



## Standard Guide for Pore-Liquid Sampling from the Vadose Zone<sup>1</sup>

This standard is issued under the fixed designation D 4696; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

ε<sup>1</sup> NOTE—Paragraph 1.6 was added editorially in October 1998.

### 1. Scope

1.1 This guide discusses equipment and procedures used for sampling pore-liquid from the vadose zone (unsaturated zone). The guide is limited to in-situ techniques and does not include soil core collection and extraction methods for obtaining samples.

1.2 The term “pore-liquid” is applicable to any liquid from aqueous pore-liquid to oil. However, all of the samplers described in this guide were designed, and are used to sample aqueous pore-liquids only. The abilities of these samplers to collect other pore-liquids may be quite different than those described.

1.3 Some of the samplers described in this guide are not currently commercially available. These samplers are presented because they may have been available in the past, and may be encountered at sites with established vadose zone monitoring programs. In addition, some of these designs are particularly suited to specific situations. If needed, these samplers could be fabricated.

1.4 The values stated in SI units are to be regarded as the standard.

1.5 *This standard does not purport to address all of the safety problems, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

1.6 *This guide offers an organized collection of information or a series of options and does not recommend a specific course of action. This document cannot replace education or experience and should be used in conjunction with professional judgment. Not all aspects of this guide may be applicable in all circumstances. This ASTM standard is not intended to represent or replace the standard of care by which the adequacy of a given professional service must be judged, nor should this document be applied without consideration of a project's many unique aspects. The word “Standard” in the title of this document means only that the document has been approved through the ASTM consensus process.*

### 2. Referenced Documents

#### 2.1 ASTM Standards:

D 653 Terminology Relating to Soil, Rock, and Contained Fluids<sup>2</sup>

### 3. Terminology

#### 3.1 Definitions:

3.1.1 Where reasonable, precise terms and names have been used within this guide. However, certain terms and names with varying definitions are ubiquitous within the literature and industry of vadose zone monitoring. For purposes of recognition, these terms and names have been included in the guide with their most common usage. In these instances, the common definitions have been included in Appendix X1. Examples of such terms are soil, lysimeter, vacuum and pore-liquid tension.

#### 3.2 Definitions of Terms Specific to This Standard:

3.2.1 Appendix X1 is a compilation of those terms used in this guide. More comprehensive compilations, that were used as sources for Appendix X1, are (in decreasing order of their usage):

3.2.1.1 Terminology D 653,

3.2.1.2 *Compilation of ASTM Terminology*,<sup>3</sup>

3.2.1.3 *Glossary of Soil Science Terms*, Soil Science Society of America,<sup>4</sup> and,

3.2.1.4 *Webster's New Collegiate Dictionary*,<sup>5</sup>

### 4. Summary of Guide

4.1 Pores in the vadose zone can be saturated or unsaturated. Some samplers are designed to extract liquids from unsaturated pores; others are designed to obtain samples from saturated pores (for example, perched ground water) or saturated macropores (for example, fissures, cracks, and burrows). This guide addresses these categories. The sampler types discussed are:

4.1.1 Suction samplers (unsaturated sampling), (see Section 7),

4.1.2 Free drainage samplers (saturated sampling), (see Section 8),

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<sup>2</sup> *Annual Book of ASTM Standards*, Vol 04.08.

<sup>3</sup> *Compilation of ASTM Terminology*, Sixth edition, ASTM, 1916 Race Street, Philadelphia, PA 19103, 1986.

<sup>4</sup> *Glossary of Soil Science Terms*, Soil Science Society of America, 1987.

<sup>5</sup> *Webster's New Collegiate Dictionary*, Fifth edition, 1977.

4.1.3 Perched ground water samplers (saturated sampling), (see Section 9), and

4.1.4 Experimental absorption samplers (unsaturated sampling), (see Section 10).

4.2 Most samplers designed for sampling liquid from unsaturated pores may also be used to sample from saturated pores. This is useful in areas where the water table fluctuates, so that both saturated and unsaturated conditions occur at different times. However, samplers designed for sampling from saturated pores cannot be used in unsaturated conditions. This is because the liquid in unsaturated pores is held at less than atmospheric pressures (see *Richard's outflow principle*, in Appendix X1).

4.3 The discussion of each sampler is divided into specific topics that include:

- 4.3.1 Operating principles,
- 4.3.2 Description,
- 4.3.3 Installation,
- 4.3.4 Operation, and
- 4.3.5 Limitations.

## 5. Significance and Use

5.1 Sampling from the vadose zone may be an important component of some ground water monitoring strategies. It can provide information regarding contaminant transport and attenuation in the vadose zone. This information can be used for mitigating potential problems prior to degradation of a ground water resource (1).<sup>6</sup>

5.2 The choice of appropriate sampling devices for a particular location is dependent on various criteria. Specific guidelines for designing vadose zone monitoring programs have been discussed by Morrison (1), Wilson (2), Wilson (3), Everett (4), Wilson (5), Everett et al (6), Wilson (7), Everett et

al (8), Everett et al (9), Robbins et al (10), Merry and Palmer (11), U.S. EPA (12), Ball (13), and Wilson (14). In general, it is prudent to combine various unsaturated and free drainage samplers into a program, so that the different flow regimes may be monitored.

5.3 This guide does not attempt to present details of installation and use of the equipment discussed. However, an effort has been made to present those references in which the specific techniques may be found.

## 6. Criteria for Selecting Pore-Liquid Samplers

6.1 Decisions on the types of samplers to use in a monitoring program should be based on consideration of a variety of criteria that include the following:

- 6.1.1 Required sampling depths,
- 6.1.2 Required sample volumes,
- 6.1.3 Soil characteristics,
- 6.1.4 Chemistry and biology of the liquids to be sampled,
- 6.1.5 Moisture flow regimes,
- 6.1.6 Required durability of the samplers,
- 6.1.7 Required reliability of the samplers,
- 6.1.8 Climate,
- 6.1.9 Installation requirements of the samplers,
- 6.1.10 Operational requirements of the samplers,
- 6.1.11 Commercial availability, and
- 6.1.12 Costs.

6.2 Some of these criteria are discussed in this guide. However, the ability to balance many of these factors against one another can only be obtained through field experience.

## 7. Suction Samplers

7.1 Table 1 presents the various types of suction samplers. The range of operating depths is the major criterion by which suction samplers are differentiated. Accordingly, the categories of suction samplers are as follows:

7.1.1 *Vacuum Lysimeters*—These samplers are theoretically operational at depths less than about 7.5 m. The practical

<sup>6</sup> The boldface numbers in parentheses refer to the list of references at the end of this standard.

**TABLE 1 Suction Sampler Summary**

Sampler Type	Porous Section Material	Maximum <sup>A</sup> Pore Size (µm)	Air Entry Value (cbar)	Operational Suction Range (cbar)	Maximum Operation Depth (m)
Vacuum lysimeters	Ceramic	1.2 to 3.0 (1) <sup>A</sup>	>100	<60 to 80	<7.5
	PTFE	15 to 30 (2) <sup>A</sup>	10 to 21	<10 to 21	<7.5
	Stainless steel	NA <sup>B</sup>	49 to 5	49 to 5	<7.5
Pressure-vacuum lysimeters	Ceramic	1.2 to 3.0 (1) <sup>A</sup>	>100	<60 to 80	<15
	PTFE	15 to 30 (2) <sup>A</sup>	10 to 21	<10 to 21	<15
High pressure-vacuum lysimeters	Ceramic	1.2 to 3.0 (1) <sup>A</sup>	>100	<60 to 80	<91
	PTFE	15 to 30 (2) <sup>A</sup>	10 to 21	<10 to 21	<91
Filter tip samplers	Polyethylene	NA <sup>B</sup>	NA <sup>B</sup>	NA <sup>B</sup>	None
	Ceramic	2 to 3 (1)	>100	<60 to 80	<7.5
	Stainless steel	NA <sup>B</sup>	NA <sup>B</sup>	NA <sup>B</sup>	none
Cellulose-acetate hollow-fiber samplers	Cellulose	<2.8	>100	<60 to 80	<7.5
	Acetate				
	Non cellulosic				
Membrane filter samplers	Polymer	<2.8	>100	<60 to 80	<7.5
	Cellulose	<2.8	>100	<60 to 80	<7.5
	Acetate				
Vacuum plate samplers	PTFE	2 to 5	NA <sup>B</sup>	NA <sup>B</sup>	<7.5
	Alundum	NA <sup>B</sup>	NA <sup>B</sup>	NA <sup>B</sup>	<7.5
	Ceramic	1.2 to 3.0	>100	60 to 80	<7.5
	Fritted glass	4 to 5.5	NA <sup>B</sup>	NA <sup>B</sup>	<7.5
	Stainless steel	NA <sup>B</sup>	49 to 5	49 to 5	<7.5

<sup>A</sup>Pore size determined by bubbling pressure (1) or mercury intrusion (2).

<sup>B</sup>NA = Not available.

operational depth is 6 m under ideal conditions.

**7.1.2 Pressure-Vacuum Lysimeters**—These samplers are operational at depths less than about 15 m.

**7.1.3 High Pressure-Vacuum Lysimeters**—(Also known as pressure-vacuum lysimeters with transfer vessels.) These samplers are normally operational down to about 46 m, although installations as deep as 91 m have been reported (15).

**7.1.4 Suction Lysimeters With Low Bubbling Pressures (Samplers With PTFE Porous Sections)**—These samplers are available in numerous designs that can be used to maximum depths varying from about 7.5 to 46 m.

NOTE 1—The samplers of 7.1.1, 7.1.2, 7.1.3, and 7.1.4 are referred to collectively as suction lysimeters. Within this standard, lysimeter is defined as a device used to collect percolating water for analyses (16).

**7.1.5 Filter Tip Samplers**—These samplers theoretically have no maximum sampling depth.

**7.1.6 Experimental Suction Samplers**—The samplers have limited field applications at the present time. They include cellulose-acetate hollow-fiber samplers, membrane filter samplers, and vacuum plate samplers. They are generally limited to depths less than about 7.5 m.

## 7.2 Operating Principles:

### 7.2.1 General:

**7.2.1.1 Suction lysimeters** consist of a hollow, porous section attached to a sample vessel or a body tube. Samples are obtained by applying suction to the sampler and collecting pore-liquid in the body tube. Samples are retrieved by a variety of methods.

**7.2.1.2 Unsaturated portions of the vadose zone** consist of interconnecting soil particles, interconnecting air spaces, and interconnecting liquid films. Liquid films in the soil provide hydraulic contact between the saturated porous section of the sampler and the soil (see Fig. 1). When suction greater than the soil pore-liquid tension is applied to the sampler, a pressure potential gradient towards the sampler is created. If the menisci of the liquid in the porous segment are able to withstand the applied suction (depending on the maximum

pore sizes and hydrophobicity/hydrophilicity), liquid moves into the sampler. The ability of the menisci to withstand a suction decreases with increasing pore size and also with increasing hydrophobicity of the porous segment (see 7.6). If the maximum pore sizes are too large and hydrophobicity too great, the menisci are not able to withstand the applied suction. As a result, they break down, hydraulic contact is lost, and only air enters the sampler. As described in 7.6, ceramic porous segments are hydrophilic and the maximum pore sizes are small enough to allow menisci to withstand the entire range of sampling suctions. Presently available polytetrafluoroethylene (PTFE) porous segments are hydrophobic, the maximum pore sizes are larger, and only a very limited range of sampling suction can be applied before menisci break down and sampling ends (see 7.6.1.3). Therefore, samplers made with PTFE porous segments may be used only for sampling soils with low pore-liquid tensions (12, 17).

**7.2.1.3** The ability of a sampler to withstand applied suctions can be directly measured by its bubbling pressure. The bubbling pressure is measured by saturating the porous segment, immersing it in water, and pressurizing the inside of the porous segment with air. The pressure at which air starts bubbling through the porous segment into the surrounding water is the bubbling pressure. The magnitude of the bubbling pressure is equal to the magnitude of the maximum suction that can be applied to the sampler before air entry occurs (air entry value). Because the bubbling pressure is a direct measure of how a sampler will perform, it is more useful than measurement of pore size distributions.

**7.2.1.4** As soil pore-liquid tensions increase (low pore-liquid contents), pressure gradients towards the sampler decrease. Also, the soil hydraulic conductivity decreases exponentially. These result in lower flow rates into the sampler. At pore-liquid tensions above about 60 (for coarse grained soils) to 80 cbar (for fine grained soils), the flow rates are effectively zero and samples cannot be collected.

### 7.2.2 Suction Lysimeters:

**7.2.2.1** Vacuum lysimeters directly transfer samples to the surface via a suction line. Because the maximum suction lift of water is about 7.5 m, these samplers cannot be operated below this depth. In reality, suction lifts of 6 m should be considered a practical maximum depth.

**7.2.2.2** Samples may be retrieved using the same technique as for vacuum lysimeters or, for deeper applications, the sample is retrieved by pressurizing the sampler with one line; this pushes the sample up to the surface in a second line.

**7.2.2.3** High pressure-vacuum lysimeters operate in the same manner as pressure-vacuum lysimeters. However, they include an inbuilt check transfer vessel or a chamber between the sampler and the surface. This prevents sample loss through the porous section during pressurization, and prevents possible cup damage due to overpressurization.

**7.2.2.4** Suction lysimeters with low bubbling pressures are available in each of the three previous designs. The only difference between these samplers and the three previous designs is that these porous sections are made with PTFE. The low bubbling pressure (and hence large pore size or hydrophobicity, or both) of PTFE constrains these samplers to soils that

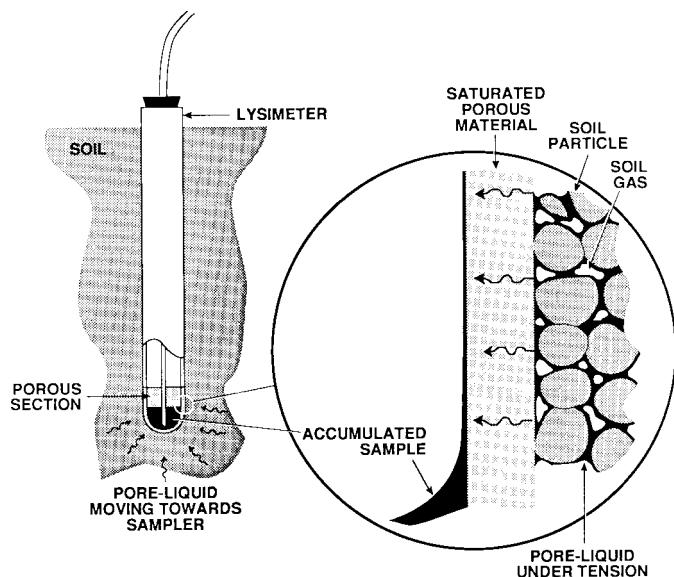


FIG. 1 Pore Section/Soil Interactions

are nearly saturated (see 7.2.1.2 and 7.6.1.3).

**7.2.3 Filter Tip Samplers**—Samples are collected from a filter tip sampler by lowering an evacuated sample vial down an access tube to a permanently emplaced porous tip. The vial is connected to the porous tip and sample flows through the porous section and into the vial. Once full, the vial is retrieved.

**7.2.4 Experimental Suction Samplers**—Experimental suction samplers generally operate on the same principle as vacuum lysimeters with different combinations of porous materials to enhance hydraulic contact. The samplers are generally fragile and difficult to install. As with vacuum lysimeters, they are generally limited to depths of less than about 7.5 m.

### 7.3 Description:

#### 7.3.1 Vacuum Lysimeters:

**7.3.1.1** Vacuum lysimeters generally consist of a porous cup mounted on the end of a tube, similar to a tensiometer. The cup is attached to the tube with adhesives (18<sup>7</sup>) or with “V” shaped flush threading sealed with an “O” ring. A stopper is inserted into the upper end of the body tube and fastened in the same manner as the porous cup or, in the case of rubber stoppers, inserted tightly (12). To recover samples, a suction line is inserted through the stopper to the base of the sampler. The suction line extends to the surface and connects to a sample bottle and suction source in series. Body tubes up to 1.8 m long have been reported (15) (see Fig. 2).

**7.3.1.2** Harris and Hansen (19) described a vacuum lysimeter with a 6 mm by 65 mm ceramic porous cup designed for intensive sampling in small areas.

**7.3.1.3** A variety of materials have been used for the porous segment including nylon mesh (20), fritted glass (21), sintered glass (22), Alundum (manufacturer name), stainless steel (23<sup>7</sup>), and ceramics (1.2 to 3.0  $\mu\text{m}$  max pore size) (18<sup>7</sup>). The sampler body tube has been made with PVC, ABS, acrylic, stainless steel (24) and PTFE (18<sup>7</sup>, 25<sup>7</sup>). Ceramic porous segments are attached with epoxy adhesives or with flush threading. The stopper is typically made of rubber (12), neoprene, or PTFE. The outlet lines are commonly PTFE, rubber, polyethylene, polypropylene, vinyl, nylon, and historically, copper. Fittings and valves are available in brass or stainless steel.

#### 7.3.2 Pressure-Vacuum Lysimeters:

**7.3.2.1** These samplers were developed by Parizek and Lane (26) for sampling deep moving pollutants in the vadose zone. The porous segment is usually a porous cup at the bottom of a body tube. The porous cup is attached with epoxy adhesives (18<sup>7</sup>) or with “V” shaped flush threading sealed with an “O” ring (25<sup>7</sup>). Two lines are forced through a two-hole stopper sealed into the upper end of the body tube. The discharge line extends to the base of the sampler and the pressure-vacuum line terminates a short distance below the stopper. At the surface, the discharge line connects to a sample bottle and the pressure-vacuum line connects to a pressure-vacuum pump. Designs are available that do not use a stopper but rather an “O” ring sealed, flush threaded top plug (25<sup>7</sup>). Tubing lines to the surface are attached to the top plug with threaded tubing

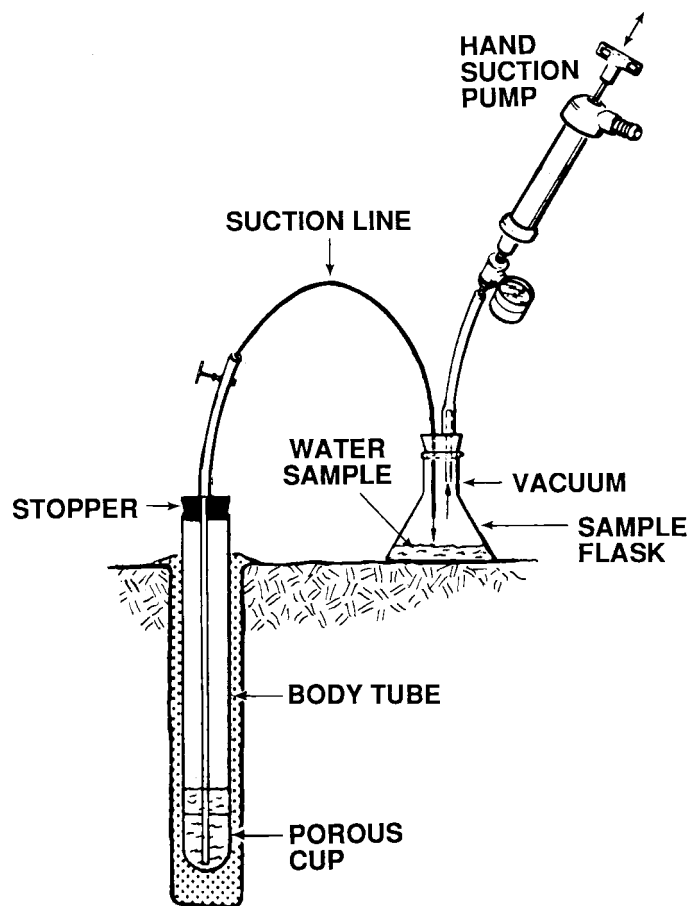


FIG. 2 Vacuum Lysimeter

fittings of appropriate materials. Body tubes are commonly available with 2.2 and 4.8 cm diameters and in a variety of lengths (see Fig. 3). The sampler and its components have been made out of the same materials used for vacuum lysimeters.

**7.3.2.2** These samplers can retrieve samples from depths below 7.5 m because pressure is used for retrieval. However, during pressurization some of the sample is forced back out of the cup. At depths over about 15 m, the volume of sample lost in this fashion may be significant. In addition, at depths over about 15 m, pressures required to bring the sample to the surface may be high enough to damage the cup or to reduce its hydraulic contact with the soil (27, 28). Rapid pressurization causes similar problems. Morrison and Tsai (29) developed a tube lysimeter with the porous section located midway up the body tube instead of at the bottom (see Fig. 4). This design mitigates the problem of sample being forced back through the cup. However, it does not prevent problems with porous segment damage due to overpressurization or rapid pressurization. The sleeve lysimeter (that is no longer available) was a modification to this design for use with a monitoring well (1) (see Fig. 5). Another modification is the casing lysimeter that consists of several tube lysimeters threaded into one unit (see Fig. 6). This arrangement allows precise spacing between units (30).

**7.3.2.3** Nightingale et al (31) described a design that allows incoming samples to flow into a portion of the sampler not in contact with the basal, porous ceramic cup (see Fig. 7). The

<sup>7</sup> This reference is manufacturer's literature, and it has not been subjected to technical review.

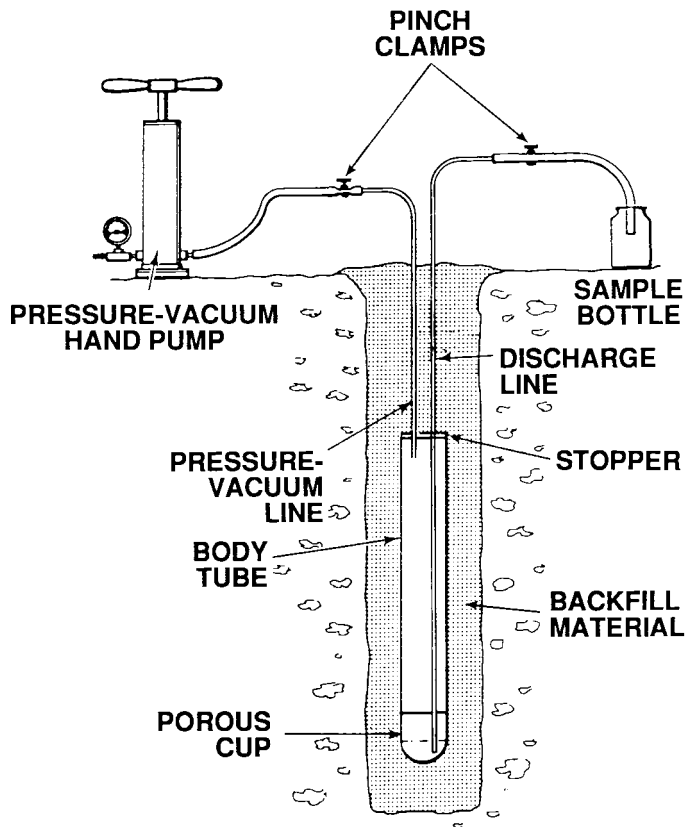


FIG. 3 Pressure-Vacuum Lysimeter

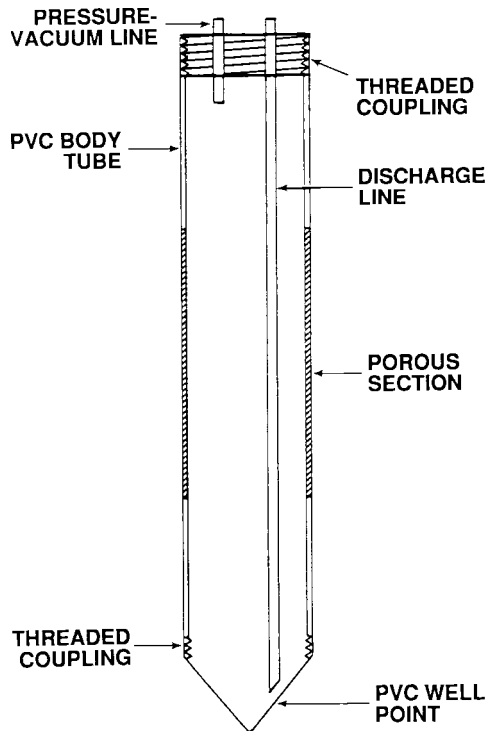


FIG. 4 Tube Pressure-Vacuum Lysimeter

ceramic cup is wedged into the body tube without adhesives or threading. The sampler was used to sample the vadose zone, the capillary fringe, and the fluctuating water table in a recharge area. Knighton and Streblow (32) reported a sampler

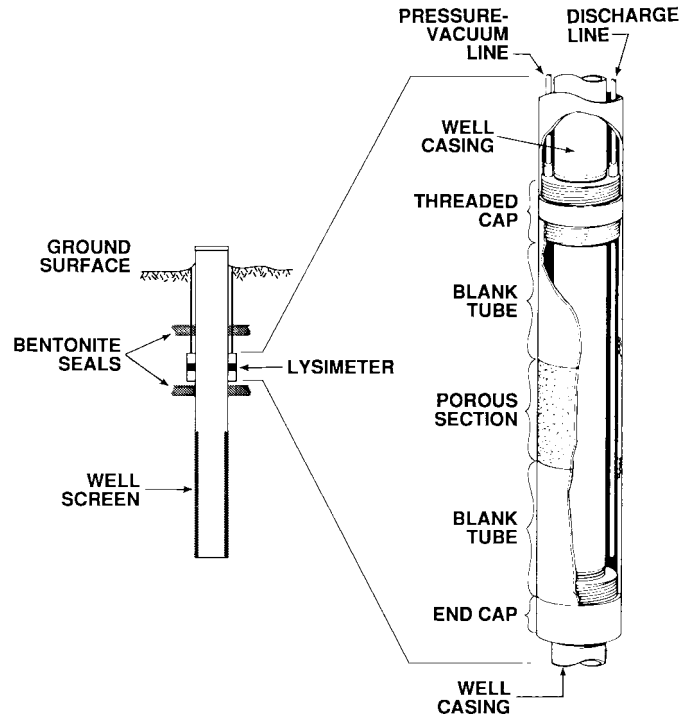


FIG. 5 Sleeve Lysimeter

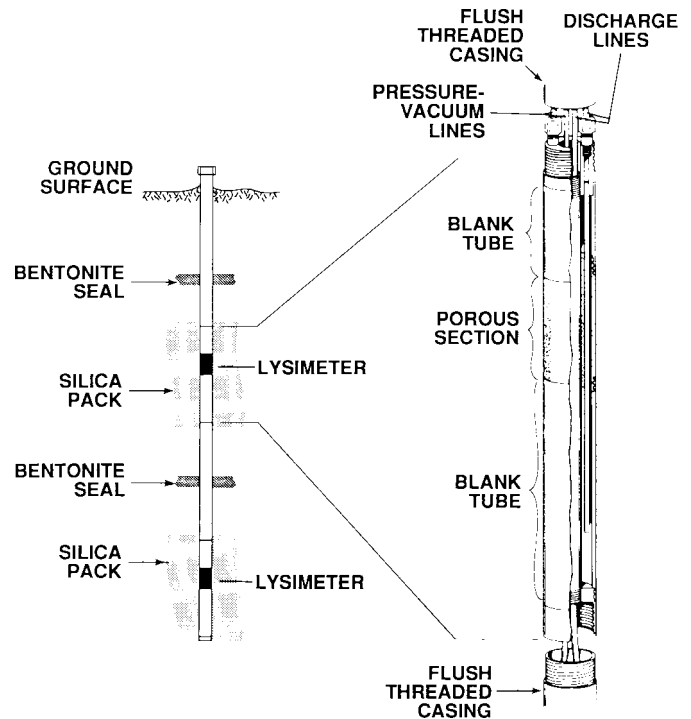


FIG. 6 Casing Lysimeter

with the porous cup upon the top of a chamber. This design was used with cup diameters ranging from 7.6 to 12.7 cm (see Fig. 8). These designs also allow pressurization for sample retrieval without significant liquid loss. However, because the porous cups are open to the rest of the samplers, possible damage due to overpressurization or rapid pressurization is still a problem.



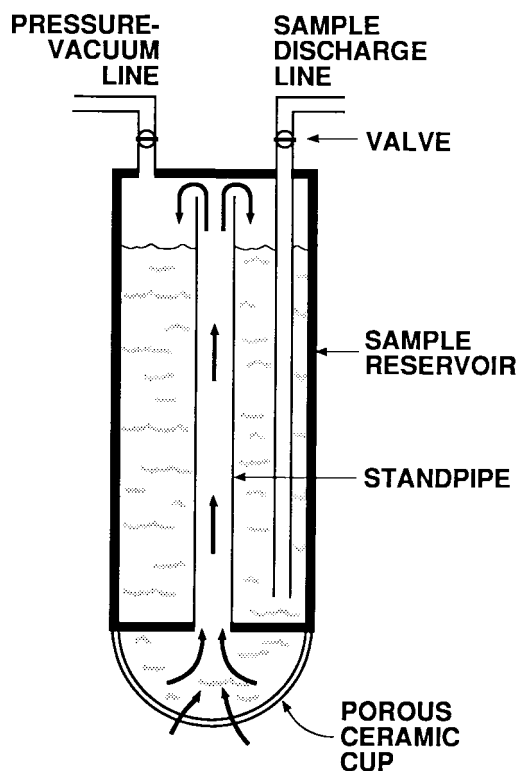


FIG. 7 Modified Pressure-Vacuum Lysimeter

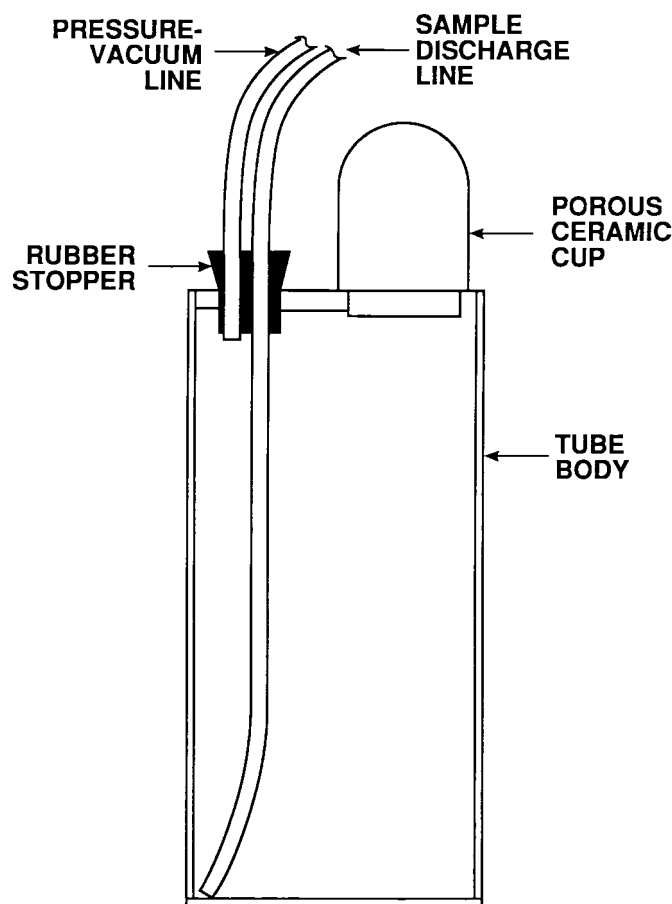


FIG. 8 Knighton and Streblow-Type Vacuum Lysimeter

**7.3.3 High Pressure-Vacuum Lysimeters (Lysimeters With a Transfer Vessel)**—High pressure-vacuum lysimeters overcome the problems of fluid loss and overpressurization through the use of an attached chamber or a connected transfer vessel (see Fig. 9). The porous segment is usually a porous cup at the bottom of the body tube. The cup is attached with epoxy adhesives (18<sup>7</sup>) or with “V” shaped flush threading sealed with an “O” ring (25<sup>7</sup>). In the attached chamber design, the body tube is separated into two chambers connected by a one-way check valve. A pressure-vacuum line and a discharge line enter through a two-hole plug at the top of the body tube. The pressure-vacuum line terminates below the plug. The discharge line extends to the bottom of the upper chamber. The transfer vessel design is similar. However, the vessel and body tube are integral components joined by a common double threaded, “O” ring sealed plug containing a check valve. Body tube diameters range from 2.7 to 8.9 cm outside diameter. Total sampler lengths commonly range from 15.2 to 182.9 cm. A threaded top plug allows attachment of casing to the lysimeter. This facilitates accurate placement and provides long-term protection for the tubing lines. The samplers and their components have been made out of the same materials as vacuum lysimeters.

**7.3.4 Suction Lysimeters with Low Bubbling Pressures (Samplers With PTFE Porous Sections)**—Designs are available in each of the three categories described in 7.3.1, 7.3.2, and 7.3.3. The only difference between this group of samplers and the previous three samplers is that PTFE is used for the porous sections of this group of samplers (25<sup>7</sup>). The porous PTFE is attached with “V” shaped flush threading sealed with an “O” ring.

#### 7.3.5 Filter Tip Samplers:

7.3.5.1 Filter tip samplers consist of two components: a

permanently installed filter tip, and a retrievable glass sample vial. The filter tip includes a pointed end to help with installation, a porous section, a nozzle, and a septum. The tip is threaded onto extension pipes that extend to the surface. The sample vial includes a second septum. When in use, the vial is seated in an adaptor that includes a disposable hypodermic needle to penetrate both the septa, allowing sample to flow from the porous segment into the vial (see Fig. 10). Extension pipes vary from 2.5 to 5.1 cm inside diameter. Vial volumes range from 35 to 500 mL (32<sup>7</sup>).

7.3.5.2 The body of the filter tip is made of thermoplastic, stainless steel, or brass. The attached porous section is available in high density polyethylene, sintered ceramic, or sintered stainless steel. The septum is made of natural rubber, nitrile rubber, or fluororubber (32<sup>7</sup>).

#### 7.3.6 Experimental Suction Samplers:

7.3.6.1 Cellulose-acetate, hollow-fiber samplers were described by Jackson et al (33) and Wilson (3). A sampler consists of a bundle of these flexible, hollow fibers (<2.8  $\mu$ m max pore size) pinched shut at one end and attached to a suction line at the other end. The suction line leads to the surface and attaches to a sample bottle and source of suction in the same manner as a vacuum lysimeter (see Fig. 11). The fibers, that are analogous to the porous sections of vacuum lysimeters, have outside diameters of up to 250  $\mu$ m (33). Levin and Jackson (34) described similar fibers made from a noncellulosic polymer solution (max pore size <2.8  $\mu$ m). Those fibers

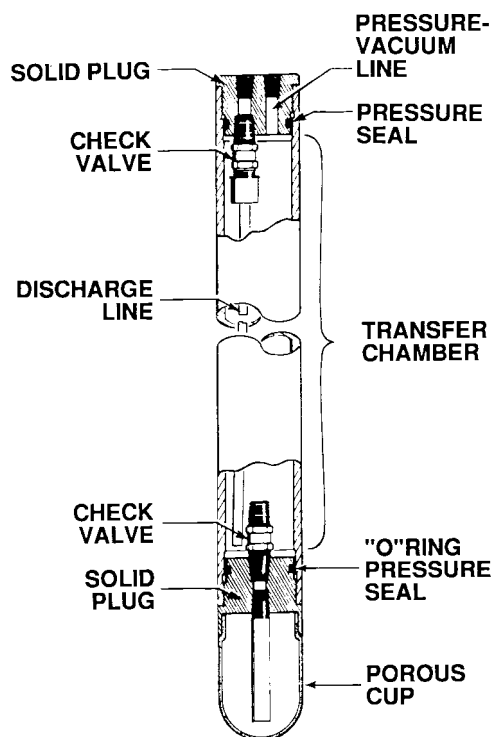


FIG. 9 High Pressure-Vacuum Lysimeter

have dense inner layers surrounded by open celled, spongy layers with diameters ranging from 50 to 250  $\mu\text{m}$ .

7.3.6.2 Membrane filter samplers were described by Morrison (1), Everett and Wilson (6), U.S. EPA (12) and Stevenson (35). A sampler consists of a membrane filter of polycarbonate, cellulose acetate ( $<2.8 \mu\text{m}$  max pore size), cellulose nitrate or PTFE (2 to 5  $\mu\text{m}$  max pore size); mounted in a "swinnex" type filter holder (35, 36, 37). The filter rests on a glass fiber prefilter. The prefilter rests on a glass fiber "wick" that in turn sits on a glass fiber collector. The collector is in contact with the soil and extends the sampling area of the small diameter filter (see Fig. 12 and 7.5.1.6). A suction line leads from the filter holder to the surface. At the surface, the suction line is attached to a sample bottle and suction source in a manner similar to vacuum lysimeters.

7.3.6.3 A vacuum plate sampler consists of a flat porous disk fitted with a nonporous backing attached to a suction line that leads to the surface (see Fig. 13). Plates are available in diameters ranging from 4.3 to 25.4 cm and custom designs are easily arranged (1, 18). Plates are available in alundum, porous stainless steel (23), ceramic (1.2 to 3.0  $\mu\text{m}$  max pore size) or fritted glass (4 to 5.5  $\mu\text{m}$  max pore size) (38, 6, 39, 40, 41, 42, 43, 44). The nonpermeable backing can be a fiberglass resin, glass, plastic, or butyl rubber.

#### 7.3.7 Comments:

7.3.7.1 When some ceramic cups are glued to the inner wall of the body tube in a suction lysimeter, an inner lip is formed (45). As the discharge line is pushed through the stopper at the top of the sampler, it may catch on this lip and the operator may conclude that the line has reached the bottom of the ceramic cup (see Fig. 14). As a result, an 80 mL error can occur in sampling rate determinations. This 80 mL of fluid accumulates in the cup, is not removed during sampling, and will cause

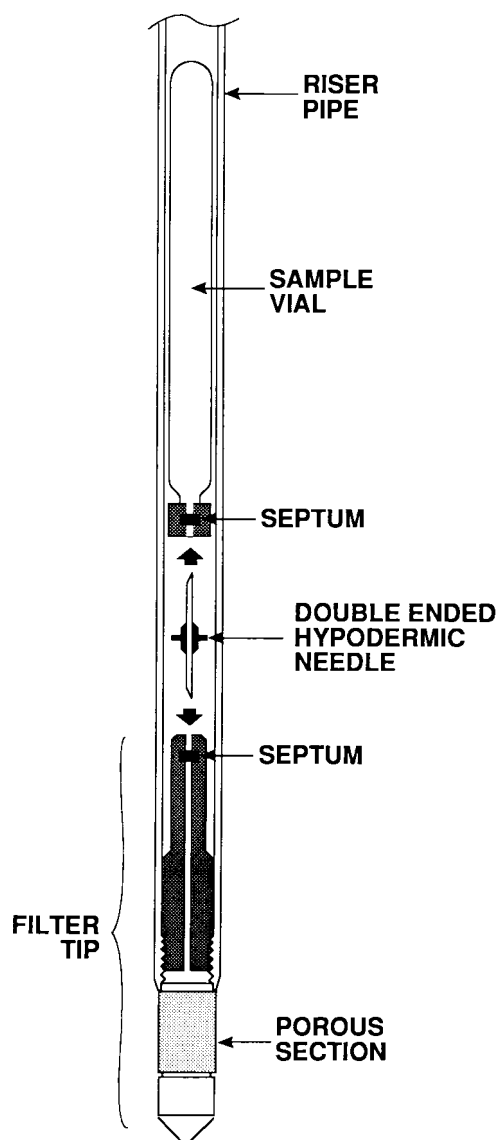


FIG. 10 Filter Tip Sampler

cross contamination between sampling events. Soilmoisture (18) suggested that the line can be kept from catching by cutting its tip at an angle. In all-PTFE suction lysimeters, the discharge line is a rigid PTFE tube extending to the bottom of the cup. This results in a zero accumulation of fluid. Older samplers with PTFE porous segments and PVC body tubes have a discharge line that does not extend all the way to the bottom. This problem has been corrected in newer PTFE and PVC samplers (25). This results in a 34 mL accumulation of fluid (12). Filter tip samplers develop an 8 mL accumulation of fluid. Haldorsen et al. (46) suggested collecting and discarding an initial sample to purge this accumulated fluid.

7.3.7.2 Because samplers are often handled roughly during installation, durability and ruggedness are important. It has been shown that PTFE has a higher impact strength than ceramics which need to be installed with care (25). It has also been found that PTFE threads and ceramic threads (when used) are susceptible to leakage, and must be securely sealed with pipe threading tape (45). TFE-fluorocarbon (PTFE) tape is not

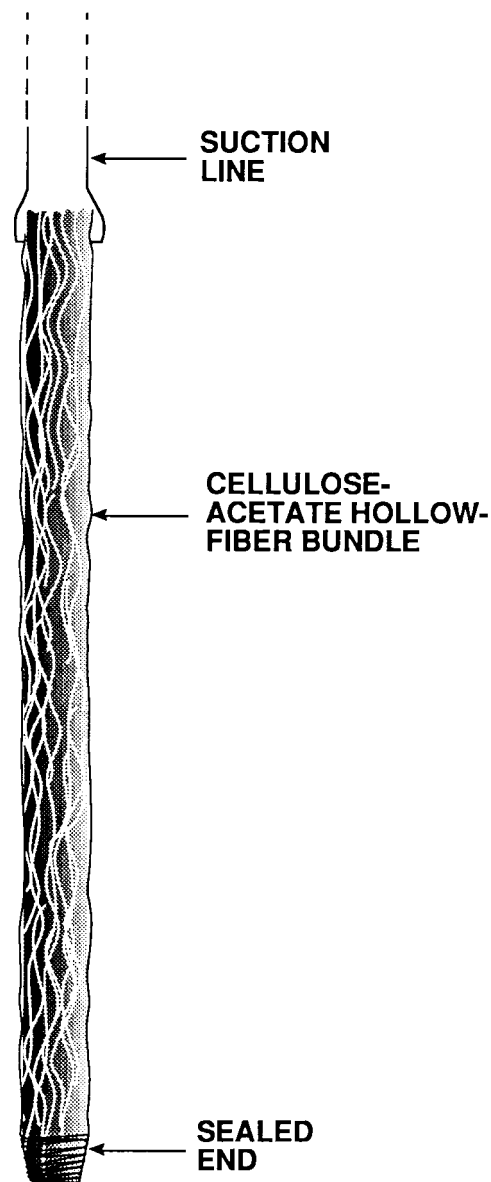


FIG. 11 Cellulose-Acetate Hollow-Fiber Sampler

recommended in square threaded joints since the tape is designed for tapered "V" threaded compression joints.

7.3.7.3 As described above, porous sections can be made from various materials. These materials have physical and chemical limitations that must be considered when designing a monitoring program. Physical limitations are described in 7.6.1. Chemical limitations are described in 7.6.2.

#### 7.4 Installation Methods:

##### 7.4.1 Pre-Installation:

7.4.1.1 As demonstrated by Neary and Tomassini (47), new samplers may be contaminated with dust during manufacturing. In order to reduce chemical interferences from substances on the porous sections, U.S. EPA (12) recommended preparation of ceramic units prior to installation following procedures originally developed by Wolff (48), modified by Wood (49) and recommended by Neary and Tomassini (47). The process involves passing hydrochloric acid (HCl) (for example, 8N) through the porous sections. This is followed by flushing with

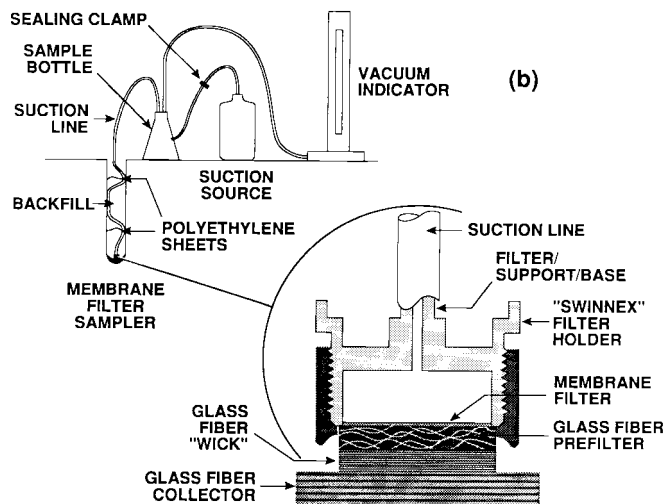
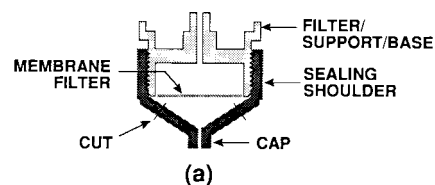


FIG. 12 Membrane Filter Sampler; (a) Preparation of Filter Sampler; and (b) Installation of Filter Sampler

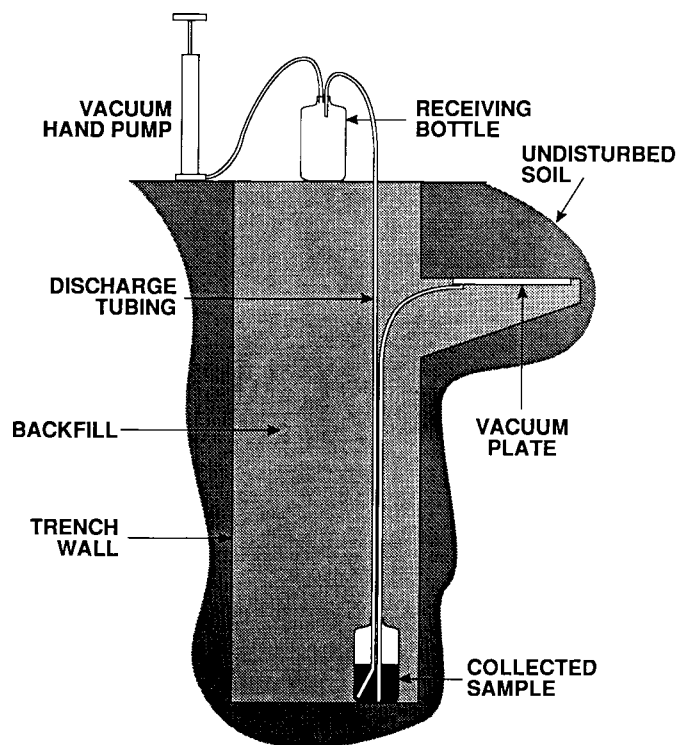


FIG. 13 Vacuum Plate Sampler Installation

distilled water until the specific conductance of the outflowing water is within 2 % of the inflowing water. Debyle et al (50) found (in agreement with 49 and 51) that flushing with HCl



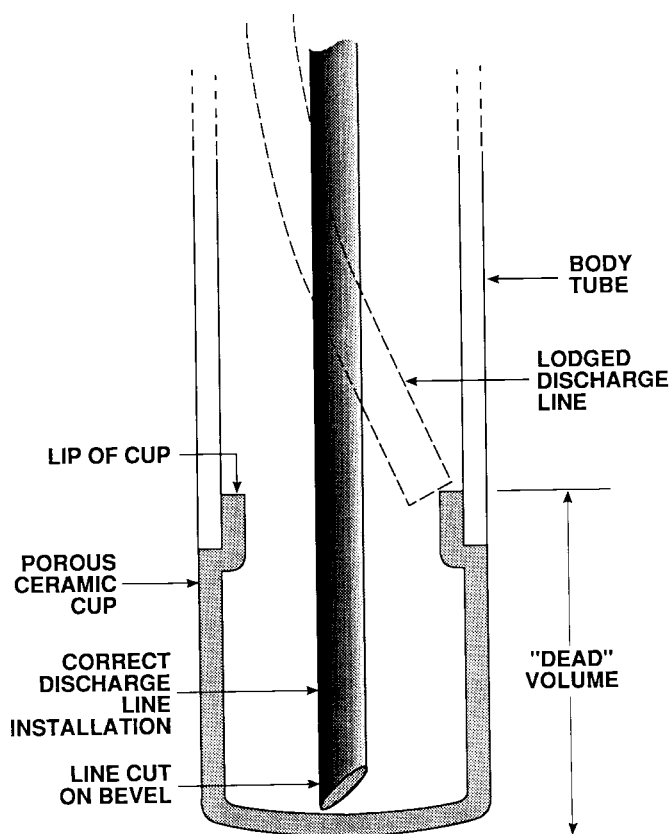


FIG. 14 Location of Potential Dead Volume in Suction Lysimeter

strips cations off of the ceramic. This results in an initial adsorption of cations from pore-liquid onto the ceramic surface. This continues until the cation exchange capacity (CEC) of the ceramic has been satisfied. The effect is not reduced by distilled water flushing after the acid flushing. Therefore, they suggested that the sampler also be flushed, prior to installation, with a solution similar in composition to the expected soil solution. Alternately, the first sample after installation could be discarded (see 7.5.2.1). Bottcher et al (52) attributed increased adsorption of  $\text{PO}_4$  to the acid leaching process. Therefore, they recommended a thorough flushing with a  $\text{PO}_4$  solution of approximately the same concentration as that found in the soil solution, rather than the acid leaching procedure, when sampling for  $\text{PO}_4$ . Peters and Healy (53) used  $\text{H}_2\text{SO}_4$  rather than HCl.

7.4.1.2 Hydrochloric acid may corrode valves within PVC and ceramic high pressure-vacuum lysimeters. Therefore, the porous segment flushing for these designs should be performed prior to attachment if possible. The maximum suction which can be applied is one atmosphere, therefore the flushing process will be slow if suction is used to draw HCl through the porous segment. The flushing can be performed more rapidly if the porous segment is filled with HCl and pressurized to force the acid out of the porous segment since more than one atmosphere of pressure can be applied. This procedure can only be used if the cups are not attached. Care must be taken to prevent overpressurization that might damage the porous section.

7.4.1.3 Corning Laboratories (38<sup>7</sup>) recommended washing fritted glass with hot HCl followed by a distilled water rinse.

Cleaning procedures for Alundum have not been reported, although an acid and water rinse procedure similar to that for ceramic would appear to be appropriate (1). Timco (25<sup>7</sup>) described cleaning procedures for PTFE. The method includes passing 0.5 L of distilled water through the material. An I.P.A. bath followed by another in hydrogen peroxide or rinsing with HCl followed by a distilled water rinse.

7.4.1.4 The use of HCl to wash/flush porous segments of lysimeters, that are to be used in sanitary landfills, may cause water quality interpretation problems. Sanitary landfills are notorious generators of methane gas. Reaction of methane with free chloride ion may result in the generation of di- and trichloromethane (also known as methylene chloride and chloroform). Because of the small liquid volumes in lysimeters and the sensitivity of EPA methods (including 601), false positives for one or both of these constituents may occur.

7.4.1.5 Stevenson (35) recommended treating cellulose-acetate hollow-fibers with silver nitrate and sodium chloride to prevent biofilm growths. Morrison (1) suggested rinsing membrane filters with distilled water.

7.4.1.6 The porous section and fittings of individual samplers may have defects that could cause air entry during sampling. Therefore, prior to taking samplers to the field, each unit should be checked for its bubbling pressure, pressure tested and vacuum tested for leaks. Procedures for these tests are given in U.S. EPA (12) and Timco (25<sup>7</sup>). Washers or "O" rings are used to seal the plugs at the tops of body tubes. However, the accesses for pressure-vacuum and discharge lines passing through these plugs are not sealed. These accesses may leak, and should also be sealed. In the past, lubricants have been used when cutting threads into body tubes, porous segments and fittings. In addition, lubricants have been used in various pressure-vacuum pumps. The user should contact the manufacturer to determine if these lubricants are still used. If present, these lubricants should be removed.

7.4.1.7 After cleaning and testing, samplers should be bagged to prevent contamination during transport to the field. Compatibility of bag material and analytical parameters should be considered. Upon arrival at the installation location, and immediately prior to installation, the porous section should be placed in distilled water for about 30 min to ensure saturation of the porous section (1). Timco (25<sup>7</sup>) indicated that applying a suction of about 50 cbar to a submerged PTFE sampler for about an hour would ensure saturation. Finally, immediately prior to installation, the sampler and associated lines should be assembled and inspected for defects (for example, crimped lines).

#### 7.4.2 Suction Lysimeter and Filter Tip Sampler Installation:

7.4.2.1 Suction lysimeter installation procedures have been described by U.S. EPA (12), Soilmoisture (18<sup>7</sup>), Timco (25<sup>7</sup>), Linden (54), and Rhoades and Oster (55). Filter tip sampler installation procedures were described by Torstensson and Peterson (32).

7.4.2.2 The goals of installation are to ensure good hydraulic contact between the porous segment and the surrounding soil, and to minimize leakage of liquid along the outside of the sampler. U.S. EPA (12) recommended a silica flour/bentonite clay method to achieve these goals for suction lysimeters. A

silica flour layer (installed as a slurry, see 7.4.2.6) placed around the porous segment increases hydraulic contact with the surrounding soil. Screened native backfill is placed above the silica flour, and a bentonite plug above the body tube prevents liquid leakage down the installation hole and along the body tube (see Fig. 15 and Fig. 16). Klute (56) indicated that a screened native soil slurry could be used in place of silica flour for shallow installations.

7.4.2.3 Samplers may be installed in the sidewall of an excavation or, for deeper applications, in a borehole preferably advanced with a hollow stem auger (12). U.S. EPA (12) suggested that suction lysimeters should be installed at an angle of 30 to 45° from vertical whenever possible. This ensures that an undisturbed column of soil is retained above the porous cup. Accordingly, pore liquid samples will reflect flow through pore sequences that have not been disturbed by sampler installation. This angular placement also improves the sampler's ability to collect macropore flow. When installed in the sidewall of a trench, the angled emplacement is simple (see Fig. 15). However, when installed in a borehole, angular emplacement entails angled drilling. Where soils permit, filter tip samplers can be installed by pushing the filter tip into the ground by applying a static load to the extension pipe (32).

7.4.2.4 When suction lysimeters are installed in a borehole advanced by a drill rig, the hole is usually advanced 15 to 20 cm below the desired location of the porous section. Morrison and Szecsody (30) found that the radius of sampling influence is maximized if the borehole diameter is only slightly larger than that of the sampler and if silica flour pack is used. U.S. EPA (12) recommended that the hole have a diameter at least 5 cm larger than the sampler. Timco (25<sup>7</sup>) recommended that the hole have a diameter at least 8 cm greater than that of the sampler to facilitate installation of the silica flour.

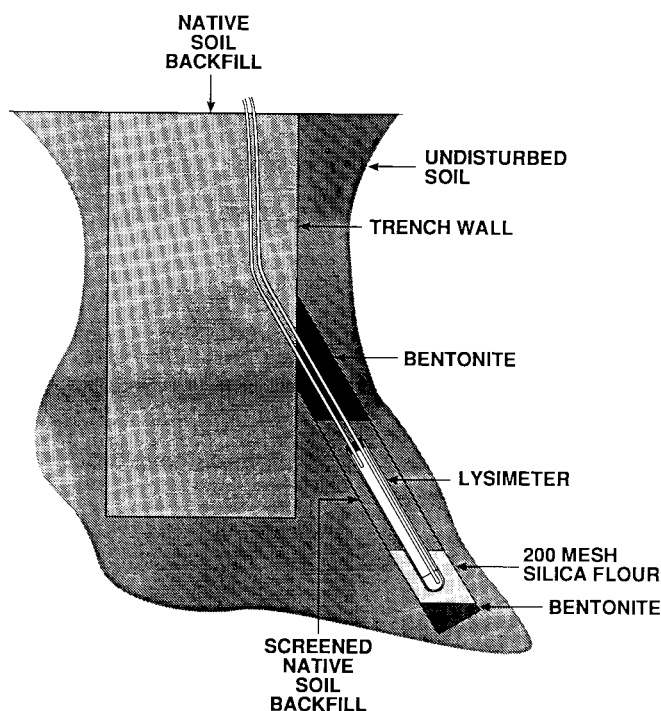


FIG. 15 Pressure-Vacuum Lysimeter Installation in the Sidewall of a Trench

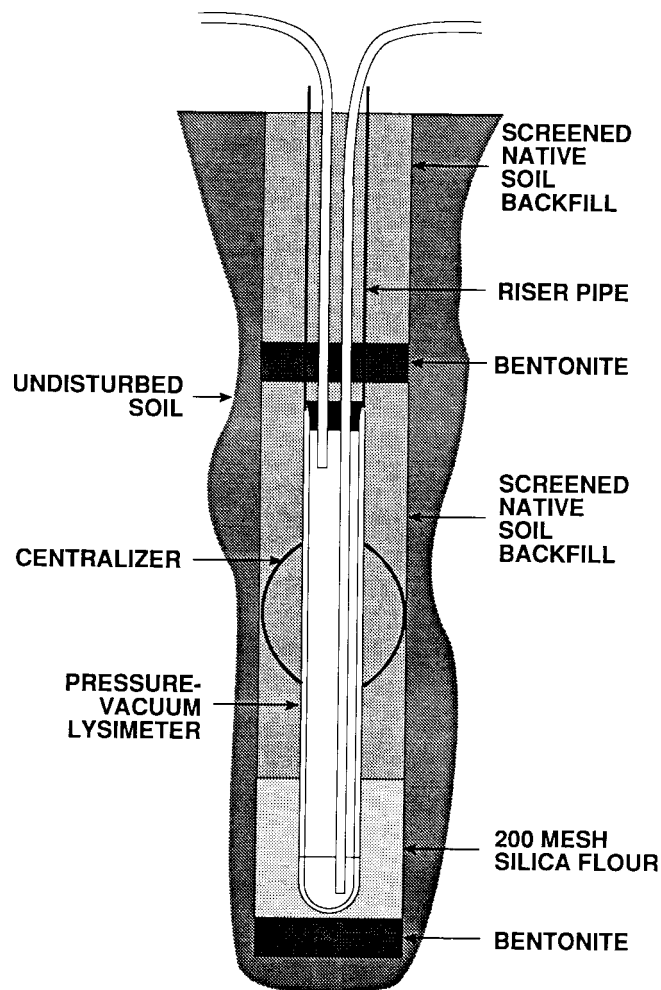


FIG. 16 Pressure-Vacuum Lysimeter Installation in a Borehole

7.4.2.5 Suction lysimeters are preferably lowered into place attached to risers. These protect the lines and ensure exact placement at the desired depth. Centralizers are often used to center the sampler in the hole. Suction lysimeters float in the silica flour that is installed as a slurry. Therefore, the samplers should be installed full of distilled water or held in place by rigid risers.

7.4.2.6 The silica flour slurry (for example 200 to 75  $\mu\text{m}$  mesh opening, silica to distilled water ratio of 0.45 kg to 150 mL) is usually installed using the tremie method (side discharge). Alternately, Brose et al (57) described a method for freezing the silica slurry around the sampler prior to placement. The sampler and frozen pack are then lowered to the sampling location in the borehole. They cited advantages of this technique as including ensurance of proper sampler placement in the flour pack and elimination of pack contamination by soils which slough down the borehole. U.S. EPA (12) recommended filling the borehole to about 30 cm above the suction lysimeter body with the silica. In addition, it was recommended that the powdered bentonite plug placed on top of the silica be about 15 cm thick. The bentonite is also sometimes installed as a slurry, being allowed to hydrate before emplacement. Mixing the bentonite with fine sand at a 1 to 9 ratio, respectively, reduces the potential for shrinking and swelling inherent with pure

bentonite (1). The excavated soil should be backfilled above the bentonite in the order in which it was withdrawn. An effort to compact the soil to its original bulk density should be made. When more than one suction lysimeter is installed in one borehole, these procedures are repeated at the various desired sampling depths (see Fig. 17). Care must be taken with these installations to ensure that lines from lower samplers do not interfere with the hydraulic contact of shallower samplers. Designs are available to avert these problems (25<sup>7</sup>).

7.4.2.7 U.S. EPA (12) recommended removal of the water within the sampler and silica slurry after installation. Litaor (58) recommended installation of samplers a year before sampling is to begin, in order to allow them to equilibrate with the surrounding soil. The lines at the surface should be labeled, clamped and housed in locked containers such as valve boxes or casing (1). Methods for cutting and splicing tubing may be found in Timco (25<sup>7</sup>). The user should be careful when using clamps and tubing provided by different manufacturers, inappropriate clamps may damage tubing. Clamps must be restricted to permanently flexible tubing otherwise stopcocks should be used.

#### 7.4.3 Experimental Suction Sampler Installation:

##### 7.4.3.1 Cellulose-acetate hollow-fiber sampler installation

procedures were described by Everett et al (9). Membrane filter sampler installation procedures were described by Stevenson (35), Everett et al (9), and Morrison (1). Vacuum plate sampler installation procedures were described by Everett et al (9) and Morrison (1).

7.4.3.2 Cellulose-acetate hollow-fiber samplers have been used almost exclusively in laboratory studies (34). Because the samplers operate on the same principles as vacuum lysimeters, the goals and concerns of installation are similar. Good hydraulic contact between the hollow-fiber and the soil is critical. However, the fibers are too thin and fragile to be pushed into place. Therefore, the fibers must be placed in a predrilled hole (vertical or horizontal). Silkworth and Grigal (59) installed these samplers within a length of perforated, protective PVC tubing filled with soil slurry.

7.4.3.3 Membrane filter samplers are placed in a hole dug to the top of the selected sampling depth. First, sheets of the glass fiber "collectors" are placed at the bottom of the hole. These develop the necessary hydraulic contact between the sampler and the soil. In addition, the "collectors" extend the area of sampling as they cover a larger area than the filter holder alone. Second, two or three smaller glass fiber "wick" discs that fit within the filter holder are placed on the "collectors." Third, the filter holder fitted with a glass fiber prefilter and the membrane filter is placed on top of the "wick" discs. The suction line leads to the surface. Finally, the hole is backfilled (1, 9).

7.4.3.4 Vacuum plate lysimeters are normally installed on the ceiling of a cavity cut into the side of a trench. In order to obtain the necessary contact between the porous plate and the soil, pneumatic bladders, inner tubes, or similar devices are placed beneath the sampler and are used to force it against the cavity ceiling (1). The cavity ceiling is not a smooth surface. Therefore, a layer of silica flour between the plate and the soil is sometimes used to enhance hydraulic contact.

#### 7.4.4 Maintenance:

7.4.4.1 The major causes of sampler failure are line damage and leaks (caused by freezing, installation, rodents, etc.), connection leaks, and clogging of the porous material. Freeze damage to the lines can be minimized if the lines are emptied of sample prior to applying a vacuum. Care needs to be taken that the tubing line closure devices are freeze proof.

7.4.4.2 The possibility of line and connection leaks is minimized by rigorously sealing and pressure testing all connections and lines before installation. A common precaution to assist in repairing surface damage to lines is to store excess line below the surface (within the riser pipe when used) when backfilling the borehole. In the event of severed lines, an excavation to this buried length allows restoration of an operational system (1). Lines should be clamped shut when not in use to prevent foreign objects or insects from entering them. The lines should be protected from weather, sunlight exposure, and vandalism with a locked housing. The use of riser pipe around the sampler lines prevents punctures by backfill materials and prevents rodents from damaging the lines.

7.4.4.3 When shallow samplers are used, the ground surface above the sampler should be maintained in a fairly representative state. Large line housings and excessive traffic around the sampler (causing compaction of the soil) will reduce the

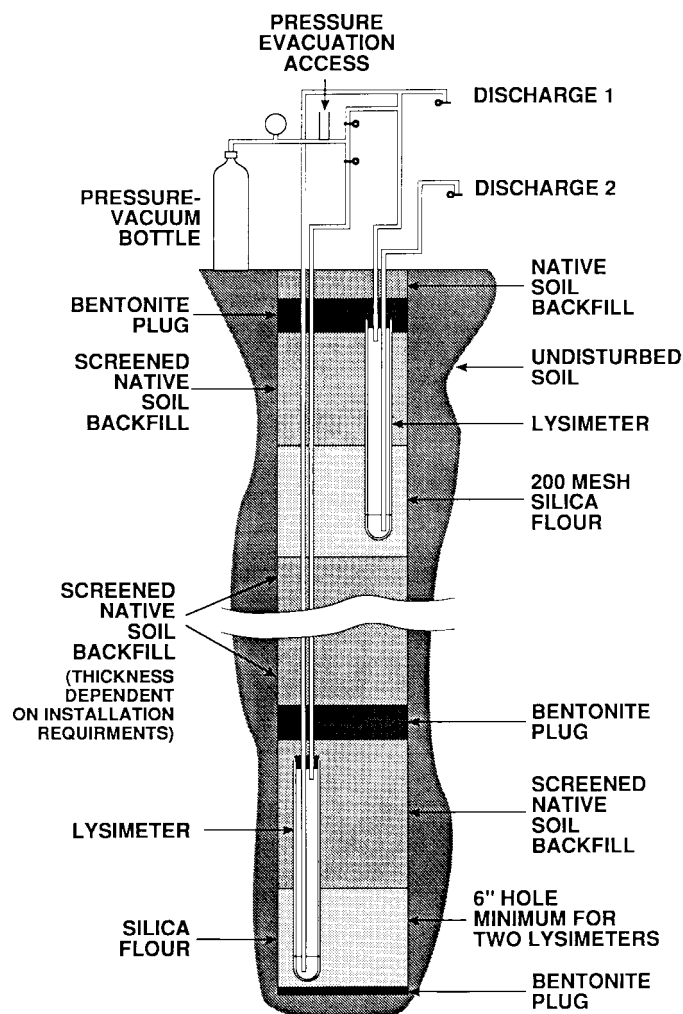


FIG. 17 Multiple Pressure-Vacuum Lysimeter Installations in a Borehole



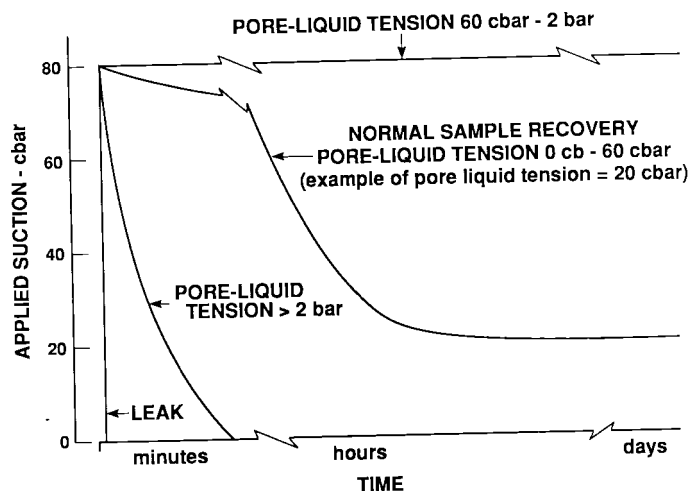
amount of infiltration in that area. This will affect the representativeness of the pore-liquid samples. Methods to avoid these effects include angled installations, and remote operation of sampler lines.

7.4.4.4 Porous sections may clog as a function of soil composition, type of porous section material, biofilm growth, suction application, and pore-liquid content (1, 17, 20, 50). However, porous section clogging appears to be less severe than once thought (12, 17). Soils and the 200 mesh silica flour filter out fine materials before they reach the porous section (60, 61, 62). Clogging can be further reduced by periodically filling the sampler with distilled water and allowing it to drain out of the sampler. Debyle et al (50) suggested removing shallow samplers on a seasonal basis for flushing with HCl and distilled water. This process restores samplers to their original operational and chemical states. A “clogged” lysimeter may be cleaned out by filling the lysimeter with pure water and applying a pressure of 5 psi for 30 min. However, reinstallation at the same location and depth does not guarantee resumption of sampling from the same soil volume.

7.4.4.5 Often no sample is retrieved during a sampling attempt. The first check should be a continuity test of the lines and connections. This test can be done by applying a gentle pressure or vacuum to the V/P line and detecting air movement from the open sample line. This could be due to sampler failure or high pore-liquid tensions. Because of this, it is prudent to install a tensiometer near the sampler at a similar depth. The tensiometer that measures pore-liquid tensions allows the operator to determine if failure to obtain a sample is due to high pore-liquid tensions or due to sampler damage. The tensiometer can also be used to gage the effect of sampling on local pore-liquid flow regimes. Pore-liquid tension should be determined as an initial condition during lysimeter installation.

7.4.4.6 If a tensiometer is not available to measure pore-liquid tensions, the lysimeter can be tested to help determine reasons for failure to recover a sample. The sampler is tested by applying a suction of 80 cbar, and monitoring the decay of suction with time. Fig. 18 depicts the various types of suction decay that might be found in a suction lysimeter with a 200 cbar bubbling pressure ceramic section. An almost instantaneous decay of suction is associated with lysimeter leakage. A suction decay over a period of minutes is associated with pore-liquid tensions greater than 200 cbar. Under these conditions, the porous section is desaturated and air enters the sampler. A suction decay over a period of hours reflects normal sample collection. This suggests that failure to retrieve a sample is related to damage of the sample retrieval system (for example, discharge line damage). When suction does not decay, or does so over a period of days, the pressure-vacuum line may be clogged or pore-liquid tensions may be greater than 60 cbar (but less than 200 cbar) causing liquid inflow rates that are too low for sample collection.

7.4.4.7 Morrison and Szecsody (63) described devices that could be used as tensiometers and then converted to pressure-vacuum lysimeters. However, they found that gases entering the devices prevented accurate measurement of pore-liquid tensions. Baier et al (64) described methods for converting tensiometers to pressure-vacuum lysimeters. It would also



NOTE 1—Also shown is the almost instantaneous decay associated with an appreciable leak in the instrument.

**FIG. 18 Decay Characteristics of Suction Applied to a Two Bar (Bubbling Pressure) Ceramic Cup Lysimeter in Equilibrium With Soils in Varying Ranges of Pore-Liquid Tension**

appear reasonable to convert suction lysimeters to tensiometers. However, Taylor and Ashcroft (65) found that the volume of water drawn from a converted lysimeter into the surrounding soil would significantly affect natural pore-liquid tensions. In addition, they found that the larger porous section of a lysimeter would cause more diffusion of dissolved air into the device, and that the time constant for measurement would be significantly increased over that of a tensiometer. Filter tip samplers can be converted to tensiometers with pressure transducers (32).

7.4.4.8 Operational lifetimes of suction samplers are dependent on installation, subsurface conditions, maintenance, and sampling frequency. Some samplers have been reported to be operational for as long as 25 years (64).

#### 7.4.5 Comments:

7.4.5.1 Vacuum lysimeters and experimental samplers use suction to retrieve samples. Therefore, the maximum sampling depth is limited by the maximum suction lift of water (about 7.5 m) (12). In practice, these samplers are generally used to about 2 m below the surface (12). They are primarily used to monitor near-surface movement of pollutants such as those from land disposal facilities and those from irrigation return flow.

7.4.5.2 Pressure-vacuum lysimeters are generally not used at depths below about 15 m. At greater depths, sample loss and overpressurization problems are considered significant enough to warrant the use of high pressure-vacuum lysimeters that do not have these limitations. High pressure-vacuum lysimeters are not preferred at the shallower depths because they are more expensive. In addition, high pressure-vacuum units have more moving parts than pressure-vacuum units, and as a result, the possibility of failure for the former is higher.

7.4.5.3 As discussed in 7.6, two problems with suction samplers are that they may not sample from macropores (under unsaturated conditions; unless the macropores are directly intercepted) and that their results cannot be used in quantitative

mass balance studies. Hornby et al (66) described an installation that could be used to surmount these problems. A barrel-sized casing (for example, 57 cm outside diameter by 85.7 cm high) is placed in a support device and gently pushed into the soil with a backhoe. As the casing is pushed, soil is excavated around it to help with insertion. The process results in an encased monolith of undisturbed soil. The monolith is then rotated and lifted, pressure-vacuum lysimeters are placed in its base, and the bottom is sealed. Subsequently the assembly is placed back into the ground at the monitoring site (see Fig. 19). All fluid draining through the monolith is collected by the samplers. Inasmuch as the boundaries of the system are sealed, the flux of liquid through the system requires maintaining a vertical hydraulic gradient by applying continual suction to the samplers.

#### 7.5 Operation:

##### 7.5.1 Methods:

7.5.1.1 *Vacuum Lysimeters*—Sampling methods are described by the U.S. EPA (12), by Soilmoisture (18<sup>7</sup>) and by Timco (25<sup>7</sup>). To collect a sample, suction is applied to the sampler, and the suction line is clamped shut. After sample has collected in the body tube, it is retrieved through a discharge line extending to the base of the porous cup. In shallow installations, with the body tube extending above the soil surface, the discharge line is sometimes inserted and removed as needed. For deeper installations, the discharge line is permanently installed. At the surface, the line is connected to a sample collection flask. Suction is applied to the flask, and liquid is pulled from the sampler, up the discharge line, and into the collection flask. Cole (42) constructed an array of samplers that were attached to a vacuum tank connected to an

electric power source. This system allowed remote operation at a constant suction. Wengel and Griffen (67) described methods by which samplers can be connected to a central control board and operated remotely. Brown et al (68) employed a solar panel to power a similar setup. Chow (44) described a sampler that shuts off automatically when the desired sample volume has been collected.

7.5.1.2 *Pressure-Vacuum Lysimeters*—Sampling methods are described in U.S. EPA (12), by Soilmoisture (18<sup>7</sup>) and by Timco (25<sup>7</sup>). To sample, suction is applied to the system via the pressure-vacuum line. The discharge line to the sample bottle is clamped shut during this time. When sufficient time has been allowed for the unit to fill with pore-liquid, suction is released and the clamp on the discharge line is opened. Gas pressure (for example, air or nitrogen; see 7.6.2) is then applied through the pressure-vacuum line. This forces the sample through the discharge line and into the collection flask at the surface (12). A variety of systems have been developed by which the pressure, suction, and sample volume can be controlled remotely or manually (44, 49, 67, 69).

7.5.1.3 *High Pressure-Vacuum Lysimeters (Lysimeters With a Transfer Vessel)*—Sampling methods may be found in U.S. EPA (12), in Soilmoisture (18<sup>7</sup>) and in Timco (25<sup>7</sup>). When suction is applied to the system, it extends to the porous section through an open, one-way check valve at the bottom of the transfer vessel or chamber. A second one-way check valve in the discharge line is closed during this time. As soil solution enters the sampler it is pulled by the suction into the transfer vessel or chamber through a line attached to the open valve at its base. The sample is brought to the surface by releasing the suction and applying pressure (for example, air or nitrogen) through the pressure-vacuum line. This shuts the one-way valve to the porous segment and opens the one-way valve in the discharge line. The sample is then pushed to the surface (12). A variety of systems have been developed to control pressure, suction and sample volume remotely or manually (44, 49, 67, 69).

7.5.1.4 *Suction Lysimeters With Low Bubbling Pressures (Samplers With PTFE Porous Sections)*—Sampling methods for this group of samplers are a bit different than those for the three designs described in 7.5.1.1, 7.5.1.2, and 7.5.1.3. This system is designed to allow the soil pore water extraction process to occur separately from the movement of the collected pore water to the transfer vessel. The only difference is that maximum sampling suctions for these units are much lower (see 7.6.1.3).

7.5.1.5 *Filter Tip Samplers*—Sampling methods may be found in Torstensson and Petsonk (32). Samples are collected by first evacuating the sample vial. The vial is then inserted in the sampling adaptor that contains a two way hypodermic needle. The adaptor is then lowered down the extension pipe. When the adaptor connects with the nozzle of the filter tip, the needle penetrates the septa in the vial and in the filter tip. Sample then flows through the porous segment and into the sample vial due to the negative pressure in the vial. As sample is collected, the negative pressure in the vial falls towards that of the pore-liquid tension. When these negative pressures are equal, sampling ends and the sample vial is retrieved. The

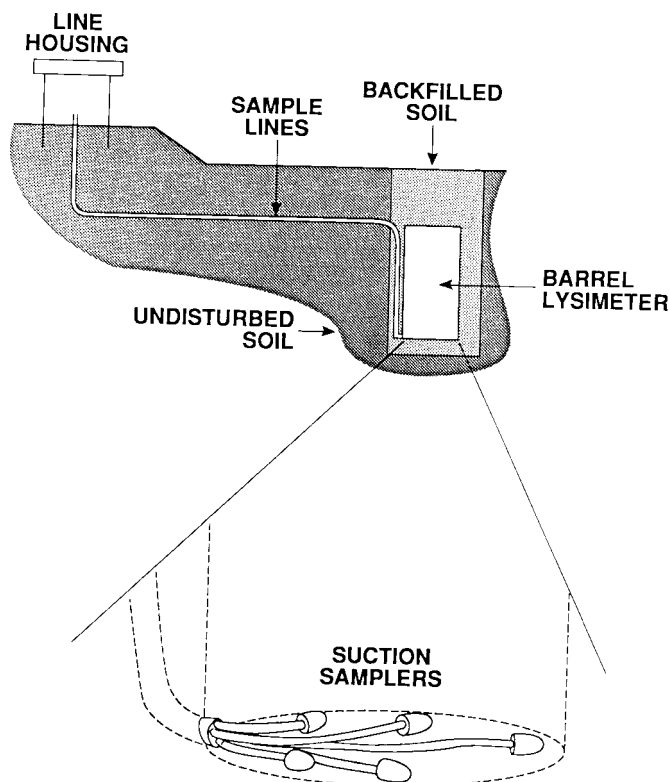


FIG. 19 Barrel Lysimeter



standard sample volume is about 35 mL. However, by connecting several vials in series, sample volumes of up to 500 mL can be obtained.

**7.5.1.6 Experimental Suction Samplers**—Cellulose-acetate hollow-fiber samplers, membrane filter samplers, and vacuum plate samplers are operated using the same general technique as for vacuum lysimeters. Jackson et al (33) sampled from soil columns using cellulose-acetate hollow-fiber samplers subjected to a constant suction of 81 cbar. At this suction, they were able to extract samples for chemical analyses from silty loams with moisture contents ranging from 20 to 50 %. Silkworth and Grigal (59) compared the performance of these samplers to suction lysimeters. They found that cellulose-acetate hollow-fiber samplers fail more often than suction lysimeters. In membrane filter samplers, the “collectors” provide hydraulic contact between the soil and the samplers. Liquid is drawn by capillarity into the “collectors.” When suction is applied, liquid flows through the “wick,” the prefilter, and finally the membrane filter. The prefilter reduces clogging of the membrane filter by fine soil materials (9). Stevenson (35) recommended using a suction of between 50 and 60 cbar when sampling with membrane filter samplers. A variety of constant suction methods for sampling with vacuum plates are described by Morrison (1). An advantage of the larger plates is that they have large contact areas with the soil. Therefore, larger sample volumes can be collected in shorter times than with vacuum lysimeters which have porous sections with smaller surface areas.

#### 7.5.2 Comments:

**7.5.2.1** Nagpal (70) recommended several consecutive extractions of liquids during a sampling event and use of only the last one for chemical analyses. The purpose of this is to flush out cross contaminants from previous sampling periods, and to ensure that any porous segment/soil solution interactions have reached equilibrium. Debye et al (50) also suggested discarding the first one or two sample volumes when sampling dilute solutions with newly flushed (HCL method) and installed samplers. The purpose of this is to allow cation exchange between the porous segment and the pore-liquid (caused by the HCL flushing) to equilibrate.

**7.5.2.2** Factors which affect the volume and source of a pore-liquid sample include the amount of suction applied, the schedule of suction application, the pore-liquid content, the distribution of pore-liquid, the soil grain size distribution, the soil structure, the porous section design, and the porous section age.

**7.5.2.3** Samples collected with lower suctions (about 10 cbar or less) tend to come from liquids migrating through soil macropores (1). Samples collected with higher suctions (greater than about 10 cbar) also include fluids held at higher tensions in micropores. The sampler may disrupt normal flow patterns due to the applied suctions. The effects may extend several meters from the sampler although the area nearest the sampler is most disturbed (71, 72, 73). This disturbance causes samples to be averages of the affected flow area rather than point samples (1). Warrick and Amoozegar-Fard (72) developed an approximate analytical equation which can be used to estimate the maximum radius of influence on the flow regime

by a suction sampler. Narasimhan and Driess (74) developed a numerical technique to simulate the effects of suction samplers on the pore-liquid regime.

**7.5.2.4** Sampling with falling suction produces samples with compositions that are “averages” of the liquids held at the range of tensions applied. Because suctions and therefore inflow rates decrease with time, these “averages” are weighted toward those portions of the samples obtained in early times. Samples collected over prolonged periods (due to slow inflow rates) are “averages” of the liquids fluxing past the sampling region during those times.

**7.5.2.5** During wet periods, samplers affect a small volume of soil and pull liquids from a sequence of pores that may include macropores. During dry periods samplers affect a larger volume of soil, draw from micropores because the macropores have been drained, and collect less liquid (75, 76). The net result of this is that sampled soil solutions are “averaged” over different volumes and derived from different pores as a function of the soil moisture content and distribution.

**7.5.2.6** Soil textures and pore-liquid tensions control the amount of liquid that can be removed by a sampler and its radius of influence. The slope on the pore-liquid release curve for a sand is greater than that for a clay at low pore-liquid tensions (see Fig. 20). This indicates that there will be a larger quantity of pore-liquid released from a sand than from a clay for an equal change of pore-liquid tension at these low tensions. At higher tensions, the slope of a clay pore-liquid release curve is greater than that for a sand (see Fig. 20). This

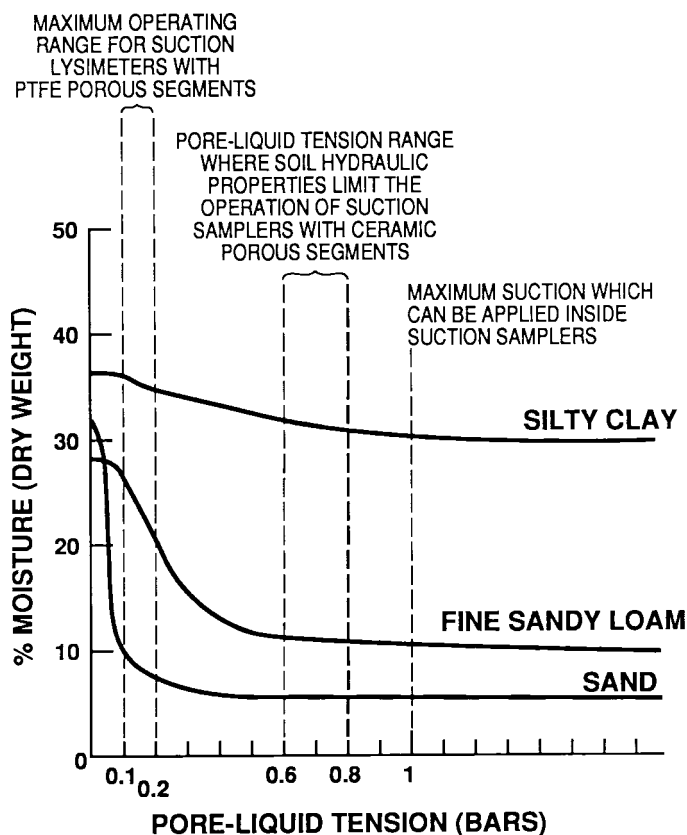


FIG. 20 Water Release Curves for Three Soils, Showing Operating Conditions for Suction Samplers

indicates that more pore-liquid will be released from a clay than from a sand for an equal change in pore-liquid tension at the higher tensions. A consequence of this is that suction samplers may not obtain samples from coarse grained soils at higher pore-liquid tensions. Morrison and Szecsody (30) found that (under the conditions of their study) radii of influence for suction lysimeters ranged from 10 cm in coarse soils up to 92 cm in fine grained soils.

7.5.2.7 Hansen and Harris (30), demonstrated that intake rates may vary substantially due to variability in the ceramic sections from one manufacturer's batch to another. As discussed in 7.4, the intake rate of a sampler is also a function of the degree of clogging. As discussed in 7.6, the range of pore-liquid tensions over which a sampler can operate is a direct function of the maximum pore size of the porous section and the surrounding silica flour pack. Finally, Morrison and Szecsody (30) found that the radius of influence of a sampler increases with the diameter of the porous section.

7.5.2.8 Because of these factors the following recommendations have been made for sampling with suction lysimeters. Hansen and Harris (19), suggested using uniform initial suctions, short sampling intervals, and uniform sampling times for different sampling events and locations to increase the uniformity of samples. Debye et al (50) also recommended sampling with uniform suctions that do not significantly exceed the tension at which the percolating soil solution is being held. U.S. EPA (12) suggested sampling after infiltration events such as rain storms, spring melts, or irrigations as these periods of high pore-liquid content are accompanied by higher pore-liquid flow rates and contaminant transport. For sampling these events, it is useful to install samplers at interfaces between coarse and fine materials to take advantage of any liquid perching which might occur. Silkworth and Grigal (59) recommended using samplers with large diameter ceramic porous sections (as opposed to small diameter ceramic samplers, or hollow cellulose fiber samplers) since they showed less of a tendency to alter the pore-liquid, they had lower failure rates, and they collected larger sample volumes. These recommendations were reinforced by van der Ploeg and Beese (73) who concluded that samplers with large cross sectional area porous sections used with low extraction rates (suctions approaching those of the pore-liquid tensions) reduce the effects of sampling on compositions of samples. Finally, U.S. EPA (12) recommended that porous section material types be carefully chosen based on pore-liquid tensions expected in the sampling area. Operational ranges of various porous section types are discussed in 7.6 and are presented in Table 1.

#### 7.6 Limitations:

##### 7.6.1 Physical Limitations:

7.6.1.1 The most severe constraint on the operation of suction samplers involves soil around the porous sections becoming so dry (and pore-liquid tensions so high) that samples cannot be collected. The limiting factors in these conditions will be the porous segment or the soil hydraulic properties. For porous segments with bubbling pressures less than 60 cbar (for example, PTFE), the porous segment will be the limiting factor because the high suction required to move liquids into the samplers will cause menisci in the porous

segments to break down and air to enter. Soil hydraulic properties will be the limiting factors for porous segments with bubbling pressures greater than 60 cbar (for example, ceramics) because unsaturated hydraulic conductivity of the soil and pressures gradients across the porous segments will be so low (due to high pore-liquid tensions) that flow into the samplers will be negligible.

7.6.1.2 The maximum suction that the saturated porous section of a sampler can withstand before air enters is a function of the pore configuration and size, and its hydrophilicity/hydrophobicity (see Appendix X1 and (65)). The following variation of the capillary rise equation (8, 12, 18, 77) combines these factors:

$$P_b = \frac{-2\delta \cos\alpha}{r} \quad (1)$$

where:

$P_b$  = bubbling pressure (gage), units –  $FLT^{-2}$ ,

$\delta$  = surface tension between pore-liquid and air, units –  $FT^{-1}$ ,

$\alpha$  = contact angle between the liquid and the material of the porous segment,  $D$ , and

$r$  = maximum pore radius of the pore segment, units –  $L$ .

This equation shows that the bubbling pressure decreases with increasing contact angle and with increasing maximum pore radius. The maximum sampling suction that can be applied is 100 cbar (1 atmosphere). For a hydrophilic material (that has an acute contact angle) the maximum pore size that will allow the application of 100 cbar of suction is 2.8  $\mu m$ . For a hydrophobic material (that has an obtuse contact angle) a smaller pore size will be required (65). The maximum pore sizes of presently available ceramics (that are hydrophilic) used for suction lysimeters and filter tip samplers vary from 1.2 to 3  $\mu m$  (as measured by the bubbling pressures) (18<sup>7</sup>, 45, 78<sup>7</sup>). The maximum pore sizes of cellulose-acetate hollow-fibers and membrane filters range from less than 2.8  $\mu m$  and 0.4 to 5.0  $\mu m$  respectively (1, 35, 36). These pore sizes result in maximum sampling suctions near 100 cbar. Therefore, these materials will not allow air to enter during sampling, and the limiting factors will be the soil hydraulic properties. The combination of soil limiting effects result in negligible sampling rates when pore-liquid tensions are above 60 cbar (for coarse grained soils) to 80 cbar (for finer grained soils) (45). At tensions above these levels, inflow rates are too low to allow sampling.

7.6.1.3 The maximum pore sizes of presently available porous PTFE segments for suction lysimeters range from about 15 to 30  $\mu m$  (calculated from bubbling pressures) (25<sup>7</sup>). These pore sizes allow maximum sampling suctions of about 10 to 21 cbar (25). The hydrophobicity of PTFE will further reduce the magnitude of the maximum sampling suction. Applied suctions of greater than 10 to 21 cbar (or less) will cause air to enter, and sampling to cease. Because a suction greater than 10 to 21 cbar cannot be applied to these samplers, pore-liquids held at tensions greater than 10 to 21 cbar cannot be sampled with these devices. Because of this, PTFE will be the limiting factor when it is used for the porous segment. A consequence of the small suction range available to PTFE porous sections is that

only very moist soils approaching saturation may be sampled (17).

7.6.1.4 The silica flour pack, that has smaller pore sizes than PTFE, can act as an extension of the porous segment, and may extend the range of suctions that can be applied to the sampler. Everett and McMillion (45) found that the pack extended the suction range of earlier, larger pore size PTFE (70 to 90  $\mu\text{m}$ ) from less than 4 to 7 cbar. Timco (25<sup>7</sup>) suggested that the operational range of the presently available PTFE samplers (15 to 30  $\mu\text{m}$ ) can be extended from 10 to 21 cbar to between 61 to 71 cbar when “properly” installed within a silica flour pack (this has not been verified in peer reviewed literature). For this to be true, the silica flour pack must be able to remain saturated over the range of applied suctions. However, the results of Everett and McMillion (45) suggest that the bubbling pressure of the silica flour pack is only 7 cbar. Trainor (27) found that even if these samplers are “properly” installed, air may still enter if applied suctions exceed pore-liquid tensions by more than 30 %. Pore-liquid tensions are not always known, and technicians may not carefully control applied suctions. In addition, pressurization of pressure-vacuum lysimeters for sample retrieval appears to damage the silica flour pack (27, 28). Thus, dependency on the silica flour pack to provide the needed suction range is an extremely limited option. Because of this, suction lysimeters with PTFE porous sections are limited to near saturated sampling and have been classified separately (see Fig. 20).

7.6.1.5 Samples can be collected (using ceramic porous sections) from clays with high pore-liquid tensions (approaching 60 to 80 cbar). However, because liquid inflow rates are low at higher tensions, the amount of time required to collect sufficient sample volumes may exceed the maximum allowable holding time for many chemical analyses. Law (76) pointed out that when soils have liquid contents that allow little or no sample collection (high pore-liquid tensions), there is little or no liquid movement in the soil. Consequently, there will be little or no contaminant migration. If samples of pore-liquids held at tensions above 60 to 80 cbar are desired, soil core sampling with subsequent laboratory liquid extraction may be used (76). However, Law (76), and Brown (79) concluded that results from the two sampling methods are not comparable. Liquid from soil core samples will include constituents that are held at tensions greater than 60 to 80 cbar and that would not be picked up by suction samplers. Because of this and because samples removed from soil cores may include some of the constituents from the soil itself (for example, cations preferentially adsorbed in electrical double layers) or sorbed organics, Law (76) concluded that soil cores are more conservative estimators of cation contaminant presence in soil. Brown (79) concluded that organic contaminant concentrations derived from soil cores and pore-liquid samplers are not comparable because of preferential sorption of some compounds. Amter (80) developed an alternative to extraction of samples from soil cores. The method involves injecting a chemically blank fluid through an existing lysimeter. After a time, the fluid (now containing dilute pore-liquid) is recovered through the sampler and analyzed. The results, although qualitative, were shown to

correlate well with known relative pore-liquid constituent concentrations.

7.6.1.6 Suction samplers may not intercept macropores because of the small size of their porous sections. Because of this, they may miss the majority of flow at high moisture contents in structured soils (81). The ability to intercept this flow can be enhanced by installing the samplers in large diameter silica flour packs. However, this involves drilling larger holes. Because suction samplers only sample when suction is applied, they may miss infiltration events unless a constant suction is applied. Therefore, under conditions of high moisture content in structured soils, free drainage samplers are recommended (see Section 8) (81). Pore-liquid composition changes with time. Because suction samplers sample over an extended period (especially in drier soils), the resulting sample should be considered an average of the total flux past the sampler during the sampling interval.

7.6.1.7 A major factor limiting the operation of shallow suction samplers in cold climates is that pore-liquid may freeze near the porous segments. In addition, liquid may freeze within porous segments and lines, preventing sample retrieval and perhaps fracturing the sampler during ice expansion. Because of this, lines should be emptied before the onset of cold weather. Additionally, some soils tend to heave during freezing and thawing. Consequently, the samplers may be displaced in the soil profile, resulting in a break of hydraulic contact (12).

#### 7.6.2 Chemical Limitations:

7.6.2.1 The inherent heterogeneities of unsaturated pore-liquid movement and chemistry limit the degree to which samples collected with suction samplers can be considered representative. This is because the small cross sectional areas of suction samplers may not adequately integrate for spatial variability in liquid movement rates and chemistry (51, 82, 83). Biggar and Nielsen (84) suggested that results of analyses from suction lysimeter samples are good for qualitative comparisons, but that they cannot be used for quantitative analysis unless the variabilities of the parameters involved are established. Law (76) came to similar conclusions, stating that results from suction lysimeter sampling could not be used for quantitative mass balance studies.

7.6.2.2 Well structured soils have two distinct flow regions including macropores (for example, cracks, burrows, and root traces) and micropores. Under saturated conditions, liquids move more rapidly through macropores than through micropores. Because of this, the movement of liquid-borne pollutants into the finer pores may be limited. Consequently, pore-liquids in macropores may have different chemistries than those in micropores (85). This is enhanced by the fact that oxygen content in macropores can change in a matter of hours during an infiltration event, whereas micropores may remain suboxic regardless of flow conditions (75). In addition, micropores are less susceptible to leaching than macropores (1, 86, 87, 88). Because of these differences, sample chemistry can vary widely from location to location and from time to time depending on the amount of liquid drawn from these two flow systems.

7.6.2.3 Suction samplers may affect pore-liquid chemistry as it is being sampled. The major sampler related factors that

can affect the sample chemistry are the porous segment material and sample storage time. The degree of chemical interaction may also be affected by the amount of porous section clogging **(1)**. Clogging slows the rate of flow through

the porous section so that contact time and chances for chemical interaction are increased **(50)**. In addition, the types of adhesives used to attach porous segments (for example, epoxy) may alter the pore-liquid chemistry.

**TABLE 2 Porous Material Interactions<sup>A,B</sup>**

	Material Sorbs Species	Material Desorbs Species <sup>C,D</sup>	Material Screens Species <sup>C,D</sup>	No Significant Interaction <sup>C,D</sup>	No Interaction
Al	...	C(2) A(14)	...	C(16) A(14)	...
Alkalinity	...	...	...	SF(11)	...
Ca	...	C(1,2,18) CAF(18) A(14)	...	C(3,6,10,11,25) PTFE (3) A(3) FG(18,22) CAF(10)	PTFE(13) ... ... ...
C	...	FG(22)	...	...	...
CO <sub>3</sub>	...	C(2)	...	...	...
HCO <sub>3</sub>	...	C(2)	...	...	...
Cd	C(11)	...	...	C(3) PTFE(3) A(3)	... ... ...
Cl	...	...	...	C(11,25) SF(11)	PTFE(13) ...
Cr	C(19)	C(3) PTFE(3) A(3)	...	...	... ...
Cu	C(11)	C(3) PTFE(3)	...	A(3)	... ...
Fe	C(11)	PTFE(3) A(3)	...	C(3,25) A(14)	PTFE(13) ...
H	...	...	...	SF(11)	...
K	C(5,6,15)	C(18) A(14)	...	C(1,25) CAF(18) <sup>E</sup> FG(18,22)	... ... ...
Mg	C(6)	C(2,3,11,18) A(3,14) CAF(18)	...	C(10,25) PTFE(3) CAF(10) FG(18,22)	PTFE(13) ... ... ...
Mn	C(11)	A(3)	...	C(3) PTFE(3) A(14)	PTFE(13) ... ...
Na	C(6)	C(2,18) A(14) CAF(18) FG(18,22)	...	C(1,11,25)	PTFE(13)
NH <sub>4</sub>	C(4,12)	...	...	PTFE(4)	...
N	...	FG(22)	...	...	...
NO <sub>2</sub>	...	...	...	C(4,5) PTFE(4)	... ...
NO <sub>3</sub>	...	...	...	C(4,8) PTFE(4)	... ...
NO <sub>3</sub> -N	...	...	CAF(10) C(10) CAF(10)	...	... ... ...
(NO <sub>2</sub> +NO <sub>3</sub> )-N	...	...	...	C(5) <sup>F</sup> CAF(18) FG(18)	... ... ...
P	C(1,5,8,15,18)	...	...	PTFE(4) CAF(10) C(10) CAF(10)	... ... ... ...
PO <sub>4</sub>	C(4,5,7)	...	...	...	PTFE(13)
PO <sub>4</sub> -P	...	...	...	...	...
Pb	...	...	...	...	...
SiO <sub>2</sub>	...	C(2)	...	...	...
Si	...	...	...	C(4) PTFE(4) C(11)	... ... ...
SO <sub>4</sub>	...	...	...	...	...
Sr	...	C(11)	...	...	...
Zn	...	C(11)	...	...	PTFE(13)
High Molecular Weight Compounds	...	...	C(17,21) CAF(10)	...	...
4-nitrophenol	PTFE(23)	...	...	...	...
Chlorinated hydrocarbons	PTFE(23,24)	...	...	...	...
Diethyl phthalate	...	...	...	PTFE(23)	...
Naphthalene	PTFE(23)	...	...	...	...
Acenaphthene	PTFE(23)	...	...	...	...



TABLE 2 Continued

References and Notes on Experimental Techniques<sup>G</sup>

Reference Number in Table	Cited Reference	Porous Section was Washed	Results are a Function of Several Factors	Dilute Solutions were Tested	Experiments were Performed on Nonporous Materials
1	Ref (137)	X	...	...	...
2	Ref (48)	X	...	...	...
3	Ref (93)	X	...	...	...
4	Ref (138)	X	...	...	...
5	Ref (139)	...	X	...	...
6	Ref (50)	X	X	...	...
7	Ref (52)	X	...	...	...
8	Ref (19)	X	...	...	...
9	Ref (56)	...	X	...	...
10	Ref (34)	...	...	X	...
11	Ref (53)	X	...	...	...
12	Ref (69)	...	...	...	...
13	Ref (1)	X	...	...	...
14	Ref (47)	X	X	X	...
15	Ref (139)	X	...	...	...
16	Ref (58)	...	X	...	...
17	Ref (76)	...	...	...	...
18	Ref (59)	X	X	...	...
19	Ref (75)	...	X	...	...
20	Ref (140)	X	...	...	...
21	Ref (57)	...	...	...	...
22	Ref (36)	X	X	...	...
23	Ref (142)	...	X	X	X
24	Ref (143)	...	X	X	X
25	Ref (17)	X	X	...	...

<sup>A</sup>Abbreviations:

C = porous ceramic,

PTFE = porous PTFE,

A = porous alundum,

CAF = cellulose-acetate fibers,

FG = fritted glass or glass fibers, and

SF = silica flour.

<sup>B</sup>Comparisons of materials based on this table should be made cautiously. Differing experimental techniques should be considered as a source of differing conclusions.

Undocumented factors often include material age and sampling history.

<sup>C</sup>Numbers in parenthesis refer to references cited in the second part of this table. This is indicated by (2).

<sup>D</sup>Valence states are often not reported in studies. This is indicated by (3).

<sup>E</sup>Reference (59) found that there is no significant interaction of cellulose-acetate fibers with potassium in solution. The porous section was washed prior to testing and results were found to be a function of several factors.

<sup>F</sup>Reference (139) found that there was no significant interaction of porous ceramic with nitrate plus nitrite nitrogen in solution. The results are a function of several factors.

<sup>G</sup>Absence of information on experimental techniques means that the techniques were not specified in the citation.

7.6.2.4 Interactions between porous materials and liquid can include sorption, desorption, cation exchange, precipitation (for example, ferric precipitation), and screening (20). These interactions can also occur with all other parts of the samplers that the samples contact. However, the much higher surface area of pores within porous segments makes them the most critical element chemically. Table 2 presents the results of a literature review for porous section/pore-liquid interactions. An attempt has been made to document the pertinent features of the listed studies. However, the reader should refer to the original papers to determine if experimental techniques are applicable to the situation of interest. The absence of entries for a constituent relative to a material does not infer absence of interactions. Although studies for membrane filter interactions have been performed, the results have not been included in Table 2. This is because membrane filters are made from a variety of materials that have differing chemical characteristics (36).

7.6.2.5 Suarez (89) showed that the pH of a sample may be affected by 0.28 to 0.44 pH units due to CO<sub>2</sub> degassing during sampling. He reduced this error by reducing the gas-liquid ratio

in the sampler, and by flushing several sampler volumes of soil solution through the sampler before collecting a sample. Alternately, Suarez (89) developed a model by which pH values could be corrected. He noted that multichamber samplers had minimal pH errors and that pH corrections due to CO<sub>2</sub> degassing were not necessary. Peters and Healy (53) found that there was no significant change in pH due to CO<sub>2</sub> degassing during long sampling times, although they recognized that pH changes could occur when the solution is originally more acidic than that which they tested. Ransom and Smeck (90) and Anderson (75) suggested purging the sampler with N<sub>2</sub> to preserve the subsurface redox states when sampling for redox dependent ions. Filter tip samplers do not use a purging gas, therefore, pore-liquid redox states are preserved in the samples.

7.6.2.6 Nightingale et al (31) indicated that normal suction sampling techniques are not suitable for sampling volatile organic compounds due to potential loss. Wood et al (91) devised a body tube connected to a purging chamber that is in turn connected to a trap packed with resin. Compounds that volatilize during sampling are captured in the trap. Pettyjohn et

al (92) described a suitable system for sampling highly volatile organics. However, the reported system was limited to a maximum sampling depth of 6 m and a small sample volume (5 to 10 mL). Torstensson and Petsonk (32) described methods that can be used to collect samples with filter tip samplers that result in no head space in the sample vial and consequently no loss of volatile compounds.

7.6.2.7 A newly forming consensus is that the effects of suction samplers (when properly pre-treated) on sample chemistry of non-dilute solutions are generally less significant than the inherent uncertainties of sampling discussed in 7.6.2.1 and 7.6.2.2 (17, 53, 93).

7.6.3 *Microbial Limitations*—Viruses or bacteria are sometimes monitored in areas where there are livestock lots, leach fields, septic tanks, or sewage sludge spreading plots. However, it has been found that although porous ceramics will allow viruses to pass, they will screen out bacteria (for example, *escherichia coli* and fecal coliform) (12, 26, 94, 95).

## 8. Free Drainage Samplers

8.1 Free drainage samplers are classified differently by various authors, depending on the installation methods. Many free drainage samplers are installed in the side walls of trenches and are referred to as trench lysimeters. However, free drainage samplers are also installed in the walls of vertical caissons. The principle behind each of the samplers is essentially the same. However, the materials and construction differ. The general types of free drainage samplers include:

- 8.1.1 *Pan lysimeters,*
- 8.1.2 *Glass block lysimeters,*
- 8.1.3 *Trough lysimeters,*
- 8.1.4 *Vacuum trough lysimeters,*
- 8.1.5 *Caisson lysimeters,*
- 8.1.6 *Wicking soil pore-liquid samplers,* and
- 8.1.7 *Sand filled funnel samplers.*

### 8.2 Operating Principles:

8.2.1 A free drainage sampler consists of some sort of collection chamber that is placed at depth in the soil. Pore-liquid in excess of field capacity is free to drain through soil (usually through macropores) under the influence of gravity. This gravity drainage creates a slightly positive pressure at the soil-sampler interface causing fluid to drip into the sampler. Hence, these samplers collect liquid from those portions of the vadose zone that are intermittently saturated due to events such as rainfall, flooding, or irrigation. Some free drainage samplers apply a small suction in order to break the initial surface tension at the soil-sampler interface. Samples are retrieved either by accessing the samplers at depth or by drawing samples to the surface through a suction line.

8.2.2 As described in 4.2, suction samplers can also be used to sample free drainage flow. However, the small area of those samplers compared to the spacing of macropores limits their usefulness for this application. As described in 7.4.5.3, Hornby et al (66) developed an installation that includes pressure-vacuum lysimeters within an encased monolith. This enhances collection of macropore flow with these samplers.

### 8.3 Description:

#### 8.3.1 *Pan Lysimeters:*

8.3.1.1 A pan lysimeter generally consists of a galvanized,

metal pan of varying dimensions. A copper tube is soldered to a raised edge of the pan. Plastic or vinyl tubing connects the copper tube to a collection vessel. Any liquid that accumulates on the pan drains through the tubing into the vessel (see Fig. 21) (26).

8.3.2 *Glass Block Lysimeters*—Barbee and Brown (81) developed free drainage samplers made from hollow glass bricks. These glass bricks, that are produced for ornamental masonry work, have dimensions of 30 by 30 by 10 cm and have a capacity of 5.5 L. To build a sampler, nine holes, 0.47 cm in diameter, are drilled along the perimeter of one of the square surfaces of a brick. Nylon tubing is inserted into one of the holes to allow for sample removal. The collecting surface is fitted with a fiberglass sheet to improve contact with the soil. Pore-liquid collection is enhanced by a raised lip along the edge of the surface (see Fig. 22).

#### 8.3.3 *Trough Lysimeters:*

8.3.3.1 Trough lysimeters, also known as Ebermayer lysimeters, rely on a trough or pail to collect pore-liquid. In order for the edges of the sampler to maintain a firm contact with the soil, a fiberglass screen is suspended inside the trough. The screen is lined with glass wool and covered with soil until the soil is even with the top of the trough (96).

8.3.3.2 Morrison (1) reported a trough lysimeter in which two parallel metal rods are inside the trough, in contact with the bottom side of the screen, and bent toward the collection tube. Liquid that enters the trough migrates along these rods towards the collection tube in response to capillary forces (see Fig. 23). A modification of this design consists of a metal trough with a length of perforated PVC pipe mounted inside. The trough is filled with graded gravel so that coarse material is immediately adjacent to the PVC pipe and fine sand is at the

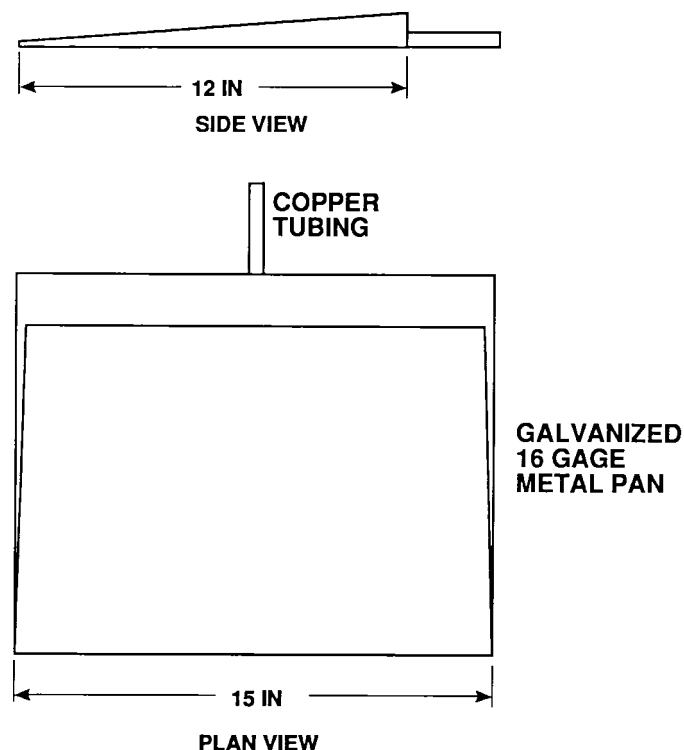


FIG. 21 Example of a Pan Lysimeter

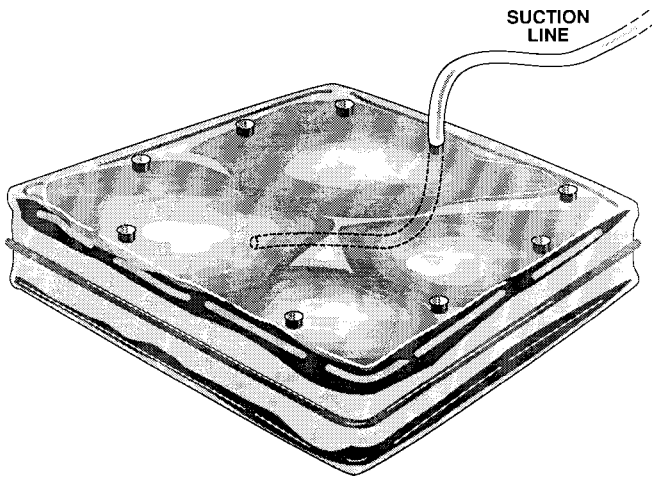


FIG. 22 Glass Block Lysimeter

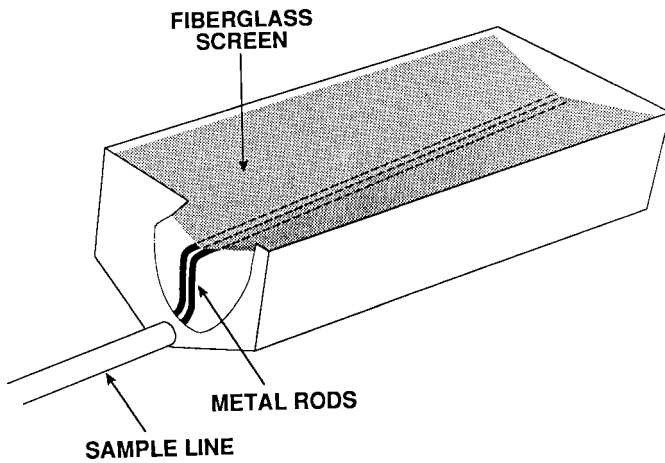


FIG. 23 Trough Lysimeter

edges and the top of the trough. The pipe is capped at one end while the other end is connected to a sample container via a drainage tube (1).

**8.3.4 Vacuum Trough Lysimeters**—Montgomery et al (97) described a vacuum trough lysimeter consisting of a metal trough equipped with two independent strings of ceramic pipe, each 13 mm in diameter. The design, otherwise similar to trough lysimeters, allowed extraction of samples under applied suctions of up to 50 cbar. The ceramic pipes act as a vacuum system, and samples are extracted through a suction line.

**8.3.5 Caisson Lysimeters**—A caisson lysimeter consists of collector pipes, radiating from a vertical chamber (1). A design used by Schmidt and Clements (98) consists of a nearly horizontal, half-screened PVC casing (see Fig. 24). Schneider et al (99) designed a similar system consisting of: a 15.2 cm diameter stainless steel tube extending diagonally upward through the caisson wall into the native soil; a screened plate assembly within the tube to retain the soil; a purging system that can be used to redevelop the sampler when it becomes clogged; and an airtight cap that prevents exchange between the air in the caisson and the soil air.

**8.3.6 Wicking Soil Pore-Liquid Samplers**—Hornby et al (66) described a wicking sampler, alleged to combine the attributes of free drainage samplers and pressure-vacuum

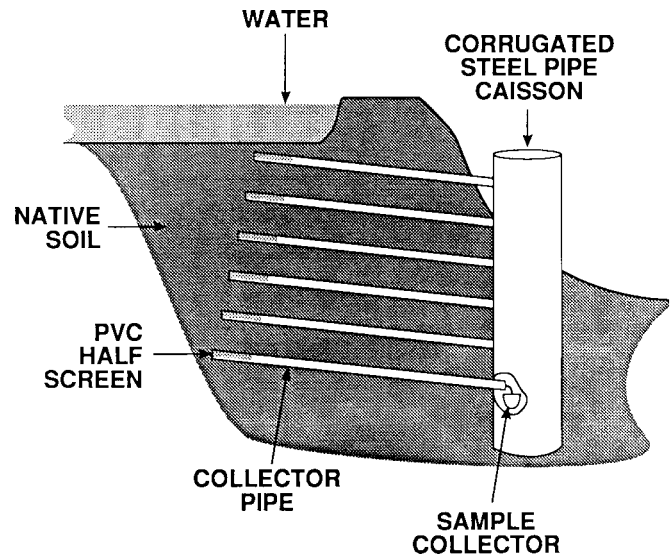


FIG. 24 Example of a Caisson Lysimeter

lysimeters. The sampler collects both free drainage liquid and liquid held at tensions to about 4 cbar. A hanging “Hurculon” fibrous column acts as a wick to exert a tension on the soil pores in contact with a geotextile fiber which serves as a plate covering a 30.5 by 30.5 by 1.3 cm pan. The terminus of the fibrous column is sealed into the cap of a tubular sample collector. The collection tube also contains an inlet pressure-vacuum line and a sample collection tube. Materials for the sample collector depend on the constituents being sampled. Glass and PTFE were recommended materials when sampling for organics (see Fig. 25) (66).

**8.3.7 Sand Filled Funnel Samplers**—K. W. Brown and Associates (100) discussed a sand-filled funnel for collecting freely draining liquid. The funnel is filled with clean sand and inserted into the sidewall of a trench. The funnel is connected through tubing to a collection bottle. Application of suction to

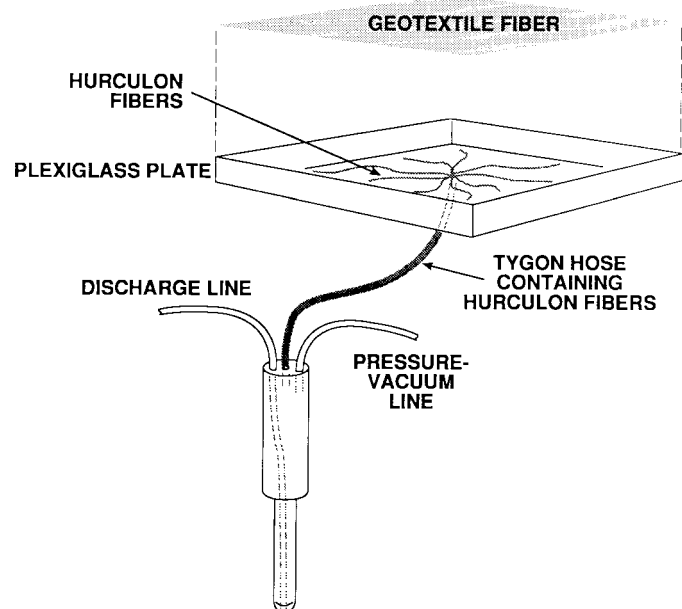


FIG. 25 Wicking Type Soil Pore-Liquid Sampler



a separate collection tube pulls the sample to land surface (see Fig. 26).

**8.3.8 Comments**—The dimensions of the free drainage samplers discussed are purposely left vague. Because the samplers collect fluid flowing primarily through macropores, the dimensions are often dictated on a site-by-site basis by the configurations and spacings of the macropores.

#### 8.4 Installation Methods:

##### 8.4.1 Installation:

**8.4.1.1 Free drainage samplers** are commonly installed into the side walls of trenches or caissons. The trenches can be dug either by hand or with backhoes. But they should be stabilized with timbers and siding if deeper than 1.5 m, in order to allow safe access (26) (see Occupational Health and Safety Administration Regulations). All wood used in the construction of permanent trenches should be treated with preservatives to protect it against degradation due to semi-saturated conditions. This may pose a problem when monitoring for organics. Any spaces between the bare trench side walls and the siding are filled with soil and peagravel to allow for free drainage. The excavations should be covered to provide positive surface drainage away from the area. Some free drainage samplers require only temporary excavations for installation. After the samplers have been installed, the excavations are backfilled with native soil.

**8.4.1.2 Caissons for housing free drainage samplers** are constructed with corrugated culverts or concrete drainage pipes. Schneider and Oaksford (101) installed caissons by excavating soil from within a concrete pipe using a crane operated shovel and manual labor. Each concrete pipe section, weighing 222.5 kN (25 tons), was set in place with a crane. As excavation inside the pipe progressed, the pipe advanced downward under its own weight.

**8.4.1.3 Pan Lysimeters**—A pan lysimeter can often be pushed or driven directly into the side wall of a trench. However, if the soil is resistant, an opening for the sampler can be created by hammering a sheet metal blade into the soil

profile with a sledge hammer. The pan is placed in the side wall so that it slopes gently toward the trench. Any voids above or below the pan are filled with soil (26). The end of the copper tubing is allowed to project through the trench siding and is connected to plastic tubing and a sample bottle (see Fig. 27).

**8.4.1.4 Glass Block Lysimeters**—A glass block lysimeter is installed in a cavity that is excavated in the side of a trench. Barbee and Brown (81) used a wooden model of the sampler in order to achieve the correct cavity size during excavation. They used a small knife to score the ceiling of the cavity in order to expose any pores that may have been smeared shut during excavation. Care should be taken to keep the ceiling of the cavity smooth and level so that liquids will not run off the upper surface of the glass block. Jordan (96) found that the edges of the sampler had to be in contact with the soil for the entire perimeter of the sampler in order to prevent liquid from running out through any spaces between the soil and the sampler. Level blocks are important so that the majority of the collected sample can be retrieved. However, the inside glass surface is uneven and has “low spots” where residual sample collects between sampling cycles. The glass block is pushed to the end of the cavity and wedges are used to hold its collecting surface firmly against the ceiling of the tunnel. Both the cavity and trench are partially backfilled. Barbee and Brown (81) recommend pressing a sheet of aluminum foil against the wall of the trench, extending below the top of the brick, before final backfilling in order to minimize any lateral migration of liquid from the disturbed portion of the soil profile to the undisturbed portion (see Fig. 28). It should be noted that aluminum foil is often coated with oil.

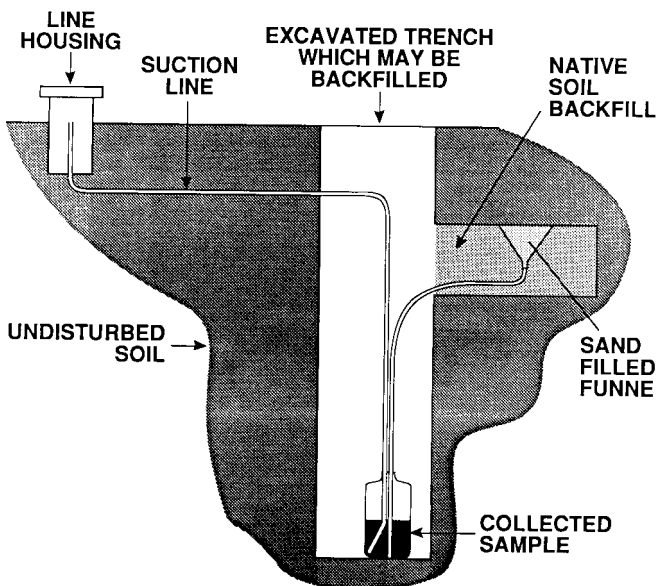


FIG. 26 Sand Filled Funnel Sampler Installation

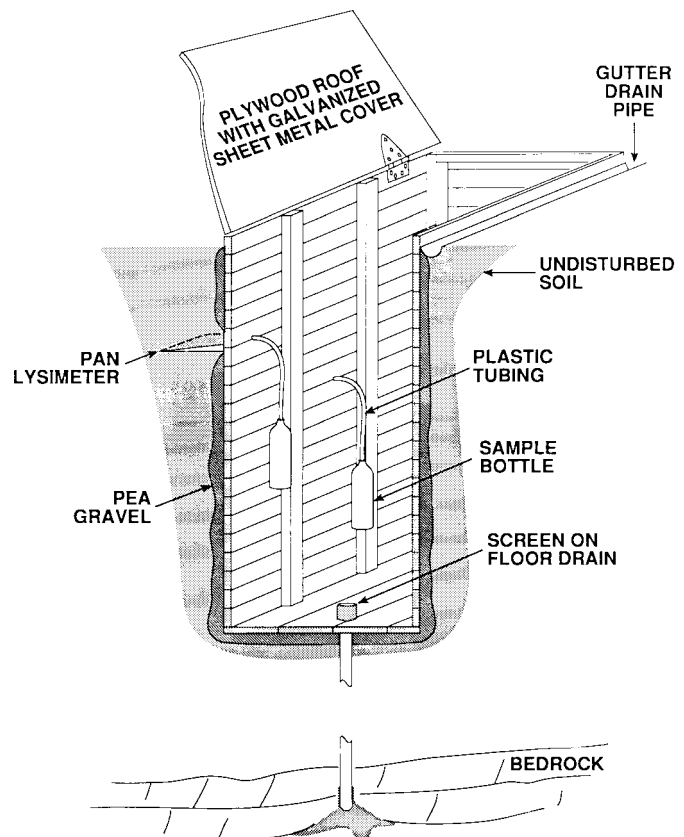


FIG. 27 Example of Pan Lysimeter Installation

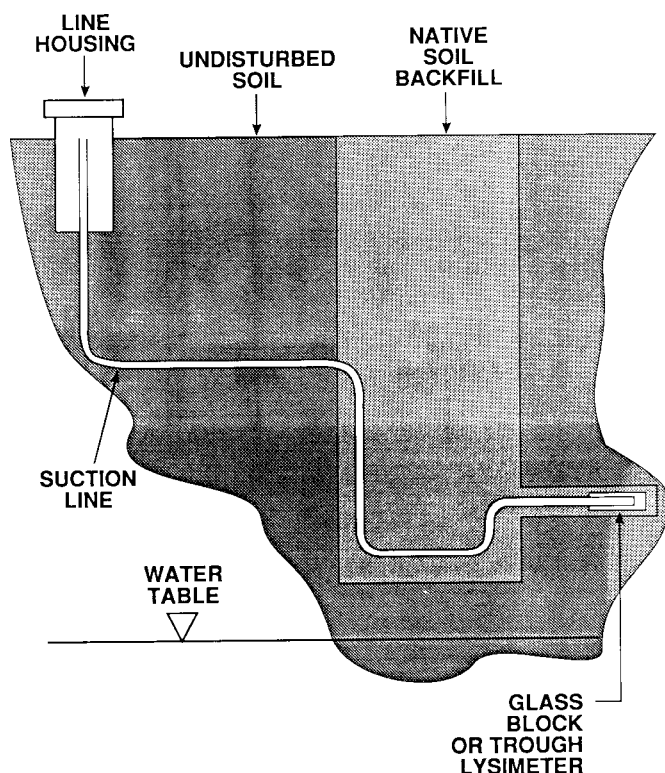


FIG. 28 Example of Glass Block or Trough Lysimeter Installation

the hanging wick and the collection tube. The pressure-vacuum line and the sample collection line are extended to the surface. During backfilling of the tunnel and trench, the bulk density of the fill should be equal to or greater than the native soil.

8.4.1.9 *Sand Filled Funnel Samplers*—The installation procedure for these samplers is similar to that used for glass block lysimeters (see Fig. 26).

#### 8.4.2 Maintenance:

8.4.2.1 Where samplers are accessed through permanent trenches and caissons, the sampling station must be protected against flooding due to excessive infiltration. Parizek and Lane (26) drilled a floor drain about 27 m through underlying soil into the unsaturated bedrock. This allowed drainage of excess liquid from the floor of the sampling trench, and also decreased the chances of contamination of soil surrounding the structure. Alternately, a sump pump can be used if a drain is not feasible. Parizek and Lane (26) also found that stratified soils intensified lateral flow of pore-liquid, thus aggravating any flooding problems. They concluded that flooding may be a problem in humid areas where more than about 5 cm of liquid per week is applied to the land surface.

8.4.2.2 The ground surface above the sampler should be maintained in a fairly representative state. Large housings and excessive traffic around the sampler (causing compaction of the soil) will reduce the amount of infiltration in that area. This will affect the representativeness of the pore-liquid samples.

8.4.3 *Comments*—A significant advantage of samplers installed in the sidewalls of trenches is that in sufficiently cohesive soils, installation produces no disturbance in the overlying soil. In cohesionless, sandy soils, stable cavities may not be possible. As a result, backfilling of the fallen material may be required. This disturbs the soil profile and macropores are not preserved (97).

#### 8.5 Operation:

##### 8.5.1 Methods:

8.5.1.1 Since pore-liquid flows into free drainage samplers under the influence of gravity, sampling is a relatively simple procedure. Liquid accumulates in the collection device and then drains through tubing into a sample bottle. The sample can be retrieved either through access to the sampling trench or by pulling it to the surface by a suction pump. The wicking pore-liquid sampler allows the application of a slight suction (4 cbar) to improve sampling. However, this design also has a tendency to clog.

8.5.1.2 Jordan (96) found that surface tension develops in trough lysimeters at the soil-air interface and prevents some of the liquid from entering the collector. Cole (41) addressed this problem by inserting an aluminum oxide disc between the soil and the collection surface, and then applying suction to break the surface tension and draw liquid out of the soil. The problem with this approach is that it requires the soil adjacent to the aluminum oxide disc to be free of roots, cracks, and channels (96). The two parallel rods included in the trough lysimeter design overcome this problem. If one end of the metal rod touches the fiberglass screen, then the surface film of liquid surrounds the rod and the liquid moves down the rod toward the sample container. Two rods, barely touching, facilitate this migration by allowing the liquid to move in response to the

8.4.1.5 *Trough Lysimeters*—Trough lysimeters are installed in the same manner as glass block lysimeters (see Fig. 28).

8.4.1.6 *Vacuum Trough Lysimeters*—The vacuum trough lysimeter described by Montgomery et al (97) is housed in a box-like structure, with four walls and a floor, but no ceiling. The floor of the structure and the lower portions of the walls are made of steel. The upper portions of the walls are composed of fiberglass coated plywood. A slotted, plastic drain pipe is set 20 mm above the floor of the structure and is surrounded by gravel. The soil profile surrounding the trough lysimeter is reconstructed incrementally in an attempt to recreate natural conditions. The structure is filled with soil in increments of 0.5 m or less. After each increment is added, liquid is piped slowly into the structure through the drain pipe and allowed to drain back out for 24 h before the next increment is added. This working of the soil particles by liquid is believed to produce bulk densities that are fairly representative of the undisturbed conditions. However, soil macropores are not reproduced.

8.4.1.7 *Caisson Lysimeters*—Lateral collectors or free drainage samplers are installed in cavities augered by hand or by power-driven equipment through holes in the caisson walls (see Fig. 24).

8.4.1.8 *Wicking Soil Pore-Liquid Samplers*—These units are installed by the trench and cavity method similar to that for glass block lysimeters (66). A backhoe excavates the trench to the desired depth. A cavity is then dug into the wall of the trench to the dimensions of the sampler. The roof of the cavity is sometimes scarified (depending on the soil type) to remove smearing caused during excavation. The sampler is then forced tightly into place to ensure good contact with the roof of the cavity. The cavity is large enough to accommodate the sampler,



capillary forces between them (96).

**8.5.2 Comments**—Under near saturated conditions with macropore flow, free drainage samplers tend to collect larger and more consistent samples than suction samplers. Since free drainage samplers are continuous samplers, they need to be emptied after each infiltration event in order to ensure sample integrity, to prevent sample container overflows, and to prevent cross contamination between hydrologic events (12).

#### 8.6 *Advantages and Limitations:*

**8.6.1 Physical Advantages and Limitations**—A major advantage of free drainage samplers is that they are essentially passive, thus they do not alter pore-liquid flow paths. The major disadvantage of free drainage samplers is that samples can only be obtained when soil moisture conditions are in excess of field capacity. Such saturated conditions usually require constant application of surface liquid, as in the case of agricultural irrigation or at land treatment sites. Under drier conditions, free drainage samplers fail to yield any liquid and suction samplers are required.

#### 8.6.2 *Chemical Advantages and Limitations:*

**8.6.2.1** There are both advantages and disadvantages to using free drainage samplers to collect pore-liquid for chemical analysis. A major advantage is that the samplers do not distort natural flow patterns as do suction samplers. Because samples are collected over known areas, quantitative mass balance estimates are possible. Because the samplers are continuous collectors, infiltration events can be sampled without having to go to the field. The major limitation of free drainage samplers is that they cannot sample pore-liquids held at tensions greater than the field capacity. As with all samplers, analytical parameters/sampler material compatibilities should be considered. As an example, samples collected from pan lysimeters should not be analyzed for copper or zinc, particularly if the pH of the collected fluid is below seven.

**8.6.2.2** Free drainage samplers tend to collect pore-liquids that drain rapidly through macropores. Since the residence time of this liquid is less than that of liquid moving under tension, the major ion chemistry appears more dilute than the fluid sampled from unsaturated pores with suction samplers. In some cases, this decrease in residence time in combination with other factors can result in an actual change in the chemical signature rather than just an overall dilution. This is because insufficient time may be available for reactions to occur with soil components that act as chemical sources or sinks (102). However, free drainage samplers have large cross-sectional areas and they are cumulative collectors. As a result, they collect samples which average soil heterogeneities and therefore give a more representative picture than suction samplers of chemical movement through wet soil, particularly through well-structured soils (81). In addition, the samplers use suction only to retrieve samples. As a result there is less potential for loss of volatile compounds than with suction samplers (12).

**8.6.3 Microbial Advantages and Limitations**—Since free drainage samplers do not have the minute openings that porous ceramic suction samplers have, they do not screen out colloidal-sized particles and soil bacteria. Consequently, they yield more representative values for suspended solids or BOD measurements (26).

## 9. Perched Ground Water Samplers

**9.1** Perched water occurs where varying permeability layers in the vadose zone retard downward movement of liquid. Over time, liquid collects above lower permeability layers and moisture contents rise until the soil becomes saturated with liquid (9, 103). Once soil becomes saturated, wells and other devices normally installed below the water table can be used to collect samples. Separate guides are available for ground water sampling, therefore, the topics are covered briefly, with reference to appropriate documents.

**9.2** Sampling perched liquid is attractive because the perching layer collects liquid over a large area. This integration allows samples to be more representative of areal conditions than suction samples (103). This also allows the sampler to potentially detect contaminants that may not be moving downward immediately adjacent to the sampler. In addition, larger sample volumes can be collected than those that can be obtained by suction samplers. Everett et al (6) and Everett et al (9) discussed the incorporation of perched ground water sampling into monitoring programs. There are a variety of systems that can be used. These include the following:

9.2.1 *Point samplers,*

9.2.2 *Wells,*

9.2.3 *Cascading water samplers,* and

9.2.4 *Drainage samplers.*

#### 9.3 *Operating Principles:*

**9.3.1 Point Samplers**—Point samplers are open ended pipes or tubes, such as piezometers or wells with short screened intervals, installed for the purpose of collecting samples from a discrete location in saturated material. Samples are collected by bringing liquid that flows freely into the device to the surface by one of a variety of methods.

**9.3.2 Wells**—A monitoring well is similar to a point sampler except the screened interval is longer. Therefore, samples are averaged over the screened length (104). Samples are collected by bringing liquid that flows freely into the well to the surface by one of a variety of methods.

**9.3.3 Cascading Water Samplers**—Cascading water occurs when a well is screened across a perched layer and the underlying water table or when water leaks through casing joints at the perched layer. Because the water table is lower than the perched layer, water flows into the well in the portion open to the perched layer, and cascades downward to the water table. This situation is common in some areas where the practice has been to install water wells with large screened intervals (105). Samples are collected by capturing liquid flowing into the well from the perched layer before it cascades down to the water table.

**9.3.4 Drainage Samplers**—Shallow perched systems may spread contamination, cause problems with structures, or interfere with agriculture. Therefore, drainage systems are sometimes installed. These systems usually funnel liquid via gravity flow to a ditch or sump from which it is pumped out. This outflow can be sampled. Typical drainage systems include tile lines or manifold collectors. Depending on the design of the system, it may be possible to sample outflows that drain different areas.

#### 9.4 *Description:*

#### 9.4.1 Point Samplers:

9.4.1.1 Point samplers can be installed in separate boreholes or clustered together in one borehole at different depths. Fig. 29 presents different configurations that have been used. Piezometers, that are often used as point samplers, are similar to wells, in that they consist of a small diameter casing open at one end or connected to a short screened interval (106). Reeve (107), Patton and Smith (108) and Morrison (1) discussed different designs.

9.4.1.2 Point samplers can be made from a variety of materials including steel, PVC, PTFE, ABS, fiberglass, and additional materials for joints, seals and other components (106, 107).

#### 9.4.2 Wells:

9.4.2.1 Monitoring wells (as depth averaged samplers) are normally installed with one well in each borehole. Components of a well generally include a bottom plug, a length of screen, a length of blank casing, a cap, and a protective cover. Different monitoring well designs are presented in Fig. 30, Fig. 31, and Fig. 32. Authors who described methods for designing and installing monitoring wells include U.S. EPA (109), Driscoll (110), Gass (111), Keely (112), Minning (113), Richter and Collentine (114), Riggs (115), Riggs and Hatheway (116), Scalf et al (117), Morrison (1), Everett et al (9), Campbell and Lehr (118). Hackett (119, 120) summarized methods for designing and installing monitoring wells with hollow stem augers. Screened hollow stem augers can also be used as temporary wells for sampling (Taylor and Serafini, (121)).

9.4.2.2 Monitoring wells can be made from a variety of materials including steel, PVC, PTFE, ABS, fiberglass, and additional materials for joints, seals, and other components. Details are provided by Barcelona et al (122), U.S. EPA (109), Morrison (1) and many of the references listed above.

#### 9.4.3 Cascading Water Samplers:

9.4.3.1 Cascading water is most often seen in production wells in areas with extensive ground water pumpage. Samplers simply consist of a bucket or bailer lowered to a point below the inflow of cascading water. Wilson and Schmidt (103) described methods for developing cascading water samplers (see Fig. 33).

9.4.3.2 Cascading wells differ from other wells only by the way in which water flows into them. Otherwise, the materials used for these wells are identical to those used for other types

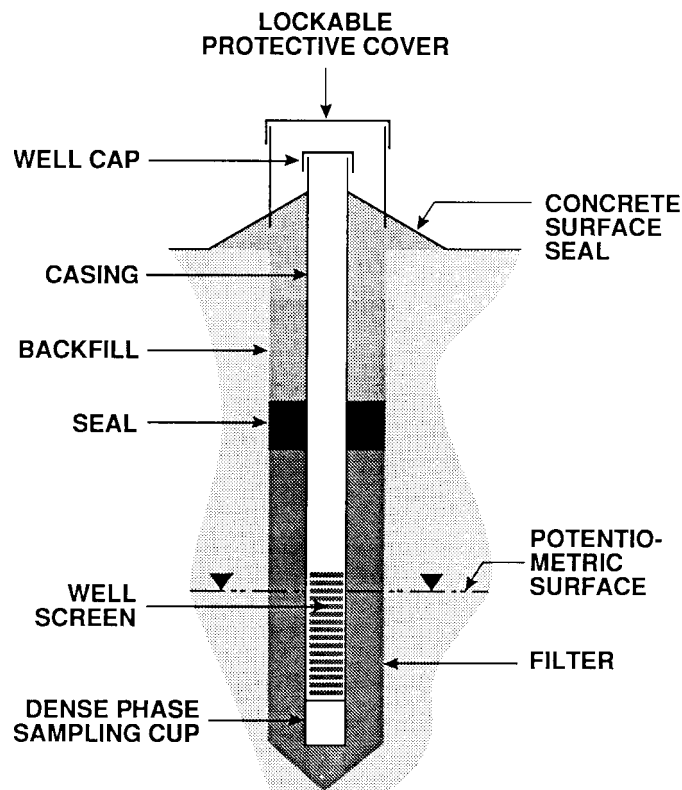


FIG. 30 A Monitoring Well With the Uppermost Ground-Water Level Intersecting the Slotted Well Screen

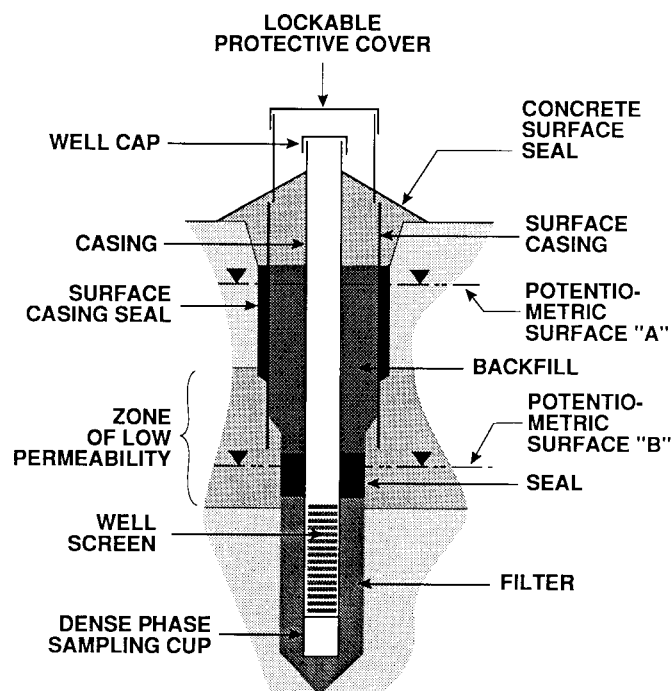


FIG. 31 A Monitoring Well Installed to Sample From the Lower of Two Ground-Water Zones

of wells. Bailers or buckets used to collect samples are also available in steel, PVC, PTFE, acrylic and other materials.

#### 9.4.4 Drainage Samplers:

9.4.4.1 Drainage systems consist of conduits installed within the perched zone at sufficient slopes for water to flow to

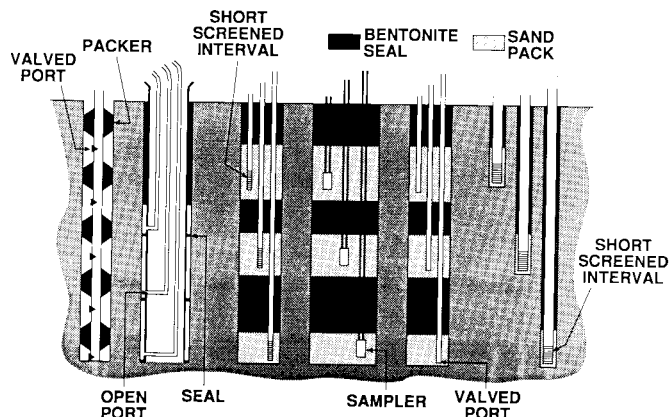


FIG. 29 Examples of Point Sampling Systems

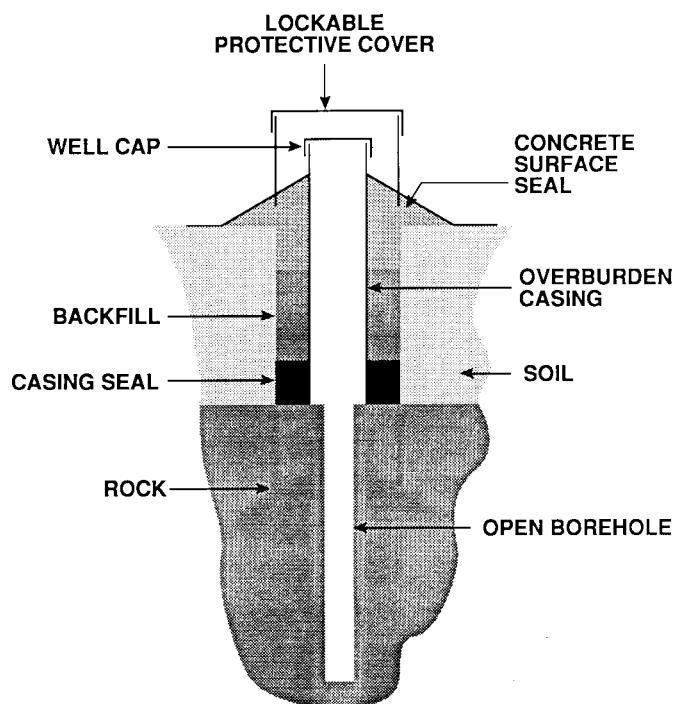


FIG. 32 An Open-Hole Ground-Water Monitoring Well in Rock

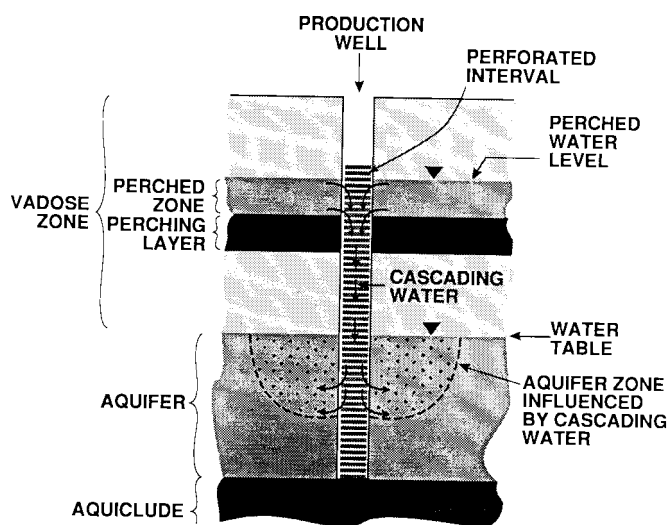


FIG. 33 Conceptualized Cross Section of a Well Showing Cascading Water from Perched Zone

a central ditch or drain. The conduits can be tile drains, half perforated pipes, synthetic sheeting, or even layers of gravel and sand. Schilfgaarde ed. (123), contains numerous papers on the design and construction of drainage systems. Donnan and Schwab (124) described sampling from agricultural drainage systems. Gilliam et al (125), Gambrell et al (126), Eccles and Gruenberg (127) and Gilliam et al (128) described sampling from tile drains. Gilliam et al (125) and Jacobs and Gilliam (129) described sampling from drainage ditches. Wilson and Small (130) described a lateral drain sampler installed beneath a new sanitary landfill. A perforated pipe collected liquid that was funneled to a sump via a drain line. In most of these systems, a thin layer of high permeability sand or gravel is installed around the drain to promote flow into the collector,

and to sieve out fine materials.

9.4.4.2 Because drainage systems often require large quantities of materials, less exotic, cheaper materials such as baked clay tiles and PVC are often used.

9.4.5 *Comments*—As with all samplers, potential chemical interaction between the sampler material and the constituents of interest should be considered. Because these samplers are usually installed for other purposes, incompatibility of materials with monitoring objectives is often a problem. Everett et al (9), Dunlap (131), and U.S. EPA (109) discussed this topic.

#### 9.5 Installation Methods:

9.5.1 *Point Samplers*—Reeve (107), Patton and Smith (108), and Morrison (1) discussed procedures for installing and maintaining point samplers.

9.5.2 *Wells*—Most of the references listed in 9.4.2.1 describe methods for installing monitoring wells.

9.5.3 *Cascading Water Samplers*—Wilson and Schmidt (103) discussed methods for installing cascading water samplers.

9.5.4 *Drainage Samplers*—Schilfgaarde ed. (123) contains articles that discussed installation of agricultural drainage systems. Associated hazards and costs often prohibit the installation of these systems at existing landfills. As a result, inclusion of these systems, as leachate collectors, in new landfills is more common. Everett et al (9) discussed methods for installing drainage sampling systems at hazardous waste sites.

#### 9.6 Operation:

9.6.1 *Point Samplers*—Point samplers usually have diameters that are too small to allow the use of submersible pumps. As a result, suction methods are usually required (1). Sampling techniques are described in Pickens et al (106), Reeve (107), Patton and Smith (108) and Morrison (1).

9.6.2 *Wells*—Samples may be retrieved from wells in the same manner as from piezometers. However, because wells are designed for sampling or pumpage, diameters are usually large enough to accommodate most pumps. Samples can be brought to the surface by a variety of systems including bailers, suction pumps (for example, peristaltic pumps), air lift pumps, piston pumps, submersible pumps and swabbing. Each of these methods have advantages and disadvantages relating to considerations such as depth to water, required sample volume, sampling speed, alteration of the sample chemistry, equipment requirements, manpower requirements, and cost. These considerations were discussed by Everett et al (9), Fenn et al (132), Gibb et al (133), U.S. EPA (109), Dunlap et al (131), Driscoll (110), and Anderson (75). Sampling methods were described in most of the references of 9.4.2.1. As described in 9.3.2, samples from wells are averaged over the screened interval. However, samples from discrete depths along the screened interval can also be obtained using packer-pump setups such as those described by Fenn et al (132).

#### 9.6.3 Cascading Water Samplers:

9.6.3.1 Wilson and Schmidt (103) described techniques for sampling from cascading wells. A bailer or bucket is decontaminated and then lowered to a position in the well below the cascading water but above the water table. When the sampler is full, it is pulled back to the surface. Alternately, as shown in



Fig. 33, the chemistry of the water table immediately around a well that has been shut down will be dominated by the cascading water. Therefore, a sample can also be collected from the water table during the initial stages of pumping.

9.6.3.2 Cascading wells are usually production wells in which drawdown has lowered the water table sufficiently to cause cascading. Because of this, there is usually a pump installed in the well that will prevent access for sampling. However, the pumps are periodically removed for maintenance. Therefore, it should be possible to coordinate sampling with pump maintenance personnel.

9.6.4 *Drainage Samplers*—Samples may be collected where tile lines or drainage pipes discharge to ditches or sumps (125, 126, 127, 128, 129, 125, 125, 125, 130). Willardson et al (134) described a “flow-path ground water sampler” that allows collection of water following different flowpaths along a tile drainage system.

#### 9.7 Limitations:

9.7.1 Perched water systems can be difficult to find and delineate. Surface and borehole geophysical methods (for example, neutron logging) and video logging of existing wells are often used. Also, perched systems tend to be ephemeral. Therefore, suction samplers are sometimes required as backups.

9.7.2 *Point Samplers*—The major problem with point sampling systems is that their diameters are often too small to allow adequate development after installation or to allow sampling by any method other than suction. Because the maximum suction lift of water is about 7.5 m, this is the maximum sampling depth for many of the small diameters systems. Systems such as those depicted in Fig. 29 require tight contact with the surrounding material to prevent side leakage of liquid. Depending on the material, this tight contact may not be achievable (9).

9.7.3 *Wells*—Wells provide samples that are averaged over the screened interval. As a result, when contaminants are detected, packer-pump arrangements must be used if zonation of the contaminants is to be delineated. When separate phases of water-immiscible fluid (for example, oil) are found floating in the well, it is difficult to obtain samples of the underlying water without contamination from the overlying fluid. As with all samplers, care should be taken to ensure that materials used to construct a well are compatible with the chemical analyses to be performed.

9.7.4 *Cascading Water Samplers*—Cascading water may enter a well from several distinct perched systems (103). As a result, the sample may be a mixture of water from several depths. Cascading water is most often sampled from pre-existing wells used for other purposes. As a result, materials used in the well construction may alter those chemical constituents of interest. Wells used for irrigation and water supply often have lubricant oils from the pump floating in them. With fluctuating water levels, these oils become smeared along the casing, and may even move out into the surrounding soils. Therefore, traces of these oils may appear in samples.

9.7.5 *Drainage Samplers*—Because of the limitations of excavation equipment, drainage samplers are limited to shallow depths. In addition, the systems are difficult to install in

rocky or steep terrains. In areas that experience freeze-thaw cycles, they may be damaged by soil heaving (9). Drainage systems are often susceptible to clogging over time as fine particles and chemically precipitated material accumulate on the drain openings. Collected samples may or may not be representative of average conditions, depending on the distribution of soil types and contaminants in the drained area. If the area of contamination is small compared to the drained area, dilution may prevent the detection of contaminants. In addition, pollutants that are heavier than water may move below the drain if it is not located at the bottom of the perched zone. As with all samplers, there is the possibility of chemical interaction between the sampling system and the chemical constituents of interest. In the case of drainage sampling systems, this effect is amplified as contaminants may have to travel considerable distances through drains before being sampled. In addition, normally non-aerated solutions may be aerated and chemically altered as they travel through drains.

## 10. Experimental Absorption Samplers

10.1 *Operating Principles*—Absorbent samplers depend on the ability of the material to absorb pore-liquid (1). Samples are collected by placing the sampler in contact with soil. Liquid is allowed to absorb into the sampler material over time. The sampler is then removed, and the sample liquid is extracted for analyses.

#### 10.2 Description:

10.2.1 Two designs have been described. The first design includes a cellulose-nylon sponge (0.5 by 4.8 by 30 cm) seated in a galvanized iron trough. The trough is pressed against a soil surface with a series of lever hinges (135).

10.2.2 The second absorbent sampler design consists of tapered ceramic rods that are driven into soil (136). The rods are made from unglazed ceramic similar to that used as the porous segments of suction samplers.

#### 10.3 Installation:

##### 10.3.1 Pre-Installation:

10.3.1.1 Sponge samplers are prepared by soaking them for 24 h in a 1 to 5 % NaOH solution containing a washing powder, Tadros and McGarity (135). Sponges are then pressed dry using rollers, stored in a moisture tight container, and taken to the field.

10.3.1.2 Ceramic rod samplers are weighed, boiled in distilled water, oven dried, and stored in a desiccator. The rods are weighed again and then taken to the field.

##### 10.3.2 Installation:

10.3.2.1 A sponge is placed in the sampling trough. The trough is then placed in a horizontal cavity cut into the side of a trench. The trough is then pressed against the cavity ceiling with the lever hinges.

10.3.2.2 A ceramic rod sampler is installed by simply driving it into the soil.

##### 10.3.3 Maintenance:

10.3.3.1 The only field maintenance required for sponge samplers is the preservation of the sampling trench if future sampling is desired at that location.

10.3.3.2 There is no field maintenance for ceramic rod samplers as they are completely removed to retrieve the samples.

**10.3.4 Comments**—Theoretically, there is no maximum installation depth for sponge samplers. However, because access trenches are required for operation, installations are restricted to shallow depths dictated by excavation equipment and safety considerations. Given the NaOH treatment (without rinsing) of the sponge, measurements of pH, conductivity, TDS, metals and major cations and anions might be affected by the residual NaOH. Depending on the composition of the “washing powder”, phosphate, BOD, and MBAS might also be affected.

**10.3.4.1 Ceramic rod samplers** will have maximum installation depths if pushed or driven from the surface. This maximum depth will generally decrease with increasing soil grain size. However, deeper installations can be achieved by drilling to the top of the interval to be sampled, lowering a rod down the hole and pushing or driving the rod into the sampling interval (**136**).

#### 10.4 Operation:

##### 10.4.1 Methods:

**10.4.1.1 Sponge samplers** are pressed against the soil until a sufficient volume of liquid for planned analyses has been absorbed. The sampler is then removed and the sponge is placed in a moisture proof container. The sample is extracted from the sponge with rollers.

**10.4.1.2 Ceramic rod samplers** are pushed or driven into the soil and left in place for a period of time. The rods are then withdrawn, and weighed. The rods are leached by boiling in a known volume of distilled water. This solution is then analyzed. The concentrations of constituents in the original pore-liquid are estimated by using the ratio of the volume of absorbed water to the volume of the boiling water.

**10.4.2 Comments**—The amount of liquid that can be sampled is dependent on time, soil type, moisture content,

absorbency of the sampler material, volume of the absorbent material, and surface area of the absorbent material in contact with the soil. Generally, sponge samplers function only at higher moisture contents approaching saturation (**1**). Shimshi (**136**) used ceramic rod samplers to sample from a sandy loam with moisture contents varying from 7 to 20 %.

#### 10.5 Limitations:

**10.5.1 Physically**, absorbent methods are limited to soils approaching saturation. Sampling requires removal of the absorbent material. Because of this, repeat sampling at the same location is difficult. Although the sampler may be placed back at its original location, identical hydraulic contact with the soil cannot be guaranteed.

**10.5.2 Chemically**, as with other samplers, there are problems with absorption, desorption, precipitation, cation exchange, and screening of various pore-liquid components as a function of the sampler materials. Tadros and McGarity (**135**) discussed these concerns in relation to sponge samplers. Shimshi (**136**) provided a good discussion of the limitations of sampling for nitrate with ceramic rod samplers. Specifically, he found that at lower moisture contents, sampled solutions became less representative due to vapor transfer and chromatographic separation. However, he suggested that these effects could be reduced by increasing the length of the rod insertion period. Clearly, boiling will affect analyses for organics, BOD, COD,  $\text{NH}_3$ , and some other species of nitrogen, among others.

## 11. Keywords

11.1 pore fluids; pressure vacuum lysimeters; soil moisture; soil water; suction lysimeters; unsaturated; vadose zone sampling

## APPENDIX

### (Nonmandatory Information)

#### X1. DESCRIPTIONS OF TERMS SPECIFIC TO THIS STANDARD

**X1.1 air entry value**—the applied suction at which water menisci of the porous segment of a suction sampler break down, and air enters.

**X1.2 bubbling pressure**—the applied air pressure at which water menisci of the porous segment of a suction sampler break down, and air exits.

**X1.3 cascading water**—perched ground water that enters a well casing via cracks or uncovered perforations, trickling, or pouring down the inside of the casing.

**X1.4 cation exchange capacity (CEC)**—the total capacity of a porous system to adsorb cations from a solution.

**X1.5 hydraulic gradient**—the change in total hydraulic head of water per unit distance of flow.

**X1.6 hydrophobicity**—the property that defines a material as being water repellent. Water exhibits an obtuse contact angle

with hydrophobic materials.

**X1.7 hydrophelicity**—the property that defines a material as attracting water. Water exhibits an acute contact angle with hydrophilic materials.

**X1.8 lysimeter**—a device to measure the quantity or rate of water movement through a block of soil, usually undisturbed or in-situ; or to collect such percolated water for analyses.

**X1.9 macropore**—interaggregate cavities that serve as the principal avenues for the infiltration and drainage of water and for aeration.

**X1.10 matric potential**—the energy required to extract water from a soil against the capillary and adsorptive forces of the soil matrix.

**X1.11 matric suction**—for isothermal soil systems, matric suction is the pressure difference across a membrane separating



soil solution, in-place, from the same bulk (see soil-water pressure).

X1.12 *micropore*—intraaggregate capillaries responsible for the retention of water and solutes.

X1.13 *percolation*—the movement of water through the vadose zone, in contrast to infiltration at the land surface and recharge across a water table.

X1.14 *pore-liquid*—Liquid that occupies an open space between solid soil particles. Within this guide, pore-liquid is limited to aqueous pore-liquid; that includes water and its solutes.

X1.15 *pore-liquid tension*—see *matric-suction* or *soil-water pressure*.

X1.16 *pressure head*—the head of water at a point in a porous system; negative for unsaturated systems, positive for saturated systems. Quantitatively, it is the water pressure divided by the specific weight of water.

X1.17 *Richard's outflow principle*—the principle that states that pore-liquid will not generally flow into an air-filled cavity (at atmospheric pressure) in unsaturated soil.

X1.18 *soil-water pressure*—the pressure on the water in a soil-water system, as measured by a piezometer for a saturated soil, or by a tensiometer for an unsaturated soil.

X1.19 *tensiometer*—a device for measuring soil-water matric potential (or tension or suction) of water in soil in-situ; a porous, permeable ceramic cup connected through a water filled tube to a pressure measuring device.

X1.20 *total soil-water potential*—the sum of the energy-related components of a soil-water system; for example, the sum of the gravitational, matric and osmotic potentials.

X1.21 *tremie method*—the method whereby materials are emplaced in the bottom of a borehole with a small diameter pipe.

X1.22 *vacuum*—a degree of rarefaction below atmospheric pressure: negative pressure.

X1.23 *Vadose zone*—the hydrogeological region extending from the soil surface to the top of the principle water table; commonly referred to as the “unsaturated zone” or “zone of aeration”. These alternate names are inadequate as they do not take into account locally saturated regions above the principle water table (for example, perched water zones).

X1.24 *water content*—the amount of water stored within a porous matrix, expressed as either a volume (volume per unit volume) or a mass (mass per unit mass) of a given solid.

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