

Energetic mismatch induced by warming decreases leaf litter decomposition by aquatic detritivores

Theme08 - Introduction to Systems Biology
Reproducing a Research Article



Figure 1: Gammarus fossarum

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Abstract

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List of Tables

1 Introduction

1.1 Goal

1.2 Theory

2 Methods

2.1 The software model

2.2 Model configuration

RMR was calculated as the slope of oxygen depletion over a ca 35 min timeframe, corrected for the mean linear trend over time of oxygen concentration in controls as follows: $RMR = (ci - cf)/(ti - tf)$, where ci and cf are initial and final oxygen concentrations ($\mu\text{mol O}_2/\text{L}$) in the wells and ti and tf are initial and final times points (day-1) of the respiration experiment. Values were converted into C release rate ($\mu\text{g C}/\text{day}$) assuming a respiratory coefficient of 0.78 (as estimated for *Gammarus pulex*, Wright & Wright, 1979).

Statistical Analysis Equations

The following equations were used to express the mass (M in mg) and temperature (T in Kelvin) dependence of individual RMR and IR:

$$I = \alpha M^b e^{Ea \left(\frac{T-T_0}{k_B T_0 T} \right)} \quad (1a)$$

$$I = \alpha M^b e^{p \left(\frac{T-T_0}{k_B T_0 T} \right) - q \left(\frac{T-T_0}{k_B T_0 T} \right)^2} \quad (1b)$$

The standard MTE formulation (1a) is simply a particular case of the quadratic formulation (1b) where $q = 0$ and the equation is reduced to the MTE model where p can thus be interpreted as the activation energy.

Energetic efficiency was also calculated as follows: $E = (IR/RMR) * A_T$, where the ratio of IR to RMR is the ingestion to metabolism efficiency and A_T is the assimilation efficiency at temperature T . The temperature (T in Kelvin) dependence of assimilation efficiency was expressed, using empirical equations and values for detritivores Assimilation efficiency was following a logistic equation with the MTE equation both at the numerator and the denominator With the following formulation, assimilation efficiency is confined between 0 and 1 (no or complete assimilation):

$$A_T = \frac{\alpha e^{Ea \left(\frac{T-T_0}{k_B T_0 T} \right)}}{1 + \alpha e^{Ea \left(\frac{T-T_0}{k_B T_0 T} \right)}} \quad (2)$$

Consumer-Resource Dynamics Model

Below are the ordinary differential equations describing temporal change in leaf litter standing stocks (L) and *Gammarus* population biomass (G) (Equation 3a and 3b).

$$\frac{dL}{dt} = I - f(L)_T G - k_T L \quad (3a)$$

$$\frac{dG}{dt} = G [f(L)_T A - RMR_T] \quad (3b)$$

3 Results

4 Discussion and Conclusion

4.1 Discussion

- Basically unreproducible in general, long pieces of code that were repeated over and over that could not be understood. No easy way to reproduce other than to immediately copy and paste multiple hundreds lines of code where only few values would be changed.
- Formulas for metabolic and ingestion rates were heavily rewritten from the base formula. They were unrecognizable so they were cleaned up to be understandable and resemble the actual formula more. It should also be noted that the position where the mean in the quadratic portion of the exponent is actually different from what would be done following the formula.

4.2 General conclusion and perspective

5 References

- R Core Team. 2019. *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing. <https://www.R-project.org>.
- Réveillon, Tom, Thibaut Rota, Éric Chauvet, Antoine Lecerf, and Arnaud Sentis. 2022. “Energetic Mismatch Induced by Warming Decreases Leaf Litter Decomposition by Aquatic Detritivores.” *Journal of Animal Ecology* n/a (n/a). <https://doi.org/https://doi.org/10.1111/1365-2656.13710>.

6 Appendices

Appendix A: ‘model.R’

```
1  ## -----
2  ##
3  ## Script name: model.R
4  ##
5  ## Purpose of script: Implements the biological model of consumer-resource dynamics
6  ##
7  ## Author: Vincent Talen
8  ##
9  ## Date Created: 20 June 2022
10 ##
11 ## Email: v.k.talen@st.hanze.nl
12 ##
13 ## -----
14 ##
15 ## Notes:
16 ##   - Goal: formulate formula functions in a more readable/recognizable manner
17 ##
18 ## -----
19
20
21 #####
22 #   Libs   #
23 #####
24 library(deSolve)
25
26
27 #####
28 #   Code   #
29 #####
30 ## ---- loading in data ----
31 # Dataframe with respiration rate and ingestion rate ( $\mu\text{g C/hour}$ )
32 input_file <- "Data_Mismatch.txt"
33 data <- read.table(input_file, header = TRUE, dec = ".")
34 data <- na.omit(data)
35
36 # Consider individuals as a qualitative variable
37 data$Indiv <- as.factor(data$Indiv)
38
39 # Suppress outlier individuals and rows with negative values
40 data <- data[!data$Indiv %in% c("12", "30", "76", "78"), ]
41 data <- data[data$Respi > 0 & data$Nutri > 0, ]
42
43
44 ## ---- mte formulations ----
45 # Convert temperature into the Boltzmann term ( $^{\circ}\text{K}$ )
46 boltz_const <- 8.62 * 10-5
47 inverse_temps <- 1 / ((data$Temp + 273.15) * boltz_const)
48 mean_inverse_temp <- mean(inverse_temps)
49
```

```

50 # Quadratic function for metabolic rate (µg C/day)
51 calcMetabolicRate <- function(T.C, M) {
52   alpha <- exp(2.41599)      # metabolic expression level at reference temperature
53   b <- 0.62308              # body mass-scaling exponent
54   p <- 0.66731              # curve steepness (of the relationship)
55   q <- 0.21153              # quadratic term
56   T.K <- T.C + 273.15       # convert temperature from Celsius to Kelvin
57
58   # Repeating part with temperatures
59   temp_dependency_part <- (1 / (T.K * boltz_const)) - mean_inverse_temp
60
61   # Calculate metabolic rate with full formula
62   metabolic_rate <- alpha * M^b * exp(-p * temp_dependency_part) * exp(-q * temp_dependency_part^2)
63   return(metabolic_rate)
64 }
65
66 # Quadratic function for ingestion rate (µg C/day)
67 calcIngestionRate <- function(T.C, M) {
68   alpha <- exp(5.26814)      # ingestion expression level at reference temperature
69   b <- 0.81654              # body mass-scaling exponent
70   p <- 0.31876              # curve steepness (of the relationship)
71   q <- 0.18909              # quadratic term
72   T.K <- T.C + 273.15       # convert temperature from Celsius to Kelvin
73
74   # Repeating part with temperatures
75   temp_dependency_part <- (1 / (T.K * boltz_const)) - mean_inverse_temp
76
77   # Calculate ingestion rate with full formula
78   ingestion_rate <- alpha * M^b * exp(-p * temp_dependency_part) * exp(-q * temp_dependency_part^2)
79   return(ingestion_rate)
80 }
81
82
83 ## ---- assimilation efficiency ----
84 # Assimilation efficiency function based on exponential decay (quadratic model)
85 # Is a logistic equation with the MTE equation both at the numerator and the denominator
86 calcAssimEff <- function(temp) {
87   mte_equation <- (exp(-0.84730) * exp(0.16400 * ((temp + 273.15) - 285.65) / (boltz_const * 285.65 * (
88     return(mte_equation / (1 + mte_equation))
89   })
90
91
92 ## ---- attack rate parameter ----
93 leaf_mass <- 10.25 * 1000    # Mean initial leaf discs mass: 10.25 ± 0.68 mg
94 leaf_mass <- leaf_mass * 0.45 # Converted from mg to in µgC with a factor: 0.45
95
96 # Attack rate function based on exponential decay (quadratic model)
97 calcAttackRate <- function(temp, mass) {
98   expt_duration <- 2
99   decay_rate <- -log(1 - calcIngestionRate(temp, mass) * expt_duration / leaf_mass)
100   attack_rate <- decay_rate / expt_duration
101   return(attack_rate)
102 }

```

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```
## ---- handling time parameter ----
# Handling time function based on exponential decay (quadratic model)
calcHandlingTime <- function(temp, mass) {1 / (calcIngestionRate(temp, mass) / 1000) }

## ---- leaf decomposition- and respiration rate ----
# Reference temperature
ref_temp <- 10

# Function for the leaf litter microbial decomposition rate
calcLeafDecomp <- function(temp) { 0.00956 * exp(-0.37000 * (1 / ((temp + 273.15) * boltz_const) - ref_

# Function for the leaf litter respiration rate
calcLeafRespi <- function(temp) { 2.5 / 0.45 * 10^-3 * exp(-0.65000 * (1 / ((temp + 273.15) * boltz_con

## ---- consumer-resource model ----
GammLeafModel <- function(temp, gamm_indv_mass, leaf_fall, gamm_start_biomass) {
  Nutri <- function(time, state, parms) {
    with(as.list(c(state, parms)), {
      fL <- a * L / (1 + a * h * L)          # Holling type II functional response
      dL <- -fL * G - k * L                  # Biomass changes of leaf litter stock
      dG <- G * (fL * A - M)                 # Biomass changes of Gammarus population
      list(c(dL, dG))
    })
  }
}

# Leaf litter fall event function
leafFallEvent <- function(time, state, parms) {
  with(as.list(c(state, parms)), {
    return(c(L + leaf_fall, G))
  })
}

# Get time points to trigger litter fall event (first 15 days of the year)
getFallTimesYearX <- function(year) { seq(year * 365 + 1, year * 365 + 15) }
leaf_fall_times <- unlist(lapply(seq(0, 6), getFallTimesYearX))

# Model parameters
parameters <- c(
  M = calcMetabolicRate(temp, gamm_indv_mass) / 1000, # Gammarus metabolic rate (in mgC/day)
  a = calcAttackRate(temp, gamm_indv_mass),           # Gammarus attack rate (in mgC/day)
  h = calcHandlingTime(temp, gamm_indv_mass),         # Gammarus handling time (in 1/day)
  A = calcAssimEff(temp),                             # Gammarus assimilation efficiency
  k = calcLeafDecomp(temp),                           # Leaf microbial decomposition (in 1/day)
  Rl = calcLeafRespi(temp)                           # [UNUSED??] Leaf respiration (in mgC/mgleaf/

)

# Times and starting conditions
times <- seq(0, 365 * 7, by = 1)                      # Times in days for 7 years
state <- c(L = leaf_fall, G = gamm_start_biomass)      # Starting biomasses (in g/m2)

# Model output
```

```

157 out <- ode(time = times, func = Nutri, y = state, parms = parameters,
158           events = list(func = leafFallEvent, time = leaf_fall_times))
159
160 # Turn deSolve class object into dataframe and change very low and negative values to 0
161 data_table <- as.data.table(out) %>% mutate(across(c(L, G), ~ fifelse(.x < 10^-3, 0, .x)))
162 return(data_table)
163 }
164
165
166 ## ---- temperature-size rule models ----
167 # Average TSR response
168 calcTSR.Avg <- function(temp, mass) {
169   conv_fact <- 6.5 # Avg. conversion factor from dry to fresh mass
170   change_slope <- -3.90 - 0.53 * log10(mass) # Slope of change in mass per carbon
171   change_prop <- log(1 + change_slope / 100) # Proportion of change in mass per C
172   change_const <- exp(log(mass) - 12.5 * change_prop) # Constant of change in mass at 12.5°C
173
174   dry_mass <- change_const * exp(change_prop * (temp)) # Dry body mass (mg)
175   fresh_mass <- dry_mass / conv_fact # Fresh body mass (mg)
176   return(dry_mass)
177 }
178
179 # Maximum TSR response
180 calcTSR.Max <- function(temp, mass) {
181   conv_fact <- 6.5 # Avg. conversion factor from dry to fresh mass
182   change_slope <- -8.0 # Slope of change in mass per carbon
183   change_prop <- log(1 + change_slope / 100) # Proportion of change in mass per C
184   change_const <- exp(log(mass) - 12.5 * change_prop) # Constant of change in mass at 12.5°C
185
186   dry_mass <- change_const * exp(change_prop * (temp)) # Dry body mass (mg)
187   fresh_mass <- dry_mass / conv_fact # Fresh body mass (mg)
188   return(dry_mass)
189 }

```

Appendix B: ‘functions.R’

```

1  ## -----
2  ##
3  ## Script name: functions.R
4  ##
5  ## Purpose of script: Functions
6  ##
7  ## Author: Vincent Talen
8  ##
9  ## Date Created: 20 June 2022
10 ##
11 ## Email: v.k.talen@st.hanze.nl
12 ##
13 ## -----
14 ##
15 ## Notes:
16 ## - x

```

```

17 ##
18 ## -----
19
20
21 #####
22 # Libs #
23 #####
24 library(data.table)
25 library(ggpubr)
26 library(reshape2)
27 library(tidyverse)
28 source("r_code/model.R")
29
30
31 #####
32 # Functions #
33 #####
34 # ---- scenario data gathering and preparations ----
35 getScenarioDataList <- function(gamm_indv_mass, leaf_fall, gamm_start_biomass) {
36   # Get data for given values for each temperature using the model function that performs an ode
37   data_list <- lapply(temperatures, GammLeafModel, gamm_indv_mass, leaf_fall, gamm_start_biomass) %>%
38     setNames(temperatures)
39   return(data_list)
40 }
41
42 prepareBigDataFrame <- function(df_list) {
43   # Function to get the population metabolism for a temperature with the population biomass
44   getPopMetabolism <- function(cur_temp, gamm_pop_biomass) {
45     # Get metabolic rate for current temperature
46     meta_rate <- calcMetabolicRate(cur_temp, gamm_indv_mass) / 1000 # Gammarus metabolic rate (in mg
47     # Calculate population metabolism
48     pop_metabolism <- meta_rate * gamm_pop_biomass
49     return(fifelse(pop_metabolism < 0, 0, pop_metabolism))
50   }
51   # Function to get the population ingestion for a temperature with the population- and leaf biomasses
52   getPopIngestion <- function(cur_temp, leaf_biomass, gamm_pop_biomass) {
53     # Get ingestion- and attack rates for current temperature
54     ingest_rate <- calcIngestionRate(cur_temp, gamm_indv_mass) / 1000 # Gammarus ingestion rate (in mg
55     attack_rate <- calcAttackRate(cur_temp, gamm_indv_mass) # Gammarus attack rate (in mg C/
56     # Calculate population leaf ingestion
57     pop_ingestion <- (attack_rate * leaf_biomass / (1 + attack_rate * 1 / (ingest_rate / 1000) * leaf_b
58     return(fifelse(pop_ingestion < 0, 0, pop_ingestion))
59   }
60
61   # Bind all dataframes from list to single big one and
62   # drop last days to have 2555 days/rows left per temperature
63   big_df <- rbindlist(df_list)[!time == 2555] %>%
64     # Rename 'time' column to conform to naming scheme
65     setnames("time", "Time") %>%
66     # Add temperature and year columns to facilitate future calculations
67     "$<-"(Temp, rep(temperatures, each = 2555)) %>%
68     "$<-"(Year, rep(rep(1:7, each = 365), 5)) %>%
69     # Add population metabolism and leaf ingestion columns

```

```

70   "$<-"(M, getPopMetabolism(. $Temp, . $G)) %>%
71   "$<-"(I, getPopIngestion(. $Temp, . $L, . $G)) %>%
72   # Set column order to a nicer one
73   setcolorder(c("Time", "L", "G", "M", "I", "Temp", "Year"))
74   return(big_df)
75 }
76
77 # ---- plotting list of scenario dataframes ----
78 createPlotForTemp <- function(cur_temp, cur_data) {
79   # Divide L & G values to create a better readable plot
80   divided_data <- copy(cur_data)
81   set(divided_data, i = NULL, "L", divided_data$L / 10^5)
82   set(divided_data, i = NULL, "G", divided_data$G / 10^5)
83
84   # Create plot
85   plot <- ggplot(divided_data, aes(x = time, y = value)) +
86     # Set axis limits and step size
87     scale_x_continuous(breaks = seq(0, 7 * 365, 365)) +
88     ylim(NA, ceiling(max(divided_data[, -1])) + 1) +
89     # Add the data (lines)
90     geom_line(aes(y = L, color = "Leaf Litter Biomass")) +
91     geom_line(aes(y = G, color = "Gammarus Fossarum Biomass")) +
92     # Add styling
93     labs(title = sprintf("%s°C", cur_temp), x = "", y = "") +
94     # theme(plot.title = element_text(hjust = 0.075, vjust = -11)) +
95     # theme(plot.margin = margin(0.1, 0.25, 0, 0, "cm")) +
96     scale_color_manual(name = "", values = c("black", "tomato2"),
97                       limits = c("Leaf Litter Biomass", "Gammarus Fossarum Biomass"))
98   return(plot)
99 }
100
101 plotScenario <- function(data, image_title, file_out) {
102   # Use lapply to create plots for each temperature in the list and collect the legend from a plot
103   plot_list <- lapply(seq_along(data), function(i) { createPlotForTemp(names(data)[i], data[[i]]) })
104   plot_legend <- get_legend(plot_list[[1]])
105
106   # Remove legends from the plots and add extracted legend to end of the list
107   plot_list <- lapply(plot_list, function(cur_plot) { cur_plot + theme(legend.position = "none") }) %>%
108     "[<-"(length(plot_list) + 1, plot_legend)
109
110   # Place plots and legend in an arranged grid
111   col_num <- 3
112   my_grid <- ggarrange(plotlist = plot_list, ncol = col_num, nrow = ceiling(length(plot_list) / col_num),
113     annotate_figure(top = text_grob(image_title), bottom = text_grob("Time (d)",
114       left = text_grob(bquote("Biomass "(10^5~ mg~ C~ m^-2)), rot = 90))
115
116   # Save the created arranged grid with the lossless 'lzw' compression that greatly reduces file size
117   ggsave(file_out, bg = "white", width=15, height=8, units="in", dpi=300, compression = "lzw")
118   dev.off()
119 }

```


Appendix C: ‘scenarios.R’

```
1  ## -----
2  ##
3  ## Script name: scenarios.R
4  ##
5  ## Purpose of script: Simulate scenarios using functions from the functions.R module
6  ##
7  ## Author: Vincent Talen
8  ##
9  ## Date Created: 20 June 2022
10 ##
11 ## Email: v.k.talen@st.hanze.nl
12 ##
13 ## -----
14 ##
15 ## Notes:
16 ##   - x
17 ##
18 ## -----
19 #setwd("BIN2-P8/endreport")
20
21
22 #####
23 #   Libs   #
24 #####
25 source("r_code/functions.R")
26
27
28 #####
29 #   Code   #
30 #####
31 # Temperatures to do simulations of
32 temperatures <- c(5, 10, 15, 20, 25)
33
34 #####
35 ### SCENARIO 0: STANDARD SCENARIO ###
36 #####
37 # Gammarus mean body mass = 4.26 mgDM
38 # Annual leaf fall        = 300 gC/m2/an = 300 000 mgC/m2/an
39 # Gammarus density        = 30 mgDM/m2   = 15 mgC/m2
40
41 # Duration of the leaf fall in days
42 fall_duration_in_days <- 15
43
44 # Define scenario values
45 gamm_indv_mass <- 4.26
46 leaf_fall <- 300000 / fall_duration_in_days
47 gamm_start_biomass <- 15
48
49 # Get data for temperatures with values of current scenario
50 scen_df_list <- getScenarioDataList(gamm_indv_mass, leaf_fall, gamm_start_biomass)
51
```

```

52 # Create plots in an arranged grid
53 file_out <- "figures/Population Dynamics Standard Scenario.tiff"
54 image_title <- "Standard Scenario: Population Dynamics over 7 years"
55 plotScenario(scen_df_list, image_title, file_out)
56
57 # Combine dataframes into one and add temperature, year, population metabolism- and ingestion columns
58 TestSD <- prepareBigDataFrame(scen_df_list)
59
60 #####
61 CutSD <- as.data.frame(setDT(TestSD)[TestSD[, tail(.I, -16), by = Year]$V1])
62 CutSD <- CutSD[!CutSD$Year == "1", ]
63
64 #####
65 ### Mean annual biomasses ###
66 #####
67 # Calculate litter and Gammarus mean annual biomasses
68 MeanL=as.data.frame(setDT(CutSD)[, .(MeanL=mean(L)), by=list(Temp,Year)])
69 MeanG=as.data.frame(setDT(CutSD)[, .(MeanG=mean(G)), by=list(Temp,Year)])
70
71 # Calculate means and deviations over 6 years for annual biomasses
72 MeanLSD=as.data.frame(setDT(MeanL)[, .(MeanL=mean(MeanL), SdL=sd(MeanL)), by=list(Temp)])
73 MeanGSD=as.data.frame(setDT(MeanG)[, .(MeanG=mean(MeanG), SdG=sd(MeanG)), by=list(Temp)])
74
75 #####
76 ### Mean annual persistence times above biomass thresholds ###
77 #####
78 # Calculate litter and Gammarus mean annual persistence times
79 ThLSD=CutSD[CutSD$L>60000,]
80 ThGSD=CutSD[CutSD$G>5000,]
81
82 TimeLSD=as.data.frame(setDT(ThLSD)[, .(TimeL=length(Time)), by=list(Temp,Year)])
83 TimeGSD=as.data.frame(setDT(ThGSD)[, .(TimeG=length(Time)), by=list(Temp,Year)])
84
85 # Calculate means and deviations over 6 years for the thresholds
86 TimeLSD=as.data.frame(setDT(TimeLSD)[, .(MeanTimeL=mean(TimeL), SdTimeL=sd(TimeL)), by=list(Temp)])
87 TimeGSD=as.data.frame(setDT(TimeGSD)[, .(MeanTimeG=mean(TimeG), SdTimeG=sd(TimeG)), by=list(Temp)])
88
89 #####
90 ### Mean biomass maximums and biomass decreases ###
91 #####
92 # Find biomass maximums and minimums
93 CycleLSD=as.data.frame(setDT(TestSD)[, .(MaxLSD=findPeaks(L), MinLSD=findValleys(L)[seq(2,14,2)], by=1)]
94 CycleGSD=as.data.frame(setDT(TestSD)[, .(MaxGSD=findPeaks(G), MinGSD=c(findValleys(G),2555)), by=list(T

```