In-class worksheet 11

Feb 26, 2019

In this worksheet, we will use the library tidyverse and ggthemes:

```
library(tidyverse)
theme_set(theme_bw(base_size = 12)) # set default ggplot2 theme
library(ggthemes)
```

1. Fitting a logistic regression model to the iris data set

We will work with the iris data set. Specifically, with a subset of the data that consists only of the species virginica and versicolor:

```
# make a reduced iris data set that only contains virginica and versicolor spec
ies
iris_small <-
   iris %>%
   filter(Species %in% c("virginica", "versicolor"))
```

Fit a logistic regression model to the iris_small data set. Then successively remove predictors until only predictors with a p value less than 0.1 remain.

```
# logistic regression
glm_out <- glm(
   Species ~ Sepal.Width + Sepal.Length + Petal.Width + Petal.Length,
   data = iris_small,
   family = binomial
) # family = binomial required for logistic regression
summary(glm_out)</pre>
```

```
##
## Call:
## glm(formula = Species ~ Sepal.Width + Sepal.Length + Petal.Width +
      Petal.Length, family = binomial, data = iris small)
##
##
## Deviance Residuals:
##
       Min
                  10
                        Median
                                     30
                                              Max
## -2.01105 -0.00541 -0.00001
                                0.00677
                                          1.78065
## Coefficients:
##
               Estimate Std. Error z value Pr(>|z|)
                -42.638
                        25.707 -1.659
## (Intercept)
                                            0.0972 .
                            4.480 -1.491
## Sepal.Width
                 -6.681
                                            0.1359
## Sepal.Length -2.465
                            2.394 -1.030
                                            0.3032
## Petal.Width
                18.286
                             9.743 1.877
                                            0.0605
## Petal.Length 9.429
                             4.737 1.991
                                            0.0465 *
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## (Dispersion parameter for binomial family taken to be 1)
##
##
      Null deviance: 138.629 on 99 degrees of freedom
## Residual deviance: 11.899 on 95 degrees of freedom
## AIC: 21.899
## Number of Fisher Scoring iterations: 10
```

We remove Sepal. Length as it has the highest P value:

```
glm_out <- glm(
   Species ~ Sepal.Width + Petal.Width + Petal.Length,
   data = iris_small,
   family = binomial
)</pre>
```

```
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
```

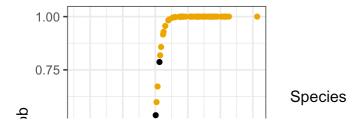
```
summary(glm_out)
```

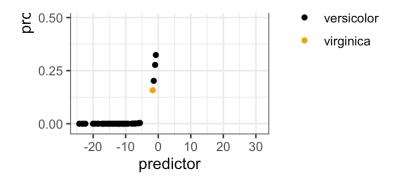
```
##
## Call:
## glm(formula = Species ~ Sepal.Width + Petal.Width + Petal.Length,
##
       family = binomial, data = iris small)
##
## Deviance Residuals:
##
        Min
                   10
                         Median
                                       30
                                                Max
## -1.75795 -0.00412
                        0.00000
                                  0.00290
                                            1.92193
## Coefficients:
##
                Estimate Std. Error z value Pr(>|z|)
                 -50.527
                                     -2.106
## (Intercept)
                             23.995
                                              0.0352 *
## Sepal.Width
                  -8.376
                              4.761 -1.759
                                              0.0785 .
## Petal.Width
                  21.430
                                      2.001
                                              0.0453 *
                             10.707
## Petal.Length
                  7.875
                              3.841
                                      2.050
                                              0.0403 *
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for binomial family taken to be 1)
##
##
       Null deviance: 138.629 on 99
                                      degrees of freedom
## Residual deviance: 13.266 on 96
                                      degrees of freedom
## AIC: 21.266
##
## Number of Fisher Scoring iterations: 10
```

Make a plot of the fitted probability as a function of the linear predictor, colored by species identity. Hint: you will have to make a new data frame combining data from the fitted model with data from the iris_small data frame.

```
lr_data <- data.frame(
   predictor=glm_out$linear.predictors,
   prob=glm_out$fitted.values,
   Species=iris_small$Species
)

ggplot(lr_data, aes(x = predictor, y = prob, color = Species)) +
   geom_point() +
   scale_color_colorblind()</pre>
```



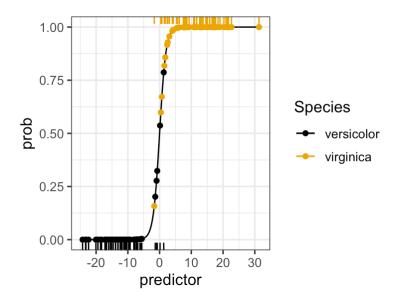


If we want to plot the original observations as "rug" along the bottom or top of the plot, we first need to make two separate data frames with the data points for either species, and then feed them individually into the <code>geom_rug()</code> function. In addition, we make a data frame that contains the logistic function, so we get a smooth black line underneath the points.

```
# extract virginica and versicolor data
virg_data <- filter(lr_data, Species == "virginica")
vers_data <- filter(lr_data, Species == "versicolor")

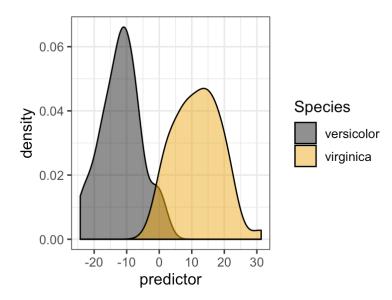
# make data frame with logistic function
predictor <- (-250:320)/10
prob <- exp(predictor)/(1 + exp(predictor))
log_fun_data <- data.frame(predictor, prob)

# plot
ggplot(lr_data, aes(x = predictor, y = prob, color = Species)) +
    geom_line(data = log_fun_data, color = "black") +
    geom_rug(data = vers_data, sides = 'b') +
    geom_rug(data = virg_data, sides = 't') +
    scale_color_colorblind()</pre>
```



Make a density plot that shows how the two species are separated by the linear predictor.

```
ggplot(lr_data, aes(x = predictor, fill = Species)) +
  geom_density(alpha = .5) +
  scale_fill_colorblind()
```



2. Predicting the species

Assume you have obtained samples from three plants, with measurements as listed below. Predict the likelihood that each of these plants belongs to the species virginica.

```
plant1 <- data.frame(</pre>
 Sepal.Length = 6.4,
 Sepal.Width = 2.8,
 Petal.Length = 4.6,
  Petal.Width = 1.8
)
plant2 <- data.frame(</pre>
 Sepal.Length = 6.3,
  Sepal.Width = 2.5,
 Petal.Length = 4.1,
 Petal.Width = 1.7
)
plant3 <- data.frame(</pre>
 Sepal.Length = 6.7,
 Sepal.Width = 3.3,
 Petal.Length = 5.2,
  Petal.Width = 2.3
)
```

```
predict(glm_out, plant1, type = "response")
```

```
## 1
## 0.6934611
```

Plant 1 has a 69% probability of being virginica. It may be a virginica.

```
predict(glm_out, plant2, type = "response")
```

```
## 1
## 0.06002675
```

Plant 2 has a 6% probability of being virginica. It is very likely versicolor.

```
predict(glm_out, plant3, type = "response")
```

```
## 1
## 0.999943
```

Plant 3 has a 99.999% probability of being virginica. It is very likely virginica.

3. If this was easy

Pick a cutoff predictor value at which you would decide that a specimen belongs to virginica rather than versicolor. Calculate how many virginicas you call correctly and how many incorrectly given that choice.

```
cutoff <- 0
virg_true <- sum(lr_data$predictor > cutoff & lr_data$Species == "virginica")
virg_false <- sum(lr_data$predictor <= cutoff & lr_data$Species == "virginica")
virg_true</pre>
```

```
## [1] 49
```

```
virg_false
```

```
## [1] 1
```

Now do the same calculation for versicolor.

```
vers_true <- sum(lr_data$predictor <= cutoff & lr_data$Species == "versicolor")
vers_false <- sum(lr_data$predictor > cutoff & lr_data$Species == "versicolor")
vers_true
```

```
## [1] 48
```

```
vers_false
```

```
## [1] 2
```

If we define a call of virginica as a positive and a call of versicolor as a negative, what are the true positive rate (sensitivity, true positives divided by all possible positives) and the true negative rate (specificity, true negatives divided by all possible negatives) in your analysis?

```
tp <- virg_true/(virg_true + virg_false)
tn <- vers_true/(vers_true + vers_false)
tp</pre>
```

```
## [1] 0.98
```

tn

```
## [1] 0.96
```

The true-positive rate (sensitivity) is 98%.

The true-negative rate (specificity) is 96%.