

## Phenotypic data

### Learning objectives:

- Explore how to combine different phenotypic data
- Explore high throughput mutagenesis data
- Explore curated phenotypic data
- Explore high throughput subcellular localization data

#### 1. Identify genes that are targeted to the ciliary tip of *Trypanosoma brucei* that are also essential for parasite fitness.

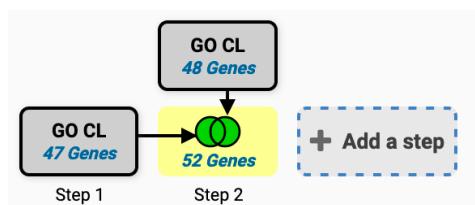
Note for this exercise use <http://tritrypdb.org>

- a. TriTrypDB integrates data from the TrypTag project (<http://tryptag.org>). Genes from *T. brucei* were N- and C-terminally tagged with a fluorescent protein and subcellular localization determined by microscopy. The description of the localization was done using gene ontology terms.
  - Start by finding the “Cellular Localization Imaging” search.

#### Identify Genes based on Cellular Localization Imaging

The screenshot shows the 'Identify Genes based on Cellular Localization Imaging' search page. On the left, there is a search bar with 'cellul' typed in, a dropdown menu showing 'Genes' selected under 'Protein targeting and localization', and a link to 'Cellular Localization Imaging'. On the right, there are filter options: 'Organism' set to 'Trypanosoma brucei brucei TREU927', 'Location of tag' set to 'C-terminal', and a search bar containing 'GO:0097542 : ciliary tip ; 3'. A red arrow points from the 'Cellular Localization Imaging' link to the search bar, and another red arrow points to the search bar itself.

- Configure the search to identify the GO term “Ciliary Tip” – notice that when you start typing the autocomplete function offers you selectable options.
- Since the experiment examined both N and C terminal fusions proteins, you will have to run the search twice and combine the results from both searches. Did you use a union or an intersect to combine the results?



- Explore the results you got. Scroll down to the results section, then scroll to the right of the results window to reveal the subcellular localization

The screenshot shows a bioinformatics interface with a table of gene results and a large image of subcellular localization. The table includes columns for product description, # Transcripts, EC numbers, and Cellular localization images. A red arrow points to one of the small localization images in the grid.

images. These are very small, but you can right click on them to open a larger image in a new window.

- b. Add a step to identify how many genes are essential for the fitness of the parasite. Click on Add step, then search for the phenotype searches. Click on the Phenotype Evidence option.

The screenshot shows a search strategy builder interface. It includes a 'Combine with other Genes' section and a 'Transform into related records' section. To the right, there are two numbered steps: 1. Choose how to combine with other Genes (radio button selected for '2 INTERSECT 3') and 2. Choose which Genes to combine. From... (radio button selected for 'A new search'). A search bar contains the term 'phen'.

- Select the “High-throughput phenotyping using RNAi target sequencing (David Horn)”.

Add a step to your search strategy

#### Search for Genes by Phenotype Evidence

The results will be intersected with the results of Step 2.

The screenshot shows the 'Search for Genes by Phenotype Evidence' interface. It includes a legend at the top right: CP (Curated Phenotype), PQ (Quantitative Phenotype), and PT (Phenotype Text). Below the legend, there are sections for 'Organism' (Trypanosoma brucei brucei TREU927) and 'Data Set' (High-throughput phenotyping using RNAi target sequencing (David Horn)). A red circle highlights the 'PO' button under 'Choose a Search'.

- Configure the search to return genes that are decreased in coverage by 1.5 fold when comparing the maximum expression value of all induced samples to the uninduced sample.

**For the Experiment**

Quantitated from the CDS Sequence  
 Quantitated from gene model (5 prime UTR + CDS)

[select all](#) | [clear all](#)

return [protein coding](#) [Genes](#)

that are [Decrease in coverage](#) [?](#)

with a Fold change  $\geq$   [?](#)

between each gene's maximum [expression value](#) [?](#)

in the following [Reference Samples](#) [?](#)

Uninduced sample  

[select all](#) | [clear all](#)

and its maximum [expression value](#) [?](#)

in the following [Comparison Samples](#) [?](#)

Induced in bloodstream (BS) forms, 3 days (10 doublings)  
 Induced in bloodstream (BS) forms, 6 days (20 doublings)  
 Induced in procyclic forms (PS) forms, 9 days (9 doublings)  
 Induced throughout differentiation (DIF = 7 BS doublings + 6 PS doublings)

[select all](#) | [clear all](#)

**Example showing one gene that would meet search criteria**  
(Dots represent this gene's expression values for selected samples)

**Down-regulated**

Expression

Reference Samples      Comparison Samples

1.5 fold

Expression Value Reference

Maximum Expression Value Comparison

For each gene, the search calculates:

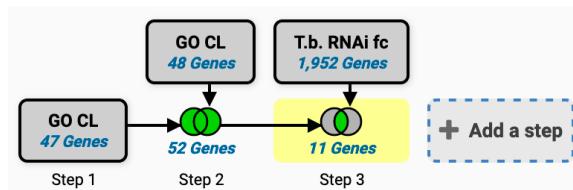
$$\text{fold change} = \frac{\text{reference expression value}}{\text{maximum expression value in comparison}}$$

and returns genes when fold change  $\geq 1.5$ .

You are searching for genes that are down-regulated between one reference sample and at least two comparison samples.

This calculation creates the narrowest window of expression values in which to look for genes that meet your fold change cutoff. To broaden the window, use the average or minimum comparison value.

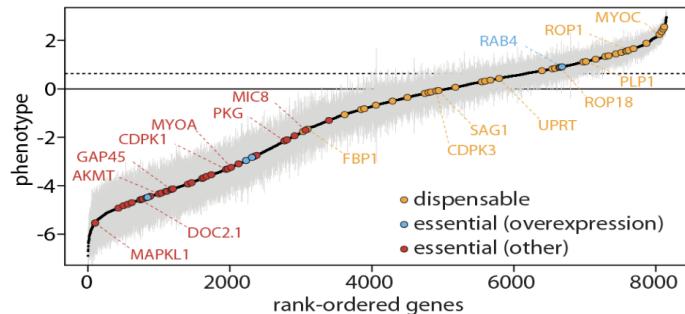
- How many genes did you get?



## 2. Finding genes based on high throughput mutagenesis and fitness analysis.

Note for this exercise use <http://toxodb.org>

- Navigate to the CRISPR phenotype search. Note that this search form is quite simple just requiring a range of fitness values. The defaults return all genes not limiting the search at all. This is only useful in as much as it tells you which genes were assayed which is nearly the entire genome. The tricky bit is deciding where to make the cutoffs. Again, the description on the search form is very helpful in this regard (as is the link to the paper ... remember these phenotypes were assayed under specific conditions so just because a particular gene doesn't show a phenotype doesn't mean it wouldn't in other conditions (or infecting an



actual host). The plot showing the phenotype score (fitness) is particularly useful. Red points along the plot are genes known to be essential under these conditions while yellow are known to be expendable. This will help you determine where to set the values. The scores range from 2.96 (least “essential”) to -6.89 (most “essential”). Try it running this search by limiting the range from -6.89 to -4. Do you get the expected results based on the above graph and the number of genes returned in your search results?

**Identify Genes based on CRISPR Phenotype**

**Phenotype Score >=**  
-6.89

**Phenotype Score <=**  
-4

**CRISPR 1,343 Genes**

**Add a step**

Step 1

- What kinds of genes are in your results? What kinds of genes would you expect to be essential? One way to explore the data is to run a GO enrichment analysis to determine if any biological processes are enriched in your results. Give this a try. What do your results look like and do they make sense?

GO ID	GO Term	Genes in the bkgd with this term	Genes in your result with this term	Percent of bkgd genes in your result	Fold enrichment	Odds ratio	P-value	Benjamini
GO:0010467	gene expression	493	235	47.7	2.35	4.38	7.07e-48	6.50e-45
GO:0034645	cellular macromolecule homeostatic process	385	194	50.4	2.49	4.72	1.82e-43	8.36e-41

- How many of these genes are upregulated in *in vivo* chronic stages of *T. gondii*?
- Click on add step and elect the RNAseq searches under the Transcriptomics category

- Find the experiment with chronic stages and run a search based on differentially expressed genes (DE).

- Intersect genes that are 2-fold upregulated in chronic stages compared to acute stages.

Add a step to your search strategy

### Experiment

Acute and chronic *T.gondii* infection of mouse. unstranded

### Reference Sample

acute infection 10 days p.i.  
 chronic infection 28 days p.i.

### Comparator Sample

acute infection 10 days p.i.  
 chronic infection 28 days p.i.

### Direction

up-regulated

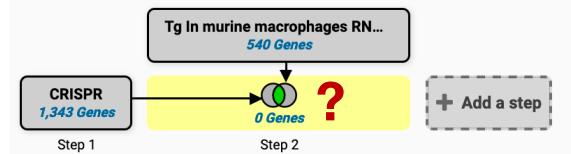
### fold difference >=

2

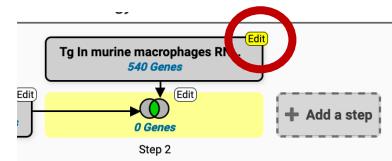
### adjusted P value less than or equal to

0.1

- Did you get zero results? This is to be expected since the CRISPR data was analyzed using the GT1 strain of *Toxoplasma* and the RNA-Seq data is from the ME49 strain. How can you fix this?



- Hint: transform the results in step 2 from *T. gondii* ME49 to *T. gondii* GT1. Click on the step edit button (move your mouse over the step and select edit).



- Select **orthologs** from the menu items at the top of the pop window.

Details for step *Tg In murine macrophages RNA-Seq (de)* 🔗  
540 Genes

Experiment Acute and chronic *T.gondii* infection of mouse, unstranded  
Reference Sample acute infection 10 days p.i.  
Comparator Sample chronic infection 28 days p.i.  
Direction up-regulated  
fold difference >= 2  
adjusted P value less than or equal to 0.1

▶ Give this search a weight

- Select *T. gondii* GT1 from the list of organisms and click on Run Step.

### Organism

1 selected, out of 31

[add these](#) | [clear these](#) | [select only these](#)  
[select all](#) | [clear all](#)

gt1 gt1 🔗

**Sarcocystidae**  
 **Toxoplasma**  
➡  **Toxoplasma gondii GT1**

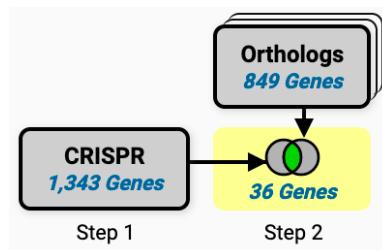
[add these](#) | [clear these](#) | [select only these](#)  
[select all](#) | [clear all](#)

### Syntenic Orthologs Only?

no

Run Step

- Now what do your results look like?



### 3. Identify essential *Plasmodium falciparum* genes that are highly expressed in schizont stages of the parasite.

Note for this exercise use <https://plasmodb.org>

- You can start by exploring the phenotype data in PlasmoDB.
- Select and run the search associated with the dataset: piggyBac insertion mutagenesis (John Adams).

The screenshot shows the PlasmoDB search interface. In the top left, there's a search bar with 'phen' typed in. Below it, a sidebar has 'Genes' selected. Under 'Phenotype', there's a link to 'Phenotype Evidence'. The main area is titled 'Identify Genes based on Phenotype Evidence'. It shows a table of datasets, with the last one, 'piggyBac insertion mutagenesis (John Adams)', highlighted. The legend at the top right includes a 'CP' button, which is circled in red.

- Configure the search to identify genes with a *mutant fitness score* of less than -3. Note that you can select the range by either clicking and dragging your mouse over the histogram or by typing the values in the selection boxes.

Identify Genes based on piggyBac insertion mutagenesis (mutant fitness and mutagenesis index scores)



- How many genes did you identify? Which gene has the lowest fitness score? Note that you might need to add the fitness score column, by clicking on add columns then filtering the options with the word “fitness”.

The screenshot shows the PlasmoDB search interface with a search strategy titled "Unnamed Search Strategy". Step 1 is set to "pB MIS/MFS 856 Genes". A modal window titled "Select Columns" is open, showing a list of columns: "Gene ID", "Phenotype", "fitn", "Location", "Product Description", and "Score". The "fitn" checkbox is checked. A red arrow points from this checkbox to the "fitn" column in the main results table below. The results table lists three genes: PF3D7\_0914400, PF3D7\_1144100, and PF3D7\_0728400, along with their descriptions and fitness scores (-4.094, -4.036, and -4.024 respectively).

- Click on Add Step and find the RNA-Seq searches.

The screenshot shows the "Add a step to your search strategy" dialog. It includes sections for "Combine with other Genes", "Transform into related records", and "Use Genomic Colocation to combine with other features". In the "Transform into related records" section, there is a "Gene models" list with checkboxes for "Gene Model Characteristics", "Transcriptomics", "Microarray Evidence", and "RNA-Seq Evidence". A red arrow points from the "RNA-Seq Evidence" checkbox in this list to the "RNA-Seq Evidence" checkbox in the "Choose which Genes to combine. From..." section above it.

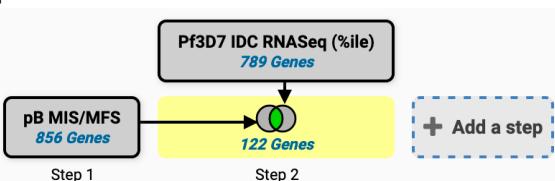
- Find the search called “Intraerythrocytic development cycle transcriptome (2019)” and select the percentile search.

#### Search for Genes by RNA-Seq Evidence

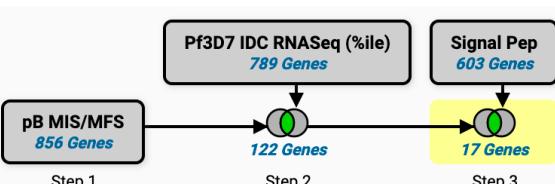
The results will be  intersected with |  the results of Step 2.

Filter Data Sets:	<input style="width: 150px; border: 1px solid black; border-radius: 5px; padding: 2px 5px;" type="text" value="intraer"/> 	Legend:	 DE Differential Expression  FC Fold Change  P Percentile  SA SenseAntisense
Organism	Data Set	Choose a Search	
<i>Plasmodium falciparum</i> 3D7	 Intraerythrocytic development cycle transcriptome (2019) (Wichers et al. 2019)	            	
<i>Plasmodium falciparum</i> 3D7	 Intraerythrocytic development cycle transcriptome (2018) (Toenhake et al.)	 	
<i>Plasmodium falciparum</i> 3D7	 Transcriptome during intraerythrocytic development (Bartfai et al.)	 	
<i>Plasmodium falciparum</i> 3D7	 Blood stage transcriptome (3D7) (Otto et al.)	 	
<i>Plasmodium falciparum</i> 3D7	 Intraerythrocytic cycle transcriptome (3D7) (Hoeijmakers et al.)	  	
<i>Plasmodium falciparum</i> 3D7	 Strand specific transcriptome of the intraerythrocytic developmental cycle (Siegel et al.)	  	
<i>Plasmodium vivax</i> P01	 Transcription profile of intraerythrocytic cycle (Zhu et al.)	 	

- Configure the search to identify all genes that are in the 80-100 percentile in all three available schizont samples. Remember to change the parameter to require matching all samples.
- How many genes did you get? Are any of these genes interesting? How many are predicted to be secreted?



- How did you identify the secreted genes? Hint, add a step and search for genes that have a predicted secretory signal peptide.



#### Samples

- young ring 8 hpi  
 late ring\_early trophozoite 16 hpi  
 mid trophozoite 24 hpi  
 late trophozoite 32 hpi  
 early schizont 40 hpi  
 schizont 44 hpi  
 late schizont 48 hpi  
 purified merozoites 0 hpi
- [select all](#) | [clear all](#)

#### Minimum expression percentile

80 

#### Maximum expression percentile

100

#### Matches Any or All Selected Samples?

all 

#### 4. Identify *Neurospora crassa* genes that affect conidia formation.

Note for the exercise use <https://fungidb.org>

- Start by locating the phenotype searches.

The screenshot shows the FungiDB search interface. At the top, it says "Release 52 20 May 2021" and "Fungal & Oomycete Informatics Resources". Below that is a search bar with "Search for..." placeholder text. A red arrow points from the text "Start by locating the phenotype searches." to the search bar. The search term "pheno" is entered. To the right of the search bar is a dropdown menu labeled "Genes" which is currently selected. Below the search bar is a "Phenotype" section with a "Phenotype Evidence" link. On the right side of the search results, there is a grid of links for various species and their phenotypes. One link for "Neurospora crassa QR7AA" is circled in red.

- This search provides you the option to filter based on categories on the left. Notice how when you select a different category on the left the filtering options in the middle change. Select the **Conidia number** category. Next select the “Reduced” value.

The screenshot shows the "Identify Genes based on Knockout Mutants" search interface. At the top, it says "Curated Phenotype". Below that is a search bar with "Identify Genes based on Knockout Mutants". A red arrow points from the text "Select the Conidia number category." to the "Conidia Number" section. The "Conidia Number" section has a dropdown menu with "Conidia Number" selected. Another red arrow points from the text "Next select the ‘Reduced’ value." to the "Reduced" checkbox in the list of options. The list includes: Increased, Normal, Not formed, Not formed, Reduced (which is checked), Severely reduced, and Not specified. To the right is a table showing the distribution of genes for each conidia number category. A red circle highlights the "Reduced" checkbox in the list of options.

Conidia Number	Remaining Genes	Genes	Distribution	%
Increased	12 (1%)	12 (1%)	1	(100%)
Normal	1,154 (90%)	1,154 (90%)	10	(100%)
Not formed	1 (< 1%)	1 (< 1%)	1	(100%)
Reduced	99 (8%)	99 (8%)	1	(100%)
Severely reduced	3 (< 1%)	3 (< 1%)	1	(100%)
Not specified	4 (< 1%)	4 (< 1%)	1	(100%)

- Notice that this search allows you to explore your results even before you click on the “Get Answer” button! Click around on the other categories on the left and see if the genes that are involved in a reduced number of conidia may also be involved in other phenotypes. For example, click on the **Ascospore Number** category, how maybe of your genes also have a phenotype with no ascospore formation?

**Genes**

1,283 Genes Total      99 of 1,283 Genes selected      Conidia Number X

expand all | collapse all      Find a variable

Ascospore Number		Check items below to apply this filter			1,283 (100%) of 1,283 Genes have data for this variable		
		Remaining Genes	Genes	Distribution	%		
<input type="checkbox"/>	Ascospore Number	99 (100%)	1,283 (100%)				
<input type="checkbox"/>	Normal	32 (22%)	1,043 (81%)		(3%)		
<input type="checkbox"/>	Not formed	56 (37%)	169 (13%)		(33%)		
<input type="checkbox"/>	Reduced	11 (11%)	65 (5%)		(17%)		
<input type="checkbox"/>	Increased	0 (0%)	2 (< 1%)		(0%)		
<input type="checkbox"/>	Severely Reduced	0 (0%)	5 (< 1%)		(0%)		
<input type="checkbox"/>	Severely reduced	0 (0%)	1 (< 1%)		(0%)		

Aerial Hyphae Height  
Basal Hyphae Growth Rate  
Conidia Morphology  
Conidia Number  
Perithecia Morphology  
Perithecia Number  
Protopheretia Number  
Protophereticial Morphology

Ascospore Number:  Ascospore Number:  Basal Hyphae Growth Rate:  Conidia Morphology:  Conidia Number:  Perithecia Morphology:  Perithecia Number:  Protopheretia Number:  Protophereticial Morphology:

- Click on get answer. What kinds of genes are in your results? Try analysing the results to see if there are any biological processes enriched in your results.

Step 1

KO Map      Add a step

99 Genes (98 ortholog groups)      Revise this search

Gene Results      Genome View      Gene Ontology Enrichment X      Analyze Results [ Rename This Analysis | Duplicate ]

Organism Filter select all | clear all | expand all | collapse all Hide zero counts Search organisms... Fungi 99 Oomycota 0

Gene Ontology Enrichment Find Gene Ontology terms that are enriched in your gene result. [Read More](#)

Parameters

Organism: **Neurospora crassa OR74A**      Ontology: Biological Process, Cellular Component, Molecular Function

Evidence: Computed, Curated

Limit to GO Slim terms: No, Yes

P-Value cutoff: 0.05 S (0 - 1)

Submit

Analysis Results:

361 rows

Open in Revigo      Show Word Cloud      Download

GO ID	GO Term	Genes in the bkgd with this term	Genes in your result with this term	Percent of bkgd genes in your result	Fold enrichment	Odds ratio	P-value	B
GO:0070787	conidiophore development	84	26	31.0	22.87	44.43	1.32e-29	1.28e-2
GO:0032501	multicellular organismal process	194	33	17.0	12.57	22.24	2.22e-28	1.08e-2
GO:0061458	reproductive system development	184	32	17.4	12.85	22.51	8.32e-28	1.61e-2
GO:0048608	reproductive structure development	184	32	17.4	12.85	22.51	8.32e-28	1.61e-2
GO:0075259	spore-bearing structure development	184	32	17.4	12.85	22.51	8.32e-28	1.61e-2
GO:0048731	system development	185	32	17.3	12.78	22.36	9.97e-28	1.61e-2
GO:0007275	multicellular organism development	187	32	17.1	12.64	22.07	1.43e-27	1.98e-2