# Linear model parameters

#### **Objectives**

- Understand how to interpret R output and parameters in linear models
- Be able to describe the difference between an interactive and additive model
- Plot predictions from additive and interactive linear models

#### Model parameters: definitions

- Parameters of a linear model typically characterize differences in means
- These are differences per unit of change for continuous predictors,
- These are differences between groups (or between group averages) for categorical predictors
- Interactions are differences between differences

### Coding for categorical predictors: contrasts

- What do the parameters of a linear model mean?
- Start with categorical variables, because they're potentially more confusing ("intercept and slope" isn't too hard)
- Default R behaviour: treatment contrasts
  - $\beta_1$  = expected value in baseline group (= first level of the factor variable, by default the first in alphabetical order);
  - $\beta_i$  = expected difference between group i and the first group.

#### **Example**

- · All model building is about hypothesis testing
- It important to understand which variables go on the x-axis and might determine patterns in your y (response variable)
- I want to test the hypothesis that the number of ant colonies (y) is higher in one place (x) than another.
- My places are field and forest two categorical parameters of the place variable

### The previously explored antcolony example:

#### Define data:

```
pr(lm1 <- lm(colonies~place,data=ants))</pre>
```

```
## Estimate Std. Error t value Pr(>|t|)
## (Intercept) 10.75 0.98 10.97 4.2e-06
## placeforest -3.75 1.27 -2.96 0.018
```

- The (Intercept) row refers to  $\beta_1$ , which is the mean density in the "field" sites ("field" comes before "forest").
- The placeforest row indicates we are looking at the effect of forest level of the place variable, i.e. the difference between "forest" and "field" sites. (To know that "field" is the baseline level we must (1) remember, or look at unique(ants\$place) or (2) notice which level is missing from the list of parameter estimates.)

### Figuring out the estimated values, not the differences

R's behaviour may seem annoying at first – it seems like the estimated values of the groups are what we're really interested in – but it is really designed for testing *differences among groups*. To get the estimates per group, you could:

- predict (base R)
- emmeans package emmeans
- plot(allEffects) package effects
- For your assignment this week, you will can you use the others, but your final graph should use predict

#### Interpretation using predict

Use the predict function:

# Interpretation using effects package

· Use the effects package:

```
library("effects")
summary(allEffects(lm1))
   model: colonies ~ place
##
##
## place effect
## place
## field forest
## 10.75 7.00
##
## Lower 95 Percent Confidence Limits
## place
## field forest
## 8.489484 5.154296
##
   Upper 95 Percent Confidence Limits
## place
      field forest
##
## 13.010516 8.845704
```

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# Interpretation using emmeans package

Use the emmeans package:

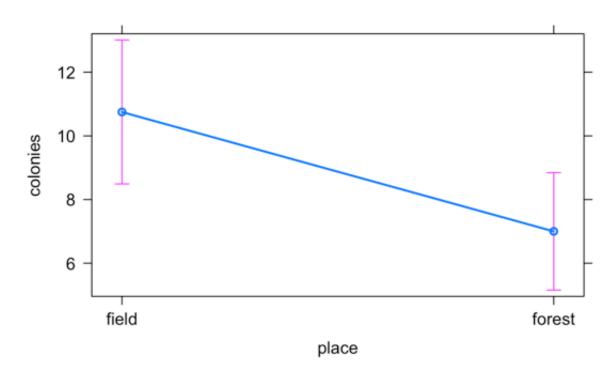
```
library("emmeans")
emmeans(lm1,specs=~place)

## place emmean SE df lower.CL upper.CL
## field 10.8 0.98 8 8.49 13.01
## forest 7.0 0.80 8 5.15 8.85
##
## Confidence level used: 0.95
```

# Graphical summaries from effects package

plot(allEffects(lm1))

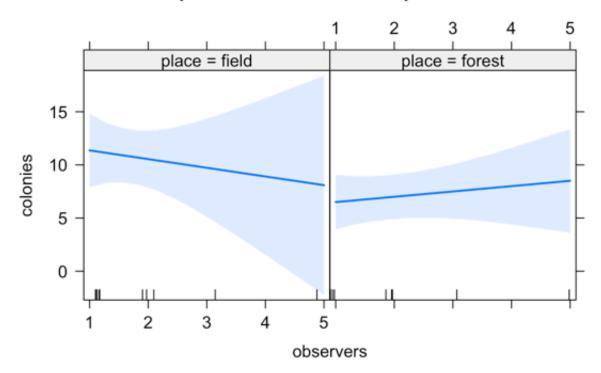
#### place effect plot



# The effects package works on more complicated models

lm3 <- lm(colonies~place\*observers,data=ants)
plot(allEffects(lm3))</pre>

#### place\*observers effect plot

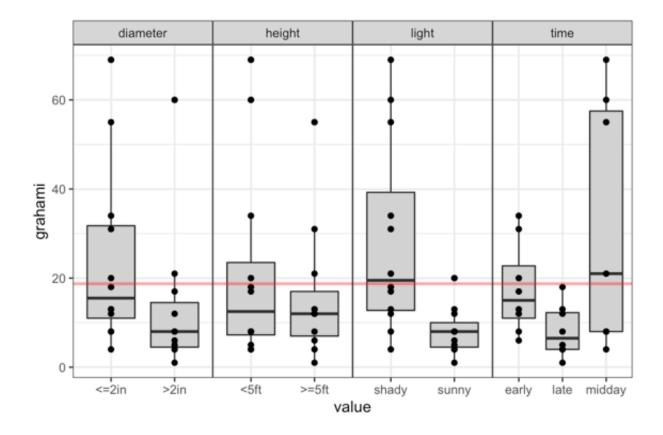


### Switching to a dataset with more than two levels

Some data on lizard perching behaviour (brglm package; Schoener 1970 *Ecology* **51**:408-418).

```
lizards <- read.csv("lizards.csv")</pre>
```

Response is number of *Anolis grahami* lizards found on perches in particular conditions.



### What is the effect of time of day on lizard perching?

```
pr(lmX <- lm(grahami~time,data=lizards))</pre>
```

```
## (Intercept) 17.63 5.97 2.95 0.0079
## timelate -9.50 8.44 -1.13 0.2739
## timemidday 14.52 8.74 1.66 0.1123
```

If we leave the factors alphabetical then  $\beta_1$  ="early",  $\beta_2$  ="late"-"early",  $\beta_3$  ="midday"-"early". It might be more sensible to change the levels in accordance with time progression.

### Change the order of the levels:

This just swaps the definitions of  $\beta_2$  and  $\beta_3$ .

```
pr(lmX <- lm(grahami~time,data=lizards))</pre>
```

```
## (Intercept) 17.63 5.97 2.95 0.0079
## timemidday 14.52 8.74 1.66 0.1123
## timelate -9.50 8.44 -1.13 0.2739
```

### Multiple treatments and interactions

#### Additive model

Consider the light variable in addition to time.

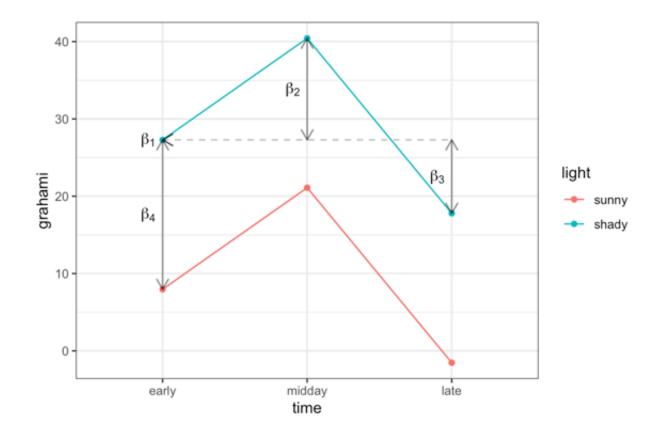
```
pr(lmTL1 <- lm(grahami~time+light,data=lizards))</pre>
```

```
## Estimate Std. Error t value Pr(>|t|)
## (Intercept) 27.29 5.63 4.85 0.00011
## timemidday 13.14 7.11 1.85 0.08010
## timelate -9.50 6.85 -1.39 0.18174
## lightsunny -19.32 5.73 -3.37 0.00321
```

- $\beta_1$  is the intercept ("early", "shady");
- $\beta_2$  and  $\beta_3$  are the differences from the baseline level ("early") of the *first* variable (time) in the *baseline* level of the other parameter(s) (light="shady");
- $\beta_4$  is the difference from the baseline level ("sunny") of the *second* variable (light) in the *baseline* level of time ("early").

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### **Graphical interpretation**



#### What are the p-values?

```
pr(lmTL2 <- lm(grahami~time+light,data=lizards))</pre>
```

```
## (Intercept) 27.29 5.63 4.85 0.00011

## timemidday 13.14 7.11 1.85 0.08010

## timelate -9.50 6.85 -1.39 0.18174

## lightsunny -19.32 5.73 -3.37 0.00321
```

The p-values tell us the difference from the baseline level, not from each other

# Assessing differences among pairs of variable levels - load packages

```
library(emmeans)
library(multcompView)
library(multcomp)
## Loading required package: mvtnorm
## Loading required package: survival
## Loading required package: TH.data
## Loading required package: MASS
##
## Attaching package: 'MASS'
## The following object is masked from 'package:dplyr':
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##
```

### Assessing differences among pairs of variable levels

```
emmeans(lmTL1, specs = "time", contr = "pairwise")
## $emmeans
## time emmean SE df lower.CL upper.CL
## early 17.62 4.85 19 7.48
                                   27.8
   midday 30.76 5.20 19 19.89
##
                                   41.6
   late 8.12 4.85 19 -2.02
##
                                   18.3
##
## Results are averaged over the levels of: light
## Confidence level used: 0.95
##
## $contrasts
## contrast estimate SE df t.ratio p.value
## early - midday -13.1 7.11 19 -1.849 0.1810
                     9.5 6.85 19 1.386 0.3677
   early - late
##
   midday - late 22.6 7.11 19 3.186 0.0129
##
##
## Results are averaged over the levels of: light
## P value adjustment: tukey method for comparing a family o
```

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#### Getting an ABCDEF.. list

```
lsm1<-emmeans(lmTL1,pairwise~time)</pre>
cld(lsm1$emmeans,by = NULL, Letters = "ABCDEFGHIJ")
   time emmean SE df lower.CL upper.CL .group
##
##
   late 8.12 4.85 19
                           -2.02
                                     18.3 A
## early 17.62 4.85 19
                           7.48
                                     27.8 AB
   midday 30.76 5.20 19 19.89
##
                                     41.6 B
##
## Results are averaged over the levels of: light
## Confidence level used: 0.95
## P value adjustment: tukey method for comparing a family o
## significance level used: alpha = 0.05
## NOTE: Compact letter displays can be misleading
        because they show NON-findings rather than findings
##
        Consider using 'pairs()', 'pwpp()', or 'pwpm()' ins
##
```

### Notes about compact letter displays

- This ability may be deprecated
- Compact-letter displays (CLDs) encourage a misleading interpretation of significance testing by visually grouping means whose comparisons have P > 0.05 as though they are equal. (Both get the same letter)
- Failing to prove two means are different does not prove that they are the same.

#### **Interactions**

- Interactions allow the value of one predictor to affect the relationship between another predictor and the response variable
- Interpreting main effects in the presence of interactions is tricky
- Your estimate of the effect of variable  $X_1$  is no longer constant
- You need to pick a fixed point, or average in some way
- · Example:

$$Y = a + b_1 X_1 + b_2 X_2 + b_{12} X_1 * X_2$$

- The response to  $X_1$  is  $Y = (a + b_2X_2) + (b_1 + b_{12}X_2)X_1$
- The response to  $X_1$  depends on the value of  $X_2$ .

#### An example

- You think that the number of lizards on a perch on a sunny day might depend on the time day
- For example, on a very sunny day, there might be fewer lizards perching at noon; whereas on a cloudy day, the number of lizards might be highest at noon

#### **Testing interactions**

- Time is an important factor
- · Use an interaction:

$$M = a + B_x X + B_t t + B_{xt} X t$$

- The interaction term  $B_{xt}$  represents the difference in the response between the two groups.
- It asks: does the number of lizards perching depend on time of day and light?
- Could also write:

$$M = a + B_1 light + B_2 time + B_3 light * time$$

#### Treatment and interactions

- We previously discussed the wrong construction of linear models to compare drug treatments in mutant and non-mutant mice (erroneous interactions)
- You want to know whether a drug that significantly reduces neuronal firing affects mutant or control mice differently
- You find that the drug sig. decreases neuronal firing in mutant mice
- The drug doesn't decrease sig. dec neuronal firing in non-mutants
- You need mouse\_type\*trmt to understand whether the treatment affects mice differently!

#### Interactions and parameters

- We can use CIs, and coefficient plots, and get a pretty good idea what's going on
- In more complicated cases, interaction terms may have many parameters
- These have all the interpretation problems of other multi-parameter variables
- Think about "differences in differences"

#### Interaction model

pr(lmTL2 <- lm(grahami~time\*light,data=lizards))</pre>

##		Estimate	Std. Erro	r t value	Pr(> t
##	(Intercept)	23.50	5.3	8 4.37	0.0004
##	timemidday	27.75	7.6	0 3.65	0.0019
##	timelate	-12.75	7.6	0 -1.68	0.1118
##	lightsunny	-11.75	7.6	0 -1.55	0.1406
##	timemidday:lightsunny	-32.83	11.1	9 –2.93	0.0092
##	timelate:lightsunny	6.50	10.7	5 0.60	0.5534

- Parameters  $\beta_1$  to  $\beta_4$  have the same meanings as before.
- $\beta_5$  and  $\beta_6$ , labelled "timemidday:lightsunny" and "timelate:lightsunny" describe the difference between the expected mean value of these treatment combinations based on the additive model

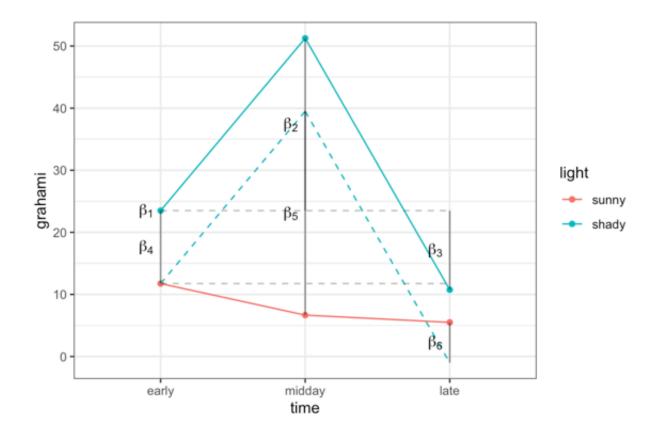
#### Interaction model cont

pr(lmTL2 <- lm(grahami~time\*light,data=lizards))</pre>

##		Estimate Std.	Error t	value	Pr(> t
##	(Intercept)	23.50	5.38	4.37	0.0004
##	timemidday	27.75	7.60	3.65	0.0019
##	timelate	-12.75	7.60	-1.68	0.1118
##	lightsunny	-11.75	7.60	-1.55	0.1406
##	timemidday:lightsunny	-32.83	11.19	-2.93	0.0092
##	timelate:lightsunny	6.50	10.75	0.60	0.5534

- num lizards when sunny at midday =  $\beta_1 + \beta_2 + \beta_4 + \beta_5$
- num lizards when sunny at late =  $\beta_1 + \beta_3 + \beta_4 + \beta_6$

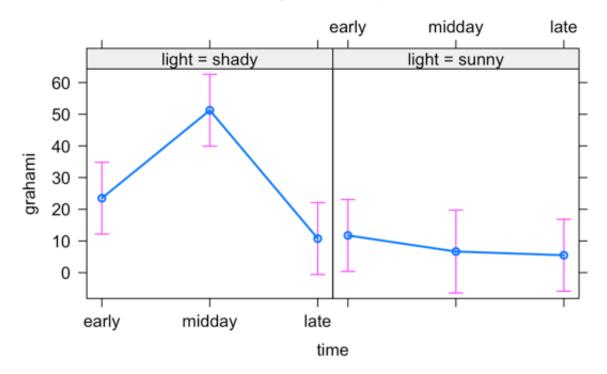
### **Graphical version**



### Effects plot

plot(allEffects(lmTL2))

#### time\*light effect plot



#### Other refs

- http://sas-andr.blogspot.com/2010/10/example-89contrasts.html
- gmodels::fit.contrast() (show parameter estimates based on re-fitting models with new contrasts), rms::contrast.rms() (ditto, for rms-based fits)
- http://www.ats.ucla.edu/stat/r/library/contrast\_codir

#### Assignment - PART 1

- 1. Make a univariate linear model for one of your hypotheses
- 2. Examine the assumptions of linearity (using tests or diagnostic plots) and explain
- Plot the relationship in ggplot using stat\_smooth (continuous) or stat\_summary (discrete)

#### Assignment - PART 2

- Make a linear model (with more than one variable) for one of your hypotheses.
   Articulate which hypothesis you are testing.
- 2. Use an interactive model and an additive model. Explain what hypothesis each of these is testing, and what the R output is telling you about your data. (Hint: you can use emmeans, effects, relevel, or predict to help you.) You should include this explanation in your code.
- 3. Plot your model (e.g. using predict) and overlay the model on top of the underlying data. See code for example to plot both model and data (on github).