```
In [3]: import numpy as np
        import matplotlib
        import skimage
        import IPython
        import imagecodecs #(New 2025)
        import matplotlib.pyplot as plt
        from IPython.display import HTML
        from matplotlib import animation, rc
        from skimage.color import rgb2gray
        from skimage import data
        from skimage.filters import gaussian
        from skimage.segmentation import active_contour # For active_contour functid
        from skimage.util import random_noise
        # For active_contour function
        from skimage.segmentation import chan_vese, morphological_chan_vese, checker
        # For some image filtering
        from skimage.morphology import white_tophat, black_tophat, disk
        import skimage.io
        from IPython.core.interactiveshell import InteractiveShell
        InteractiveShell.ast_node_interactivity = "all"
        # PRINT VERSIONS
        print("np.__version__",np.__version__)
        print("matplotlib.__version__", matplotlib.__version__)
        print("skimage.__version__",skimage.__version__)
        print("IPython.__version__",IPython.__version__)
        print("imagecodecs.__version__",imagecodecs.__version__)
       np.__version__ 2.2.1
       matplotlib.__version__ 3.10.0
       skimage.__version__ 0.25.0
       IPython.__version__ 8.31.0
       imagecodecs.__version__ 2024.12.30
In [4]: def edge_map(img,sigma):
            blur = skimage.filters.gaussian(img,sigma)
            return skimage.filters.sobel(blur)
        def edge_map2(img,sigma):
            blur = skimage.filters.gaussian(img,sigma)
            return skimage.filters.scharr(blur)
        def subtract background(image, radius=5, light bg=False):
                str_el = disk(radius)
                if light_bg:
                    return black_tophat(image, str_el)
                    return white_tophat(image, str_el)
```

```
def define initial circle(R0,r0,c0,Nber pts=400):
    # Define initial contour shape
           = np.linspace(0, 2*np.pi, Nber_pts)
    Radius = R0
         = r0 + Radius*np.sin(s)
           = c0 + Radius*np.cos(s) #col
    init = np.array([r, c]).T
    return init
## Create slides for animation
def animate_cv(image, segs, interval=1000):
    fig, ax = plt.subplots(figsize=(8, 8))
    ax.imshow(image, cmap='gray');
    im = ax.imshow(segs[0], alpha=0.5, cmap='inferno');
    ax.axis('off')
    def init():
        im.set_data(segs[0])
        return [im]
    def animate(i):
        im.set_array(segs[i])
        return [im]
    anim = animation.FuncAnimation(fig, animate, init_func=init,
                                    frames=len(segs), interval=1000, blit=Tru
    return anim
def animate_snake(image, segs, interval=500):
    fig, ax = plt.subplots(figsize=(6, 6))
    ax.imshow(image, cmap='gray');
         im = ax.imshow(segs[0], alpha=0.5, cmap='inferno');
    #ax.plot(segs[0][:, 1], segs[0][:, 0], '--r', lw=3)
    ax.axis('off')
    line, = ax.plot([], [], '-r', lw=2)
    def init():
        line.set_data(segs[0,:,1],segs[0,:,0])
        return [line,]
    def animate(i):
        line.set_data(segs[i,:,1],segs[i,:,0])
        return [line,]
    anim = animation.FuncAnimation(fig, animate, init_func=init,
                                    frames=len(segs), interval=1000, blit=Tru
    return anim
###################################
def store evolution in(lst):
```

```
"""Returns a callback function to store the evolution of the level sets
the given list.
"""

def _store(x):
    lst.append(np.copy(x))

return _store
```

# Read images

This part reads a series of images that you can then use in various tests.

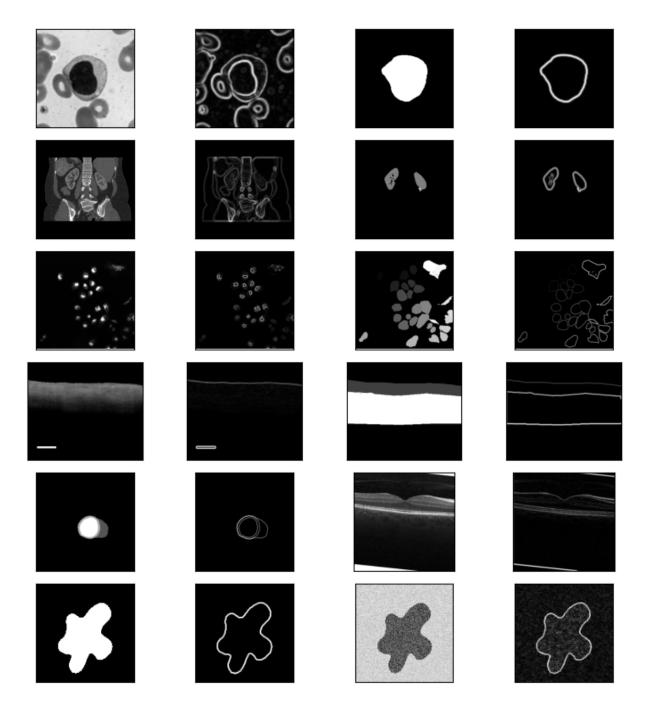
Note that some images are provided with ground-truth masks of structures of interest:

- 1. OCT\_tissue
- 2. CTabd (CT of the abdomen)
- 3. US nodule (Ultrasound image with a nodule)
- 4. images\_blood\_cells/000016.png [several images available]

```
In [5]: # import warnings
        # warnings.filterwarnings( "ignore", module = "matplotlib\..*" )
        # skimage.io.imshow(img_mask)
        # Binary images - w/o ground truth
                  = skimage.io.imread('./images_misc/smooth_star.png', as_gr
        img_star
        edge_star = edge_map(img_star, sigma=0)
        img_star_noisy = skimage.io.imread('./images_misc/smooth_star_noisy.png',
        edge_star_noisy = edge_map(img_star_noisy, sigma=0)
        img_binshape = skimage.io.imread('./images_misc/binary_shape_2024.png',
        edge_binshape = edge_map(img_binshape, sigma=0)
        img_cardiacshape = skimage.io.imread('./images_misc/cardiac_mri_mask.png',
        edge_cardiacshape = edge_map(img_cardiacshape, sigma=0)
        # Microscopy images - w/o ground truth # line changed 2025
        img_hela
                         = skimage.io.imread('./images_misc/hela_big.png')
        # OCT eye images - w/o ground truth
        img_oct_eye = skimage.io.imread('./images_misc/OCT_normal.jpeg', as_gray =
        img_oct_eye = np.squeeze(img_oct_eye)
        img_oct_eye = img_oct_eye.astype('float64')
        img_oct_eye = img_oct_eye/np.max(img_oct_eye)
        edge_oct_eye = edge_map(img_oct_eye, sigma=2)
        # CT abdo images - with ground truth
        img_CTabd = skimage.io.imread('./images_misc/CT_kidney_im.png', as_gray
        edge_CTabd = edge_map(img_CTabd, sigma=2)
                  = skimage.io.imread('./images_misc/CT_kidney_mask.png', as_gra
        gt_CTabd
        edge_gt_CTabd = edge_map(gt_CTabd, sigma=2)
```

```
# Cell images — with ground truth
img_cell = skimage.io.imread('./images_blood_cells/0000152.png', as_gray =
edge cell = edge map(img cell, sigma=2)
#skimage.io.imshow(img cell)
gt_cell = skimage.io.imread('./masks_blood_cells/0000152.png', as_gray = Tru
edge_gt_cell = edge_map(gt_cell, sigma=2)
# Cell image - challenge multi
             = skimage.io.imread('./images misc/cell 00236.tif', as gray =
img cell2
edge_cell2
             = edge_map(img_cell2, sigma=2)
gt_cell2
             = skimage.io.imread('./images_misc/cell_00236_label.tiff', as_
edge_gt_cell2 = edge_map(gt_cell2, sigma=2)
# Hela Cell image - Cell tracking challenge
img helat1
              = skimage.io.imread('./images misc/hela t001.tif', as gray =
              = edge map(img helat1, sigma=2)
edge helat1
              = skimage.io.imread('./images_misc/hela_mask001.tif', as_gray
gt_helat1
edge_gt_helat1 = edge_map(gt_helat1, sigma=2)
# Fluo Cell image - Cell tracking challenge
            = skimage.io.imread('./images_misc/fluo000.tif', as_gray = True
img_fluo
edge fluo
             = edge map(img fluo, sigma=2)
             = skimage.io.imread('./images_misc/fluo000_seg.tif', as_gray =
gt_fluo
edge_gt_fluo = edge_map(gt_fluo, sigma=2)
# OCT image of tissue - with ground truth
img oct tissue = skimage.io.imread('./OCT myocardium/case272.tif', as gray
edge_oct_tissue = edge_map(img_oct_tissue, sigma=2)
gt_oct_tissue = skimage.io.imread('./OCT_myocardium/case272_label.tiff', as
edge qt oct tissue = edge map(qt oct tissue, sigma=2)
# US image of a nodule - with ground truth
img USnodule = skimage.io.imread('./thyroid nodule/1074.png', as gray = Tru
edge_USnodule = edge_map(img_USnodule, sigma=2)
gt_USnodule = skimage.io.imread('./thyroid_nodule/1074_mask.png', as_gray
edge gt USnodule = edge map(gt USnodule, sigma=2)
# PLOTS
fig, axes = plt.subplots(6,4, figsize=(8, 8))
ax = axes.ravel()
ax[0].imshow(img_cell, cmap=plt.cm.gray);
ax[1].imshow(edge_cell, cmap=plt.cm.gray);
ax[2].imshow(gt_cell, cmap=plt.cm.gray);
ax[3].imshow(edge_gt_cell, cmap=plt.cm.gray);
ax[4].imshow(img_CTabd, cmap=plt.cm.gray);
ax[5].imshow(edge CTabd, cmap=plt.cm.gray);
ax[6].imshow(gt_CTabd, cmap=plt.cm.gray);
ax[7].imshow(edge_gt_CTabd, cmap=plt.cm.gray);
ax[8].imshow(img_fluo, cmap=plt.cm.gray);
ax[9].imshow(edge_fluo, cmap=plt.cm.gray);
ax[10].imshow(gt fluo, cmap=plt.cm.gray);
```

```
ax[11].imshow(edge_gt_fluo, cmap=plt.cm.gray);
ax[12].imshow(img_oct_tissue, cmap=plt.cm.gray);
ax[13].imshow(edge_oct_tissue, cmap=plt.cm.gray);
ax[14].imshow(gt_oct_tissue, cmap=plt.cm.gray);
ax[15].imshow(edge_gt_oct_tissue, cmap=plt.cm.gray);
ax[16].imshow(img_cardiacshape, cmap=plt.cm.gray);
ax[17].imshow(edge_cardiacshape, cmap=plt.cm.gray);
ax[18].imshow(img_oct_eye, cmap=plt.cm.gray);
ax[19].imshow(edge_oct_eye, cmap=plt.cm.gray);
ax[20].imshow(img_star, cmap=plt.cm.gray);
ax[21].imshow(edge_star, cmap=plt.cm.gray);
ax[22].imshow(img_star_noisy, cmap=plt.cm.gray);
ax[23].imshow(edge_star_noisy, cmap=plt.cm.gray);
for i in range(0,24):
    ax[i].set_xticks([]), ax[i].set_yticks([]);
fig.tight_layout()
plt.show();
```



# Image properties:

## Range of values and data type matter ...

Some routines won't work if your image type is int8 or uint8... Here is how to check your image data type

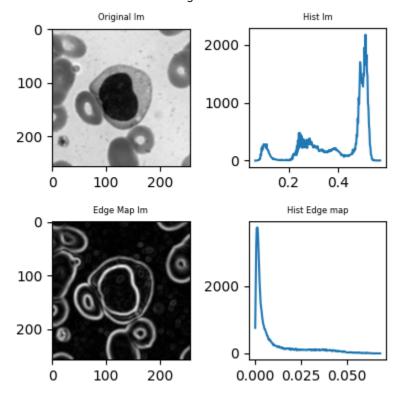
And regularly check your image content in terms of:

- intensities range of values
- distributions of intensities via its histogram

```
imq test = imq cell #imq star noisy#imq oct eye #imq CTabd #img cell
In [6]:
        Sigma val = 2
         edge_test = edge_map(img_test, sigma=Sigma_val)
        ## Print some basic image properties
        print("Image size of img_test is: ", img_test.shape)
        print("Data type of img_test is: ", img_test.dtype)
print("min - max value in image:" , np.min(img_test), np.max(img_test))
        ## Hot to plot a Histogram
        hist test, bins test
                                            = np.histogram(img test.flatten(), bins=256
        hist_edge_test, bins_edges_test = np.histogram(edge_test.flatten(), bins=25
        fig, axes = plt.subplots(2,2, figsize=(4, 4))
                   = axes.ravel()
        ax[0].imshow(img_test, cmap=plt.cm.gray);
        ax[0].set title("Original Im", fontsize=6);
        ax[1].plot(bins_test[0:-1],hist_test);
        ax[1].set_title("Hist Im", fontsize=6);
        ax[2].imshow(edge_test, cmap=plt.cm.gray);
        ax[2].set_title("Edge Map Im", fontsize=6);
        ax[3].plot(bins_edges_test[0:-1],hist_edge_test);
        ax[3].set title("Hist Edge map", fontsize=6);
        fig.tight_layout()
        plt.show();
```

Image size of img\_test is: (256, 256)
Data type of img\_test is: float64

min - max value in image: 0.06550980392156862 0.5724509803921568



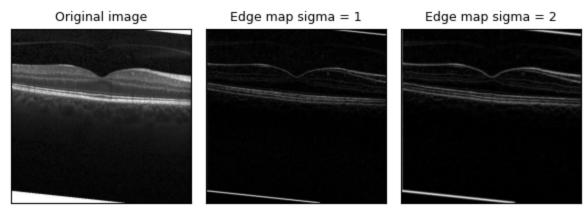
## Edge maps

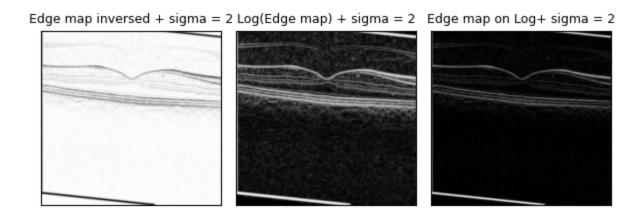
Deformable models rely on edge maps. Most routines have their own strategy coded to compute the edge map.

- Edge maps usually involve smoothing of the image, to be robust to noise. Make sure you understand how this is controlled in the routine you use.
- Edge maps usually show pixels with high gradient magnitudes in white (high values)
- Most deformable model routines can be fed directly with an Edge Map rather than the original image as its input
- Some routine expect to be fed with an inverse edge map where high gradient locations have small values, to stop the contour via a velocity set to ~zero.

```
In [7]:
        img_to_test = img_oct_eye
        print("Data type of img_test is: ", img_test.dtype)
        print("min - max value in image:" , np.min(img_test), np.max(img_test))
        Font_size = 9
        # Classic Edge map with Gaussian smoothing controled by sigma
        edge_test1 = edge_map(img_to_test, sigma=1)
edge_test2 = edge_map(img_to_test, sigma=2)
edge_test2_l = np.log2((edge_test2*100)+1)
        edge_testl_2 = edge_map(np.log2((img_to_test+1)*100), sigma=2)
        # Inversed Edge map
        # Returns Edge map = 1.0 / np.sqrt(1.0 + alpha * gradnorm)
        edge_inv_test = skimage.segmentation.inverse_gaussian_gradient(img_to_test,
         fig, axes = plt.subplots(2,3, figsize=(6, 6))
         ax = axes.ravel()
        ax[0].imshow(img_to_test, cmap=plt.cm.gray);
         ax[0].set_title("Original image", fontsize=Font_size);
        ax[1].imshow(edge_test1, cmap=plt.cm.gray);
         ax[1].set_title("Edge map sigma = 1", fontsize=Font_size);
        ax[2].imshow(edge_test2, cmap=plt.cm.gray);
        ax[2].set_title("Edge map sigma = 2", fontsize=Font_size);
        ax[3].imshow(edge_inv_test, cmap=plt.cm.gray);
        ax[3].set_title("Edge map inversed + sigma = 2", fontsize=Font_size);
         ax[4].imshow(edge_test2_l, cmap=plt.cm.gray);
        ax[4].set_title("Log(Edge map) + sigma = 2", fontsize=Font_size);
         ax[5].imshow(edge_testl_2, cmap=plt.cm.gray);
        ax[5].set_title("Edge map on Log+ sigma = 2", fontsize=Font_size);
        for i in range(0,6):
             ax[i].set_xticks([]), ax[i].set_yticks([]);
         fig.tight layout()
         plt.show();
```

Data type of img\_test is: float64 min - max value in image: 0.06550980392156862 0.5724509803921568





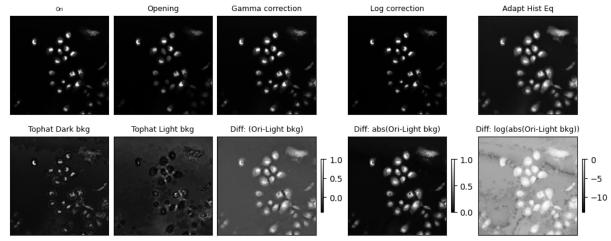
# Image transforms

Let you test some image transformations based on morphological operators and histogram manipulation. When transforming image contrast, it is always interesting to look at the differences between the original image and the transformed version.

```
In [8]:
       img_ori_to_test = img_fluo #img_CTabd #img_oct_eye #img_CTabd
        img_to_test = img_ori_to_test
        epsilon
                      = 0.000001 #to prevent log on 0
        img_eps
                      = np.full_like(img_to_test, epsilon)
        PRE_ENHANCE = 1
        OPTION_ENHANCE = 4 # can be 0 (nothing) OR 1,2,3,4 for different enchanceme
        Font_size = 9
        # Run all OPTION_ENHANCE for display here
        gamma_corrected = skimage.exposure.adjust_gamma(img_to_test, 0.8)
        logarithmic_corrected = skimage.exposure.adjust_log(img_to_test, gain= 1,inv
                     = skimage.morphology.diameter_opening(img_to_test, 40,
        img_open
                             = skimage.exposure.equalize_adapthist(img_to_test, cli
        img_adapteq
        # PRE ENHANCEMENT OPTIONS:
        if PRE ENHANCE==1:
```

```
if OPTION ENHANCE==1:
        # Gamma
        img to test
                       = gamma corrected
    elif OPTION ENHANCE==2:
        # Logarithmic (0 = gain*log(1 + I)) or if Inv(0 = gain*(2**I - 1))
        img to test
                             = logarithmic corrected
    elif OPTION_ENHANCE==3:
        # Morpho Opening
        img to test
                             = img open
   elif OPTION ENHANCE==4:
        # Contrast Limited Adaptive Histogram Equalization (CLAHE).
        img_to_test
                      = img adapteg
# Enhance details either dark around light background of vice versa with the
Radius val = 15
img_test1 = subtract_background(img_to_test, radius=Radius_val, light_bg=Fa
img_test2 = subtract_background(img_to_test, radius=Radius_val, light_bg=Tr
# SHOW OUTPUTS
fig, axes = plt.subplots(2,5, figsize=(10, 4),constrained_layout=True)
         = axes.ravel()
Shrink factor colormap = 0.5
ax[0].imshow(img_ori_to_test, cmap=plt.cm.gray);
ax[0].set_title("Ori", fontsize=6);
ax[1].imshow(img open, cmap=plt.cm.gray);
ax[1].set_title("Opening", fontsize=Font_size);
ax[2].imshow(gamma_corrected, cmap=plt.cm.gray);
ax[2].set_title("Gamma correction", fontsize=Font_size);
ax[3].imshow(logarithmic_corrected, cmap=plt.cm.gray);
ax[3].set_title("Log correction", fontsize=Font_size);
ax[4].imshow(img_adapteq, cmap=plt.cm.gray);
ax[4].set_title("Adapt Hist Eq", fontsize=Font_size);
ax[5].imshow(img_test1, cmap=plt.cm.gray);
ax[5].set_title("Tophat Dark bkg", fontsize=Font_size);
ax[6].imshow(img_test2, cmap=plt.cm.gray);
ax[6].set_title("Tophat Light bkg", fontsize=Font_size);
tmp_show = ax[7].imshow(img_to_test-img_test2, cmap=plt.cm.gray);
ax[7].set_title("Diff: (Ori-Light bkg)", fontsize=Font_size);
plt.colorbar(tmp_show,ax=ax[7], shrink=Shrink_factor_colormap, location='rig
tmp show = ax[8].imshow(abs(imq to test-imq test2), cmap=plt.cm.gray);
ax[8].set_title("Diff: abs(Ori-Light bkg)", fontsize=Font_size);
plt.colorbar(tmp_show,ax=ax[8], shrink=Shrink_factor_colormap, location='rig
tmp\_show = ax[9].imshow(np.log2(abs(img\_to\_test-img\_test2+img\_eps)), cmap=pl
ax[9].set_title("Diff: log(abs(Ori-Light bkg))", fontsize=Font_size);
plt.colorbar(tmp show,ax=ax[9], shrink=Shrink factor colormap, location='rig
for i in range(0,10):
    ax[i].set_xticks([]), ax[i].set_yticks([]);
#fig.tight_layout() # not compatible with option constrained_layout=True in
plt.show();
```

/var/folders/81/3v\_d9rcn7wd1szht19l1v89c0000gn/T/ipykernel\_48152/293332637
7.py:64: RuntimeWarning: divide by zero encountered in log2
 tmp\_show = ax[9].imshow(np.log2(abs(img\_to\_test-img\_test2+img\_eps)), cmap=
plt.cm.gray);



## Seg #1:

#### Snake on a binary shape + noise effects

This part of the practical work uses the routine **active\_contour** from skimage. Default **parameter values** are:

- alpha=0.01 (Snake length shape parameter. Higher values makes snake contract faster.)
- beta=0.1 (Snake smoothness shape parameter. Higher values makes snake smoother.)
- gamma=0.01 (Explicit time stepping parameter Equivalent to the viscosity of the environment)
- max\_px\_move=1.0

There are two **other parameters** that define the final image information used to define external forces used to define regions.img =  $w_line x img + w_edge x edge$ :

- w\_line\_val= 0 (default) | =1 if want to input\_edge map directly. Use negative values to attract toward dark
- w\_edge\_val= 1 (default) | = 0 if do not want to use internal edge map. Use negative values to repel snake from edges

## TODO:

#### Provide answers in text boxes

Q1.1. Run the code for img\_to\_seg=img\_cardiacshape using img\_ori and all

parameter values as provided, except for changing the R0 value. Comment on behavior for:

- R0=10:
- R0=20:
- R0=30:
- R0=50:

**Q1.2**. For R0=30 test the segmentation without smoothing and then with Niter\_smooth = 1 and 2. Comment on the segmentation quality for:

- · no smoothing:
- Niter\_smooth = 1:
- Niter\_smooth = 2:
- **Q1.3**. Now run the segmentation on the **noisy version** or the image. 2 types of noise are simulated: (1) Additive Gaussian noise, (2) Speckle (multiplicative) noise.
- **Q1.3.1** Check **appearance** of the 2 noisy images. Why is there no noise in the background in the speckle case?

#### Answer:

Q1.3.2 Using R0=50, run the segmentation on the noisy images without and with smoothing (Niter\_smooth= 1). Comment on segmentation quality or issues for the 4 observations:

- Speckle noise + no smoothing:
- Speckle noise + smoothing:
- Gaussian noise + no smoothing:
- Gaussian noise + smoothing:

```
# Pre-smooth the image
Niter_smooth = 1 # set to 0 for no smoothing
                = gaussian(img_to_seg, Niter_smooth, preserve_range=False)
img_to_seg
# Run active contour
snake1 = active_contour(img_to_seg,
                       init, max_num_iter=1, convergence=convergence_val,
                         alpha=alpha_val, beta=beta_val, gamma=gamma_val)
snake10 = active contour(img to seg,
                       init, max_num_iter=10, convergence=convergence_val,
                         alpha=alpha_val, beta=beta_val, gamma=gamma_val)
snake_max = active_contour(img_to_seg,
                       init, max_num_iter=Niter_snake, convergence=convergen
                           alpha=alpha_val, beta=beta_val, gamma=gamma_val)
# Display results
fig, axes = plt.subplots(1,2, figsize=(8, 4),constrained_layout=True)
          = axes.ravel()
ax
Font size = 9
ax[0].imshow(img_to_seg_raw, cmap=plt.cm.gray);
ax[0].set_xticks([]), ax[0].set_yticks([]);
ax[0].set_title("Image to segment", fontsize=Font_size);
ax[1].imshow(img_to_seg, cmap=plt.cm.gray);
ax[1].plot(init[:, 1], init[:, 0], '--y', lw=1);
ax[1].plot(snake10[:, 1], snake1[:, 0], '-b', lw=1);
ax[1].plot(snake10[:, 1], snake10[:, 0], '-g', lw=1);
ax[1].plot(snake_max[:, 1], snake_max[:, 0], '-r', lw=2);
ax[1].set_xticks([]), ax[1].set_yticks([]);
ax[1].axis([0, img_to_seg.shape[1], img_to_seg.shape[0], 0]);
ax[1].set_title("Smoothed image + Seg", fontsize=Font_size);
plt.show();
```

## Seg #2:

#### Snake on Cell image

We are using here **img\_to\_seg = img\_cell** for which you have a ground truth mask **gt\_cell** of the target segmentation for the bright right cell.

### **TODO**

**Q2.1 Segment large right cell**: Run with code as provided and check quality of the segmentation versus the ground-truth. Did it work?

Answer:

Q2.2 Now aiming to segment the internal dark part of the cell: change only values for

Niter\_smooth and R0 and propose a solution that works.

**Answer**: managed to obtain a correct segmentation with Niter\_smooth = XX and R0 = XX

Q2.3 Segment small left cell: Run with the proposed initialisation and check correct segmentation of the whole left cell. Now change ONLY some initial contour parameter(s) [r0 = 153; c0=66; R0 = 25] to obtain a perfect segmentation of the internal bright center of the cell.

**Answer**: managed to obtain a correct segmentation with [r0 = XX; c0=XX; R0 = XX]

```
In []: # Input image and parameter values
        img_to_seg = img_cell;
        img_gt
                   = gt_cell;
        # Large rigt cell - ground truth provided
        r0 = 128; c0=128; R0 = 53
        # Small left cell - no ground truth
        \#r0 = 153; c0=66; R0 = 25
        alpha val = 0.01; beta val = 0.1; gamma val = 0.01;
        convergence_val = 1e-4; Niter_snake = 1200;
        # Pre smooth the image
        Niter smooth = 1
        img_to_seg = gaussian(img_to_seg, Niter_smooth, preserve_range=False)
        # Initialise contour
        init = define_initial_circle(R0,r0,c0)
        # Run active contour
        snake30 = active_contour(img_to_seg,
                               init, max_num_iter=30, convergence=convergence_val,
                                 alpha=alpha_val, beta=beta_val, gamma=gamma_val)
        snake = active_contour(img_to_seg,
                               init, max_num_iter=Niter_snake, convergence=convergen
                               alpha=alpha_val, beta=beta_val, gamma=gamma_val)
        # Display results
        fig, axes = plt.subplots(1,2, figsize=(8, 4),constrained_layout=True)
             = axes.ravel()
        Font_size = 9
        ax[0].imshow(img_to_seg, cmap=plt.cm.gray)
        ax[0].plot(init[:, 1], init[:, 0], '--y', lw=1)
        ax[0].plot(snake30[:, 1], snake30[:, 0], '-b', lw=1.5)
        ax[0].plot(snake[:, 1], snake[:, 0], '-r', lw=2)
        ax[0].set_xticks([]), ax[0].set_yticks([])
        ax[0].axis([0, img_to_seg.shape[1], img_to_seg.shape[0], 0])
```

```
ax[1].imshow(img_gt, cmap=plt.cm.gray)
ax[1].plot(init[:, 1], init[:, 0], '--y', lw=1)
ax[1].plot(snake30[:, 1], snake30[:, 0], '-b', lw=1.5)
ax[1].plot(snake[:, 1], snake[:, 0], '-r', lw=2)
ax[1].set_xticks([]), ax[1].set_yticks([])
ax[1].axis([0, img_to_seg.shape[1], img_to_seg.shape[0], 0])
plt.show();
```

## Seg # 3

A tool to visualise the deformations of the snake over iterations

#### TO DO:

Run the code with provided parameter values.

**Q3.1** Checking the video, would you confirm that the snake has converged and is stable? **Answer**:

Q3.2 Change R0 to R0 = 52. Has convergence time been shorter or longer? Did you expect such observation (yes/no)?

Answer: convergence time shorter/longer. It was (not?) expected since XX...

```
In [ ]: |img_to_seg
                          = img_cell
        img_to_seg_ori = img_to_seg
        # Init to segment cell
        r0 = 128; c0=128; R0 = 53
        alpha_val = 0.01 ; beta_val = 0.1 ;gamma_val = 0.01;
        convergence val = 1e-4; Niter snake = 800;
        # Pre filter the image
        img_adapteq = skimage.exposure.equalize_adapthist(img_to_seg, clip_limit=0.0
        #img_to_seg = img_adapteq
        # Pre smooth the image
        Niter smooth = 1
                    = gaussian(img_to_seg, Niter_smooth, preserve_range=False)
        img_to_seg
        # Initialise contour
        init = define_initial_circle(R0,r0,c0)
        # Run active contour while saving intermediate contours to see deformations
        segs = []
```

## Seg # 4

### Snake with Gradient Vector Flow (GVF)

This implementation of the GVF is performed by computing the edge map, diffusing the gradient over the whole image and directly input the GVF\_edge\_map to be used as external forces by setting w\_line=1 and w\_edge=0 in the active\_contour function.

### TODO:

**Q4.1.1** Report the visual differences in the GVF\_map between mu=5 and mu=15.

Answer:

**Q4.1.2** What is the mu parameter controling?

**Answer**: The mu parameter controls for XX.

Q4.2 Why does mu=15 enable to obtain a correct segmentation?

Answer:

**Q4.3** Report what happens when segmenting with the classic Edge\_map rather than the GVF\_map.

Answer:

```
In [ ]: import gvf_elsa2
from gvf_elsa2 import gradient_field, gradient_vector_flow

# Image to seg
img_to_seg = img_star
```

```
r0 = 64; c0=64; R0 = 50
alpha val = 0.01; beta val = 0.1; gamma val = 0.01;
convergence_val = 1e-4; Niter_snake = 200;
# Initialise contour
init
            = define_initial_circle(R0, r0, c0, Nber_pts=400)
# Compute edge map and gvf
img_to_seg = img_to_seg.astype(np.float32) / np.max(img_to_seg)
Edge_map = edge_map(img_to_seg,sigma=1)
fx, fy = gradient_field(img_to_seg)
gx, gy = gradient_vector_flow(fx, fy, mu=5)
GVF_map = np.sqrt(gx**2 + gy**2)
# Run active contour while saving intermediate contours to see deformations
Map_to_seg = Edge_map
# Run active contour while saving intermediate contours to see deformations
seqs = []
print('start')
for i in range(1,Niter_snake,10):
    print(i, " ", end='')
    segs.append(active_contour(Map_to_seg, init, max_num_iter=i, convergence
                alpha=alpha_val, beta=beta_val, gamma=gamma_val,
                    w_line=1,w_edge=0))
print('stop')
np.save('ANIM_contours.npy', np.array(segs))
# display animation
segs = np.load('ANIM_contours.npy')
anim = animate_snake(Map_to_seg, segs);
HTML(anim.to_html5_video())
```

## Seg # 5:

## The active contour with fixed end points

You will now run the active\_contour with the option to maintain some points from the inital contour fixed. You are working with the **img\_oct\_eye** which shows different layers of the retina. The **active\_contour** routine is called to used directly the **Edge\_map** as input.

#### TO DO:

**Q5.1** Rank the 4 options for the Edge\_map options from top to worst to segment the two layers wrt to segmentation quality and robustness to leyer selection.

Answer: Ranked Edge\_map options from best to worst:

```
    edge_test1
    edge_test2
    edge_test2_l
    edge_test1_2
```

```
In [ ]: img_to_seg = img_oct_eye
        # init for 1st layer
        r_left = 103; r_right=138; c_left=0; c_right = 510
        # init for 2nd layer
        #r_left = 158; r_right=204; c_left=0; c_right = 510
        alpha_val = 0.01; beta_val = 0.1; gamma_val = 0.01;
        convergence_val = 1e-4; Niter_snake = 500;
        w_line_val=1; w_edge_val=0;
        # Computation of edge maps
                     = edge_map(img_to_seg, sigma=1)
= edge_map(img_to_seg, sigma=2)
        edge test1
        edge_test2
        edge_test2_l
                          = np.log2((edge_test2*100)+1)
        edge_testl_2 = edge_map(np.log2((img_to_seg+1)*100), sigma=2)
        # Selection of edge_map to use
        Edge_map
                           = edge_test1
        # Initialise contour
        Nber_pts_contour = 200
                   = np.linspace(r_left, r_right, Nber_pts_contour)
        С
                   = np.linspace(c_left, c_right, Nber_pts_contour)
                   = np.array([r, c]).T
        init
        snake = active_contour(Edge_map,
                                init, boundary_condition='fixed-fixed',max_num_iter=N
                                alpha=alpha_val, beta=beta_val, gamma=gamma_val,
                                w_line=w_line_val, w_edge=w_edge_val)
        # FIGURE
        fig, ax = plt.subplots(figsize=(9, 5));
        ax.imshow(Edge_map[0:300,:], cmap=plt.cm.gray);
        ax.plot(init[:, 1], init[:, 0], '--y', lw=2);
        ax.plot(snake[:, 1], snake[:, 0], '-r', lw=3);
        ax.set_xticks([]), ax.set_yticks([]);
        ax.set(xlim=(0, 500));
        plt.show();
```

## Seg # 6

**BONUS - Optional** 

# Your turn on proposing a motivated pipeline using the snake capabilities from the active\_contour function

#### TODO:

Choose a new image in the pool provided and propose a segmentation pipeline using the active\_contour approach. Options on points to work on include:

- Pre-filter the image as you wish
- Manually or automatically position the initial contour
- Provide one segmentation result or merge several solutions in a probability map
- Detect issues in contour shape during deformations and propose an early stop criteria.

Q6 Provide code + visual illustrations of results

Answer

# Seg #7

#### Test on the Geometric Level-Set formulation using the Chan-Vese model.

Skimage provides two implementations of the Chan-Vese approach:

[1] chan\_vese: |implementation or original paper and

[2] morphological\_chan\_vese: faster implementation but less precise. "Active contours without edges implemented with morphological operators. It is required that the inside of the object looks different on average than the outside (i.e., the inner area of the object should be darker or lighter than the outer area on average)."

The contours of objects are now encoded in a level set function **Phi**.

The **initialisation** tested here is with a "checkerboard" pattern for 2 classes (object and background).

For the **chan\_vese** original implementation, the **hyper-parameters** include:

- mu = 0.25 (default) | edge regularisation terms. Similar to 'edge length' weight parameter. Higher mu values will produce 'smoother' contours.
- dt = 0.5 (default) | delta time step for each optimisation step.
- lambda1=1, lambda2=1 (default) | weights in the cost metric to balance inside and

outside homogeneity terms.

• tol=1e-3 (default) | Tolerance to test if the contours are "stable" and stop early.

The output contains: cv[0]=Seg and cv[1]=Phi

For the **morphological\_chan\_vese** implementation, the only **hyper-parameter** is the number of smoothing iterations (1 to 4 recommended).

#### TO DO:

Using **Chan-Vese** original implementation:

**Q7.1** Run the code on **img\_hela** with 200 iterations using **raw image** and the same image after **histogram equalisation**. What are differences observed in the Segmentation results and in the numerical values of the Phi level set function?

#### Answer:

Differences in the segmentation results: The segmented region is approximately the same, but the histogram equalisation makes the region selected smoother and more accurate (the segmentatios without filtering shows some regions outside the main area). Differences in the Phi values: The functions values without histogram equalisation appear with aliasing. The same figure with the pre filtering shows a much smoother function, without aliasing

**Q7.2** Run the code on img\_hela with **2** iterations using raw image. Why does the Segmentation image show the structures to segment but with "gray" values while this is a binary image?

**Answer**: This happens because of the initialization parameter checkboard . Then, the function  $\phi$  follows a checkboard pattern, varying from positive to negative. This creates the optical illusion of appearing gray, but in fact, it is a grid of black and white squares.

**Q7.3** Run the code on img\_hela with 200 iterations using raw image but changing the tol\_val = 10-5. Comment on major differences observed compared to tol\_val = 10-3 and propose an explanation:

**Answer**: The biggest difference is on the contours of the segmented region. With a lower tol\_val, the contours are smoother and with less abrubt changes.

**Q7.4** Run the code on **img\_cell** with 200 iterations using raw image. Comment on the issue observed with this method if you compare to the segmentation targeted given in **gt\_cell**:

Answer: The method creates holes in the segmentation that should not have appeared.

It also segments other cells that partially appear in the image. Moreover, it seems like the method segments the background instead of the cell (the cell appears black).

Using morphological\_chan\_vese implementation:

**Q7.5** Run the code on **img\_cell** with 100 iterations using raw image. Compare using Init\_method\_cv\_morpho= "checkerboard" or "disk". :

#### Answer:

General benefits from this implementation of Chan-Vese: This method correctly tries to segment the cell from the background (instead of the other way around).

Common issue seen on both segmentation results: Other objects are segmented with the cell. In this implementation, holes keep appearing inside the segmented area. Issue seen when using Morpho CV + disk: With the disk initialization, the cell in the middle of the image seems to have been correctly segmented (without holes), but it is black, which suggests the function is segmenting the background. Moreover, there seems to be small particles that were mistakenly segmented with the cell. Finally, the segmentation of the cell joins with the segmentation of another cell in the bottom of the image.

Using one of the **chan\_vese** implementation:

**Q7.6** Segment the image: **img\_fluo** and report the setup that lead to the best result. Include a display of the prefiltered-image and the segmentation results in the notebook. You can use any filtering you want :

#### Answer:

Parameters used:  $tol_val = 10^{-5}$ , 200 iterations, morphological implementation Initialisation used: checkerboard

Pre-processing used: Gaussian filtering ( $\sigma = 5$ ) followed by histogram equalization

## **Bonus points**

**Q7.7** Evaluate the quality of the segmentation of **img\_fluo** Display together your segmentation results and the ground-truth provided. Propose a measure to compare these segmentations. Provide code and display results in a new cell below:

Answer: Add here any comment you would like to add

```
In [85]: img_raw = img_fluo # img_hela[0] changed 2025 for some environments - img
img_to_seg = img_raw
# PARAMETERS
```

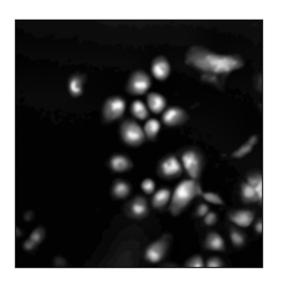
```
mu_val=0.5 ; lambda1_val=1; lambda2_val=1;
tol_val=1e-5; dt_val=0.5
smoothing_val = 1
PRE_FILTER
Num_iter_cv_ori = 200
Num_iter_cv_morpho = 200
CHAN_VESE_ORI = 0
Init_method_cv_ori = "checkerboard" # "checkerboard" or "disk" or "sma
Init_method_cv_morpho = "checkerboard" # "disk" # or "disk" or "small di
# Pre-filter (TO TURN ON IF ASKED)
if PRE FILTER:
    img_gauss = skimage.filters.gaussian(img_raw, sigma=5)
    img_adapteq = skimage.exposure.equalize_adapthist(img_gauss, clip_limit=
    img_to_seg = img_adapteq
# Select image to segment and print information
print("min - max value in image:" , np.min(img_to_seg), np.max(img_to_seg))
print("size of image:" , img_to_seg.shape)
# run segmentation
if CHAN_VESE_ORI == 1:
    # STANDARD implementation from original paper
    init_ls = checkerboard_level_set(img_to_seg.shape, 45)
    cv = chan vese(img to seq, mu=mu val, lambda1=lambda1 val, lambda2=lambd
                   tol=tol val, dt=dt val,
                   max_num_iter=Num_iter_cv_ori, init_level_set=Init_method_
                   extended output=True)
    # Show results
   Nber plots = 4
    fig, axes = plt.subplots(2,2,figsize=(7, 7))
             = axes.ravel()
    ax[0].imshow(img_to_seg, cmap=plt.cm.gray)
    ax[0].set_title("Image to segment", fontsize=12);
    ax[1].imshow(1-cv[0], cmap=plt.cm.gray)
    title = f'C-V with - {len(cv[2])} iterations'
    ax[1].set_title(title, fontsize=12)
    ax[2].imshow(1-cv[0][0:64,0:64], cmap=plt.cm.gray)
    ax[2].set_title("Zoom on segmentation result", fontsize=12);
    tmp\_show = ax[3].imshow(cv[1][0:64,0:64], cmap=plt.cm.jet)
    ax[3].set_title("Zoom on final Phi", fontsize=12);
    plt.colorbar(tmp_show,ax=ax[3], shrink=0.75, location='right')
    print("min - max value in Seg (cv[0]):" , np.min(cv[0]), np.max(cv[0]))
else:
    # FASTER implementation implemented with morphological operators BUT LES
            = morphological_chan_vese(img_to_seg, num_iter=Num_iter_cv_morph
```

```
smoothing=smoothing_val, init_level_se
# Show results
Nber_plots = 2
fig, ax = plt.subplots(1,2,figsize=(7, 7))
ax[0].imshow(img_to_seg, cmap=plt.cm.gray)
ax[1].imshow(1-cv, cmap=plt.cm.gray)
title = f'C-V_morph with - {Num_iter_cv_morpho} iterations'
ax[1].set_title(title, fontsize=12)

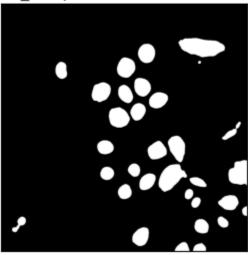
for i in range(0,Nber_plots):
    ax[i].set_xticks([]), ax[i].set_yticks([]);

plt.show();
```

min - max value in image: 0.0 1.0 size of image: (1024, 1024)

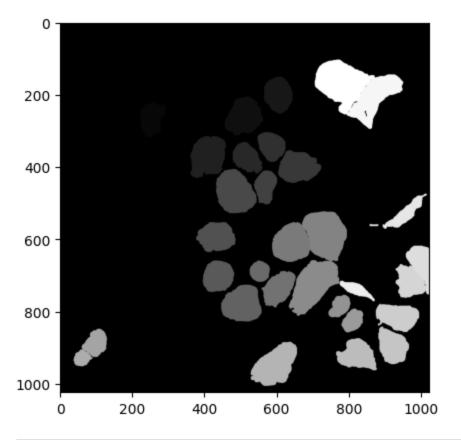


C-V\_morph with - 200 iterations



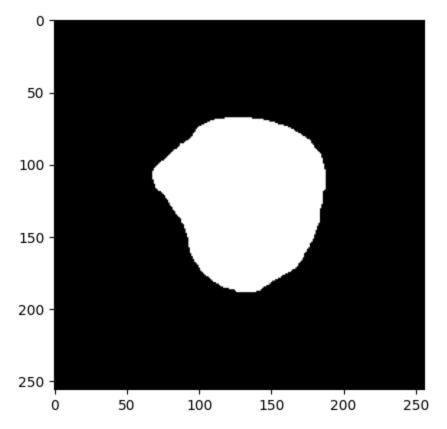
```
In [59]: plt.imshow(gt_fluo, cmap="gray")
```

Out[59]: <matplotlib.image.AxesImage at 0x12d0c0cd0>



In [30]: plt.imshow(gt\_cell, cmap="gray")

Out[30]: <matplotlib.image.AxesImage at 0x11e26bd90>



## Seg # 8

### Geometric active contours with balloon force

Controling level-set deformable model with speed values acting on the contour. You are provided with a tool to track the deformation patterns of the active contour over iterations.

The geometric active contour routine is **morphological\_geodesic\_active\_contour** which deforms a level set function with local speed values. It has the following **hyper-parameters**:

- Thresh\_cont\_val = 'auto'=> np.percentile(image, 40) (default if 'auto') | pixels <
   Thresh\_cont\_val are considered borders. The evolution of the contour will stop on
   these pixels. (It is used in the code to cancel the balloon speed using:
   Threshold\_mask\_balloon = image > threshold / np.abs(Balloon\_weight))
- Balloon\_weight = 1 (default) | weight of the balloon force. Can be negative to inflate/deflate
- **Smooth\_cont\_iter** = 1 (default) | Number of times a smoothing operator is applied per iteration

## TO DO:

**Q8.1**: Segment the **img\_cell** with the provided configuration to **inflate** the initial contour using Conf #1. What is the issue when compared to the targeted ground-truth (GT) segmentation?

**Answer**: The method segmented only the center of the cell, instead of the whole cell.

**Q8.2**: Segment the **img\_cell** with the provided configuration to **inflate** the initial contour using Conf #2.

- What is the issue when compared to the targeted ground-truth (GT) segmentation?
   Answer: It also segments another cell with the one begin analysed. This extra segmentations shows a hole as well.
- What benefit(s) do you observe when using this "implicit" formulation of a
  deformable model versus a snake-like "explicit" model using the same initialisation
  set-up and a balloon force: ?

Answer: One main advantage is that the implicit (level-set) formulation naturally

allows the contour to split or merge during deformation without extra work. An explicit (snake-like) model would need special handling whenever its topology changes.

**Q8.3**: Now Segment the **img\_CTabd** with the provided configuration Conf #3 to **deflate** the initial contour (Adjust balloon parameter accordingly). Comment with your own words the quality and properties of the obtained segmentation:

**Answer**: The kidneys are correctly segmented, but the function segments other parts of the image, like the spine. It also wrongly adds some particles to the segmentation result.

## **Bonus points**

**Q8.4a**: Check by yourself and explain why you think pre-processing using skimage.morphology.diameter\_closing help the segmentation on **img\_CTabd**? **Answer**:

**Q8.4b**: Provide a different set up (with printed code and results) with different initialisation and/or pre-processing set-up that leads to a correct segmentation of the 2 kidneys (as in the ground truth).

Answer:

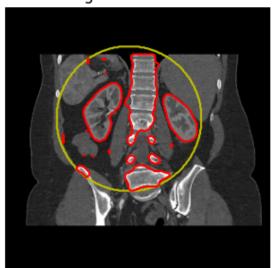
```
In [91]: # img_cell:
         # img_raw = img_cell;
         # gt_seg
                     = gt_cell
         # img_CTabd:
         img_raw = img_CTabd ;
         gt_seg = gt_CTabd
         # Select img to segment
         img_to_seg = img_raw;
         # Conf #1: Disk parameters to initialise shape for INFLATION on img_cell
         \# \ r0 = 130; \ c0 = 125 ; \ R0 = 30 \ \# \ inflate
         # Conf #2: Disk parameters to initialise shape for INFLATION on img on img_c
         \# r0 = 130; c0 = 125; R0 = 45 \# inflate
         # Conf #3: Disk parameters to initialise shape for DEFLATION on img
         r0 = 130; c0 = 145 ; R0 = 85 # deflate TO KEEP ON CTabd
         # Hyper-parameter for Balloon velocity:
         Balloon weight = -1; # +1 or -1 to Inflate or deflate
         # Additional Hyper-parameters for geodesic deformable model:
         Threshold_contour_level = 20;
```

```
Smooth_cont_iter = 1 ; # Number of times a smoothing operator is appl
Niter_snake
# Hyper-parameter for preparing the input data:
INV_EDGE_MAP = 1; # needed when using the Balloon force
# Pre-processing the image with closing
img_to_seg = skimage.morphology.diameter_closing(img_to_seg, 40, connectivit
# Inverse edge image: Compute the magnitude of the gradients in the image an
# then inverts the result in the range [0, 1]
if INV EDGE MAP:
     img to seg = skimage.segmentation.inverse gaussian gradient(
                                    img_to_seg,alpha=50,sigma=2)
# Print information
print("min - max value in image to seg:" , np.min(img_to_seg), np.max(img_to
print("size of image to seg:" , img_to_seg.shape)
# Set and print threshold used in morphological_geodesic_active_contour
# Areas of the inverse edge map with value < Threshold will be considered bo
Threshold_contour = np.percentile(img_to_seg, Threshold_contour_level);
Thresh_cont_val = Threshold_contour ; # 'auto' ; # pixels < Thresh_cont_va
print("threshold used in morphological_geodesic", Thresh_cont_val)
# initialise call back
evolution = []
callback = store_evolution_in(evolution)
# Initialise contour as a disk
init_ls = skimage.segmentation.disk_level_set(img_to_seg.shape, center=[r0,
# Run geodesic active contour
        = morphological_geodesic_active_contour(
            img_to_seg, Niter_snake, init_ls,
            smoothing=Smooth cont iter, balloon=Balloon weight,
            threshold=Thresh cont val,
            iter_callback=callback);
# Figure with plots of level-set evolution
fig, axes = plt.subplots(2, 2, figsize=(8, 8));
ax = axes.flatten();
ax[0].imshow(img_raw, cmap="gray");
ax[0].set_axis_off();
contour = ax[0].contour(evolution[0], [0.5], colors='y');
#contour.collections[0].set_label("Contours"); #removed 2025
ax[0].contour(ls, [0.5], colors='r');
ax[0].set_title("Img ori + contours", fontsize=12);
ax[1].imshow(img_to_seg, cmap="gray");
ax[1].set_axis_off();
contour = ax[1].contour(evolution[0], [0.5], colors='y');
#contour.collections[0].set_label("Contours"); #removed 2025
#contour.set_label("Contours"); # option to update 2025
```

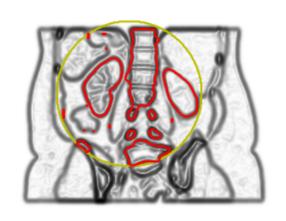
```
#ax[0].contour(ls, [0.5], colors='r');#removed 2025
ax[1].contour(ls, [0.5], colors='r');
ax[1].set_title("Img to seg + contours", fontsize=12);
#ax[1].contour(ls, [0.5], colors='r');#removed 2025
ax[2].imshow(img_to_seg, cmap="gray");
ax[2].set_axis_off();
contour = ax[2].contour(evolution[0], [0.5], colors='y');
#contour.collections[0].set label("Contours"); # removed 2025
title = f'Morphological GAC Curve evolution';
ax[2].set_title(title, fontsize=12);
for i in range(1, Niter_snake-1, 5):
    contour = ax[2].contour(evolution[i], [0.5], linewidths=0.5, colors='r')
ax[3].imshow(gt seg, cmap="gray");
ax[3].set_axis_off();
ax[3].contour(ls, [0.5], colors='r');
ax[3].set_title("GT segmentation mask", fontsize=12);
plt.show();
```

min - max value in image to seg: 0.3062454376394709 1.0 size of image to seg: (311, 311) threshold used in morphological\_geodesic 0.7199191982831742

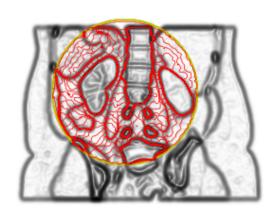
Img ori + contours



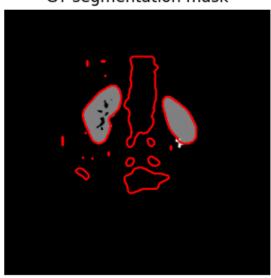
Img to seg + contours



Morphological GAC Curve evolution



GT segmentation mask



In [ ]: