Tradict enables high fidelity reconstruction of the eukaryotic transcriptome from 100 marker genes

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Abstract

Transcript levels are a critical determinant of the proteome and hence cellular function. Because the transcriptome is an outcome of the interactions between genes and their products, we reasoned it might be accurately represented by a subset of transcript abundances. We develop a method, Tradict (transcriptome predict), capable of learning and using the expression measurements of a small subset of 100 marker genes to reconstruct entire transcriptomes. By analyzing over 23,000 publicly available RNA-Seq datasets, we show that Tradict is robust to noise and accurate, especially for predicting the expression of a comprehensive, but interpretable list of transcriptional programs that represent the major biological processes and cellular pathways. Coupled with targeted RNA sequencing, Tradict may therefore enable simultaneous transcriptome-wide screening and mechanistic investigation at large scales. Thus, whether for performing forward genetic, chemogenomic, or agricultural screens or for profiling single-cells, Tradict promises to help accelerate genetic dissection and drug discovery.

Introduction

As the critical determinant of the proteome and therefore cellular status, the transcriptome represents a key node of regulation for all life[1]. Transcriptional control is managed by a finely tuned network of transcription factors that integrate environmental and developmental cues in order to actuate the appropriate responses in gene expression[2–4]. Importantly, the transcriptomic state space is constrained. Pareto optimality suggests that no gene expression profile or phenotype can be optimal for all tasks, and consequently, that some expression profiles or phenotypes must come at the expense of others[5,6]. Furthermore, across all major studied kingdoms of life, cellular networks demonstrate remarkably conserved scale-free properties that are topologically characterized by a small minority of highly connected regulatory nodes that link the remaining majority of sparsely connected nodes to the network[7–9]. These theories suggest that the effective dimension of the transcriptome should be far less than the total number of genes it contains. If true to a large enough extent, it may be possible to
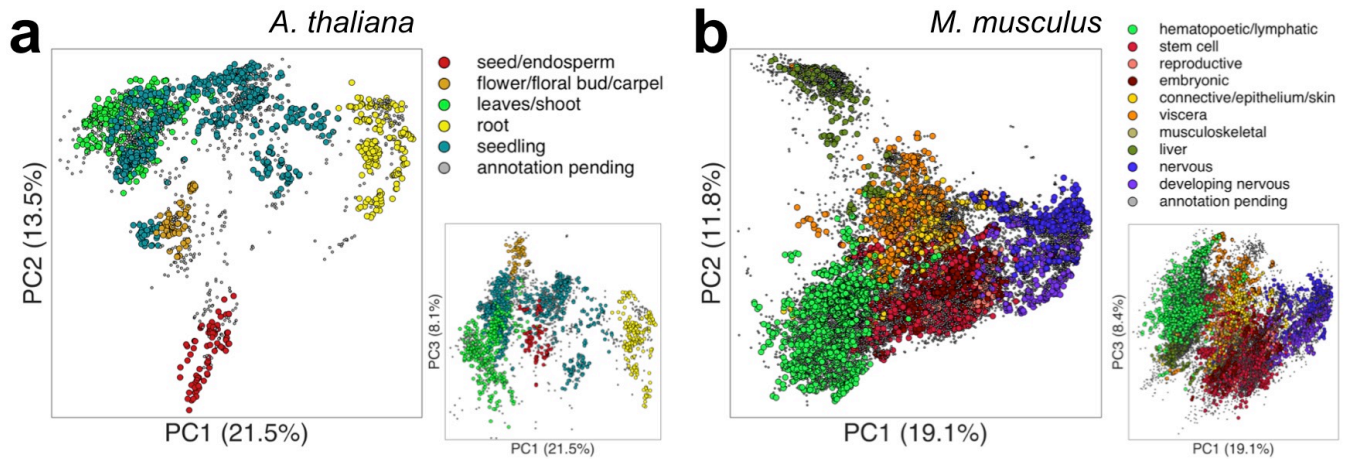
Figure 1. The primary drivers of variation in our training transcriptome collection are developmental stage and tissue. a) *A. thaliana*, b) *M. musculus*. Also shown are plots of PC3 vs. PC1 to provide another perspective.

51 faithfully compress and prospectively reconstruct the entire transcriptomes using only a small, carefully
52 chosen subset of it.
53   Indeed, previous studies have exploited this reduced dimensionality to perform gene expression
54 imputation for microarray data for missing or corrupted values[10–12]. Others have extended these
55 intuitions to predict expression from probe sets containing a few hundred genes[13,14]. However,
56 prediction accuracies have been modest and usually limited to 4,000 target probes/genes. Recently,
57 several studies examined the transcriptomic information recoverable by shallow sequencing especially
58 as it applies to single-cell experiments[15–18]. Jaitin *et al.* (2014) and Pollen *et al.* (2014) demonstrated
59 that only tens of thousands of reads are required per cell to learn and classify cell types *ab initio*[16,18].
60 Heimberg *et al.* (2016) extended these findings and demonstrated that the major principal components
61 of a typically sequenced mouse bulk or single-cell expression dataset may be estimated with little error
62 at even 1% of the depth[15]. Though these approaches advance the notion of strategic transcriptome
63 undersampling, they only recover broad transcriptional states and are restricted to measuring only the
64 most abundant genes. During sample preparation -- arguably the most expensive cost of a multiplexed
65 sequencing experiment -- shallow sequencing-based approaches still utilize protocols meant for
66 sampling the entire transcriptome and thus consume more resources than necessary. Furthermore,
67 given that the expression of even the most abundant genes is highly skewed, sequencing effort is
68 wastefully distributed compared to an approach that chooses which genes to measure more wisely.
69 Finally, it is still not clear from sample sizes and biological contexts previously studied whether the low
70 dimensionality of the transcriptome may be leveraged unconditionally (or nearly so) across organism
71 and application.
72   In this work, we introduce Tradict (transcriptome predict), a novel, robust-to-noise, and
73 probabilistically sound algorithm for inferring the transcriptome using only the expression
74 measurements of a single, context-independent, machine-learned subset of 100 marker genes. Using a
75 transcriptionally representative sampling of over 23,000 publicly available, transcriptome-wide RNA-
76 Seq datasets for *Arabidopsis thaliana* and *Mus musculus*, we train Tradict to prospectively reconstruct
77 gene expression, and to predict, with a striking degree of accuracy, the expression of a comprehensive,
78 but quickly interpretable collection of transcriptional programs that represent the major biological
79 processes and pathways of the cell. Our work demonstrates the development and large-scale
80 application of a multivariate count/non-negative data model, and highlights the power of directly
81 modeling the expression of transcriptional programs in a supervised manner. We believe that Tradict,
82 coupled with targeted RNA sequencing[19–22], can rapidly illuminate biological mechanism and improve
83 the time and cost of performing large forward genetic, breeding, or chemogenomic screens, accurately
84 profiling single-cells, and performing gene expression based clinical diagnostics.
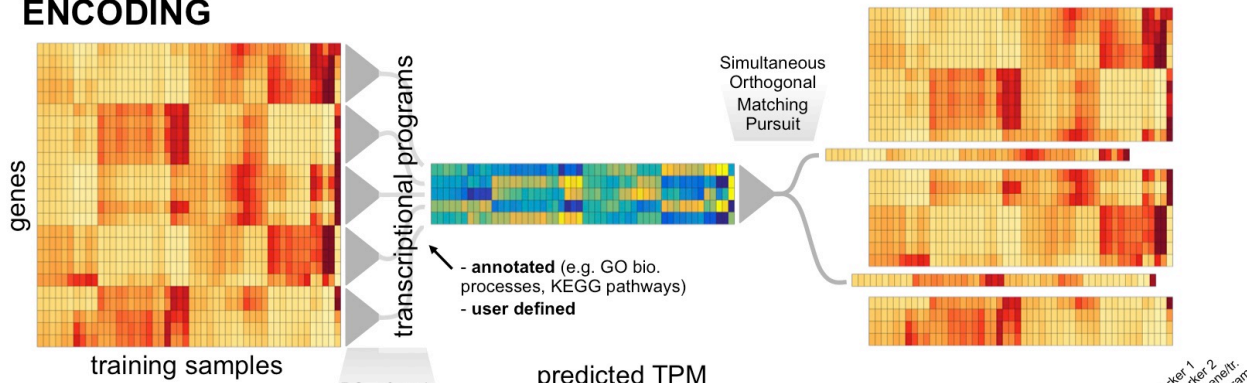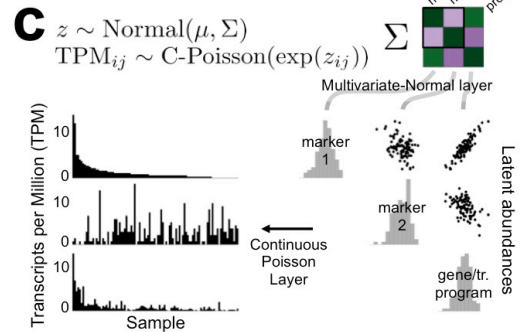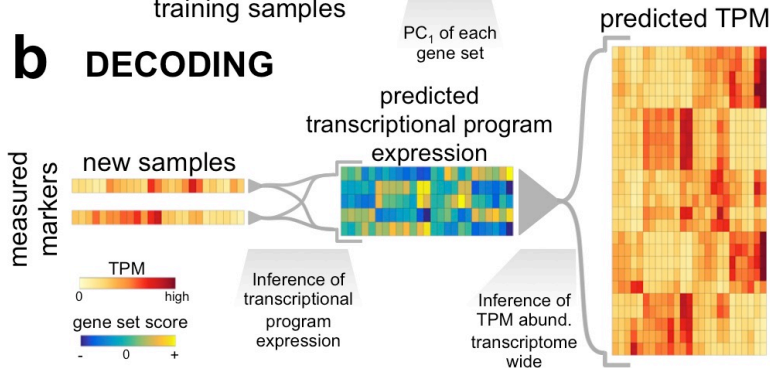
Figure 2. Tradict's algorithmic workflow. a) During encoding the transcriptome is first quantitatively summarized in terms of a collection of a few hundred, biologically comprehensive transcriptional programs. These are then decomposed into a subset of marker genes using an adaptation of the Simultaneous Orthogonal Matching Pursuit algorithm, and a Multivariate Normal Continuous-Poisson hierarchical model is used as a predictive model to capture covariance relationships between markers, transcriptional programs, and all genes. b) During decoding, Tradict predicts the expression of transcriptional programs and all genes in the transcriptome using only expression measurements of the marker genes. c) The Multivariate Normal Continuous-Poisson hierarchy enables Tradict to efficiently model statistical coupling between the non-negative expression measurements typical of sequencing. This is done by assuming that associated with each observed, noisy TPM measurement, there is an unmeasured, denoised latent abundance the logarithm of which comes from a Multivariate Normal distribution over all genes and transcriptional programs.

85
86 **Results**
87
88 **Assembly of a comprehensive training collection of transcriptomes**
89   We attempted to download all available Illumina sequenced publicly deposited RNA-Seq
90 samples (transcriptomes) on NCBI's Sequence Read Archive (SRA). Among samples with at least 4
91 million reads, we successfully downloaded and quantified the raw sequence data of 3,621 and 27,450
92 transcriptomes for *A. thaliana* and *M. musculus,* respectively. After stringent quality filtering, we
93 retained 2,597 (71.7%) and 20,847 (76.0%) transcriptomes comprising 225 and 732 unique SRA
94 submissions for *A. thaliana* and *M. musculus*, respectively. An SRA 'submission' consists of multiple,
95 experimentally linked samples submitted concurrently by an individual or lab. We defined 21,277 (*A.*
96 *thaliana*) and 21,176 (*M. musculus)* measurable genes with reproducibly detectable expression given
97 our tolerated minimum sequencing depth and mapping rates (see Supplemental Information "Materials
98 and Methods" for further information regarding data acquisition, transcript quantification, quality filtering,
99 and expression filtering). We hereafter refer to the collection of quality and expression filtered
100 transcriptomes as our *training transcriptome collection*.
101   In order to assess the quality and comprehensiveness of our training collection, we performed a
102 deep characterization of the expression spaced spanned by these transcriptomes. We found that the

103 transcriptome of both organisms was highly compressible and that the primary drivers of variation were
104 tissue and developmental stage (Figure 1a-b, Figure S1), with many significant, biologically realistic
105 trends within each cluster (Supplemental Note 1). We additionally examined the distribution of
106 submissions across the expression space, compared inter-submission variability within and between
107 tissues, inspected expression correlations among genes with well-established regulatory relationships,
108 and assessed the evolution of the expression space across time. These investigations revealed our
109 training collection is of high and reproducible technical quality, reflective of known biology, and stable
110 (Supplemental Note 1, Figures S2-S4). Given additionally the diversity of tissues, genetic perturbations,
111 and environmental stimuli represented in the SRA, these results, taken together, suggest that our
112 training collection is an accurate and representative sampling of the transcriptomic state space that is of
113 experimental interest for both organisms.
114
115 **Tradict - algorithm overview**
116   Given a training sample of transcriptomes, Tradict encodes the transcriptome into a single
117 subset of globally representative *marker* genes and learns their predictive relationship to the expression
118 of a comprehensive collection of transcriptional programs (e.g. pathways, biological processes) and to
119 the rest of the transcriptome. Tradict's key innovation lies in using a Multivariate Normal Continuous-
120 Poisson (MVN-CP) hierarchical model to model marker latent abundances -- rather than their
121 measured, noisy abundances -- jointly with the expression of transcriptional programs, and ultimately,
122 the latent abundances of the remaining non-marker genes in the transcriptome. In so doing, Tradict is
123 able to 1) efficiently capture covariance structure within the non-negative, right-skewed output typical of
124 sequencing experiments, and 2) perform robust inference of gene set and non-marker expression even
125 in the presence of significant noise.
126   Figure 2 illustrates Tradict's general workflow. Estimates of expression are noisy, especially for
127 low to moderately expressed genes. Given samples are often explored unevenly and that the *a priori*
128 abundance of each gene differs, the level of noise in a gene's measured expression for a given sample
129 varies, but it can be estimated. Therefore, during training, Tradict first learns the log-latent, denoised
130 abundances for each gene in every sample in the training collection using the lag transformation[23]. It
131 then collapses this latent transcriptome into a collection of predefined, globally comprehensive
132 collection of *transcriptional programs* that represent the major processes and pathways of the cell
133 related growth, development, and response to the environment (Supplemental Tables 3-4). In this work,
134 we focus on creating a Gene Ontology derived panel of transcriptional programs, in which the first
135 principal component of all genes contained within an appropriately sized and representative GO term is
136 used to define the similarly named transcriptional program[24,25]. The expression values of these
137 programs are then encoded using an adapted version of the Simultaneous Orthogonal Matching Pursuit
138 into a small subset of marker genes selected from the transcriptome[26,27]. Tradict finally stores the mean
139 and covariance relationships between the log-latent expression of the selected markers, the
140 transcriptional programs, and the log-latent expression of the remaining non-marker genes at the
141 Multivariate Normal layer of the underlying MVN-CP hierarchical model for use in future decoding
142 (Figure 2a).
143   Prospectively, only the expression of these marker genes needs to be measured and the
144 expression of transcriptional programs and/or the rest of the transcriptome can be inferred as needed.
145 During this process of decoding, Tradict first utilizes an iterative Bayesian updating algorithm to learn
146 the log-latent abundances associated with each measured maker for every sample. Though a simply a
147 consequence of proper inference of our model, this denoising step adds considerable robustness to
148 Tradict's predictions. Tradict then uses the covariance relationships learned during training to formulate
149 a prediction for the expression of transcriptional programs and the most likely expression values for all
150 remaining non-marker genes (Figure 2b).
151
152

Tradict prospectively predicts unseen transcriptomes and transcriptional programs with robust accuracy

An SRA submission consists of multiple, experimentally linked samples submitted concurrently by an individual or lab. Consequently, high *intra*-submission prospective prediction accuracy is most indicative of a method's performance.

To more completely understand Tradict's prospective predictive performance, we performed 20-fold cross validation on the training transcriptome collections for both *A. thaliana* and *M. musculus* and evaluated Pearson correlation coefficients (PCC) between predicted and actual expression for each fold when the remaining 95% of folds were used for training. To make this experiment as reflective of reality as possible, folds were divided by submission so that samples from the same set of experiments would not appear both in training and test sets. Because submissions to the SRA span a broad array of biological contexts, the total biological signal contained in the test set exceeds that of what would be expected for typical application, which in turn would lead to overly optimistic estimates of prediction accuracy. To therefore evaluate *intra-submission* accuracy, PCC calculations were performed on 'submission-adjusted' expression values in which each submission's mean expression was subtracted from the expression values of all associated samples.

Figures 3a and 3c illustrate that the reconstruction performance for transcriptional programs in both organisms is strikingly accurate across all collected submissions. Quantitatively speaking, the average intra-submission PCCs for transcriptional programs are 0.94 and 0.93 for *A. thaliana* and *M. musculus*, respectively. This is despite lower, but still accurate prediction performance on gene expression (Figures 3b and 3d). Intuitively, this is because transcriptional programs are measured as linear combinations of the log-latent TPMs of the genes that comprise them, effectively smoothing over the orthogonal noise present in each gene's expression prediction.

Gene expression prediction error was negatively correlated with mean expression (ρ = -0.496 *A. thaliana*, ρ = -0.607 *M. musculus*; Spearman correlation) consistent with the findings of Donner *et al.* 2012. Similarly, for transcriptional programs, prediction error was negatively correlated with the mean expression of genes (ρ = -0.325 *A. thaliana*, ρ = -0.577 *M. musculus*; Spearman correlation) and the number of genes (ρ = -0.545 *A. thaliana*, ρ = -0.5826 *M. musculus*; Spearman correlation) contained within each program (Supplemental Tables 3-4). We did not find any transcriptional programs with relatively low (PCC < 0.9) prediction accuracy that were not composed of a few or lowly expressed genes.

We additionally found Tradict's performance to be superior to several baselines. These include two approaches developed in Donner *et al.* (2012) for microarray[14], and a version of Tradict that uses the 100 most abundant genes as it's selected markers (Figure S5a, Supplemental Note 2). The latter baseline examines the utility of simple shallow sequencing. We further found Tradict's predictions were highly robust to noise (Figure S5b, Supplemental Note 2) -- a consequence of its probabilistic framework, in which training and prediction is performed in the space of denoised latent abundances.

Case studies reveal the power of predicting and studying predefined transcriptional programs

To demonstrate how Tradict may be applied in practice, we focused on two case studies related to innate immune signaling -- one performed using bulk *A. thaliana* seedlings (detailed below), and the other using primary immune *M. musculus* cell lines (detailed in Supplemental Note 3; Figure S6). We trained Tradict on our full collection of training transcriptomes for each organism to produce two organism-specific Tradict models. Each was based on the selection of 100 markers learned from the full training transcriptome collection (Supplemental Tables 5-6) that we assert are globally representative, and context-independent. The case study samples do not, of course, appear in the collection of training transcriptomes.
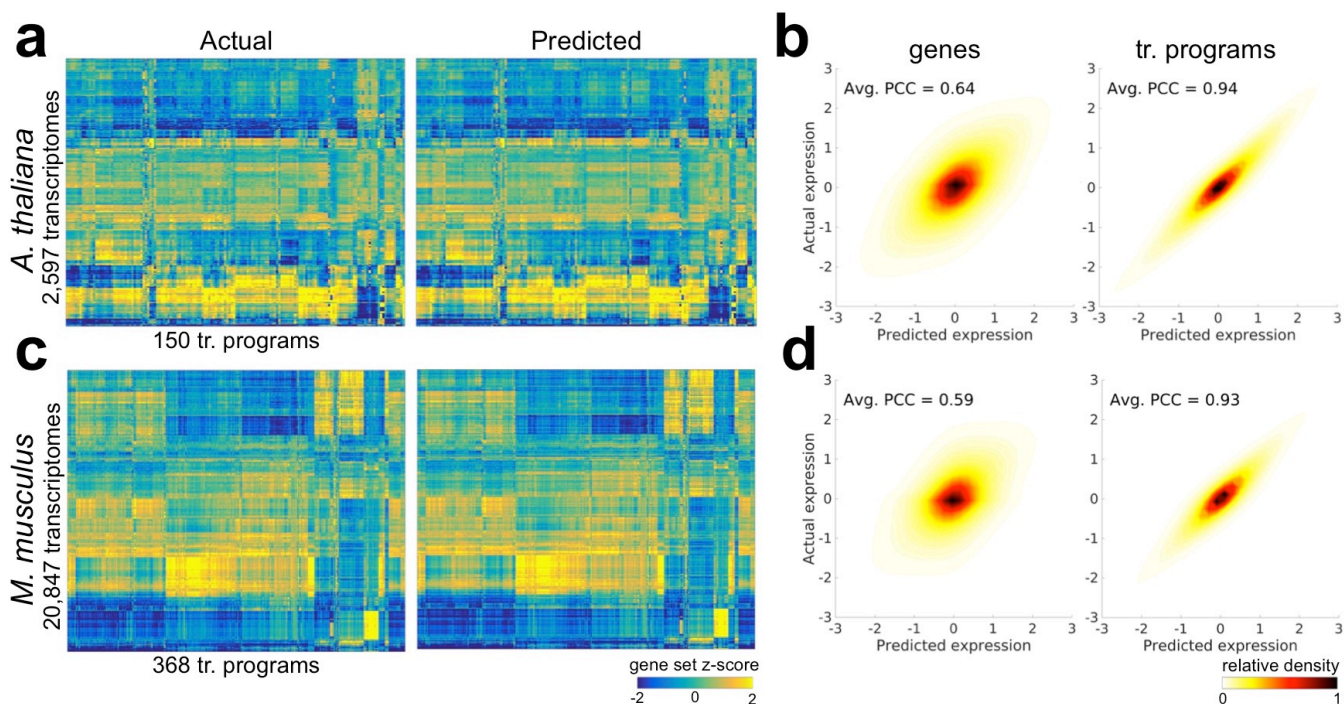
Figure 3. Tradict prospectively predicts unseen transcriptomes with robust accuracy. Tradict's prospective prediction accuracy during 20-fold cross validation of the entire training collection for both organisms. a) Heatmaps illustrating test-set reconstruction performance of all transcriptional programs for *A. thaliana*. b) Density plots of predicted vs. actual test-set expression for all genes (left) and transcriptional programs (right) for *A. thaliana*, after controlling for inter-sumbission biological signal. The intra-submission expression of each gene and transcriptional program was z-score transformed to make their expression comparable. c & d) Same as a & b, but for *M. musculus*.

201
202 **Tradict accurately predicts temporal transcriptomic expression patterns for a diverse panel of**
203 ***A. thaliana* immune signaling mutants under different hormone perturbations -** The hormones
204 salicylic acid (SA) and jasmonic acid (JA) play a major, predominantly antagonistic regulatory role in the
205 activation of plant defense responses to pathogens. Yang *et al.* (2016) investigated the effect of a
206 transgenically expressed bacterial effector, HopBB1, on immune signaling in *A. thaliana*[28]. In their
207 study, they performed a time course experiment, treating plants with MeJA (a JA response inducer),
208 BTH (an SA mimic and SA response inducer), or mock buffer and monitored the transcriptome of bulk
209 seedlings at 0 hr, 1 hr, 5 hr, and 8 hr post treatment. These experiments included several immune
210 signaling mutants with differing degrees of response efficiency to MeJA and BTH treatment. Among
211 other findings, they conclude that HopBB1 enhances the JA response, thereby repressing the SA
212 response and facilitating biotrophic pathogen infection.
213 We asked to what extent strategic undersampling of the transcriptome and application of Tradict
214 could quantitatively recapitulate the findings of Yang *et al.* (2016). Given Tradict's near perfect accuracy
215 on predicting the expression of transcriptional programs, we took a top down, but hypothesis driven
216 approach to our analysis which first examined the expression of all transcriptional programs. Figure 4a
217 illustrates the actual and predicted expression of all transcriptional programs in *A. thaliana* as a function
218 of time and treatment. Here, Tradict reconstructs the expression of all transcriptional programs with an
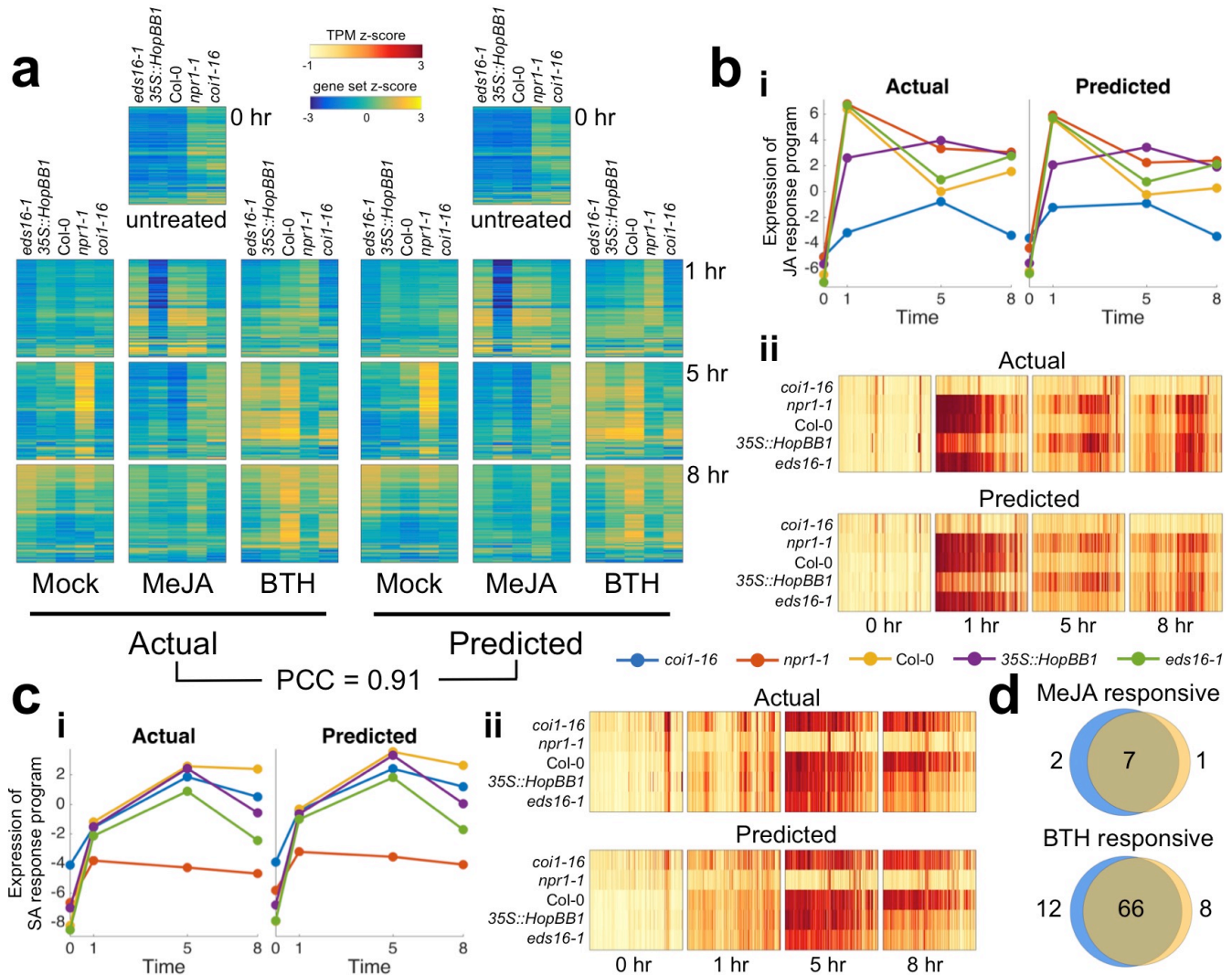219 average PCC of 0.91.

Figure 4. Tradict accurately predicts transcriptional responses across time in response to hormone perturbation in an *A. thaliana* innate immune signaling dataset. After being trained on the full *A. thaliana* training transcriptome collection, the selected set of 100 globally representative and context-independent markers were used to predict the expression of transcriptional programs and all genes for the transcriptomes presented in Yang *et al.* (2016). a) Actual vs. predicted heatmaps for the expression of all 150 transcriptional programs in *A. thaliana* across genotype, time, and hormone treatment. b) Predicted vs. actual expression of i) the JA response transcriptional program, and ii) the genes involved in the JA response program. c i-ii) Same as b, but for the SA response transcriptional program. d) Hypothesis free, differential transcriptional program expression analysis as performed on the actual expression of transcriptional programs vs those predicted by Tradict. Blue circles represent the actual and orange represent the predicted. All heatmaps are clustered in the same order across time, treatment, genotype, and between predicted and actual.

220   Recall that the genes participating in each of our transcriptional programs are pre-defined, in
221 this work, by a carefully chosen, interpretable, but maximally representative set of GO biological
222 processes. Therefore, given the goals of this study, we next examined the expression of the "response
223 to jasmonic acid" and "response to salicylic acid" transcriptional programs. Figure 4b shows the
224 expression behavior for the "response to jasmonic acid" transcriptional program across all the
225 genotypes and time points upon MeJA treatment. More specifically, part (i) shows that the predicted
226 expression and actual expression are qualitatively and quantitatively in agreement, both in magnitude
227 and rank across the different genotypes. For example, as expected, *coi1-16*, which cannot sense JA,
228 does not respond to the MeJA stimulus, while wildtype Col-0 does. However, even more subtle
229 expression dynamics are captured by Tradict's predictions. For example, *eds16-1* and *npr1-1* -- slightly

230 and strongly impaired SA responders, respectively -- are slightly and strongly hyper-responsive to
231 MeJA, respectively -- just as expected from the JA-SA antagonism. Furthermore, as demonstrated in
232 Yang *et al.* (2016), the *35S::HopBB1* transgenic line exhibits a prolonged and sustained JA response
233 for both the actual and predicted expression for this transcriptional program. Part (ii) of Figure 4b
234 illustrates the expression of all the MeJA responsive genes in this transcriptional program. Again
235 Tradict's predictions are in good agreement with actuality, achieving a PCC of 0.72, and it's visually
236 clear that the expression magnitude of these genes positively correlates with the registered expression
237 magnitude of the "response to jasmonic acid" transcriptional program. Figure 4c parts (i) and (ii) are
238 presented in the same light as Figure 4b, but are instead illustrated for the SA response transcriptional
239 program and constituent genes under BTH treatment. Again predictions match actuality, and the
240 observed trends make biological sense[29].
241   In order to illustrate Tradict's use in hypothesis-free investigation, we performed a differential
242 transcriptional program expression analysis for transcriptional programs affected by MeJA or BTH
243 treatment (Figure 4d, see Methods). Differentially expressed transcriptional programs based on
244 Tradict's predictions versus actual measurements were highly concordant and biologically reasonable.
245 Transcriptional programs differentially expressed with respect to MeJA treatment included "response to
246 bacterium," "defense response to fungus", "response to wounding," and "response to jasmonic acid" as
247 expected. Transcriptional programs differentially expressed with respect to BTH treatment included
248 various abiotic stress responses, "defense response to fungus", "response to jasmonic acid" (via
249 antagonism), and "response to salicylic acid," again, as expected.
250
251 **Discussion**
252   Tradict is an accurate, robust-to-noise algorithm for high fidelity transcriptome reconstruction
253 given the expression measurements of a small, machine-learned subset of 100 marker genes. Given
254 the comprehensiveness, stability, and exponentially growing size of the training datasets we have
255 assembled from publicly available sources, the 100 markers Tradict learns are likely to be predictive
256 independent of most contexts and applications. Furthermore, Tradict's ability to near perfectly model
257 the expression of a biologically comprehensive, but interpretable list of annotated transcriptional
258 programs enables one to rapidly generate hypotheses and dissect mechanism.
259   When coupled with target RNA sequencing, we believe Tradict can enable transcriptome-wide
260 screening cheaply at scale. Commercial[19,20] and non-commercial[21,22] methods exist for targeted RNA
261 sequencing in a multiplexed manner, and they are able to measure the expression of 10's-100's of
262 genes with accuracy, making their use compatible with Tradict. Nevertheless, we see improvements in
263 these approaches that could increase multiplexibility and reduce cost. More specifically, we estimate
264 that Tradict coupled with a time and resource efficient targeted RNA-sequencing protocol could bring
265 the cost of obtaining actionable transcriptome-wide information for thousands to tens of thousands of
266 samples to close to $1 a sample (Supplemental Note 4).
267   This scale is exactly what is needed for many high-throughput profiling and screening
268 applications. Many single-cell experiments now profile thousands of cells, and instead of using shallow
269 sequencing (Figure S5), these investigations may benefit from a more targeted effort to query easier-to-
270 measure, but statistically informative transcripts. Tradict could then use these measurements to
271 generate predictions about the status of all transcriptional programs in the cell. With respect to
272 screening, forward genetic screens in most eukaryotic organisms require assaying 10^3-10^4 mutants.
273 Small molecule, or more generally chemogenomic, drug screens often require screening thousands of
274 molecules against multiple cell lines in multiple conditions. Though in these cases the screen is made
275 cheap and scalable by monitoring an easily selectable phenotype, new phenotyping architectures must
276 be developed and optimized for each new screen (e.g. reporter lines, imaging hardware/software).
277 Given the ubiquity of RNA, a transcriptome-wide screening approach does not suffer from such a
278 drawback.
279   Furthermore, and more importantly, though quickly interpretable, the phenotype being screened
280 for is usually a uni-dimensional datum that offers little immediate insight into mechanism. In contrast,

using Tradict to help perform transcriptome-wide screening could strongly couple the process of hypothesis generation and mechanistic investigation. Here, we argue that the scalable monitoring of the expression of a comprehensive list of just a few hundred transcriptional programs affords just the right balance of nuance and interpretability. Consequently, this efficient investigation, largely facilitated by Tradict, could greatly accelerate the pace of genetic dissection, breeding, and drug discovery.

Code availability
A MATLAB implementation of Tradict is available at https://github.com/surgebiswas/tradict. All code to perform data downloads, analysis, and generate figures are available at https://github.com/surgebiswas/transcriptome_compression. Note that in order to make Tradict open-source and reach a wider audience we are developing user-friendly, unit tested R-package of Tradict that will be made available before publication.

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