Classifier(s) (legacy)

```
set.seed(42)
library(caret)
## Loading required package: ggplot2
## Loading required package: lattice
library(party)
## Loading required package: grid
## Loading required package: mvtnorm
## Loading required package: modeltools
## Loading required package: stats4
## Loading required package: strucchange
## Loading required package: zoo
## Attaching package: 'zoo'
## The following objects are masked from 'package:base':
##
##
       as.Date, as.Date.numeric
## Loading required package: sandwich
library(tidyverse)
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr
              1.1.4 v readr
                                     2.1.5
## v forcats 1.0.0 v stringr 1.5.1
## v lubridate 1.9.3 v tibble
                                   3.2.1
              1.0.2 v tidyr
## v purrr
                                    1.3.1
## -- Conflicts ----- tidyverse_conflicts() --
## x stringr::boundary() masks strucchange::boundary()
## x dplyr::filter() masks stats::filter()
## x dplyr::lag() masks stats::lag()
## x purrr::lift() masks caret::lift()
## x dplyr::where() masks party::where()
## i Use the conflicted package (<a href="http://conflicted.r-lib.org/">http://conflicted.r-lib.org/</a>) to force all conflicts to become error
library(tidymodels)
## -- Attaching packages ------ tidymodels 1.2.0 --
## v broom 1.0.5 v rsample
                                          1.2.1
## v dials
                 1.3.0 v tune
                                          1.2.1
## v diais 1.0.7 v workflows 1.1.4
```

v modeldata 1.4.0 v workflowsets 1.1.0

```
1.2.1 v yardstick 1.3.2
## v parsnip
                 1.1.0
## v recipes
## -- Conflicts ----- tidymodels conflicts() --
## x scales::discard()
                            masks purrr::discard()
                            masks stats::filter()
## x dplyr::filter()
## x parsnip::fit()
                            masks infer::fit(), party::fit(), modeltools::fit()
## x recipes::fixed()
                          masks stringr::fixed()
## x dplyr::lag()
                            masks stats::lag()
## x purrr::lift()
                            masks caret::lift()
## x tune::parameters()
                            masks dials::parameters(), modeltools::parameters()
## x yardstick::precision() masks caret::precision()
## x yardstick::recall()
                            masks caret::recall()
## x yardstick::sensitivity() masks caret::sensitivity()
                            masks readr::spec()
## x yardstick::spec()
## x yardstick::specificity() masks caret::specificity()
## x recipes::step()
                            masks stats::step()
## x recipes::update()
                            masks stats4::update(), stats::update()
## x dplyr::where()
                            masks party::where()
## * Dig deeper into tidy modeling with R at https://www.tmwr.org
```

Load and tidy data

```
pretty_names <- read_csv("../feat_name_mapping.csv")</pre>
## Rows: 85 Columns: 2
## -- Column specification -----
## Delimiter: ","
## chr (2): name_orig, name_pretty
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
data <- read csv("../measurements/measurements.csv")</pre>
## Rows: 754 Columns: 108
## -- Column specification -------
## Delimiter: ","
## chr (20): fpath, KUK_ID, FileName, FileFormat, FolderPath, subcorpus, Source...
## dbl (85): RuleAbstractNouns, RuleAmbiguousRegards, RuleAnaphoricReferences, ...
## lgl (3): ClarityPursuit, SyllogismBased, Bindingness
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
data_no_nas <- data %>%
 select(!c(
   fpath,
   # KUK_ID,
   # FileName,
   FolderPath,
   # subcorpus,
   DocumentTitle,
   ClarityPursuit,
   Readability,
```

```
SyllogismBased,
 SourceDB
)) %>%
# replace -1s in variation coefficients with NAs
mutate(across(c(
  `RuleDoubleAdpos.max_allowable_distance.v`,
  `RuleTooManyNegations.max_negation_frac.v`,
  `RuleTooManyNegations.max_allowable_negations.v`,
  RuleTooManyNominalConstructions.max noun frac.v,
  `RuleTooManyNominalConstructions.max_allowable_nouns.v`,
  `RuleCaseRepetition.max_repetition_count.v`,
  `RuleCaseRepetition.max_repetition_frac.v`,
  `RulePredSubjDistance.max_distance.v`,
  `RulePredObjDistance.max_distance.v`,
  `RuleInfVerbDistance.max_distance.v`,
  `RuleMultiPartVerbs.max distance.v`,
  `RuleLongSentences.max_length.v`,
  `RulePredAtClauseBeginning.max_order.v`,
  `mattr.v`,
  `maentropy.v`
), ~ na if((x, -1))) %>%
# replace NAs with Os
replace_na(list(
 RuleGPcoordovs = 0,
 RuleGPdeverbaddr = 0,
 RuleGPpatinstr = 0,
 RuleGPdeverbsubj = 0,
 RuleGPadjective = 0,
 RuleGPpatbenperson = 0,
 RuleGPwordorder = 0,
 RuleDoubleAdpos = 0,
 RuleDoubleAdpos.max_allowable_distance = 0,
  RuleDoubleAdpos.max allowable distance.v = 0,
 RuleAmbiguousRegards = 0,
 RuleReflexivePassWithAnimSubj = 0,
 RuleTooManyNegations = 0,
  RuleTooManyNegations.max_negation_frac = 0,
 RuleTooManyNegations.max negation frac.v = 0,
  RuleTooManyNegations.max allowable negations = 0,
 RuleTooManyNegations.max_allowable_negations.v = 0,
 RuleTooManyNominalConstructions.max_noun_frac.v = 0,
 RuleTooManyNominalConstructions.max_allowable_nouns.v = 0,
  RuleFunctionWordRepetition = 0,
  RuleCaseRepetition.max_repetition_count.v = 0,
 RuleCaseRepetition.max_repetition_frac.v = 0,
 RuleWeakMeaningWords = 0,
 RuleAbstractNouns = 0,
  RuleRelativisticExpressions = 0,
 RuleConfirmationExpressions = 0,
  RuleRedundantExpressions = 0,
 RuleTooLongExpressions = 0,
 RuleAnaphoricReferences = 0,
 RuleLiteraryStyle = 0,
```

```
RulePassive = 0.
   RulePredSubjDistance = 0,
   RulePredSubjDistance.max distance = 0,
   RulePredSubjDistance.max_distance.v = 0,
    RulePredObjDistance = 0,
   RulePredObjDistance.max_distance = 0,
   RulePredObjDistance.max_distance.v = 0,
   RuleInfVerbDistance = 0,
   RuleInfVerbDistance.max distance = 0,
   RuleInfVerbDistance.max_distance.v = 0,
   RuleMultiPartVerbs = 0,
   RuleMultiPartVerbs.max_distance = 0,
   RuleMultiPartVerbs.max_distance.v = 0,
   RuleLongSentences.max_length.v = 0,
   RulePredAtClauseBeginning.max_order.v = 0,
   RuleVerbalNouns = 0.
   RuleDoubleComparison = 0,
   RuleWrongValencyCase = 0,
   RuleWrongVerbonominalCase = 0,
   RuleIncompleteConjunction = 0
  ))
data_clean <- data_no_nas %>%
  # norm data expected to correlate with text length
  mutate(across(c(
   RuleGPcoordovs,
   RuleGPdeverbaddr,
   RuleGPpatinstr,
   RuleGPdeverbsubj,
   RuleGPadjective,
   RuleGPpatbenperson,
   RuleGPwordorder,
   RuleDoubleAdpos,
   RuleAmbiguousRegards,
   RuleFunctionWordRepetition,
   RuleWeakMeaningWords,
   RuleAbstractNouns,
   RuleRelativisticExpressions,
   RuleConfirmationExpressions,
   RuleRedundantExpressions,
   RuleTooLongExpressions,
   RuleAnaphoricReferences,
   RuleLiteraryStyle,
   RulePassive,
   RuleVerbalNouns,
   RuleDoubleComparison,
   RuleWrongValencyCase,
   RuleWrongVerbonominalCase,
   RuleIncompleteConjunction,
    num hapax,
   RuleReflexivePassWithAnimSubj,
   RuleTooManyNominalConstructions,
   RulePredSubjDistance,
```

```
RuleMultiPartVerbs,
   RulePredAtClauseBeginning
  ), ~ .x / word count)) %>%
  mutate(across(c(
   RuleTooFewVerbs.
   RuleTooManyNegations,
   RuleCaseRepetition,
   RuleLongSentences,
   RulePredObjDistance,
   RuleInfVerbDistance
  ), ~ .x / sent_count)) %>%
  # remove variables identified as "u counts"
  select(!c(
    RuleTooFewVerbs,
   RuleTooManyNegations,
   RuleTooManyNominalConstructions,
   RuleCaseRepetition,
   RuleLongSentences,
   RulePredAtClauseBeginning,
    sent_count,
   word count,
    syllab count,
    char_count
  )) %>%
  # remove variables identified as unreliable
  select(!c(
   RuleAmbiguousRegards,
    RuleFunctionWordRepetition,
   RuleDoubleComparison,
   RuleWrongValencyCase,
    RuleWrongVerbonominalCase
  )) %>%
  # remove artificially limited variables
  select(!c(
   RuleCaseRepetition.max repetition frac,
   RuleCaseRepetition.max_repetition_frac.v
  )) %>%
  # remove further variables belonging to the 'acceptability' category
  select(!c(RuleIncompleteConjunction)) %>%
  unite("strata", c(subcorpus, class), sep = "_", remove = FALSE) %>%
  mutate(across(c(class), ~ as.factor(.x)))
# no NAs should be present now
data_clean[!complete.cases(data_clean), ]
## # A tibble: 754 x 84
##
      KUK_ID
                      FileName FileFormat strata subcorpus SourceID DocumentVersion
                                          <chr> <chr>
##
      <chr>
                      <chr>
                               <chr>
                                                            <chr>
                                                                     <chr>
## 1 673b7a37c6537d~ 002_Kom~ TXT
                                          KUKY_~ KUKY
                                                            <NA>
                                                                     Original
                                          KUKY_~ KUKY
## 2 673b7a37c6537d~ 006 Chc~ TXT
                                                            <NA>
                                                                     Redesign
## 3 673b7a37c6537d~ 004 Nev~ TXT
                                          KUKY ~ KUKY
                                                            <NA>
                                                                     Original
                                          KUKY_~ KUKY
## 4 673b7a37c6537d~ 008_Pol~ TXT
                                                            <NA>
                                                                     Original
## 5 673b7a37c6537d~ 005_Och~ TXT
                                          KUKY_~ KUKY
                                                            <NA>
                                                                     Original
## 6 673b7a37c6537d~ 016_Obc~ TXT
                                          KUKY_~ KUKY
                                                            <NA>
                                                                     Original
```

```
## 7 673b7a37c6537d~ 019_Dět~ TXT
                                           KUKY_~ KUKY
                                                             <NA>
                                                                      Redesign
                                           KUKY_~ KUKY
## 8 673b7a37c6537d~ 007_D\u00fc\u00fc~ TXT
                                                             < NA >
                                                                      Redesign
                                           KUKY ~ KUKY
## 9 673b7a37c6537d~ 024 Opa~ TXT
                                                             < NA >
                                                                      Original
                                           KUKY_~ KUKY
## 10 673b7a37c6537d~ 047_Dav~ TXT
                                                                      Original
                                                             <NA>
## # i 744 more rows
## # i 77 more variables: ParentDocumentID <chr>, LegalActType <chr>,
       Objectivity <chr>, Bindingness <lgl>, AuthorType <chr>,
       RecipientType <chr>, RecipientIndividuation <chr>, Anonymized <chr>,
## #
## #
       `Recipient Type` <chr>, class <fct>, RuleAbstractNouns <dbl>,
## #
       RuleAnaphoricReferences <dbl>,
       RuleCaseRepetition.max_repetition_count <dbl>, ...
```

Filter for features identified as important

This may not be necessary, as the identification was crucial to the EFA above all, so that features irrelevant for readability would not appear in the model. It may be useful to compare the importances of a model trained on all features and on a selected-feature model.

```
## Rows: 67 Columns: 3
## -- Column specification ------
## Delimiter: ","
## chr (1): feat_name
## dbl (1): p_value
## lgl (1): selected
##
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
formula_all <- reformulate(
    selected_features_tibble %>% pull(feat_name), "class"
)
formula_selected <- reformulate(
    selected_features_tibble %>% filter(selected) %>% pull(feat_name), "class"
)
```

Split and folds

[1] 601

```
.split_prop <- 4 / 5
.no_folds <- 10

split <- data_clean %>% initial_split(prop = .split_prop, strata = strata)

training_set <- training(split)

testing_set <- testing(split)

folds <- vfold_cv(training_set, v = .no_folds, strata = strata)

nrow(training_set)</pre>
```

```
training_set %>%
  select(subcorpus, class) %>%
  table()
##
               class
## subcorpus
                bad good
##
     CzCDC
                170
     FrBo
                 62 183
##
##
    KUKY
                 65
                     87
##
    LiFRLaw
                 2
                       0
##
     OmbuFlyers 32
                       0
nrow(testing set)
## [1] 153
testing_set %>%
  select(subcorpus, class) %>%
 table()
##
               class
## subcorpus
                bad good
##
    CzCDC
                 44
##
    FrBo
                 16
                      46
##
    KUKY
                 17
                      23
    LiFRLaw
                       0
##
                  1
     OmbuFlyers
                  6
```

Experimental model

To familiarize myself with the library and CRFs.

```
training_split <- training_set %>%
  initial_split(prop = .split_prop, strata = strata)
train_subset <- training(training_split)</pre>
devtest_subset <- testing(training_split)</pre>
model_rf_exp <- cforest(</pre>
 formula selected,
  data = train_subset, controls = cforest_control(ntree = 1000)
predictions_exp <- predict(model_rf_exp, newdata = devtest_subset)</pre>
confusionMatrix(predictions_exp, devtest_subset$class, positive = "good")
## Confusion Matrix and Statistics
##
##
             Reference
## Prediction bad good
##
               49
                     18
         bad
##
         good 19
                     37
##
##
                   Accuracy : 0.6992
##
                     95% CI: (0.61, 0.7786)
##
       No Information Rate: 0.5528
       P-Value [Acc > NIR] : 0.00063
##
```

```
##
##
                      Kappa: 0.3926
##
   Mcnemar's Test P-Value : 1.00000
##
##
               Sensitivity: 0.6727
##
##
               Specificity: 0.7206
            Pos Pred Value: 0.6607
##
##
            Neg Pred Value: 0.7313
                Prevalence: 0.4472
##
##
            Detection Rate: 0.3008
##
      Detection Prevalence: 0.4553
##
         Balanced Accuracy: 0.6967
##
##
          'Positive' Class : good
##
importances_exp <- varimp(model_rf_exp)</pre>
# computationally expensive
# cimportances_exp <- varimp(model_rf_exp, conditional = TRUE)</pre>
```

MFV model

```
(nrow(data_clean %>% filter(class == "bad")) / nrow(data_clean)) %>%
    round(3)

## [1] 0.55
(nrow(training_set %>% filter(class == "bad")) / nrow(training_set)) %>%
    round(3)

## [1] 0.551
(nrow(testing_set %>% filter(class == "bad")) / nrow(testing_set)) %>%
    round(3)

## [1] 0.549
```

Helpers

```
ntree_tune_levels <- 500 + 0:8 * 250

tune_crf <- function(formula, folds, ntree_tune_levels) {
   accuracy_column <- numeric()
   ntree_column <- numeric()
   fold_column <- numeric()

for (ntree_ in ntree_tune_levels) {
   message(paste0(c("ntree_ ", ntree_), collapse = " "))
   ctrl <- cforest_control(ntree = ntree_)

for (i in seq_len(nrow(folds))) {
   alldata <- pull(folds[i, 1])[[1]]$data
   trindices <- pull(folds[i, 1])[[1]]$in_id</pre>
```

```
trdata <- alldata[trindices, ]</pre>
      tsdata <- alldata[-trindices, ]</pre>
      model <- cforest(formula, data = trdata, controls = ctrl)</pre>
      pred <- predict(model, newdata = tsdata)</pre>
      cm <- confusionMatrix(pred, tsdata$class, positive = "good")</pre>
      ntree_column <- c(ntree_column, ntree_)</pre>
      fold_column <- c(fold_column, i)</pre>
      accuracy_column <- c(accuracy_column, cm$overall["Accuracy"])</pre>
    }
  }
  data.frame(
    ntree = ntree_column,
    fold = fold_column,
    accuracy = accuracy_column
  )
}
```

Selected-features model

Tune

```
tune_df_sel <- tune_crf(formula_selected, folds, ntree_tune_levels)</pre>
## ntree_ 500
## ntree_ 750
## ntree_ 1000
## ntree 1250
          1500
## ntree_
          1750
## ntree_
## ntree_
          2000
## ntree_
          2250
## ntree_ 2500
tune_df_sel %>%
  group_by(ntree) %>%
  summarize(mean_acc = mean(accuracy), sd_acc = sd(accuracy))
## # A tibble: 9 x 3
    ntree mean_acc sd_acc
##
##
   <dbl> <dbl> <dbl>
## 1 500 0.759 0.0226
     750 0.760 0.0361
## 2
## 3 1000 0.757 0.0362
## 4 1250 0.759 0.0346
## 5 1500
             0.762 0.0308
```

```
## 6 1750
             0.762 0.0358
## 7 2000
             0.759 0.0336
## 8 2250
             0.760 0.0393
## 9 2500
             0.760 0.0430
tune_df_sel %>%
  group_by(fold) %>%
  summarize(mean_acc = mean(accuracy), sd_acc = sd(accuracy))
## # A tibble: 10 x 3
##
      fold mean acc sd acc
##
      <dbl>
              <dbl>
                     <dbl>
## 1
         1
              0.806 0.0106
## 2
         2
            0.738 0.00711
              0.750 0.00723
## 3
         3
## 4
         4
             0.741 0.0206
## 5
             0.772 0.00833
## 6
         6
              0.702 0.0194
## 7
         7
              0.748 0.0130
## 8
            0.746 0
         8
## 9
         9 0.805 0.00862
              0.791 0.00575
## 10
        10
best_ntree_sel <- tune_df_sel %>%
 group_by(ntree) %>%
  summarize(mean_acc = mean(accuracy)) %>%
  arrange(-mean_acc) %>%
 head(n = 1) \%
 pull(ntree)
Fit
model_crf_sel <- cforest(</pre>
```

```
formula_selected, training_set,
  controls = cforest_control(ntree = best_ntree_sel)
)
predictions_sel_prob <- predict(</pre>
 model_crf_sel,
 newdata = testing_set, type = "prob"
) %>%
  map(function(x) x[1, 2]) %>%
  unlist() %>%
  as.vector()
predictions_sel <- if_else(predictions_sel_prob > 0.5, "good", "bad") %>%
  as.factor()
confusionMatrix(predictions_sel, testing_set$class, positive = "good")
## Confusion Matrix and Statistics
##
##
             Reference
## Prediction bad good
##
         bad
               62
                    14
##
         good 22
                    55
```

```
##
##
                  Accuracy : 0.7647
                    95% CI: (0.6894, 0.8294)
##
##
       No Information Rate: 0.549
       P-Value [Acc > NIR] : 2.577e-08
##
##
##
                     Kappa: 0.5297
##
##
    Mcnemar's Test P-Value: 0.2433
##
##
               Sensitivity: 0.7971
               Specificity: 0.7381
##
            Pos Pred Value: 0.7143
##
            Neg Pred Value: 0.8158
##
##
                Prevalence: 0.4510
            Detection Rate: 0.3595
##
##
      Detection Prevalence: 0.5033
         Balanced Accuracy: 0.7676
##
##
          'Positive' Class : good
##
##
testing_set_sel <- testing_set %>%
 mutate(.prob = predictions_sel_prob, .pred = predictions_sel)
testing_set_sel %>% ggplot(aes(x = .prob, y = class, color = subcorpus)) +
  geom_jitter(width = 0, height = 0.1)
  good -
                                                                           subcorpus
                                                                               CzCDC
                                                                               FrBo
class
                                                                               KUKY
                                                                               LiFRLaw
                                                                               OmbuFlyers
   bad -
                                                        0.75
       0.00
                        0.25
                                        0.50
```

.prob

```
testing_set_sel %>%
  mutate(abs_dev = abs(0.5 - .prob)) %>%
  filter(class != .pred & abs_dev > 0.25) %>%
  arrange(-abs_dev) %>%
  select(subcorpus, FileName, class, .prob, abs_dev) %>%
  mutate(across(c(.prob, abs_dev), ~ round(.x, 3))) %>%
  as.data.frame()
##
     subcorpus
## 1
         KUKY
## 2
          KUKY
## 3
         FrBo
## 4
         KUKY
## 5
         FrBo
##
                                                                                          FileName
## 1
                                                              0217_6Afs_2000035_20210219141328__1_
## 2
                                                                                     11_vizum_pred
## 3 orig_Co můžete dělat, pokud obec postupuje při prodeji nebo pronájmu pozemků nezákonně_final
## 4
                                                                                          Odvolani
## 5
                                                                   orig_Jak probíhá správní řízení
##
     class .prob abs_dev
## 1 good 0.107
                   0.393
## 2
     good 0.124
                   0.376
## 3
      bad 0.860
                   0.360
## 4 good 0.181
                   0.319
## 5
      bad 0.788
                   0.288
```

All-features model

Tune

```
tune_df_all <- tune_crf(formula_all, folds, ntree_tune_levels)</pre>
## ntree_
           500
## ntree_
           750
## ntree_
           1000
## ntree_
           1250
## ntree_
           1500
## ntree
           1750
## ntree_
           2000
## ntree
           2250
## ntree_ 2500
tune_df_all %>%
  group_by(ntree) %>%
  summarize(mean_acc = mean(accuracy), sd_acc = sd(accuracy))
## # A tibble: 9 x 3
##
     ntree mean_acc sd_acc
     <dbl>
              <dbl> <dbl>
##
```

```
## 1
      500
             0.757 0.0347
## 2
      750
             0.759 0.0438
## 3 1000
             0.759 0.0382
## 4 1250
             0.760 0.0351
## 5 1500
             0.757 0.0339
## 6 1750
             0.757 0.0382
## 7 2000
             0.759 0.0345
## 8 2250
             0.760 0.0340
## 9 2500
             0.759 0.0330
tune_df_all %>%
  group_by(fold) %>%
  summarize(mean_acc = mean(accuracy), sd_acc = sd(accuracy))
## # A tibble: 10 x 3
##
      fold mean_acc sd_acc
##
      <dbl>
              <dbl>
                      <dbl>
##
   1
         1
              0.799 0.00794
##
   2
         2
              0.729 0.00711
## 3
         3
              0.754 0
## 4
         4
              0.743 0.0147
## 5
              0.802 0.0130
         5
## 6
         6
              0.704 0.0162
## 7
         7
              0.741 0.00878
              0.731 0.0102
## 8
         8
              0.793 0
## 9
         9
## 10
        10
              0.791 0.00575
best_ntree_all <- tune_df_all %>%
 group by(ntree) %>%
 summarize(mean_acc = mean(accuracy)) %>%
  arrange(-mean_acc) %>%
 head(n = 1) \%
 pull(ntree)
```

\mathbf{Fit}

```
model_crf_all <- cforest(
  formula_all, training_set,
    controls = cforest_control(ntree = best_ntree_all)
)

predictions_all_prob <- predict(
  model_crf_all,
  newdata = testing_set, type = "prob"
) %>%
  map(function(x) x[1, 2]) %>%
  unlist() %>%
  as.vector()
predictions_all <- if_else(predictions_all_prob > 0.5, "good", "bad") %>%
  as.factor()

confusionMatrix(predictions_all, testing_set$class, positive = "good")
```

Confusion Matrix and Statistics

```
##
##
            Reference
## Prediction bad good
##
        bad 62
         good 22
                   55
##
##
##
                 Accuracy: 0.7647
                    95% CI : (0.6894, 0.8294)
##
##
       No Information Rate: 0.549
##
       P-Value [Acc > NIR] : 2.577e-08
##
##
                     Kappa: 0.5297
##
##
    Mcnemar's Test P-Value: 0.2433
##
##
              Sensitivity: 0.7971
##
              Specificity: 0.7381
           Pos Pred Value: 0.7143
##
##
           Neg Pred Value: 0.8158
               Prevalence: 0.4510
##
           Detection Rate: 0.3595
##
##
      Detection Prevalence: 0.5033
##
         Balanced Accuracy: 0.7676
##
##
          'Positive' Class : good
testing_set_all <- testing_set %>%
  mutate(.prob = predictions_all_prob, .pred = predictions_all)
testing_set_all %>% ggplot(aes(x = .prob, y = class, color = subcorpus)) +
geom_jitter(width = 0, height = 0.1)
```

