



JASP

STATISTICAL ANALYSIS IN JASP

A GUIDE FOR STUDENTS

JASP 2025

Professor Mark A Goss-Sampson



JASP

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PREFACE

JASP stands for **Jeffrey's Amazing Statistics Program** in recognition of the pioneer of Bayesian inference Sir Harold Jeffreys. This is a **free** multi-platform open-source statistics package, developed and continually updated by a group of researchers at the University of Amsterdam. They aimed to develop a free, open-source programme that includes both standard and more advanced statistical techniques with a major emphasis on providing a simple intuitive user interface.

In contrast to many statistical packages, JASP provides a simple drag-and-drop interface, easy access menus, intuitive analysis with real-time computation and display of all results. All tables and graphs are presented in APA format and can be copied directly and/or saved independently. Tables can also be exported from JASP in LaTeX format

JASP can be downloaded free from the website <https://jasp-stats.org/> and is available for Windows, Mac OS X and Linux. You can also download a pre-installed Windows version that will run directly from a USB or external hard drive without the need to install it locally. The WIX installer for Windows enables you to choose a path for the installation of JASP – however, this may be blocked in some institutions by local Administrative rights.

The programme also includes a data library with an initial collection of over 50 datasets from Andy Fields' book, Discovering Statistics using IBM SPSS statistics¹ and The Introduction to the Practice of Statistics² by Moore, McCabe and Craig.

Keep an eye on the JASP site since there are regular updates as well as helpful videos and blog posts!!

This book is a collection of standalone handouts covering the most common standard (frequentist) statistical analyses used by students studying Human Sciences. Datasets used in this document are available for download from <https://osf.io/bx6uv/>

I would also like to thank Per Palmgren from the Karolinska Institutet in Sweden for his helpful comments, suggestions and proofreading of the original guide.

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2025

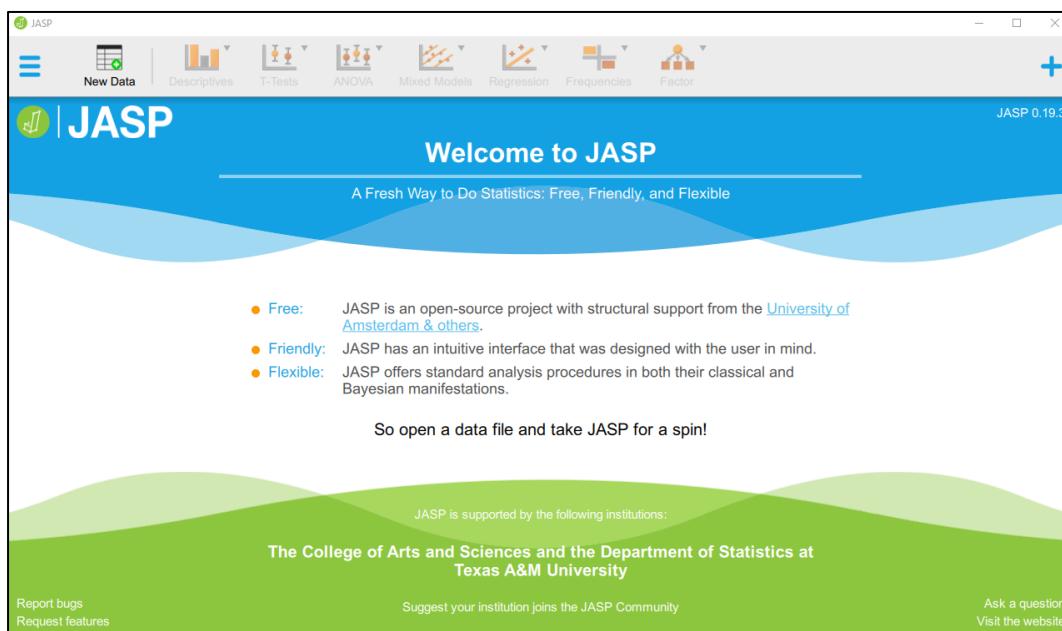
¹ A Field. (2017) Discovering Statistics Using IBM SPSS Statistics (5th Ed.) SAGE Publications.

² D Moore, G McCabe, B Craig. (2011) Introduction to the Practice of Statistics (7th Ed.) W H Freeman.



USING THE JASP ENVIRONMENT

Open JASP.



The main menu can be accessed by clicking on the top-left icon.

The image shows the "Open" dialog box in JASP. On the left is a sidebar with options: Open, Save, Save As, Export Results, Export Data, Sync Data, Close, Preferences, and About. The main area is titled "Open:" and lists "Recent Files" and "Computer", "OSF", and "Data Library". To the right of the dialog box is a list of supported file formats:

Open:

JASP has its own **.jasp** format but can open a variety of different dataset formats, such as:

- .xls/xlsx** (Excel files)
- .csv** (comma-separated values) can be saved in Excel
- .txt** (plain text) can also be saved in Excel
- .tsv** (tab-separated values) can also be saved in Excel
- .sav** (IBM SPSS data file)
- .ods** (Open Document spreadsheet)
- .dta** (Stata data file)
- .por** (SPSS ASCII file)
- .Sas7bdat/cat** (SAS data files)
- .xpt** (SAS transport file)

You can open recent files, browse your computer files, access the Open Science Framework (OSF) or open the wide range of examples that are packaged with the Data Library in JASP.

**Save/Save as:**

The screenshot shows the JASP menu bar with the following structure:

- Open
- Save
 - Save As
 - Export Results
 - Export Data
 - Sync Data
- Close
- Preferences
- About

The "Save As" option is highlighted in grey.

Using these options for the data file, any annotations and the analysis can be saved in the .jasp format

Export:

Results can be exported to either an HTML file or a PDF file

Data can be exported to either a .csv, .tsv or .txt file

Sync data:

Used to synchronise with any updates in the current data file (also can use Ctrl-Y)

Close:

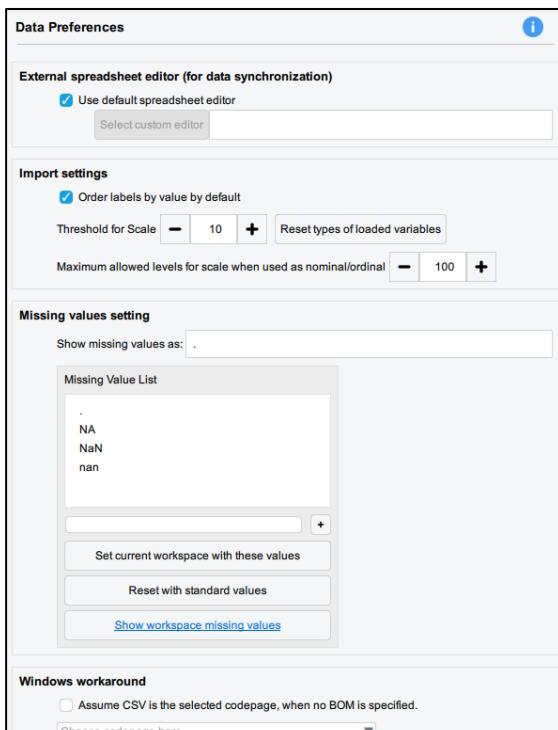
As it states, it closes the current file but not JASP.

Preferences:

There are four sections that users can use to tweak JASP to suit their needs

In the **Data Preferences** section, users can:

- Set the default spreadsheet editor (i.e. Excel, SPSS, etc.)
- Change the threshold so that JASP more readily distinguishes between nominal and scale data.
- Add a custom missing value code.





In the **Results Preferences** section, users can:

- Set JASP to return exact p values, i.e. P=0.00087 rather than P<.001
- Fix the number of decimals for data in tables – makes tables easier to read/publish
- Change the pixel resolution of the graph plots
- Select when copying graphs whether they have a white or transparent background.
- PDF output settings

The screenshot shows the 'Results Preferences' dialog box. It includes the following sections:

- Table options:** Includes checkboxes for 'Display exact p-values' (unchecked), 'Use exponent notation' (unchecked), and 'Fix the number of decimals' (checked). A numeric input field with a minus sign, the value '3', and a plus sign is shown.
- Plot options:** Includes a checkbox for 'Use PPI of screen in plots: (96)' (unchecked) and a 'Custom PPI:' input field set to '300'. Below this are options for 'Image background color': 'White' (selected) and 'Transparent'.
- PDF Settings:** Includes 'Orientation' options ('Portrait' selected, 'Landscape' unselected) and a 'Page size' dropdown menu currently set to 'A4'.
- Miscellaneous options:** Includes a checkbox for 'Show R syntax' (unchecked).

In the **Interface Preferences** section, users can now define a user font and pick between two different themes: a light theme (default) and a dark theme.

The preferred language currently supports 13 languages: English, Spanish, German, Dutch, French, Indonesian, Japanese, Portuguese, Chinese and Galician.

In this section, there is also the ability to change the system size (zoom) for accessibility and scroll speeds.



User Interface Options

Fonts

Interface: default: Segoe UI
R, JAGS, or lavaan code: default: Segoe UI
Result & help: default: Segoe UI

Use Qt's text rendering

Themes

Light theme
 Dark theme

Preferred language

Choose language en - English

[Help us translate or improve JASP in your language](#)

Accessibility options

ALT-Navigation mode

Check for updates

Daily automatic check for updates & known issues

Miscellaneous options

Zoom (%):

Ribbon scale (%):

Scroll speed (pix/s):

Safe graphics mode
 Disable animations
 Use native file dialogs

Display

Descriptive statistics
 Estimates of effect size
 η^2 partial η^2 ω^2
 Vovk-Sellke maximum p-ratio

► Model

▼ Assumption Checks

Homogeneity tests
Homogeneity corrections
 None Brown-Forsythe Welch
 Q-Q plot of residuals

Display

Descriptive statistics
 Estimates of effect size
 η^2 partial η^2 ω^2
 Vovk-Sellke maximum p-ratio

► Model

▼ Assumption Checks

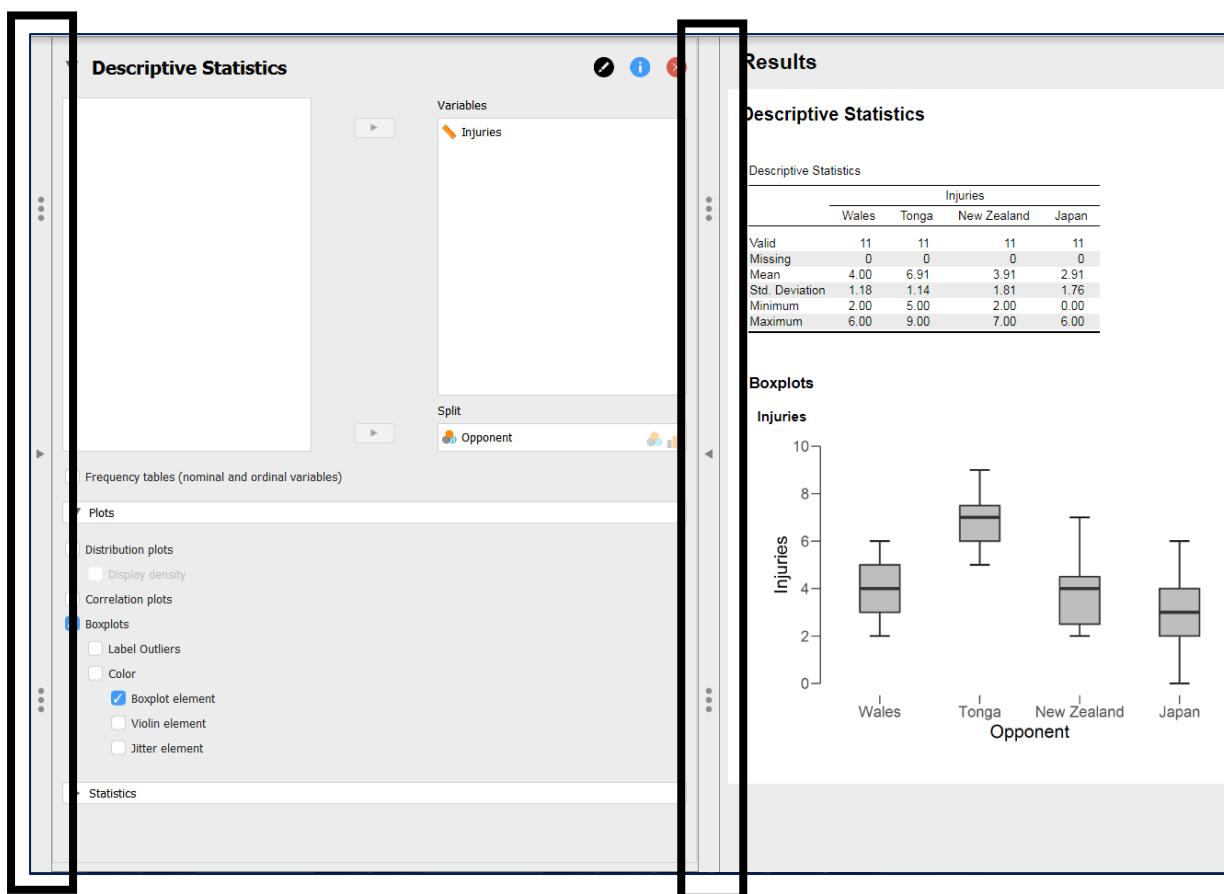
Homogeneity tests
Homogeneity corrections
 None Brown-Forsythe Welch
 Q-Q plot of residuals

Comparison of the dark and light themes in JASP

In the **Advanced Preferences** section, most users will probably never have to change any of the default settings.



JASP has a streamlined interface to switch between the spreadsheet, analysis and results views.



The vertical bars highlighted above allow for the windows to be dragged right or left by clicking and dragging the three vertical dots.



The individual windows can also be completely collapsed using the right or left arrow icons. ➤ ➙

If you click the Results ▼ icon, a range of options is provided, including:

- **Edit title**
- **Copy**
- **Export results**
- **Add notes**
- **Remove all**
- **Refresh all**

The 'add notes' option allows the results output to be easily annotated and then exported to an HTML or PDF file by going to File > Export Results.



ANOVA - Number of England Injuries

Cases	Sum of Squares	df	Mean Square	F	p
Country code	97.09	3	32.364	13.23	< .001
Residual	97.82	40	2.445		

Note. Type III Sum of Squares

One way ANOVA of injuries received by England rugby players against Tonga, New Zealand, France and Wales



The Add notes menu provides many options to change text font, colour, size, etc.

You can change the size of all the tables and graphs using Ctrl+ (increase), Ctrl- (decrease), Ctrl= (back to default size). Graphs can also be resized by dragging the bottom right corner of the graph.

As previously mentioned, all tables and figures are APA standard and can just be copied into any other document. Since all images can be copied/saved with either a white or transparent background. This can be selected in Preferences > Advanced as described earlier.

There are many further resources on using JASP on the website <https://jasp-stats.org/>



DATA HANDLING IN JASP

For this section, open England injuries.csv

All files must have a header label in the first row. Once loaded, the dataset appears in the window:

	Opponent	Injuries	
1	Japan	4	
2	Japan	1	
3	Japan	3	
4	Japan	6	
5	Japan	2	
6	Japan	3	
7	Japan	4	
8	Japan	0	
9	Japan	5	
10	Japan	2	
11	Japan	2	
12	New Zealand	2	
13	New Zealand	4	

For large datasets, there is a hand icon that allows easy scrolling through the data.



On import, JASP makes a best guess at assigning data to the different variable types:



If JASP has incorrectly identified the data type, just click on the appropriate variable data icon in the column title to change it to the correct format.

	Opponent	Injuries
2	Japan	

A context menu is open over the 'Injuries' column header, showing options: Scale (highlighted), Ordinal, and Nominal.

If you have coded the data, you can click on the variable name to open the following window in which you can label each code. These labels now replace the codes in the spreadsheet view. If you save this as a .jasp file, these codes, as well as all analyses and notes, will be saved automatically. This makes the data analysis fully reproducible.



Filter	Value	Label	
<input checked="" type="checkbox"/>	1	Tonga	
<input checked="" type="checkbox"/>	2	New Zealand	
<input checked="" type="checkbox"/>	3	France	
<input checked="" type="checkbox"/>	4	Wales	

In this window, you can also carry out simple filtering of data, for example, if you untick the Wales label, it will not be used in subsequent analyses.



Clicking this icon in the spreadsheet window opens up a much more comprehensive set of data filtering options:

+ - * ÷ / ^ √ % = ≠ < ≤ > ≥ ∧ ∨ | -

G...e
Cou...ode
Number of... Injuries

|y|
 σ_y
 σ^2_y
 $\sum y$
 $\prod y$
 $\min(y)$
 $\max(y)$
 $\text{mean}(y)$
 $\text{round}(y)$
 $\text{length}(y)$
 $\text{median}(y)$

Welcome to the drag and drop filter!

Using this option will not be covered in this document. For detailed information on using more complex filters, refer to the following link: <https://jasp-stats.org/2018/06/27/how-to-filter-your-data-in-jasp/>



By default, JASP plots data in the Value order (i.e. 1-4). The order can be changed by highlighting the label and moving it up or down using the appropriate arrows:

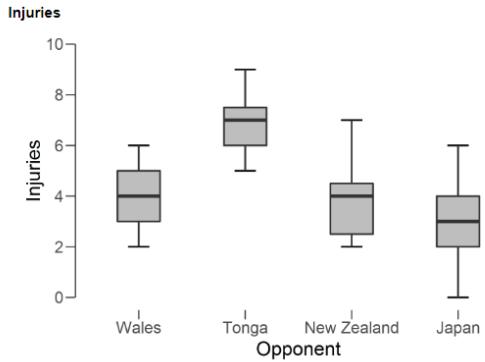
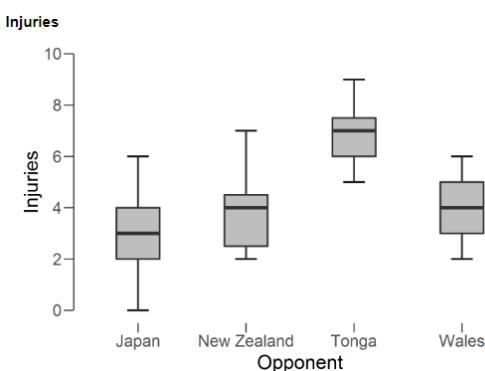
Filter	Value	Label
<input checked="" type="checkbox"/>	1	Tonga
<input checked="" type="checkbox"/>	2	New Zealand
<input checked="" type="checkbox"/>	3	France
<input checked="" type="checkbox"/>	4	Wales

Move up

Move down

Reverse order

Close



Filter	Value	Label
<input checked="" type="checkbox"/>	Japan	Japan
<input checked="" type="checkbox"/>	New Zealand	New Zealand
<input checked="" type="checkbox"/>	Tonga	Tonga
<input checked="" type="checkbox"/>	Wales	Wales

Filter	Value	Label
<input checked="" type="checkbox"/>	Wales	Wales
<input checked="" type="checkbox"/>	Tonga	Tonga
<input checked="" type="checkbox"/>	New Zealand	New Zealand
<input checked="" type="checkbox"/>	Japan	Japan

If you need to edit the data in the spreadsheet, just double-click on a cell and the data should open up in the original spreadsheet, i.e. Excel. Once you have edited your data and saved the original spreadsheet, JASP will automatically update to reflect the changes that were made, provided that you have not changed the file name.



DATA EDITING IN JASP

JASP has its internal data editor, which allows users to either enter new data directly,



or to edit existing (external) data.



Clicking on this opens a new ribbon allowing the user to add new data directly or edit an open dataset. If an external dataset has been modified, the Synchronisation option will save those changes to the dataset directly.



Once the editing has been completed, you can return directly to the data analysis interface.



JASP ANALYSIS MENU



The **main** analysis options can be accessed from the main toolbar. Currently, JASP offers the following frequentist (parametric and non-parametric standard statistics) and alternative Bayesian tests:

Descriptives <ul style="list-style-type: none">• Descriptive stats• Raincloud plots• Time Series Descriptives• Flexplot	Regression <ul style="list-style-type: none">• Correlation• Linear regression• Logistic regression• Generalised Linear Model*
T-Tests <ul style="list-style-type: none">• Independent• Paired• One sample	Frequencies <ul style="list-style-type: none">• Binomial test• Multinomial test• Contingency tables• Log-linear regression*
ANOVA <ul style="list-style-type: none">• Independent• Repeated measures• ANCOVA• MANOVA *	Factor <ul style="list-style-type: none">• Principal Component Analysis (PCA)*• Exploratory Factor Analysis (EFA)*• Confirmatory Factor Analysis (CFA)*
Mixed Models* <ul style="list-style-type: none">• Linear Mixed Models• Generalised linear mixed models	Meta-Analysis <ul style="list-style-type: none">• Effect size computation• Meta-Analysis• Forest Plots

* Not covered in this guide

BY clicking on the icon on the top-right menu bar, you can also access advanced options that allow the addition of optional advanced modules. Once ticked, they will be added to the main analysis ribbon.

See the JASP website for more information on these advanced modules.



Once you have selected your required analysis, all the possible statistical options appear in the left window and the output appears in the right window.

JASP provides the ability to rename and ‘stack’ the results output, thereby organising multiple analyses.

The screenshot shows the JASP interface with the left sidebar expanded. It lists three analyses: "Descriptive Statistics", "ANOVA", and "Correlation". Each analysis has a set of four icons at the end of its row: a pen (edit), a green plus sign (copy), a blue information icon (info), and a red cross (delete). The "Correlation" entry is currently selected, indicated by a grey background.

- ▶ **Descriptive Statistics**
- ▶ **ANOVA**
- ▼ **Correlation**

The screenshot shows the JASP interface with the left sidebar expanded. The analyses listed are now renamed: "Descriptive Statistics England injuries", "ANOVA 5 countries played", and "Correlation -Spearmans injury correlation". The "Correlation" entry is still selected. The icons at the end of each row remain the same: edit, copy, info, and delete.

- ▶ **Descriptive Statistics England injuries**
- ▶ **ANOVA 5 countries played**
- ▼ **Correlation -Spearmans injury correlation**

The individual analyses can be renamed using the pen icon or deleted using the red cross.

Clicking on the analysis in this list will then take you to the appropriate part of the results output window. They can also be rearranged by dragging and dropping each of the analyses.

The green + icon produces a copy of the chosen analysis.

The screenshot shows the JASP interface with the left sidebar expanded. The analyses listed are: "Descriptive Statistics England injuries", "ANOVA 5 countries played", "Correlation -Spearmans injury correlation", and "Copy of Correlation -Spearmans injury correlation". The last entry is selected. The icons at the end of each row are identical: edit, copy, info, and delete.

- ▶ **Descriptive Statistics England injuries**
- ▶ **ANOVA 5 countries played**
- ▶ **Correlation -Spearmans injury correlation**
- ▼ **Copy of Correlation -Spearmans injury correlation**

The blue information icon provides detailed information on each of the statistical procedures used and includes a search option.



JASP Help

Correlation

The Correlation analysis allows estimation of the population correlation, as well as testing the null hypothesis that the population correlation between pairs of variables equals 0. All possible pairs of the specified variables are analyzed.

Assumptions (Pearson's rho)

- Continuous variables.
- The data are a random sample from the population.
- The pairs of variables follow a bivariate normal distribution in the population.
- The relationship between the pairs of variables is linear.

Assumptions (Spearman's rho & Kendall's tau)

- Ordinal or continuous variables.
- The data are a random sample from the population

Search for:



DESCRIPTIVE STATISTICS

Presentation of all the raw data is very difficult for a reader to visualise or draw any inference from. Descriptive statistics and related plots are a succinct way of describing and summarising data, but do not test any hypotheses. Various types of statistics are used to describe data:

- Measures of central tendency
- Measures of dispersion
- Percentile values
- Measures of distribution
- Descriptive plots

To explore these measures, load **Descriptive data.csv** into JASP. Go to Descriptives > Descriptive statistics and move the Variable data to the Variables box on the right.



You also have options to change and add tables in this section:

- Split analyses by a categorical variable (i.e., group)
- Transpose the main descriptive table (switch columns and rows)



The Statistics menu can now be opened to see the various options available.

▼ Statistics

Sample size

Valid
 Missing

Quantiles

Quartiles
 Cut points for: 4 equal groups
 Percentiles:

Central tendency

Mode
 Median
 Mean

Distribution

Skewness
 Kurtosis
 Shapiro-Wilk test
 Sum

Dispersion

Std. deviation
 MAD
 IQR
 Range
 Maximum

Coefficient of variation
 MAD robust
 Variance
 Minimum

Inference

S.E. mean
 Confidence interval for mean

Width 95.0 %
Method T model ▾

Confidence interval for std. deviation

Width 95.0 %
Method Analytical (chi-square) ▾

Confidence interval for variance

Width 95.0 %
Method Analytical (chi-square) ▾

Association matrix

Covariance
 Correlation
Use Everything ▾

CENTRAL TENDENCY.

This can be defined as the tendency for variable values to cluster around a central value. The three ways of describing this central value are mean, median and mode. If the whole population is considered, the term **population** mean / median/mode is used. If a sample/subset of the population is being analysed, the term **sample** mean/ median/mode is used. The measures of central tendency move toward a constant value when the sample size is sufficient to be representative of the population.



Central Tendency

- Mode
- Median
- Mean

Descriptive Statistics	
	Variable
Valid	810
Missing	0
Mean	17.71
Median	17.90
Mode	20.00

The **mean, M or \bar{x}** (17.71), is equal to the sum of all the values divided by the number of values in the dataset, i.e. the average of the values. It is used for describing continuous data. It provides a simple statistical model of the centre of distribution of the values and is a theoretical estimate of the ‘typical value’. However, it can be influenced heavily by ‘extreme’ scores.

The **median, Mdn** (17.9), is the middle value in a dataset that has been ordered from the smallest to the largest value and is the normal measure used for ordinal or non-parametric continuous data. Less sensitive to outliers and skewed data

The **mode** (20.0) is the most frequent value in the dataset and is usually the highest bar in a distribution histogram.

DISPERSION

Dispersion

- Std. deviation Coefficient of variation
- MAD MAD robust
- IQR Variance
- Range Minimum
- Maximum

Inference

- S.E. mean

Descriptive Statistics	
	Variable
Valid	810
Missing	0
Mean	17.711
Std. Error of Mean	0.244
Std. Deviation	6.935
Coefficient of variation	0.392
MAD	4.700
MAD robust	6.968
IQR	9.175
Variance	48.100

The **standard error of the mean, SE** (0.244), is a measure of how far the sample mean of the data is expected to be from the true population mean. As the size of the sample data grows larger, the SE decreases compared to S, and the true mean of the population is known with greater specificity.

Standard deviation, S or SD (6.935), is used to quantify the amount of dispersion of data values around the mean. A low standard deviation indicates that the values are close to the mean, while a high standard deviation indicates that the values are dispersed over a wider range.



The coefficient of variation (0.392) provides the relative dispersion of the data, in contrast to the standard deviation, which gives the absolute dispersion.

MAD, (4.7) median absolute deviation, a robust measure of the spread of data. It is relatively unaffected by data that is not normally distributed. Reporting median +/- MAD for data that is not normally distributed is equivalent to mean +/- SD for normally distributed data.

MAD Robust: (6.968) median absolute deviation of the data points, adjusted by a factor for asymptotically normal consistency.

IQR (9.175) Interquartile Range is similar to the MAD but is less robust (see Boxplots).

Variance (48.1) is another estimate of how far the data is spread from the mean. It is also the square of the standard deviation.

Confidence intervals (CI), although not shown in the general Descriptive statistics output, are used in many other statistical tests. When sampling from a population to get an estimate of the mean, confidence intervals are a range of values within which you are n% confident the true mean is included. A 95% CI is, therefore, a range of values that one can be 95% certain contains the true mean of the population. This is **not** the same as a range that contains 95% of **ALL** the values.

For example, in a normal distribution, 95% of the data are expected to be within ± 1.96 SD of the mean and 99% within ± 2.576 SD.

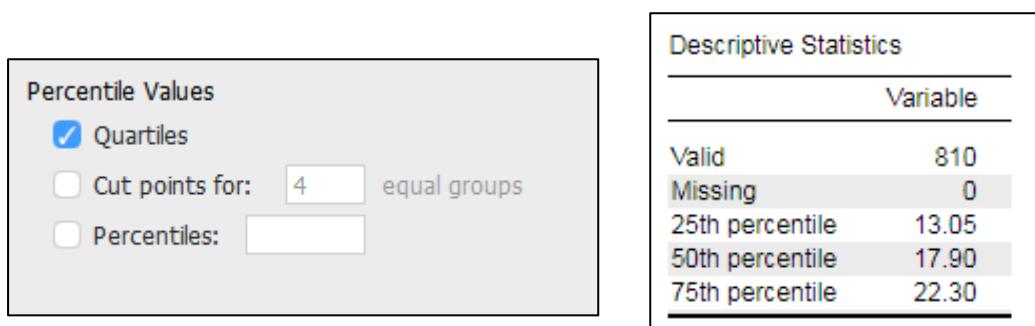
$$95\% \text{ CI} = M \pm 1.96 * \text{the standard error of the mean.}$$

Based on the data so far, $M = 17.71$, $SE = 0.24$; this will be $17.71 \pm (1.96 * 0.24)$ or 17.71 ± 0.47 .

Therefore, the 95% CI for this dataset is 17.24 - 18.18 and suggests that the true mean is likely to be within this range 95% of the time

QUARTILES

In the Statistics options, make sure that everything is unticked apart from Quartiles.





Quartiles are where datasets are split into 4 equal quarters, normally based on the rank ordering of median values. For example, in this dataset

1	1	2	2	3	3	4	4	4	4	5	5	5	6	7	8	8	9	10	10
				25%					50%					75%					

The median value that splits the data by 50% = 50th percentile = 5

The median value of left side = 25th percentile = 3

The median value of right side = 75th percentile = 8

From this, the Interquartile range (IQR) can be calculated, which is the difference between the 75th and 25th percentiles, i.e. 5. These values are used to construct the descriptive box plots later. The IQR can also be shown by ticking this option in the Dispersion menu.

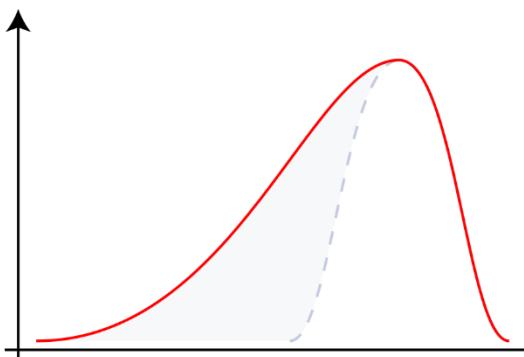
DISTRIBUTION

Distribution

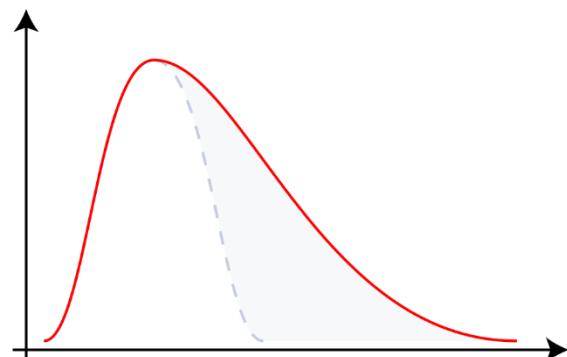
- Skewness
- Kurtosis
- Shapiro-Wilk test
- Sum

Descriptive Statistics		
	Variable	
Skewness	-0.004	*
Std. Error of Skewness	0.086	
Kurtosis	-0.410	
Std. Error of Kurtosis	0.172	
Shapiro-Wilk	0.996	
P-value of Shapiro-Wilk	0.032	
Sum	14346.200	

Skewness describes the shift of the distribution away from a normal distribution. Negative skewness shows that the mode moves to the right, resulting in a dominant left tail. Positive skewness shows that the mode moves to the left, resulting in a dominant right tail.



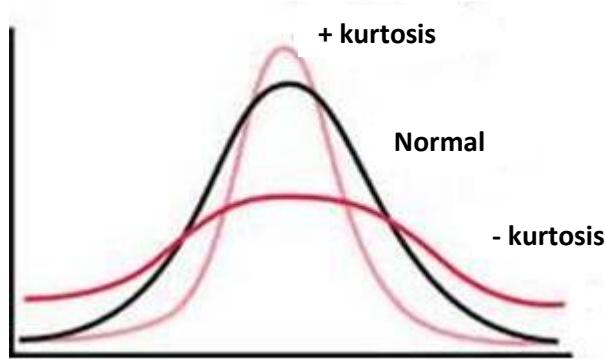
Negative skewness



Positive skewness



Kurtosis describes how heavy or light the tails are. Positive kurtosis results in an increase in the “pointiness” of the distribution with heavy (longer) tails, while negative kurtosis exhibits a much more uniform or flatter distribution with light (shorter) tails.



In the Statistics options, make sure that everything is unticked apart from skewness, kurtosis and the Shapiro-Wilk test.

We can use the Descriptives output to calculate skewness and kurtosis. For a normal data distribution, both values should be close to zero. The Shapiro-Wilk test is used to assess whether the data is significantly different from a normal distribution. (see - Exploring data integrity in JASP for more details).

INFERENCE

Confidence intervals provide a measure of how the data sample represents the population being studied. The probability that the confidence interval includes the true mean of the population is termed the confidence level and is typically set at 95%.

Inference

S.E. mean

Confidence interval for mean
Width 95.0 %
Method T model ▾

Confidence interval for std. deviation
Width 95.0 %
Method Analytical (chi-square) ▾

Confidence interval for variance
Width 95.0 %
Method Analytical (chi-square) ▾



SPLITTING DATA FILES

If there is a grouping variable (categorical or ordinal), descriptive statistics and plots can be produced for each group. Using **Descriptive data.csv** with the variable data in the Variables box, now add Group to the Split box.

The screenshot shows the JASP interface with two main panels. On the left is the 'Variables' panel, which contains a 'Variable' section with a pencil icon, a 'Split' section with a gear icon, and a 'Group' section with a gear and bar chart icon. On the right is the 'Descriptive Statistics' panel, which displays a table comparing Group 1 and Group 2 across various statistical measures:

	Variable	
	Group 1	Group 2
Mean	16.021	18.787
Std. Error of Mean	0.362	0.316
Std. Deviation	6.424	7.040
Skewness	0.200	-0.176
Std. Error of Skewness	0.137	0.110
Kurtosis	-0.101	-0.397
Std. Error of Kurtosis	0.274	0.219
Shapiro-Wilk	0.993	0.993
P-value of Shapiro-Wilk	0.119	0.029

DESCRIPTIVE DATA VISUALISATION

JASP produces a comprehensive range of descriptive and analysis-specific plots. These plots will be explained in their relevant chapters.

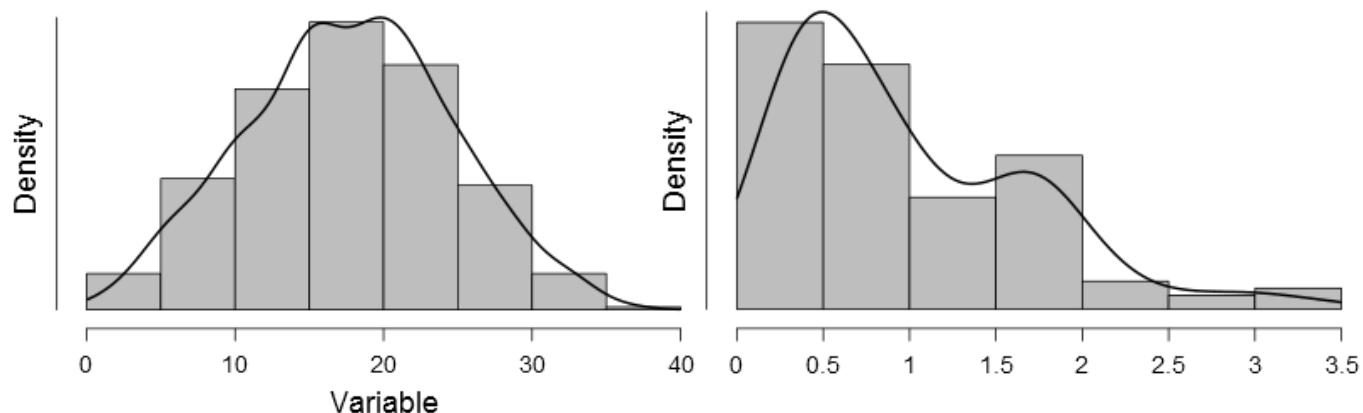
BASIC PLOTS

Firstly, to look at examples of the basic plots, open **Descriptive data.csv** with the variable data in the Variables box, go to Plots and tick Distribution plots, Display density, Interval plots, Q-Q plots, and dot plots.

The screenshot shows the 'Basic plots' settings dialog. It includes sections for 'Distribution plots' (checked), 'Correlation plots' (unchecked), 'Display density' (checked), 'Display rug marks' (unchecked), 'Bin width type' (set to 'Sturges'), 'Number of bins' (set to 30), 'Interval plots' (checked), 'Q-Q plots' (checked), 'Pie charts' (unchecked), 'Dot plots' (checked), 'Categorical plots' (unchecked), 'Pareto plots' (unchecked), 'Pareto rule' (set to 95.0%), 'Likert plots' (unchecked), 'Assume all variables share the same levels' (unchecked), and 'Adjustable font size for vertical axis' (set to 'Normal').

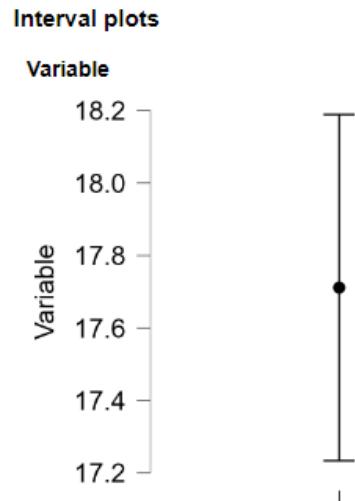
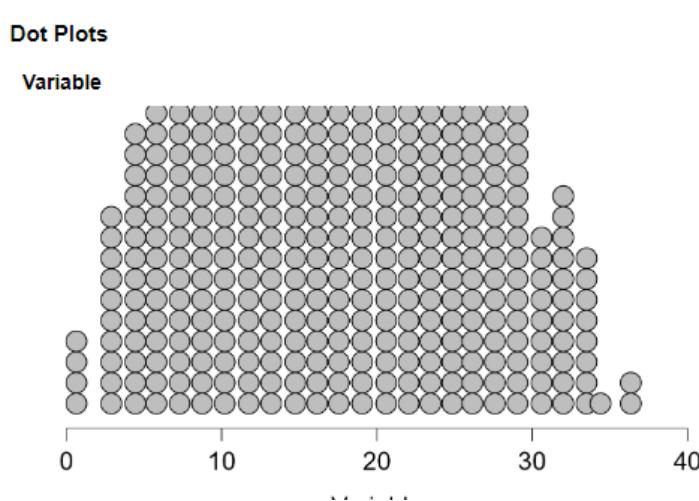


The Distribution plot is based on splitting the data into frequency bins, which are then overlaid with the distribution curve. As mentioned before, the highest bar is the mode (the most frequent value of the dataset). In this case, the curve looks approximately symmetrical, suggesting that the data is approximately normally distributed. The second distribution plot is from another dataset, which shows that the data is positively skewed.

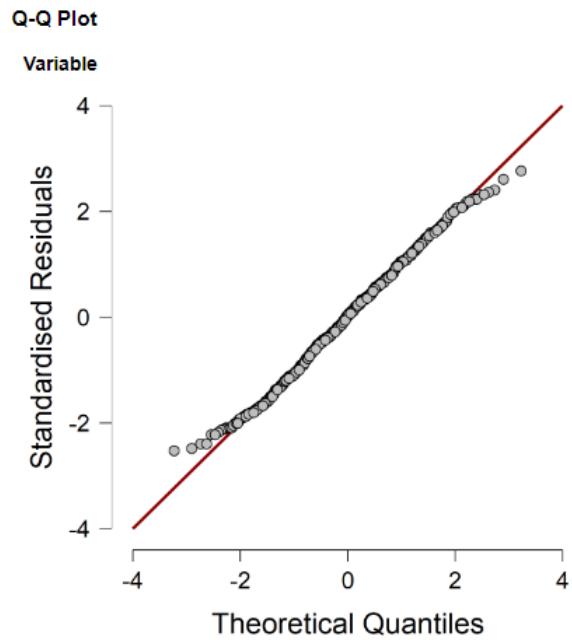


The dot plot displays the distribution where each dot represents a value. If a value occurs more than once, the dots are placed one above the other so that the height of the column of dots represents the frequency for that value.

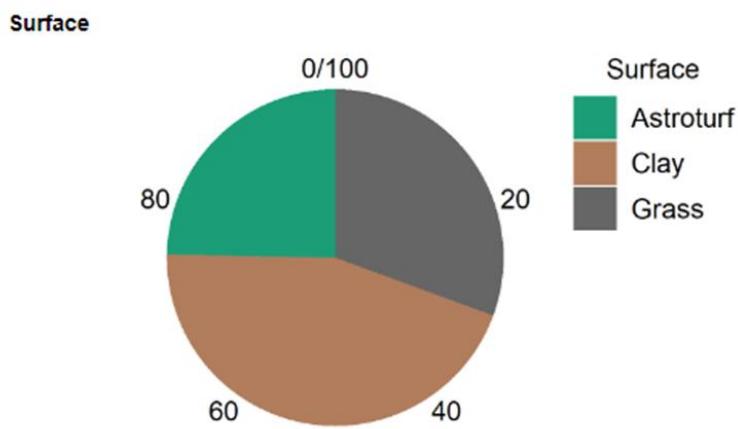
The interval plot shows a 95% confidence interval for the mean of each variable.



The Q-Q plot (quantile-quantile plot) can be used to visually assess if a set of data comes from a normal distribution. Q-Q plots take the sample data, sort it in ascending order, and then plot it against quantiles (percentiles) calculated from a theoretical distribution. If the data is normally distributed, the points will fall on or close to the 45-degree reference line. If the data is not normally distributed, the points will deviate from the reference line.



Depending on the data sets, basic correlation graphs and pie charts for non-scale data can also be produced, as can Pareto and Likert plots.





CUSTOMISABLE PLOTS

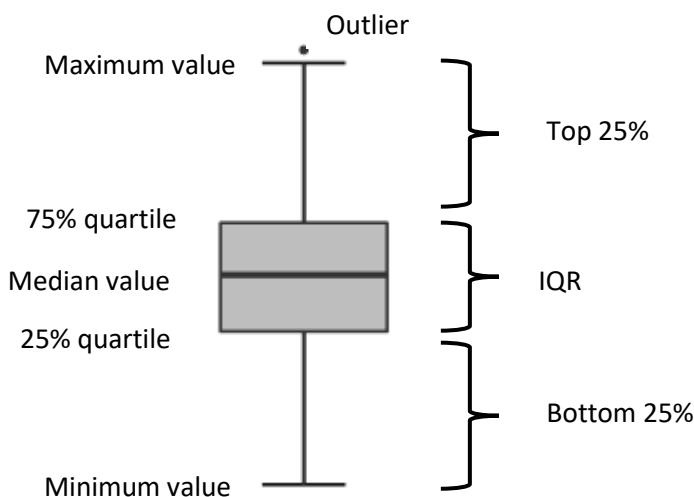
The screenshot shows the 'Customizable plots' settings window in JASP. It includes sections for Boxplots, Scatter plots, Density plots, and Tile heatmaps.

- Boxplots:** Includes options for Boxplot element (checked), Use color palette (checked), Violin element (unchecked), Label outliers (unchecked), and Jitter element (unchecked).
- Scatter plots:** Includes Graph above scatter plot (Density selected), Graph right of scatter plot (Density selected), Show legend (checked), Add regression line (checked), Smooth (selected), Linear (unchecked), and Show confidence interval (checked) with a value of 95.0%.
- Density plots:** Includes a checkbox for Display density plots (unchecked) and a Transparency slider set to 20.
- Separate densities:** A section with a button to add variables to the list.
- Tile heatmaps for selected variables:** Includes a checkbox for Display legend (unchecked), a checkbox for Display value (unchecked), a Relative text size slider set to 1, and a Width to height ratio of tiles slider set to 1.
- Horizontal axis:** A list for selecting variables.
- Vertical axis:** A list for selecting variables.

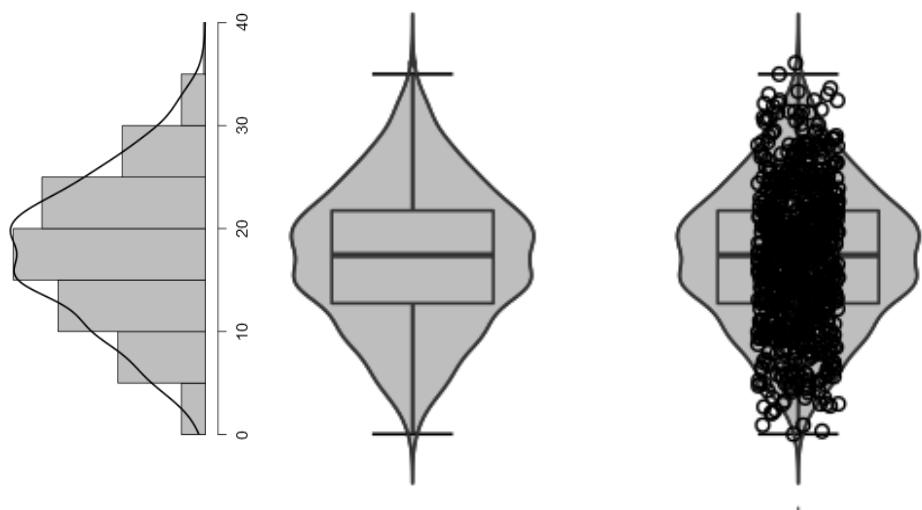
There are a variety of options depending on your datasets.

The boxplots visualise several statistics described above in one plot:

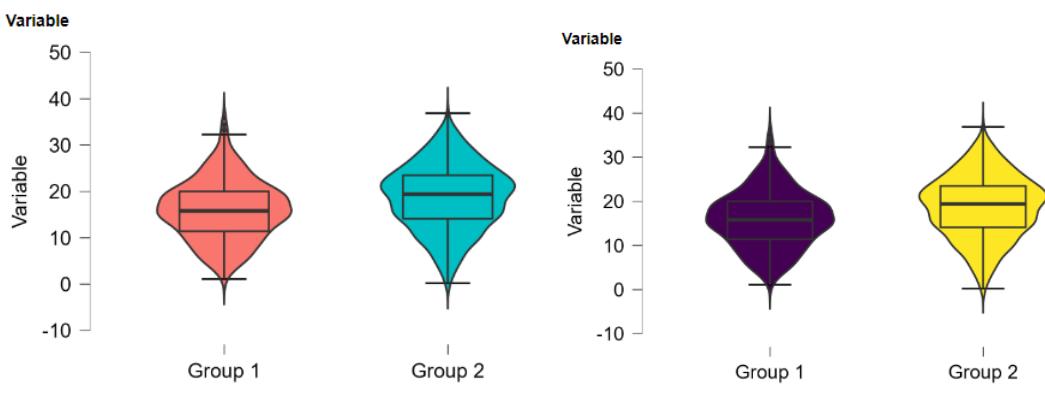
- Median value
- 25 and 75% quartiles
- Interquartile range (IQR) i.e., 75% - 25% quartile values
- Maximum and minimum values plotted with outliers excluded
- Outliers are shown if requested



Go back to the statistics options, in Descriptive plots, tick both Boxplot and Violin Element, and look at how the plot has changed. Next, tick Boxplot, Violin and Jitter Elements. The Violin plot has taken the smoothed distribution curve from the Distribution plot, rotated it 90° and superimposed it on the boxplot. The jitter plot has further added all the data points.



If your data is split by group, for example, the boxplots for each group will be shown on the same graph, and the colours of each will be different if the Colour palette is ticked. 5 colour palettes are available.

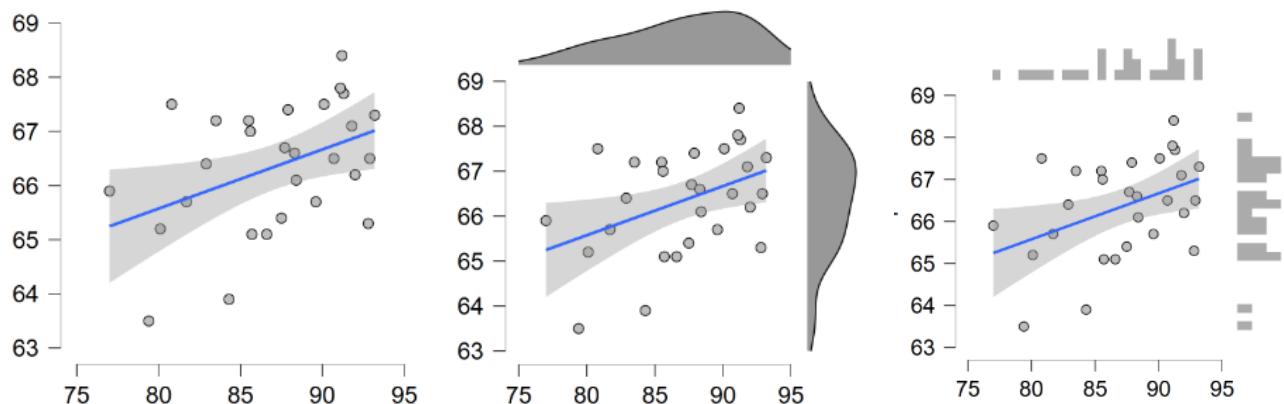


GGplot2 palette

Viridis palette

Scatter Plots

JASP can produce scatterplots of various types and can include smooth or linear regression lines. There are also options to add distributions to these, either in the form of density plots or histograms.





Tile Heatmap

These plots provide an alternative way of visualising data. For example, using the Titanic survival dataset to look at the relationship between the class of passage and survival.

Tile Heatmap

Horizontal axis: **survived**

Vertical axis: **Class**

Display legend

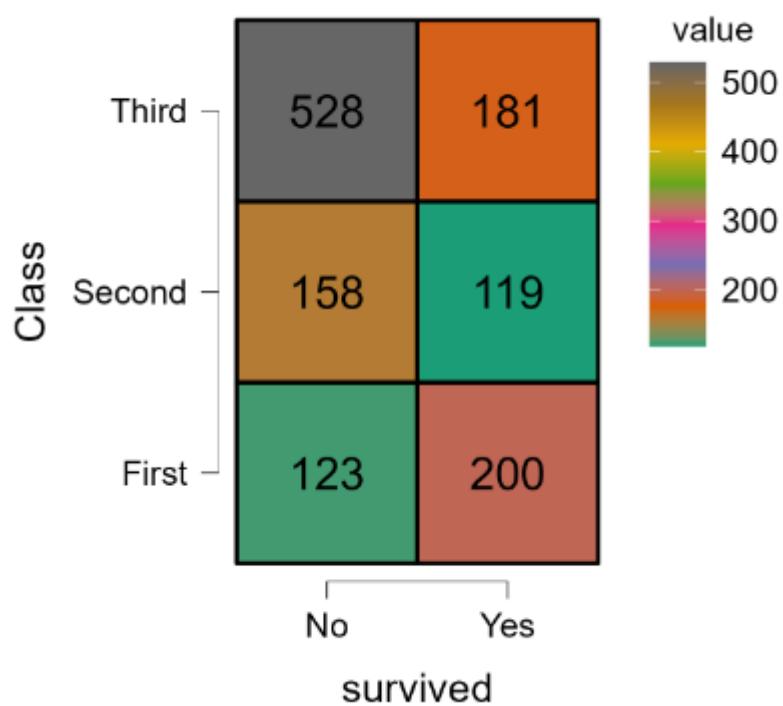
Display value

Relative text size

Width to height ratio of tiles

Statistic to plot

For scale variables	For nominal and ordinal variables
<input checked="" type="radio"/> Mean	<input type="radio"/> Mode
<input type="radio"/> Median	<input type="radio"/> Value itself
<input type="radio"/> Value itself	<input checked="" type="radio"/> Number of observations
<input type="radio"/> Number of observations	





EDITING PLOTS

Clicking on the drop-down menu provided access to a range of options, including Edit Image. Selecting this option provides some customisation for each graph.

Boxplots

Variable ▾

- 40 Copy
- Copy Citations
- Save Image As
- 30 Edit Image

This will open the plot in a new window, which allows some modifications of each axis in terms of axis title and range.

The image shows two vertically stacked dialog boxes for modifying plot axes. Both boxes have tabs for 'x-axis' and 'y-axis' at the top, and a blue info icon on the right.

Top Dialog (Initial State):

- Show title
- Title: Group
-

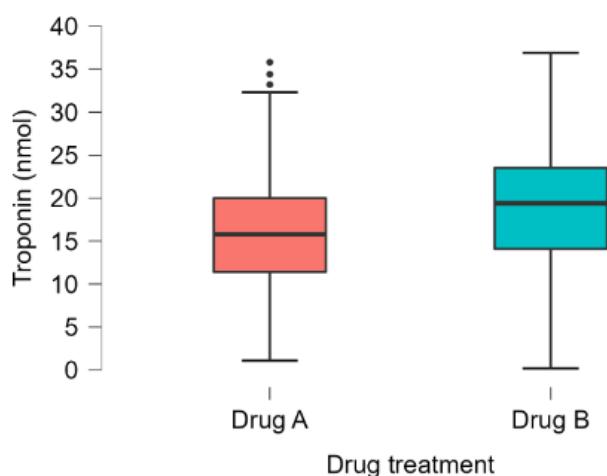
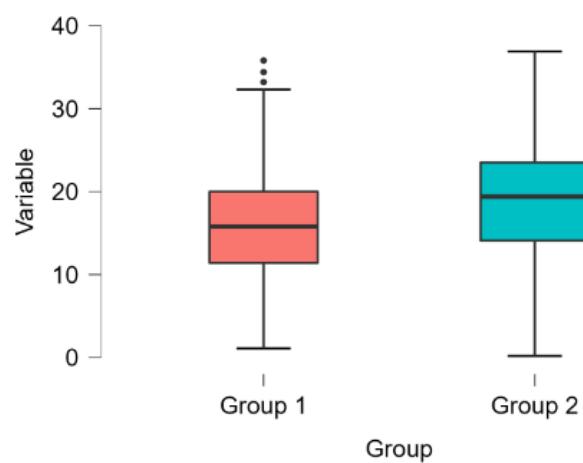
Bottom Dialog (Advanced State):

- Show title
- Title: Variable
- Show axis
- Ticks:
 - Specify sequence
 - Set manually
- from: 0
- to: 40
- steps: 10
-

Any changes are then updated in the results window. The new plot can be saved as an image or can be reset to default values.



Do not forget that group labels can be changed in the spreadsheet editor.





EXPLORING DATA INTEGRITY

Sample data is used to estimate parameters of the population, whereby a parameter is a measurable characteristic of a population, such as a mean, standard deviation, standard error or confidence intervals, etc.

What is the difference between a statistic and a parameter? If you randomly polled a selection of students about the quality of their student bar, you may find that 75% of them were happy with it. That is a sample **statistic** since only a sample of the population was asked. You calculated what the population was likely to do based on the sample. If you asked **all** the students in the university, and 90% were happy, you have a **parameter** since you asked the whole university population.

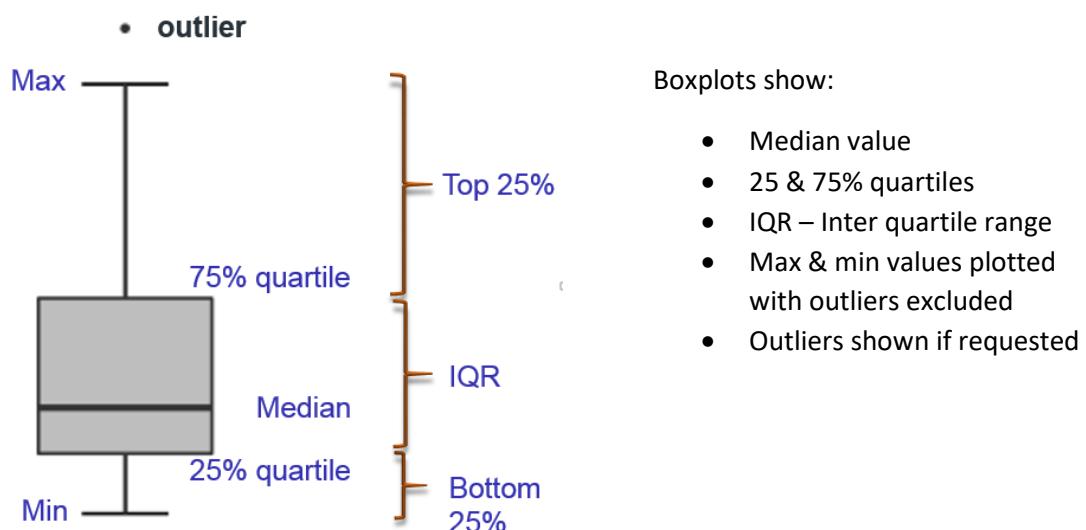
Bias can be defined as the tendency of a measurement to over- or underestimate the value of a population parameter. Many types of bias can appear in research design and data collection, including:

- Participant selection bias – some being more likely to be selected for study than others
- Participant exclusion bias - due to the systematic exclusion of certain individuals from the study
- Analytical bias - due to the way that the results are evaluated

However, statistical bias can affect a) parameter estimates, b) standard errors and confidence intervals or c) test statistics and p values. So, how can we check for bias?

IS YOUR DATA CORRECT?

Outliers are data points that are abnormally outside all other data points. Outliers can be due to a variety of things, such as errors in data input or analytical errors at the point of data collection. Boxplots are an easy way to visualise such data points where outliers are outside the upper ($75\% + 1.5 * \text{IQR}$) or lower ($25\% - 1.5 * \text{IQR}$) quartiles





Load **Exploring Data.csv** into JASP. Under Descriptives > Descriptive Statistics, add Variable 1 to the Variables box. In Plots, tick the following: Boxplots, Label Outliers, and BoxPlot Element.

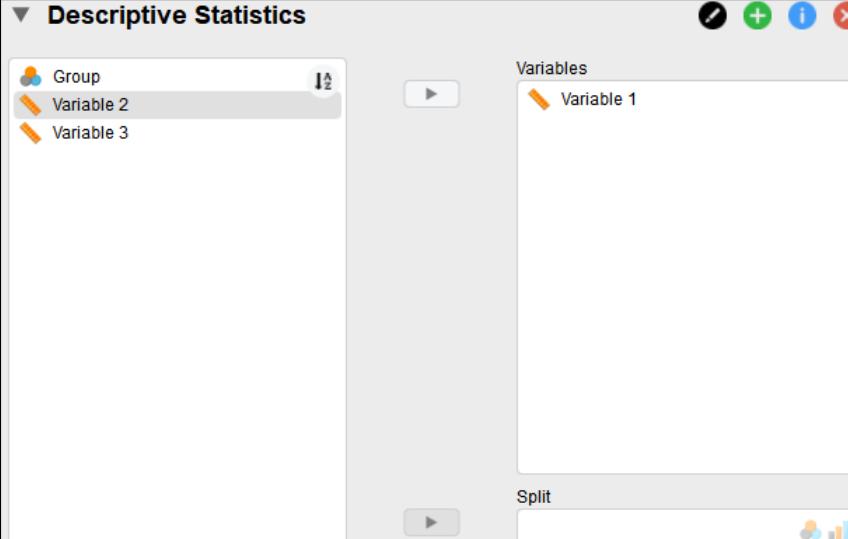
Descriptive Statistics

Variables

Group
Variable 2
Variable 3

Variable 1

Split



Customizable plots

Color palette Colorblind ▾

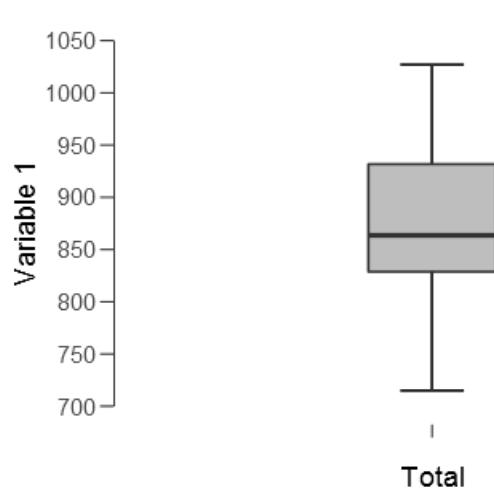
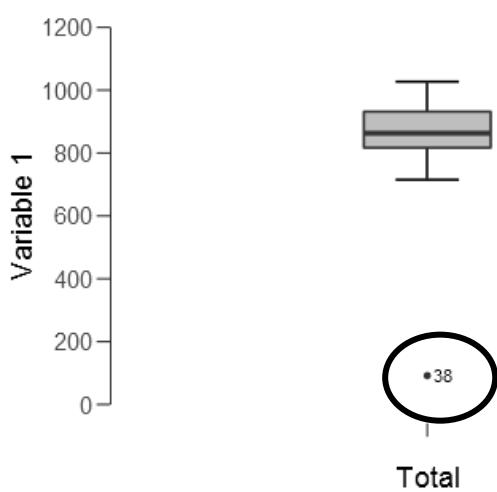
Boxplots

Boxplot element Use color palette

Violin element

Label outliers

Jitter element



The resulting Boxplot on the left looks very compressed, and an obvious outlier is labelled as being in row 38 of the dataset. This can be traced back to a data input error in which 91.7 was input instead of 917. The graph on the right shows the BoxPlot for the 'clean' data.



How you deal with an outlier depends on the cause. Most parametric tests are highly sensitive to outliers, while non-parametric tests are generally not.

Correct it? – Check the original data to make sure that it isn't an input error; if it is, correct it, and rerun the analysis.

Keep it? - Even in datasets of normally distributed data, outliers may be expected for large sample sizes and should not automatically be discarded if that is the case.

Delete it? – This is a controversial practice in small datasets where a normal distribution cannot be assumed. Outliers resulting from an instrument reading error may be excluded, but they should be verified first.

Replace it? – Also known as winsorizing. This technique replaces the outlier values with the relevant maximum and/or minimum values found after excluding the outlier.

Whatever method you use must be justified in your statistical methodology and subsequent analysis.

WE MAKE MANY ASSUMPTIONS ABOUT OUR DATA.

When using parametric tests, we make a series of assumptions about our data and bias will occur if these assumptions are violated:

- Normality
- Homogeneity of variance or homoscedasticity

Many statistical tests are an omnibus of tests, of which some will check these assumptions.

TESTING THE ASSUMPTION OF NORMALITY

Normality does not necessarily mean that the data is normally distributed per se, but it is whether or not the dataset can be modelled by a normal distribution. Normality can be explored in a variety of ways:

- Numerically
- Visually / graphically
- Statistically

Numerically, we can use the Descriptives output to calculate skewness and kurtosis. For a normal data distribution, both values should be close to zero. To determine the significance of skewness or kurtosis, we calculate their z-scores by dividing them by their associated standard errors:

$$\text{Skewness } z = \frac{\text{skewness}}{\text{Skewness standard error}} \qquad \text{Kurtosis } z = \frac{\text{kurtosis}}{\text{kurtosis standard error}}$$

Z score significance: $p < 0.05$ if $z > 1.96$ $p < 0.01$ if $z > 2.58$ $p < 0.001$ if $z > 3.29$



Using **Exploring data.csv**, go to Descriptives>Descriptive Statistics move Variable 3 to the Variables box, in the Statistics drop-down menu select Mean, Std Deviation, Skewness and Kurtosis as shown below with the corresponding output table.

▼ Statistics

Sample size

Valid

Missing

Central tendency

Mode

Median

Mean

Quantiles

Quartiles

Cut points for: equal groups

Percentiles:

Distribution

Skewness

Kurtosis

Shapiro-Wilk test

Sum

Descriptive Statistics

	Variable 3
Valid	50
Missing	0
Mean	0.893
Std. Deviation	0.673
Skewness	0.839
Std. Error of Skewness	0.337
Kurtosis	-0.407
Std. Error of Kurtosis	0.662

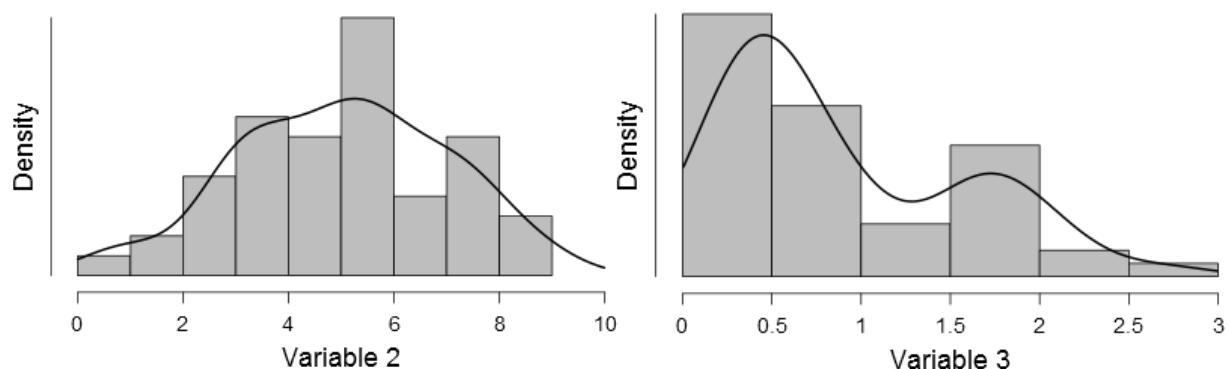
Neither skewness nor kurtosis is close to 0. The positive skewness suggests that the data is distributed more on the left (see graphs later), while the negative kurtosis suggests a flat distribution. When calculating their z-scores, it can be seen that the data is significantly skewed, $p < 0.05$.

$$\text{Skewness } Z = \frac{0.839}{0.337} = 2.49$$

$$\text{Kurtosis } Z = \frac{-0.407}{0.662} = 0.614$$

[As a note of caution, skewness and kurtosis may appear significant in large datasets even though the distribution is normal.]

Now add Variable 2 to the Variables box, and in Plots, tick Distribution plot. This will show the following two graphs:

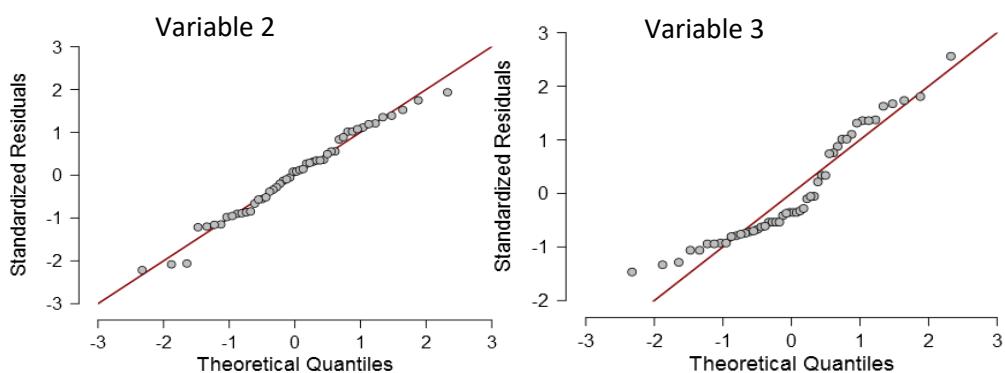




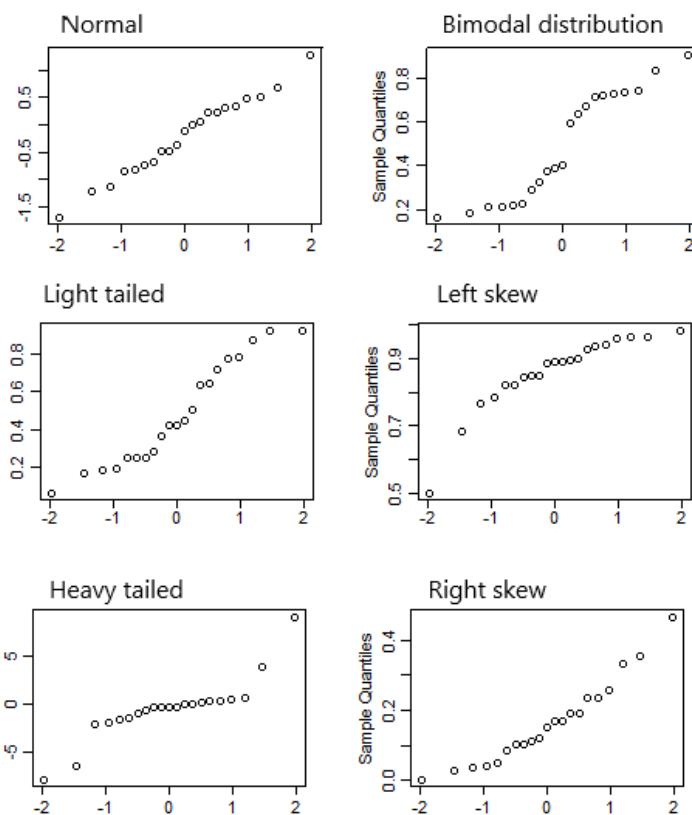
It is quite easy to visualise that Variable 2 has a symmetrical distribution. Variable 3 is skewed to the left, as confirmed by the skewness Z score.

Another graphical check for normality is a Q-Q plot. Q-Q plots are available in Descriptives and are also produced as part of the Assumption Checks used in **linear regression** and **ANOVA**. Q-Q plots show the quantiles of the actual data against those expected for a normal distribution.

If the data are normally distributed, all the points will be close to the diagonal reference line. If the points 'sag' above or below the line, there is a problem with kurtosis. If the points snake around the line, then the problem is skewness. Below are Q-Q plots for Variables 2 and 3. Compare these to the previous distribution plots and the skewness/kurtosis z scores above.



The following Q-Q plot scenarios are possible:





The Shapiro-Wilk test is a statistical test used by JASP to check the assumption of normality. It is also used in the **Independent** (distribution of the two groups) and **Paired** (distribution of differences between pairs) **t-tests**. The test results in a W value, where small values indicate your sample is not normally distributed (the null hypothesis that your population is normally distributed, if your values are under a certain threshold, can, therefore, be rejected).

Descriptive Statistics		
	Variable 2	Variable 3
Valid	50	50
Missing	0	0
Shapiro-Wilk	0.982	0.885
P-value of Shapiro-Wilk	0.650	< .001

In Descriptives, the Shapiro-Wilk test can be selected in the Distribution tests. The Shapiro-Wilk output table shows no significant deviation in normality for Variable 2 but a significant deviation ($p<.001$) for Variable 3.

The most important limitation is that the test can be biased by sample size. The larger the sample, the more likely you'll get a statistically significant result.

Testing the assumption of normality – A cautionary note!

For most parametric tests to be reliable, one of the assumptions is that the data is **approximately** normally distributed. A normal distribution peaks in the middle and is symmetrical about the mean. However, data does not need to be perfectly normally distributed for the tests to be reliable.

So, having gone on about testing for normality, is it necessary?

The Central Limit Theorem states that as the sample size gets larger, i.e. >30 data points, the distribution of the sampling means approaches a normal distribution. So the more data points you have, the more normal the distribution will look and the closer your sample mean approximates the population mean.

Large datasets may result in significant tests of normality, i.e. Shapiro-Wilk or significant skewness and kurtosis z-scores when the distribution graphs look fairly normal. Conversely, small datasets will reduce the statistical power to detect non-normality.

However, data that does not meet the assumption of normality is going to result in poor results for certain types of tests (i.e. ones that state that the assumption must be met!). How closely does your data need to be normally distributed? This is a judgment call best made by eyeballing the data.



WHAT DO I DO IF MY DATA IS NOT NORMALLY DISTRIBUTED?

Transform the data and redo the normality checks on the transformed data. Common transformations include taking the log or square root of the data. Please refer to the next chapter.

Use non-parametric tests since these are distribution-free tests and can be used instead of their parametric equivalent.

TESTING HOMOGENEITY OF VARIANCE

Levene's test is commonly used to test the null hypothesis that variances in different groups are equal. The result from the test (F) is reported as a p-value; if not significant, then you can say that the null hypothesis stands — that the variances are equal; if the p-value is significant, then the implication is that the variances are unequal. Levene's test is included in the **Independent t-test** and **ANOVA** in JASP as part of the Assumption Checks.

Using **Exploring data.csv**, go to T-Tests>Independent Samples t-test, move Variable 1 to the Variables box and Group to the Grouping variable and tick Assumption Checks > Equality of variances.

▼ Independent Samples T-Test

Variable 2
Variable 3

▶

Dependent Variables

Variable 1

Grouping Variable

Group

Assumption Checks

Normality

Equality of variances

Brown-Forsythe

Levene's

Missing Values

Exclude cases per dependent variable

Exclude cases listwise

Test of Equality of Variances (Levene's)			
	F	df	p
Variable 1	0.218	1	0.643



In this case, there is no significant difference in variance between the two groups, $F(1) = 0.218$, $p = .643$.

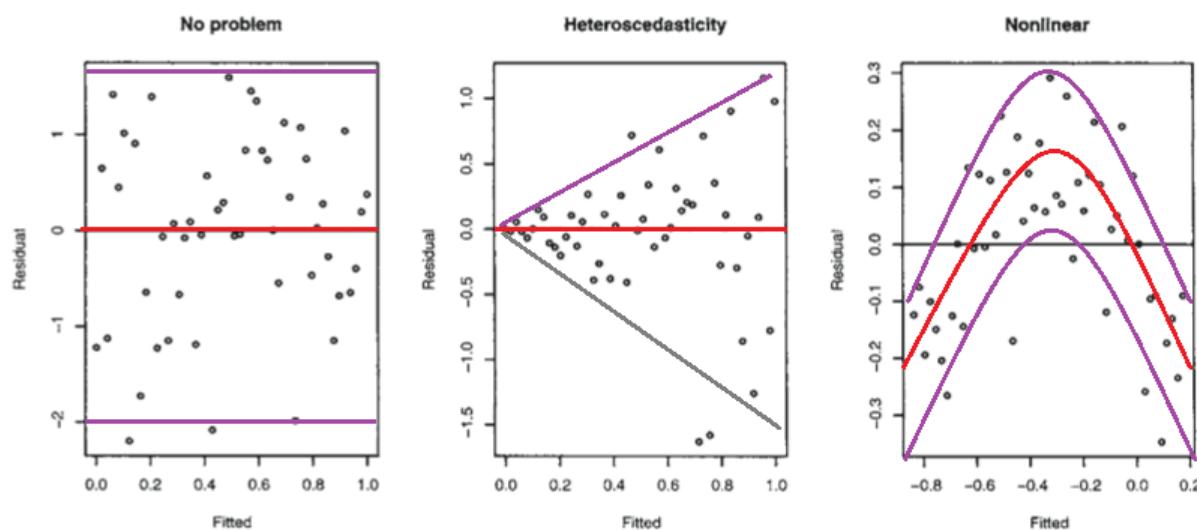
The assumption of homoscedasticity (equal variance) is important in **linear regression** models, as is linearity. It assumes that the variance of the data around the regression line is the same for all predictor data points. Heteroscedasticity (the violation of homoscedasticity) is present when the variance differs across the values of an independent variable. This can be visually assessed in linear regression by plotting actual residuals against predicted residuals.

Plots

Residuals Plots

- Residuals vs. dependent
- Residuals vs. covariates
- Residuals vs. predicted
- Residuals vs. histogram
- Standardized residuals
- Q-Q plot standardized residuals
- Partial plots

If homoscedasticity and linearity are not violated, there should be no relationship between what the model predicts and its errors, as shown in the graph on the left. Any sort of funnelling (middle graph) suggests that homoscedasticity has been violated, and any curve (right graph) suggests that linearity assumptions have not been met.





DATA TRANSFORMATION

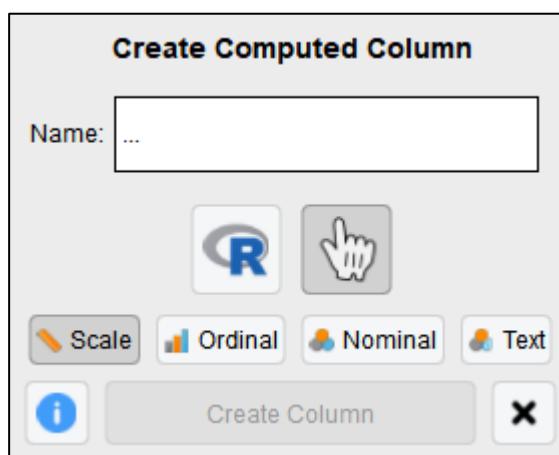
JASP provides the ability to compute new variables or transform data. In some cases, it may be useful to compute the differences between repeated measures, or, to make a dataset more normally distributed, you can apply a log transform, for example.

When a dataset is opened, there will be a plus sign (+) at the end of the columns.

	Group	Variable 1	Variable 2	Variable 3	
1	1	912	2.78	0.29	
2	1	826	4.89	0.55	
3	1	1004	6.79	0.47	
4	1	982	6.24	1.58	
5	1	920	8.59	0.76	
6	1	814	5.86	0.76	

Clicking on the + opens a small dialogue window where you can;

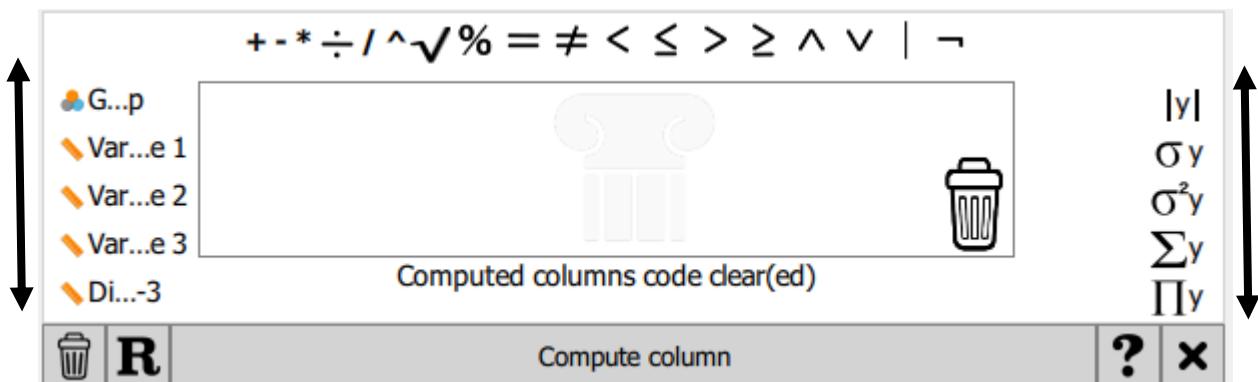
- Enter the name of a new variable or the transformed variable
- Select whether you enter the R code directly or use the commands built into JASP
- Select what data type is required



Once you have named the new variable and chosen the other options, click create.



If you choose the manual option rather than the R code, this opens all the built-in create and transform options. Although not obvious, you can scroll the left and right-hand options to see more variables or more operators, respectively.



For example, we want to create a column of data showing the difference between variable 2 and variable 3. Once you have entered the column name in the Create Computed Column dialogue window, its name will appear in the spreadsheet window. The mathematical operation now needs to be defined. In this case, drag variable 2 into the equation box, drag the 'minus' sign down and then drag in variable 3.

The screenshot shows the 'Compute column' dialog box with the title 'Diff 2-3'. The equation editor at the top shows the expression $\text{Var...e 2} - \text{Var...e 3}$. Red arrows point from the variable names 'Var...e 2' and 'Var...e 3' in the list on the left towards the minus sign in the equation editor. The rest of the interface is identical to the first screenshot, including the toolbar, variable list, trash bin, and bottom buttons.

	Group	Variable 1	Variable 2	Variable 3	f _x Diff 2-3	
1	1	912	2.78	0.29		
2	1	826	4.89	0.55		
3	1	1004	6.79	0.47		

If you have made a mistake, i.e. used the wrong variable or operator, remove it by dragging the item into the dustbin in the bottom right corner.



When you are happy with the equation/operation, click ‘compute column’ and the data will be entered.

Diff 2-3

$+ - * \div / ^ \sqrt \% = \neq < \leq > \geq \wedge \vee | \neg$

G...p
Var...e 1
Var...e 2
Var...e 3
Di...-3

Var...e 2 - Var...e 3

Computed columns code applied

|y|
 σ_y
 σ^2_y
 $\sum y$
 $\prod y$

	R	Compute column				?	X
T	Group	Variable 1	Variable 2	Variable 3	Diff 2-3	+	
1	1	912	2.78	0.29	2.49		
2	1	826	4.89	0.55	4.34		
3	1	1004	6.79	0.47	6.32		

If you decide that you do not want to keep the derived data, you can remove the column by clicking the other dustbin icon next to the R.

Another example is to do a log transformation of the data. In the following case, variable 1 has been transformed by scrolling the operators on the left and selecting the $\log(y)$ option. Replace the “y” with the variable that you want to transform, and then click ‘Compute column’. When finished, click the X to close the dialogue.

Log10 Variable 1

$+ - * \div / ^ \sqrt \% = \neq < \leq > \geq \wedge \vee | \neg$

G...p
Var...e 1
Var...e 2
Var...e 3
Log10...ble 1

$\log(\text{Var...e 1})$

Computed columns code applied

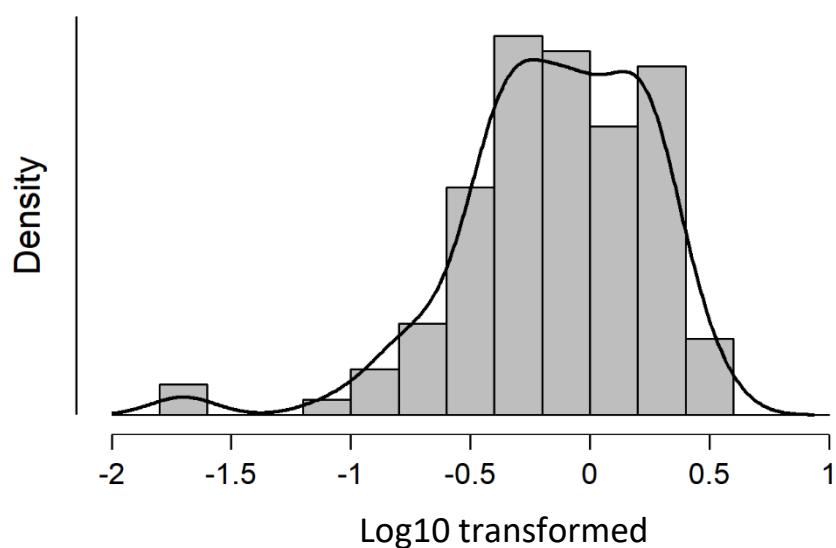
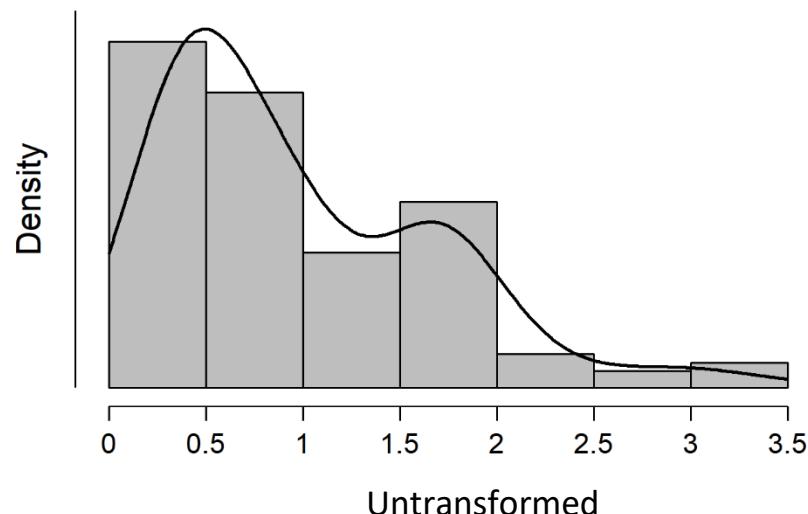
log(y)
log2(y)
log10(y)
logb(y)
exp(y)

	R	Compute column				?	X
T	Group	Variable 1	Variable 2	Variable 3	Log10 Variable 1	+	
1	1	912	2.78	0.29	2.95999		
2	1	826	4.89	0.55	2.91698		
3	1	1004	6.79	0.47	3.00173		

The Export function will also export any new data variables that have been created.



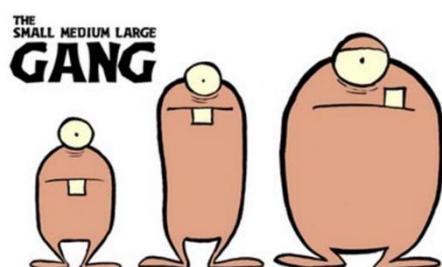
The two graphs below show the untransformed and the log10-transformed data. The skewed data has been transformed into a profile with a more normal distribution.





EFFECT SIZE

When performing a hypothesis test on data, we determine the relevant statistic (r , t , F , etc) and p-value to decide whether to accept or reject the null hypothesis. A small p-value, <0.05 in most analyses, provides evidence against the null hypothesis, whereas a large p-value >0.05 only means that there is not enough evidence to reject the null hypothesis. A lower p-value is sometimes incorrectly interpreted as meaning there is a stronger relationship or difference between variables. So what is needed is not just null hypothesis testing but also a method of determining precisely how large the effects seen in the data are.



An effect size is a statistical measure used to determine the strength of the relationship or difference between variables. Unlike a p-value, effect sizes can be used to quantitatively compare the results of different studies.

For example, comparing heights between 11 and 12-year-old children may show that the 12-year-olds are significantly taller, but it is difficult to visually see a difference, i.e. small effect size. However, a significant difference in height between 11 and 16-year-old children is obvious (large effect size).

The effect size is usually measured in three ways:

- The standardised mean difference
- correlation coefficient
- odds ratio

When looking at differences between groups, most techniques are primarily based on the differences between the means divided by the average standard deviations. The values derived can then be used to describe the magnitude of the differences. The effect sizes calculated in JASP for t-tests and ANOVA are shown below:

Test	Measure	Trivial	Small	Medium	Large
Between means Parametric	Cohen's d	<0.2	0.2	0.5	0.8
	Hedge's g	<0.2	0.2	0.5	0.8
	Rank -biserial (r_B)	<0.1	0.1	0.3	0.5
ANOVA	Eta squared (η^2)	<0.1	0.1	0.25	0.37
	Partial Eta squared (η^2_p)	<0.01	0.01	0.06	0.14
	Omega squared (ω^2)	<0.01	0.01	0.06	0.14



When analysing bivariate or multivariate relationships, the effect sizes are the correlation coefficients:

Test	Measure	Trivial	Small	Medium	Large
Correlation	Correlation coefficient (r)	<0.1	0.1	0.3	0.5
	Spearman's rho	<0.1	0.1	0.3	0.5
	Kendall's tau	<0.1	0.1	0.3	0.5
Multiple Regression	Multiple correlation coefficient (R)	<0.1	0.1	0.3	0.5

When analysing categorical relationships via contingency tables, i.e. chi-square test, Phi is only used for 2x2 tables while Cramer's V can be used for any table size.

Contingency tables	df	Small	Medium	Large
Phi (2x2 only)	1	0.1	0.3	0.5
Cramer's V	2	0.07	0.21	0.35
Cramer's V	3	0.06	0.17	0.29
Cramer's V	4	0.05	0.15	0.25
Cramer's V	5	0.04	0.13	0.22

For a 2×2 contingency table, we can also define the odds ratio measure of effect size.



ONE-SAMPLE T-TEST

Research is normally carried out in sample populations, but how closely does the sample reflect the whole population? The parametric one-sample t-test determines whether the sample mean is statistically different from a known or hypothesised population mean.

The null hypothesis (H_0) tested is that the sample mean is equal to the population mean.

ASSUMPTIONS

Three assumptions are required for a one-sample t-test to provide a valid result:

- The test variable should be measured on a **continuous** scale.
- The test variable data should be **independent**, i.e. no relationship between any of the data points.
- The data should be approximately **normally distributed**.
- There should be no significant **outliers**.

The screenshot shows the JASP software interface for performing a One Sample T-Test. The main window title is "One Sample T-Test". On the left, there is a list of variables: "mass" and "height". The "Variables" section is currently empty. Below the variables, the "Tests" section is expanded, showing the following options:

- Student
- Wilcoxon signed-rank
- Z Test

Below these are input fields for "Test value" (set to 0) and "Std. deviation" (set to 1). The "Alternative Hypothesis" section contains three radio buttons:

- ≠ Test value
- > Test value
- < Test value

The "Assumption checks" section includes two checkboxes:

- Normality
- Q-Q plot residuals

The "Missing Values" section has two radio buttons:

- Exclude cases per variable
- Exclude cases listwise

On the right side of the dialog, the "Additional Statistics" section is expanded, showing:

- Location estimate
 - Confidence interval 95.0 %
- Effect size
 - Confidence interval 95.0 %
- Descriptives
- Vovk-Sellke maximum p-ratio

The "Plots" section is also expanded, showing:

- Descriptives plots
 - Confidence interval 95.0 %
- Raincloud plots
 - Horizontal display
- Bar plots
 - Confidence interval 95.0 %
 - Standard error
 - Fix horizontal axis to 0

At the top right of the dialog are several icons: a blue "R", a black pencil, a green circular arrow, a blue info icon, and a red X.



RUNNING THE ONE-SAMPLE T-TEST

Open **one sample t-test.csv**, which contains two columns of data representing the height (cm) and body mass (kg) of a sample population of males used in a study. In 2017, the average adult male in the UK population was **178 cm** tall and had a body mass of **83.4 kg**.

Go to T-Tests > One-Sample t-test and in the first instance, add height to the analysis box on the right. Then tick the following options above and add **178** as the test value.

UNDERSTANDING THE OUTPUT

The output should contain three tables and two graphs.

Test of Normality (Shapiro-Wilk)

	W	p
height	0.983	0.954

Note. Significant results suggest a deviation from normality.

The assumption check of normality (Shapiro-Wilk) is not significant, suggesting that the heights are normally distributed; therefore, this assumption is not violated. If this showed a significant difference, the analysis should be repeated using the non-parametric equivalent, **Wilcoxon's signed-rank test**, tested against the population median height.

One Sample T-Test

	t	df	p	Mean Difference	Cohen's d	SE Cohen's d
height	-0.382	22	0.706	-0.391	-0.080	0.209

Note. For the Student t-test, effect size is given by Cohen's d.

Note. For the Student t-test, location difference estimate is given by the sample mean difference d.

Note. For the Student t-test, the alternative hypothesis specifies that the mean is different from 178.

Note. Student's t-test.

This table shows that there are no significant differences between the means, $p = .706$

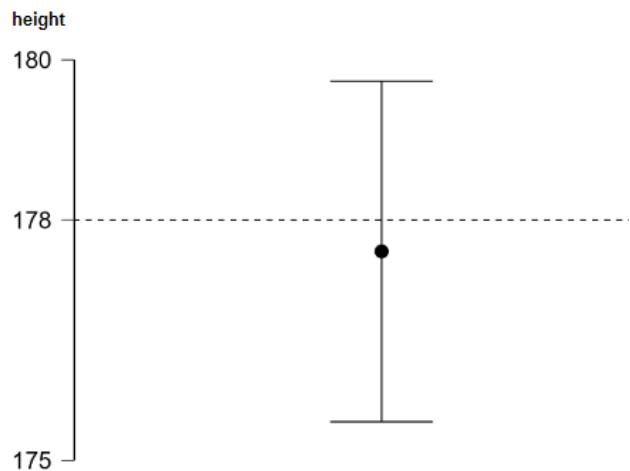
Descriptives

	N	Mean	SD	SE
height	23.000	177.609	4.915	1.025

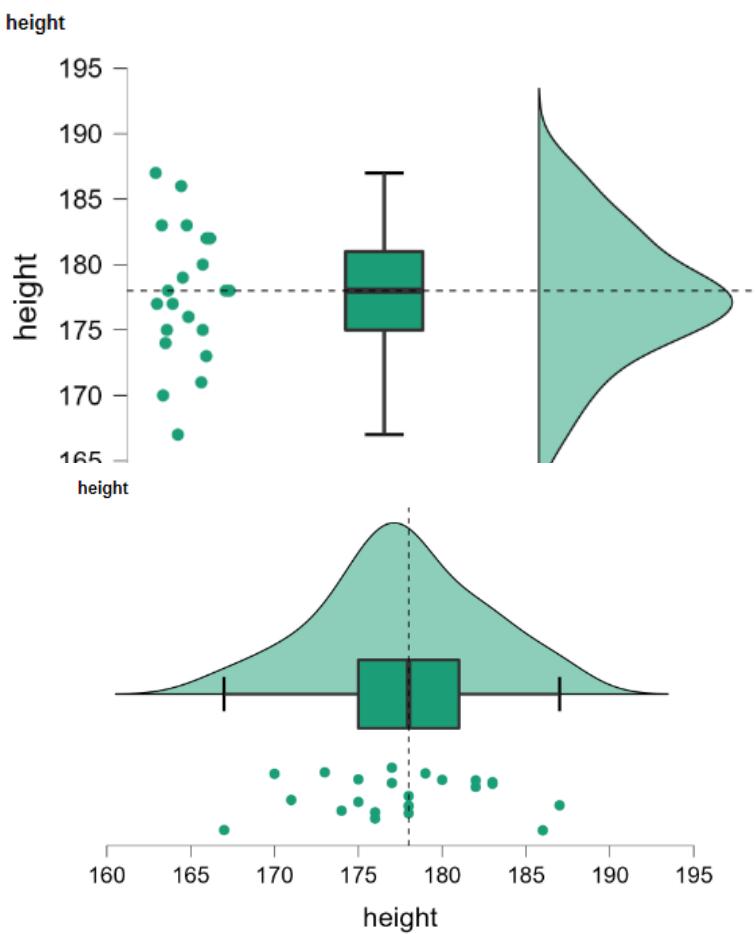
The descriptive data shows that the mean height of the sample population was 177.6 cm compared to the average of 178 cm for UK males.



The two plots show essentially the same data but in different ways. The standard Descriptive plot is an Interval plot showing the sample mean (black bullet), and the 95% confidence interval (whiskers), relative to the test value (dashed line).



The Raincloud Plot shows the data as individual data points, a box plot, and the distribution plot. This can be shown as either a vertical or horizontal display.





Repeat the procedure by replacing height with mass and changing the test value to 83.4.

Test of Normality (Shapiro-Wilk)

	W	p
mass	0.941	0.185

Note. Significant results suggest a deviation from normality.

The assumption check of normality (Shapiro-Wilk) is not significant, suggesting that the masses are normally distributed.

One Sample T-Test

	t	df	p	Mean Difference	Cohen's d	SE Cohen's d
mass	-7.159	22	< .001	-10.487	-1.493	0.303

Note. For the Student t-test, effect size is given by Cohen's d.

Note. For the Student t-test, location difference estimate is given by the sample mean difference d.

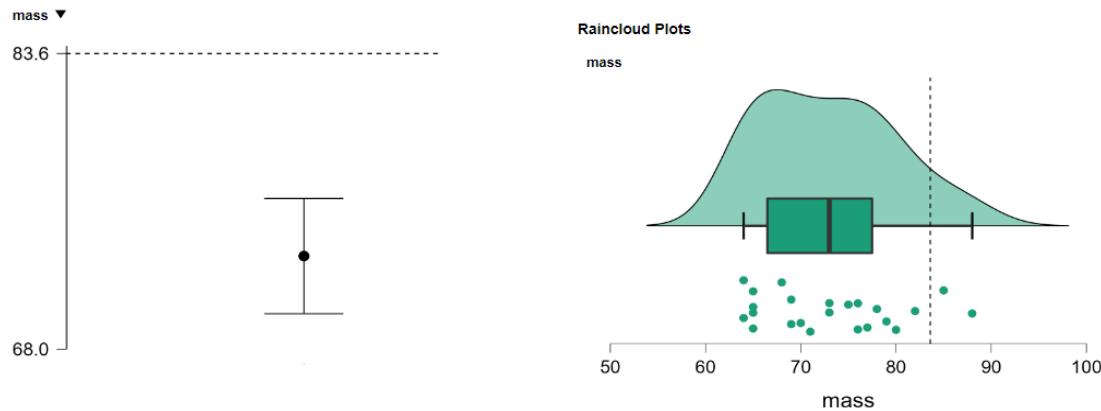
Note. For the Student t-test, the alternative hypothesis specifies that the mean is different from 83.4.

Note. Student's t-test.

This table shows that there is a significant difference between the mean sample (72.9 kg) and population body mass (83.4 kg), $p < .001$

Descriptives

	N	Mean	SD	SE
mass	23.000	72.913	7.025	1.465



REPORTING THE RESULTS

A one-sample t-test showed no significant difference in height compared to the population mean ($t(22) = -0.382, p = .706$); however, the participants were significantly lighter than the UK male population average ($t(22) = -7.159, p < .001$).



BINOMIAL TEST

The binomial test is effectively a non-parametric version of the one-sample t-test for use with dichotomous (i.e. yes/no) categorical datasets. This tests if the sample frequency is statistically different from a known or hypothesised population frequency.

The null hypothesis (H_0) tested is that the sample data frequency is equal to the expected population frequency.

ASSUMPTIONS

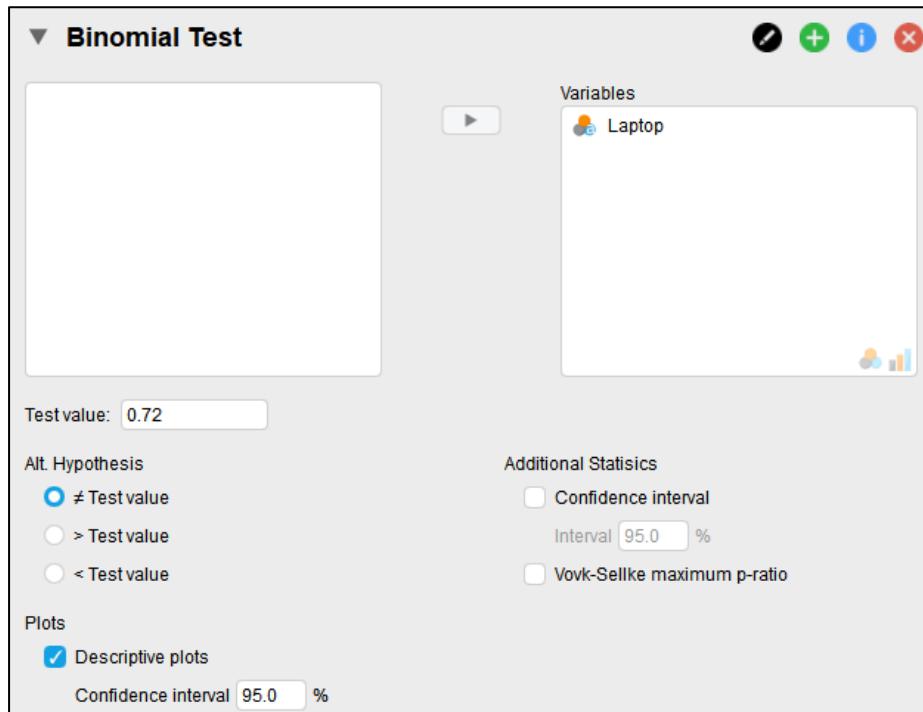
Three assumptions are required for a binomial test to provide a valid result:

- The test variable should be a dichotomous scale (such as yes/no, male/female, etc.).
- The sample responses should be independent
- The sample size is smaller, but representative of the population

RUNNING THE BINOMIAL TEST

Open **binomial.csv**; this contains one column of data showing the number of students using either a Windows laptop or a MacBook at university. In January 2021, when comparing just the two operating systems, the relative UK market share of Windows was 72% and macOS 28%.³

Go to Frequencies >Binomial test. Move the Laptop variable to the data window and set the Test value to 0.72 (72%). Also, tick the Descriptive plots.



³<https://www.statista.com/statistics/487002/market-share-of-desktop-operating-systems-uk/#:~:text=Apple%20and%20Microsoft%20own%20a,26.48%20percent%20of%20the%20market.>

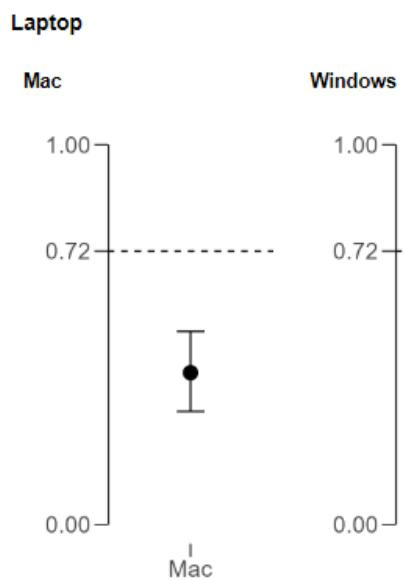


The following table and graph show that the frequencies of both laptops are significantly less than 72%. In particular, students are using significantly fewer Windows laptops than was expected compared to the UK market share.

Binomial Test

Variable	Level	Counts	Total	Proportion	p
Laptop	Mac	36	90	0.400	< .001
	Windows	54	90	0.600	0.014

Note. Proportions tested against value: 0.72.



Is this the same for MacBook users? Go back to the Options window and change the test value to 0.28 (14%). This time, both frequencies are significantly higher than 28%. This shows that students are using significantly more MacBooks than was expected compared to the UK market share.

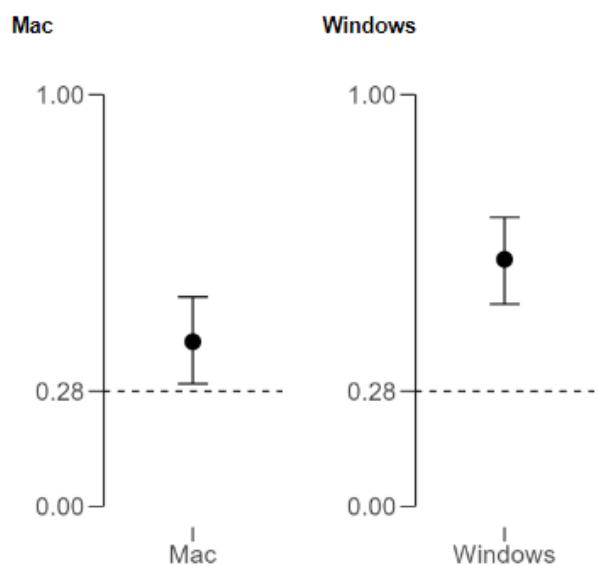
Binomial Test

Variable	Level	Counts	Total	Proportion	p
Laptop	Mac	36	90	0.400	0.014
	Windows	54	90	0.600	< .001

Note. Proportions tested against value: 0.28.



Laptop



REPORTING THE RESULTS

The UK relative proportion of Windows and MacBook users was reported to be 72% and 28% respectively. In a cohort of university students ($N=90$), a Binomial test revealed that the proportion of students using Windows laptops was significantly less (60%, $p=.014$) and those using MacBooks significantly more (40%, $p=.014$) than expected.



MULTINOMIAL TEST

The multinomial test is effectively an extended version of the Binomial test for use with categorical datasets containing three or more factors. This tests whether or not the sample frequency is statistically different from a hypothesised population frequency (multinomial test) or a known (Chi-square ‘goodness-of-fit’ test).

The null hypothesis (H_0) tested is that the sample frequency is equal to the expected population frequency.

ASSUMPTIONS

Three assumptions are required for a multinomial test to provide a valid result:

- The test variable should be a categorical scale containing 3 or more factors
- The sample responses should be independent
- The sample size is smaller, but representative of the population

RUNNING THE MULTINOMIAL TEST

Open **multinomial.csv**. This contains three columns of data showing the number of different coloured M&Ms counted in five bags. Without any prior knowledge, it could be assumed that the different coloured M&Ms are equally distributed.

Go to Frequencies > Multinomial test. Move the colour of the M&Ms to Factor and the observed number of M&Ms to counts. Tick Descriptives and Descriptives Plots.

The screenshot shows the 'Multinomial Test' dialog box. In the 'Expected' section, 'Colour' is selected as the factor. In the 'Counts' section, 'Observed' is selected. Under 'Test Values', 'Equal proportions (multinomial test)' is selected. Under 'Additional Statistics', 'Descriptives' is checked, with 'Confidence interval' set to 95.0%. Under 'Display', 'Counts' is selected. Under 'Plots', 'Descriptives plot' is checked, with 'Confidence interval' set to 95.0%.



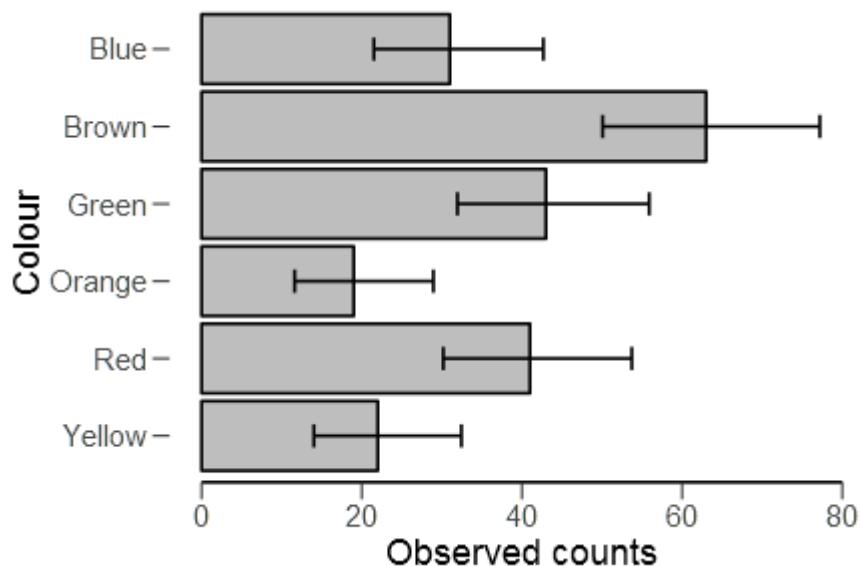
As can be seen in the Descriptive table, the test assumes an equal expectation for the proportions of coloured M&Ms (36 of each colour). The Multinomial test results show that the observed distribution is significantly different ($p < .001$) from an equal distribution.

Multinomial Test

	χ^2	df	p
Multinomial	35.932	5	< .001

Descriptives

Colour	Observed	Expected: Multinomial
Blue	31	36
Brown	63	36
Green	43	36
Orange	19	36
Red	41	36
Yellow	22	36



REPORTING THE RESULTS

A multinomial test showed that there was a significant difference between the observed and expected M&M colour distribution: $\chi^2 (5) = 35.9$, $p < .001$.



CHI-SQUARE ‘GOODNESS-OF-FIT’ TEST.

However, further research shows that the manufacturer produces coloured M&Ms in different ratios:

Colour	Blue	Brown	Green	Orange	Red	Yellow
Proportion	24	13	16	20	13	14

These values can now be used as the expected counts, so move the Expected variable to the Expected Counts box. This automatically runs the χ^2 ‘goodness-of-fit’ test, leaving the Hypothesis options greyed out.

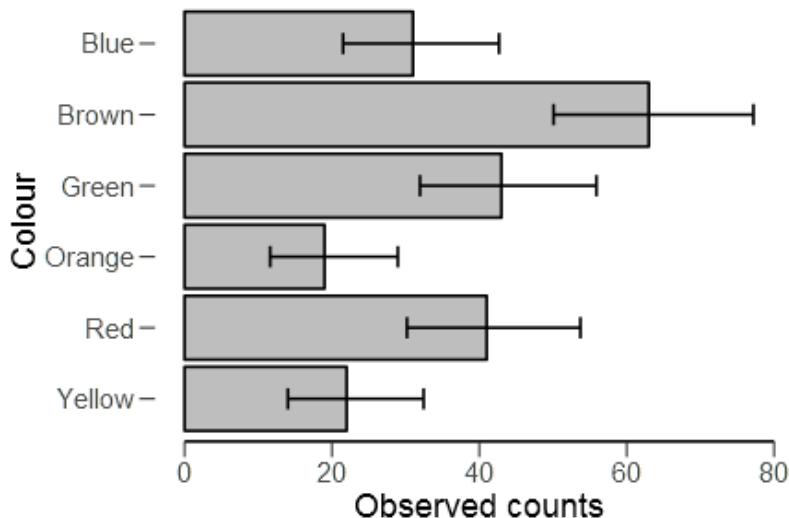
As can be seen in the Descriptives table, JASP has calculated the expected numbers of the different coloured M&Ms based on the manufacturers' reported production ratio. The results of the test show that the observed proportions of the different coloured M&Ms are significantly different ($\chi^2 = 74.5$, $p < .001$) from those proportions stated by the manufacturer.

Multinomial Test

	χ^2	df	p
Expected	74.535	5	< .001

Descriptives

Colour	Observed	Expected: Expected
Blue	31	52
Brown	63	28
Green	43	35
Orange	19	43
Red	41	28
Yellow	22	30





MULTINOMIAL AND χ^2 'GOODNESS-OF-FIT' TEST.

JASP also provides another option whereby both tests can be run at the same time. Go back to the Options window and only add Colour to the Factor and Observed to the Counts boxes, and remove the expected counts if the variable is still there. In Hypotheses, now tick the χ^2 test. This will open a small spreadsheet window showing the colour and H_0 (a) with each cell having 1 in it. This is assuming that the proportions of each colour are equal (multinomial test).

In this window, add another column which will automatically be labelled H_0 (b). The expected proportions of each colour can now be typed in.

Test Values

Equal proportions (multinomial test)

Custom expected proportions (χ^2 test)

Add Column	Row #	H_0 (a)	H_0 (b)
	Blue	1	24
	Brown	1	13
	Green	1	16
	Orange	1	20

Now, when the analysis is run, the results of the tests for the two hypotheses are shown. H_0 (a) is testing the null hypothesis that the proportions of each colour are equally distributed, while H_0 (b) is testing the null hypothesis that the proportions are the same as those expected. As can be seen, both hypotheses are rejected. Evidence indicates that the colours of M&M's do not match the manufacturer's published proportions.

Multinomial Test

	X ²	df	p
H_0 (a)	35.932	5	< .001
H_0 (b)	74.535	5	< .001

Descriptives

Colour	Observed	Expected	
		H_0 (a)	H_0 (b)
Blue	31	36	52
Brown	63	36	28
Green	43	36	35
Orange	19	36	43
Red	41	36	28
Yellow	22	36	30



Reporting the results

A χ^2 'goodness of fit' test was carried out to assess the distributions of M&Ms in retail packets. This showed that firstly, the proportions of each colour are not equally distributed, $\chi^2 (5) = 35.9, p < .001$. Secondly, the distribution of colours was significantly different to the manufacturer's stated distribution, $\chi^2 (5) = 74.5, p < .001$.



COMPARING TWO INDEPENDENT GROUPS

INDEPENDENT T-TEST

The parametric independent t-test, also known as Student's t-test, is used to determine if there is a statistical difference between the means of two independent groups. The test requires a continuous dependent variable (i.e. body mass) and an independent variable comprising 2 groups (i.e. males and females).

This test produces a t-score, which is a ratio of the differences between the two groups and the differences within the two groups:

$$t = \frac{\text{mean group 1} - \text{mean group 2}}{\text{standard error of the mean differences}}$$

$$t = \frac{(X_1 - X_2)}{\sqrt{\frac{(S_1)^2}{n_1} + \frac{(S_2)^2}{n_2}}}$$

X = mean
S = standard deviation
n = number of data points

A large t-score indicates that there is a greater difference between groups. The smaller the t-score, the more similarity there is between groups. A t-score of 5 means that the groups are five times as different from each other as they are within each other.

The null hypothesis (H_0) tested is that the population means from the two unrelated groups are equal.

ASSUMPTIONS OF THE PARAMETRIC INDEPENDENT T-TEST

Group independence:

Both groups must be independent of each other. Each participant will only provide one data point for one group. For example, participant 1 can only be in either a male or female group, not both. Repeated measures are assessed using the **Paired t-test**.

Normality of the dependent variable:

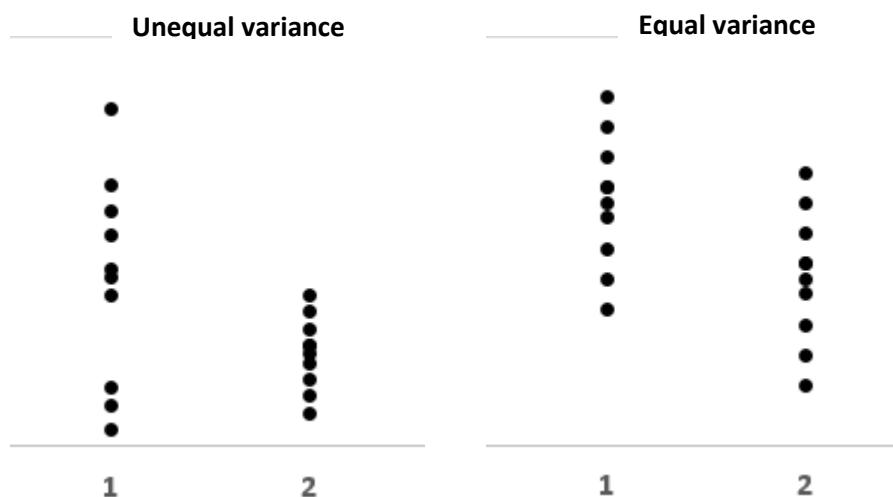
The dependent variable should also be measured on a continuous scale and be approximately normally distributed with no significant outliers. This can be checked using the Shapiro-Wilk test. The t-test is fairly robust, and small deviations from normality are normally acceptable. However, this is not the case if the group sizes are very different. A rule of thumb is that the ratio between the group sizes should be <1.5 (i.e. group A = 12 participants and group B = >8 participants).

If normality is violated, you can try transforming your data (for example, log values, square root values) or, if the group sizes are very different, use the **Mann-Whitney U** test, which is a non-parametric equivalent that does not require the assumption of normality (see later).



Homogeneity of variance:

The variances of the dependent variable should be equal in each group. This can be tested using Levene's Test of Equality of Variances.



If Levene's test is statistically significant, indicating that the group variances are unequal, we can correct for this violation by using an adjusted t-statistic based on the **Welch** method.

RUNNING THE INDEPENDENT T-TEST

Open **Independent t-test.csv**, which contains weight loss on a self-controlled 10-week diet between men and women. It's good practice to check the Distribution and boxplots in Descriptives to visually check for distribution and outliers.

Go to T-Tests > Independent Samples t-test and put weight loss in the Dependent variable box and gender (independent variable) in the Grouping Variable box.

The screenshot shows the 'Independent Samples T-Test' dialog in JASP. In the 'Dependent Variables' section, 'Weight loss' is selected. In the 'Grouping Variable' section, 'Gender' is selected. The top right corner of the dialog has four small circular icons: a black circle with a pencil, a green circle with a plus sign, a blue circle with an info symbol, and a red circle with a minus sign.



In the analysis window, tick the following options:

The screenshot shows the JASP software interface for a statistical analysis. The dependent variable is 'Weight loss' and the grouping variable is 'Gender'. In the 'Tests' section, 'Student' and 'Welch' are checked, while 'Mann-Whitney' is unchecked. Under 'Alternative Hypothesis', 'Group 1 ≠ Group 2' is selected. In 'Assumption Checks', 'Normality' and 'Equality of variances' (Levene's test) are checked. Under 'Missing Values', 'Exclude cases per dependent variable' is selected. In the 'Additional Statistics' section, 'Location parameter' and 'Effect size (Cohen's d)' are checked. Descriptive statistics and raincloud plots are also selected. Plots include bar charts with confidence intervals and horizontal displays.

UNDERSTANDING THE OUTPUT

The output should consist of four tables and three graphs. Firstly, we need to check that the parametric assumptions required are not violated.

Test of Normality (Shapiro-Wilk)

		W	p
Weight loss	Females	0.968	0.282
	Males	0.971	0.310

Note. Significant results suggest a deviation from normality.

The Shapiro-Wilk test shows that both groups have normally distributed data; therefore, the assumption of normality is not violated. If one or both were significant, you should consider using the non-parametric equivalent **Mann-Whitney** test.



Test of Equality of Variances (Levene's)

	F	df	p
Weight loss	2.278	1	0.135

Levene's test shows that there is no difference in the variance; therefore, the assumption of homogeneity of variance is not violated. If Levene's test was significant, **Welch's** adjusted t-statistic, degrees of freedom and p-values should be reported.

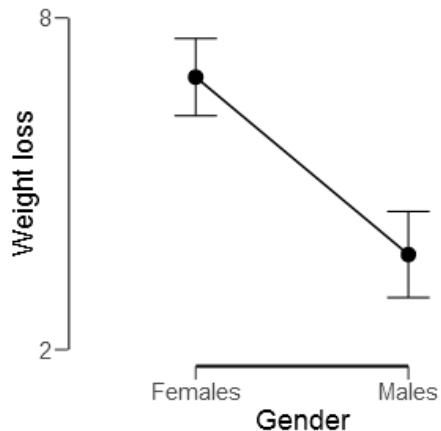
This table shows the two computed t-statistics (**Student and Welch**). Remember, the t-statistic is derived from the mean difference divided by the standard error of the difference. Both show that there is a significant statistical difference between the two groups ($p < .001$), and Cohen's d suggests that this is a large effect.

Independent Samples T-Test

	Test	Statistic	df	p	Mean Difference	SE Difference	Cohen's d	SE Cohen's d
Weight loss	Student	6.160	85.000	< .001	3.209	0.521	1.322	0.259
	Welch	6.191	84.544	< .001	3.209	0.518	1.325	0.259

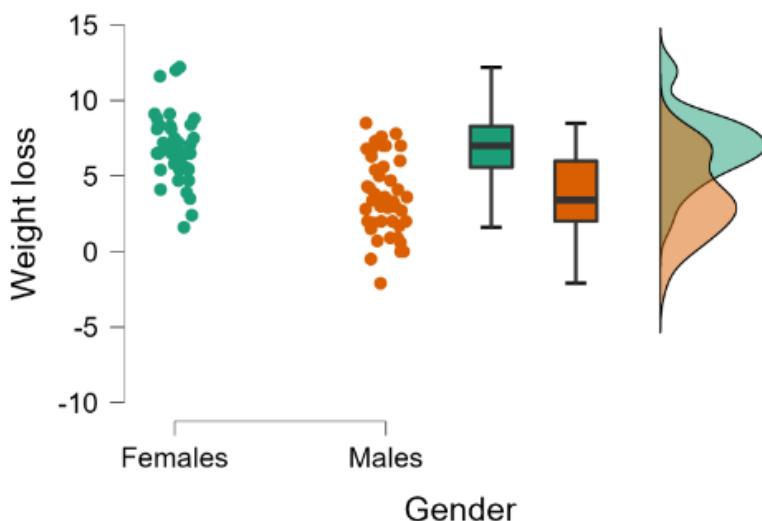
Group Descriptives

	Group	N	Mean	SD	SE
Weight loss	Females	42	6.929	2.242	0.346
	Males	45	3.720	2.588	0.386

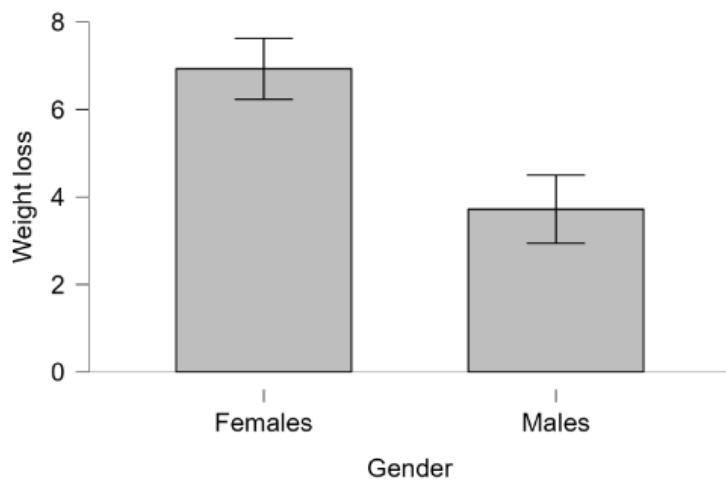




Weight loss



Weight loss



From the descriptive data and plots (use whichever you want), it can be seen that females had a higher weight loss than males.

REPORTING THE RESULTS

A Levene test found that the assumption of homogeneity of variance was met: $F(1) = 2.28, p=.135$. A Shapiro-Wilk test shows that both groups have normally distributed data: Females ($W=0.97, p=.282$) and males ($W=0.97, p=0.31$). Therefore, an independent sample Student t-test based on equal variances was carried out. This showed that females lost significantly more weight ($M = 6.93, SD = 2.24$) over 10 weeks of dieting than males ($M = 3.72, SD = 2.56$): $t(85)=6.16, p<.001$. Cohen's d (1.322) suggests that this is a large effect.

NB if the Levene test is significant, the Welch t-statistic and corrected df should be reported.



MANN-WITNEY U TEST

If you find that your data is not normally distributed (such as a significant Shapiro-Wilk test result) or is ordinal by nature, the equivalent non-parametric independent test is the **Mann-Whitney U test**.

Open **Mann-Whitney pain.csv**, which contains subjective pain scores (0-10) with and without ice therapy. NOTE: Make sure that Treatment is categorical, and the pain score is ordinal. Go to T-Tests > Independent t-tests, put pain score in the Dependent variable box and use Treatment as the grouping variable.

In the analysis options, only tick:

- ✓ Mann-Whitney
- ✓ Location parameter
- ✓ Effect size

There is no reason to repeat the assumption checks since Mann-Whitney does not require the assumption of normality or homogeneity of variance required by parametric tests.

UNDERSTANDING THE OUTPUT

This time, you will only get one table:

Independent Samples T-Test

	U	df	p	Hodges-Lehmann Estimate	Rank-Biserial Correlation	SE Rank-Biserial Correlation
Pain score	207.00		< .001	3.00	0.84	0.21

Note. For the Mann-Whitney test, effect size is given by the rank biserial correlation.

Note. Mann-Whitney U test.

The Mann-Whitney U-statistic is highly significant. **W=207, p<.001**.

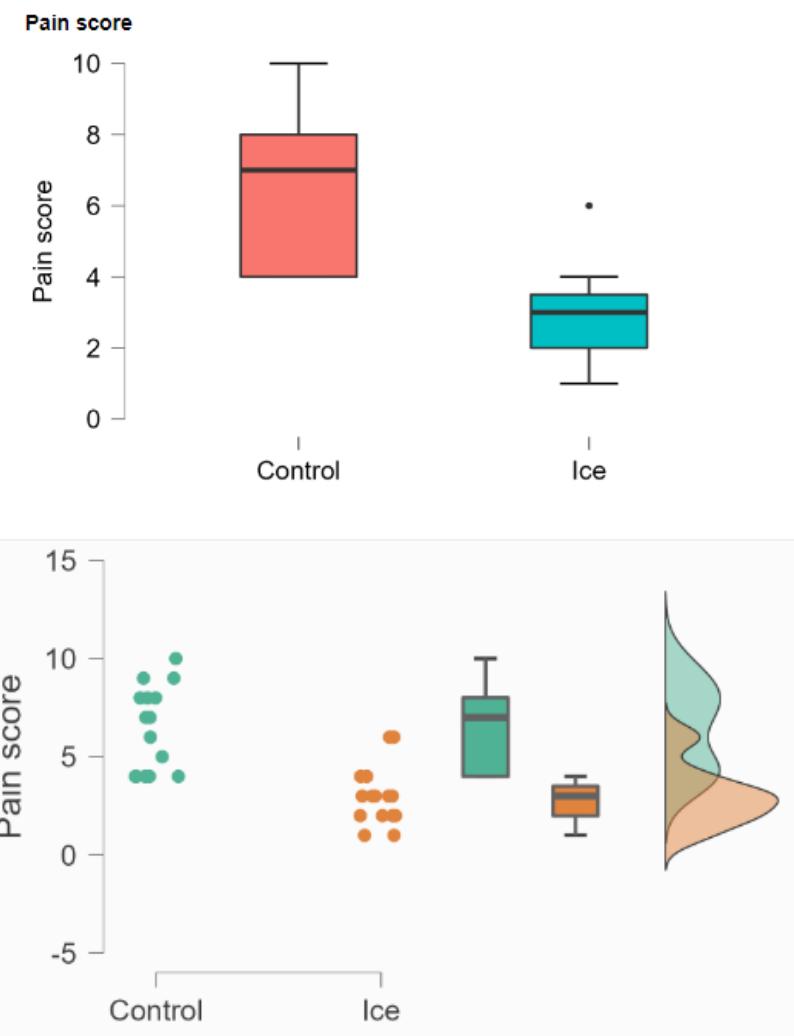
The location parameter, the Hodges–Lehmann estimate, is a measure of the **median** difference between the two groups. The rank-biserial correlation (r_B) can be considered as an effect size and is interpreted the same as Pearson's r, so 0.84 is a large effect size.

For non-parametric data, you should report **median** values as your descriptive statistics and use boxplots instead of line graphs and confidence intervals, SD/SE bars. Go to Descriptive statistics, put the Pain score into the variable box and split the file by Treatment.

You can alternatively show the Raincloud plots for more information on the differences and distributions.

Descriptive Statistics

	Pain score	
	Control	Ice
Median	7.000	3.000
Std. Deviation	2.167	1.512
MAD	2.000	1.000
IQR	4.000	1.500



REPORTING THE RESULTS

A Mann-Whitney test showed that Ice therapy significantly reduces pain scores ($Mdn = 3$) compared to the control group ($Mdn = 7$): $W=207$, $p<.001$. The $r_B= 0.84$ suggests that this is a large effect size.



COMPARING TWO RELATED GROUPS

PAIRED SAMPLES T-TEST

As with the Independent t-test, there are both parametric and non-parametric options available in JASP. The parametric paired-samples t-test (also known as the dependent sample t-test or repeated measures t-test) compares the means between two related groups on the same continuous, dependent variable. For example, looking at weight loss pre- and post-10 weeks of dieting.

$$\text{The paired } t\text{-statistic} = \frac{\text{mean of the differences between group pairs}}{\text{the standard error of the mean differences}}$$

With the paired t-test, the null hypothesis (H_0) is that the pairwise difference between the two groups is zero.

ASSUMPTIONS OF THE PARAMETRIC PAIRED SAMPLES T-TEST

Four assumptions are required for a paired t-test to provide a valid result:

- The **dependent variable** should be measured on a continuous scale.
- The **independent variable** should consist of 2 categorical related/matched groups, i.e. each participant is matched in both groups.
- The differences between the matched pairs should be approximately **normally distributed**.
- There should be no significant **outliers** in the differences between the 2 groups.

RUNNING THE PAIRED SAMPLES T-TEST

Open **Paired t-test.csv** in JASP. This contains two columns of paired data, pre-diet body mass and post-weeks of dieting. Go to T-Tests > Paired Samples t-test. Ctrl-click both variables and add them to the analysis box on the right.





In the analysis options, tick the following:

Tests <input checked="" type="checkbox"/> Student <input type="checkbox"/> Wilcoxon signed-rank	Additional Statistics <input checked="" type="checkbox"/> Location parameter <input type="checkbox"/> Confidence interval 95.0 % <input checked="" type="checkbox"/> Effect size <input type="checkbox"/> Confidence interval 95.0 % <input type="checkbox"/> Correct for correlation <input checked="" type="checkbox"/> Descriptives <input type="checkbox"/> Vovk-Sellke maximum p-ratio
Alternative Hypothesis <input checked="" type="radio"/> Measure 1 ≠ Measure 2 <input type="radio"/> Measure 1 > Measure 2 <input type="radio"/> Measure 1 < Measure 2	
Assumption Checks <input checked="" type="checkbox"/> Normality <input checked="" type="checkbox"/> Q-Q plot residuals	Plots <input checked="" type="checkbox"/> Descriptives plots Confidence interval 95.0 % <input checked="" type="checkbox"/> Raincloud plots <input type="checkbox"/> Raincloud difference plots <input type="checkbox"/> Horizontal display <input type="checkbox"/> Bar plots Confidence interval 95.0 % <input type="checkbox"/> Standard error <input checked="" type="checkbox"/> Fix horizontal axis to 0
Missing Values <input checked="" type="radio"/> Exclude cases per variable <input type="radio"/> Exclude cases listwise	

UNDERSTANDING THE OUTPUT

The output should consist of three tables and three graphs.

Test of Normality (Shapiro-Wilk)

		W	p
Pre diet body mass	-	Post 4 weeks diet	0.975 0.124

Note. Significant results suggest a deviation from normality.

The assumption check of normality (Shapiro-Wilk) is not significant, suggesting that the pairwise differences are normally distributed; therefore, the assumption is not violated. If this showed a significant difference, the analysis should be repeated using the non-parametric equivalent, **Wilcoxon's signed-rank test**.



Paired Samples T-Test

Measure 1		Measure 2	t	df	p	Mean Difference	SE Difference	Cohen's d	SE Cohen's d
Pre diet body mass	-	Post 4 weeks diet	13.04	77	< .001	3.78	0.29	1.48	0.05

Note. Student's t-test.

This shows that there is a significant difference in body mass between the pre- and post-dieting conditions, with a mean difference (location parameter) of 3.78kg. Cohen's *d* states that this is a large effect.

The descriptive statistics and plot show that there was a reduction in body mass following 4 weeks of dieting.

Descriptives

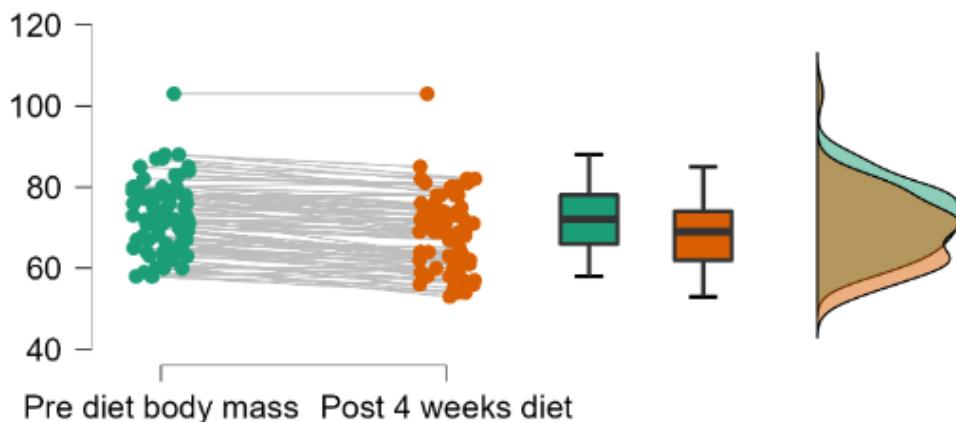
	N	Mean	SD	SE
Pre diet body mass	78	72.526	8.723	0.988
Post 4 weeks diet	78	68.744	9.009	1.020



The Raincloud plot shows the data for both groups and the distribution of the differences between the two groups.



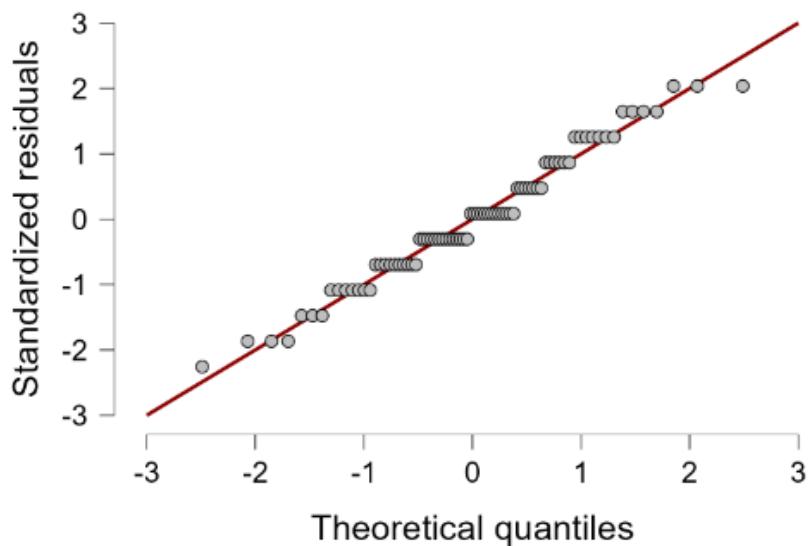
Pre diet body mass - Post 4 weeks diet



The Q-Q plot reinforces the fact that the data is normally distributed; the closer to the diagonal line the points lie, the more normally distributed the data.

Q-Q Plots

Pre diet body mass - Post 4 weeks diet



REPORTING THE RESULTS

The Shapiro-Wilk test was not significant, suggesting that the pairwise differences were normally distributed; therefore, the assumption is not violated: $W=0.98, p=.124$.

Post-diet body mass was lower ($M = 68.7, SD = 9.0 \text{ kg}$) than at the start of the diet ($M = 72.5, SD = 8.7 \text{ kg}$). This mean difference ($M = 3.78, SE = 0.29 \text{ kg}$) was shown to be significant: $t(77) = 13.04, p < .001$. Cohen's d (1.48) suggests that this is a large effect.



RUNNING THE NON-PARAMETRIC PAIRED SAMPLES TEST

WILCOXON'S SIGNED RANK TEST

If you find that your data is not normally distributed (significant Shapiro-Wilk test result) or is ordinal by nature, the equivalent non-parametric independent test is Wilcoxon's signed-rank test. Open **Wilcoxon's rank.csv**. This has two columns, one with pre-anxiety and post-hypnotherapy anxiety scores (from 0 - 50). In the dataset view, make sure that both variables are assigned to the ordinal data type.

Go to T-Tests > Paired Samples t-test and follow the same instructions as above, but now only tick the following options:

The dialog box shows the following settings:

- Variable Pairs:** Pre diet body mass, Post 4 weeks diet
- Tests:** Student (unchecked), Wilcoxon signed-rank (checked)
- Additional Statistics:** Location parameter (checked), Confidence interval (unchecked, 95.0%), Effect size (checked), Confidence interval (unchecked, 95.0%)

There will be only one table in the output:

Paired Samples T-Test

Paired Samples T-Test								
Measure 1	Measure 2	W	z	df	p	Hodges-Lehmann Estimate	Rank-Biserial Correlation	SE Rank-Biserial Correlation
Pre-anxiety	- Post-anxiety	322.000	4.292		< .001	8.000	0.982	0.225

Note: Wilcoxon signed-rank test.

The Wilcoxon W-statistic is highly significant, $p<0.001$.

The location parameter, the Hodges–Lehmann estimate, is the median difference between the two groups. The rank-biserial correlation (r_B) can be considered as an effect size and is interpreted the same as Pearson's r , so 0.48 is a medium to large effect size.

Effect size	Trivial	Small	Medium	Large
Rank -biserial (r_B)	<0.1	0.1	0.3	0.5



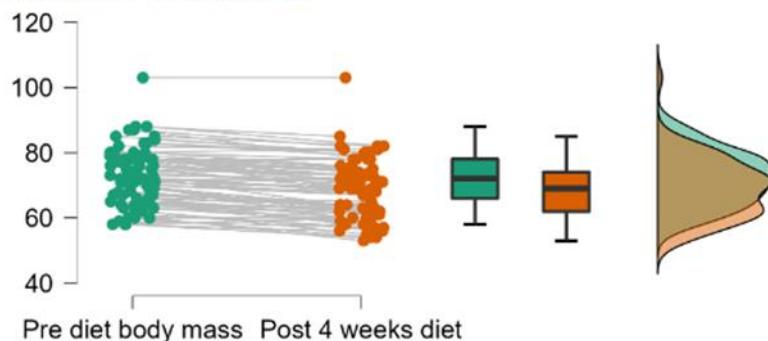
For non-parametric data, you should report median values as your descriptive statistics and use boxplots or Rainbow plots instead of line graphs and confidence intervals, SD/SE bars.

Descriptive Statistics

	Pre-anxiety	Post-anxiety
Median	22.000	15.000
MAD	5.000	2.000



Pre diet body mass - Post 4 weeks diet



REPORTING THE RESULTS

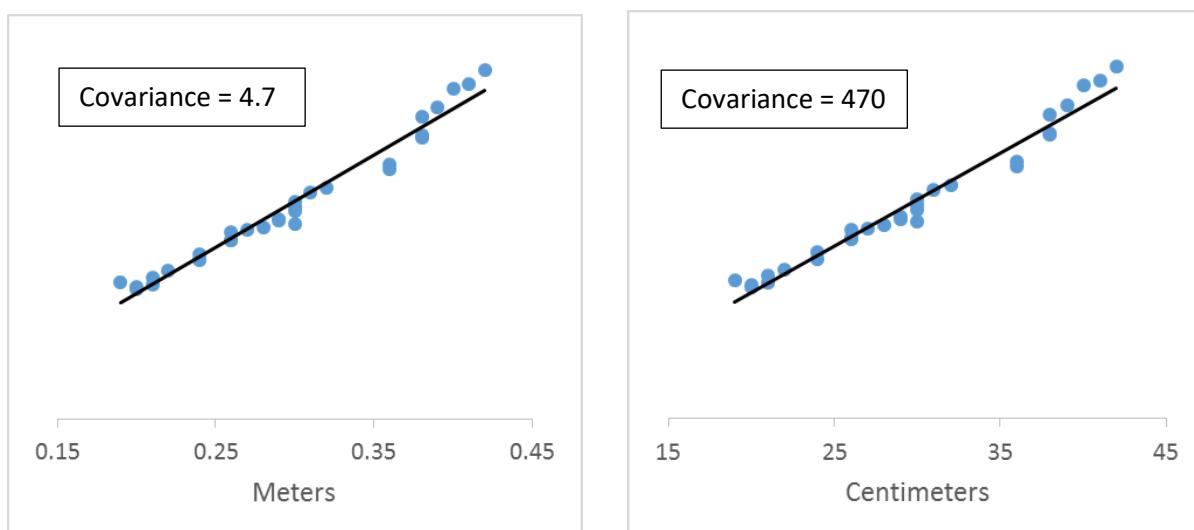
A Wilcoxon signed-rank test showed that hypnotherapy significantly reduces anxiety scores ($Mdn = 15$) compared to pre-therapy ($Mdn = 22$) scores, $W=322$, $p<.001$. The rank-biserial correlation (r_B) = 0.982 suggests that this is a large effect size.



CORRELATION ANALYSIS

Correlation is a statistical technique that can be used to determine if, and how strongly, pairs of variables are associated. Correlation is only appropriate for quantifiable data in which numbers are meaningful, such as continuous or ordinal data. It cannot be used for purely categorical data, for which we have to use contingency table analysis (see Chi-square analysis in JASP).

Essentially, do different variables co-vary? i.e. are changes in one variable reflected in similar changes to another variable? If one variable deviates from its mean, does the other variable deviate from its mean in either the same or opposite direction? This can be assessed by measuring covariance; however, this is not standardised. For example, we can measure the covariance of two variables that are measured in meters; however, if we convert the same values to centimetres, we get the same relationship but with a completely different covariance value.



To overcome this, standardised covariance is used, which is known as **Pearson's correlation coefficient** (or "r"). It ranges from -1.0 to +1.0. The closer r is to +1 or -1, the more closely the two variables are related. If r is close to 0, there is no relationship. If r is (+), then as one variable increases, the other also increases. If r is (-), then as one increases, the other decreases (sometimes referred to as an "inverse" correlation).

The correlation coefficient (r) should not be confused with R^2 (coefficient of determination) or R (multiple correlation coefficient as used in the regression analysis).

The main assumption in this analysis is that the data have a normal distribution and are linear. This analysis will not work well with curvilinear relationships.



RUNNING CORRELATION

The analysis tests the null hypothesis (H_0) that there is no association between the two variables.

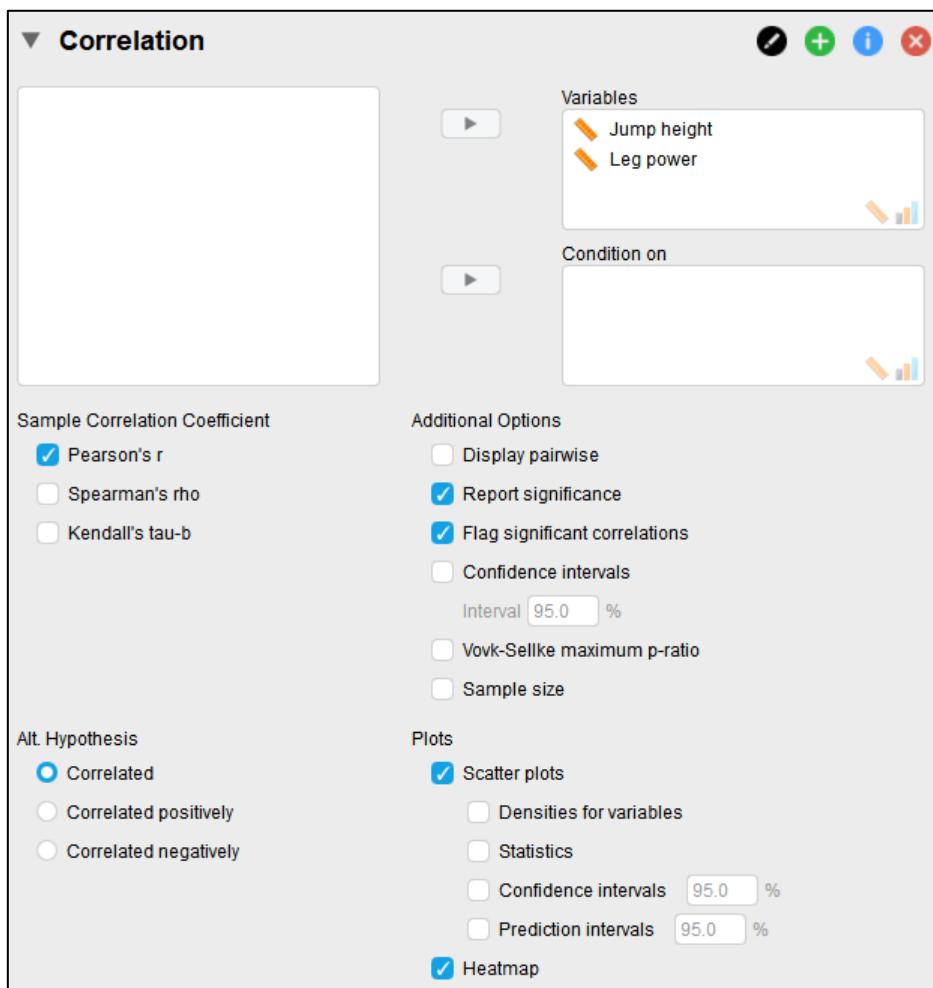
From the example data, open **Jump height correlation.csv**, which contains 2 columns of data, jump height (m) and explosive leg power (W). Firstly, run the Descriptive statistics and check the box plots for any outliers.

To run the correlation analysis, go to Regression > Correlation. Move the 2 variables to the analysis box on the right. Tick

- ✓ Pearson,
- ✓ Report significance,
- ✓ Flag significant correlations

Under Plots

- ✓ Scatter plots
- ✓ Heatmap





UNDERSTANDING THE OUTPUT

The first table shows the correlation matrix with Pearson's r-value and its p-value. This shows a highly significant correlation ($p < .001$) with a large r value close to 1 ($r = 0.984$), and we can reject the null hypothesis.

Pearson Correlations

		Jump height	Leg power
Jump height	Pearson's r p-value	—	—
Leg power	Pearson's r p-value	0.984*** $< .001$	—

* $p < .05$, ** $p < .01$, *** $p < .001$

For simple correlations like this, it is easier to look at the pairwise table (go back to analysis and tick the Display pairwise table option. This replaces the correlation matrix in the results, which may be easier to read.

Pearson Correlations

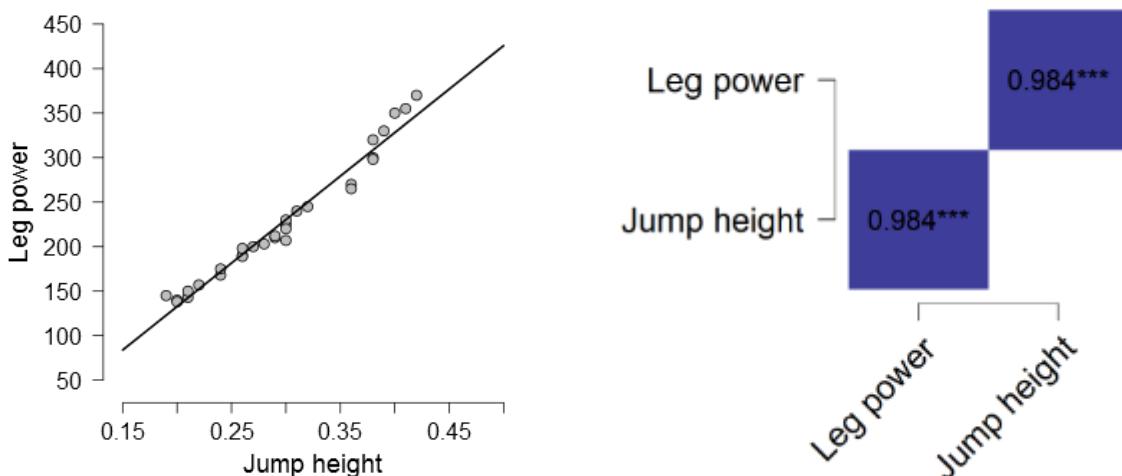
		Pearson's r	p
Jump height	-	Leg power	0.984*** $< .001$

* $p < .05$, ** $p < .01$, *** $p < .001$

Pearson's r value is an effect size where <0.1 is trivial, $0.1 - 0.3$ is a small effect, $0.3 - 0.5$ is a moderate effect, and >0.5 is a large effect.

The plot provides a simple visualisation of this strong positive correlation ($r = 0.984$, $p < .001$), which is also highlighted by the heatmap (more relevant when looking at multiple correlations).

Pearson's r heatmap





GOING ONE STEP FURTHER.

If you take the correlation coefficient r and square it, you get the coefficient of determination (R^2). This is a statistical measure of the proportion of variance in one variable that is explained by the other variable. Or:

$$R^2 = \text{Explained variation} / \text{Total variation}$$

R^2 is always between 0 and 100% where:

- 0% indicates that the model explains none of the variability of the response data around its mean, and
- 100% indicates that the model explains all the variability of the response data around its mean.

In the example above, $r = 0.984$, so $R^2 = 0.968$. This suggests that jump height accounts for 96.8% of the variance in explosive leg power.

REPORTING THE RESULTS

Pearson's correlation showed a significant correlation between jump height and leg power ($r = 0.984$, $p < .001$), with jump height accounting for 96.8% of the variance in leg power.



RUNNING NON-PARAMETRIC CORRELATION – Spearman’s and Kendall’s tau

If your data is ordinal or is continuous data that has violated the assumptions required for parametric testing (normality and/or variance), you need to use the non-parametric alternatives to Pearson’s correlation coefficient.

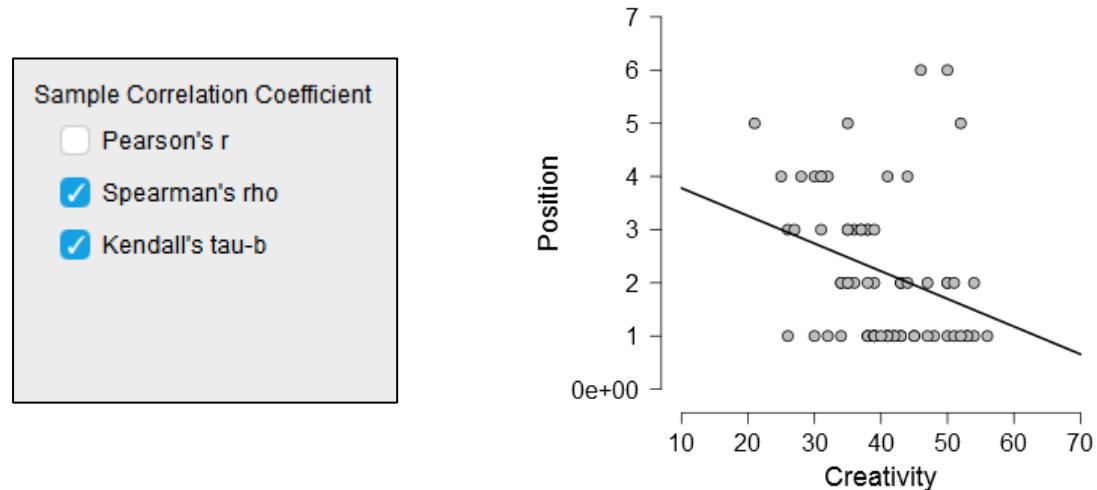
The alternatives are Spearman’s (ρ) or Kendall’s (τ) correlation coefficients. Both are based on ranking data and are not affected by outliers or normality/variance violations.

Spearman’s ρ is usually used for ordinal scale data, and Kendall’s τ is used in small samples or when many values with the same score (ties). In most cases, Kendall’s τ and Spearman’s rank correlation coefficients are very similar and thus invariably lead to the same inferences.

The effect sizes are the same as Pearson’s r . The main difference is that ρ^2 can be used as an approximate non-parametric coefficient of determination, but the same is not true for Kendall’s τ .

From the example data, open **Non-parametric correlation.csv**, which contains 2 columns of data, a creativity score and position in the ‘World’s biggest liar’ competition (thanks to Andy Field).

Run the analysis as before, but now using Spearman and Kendall’s τ - b coefficients instead of Pearson’s.



Correlation Table

	Spearman		Kendall			
	rho	p	tau B	p		
Creativity	-	Position	-0.373**	0.002	-0.300**	0.001

* p < .05, ** p < .01, *** p < .001

REPORTING THE RESULTS

The relationship between creativity scores and final position in the ‘World’s biggest liar’ competition was assessed. A Spearman’s correlation test showed a significant relationship: $\rho = -0.373$, $p=.002$.

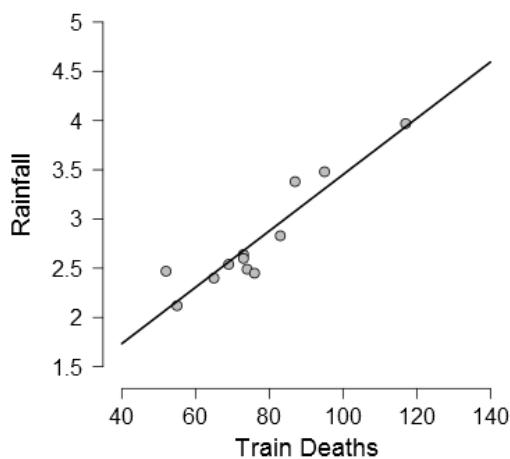
**NOTE OF CAUTION.**

The correlation only gives information on the strength of association. It gives no information on the direction, i.e. which variable causes the other to change. So, it cannot be used to state that one thing causes the other. Often, a significant correlation means absolutely nothing and is purely by chance, especially if you correlate thousands of variables. This can be seen in the following strange correlations:

Pedestrians killed in a collision with a railway train correlate with rainfall in Missouri:

Pearson Correlations

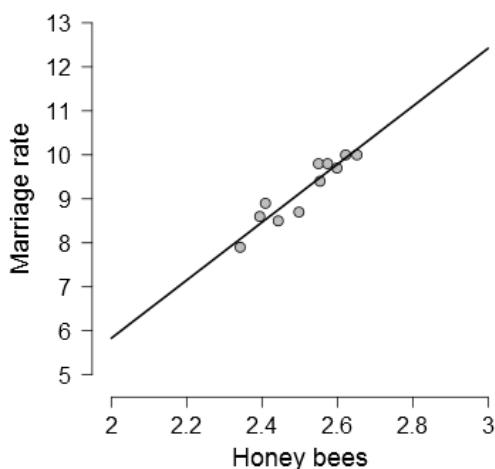
		Pearson's r	p
Train Deaths	- Rainfall	0.928	< .001



The number of honey-producing bee colonies (1000s) correlates strongly with the marriage rate in South Carolina (per 1000 marriages)

Pearson Correlations

		Pearson's r	p
Honey bees	- Marriage rate	0.938	< .001





REGRESSION

Whereas correlation tests for associations between variables, regression is the next step commonly used for predictive analysis, i.e. to predict a dependent outcome variable from one (simple regression) or more (multiple regression) independent predictor variables.

Regression results in a hypothetical model of the relationship between the outcome and predictor variable(s). The model used is a linear one defined by the formula;

$$y = c + b * x + \varepsilon$$

- y = estimated dependent outcome variable score,
- c = constant,
- b = regression coefficient and
- x = score on the independent predictor variable
- ε = random error component (based on residuals)

Linear regression provides both the constant and regression coefficient(s).

Linear regression makes the following assumptions:

- **Data** (1 dependent variable of continuous level, independent variables that are measured either at the continuous or nominal level, sample size, etc.)
- **Outliers**, there should be no significant outliers, high-leverage points or highly influential points.
- **Linear relationship**, there needs to be a linear relationship between (a) the dependent variable and each of your independent variables, and (b) the dependent variable and the independent variables collectively.
- **Independence**, there should be independence of observations (i.e., independence of residuals)
- **Normality**, a) check that the residuals (errors) are approximately normally distributed; and b) check that the dependent variable is approximately normally distributed.
- **Equal error variances**, data needs to show homoscedasticity of residuals (equal error variances)
- **Multicollinearity**, data must show minimal multicollinearity. This occurs when you have two or more independent variables that are highly correlated with each other.

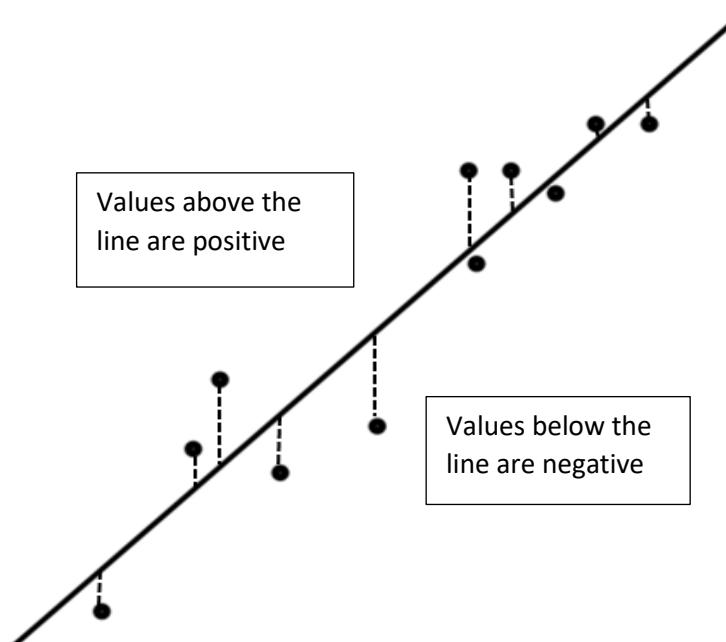
Concerning sample sizes, there are many different ‘rules of thumb’ in the literature, ranging from 10-15 data points per predictor in the model, i.e. 4 predictor variables will each require between 40 and 60 data points each, to $50 + (8 * \text{number of predictors})$ for each variable. So, for 4 variables, that would require 82 data points for each variable. Effectively, the bigger your sample size, the better your model.



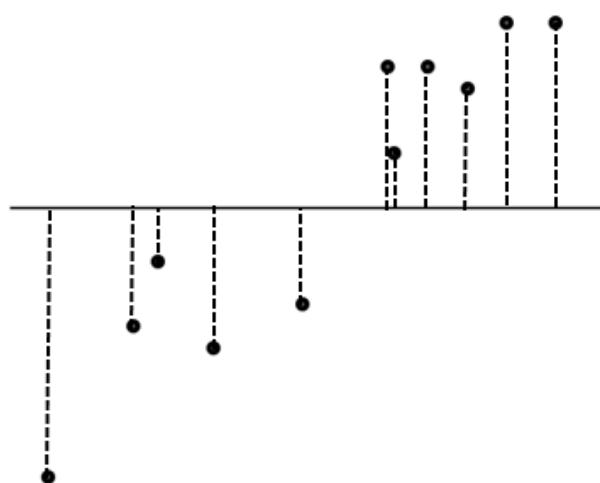
SUMS OF SQUARES (Boring, but the basis of evaluating the regression model.)

Most regression analyses will produce the best model available, but how good is it actually, and how much error is in the model?

This can be determined by looking at ‘the goodness of fit’ using the sums of squares. This is a measure of how close the actual data points are to the modelled regression line.

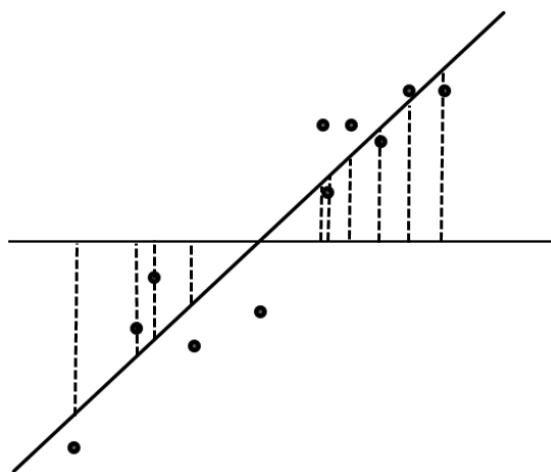


The vertical difference between the data points and the predicted regression line is known as the **residuals**. These values are squared to remove the negative numbers and then summed to give SS_R . This is effectively the error of the model or the ‘**goodness of fit**’, obviously, the smaller the value, the less error in the model.





The vertical difference between the data points and the mean of the outcome variable can be calculated. These values are squared to remove the negative numbers and then summed to give the **total** sum of the squares, SS_T . This shows how good the mean value is as a model of the outcome scores.



The vertical difference between the mean of the outcome variable and the predicted regression line is now determined. Again, these values are squared to remove the negative numbers and then summed to give the **model** sum of squares (SS_M). This indicates how better the model is compared to just using the mean of the outcome variable. SS_T is the total sum of the squares.

So, the larger the SS_M , the better the model is at predicting the outcome compared to the mean value alone. If this is accompanied by a small SS_R , the model also has a small error.

R^2 is similar to the coefficient of determination in correlation in that it shows how much of the variation in the outcome variable can be predicted by the predictor variable(s).

$$R^2 = \frac{SS_M}{SS_T}$$

In regression, the model is assessed by the F statistic based on the improvement in the prediction of the model SS_M and the residual error SS_R . The larger the F value, the better the model.

$$F = \frac{\text{Mean } SS_M}{\text{Mean } SS_R}$$



SIMPLE REGRESSION

Regression tests the null hypothesis (H_0) that there will be no significant prediction of the dependent (outcome) variable by the predictor variable(s).

Open **Rugby kick regression.csv**. This dataset contains rugby kick data, including distance kicked, right/left leg strength and flexibility and bilateral leg strength. Firstly, go to Descriptives > Descriptive statistics and check the boxplots for any outliers. In this case, there should be none, though it is good practice to check.

For this simple regression, go to Regression > Linear regression and put distance into the Dependent Variable (outcome) and R_Strength into the Covariates (Predictor) box. Tick the following options in the Statistics options:

The screenshot shows the 'Statistics' dialog box in JASP. Under 'Coefficients', 'Estimates' and 'Model fit' are checked. Under 'Residuals', 'Durbin-Watson' is checked. Other options like 'Confidence intervals', 'Covariance matrix', 'Vovk-Sellke maximum p-ratio', 'R squared change', 'Descriptives', 'Part and partial correlations', and 'Collinearity diagnostics' are unchecked.

UNDERSTANDING THE OUTPUT

You will now get the following outputs:

Model Summary - Distance

Model	R	R^2	Adjusted R^2	RMSE	Durbin-Watson		
					Autocorrelation	Statistic	p
M_0	0.000	0.000	0.000	85.250	0.588	0.762	0.010
M_1	0.784	0.614	0.579	55.285	0.225	1.524	0.287

Note. M_1 includes R_Strength

Here it can be seen that the correlation (R) between the two variables is high (0.784), while that for the null model M_0 is 0. The R^2 value of 0.614 tells us that right leg strength accounts for 61.4% of the variance in kick distance. Durbin-Watson checks for correlations between residuals, which can invalidate the test. This should be above 1 and below 3, and ideally around 2.



ANOVA

Model		Sum of Squares	df	Mean Square	F	p
M ₁	Regression	53589.863	1	53589.863	17.533	0.002
	Residual	33621.061	11	3056.460		
	Total	87210.923	12			

Note. M₁ includes R_Strength

Note. The intercept model is omitted, as no meaningful information can be shown.

The ANOVA table shows all the sums of squares mentioned earlier. With regression being the model and Residual being the error. The F-statistic is significant - $p=.002$. This tells us that the model is a significantly better predictor of kicking distance than the mean distance.

Report as **F(1, 11) = 17.53, p=.002**.

Coefficients

Model		Unstandardized	Standard Error	Standardized	t	p
M ₀	(Intercept)	486.077	23.644		20.558	< .001
	R_Strength	6.425	1.534	0.784	4.187	0.002

This table gives the coefficients (unstandardized) that can be put into the linear equation.

$$y = c + b * x$$

y = estimated dependent outcome variable score,

c = constant (**intercept**)

b = regression coefficient (**R_strength**)

x = score on the independent predictor variable

For example, for a leg strength of 60 kg, the distance kicked can be predicted by the following:

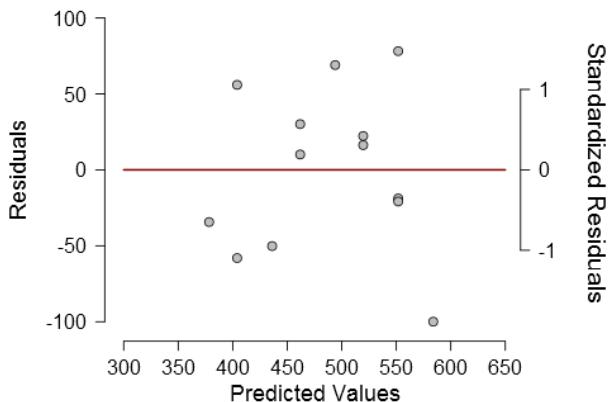
$$\text{Distance} = 57.105 + (6.452 * 60) = 454.6 \text{ m}$$

FURTHER ASSUMPTION CHECKS

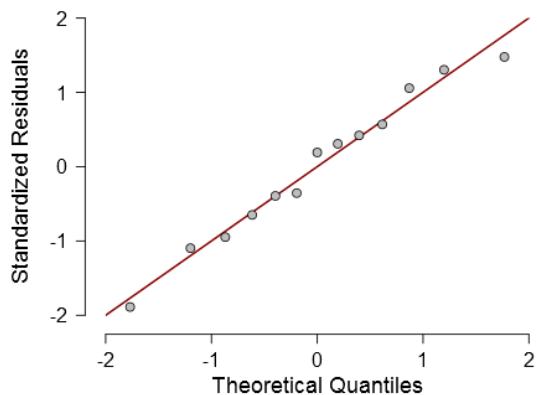
In Plots checks, tick the following two options:



This will result in two graphs:



This graph shows a balanced random distribution of the residuals around the baseline, suggesting that the assumption of homoscedasticity has not been violated. (See Exploring data integrity in JASP for further details.)



The Q-Q plot shows that the standardised residuals fit nicely along the diagonal, suggesting that both assumptions of normality and linearity have also not been violated.

REPORTING THE RESULTS

A simple linear regression was used to predict rugby kicking distance from right leg strength. Leg strength was shown to explain a significant amount of the variance in the kicking distance: $F(1, 11) = 17.53$, $p=0.002$, $R^2 = 0.614$. The regression coefficient ($b = 6.452$) allows the kicking distance to be predicted using the following regression equation:

$$\text{Distance} = 57.105 + (6.452 * \text{Right leg strength})$$



MULTIPLE REGRESSION

The model used is still a linear one defined by the formula.

$$y = c + b * x + \epsilon$$

- y = estimated dependent outcome variable score,
- c = constant,
- b = regression coefficient and
- x = score on the independent predictor variable
- ϵ = random error component (based on residuals)

However, we now have more than 1 regression coefficient and predictor score, i.e.

$$y = c + b_1 * x_1 + b_2 * x_2 + b_3 * x_3 \dots \dots \dots b_n * x_n$$

Data entry methods.

If predictors are uncorrelated, their order of entry has little effect on the model. In most cases, predictor variables are correlated to some extent, and thus, the order in which the predictors are entered can make a difference. The different methods are subject to much debate in the area.

Enter: This is the **default method** in which all the predictors are forced into the model in the order they appear in the Covariates box. This is considered to be the best method.

Blockwise entry (Hierarchical entry): The researcher, normally based on prior knowledge and previous studies, decides the order in which the known predictors are entered first, depending on their importance in predicting the outcome. Additional predictors are added in further steps.

Backwards: All predictors are initially entered into the model, and then the contribution of each is calculated. Predictors with less than a given level of contribution ($p < 0.1$) are removed. This process repeats until all the predictors are statistically significant.

Forward: The predictor with the highest simple correlation with the outcome variable is entered first. Subsequent predictors are selected based on the size of their semi-partial correlation with the outcome variable. This is repeated until all predictors that contribute significant unique variance to the model have been included in the model.

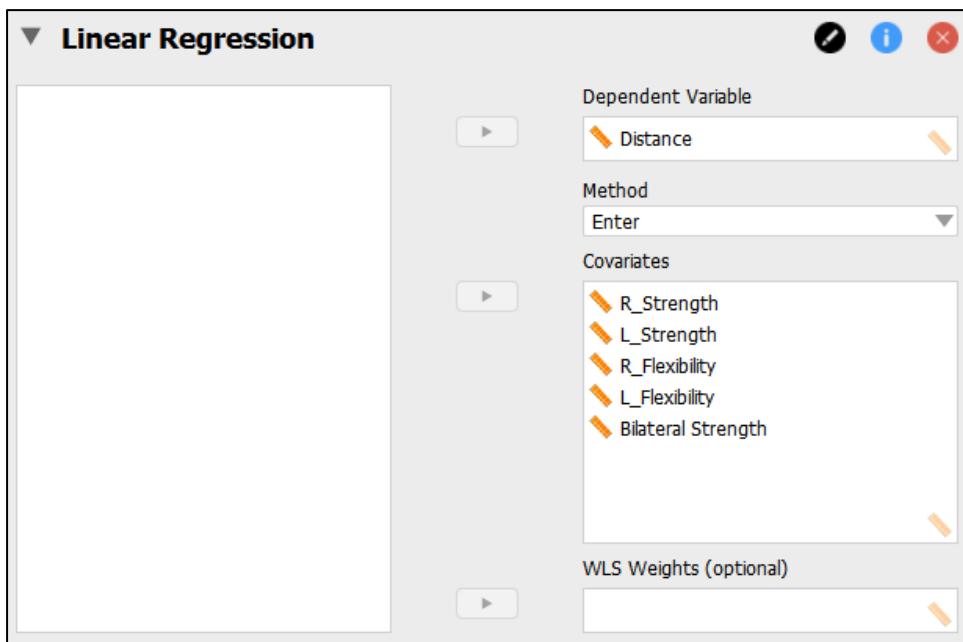
Stepwise entry: Same as the Forward method, except that every time a predictor is added to the model, a removal test is made of the least useful predictor. The model is constantly reassessed to see whether any redundant predictors can be removed.

There are many reported disadvantages of using stepwise data entry methods; however, **backwards** entry methods can be useful for exploring previously unused predictors or for fine-tuning the model to select the best predictors from the available options.

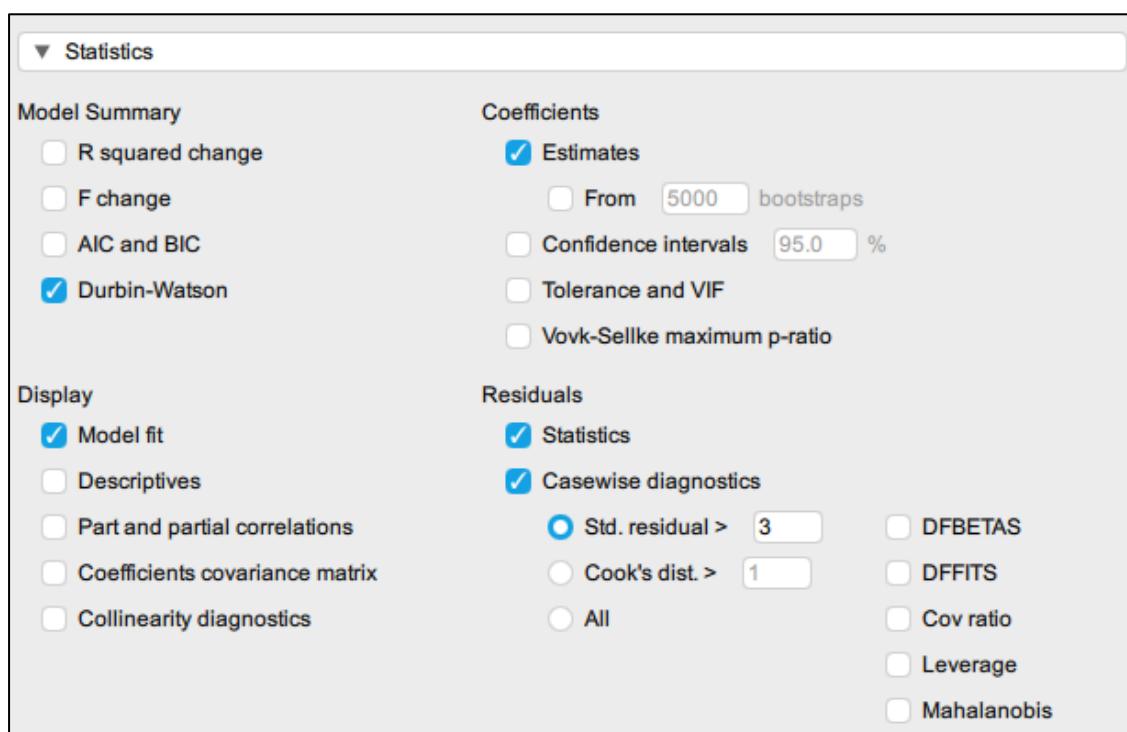


RUNNING MULTIPLE REGRESSION

Open **Rugby kick regression.csv** that we used for simple regression. Go to Regression > Linear regression and put distance into the Dependent Variable (outcome), and now add all the other variables into the Covariates (Predictor) box.



In the Variable section, leave the Method as **Enter**. Tick the following options in the Statistics options, Durbin-Watson, Model fit, Estimates and Casewise diagnostics.





UNDERSTANDING THE OUTPUT

You will now get the following outputs:

Model Summary - Distance

Model	R	R ²	Adjusted R ²	RMSE	Durbin-Watson		
					Autocorrelation	Statistic	p
M ₀	0.000	0.000	0.000	85.250	0.588	0.762	0.010
M ₁	0.902	0.814	0.681	48.132	0.287	1.328	0.133

Note. M₁ includes R_Strength, L_Strength, R_Flexibility, L_Flexibility, Bilateral Strength

This provides information on a model based on the M₀ (no predictors) and the alternative H¹. The adjusted R² (used for multiple predictors) shows that they can predict 68.1% of the outcome variance. Durbin-Watson checks for correlations between residuals are between 1 and 3 as required.

ANOVA

Model		Sum of Squares	df	Mean Square	F	p
M ₁	Regression	70994.078	5	14198.816	6.129	0.017
	Residual	16216.845	7	2316.692		
	Total	87210.923	12			

Note. M₁ includes R_Strength, L_Strength, R_Flexibility, L_Flexibility, Bilateral Strength

Note. The intercept model is omitted, as no meaningful information can be shown.

The ANOVA table shows the F-statistic to be significant at p=0.017, suggesting that the model is a significantly better predictor of kicking distance than the mean distance.

Coefficients

Model		Unstandardized	Standard Error	Standardized	t	p
M ₀	(Intercept)	486.077	23.644		20.558	< .001
M ₁	(Intercept)	-92.367	218.389		-0.423	0.685
	R_Strength	1.747	3.321	0.213	0.526	0.615
	L_Strength	0.703	3.590	0.086	0.196	0.850
	R_Flexibility	4.078	4.759	0.373	0.857	0.420
	L_Flexibility	-1.339	2.447	-0.135	-0.547	0.601
	Bilateral Strength	1.665	0.946	0.423	1.759	0.122

This table shows both the M₀ and M₁ models and the constant (intercept) and regression coefficients (unstandardized) for all the predictors forced into the model. Even though the ANOVA shows the model to be significant, none of the predictor regression coefficients is significant!



The casewise diagnostics table is empty! This is good news. This will highlight any cases (rows) that have residuals that are 3 or more standard deviations away from the mean. These cases with the largest errors may well be outliers. Too many outliers will have an impact on the model and should be dealt with in the usual way (see Exploring Data Integrity).

Influential Cases

Case Number	Std. Residual	Distance	Predicted Value	Residual	Cook's Distance
1	-0.100	0.000	48.132	48.132	0.000

Note: No influential cases found.

As a comparison, re-run the analyses but now choose Backwards as the method of data entry.

The outputs are as follows:

Model Summary - Distance

Model	R	R ²	Adjusted R ²	RMSE	Durbin-Watson		
					Autocorrelation	Statistic	p
M ₀	0.902	0.814	0.681	48.132	0.287	1.328	0.133
M ₁	0.902	0.813	0.720	45.146	0.303	1.296	0.088
M ₂	0.897	0.805	0.740	43.505	0.284	1.313	0.144
M ₃	0.884	0.782	0.738	43.618	0.132	1.676	0.439

JASP has now calculated 4 potential regression models. It can be seen that each consecutive model increases the adjusted R^2 , with model 4 accounting for 73.5% of the outcome variance.

The Durbin-Watson score is also higher than the forced entry method.

ANOVA

Model		Sum of Squares	df	Mean Square	F	p
M ₀	Regression	70994.078	5	14198.816	6.129	0.017
	Residual	16216.845	7	2316.692		
	Total	87210.923	12			
M ₁	Regression	70905.329	4	17726.332	8.697	0.005
	Residual	16305.594	8	2038.199		
	Total	87210.923	12			
M ₂	Regression	70176.855	3	23392.285	12.359	0.002
	Residual	17034.068	9	1892.674		
	Total	87210.923	12			
M ₃	Regression	68185.712	2	34092.856	17.920	< .001
	Residual	19025.211	10	1902.521		
	Total	87210.923	12			

The ANOVA table indicates that each successive model is better, as shown by the increasing F-value and improving p-value.



Coefficients

Model		Unstandardized	Standard Error	Standardized	t	p
M ₀	(Intercept)	-92.367	218.389		-0.423	0.685
	R_Strength	1.747	3.321	0.213	0.526	0.615
	L_Strength	0.703	3.590	0.086	0.196	0.850
	R_Flexibility	4.078	4.759	0.373	0.857	0.420
	L_Flexibility	-1.339	2.447	-0.135	-0.547	0.601
	Bilateral Strength	1.665	0.946	0.423	1.759	0.122
M ₁	(Intercept)	-110.347	185.840		-0.594	0.569
	R_Strength	2.218	2.148	0.271	1.033	0.332
	R_Flexibility	4.501	3.978	0.411	1.131	0.291
	L_Flexibility	-1.370	2.291	-0.138	-0.598	0.566
	Bilateral Strength	1.605	0.840	0.408	1.910	0.092
M ₂	(Intercept)	-116.892	178.772		-0.654	0.530
	R_Strength	2.710	1.911	0.331	1.418	0.190
	R_Flexibility	2.886	2.814	0.264	1.026	0.332
	Bilateral Strength	1.642	0.807	0.418	2.033	0.073
M ₃	(Intercept)	46.251	81.820		0.565	0.584
	R_Strength	3.914	1.512	0.478	2.588	0.027
	Bilateral Strength	2.009	0.725	0.511	2.770	0.020

Model 1 is the same as the forced entry method first used. The table shows that as the least significantly contributing predictors are sequentially removed, we end up with a model with two significant predictor regression coefficients, right leg strength and bilateral leg strength.

We can now report the Backwards predictor entry results in a highly significant model $F(2, 10) = 17.92$, $p < .001$ and a regression equation of

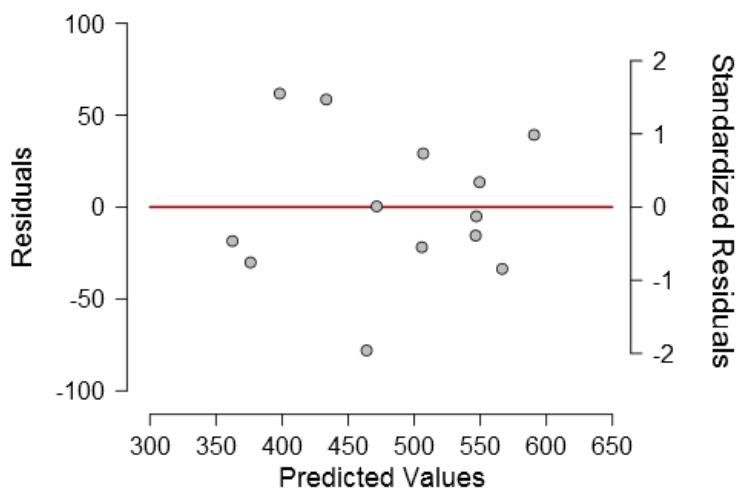
$$\text{Distance} = 46.251 + (3.914 * \text{R_Strength}) + (2.009 * \text{Bilateral Strength})$$

TESTING FURTHER ASSUMPTIONS.

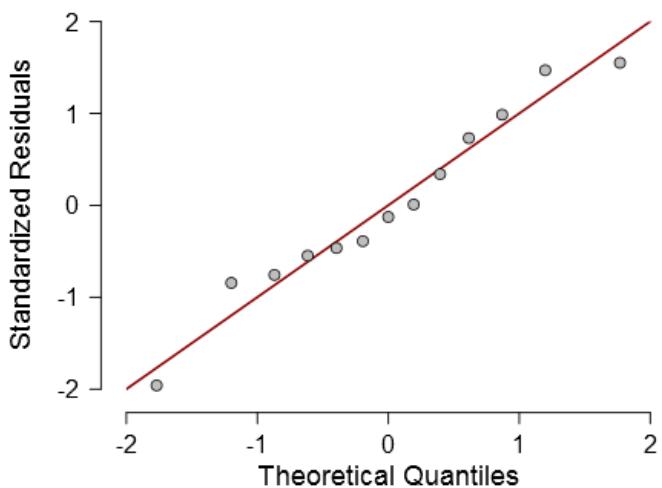
As for the simple linear regression example, tick the following options.



Residuals vs. Predicted



Q-Q Plot Standardized Residuals ▾



The balanced distribution of the residuals around the baseline suggests that the assumption of homoscedasticity has not been violated.

The Q-Q plot shows that the standardised residuals fit along the diagonal, suggesting that both assumptions of normality and linearity have also not been violated.



IN SUMMARY

R^2 provides information on how much variance is explained by the model using the predictors provided.

F-statistics provide information as to how good the model is.

The unstandardized (b)-value provides a constant which reflects the strength of the relationship between the predictor(s) and the outcome variable.

Violation of assumptions can be checked using the Durbin-Watson value, tolerance/VIF values, Residual vs predicted and Q-Q plots.

REPORTING THE RESULTS

Multiple linear regression using backwards data entry shows that right leg and bilateral strength can significantly explain 78.2% of the variance in kicking distance, $F(2,10) = 17.92, p<.001, R^2 = 0.782$. Two significant regression coefficients were found, right leg strength ($b_1=3.91, t=2.59, p=.027$) and bilateral strength ($b_2 = 2.009, t =2.77, p=0.02$). Kicking distance could be predicted using a regression equation of;

$$\text{Distance} = 57.105 + (3.914 * \text{R_Strength}) + (2.009 * \text{Bilateral Strength})$$

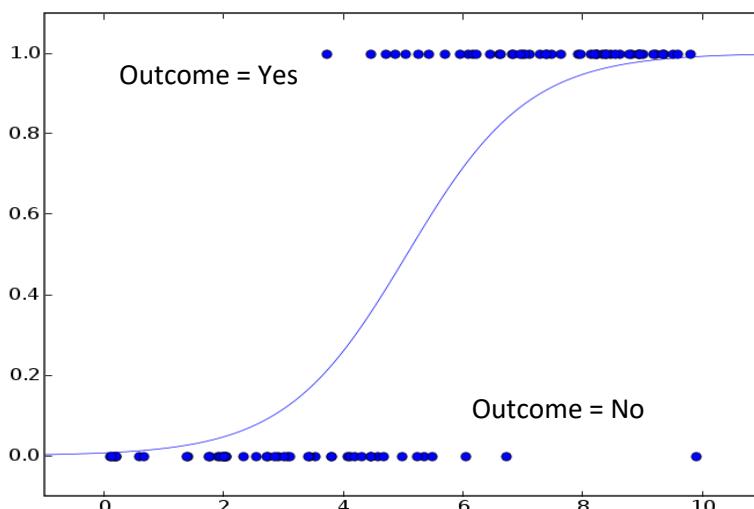


LOGISTIC REGRESSION

In simple and multiple linear regression outcome and predictor variable(s) were continuous data. What if the outcome was a binary/categorical measure? Can, for example, a yes or no outcome be predicted by other categorical or continuous variables? The answer is yes if binary logistic regression is used. This method is used to predict the probability of a binary yes or no outcome.

The null hypothesis tested is that there is no relationship between the outcome and the predictor variable(s).

As can be seen in the graph below, a linear regression line between the yes and no responses would be meaningless as a prediction model. Instead, a sigmoidal logistic regression curve is fitted with a minimum of 0 and a maximum of 1. Some predictor values overlap between yes and no. For example, a prediction value of 5 would give an equal 50% probability of being a yes or no outcome. Thresholds are therefore calculated to determine if a predictor data value will be classified as a yes or no outcome.



ASSUMPTIONS FOR BINARY LOGISTIC REGRESSION

- The dependent variable must be binary, i.e. yes or no, male or female, good or bad.
- One or more independent (predictor variables) can be continuous or categorical variables.
- A linear relationship exists between any continuous independent variables and the logit transformation (natural log of the odds that the outcome equals one of the categories) of the dependent variable.

LOGISTIC REGRESSION METRICS

AIC (Akaike Information Criteria) and **BIC** (Bayesian Information Criteria) are measures of fit for the model; the best model will have the lowest AIC and BIC values.



Four **pseudo R²** values are calculated in JASP: McFadden, Nagelkerke, Tjur and Cox & Snell. These are analogous to R² in linear regression and all give different values. What constitutes a good R² value varies; however, they are useful when comparing different models for the same data. The model with the largest R² statistic is considered to be the best.

The Wald test is used to determine the statistical significance of each of the independent variables.

The **confusion matrix** is a table showing actual vs predicted outcomes and can be used to determine the accuracy of the model. From this, sensitivity and specificity can be derived.

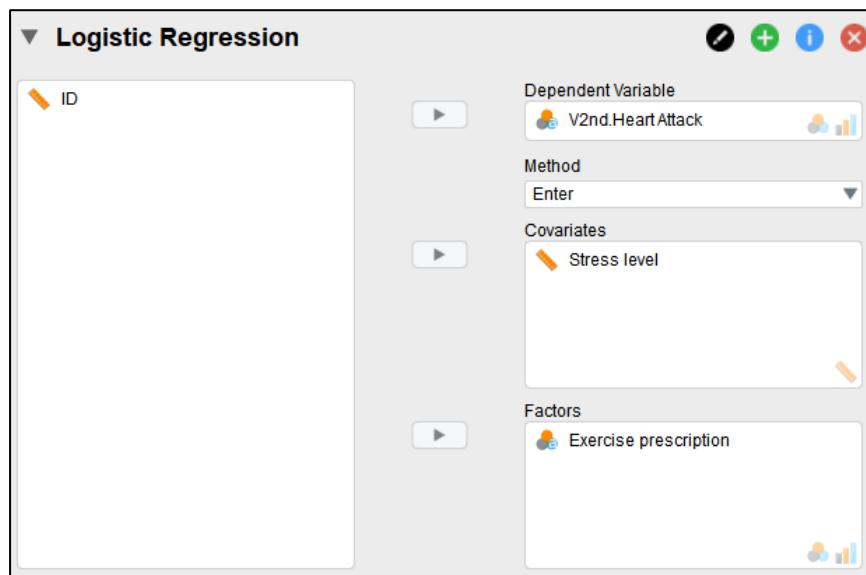
Sensitivity is the percentage of cases in which the observed outcome was correctly predicted by the model (i.e., true positives).

Specificity is the percentage of observations that were also correctly predicted as not having the observed outcome (i.e., true negatives).

RUNNING LOGISTIC REGRESSION

Open **Heart attack.csv** in JASP. This contains 4 columns of data: Patient ID, did they have a second heart attack (yes/no), whether they were prescribed exercise (yes/no) and their stress levels (high value = high stress).

Put the outcome variable (2nd heart attack) into the Dependent variable, add the stress levels to the Covariates and Exercise prescription to Factors. Leave the data entry method as Enter.



In the Statistics options, tick Estimates, Odds ratios, Confusion matrix, Sensitivity and Specificity.



▼ Statistics

Descriptives

Factor descriptives

Regression Coefficients

Estimates

From bootstraps

Standardized coefficients

Odds ratios

Confidence intervals

Interval %

Odds ratio scale

Robust standard errors

Vovk-Sellke maximum p-ratio

Multicollinearity Diagnostics

Performance Diagnostics

Confusion matrix

Performance Metrics

Accuracy

AUC

Sensitivity / Recall

Specificity

Precision

F-measure

Brier score

H-measure

UNDERSTANDING THE OUTPUT

The initial output should comprise 4 tables.

The model summary shows that H₁ (with the lowest AIC and BIC scores) suggests a significant relationship ($\chi^2(37) = 21.257$, $p < .001$) between the outcome (2nd heart attack) and the predictor variables (exercise prescription and stress levels).

McFadden's $R^2 = 0.383$. It is suggested that a range from 0.2 to 0.4 indicates a good model fit.

Model Summary - 2nd.Heart Attack

Model	Deviance	AIC	BIC	df	$\Delta\chi^2$	p	McFadden R ²	Nagelkerke R ²	Tjur R ²	Cox & Snell R ²
M ₀	55.452	57.452	59.141	39			0.000		0.000	
M ₁	34.195	40.195	45.261	37	21.257	< .001	0.383	0.550	0.446	0.412

Note. M₁ includes Stress level, Exercise prescription

Coefficients

Model		Estimate	Standard Error	Odds Ratio	z	Wald Test		
						Wald Statistic	df	p
M ₀	(Intercept)	-0.000	0.316	1.000	-4.441×10 ⁻¹⁶	1.972×10 ⁻³¹	1	1.000
M ₁	(Intercept)	-4.368	2.550	0.013	-1.713	2.933	1	0.087
	Stress level	0.089	0.041	1.093	2.159	4.662	1	0.031
	Exercise prescription (Yes)	-2.043	0.890	0.130	-2.295	5.268	1	0.022

Note. 2nd.Heart Attack level 'Yes' coded as class 1.



Both stress level and exercise prescription are significant predictor variables ($p=.031$ and $.022$, respectively). The most important values in the coefficients table are the odds ratios. For the continuous predictor, an odds ratio of greater than 1 suggests a positive relationship, while < 1 implies a negative relationship. This suggests that high-stress levels are significantly related to an increased probability of having a second heart attack.

Thus, the odds ratio of 1.093 means that for each additional point on the stress scale, the odds of having a second heart attack are increased 1.1 times. Or presented in percentage, for every additional point on the stress scale, the odds of having a second heart attack increase by 9.3%. The odds ratio of 0.130 regarding the exercise prescription variable can be interpreted as having a second heart attack undergoing an exercise intervention is 0.13 times the odds of not having such an intervention. Or presented in percentage, those having exercise prescription are 87% less likely to have a second heart attack compared to those not having such an intervention.

Confusion matrix

Observed	Predicted		% Correct	Performance metrics
	No	Yes		
No	15	5	75.000	
Yes	5	15	75.000	
Overall % Correct		75.000		

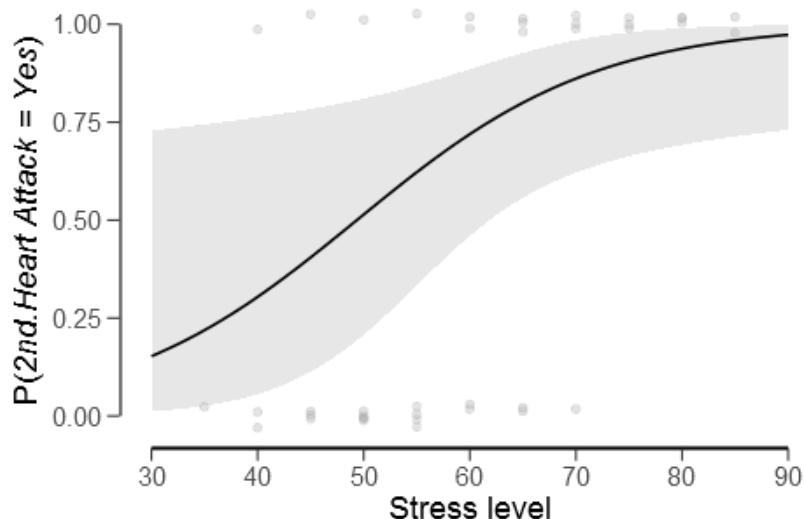
Note. The cut-off value is set to 0.5

The confusion matrix shows (where agreement (yes/yes) or disagreement (yes/no) occurred between the predicted and observed) that the 15 true negative and positive cases were predicted by the model, while the error, false negatives and positives, were found in 5 cases. This is confirmed in the Performance metrics, where both sensitivity (% of cases that had the outcome correctly predicted) and specificity (% of cases correctly predicted as not having the outcome (i.e., true negatives) are both 75%.

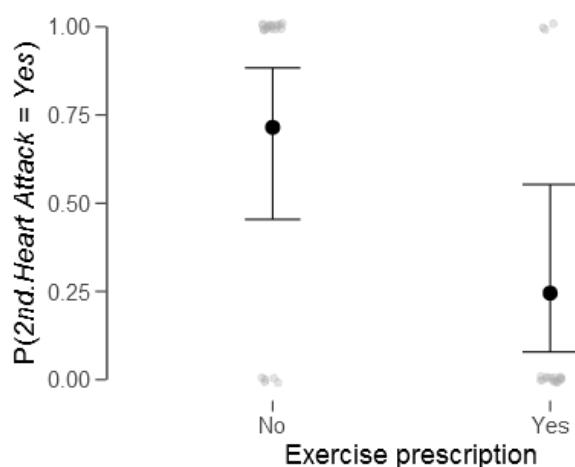
PLOTS

These findings can be easily visualised through the inferential plots.

The screenshot shows the 'Plots' section of the JASP interface. Under 'Inferential Plots', 'Conditional estimates plots' is checked, and 'Confidence interval' is set to 95.0%. 'Show data points' is also checked. Under 'Residual Plots', three types are listed: 'Predicted - residual plot', 'Predictor - residual plots', and 'Squared Pearson residuals plot'. Under 'Residual Type', 'Deviance' is selected. Under 'Performance Plots', there are two sections: 'ROC plot' with 'Cutoff step' at 0.2 and 'PR plot' with 'Cutoff step' at 0.2. Both sections have 'Add cutoff labels' options.



As stress levels increase, the probability of having a second heart attack increases.



No exercise intervention increases the probability of a 2nd heart attack, while it is reduced when it has been put in place.

REPORTING THE RESULTS

Logistic regression was performed to ascertain the effects of stress and exercise intervention on the likelihood that participants have a 2nd heart attack. The logistic regression model was statistically significant, $\chi^2 (37) = 21.257$, $p < .001$. The model correctly classified 75.0% of cases. Increasing stress was associated with an increased likelihood of a 2nd heart attack, but decreasing stress was associated with a reduction in the likelihood. The presence of an exercise intervention programme reduced the probability of a 2nd heart attack to 13%.



COMPARING MORE THAN TWO INDEPENDENT GROUPS

ANOVA

Whereas t-tests compare the means of two groups/conditions, one-way analysis of variance (**ANOVA**) compares the means of 3 or more groups/conditions. There are both independent and repeated measures ANOVAs available in JASP. ANOVA has been described as an 'omnibus test' which results in an F-statistic that compares whether the dataset's overall explained variance is significantly greater than the unexplained variance. **The null hypothesis tested is that there is no significant difference between the means of all the groups.** If the null hypothesis is rejected, ANOVA just states that there is a significant difference between the groups, but not where those differences occur. To determine where the group differences are, post hoc (From the Latin *post hoc*, "after this") tests are subsequently used.

Why not just multiple pairwise comparisons? If there are 4 groups (A, B, C, D), for example and the differences were compared using multiple t-tests:

- | | | |
|-----------|--------|---------------------|
| • A vs. B | P<0.05 | 95% no type I error |
| • A vs. C | P<0.05 | 95% no type I error |
| • A vs. D | P<0.05 | 95% no type I error |
| • B vs. C | P<0.05 | 95% no type I error |
| • B vs. D | P<0.05 | 95% no type I error |
| • C vs. D | P<0.05 | 95% no type I error |

Assuming that each test was independent, the overall probability would be:

$$0.95 * 0.95 * 0.95 * 0.95 * 0.95 * 0.95 = 0.735$$

This is known as familywise error or, cumulative Type I error, and in this case, results in only a 73.5% probability of no Type I error whereby the null hypothesis could be rejected when it is true. This is overcome by using post hoc tests that make multiple pairwise comparisons with stricter acceptance criteria to prevent familywise error.

ASSUMPTIONS

The independent ANOVA makes the same assumptions as most other parametric tests.

- The independent variable must be categorical, and the dependent variable must be continuous.
- The groups should be independent of each other.
- The dependent variable should be approximately normally distributed.
- There should be no significant outliers.
- There should be homogeneity of variance between the groups; otherwise, the p-value for the F-statistic may not be reliable.

The first 2 assumptions are usually controlled using an appropriate research method design.

If the last three assumptions are violated, then the non-parametric equivalent, Kruskal-Wallis, should be considered instead.



CONTRASTS

Contrasts are ‘a priori’ tests (i.e. planned comparisons before any data were collected). As an example, researchers may want to compare the effects of some new drugs to the currently prescribed ones. These should only be a small set of comparisons to reduce family-wise error. The choice must be based on the scientific questions being asked and chosen during the experimental design. Hence, the term planned comparisons. Therefore, they are looking at specified mean differences and can be used if the ANOVA F test is insignificant.

JASP provides 6 planned contrasts enabling different types of comparisons:

Deviation: the mean of each level of the independent variable is compared to the overall mean (the mean when all the levels are taken together).

Simple: the mean of each level is compared to the mean of a specified level, for example, with the mean of the control group.

Difference: the mean of each level is compared to the mean of the previous levels.

Helmert: the mean of each level is compared to the mean of the subsequent levels.

Repeated: By selecting this contrast, the mean of each level is compared to the mean of the following level.²

Polynomial: tests polynomial trends in the data.

POST HOC TESTING

Post hoc tests are tests that were decided upon after the data had been collected. They can only be carried out if the ANOVA F test is significant.

JASP provides 4 types of post hoc testing

Standard

Games-Howell – used when you are unsure about the equality of group variances.

Dunnett’s – used to compare all the groups to one group, i.e. the control group

Dunn – a non-parametric post hoc test used for testing small sub-sets of pairs.

JASP also provides 5 types of corrections for use with the independent group ANOVA tests:

Bonferroni – can be very conservative but gives guaranteed control over Type I errors at the risk of reducing statistical power. Does not assume independence of the comparisons.

Holm – the Holm-Bonferroni test is a sequential Bonferroni method that is less conservative than the original Bonferroni test.



Tukey – is one of the most commonly used tests and provides controlled Type I error for groups with the same sample size and equal group variance.

Scheffe – controls for the overall confidence level when the group sample sizes are different.

Sidak – is similar to Bonferroni but assumes that each comparison is independent of the others. Slightly more powerful than Bonferroni.

EFFECT SIZE

JASP provides 3 alternative effect size calculations for use with the independent group ANOVA tests:

Eta squared (η^2) – accurate for the sample variance explained but overestimates the population variance. This can make it difficult to compare the effect of a single variable in different studies.

Partial Eta squared (η_p^2) – this solves the problem relating to population variance overestimation, allowing for comparison of the effect of the same variable in different studies.

Omega squared (ω^2) – Normally, statistical bias gets very small as sample size increases, but for small samples ($n < 30$), ω^2 provides an unbiased effect size measure.

Test	Measure	Trivial	Small	Medium	Large
ANOVA	Eta	<0.1	0.1	0.25	0.37
	Partial Eta	<0.01	0.01	0.06	0.14
	Omega squared	<0.01	0.01	0.06	0.14

RUNNING THE INDEPENDENT ANOVA

Load **Independent ANOVA diets.csv**. This contains A column containing the 3 diets used (A, B and C) and another column containing the absolute amount of weight loss after 8 weeks on one of 3 different diets. For good practice, check the descriptive statistics and the boxplots for any extreme outliers.

Go to ANOVA > ANOVA, put weight loss into the Dependent Variable and the Diet groupings into the Fixed Factors box. In the first instance, tick Descriptive statistics and ω^2 as the effect size.

The screenshot shows the JASP ANOVA dialog box. At the top, there's a 'Dependent Variable' section with 'Weight loss kg' selected. Below that is a 'Fixed Factors' section. The main area is titled 'Display' and contains several checkboxes:

- Descriptive statistics
- Estimates of effect size
 - ω^2
 - partial ω^2
 - η^2
 - partial η^2



In Assumptions checks, tick all options:

▼ Assumption Checks

Homogeneity tests

Homogeneity corrections

None Brown-Forsythe Welch

Q-Q plot of residuals

This should result in 3 tables and one Q-Q plot.

UNDERSTANDING THE OUTPUT

ANOVA - Weight loss kg

Homogeneity Correction	Cases	Sum of Squares	df	Mean Square	F	p	ω^2
None	Diet	92.369	2.000	46.184	10.826	< .001	0.214
	Residuals	294.371	69.000	4.266			
Brown-Forsythe	Diet	92.369	2.000	46.184	10.826	< .001	0.214
	Residuals	294.371	64.352	4.574			
Welch	Diet	92.369	2.000	46.184	11.449	< .001	0.214
	Residuals	294.371	44.987	6.544			

Note. Type III Sum of Squares

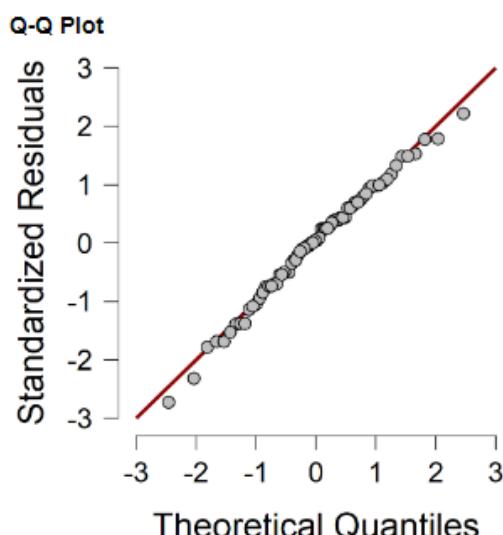
The main ANOVA table shows that the F-statistic is significant ($p<.001$) and that there is a large effect size. Therefore, there is a significant difference between the means of the 3 diet groups.

TESTING ASSUMPTIONS

Before accepting this, any violations in the assumptions required for an ANOVA should be checked.

Test for Equality of Variances (Levene's)			
F	df1	df2	p
1.298	2.000	69.000	0.280

Levene's test shows that the homogeneity of variance is not significant. However, if Levene's test shows a significant difference in variance, the Brown-Forsythe or Welch correction should be reported.



The Q-Q plot shows that the data appear to be normally distributed and linear.

Descriptives - Weight loss kg

Diet	N	Mean	SD	SE	Coefficient of variation
Diet A	24	3.008	1.668	0.340	0.554
Diet B	24	3.413	2.361	0.482	0.692
Diet C	24	5.587	2.108	0.430	0.377

The descriptive statistics suggest that Diet 3 results in the highest weight loss after 8 weeks.

CONTRAST EXAMPLE

If, for example, one planned to compare the effects of diets B and C to diet A, click on the drop-down menu and select 'simple' next to diet. This will test the significance between the first category in the list with the remaining categories.

▼ Contrasts

Factors

Diet simple ▾

Simple Contrast - Diet

Comparison	Estimate	SE	df	t	p
Diet B - Diet A	0.404	0.596	69.000	0.678	0.500
Diet C - Diet A	2.579	0.596	69.000	4.326	< .001

As can be seen, only diet C is significantly different from diet A ($t(69) = 4.326, p < .001$).



If the ANOVA reports no significant difference, you can go no further in the analysis.

POST HOC TESTING

If the ANOVA is significant, post hoc testing can now be carried out. In Post Hoc Tests, add Diet to the analysis box on the right, tick Standard type, use Tukey for the post hoc correction and tick the flag significant comparisons.

The screenshot shows the 'Post Hoc Tests' dialog in JASP. On the right side, under 'Analysis', 'Diet' is selected. In the 'Type' section, 'Standard' is checked. Under 'Correction', 'Tukey' is checked. At the bottom, 'Flag Significant Comparisons' is checked. Other options like 'From 1000 bootstraps' and 'Effect size' are available but unchecked.

Post hoc testing shows that there is no significant difference between weight loss in diets A and B. However, It is significantly higher in diet C compared to diet A ($p<.001$) and diet B ($p=.001$). Cohen's d shows that these differences have a large effect size.

Post Hoc Comparisons - Diet

		Mean Difference	SE	t	p _{tukey}
Diet A	Diet B	-0.404	0.596	-0.678	0.777
	Diet C	-2.579	0.596	-4.326	< .001***
	Diet B	-2.175	0.596	-3.648	0.001**

** $p < .01$, *** $p < .001$

Note. P-value adjusted for comparing a family of 3

Also, in Descriptive Plots, add the Factor 'Diet' to the horizontal axis and tick display error bars and in Raincloud plots, do the same but tick Horizontal display.

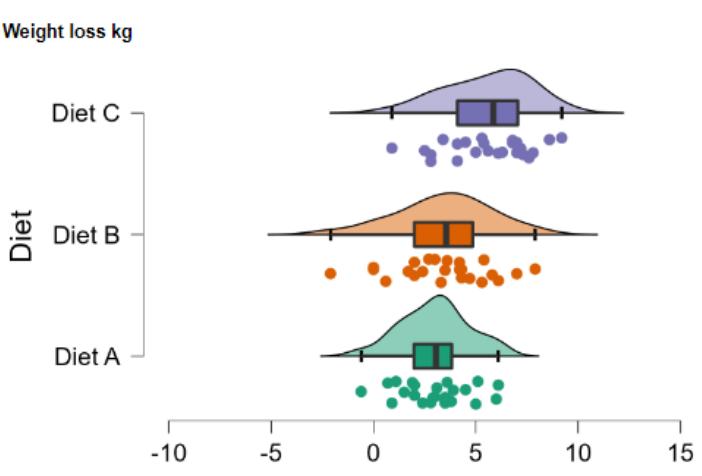
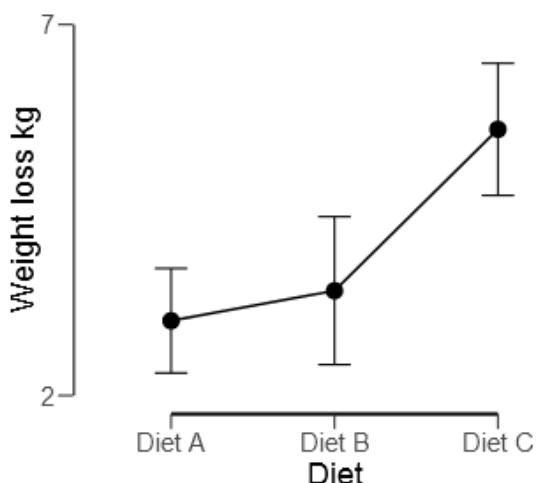


Descriptives Plots

Factors: Diet

Horizontal Axis: Diet

Display: Display error bars (checked), Confidence interval 95.0 %



REPORTING THE RESULTS

Levene's test showed that there was equal variance for all three groups: $F(2,69) = 1.3$, $p=.28^*$.

Independent one-way ANOVA showed a significant effect of the type of diet on weight loss after 10 weeks ($F(2, 69) = 46.184$, $p < .001$, The $\omega^2 = 0.214$, suggesting that this is a large effect size).

Post hoc testing using pairwise comparisons of the estimated marginal means with Tukey's correction revealed that diet C resulted in significantly greater weight loss ($M = 5.59$, $SD = 2.1$ kg) than diet A ($M = 3.0$, $SD = 2.36$ kg, $p < .001$) or diet B ($M = 3.41$, $SD = 2.36$ kg, $p = .001$). There were no significant differences in weight loss between diets A and B ($p = .777$).

* If Levene's test shows a significant difference in variance, the Brown-Forsythe or Welch correction should be reported.



KRUSKAL-WALLIS – NON-PARAMETRIC ANOVA

If your data fails parametric assumption tests or is nominal, the Kruskal-Wallis H test is a non-parametric equivalent to the independent samples ANOVA. It can be used for comparing two or more independent samples of equal or different sample sizes. Like the Mann-Whitney and Wilcoxon tests, it is a rank-based test.

As with the ANOVA, the Kruskal-Wallis H test (also known as the "one-way ANOVA on ranks") is an omnibus test that does not specify which specific groups of the independent variable are statistically significantly different from each other. To do this, JASP provides the option for running Dunn's post hoc test. These multiple comparison tests can be very conservative, particularly for large numbers of comparisons.

Load the **Kruskal-Wallis ANOVA.csv** dataset into JASP. This dataset contains subjective pain scores for participants undergoing no treatment (control), cryotherapy or combined cryotherapy-compression for delayed onset muscle soreness after exercise.

RUNNING THE KRUSKAL-WALLIS TEST

Go to ANOVA >ANOVA. In the analysis window, add the Pain score to the dependent variable and treatment to the fixed factors. Check that the pain score is set to ordinal. This will automatically run the normal independent ANOVA. Under Assumption Checks, tick both Homogeneity tests and Q-Q plots.

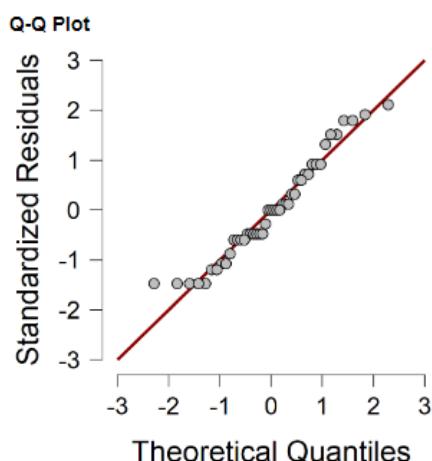
ANOVA - Pain Score

Cases	Sum of Squares	df	Mean Square	F	p
Treatment	98.844	2.000	49.422	16.457	< .001
Residual	126.133	42.000	3.003		

Note. Type III Sum of Squares

Test for Equality of Variances (Levene's)

F	df1	df2	p
3.832	2.000	42.000	0.030





Although the ANOVA indicates a significant result, the data have not met the assumptions of homogeneity of variance as seen by the significant Levene's test and only shows linearity in the middle of the Q-Q plot and curves off at the extremities, indicating more extreme values. Added to the fact that the dependent variable is based on subjective pain scores suggests the use of a non-parametric alternative.

Return to the statistics options and open the Nonparametric option at the bottom. For the Kruskal-Wallis test, move the Treatment variable to the box on the right and tick Dunn's post hoc test.

The screenshot shows the JASP software's 'Nonparametrics' section. Under the 'Kruskal-Wallis Test' heading, the 'Treatment' variable is listed in the right-hand selection box. At the bottom of the dialog, there are two checkboxes: 'Estimates of effect size' (unchecked) and 'Dunn's post hoc tests' (checked, indicated by a blue checkmark).

UNDERSTANDING THE OUTPUT

Two tables are shown in the output. The Kruskal-Wallis test shows that there is a significant difference between the three treatment modalities.

Kruskal-Wallis Test

Factor	Statistic	df	p
Treatment	19.693	2	< .001

Dunn

Dunn's Post Hoc Comparisons - Treatment

Comparison	z	W _i	W _j	p	p _{bonf}	p _{holm}
Control - Cryo+Compression	4.317	34.600	14.200	< .001***	< .001***	< .001***
Control - Cryotherapy	3.048	34.600	20.200	0.001**	0.003**	0.002**
Cryo+Compression - Cryotherapy	-1.270	14.200	20.200	0.102	0.306	0.102

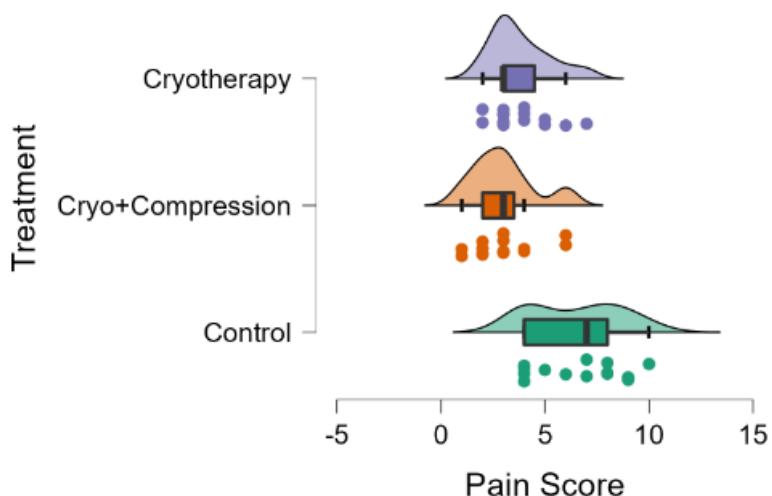
** p < .01, *** p < .001

Dunn's post hoc test provides its p-value as well as those for Bonferroni and Holm's Bonferroni correction. As can be seen, both treatment conditions are significantly different from the controls, but not from each other.



Descriptive Statistics

	Pain Score		
	Control	Cryo+Compression	Cryotherapy
Median	7.000	3.000	3.000
MAD	2.000	1.000	1.000
IQR	4.000	1.500	1.500



REPORTING THE RESULTS

Pain scores were significantly affected by treatment modality, $H(2) = 19.693$, $p < .001$. Pairwise comparisons using Dunn's post hoc showed that both cryotherapy ($Mdn = 3$) and cryotherapy with compression ($Mdn = 3$) significantly reduced pain scores ($p = .002$ and $p < .001$, respectively) compared to the control group ($Mdn = 7$). There was no significant difference between cryotherapy and cryotherapy with compression ($p = .102$).



COMPARING MORE THAN TWO RELATED GROUPS

RMANOVA

The one-way repeated measures ANOVA (**RMANOVA**) is used to assess if there is a difference in means between 3 or more groups (where the participants are the same in each group) that have been tested multiple times or under different conditions. Such a research design, for example, could be that the same participants were tested for an outcome measure at 1, 2 and 3 weeks or that the outcome was tested under conditions 1, 2 and 3.

The null hypothesis tested is that there is no significant difference between the means of the differences between all the groups.

The independent variable should be categorical, and the dependent variable needs to be a continuous measure. In this analysis, the independent categories are termed **levels**, i.e. these are the related groups. So in the case where an outcome was measured at weeks 1, 2 and 3, the 3 levels would be week 1, week 2 and week 3.

The **F-statistic** is calculated by dividing the mean squares for the variable (variance explained by the model) by its error mean squares (unexplained variance). The larger the F-statistic, the more likely it is that the independent variable will have had a significant effect on the dependent variable.

ASSUMPTIONS

The RMANOVA makes the same assumptions as most other parametric tests.

- The dependent variable should be approximately normally distributed.
- There should be no significant outliers.
- Sphericity relates to the equality of the variances of the differences between levels of the repeated measures factor.

If the assumptions are violated, then the non-parametric equivalent, **Friedman's test**, should be considered instead and is described later in this section.

SPHERICITY

If a study has 3 levels (A, B and C), sphericity assumes the following:

$$\text{Variance (A-B)} \approx \text{Variance (A-C)} \approx \text{Variance (B-C)}$$

RMANOVA checks the assumption of sphericity using Mauchly's (pronounced Mockley's) test of sphericity. This tests **the null hypothesis that the variances of the differences are equal**. In many cases, repeated measures violate the assumption of sphericity, which can lead to Type I error. If this is the case, corrections to the F-statistic can be applied.

JASP offers two methods of correcting the F-statistic, the **Greenhouse-Geisser** and the **Huynh-Feldt** epsilon (ϵ) corrections. A general rule of thumb is that if the ϵ values are <0.75 , then use the Greenhouse-Geisser correction and if they are >0.75 then use the Huynh-Feldt correction.



POST HOC TESTING

Post hoc testing is limited in RMANOVA. JASP provides two alternatives:

Bonferroni – can be very conservative but gives guaranteed control over Type I errors at the risk of reducing statistical power.

Holm – the Holm-Bonferroni test is a sequential Bonferroni method that is less conservative than the original Bonferroni test.

If you ask for either Tukey or Scheffe post hoc corrections, JASP will return a NaN (not a number) error.

EFFECT SIZE

JASP provides the same alternative effect size calculations that are used with the independent group ANOVA tests:

Eta squared (η^2) – accurate for the sample variance explained, but overestimates the population variance. This can make it difficult to compare the effect of a single variable in different studies.

Partial Eta squared (η_p^2) – this solves the problem relating to population variance overestimation, allowing for comparison of the effect of the same variable in different studies. This appears to be the most commonly reported effect size in repeated measures ANOVA.

Omega squared (ω^2) – Normally, statistical bias gets very small as sample size increases, but for small samples ($n < 30$), ω^2 provides an unbiased effect size measure.

Levels of effect size:

Test	Measure	Trivial	Small	Medium	Large
ANOVA	Eta	<0.1	0.1	0.25	0.37
	Partial Eta	<0.01	0.01	0.06	0.14
	Omega squared	<0.01	0.01	0.06	0.14

RUNNING THE REPEATED MEASURES ANOVA

Load **Repeated ANOVA cholesterol.csv**. This contains one column with the participant IDs and 3 columns, one for each repeated measurement of blood cholesterol following an intervention. For good practice, check the descriptive statistics and the boxplots for any extreme outliers.

Go to ANOVA > Repeated measures ANOVA. As stated above, the independent variable (repeated measures factor) has levels; in this case, there are 3 levels. Rename RM Factor 1 to Time post-intervention, and then rename 3 levels to Week 0, Week 3 and Week 6 accordingly.



Once these have been done, they will appear in the Repeated Measures Cells. Now add the appropriate data to the appropriate level.

The screenshot shows the JASP interface for a Repeated Measures ANOVA. On the left, under 'Repeated Measures ANOVA', there is a 'Participant' section with 'Week 6' selected. A large black arrow points from this section to the 'Repeated Measures Cells' section on the right. The 'Repeated Measures Factors' section contains 'Time post intervention' with levels 'Week 0', 'Week 3', 'Week 6', and 'Level 4'. An 'RM Factor 2' button is also present. The 'Repeated Measures Cells' section shows a 3x2 grid of cells: Week 0, Week 0; Week 3, Week 3; Week 6, Week 6. There is a small orange pencil icon at the bottom right of this section.

Tick Descriptive Statistics, Estimates of effect size and ω^2 .

The screenshot shows the 'Display' settings in JASP. Under 'Descriptive statistics', all checkboxes are checked: 'Descriptive statistics', 'Estimates of effect size', ' ω^2 ' (with a checked checkbox), ' η^2 ' (unchecked), 'partial ω^2 ' (unchecked), 'partial η^2 ' (unchecked), 'general η^2 ' (unchecked), and 'Vovk-Sellke maximum p-ratio' (unchecked).

Under Assumption Checks, tick Sphericity tests and all Sphericity correction options.

The screenshot shows the 'Assumption Checks' settings in JASP. Under 'Sphericity tests', 'Sphericity corrections' is checked. Under 'Sphericity corrections', 'None' (unchecked), 'Greenhouse-Geisser' (checked), and 'Huynh-Feldt' (checked) are listed. 'Homogeneity tests' (unchecked) is also present.

The output should consist of 4 tables. **The third table, between-subject effects, can be ignored for this analysis.**



UNDERSTANDING THE OUTPUT

Within Subjects Effects

	Sphericity Correction	Sum of Squares	df	Mean Square	F	p	ω^2
Time post intervention	None	4.320*	2.000*	2.160*	212.321*	< .001*	0.058
	Greenhouse-Geisser	4.320*	1.235*	3.497*	212.321*	< .001*	0.058
	Huynh-Feldt	4.320*	1.284*	3.365*	212.321*	< .001*	0.058
Residual	None	0.346	34.000	0.010			
	Greenhouse-Geisser	0.346	21.001	0.016			
	Huynh-Feldt	0.346	21.822	0.016			

Note. Type III Sum of Squares

* Mauchly's test of sphericity indicates that the assumption of sphericity is violated ($p < .05$).

The within-subjects effects table reports a large F-statistic, which is highly significant ($p<.001$) and has a small to medium effect size (0.058). This table shows the statistics for sphericity assumed (none) and the two correction methods. The main differences are in the degrees of freedom (df) and the value of the mean square. Under the table, it is noted that the assumption of sphericity has been violated.

The following table gives the results of Mauchly's test of sphericity. There is a significant difference ($p<.001$) in the variances of the differences between the groups. Greenhouse-Geisser and the Huynh-Feldt epsilon (ϵ) values are below 0.75. Therefore, the ANOVA result should be reported based on the Greenhouse-Geisser correction:

Assumption Checks

Test of Sphericity

	Mauchly's W	Approx. χ^2	dfSphericity	p-value	Greenhouse-Geisser ϵ	Huynh-Feldt ϵ	Lower Bound ϵ
Time post intervention	0.381	15.440	2	< .001	0.618	0.642	0.500

To provide a cleaner table, go back to Assumption Checks and only tick Greenhouse-Geisser for sphericity correction.

Within Subjects Effects

	Sphericity Correction	Sum of Squares	df	Mean Square	F	p	ω^2
Time post intervention	Greenhouse-Geisser	4.320*	1.235*	3.497*	212.321*	< .001*	0.058
Residual	Greenhouse-Geisser	0.346	21.001	0.016			

Note. Type III Sum of Squares

* Mauchly's test of sphericity indicates that the assumption of sphericity is violated ($p < .05$).

There is a significant difference between the means of the differences between all the groups, $F(1.235, 21.0) = 212.3, p < .001, \omega^2 = 0.058$.



Descriptives

Time post intervention	N	Mean	SD	SE	Coefficient of variation
Week 0	18	6.408	1.191	0.281	0.186
Week 3	18	5.842	1.123	0.265	0.192
Week 6	18	5.779	1.102	0.260	0.191

The descriptive data suggest that blood cholesterol levels were higher at week 0 compared to weeks 3 and 6.

However, if the ANOVA reports no significant difference, you can go no further in the analysis.

POST HOC TESTING

If the ANOVA is significant, post hoc testing can now be carried out. In Post Hoc Tests, add Time post-intervention to the analysis box on the right, tick Effect size, and, in this case, use Holm for the post hoc correction.

The screenshot shows the 'Post Hoc Tests' dialog box in JASP. At the top, there is a dropdown menu labeled 'Post Hoc Tests'. Below it, a large rectangular box contains the variable 'Time post intervention' with a right-pointing arrow button above it. Underneath this box are several configuration options:

- Effect size
- Pool error term for follow-up tests
- Conditional comparisons for interactions

Below these are sections for 'Correction' and 'Display':

- Correction:** A checkbox for 'Holm' is checked, while 'Bonferroni', 'Tukey', and 'Scheffé' are unchecked.
- Display:** A checkbox for 'Flag significant comparisons' is checked. Next to it is a 'Confidence intervals' field set to 95.0%.



Also, in Descriptive Plots, add the Factor – Time post-intervention to the horizontal axis and tick display error bars.

Post hoc testing shows that there are significant differences in blood cholesterol levels between all of the time point combinations and are associated with large effect sizes.

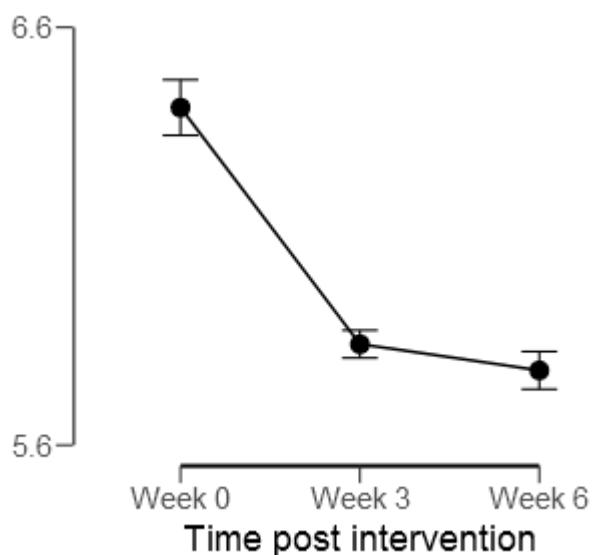
Post Hoc Comparisons - Time post intervention

		Mean Difference	SE	t	Cohen's d	p _{holm}
Week 0	Week 3	0.566	0.037	15.439	0.497	< .001***
	Week 6	0.629	0.042	14.946	0.552	< .001***
Week 3	Week 6	0.063	0.017	3.781	0.055	0.001**

** p < .01, *** p < .001

Note. Computation of Cohen's d based on pooled error.

Note. P-value adjusted for comparing a family of 3



REPORTING THE RESULTS

Mauchly's test of sphericity showed a significant difference in variation between the group differences: $W (2) = 0.381, p<.001$. Therefore, the Greenhouse-Geisser correction was used. This showed that cholesterol levels differed significantly between time points, $F(1.235, 21.0) = 212.3, p<.001$. $\omega^2 = 0.058$, suggesting that this is a small to medium effect.

Post hoc testing using the Holm correction revealed that cholesterol levels decreased significantly as time increased, weeks 0 – 3 ($M = 0.566, SE = 0.04, p<.001$), weeks 1 – 6 ($M = 0.63 SE = 0.04, p=.004$) and weeks 3-6 ($M=0.06, SE = 0.016$).



FRIEDMAN'S REPEATED MEASURES ANOVA

If parametric assumptions are violated or the data is ordinal, you should consider using the non-parametric alternative, Friedman's test. Similar to the Kruskal-Wallis test, Friedman's test is used for one-way repeated measures analysis of variance by ranks and doesn't assume the data comes from a particular distribution. This test is another omnibus test that does not specify which specific groups of the independent variable are statistically significantly different from each other. To do this, JASP provides the option for running Conover's post hoc test if Friedman's test is significant.

Load **Friedman RMANOVA.csv** into JASP. This has 3 columns of subjective pain ratings measured at 18-, 36- and 48-hours post-exercise. Check that the pain scores are set to ordinal data.

RUNNING THE FRIEDMAN'S TEST

Go to ANOVA > Repeated measures ANOVA. The independent variable (repeated measures factor) has 3 levels. Rename RM Factor 1 to Time, and then rename the 3 levels to 18 hours, 36 hours and 48 hours accordingly.

Once these have been done, they will appear in the Repeated Measures Cells. Now add the appropriate dataset to the appropriate level.

The screenshot shows the JASP interface for a Repeated Measures ANOVA. On the left, there is a tree view under 'Subject'. In the center, the 'Repeated Measures Factors' section lists 'Time' with levels '18 hours', '36 hours', and '48 hours'. Below it, the 'Repeated Measures Cells' section displays a 3x2 grid of cells, each containing a pain rating and its corresponding time point: Pain 18 hours / 18 hours, Pain 36 hours / 36 hours, and Pain 48 hours / 48 hours.

This will automatically produce the standard repeated measures within-subjects ANOVA table. To run a Friedman's test, expand the Nonparametrics tab, move Time to the RM factor box and tick Conover's post hoc tests.

The screenshot shows the 'Nonparametrics' dialog box. Under 'Factors', there is an empty list. Under 'RM Factor', 'Time' is selected. Under 'Optional Grouping Factor', there is an empty list. At the bottom, a checkbox labeled 'Conover's post hoc tests' is checked.



UNDERSTANDING THE OUTPUT

Two tables should be produced.

Nonparametrics

Friedman Test

Factor	χ^2_F	df	p	Kendall's W
Time	26.772	2	< .001	0.892

Conover Test

Conover's Post Hoc Comparisons - Time

		T-Stat	df	W_i	W_j	r_{rb}	p	p_{bonf}	p_{holm}
18 hours	36 hours	15.171	28	17.000	44.500	-1.000	< .001	< .001	< .001
	48 hours	6.344	28	17.000	28.500	-0.857	< .001	< .001	< .001
36 hours	48 hours	8.827	28	44.500	28.500	1.000	< .001	< .001	< .001

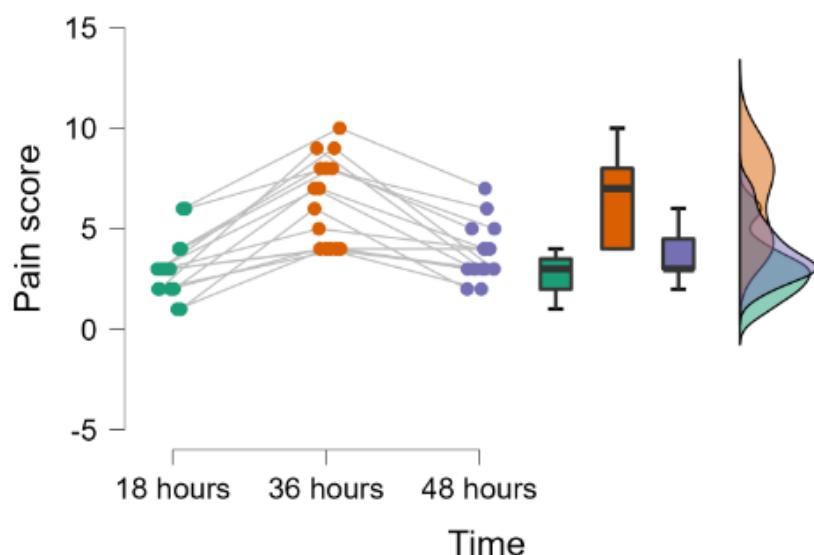
Note. Grouped by subject.

Note. Rank-biserial correlation based on individual signed-rank tests.

Friedman's test shows that time has a significant effect on pain perception. Connor's post hoc pairwise comparisons show that all pain perception is significantly different between each time point.

Descriptive Statistics

	Pain 18 hours	Pain 36 hours	Pain 48 hours
Median	3.000	7.000	3.000
MAD	1.000	2.000	1.000
IQR	1.500	4.000	1.500





REPORTING THE RESULTS

Friedmann's test showed that time has a significant effect on subjective pain scores $\chi^2_F(2) = 26.77$, $p < .001$. Conover's pairwise post hoc comparisons showed that pain perception peaked at 36 hours ($Mdn = 7$) and was significantly higher than at 18 hours ($Mdn = 3$: $T(28) = 15.17$, $p < .001$) and then decreased by 48 hours ($Mdn = 3$: $T(28) = 8.82$ $p < .001$).



COMPARING INDEPENDENT GROUPS AND THE EFFECTS OF COVARIATES

ANCOVA

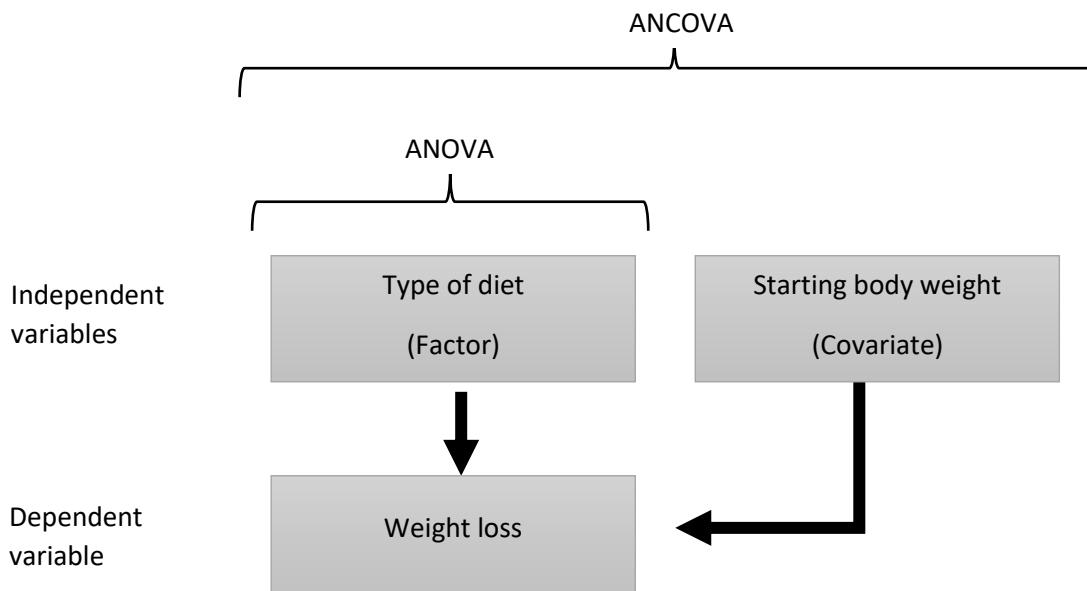
ANOVA can be used to compare the means of one variable (dependent) in two or more groups, whereas analysis of covariance (**ANCOVA**) sits between ANOVA and regression and compares the means of one (dependent) variable in two or more groups while considering the variability of other continuous variables (**COVARIATES**). ANCOVA checks for differences in ‘adjusted’ means (i.e. adjusted for the effects of the covariate). A covariate may not usually be part of the main research question, but could influence the dependent variable and therefore needs to be adjusted or controlled for. If a good covariate is used, ANCOVA will have improved statistical power and control over error.

Control for – to subtract statistically the effects of a variable (a control variable) to see what a relationship would be without it (Vogt 1977).

Hold constant – to “subtract” the effects of a variable from a complex relationship to study what the relationship would be if the variable were, in fact, a constant. Holding a variable constant essentially means assigning it an average value (Vogt 1977).

Statistical control – using statistical techniques to isolate or “subtract” variance in the dependent variable attributable to variables that are not the subject of the study (Vogt, 1999).

For example, when looking for a difference in weight loss between the three diets, it would be appropriate to take into account the individuals' pre-trial bodyweight since heavier people may lose proportionately more weight.





The null hypothesis tested is that there is no significant difference between the ‘adjusted’ means of all the groups.

ASSUMPTIONS

ANCOVA makes the same assumptions as the independent ANOVA. However, there are two further assumptions:

- The relationship between the dependent and covariate variables is linear.
- **Homogeneity of regression**, i.e. the regression lines for each of the independent groups are parallel to each other.



POST HOC TESTING

JASP provides 4 Types

- **Standard** – as above
- **Games-Howell** – used when you are unsure about the equality of group variances.
- **Dunnett’s** – used to compare all the groups to one group, i.e. the control group
- **Dunn** – a non-parametric post hoc test used for testing small sub-sets of pairs.

JASP additionally provides 4 corrections for use with the independent group ANOVA tests:

- **Bonferroni** – can be very conservative but gives guaranteed control over Type I errors at the risk of reducing statistical power.
- **Holm** – the Holm-Bonferroni test is a sequential Bonferroni method that is less conservative than the original Bonferroni test.
- **Tukey** – is one of the most commonly used tests and provides controlled Type I error for groups with the same sample size and equal group variance.
- **Scheffe** – controls for the overall confidence level when the group sample sizes are different.



EFFECT SIZE

JASP provides 3 alternative effect size calculations for use with the independent group ANOVA tests:

- **Eta squared (η^2)** - accurate for the sample variance explained, but overestimates the population variance. This can make it difficult to compare the effect of a single variable in different studies.
- **Partial Eta squared (η_p^2)** – this solves the problem relating to population variance overestimation, allowing for comparison of the effect of the same variable in different studies.
- **Omega squared (ω^2)** – Normally, statistical bias gets very small as sample size increases, but for small samples ($n < 30$), ω^2 provides an unbiased effect size measure.

Test	Measure	Trivial	Small	Medium	Large
ANOVA	Eta	<0.1	0.1	0.25	0.37
	Partial Eta	<0.01	0.01	0.06	0.14
	Omega squared	<0.01	0.01	0.06	0.14

RUNNING THE INDEPENDENT ANCOVA

Load **ANCOVA hangover.csv**. This dataset has been adapted from the one provided by Andy Field (2017). The morning after a Fresher's ball, students were given either water, coffee or a Barocca to drink. Two hours later, they reported how well they felt (from 0 – awful to 10 –very well). At the same time, data were collected on how drunk they were the night before (0-10).

Initially, run an ANOVA with wellness as the dependent variable and the type of drink as the fixed factor.

ANCOVA - wellness

Cases	Sum of Squares	df	Mean Square	F	p
drink	4.267	2	2.133	1.714	0.199
Residuals	33.600	27	1.244		

Note. Type III Sum of Squares

As can be seen from the results, homogeneity of variance has not been violated, while the ANOVA shows that there is no significant difference in the wellness scores between any of the morning drinks. $F(2,27)=1.714$, $p=.199$. However, this may be related to how drunk the students were the night before!

Go to ANOVA > ANCOVA, put wellness as the dependent variable and the type of drink as the fixed factor. Now add drunkenness to the Covariate(s) box. In the first instance, tick Descriptive statistics and ω^2 as the effect size;



Dependent Variable: wellness

Fixed Factors: drink

Covariates: drunkeness

WLS Weights

Display:

- Descriptive statistics
- Estimates of effect size
 - ω^2
 - partial ω^2
 - η^2
 - partial η^2

In Assumption Checks, tick both options.

Assumption Checks

- Homogeneity tests
- Q-Q plot of residuals

In Marginal Means, move drink to the right.

Marginal Means

drink

From 1000 bootstraps

Compare marginal means to 0

Confidence interval adjustment: None

This should result in 4 tables and one Q-Q plot.



UNDERSTANDING THE OUTPUT

ANCOVA - wellness

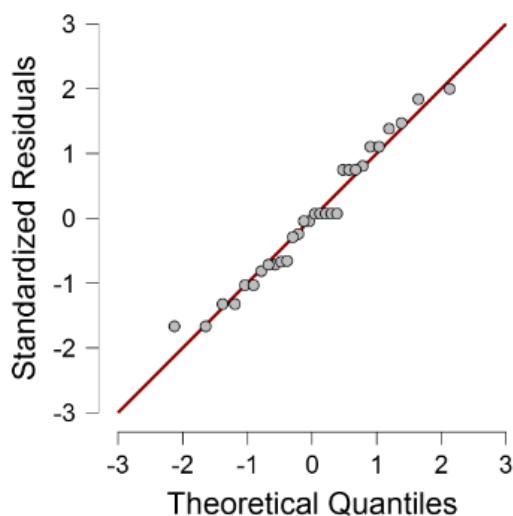
Cases	Sum of Squares	df	Mean Square	F	p	ω^2
drink	8.503	2	4.252	7.470	0.003	0.173
drunkeness	18.801	1	18.801	33.032	< .001	0.427
Residuals	14.799	26	0.569			

Note. Type III Sum of Squares

Test for Equality of Variances (Levene's)

F	df1	df2	p
3.580	2	27	0.042

The covariate (drunkenness) significantly predicts wellness ($p<.001$). The effects of the type of drink on wellness, when adjusted for the effects of drunkenness, are now significant ($p=.003$).



Levene's test is significant, unlike in ANOVA, where no homogeneity of variance corrections (i.e. Welch) are provided. For ANCOVA, this can be ignored. The Q-Q plot appears to be normal.

Descriptives - wellness

drink	N	Mean	SD	SE	Coefficient of variation
Barocca	10	6.000	1.414	0.447	0.236
Coffee	10	6.000	0.667	0.211	0.111
Water	10	5.200	1.135	0.359	0.218



The descriptive statistics show the unadjusted means for wellness in the three drink groups.

Marginal Means - drink

drink	Marginal Mean	95% CI for Mean Difference		
		Lower	Upper	SE
Barocca	6.571	6.040	7.103	0.258
Coffee	5.273	4.718	5.828	0.270
Water	5.356	4.862	5.849	0.240

The marginal means are now the wellness means, having been adjusted for the effects of the covariate (drunkenness).

TESTING FURTHER ASSUMPTIONS

As previously mentioned, the assumption of homogeneity of regression is important in ANCOVA. This can be tested by looking at the interaction between the type of drink and the drunkenness scores. Go to Model, drink, and drunkenness will have been automatically added as individual Model terms. Now highlight both drink and drunkenness and add them both to Model terms.

ANCOVA - wellness

Cases	Sum of Squares	df	Mean Square	F	p	ω^2
drink	2.290	2	1.145	1.876	0.175	0.047
drunkenness	4.865	1	4.865	7.971	0.009	0.189
drink * drunkenness	0.150	2	0.075	0.123	0.885	0.000
Residual	14.649	24	0.610			

Note. Type III Sum of Squares

The ANOVA table now has an extra row showing the interaction between the type of drink and drunkenness. This is not significant ($p=.885$), i.e. the relationships between drunkenness and wellness are the same in each drink group. If this is significant, there will be concerns over the validity of the main ANCOVA analysis.

Having checked this, go back and remove the interaction term from the Model terms.

If the ANCOVA reports no significant difference, you can go no further in the analysis.



POST HOC TESTING

If the ANCOVA is significant, post hoc testing can now be carried out. In Post Hoc Tests, add Drink to the analysis box on the right, tick Effect size, and, in this case, use Tukey for the post hoc correction. Also, tick flag significant comparisons.

The screenshot shows the 'Post Hoc Tests' dialog box. On the left, there is a large empty list box. To its right is a smaller box containing a blue circular icon with a white 'd' and the word 'drink'. Below these are two sections: 'Type' and 'Display'. Under 'Type', 'Standard' is checked, along with 'Effect size'. Under 'Display', 'Flag significant comparisons' is checked. To the right of these sections is a 'Correction' section where 'Tukey' is selected, along with several other options that are not checked: Scheffé, Bonferroni, Holm, and Šidák.

Standard (HSD)

Post Hoc Comparisons - drink

		Mean Difference	SE	df	t	Cohen's d	P _{tukey}
Barocca	Coffee	1.298	0.406	26	3.198	1.721	0.010**
	Water	1.215	0.345	26	3.523	1.611	0.004**
Coffee	Water	-0.083	0.371	26	-0.224	-0.110	0.973

** p < .01

Note. P-value adjusted for comparing a family of 3 estimates.

Post hoc testing shows that there is no significant difference between coffee and water on wellness. However, wellness scores were significantly higher after drinking a Barocca.

This can be seen from the Descriptive plots.



Descriptives Plots

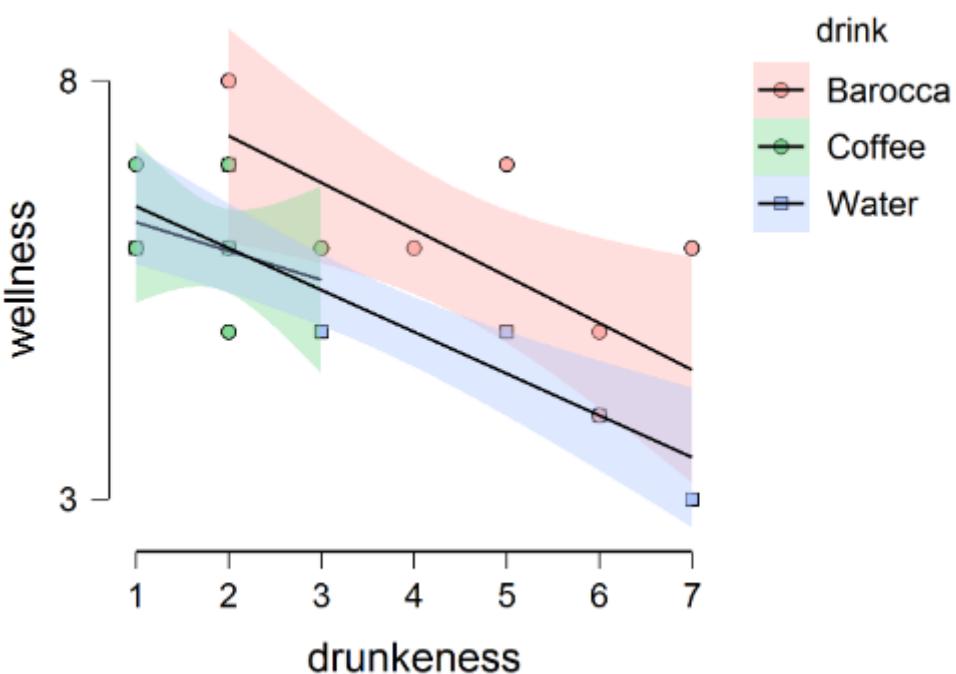
Factors: drunkenness

Horizontal Axis: drink

Separate Lines: Separate Plots

Display:

- Display error bars
- Confidence intervals %
- Standard error



REPORTING THE RESULTS

The covariate, drunkenness, was significantly related to the morning after wellness, $F(1,26) = 33.03$, $p < .001$, $\omega^2 = 0.427$. There was also a significant effect of the drink on wellness after controlling for drunkenness, $F(2, 26) = 7.47$, $p = .003$, $\omega^2 = 0.173$.

Post hoc testing using Tukey's correction revealed that drinking a Barocca resulted in significantly greater wellness compared to water ($p = .004$) and compared to coffee ($p = .01$). There were no significant differences in wellness between water and coffee ($p = .973$).



TWO-WAY INDEPENDENT ANOVA

One-way ANOVA tests situations when only one independent variable is manipulated, and two-way ANOVA is used when more than 1 independent variable has been manipulated. In this case, independent variables are known as factors.

FACTOR 1	FACTOR 2	
CONDITION 1	Group 1	Dependent variable
	Group 2	Dependent variable
CONDITION 2	Group 1	Dependent variable
	Group 2	Dependent variable
CONDITION 3	Group 1	Dependent variable
	Group 2	Dependent variable

The factors are split into levels; therefore, in this case, Factor 1 has 3 levels and Factor 2 has 2 levels.

A “main effect” is the effect of one of the independent variables on the dependent variable, ignoring the effects of any other independent variables. There are 2 main effects tested, both of which are “**between-subjects**”: in this case, comparing differences between factor 1 (i.e. condition) and differences between factor 2 (i.e. groups). **Interaction** is where one factor influences the other factor.

The two-way independent ANOVA is another omnibus test that is used to test 2 null hypotheses:

1. *There is no significant between-subject effect, i.e. no significant difference between the means of the groups in either of the factors.*
2. *There is no significant interaction effect, i.e. no significant group differences across conditions.*

ASSUMPTIONS

Like all other parametric tests, mixed factor ANOVA makes a series of assumptions that should either be addressed in the research design or can be tested for.

- The independent variables (factors) should have at least two categorical independent groups (levels).
- The dependent variable should be continuous and approximately normally distributed for all combinations of factors.
- There should be homogeneity of variance for each of the combinations of factors.
- There should be no significant outliers.

RUNNING TWO-WAY INDEPENDENT ANOVA

Open **2-way independent ANOVA.csv** in JASP. This comprises 3 columns of data, Factor 1 – gender with 2 levels (male and female), Factor 2 - supplement with 3 levels (control, carbohydrate CHO and protein) and the dependent variable (explosive jump power). In Descriptive statistics, check the data for significant outliers. Go to ANOVA >ANOVA, add Jump power to the Dependent variable, Gender and Supplement to the Fixed factors.



▼ ANOVA

Dependent Variable
Jump power

Fixed Factors
Gender
Supplement

Display

Descriptive statistics

Estimates of effect size

ω^2 partial ω^2

η^2 partial η^2

Confidence intervals 95.0 %

Tick Descriptive statistics and Estimates of effect size (ω^2).

In Descriptive plots, add the supplement to the horizontal axis and Gender to separate lines. In Additional Options,

▼ Descriptives Plots

Factors

Horizontal Axis
Supplement

Separate Lines
Gender

Separate Plots

Display

Display error bars

Confidence interval 95 %

Standard error



UNDERSTANDING THE OUTPUT

The output should comprise 2 tables and one plot.

ANOVA - Jump power

Cases	Sum of Squares	df	Mean Square	F	p	ω^2
Gender	119108.037	1.000	119108.037	9.589	0.003	0.058
Supplement	896116.137	2.000	448058.068	36.071	< .001	0.477
Gender * Supplement	275806.438	2.000	137903.219	11.102	< .001	0.138
Residual	521712.054	42.000	12421.716			

Note. Type III Sum of Squares

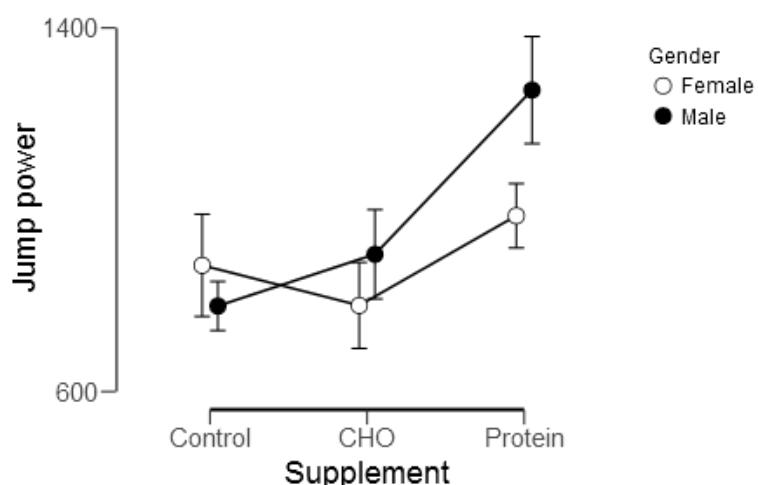
The ANOVA table shows that there are significant main effects for both Gender and Supplement ($p=0.003$ and $p<.001$, respectively) with medium and large effect sizes, respectively. This suggests that there is a significant difference in jump power between genders, irrespective of Supplement, and significant differences between supplements, irrespective of Gender.

There is also a significant interaction between Gender and Supplement ($p<.001$), which also has a medium to large effect size (0.138). This suggests that the differences in jump power between genders are affected somehow by the type of supplement used.

The Descriptive statistics and plot suggest that the main differences are between genders when using a protein supplement.

Descriptives - Jump power

Gender	Supplement	N	Mean	SD	SE	Coefficient of variation
Female	CHO	7	789.286	102.283	38.660	0.130
	Control	8	877.500	134.563	47.575	0.153
	Protein	9	986.667	91.924	30.641	0.093
Male	CHO	8	901.875	117.502	41.543	0.130
	Control	8	788.125	64.417	22.775	0.082
	Protein	8	1263.125	140.863	49.803	0.112





TESTING ASSUMPTIONS

In Assumption Checks, tick Homogeneity tests and Q-Q plot of residuals.

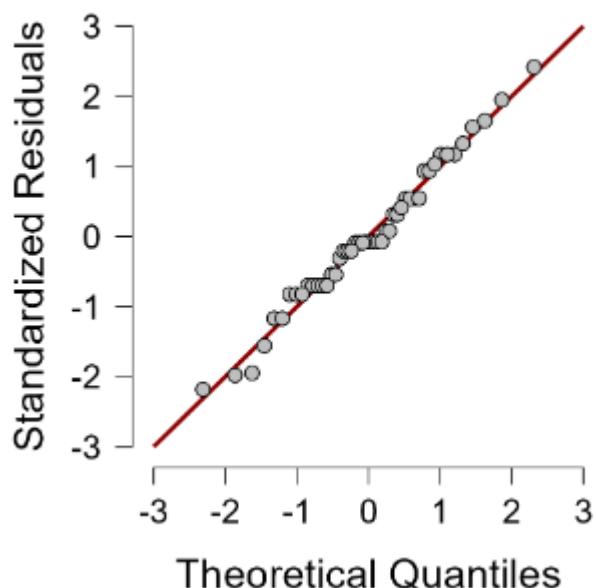
Assumption Checks

Test for Equality of Variances (Levene's)

F	df1	df2	p
1.100	5.000	42.000	0.375

Levene's test shows no significant difference in variance within the dependent variable groups; thus, homogeneity of variance has not been violated.

Q-Q Plot



The Q-Q plot shows that the data appear to be normally distributed and linear. We can now accept the ANOVA result since none of these assumptions has been violated.

However, if the ANOVA reports no significant difference, you can go no further with the analysis.

SIMPLE MAIN EFFECTS

Go to the analysis options and Simple Main Effects. Here, add Gender to the Simple effect factor and Supplement to the Moderator Factor 1. Simple main effects are effectively limited to pairwise comparisons.



▼ Simple Main Effects

Factors

Simple Effect Factor

Gender

Moderator Factor 1

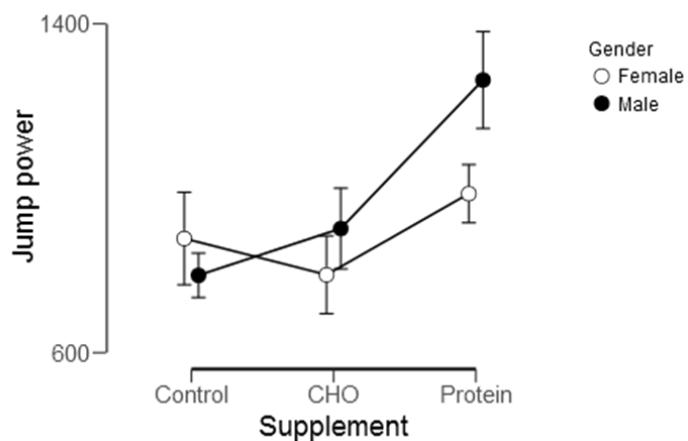
Supplement

Moderator Factor 2

Simple Main Effects - Gender

Level of Supplement	Sum of Squares	df	Mean Square	F	p
Control	31951.563	1	31951.563	2.572	0.116
CHO	47325.030	1	47325.030	3.810	0.058
Protein	323700.184	1	323700.184	26.059	< .001

This table shows that there are no gender differences in jump power between the control and CHO groups ($p=.116$ and $p=0.058$, respectively). However, there is a significant difference ($p<.001$) in jump power between genders in the protein supplement group.



CONTRAST TESTING

There are two ways of testing for a difference between (combinations of) cells: post hoc tests and contrast analysis. JASP has a range of different contrast tests available, including custom contrasts. For example, we can contrast the three different supplements. Open up the Contrasts menu, and next to Supplement, click on the drop-down menu and select custom. This will add another series of options to this window.



▼ Contrasts

Factors

- Gender
- Supplement

none ▾
custom ▾

Custom contrast for Supplement

Add Contrast		CHO	Control	Protein
Delete Contrast	Contrast 1	0	0	0
Reset				

In this window, contrasts can be added; in this case, three contrasts can be defined:

Custom for Supplement

Add Contrast	Delete Contrast	Reset		
	Supplement	Contrast 1	Contrast 2	Contrast 3
1	Control	0	-1	1
2	CHO	1	1	0
3	Protein	-1	0	-1

Contrast 1 = CHO vs Protein

Contrast 2 = CHO vs Control

Contrast 3 = Control vs Protein



This will result in the following tables:

Custom Contrast - Supplement

Comparison	Estimate	SE	df	t	p
1	-273.899	47.755	44	-5.736	< .001
2	13.085	48.403	44	0.270	0.788
3	-286.984	46.908	44	-6.118	< .001

Custom Contrast Coefficients - Supplement

Supplement	Comparison 1	Comparison 2	Comparison 3
Control	0	-1	1
CHO	1	1	0
Protein	-1	0	-1

Comparing this table to the post hoc analysis below, the estimates of the differences in marginal means are the same, as well as their standard errors and t-statistics. However, both the p-values and confidence intervals vary: the corrected p-values are typically higher, and the confidence intervals are wider, for the post hoc analysis.

POST HOC TESTING

If the ANOVA is significant, post hoc testing can now be carried out. In Post Hoc Tests, add Supplement and the Gender*Supplement to the analysis box on the right, tick Effect size, and, in this case, use Tukey for the post hoc correction. Also, for ease of viewing, tick Flag significant comparisons.

Post hoc testing is not done for Gender since there are only 2 levels.

Standard

Post Hoc Comparisons - Supplement

		Mean Difference	SE	t	p _{Tukey}
CHO	Control	12.77	40.10	0.32	0.95
	Protein	-279.32	39.56	-7.06	< .001***
Control	Protein	-292.08	38.85	-7.52	< .001***

*** p < .001

Note. P-value adjusted for comparing a family of 3

Note. Results are averaged over the levels of: Gender

Post hoc testing shows no significant difference between the control and CHO, supplement group, irrespective of Gender, but significant differences between Control and Protein ($p < .001$) and between CHO and Protein ($p < .001$).



The post hoc comparisons for the interactions decompose the results further:

Post Hoc Comparisons - Gender * Supplement

		Mean Difference	SE	t	p _{Tukey}
Female, CHO	Male, CHO	-112.59	57.68	-1.95	0.39
	Female, Control	-88.21	57.68	-1.53	0.65
	Male, Control	1.16	57.68	0.02	1.00
	Female, Protein	-197.38	56.17	-3.51	0.01*
	Male, Protein	-473.84	57.68	-8.21	< .001***
Male, CHO	Female, Control	24.38	55.73	0.44	1.00
	Male, Control	113.75	55.73	2.04	0.34
	Female, Protein	-84.79	54.16	-1.57	0.62
	Male, Protein	-361.25	55.73	-6.48	< .001***
Female, Control	Male, Control	89.37	55.73	1.60	0.60
	Female, Protein	-109.17	54.16	-2.02	0.35
	Male, Protein	-385.63	55.73	-6.92	< .001***
Male, Control	Female, Protein	-198.54	54.16	-3.67	8.41e-3**
	Male, Protein	-475.00	55.73	-8.52	< .001***
	Female, Protein	-276.46	54.16	-5.10	< .001***

* p < .05, ** p < .01, *** p < .001

Note. P-value adjusted for comparing a family of 6

REPORTING THE RESULTS

A two-way ANOVA was used to examine the effect of gender and supplement type on explosive jump power. There were significant main effects for both gender ($F(1, 42) = 9.59, p=.003, \omega^2 = 0.058$) and Supplement ($F(2, 42) = 36.1, p<.001, \omega^2 = 0.477$). There was a statistically significant interaction between the effects of gender and supplement on explosive jump power ($F(2, 42) = 11.1, p<.001, \omega^2 = 0.138$).

Tukey's post hoc correction showed that explosive leg power was significantly higher in the protein group compared to the control or CHO groups ($t=-7.52, p<.001$ and $t=-7.06, p<.001$ respectively).

Simple main effects showed that jump power was significantly higher in males on a protein supplement ($M = 1263, SD = 140$) compared to females ($M = 987, SD = 92, F(1) = 26.06, p<.001$).



TWO-WAY REPEATED MEASURES ANOVA

Two-way repeated Measures ANOVA means that there are two factors in the experiment, for example, different treatments and different conditions. "Repeated measures" means that the same subject received more than one treatment and/or more than one condition.

<i>Independent variable (Factor 2)</i>		<i>Independent variable (Factor 1) = time</i>		
	Participant	Time 1	Time 2	Time 3
Condition 1	1	Dependent variable	Dependent variable	Dependent variable
	2	Dependent variable	Dependent variable	Dependent variable
	3	Dependent variable	Dependent variable	Dependent variable
Condition 2	1	Dependent variable	Dependent variable	Dependent variable
	2	Dependent variable	Dependent variable	Dependent variable
	3	Dependent variable	Dependent variable	Dependent variable

The factors are split into levels; therefore, in this case, Factor 1 has 3 repeated levels and Factor 2 has 2 repeated levels.

A "main effect" is the effect of one of the independent variables on the dependent variable, ignoring the effects of any other independent variables. There are 2 main effects tested, both of which are "**between-subjects**": in this case, comparing differences between factor 1 (i.e. condition) and differences between factor 2 (i.e. groups). **Interaction** is where one factor influences the other factor.

The two-way repeated ANOVA is another omnibus test that is used to test the following main effect null hypotheses:

H01: The dependent variable scores are the same for each level in factor 1 (ignoring factor 2).

H02: The dependent variable scores are the same for each level in factor 2 (ignoring factor 1).

The null hypothesis for the interaction between the two factors is:

H03: The two factors are independent, or that an interaction effect is not present.

ASSUMPTIONS

Like all other parametric tests, two-way repeated ANOVA makes a series of assumptions that should either be addressed in the research design or can be tested for.

- The independent variables (factors) should have at least two categorical related groups (levels).



- The dependent variable should be continuous and approximately normally distributed for all combinations of factors.
- Sphericity, i.e. the variances of the differences between all combinations of related groups must be equal.
- There should be no significant outliers.

RUNNING TWO-WAY REPEATED MEASURES ANOVA

Open **2-way repeated ANOVA.csv** in JASP. This comprises 4 columns of data (“sit and reach” flexibility scores for two factors, Factor 1 with 2 levels (stretch and no stretch) and Factor 2 with 2 levels (warm-up and no warm-up). In Descriptive statistics, check the data for significant outliers. Go to ANOVA > Repeated Measures ANOVA. Firstly, each Factor and its levels should be defined. For RM Factor 1 – define this as Stretching and its levels as stretch and no stretch. Then define RM Factor 2 as Warm-up and its levels as warm-up and no warm-up. Then add the appropriate column of data to the assigned repeated measures cells.

The screenshot shows the 'Descriptives Plots' dialog in JASP. Under 'Factors', 'Horizontal Axis' is set to 'Stretching' and 'Separate Lines' is checked. Under 'Display', 'Error bars' is checked and 'Confidence interval' is set to 95.0%.

Also, tick Descriptive statistics and estimates of effect size - ω^2 .

The screenshot shows the 'Display' dialog in JASP. 'Descriptive statistics' and 'Estimates of effect size' are checked. Under 'Estimates of effect size', 'omega squared' is checked, while 'eta squared' and 'general eta squared' are unchecked. 'Vovk-Sellke maximum p-ratio' is also unchecked.



In Descriptive plots, add the Stretching factor to the horizontal axis and the Warm-up factor to separate lines. Tick the display error bars option.

Descriptives Plots

Factors

Horizontal Axis

- ▶ Stretching
- ▶ Separate Lines
- ▶ Warm up
- ▶ Separate Plots

Label y-axis

Display

Display error bars Average across unused RM factors

Confidence interval 95 %

Standard error

UNDERSTANDING THE OUTPUT

The output should comprise 3 tables and one plot. The Between-Subjects Effects table can be ignored in this analysis.

Within Subjects Effects

	Sum of Squares	df	Mean Square	F	p	ω^2
Stretch	246.43	1	246.432	123.40	< .001	0.647
Residual	21.97	11	1.997			
Warm up	85.71	1	85.707	68.69	< .001	0.404
Residual	13.72	11	1.248			
Stretch * Warm up	34.99	1	34.987	29.64	< .001	0.215
Residual	12.98	11	1.180			

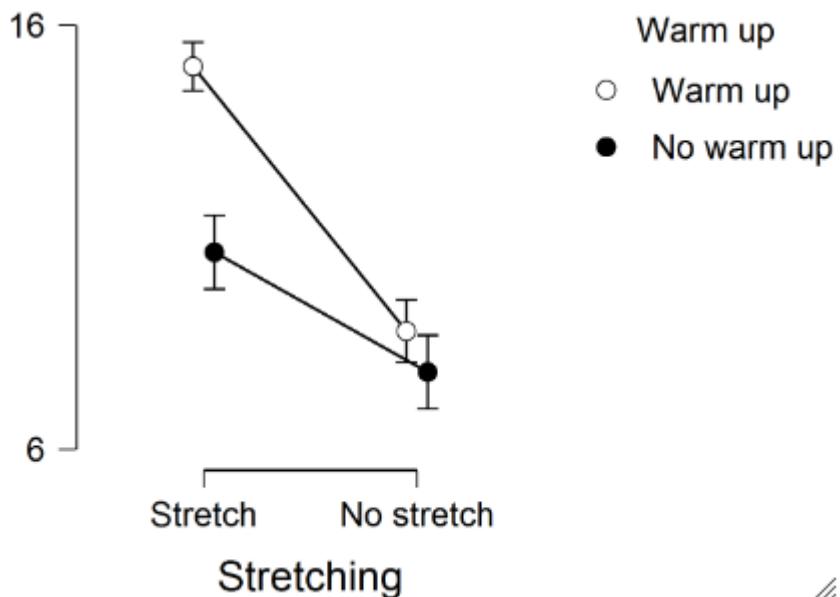
Note. Type III Sum of Squares

The ANOVA within-subjects effects table shows that there are significant main effects for both stretch ($p<.001$) and warm-up ($p<.001$) on sit and reach distance. Both are associated with large effect sizes. There is also a significant interaction between stretch and warm-up ($p<.001$), which suggests that the effects of performing a stretch on sit and reach distance are different depending on whether a warm-up had been performed. These findings can be seen in both the descriptive data and the plot.



Descriptives

Stretching	Warm up	N	Mean	SD	SE	Coefficient of variation
Stretch	Warm up	12	15.029	1.551	0.448	0.103
	No warm up	12	10.649	2.121	0.612	0.199
No stretch	Warm up	12	8.790	1.892	0.546	0.215
	No warm up	12	7.825	1.750	0.505	0.224



TESTING ASSUMPTIONS

In this case, there are no assumption checks. Sphericity can only be tested when there are at least three levels, and homogeneity requires at least two unrelated data sets. If a factor has more than 2 levels, Mauchly's test of Sphericity should also be run, and the appropriate corrected F value used if necessary (See Repeated Measures ANOVA for details).

However, if the ANOVA reports no significant difference, you can go no further with the analysis.



SIMPLE MAIN EFFECTS

Now go to the analysis options and Simple Main Effects. Here, add Warm up to the Simple effect factor and Stretch to the Moderator Factor 1. Simple main effects are effectively pairwise comparisons.

Simple Main Effects

Factors

Simple Effect Factor

Warm up

Moderator Factor 1

Stretching

Moderator Factor 2

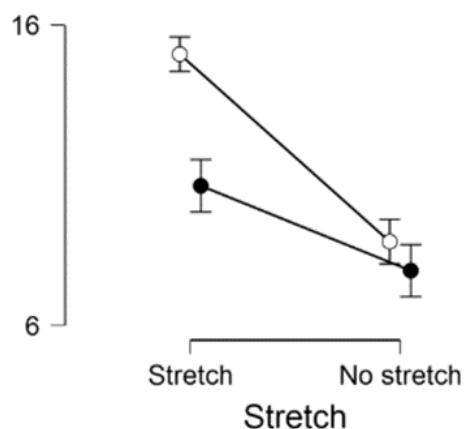
Pool error terms

Simple Main Effects - Warm up

Level of Stretch	Sum of Squares	df	Mean Square	F	p
Stretch	115.106	1	115.106	112.673	< .001
No stretch	5.587	1	5.587	3.973	0.072

Note. Type III Sum of Squares

This table shows that when moderating for warm-up there is a significant difference ($p<.001$) in sit and reach performance when a stretch was also carried out but not without a stretch ($p=0.072$).





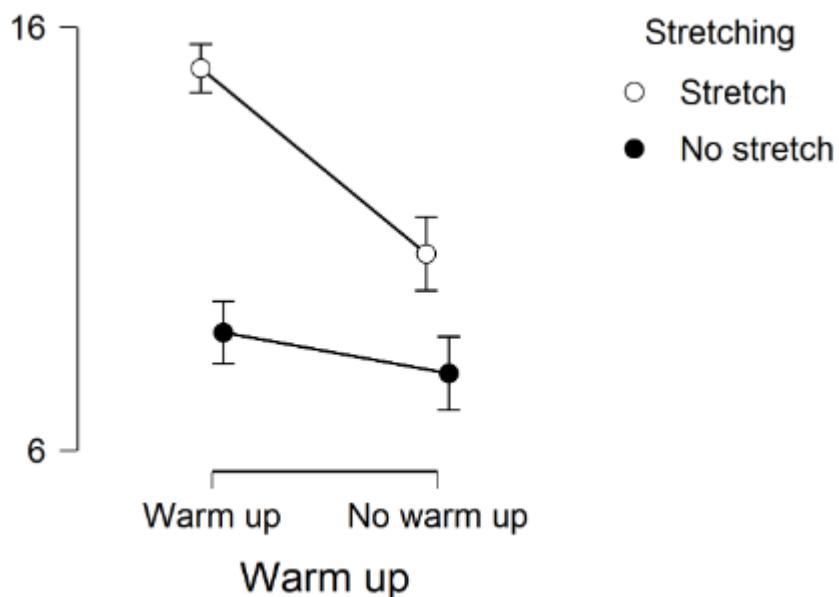
We can now moderate for stretch by changing the Simple Main Effects to use Stretch as the simple effect factor and warm-up as the moderator factor. We can also replot the descriptives with a warm-up on the horizontal axis and stretch as separate lines.

Simple Main Effects - Stretch

Level of Warm up	Sum of Squares	df	Mean Square	F	p
Warm up	233.56	1	233.56	234.18	< .001
No warm up	47.86	1	47.86	21.95	< .001

Note. Type III Sum of Squares

In this case, when controlling for Stretch, there were significant differences between both warm-up and no warm-up.



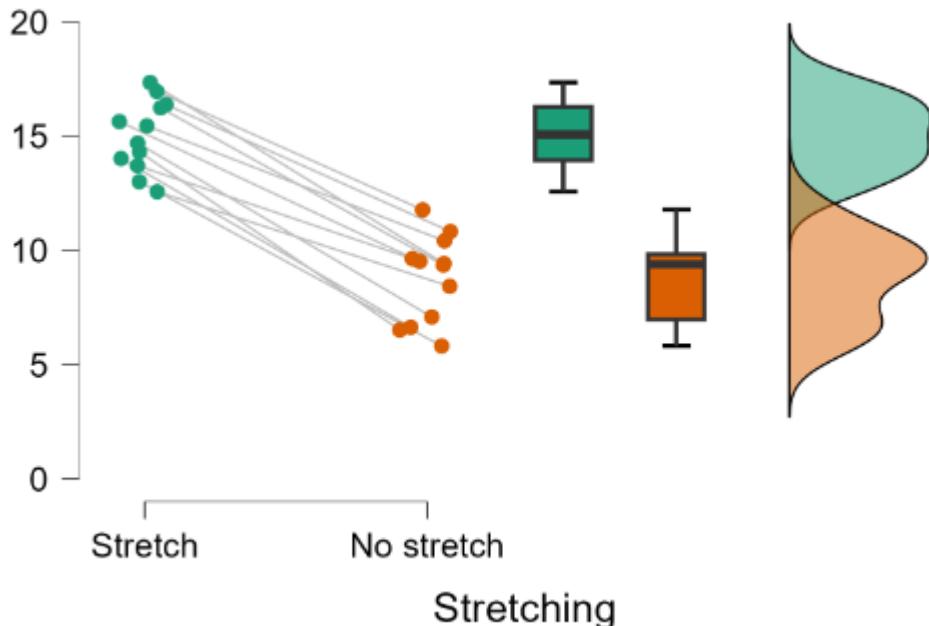
Both simple main effects can be visualised in their descriptive plots.

The data can be further visualised using Raincloud plots.

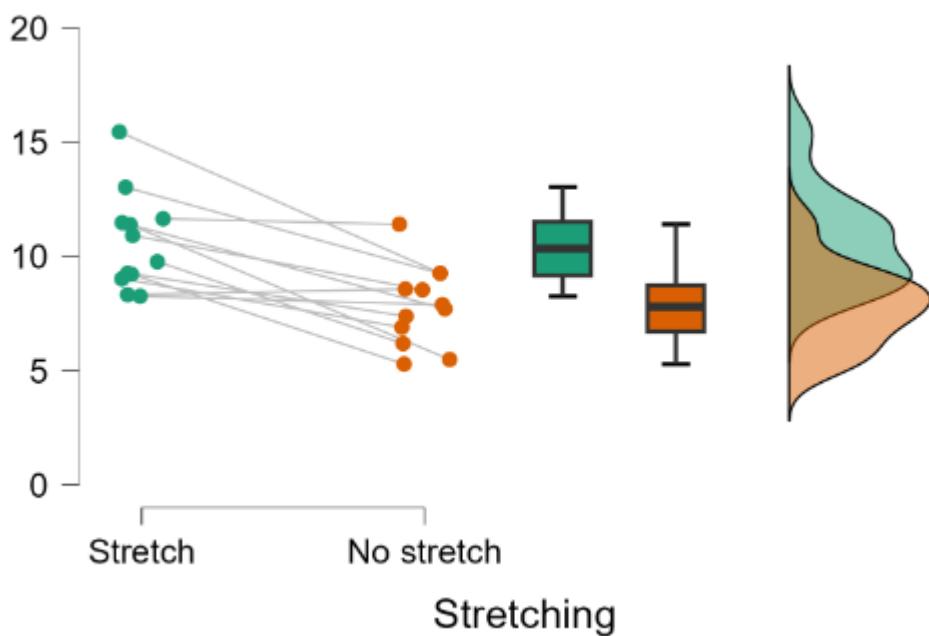
The screenshot shows the JASP software interface with the "Raincloud Plots" section selected. Under "Factors", there is a list box. On the right, under "Horizontal Axis", "Stretching" is selected. Under "Separate Plots", "Warm up" is selected.



Dependent: Warm up: Warm up



Dependent: Warm up: No warm up



POST HOC TESTING

If the ANOVA is significant, post hoc testing can now be carried out. In Post Hoc Tests, add stretch, warm-up and the Stretching*warm-up interaction to the analysis box on the right, tick Effect size, and, in this case, use Holm for the post hoc correction. Tick Flag significant comparisons.



▼ Post Hoc Tests

▶

Stretching
Warm up
Stretching * Warm up

Effect size Pool error term for follow-up tests

Conditional comparisons for interactions

Correction

Holm
 Bonferroni
 Tukey
 Scheffé

Display

Confidence intervals 95.0 %
 Flag significant comparisons
 Letter-based grouping table a-level: 0.05

Post hoc testing for the main effects confirms that there are significant differences in sit and reach distance when comparing the two levels of each factor. This is further decomposed in the Post hoc comparisons for the interaction.

Post Hoc Comparisons - Stretching

		Mean Difference	SE	t	Cohen's d	p _{holm}
Stretch	No stretch	4.532	0.408	11.109	2.463	< .001***

*** p < .001

Note. Computation of Cohen's d based on pooled error.

Note. Results are averaged over the levels of: Warm up

Post Hoc Comparisons - Warm up

		Mean Difference	SE	t	Cohen's d	p _{holm}
Warm up	No warm up	2.672	0.322	8.288	1.452	< .001***

*** p < .001

Note. Computation of Cohen's d based on pooled error.

Note. Results are averaged over the levels of: Stretching



Post Hoc Comparisons - Stretching * Warm up

		Mean Difference	SE	t	p _{holm}
Stretch, Warm.up	No.Stretch, Warm.up	6.239	0.515	12.125	< .001***
	Stretch, No.warm.up	4.380	0.450	9.737	< .001***
	No.Stretch, No.warm.up	7.204	0.520	13.854	< .001***
No.Stretch, Warm.up	Stretch, No.warm.up	-1.859	0.520	-3.575	0.004**
	No.Stretch, No.warm.up	0.965	0.450	2.145	0.043*
Stretch, No.warm.up	No.Stretch, No.warm.up	2.824	0.515	5.488	< .001***

* p < .05, ** p < .01, *** p < .001

Note. P-value adjusted for comparing a family of 6

REPORTING THE RESULTS

A two-way ANOVA was used to examine the effect of stretch and warm-up type on sit and reach performance. There were significant main effects for stretch ($F(1, 11) = 123.4, p < .001, \omega^2 = 0.647$) and warm-up ($F(1, 11) = 68.69, p < .001, \omega^2 = 0.404$). There was a statistically significant interaction between the effects of stretch and warm-up on sit and reach performance ($F(1, 11) = 29.64, p < .001, \omega^2 = 0.215$).

Simple main effects showed that sit and reach performance was significantly higher when both a stretch and warm-up had been done ($F(1) = 234, p < .001$).



MIXED FACTOR ANOVA

Mixed factor ANOVA (another two-way ANOVA) is a combination of both independent and repeated measures ANOVA involving more than 1 independent variable (known as factors).

<i>Independent variable (Factor 2)</i>	<i>Independent variable (Factor 1) = time or condition</i>		
	Time/condition 1	Time/condition 2	Time/condition 3
<i>Group 1</i>	Dependent variable	Dependent variable	Dependent variable
<i>Group 2</i>	Dependent variable	Dependent variable	Dependent variable

The factors are split into levels; therefore, in this case, Factor 1 has 3 levels and Factor 2 has 2 levels. This results in 6 possible combinations.

A “main effect” is the effect of one of the independent variables on the dependent variable, ignoring the effects of any other independent variables. There are 2 main effects tested: in this case, comparing data across factor 1 (i.e. time) is known as the “**within-subjects**” factor, while comparing differences between factor 2 (i.e. groups) is known as the “**between-subjects**” factor. **Interaction** is where one factor influences the other factor.

The main effect of time or condition tests the following, i.e. irrespective of which group is in

<i>Independent variable (Factor 2)</i>	<i>Independent variable (Factor 1) = time or condition</i>		
	Time/condition 1	Time/condition 2	Time/condition 3
<i>Group 1</i>	All data	All data	All data
<i>Group 2</i>			

The main effect of group tests is the following, i.e. irrespective of which condition the data is in:

<i>Independent variable (Factor 2)</i>	<i>Independent variable (Factor 1) = time or condition</i>		
	Time/condition 1	Time/condition 2	Time/condition 3
<i>Group 1</i>	All data		
<i>Group 2</i>	All data		

Simple main effects are effectively pairwise comparisons:

<i>Independent variable (Factor 2)</i>	<i>Independent variable (Factor 1) = time or condition</i>		
	Time/condition 1	Time/condition 2	Time/condition 3
<i>Group 1</i>	Data		Data
<i>Group 2</i>	Data		Data



A mixed factor ANOVA is another omnibus test that is used to test 3 null hypotheses:

3. ***There is no significant within-subject effect, i.e. no significant difference between the means of the differences between all the conditions/times.***
4. ***There is no significant between-subject effect, i.e. no significant difference between the means of the groups.***
5. ***There is no significant interaction effect, i.e. no significant group differences across conditions/time.***

ASSUMPTIONS

Like all other parametric tests, mixed factor ANOVA makes a series of assumptions that should either be addressed in the research design or can be tested for.

- The “**within-subjects**” factor should contain at least two related (repeated measures) categorical groups (levels)
- The “**between-subjects**” factor should have at least two categorical independent groups (levels).
- The dependent variable should be continuous and approximately normally distributed for all combinations of factors.
- There should be homogeneity of variance for each of the groups, and, if more than 2 levels) sphericity between the related groups.
- There should be no significant outliers.

RUNNING THE MIXED FACTOR ANOVA

Open **2-way Mixed ANOVA.csv** in JASP. This contains 4 columns of data relating to the type of weightlifting grip and speed of the lift at 3 different loads (%1RM). Column 1 contains the grip type, and columns 2-4 contain the 3 repeated measures (30, 50 and 70%). Check for significant outliers using boxplots, then go to ANOVA > Repeated measures ANOVA.

Define the Repeated Measures Factor, %1RM, and add 3 levels (30, 50 and 70%). Add the appropriate variable to the Repeated measures Cells and add Grip to the Between-Subjects Factors:



Repeated Measures ANOVA

Repeated Measures Factors

%1RM

30%
50%
70%
Level 4

RM Factor 2

Repeated Measures Cells

RM30	30%
RM50	50%
RM70	70%

Between Subject Factors

Grip

Additionally, tick Descriptive statistics and Estimates of effect size (ω^2).

Display

Descriptive statistics

Estimates of effect size

ω^2 partial ω^2

η^2 partial η^2

general η^2

Vovk-Sellke maximum p-ratio

In Descriptive plots, move %1RM to the horizontal axis and Grip to separate lines. It is now possible to add a title for the vertical axis.

Descriptives Plots

Factors

Horizontal Axis

%1RMMax

Separate Lines

Grip

Separate Plots

Label y-axis: Velocity

Display

Display error bars Average across unused RM factors

Confidence interval 95 %

Standard error



UNDERSTANDING THE OUTPUT

The output should initially comprise 3 tables and 1 graph.

Within Subjects Effects

	Sum of Squares	df	Mean Square	F	p	ω^2
%1RM	5.605*	2*	2.803*	115.450*	< .001*	0.744
%1RM * Grip	0.583*	2*	0.291*	12.003*	< .001*	0.218
Residual	0.874	36	0.024			

Note. Type III Sum of Squares

* Mauchly's test of sphericity indicates that the assumption of sphericity is violated ($p < .05$).

For the main effect for %1RM, the within-subjects effects table reports a large F-statistic, which is highly significant ($p < .001$) and has a large effect size (0.744). Therefore, irrespective of grip type, there is a significant difference between the three %1RM loads.

However, JASP has reported under the table that the assumption of sphericity has been violated. This will be addressed in the next section.

Finally, there is a significant interaction between %1RM and grip ($p < .001$), which also has a large effect size (0.499). This suggests that the differences between the %1RM loads are affected somehow by the type of grip used.

Between Subjects Effects

Cases	Sum of Squares	df	Mean Square	F	p	ω^2
Grip	1.095	1	1.095	20.925	< .001	0.344
Residuals	0.942	18	0.052			

Note. Type III Sum of Squares

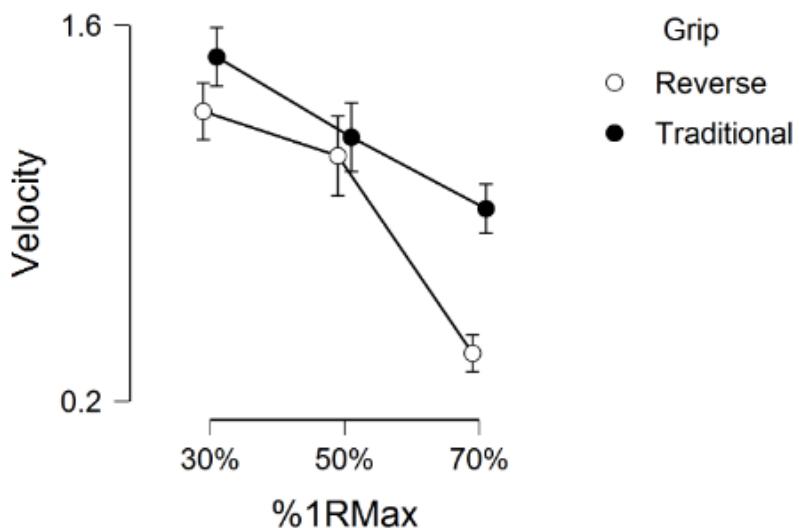
For the main effect of grip, the between-subjects table shows a significant difference between grips ($p < .001$), irrespective of %1RM.

From the descriptive data and the plot, it appears that there is a larger difference between the two grips at the high 70% RM load.



Descriptives

%1RM	Grip	N	Mean	SD	SE	Coefficient of variation
30%	Reverse	10	1.279	0.178	0.056	0.139
	Traditional	10	1.482	0.217	0.069	0.146
50%	Reverse	10	1.114	0.198	0.063	0.178
	Traditional	10	1.183	0.256	0.081	0.216
70%	Reverse	10	0.379	0.105	0.033	0.278
	Traditional	10	0.917	0.086	0.027	0.093



TESTING ASSUMPTIONS

In Assumptions Checks, tick Sphericity tests, Sphericity corrections and Homogeneity tests.

▼ Assumption Checks

Sphericity tests
 Sphericity corrections
 None Greenhouse-Geisser Huynh-Feldt
 Homogeneity tests

Test of Sphericity

Mauchly's W	Approx. χ^2	df	p-value	Greenhouse-Geisser ϵ	Huynh-Feldt ϵ	Lower Bound ϵ
0.649	7.360	2	0.025	0.740	0.791	0.500

Mauchly's test of sphericity is significant, so that assumption has been violated; therefore, the Greenhouse-Geisser correction should be used since epsilon is <0.75. Go back to Assumption Checks and in Sphericity corrections, leave Greenhouse-Geisser only ticked. This will result in an updated Within-Subjects Effects table:



Within Subjects Effects

Cases	Sphericity Correction	Sum of Squares	df	Mean Square	F	p	ω^2
%1RM	Greenhouse-Geisser	5.605	1.480	3.787	115.450	<.001	0.744
%1RM * Grip	Greenhouse-Geisser	0.583	1.480	0.394	12.003	<.001	0.218
Residuals	Greenhouse-Geisser	0.874	26.639	0.033			

Note. Type III Sum of Squares

* Mauchly's test of sphericity indicates that the assumption of sphericity is violated ($p < .05$).

Levene's test shows that there is no difference in variance in the dependent variable between the two grip types.

Test for Equality of Variances (Levene's)

	F	df1	df2	p
RM30	0.523	1.000	18.000	0.479
RM50	0.346	1.000	18.000	0.564
RM70	0.183	1.000	18.000	0.674

However, if the ANOVA reports no significant difference, you can go no further in the analysis.

POST HOC TESTING

If the ANOVA is significant, post hoc testing can now be carried out. In Post Hoc Tests, add %1RM to the analysis box on the right, tick Effect size, and, in this case, use Holm for the post hoc correction. Only Bonferroni or Holm's corrections are available for repeated measures.

The screenshot shows the 'Post Hoc Tests' dialog in JASP. The 'Analysis' dropdown is set to '%1RM'. The 'Effect size' and 'Pool error term for RM factors' checkboxes are checked. Under 'Correction', 'Holm' is selected. The 'Display' section has 'Flag Significant Comparisons' checked.



Post Hoc Tests

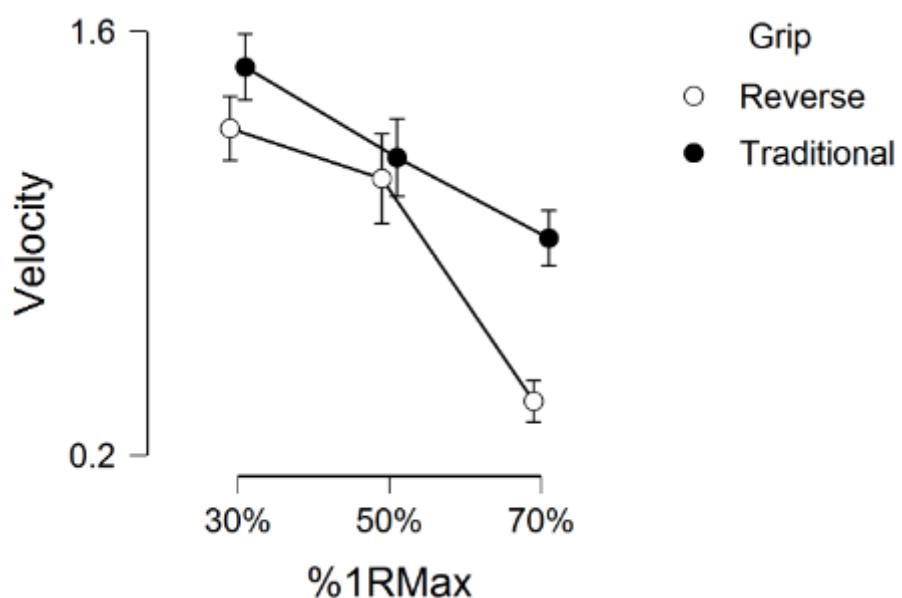
Post Hoc Comparisons - %1RM

		Mean Difference	SE	df	t	Cohen's d	P _{holm}
30%	50%	0.232	0.060	18	3.881	1.266	0.001
	70%	0.733	0.033		22.040	3.995	< .001
50%	70%	0.500	0.051	18	9.814	2.728	< .001

Note. P-value adjusted for comparing a family of 3 estimates.

Note. Results are averaged over the levels of: Grip

The post hoc tests show that, irrespective of grip type, each load is significantly different from each of the other loads, and as seen from the plot, lift velocity significantly decreases as the load increases.



Finally, in Simple main effects, add Grip to the Simple effect factor and %1RM to the Moderator factor 1.

▼ Simple Main Effects

Factors	Simple Effect Factor
<input type="checkbox"/> Grip	Grip
<input type="checkbox"/> %1RMax	Moderator Factor 1
<input type="checkbox"/>	Moderator Factor 2
<input type="checkbox"/> Pool error terms	



Simple Main Effects - Grip

Level of %1RM	Sum of Squares	df	Mean Square	F	p
30%	0.206	1	0.206	5.229	0.035
50%	0.024	1	0.024	0.461	0.506
70%	1.447	1	1.447	157.212	< .001

These results show that there is a significant difference in lift speed between the two grips at 30% 1RM and the higher 70% 1RM loads ($p=0.035$ and $p<0.001$, respectively).

REPORTING THE RESULTS

Using the Greenhouse-Geisser correction, there was a significant main effect of load: $F(1.48, 26.64) = 115.45, p<.001$. Bonferroni corrected post hoc testing showed that there was a significant sequential decline in lift speed from 30-50% 1RM ($p=.035$) and 50-70% 1RM ($p<.001$).

There was a significant main effect for grip type: $F(1, 18) = 20.925, p<.001$, showing an overall higher lift speed using the traditional rather than the reverse grip.

Using the Greenhouse-Geisser correction, there was a significant %1RM x Grip interaction: $F(1.48, 26.64) = 12.00, p<.001$ showing that the type of grip affected lift velocity over the %1RM loads.



CHI-SQUARE TEST FOR ASSOCIATION

The chi-square (χ^2) test for independence (also known as Pearson's χ^2 test or the χ^2 test of association) can be used to determine if a relationship exists between two or more categorical variables. The test produces a contingency table, or cross-tabulation, which displays the cross-grouping of the categorical variables.

The χ^2 test checks the null hypothesis that there is no association between two categorical variables. It compares the observed frequencies of the data with frequencies that would be expected if there were no association between the two variables.

The analysis requires two assumptions to be met:

1. The two variables must be categorical data (nominal or ordinal)
2. Each variable should comprise two or more independent categorical groups

Most statistical tests fit a model to the observed data with a null hypothesis that there is no difference between the observed and modelled (expected) data. The error or deviation of the model is calculated as:

$$\text{Deviation} = \sum (\text{observed} - \text{model})^2$$

Most parametric models are based on population means and standard deviations. The χ^2 model, however, is based on expected frequencies.

How are the expected frequencies calculated? For example, we categorised 100 people into male, female, short and tall. If there was an equal distribution between the 4 categories expected frequency = 100/4 or 25% but the actual observed data does not have an equal frequency distribution.

Equal Distribution	Male	Female	Row Total
Tall	25	25	50
Short	25	25	50
Column Total	50	50	

Observed Distribution	Male	Female	Row Total
Tall	57	24	81
Short	14	5	19
Column Total	71	29	

The model based on expected values can be calculated by:

Model (expected)	= (row total x column total)/100
Model – tall male	= (81 x 71) /100 = 57.5
Model – tall female	= (81 x 29) /100 = 23.5
Model – small male	= (19 x 71) /100 = 13.5
Model – small female	= (19 x 29) /100 = 5.5

These values can then be added to the contingency table:



	Male (M)	Female (F)	Row Total
Tall (T)	57	24	81
Expected	57.5	23.5	
Short (S)	14	5	19
Expected	13.5	5.5	
Column Total	71	29	

The χ^2 statistic is derived from $\sum \frac{(\text{observed} - \text{expected})^2}{\text{expected}}$

Validity

χ^2 tests are only valid when you have a reasonable sample size, that is, less than 20% of cells have an expected count of less than 5, and none have an expected count of less than 1.

RUNNING THE ANALYSIS

The dataset **Titanic survival** is a classic dataset used for machine learning and contains data on 1309 passengers and crew who were on board the Titanic when it sank in 1912. We can use this to look at associations between survival and other factors. The dependent variable is 'Survival', and possible independent variables are all the other values.

Class	survived	name	sex	age
Third	No	Abbing, Mr. Anthony	male	42
Third	No	Abbott, Master. Eugene Joseph	male	13
Third	No	Abbott, Mr. Rossmore Edward	male	16
Third	Yes	Abbott, Mrs. Stanton (Rosa Hunt)	female	35
Third	Yes	Abelseth, Miss. Karen Marie	female	16
Third	Yes	Abelseth, Mr. Olaus Jorgensen	male	25
Second	No	Abelson, Mr. Samuel	male	30
Second	Yes	Abelson, Mrs. Samuel (Hannah Wizosky)	female	28
Third	Yes	Abrahamsson, Mr. Abraham August Johannes	male	20
Third	Yes	Abrahim, Mrs. Joseph (Sophie Halaut Easu)	female	18
Third	No	Adahl, Mr. Mauritz Nils Martin	male	30
Third	No	Adams, Mr. John	male	26



By convention, the independent variable is usually placed in the contingency table columns, and the dependent variable is placed in the rows.

Open **Titanic survival chi square.csv** in JASP, in the spreadsheet tab and double click on Survived. This will open up the Label editor, where you can add labels to the code. Survival has been coded as no = 0 and yes = 1.

The screenshot shows the 'Label editor' tab selected in the JASP interface. The 'survived' column is being edited. The 'Value' column contains two entries: 0 (labeled 'No') and 1 (labeled 'Yes').

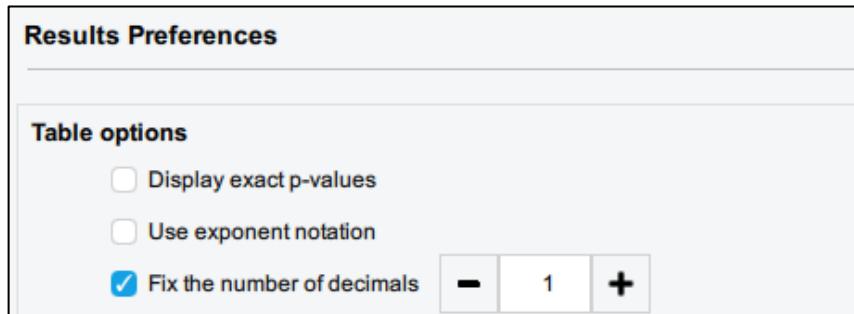
Filter	Value	Label
✓	0	No
✓	1	Yes

Go to Frequencies > Contingency tables and add survived to rows as the dependent variable and sex into columns as the independent variable.

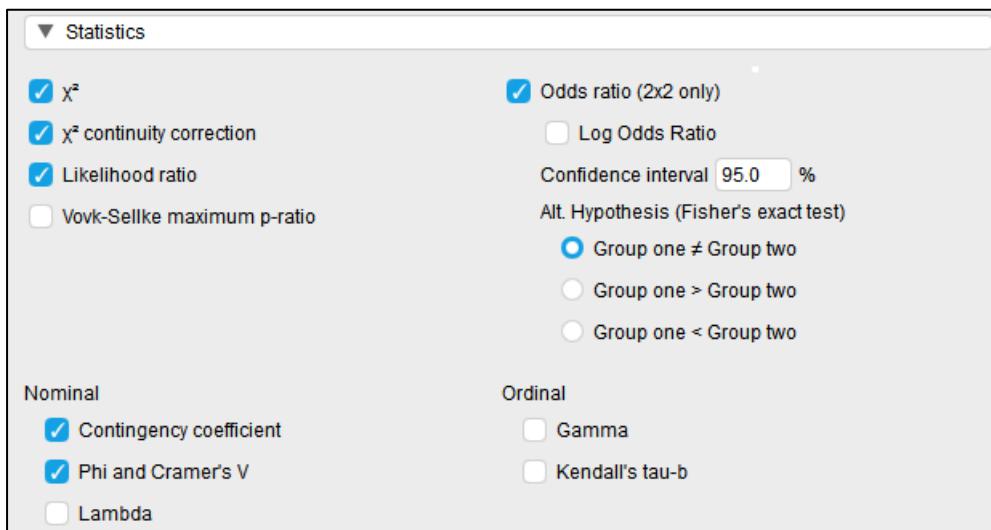
The 'Contingency Tables' dialog is shown. The 'Rows' section contains 'survived' (selected). The 'Columns' section contains 'sex'. The 'Independent variables' list on the left includes 'age' (selected), 'spouses on board', 'parents with children', and 'embarked'.



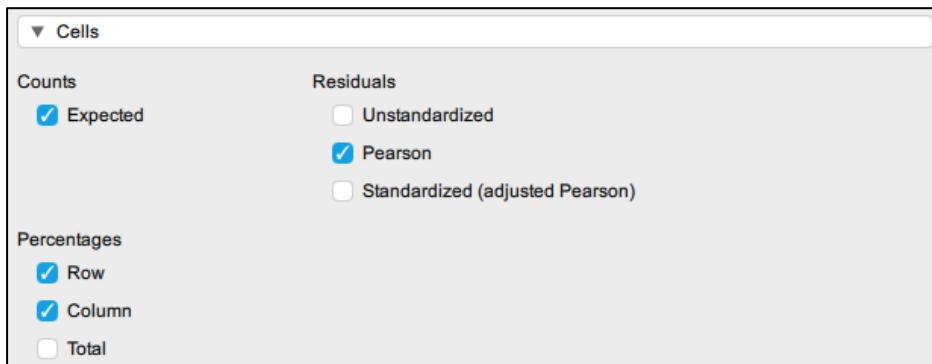
To make the tables easier to read, in Preferences, I suggest that you reduce the number of decimals to 1.



In statistics, tick all the following options:



In Cells, tick the following:





UNDERSTANDING THE OUTPUT

First, look at the Contingency table output.

Contingency Tables

		sex		Total
survived		female	male	
No	Count	127.0	682.0	809.0
	Expected count	288.0	521.0	809.0
	% within row	15.7 %	84.3 %	100.0 %
	% within column	27.3 %	80.9 %	61.8 %
	Pearson residuals	-9.5	7.1	
Yes	Count	339.0	161.0	500.0
	Expected count	178.0	322.0	500.0
	% within row	67.8 %	32.2 %	100.0 %
	% within column	72.7 %	19.1 %	38.2 %
	Pearson residuals	12.1	-9.0	
Total	Count	466.0	843.0	1309.0
	Expected count	466.0	843.0	1309.0
	% within row	35.6 %	64.4 %	100.0 %
	% within column	100.0 %	100.0 %	100.0 %

Remember that χ^2 tests are only valid when you have a reasonable sample size, i.e. less than 20% of cells have an expected count of less than 5, and none have an expected count of less than 1.

From this table, looking at % within rows, more males died on the Titanic compared to females, and more females survived compared to males. But is there a significant association between gender and survival?

The statistical results are shown below:

Chi-Squared Tests

	Value	df	p
χ^2	365.9	1	< .001
χ^2 continuity correction	363.6	1	< .001
Likelihood ratio	372.9	1	< .001
N	1309		

The χ^2 statistic ($\chi^2(1) = 365.9, p < .001$) suggests that there is a significant association between gender and survival.

χ^2 continuity correction can be used to prevent the overestimation of statistical significance for small datasets. This is mainly used when at least one cell of the table has an expected count smaller than 5.



As a note of caution, this correction may overcorrect and result in an overly conservative result that fails to reject the null hypothesis when it should (a type II error).

The likelihood ratio is an alternative to the Pearson chi-square. It is based on the maximum-likelihood theory. For large samples, it is identical to Pearson χ^2 . It is recommended for small sample sizes, i.e. <30.

Nominal measures, Phi (2 x 2 contingency tables only) and Cramer's V (most popular) are both tests of the strength of association (i.e. effect sizes). Both values are in the range of 0 (no association) to 1 (complete association). The strength of association between the variables has a large effect size.

Nominal

	Value
Contingency coefficient	0.47
Phi-coefficient	0.53
Cramer's V	0.53

The Contingency coefficient is an adjusted Phi value and is only suggested for large contingency tables, such as 5 x 5 tables or larger.

Effect size ⁴	df	Small	Moderate	Large
Phi and Cramer's V (2x2 only)	1	0.1	0.3	0.5
Cramer's V	2	0.07	0.21	0.35
Cramer's V	3	0.06	0.17	0.29
Cramer's V	4	0.05	0.15	0.25
Cramer's V	5	0.04	0.13	0.22

JASP also provides the Odds ratio (OR), which is used to compare the relative odds of the occurrence of the outcome of interest (survival), given exposure to the variable of interest (in this case, gender).

In Preferences, I suggest that you return the number of decimals to 3.

Odds Ratio

Odds Ratio	95% Confidence Intervals			p
	Lower	Upper		
Odds ratio	0.088	0.068	0.115	
Fisher's exact test	0.089	0.067	0.116	< .001

For ease of understanding, take the reciprocal of the OR (1/0.088), i.e. 11.36. This suggests that male passengers had 11.36 times more chance of dying than females.

⁴ Kim HY. Statistical notes for clinical researchers: Chi-squared test and Fisher's exact test. Restor. Dent. Endod. 2017; 42(2):152-155.



How is this calculated? Use the counts from the contingency table in the following:

$$\begin{array}{llll} \text{Odds[males]} & = \text{Died}/\text{Survived} & = 682/162 & = 4.209 \\ \text{Odds[females]} & = \text{Died}/\text{Survived} & = 127/339 & = 0.374 \end{array}$$

$$\text{OR} = \text{Odds[males]} / \text{Odds [females]} = 11.3$$

GOING ONE STEP FURTHER.

We can also further decompose the contingency table as a form of post hoc testing by converting the counts and expected counts in each cell to a standardised residual. This can tell us if the observed counts and expected counts are significantly different in each cell.

The Pearson's residual for a cell in a table is a version of the standard z-score, calculated as

$$z = \frac{\text{observed} - \text{expected}}{\sqrt{\text{expected}}}$$

The resulting value of z is then given a positive sign if $\text{observed} > \text{expected}$ and a negative sign if $\text{observed} < \text{expected}$. Z-score significances are shown below.

z-score	P-value
<-1.96 or > 1.96	<0.05
<-2.58 or > 2.58	<0.01
<-3.29 or > 3.29	<0.001

Contingency Tables

		sex		
		survived	female	male
		No	Count	809.0
			Expected count	288.0
			% within row	15.7 %
			% within column	27.3 %
			Pearson residuals	-9.5
		Yes	Count	500.0
			Expected count	178.0
			% within row	67.8 %
			% within column	72.7 %
			Pearson residuals	12.1
		Total	Count	1309.0
			Expected count	466.0
			% within row	35.6 %
			% within column	100.0 %
			Pearson residuals	-9.0



When Pearson's residual z-scores are calculated for each cell in the contingency table, we can see that the z-scores are significant in all cells.

UNDERSTANDING THE OUTPUT

There is a significant association between gender and survival ($\chi^2(1) = 365.9, p < .001$). The odds ratio suggests that male passengers had 11.36 times more chance of dying than females.

Pearson's residuals suggest that significantly fewer women died than expected and significantly more males died than expected.



META-ANALYSIS IN JASP

Background

A meta-analysis is a statistical analysis that integrates results from multiple studies, providing a single numerical value of the overall treatment effect for that group of studies. The difference between statistical data analysis and meta-analysis is shown below. Effectively, each study becomes a “participant” in the meta-analysis.

Statistical analysis		Meta-analysis	
Participant 1	Individual data	Study 1	Study data
Participant 2	Individual data	Study 2	Study data
Participant 3	Individual data	Study 3	Study data
Participant 4	Individual data	Study 4	Study data
Participant 5	Individual data	Study 5	Study data
Overall group data & statistics		Overall study group data & statistics	

In JASP, click on the blue cross in the top right corner, and then tick the Meta-Analysis module. This will add meta-analysis to the main ribbon.

Effect size and calculations

To perform a Meta-analysis in JASP, the overall effect size (ES) and standard error (SE) of the study need to be provided. An ES is a dimensionless estimate (no units) that indicates both the direction and magnitude of the effect/outcome. The standard error measures the dispersion of different sample means taken from the same population and can be estimated from a single sample standard deviation. Some studies will provide this information, although many do not. However, they will provide results such as:

- Central tendencies and dispersion
- T or F statistics
- P-values
- Correlation coefficients
- Chi-square

All of these can be converted to an ES and the SE determined using JASP's Effect size computation, which is described later.

For example, a study comparing a treatment to a control group may provide the variable mean, SD and n of each group. From this, the ES, a standardised mean difference (d), and estimated standard error can be calculated.

To interpret the meta-analysis output, one needs to understand the following concepts: heterogeneity, the model, effect size, and the forest plot.

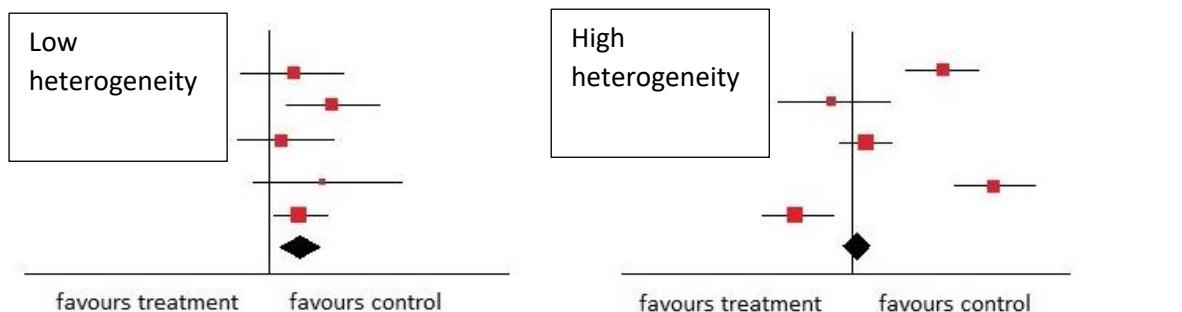
Heterogeneity:

Heterogeneity describes any variability that may exist between the different studies. It is the opposite of homogeneity, which means that the population/data/results are the same. There are 3 types:



- Clinical: Differences in participants, interventions, or outcomes
- Methodological: Differences in study design, risk of bias
- Statistical: Variation in intervention effects or results

If there is no variability, then the data can be described as homogeneous. Meta-analysis is concerned with statistical heterogeneity. Statistical heterogeneity is used to describe variability among data/studies and occurs when the treatment effect estimates of a set of data/studies vary among one another. Studies with methodological flaws and small studies may overestimate treatment effects and can contribute to statistical heterogeneity.



Meta-analysis provides quantitative statistical methods to measure heterogeneity.

Tests for heterogeneity

Q Cochran Q test - values are given for 2 tests outlined below, (these tests have low statistical power when only a few studies are included in the meta-analysis):

'Test for residual heterogeneity' tests the null hypothesis that all the effect sizes in the studies are equal (homogeneity of effect sizes).

τ^2 Tau square is an estimate of the total amount of heterogeneity. This is interpreted as systematic, unexplained differences between the observed effects of the separate studies. It is not affected by the number of studies, but it is often hard to interpret how relevant the value is from a practical standpoint.

I^2 This measures the extent of heterogeneity not caused by sampling error. If there is high heterogeneity, a possible subgroup analysis could be done. If the value is very low, there is no point in doing further subgroup analyses. I^2 is not sensitive to changes in the number of studies and is therefore used extensively in medical and psychological research, especially since there is a "**rule of thumb**" to interpret it. A rough guide for interpretations has been suggested as follows⁶:

- 0% to 40%: might not be important.
- 30% to 60%: may represent moderate heterogeneity.
- 50% to 90%: may represent substantial heterogeneity.
- 75% to 100%: considerable heterogeneity.

⁵ <https://s4be.cochrane.org/blog/2018/11/29/what-is-heterogeneity/>

⁶ Cochrane Handbook for Systematic Reviews of Interventions



- H^2 The between-study variance is determined by equating the Q statistic to its expected value. H^2 has a value of 1 in the case of homogeneity, and heterogeneity is assumed to be present when $H^2 > 1$.

Meta-analysis models

There are two models commonly used in meta-analysis, and each makes different assumptions relating to the observed differences among the studies.

Fixed effects model: this assumes that all studies share a common true ES, i.e. the data is homogeneous. All factors that could influence the ES are the same in all the study samples, and therefore very little heterogeneity. Between-study differences are assumed to be due to chance and thus not incorporated into the model. Therefore, each study included in the meta-analysis estimates the same population treatment effect, which, in theory, represents the true population treatment effect. Each study is weighted where more weight is given to studies with large sample sizes, i.e. more information.

This model answers the following question: “**What is the best estimate of the population effect size?**”

Random effects model: This assumes a distribution of the treatment effect for some populations. i.e. the different studies are estimating different, yet related, intervention effects. Therefore, heterogeneity cannot be explained because it is due to chance. This model assigns a more balanced weighting between studies.

This model answers the question “**What is the average treatment effect?**”.

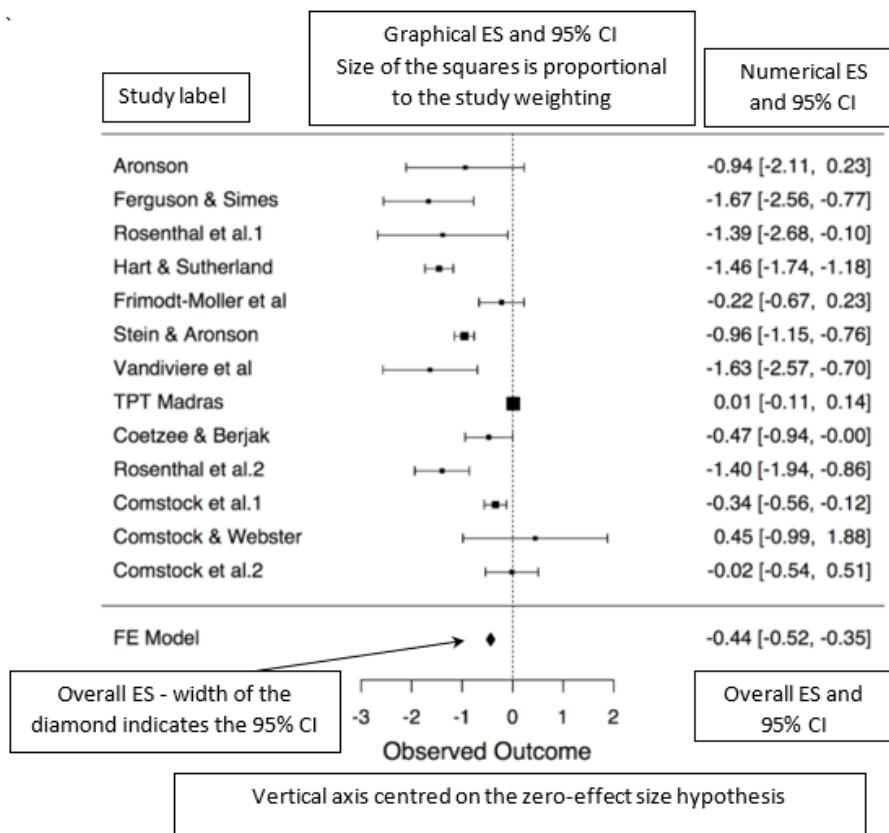
It is therefore important to check for significant heterogeneity to select the correct model for the meta-analysis.

Random effects model selection.

JASP provides 8 random effects estimators to estimate the 4 indices of heterogeneity (see tests for heterogeneity above). All use a slightly different approach, resulting in different pooled ES estimates and CIs. To date, the most often used estimator in medical research is the DerSimonian-Laird estimator; however, recent studies have shown better estimates of between-study variance using Maximum-Likelihood, Restricted ML (**Default in JASP**), Empirical Bayes and Sidak-Jonkman methods.

The Forest Plot.

This plot provides a visual summary of the meta-analysis findings. Graphically, it represents the ES and 95% CI for each study and an estimate of the overall ES and 95% CI for all the studies that were included. As previously mentioned, it can be used to visually assess the level of heterogeneity.



As a rule of thumb, if all the individual study CIs cover the final combined ES and its 95% CI (diamond), then there is low heterogeneity (or high homogeneity). In this case, 6 studies do not intersect with the diamond.

Funnel plot

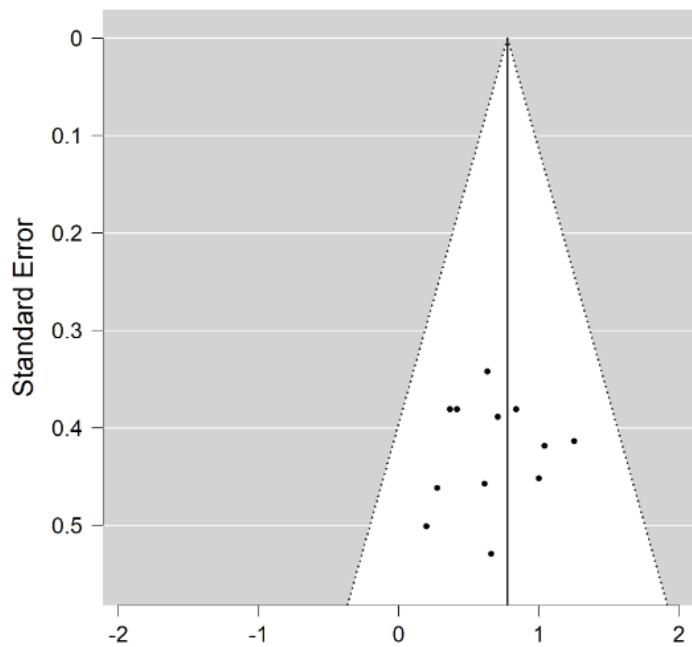
Looking like an inverted funnel, this is a scatter plot of the intervention effect estimates from individual studies against each study's size or precision. The intervention effect should increase as the size of the study increases. Studies with low power will cluster at the bottom, while higher power studies will be near the top. Ideally, the data points should have a symmetrical distribution. Funnel plot asymmetry indicates that the meta-analysis results may be biased. Potential reasons for bias include:

- Publication bias (the tendency of authors/publishers to only publish studies with significant results)
- True heterogeneity
- Artefact (wrong choice of effect measure)
- Data irregularities (methodological design, data analysis, etc.)

Funnel plot asymmetry can be analysed statistically using either meta-regression, weighted regression or rank correlation.



Funnel Plot





EFFECT SIZE COMPUTATION IN JASP

Before running a meta-analysis, you will have to calculate the effect sizes and standard errors based on the data extracted from the studies.

Open the Meta-Analysis fatigue.csv dataset. This is from papers comparing the effects of a new treatment on the force recovery from eccentric exercise. The data comprises of the means, SD and N for control and treatment groups as well as the authors and publication date.

T	Author	date	Control N	Control mean	Control SD	Treatment N	Treatment mean	Treatment SD
1	Bae	2013	88	3.1	10.42	193	8.1	10.42
2	Choy	2012	102	3.74	17.833	119	14.72	19.165
3	Emery	2008	263	16.7	21.9	265	21.4	22.9
4	Horslev-Petersen	2014	91	23.3	31.3	89	33.1	32.9
5	Keystone	2004	96	7.87	21.3	206	15.1	21.94

Click on the Meta-Analysis icon (this module may need to be added by clicking the blue plus sign). Then click on Effect size computation.

This allows you to calculate effect sizes and standard errors from a variety of research designs and

The screenshot shows the 'Effect size' tab in JASP. Under 'Step 1', the 'Design' is set to 'Independent groups', 'Measurement' is 'Quantitative', and 'Effect size' is 'SMD'. A green '+' button is visible at the bottom right of the panel.

measurements. In this case, we can use the default settings. Next, populate the appropriate cells:

The screenshot shows the 'Step 1: SMD (Independent groups/Quantitative)' panel. On the left, under 'Available variables', 'Treatment N' is selected. On the right, under 'Mean Group 1', 'Control mean' and 'Treatment mean' are listed. Similar panels for 'Mean Group 2', 'SD Group 1', 'SD Group 2', 'Sample Size Group 1', and 'Sample Size Group 2' are also shown.

You will see in the results that JASP has computed 21 effect sizes.



If you return to the spreadsheet viewer, you will see that 3 extra columns have been added:

- Computed effect size
- Computed standard error
- Computed effect size type – in this case, the standardised mean difference (SMD)

These are used to run the meta-analysis.

RUNNING THE META-ANALYSIS IN JASP

Go to Meta-Analysis and click on Classical Meta-Analysis. Add the computed effect size, computed standard error and finally the Author to the Study Labels. Keep the method as Restricted ML.

The screenshot shows the JASP Classical Meta-Analysis dialog box. On the left, a list of variables is displayed: date, Control N, Control mean, Control SD, Treatment N, Treatment mean, Treatment SD, and computed effect size type. To the right, various settings are configured:

- Effect Size:** Includes 'computed effect size' and 'computed standard error'.
- Method:** Set to 'Restricted ML'.
- Fixed effect test:** Set to 'knha'.
- Predictors:** An empty list.
- Clustering:** An empty list.
- Study Labels:** Set to 'Author'.

In the Statistics tab, tick all 4 Heterogeneity measures, T , T^2 , I^2 , H^2 .

This will result in 3 tables.



Model Summary

Residual Heterogeneity Test

Q _e	df	p
49.439	20	< .001

Pooled Effect Size Test

Estimate	Standard Error	t	df	p
-0.352	0.045	-7.900	20.000	< .001

Meta-Analytic Estimates

	Estimate	95% CI		95% PI	
		Lower	Upper	Lower	Upper
Effect Size	-0.352	-0.445	-0.259	-0.668	-0.036
τ	0.145	0.081	0.279		
τ^2	0.021	0.007	0.078		
I^2	59.313	31.196	84.394		
H^2	2.458	1.453	6.408		

The Residual Heterogeneity test confirms significant heterogeneity in the data and that the random-effects model (Restricted ML) is appropriate in this case. If this were not significant, the method should be changed to fixed effects.

The Pooled Effect Size tests the H_0 that all the estimates are zero, i.e. the interventions have no significant effect. As can be seen, this is significant and H_0 can be rejected, thus the intervention has a significant effect with an effect size difference of -0.352.

Looking at the Meta-Analytic Estimates, both H^2 and I^2 show excess variance (heterogeneity) between the studies, thus supporting the use of a random-effects model.

Forest Plots

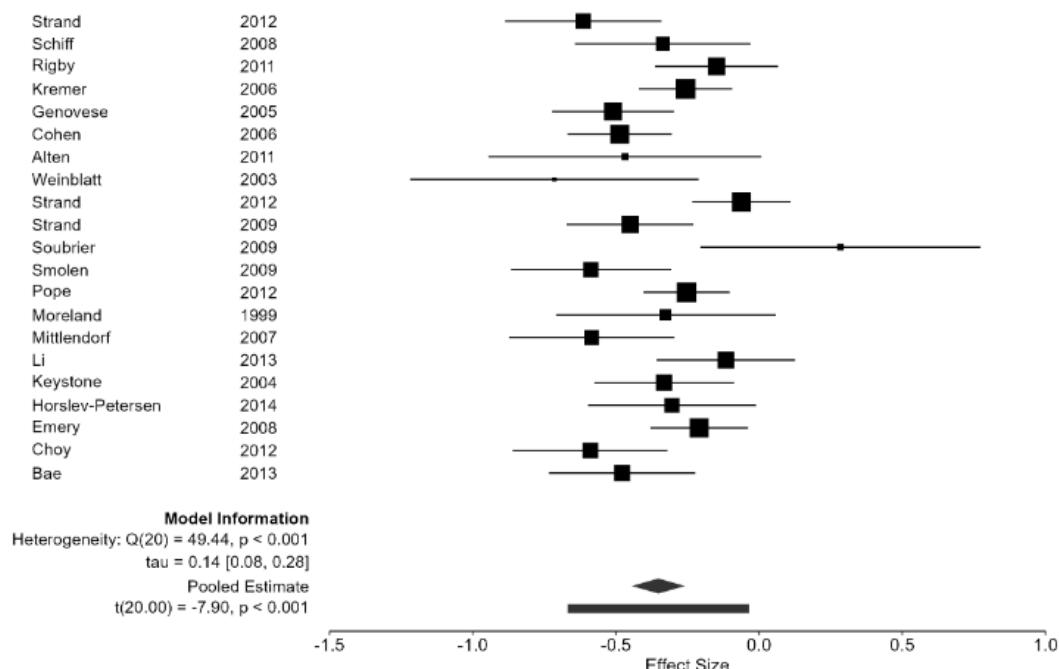
Return to the Statistics options and Forest Plot, tick Study information, and add both Author and Date to the selected variables. Scroll down and tick Model information.

The screenshot shows the JASP software interface for creating a forest plot. The top navigation bar has 'File', 'Edit', 'Statistics', 'Plots', 'Reports', 'Help' menu items, and a search bar. Below the menu is a toolbar with icons for 'Forest Plot', 'Divergence Plot', 'Forest Plot (ML)', 'Divergence Plot (ML)', 'Forest Plot (REML)', 'Divergence Plot (REML)', 'Forest Plot (ML, REML)', 'Divergence Plot (ML, REML)', 'Forest Plot (ML, REML, QI)', and 'Divergence Plot (ML, REML, QI)'. The main area has tabs for 'Forest Plot' (selected), 'Divergence Plot', 'Forest Plot (ML)', 'Divergence Plot (ML)', 'Forest Plot (REML)', 'Divergence Plot (REML)', 'Forest Plot (ML, REML)', 'Divergence Plot (ML, REML)', 'Forest Plot (ML, REML, QI)', and 'Divergence Plot (ML, REML, QI)'. Under 'Forest Plot' settings, 'Study information' is checked. In the 'Selected Variables' panel, 'Author' and 'date' are selected. The bottom status bar shows 'JASP 0.19.3 - Professor Mark Goss-Sampson'.



The Forest plot shows the weighted effect sizes (the size of the squares reflects the weight of each study) and CIs used to determine the overall pooled ES (diamond).

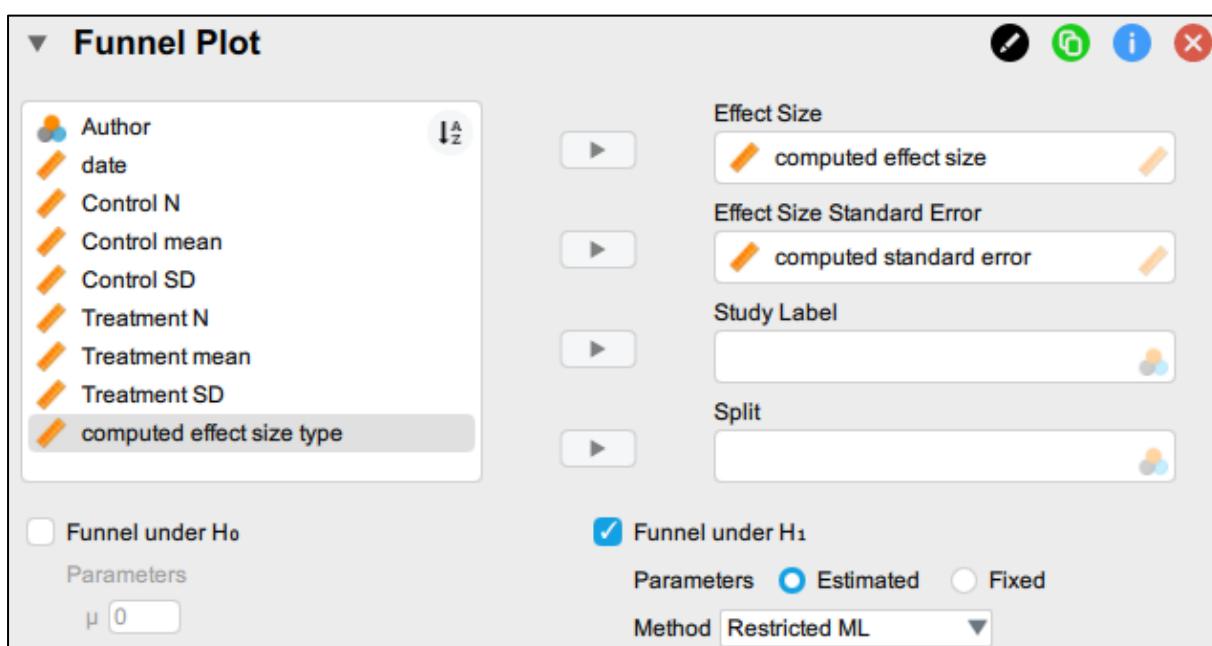
Forest Plot

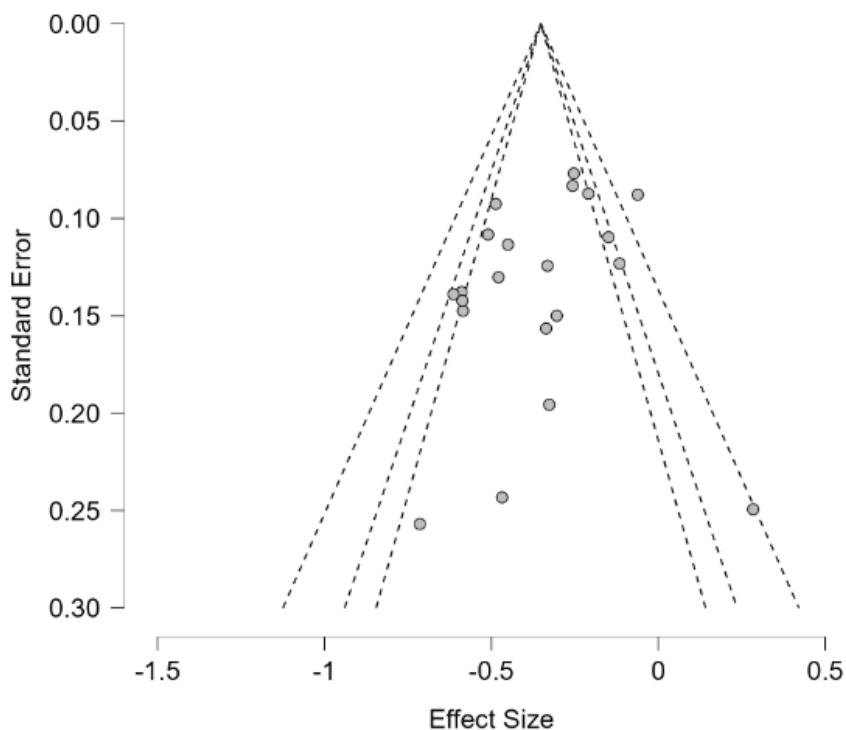


The model information is just a summary of the first 3 tables above.

Funnel Plot

From the Meta-Analysis icon, click on Funnel Plot. Add the computed effect size and computed standard error to their relevant boxes. Also, click on Funnel under H1 and click Funnel plot asymmetry tests.



**Funnel Plot****Funnel Plot Asymmetry Tests***Meta-Regression Test for Funnel Plot Asymmetry*

Estimates	Asymmetry Test		Limit Estimate		
	z	p	Estimate	Lower 95% CI	Upper 95% CI
21	-0.865	0.387	-0.245	-0.499	0.009

The funnel plot shows that the observed effect sizes appear to be symmetrically distributed around the vertical axis (based on the pooled effect size estimate, in this case, -0.352) and lie within the 95% confidence triangle. Asymmetry is often reported as being indicative of publication bias. This plot is accompanied by the 'Meta-Regression Test' for funnel plot asymmetry, which in this case is non-significant ($p=.387$).

Reporting the results.

A random-effects model (Restricted ML) was used in the analysis. The Residual Heterogeneity test confirms significant heterogeneity in the data $Q(20) = 49.4$, $p<.001$. This was supported with an I^2 of 59.3%. The Pooled Effect Size test showed a significant difference of -0.352 between the control and treatment studies, $t(20) = -7.9$, $p<.001$. Meta-regression showed no Funnel plot asymmetry, $Z=-0.865$, $p=0.387$.

Also, show the Forest plot.



JASP



EXPERIMENTAL DESIGN AND DATA LAYOUT IN EXCEL FOR JASP IMPORT.

Independent t-test

Design example:

Independent variable	Group 1	Group 2
Dependent variable	Data	Data

Independent variable

Categorical

Dependent variable

Continuous

	A	B
1	Group	Data
2	1	0
3	1	0
4	1	3.8
5	1	6
6	1	0.7
7	1	2.9
8	1	2.8
9	1	2
10	1	2
11	1	8.5
12	1	1.9
13	1	3.1
14	1	1.5
15	1	3
16	1	3.6
17	1	0.9
18	1	-2.1
19	2	2
20	2	1.7
21	2	4.3
22	2	7
23	2	0.6
24	2	2.7
25	2	3.6

More dependent variables can be added if required



Paired samples t-test

Design example:

Independent variable	Pre-test	Post-test
Participant	Dependent variable	
1	Data	Data
2	Data	Data
3	Data	Data
..n	Data	Data

Pre-test Post-test

	A	B
1	Pre-test	Post-test
2	60	60
3	103	103
4	58	54
5	60	54
6	64	63
7	64	61
8	65	62
9	66	64
10	67	65
11	69	61
12	70	68
13	70	67
14	72	71
15	72	69
16	72	68
17	82	81
18	58	60
19	58	56
20	59	57
21	61	57
22	62	55
23	63	62
24	63	60
25	63	59



Correlation

Design example:

Simple correlation

Participant	Variable 1	Variable 2	Variable 3	Variable 4	Variable ..n
1	Data	Data	Data	Data	Data
2	Data	Data	Data	Data	Data
3	Data	Data	Data	Data	Data
...n	Data	Data	Data	Data	Data

Multiple correlations

	A	B	C	D	E	F
1	Participant	Variable 1	Variable 2	Variable 3	Variable 4	Variable 5
2	1	533	77	77	106	106
3	2	472	63	59	92	93
4	3	484	82	77	93	78
5	4	536	72	72	103	93
6	5	630	77	68	104	93
7	6	563	68	68	101	87
8	7	531	77	82	108	106
9	8	344	50	50	86	92
10	9	346	54	50	90	86
11	10	386	59	54	85	80
12	11	460	54	63	89	83
13	12	492	63	59	92	94



Regression

Design example:

Simple Regression

Participant	Outcome	Predictor 1	Predictor 2	Predictor 3	Predictor ..n
1	Data	Data	Data	Data	Data
2	Data	Data	Data	Data	Data
3	Data	Data	Data	Data	Data
...n	Data	Data	Data	Data	Data

Multiple regression

	A	B	C	D	E	F
1	Participant	Outcome	Predictor 1	Predictor 2	Predictor 3	Predictor 4
2	1	533	77	77	106	106
3	2	472	63	59	92	93
4	3	484	82	77	93	78
5	4	536	72	72	103	93
6	5	630	77	68	104	93
7	6	563	68	68	101	87
8	7	531	77	82	108	106
9	8	344	50	50	86	92
10	9	346	54	50	90	86
11	10	386	59	54	85	80
12	11	460	54	63	89	83
13	12	492	63	59	92	94



Logistic Regression

Design example:

	Dependent Variable (categorical)	Factor (categorical)	Covariate (continuous)
Participant	Outcome	Predictor 1	Predictor 2
1	Data	Data	Data
2	Data	Data	Data
3	Data	Data	Data
...n	Data	Data	Data

A	B		C	D
1	ID	Outcome	Factor	Covariate
2	1	Yes	Yes	70
3	2	Yes	No	80
4	3	Yes	Yes	50
5	4	Yes	No	60
6	5	Yes	No	40
7	6	Yes	No	65
8	7	Yes	No	75
9	8	Yes	No	80
10	9	Yes	No	70
11	10	Yes	No	60
12	11	No	Yes	65
13	12	No	Yes	50
14	13	No	Yes	45
15	14	No	Yes	35
16	15	No	Yes	40
17	16	No	Yes	50
18	17	No	No	55
19	17	Yes	No	65
20	18	No	Yes	45

More factors and covariates can be added if required



One-way Independent ANOVA

Design example:

Independent variable	Group 1	Group 2	Group 3	Group...n
Dependent variable	Data	Data	Data	Data

Independent variable **Dependent variable****(Categorical)****(Continuous)**

	A	B
1	Group	Dependent variable
2	Group 1	3.8
3	Group 1	6
4	Group 1	0.7
5	Group 1	2.9
6	Group 1	2.8
7	Group 1	2
8	Group 1	2
9	Group 1	3.5
10	Group 2	1.9
11	Group 2	3.1
12	Group 2	1.5
13	Group 2	3
14	Group 2	3.6
15	Group 2	0.9
16	Group 2	-0.6
17	Group 3	1.1
18	Group 3	4.5
19	Group 3	6.1
20	Group 3	5
21	Group 3	2.4
22	Group 3	3.9
23	Group 3	3.5
24	Group 3	5.1
25	Group 3	3.5

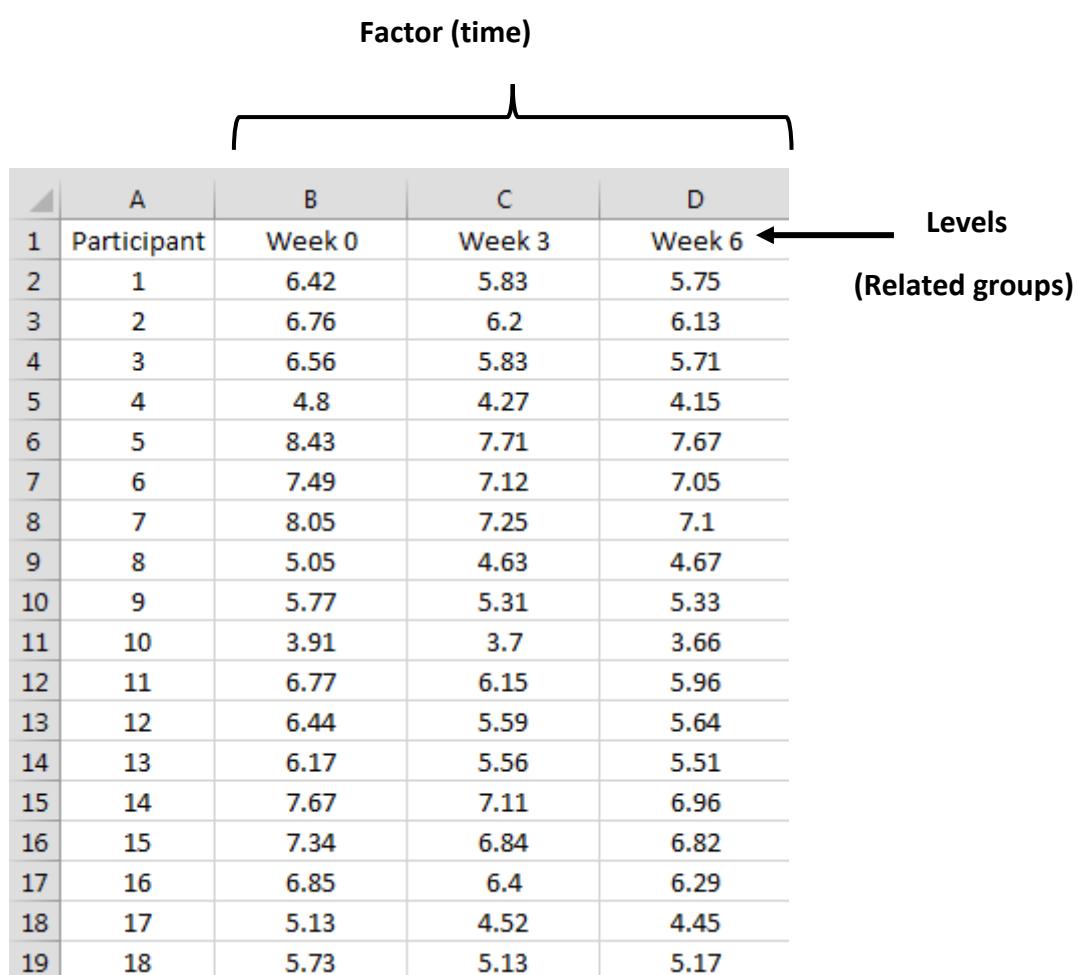
More dependent variables can be added if required



One-way repeated measures ANOVA

Design example:

Participant	Independent variable (Factor)			
	Level 1	Level 2	Level 3	Level..n
1	Data	Data	Data	Data
2	Data	Data	Data	Data
3	Data	Data	Data	Data
4	Data	Data	Data	Data
...n	Data	Data	Data	Data



More levels can be added if required



Two-way Independent ANOVA

Design example:

Factor 1	Supplement 1			Supplement 2		
Factor 2	Dose 1	Dose 2	Dose 3	Dose 1	Dose 2	Dose 3
Dependent variable	Data	Data	Data	Data	Data	Data

Factor 1 Factor 2 Dependent variable

	A	B	C
1	supp	dose	len
2	OJ	1000	19.7
3	OJ	1000	23.3
4	OJ	1000	23.6
5	OJ	1000	26.4
6	OJ	1000	20
7	OJ	1000	25.2
8	OJ	1000	25.8
9	OJ	1000	21.2
10	OJ	1000	14.5
11	OJ	1000	27.3
12	OJ	2000	25.5
13	OJ	2000	26.4
14	OJ	2000	22.4
15	OJ	2000	24.5
16	OJ	2000	24.8
17	OJ	2000	30.9
18	OJ	2000	26.4
19	OJ	2000	27.3
20	OJ	2000	29.4
21	OJ	2000	23
22	VC	1000	16.5
23	VC	1000	16.5
24	VC	1000	15.2
25	VC	1000	17.3

More factors and dependent variables can be added if required



Two-way Repeated measures ANOVA

Design example:

Factor 1 Interventions		Level 1			Level 2		
		i.e. intervention 1			i.e. intervention 2		
Factor 2 Time		Level 1 i.e time 1	Level 2 i.e time 2	Level 3 i.e time 3	Level 1 i.e time 1	Level 2 i.e time 2	Level 3 i.e time 3
		1	Data	Data	Data	Data	Data
2		Data	Data	Data	Data	Data	Data
3		Data	Data	Data	Data	Data	Data
...n		Data	Data	Data	Data	Data	Data

Factor 1 levels 1-n

Factor 2 levels 1-n

	A	B	C	D	E
1	Subject	Factor 1 level 1	Factor 1 level 2	Factor 2 level 1	Factor 2 level 2
2	A	7.38	6.52	9.27	14.32
3	B	7.71	10.83	11.48	16.38
4	C	6.19	10.42	9.77	15.45
5	D	9.27	11.78	15.45	16.96
6	E	11.41	9.52	11.65	15.64
7	F	5.29	5.82	9.22	13.01
8	G	8.54	9.43	10.92	17.35
9	H	7.89	8.43	8.26	12.57
10	I	5.49	6.64	11.39	14.02
11	J	9.26	9.36	13.03	16.24
12	K	6.9	7.09	9.02	14.7
13	L	8.57	9.64	8.33	13.71



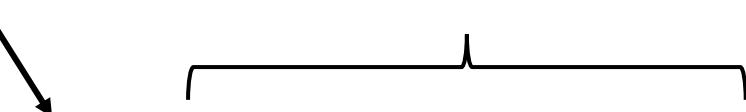
Two-way Mixed Factor ANOVA

Design example:

Factor 1 (Between subjects)		Group 1			Group 2		
Factor 2 levels (Repeated measures)		Trial 1	Trial 2	Trial 3	Trial 1	Trial 2	Trial 3
1		Data	Data	Data	Data	Data	Data
2		Data	Data	Data	Data	Data	Data
3		Data	Data	Data	Data	Data	Data
...n		Data	Data	Data	Data	Data	Data

Factor 1
(Categorical)

Factor 2 levels
(Continuous)



	A	B	C	D
1	Group	Level 1	Level 2	Level 3
2	Group 1	1.31	0.9	0.9
3	Group 1	1.29	0.89	0.72
4	Group 1	1.8	0.9	0.96
5	Group 1	1.4	1.26	0.97
6	Group 1	1.49	1.18	0.88
7	Group 1	1.35	1.15	0.92
8	Group 1	1.45	1.19	1
9	Group 1	1.21	1.2	0.85
10	Group 1	1.79	1.48	0.99
11	Group 1	1.73	1.68	0.98
12	Group 2	1.55	0.9	0.55
13	Group 2	1.27	0.95	0.41
14	Group 2	1.53	0.87	0.42
15	Group 2	1.26	1.15	0.44
16	Group 2	1.14	1.12	0.38
17	Group 2	1.11	1.08	0.34
18	Group 2	1.1	1.0758	0.18
19	Group 2	1.08	1.18	0.24
20	Group 2	1.3	1.26	0.39
21	Group 2	1.45	1.55	0.44



Chi-squared - Contingency tables

Design example:

Participant	Response 1	Response 2	Response 3	Response...n
1	Data	Data	Data	Data
2	Data	Data	Data	Data
3	Data	Data	Data	Data
..n	Data	Data	Data	Data

All data should be categorical

	A	B	C	D	E
1	Respondant	Response 1	Response 2	Response 3	Response 4
2	1	Female	clay	Morning	yes
3	2	Male	astro	Morning	No
4	3	Female	grass	Evening	No
5	4	Male	clay	Afternoon	No
6	5	Male	clay	Morning	No
7	6	Male	grass	Evening	No
8	7	Female	grass	Evening	yes
9	8	Male	clay	Morning	yes
10	9	Female	grass	Morning	No
11	10	Male	clay	Afternoon	No
12	11	Female	clay	Afternoon	No
13	12	Male	astro	Afternoon	No
14	13	Male	astro	Afternoon	No
15	14	Male	astro	Afternoon	yes
16	15	Female	clay	Morning	No
17	16	Male	astro	Afternoon	yes
18	17	Female	astro	Afternoon	yes
19	18	Male	grass	Morning	No
20	19	Male	clay	Afternoon	No



SOME CONCEPTS IN FREQUENTIST STATISTICS

The frequentist approach is the most commonly taught and used statistical methodology. It describes sample data based on the frequency or proportion of the data from repeated studies through which the probability of events is defined.

Frequentist statistics uses rigid frameworks including hypothesis testing, p values confidence intervals etc.

Hypothesis testing

A hypothesis can be defined as "***a supposition or proposed explanation made based on limited evidence as a starting point for further investigation***".

There are two simple types of hypotheses, a null hypothesis (H_0) and an alternative or experimental hypothesis (H_1). The **null hypothesis** is the default position for most statistical analyses in which it is stated that there is no relationship or difference between groups. The **alternative hypothesis** states that there is a relationship or difference between groups in a direction of difference/relationship. For example, if a study was carried out to look at the effects of a supplement on sprint time in one group of participants compared to the placebo group:

H_0 = there is **no** difference in sprint times between the two groups

H_1 = there is a difference in sprint times between the two groups

H_2 = Group 1 is greater than Group 2

H_3 = Group 1 is less than Group 2

Hypothesis testing refers to the strictly predefined procedures used to accept or reject the hypotheses and the probability that this could be purely by chance. The confidence at which a null hypothesis is accepted or rejected is called the level of significance. The level of significance is denoted by α , usually 0.05 (5%). This is the level of probability of accepting an effect as true (95%) and that there is only 5% of the result is purely by chance.

Different types of hypotheses can easily be selected in JASP, however, the null hypothesis is always the default.

Hypothesis

- Group 1 \neq Group 2
- Group 1 $>$ Group 2
- Group 1 $<$ Group 2



Type I and II errors

The probability of rejecting the null hypothesis, when it is, in fact, true, is called Type I error whereas the probability of accepting the null hypothesis when it is not true is called Type II error.

		The truth	
		Not guilty (H_0)	Guilty (H_1)
The verdict	Guilty (H_1)	Type I error An innocent person goes to prison	Correct decision
	Not guilty (H_0)	Correct decision	Type II error A guilty person goes free

Type I error is deemed the worst error to make in statistical analyses.

Statistical power is defined as the probability that the test will reject the null hypothesis when the alternative hypothesis is true. For a set level of significance, if the sample size increases, the probability of Type II error decreases, which therefore increases the statistical power.

Testing the hypothesis

The essence of hypothesis testing is to first define **the null (or alternative) hypothesis**, set the criterion level α , usually 0.05 (5%), collect and analyse sample data. Use a **test statistic** to determine how far (or the number of standard deviations) the sample mean is from the population mean stated in the null hypothesis. The test statistic is then compared to a critical value. This is a cut-off value defining the boundary where less than 5% of the sample means can be obtained if the null hypothesis is true.

If the probability of obtaining a difference between the means by chance is less than 5% when the null hypothesis has been proposed, the null hypothesis is rejected and the alternative hypothesis can be accepted.

The **p-value** is the probability of obtaining a sample outcome, given that the value stated in the null hypothesis is true. If the p-value is less than 5% ($p < .05$) the null hypothesis is rejected. When the p-value is greater than 5% ($p > .05$), we accept the null hypothesis.

Effect size

An effect size is a standard measure that can be calculated from any number of statistical analyses. If the null hypothesis is rejected the result is significant. This significance only evaluates the probability of obtaining the sample outcome by chance but does not indicate how big a difference (practical significance) is, nor can it be used to compare across different studies.

The effect size indicates the magnitude of the difference between the groups. So for example, if there was a significant decrease in 100m sprint times in a supplement compared to a



placebo group, the effect size would indicate how much more effective the intervention was. Some common effect sizes are shown below.

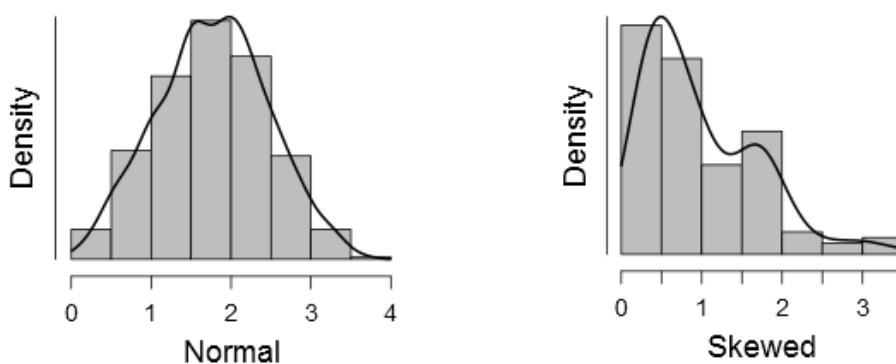
Test	Measure	Trivial	Small	Medium	Large
Between means	Cohen's d	<0.2	0.2	0.5	0.8
Correlation	Correlation coefficient (r)	<0.1	0.1	0.3	0.5
	Rank -biserial (r_B)	<0.1	0.1	0.3	0.5
	Spearman's rho	<0.1	0.1	0.3	0.5
Multiple Regression	Multiple correlation coefficient (R)	<0.10	0.1	0.3	0.5
ANOVA	Eta	<0.1	0.1	0.25	0.37
	Partial Eta	<0.01	0.01	0.06	0.14
	Omega squared	<0.01	0.01	0.06	0.14
Chi-squared	Phi (2x2 tables only)	<0.1	0.1	0.3	0.5
	Cramer's V	<0.1	0.1	0.3	0.5
	Odds ratio (2x2 tables only)	<1.5	1.5	3.5	9.0

In small datasets, there may be a moderate to large effect size but no significant differences. This could suggest that the analysis lacked statistical power and that increasing the number of data points may show a significant outcome. Conversely, when using large datasets, significant testing can be misleading since small or trivial effects may produce statistically significant results.

PARAMETRIC vs NON-PARAMETRIC TESTING

Most research collects information from a sample of the population of interest, it is normally impossible to collect data from the whole population. We do, however, want to see how well the collected data reflects the population in terms of the population mean, standard deviations, proportions etc. based on parametric distribution functions. These measures are the population **parameters**. Parameter estimates of these in the sample population are statistics. Parametric statistics require assumptions to be made of the data including the normality of distribution and homogeneity of variance.

In some cases, these assumptions may be violated in that the data may be noticeably skewed:



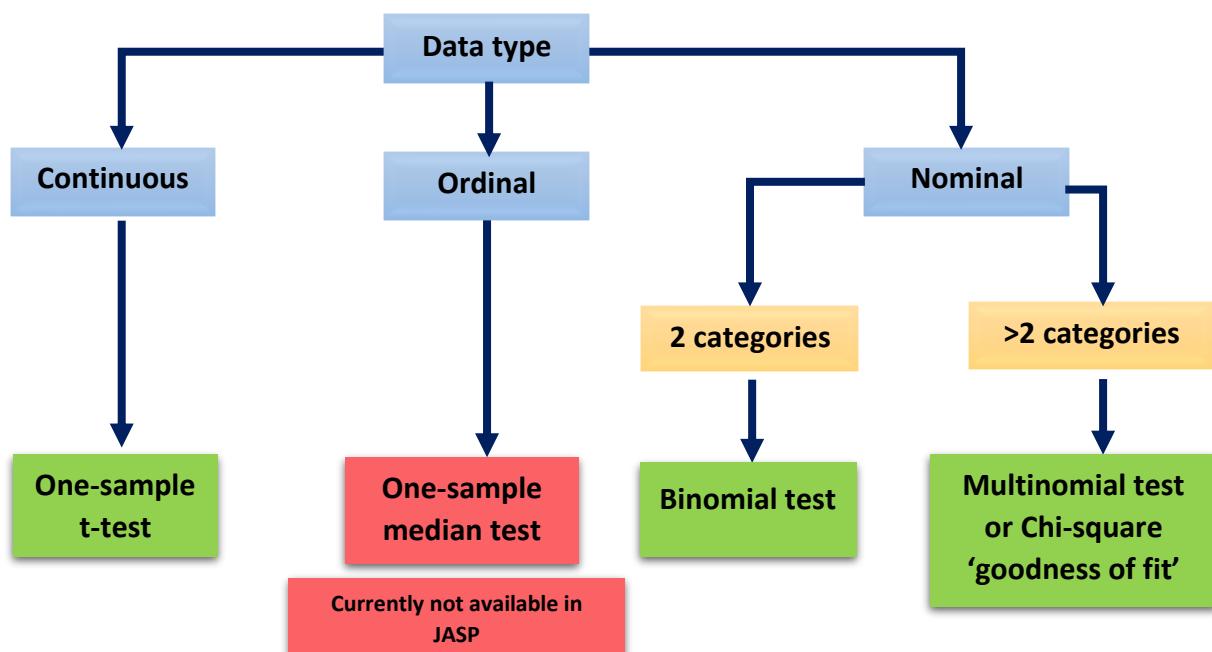
Sometimes transforming the data can rectify this but not always. It is also common to collect ordinal data (i.e. Likert scale ratings) for which terms such as mean and standard deviation are meaningless. As such there are no parameters associated with ordinal (**non-parametric**) data. The non-parametric counterparts include median values and quartiles.

In both of the cases described non-parametric statistical tests are available. There are equivalents to the most common classical parametric tests. These tests don't assume normally distributed data or population parameters and are based on sorting the data into ranks from lowest to highest values. All subsequent calculations are done with these ranks rather than with the actual data values.

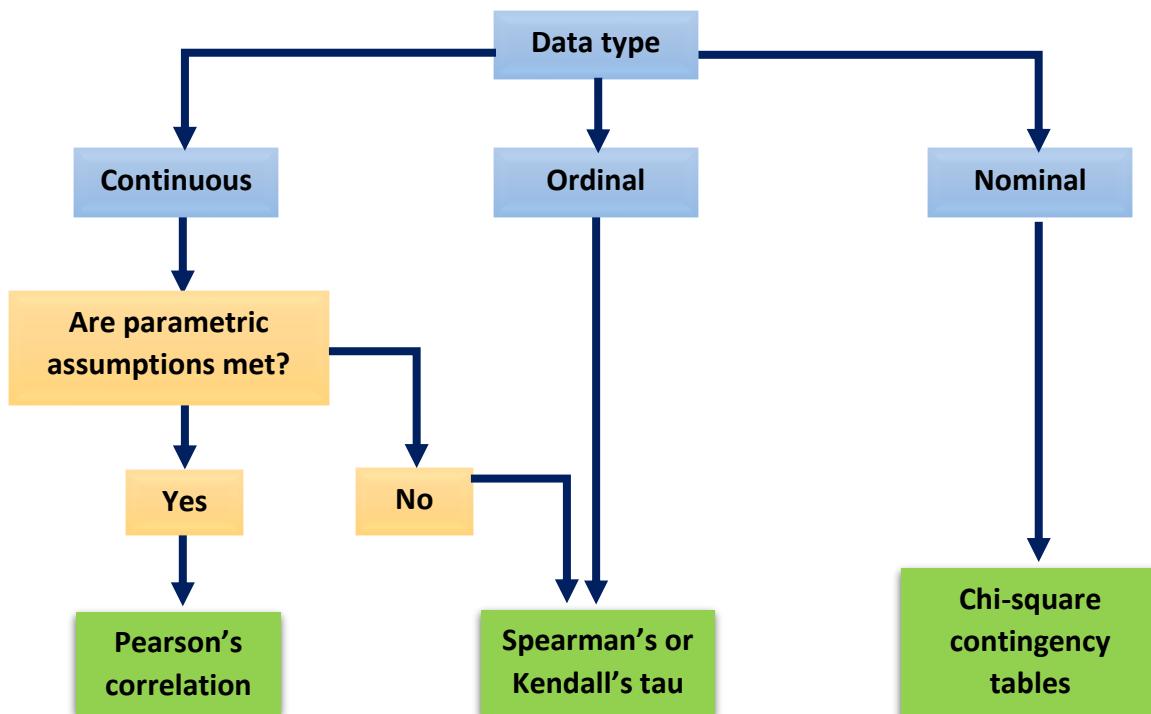


WHICH TEST SHOULD I USE?

Comparing one sample to a known or hypothesized population mean.

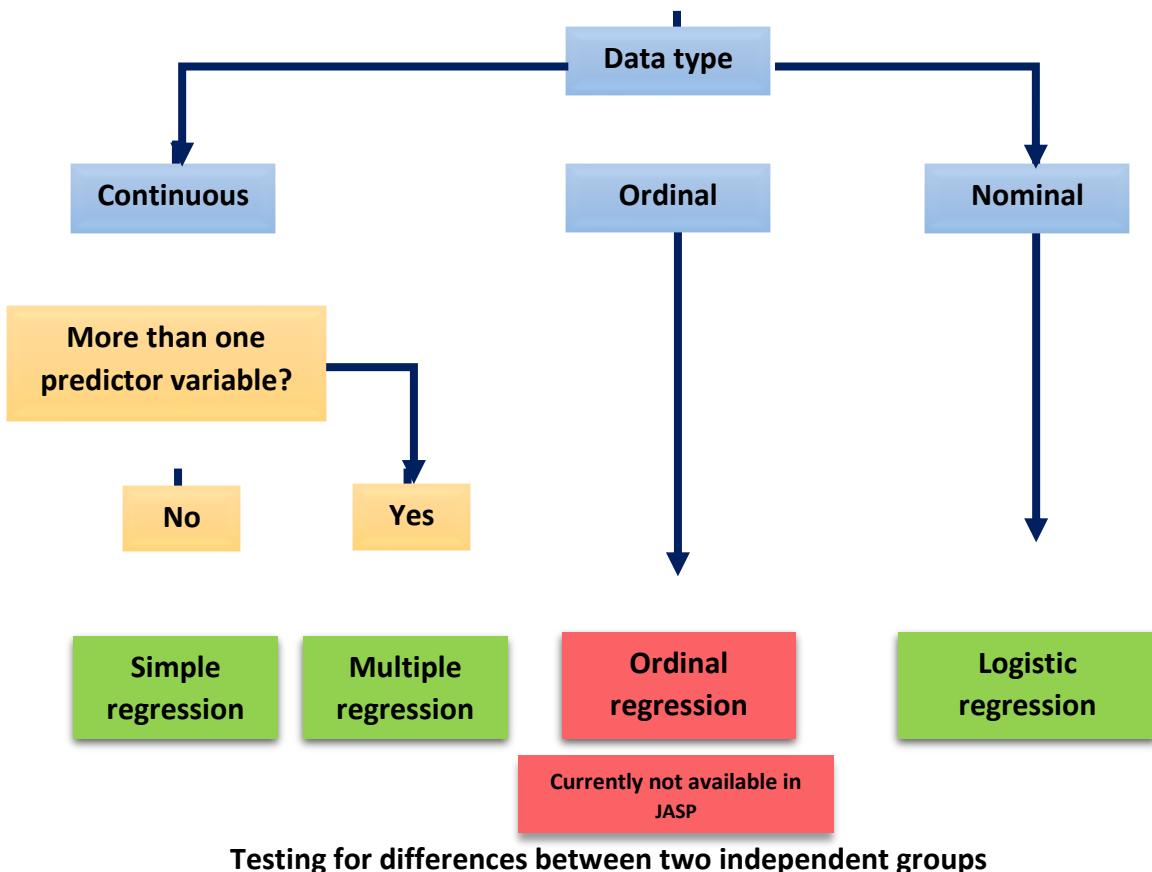


Testing relationships between two or more variables

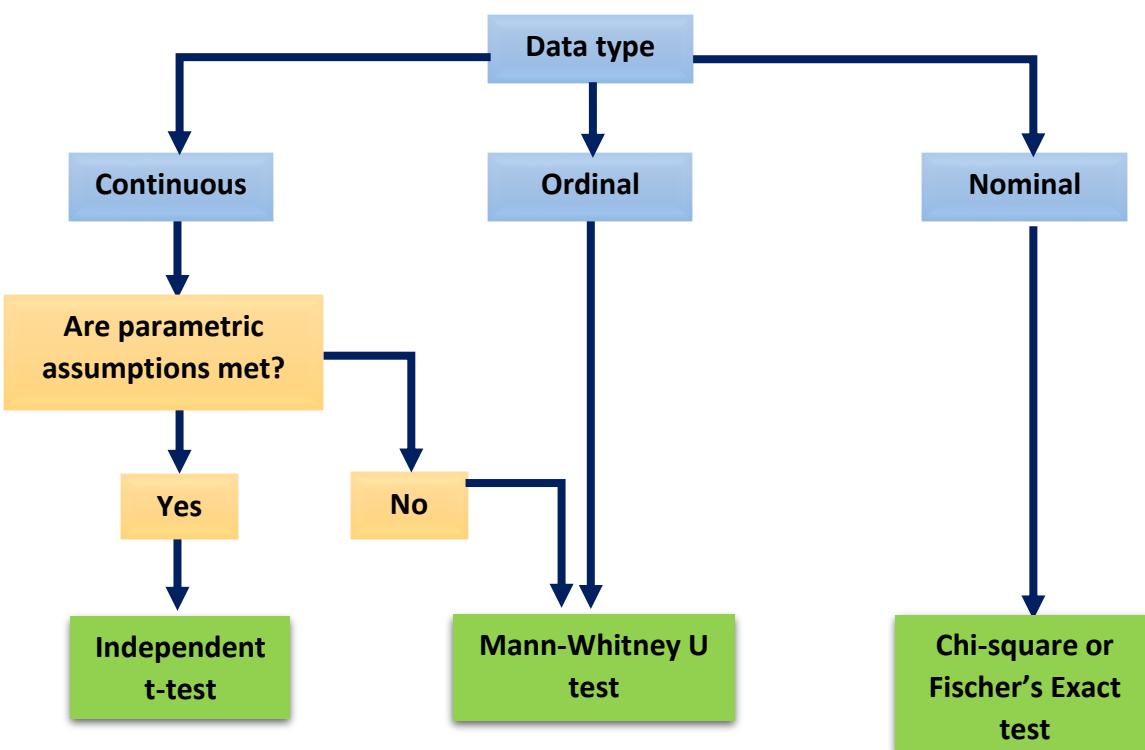




Predicting outcomes

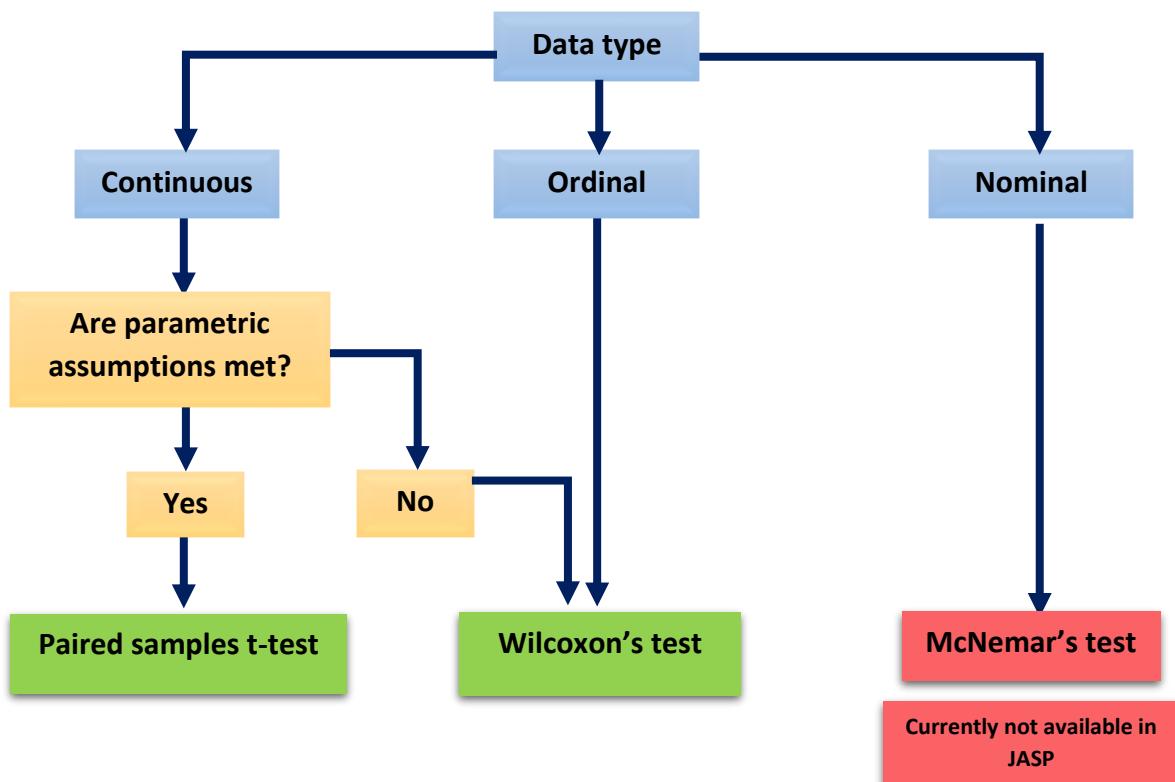


Testing for differences between two independent groups

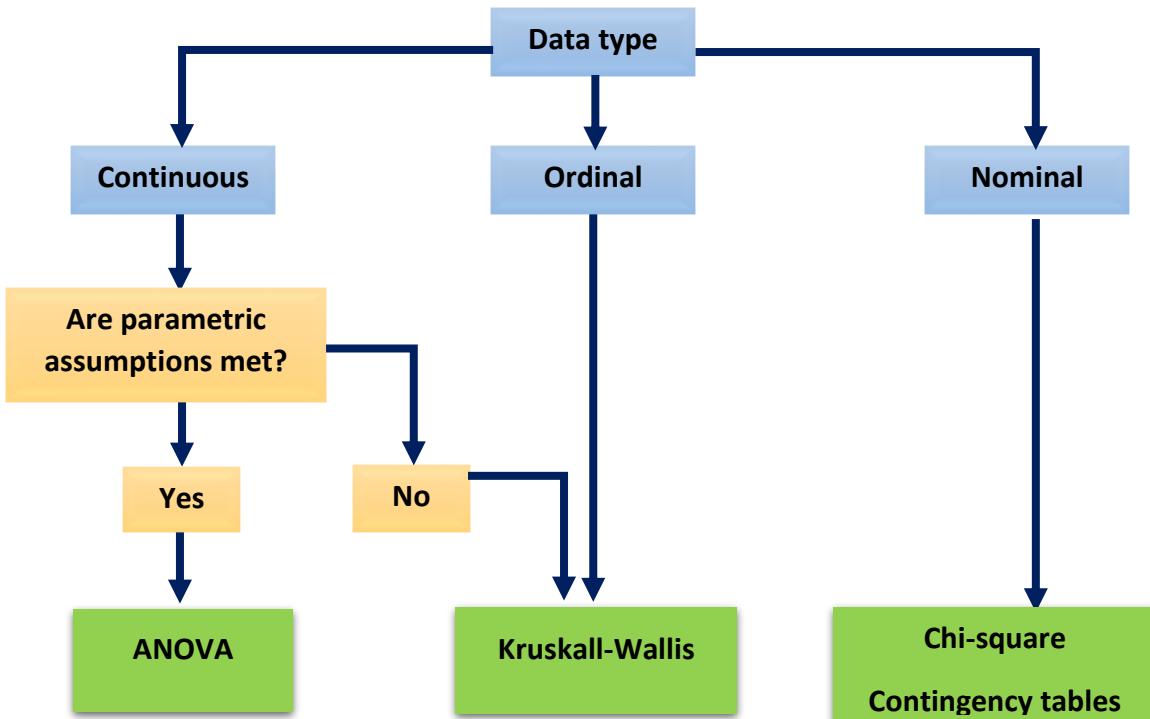




Testing for differences between two related groups

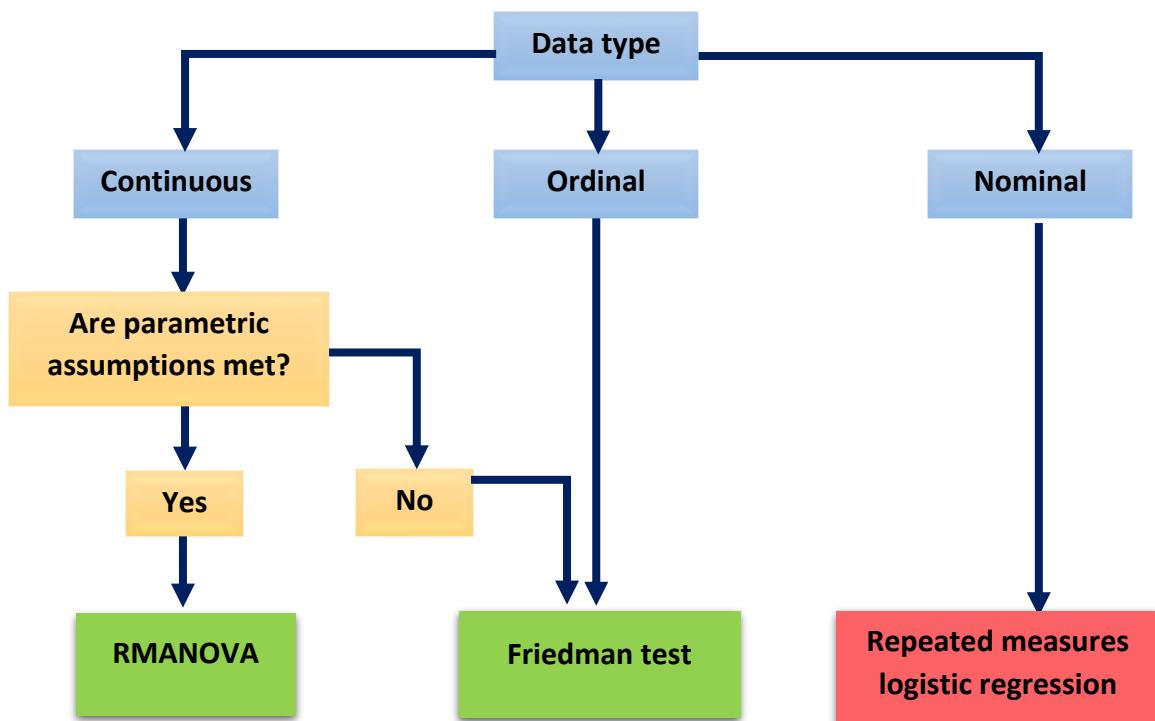


Testing for differences between three or more independent groups





Testing for differences between three or more related groups



Test for interactions between 2 or more independent variables

