

Bioinformatics Resources Project 2024

Execute the following tasks and comment the results you obtain. You can either:

- 1) Provide an R script with the code and a separate PDF report describing your analyses and the results you obtained;
- 2) Use frameworks like R markdown to provide both the report and the code together in the same document. In this case provide a final readable format (e.g. PDF or html).

Select one among the RData files available (please be sure that no more than 2 groups select the same dataset) representing RNA-seq count data extracted from different cancer datasets from the Cancer Genome Atlas (TCGA). From the original TCGA data cases (tumor samples) and controls (normal samples) were randomly selected.

1. Load the RData file. The following three data-frames are available:
 - a `raw_counts_df` = contains the raw RNA-seq counts;
 - b `c_anno_df` = contains sample names and conditions (case or control);
 - c `r_anno_df` = contains the ENSEMBL genes ids, the length of the genes and the genes symbols.
2. Update `raw_count_df` and `r_anno_df` extracting only protein coding genes:
 - a Use `biomaRt` package to retrieve the needed information;
 - b Next tasks should use the new data-frames you have created.
3. Perform a differential expression analysis using `edgeR` package and select up- and down-regulated genes using an adjusted p-value cutoff of 0.01, a log fold change ratio >1.5 for up-regulated genes and $< (-1.5)$ for down-regulated genes and a log CPM >1 . Relax the thresholds if no or few results are available.
 - a Use the workflow we developed during the course;
 - b Filter raw counts data retaining only genes with a raw count >20 in at least 5 Cases or 5 Control samples;
 - c Create a volcano plot of your results;

- d Create an annotated heatmap focusing only on up- and down-regulated genes.

4. Perform gene set enrichment analysis using clusterProfiler R package.

- a Perform both GO (BP and MF) and WP analysis;
- b Report the top 10 enriched GO terms and the top 10 enriched WP pathways; resulting from both up- and down-regulated gene lists.

5. Use the pathview R package to visualize one pathway you find enriched using the up-regulated gene list.

6. Identify which transcription factors (TFs) have enriched scores in the promoters of all up-regulated (or down-regulated if you prefer) genes.

- a use a window of 500 nucleotides upstream each gene.

7. Select one among the top enriched TFs, compute the empirical distributions of scores for all PWMs that you find in MotifDB for the selected TF and determine for all of them the distribution (log2) threshold cutoff at 99.75% (relax the threshold if needed).

8. Identify which up-regulated (or down-regulated depending on the choice you made at point 7) genes have a region in their promoter (defined as previously) with binding scores above the computed thresholds for any of the previously selected PWMs.

- a Use pattern matching as done during the course;

9. Use STRING database to find PPI interactions among differentially expressed genes and export the network in TSV format.

10. Import the network in R and using igraph package identify and plot the largest connected component.