

## APPLICATIONS NOTE

# MOBY Web services to support promoter analysis protocols

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### ABSTRACT

Summary: Here we present a computational protocol to analyse the promoter regions of a given set of co-expressed genes, and its implementation by developing a set of Web services that are compliant with MOBY specifications. The main goal of this study is to show that in-silico experiments of genomic data can be done in an automatized manner using Web resources.

Availability : {{[http://genome.imim.es/cgi-bin/moby/GeneClustering\\_DataSubmission.cgi](http://genome.imim.es/cgi-bin/moby/GeneClustering_DataSubmission.cgi)}}

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Supplementary information :

{{<http://genome.imim.es/webservices/index.html>}}

## 1 INTRODUCTION

With the completion of many sequencing projects, there are tremendous amount of data and, coming along, of analysis methods that are being made available through the Web to the scientific community. While these resources are of great help for retrieval of data, and quick hypothesis verification, they are mostly used through manual execution and can not be applied for automated tasks. This implies some drawbacks, such as slowness and being error-prone when executed repeatedly.

*In silico* experiments, on the other hands, are described in protocols that can be seen as an orchestrated execution of atomic steps. Such computational protocols are commonly implemented using a script language such as Perl. The various steps may be executed on local resources, but increasingly often using remote resources.

In this regard, Web services architecture (Web Services Architecture specifications document, {{<http://www.w3.org/TR/ws-arch/>}}) have emerged to provide programmatic access to remote resources, thus allowing users to perform *in silico* experiments through the Web in an automatic manner (Stevens *et al.*, 2004).

We have applied such technology to develop a pipeline for the characterization of the promoter regions of co-regulated genes. It is generally assumed that genes with similar transcriptional regulatory programs also exhibit similar configurations of Transcription Factor (TF) Binding Sites (TFBS) in their promoter regions upstream of the Transcription Start Site (TSS) (Wray *et al.*, 2003). Because TFBSs are short DNA motifs (8 to 15bp in range), they can occur by chance very often in DNA sequences, thus producing a high level of false positives. To differentiate false positive predictions from truly functional elements, new methods have been proposed (for a review, see Wasserman *et al.*, 2004). In addition, promoter elements bound by the same TF may not show sequence similarity and, therefore,

sequence comparisons between promoter elements of co-expressed genes often fail to reveal the underlying common regulatory domains. To address this limitation, Blanco *et al.* (2005) introduced TF-map alignments. In these, TFBS on promoter sequences are labeled according to the corresponding TF, and the comparison is performed between the sequence of labels. TF-map alignments have been shown to uncover common regulatory domains, which can not be detected by typical sequence comparisons. Here we have developed and automated a protocol which clusters a given set of co-regulated genes in subsets of genes showing similar configurations of regulatory elements as revealed by TF-map alignments.

## 2 PROTOCOL

The protocol is schematized in Figure 1. Given a set of gene identifiers, in the first step, the upstream sequences of the genes are automatically extracted from the Ensembl database (Birney *et al.*, 2006). It is also possible to directly provide the upstream sequences in FASTA format. The second step is the search for putative TFBSs in the sequences. Two public position weight matrices (PWMs) libraries are available, Jaspar (Vlieghe *et al.*, 2006) and Transfac (v6.4) (Matys *et al.*, 2006). This step is performed using MatScan software (E. Blanco, unpublished). The third step performs the pairwise alignments of the TFBSs maps using the TF-alignment software (Blanco *et al.*, 2005). In the fourth step, the pairwise alignment scores are parsed to generate a score matrix. In the fifth step, a K-means clustering is applied to partition the gene space into clusters according to the score of the alignments of the TFBSs maps. This step is performed using Cluster v3.0 software (de Hoon *et al.*, 2004). Finally, for each gene cluster, the sixth step consists in running the multiple TF-map alignment software (E. Blanco, unpublished) to define a consensus “transcriptional regulatory pattern”.

## 3 IMPLEMENTATION

At each step of this procedure corresponds a Web service that has been implemented following the BioMOBY specifications (Wilkinson *et al.*, 2002). To facilitate the execution of the procedure, a data submission page has been setup at the following URL, {{[http://genome.imim.es/cgi-bin/moby/GeneClustering\\_DataSubmission.cgi](http://genome.imim.es/cgi-bin/moby/GeneClustering_DataSubmission.cgi)}}.

The various Web services that compose our procedure can also be executed separately or integrated into other pipelines of analysis. To this effect, one can use the stand-alone application called Taverna

(Hull *et al.*, 2006) or Web-based tools such as Remora (Carrere *et al.*, 2006) or MOWServ (Navas-Delgado *et al.*, 2006).

#### 4 DISCUSSION

Web services technologies look promising to federate computational Bioinformatics resources through the Web. Here we have applied the MOBY framework to develop a set of Web services that can be combined together to allow the execution of a new pipeline of analysis of the promoter regions of co-expressed genes. The different steps of the pipeline may be executed in different distant computational resources, but this is totally transparent to the user. The pipeline encompasses many steps that the users would otherwise need to perform individually, and ensure, therefore the reproducibility of this *in silico* experiment. Furthermore, because MOBY Web services are formally described and their description is published in a central registry, this would also contribute in facilitating their discovery and their integration in other pipelines of analysis.

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**Fig. 1.** Co-expressed genes promoter analysis protocol description

