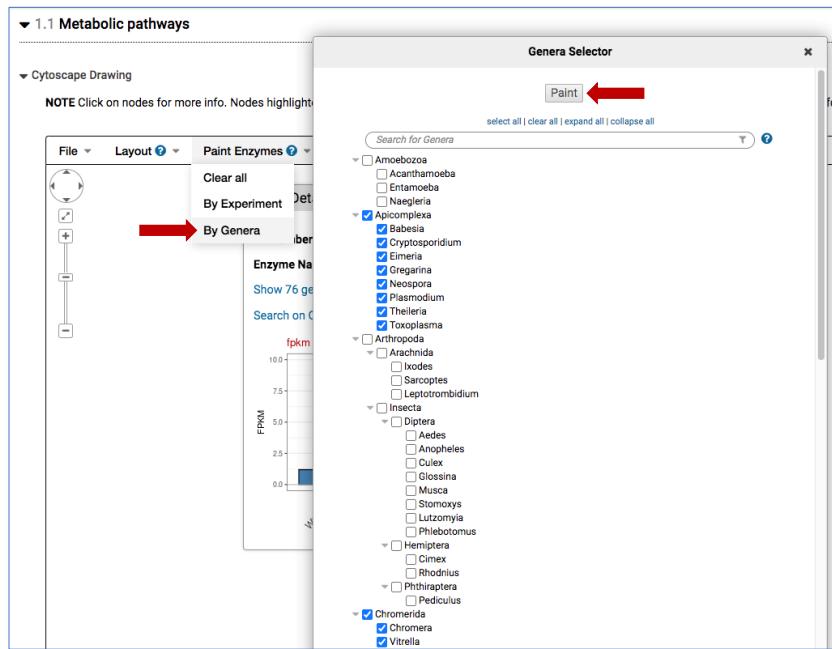
Gene Results Genome View Analyze Results Rows per page: 20 Download Add to Basket Add Columns

Gene ID	Transcript ID	Organism	Genomic Location (Gene)	Product Description	EC numbers	EC numbers from OrthoMCL
PADL01_1126600	PADL01_1126600-136_1	Plasmodium adleri	PADLG01_11:993,205,998,472(	6-phosphofructokinase	N/A	2.7.1.11 (6-phosphofructokinase)
PBANKA_0816400	PBANKA_0816400.1	Plasmodium berghei ANKA	PbANKA_08_v3:674,000..677,899(	ATP-dependent 6-phosphofructokinase, putative	2.7.1.90 (Diphosphate-fructose-6-phosphate 1-phosphotransferase)	
PBANKA_0919900	PBANKA_0919900.1	Plasmodium berghei ANKA	PbANKA_09_v3:737,188..741,888(+)	ATP-dependent 6-phosphofructokinase, putative	2.7.1.11 (6-phosphofructokinase)	
PBILCG01_1123600	PBILCG01_1123600-136_1	Plasmodium billcollinsi G01	PBILCG01_11:941,286,946,581(	6-phosphofructokinase	N/A	2.7.1.11 (6-phosphofructokinase)
PBLACG01_1126300	PBLACG01_1126300-136_1	Plasmodium blacklocki G01	PBLACG01_11:973,677..979,008(	6-phosphofructokinase	N/A	2.7.1.11 (6-phosphofructokinase)
PCHAS_0816700	PCHAS_0816700.1	Plasmodium chabaudi chabaudi	PCHAS_08_v3:703,597..707,517(	ATP-dependent 6-phosphofructokinase, putative	2.7.1.90 (Diphosphate-fructose-6-phosphate 1-phosphotransferase)	

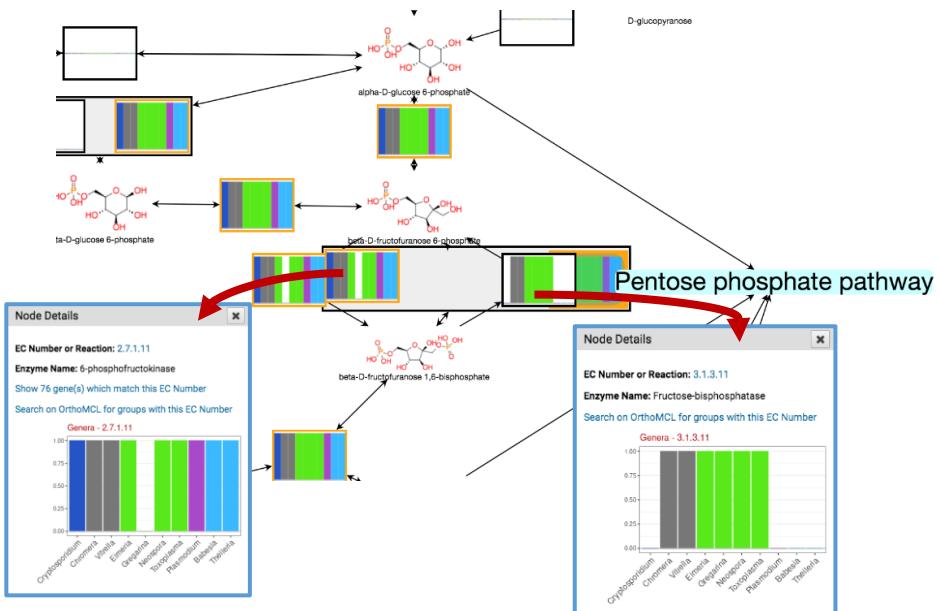
- Use your Browser's back button to return to the glycolysis pathway record page and open the Paint Experiment menu. Choose the experiment "Salivary gland sporozoite transcriptomes: WT vs Puf2-KO (Lindner et al)". Be patient while the graphs appear in place of the EC numbers.
- Does 6-phosphofructokinase appear to be expressed in salivary gland sporozoites? What enzymes in this pathway are affected in knockouts of Puf2?



- Use the Paint Genera option to determine whether 6-phosphofructokinase has orthologs across Apicomplexa and Chromerida.



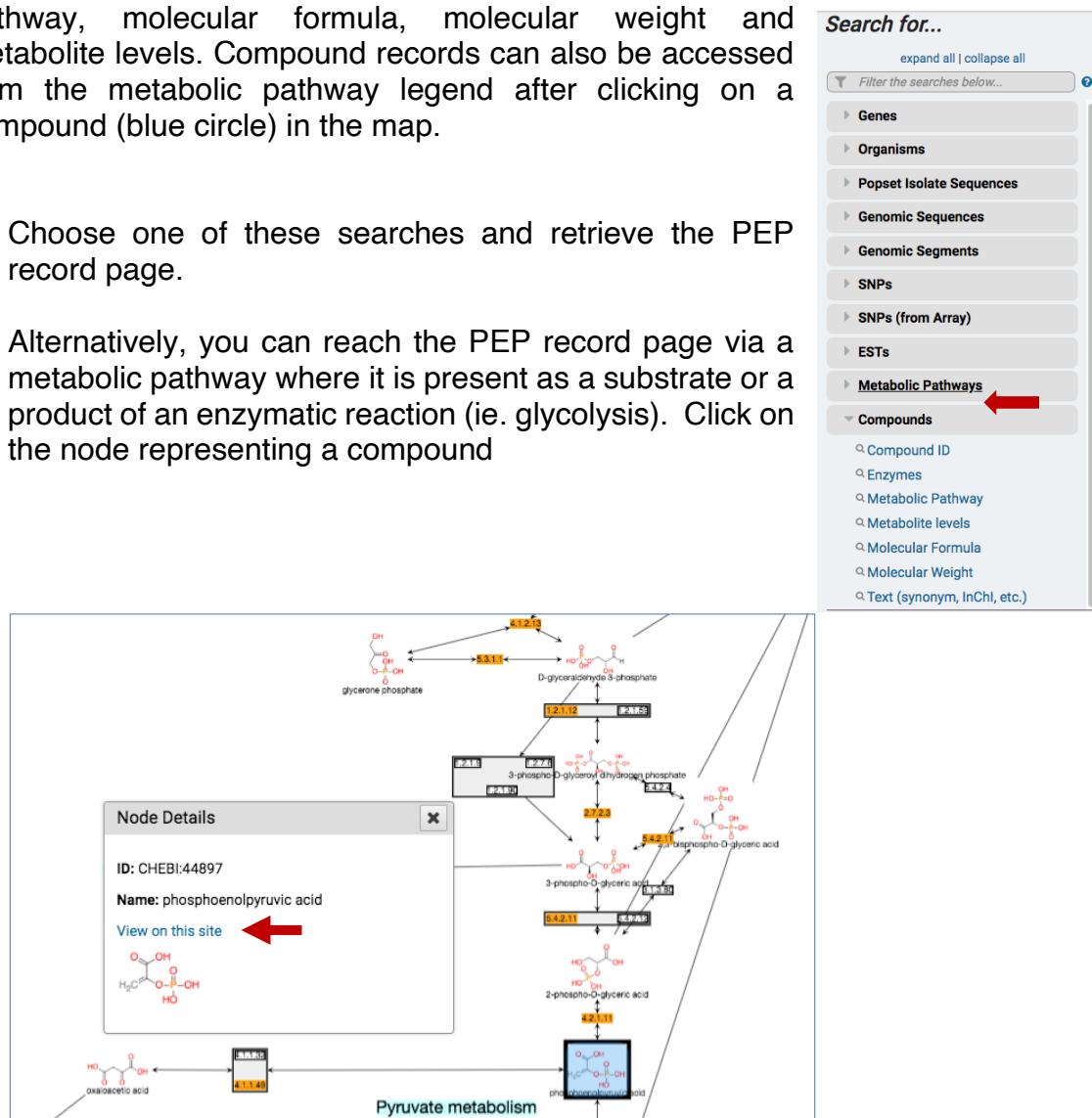
- What about the enzyme that catalyzes the reverse reaction (Fructose-bisphosphatase)?



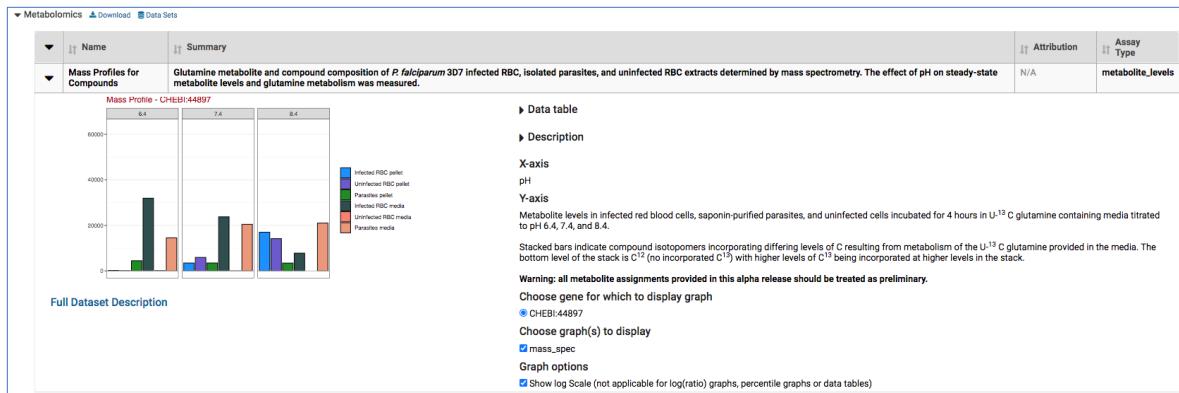
## 2. Find and explore the compound record page for phosphoenolpyruvate (phosphoenolpyruvic acid or PEP).

Compound records are accessed by running one of the compound searches available under the “Compounds” heading. Compounds may be retrieved by ID, text, metabolic pathway, molecular formula, molecular weight and metabolite levels. Compound records can also be accessed from the metabolic pathway legend after clicking on a compound (blue circle) in the map.

- Choose one of these searches and retrieve the PEP record page.
- Alternatively, you can reach the PEP record page via a metabolic pathway where it is present as a substrate or a product of an enzymatic reaction (ie. glycolysis). Click on the node representing a compound



- Which method did you use to get to the PEP record page? What compound name worked the best?
- Examine the PEP record page.
- What data sections do you see?
- Under which conditions is PEP present at highest concentrations? (Hint: navigate to the Metabolomics section)



10) The metabolite abundance experiment in PlasmoDB compares the following conditions at 3 pH levels:

- a) Parasites isolated from infected red blood cells using saponin lysis
  - b) Whole infected red blood cells isolated with Percoll
  - c) Whole uninfected red blood cells
- For both conditions, data was collected from the cell pellet and the media supernatant.

Find metabolites that are enriched in the isolated parasites (saponin) compared to infected red blood cells (Percoll) in the cell pellet at pH 7.4.

This can be done using the metabolite levels search, which looks a lot like the fold-change searches you have previously seen for transcriptomics data.

**Search for...**

expand all | collapse all

Filter the searches below...

- Genes
- Organisms
- Popset Isolate Sequences
- Genomic Sequences
- Genomic Segments
- SNPs
- SNPs (from Array)
- ESTs
- Metabolic Pathways
- Compounds

  - Compound ID
  - Enzymes
  - Metabolic Pathway
  - Metabolite levels ←
  - Molecular Formula
  - Molecular Weight
  - Text (synonym, InChI, etc.)

### Identify Compounds based on Metabolite levels

For the Experiment: Effect of pH on metabolite levels (Lewis, Baska and Llinás)

return compounds that are up-regulated with a Fold change >= 2 between each compound's maximum metabolite level in the following Reference Samples:

- infected RBC (Percoll) pH 6.4 pellet
- infected RBC (Percoll) pH 7.4 pellet
- infected RBC (Percoll) pH 8.4 pellet
- uninfected RBC pH 6.4 pellet
- uninfected RBC pH 7.4 pellet
- uninfected RBC pH 8.4 pellet

and its minimum metabolite level in the following Comparison Samples:

- uninfected RBC pH 6.4 pellet
- uninfected RBC pH 7.4 pellet
- uninfected RBC pH 8.4 pellet
- isolated parasites (saponin) pH 6.4 pellet
- isolated parasites (saponin) pH 8.4 pellet

Example showing one compound that would meet search criteria (dots represent this compound's metabolite levels for selected samples)

Up-regulated

Metabolite Level Comparison

Metabolite Level Reference

For each compound, the search calculates:  

$$\text{fold change} = \frac{\text{comparison metabolite level}}{\text{reference metabolite level}}$$

and returns compounds when fold change >= 2.

You are searching for compounds that are up-regulated between one reference sample and one comparison sample.

Get Answer

- How many compounds did you get?

207

- Add a step and use the same search to find out how many of these compounds (metabolites) are **NOT** enriched by 2-fold in isolated parasites (saponin) compared to the infected red blood cells (Percoll) in the media supernatant at pH 7.4. Make sure to use the correct operator!

**PlasmoDB** Release 48 beta 27 Aug 2020 Site search My Strategies

## My Search

Opened (1) All Unnamed Search

**Step 1**

**fold change**  
8 Compounds

**6 Metabolic Pathways**

Metabolic Pathway Results

Rows per page

Pathway

ec00230  
ec00240  
ec00790  
ec00515  
ec00740  
ec00670

Rows per page

**Identify Compounds based on Metabolite levels**

Revise your step

For the Experiment  
Effect of pH on metabolite levels (Lewis, Baska and Linas) ↗  
return compounds that are up-regulated ↗  
with a Fold change >= 2  
between each compound's maximum ↗ metabolite level  
in the following Reference Samples ↗

isolated parasites (saponin) pH 7.4 pellet  
 isolated parasites (saponin) pH 8.4 pellet  
 infected RBC (Percoll) pH 6.4 media  
 infected RBC (Percoll) pH 7.4 media  
 infected RBC (Percoll) pH 8.4 media

select all | clear all

and its minimum ↗ metabolite level  
in the following Comparison Samples ↗

uninfected RBC pH 7.4 media  
 uninfected RBC pH 8.4 media  
 isolated parasites (saponin) pH 6.4 media  
 isolated parasites (saponin) pH 7.4 media  
 isolated parasites (saponin) pH 8.4 media

select all | clear all

**Example showing one compound that would meet search criteria**  
(Dots represent this compound's metabolite levels for selected samples)

For each compound, the search calculates:  
$$\text{fold change} = \frac{\text{comparison metabolite level}}{\text{reference metabolite level}}$$

and returns compounds when fold change >= 2.

You are searching for compounds that are up-regulated between one reference sample and one comparison sample.

Revise

fold change  
10

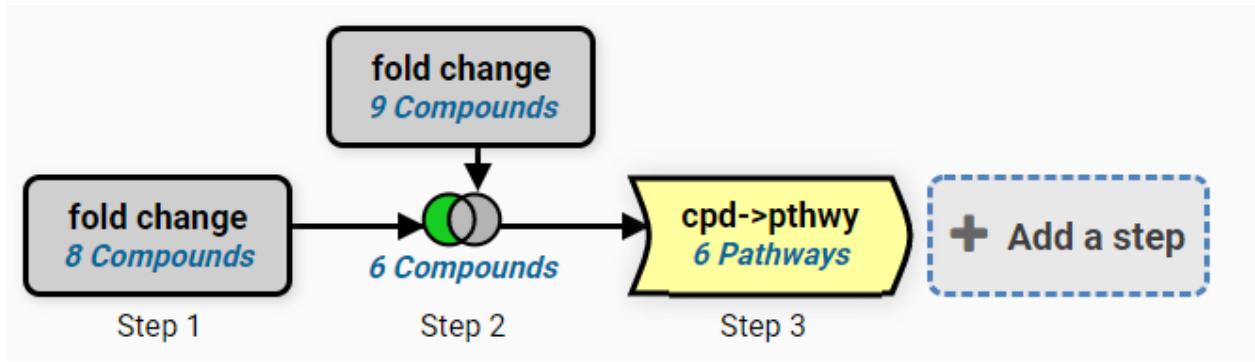
### Description

Find compounds with variation in metabolite levels. Note that when the compound has multiple isotopomers, the metabolite isotopomers levels were summed.

Infected RBCs were enriched using Percoll and parasites were isolated from infected RBCs with saponin. Intracellular metabolites were measured from pellets while secreted metabolites were measured from media. Infected RBCs, isolated parasites, and uninfected RBCs were incubated for 4 hours in RPMI containing U-13C glutamine at pH 6.4, 7.4, and 8.4. Extracts were analyzed by mass spectrometry. The effect of pH on steady-state metabolite levels and glutamine metabolism was measured.

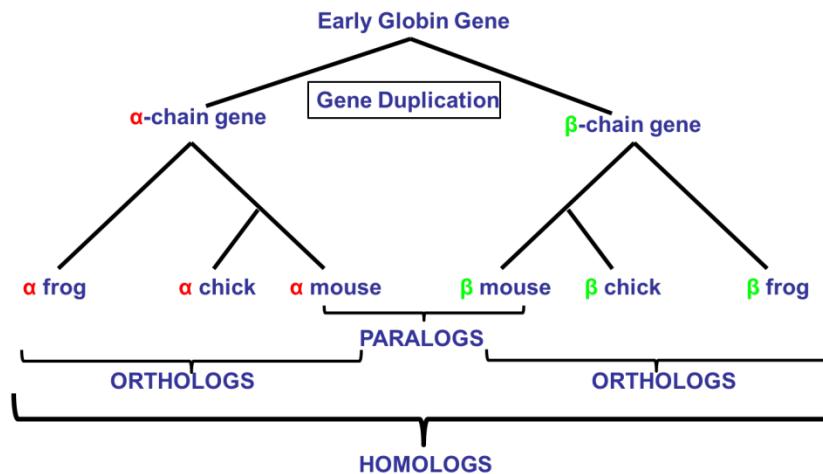
Warning: all metabolite assignments provided in this alpha release should be treated as preliminary.

How many compounds do you have now? Which metabolic pathways do these compounds belong to? Click Add a Step and transform the results to metabolic pathways.



## Orthology and Phyletic Patterns

### Homology



#### Learning objectives:

- Explore the orthology table on VEuPathDB gene pages
- Getting to OrthoMCL from VEuPathDB gene pages
- Run searches in OrthoMCL
- Explore the cluster graphs in OrthoMCL
- Leverage the phyletic pattern search
- Leverage the orthology transform tool

#### 1. Getting to OrthoMCL from VEuPathDB databases

Note: For this exercise use <http://cryptodb.org> and <http://orthomcl.org/>

- Go to the gene page for the *Cryptosporidium muris* gene with the ID: CMU\_034340

- b. What information on the gene page can you use to guess a function for this gene? It is annotated as a hypothetical protein! Hint: look at the orthologs table and the domains in the protein features graph. You may also want to visit some of the external links or take a look at InterPro domains.

▼ Proteins Properties and Features [Download](#) [Data sets](#)

	Transcript ID	Isoelectric Point	Molecular Weight	Has SignalP	Has TMHMM	Protein Length	Pro

## 7 Orthology and synteny

Ortholog Group [OG6\\_101337](#)

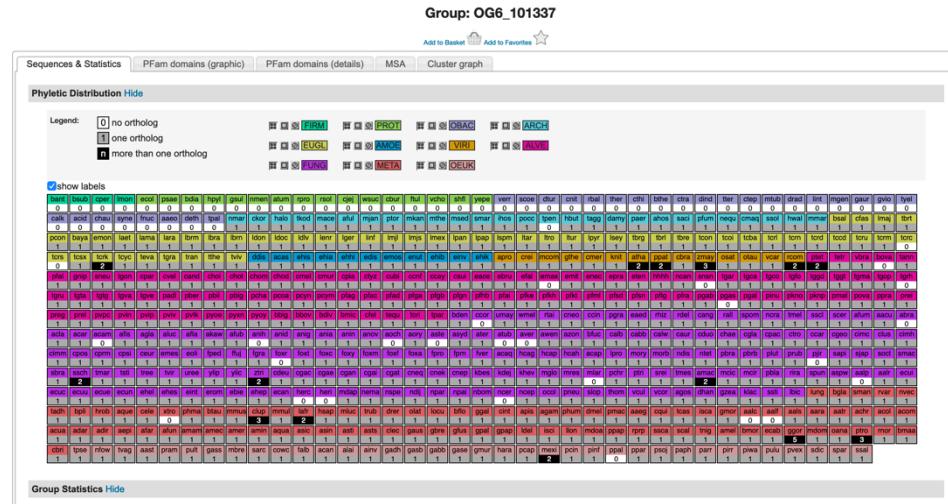
▼ Orthologs and Paralogs within CryptoDB [Data sets](#)

To run Clustal Omega, select genes from the table below. Then choose the sequence type and initiate the alignment with the 'Run Clustal Omega for selected genes' button.

Clustal Omega	Gene	Organism	Product	is syntenic	has comments
<input type="checkbox"/>	<a href="#">Cvel_467</a>	Chromera velia CCMP2878	rRNA-processing protein FCF1 homolog, putative	no	no
<input type="checkbox"/>	<a href="#">cand_030400</a>	Cryptosporidium andersoni isolate 30847	hypothetical protein	yes	no
<input type="checkbox"/>	<a href="#">Chro.70261</a>	Cryptosporidium hominis TU502	hypothetical protein	yes	no
<input type="checkbox"/>	<a href="#">CHUDEA7_2290</a>	Cryptosporidium hominis UdeA01	unspecified product	yes	no
<input type="checkbox"/>	<a href="#">GY17_00002025</a>	Cryptosporidium hominis isolate 30976	rRNA-processing protein Fcf1/Utp23	yes	no
<input type="checkbox"/>	<a href="#">ChTU502y2012_407q1140</a>	Cryptosporidium	Fcf1	yes	no

- c. Go to the Orthology and Synteny section and look at the table labeled "Orthologs and Paralogs within CryptoDB". Does this gene have orthologs in other *Cryptosporidium* species? What about other organisms? (hint: click on the Ortholog Group link above the table).

- d. What about orthologs in organisms not in VEuPathDB? (hint: click on the Ortholog Group link above the table). Does it have any orthologs in bacteria or archaea? (Hint: mouse over the colorful boxes in the table to reveal the full species and phylum names).



- e. Look at the PFAM domain architectures found under the PFam domains (graphic) tab. Do all the proteins in this group have similar domain architecture?
- f. Based on the orthologs, what do you think this protein might be doing? If you had to give this gene a name, what would you call it?

## 2. Using the phyletic pattern tool in OrthoMCL

Note: For this exercise use <http://orthomcl.org/>

How many protein groups in OrthoMCL do not have any orthologs in bacteria or archaea? (Hint: go to the “Phyletic Pattern” search in the Evolution section of the “Identify Ortholog Groups”

**Identify Groups based on Phyletic Pattern**

Find Ortholog Groups that have a particular phyletic pattern, i.e., that include or exclude taxa or species that you specify.

The search is controlled by the Phyletic Pattern Expression (PPE) shown in the text box. Use either the text box or the graphical tree display, or both, to specify your pattern. The graphical tree display is a friendly way to generate a pattern expression. You can always edit the expression directly. For PPE help see the [instructions at the bottom of this page](#).

In the graphical tree display:

- Click on  $\nearrow$  to show or hide subtaxa and species.
- Click on the  $\star$  icon to specify which taxa or species to include or exclude in the profile.
- Refer to the legend below to understand other icons.

Expression: BACT=0T AND ARCH=0T      Get Answer

Key:  $\circlearrowleft$  =no constraints |  $\checkmark$  =must be in group |  $\checkmark\text{!}$  =at least one subtaxon must be in group |  $\times$  =must not be in group |  $\diamond$  =mixture of constraints

Legend:  $\circlearrowleft$  Root (ALL);  $\circlearrowleft \times$  Bacteria (BACT);  $\circlearrowleft \diamond$  Archaea (ARCH);  $\circlearrowleft \checkmark$  Eukaryota (EUKA);

category). To specify a phyletic pattern click on the icon next to the taxonomic group or species to include or exclude it.

- a. How many protein groups do not contain orthologs from bacteria and archaea?
- b. Find all groups that contain orthologs from at least one species of *Cryptosporidium* and *Giardia* but not from bacteria or archaea. If you are getting frustrated trying to figure this one out, you have a right to be! You cannot answer this question by using the check boxes (we will discuss why). However, OrthoMCL has an added feature that allows you to enter an expression to define the phyletic pattern. This option provides additional flexibility. Can you figure out what expression to use to answer this question? (hint: scroll down to the bottom of the page to find additional information about expression parameters.)

Before looking at the answer below, try this on your own or with the people sitting next to you.

Find Ortholog Groups that have a particular phyletic pattern, i.e., that include or exclude taxa or species that you specify.

The search is controlled by the Phyletic Pattern Expression (PPE) shown in the text box. Use either the text box or the graphical tree display, or both, to specify your pattern. The graphical tree display is a friendly way to generate a pattern expression. You can always edit the expression directly. For PPE help see the [instructions at the bottom of this page](#).

In the graphical tree display:

- Click on -/+ to show or hide subtaxa and species.
- Click on the  icon to specify which taxa or species to include or exclude in the profile.
- Refer to the legend below to understand other icons.

Expression:

BACT=0T AND ARCH=0T AND cpar+cand+choi+chot+chom+chod+cmeI+cmur+cpia+ctyz+cubi>=1T AND gass+gadh+gasb+gabb+gase+gmur>=1T

All VEuPathDB sites also have a phyletic pattern search that uses OrthoMCL data under Genes > Evolution -> Orthology Phylogenetic Profile. This search is very useful to identify genes in your organism of interest that are restricted in their profile. For example, you frequently want to identify genes that are conserved among organisms in your genus but not present in the host as these genes may make good drug targets or vaccine candidates. Optional: go to your favorite VEuPathDB site and run this search to identify all genes that are not present in human or mouse.

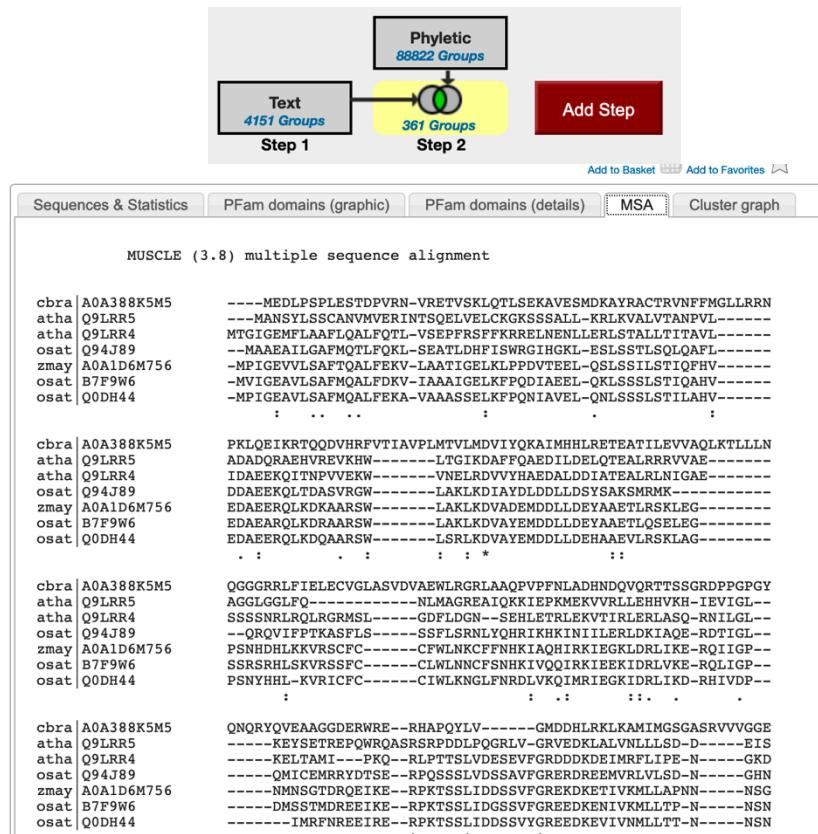
### 3. Combining searches in OrthoMCL (Use <http://orthomcl.org> for this exercise).

Find all plant proteins that are likely phosphatases that do not have orthologs outside of plants.

- a. Use the text search **to find OrthoMCL groups** that contain the word “\*phosphatase\*” (note that the search should be run without the quotation marks but with the asterisks).

The screenshot shows the OrthoMCL DB homepage with a search bar containing "phosphatase". Below the search bar, a navigation menu includes Home, New Search, My Strategies, My Basket (0), Tools, Data Summary, Downloads, and Community. A "My Favorites" button is also present. The main content area displays search results for "phosphatase", with a yellow box highlighting the "Text" step and its count of 4151 Groups. Buttons for "Add Step" and "Step 1" are visible. On the right, a context menu for the step includes options like Strategy, Rename, Duplicate, Save As, Share, and Delete. At the bottom, a summary shows 4151 Groups from Step 1 and a "Revise" button.

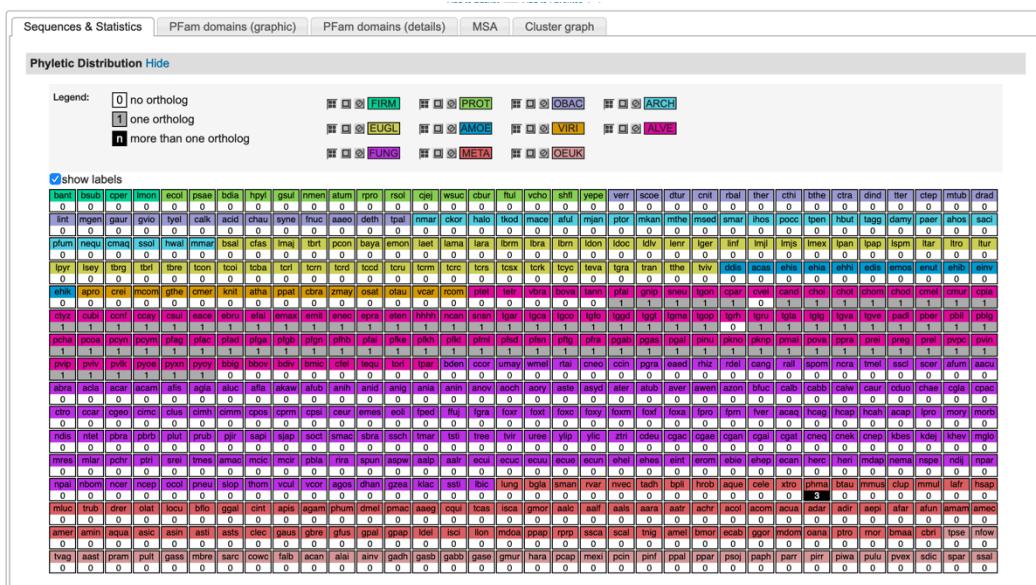
- b. Add a step and run a phyletic pattern search for groups that contain any plant protein but do not contain any other organism outside plants. (hint: make sure everything has a red x on it except for plants (Viridiplantae (VIRI)), which should be a grey circle).
  - c. How many groups did you return? Explore the multiple sequence alignments from some of these groups. (Hint: click on a group ID and open the MSA tab).



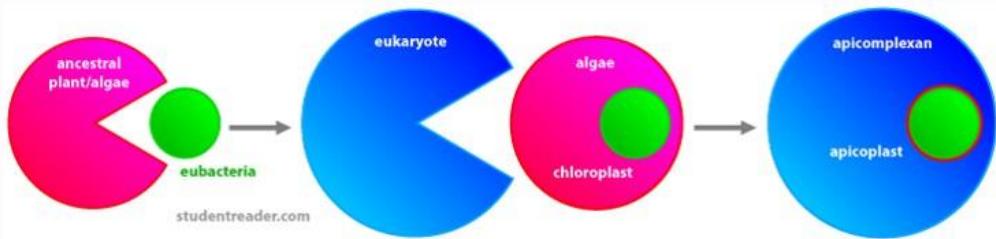
- \* Root (ALL):
- **X Bacteria (BACT):**
- **X Archaea (ARCH):**
- \* **Eukaryota (EUKA):**
  - **X Alveolates (ALVE):**
    - X Amoebozoa (AMOE):**
    - X Euglenozoa (EUGL):**
  - Viridiplantae (VIRI):**
    - Streptophyta (STRE):**
    - Chlorophyta (CHLO):**
    - Rhodophyta (RHOD):**
    - Cryptophyta (CRYP):**
    - Bacillariophyta (BACI):**
  - **X Fungi (FUNG):**
  - **X Metazoa (META):**
  - **X Other Eukaryota (OEUK):**

4. Exploring a specific OrthoMCL group - examining the cluster graph. (Use <http://orthomcl.org> for this exercise).

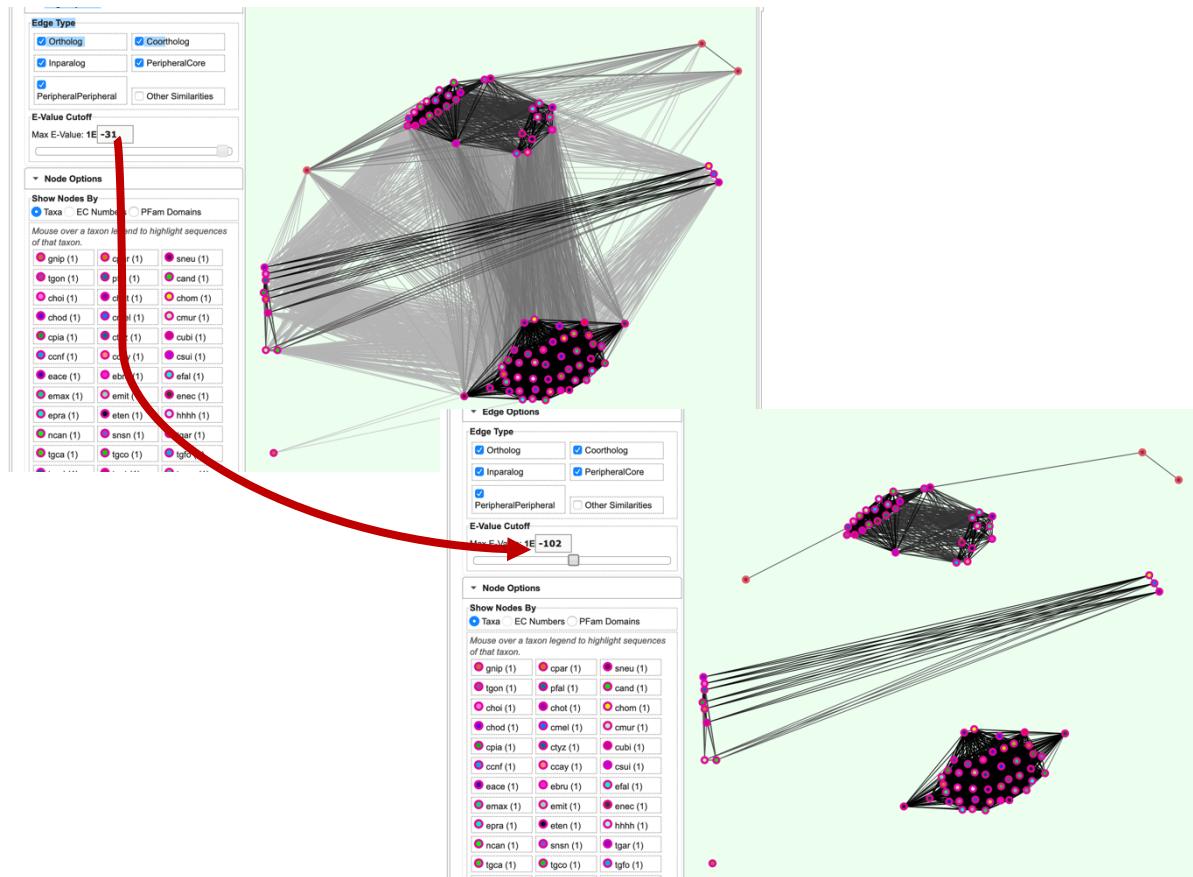
- a. Visit the orthomcl group OG6\_131670. You can either type the ID in the group quick search option at the top of the page or follow this link:  
[http://orthomcl.org/group/OG6\\_131670](http://orthomcl.org/group/OG6_131670)
  - b. Examine the "Sequences & Statistics" tab: Based on the product descriptions of the members of this group, what kind of proteins are in this group? What is the phylogenetic distribution of the members of this group?



- c. Examine the “Cluster Graph” tab: Modify the E-value cutoff slider. What happens when you increase or decrease the E-value? Can you identify subclusters?
5. Using the orthology transform tool to identify apicoplast targeted genes in *Toxoplasma* and *Neospora*.
- Note: For this exercise use <http://veupathdb.org>



The apicoplast likely became encased in four membranes via a double endosymbiotic event.



The chloroplast arose by engulfment of a cyanobacteria by a plant/algae ancestor. An algae was

then engulfed by the ancestor of all apicomplexans. Thus, an apicoplast organelle arose with four membranes.

- a. Start by finding genes in *Plasmodium* that are predicted to target to the apicoplast. Hint: click on “Protein targeting and localization” then on “P.f. Subcellular Localization”. You can further expand your list of potentially Apicoplast targeted proteins by running a GO terms search for the term “apicoplast” or the GO ID: GO:0020011 in *P falciparum* 3D7 (hint, click

The screenshot shows a search interface for a database of *Plasmodium* genes. On the left, a sidebar titled "Search for..." lists various categories: Pathways and interactions, Phenotype, Protein features and properties, Protein targeting and localization (which is expanded), Exported Protein, P.f. Subcellular Localization (which is highlighted in blue and has a red arrow pointing to it from the text above), Predicted Signal Peptide, Transmembrane Domain Count, and Proteomics. Below this is a "Get Answer" button. On the right, the main panel is titled "Identify Genes based on P.f. Subcellular Localization" and shows a dropdown menu set to "Apicoplast".

on add step the go to the function prediction category and select the GO term search). Which Boolean operation did you use? Union or intersect?

**Evidence**

---

Curated  
 Computed  
[select all](#) | [clear all](#)

**Limit to GO Slim terms**

---

Yes  
 No

```
graph LR; A[Subcell Loc  
513 Genes] --> B[GO Term  
2,297 Genes]
```

The diagram illustrates a process flow. A box labeled "Subcell Loc" containing "513 Genes" has an arrow pointing to a larger box labeled "GO Term" containing "2,297 Genes". Each box has an "Edit" button in the top right corner. The "Subcell Loc" box also has a "Step 1" label below it. The "GO Term" box has a "Step 2" label below it. To the right of the "GO Term" box is a dashed blue box with a plus sign and the text "Add a step".

GO:0020011 : apicoplast ; 6

- b. Transform the results of the above search to their *Toxoplasma* and *Neospora* orthologs.  
Hint: add a step, then select “Transform by Orthology”. On the search page , select all *Toxoplasma* and *Neospora*.

The screenshot shows a search interface with a sidebar on the left containing three search strategies:

- Combine with other Genes**: A diagram showing a green oval (GO Term) connected to a grey box (2,579 Genes) via arrows labeled "Step 2" and "Step 3".
- Transform into related records**: A diagram showing a green oval (GO Term) connected to a grey box (2,579 Genes) via arrows labeled "Step 1" and "Step 3". A red arrow points from this section to the main search results area.
- Use Genomic Colocation to combine with other features**: A diagram showing a green oval (GO Term) connected to a grey box (2,579 Genes) via arrows labeled "Step 2" and "Step 3".

The main search results area displays the following information:

**Transform 2,579 Genes into...**

**Orthologs**

**Add a step to your search strategy** ?

**Note: You must select at least 1 values for this parameter.**  
16 selected, out of 399

**select all | clear all | expand all | collapse all**

**Filter list below...** ?

- Amoebozoa
- Apicomplexa
- Aconoidasida
- Coccidia
  - Cryptosporidiidae
  - Eimeriidae
- Sarcocystidae
  - Cystoisospora
    - Cystoisospora suis* strain Wien I
  - Hammondiidae
    - Hammonia hammondi* strain H.H.34
- Neospora
  - Neospora caninum* Liverpool
- Sarco cystidae
  - Toxoplasma*
- Gregarinida
- Chromerida
- Fungi
- Heterolobosea
- Hexamitidae
- Kinetoplastida
- Metazoa
- Oomycetes
- Oxyomonadida
- Trichomonadida

**select all | clear all | expand all | collapse all**

Although *Cryptosporidium* is an apicomplexan parasite it has lost its apicoplast! Can you use this fact to refine your results from the above search? Hint: try subtracting out any orthologs present in *Cryptosporidium*. You will need to use a nested strategy and use the ortholog transform back to *Toxoplasma* and *Neospora* genes for the subtraction to complete.



## Add a step to your search strategy ?

The results will be subtracted from | ▼ the results of Step 3.

### ? Organism

Note: You must select at least 1 values for this parameter.  
11 selected, out of 399

[add these](#) | [clear these](#) | [select only these](#)  
[select all](#) | [clear all](#)

crypto

- Apicomplexa
- Coccidia
- Cryptosporidiidae
  - Cryptosporidium andersoni
  - Cryptosporidium andersoni isolate 30847
  - Cryptosporidium hominis
  - Cryptosporidium hominis TU502
  - Cryptosporidium hominis UdeA01
  - Cryptosporidium hominis isolate 30976
  - Cryptosporidium hominis isolate TU502\_2012
  - Cryptosporidium meleagridis
  - Cryptosporidium meleagridis strain UKMEL1
  - Cryptosporidium muris
  - Cryptosporidium muris RN66
  - Cryptosporidium parvum
  - Cryptosporidium parvum IOWA-ATCC
  - Cryptosporidium parvum Iowa II
  - Cryptosporidium tyzzeri
  - Cryptosporidium tyzzeri isolate UGA55
  - Cryptosporidium ubiquitum
  - Cryptosporidium ubiquitum isolate 39726

Opened (1) All (1) Public (20) Help

Unnamed Search Strategy \*



## Exploring Transcriptomic data

### 1. Exploring RNA sequence data in *Plasmodium falciparum*.

Note: For this exercise use <http://www.plasmodb.org>

- a. Find all genes in *P. falciparum* that are up-regulated during the later stages of the intraerythrocytic cycle.
- Use the fold change search for the data set “**Transcriptome during intraerythrocytic development (Bartfai et al.)**”. For this data set, synchronized Pf3D7 parasites were assayed by RNA-seq at 8 time-points during the iRBC cycle. We want to find genes that are up-regulated in the later time points (30, 35, 40 hours) using the early time points (5, 10, 15, 20, 25 hours) as reference.

The screenshot shows the Plasmodb.org search interface. On the left, a sidebar titled "Search for..." lists various categories under "Genes". A red arrow points from the text "Find all genes in *P. falciparum* that are up-regulated during the later stages of the intraerythrocytic cycle." to the "Genes" section. Another red arrow points from the text "Use the fold change search for the data set ‘Transcriptome during intraerythrocytic development (Bartfai et al.)’." to the search bar. The main search results page is titled "Identify Genes based on RNA-Seq Evidence". It shows a search bar with "development" and a dropdown menu with "Organism" selected. A legend at the top right includes "Differential Expression", "Fold Change", "Percentile", and "Sense/Antisense". Below the search bar, a list of datasets is shown, with "Plasmodium falciparum 3D7" highlighted. To the right, a "Choose a Search" section shows a grid of buttons for different search types, with "Fold Change" highlighted. The bottom half of the page displays a search form for "Identify Genes based on P. falciparum 3D7 Transcriptome during intraerythrocytic development RNA-Seq (fold change)". The form includes fields for "For the Expression" (set to "Up or down regulated"), "Reference Sample" (set to "5, 10, 15, 20, 25"), and "Fold Change" (set to "12"). To the right of the form is a "Help" section with a graph illustrating expression levels over time. A red box encloses the search form and the help section. A red arrow points from the text "Choose up-regulated" to the "Up or down regulated" field. Another red arrow points from the text "Choose 12" to the "Fold Change" field. A third red arrow points from the text "choose 5, 10, 15, 20, 25" to the "Reference Sample" field.

- There are a number of parameters to manipulate in this search. As you modify parameters on the left side note the dynamic help on the right side. See screenshots.
- **Direction:** the direction of change in expression. **Choose up-regulated**.
- **Fold Change>=** the intensity of difference in expression needed before a gene is returned by the search. **Choose 12** but feel free to modify this.
- **Reference Sample:** the samples that will serve as the reference when comparing expression between samples. **choose 5, 10, 15, 20, 25**
- **Between each gene's AVERAGE expression value:** This parameter appears once you have chosen two Reference Samples and defines the operation applied to reference samples.

Fold change is calculated as the ratio of two values (upregulated ratio = expression in comparison)/(expression in reference). When you choose multiple samples to serve as reference, we generate one number for the fold change calculation by using the minimum, maximum, or average. **Choose average**

- **(or a Floor of 10 reads):** This parameter defines a lower limit of aligned reads for a gene to avoid unreliable fold change calculations. (Low numbers of aligned reads means low expression but the low values may be technically inaccurate. Dividing by small numbers creates large numbers.  $2000\text{FPKM}/10 = 200$ ;  $2000/0.1 = 20,000$ ) If a gene has fewer than 10 aligned reads, it is assigned 10 reads before the fold change calculation is made. **Leave this as default at 10 reads.**
- **Comparison Sample:** the sample that you are comparing to the reference. In this case you are interested in genes that are up-regulated in later time points **choose 30, 35, 40**
- **And its AVERAGE expression value:** This parameter appears once you have chosen two Comparison Samples and defines the operation applied to comparison samples. See explanation above. **Choose average**

## Identify Genes based on P. falciparum 3D7 Transcriptome during intraerythrocytic development RNASeq (fold change)

[Tutorial](#) 

For the Experiment: Transcriptome during intraerythrocytic development scaled unstranded

return protein coding Genes  
that are up-regulated  
with a Fold change >= 12

between each gene's average expression value  
(or a Floor of 10 reads (1 FPKM))

in the following Reference Samples

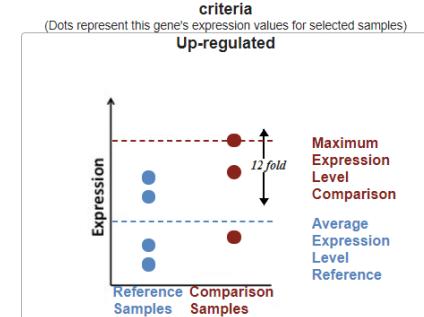
Hour 5  
 Hour 10  
 Hour 15  
 Hour 20  
 Hour 25  
[select all](#) [clear all](#)

and its maximum expression value  
(or the Floor selected above)

in the following Comparison Samples

Hour 20  
 Hour 25  
 Hour 30  
 Hour 35  
 Hour 40  
[select all](#) [clear all](#)

Example showing one gene that would meet search criteria  
(Dots represent this gene's expression values for selected samples)



A maximum of four samples are shown when more than four are selected.  
You are searching for genes that are up-regulated between at least two reference samples and at least two comparison samples.

For each gene, the search calculates:  

$$\text{fold change} = \frac{\text{maximum expression level in comparison}}{\text{average expression level in reference}}$$

and returns genes when fold change >= 12.  
To narrow the window, use the maximum reference value, or average or minimum comparison value. To broaden the window, use the minimum reference value.

See the detailed help for this search.

\* or FPKM Floor, whichever is greater

[Get Answer](#)

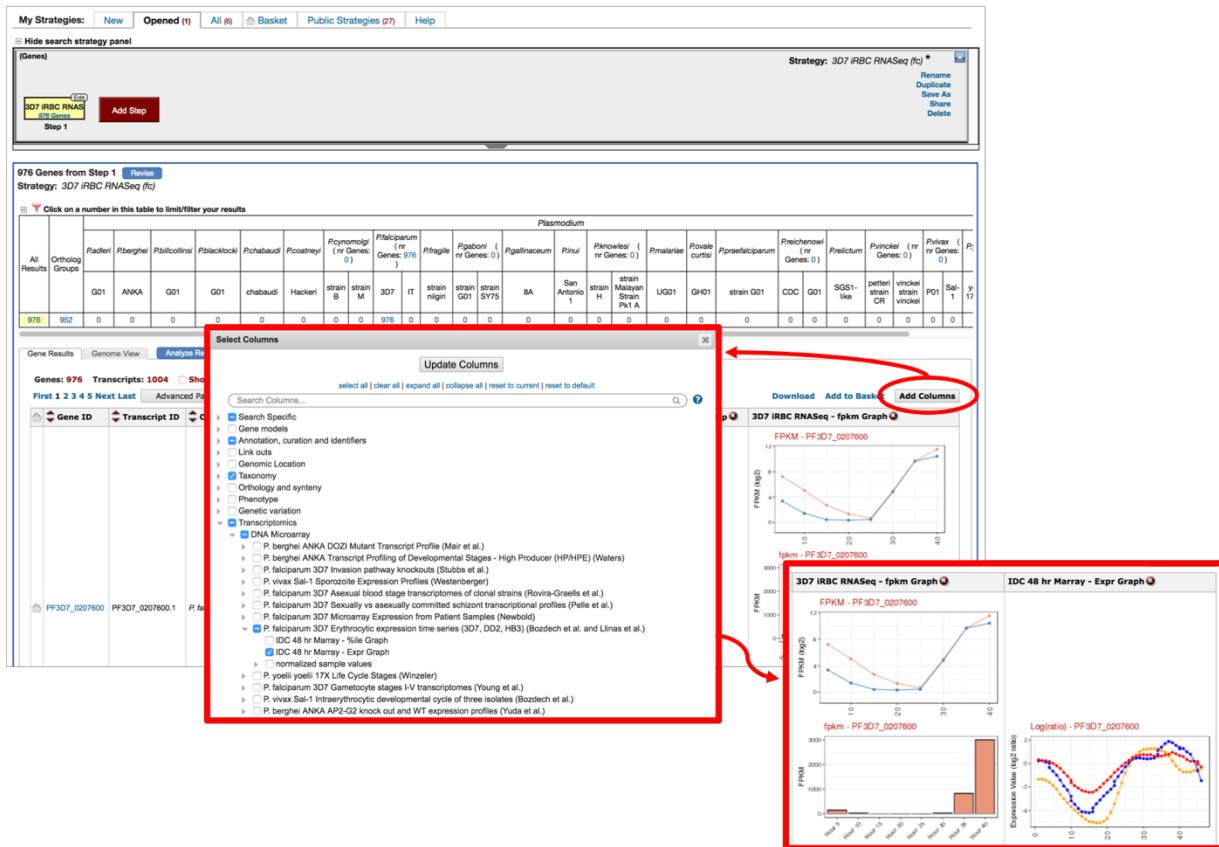
3D7 IRBC RNA-Seq (fc)  
969 Genes

+ Add a step

Step 1

b. For the genes returned by the search, how does the RNA-sequence data compare to microarray data?

- Hint: PlasmoDB contains data from a similar experiment that was analyzed by microarray instead of RNA sequencing. This experiment is called: **Erythrocytic expression time series (3D7, DD2, HB3) (Bozdech et al. and Linas et al.). IDC 48 hr Marray – Expr Graph** shows normalized expression values. To directly compare the data for genes returned by the RNA-seq search that you just ran, add the column called “Pf-iRBC 48hr - Graph”.



OPTIONAL: You can also run a fold change search using this experiment to compare results on a genome scale. Add a step to your strategy and intersect your current results (genes upregulated 12 fold in later IDC time periods) with a fold change search using the “Erythrocytic expression time series (3D7, Dd2, HB3) (Bozdech et al. and Linas et al.)” experiment (under microarray evidence). Configure it similarly to the RNA-seq experiment although you will probably need to make the fold change smaller (try 2 or 3) due to the decreased dynamic range of microarrays compared to RNA-seq.

Add a step to your search strategy [?](#)

Search for Genes by Microarray Evidence

The results will be [\(intersected with\)](#) the results of Step 1.

Filter Data Sets: [\(1\)](#) Legend: [Similarity](#) [Direct Comparison](#) [Fold Change](#) [Percentile](#)

[Organism](#) [Data Set](#) Choose a Search

[Plasmodium falciparum 3D7](#) [Erythrocytic expression time series \(3D7, D02, HB3\) \(Bozdech et al. and Llinás et al.\)](#)

[Plasmodium knowlesi strain H](#) [Intraerythrocytic cycle expression profile: in vitro and ex vivo \(Pkn6 Pk1\(A+\)\) \(Lapp et al.\)](#)

[Plasmodium vivax P01](#) [Intraerythrocytic developmental cycle of three isolates \(Bozdech et al.\)](#)

**Step 1**

**IDC 48 hr Marray (fc)  
489 Genes**

**Step 2**

**+ Add a step**

**Similarity** **Fold Change** **Percentile**

For the Experiment: [IRBC HB3 \(48 Hour scaled\)](#) [genes](#) return [protein coding](#) [genes](#) that are [up-regulated](#) [with a Fold change >= 2](#) between each gene's [average](#) [expression value](#) in the following [Reference Samples](#) [Note: You must select at least 1 values for this parameter.](#) [28 selected, out of 46](#) [select all | clear all | expand all | collapse all](#)

and its [average](#) [expression value](#) in the following [Comparison Samples](#) [Note: You must select at least 1 values for this parameter.](#) [18 selected, out of 46](#) [select all | clear all | expand all | collapse all](#)

**Up-regulated**

A maximum of four samples are shown when more than four are selected.

For each gene, the search calculates:

$$\text{fold change} = \frac{\text{average expression value in comparison}}{\text{average expression value in reference}}$$

and returns genes that are **up-regulated** between at least two **reference samples** and at least two **comparison samples**.

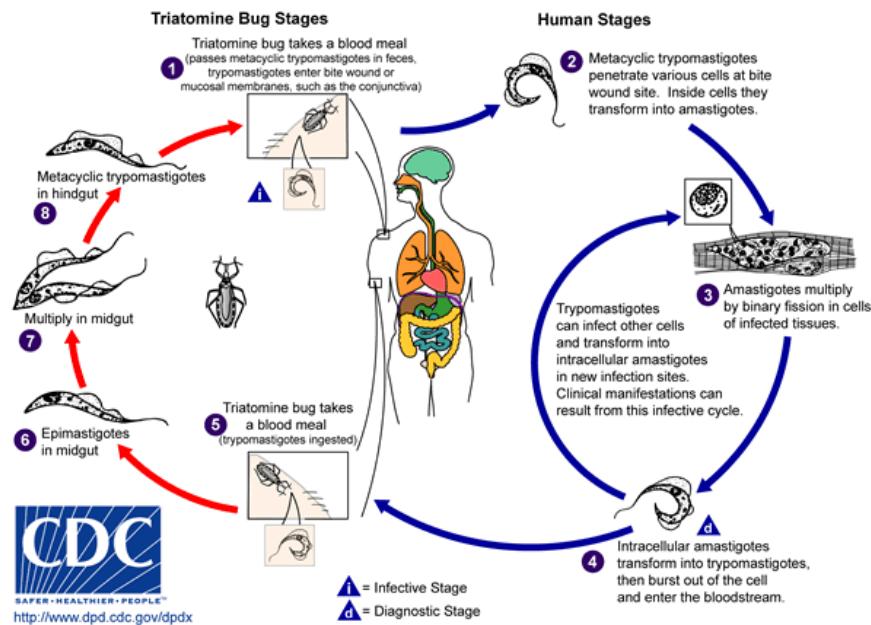
You are searching for genes that are **up-regulated** between at least two **reference samples** and at least two **comparison samples**.

To narrow the window, use the maximum reference value, or minimum comparison value. To broaden the window, use the maximum comparison value, or maximum reference value.

**Run Step**

## 2. Exploring microarray data in TriTrypDB.

Note: For this exercise use <http://www.tritrypdb.org>



- a. Find *T. cruzi* protein coding genes that are upregulated in amastigotes compared to trypomastigotes. Go to the transcript expression section then select **microarray**. Choose the fold change (FC) search for the data set called: **Transcriptomes of Four Life-Cycle Stages (Manning et al.)**.

**Identify Genes based on *T. cruzi* CL Brener Esmeraldo-like Transcriptomes of Four Life-Cycle Stages Microarray (fold change)**

**For the Experiment**  
Transcriptomes of Four Life-Cycle Stages tcruCLBrenerEsmeraldo-lif...  
return protein coding Genes  
that are up-regulated  
with a Fold change >= 2.0

between each gene's expression value in the following Reference Samples:  
 amastigotes  
 trypanomastigotes  
 epimastigotes  
 metacyclcs  
select all | clear all

and its expression value in the following Comparison Samples:  
 amastigotes  
 trypanomastigotes  
 epimastigotes  
 metacyclcs  
select all | clear all

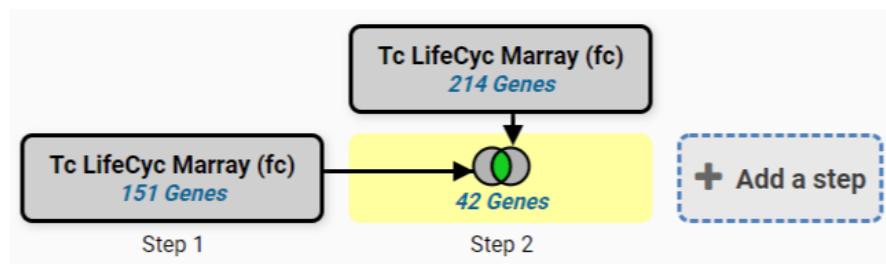
**Example showing one gene that would meet search criteria**  
(Dots represent this gene's expression values for selected samples)  
Up-regulated  
Comparison  
Reference  
Samples  
Comparison  
Samples  
2.0 fold  
Expression

You are searching for genes that are up-regulated between one reference sample and one comparison sample.  
For each gene, the search calculates:  
$$\text{fold change} = \frac{\text{comparison expression value}}{\text{reference expression value}}$$
  
and returns genes when fold change >= 2.0.  
See the detailed help for this search.

**Advanced Parameters**

**Get Answer**

- Select the direction of regulation, your reference sample and your comparison sample. For the fold change keep the default value 2.
- How many genes did you find? Do the results seem plausible?
- Are any of these genes also up-regulated in the replicative insect stage compared to the transmissive insect stage? How can you find this out? (*Hint*: add a step and run a microarray search comparing expression of epimastigotes to metacyclics).



- Do these genes have orthologs in other kinetoplastids? Transform your results into orthologs in all other organisms in TriTrypDB. This can be done by adding a step, or by editing a step, as shown in the screenshots.

Step 1: Tc LifeCyc Marray (fc)  
151 Genes

Step 2: Tc LifeCyc Marray (fc)  
214 Genes

+ Add a step

Details for step Combine Gene results

Revise as a boolean operation

1 INTERSECT 2    1 UNION 2    1 MINUS 2    2 MINUS 1

Revise as a span operation

1 RELATIVE TO 2, using genomic colocation

Ignore one of the inputs

IGNORE 2    IGNORE 1

TcChr24-S:392,078..392,896(+)  
TcChr26-S:87,624..88,389(+)  
TcChr26-S:708,934..710,118(-)

Orthologs

Orthologs 2,792 Genes

Run Step

- How many orthologs exist in *L. braziliensis* MHOM/BR/75/M2903? (*Hint:* look at the organism filter to the left of your result list. Click on the check box next to a species, then click apply to view results from a specific species). Explore your results. Scan the product descriptions for this list of genes. Did you find anything interesting? Perhaps a GO enrichment analysis would support your ideas.

My Search Strategies

Opened (1) All (1) Public (49) Help

Unnamed Search Strategy \*

Step 1: Tc LileCyc Mayay (R)  
127 Genes  
Step 2: Orthologs 28 Genes \* Add a step

50 Genes (28 ortholog groups) [Reuse this search](#)

Gene Results | Economic View | Gene Ontology Enrichment | [View all results](#)

Organism Filter (Select an organism to expand all / collapse all) Reset to current  
 Hide zero counts  
 Search organisms...  

- Treponematales** 2,746
- Trypanosomatidae** 38
- Biochemones** 38
- Leishmania brasiliensis (strain B0B-376)** 69
- Oribatida** 69
- Leishmania brasiliensis (strain LCI-01)** 69
- Endoprymata** 69
- Leishmania panamensis (strain LCI-01)** 69
- Leishmania** 1,114
- Leishmania antillarum** 49
- Leishmania aethiopica L147** 49
- Leishmania major** 36
- Leishmania amazonensis** 36
- MESM/MSM/MSM/M2903** 36
- Leishmania braziliensis** 147
- Leishmania brasiliensis MHOM/BR/75/M2903** 50
- Leishmania brasiliensis MHOM/BR/75/M2904** 55
- Leishmania brasiliensis MHOM/BR/75/M2904 2019** 42

Gene Ontology Enrichment Find Gene Ontology terms that are enriched in your gene result. [Read More](#)

Parameters

Organism Leishmania brasiliensis MHOM/BR/75/M2903  
 Ontology  Cellular Component  Molecular Function  Biological Process  
 Evidence  Compiled  Curated  
select all / clear all

Limit to GO Slim terms  No  Yes  
 P-Value cutoff 0.05 [0 - 1]

[Submit](#)

### 3. Finding genes based on RNAseq evidence and inferring function of hypothetical genes. Note: Use <http://plasmodb.org> for this exercise.

- Find all genes in *P. falciparum* that are up-regulated at least 50-fold in ookinetes compared to other stages: “**Transcriptomes of 7 sexual and asexual life stages (Lopez-Barragan et al.)**”. For this search select “average” for the operation applied on the reference samples.

## Identify Genes based on P. falciparum 3D7 Transcriptomes of 7 sexual and asexual life stages RNASeq (fold change)

[Tutorial](#)



For the **Experiment** Transcripomes of 7 sexual and asexual life stages unstranded [?](#)  
 return **protein coding** [?](#) **Genes**  
 that are **up-regulated** [?](#)  
 with a **Fold change >= 50** [?](#)

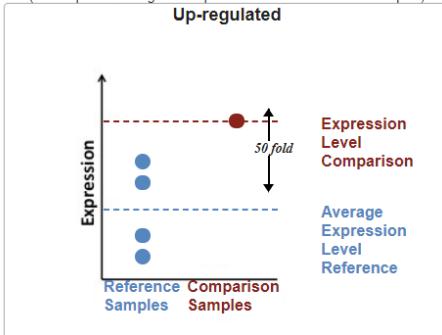
between each gene's **average** [expression value](#) [?](#)  
 (or the **Floor** of **10 reads (.88 FPKM)** [?](#) [NEW](#)  
 in the following **Reference Samples** [?](#)

Ring  
 Early Trophozoite  
 Late Trophozoite  
 Schizont  
 Gametocyte II  
[select all](#) | [clear all](#)

and its **expression value** [?](#)  
 (or the **Floor** selected above)  
 in the following **Comparison Samples** [?](#)

Late Trophozoite  
 Schizont  
 Gametocyte II  
 Gametocyte V  
 Ookinete  
[select all](#) | [clear all](#)

**Example showing one gene that would meet search criteria**  
 (Dots represent this gene's expression values for selected samples)



A maximum of four samples are shown when more than four are selected.  
 You are searching for genes that are up-regulated between at least two reference samples and one comparison sample.

For each gene, the search calculates:

$$\text{fold change} = \frac{\text{comparison expression level}}{\text{average expression level in reference*}}$$

and returns genes when **fold change >= 50**.

To narrow the window, use the maximum reference value. To broaden the window, use the minimum reference value.

See the detailed help for this search.

\* or **FPKM Floor**, whichever is greater

[Get Answer](#)

- b. The above search will give you all genes that are up-regulated by 50 fold in ookinetes compared to the average expression level of other stages. Despite the high fold change, some genes in the list may be highly expressed in the other stages. How can you remove genes from the list that are highly expressed in the other stages?

**3D7 7Stages RNA-Seq (fc)  
31 Genes**

**Step 1**

**+ Add a step**

- Hint: Add a search for genes based on RNA Seq evidence from the same experiment, but this time select the percentile search: P.f. seven stages - RNA Seq (percentile). What minimal percentile values should you choose? 40 – 100%? How does setting the any / all samples impact the result .... Which would be better in this case?



## Add a step to your search strategy ?

### Search for Genes by RNA-Seq Evidence

The results will be intersected with | the results of Step 1.

Filter Data Sets:  ? Legend: DE Differential Expression FC Fold Change P Percentile SA SenseAntisense

Organism <span style="color: blue;">?</span>	Data Set	Choose a Search
Plasmodium falciparum 3D7	Transcriptomes of 7 sexual and asexual life stages (Lopez-Barragan et al.)	<span style="background-color: #0072BD; color: white; border-radius: 5px; padding: 2px 5px;">FC</span> <span style="background-color: #0072BD; color: white; border-radius: 5px; padding: 2px 5px;">P</span>

Show All Data Sets

Fold Change Percentile

? Experiment

Transcriptomes of 7 sexual and asexual life stages unstranded

? Samples

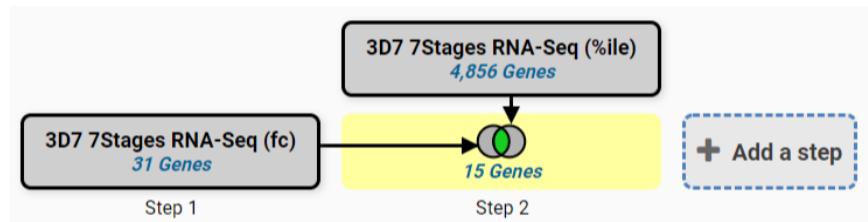
Ring  
 Early Trophozoite  
 Late Trophozoite  
 Schizont  
 Gametocyte II  
 Gametocyte V  
 Ookinete  
[select all](#) | [clear all](#)

? Minimum expression percentile

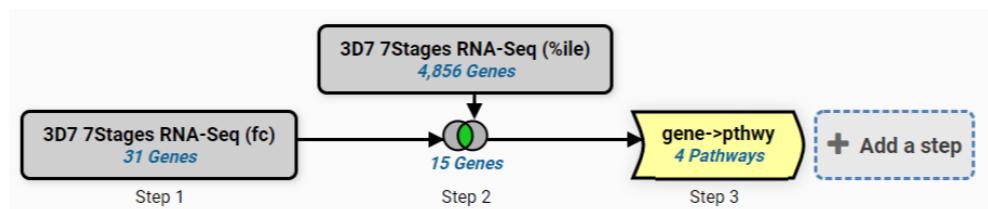
? Maximum expression percentile

? Matches Any or All Selected Samples?

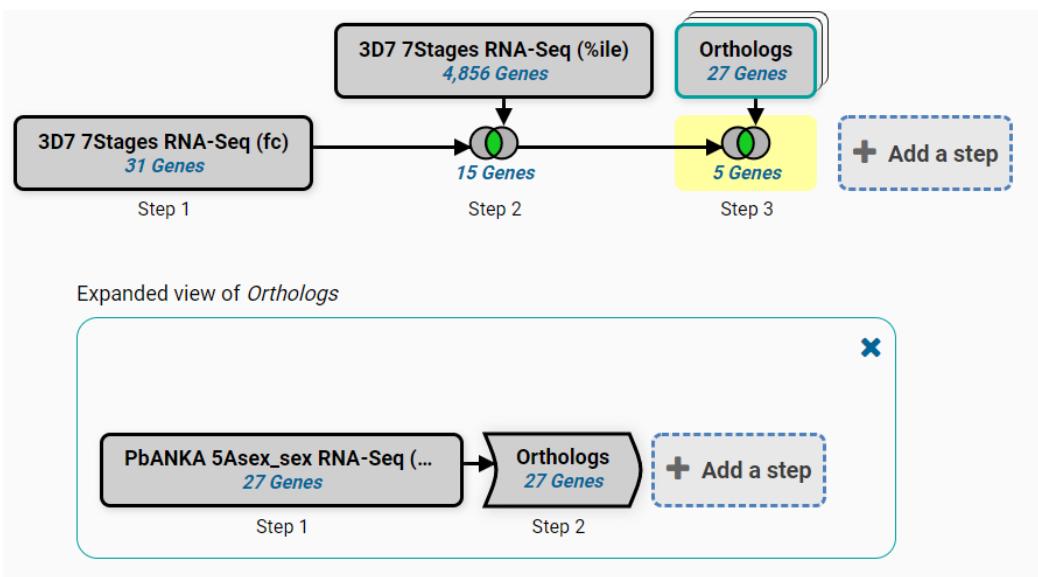
- Hint II: Try changing the operator from average to maximum for the set of non-ookinete stages in your initial fold change search. What does this do? How do the resulting genes compare with the two step strategy you generated in the first hint? Which hint do you think works better?



- c. Which metabolic pathways are represented in this gene list? Hint: add a step and transform results to pathways. How does this result compare to running a pathways enrichment on step 2?



- d. What happens if you revise the first step and modify the fold difference to a lower value - 10 for example? Compare results when you also modify the “between each genes” parameter. What happens if you set this to maximum? Which value do you think is most stringent for ensuring at 10 fold up regulation compared to the other samples?
- e. PlasmoDB also has an experiment examining gene expression during sexual development in *Plasmodium berghei* (rodent malaria). Can you determine if there are genes that are up-regulated in both human and rodent ookinetes (compared to all other stages)? Hint: start by deleting the last step you added in this exercise (transform to pathways). To do this click on edit then delete in the popup. Next, add steps for the *P. berghei* experiments “*P. berghei* ANKA 5 asexual and sexual stage transcriptomes RNASeq”. Note that you will have to use a nested strategy or by running a separate strategy then combining both strategies.



4. Find genes that are essential in procyclics but not in blood form *T. brucei*.

Note: for this exercise use <http://TriTrypDB.org>.

- Find the query for High Throughput Phenotyping. Think about how to set up this query (*Hint*: you will have to set up a two-step strategy). Remember you can play around with the parameters but there is no one correct way of setting them up –

Quantitative Phenotype

Learn more about this search

## Identify Genes based on High-Throughput Phenotyping

Tutorial YouTube

For the Experiment Quantitated from the CDS Sequence return protein coding Genes that are Decrease in coverage with a Fold change >= 1.5 between each gene's expression value in the following Reference Samples

(Uninduced sample)

and its expression value in the following Comparison Samples

Induced in bloodstream (BS) forms, 3 days (10 doublings)  
Induced in bloodstream (BS) forms, 6 days (20 doublings)  
 Induced in procyclic forms (PS) forms, 9 days (9 doublings)  
Induced throughout differentiation (DIF = 7 BS doublings + 6 PS doublings)  
select all | clear all

**Example showing one gene that would meet search criteria**  
(Dots represent this gene's expression values for selected samples)

Down-regulated

You are searching for genes that are down-regulated between one reference sample and one comparison sample.

For each gene, the search calculates:

$$\text{fold change} = \frac{\text{reference expression level}}{\text{comparison expression level}}$$

and returns genes when fold change >= 1.5.

See the detailed help for this search.

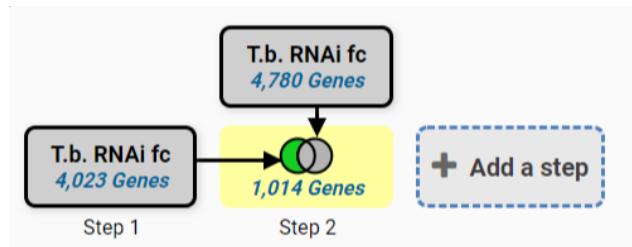
Get Answer

T.b. RNAi fc  
4,023 Genes

+ Add a step

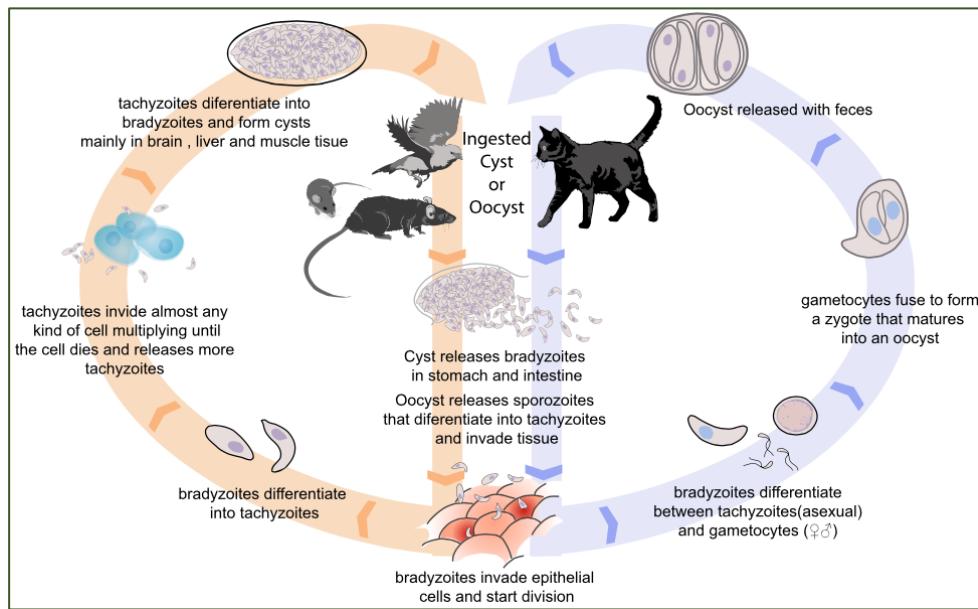
Step 1

- Next add a step and run the same search except this time select the “induced bloodstream form” samples.
- How did you combine the results? Remember you want to find genes that are essential in procyclics and not in blood form.



5. Finding oocyst expressed genes in *T. gondii* based on microarray evidence.

Note: For this exercise use <http://toxodb.org>



- a. Find genes that are expressed at 10 fold higher levels in one of the oocyst stages than in any other stage in the “Oocyst, tachyzoite, and bradyzoite developmental expression profiles (M4) (Fritz and Buchholz)” microarray experiment.

## Search for...

[expand all](#) | [collapse all](#)

**▼ Genes**

- ▶ Annotation, curation and identifiers
- ▶ Epigenomics
- ▶ Function prediction
- ▶ Gene models
- ▶ Genetic variation
- ▶ Genomic Location
- ▶ Immunology
- ▶ Orthology and synteny
- ▶ Pathways and interactions
- ▶ Phenotype
- ▶ Protein features and properties
- ▶ Protein targeting and localization
- ▶ Proteomics
- ▶ Sequence analysis
- ▶ Structure analysis
- ▶ Taxonomy
- ▶ Text
- ▼ Transcriptomics
- [Microarray Evidence](#) 
- [RNA-Seq Evidence](#)

Filter Data Sets: oocyst

Legend: S Similarity FC Fold Change P Percentile

Organism: T. gondii ME49  
(filtered from 11 total entries)

Data Set: Oocyst, tachyzoite, and bradyzoite developmental expression profiles (M4) (Fritz and Buchholz et al.)

Show All Data Sets

Learn more about this search [Tutorial](#)

Identify Genes based on T. gondii ME49 Oocyst, tachyzoite, and bradyzoite developmental expression profiles (M4) Microarray (fold change)

For the Experiment: Oocyst, tachyzoite, and bradyzoite developmental expression profiles (M4) [return protein coding genes](#) that are up or down regulated [with a Fold change >= 10](#) between each gene's average expression value in the following Reference Samples:

- 10 days sporulated
- 2 days in vitro
- 4 days in vitro
- 6 days in vitro
- 21 days in vivo

and its average expression value in the following Comparison Samples:

- unsporulated
- 4 days sporulated
- 10 days sporulated
- 2 days in vitro
- 4 days in vitro

Example showing one gene that would meet search criteria (dots represent this gene's expression values for selected samples)

Up or down regulated

You are searching for genes that are up or down regulated between at least two reference samples and at least two comparison samples.

For each gene, the search calculates:

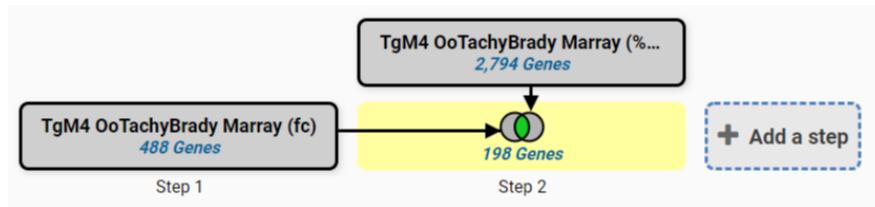
$$\text{fold change}_{\text{up}} = \frac{\text{average expression level in comparison}}{\text{average expression level in reference}}$$

$$\text{fold change}_{\text{down}} = \frac{\text{average expression level in reference}}{\text{average expression level in comparison}}$$

and returns genes when  $\text{fold change}_{\text{up}} >= 10$  or  $\text{fold change}_{\text{down}} >= 10$ . See the detailed help for this search.

[Get Answer](#)

- b. Add a step to limit this set of genes to only those for which all the non-oocyst stages are expressed below 50<sup>th</sup> percentile ... i.e., likely not expressed at those stages. (Hint: after you click on add step find the same experiment under microarray expression and chose the percentile search).
- Select the 4 non-oocyst samples.
  - We want all to have less than 50<sup>th</sup> percentile so set **minimum percentile to 0** and **maximum percentile to 50**.
  - Since we want all of them to be in this range, choose **ALL** in the “**Matches Any or All Selected Samples**”.



- To view the graphs in the final result table, turn on the columns called “TgM4 OoTachyBrady Marray - Expr Graph” and “TgM4 OoTachyBrady Marray - %ile Graph” (inside the “*T. gondii* ME49 Oocyst, tachyzoite, and bradyzoite developmental expression profiles (M4) (Fritz and Buchholz et al.)” Microarray).



## 6. Comparing RNA abundance and Protein abundance data.

Note: for this exercise use <http://TriTrypDB.org>.

In this exercise we will compare genes that show differential RNA abundance levels between procyclic and blood form stages in *T. brucei* with genes that show differential protein abundance in these same stages.

- a. Find genes that are down-regulated 2-fold in procyclic form cells. Go to the search page for Genes by Microarray Evidence and select the fold change search for the “Expression profiling of five life cycle stages (Marilyn Parsons)” experiment and configure the search to return protein-coding genes that are down-regulated 2 fold in procyclic form (PCF) relative to the Blood Form reference sample. Since there are two PCF samples, it is reasonable to choose both and average them.

**Identify Genes based on Microarray Evidence**

**Search for...**

expand all | collapse all  
Filter the searches below...

**Legend:** DC Direct Co... FC Fold Chan... P Percentile

Organism	Data Set	Choose a search
<i>L. infantum</i> JPCM5	Promastigote-to-amastigote differentiation ( <i>L.d.</i> Samples) (Lahav et al.)	FC P
<i>L. infantum</i> JPCM5	Axenic and intracellular amastigote profiles (Rochette et al.)	DC P
<i>L. major</i> strain Friedlin	Three Developmental Stages (Stephen M. Beverley)	DC P
<i>T. brucei</i> brucei TREU927	Expression profiling of in vitro differentiation (Queiroz et al.)	FC P
<i>T. brucei</i> brucei TREU927	Expression profiling of five life cycle stages (Marilyn Parsons)	FC P
<i>T. brucei</i> brucei TREU927	Procyclic trypanosomes: heat shock vs untreated control (Kramer et al.)	DC P
<i>T. brucei</i> brucei TREU	Identify Genes based on <i>T. brucei</i> Expression profiling of five life cycle stages Microarray (fold change)	FC P
<i>T. brucei</i> brucei TREU	Tutorial	DC P
<i>T. cruzi</i> CL Brener Esn		DC P

For the Experiment Expression profiling of five life cycle stages  
return protein coding Genes  
that are down-regulated  
with a Fold change >= 2.0

For each gene's average expression value in the following Reference Samples  
 Blood Form  
 Stender  
 Stunay  
 PCF Log  
 PCF Stat  
 select all | clear all

and its average expression value in the following Comparison Samples  
 Blood Form  
 Stender  
 Stunay  
 PCF Log  
 PCF Stat  
 select all | clear all

**Example showing one gene that would meet search criteria**  
 (dots represent this gene's expression values for selected samples)

You are searching for genes that are down-regulated between at least two reference samples and at least two comparison samples.

For each gene, the search calculates:  

$$\text{fold change} = \frac{\text{average expression value in reference samples}}{\text{average expression value in comparison samples}}$$

and returns genes when fold change >= 2.0. To narrow the window, use the minimum reference value, or maximum comparison value. To broaden the window, use the maximum reference value, or minimum comparison value.

See the detailed help for this search.

Get Answer

**Tb LifeCyc Marray (fc)  
378 Genes**

+ Add a step

Step 1

- b. Add a step to compare with quantitative protein expression. Select protein expression then "Quantitative Mass Spec Evidence" and the "Quantitative phosphoproteomes of bloodstream and procyclic forms (Tb427) (Urbaniak et al.)" experiment. Configure this search to return genes that are down-regulated in procyclic form relative to blood form.

Add a step to your search strategy

Search for Genes by Quantitative Mass Spec. Evidence

The results will be  intersected with |  the results of Step 2.

Filter Data Sets:

Legend: DC Direct Comparison DCC Direct Confidence Comparison QR Quantitative Ratio FC Fold Change

**Organism**: Trypanosoma brucei TREU927 **Data Set**: Quantitative phosphoproteomes of bloodstream and procyclic forms (Tb427) (Urbaniak et al.) **Choose a Search**: DC

Show All Data Sets

**Direct Comparison**

**Experiment**: Quantitative phosphoproteomes of bloodstream and procyclic forms (Tb427)

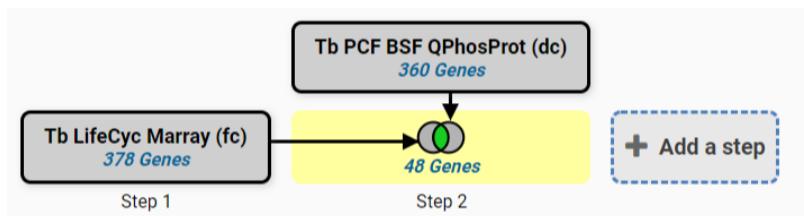
**Direction**: down-regulated

**Comparison**: Pcf-Bsf ratio

**Fold difference >=**: 2

Run Step

- c. How many genes are in the intersection? Does this make sense? Make certain that you set the directions correctly.



- d. Try changing directions and compare up-regulated genes/proteins. (*Hint:* revise the existing strategy ... you might want to duplicate it so you can keep both). When you change one of the steps but not the other do you have any genes in the intersection? Why might this be?

- e. Can you think of ways to provide more confidence (or cast a broader net) in the microarray step? (*Hint:* you could insert steps to restrict based on percentile or add a RNA Sequencing step that has the same samples).

7. Find genes with evidence of protein phosphorylation in intracellular *Toxoplasma* tachyzoites.

For this exercise use <http://www.toxodb.org>

Phosphorylated peptides can be identified by searching the appropriate experiments in the Mass Spec Evidence search page.

- 7a. Find all genes with evidence of protein phosphorylation in intracellular tachyzoites. Navigate to the Post-Translational Modification search. Select the “**Infected host cell, phosphopeptide-enriched** (peptide discovery against TgME49)” sample under the experiment called “**Tachyzoite phosphoproteome from purified parasite or infected host cell (RH) (Treeck et al.)**”

The screenshot shows the ToxoDB search interface for identifying genes based on post-translational modifications. On the left, a sidebar titled "Search for..." lists various categories under "Genes" and "Proteomics". The "Post-Translational Modification" category is circled in red and has a blue arrow pointing to the main search area. The main search area is titled "Identify Genes based on Post-Translational Modification". It includes sections for "Type of Post-Translational Modification" (set to "phosphorylation site"), "Experiments and Samples" (listing "1 selected, out of 9" experiments under "Toxoplasma gondii" and "Toxoplasma gondii ME49"), "Number of modifications is" (set to "Greater than or equal to"), and "Number of Modifications" (set to "1"). A "Get Answer" button is at the bottom right.

- 7b. Remove all genes with phosphorylation evidence from purified tachyzoites and the phosphopeptide depleted fractions.

Hint: Use the Mass Spec Evidence search to access the tachyzoite and depleted fractions. Subtract (1 minus 2) these results from your first search.



Add a step to your search strategy

## Search for Genes by Mass Spec. Evidence

The results will be subtracted from the results of Step 1.

### Experiments and Samples

Note: You must select at least 1 values for this parameter.  
3 selected, out of 92

[select all](#) | [clear all](#) | [expand all](#) | [collapse all](#)

[Filter list below...](#)

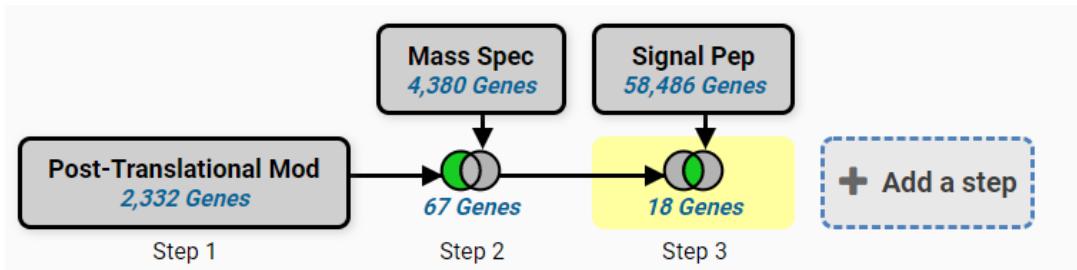


- Eimeria
- Neospora
- Toxoplasma
  - Toxoplasma gondii
    - T. gondii* Proteome During Infection in *Homo sapiens* (GT1 ME49 RH VEG) (Krishna et al.)
    - Extracellular vesicles (RH) (Wowk et al.)
    - Mitochondrial Matrix Proteome (Seidi and Muellner-Wong et al.)
    - Monomethylarginine Proteomics (RH) (Yakubu et al.)
      - monomethylarginine
    - Mouse brain bradyzoite proteomics time course (Garfoot et al.)
    - N-terminal Peptides (RH) (Dogga et al.)
    - Oocyst Partially Sporulated Proteome (VEG) (Possenti et al.)
      - Oocyst proteome
    - Oocyst proteome (M4 TypeII) (Wastling)
      - Oocyst peptides
    - Oocyst proteome - Fractionated (M4 type II) (Fritz et al.)
    - TAILS peptides (Coffey et al.)
      - TAILS N-terminal proteomics
    - Tachyzoite Intra- and Extracellular Lysine-Acetylomes (RH) (Jeffers and Xue)
    - Tachyzoite Rhoptry proteome (RH) (Bradley et al.)
      - purified rhoptries
    - Tachyzoite Ubiquitome (Simlon de Monerri et. al)
    - Tachyzoite conoid proteome (RH) (Hu et al.)
    - Tachyzoite membrane and cytosolic proteomes (RH) (Dybas et al.)
    - Tachyzoite phosphoproteome - Calcium dependent (RH) (Nebi et al.)
    - Tachyzoite phosphoproteome from purified parasite or infected host cell (RH) (Treeck et al.)
      - Infected host cell, phosphopeptide-depleted (peptide discovery against TgME49)
      - Infected host cell, phosphopeptide-enriched (peptide discovery against TgME49)
      - Purified tachyzoites phosphopeptide-depleted (peptide discovery against TgME49)
      - Purified tachyzoites phosphopeptide-enriched (peptide discovery against TgME49)
    - Tachyzoite secretome (RH) (Zhou et al.)
    - Tachyzoite subcellular fractions (Moreno)
    - Tachyzoite total proteome (RH) (Wastling)

[select all](#) | [clear all](#) | [expand all](#) | [collapse all](#)

7d. Explore your results. What kinds of genes did you find? Hint: use the Product description word column or perform a GO enrichment analysis of your results.

7e. Are any of these genes likely to be secreted? Hint: add a step searching for genes with secretory signal peptides.



7f. Pick one or two of the hypothetical genes in your results and visit their gene pages. Can you infer anything about their function? Hint: explore the protein and expression sections.

7g. What about polymorphism data? Go back to your strategy and add columns for SNP data found under the population biology section. Explore the gene page for the gene that has the most number of non-synonymous SNPs. Hint: you can sort the columns by clicking on the up/down arrows next to the column names.

The screenshot shows the ToxoDB Gene Results page. At the top, there's a header with the ToxoDB logo, a site search bar, and a 'Add a step' button. Below the header is a 'My Search Strategies' section with a 'Post-Translational Mod' strategy. The main content area displays 18 genes across 18 ortholog groups. On the left, there's a 'Organism Filter' sidebar. The main table has columns for Gene ID, Transcript ID, Gene Group, Genomic Location (Gene), Product Description, Transcripts, TgM4 OnTachibana Brady Murray - Expr Graph, Non-Coding SNPs, NonSyn/Syn SNPs All Strains, NonSynonymous SNPs All Strains, and Synonymous SNPs All Strains. Two specific genes are highlighted with bar charts: TOME49\_257595 and TOME49\_219640, showing their expression profiles across different strains.

## **8. Find *T. gondii* genes expressed in late enteroepithelial stages**

*Toxoplasma gondii* is a zoonotic pathogen for which felids serve as definitive hosts. In cats, the parasite undergoes several rounds of asexual replication before entering the sexual cycle which gives rise to oocysts that are shed into the environment. These then sporulate and become infective to humans and livestock. To understand the genes involved in the parasite development in the felid host and identify potential intervention targets, we designed a transcriptomic approach to compare the cat intestinal stages with the well characterized tachyzoites that mediate acute infection and tissue cysts that are responsible for chronic infection. Cats were infected with *T. gondii* CZ clone H3 tissue cysts from mouse brain and the intestinal stages were sampled at day 3, 5 and 7 post infection. As an input sample, we also collected tissue cysts from mouse brain. In vitro cultivated tachyzoites were also harvested. Total RNA was extracted, enriched for mRNA and used for cDNA synthesis. RNA-Seq was then performed to describe the transcriptomic repertoire of each developmental stage. RNA-seq datasets from each time point post inoculation with bradyzoites in kittens were subjected to cluster analysis and assigned to five enteroepithelial developmental stages (EES) according to their profile.

### Cat enteroepithelial stages:

- EES1 = very early enteroepithelial stages
- EES2 = early enteroepithelial stages
- EES3 = mixed enteroepithelial stages
- EES4 = late enteroepithelial stages
- EES5 = very late enteroepithelial stages
- Navigate to the RNaseq searches and identify the experiment of cat enterocyte stages. Configure the search to identify call *T. gondii* genes that are upregulated by at least 2-fold in late and very late enteroepithelial stages (EES4 and EES5) compared to all other stages available from this experiment.

## Identify Genes based on RNA-Seq Evidence

Filter Data Sets: enteroto ?

Legend: DE Differential Expression FC Fold Change P Percentile SA SenseAntisense

Organism <span style="float: right;">?</span>	Data Set <span style="float: right;">?</span>	Choose a Search
Toxoplasma gondii ME49	Feline enterocyte, tachyzoite, bradyzoite stage transcriptome (Hehl, Ramakrishnan et al.)	<span style="border: 2px solid red; padding: 2px;">DE</span> <span style="border: 2px solid red; padding: 2px;">FC</span> <span style="border: 2px solid red; padding: 2px;">P</span> <span style="border: 2px solid red; padding: 2px;">SA</span>

↓ Show All Data Sets ↓

Differential Expression    Fold Change    Percentile    SenseAntisense

Identify Genes based on T. gondii ME49 Feline enterocyte, tachyzoite, bradyzoite stage transcriptome RNA-Seq (fold change)

For the Experiment  
Feline enterocyte, tachyzoite, bradyzoite stage transcriptome - Sense

return protein coding ? Genes ?  
that are up-regulated ?  
with a Fold change >= 2 ?  
between each gene's maximum ? expression value  
(or a Floor of 10 reads ?)  
in the following Reference Samples ?

- EES3
- EES4
- EES5
- Tachyzoites
- Tissue cysts

select all | clear all

and its minimum ? expression value ?  
(or the Floor selected above)  
in the following Comparison Samples ?

- EES1
- EES2
- EES3
- EES4
- EES5

select all | clear all

Example showing one gene that would meet search criteria  
(Dots represent this gene's expression values for selected samples)

A maximum of four samples are shown when more than four are selected.

For each gene, the search calculates:

$$\text{fold change} = \frac{\text{minimum expression value in comparison}}{\text{maximum expression value in reference}}$$

and returns genes when **fold change >= 2**.

You are searching for genes that are **up-regulated** between at least two **reference samples** and at least two **comparison samples**.

This calculation creates the **narrowest** window of expression values in which to look for genes that meet your fold change cutoff. To broaden the window, use the average or maximum reference value, or average or maximum comparison value.

Get Answer

+ **Toxo Cat RNAseq (fc)**  
**238 Genes**

+ **Add a step**

Step 1

- What kinds of genes did this search identify? How can you determine if your results are enriched for a particular function? Try clicking on Analyze Results and explore the GO enrichment tool.

My Search Strategies

Search (1) Toxoplasma gondii ME49 - PubGet

Advanced Search Strategy: >

200 Genes (200 matching genes)

Gene Results

Analyze your Gene results with a tool below.

Gene Ontology Enrichment

The screenshot shows the PubGet search interface. At the top, there's a search bar with the query "Toxoplasma gondii ME49 - PubGet". Below it, a search strategy is defined: "Search (1) Toxoplasma gondii ME49 - PubGet". A red arrow points from the "Gene Results" section to the "Gene Ontology Enrichment" tool on the right. Another red arrow points from the "Analysis Results" table back to the search interface.

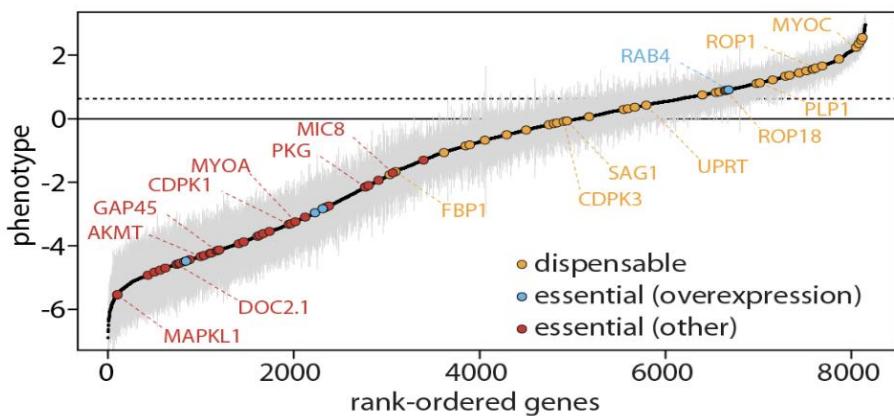
**Analysis Results:**

GO Term	Genes in the Me49 with this term	Genes in your result with this term	Percent of Me49 genes in your result	Fold enrichment	Odds ratio	P-value	Benjamini	Bonferroni
GO:0016710 phosphorylation	192	11	5.7	2.47	2.79	4.15e-3	2.20e-1	5.36e-1
GO:0004848 protein phosphorylation	167	10	6.0	2.50	2.80	4.40e-3	2.20e-1	6.57e-1
GO:0019344 cysteine biotinylation process	1	1	100.0	43.09	infinity	2.30e-2	4.20e-1	1.00e+0
GO:0019342 cysteine biotinylation process via cysteathione	1	1	100.0	43.09	infinity	2.30e-2	4.20e-1	1.00e+0
GO:0000912 galactose metabolic process	1	1	100.0	43.09	infinity	2.30e-2	4.20e-1	1.00e+0

## 9. Finding genes based on high throughput mutagenesis and fitness analysis.

In EuPathDB we have a variety of studies where genome scale phenotypic analyses were carried out. In this exercise we'll use [ToxoDB.org](#) and look at fitness following CRISPR mutagenesis. You could also explore phenotyping studies in PlasmoDB or FungiDB if you prefer, the principles are the same.

- Navigate to the CRISPR phenotype search. Note that this search form is quite simple just requiring a range of fitness values. The defaults return all genes not limiting the search at all. This is only useful in as much as it tells you which genes were assayed which is nearly the entire genome. The tricky bit is deciding where to make the cutoffs. Again, the description on the search form is very helpful in this regard (as is the link to the paper ... remember these phenotypes were assayed under specific conditions so just because a particular gene doesn't show a phenotype doesn't mean it wouldn't in other conditions (or infecting an actual host). The plot showing the phenotype score (fitness) is particularly useful. Red points along the plot are genes known to be essential under these conditions



while yellow are known to be expendable. This will help you determine where to set the values. The last essential gene has a fitness score just  $\geq$  than -4 so setting the phenotype score  $\leq$  -2 would provide a pretty stringent search but still return more than 1000 genes. Try it. Do you get the expected results based on the number of genes returned?

Search for...

expand all | collapse all

Filter the searches below...

Genes

- Annotation, curation and identifiers
- Epigenomics
- Function prediction
- Gene models
- Genetic variation
- Genomic Location
- Immunology
- Orthology and synteny
- Pathways and interactions
- Protein features and properties
- Protein targeting and localization
- Proteomics
- Sequence analysis
- Structure analysis
- Taxonomy
- Text
- Transcriptomics

Phenotype

CRISPR Phenotype

Identify Genes based on CRISPR Phenotype

Step 1

Get Answer

Phenotype Score  $\geq$   Phenotype Score  $\leq$

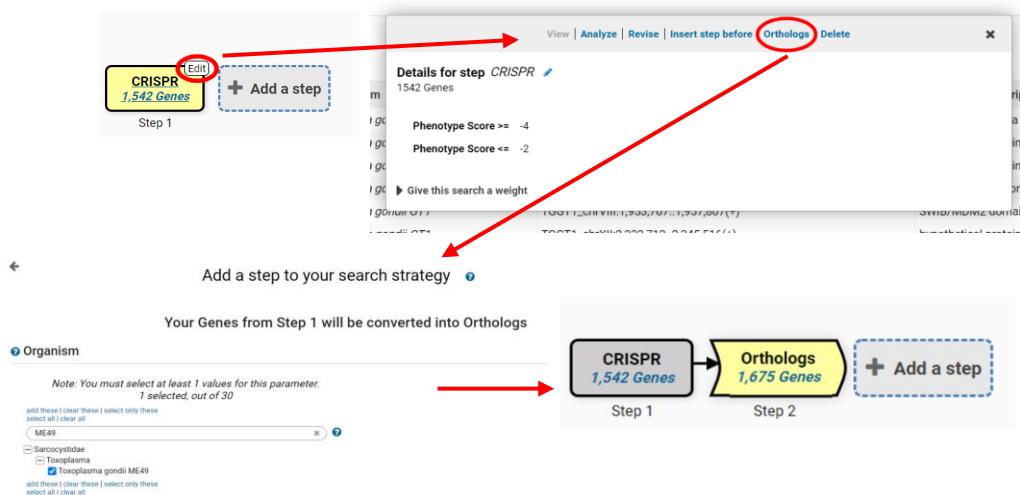
CRISPR 1,542 Genes

+ Add a step

- Can you find additional evidence that these genes are essential? One way is to use the analysis tools to assess biological process and go function. Are the results what you would expect?



- Try adding columns to show additional data or intersecting these results with other queries, perhaps expression queries, to further assess this list. NOTE: this experiment was done in GT1 while all *T. gondii* functional data in ToxoDB is mapped to ME49 so an ortholog transform to ME49 is required before adding any additional functional studies.
- To do this, click on add step and select the Transform to orthologs option and select *T. gondii* ME49 to transform to.



- How many of these genes are upregulated in *in vivo* chronic stages of *T. gondii*?
  - Click on add step
  - Select the RNAseq searches under the Transcriptomics category
  - Find the experiment with chronic stages and run a search based on differentially expressed genes (DE).

The screenshot shows the 'Add Step 3 : RNA Seq Evidence' dialog. At the top, there is a 'Filter Data Sets' input field containing 'chroni'. Below it is a legend with four buttons: 'DE' (Differential Expression, highlighted with a red circle), 'FC' (Fold Change), 'P' (Percentile), and 'SA' (SenseAntisense). The 'Organism' dropdown shows 'T. gondii ME49' and the 'Data Set' dropdown shows 'Transcriptome during acute or chronic infection in mouse brain (Pittman et al.)'. A red circle highlights the 'DE' button in the legend.

- Intersect genes that are 2-fold upregulated in chronic stages compared to acute stages.
- What do these results look like? Do you find any interesting genes?

**Add Step 3 : T. gondii ME49 Transcriptome during acute or chronic infection in mouse brain RNASeq (Differential Expression)**

**Experiment**  
Acute and chronic T.gondii infection of mouse, unstranded

**Reference Sample**  
 acute infection 10 days p.i.  
 chronic infection 28 days p.i.

**Comparator Sample**  
 acute infection 10 days p.i.  
 chronic infection 28 days p.i.

**Direction**  
up-regulated

**fold difference >=**  
2

**adjusted P value less than or equal to**  
0.1

**Tg In murine macrophages RN...  
314 Genes**

**Step 1**: CRISPR  
1,542 Genes

**Step 2**: Orthologs  
1,675 Genes

**Step 3**: 30 Genes

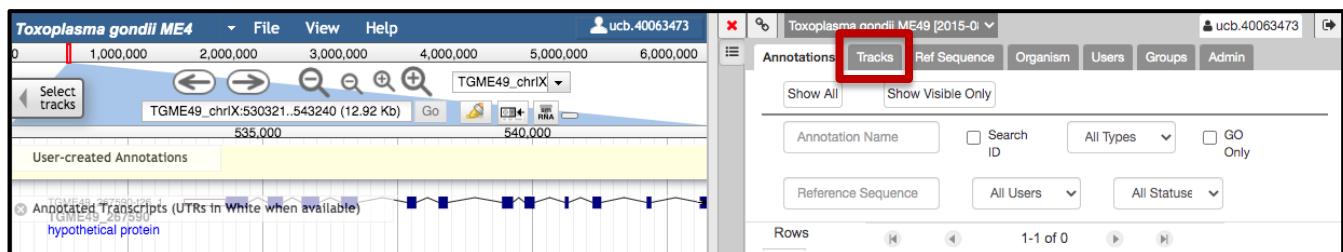
+ Add a step

**Combine Genes in Step 2 with Genes in Step 3:**

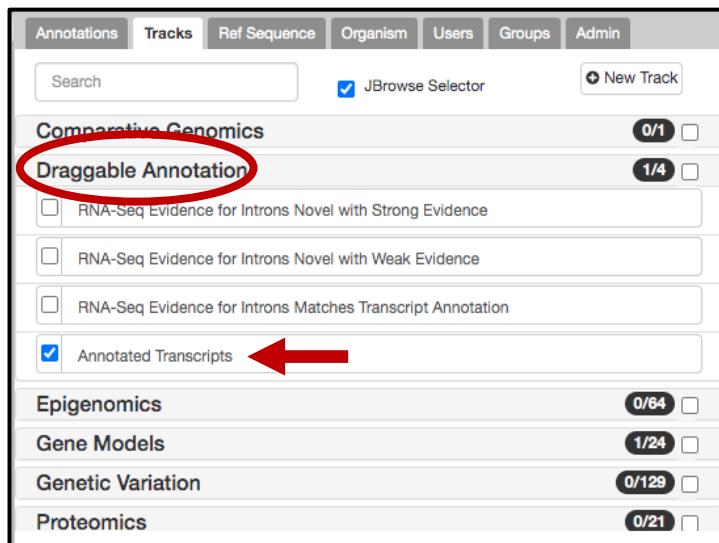
2 Intersect 3       2 Minus 3  
 2 Union 3       3 Minus 2  
 2 Relative to 3 , using genomic colocation

## 2) Dragging and dropping the annotation

Select on the menu on the right-hand side the tab **Tracks**.



Click on the menu item **Draggable Annotation** and select **Annotated Transcripts**.



You can now see your gene of interest in the Annotated Transcript track. Select the gene model by clicking on one of the introns or by double clicking on the gene model (1). The gene model will show up with red boundaries. Drag and drop the gene into the User-created Annotations area (2).





May 2020

## MapVEu: part I - An introduction

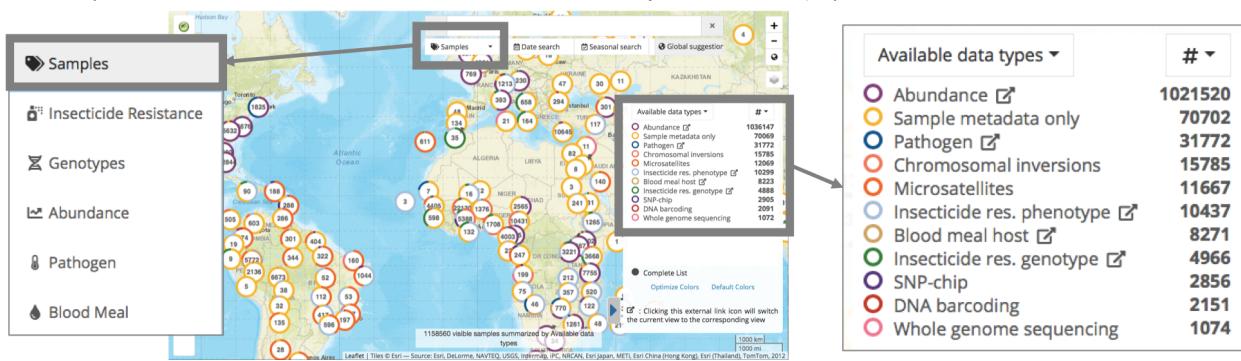
**Note:** this exercise uses VectorBase.org data as an example, but the same functionality will soon become available for other VEuPathDB resources

### Learning objectives:

- Use keywords, filters and ‘click and zoom’ to query and browse samples or assays
- Interpret the markers or points on the map, their colors and numbers
- Share a link of your current ‘interactive’ map view, not just a static screenshot
- Download the raw data
- Find the data providers/sources to cite them

Advanced features can be found in [part II](#) of this tutorial

MapVEu is part of our ongoing efforts to integrate genomic, phenotypic and population data as shown below (the # column indicates the number of samples or assays):



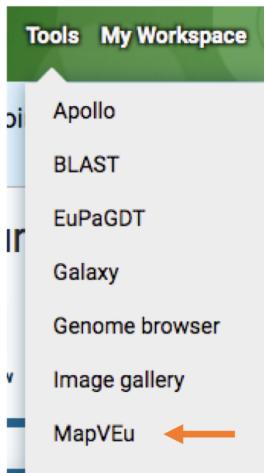
This icon indicates which data types have their own ‘map view’, click on them to switch views. An alternative way to change the view is by clicking the dropdown menu (top of the page). Currently there are six views and there are plans to include more, based on users needs. The markers in the map, such

us this one , indicate each data point and corresponding metadata including publication citation or DOI for unpublished datasets. Other menus and filters will be explained in more detail with the following [sample use case](#): Let’s explore the insecticide resistance state of *Anopheles gambiae sensu lato* in Ethiopia, for assays performed with the WHO paper kit diagnostic test (DT) for adults:

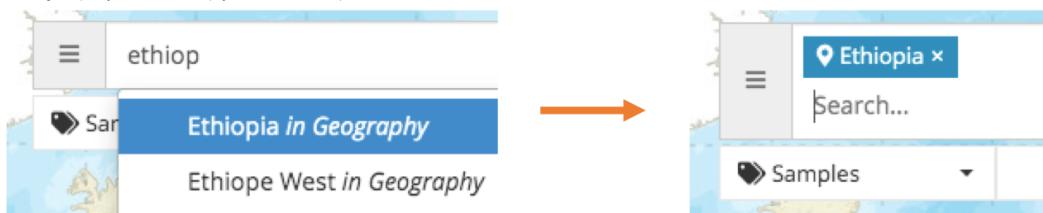
- How many different insecticides have been tested?
- What is the time span in years for the data collections?
- Are there insecticides with a mortality indicating insecticide susceptibility (i.e., ≥98% mortality)?
- Are there any insecticides with an increase in mortality suggesting possible resistance (i.e., 90–97% mortality)?
- For the last year in record, has there been any confirmed cases of resistance (i.e., <90% mortality)?
- How do I cite the data?

Here is the **step by step process** to answer these questions

1. To open the map, go to home page > Tools menu > MapVEu



2. By default, the map view is set to 'Samples', which aggregates ALL the data from the map. Use Ethiopia as a keyword. Notice how while you type, autocomplete suggests this word for the geography (ontology) category, select it



3. Before you click on the map, there are three markers. Now slowly begin to zoom in, notice the markers divide. Different alternatives to zooming include: click on the plus sign on the map (as shown in the image below), mouse wheel, double click marker, double click map, and shift-click to draw rectangle



4. The markers that will not divide any further are labeled with a pin



5. On the right hand side menu

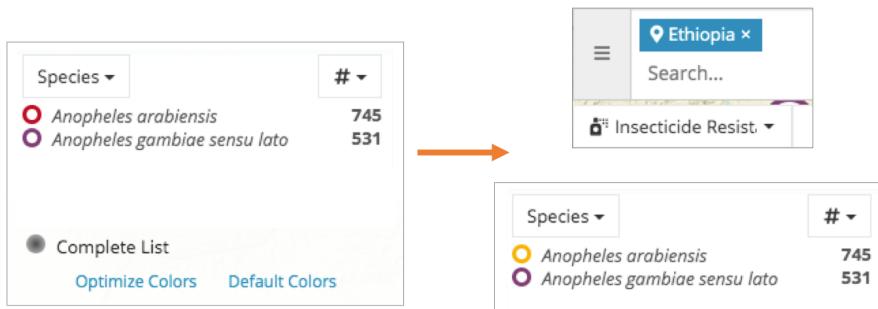
Available data types ▾

# ▾

Abundance ↗  
 Insecticide res. phenotype ↗ 1239

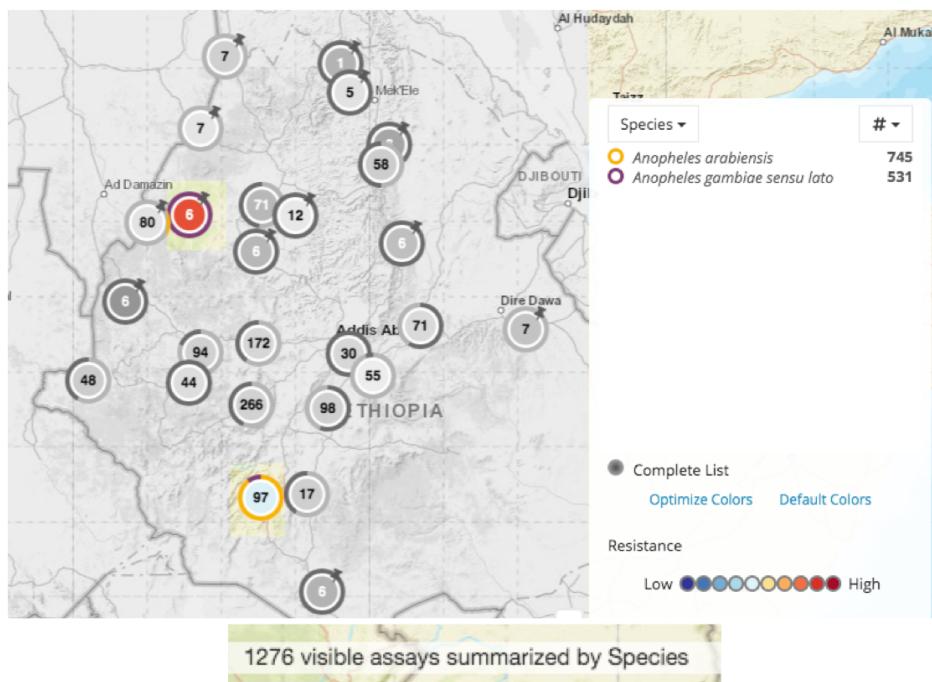
- Click on this icon ↗ next to ‘insecticide resistance phenotype’. This will switch you from ‘samples’ to ‘insecticide resistance’ (IR) view. Except for the samples view, each other ‘view’ represents a SUBSET of the data. In IR view we are now visualizing assays, not samples
- Note: this is different from a click on the text (‘insecticide resistance phenotype’), which will generate a query with these keywords in the same (Samples) view

6. The filters in the right hand menu change from ‘available data types’ to ‘species’. Click on ‘optimize colors’ to have a better color contrast of the markers



7. This is how to interpret the exterior ring and interior circle of the markers:

- There are 6 assays for *gambiae sensu lato* with insecticide resistance (red)
- There are 97 assays for *A. gambiae sensu lato* and *A. arabiensis*, with insecticide susceptibility (blue)

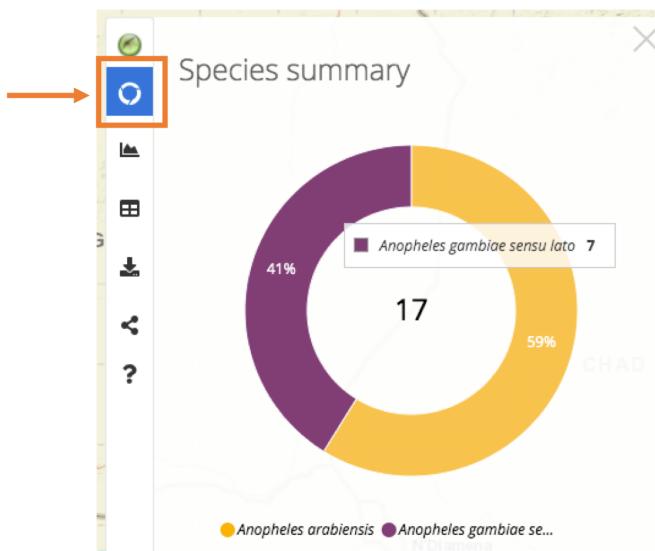


8. Click on a marker to obtain metadata about these samples and details about the values of insecticide resistance. For example, click in the one with the number 17

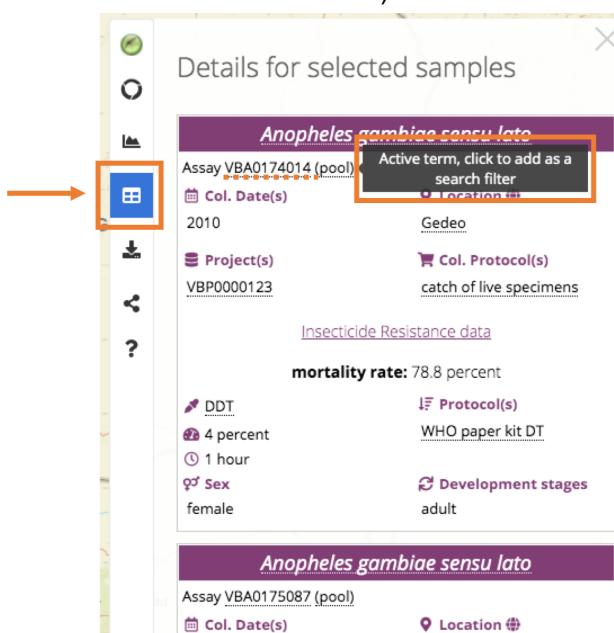


9. A side panel will open on the left of the map showing:

- a. Summary statistics for the species in percentages. Hover the mouse over the donut/plot to see the raw numbers



- b. Metadata details for the selected samples (You could hover over and click the dotted lines to add the active term as a search filter)



10. Apply filters in the right hand side menu:

- Select the '**Species**' of interest to add it as a search term in the top box (or type the name directly in the search box)

The screenshot shows a search interface with a sidebar on the left containing dropdown menus for 'Species' (with options for Anopheles arabiensis and Anopheles gambiae sensu lato), 'Sample type', 'Collection protocol', 'Project', and 'Protocol'. A red arrow points from the 'Species' dropdown to the top search bar, which now displays 'Ethiopia x Anopheles gambiae sensu lato x Search...'. To the right of the search bar are two small icons.

- Select the assay '**Protocol**' of interest to add it as a filter term in the top box (or type the name directly in the search box)

This screenshot illustrates the process of applying multiple filters. It shows a sidebar with 'Protocol' selected (indicated by a red box). A red arrow points from the 'Protocol' dropdown to the top search bar, which now includes 'WHO paper kit DT x'. Another red arrow points from the bottom search bar to the top search bar, indicating the cumulative effect of the applied filters: 'Ethiopia x Anopheles gambiae sensu lato x WHO paper kit DT x Search...'.

- Select '**Insecticide**' as a filter. As with the previous filters, you could click on any of them to add them as query terms, but the purpose here is actually to see them all

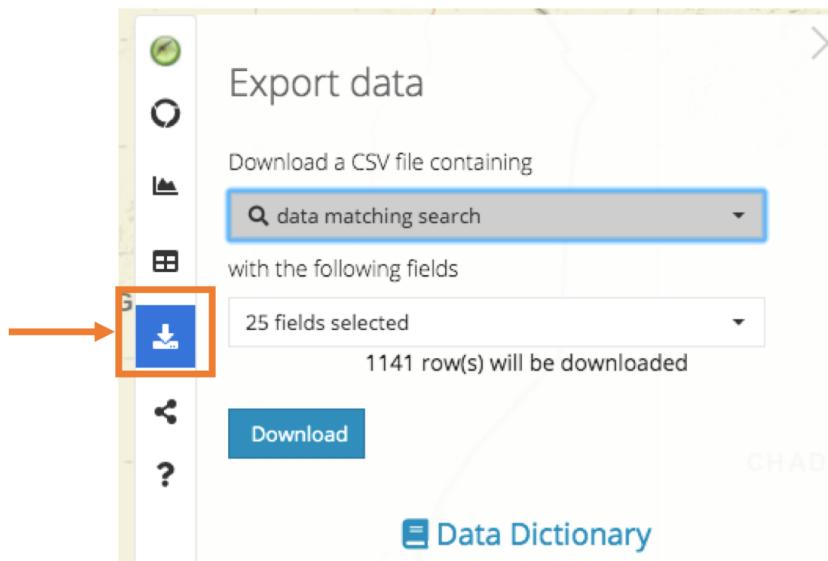
A detailed list of insecticides is shown in a sidebar, each with a color-coded circle and a count. The list includes Deltamethrin (160), DDT (156), Bendiocarb (149), Propoxur (143), Pirimiphos-methyl (129), Fenitrothion (129), Malathion (109), Lambda-cyhalothrin (50), Permethrin (42), Etofenprox (35), Alpha-cypermethrin (34), and Clothianidin (5). A red arrow points from the 'Insecticide' dropdown to the top search bar, which now includes 'WHO paper kit DT x'.

Insecticide	#
Deltamethrin	160
DDT	156
Bendiocarb	149
Propoxur	143
Pirimiphos-methyl	129
Fenitrothion	129
Malathion	109
Lambda-cyhalothrin	50
Permethrin	42
Etofenprox	35
Alpha-cypermethrin	34
Clothianidin	5

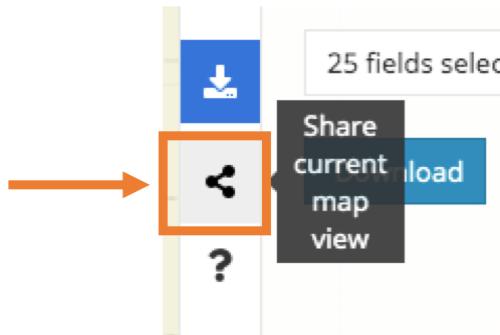
11. Click on 'Date Search'. Click on each date range. Delete (click on the x) the ones with no data. To activate all the date ranges with data click + command/control

The screenshot shows the 'Date search' section of the interface. It features a search bar with '2005-2009,2010-2014,2015-now x'. Below this are several date ranges: '1985-1989', '1990-1994', '1995-1999', '2000-2004', '2005-2009' (which is highlighted in blue), '2010-2014', and '2015-now'. There are also buttons for 'Insecticide Resistance', 'Seasonal search', and 'Global suggestions'.

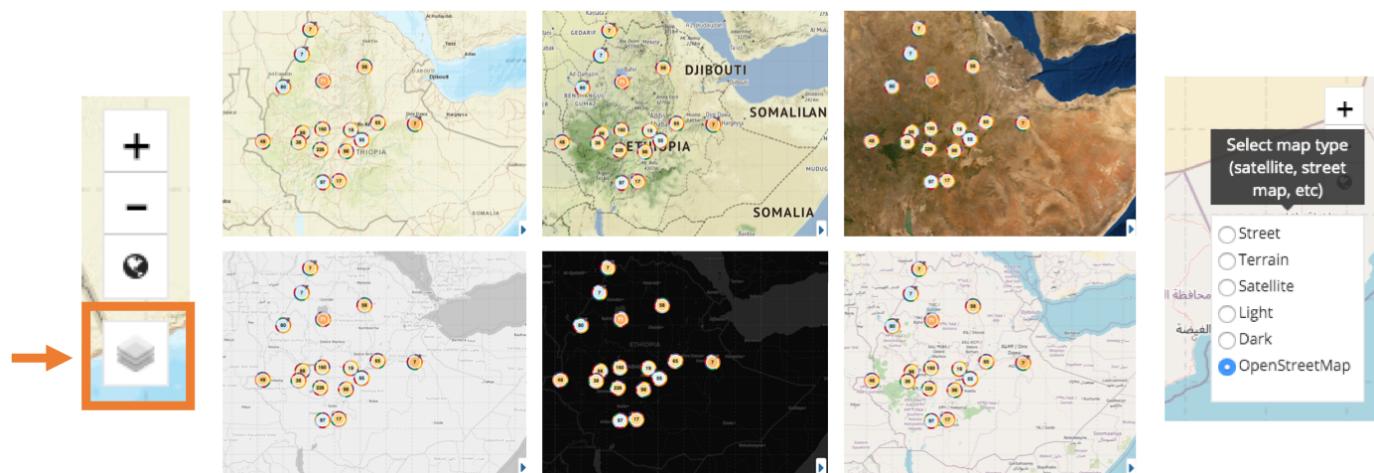
12. Download the csv file for the ‘data matching search’ and all ‘25 fields selected’, which can be opened in programs such as Excel



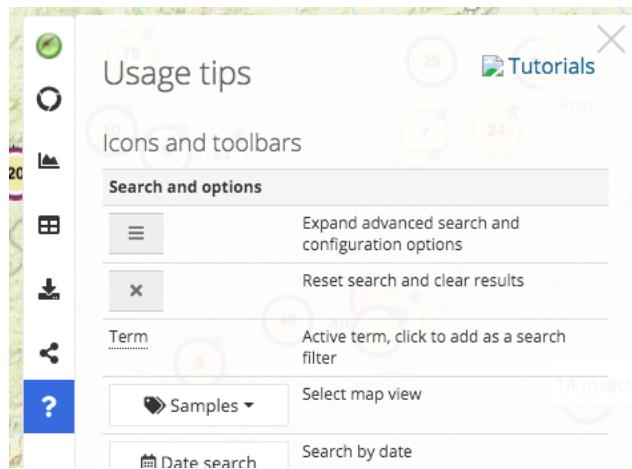
13. Share the map link in your paper or with a colleague, the link is ‘interactive’ and not just a static screenshot



- Your colleague can edit the map too, including change its background



14. The ‘Data Dictionary’ (Step 12), ‘Usage tips’ and ‘Tutorials’ options provide help about the use of the map



## Results interpretation

From the map visual interface we can see that there are 1141 assays summarized by insecticide, tested with WHO paper kit DT, for *Anopheles gambiae sensu lato* in Ethiopia. These samples have a mix of resistant and susceptible phenotypes

- How many different insecticides have been tested?: Twelve, the specific list can be found in figure 10.c
- In what year did the data collection start and finish?: 2006 to 2017, detailed years in the csv file

From the csv file we downloaded, the following questions can be answered:

- Are there insecticides with a mortality indicating insecticide susceptibility (i.e.,  $\geq 98\%$  mortality)?: There are nine insecticides with a mortality indicating insecticide susceptibility: bendiocarb, clothianidin, deltamethrin, fenitrothion, lambda-cyhalothrin, malathion, permethrin, pirimiphos-methyl and propoxur
- Are there any insecticides with an increase in mortality suggesting possible resistance (i.e., 90–97% mortality)?: No
- For the last year on record, has there been any confirmed cases of resistance (i.e., <90% mortality)?: No. In 2017, the only insecticide tested was Clotianidin and it showed 100% mortality
- How do I cite the data?: in the csv file there is a column called citations with DOIs and PubMed IDs if available

## **How to cite MapVEu data in your publications?**

- Submitting to MapVEu and referencing your own data:

## Data availability.

Microsatellite and SNP genotypes were deposited in VectorBase<sup>48 49</sup>, PopBio projects: VBP0000201 (new data), and VBP0000138, VBP0000176-177 (previously published data). Sequencing data were deposited in NCBI under accession numbers [MF371160-MF371174](#) and [MG241351-MG241354](#).

Gloria-Soria, A., Lima, A., Lovin, D.D., Cunningham, J.M., Severson, D.W., & Powell, J.R. (2018). Origin of a High-Latitude Population of *Aedes aegypti* in Washington, DC. *The American journal of tropical medicine and hygiene*, 98(2), 445-52.

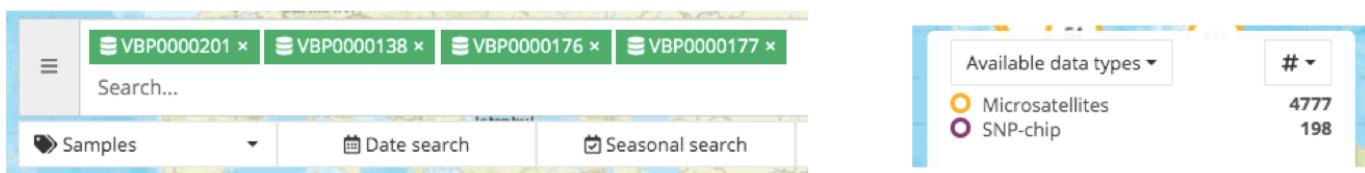
- Using and citing data submitted by others:

combinations of keywords. Additionally, we searched for data meeting our criteria in the following data repositories: BioTIME(37), GPDD(38), the LTER repository ([www.lternet.edu](#)), Knowledge Network Biocomplexity (<https://knb.ecoinformatics.org/>), the LTREB database, the Global Biodiversity Information Facility ([www.gbif.org](#)) and VectorBase (39). This yielded 166 studies from which data could be extracted. We extracted time series of insect abundance and /

Van Klink, R., Bowler, D.E., Gongalsky, K.B., Swengel, A.B., Gentile, A., & Chase, J.M. (2020). Meta-analysis reveals declines in terrestrial but increases in freshwater insect abundances. *Science* (New York, N.Y.), 368(6489), 417-20.

## How can a reader from your paper can find the data in MapVEu?

This is how a reader from Gloria-Soria et al 2018 (reference above), could query for the data: In the 'Samples' view, type the VectorBase project IDs, VBP\* in the search box. In this case, the filters show that the data are classified as microsatellites and SNPs





5 May 2020

## MapVEu: part II - Additional features

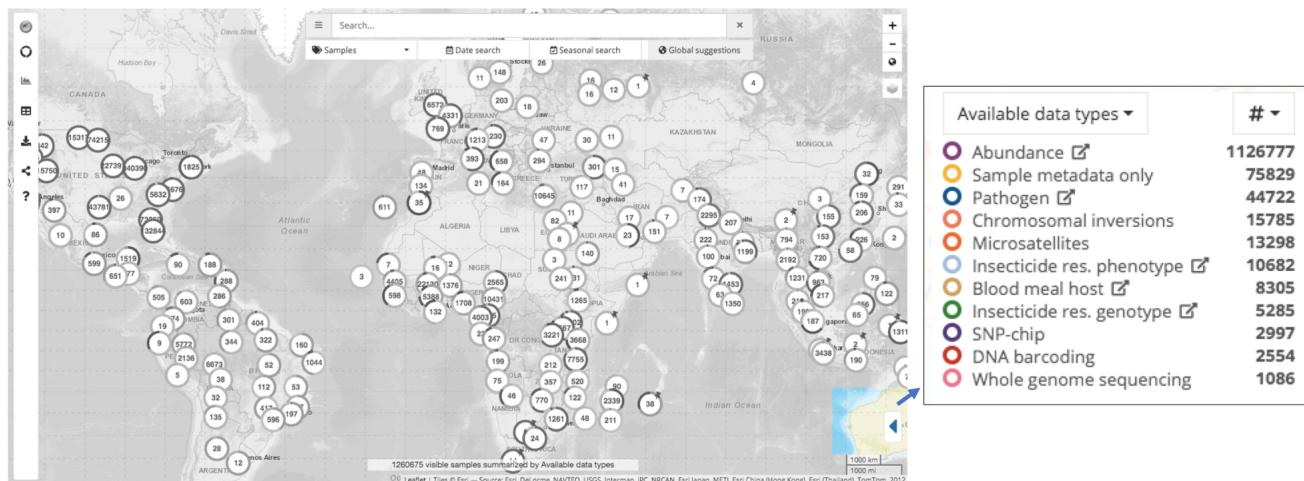
**Note:** this exercise uses VectorBase.org data as an example, but the same functionality will soon become available for other VEuPathDB resources

### Learning objectives:

Basic/introductory features have been covered in [part I](#) of this tutorial

- Use date and season search
- Construct queries with 'NOT' searches & reset queries
- Interpret different tables and figures and their corresponding metadata

The map has worldwide field collected samples, some with omic/genetic related test performed



### Omic/genetic tests:

- Sample metadata only
- Chromosomal inversions
- Microsatellites
- **Insecticide resistance genotypes:** identification of polymorphisms that confer resistance or susceptibility
- SNP chip
- DNA barcoding
- Whole genome sequencing

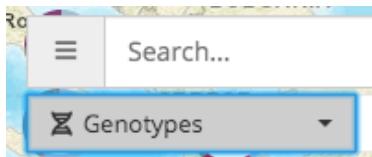
### Other test or measures:

- **Population abundance:** how many arthropods were collected in a specific location?
- **Pathogen infection status:** what virus or parasite were the arthropods infected with?
- **Insecticide resistance phenotypes:** are the arthropods exposed to insecticides susceptible or resistant?
- **Blood meal host:** what was the source of the arthropod blood meal? Did it bite a human or which other animal?

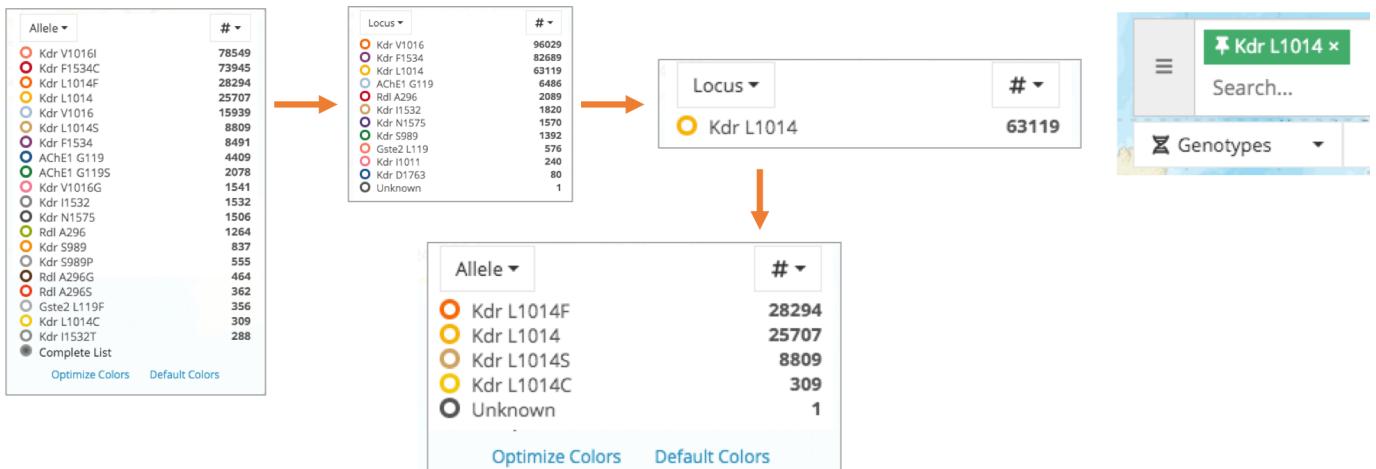
In part I of the tutorial we look at 'Samples' and 'Insecticide Resistance' phenotypes, here we will look at insecticide resistance 'Genotypes' and population 'Abundance'. In the last page there are links to [\*\*bonus cases\*\*](#) with all map views

## Use case #1: what is the distribution of the DDT and pyrethroid insecticide susceptibility based on the knockdown resistance (kdr) mutations, for the genus *Anopheles*?

- Let's go to the Genotypes view. It shows the prevalence of locus and their corresponding alleles from genotype assays. Currently mostly for insecticides but it can display any other type of genotype.



- By default it opens in Allele, change to Locus. In the Locus let's select kdr L1014, this action adds the term as query in the search box. Go back to Allele, this time it only shows the alleles for this locus.



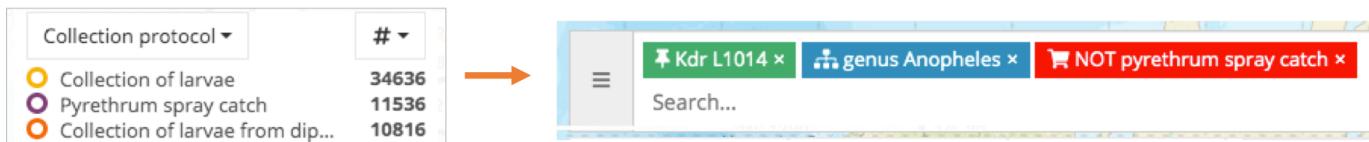
- Click on 'optimize colors' for a better color representation. The purple Allele (Kdr L1014) is the wild type (with L-leucine), the others are the mutants (with F-phenylalanine, S-serine and C-cysteine).



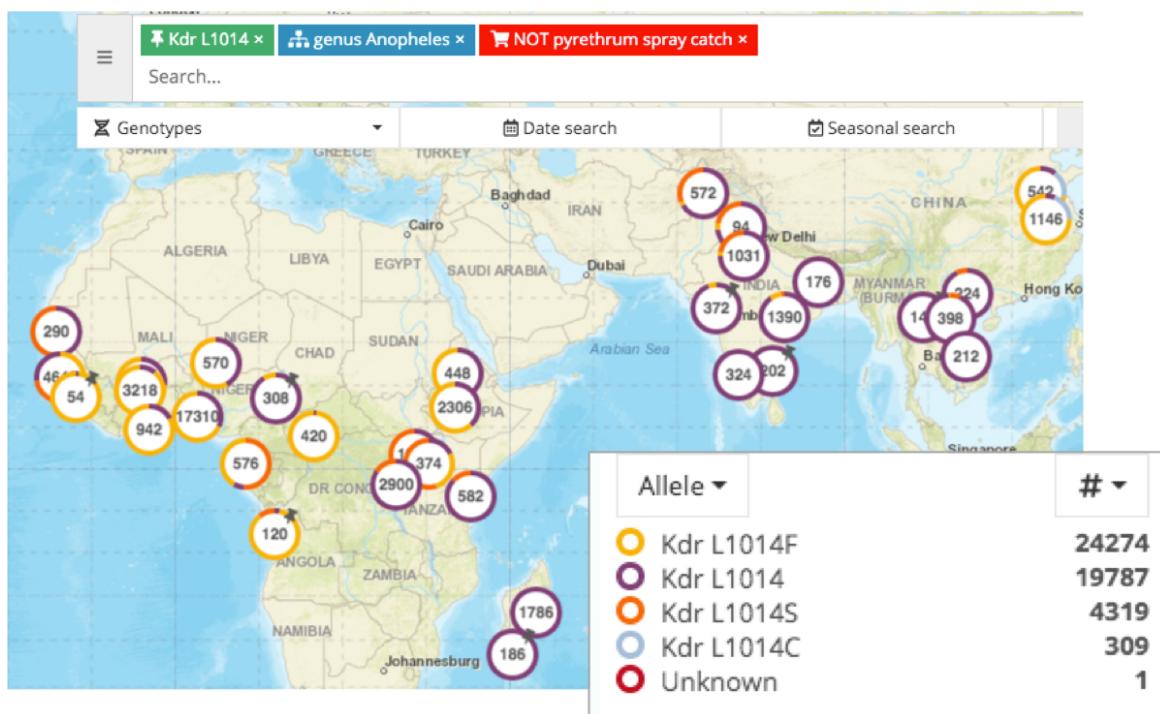
- Filter only for the genus *Anopheles*.



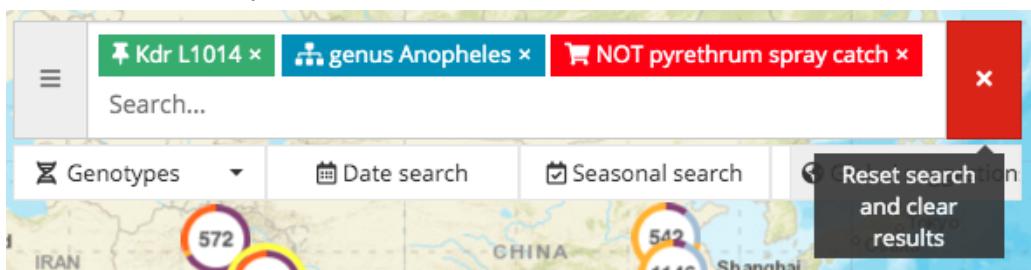
5. Filter for the collection protocol, including all except 'pyrethrum spray catch' with control (Windows) / command (Mac) + click. We perform this filter because we are looking at insecticide resistance genotypes and that collection protocol, using pyrethrum, creates a bias in our results



**Results interpretation case #1:** In India for example, results show that the wild type alleles are mostly located towards the center and South of the country, with a higher prevalence compared with the mutants in the North. In contrast, in continental Africa, the difference seems to be East to West, with a lower proportion of the wild type in the East. In Madagascar, there are only wild type alleles.

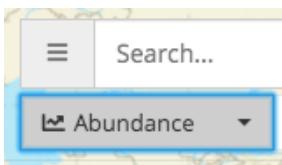


6. Clear or delete all queries and filters with a click in the search box right hand 'X'

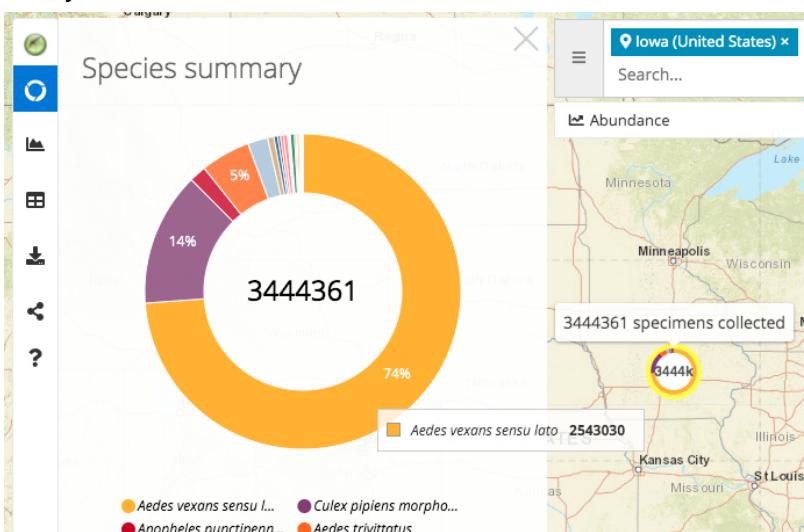


**Use case #2:** Currently most of the population abundance data come from the USA mosquito control abatement districts, and we are expecting soon to increase the mosquito data from Africa too. Most of the non-mosquito data are the *Culicoides* data coming from the UK. The map can handle any arthropod vector or related invertebrate species of interest. Let's explore some trends of the mosquito population abundance data in the USA for the state of Iowa

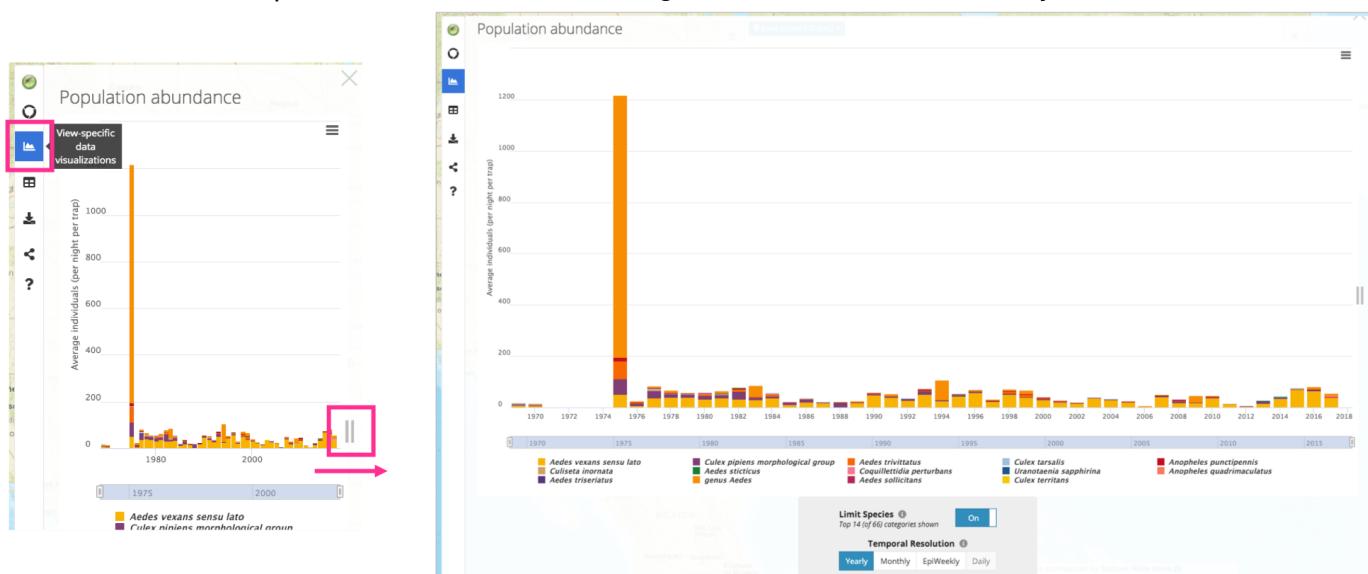
7. Let's go to the population "Abundance" view.



8. Query the data for the state of Iowa. Notice most of the data is from *Aedes vexans*.

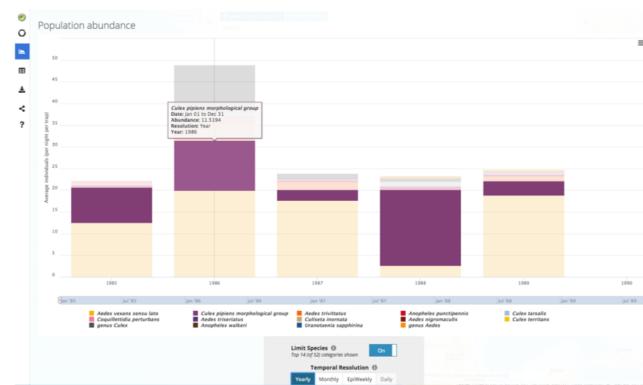
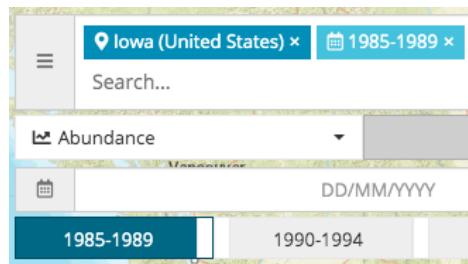


9. Now let's explore the species prevalence in time. On the left of the map, open the timeline graph. Using the double lines, expand it to show a yearly view. There is data from 1969 to 2017, with an unusual data peak in 1975 and data missing from 1971 to 74. This is 44 years of data!

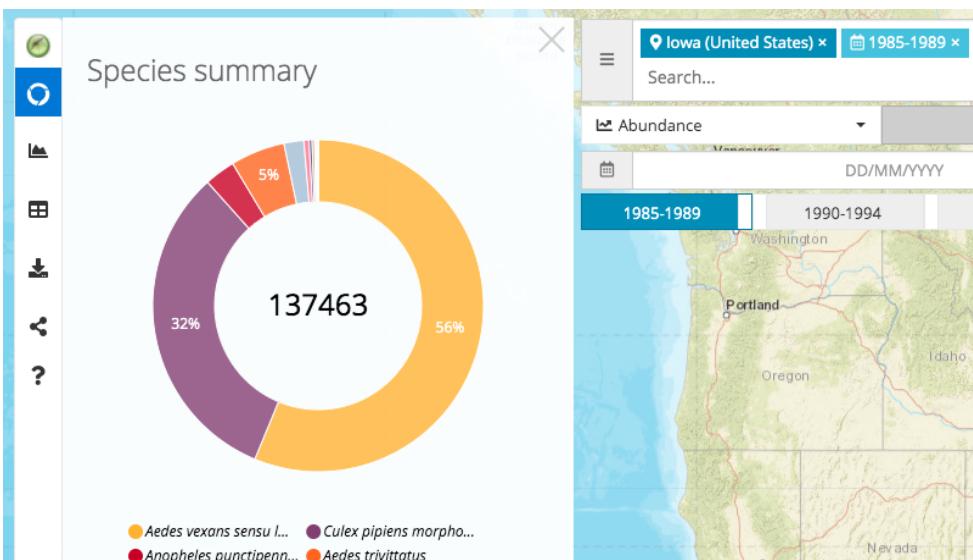


10. What are the most prevalent species in each date range? Select one range at the time, open and expand the timeline graph, and change the temporal resolution to year. Hover with the mouse over the bars, to identify the most prevalent species.

- a. 1985 - 1989: this shows that the two most prevalent records are from two species complexes, *Aedes vexans* and *Culex pipiens*.

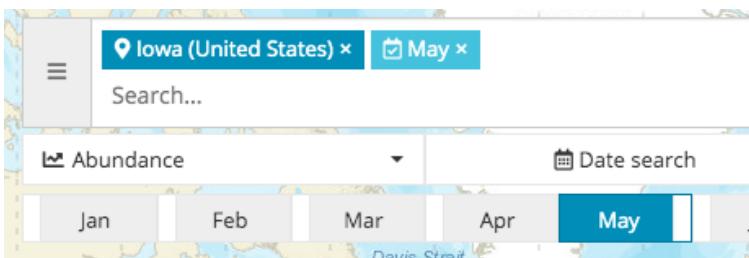


- b. 1985 - 1989: To know the exact species distribution for this date range, go back to the pie chart.



- c. We could repeat these steps for each date range, to identify any possible species patterns.

11. Data can also be explored at lower geographic resolution, looking for patterns from north to south and east to west in the State
12. To explore species abundance patterns of increase and decrease monthly, you could look at the data one month at the time



13. Metadata for each record is shown in the sample details, where the dotted lines indicate filters that can be applied in the top search box or the right hand side filter.

**Left Screenshot: Details for selected samples**

Sample VBS0773652 (pool)  
**Col. Date(s)**: Sat, May 31, 2003  
**Location**: Marshall (Iowa (United States))  
**Project(s)**: VBP0000388  
**Col. Protocol(s)**: new jersey trap catch  
**Attractant(s)**: light

**Abundance data**  
**Abundance:** 0 individual(s) per day  
**Protocol(s)**: morphological examination  
**Sex**: female  
**Development stages**: adult

**Right Screenshot: Species abundance data**

Species	#
<i>Aedes sensu lato</i>	9814
<i>Aedes morphological group</i>	365
<i>Aedes trivittatus</i>	247
<i>Aedes albifasciatus</i>	36
<i>Aedes punctipennis</i>	23
<i>Aedes tornata</i>	12
<i>Aedes sapphirina</i>	10
<i>Aedes eriatus</i>	10
<i>Aedes vexans</i>	7
<i>Aedes vexator</i>	3
<i>Aedes horrida</i>	1
<i>Aedes ciliata</i>	1
<i>Aedes pomonaculus</i>	1

## A scientific paper is a static view of the data, in contrast when you submit to MapVEu (VectorBase.org):

- the data on the map gets updated every two months
- the map allows data visualization with different filters
- the data can be analysed at different geographic levels
- the data from both scientific research and control/monitor agencies can be compared
- your papers gets more exposure and users can find it both with a literature search or with a query in MapVEu

- if a reader from your papers wishes to have your raw data, they do not need to contact you, it can be download from MapVEu

## **Bonus cases:** Try these queries using MapVEu

### Samples:

- PCR-based species identification
- animal baited net trap catch, genus *Anopheles*
- genotyping by high throughput sequencing, pyrethrum spray catch

### Insecticide Resistance:

Insecticide resistance assay results can be filtered by species, sample type, collection protocol, project, assay protocol and insecticide.

- *Anopheles gambiae*, West Africa, malathion
- WHO larvicide dose response test, temephos, collection of larvae
- Africa + sample type (individual or pool)
- Pyrethroid + genus *Aedes* + collection protocol (ovitrap, CDC light trap, ...)
- CDC bottle bioassay + permethrin + collection of larvae + project (VBP\*)

### Genotypes:

Shows the distribution and proportion of insecticide resistance alleles. It can be filtered by a single locus or allele, species, sample type, collection protocol, project or assay protocol.

- Madagascar, Kdr L1014, *Anopheles arabiensis*
- *Anopheles gambiae*, *Anopheles gambiae* sensu lato, *Anopheles gambiae* x *Anopheles coluzzii*
- AChE1 G119, Rdl A296
- India (displaying the markers as 'Allele')

### Abundance:

Temporal abundance that can be filtered by species, sample type, collection protocol, project and assay protocol. We have large multi year datasets from regular USA mosquito control district surveillance. We also have records from other countries.

- State of Iowa, genus *Culex*, CDC gravid trap, Lyric Bartholomay
- VBP0000213, morphological examination, RT-PCR assay
- pool, house resting-daytime catch
- *Aedes albopictus*
- Location + sex + identification method + taxon + date range + trap + attractant + GPS provenance and precision
- Florida + BG-Counter trap<sup>1</sup>

---

<sup>1</sup> Traps that automatically identify the collected material (to Culicidae). We obtain the data directly from the manufacturers API

Pathogen:

The pathogen status view provides vector pathogen-status testing, which can be filtered by pathogen, infection status, vector species, sample type, collection protocol, project and protocol assay.

- West Nile virus, *Culex pipiens* group
- Query with no keywords: directly zoom in and out and move around the map
- *Plasmodium*

# Functional annotation in Apollo

There are two options to add function annotation in VEuPathDB:

- 1) Adding a user comment on the gene record page
- 2) Using the community annotation tool Apollo

## Functional annotation can involve:

- Adding or changing the description of a gene or product
- Assigning or changing a gene name/symbol
- Adding Gene Ontology (GO) terms
- Adding a publication
- Adding an EC number

In this manual we are showing you step by step how to add a gene name, product description and GO term to a gene in Apollo.

## 1) Accessing Apollo

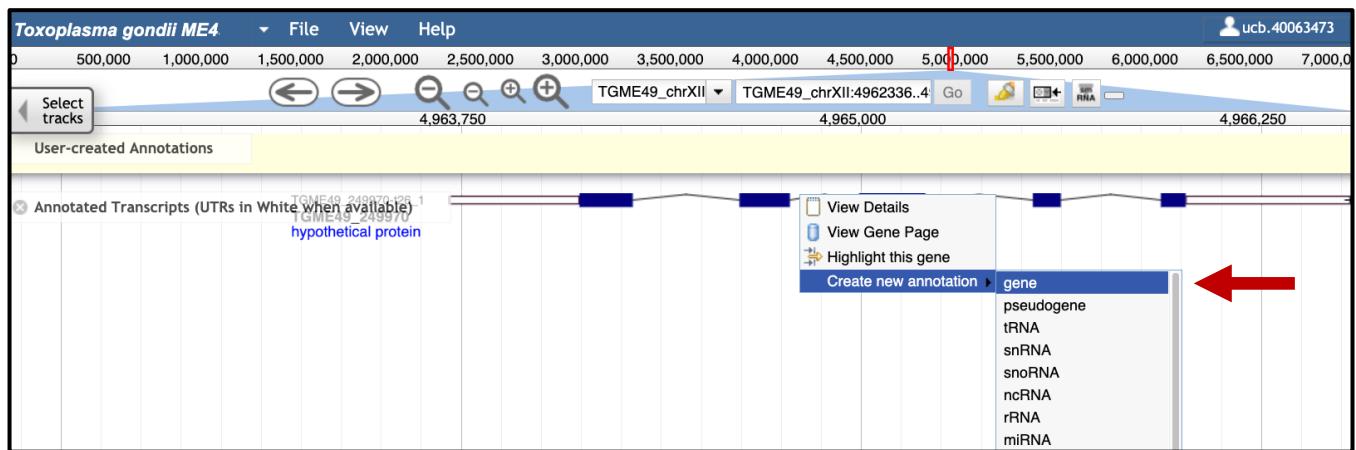
To access Apollo go to the gene record page of your gene of interest and click on the link **View and update community annotations in Apollo (1)**. You can also access Apollo from the gene models section by clicking on the button **Annotate in Apollo (2)**. Alternatively, go to the **Tools** menu and choose Apollo from the drop-down list (3).

The screenshot shows the ToxoDB gene record page for TGME49\_249970. At the top, there's a navigation bar with links like 'My Strategies', 'Search', 'Tools' (which is circled in red with '3'), 'Data', 'About', 'Help', and 'Contact Us'. Below the navigation, there's a search bar and some social media links. The main content area displays gene details: Type: protein coding gene, Chromosome: XII, Location: TGME49\_chrXII:4,963,712..4,966,931(+), Species: *Toxoplasma gondii*, Strain: ME49, and Status: Reference Strain. A red circle labeled '1' encloses the link 'View and update community annotations in Apollo'. Another red circle labeled '2' encloses the 'Annotate in Apollo' button in the 'Gene models' section. A red circle labeled '3' encloses the 'Tools' menu item. On the right side, there are various data visualization links like Synteny, Alignments, Phenotype, SNPs, Transcriptomics, Protein Features, and Proteomics.

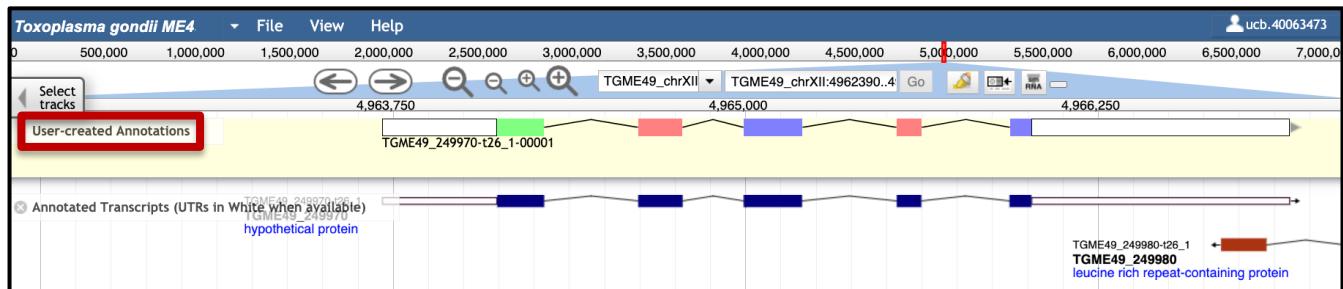
To use Apollo you need to be logged into VEuPathDB. If you have not done so yet log now into Apollo with your VEuPathDB user ID and password.

## 2) Dragging and dropping the annotation

Right click on the gene in the Annotated Transcripts track, select from the drop-down menu **Create new annotation > gene**.

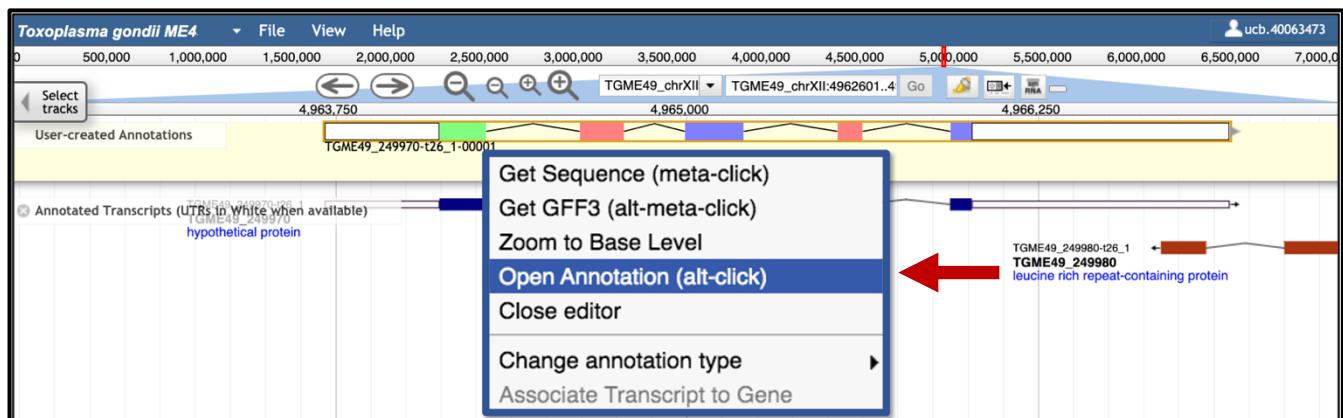


You can now see your gene of interest in the User-created Annotations track.



## 3) Opening of the Annotation editor window

Select the gene in the User-created Annotations track and with a right-click open the drop-down menu and choose **Open Annotation**. Alternatively, you can use the short-cut **alt-click**.



The annotation editor window is now shown on the right-hand side.

The screenshot shows the Toxoplasma gondii ME49 genome browser interface. On the left, a genomic track viewer displays a chromosome (TGME49\_chrXII) with various annotations, including a green bar labeled "User-created Annotations" and a red bar labeled "hypothetical protein". A red arrow points from the text "Annotation editor" to the right-hand side of the screen. On the right, the "Annotations" tab of the annotation editor is active, showing a table with one row for the gene. The table includes columns for Name, Seq, Type, Length, and Updated. The gene entry is TGME49\_249970-t26\_1-00001, with Type set to "gene". Below the table is a detailed form for editing the gene's properties. A red box highlights this form. A red arrow points from the text "You can either select the gene or the mRNA." to the "Type" field in the form, which currently has "gene" selected. The form also includes fields for Name (TGME49\_249970-t26\_1), Symbol, Aliases, Description, Location (4963712 - 4966931 strand(+)), Ref Sequence (TGME49\_chrXII), Owner (ucb.40063473), Created (May 27, 2021 03:13 PM), and Updated (May 27, 2021 03:13 PM). There are tabs for Details, GO, Gene Product, Provenance, DbXref, Comment, and Attributes.

You can either select the gene or the mRNA.

Please note functional annotation should be added to the gene. For genes with alternative transcripts add the information to the mRNA.

#### 4) Adding Functional annotation

In this example we are showing you how to improve the functional annotation of TGME49\_249970. This gene is currently annotated as hypothetical protein. It has been experimentally characterised in the following publication:

<https://pubmed.ncbi.nlm.nih.gov/26962945/>

Gene name/symbol: APH

Gene product: Acylated pleckstrin-homology domain-containing protein

GO term: microneme

PMID: 26962945

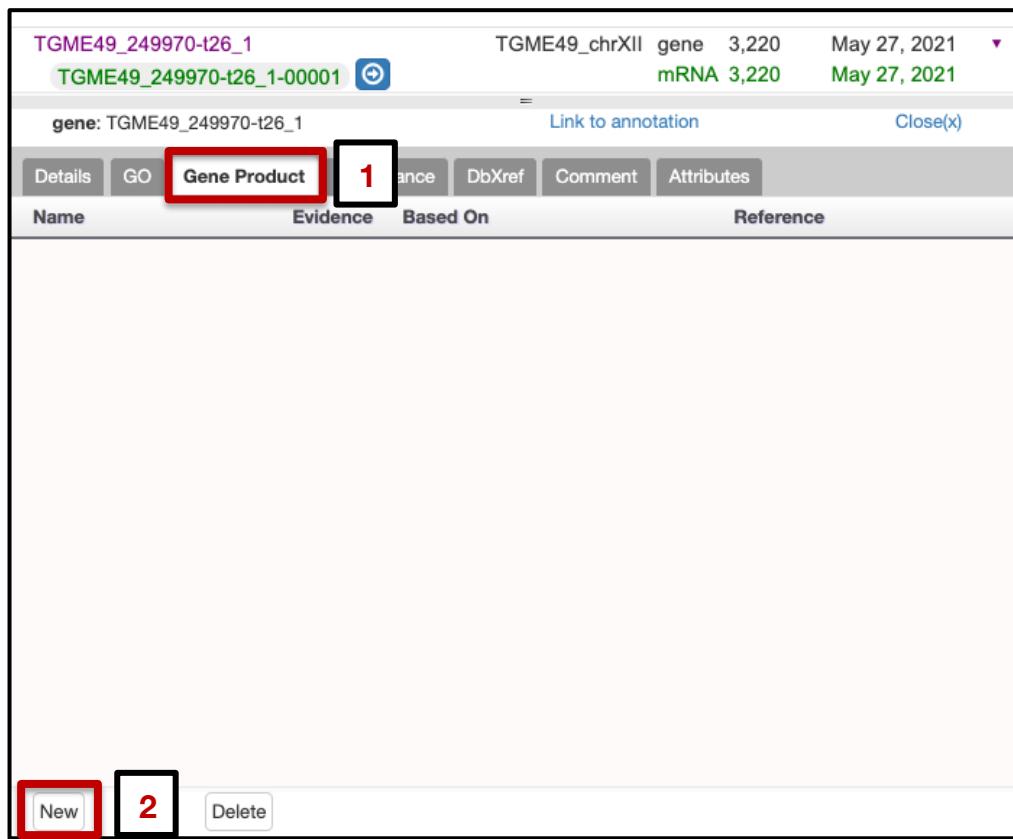
## Adding a gene name/symbol

Once the annotations panel (1) is open click on the details tab (2) and add the gene name/symbol in the field Symbol (3). In our example the new gene name/symbol is APH.

The screenshot shows the 'Annotations' panel for the 'Toxoplasma gondii ME49 [Mar 22, ]' dataset. A red box labeled '1' highlights the 'Annotations' tab in the top navigation bar. Below it, a red box labeled '2' highlights the 'Details' tab in the sub-navigation bar. A red circle with the number '3' highlights the 'Symbol' field, which contains the value 'APH'. Other fields visible include 'Name' (TGME49\_249970-t26\_1), 'Seq' (TGME49\_chrXII), 'Type' (gene), 'Length' (3,220), and 'Updated' (May 27, 2021).

## Adding a product description

To add a product description choose the tab Gene Product (1) and click on New at the bottom of the editor window (2).



Fill in the fields product, evidence and PMID. Click on Save. More information about evidence codes can be found here: <http://geneontology.org/docs/guide-go-evidence-codes>

The dialog box is titled 'Add new Gene Product to TGME49\_249970-t26\_1'. It contains several input fields and options:

- Product:** 'Acylated pleckstrin-homology domain'
- Evidence:** 'ECO:0000314' (with a checkbox labeled 'All ECO Evidence')
- ID:** 'IDA' (purple text)  
(ECO:0000314):  
direct assay  
evidence used in  
manual assertion  
Evidence Code Info
- With:** 'Prefix' and 'ID' fields, with a '+ Add' button between them.
- Reference:** 'PMID' field containing '26962945'.
- Note:** A text input field with a '+ Add' button.
- Buttons at the bottom:** 'Save' (highlighted with a red circle) and 'Cancel'.

## Adding GO terms

Choose the tab GO in the editor window, fill in the required fields and click on Save.

Add new GO Annotation to TGME49\_249970-t26\_1 X

**GO Annotation Guidance**

<b>Aspect</b> CC	<b>Go Term</b> GO:0020009 <i>microneme</i> (GO:0020009)	
cellular component		
<b>Relationship between Gene Product and GO Term</b> part of	<input type="checkbox"/> Not	<b>Evidence</b> ECO:0000314 <i>IDA</i> (ECO:0000314): direct assay evidence used in manual assertion Evidence Code Info
		<input type="checkbox"/> All ECO Evidence
<b>With</b>	<b>Prefix</b> :	<b>ID</b> <input type="button" value="+ Add"/>
<b>Reference</b>	<b>PMID</b> :	26962945
<b>Note</b>	<input type="button" value="+ Add"/>	
<b>Save</b> <input type="button" value="Cancel"/>		

## Adding a PubMed ID

Choose the Tab DbXref in the annotation editor window, add the PMID as shown in the screenshot.  
Click on the + sign.

gene: TGME49\_249970-t26\_1 Link to annotation Close(x)

Details	GO	Gene Product	Provenance	<b>DbXref</b>	Comment	Attributes
▲ Prefix	Accession		<input type="text" value="PMID"/> <input type="text" value="Accession"/> <input type="button" value="+"/> 26962945 <input type="button" value="+"/>			

A small window will come up showing the title of the Pubmed Article. Click OK.

Add article Phosphatidic Acid-Mediated Signaling Regulates Microneme Secretion in Toxoplasma. X

You can also add additional database identifiers (DbXref), i.e. EC numbers.

A screenshot of a software interface showing the 'DbXref' tab selected. Under the 'Accession' section, there is a row for 'EC' with the value '2.7.1.1' and a '+' button. Below it is a row for 'PMID' with a '+' button.

## 5) Finalising the functional annotation

Go to the **Attributes** tab in the gene section, choose from the “Select canned tag” drop-down menu annotation.

A screenshot of the 'Attributes' tab. A red box highlights the 'Attributes' tab. A dropdown menu is open under the 'Tag' field, showing options: 'Select canned tag', 'structural', 'user\_comment', and 'annotation'. The 'annotation' option is highlighted with a blue background and has a red arrow pointing to it.

From the “Select canned value” drop-down menu choose **added\_product**. Repeat this and choose **added\_symbol**, **added\_go** and **added\_pmids**. Finally click on the + sign.

A screenshot of the 'Attributes' tab. A red circle highlights the '+' button next to the 'Value' field. A dropdown menu is open under the 'Value' field, showing options: 'annotation', 'Select canned value', 'added\_comment', 'split', 'merge', 'retain\_previous', and 'added\_product'. The 'added\_product' option is highlighted with a blue background and has a red arrow pointing to it.

gene: TGME49\_249970-t26\_1

Link to annotation

Close(x)

Details	GO	Gene Product	Provenance	DbXref	Comment	Attributes
▲ Prefix Accession						Tag Value <input type="button" value="+"/>
annotation added_symbol						Select can <input type="button" value="▼"/>
annotation added_go						Select can <input type="button" value="▼"/>
annotation added_product						
annotation added_pmid						

In the last step, go back to the Details tab and add to the gene and mRNA the status **Finished**.

gene: TGME49\_249970-t26\_1

Link to annotation

Close(x)

Details	GO	Gene Product	Provenance	DbXref	Comment	Attributes
<input type="button" value="⊕ Go"/> <input type="button" value="ID"/> <input type="button" value="Sync name with transcript"/> <input type="checkbox" value="Obsolete"/>	<input type="button" value="Delete"/>					
Type gene	Status	<ul style="list-style-type: none"><li><input checked="" type="checkbox"/> No status selected</li><li>Not Finished</li><li><b>Finished</b></li><li>Requires Curator</li></ul>				
Name TGME49_249970-t26_1	Symbol APH					

Done! For additional questions, please get in touch with the VEuPathDB help desk.

# Structural annotation in Apollo

## Modifying a gene model

In this short tutorial we are showing you step-by-step how to modify an existing gene model in Apollo. Modifying a gene model can include, adding additional exons, extending exons, deleting exons, splitting exons and adding UTRs.

### 1) Accessing Apollo

To access Apollo go to the gene record page of your gene of interest and click on the link **View and update community annotations in Apollo (1)**. You can also access Apollo from the gene models section by clicking on the button **Annotate in Apollo (2)**. Alternatively, go to the **Tools** menu and choose Apollo from the drop-down list (3).

The screenshot shows the ToxoDB gene record page for TGME49\_305150. At the top, there's a navigation bar with links for 'My Strategies', 'Searcher', 'Tools' (circled with a red box labeled 3), 'Gene', 'Data', 'About', 'Help', and 'Contact Us'. Below the navigation bar, there's a site search bar and a green banner for VEuPathDB. The main content area displays gene details: Type: protein coding gene, Chromosome: IX, Location: TGME49\_chrlX:5,245,620..5,257,794(+), Species: *Toxoplasma gondii*, Strain: ME49, Status: Reference Genome. There are also 'Shortcuts' for Synteny, Alignments, Phenotype, SNPs, Transcriptomics, Protein Features, and Proteomics. A note says 'Also see TGME49\_305150 in the Genome Browser or Protein Browser'. On the left, a sidebar shows a tree view of gene models, annotations, and identifiers, with a red circle around the 'View and update community annotations in Apollo' link (1). At the bottom right, there's a 'Gene models' section with a red circle around the 'Annotate in Apollo' button (2). A red circle also highlights the 'Tools' menu item (3) in the top navigation bar.

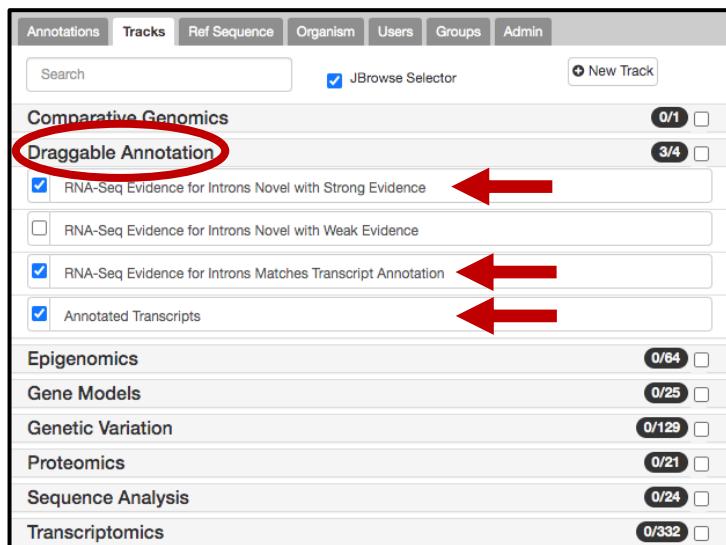
To use Apollo you need to be logged into VEuPathDB. If you have not done so yet log now into Apollo with your VEuPathDB user ID and password.

### 2) Adding draggable annotation and supporting evidence

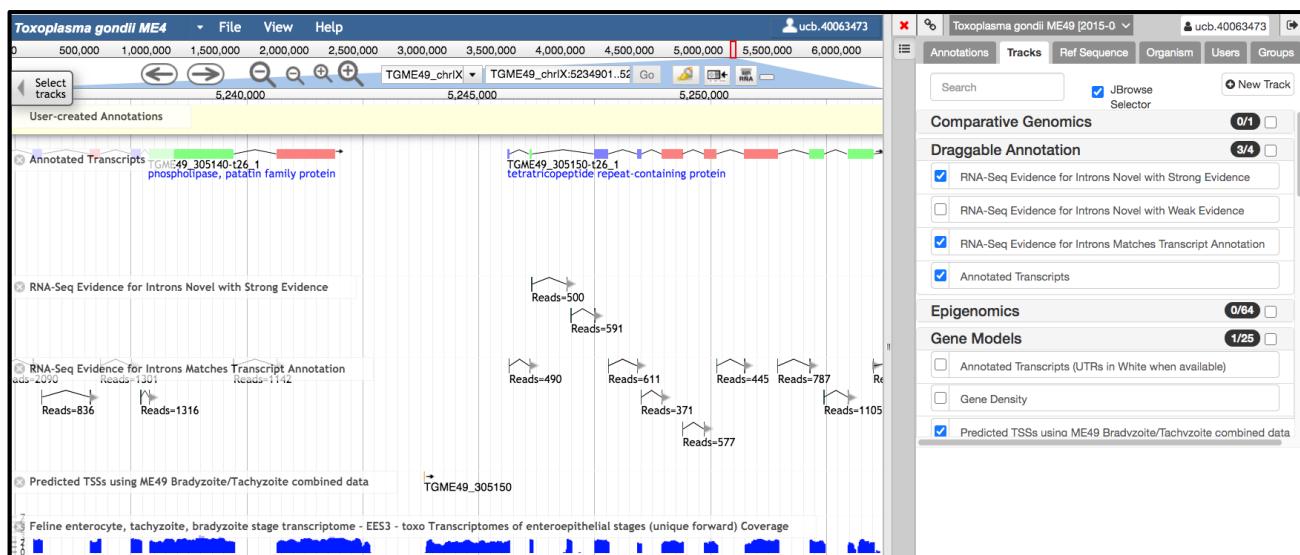
Select on the right-hand side the tab **Tracks**.

The screenshot shows the JBrowse genome browser interface for *Toxoplasma gondii* ME49. The top navigation bar includes 'File', 'View', 'Help', and a user profile. The main area shows a genomic track for chromosome IX, with a scale from 0 to 6,000,000. A red box highlights the 'Tracks' tab in the top right corner of the interface. The 'Annotations' tab is also visible. On the right, there's a panel for 'Comparative Genomics' and 'Draggable Annotation'.

Click on the menu item **Draggable Annotation** select **Annotated Transcripts, RNA-Seq Evidence for Introns Novel with Strong Evidence and RNA-Seq Evidence for Introns Matches Transcript Annotation.**

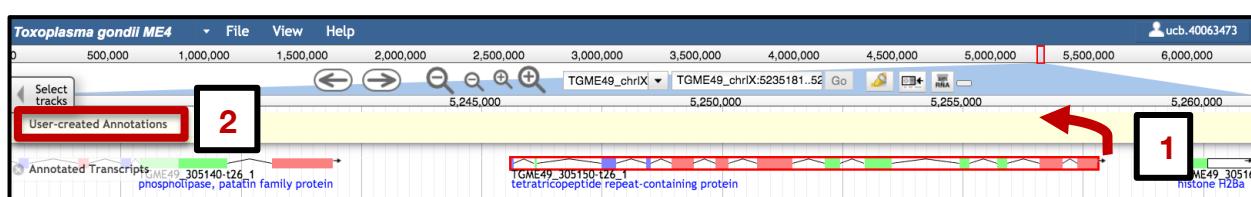


Select additional evidence, i.e. RNAseq plots or predicted TSS (transcription start sites).

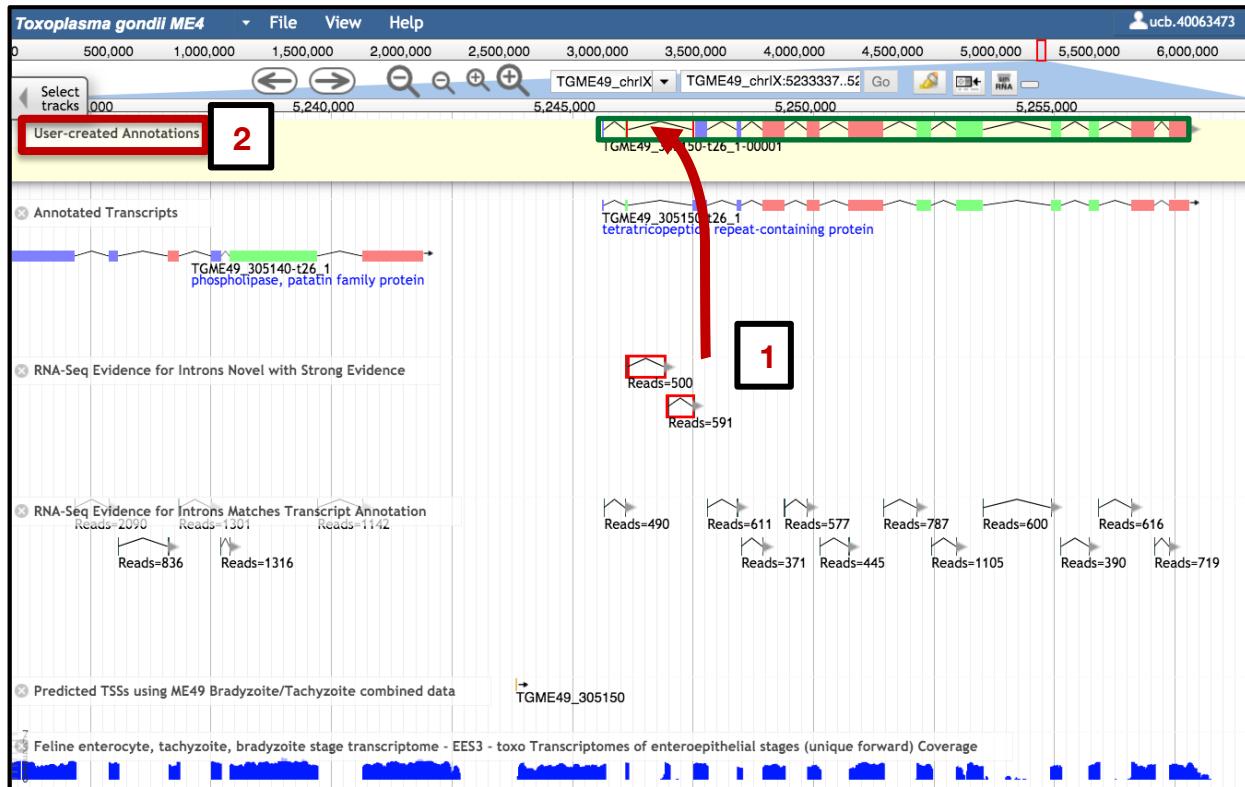


### 3) Modifying the gene

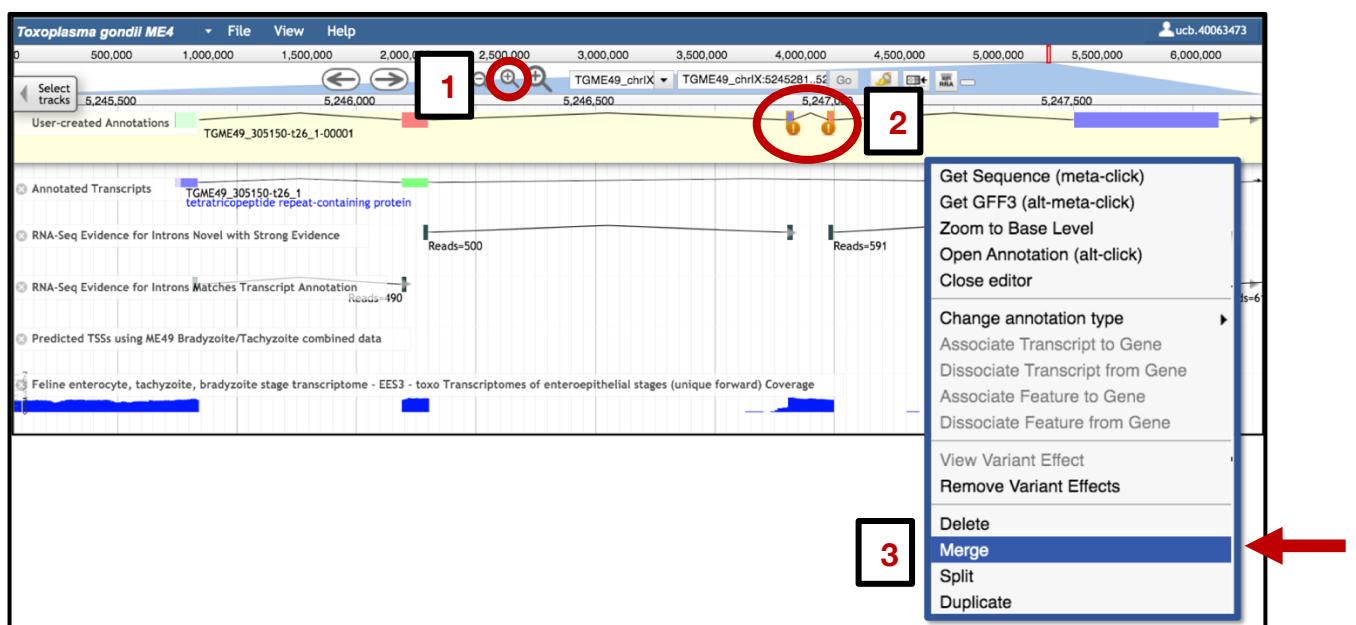
Select the gene model by clicking on one of the introns or by double clicking on the gene model (1). The gene model will show up with red boundaries. Drag and drop the gene into the User-created Annotations track (2).



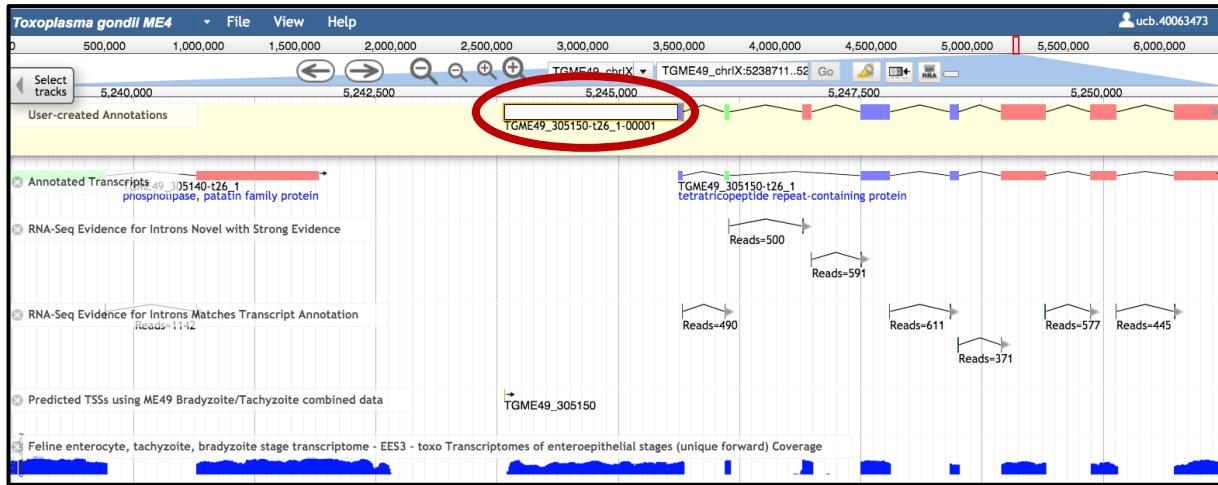
You can either select the intron junctions individually, or hold down the shift key and select both intron junctions with strong evidence (1), drag and drop them into the gene model (2). The gene will get a green box when dragging and dropping the intron evidence.



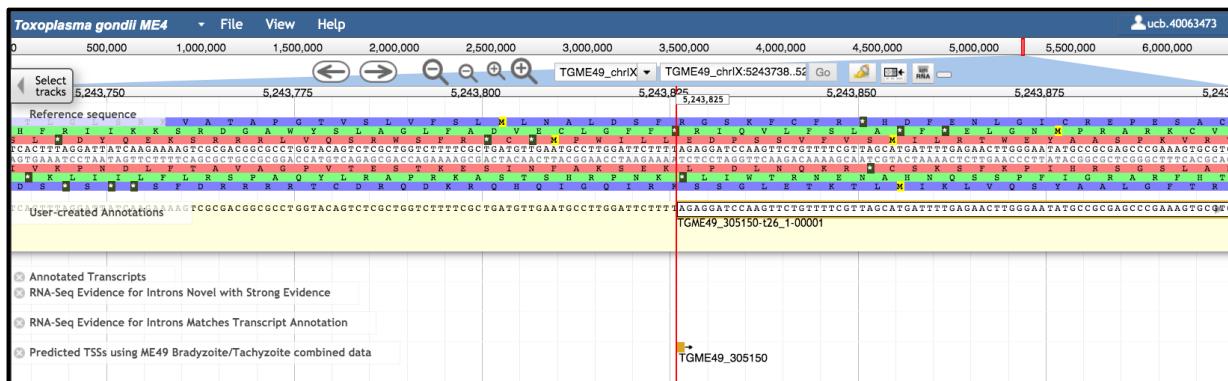
Zoom in by clicking on the + sign on the top (1). Press the Shift key and select the two small exons in the middle (2). With a right-click open the drop-down menu and choose **Merge** (3). Alternatively, select one of the exons you would like to merge, go to the edge of the feature until a little arrow appears and extend the exon until it overlaps with the second exon.



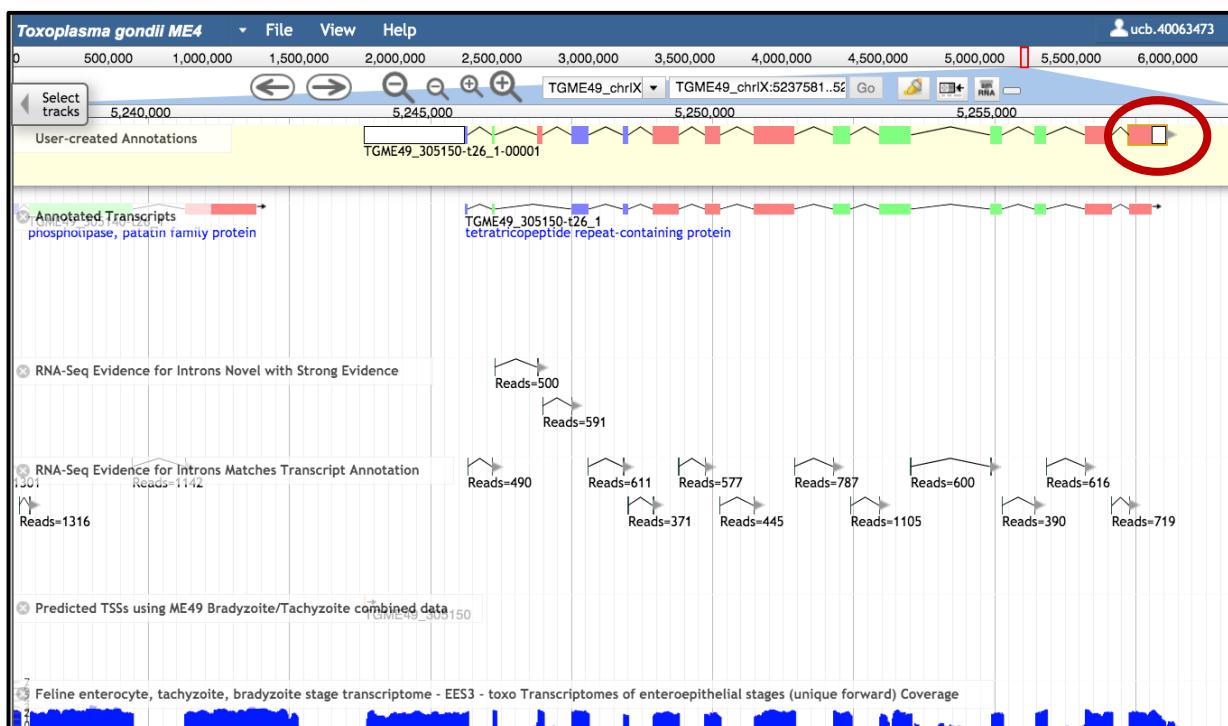
Select the first exon, point your mouse at the edge of the feature, a little arrow will appear, then extend the exon to the transcription start. Apollo will automatically create the 5'UTR!



You can zoom in to recheck the transcription start site.

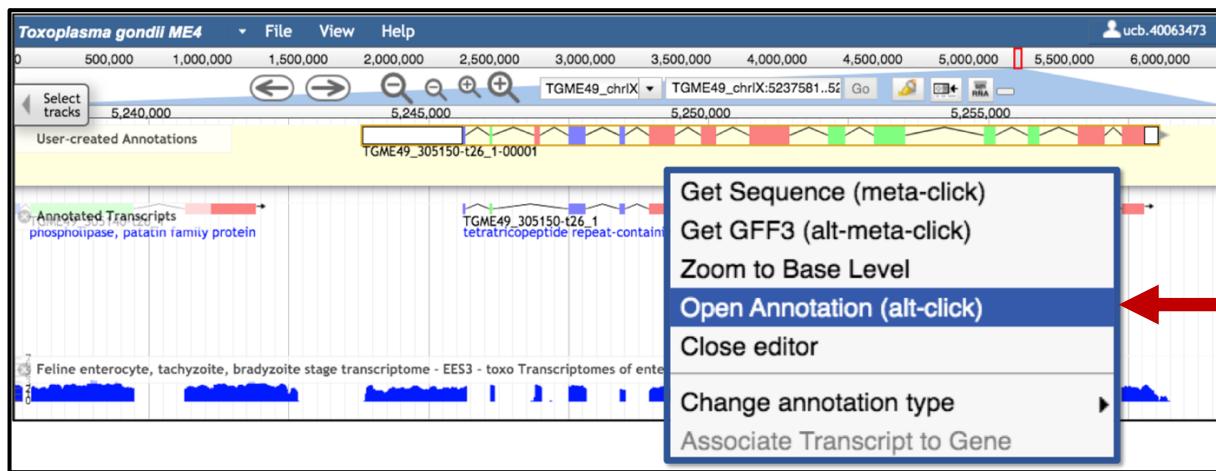


Select the last exon, point your mouse at the edge and extend the exon. Apollo will create the 3'UTR automatically!

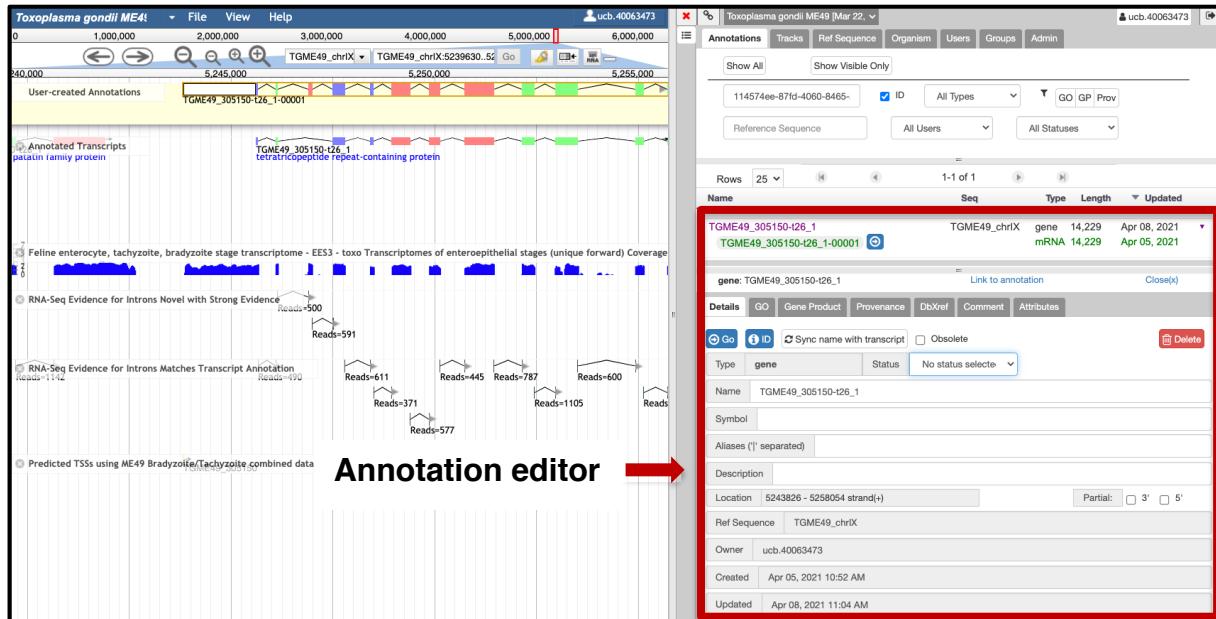


#### 4) Opening of the Annotation editor window

Select the gene in the User-created Annotation track and with a right-click open the drop-down menu and choose **Open Annotation**. Alternatively, you can use the short-cut **alt-click**.



The annotation editor window is now shown on the right-hand side.



#### 5) Finalising the structural annotation

Once the annotations panel is open click on the Attributes tab, select from the canned tag **structural** and from the canned value **modify**. Click on the + sign.

The screenshot shows the 'Attributes' tab of a gene record. The 'Attributes' tab is highlighted with a red box. Below it, there's a table with two rows: 'Prefix' and 'Accession'. To the right of the table is a dropdown menu titled 'structural' with a '+' sign next to it. The dropdown menu lists several options, each preceded by a checkmark and a brief description. The 'modify' option is highlighted with a blue background and a red arrow points to it from the right.

If there is no change in the functional annotation choose from the canned tag - **annotation** and from the canned value in the Attributes section **retain previous**. Click on the + sign.

The screenshot shows the 'Attributes' tab for a specific gene record (TGME49\_305150-t26\_1). The 'Attributes' tab is highlighted with a red box. Below it, there's a table with two rows: 'Prefix' and 'Accession'. To the right of the table is a dropdown menu with a '+' sign next to it. The dropdown menu lists two options: 'annotation' and 'retain\_previous'. The 'retain\_previous' option is highlighted with a blue background and a red arrow points to it from the right.

Finally go the Details tab and select the status **Finished** on the gene.

The screenshot shows the 'Details' tab for a gene record (TGME49\_305150-t26\_1). The 'Details' tab is highlighted with a red box. Below it, there are several input fields: 'Type' (gene), 'Status' (dropdown menu), 'Name' (TGME49\_305150-t26\_1), 'Symbol', 'Aliases', 'Description', 'Location' (5243825 - 5258054 strand(+)), 'Ref Sequence' (TGME49\_chrIX), 'Owner' (ucb.40063473), 'Created' (Apr 05, 2021 10:52 AM), and 'Updated' (Apr 05, 2021 10:52 AM). To the right of the 'Status' field is a 'Delete' button. A dropdown menu is open under the 'Status' field, listing four options: 'No status selected', 'Not Finished', 'Finished', and 'Requires Curator'. The 'Finished' option is highlighted with a blue background and a red arrow points to it from the right.

**Done!** For additional questions, please get in touch with the VEuPathDB help desk.

# Structural annotation in Apollo Merging/Splitting gene models

In this short tutorial we are showing you step-by-step how to merge/split gene models in Apollo.

## 1) Accessing Apollo

To access Apollo go to the gene record page of your gene of interest and click on the link **View and update community annotations in Apollo (1)**. You can also access Apollo from the gene models section by clicking on the button **Annotate in Apollo (2)**. Alternatively, go to the **Tools** menu and choose Apollo from the drop-down list (3).

The screenshot shows the ToxoDB gene record page for *TGME49\_319312*, a hypothetical protein. The top navigation bar includes links for My Strategies, Searcher, Tools (circled in red), My Works, Data, About, Help, and Contact Us. A guest user is logged in. Below the navigation is a search bar and a 'Site search' input field. The main content area displays gene details: Type: protein coding gene, Chromosome: IV, Location: *TGME49\_chrlV:1,041,310..1,050,287(+)*, Species: *Toxoplasma gondii*, Strain: ME49, and Status: Reference Strain. A red circle highlights the 'View and update community annotations in Apollo' link. A red box highlights the 'Annotate in Apollo' button in the gene models table. A white box highlights the 'Tools' menu item in the top navigation bar. The gene models table shows 1 Gene model with 9 exons and 1 transcript. A red number '1' is placed over the 'View and update community annotations in Apollo' link, and a red number '2' is placed over the 'Annotate in Apollo' button. A white box highlights the 'Tools' menu item in the top navigation bar.

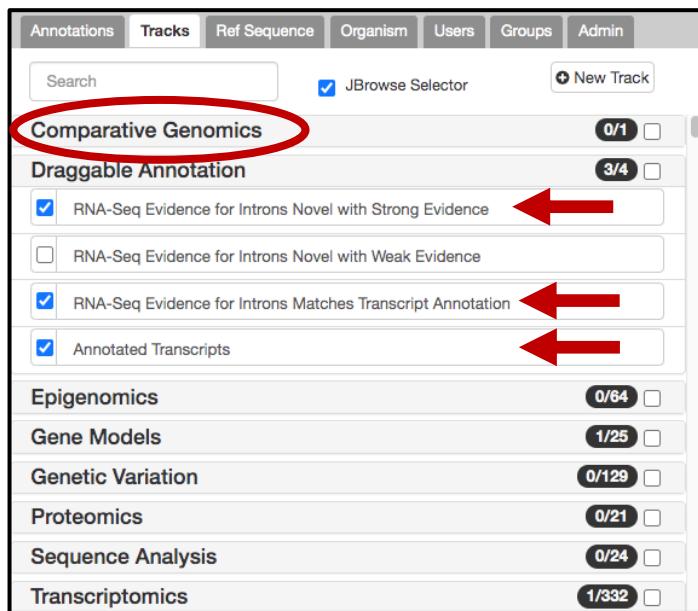
To use Apollo you need to be logged into VEuPathDB. If you have not done so yet log now into Apollo with your VEuPathDB user ID and password.

## 2) Adding draggable annotation and supporting evidence

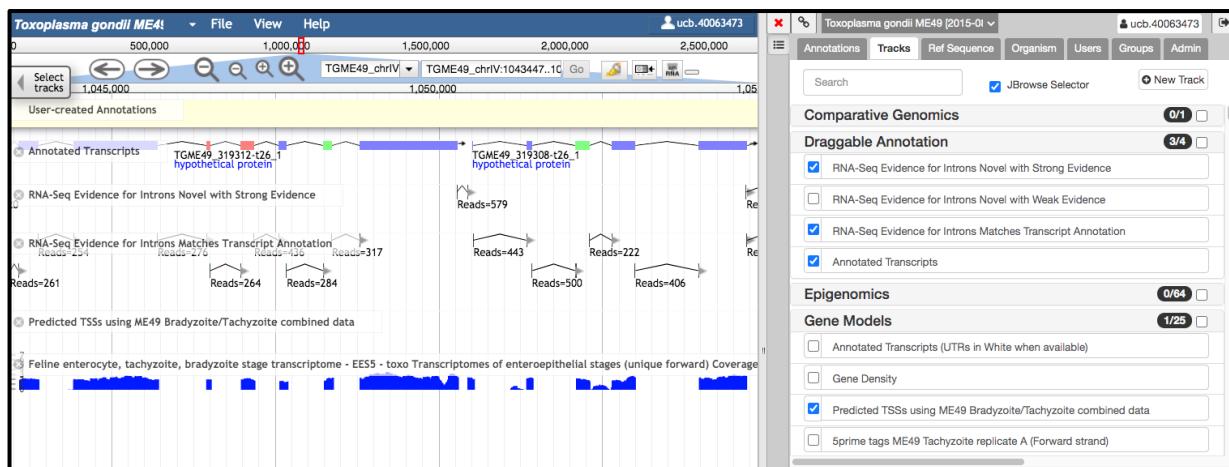
Select on the right-hand side the tab **Tracks**.



Click on the menu item **Draggable Annotation** select **Annotated Transcripts, RNA-Seq Evidence for Introns Novel with Strong Evidence and RNA-Seq Evidence for Introns Matches Transcript Annotation**.

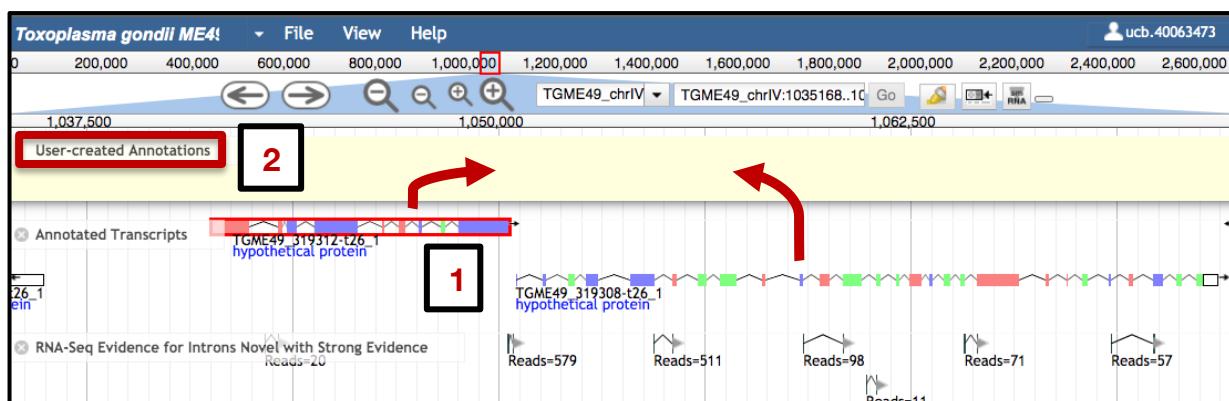


Select additional evidence, i.e. RNAseq plots and predicted TSS (transcription start sites).

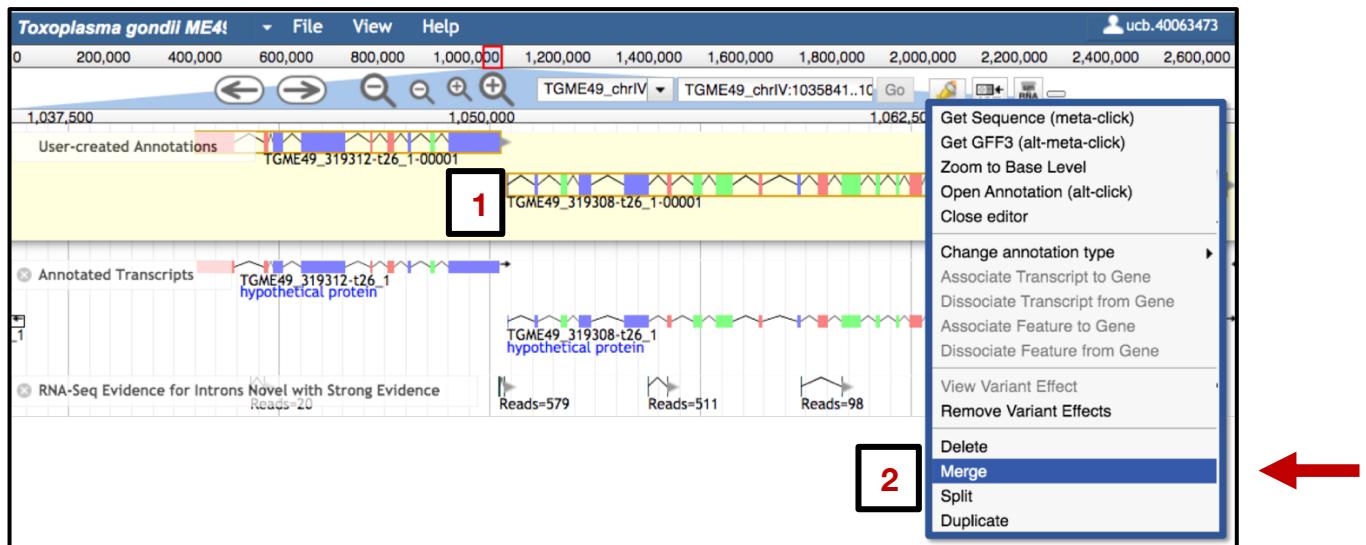


### 3) Merging genes

Select the gene models that you would like to merge by clicking on one of the introns or by double clicking on the gene model (1). Drag and drop the genes into the User-created Annotations track (2).

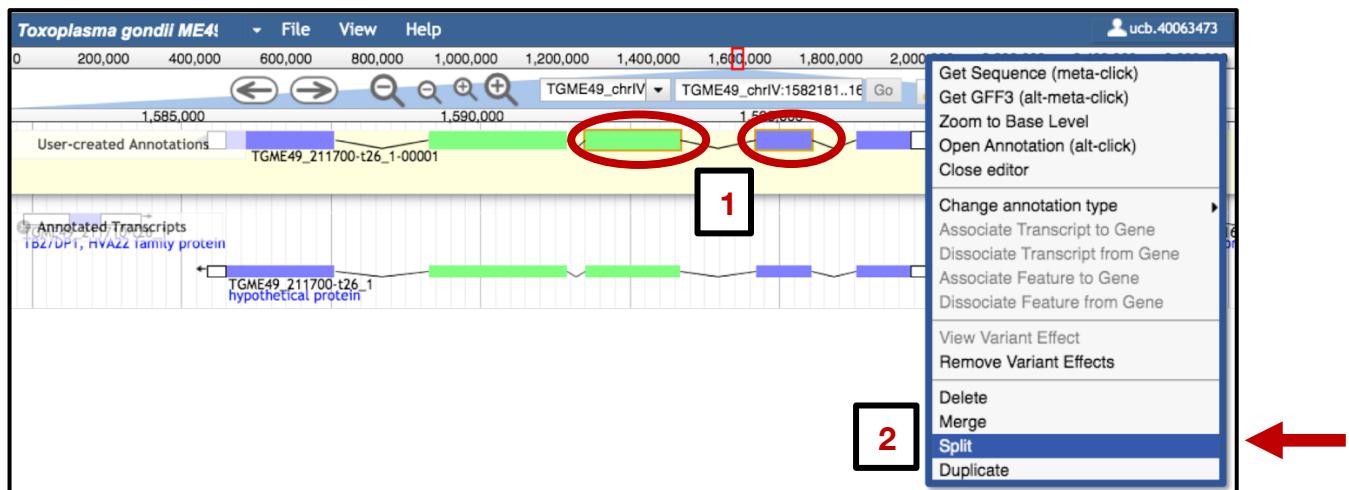


Hold down the shift key and select both gene models in the User-created Annotations track (1). With a right-click open the drop-down menu and choose **Merge** (2).



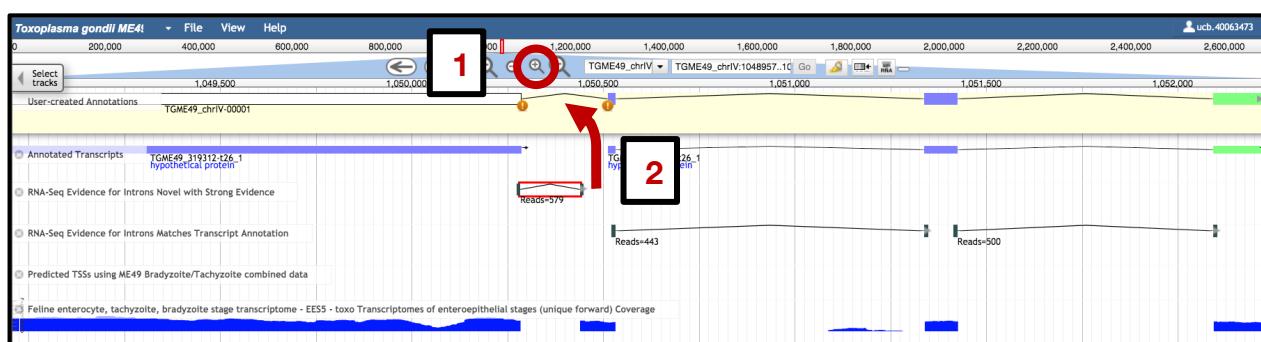
### 3.1) Splitting gene models

For splitting gene models, select the exons that border the intron that should be split. With a right-click open the annotation drop-down menu and choose split. Once you've split the gene model, recheck if the gene model has the correct start and stop.

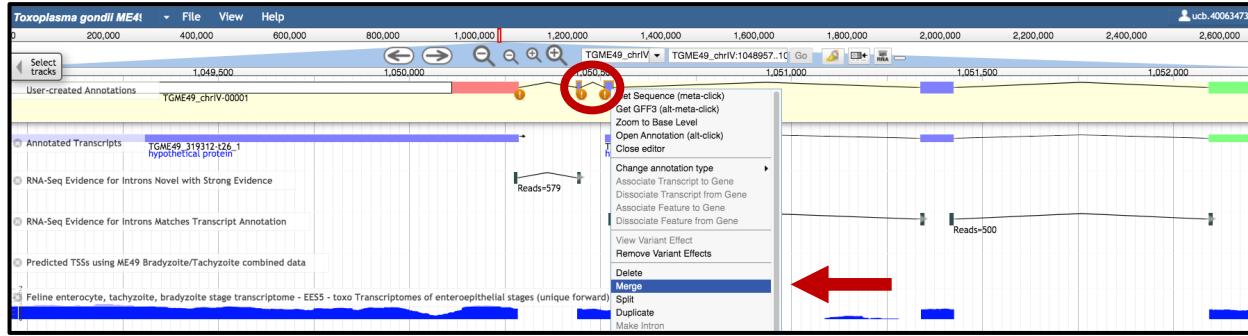


### 4) Correcting intron-exon boundaries

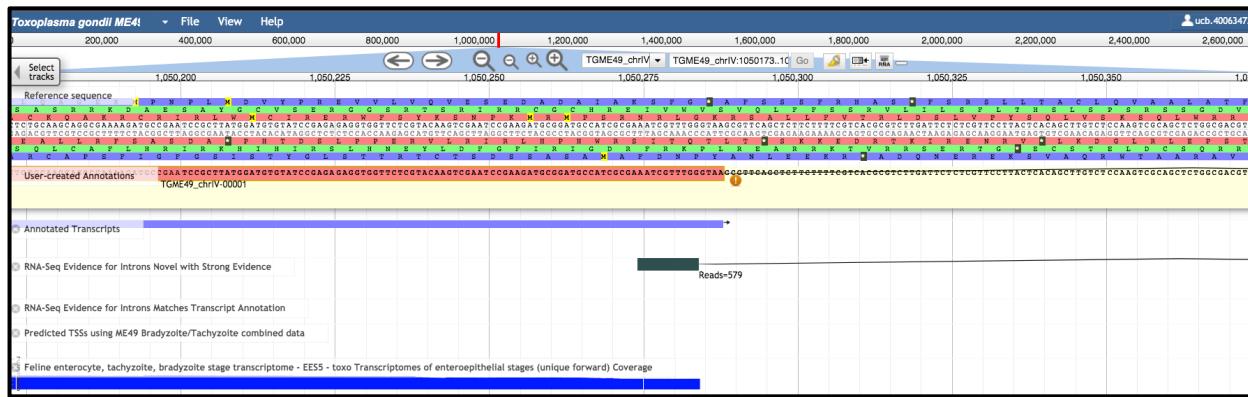
Once you've merged the gene zoom in by clicking on the + sign on the top (1). Select the new splice junction and drag it into the gene model (2).



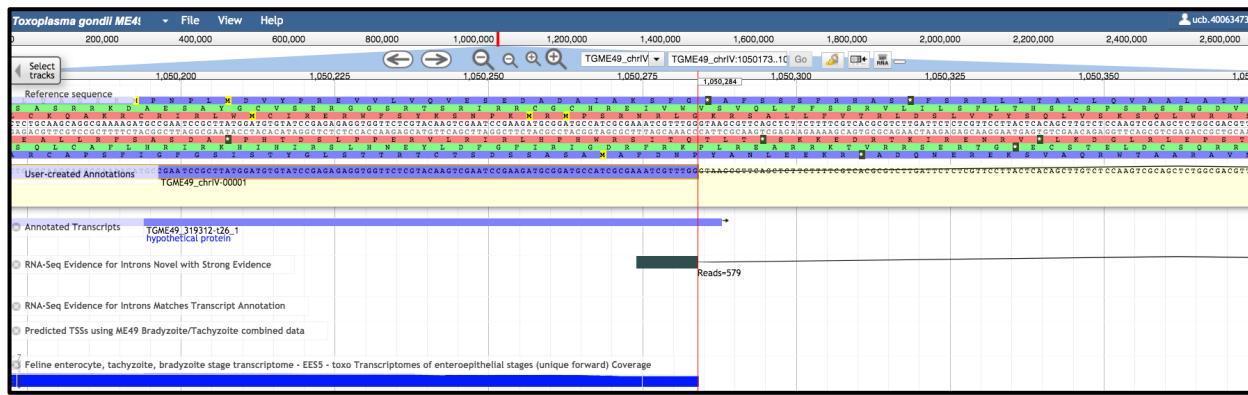
Hold down the shift key and select the two small exons. With a right-click open the drop-down menu and select merge. Hint: The exclamation mark tells you that there are non-canonical splice sites.



Zoom in, select the exon on the left side, point your mouse at the edge of the exon, a little arrow will appear.

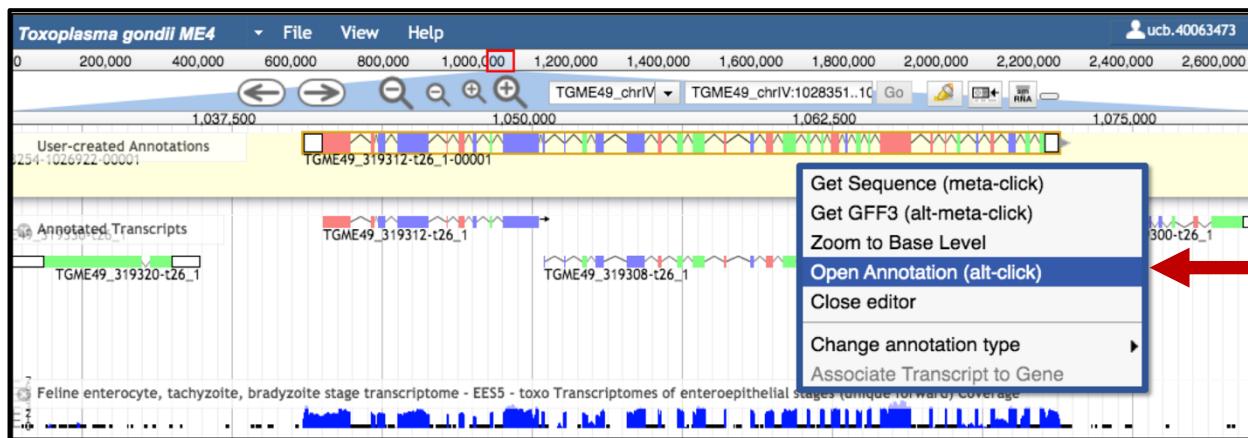


Shorten the exon to the correct splice site.

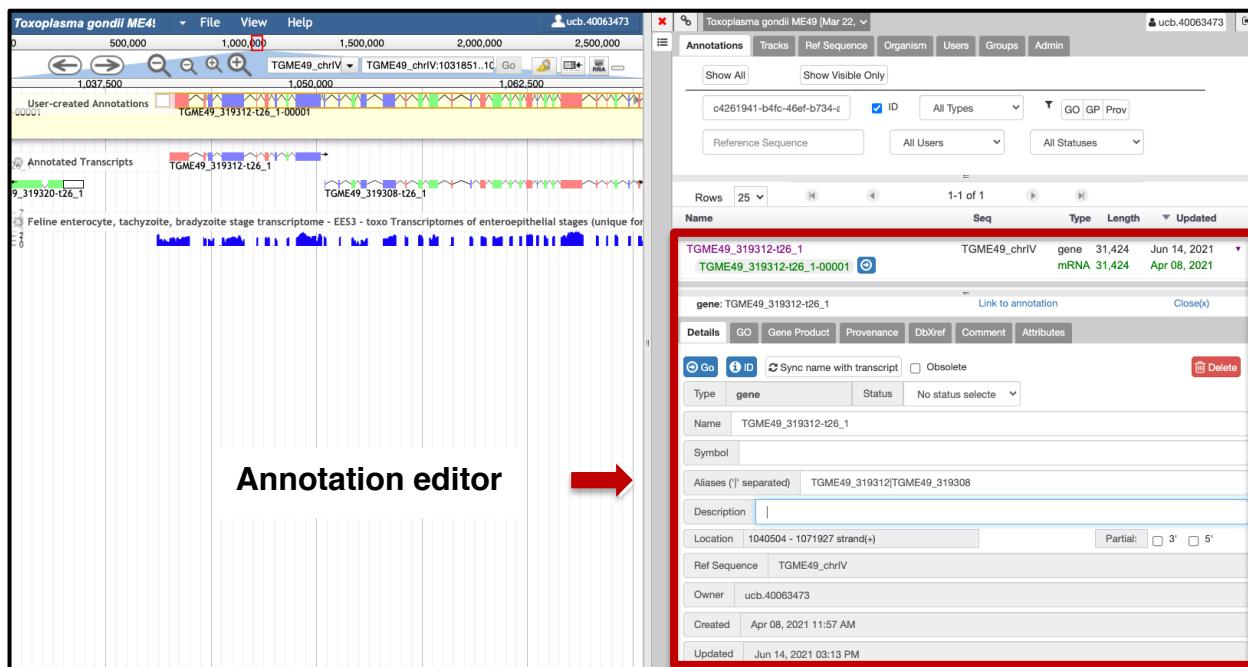


## 5) Opening of the Annotation editor window

Select the gene in the User-created Annotation track and with a right-click open the drop-down menu and choose **Open Annotation**. Alternatively, you can use the short-cut **alt-click**.



The annotation editor window is now shown on the right-hand side.



## 6) Finalising the structural annotation

Once the annotations panel is open click on the Attributes tab, select from the canned tag **structural** and from the canned value **merge**. Click on the + sign.



Open the Details section and add the gene IDs that you've merged in the Aliases section. Ideally, also add a Description/Gene Product.

The screenshot shows the 'Details' tab selected in the top navigation bar. The 'Aliases ('|' separated)' field contains 'TGME49\_319308|TGME49\_319312'. A red arrow points to this field.

TGME49_319312-t26_1	TGME49_chrlV	gene	30,508	Apr 08, 2021
TGME49_319312-t26_1-00001	mRNA	30,508	Apr 08, 2021	(dropdown menu)
gene: TGME49_319312-t26_1				
<b>Details</b> GO Gene Product Provenance DbXref Comment Attributes				
<a href="#">+ Go</a> <a href="#">ID</a>		<a href="#">Delete</a>		
Type	gene	Status	No status selected	
Name	TGME49_319312-t26_1			
Symbol				
Aliases (' ' separated)	TGME49_319308 TGME49_319312			
Description				
Location	1041309 - 1071817 strand(+)			
Ref Sequence	TGME49_chrlV			
Owner	ucb.40063473			
Created	Apr 08, 2021 11:57 AM			
Updated	Apr 08, 2021 11:57 AM			

To finalise the annotation select the status **Finished** on the gene.

The screenshot shows the 'Details' tab selected. The 'Status' dropdown menu is open, showing options: 'No status selected' (unchecked), 'Not Finished' (unchecked), 'Finished' (selected and highlighted in blue), and 'Requires Curator' (unchecked). A red arrow points to the 'Finished' option.

TGME49_319312-t26_1	TGME49_chrlV	gene	30,508	Apr 08, 2021
TGME49_319312-t26_1-00001	mRNA	30,508	Apr 08, 2021	(dropdown menu)
gene: TGME49_319312-t26_1				
<b>Details</b> GO Gene Product Provenance DbXref Comment Attributes				
<a href="#">+ Go</a> <a href="#">ID</a>		<a href="#">Delete</a>		
Type	gene	Status	✓ No status selected Not Finished <b>Finished</b> Requires Curator	
Name	TGME49_319312-t26_1			
Symbol				
Aliases (' ' separated)	TGME49_319312 TGME49_319308			
Description	Hypothetical protein, conserved			
Location	1041309 - 1071817 strand(+)			
Ref Sequence	TGME49_chrlV			
Owner	ucb.40063473			
Created	Apr 08, 2021 11:57 AM			
Updated	Apr 08, 2021 02:00 PM			

**Done!** For additional questions, please get in touch with the VEuPathDB help desk.

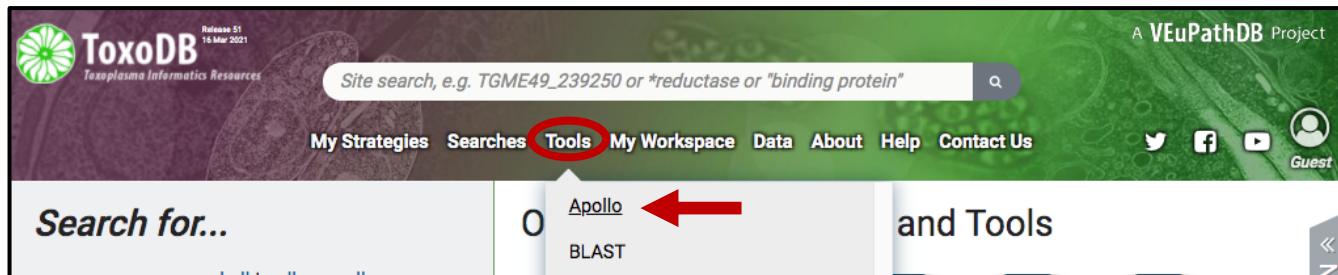
# Structural annotation in Apollo

## Adding a new gene

In this short tutorial we are showing you step-by-step how to add a new gene in Apollo.

### 1) Accessing Apollo

To access Apollo select **Tools** from the top menu and choose **Apollo** from the drop-down menu.



Click on the button **Go to Apollo**.

### Structural and functional community curation in Apollo

Welcome to the VEuPathDB Apollo service (Dunn et al. 2019), a real time collaborative genome annotation and curation platform.

Use Apollo to integrate new or update current structural and functional data, for gene models in the organisms available in VEuPathDB. Organisms in AmoebaDB, CryptoDB, FungiDB, GiardiaDB, MicrosporidiaDB, PiroplasmaDB, PlasmoDB, ToxoDB, TrichDB, TriTrypDB & VectorBase are available for community curation.

Apollo help and documentation:

- A [sandbox](#) is available for you to get familiar with all Apollo menus, tools, and tracks before you decide to use it for your real gene manual annotations. These changes will not affect any of the organism's official gene set, neither will be preserved.
- Comprehensive webinar to learn [how to use Apollo](#) (57:40 min)
- [Quick commands](#)
- [Functional annotation tutorial](#)
- [About Apollo \(Login required\)](#)
- [User Guide](#)
- [Request feature/Report a bug](#)
- [Powered by JBrowse](#)
- [Web Service API \(Login required\)](#)

A screenshot of the Apollo interface. On the left is a genome browser track for 'Amoebae albostramine STEC' showing genomic regions and various annotations. On the right is a detailed view of a gene annotation for 'AAL000027-AKA' with tabs for 'Annotations', 'Details', 'Coding', 'GO', 'Gene Product', 'Provenance', 'Editor', 'Comment', and 'Metadata'. The 'Details' tab is selected, showing information like 'Name: AAL000027-00001', 'Ref Sequence: AAL000027', 'Length: 15,629', and 'Status: Finished'. A red circle highlights the 'Go to Apollo' button at the bottom of the page.

To use Apollo you need to be logged into VEuPathDB. If you have not done so yet log now into Apollo with your VEuPathDB user ID and password.

## 2) Navigate to the genome and chromosome coordinates

Choose from the drop-down menu the genome of interest (1) and the chromosome (2).

The screenshot shows the JBrowse interface for *Toxoplasma gondii* ME4. The top navigation bar has tabs for Annotations, Tracks, Ref Sequence, Organism, Users, Groups, and Admin. The 'Annotations' tab is selected. A dropdown menu on the left shows 'Toxoplasma gondii ME4' and 'Toxoplasma gondii ME49 [2015-0]'. A red box labeled '1' highlights the 'Toxoplasma gondii ME49 [2015-0]' option. Another red box labeled '2' highlights the 'TGME49\_chrX' entry in the list of chromosomes.

Go to the coordinates with the missing gene. Use the arrows to navigate to the coordinates (1), or type the coordinates in the search box (2).

The screenshot shows the JBrowse interface for *Toxoplasma gondii* ME4. The top navigation bar has tabs for Annotations, Tracks, Ref Sequence, Organism, Users, Groups, and Admin. The 'Annotations' tab is selected. A dropdown menu on the left shows 'Toxoplasma gondii ME4' and 'Toxoplasma gondii ME49 [2015-0]'. A red box labeled '1' highlights the 'Select tracks' button. A red box labeled '2' highlights the search input field containing 'TGME49\_chrVIII:4592061'. The genome browser displays a single track for 'TGME49\_chrVIII' with a yellow highlight.

## 3) Adding draggable annotation and supporting evidence

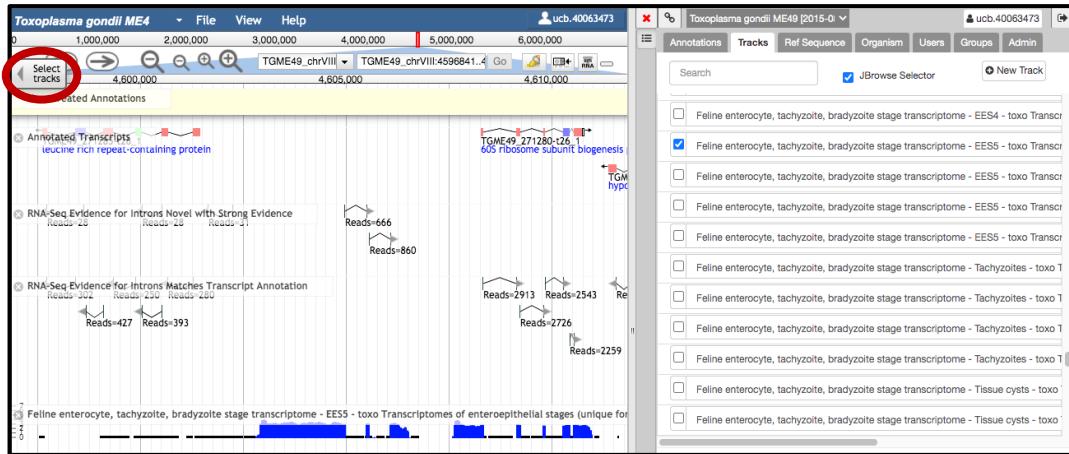
Select on the right-hand side the tab **Tracks**.

The screenshot shows the JBrowse interface for *Toxoplasma gondii* ME4. The top navigation bar has tabs for Annotations, Tracks, Ref Sequence, Organism, Users, Groups, and Admin. A red box highlights the 'Tracks' tab. The genome browser displays a single track for 'TGME49\_chrVIII' with a yellow highlight. On the right, the 'Tracks' panel is open, showing categories: Comparative Genomics, Draggable Annotation, Epigenomics, and Gene Models. Each category has a count (e.g., 0/1, 0/4, 0/64, 0/25) and a checkbox.

Click on the menu item **Draggable Annotation** select **Annotated Transcripts, RNA-Seq Evidence for Introns Novel with Strong Evidence and RNA-Seq Evidence for Introns Matches Transcript Annotation**.

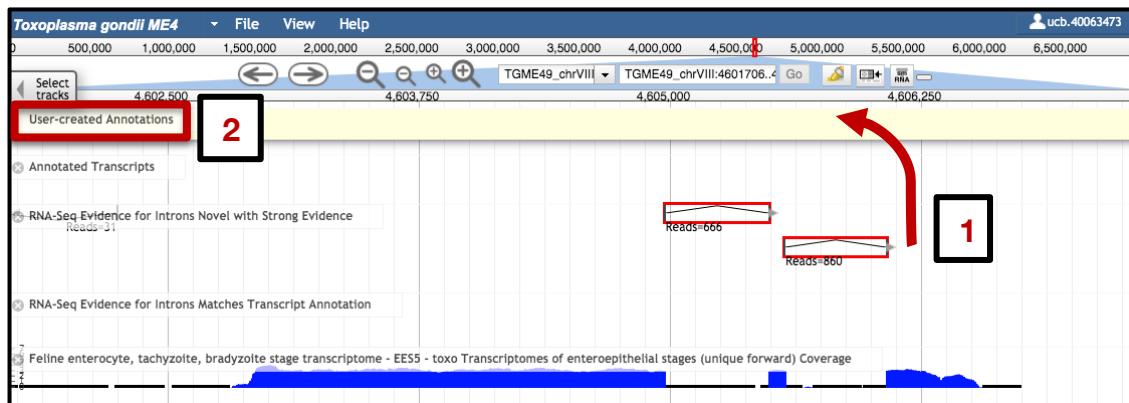
The screenshot shows the 'Draggable Annotation' section of the JBrowse Tracks panel. A red box highlights the 'Draggable Annotation' heading. Below it, four checkboxes are listed with red arrows pointing to them: 'RNA-Seq Evidence for Introns Novel with Strong Evidence', 'RNA-Seq Evidence for Introns Novel with Weak Evidence', 'RNA-Seq Evidence for Introns Matches Transcript Annotation', and 'Annotated Transcripts'. Each checkbox has a count next to it: 0/1, 0/4, 0/64, and 0/25 respectively. Other sections like Comparative Genomics, Epigenomics, Gene Models, etc., are also visible.

Select additional evidence, i.e. RNAseq plots from the Transcriptomics section. Alternatively, you can select evidence from the JBrowse menu by clicking on **Select tracks**.

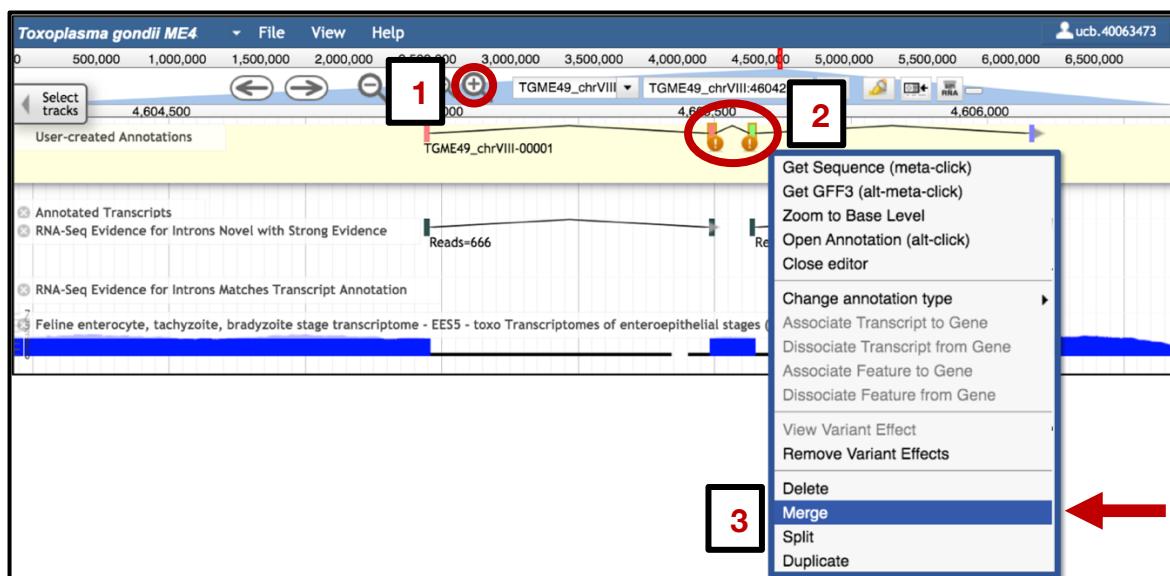


#### 4) Building the new gene

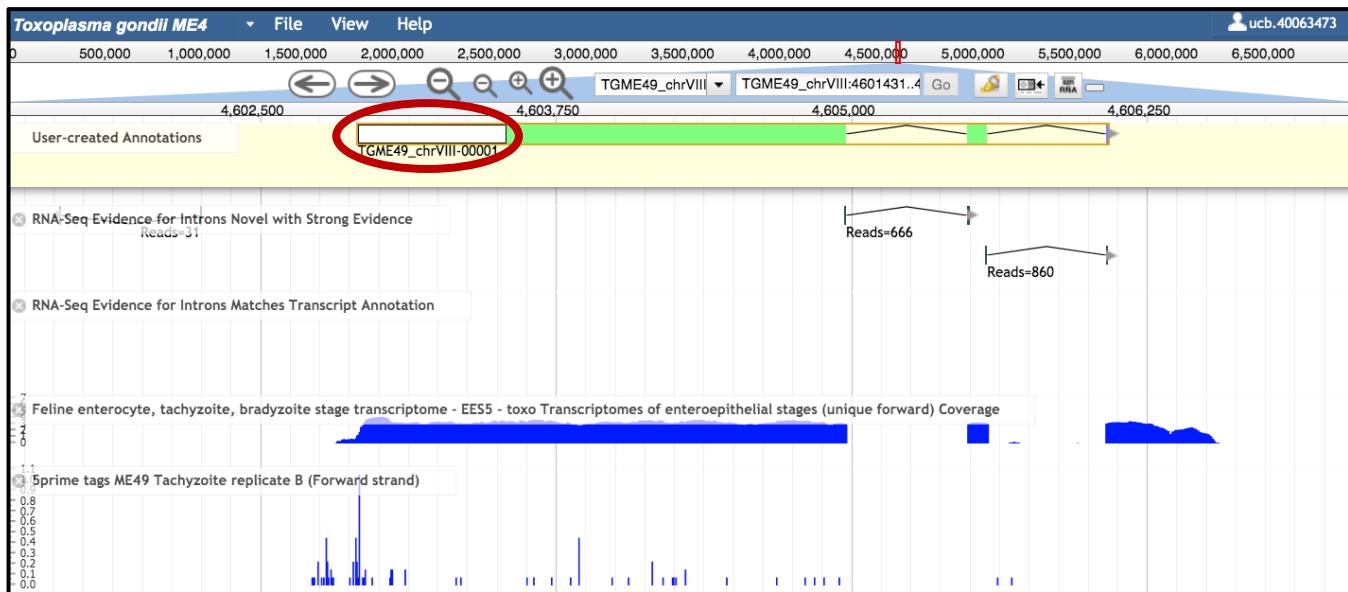
Hold down the shift key and select the introns with strong evidence (1), drag and drop them into the User-created Annotations track (2).



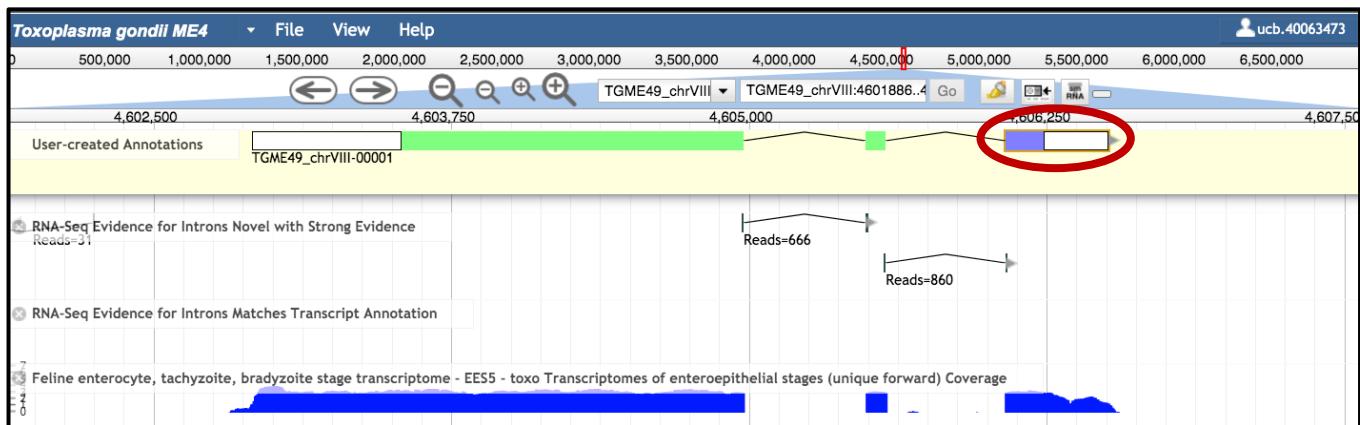
Zoom in by clicking on the + sign on the top (1). Press the Shift key and select the two small exons in the middle (2). With a right-click open the drop-down menu and choose **Merge** (3). Alternatively, select one of the exons you would like to merge, go to the edge of the feature until a little arrow appears and extend the exon until it overlaps with the second exon.



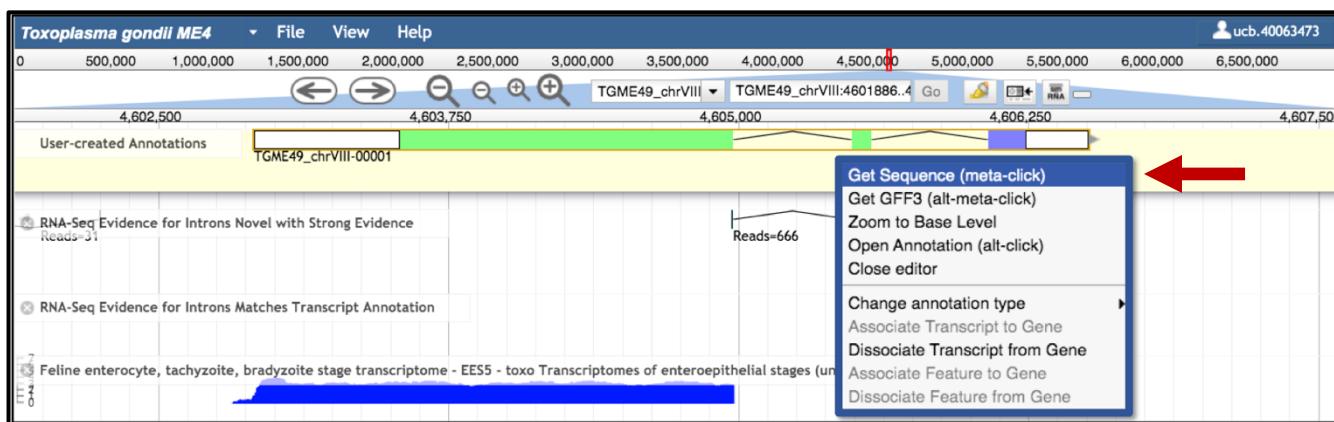
Select the first exon, point your mouse at the edge of the feature until a little arrow appears, then extend the exon to the transcription start. Apollo will automatically create the 5'UTR!



Select the last exon, point your mouse at the edge and extend the exon to the end of the gene model. Apollo will create the 3'UTR automatically!

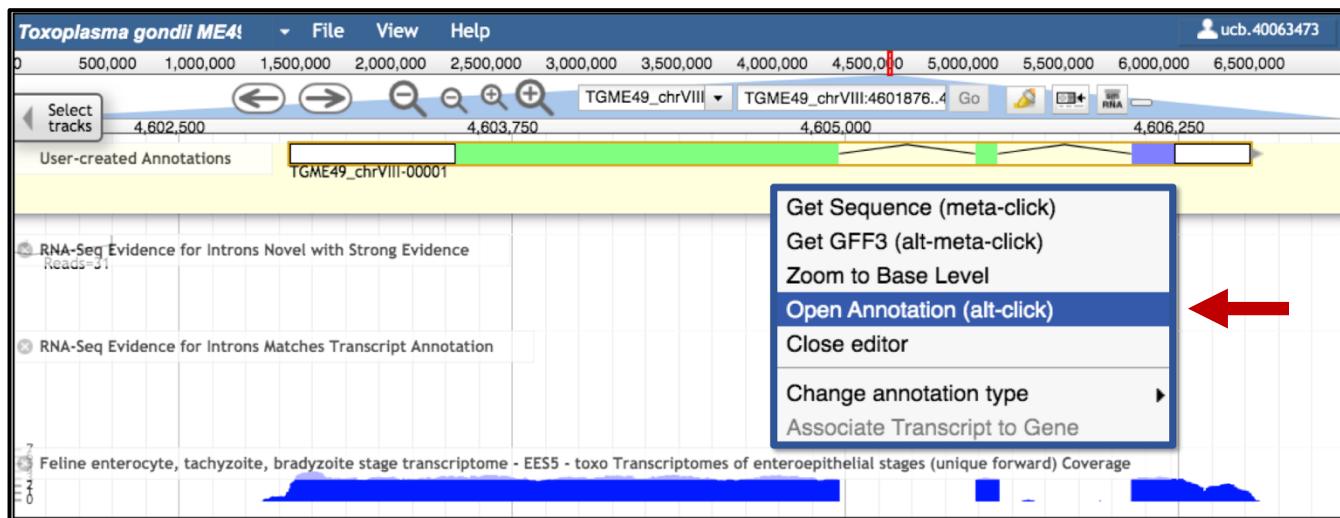


Select the new gene, with a right-click open the annotation drop-down menu and choose **Get Sequence**. Copy the sequence and run blast (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>) and InterPro (<https://www.ebi.ac.uk/interpro>) to get additional information about the new gene.

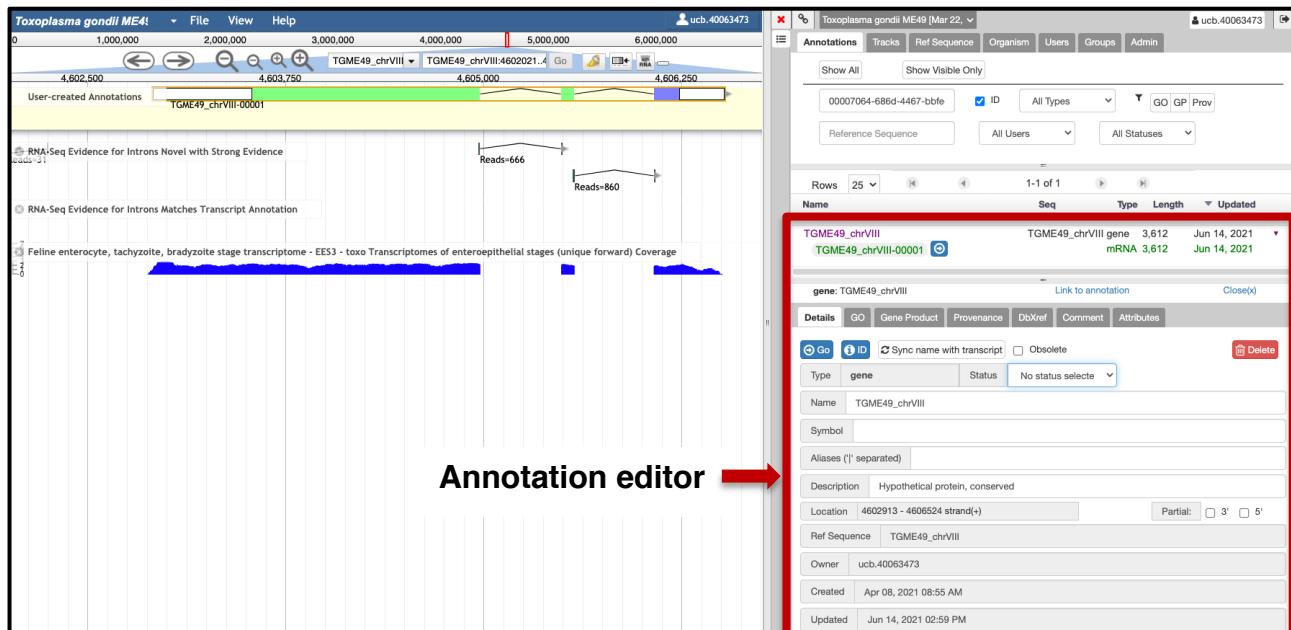


## 5) Opening of the Annotation editor window

Select the gene in the User-created Annotation track and with a right-click open the drop-down menu and choose **Open Annotation**. Alternatively, you can use the short-cut **alt-click**.



The annotation editor window is now shown on the right-hand side.



## 6) Finalising the structural annotation

Once the annotations panel is open click on the Attributes tab, select from the canned tag **structural** and from the canned value **new**. Click on the + sign.

The screenshot shows the VEuPathDB annotations interface for a gene named TGME49\_chrVIII. The top navigation bar shows the gene ID, name (TGME49\_chrVIII), version (3,603), date (Apr 04, 2021), and mRNA count (3,603). Below the navigation bar, there are tabs: Details, GO, Gene Product, Provenance, DbXref, Comment, and Attributes. The Attributes tab is currently selected. Under the Attributes tab, there is a search bar with 'structur' in the prefix field and 'Value' in the dropdown. A dropdown menu is open, listing various canned values: 'Select canned value', 'added\_comment', 'split', 'merge', 'retain\_previous', 'added\_product', 'added\_go', 'added\_symbol', 'added\_alias', 'added\_pmid', 'removed\_product', 'added\_ec\_number', 'delete', 'isofrom', 'new', 'modify', and 'added\_dbxref'. The 'new' option is highlighted with a blue background and a red arrow points to it from the right.

Go to the Details tab, add a description/gene product to your new gene and select the status **Finished** on the gene.

The screenshot shows the VEuPathDB annotations interface for the same gene. The top navigation bar and gene information are identical. The Details tab is currently selected. The gene details include: Type (gene), Status (dropdown menu open, showing 'No status selected', 'Not Finished', 'Finished' (highlighted with a blue background and a red arrow), and 'Requires Curator'), Name (TGME49\_chrVIII), Symbol, Aliases, Description (hypothetical protein), Location (4602917 - 4606520 strand(+)), Ref Sequence (TGME49\_chrVIII), Owner (ucb.40063473), Created (Apr 04, 2021 04:20 PM), and Updated (Apr 04, 2021 04:20 PM). The 'Status' dropdown menu has 'No status selected', 'Not Finished', 'Finished' (highlighted with a blue background and a red arrow), and 'Requires Curator'.

Done! For additional questions, please get in touch with the VEuPathDB help desk.

# Structural annotation in Apollo

## Alternative transcripts

In this short tutorial we are showing you step-by-step how to create alternative transcripts in Apollo.

### 1) Accessing Apollo

To access Apollo go to the gene record page of your gene of interest and click on the link **View and update community annotations in Apollo (1)**. You can also access Apollo from the gene models section by clicking on the button **Annotate in Apollo (2)**. Alternatively, go to the **Tools** menu and choose Apollo from the drop-down list (3).

The screenshot shows the ToxoDB gene record page for TGME49\_315160. At the top, there's a navigation bar with links like 'My Strategies', 'Searcher', 'Tools' (circled in red), 'My...', 'Data', 'About', 'Help', and 'Contact Us'. Below the navigation, there's a search bar and several shortcut links for Synteny, Alignments, Phenotype, SNPs, Transcriptomics, Protein Features, and Proteomics. The main content area displays gene details: Type: protein coding gene, Chromosome: XI, Location: TGME49\_chrXI:4,526,997..4,538,020(-), Species: *Toxoplasma gondii*, Strain: ME49, Status: Reference strain. It also shows user comments and a link to view and update community annotations in Apollo (1). Under 'Gene models', it lists 1 Gene model, 1 Transcript, and 1 Gene Model. A note says 'This gene is available in Apollo for community annotation.' An 'Annotate in Apollo' button is circled in red (2). A red box highlights the 'Tools' menu item (3) in the top navigation bar.

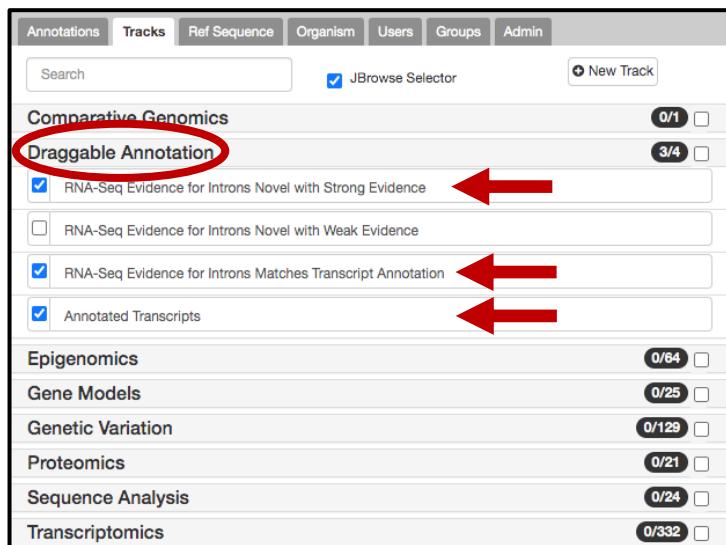
To use Apollo you need to be logged into VEuPathDB. If you have not done so yet log now into Apollo with your VEuPathDB user ID and password.

### 2) Adding draggable annotation and supporting evidence

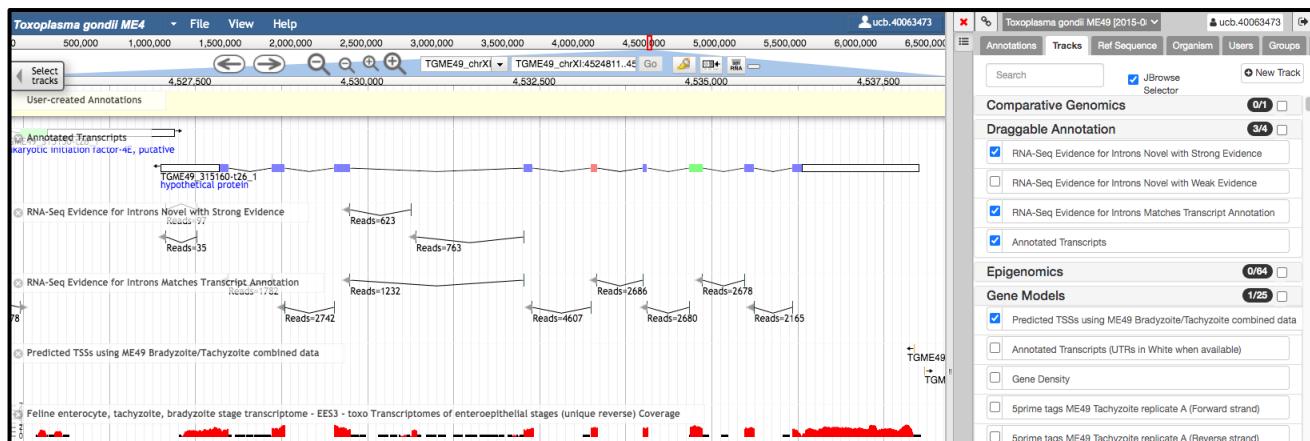
Select on the right-hand side the tab **Tracks**.

The screenshot shows the JBrowse genome browser interface for the Toxoplasma gondii ME4 genome. The top navigation bar includes 'File', 'View', 'Help', and a user account icon. The main area shows a genomic track for chromosome X, with coordinates 0 to 6,000,000. A red box highlights the 'Tracks' tab in the top right corner of the interface. Other tabs include 'Annotations', 'Ref Sequence', 'Organism', 'Users', 'Groups', and 'Admin'. The bottom right corner shows a status bar with 'ucb.40063473'.

Click on the menu item **Draggable Annotation** select **Annotated Transcripts, RNA-Seq Evidence for Introns Novel with Strong Evidence and RNA-Seq Evidence for Introns Matches Transcript Annotation**.

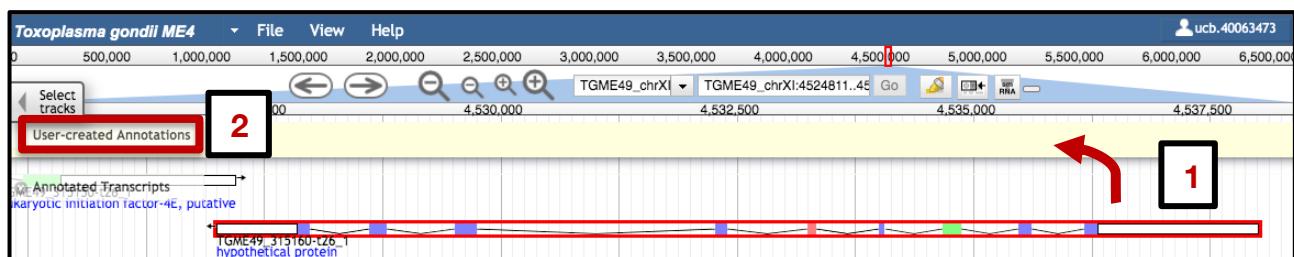


Select additional evidence, i.e. RNAseq plots and predicted TSS (transcription start sites).

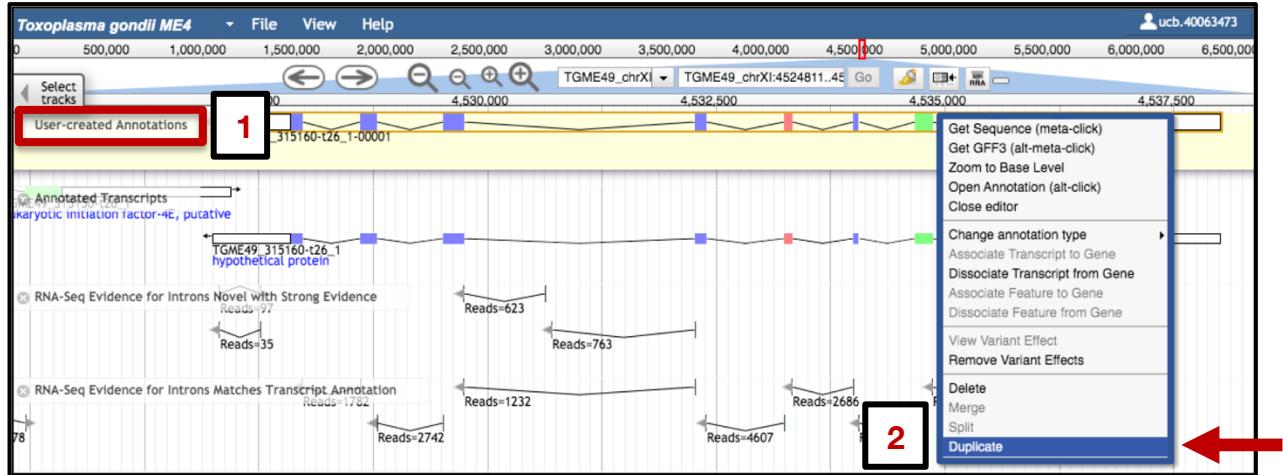


### 3) Adding alternative transcripts

Select the gene model by clicking on one of the introns or by double clicking on the gene model (1). The gene will show up with red boundaries. Drag and drop the gene into the User-created Annotations track (2).

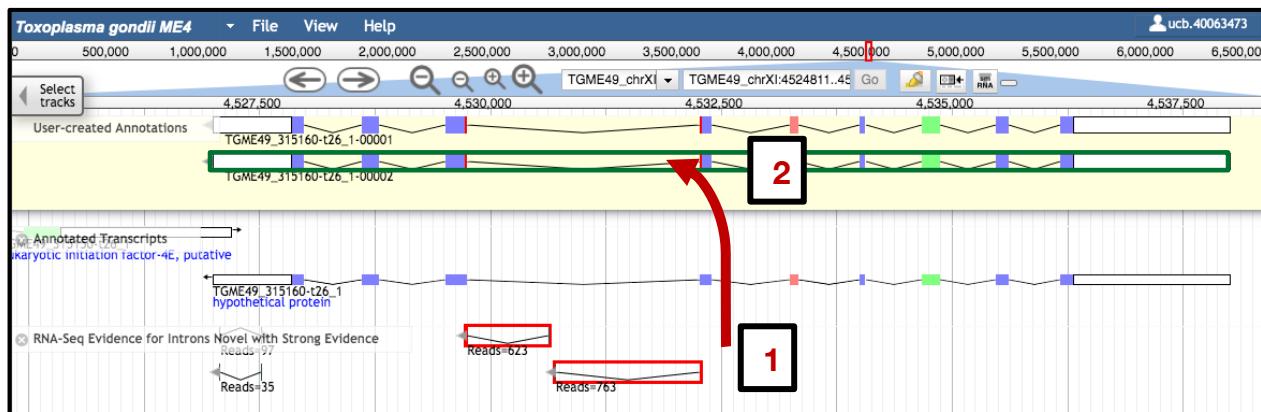


Select the gene in the User-created Annotations area (1). With a right-click open the annotation drop-down menu and choose duplicate (2).

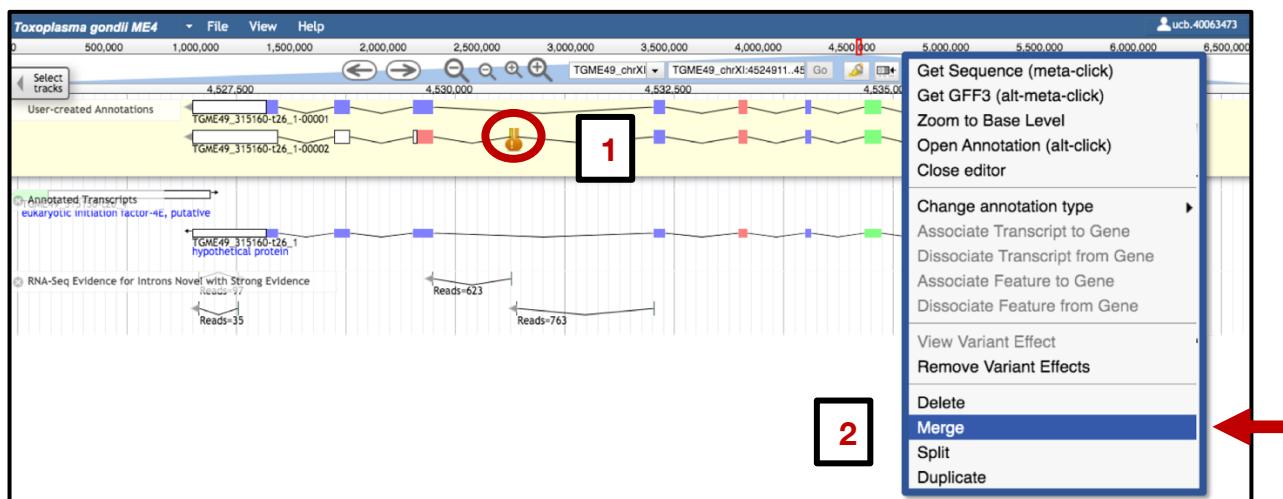


#### 4) Modifying the alternative transcript

Select the intron junctions individually, or hold down the shift key and select both intron junctions with strong evidence (1), drag and drop them into the gene model (2). The gene will get a green box when dragging and dropping the intron evidence.

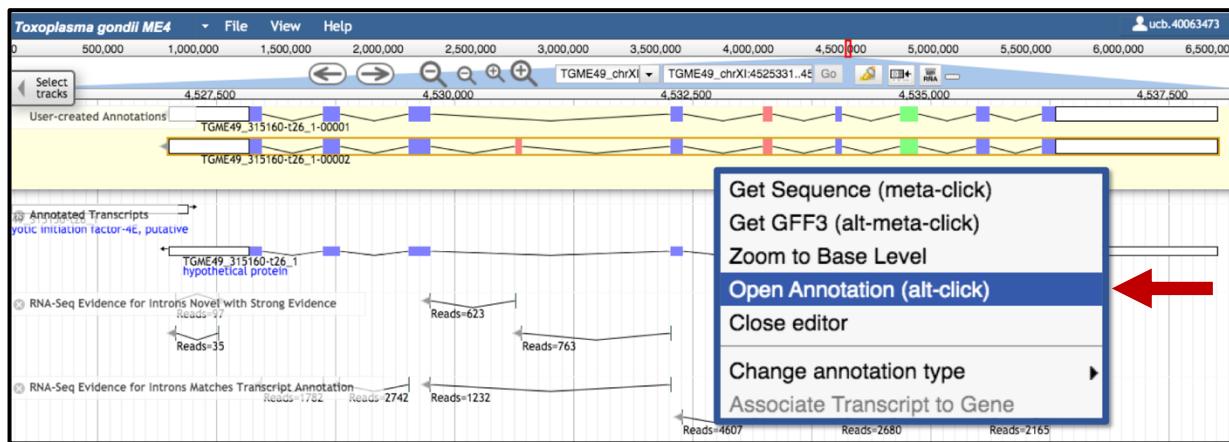


Hold down the shift key and select the two small exons. With a right-click open the drop-down menu and select merge.

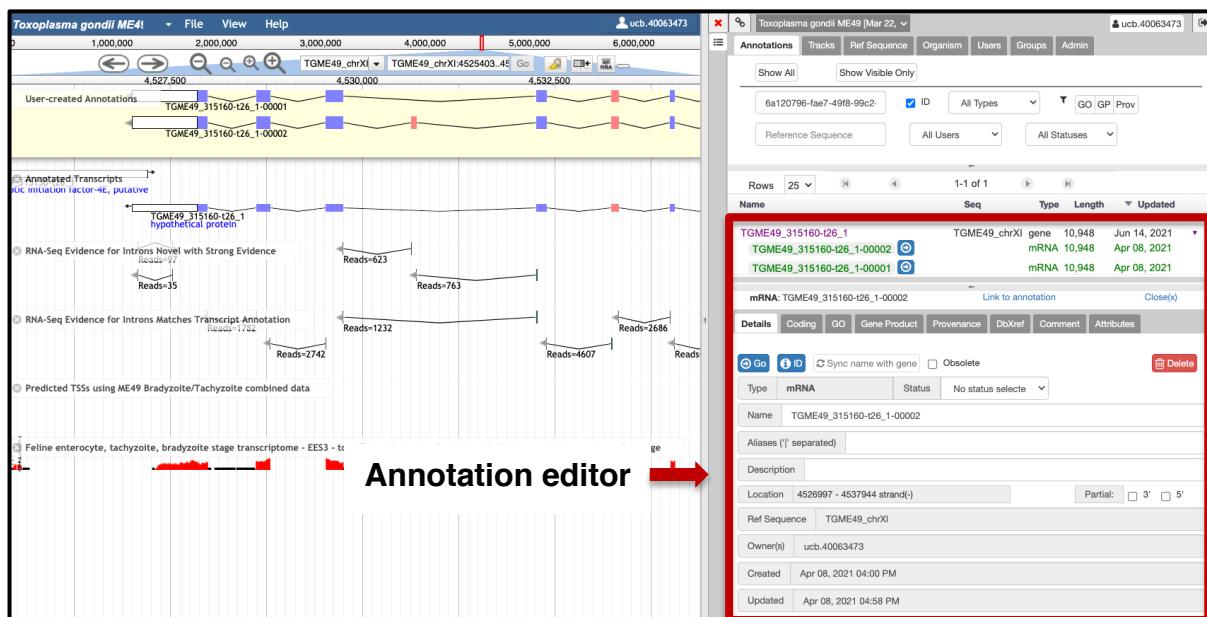


## 5) Opening of the Annotation editor window

Select one of the transcripts in the User-created Annotation track, with a right-click open the drop-down menu and choose **Open Annotation**. Alternatively, you can use the short-cut **alt-click**.



The annotation editor window is now shown on the right-hand side.



## 6) Finalising the structural annotation

Once the annotations panel is open click on the Attributes tab, select from the canned tag **structural** and from the canned value **modify**. Click on the + sign.



Add a Description/Gene Product. Finally go the Details tab and select the status **Finished** on the gene.

gene: TGME49\_315160-t26\_1

Details GO Gene Product Provenance DbXref Comment Attributes

Type gene Status ✓ No status selected  
Not Finished  
**Finished** (highlighted)  
Requires Curator

Name TGME49\_315160-t26\_1

Symbol

Aliases ('|' separated)

Description

Location 4526996 - 4538020 strand(-)

Ref Sequence TGME49\_chrXI

Owner ucb.40063473

Created Apr 08, 2021 03:51 PM

Updated Apr 08, 2021 04:00 PM

Delete

Done! For additional questions, please get in touch with the VEuPathDB help desk.