In-class worksheet 19

Apr 2, 2019

Introduction to Biopython

The Biopython package, available at <u>biopython.org</u>, (http://biopython.org) consists of a large set of helpful functions and tools to solve frequently encountered problems in computational biology. In particular, it has excellent functionality to download and analyze sequence data. It also has a useful module for carrying out analysis of protein structure.

Here, we will start by doing some basic sequence analysis. We will use the biopython modules Entrez and SeqI0:

In [1]: from Bio import Entrez, SeqIO

Entrez provides a computational interface to the widely-use entrez online database provided by the National Institutes of Health: http://www.ncbi.nlm.nih.gov/pubmed. In this database you can find almost any information of interest in biological research.

As an example, let's look at the gene of a recent influenza virus. Specifically, we look at hemagglutinin (HA) from a 2009 swine-flu strain. It is listed by ID number FJ966082.1, and it can be found online here: http://www.ncbi.nlm.nih.gov/nuccore/FJ966082.1, and it can be found online here: http://www.ncbi.nlm.nih.gov/nuccore/FJ966082.1)

We can download this record directly from python using the following Biopython code:

```
In [2]: Entrez.email = "wilke@austin.utexas.edu" # put your email here
        # Download sequence record for genbank id KT220438
        # This is HA gene of Influenza A virus, strain A/NewJersey/NHRC_93219/2015(H3N2
        handle = Entrez.efetch(db="nucleotide", id="KT220438", rettype="qb", retmode="t
        ext")
        record = SeqIO.read(handle, "genbank")
        handle.close()
        # The sequence record is now stored in the variable `record` and
        # we can print it to see what it contains
        print(record)
        ID: KT220438.1
        Name: KT220438
        Description: Influenza A virus (A/NewJersey/NHRC_93219/2015(H3N2)) segment 4 he
        magglutinin (HA) gene, complete cds
        Number of features: 5
        /structured_comment=OrderedDict([('Assembly-Data', OrderedDict([('Sequencing Te
        chnology', 'Sanger dideoxy sequencing')]))])
        /accessions=['KT220438']
        /source=Influenza A virus (A/New Jersey/NHRC_93219/2015(H3N2))
        /data file division=VRL
        /topology=linear
        /date=20-JUL-2015
        /keywords=['']
```

/organism=Influenza A virus (A/New Jersey/NHRC_93219/2015(H3N2))

/references=[Reference(title='GEISS Influenza Surveillance Response Program', .

/taxonomy=['Viruses', 'ssRNA viruses', 'ssRNA negative-strand viruses', 'Orthom

Seq('ATGAAGACTATCATTGCTTTGAGCTACATTCTATGTCTGGTTTTTCGCTCAAAAA...TGA', IUPACAmbigu

Since the record is a simple python object, we can access elements of it as usual:

yxoviridae', 'Influenzavirus A']

..), Reference(title='Direct Submission', ...)]

/molecule_type=cRNA
/sequence version=1

ousDNA())

```
In [3]: print("- ID of the record:")
    print("\n- Brief description of the record:")
    print(record.description)

    print("\n- Annotations that come with the record (given as a python dictionary)
    :")
    print(record.annotations)

    print("\n- The sequence in this record:")
    print(record.seq)
```

- ID of the record: KT220438.1
- Brief description of the record: Influenza A virus (A/NewJersey/NHRC_93219/2015(H3N2)) segment 4 hemagglutinin (HA) gene, complete cds
- Annotations that come with the record (given as a python dictionary): {'structured_comment': OrderedDict([('Assembly-Data', OrderedDict([('Sequencing Technology', 'Sanger dideoxy sequencing')]))]), 'accessions': ['KT220438'], 'so urce': 'Influenza A virus (A/New Jersey/NHRC_93219/2015(H3N2))', 'data_file_div ision': 'VRL', 'topology': 'linear', 'date': '20-JUL-2015', 'keywords': [''], 'organism': 'Influenza A virus (A/New Jersey/NHRC_93219/2015(H3N2))', 'molecule_type': 'cRNA', 'sequence_version': 1, 'references': [Reference(title='GEISS Influenza Surveillance Response Program', ...), Reference(title='Direct Submission', ...)], 'taxonomy': ['Viruses', 'ssRNA viruses', 'ssRNA negative-strand viruses', 'Orthomyxoviridae', 'Influenzavirus A']}
- The sequence in this record: ATGAAGACTATCATTGCTTTGAGCTACATTCTATGTCTGGTTTTCGCTCAAAAAATTCCTGGAAATGACAATAGCACGG CAACGCTGTGCCTTGGGCACCATGCAGTACCAAACGGAACGATAGTGAAAACAATCACAAATGACCGAATTGAAGTTAC TAATGCTACTGAGCTGGTTCAGAATTCCTCAATAGGTGAAATATGCGACAGTCCTCATCAGATCCTTGATGGAGAAAAC TGCACACTAATAGATGCTCTATTGGGAGACCCTCAGTGTGATGGCTTTCAAAATAAGAAATGGGACCTTTTTGTTGAAC GAAGCAAAGCCTACAGCAACTGCTACCCTTATGATGTGCCGGATTATGCCTCCCTTAGGTCACTAGTTGCCTCATCCGG CACACTGGAGTTTAACAATGAAAGCTTCAATTGGACTGGAGTCACTCAAAACGGAACAAGTTCTGCTTGCATAAGGAGA TCTAGTAGTAGTTTCTTTAGTAGATTAAATTGGTTGACCCACTTAAACTACACATACCCAGCATTGAACGTGACTATGC CAAACAATGAACAATTTGACAAATTGTACATTTGGGGGGGTTCACCACCCGGGTACGGACAAGGACCAAATCTTCCTGTA TGCTCAATCATCAGGAAGAATCACAGTATCTACCAAAAGAAGCCAACAAGCTGTAATCCCAAATATCGGATCTAGACCC AGAATAAGGGATATCCCTAGCAGAATAAGCATCTATTGGACAATAGTAAAACCGGGAGACATACTTTTGATTAACAGCA CAGGGAATCTAATTGCTCCTAGGGGTTACTTCAAAATACGAAGTGGGAAAAGCTCAATAATGAGATCAGATGCACCCAT TGGCAAATGCAAGTCTGAATGCATCACTCCAAATGGAAGCATTCCCAATGACAAACCATTCCAAAATGTAAACAGGATC ACATACGGGGCCTGTCCCAGATATGTTAAGCATAGCACTCTAAAATTTGGCAACAGGAATGCGAAATGTACCAGAGAAAC CAGGCATCAAAATTCTGAGGGAAGAGGACAAGCAGCAGATCTCAAAAGCACTCAAGCAGCAATCGATCAAATCAATGGG TTCAGGACCTTGAGAAATATGTTGAGGACACTAAAATAGATCTCTGGTCATACAACGCGGAGCTTCTTGTTGCCCTGGA GAACCAACATACARTTGATCTAACTGACTCAGAAATGAACAAACTGTTTGAAAAAACAAAGAAGCAACTGAGGGAAAAT GCTGAGGATATGGGAAATGGTTGTTTCAAAATATACCACAAATGTGACAATGCCTGCATAGGATCAATAAGAAATGGAA CTTATGACCACAATGTGTACAGGGATGAAGCATTAAACAACCGGTTCCAGATCAAGGGAGTTGAGCTGAAGTCAGGGTA CAAAGATTGGATCCTATGGATTTCCTYTGCCATATCATGTTTTTTGCTTTGTTGCTTTGTTGGGGTTCATCATGTGG

We can also do things like extract the DNA sequence and translate it into a protein sequence:

GCCTGCCAAAAGGGCAACATTAGGTGCAACATTTGCATTTGA

```
In [4]: # extract the sequence from the record
DNA_seq = record.seq

# translate into a protein sequence
protein_seq = DNA_seq.translate()
print(protein_seq)
```

MKTIIALSYILCLVFAQKIPGNDNSTATLCLGHHAVPNGTIVKTITNDRIEVTNATELVQNSSIGEICDSPHQILDGEN CTLIDALLGDPQCDGFQNKKWDLFVERSKAYSNCYPYDVPDYASLRSLVASSGTLEFNNESFNWTGVTQNGTSSACIRR SSSSFFSRLNWLTHLNYTYPALNVTMPNNEQFDKLYIWGVHHPGTDKDQIFLYAQSSGRITVSTKRSQQAVIPNIGSRP RIRDIPSRISIYWTIVKPGDILLINSTGNLIAPRGYFKIRSGKSSIMRSDAPIGKCKSECITPNGSIPNDKPFQNVNRI TYGACPRYVKHSTLKLATGMRNVPEKQTRGIFGAIAGFIENGWEGMVDGWYGFRHQNSEGRGQAADLKSTQAAIDQING KLNRLIGKTNEKFHQIEKEFSEVEGRIQDLEKYVEDTKIDLWSYNAELLVALENQHTXDLTDSEMNKLFEKTKKQLREN AEDMGNGCFKIYHKCDNACIGSIRNGTYDHNVYRDEALNNRFQIKGVELKSGYKDWILWISXAISCFLLCVALLGFIMW ACQKGNIRCNICI*

Problems

Problem 1:

- (a) Download the sequence record with the ID number FJ966082 and print it out. What kind of a sequence is this?
- (b) Print out the comments section of the annotation
- (c) Translate the DNA sequence into a protein sequence and print it out

```
In [5]: # Problem 1a
                 # you will need to import the appropriate modules for this to work
                 from Bio import Entrez, SeqIO
                 # always give Entrez your email
                 Entrez.email = "wilke@austin.utexas.edu"
                 # download sequence record for genbank id FJ966082
                 # This is HA gene of Influenza A virus, strain A/California/04/2009(H1N1)
                 handle = Entrez.efetch(db="nucleotide", id="FJ966082", rettype="qb", retmode="t
                 record = SeqIO.read(handle, "genbank")
                 handle.close()
                 # print the record
                 print(record)
                 ID: FJ966082.1
                Name: FJ966082
                Description: Influenza A virus (A/California/04/2009(H1N1)) segment 4 hemagglut
                 inin (HA) gene, complete cds
                Database cross-references: BioProject:PRJNA37813
                Number of features: 6
                 /structured_comment=OrderedDict([('FluData', OrderedDict([('Isolate', 'A/Califo
                 \label{eq:rnia-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-delta-d
                 ('Passage_history', 'CX'), ('Adamantane_resistance', 'resistant'), ('Zanamivir_resistance', 'sensitive'), ('Oseltamivir_resistance', 'sensitive'), ('Country',
                 'USA'), ('State/Province', 'California'), ('Collection_day', '1'), ('Collection
                  _month', '4'), ('Collection_year', '2009'), ('EPI_isolate', 'GISAID EPI_ISL_295
                 73'), ('EPI accession', 'EPI176470')]))])
                 /accessions=['FJ966082']
                 /source=Influenza A virus (A/California/04/2009(H1N1))
                 /data file division=VRL
                 /topology=linear
                 /date=02-SEP-2010
                 /keywords=['']
                 /organism=Influenza A virus (A/California/04/2009(H1N1))
                 comment=GenBank Accession Numbers FJ966079-FJ966086 represent sequences
                 from the 8 segments of Influenza A virus
                 (A/California/04/2009(H1N1)).
                 Swine influenza A (H1N1) virus isolated during human swine flu
                 outbreak of 2009. For more information, see http://www.cdc.gov/.
                 Some of the information does not have GenBank feature identifiers
                 and is being provided in the comment section.
                 /molecule_type=cRNA
                 /sequence_version=1
                 /references=[Reference(title='Emergence of a novel swine-origin influenza A (H1
                N1) virus in humans', ...), Reference(title='Antigenic and genetic characterist
                ics of swine-origin 2009 A(H1N1) influenza viruses circulating in humans', ...)
                 , Reference(title='Direct Submission', ...)]
/taxonomy=['Viruses', 'ssRNA viruses', 'ssRNA negative-strand viruses', 'Orthom'
                 yxoviridae', 'Influenzavirus A']
```

5 of 8 4/9/19, 12:01 PM

ousDNA())

Seg('ATGAAGGCAATACTAGTAGTTCTGCTATATACATTTGCAACCGCAAATGCAGAC...TAA', IUPACAmbiqu

In [6]: # Problem 1b

print(record.annotations['comment'])

GenBank Accession Numbers FJ966079-FJ966086 represent sequences from the 8 segments of Influenza A virus (A/California/04/2009(H1N1)). Swine influenza A (H1N1) virus isolated during human swine flu

outbreak of 2009. For more information, see http://www.cdc.gov/. Some of the information does not have GenBank feature identifiers and is being provided in the comment section.

In [7]: # Problem 1c

protein seq = record.seq.translate() print("Protein sequence:") print(protein_seq)

Protein sequence:

MKAILVVLLYTFATANADTLCIGYHANNSTDTVDTVLEKNVTVTHSVNLLEDKHNGKLCKLRGVAPLHLGKCNIAGWIL GNPECESLSTASSWSYIVETPSSDNGTCYPGDFIDYEELREQLSSVSSFERFEIFPKTSSWPNHDSNKGVTAACPHAGA KSFYKNLIWLVKKGNSYPKLSKSYINDKGKEVLVLWGIHHPSTSADQQSLYQNADTYVFVGSSRYSKKFKPEIAIRPKV RDQEGRMNYYWTLVEPGDKITFEATGNLVVPRYAFAMERNAGSGIIISDTPVHDCNTTCQTPKGAINTSLPFQNIHPIT ${\tt IGKCPKYVKSTKLRLATGLRNIPSIQSRGLFGAIAGFIEGGWTGMVDGWYGYHHQNEQGSGYAADLKSTQNAIDEITNK}$ VNSVIEKMNTOFTAVGKEFNHLEKRIENLNKKVDDGFLDIWTYNAELLVLLENERTLDYHDSNVKNLYEKVRSOLKNNA KEIGNGCFEFYHKCDNTCMESVKNGTYDYPKYSEEAKLNREEIDGVKLESTRIYQILAIYSTVASSLVLVVSLGAISFW MCSNGSLOCRICI*

Problem 2:

Print the record you downloaded under Problem 1 in FASTA format. This means that you need to first print a line starting with ">" plus some description of the record. Then you need to print a line containing the sequence in the record.

In [8]: print(">", record.id, record.description) print(record.seq)

> > FJ966082.1 Influenza A virus (A/California/04/2009(H1N1)) segment 4 hemagglut inin (HA) gene, complete cds

> ATGAAGGCAATACTAGTAGTTCTGCTATATACATTTGCAACCGCAAATGCAGACACATTATGTATAGGTTATCATGCGA ACAATTCAACAGACACTGTAGACACAGTACTAGAAAAGAATGTAACAGTAACACTCTGTTAACCTTCTAGAAGACAA GGAAATCCAGAGTGTGAATCACTCTCCACAGCAAGCTCATGGTCCTACATTGTGGAAACACCTAGTTCAGACAATGGAA CGTGTTACCCAGGAGATTTCATCGATTATGAGGAGCTAAGAGAGCAATTGAGCTCAGTGTCATCATTTGAAAGGTTTGA GATATTCCCCAAGACAAGTTCATGGCCCAATCATGACTCGAACAAAGGTGTAACGGCAGCATGTCCTCATGCTGGAGCA AAAAGCTTCTACAAAAATTTAATATGGCTAGTTAAAAAAGGAAATTCATACCCAAAGCTCAGCAAATCCTACATTAATG TGCAGATACATATGTTTTTGTGGGGTCATCAAGATACAGCAAGAAGTTCAAGCCGGAAATAGCAATAAGACCCAAAGTG AGGGATCAAGAAGGGAGAATGAACTATTACTGGACACTAGTAGAGCCGGGAGACAAAATAACATTCGAAGCAACTGGAA CGATTGCAATACAACTTGTCAAACACCCAAGGGTGCTATAAACACCCAGCCTCCCATTTCAGAATATACATCCGATCACA ATTGGAAAATGTCCAAAATATGTAAAAAGCACAAAATTGAGACTGGCCACAGGATTGAGGAATATCCCGTCTATTCAAT CCATCAAAATGAGCAGGGTCAGGATATGCAGCCGACCTGAAGAGCACACAGAATGCCATTGACGAGATTACTAACAAA GTAAATTCTGTTATTGAAAAGATGAATACACAGTTCACAGCAGTAGGTAAAGAGTTCAACCACCTGGAAAAAAGAATAG AGAATTTAAATAAAAAAGTTGATGATGGTTTCCTGGACATTTGGACTTACAATGCCGAACTGTTGGTTCTATTGGAAAA TGAAAGAACTTTGGACTACCACGATTCAAATGTGAAGAACTTATATGAAAAGGTAAGAAGCCAGCTAAAAAAACAATGCC AAGGAAATTGGAAACGGCTGCTTTGAATTTTACCACAAATGCGATAACACGTGCATGGAAAGTGTCAAAAATGGGACTT ATGACTACCCAAAATACTCAGAGGAAGCAAAATTAAACAGAGAAGAAATAGATGGGGTAAAGCTGGAATCAACAAGGAT TTACCAGATTTTGGCGATCTATTCAACTGTCGCCAGTTCATTGGTACTGGTAGTCTCCCTGGGGGCCAATCAGTTTCTGG ATGTGCTCTAATGGGTCTCTACAGTGTAGAATATGTATTTAA

If this was easy

Problem 3:

Write a function that takes a sequence record as input and prints it out in FASTA format. Write the function in such a way that it breaks the sequence over multiple lines, such that each line contains at most 60 characters.

```
In [9]: def print_fasta(record):
    print(">", record.id, record.description)
    seq = record.seq
    for i in range(0, len(seq), 60):
        print(seq[i:i+60])

print_fasta(record)
```

> FJ966082.1 Influenza A virus (A/California/04/2009(H1N1)) segment 4 hemagglut inin (HA) gene, complete cds

ATGAAGGCAATACTAGTTCTGCTATATACATTTGCAACCGCAAATGCAGACACATTA TGTATAGGTTATCATGCGAACAATTCAACAGACACTGTAGACACAGTACTAGAAAAGAAT GTAACAGTAACACTCTGTTAACCTTCTAGAAGACAAGCATAACGGGAAACTATGCAAA AATCCAGAGTGTGAATCACTCTCCACAGCAAGCTCATGGTCCTACATTGTGGAAACACCT AGTTCAGACATGGAACGTGTTACCCAGGAGATTTCATCGATTATGAGGAGCTAAGAGAG CAATTGAGCTCAGTGTCATCATTTGAAAGGTTTGAGATATTCCCCAAGACAAGTTCATGG CCCAATCATGACTCGAACAAAGGTGTAACGGCAGCATGTCCTCATGCTGGAGCAAAAAGC TTCTACAAAAATTTAATATGGCTAGTTAAAAAAGGAAATTCATACCCAAAGCTCAGCAAA ACTAGTGCTGACCAACAAGTCTCTATCAGAATGCAGATACATATGTTTTTGTGGGGTCA TCAAGATACAGCAAGAAGTTCAAGCCGGAAATAGCAATAAGACCCAAAGTGAGGGATCAA GAAGGGAGAATGAACTATTACTGGACACTAGTAGAGCCGGGAGACAAAATAACATTCGAA GGTATTATCATTTCAGATACACCAGTCCACGATTGCAATACAACTTGTCAAACACCCAAG GGTGCTATAAACACCAGCCTCCCATTTCAGAATATACATCCGATCACAATTGGAAAATGT CCAAAATATGTAAAAAGCACAAAATTGAGACTGGCCACAGGATTGAGGAATATCCCGTCT ATTCAATCTAGAGGCCTATTTGGGGCCATTGCCGGTTTCATTGAAGGGGGGTGGACAGGG ATGGTAGATGGATGGTACGGTTATCACCATCAAAATGAGCAGGGGTCAGGATATGCAGCC GACCTGAAGAGCACACAGAATGCCATTGACGAGATTACTAACAAAGTAAATTCTGTTATT GAAAAGATGAATACACAGTTCACAGCAGTAGGTAAAGAGTTCAACCACCTGGAAAAAAGA ATAGAGAATTTAAATAAAAAAGTTGATGATGGTTTCCTGGACATTTGGACTTACAATGCC GAACTGTTGGTTCTATTGGAAAATGAAAGAACTTTGGACTACCACGATTCAAATGTGAAG AACTTATATGAAAAGGTAAGAAGCCAGCTAAAAAACAATGCCAAGGAAATTGGAAACGGC TGCTTTGAATTTTACCACAAATGCGATAACACGTGCATGGAAAGTGTCAAAAATGGGACT TATGACTACCCAAAATACTCAGAGGAAGCAAAATTAAACAGAGAAGAAATAGATGGGGTA AAGCTGGAATCAACAAGGATTTACCAGATTTTGGCGATCTATTCAACTGTCGCCAGTTCA TTGGTACTGGTAGTCTCCCTGGGGGCAATCAGTTTCTGGATGTGCTCTAATGGGTCTCTA CAGTGTAGAATATGTATTTAA

Problem 4:

Write a function that takes a DNA sequence as input, translates the sequence in all three reading frames, and counts the number of stop codons found in each translation. Remember that stop codons are indicated by a "*".

Note that biopython doesn't like to translate sequences whose length is not a multiple of three, so you will have to pad the sequence with trailing Ns to avoid a warning or error.

Hint: The solution to this problem will be simpler if you first write a function that can count the stop codons in one sequence.

```
In [10]: def count_stop(aa_seq):
             count = 0
             for c in aa_seq:
                 if c == "*":
                     count += 1
             return count
         def translate_all_frames(seq):
             # frame 1
             count1 = count_stop(seq.translate())
             # frame 2
             seq2 = seq[1:]+"N"
             count2 = count_stop(seq2.translate())
             # frame 3
             seq3 = seq[2:]+"NN"
             count3 = count_stop(seq3.translate())
             # return all three counts
             return (count1, count2, count3)
         counts = translate_all_frames(record.seq)
         print(counts)
```

(1, 42, 33)