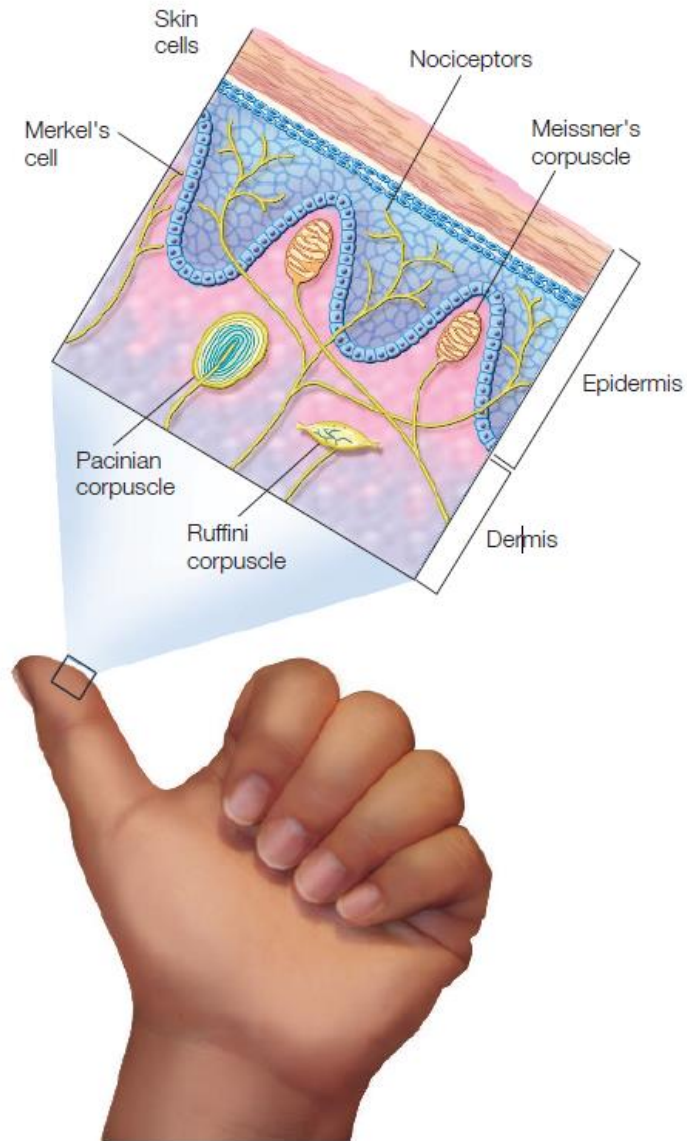




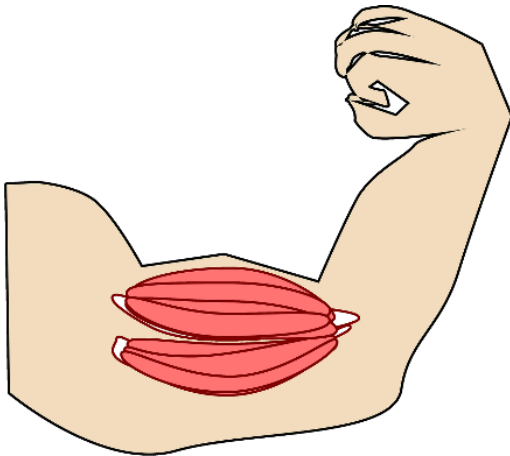
# Somatosensation



# Somatosensation subgroups

## PROPRIOCEPTION

Sensing oneself (proprius  
→ one's own).



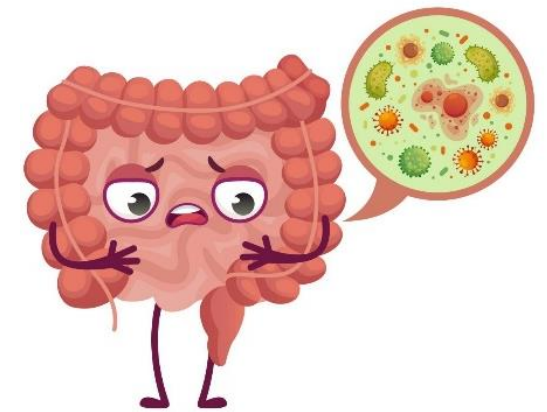
## EXTEROCEPTION

Sensing the  
outside world

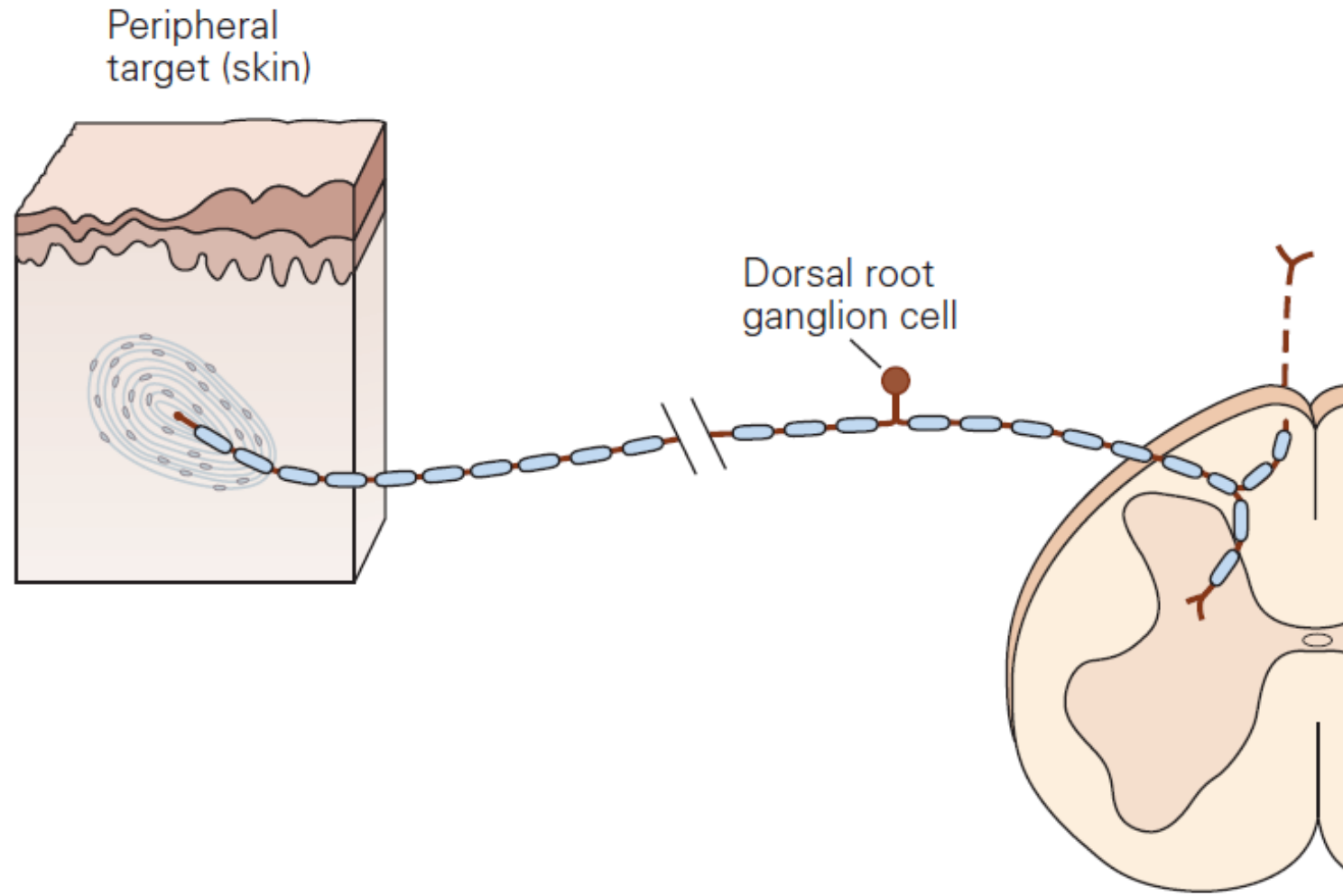


## INTEROCEPTION

Sensing the  
inside world



# DRGs

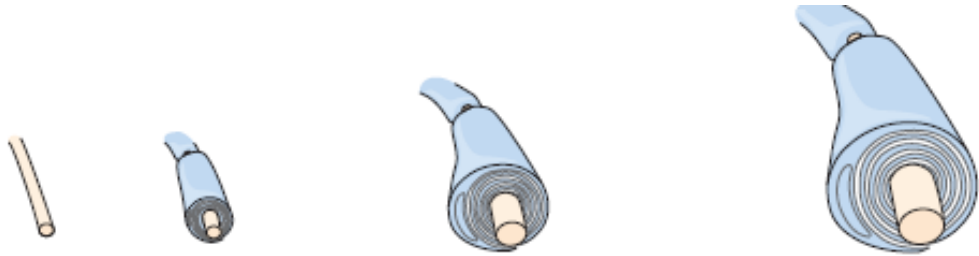


# Nerve fiber classification

	Muscle nerve	Cutaneous nerve <sup>2</sup>	Fiber diameter (μm)	Conduction velocity (m/s)
Myelinated				
Large diameter	I	Aα	12–20	72–120
Medium diameter	II	Aβ	6–12	36–72
Small diameter	III	Aδ	1–6	4–36
Unmyelinated	IV	C	0.2–1.5	0.4–2.0

<sup>1</sup>Sensory fibers from muscle are classified according to their diameter, whereas those from the skin are classified by conduction velocity.

<sup>2</sup>The types of receptors innervated by each type of fiber are listed in Table 22–2.



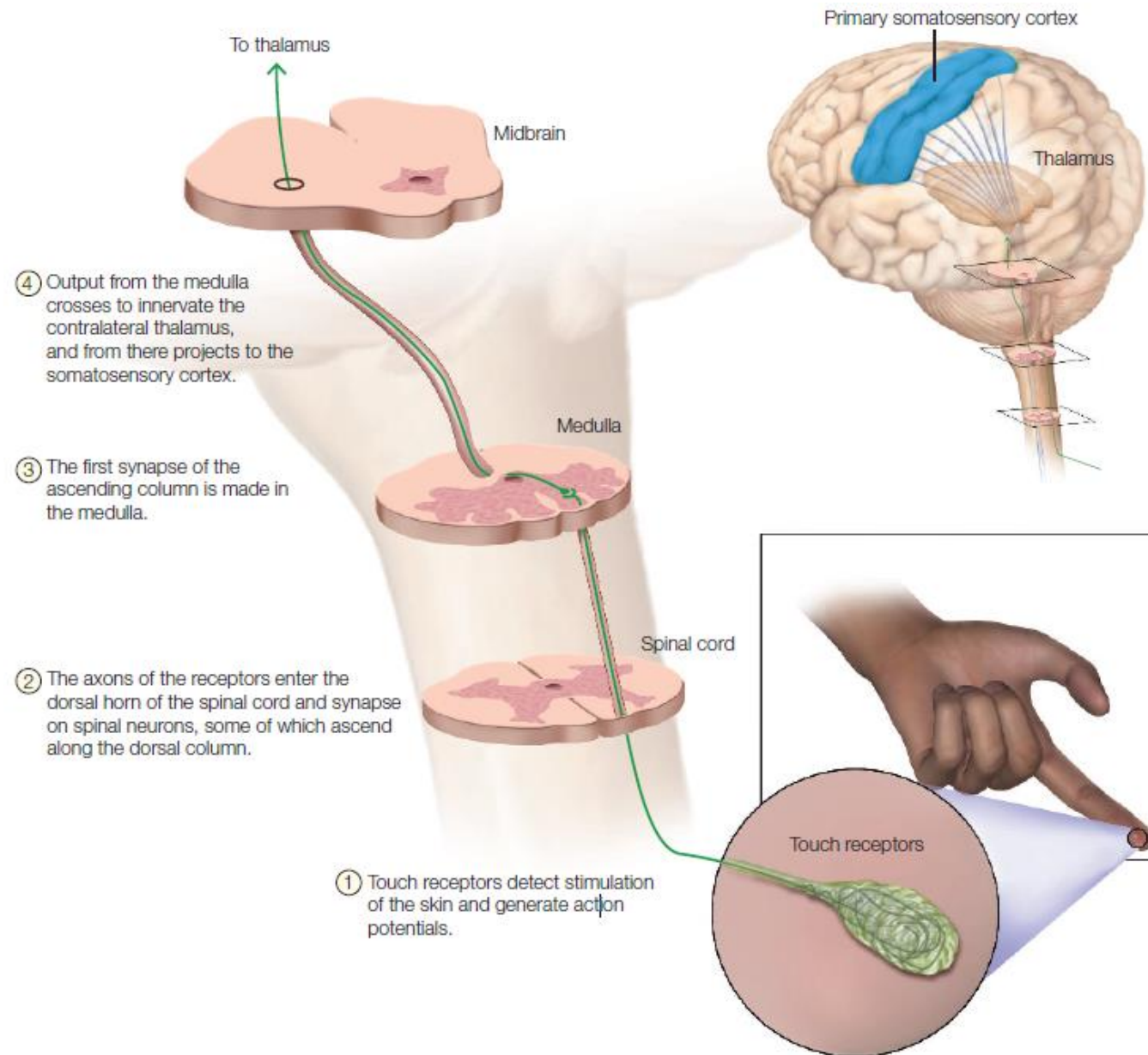
Axon diameter (μm)	1	5	12	20
Conduction velocity (m/s)	1	30	72	120

# Receptors, Fibers and Modalities

Receptor type	Fiber group <sup>1</sup>	Fiber name	Modality
Cutaneous and subcutaneous mechanoreceptors			Touch
Meissner corpuscle	A $\alpha$ , $\beta$	RA1	Stroking, flutter
Merkel disk receptor	A $\alpha$ , $\beta$	SA1	Pressure, texture
Pacinian corpuscle <sup>2</sup>	A $\alpha$ , $\beta$	RA2	Vibration
Ruffini ending	A $\alpha$ , $\beta$	SA2	Skin stretch
Hair-tylotrich, hair-guard	A $\alpha$ , $\beta$	G1, G2	Stroking, fluttering
Hair-down	A $\delta$	D	Light stroking
Field	A $\alpha$ , $\beta$	F	Skin stretch
C mechanoreceptor	C		Stroking, erotic touch
Thermal receptors			Temperature
Cool receptors	A $\delta$	III	Skin cooling (<25°C [77°F])
Warm receptors	C	IV	Skin warming (>35°C [95°F])
Heat nociceptors	A $\delta$	III	Hot temperature (>45°C [113°F])
Cold nociceptors	C	IV	Cold temperature (<5°C [41°F])
Nociceptors			Pain
Mechanical	A $\delta$	III	Sharp, pricking pain
Thermal-mechanical (heat)	A $\delta$	III	Burning pain
Thermal-mechanical (cold)	C	IV	Freezing pain
Polymodal	C	IV	Slow, burning pain
Muscle and skeletal mechanoreceptors			Limb proprioception
Muscle spindle primary	A $\alpha$	Ia	Muscle length and speed
Muscle spindle secondary	A $\beta$	II	Muscle stretch
Golgi tendon organ	A $\alpha$	Ib	Muscle contraction
Joint capsule receptors	A $\beta$	II	Joint angle
Stretch-sensitive free endings	A $\delta$	III	Excess stretch or force

<sup>1</sup>See Table 22-1

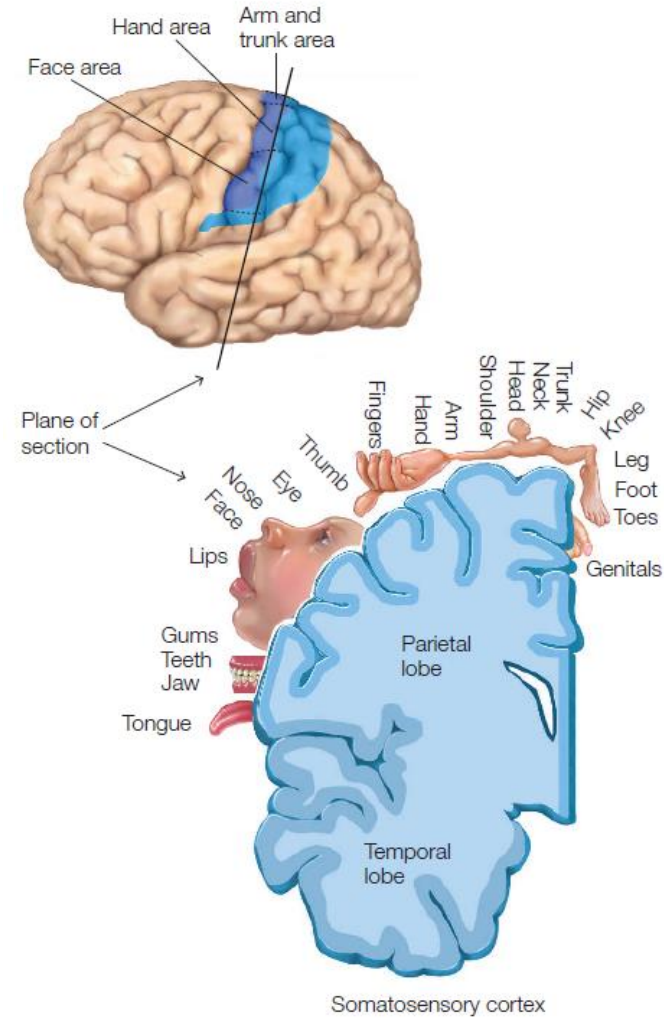
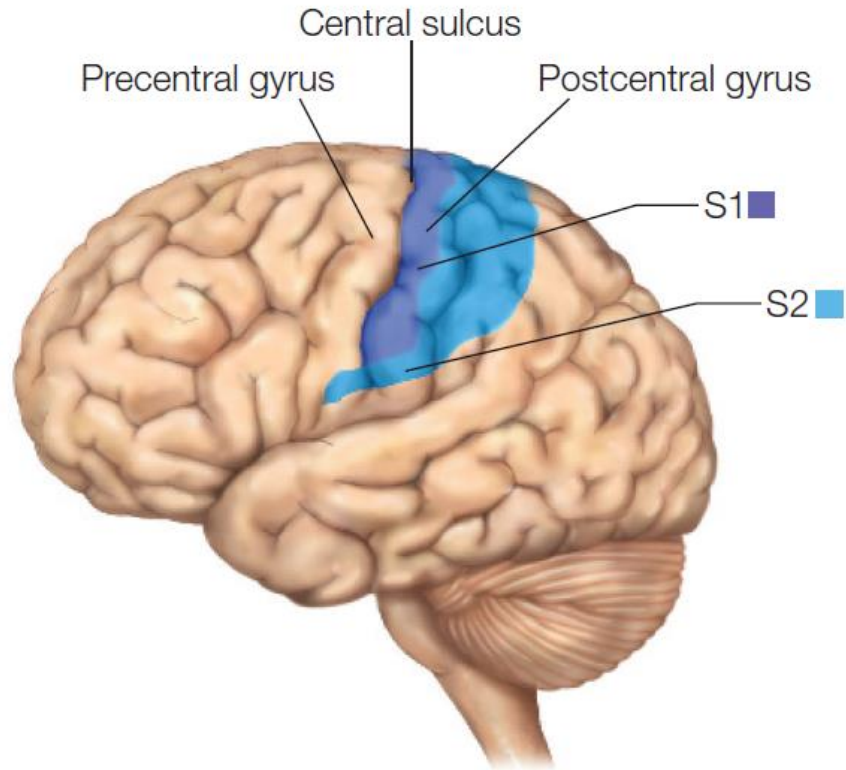
# From periphery to brain



Thalamus:  
Door to consciousness



# The primary somatosensory cortex

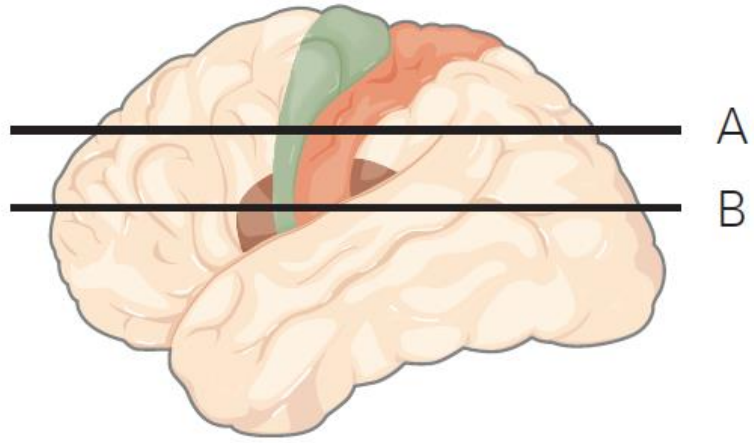


## Homunculus

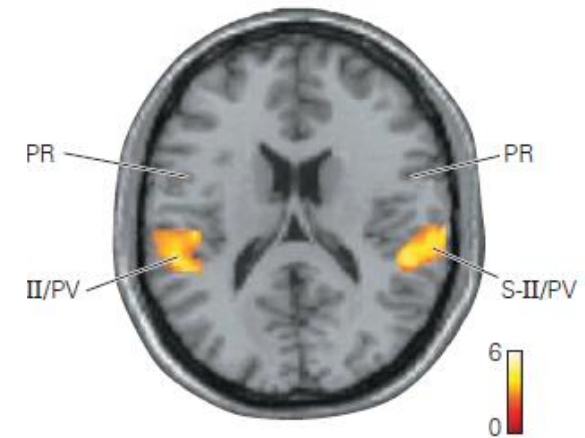
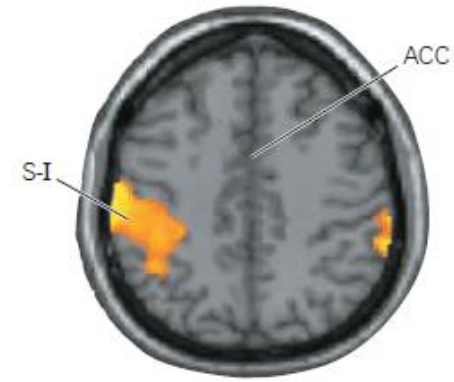




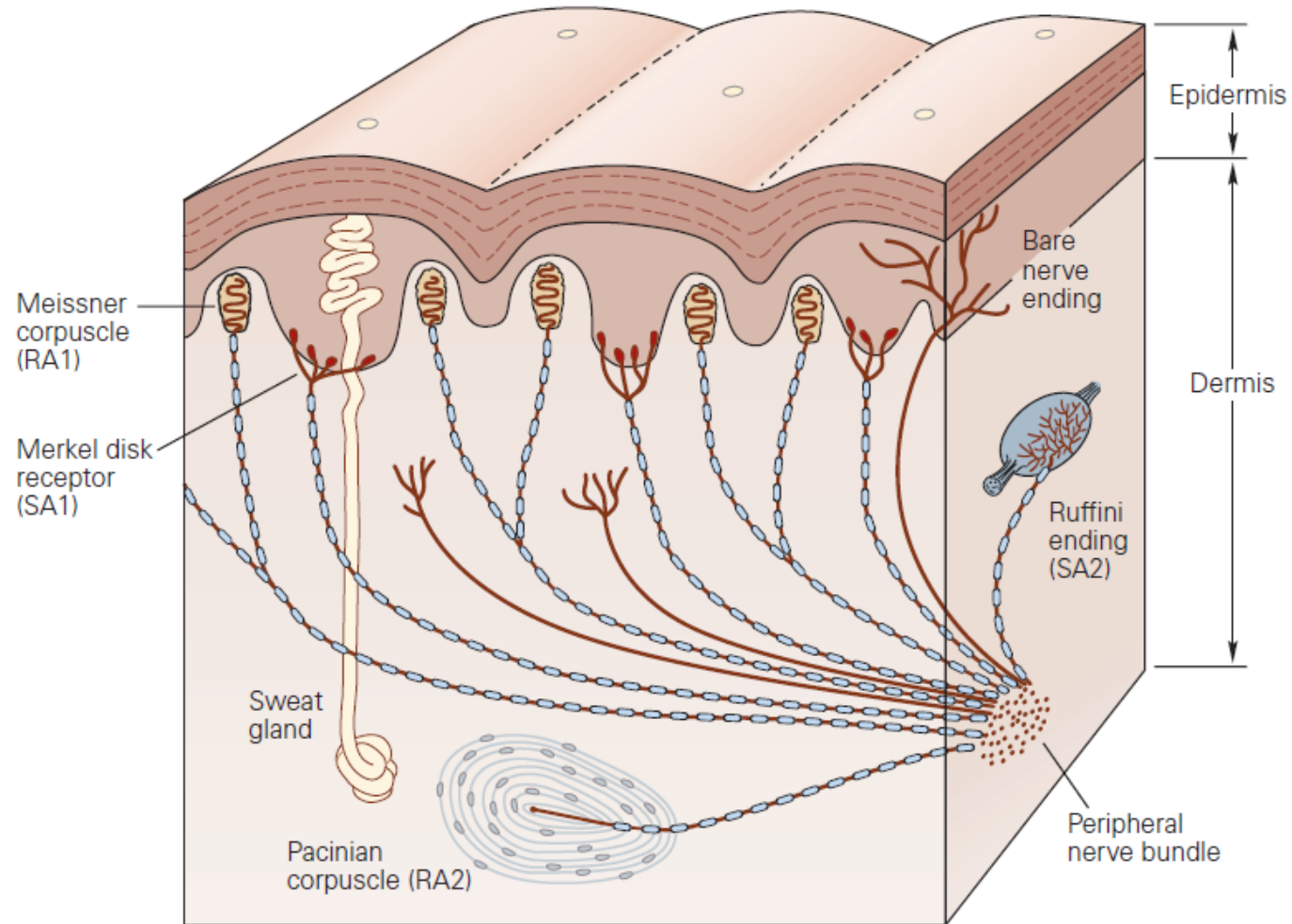
# Touch



Passive touch



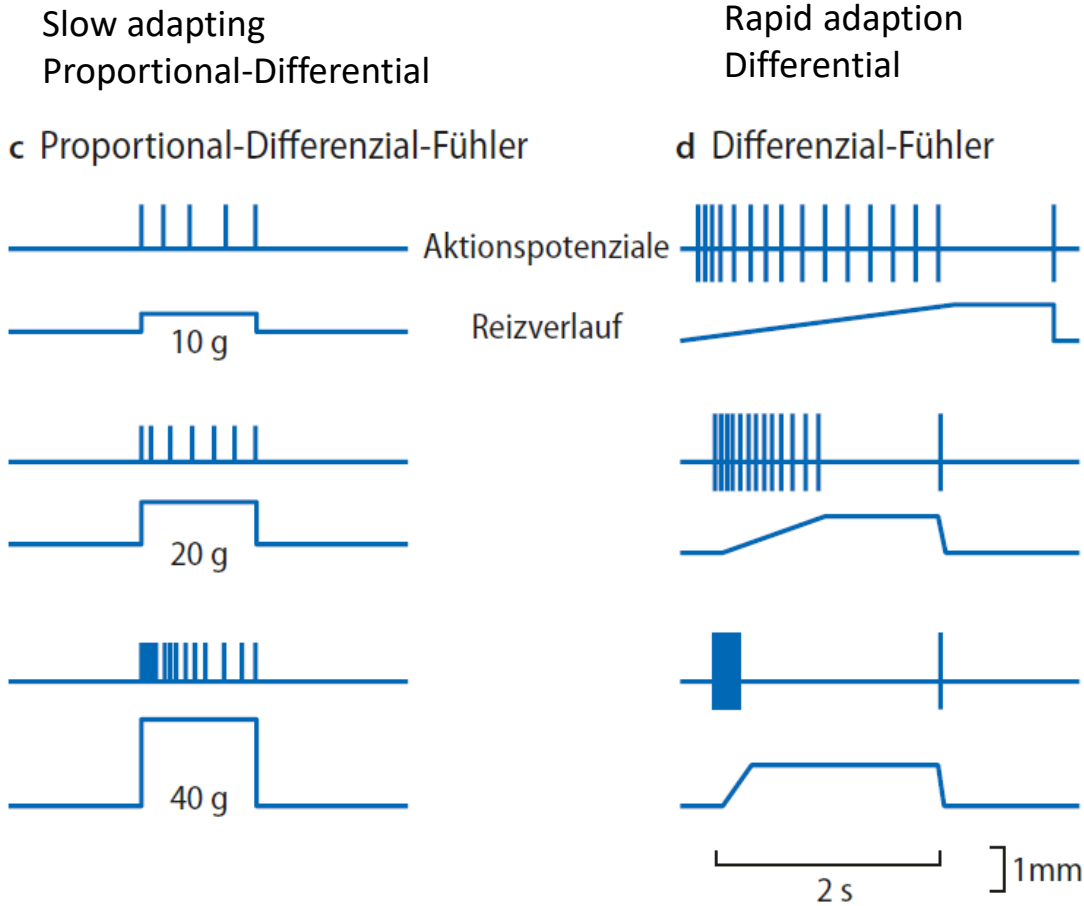
# How does „touch“ work?



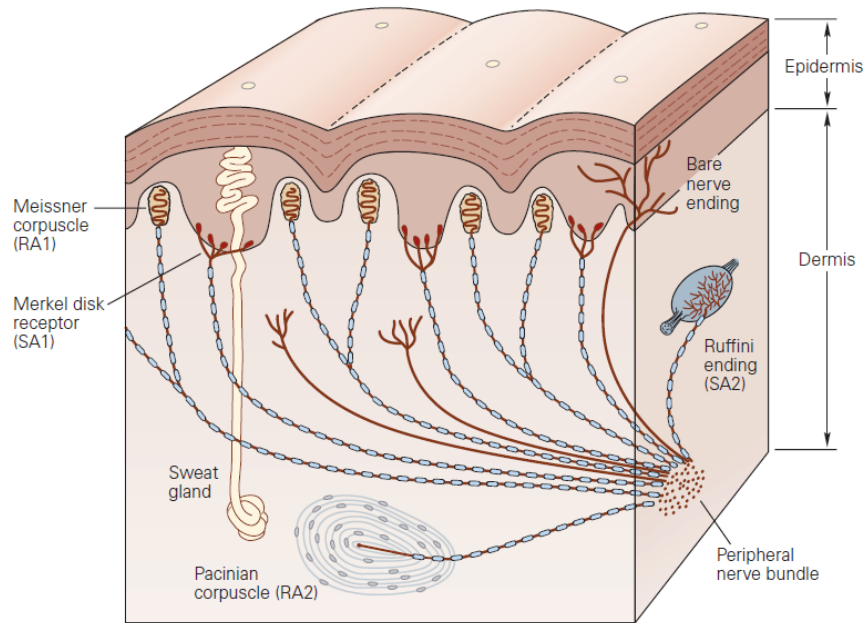
# Cutaneous mechanoreceptor systems

	Type 1		Type 2	
	SA1	RA1 <sup>1</sup>	SA2	RA2 <sup>2</sup>
Receptor	Merkel cell	Meissner corpuscle	Ruffini ending	Pacinian corpuscle
Location	Tip of epidermal sweat ridges	Dermal papillae (close to skin surface)	Dermis	Dermis (deep tissue)
Axon diameter (μm)	7–11	6–12	6–12	6–12
Conduction velocity (ms)	40–65	35–70	35–70	35–70
Best stimulus	Edges, points	Lateral motion	Skin stretch	Vibration
Response to sustained indentation	Sustained with slow adaptation	None	Sustained with slow adaptation	None
Frequency range (Hz)	0–100	1–300		5–1,000
Best frequency (Hz)	5	50		200
Threshold for rapid indentation or vibration (best) (μm)	8	2	40	0.01

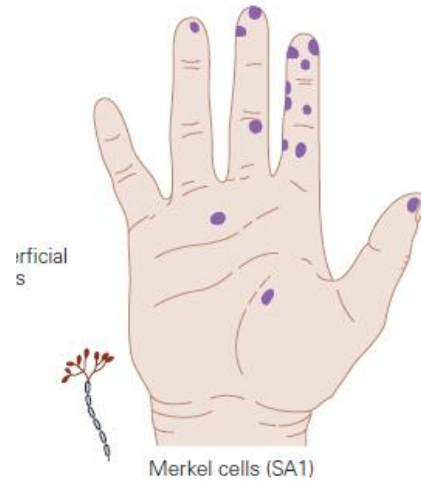
<sup>1</sup>Also called RA, QA, or FA1.  
<sup>2</sup>Also called PC or FA2.  
RA1, rapidly adapting type 1; RA2, rapidly adapting type 2; SA1, slowly adapting type 1; SA2, slowly adapting type 2.



# Receptive fields in the skin



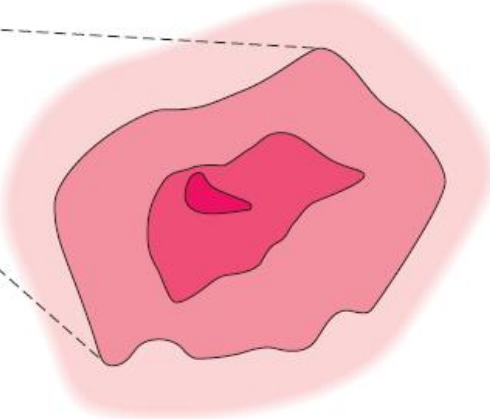
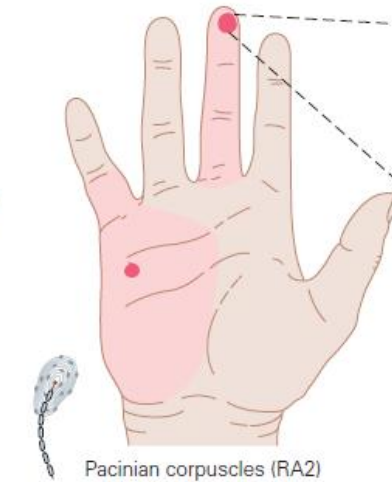
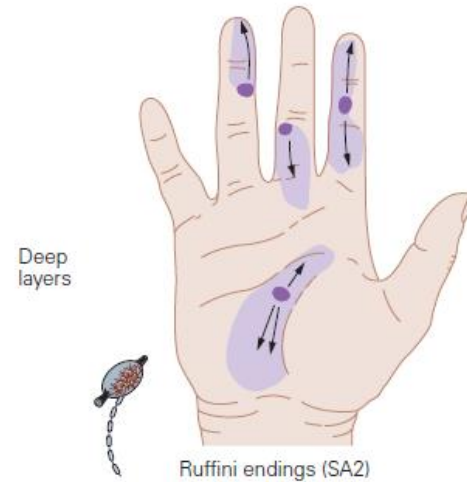
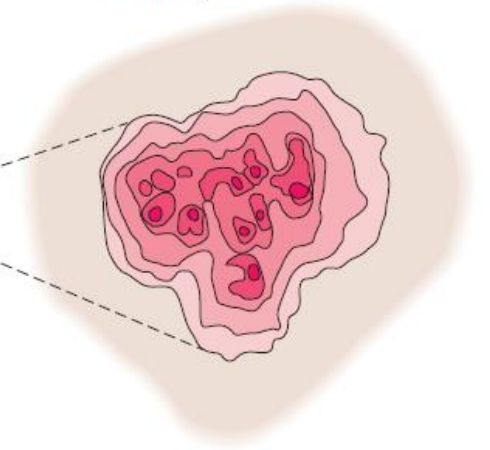
A Slowly adapting mechanoreceptors



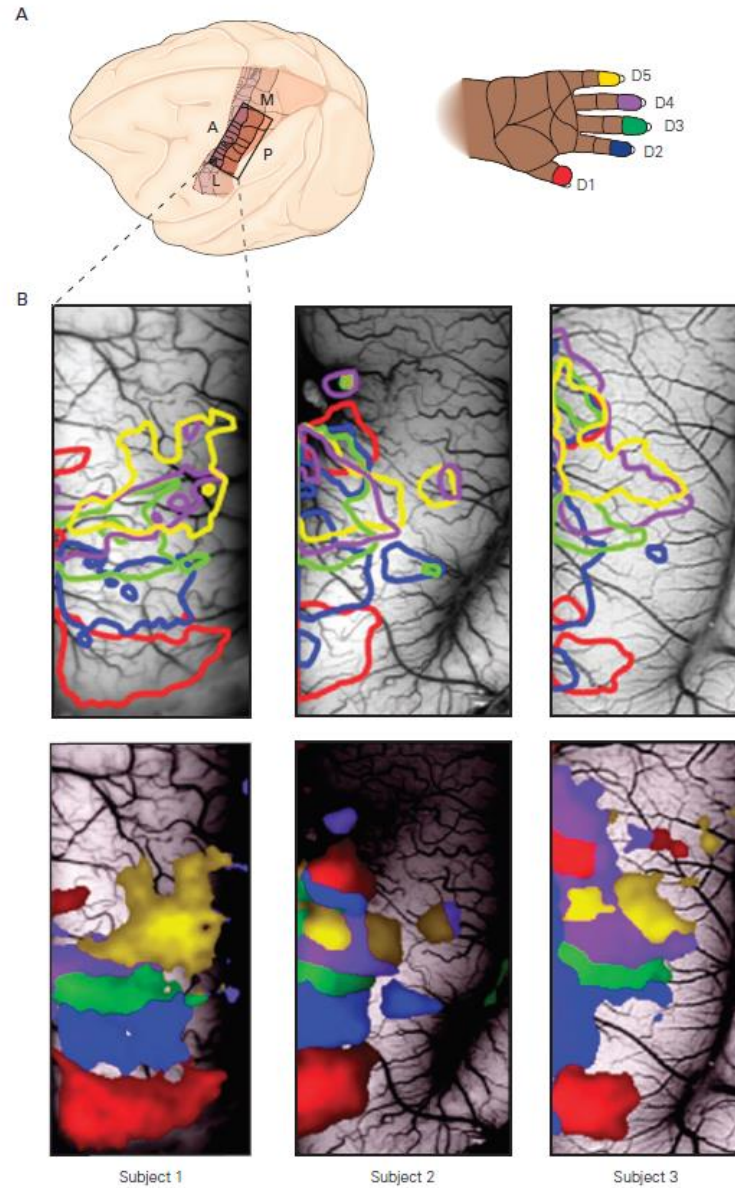
B Rapidly adapting mechanoreceptors



C Receptive field sensitivity

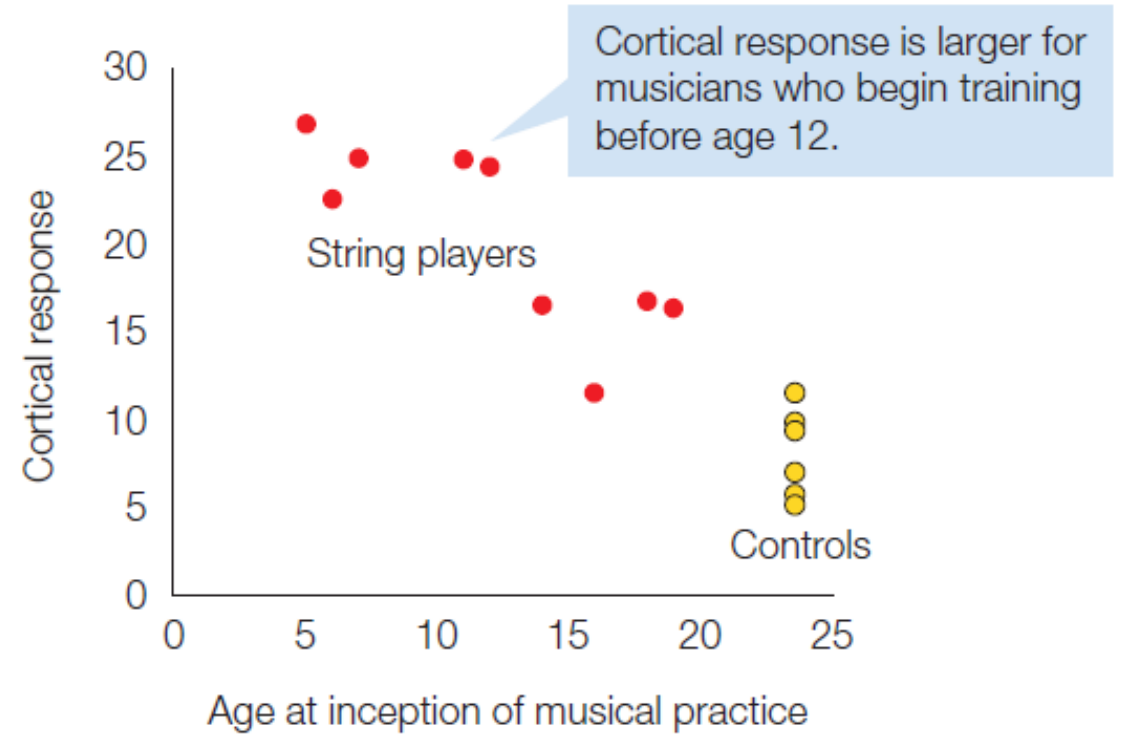
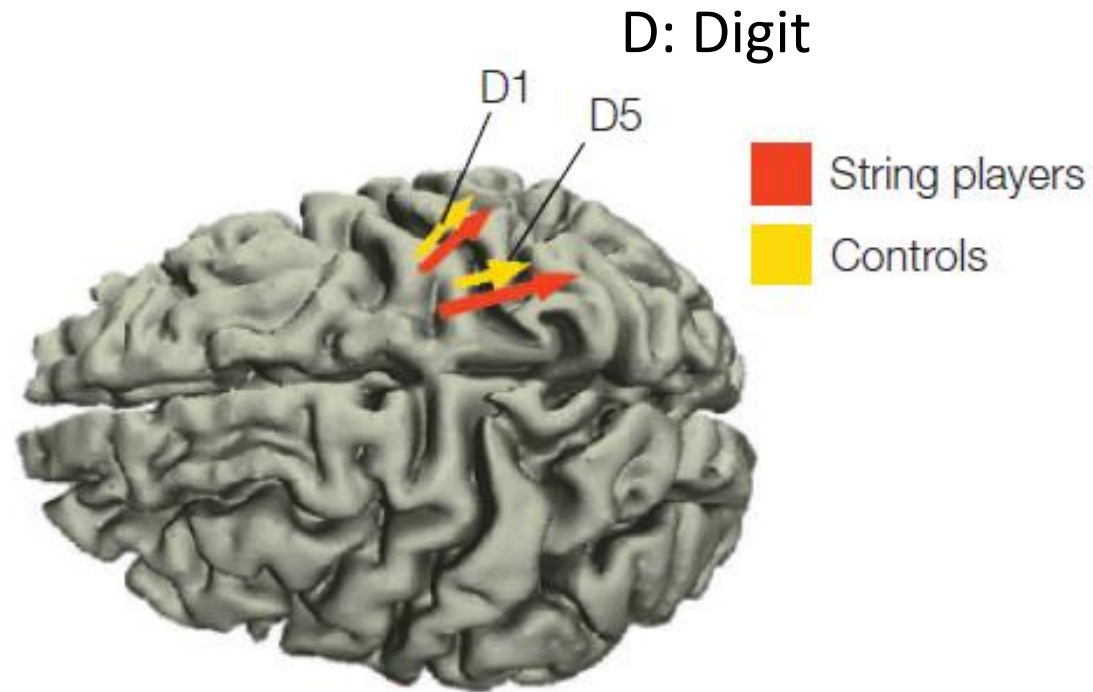


# Digit representation follows a common plan



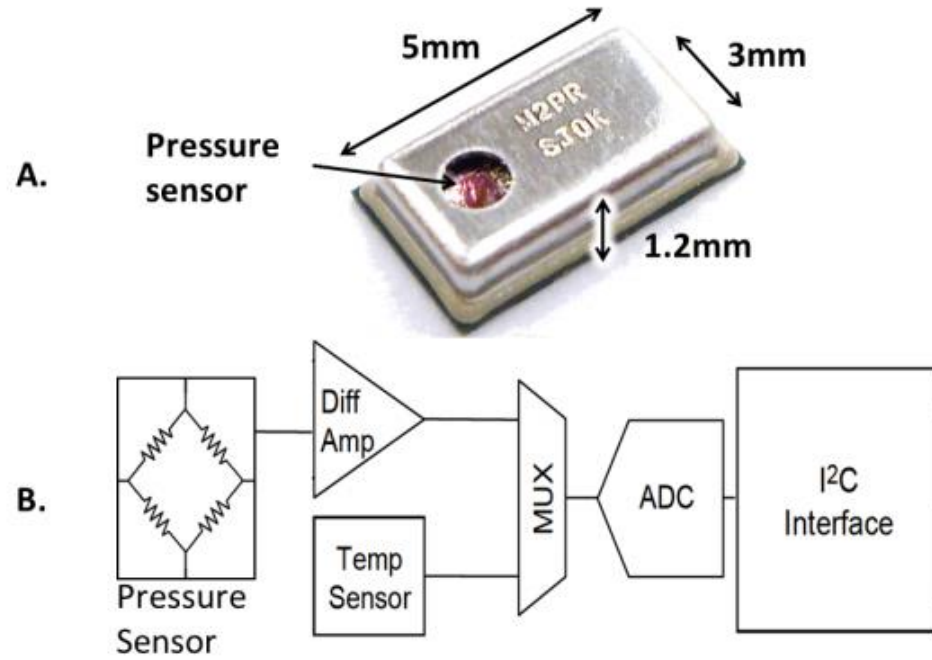


# Plasticity in the somatosensory cortex

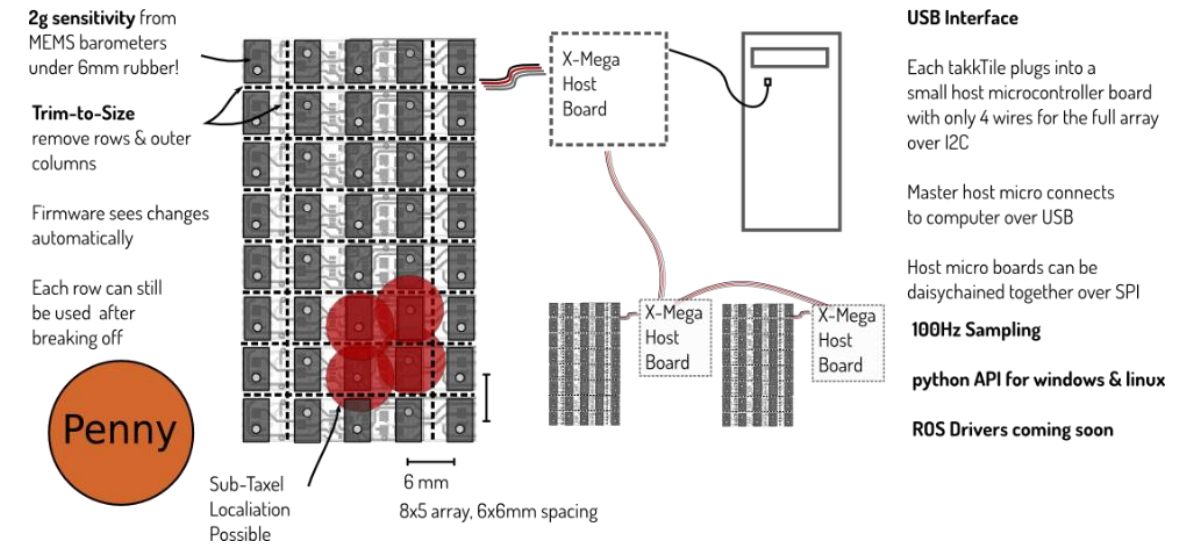


Elbert et al., 1995

# Tactile sensing in robots



## MEMS barometer



<http://www.takktile.com/main:plans>

## Inexpensive and Easily Customized Tactile Array Sensors using MEMS Barometers Chips

Yaroslav Tenzer, Leif P. Jentoft, Robert D. Howe  
Harvard School of Engineering and Applied Sciences



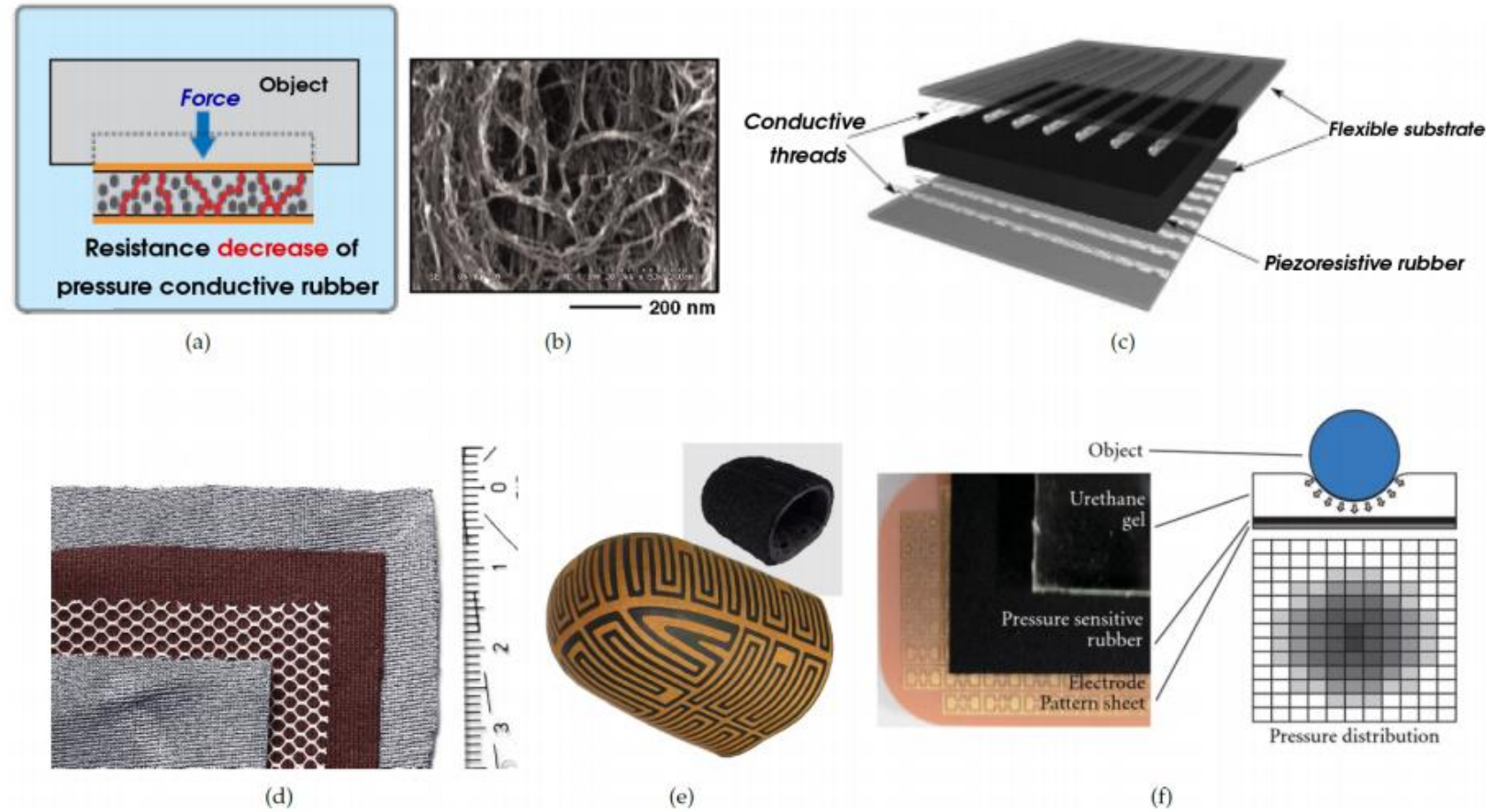
# Tactile sensing in robots

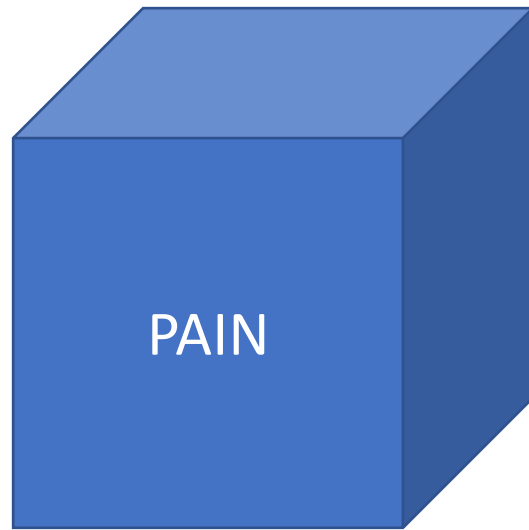
## Tactile sensing in dexterous robot hands – review

Zhanat Kappasov<sup>a,\*</sup>, Juan-Antonio Corrales<sup>b</sup>, Véronique Perdereau<sup>a</sup>

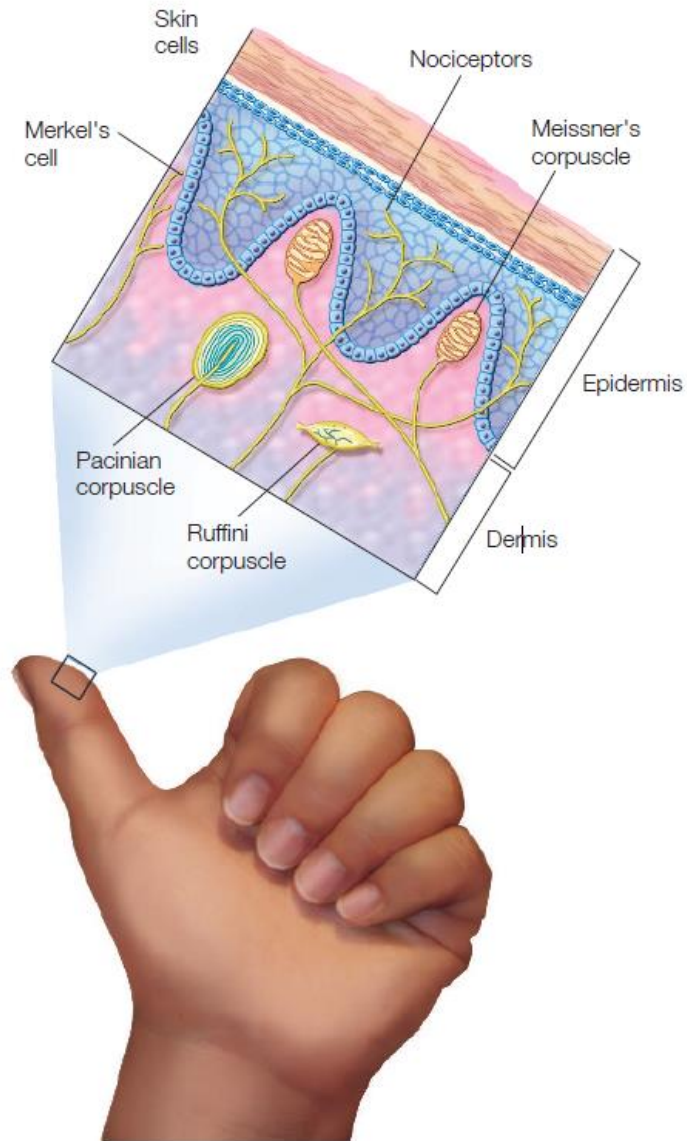
<sup>a</sup>Institute of Intelligent Systems and Robotics, University of Pierre and Marie Curie, CC 173 - 4 Place Jussieu 75005, Paris, France

<sup>b</sup>Institut Français de Mécanique Avancée, Campus de Clermont-Ferrand les Cezeaux BP265 63175 AUBIERE Cedex, France





# Somatosensation



# Pain

PAIN PERCEPTION IS CONTEXT  
AND INDIVIDUAL DEPENDENT

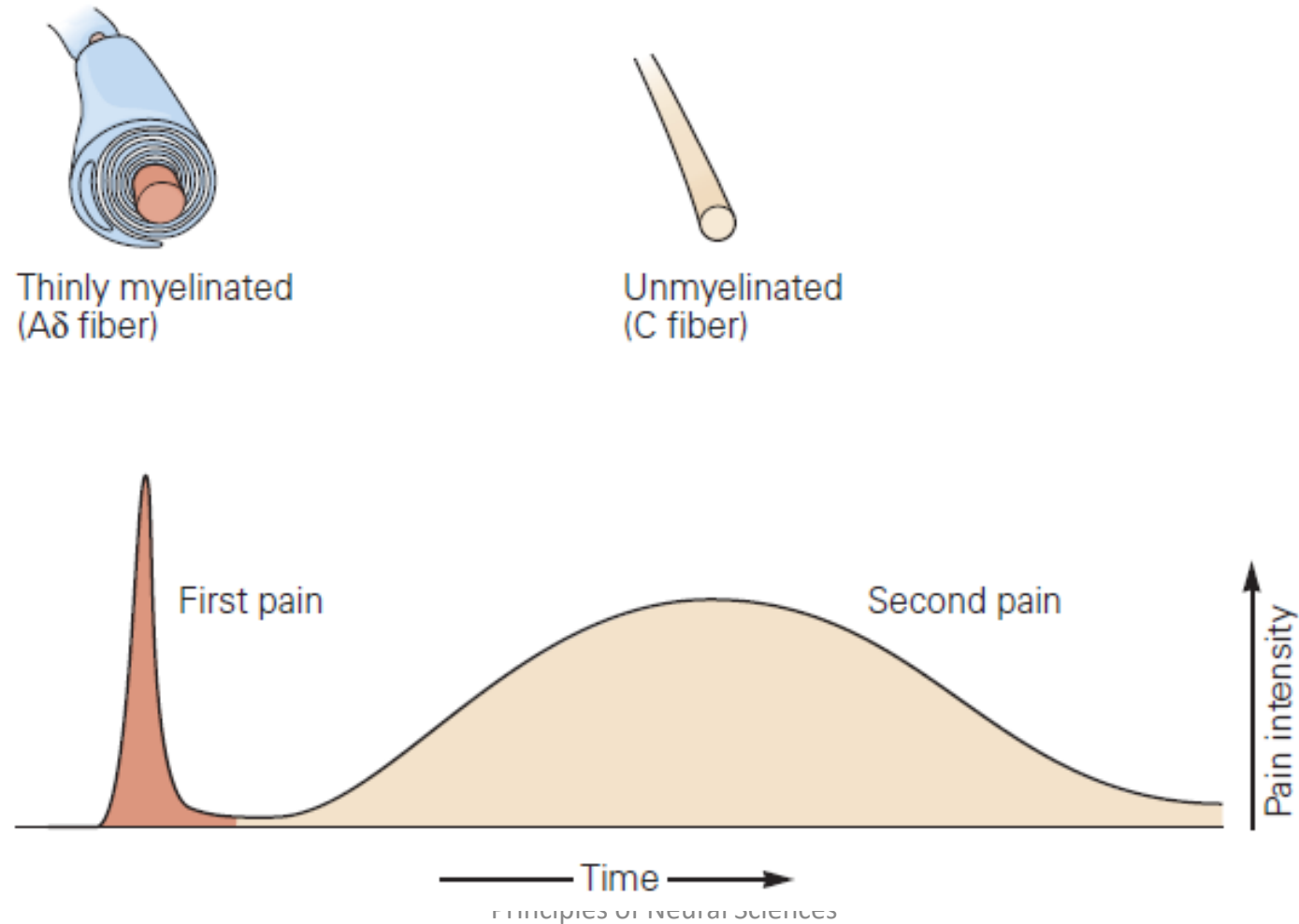
ACUTE

PERSISTANT

CHRONIC

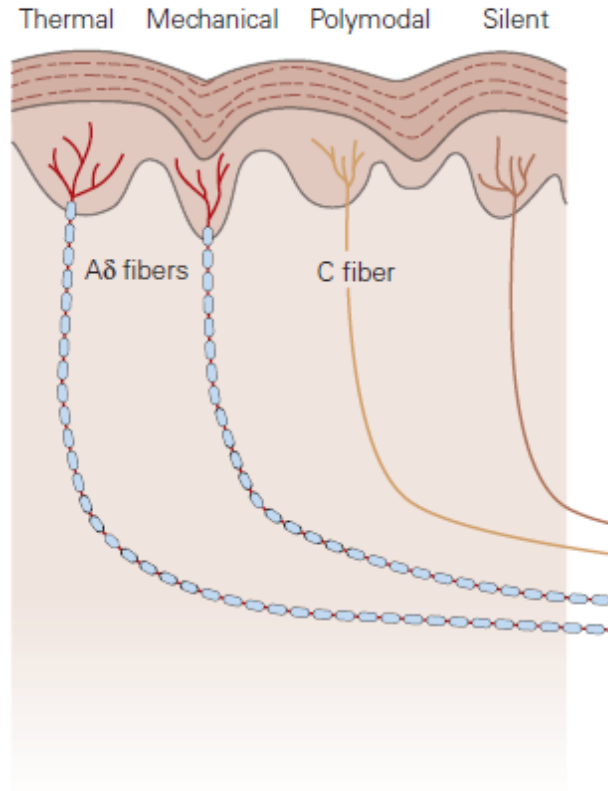
# How pain is elicited

## B First and second pain

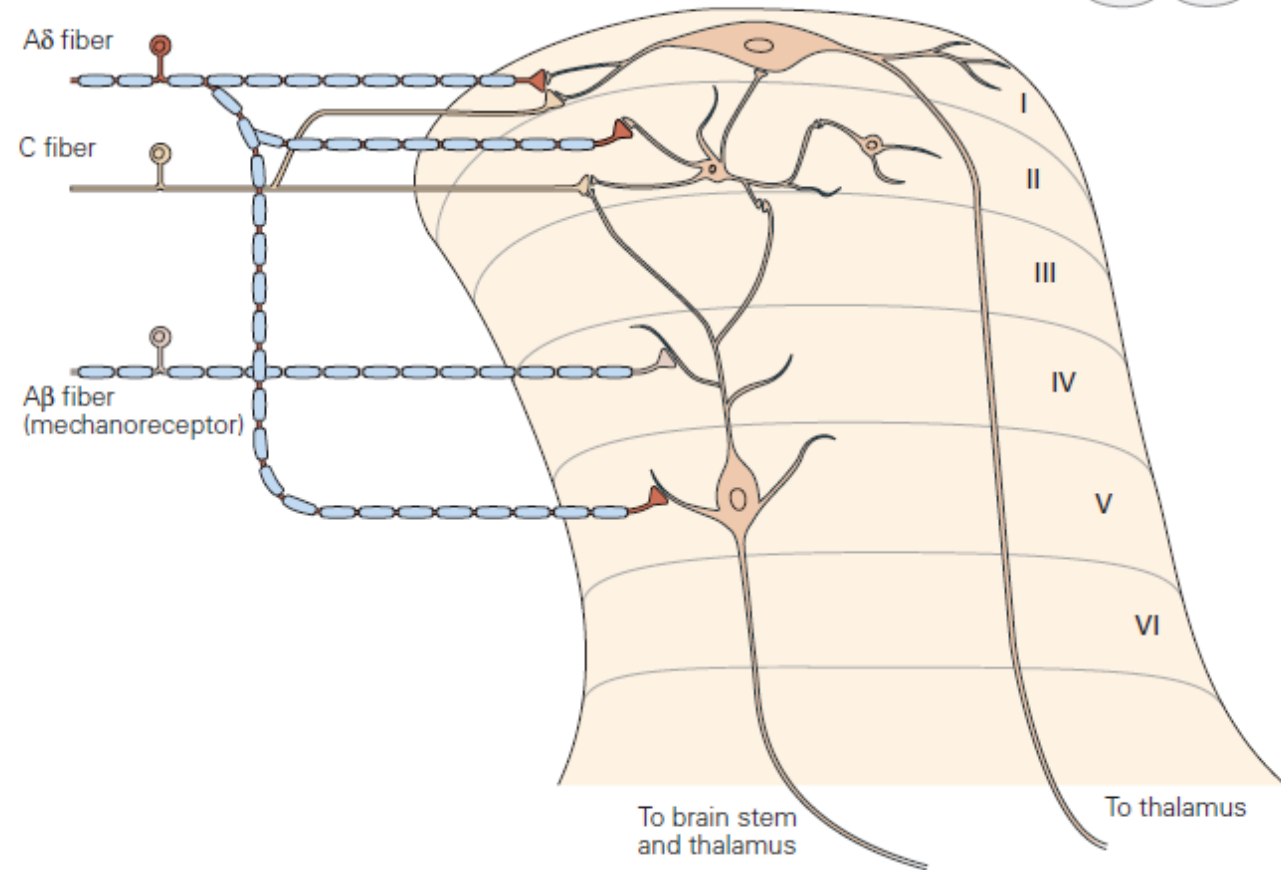


# Nociceptor types

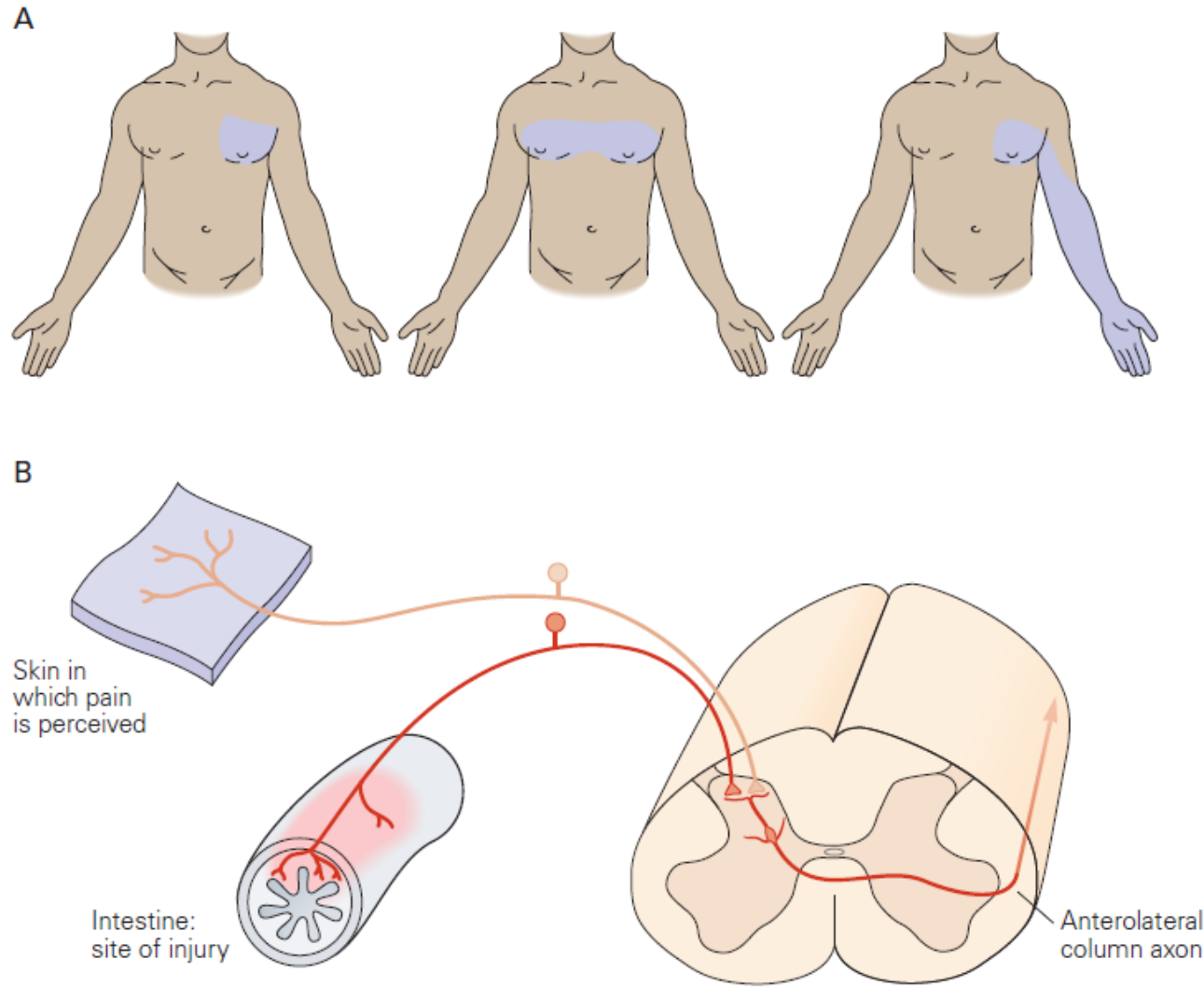
**A** Nociceptor types



**B** Spinal cord inputs

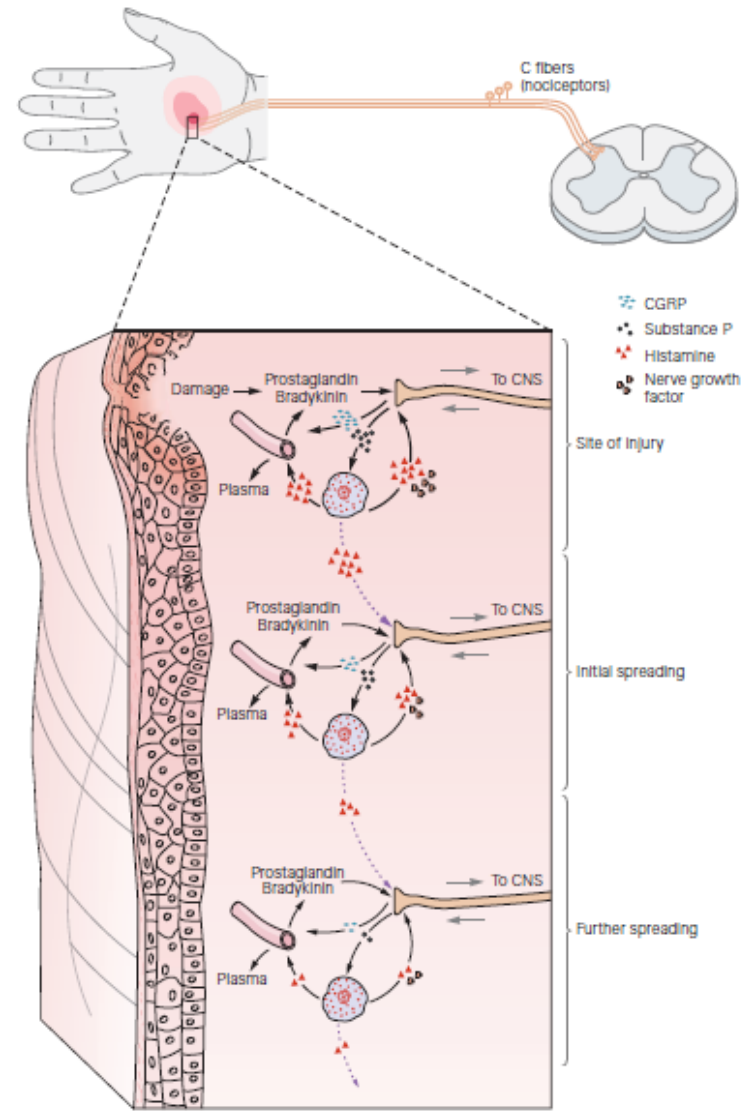


# Referred pain

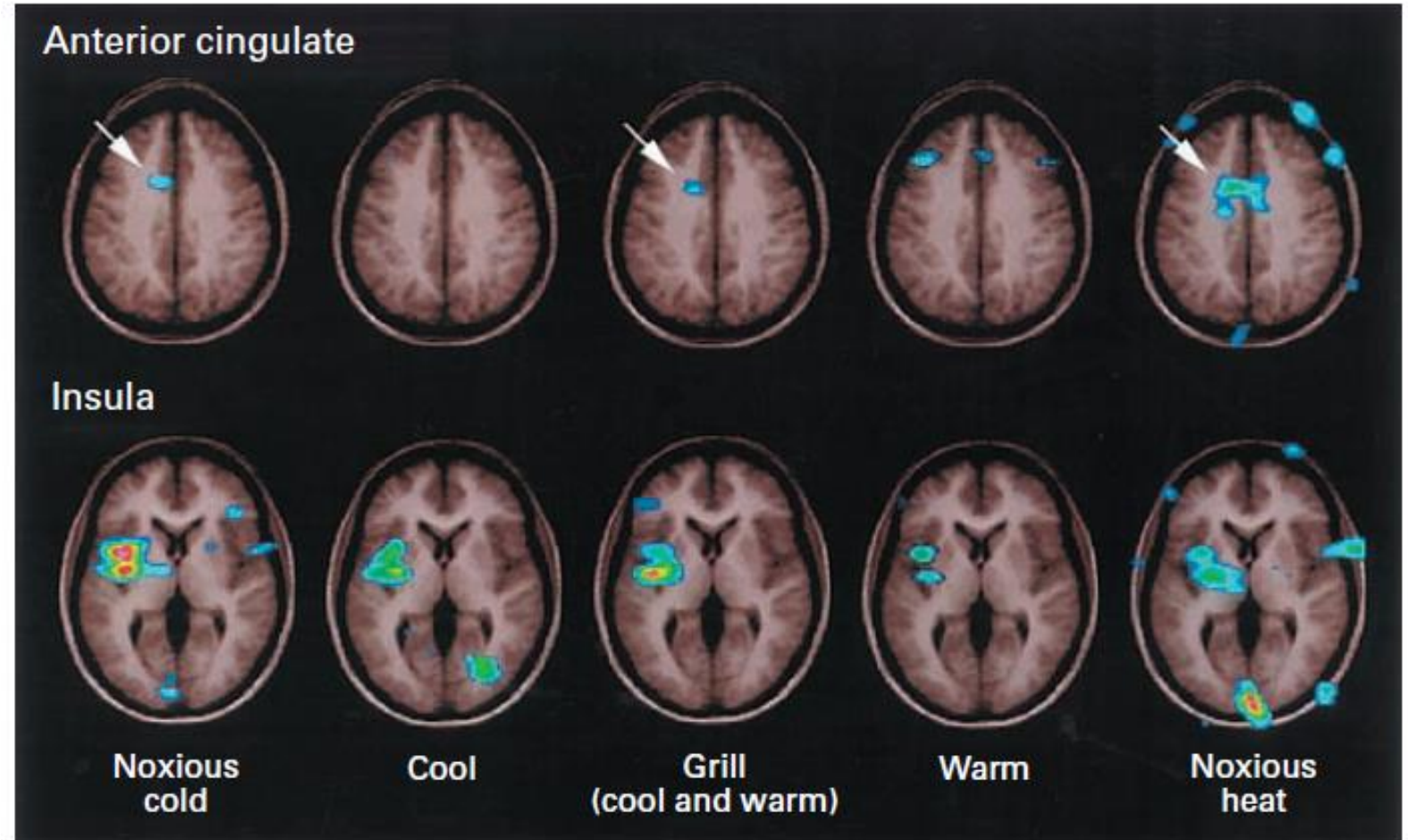
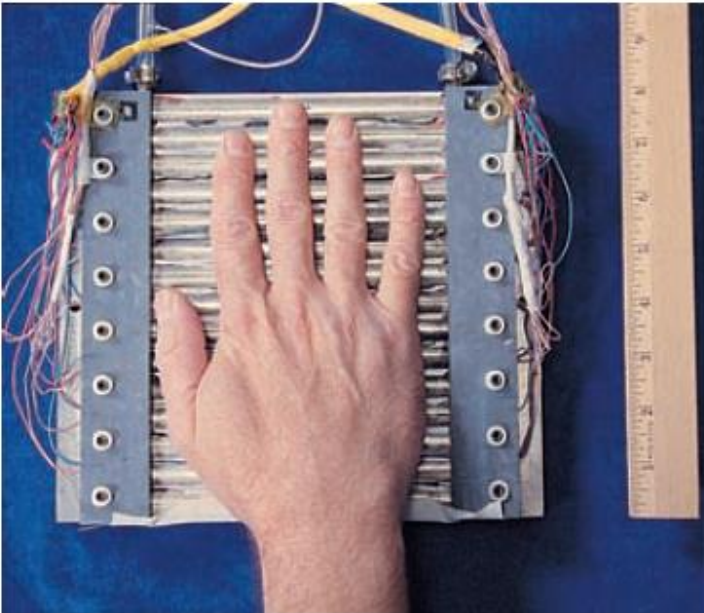




# Neurogenic inflammation



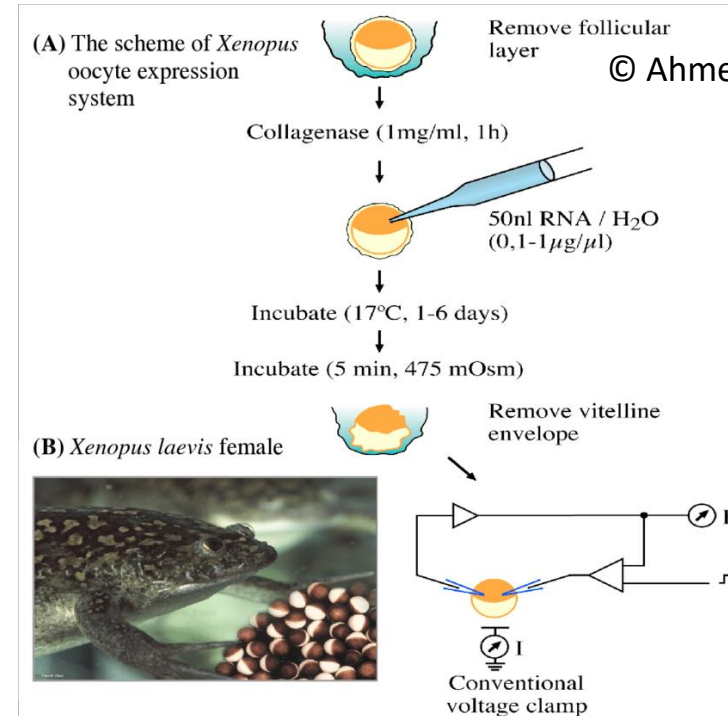
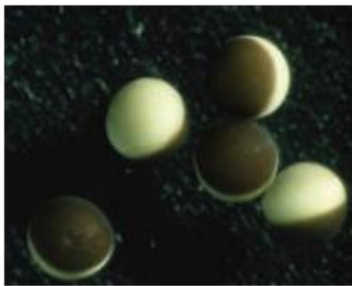
# Thunberg's illusion



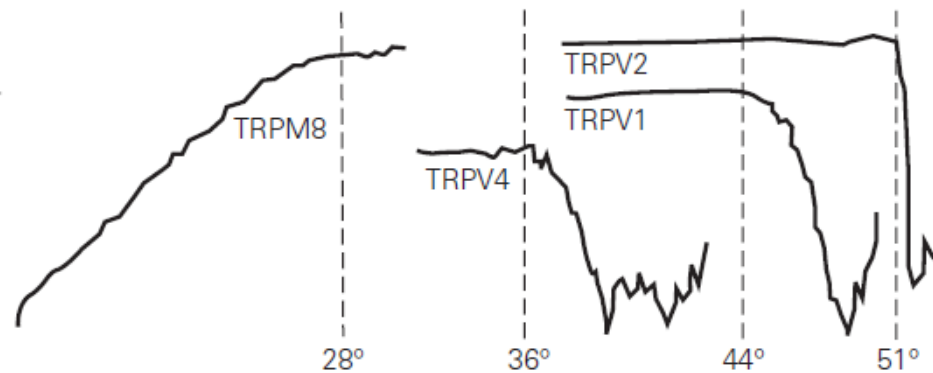
# Electrophysiology in *Xenopus* oocytes



A Thermosensitivity of TRP channels in *Xenopus* oocytes

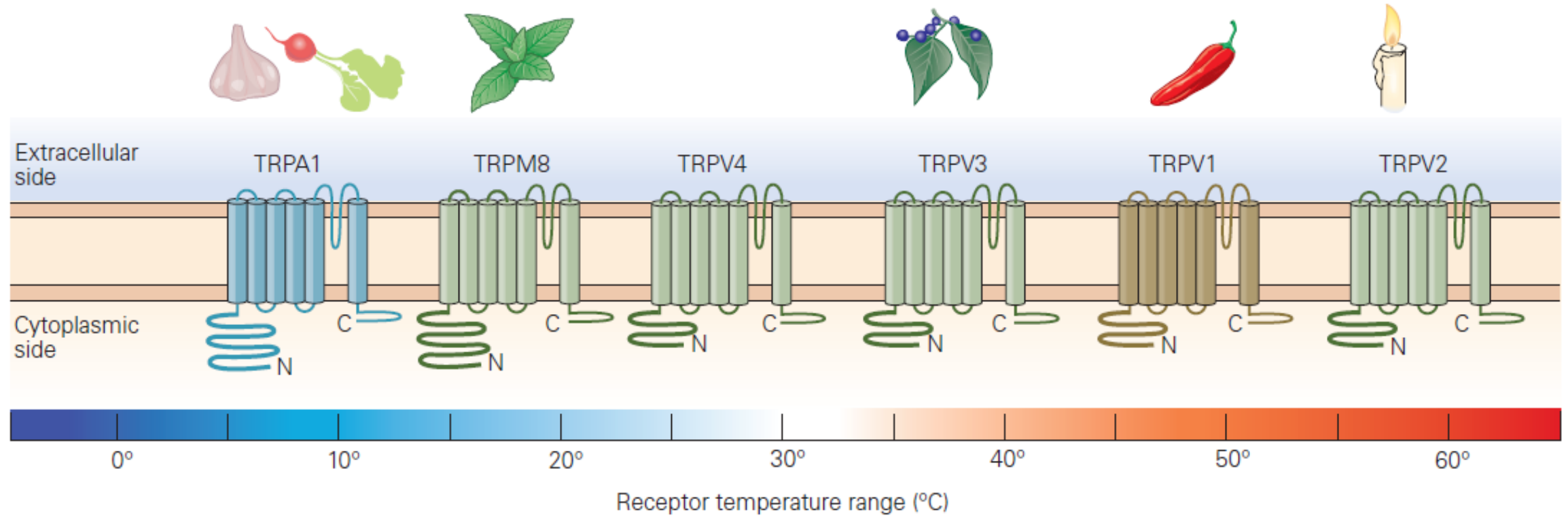


© Ahmed Al-Sabi



# Thermosensitivity of TRP channels

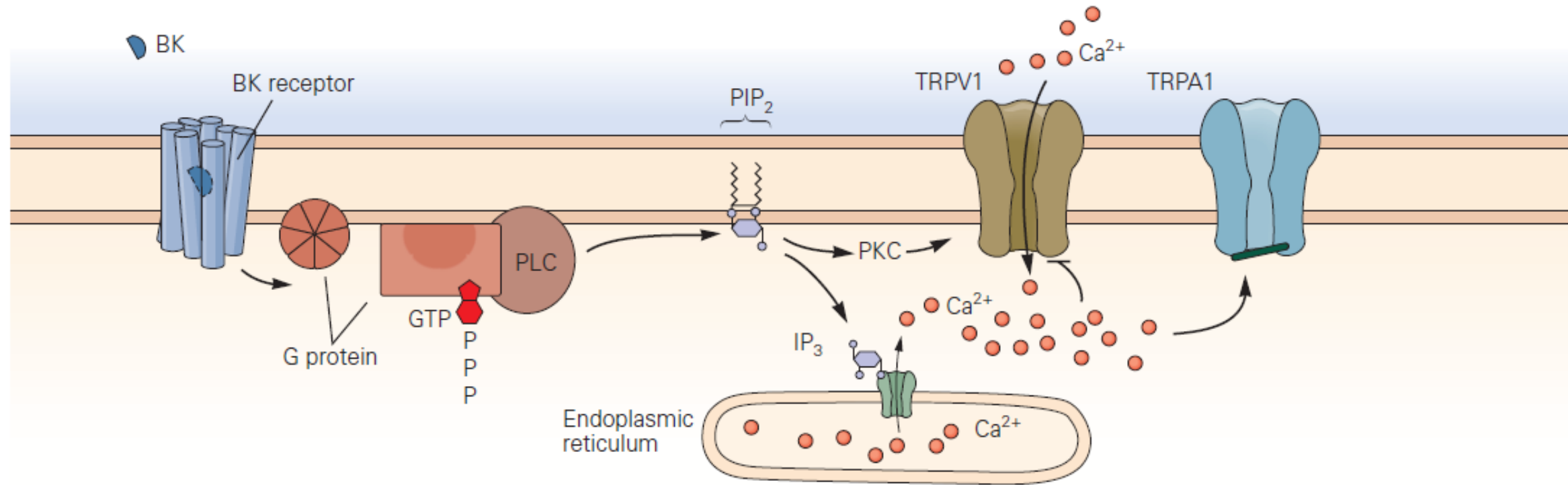
## B Thermosensitivity of TRP channels in dorsal root ganglion cells



# Pathways

## Bradykinin

C Pathway to TRP channel opening




## Sensitizing TRP channels



# Navs and cold perception

Published: 14 June 2007

## Sensory neuron sodium channel $\text{Na}_v1.8$ is essential for pain at low temperatures

Katharina Zimmermann , Andreas Leffler, Alexandru Babes, Cruz Miguel Cendan, Richard W. Carr, Jin-ichi Kobayashi, Carla Nau, John N. Wood & Peter W. Reeh

*Nature* **447**, 856–859 (2007) | [Cite this article](#)

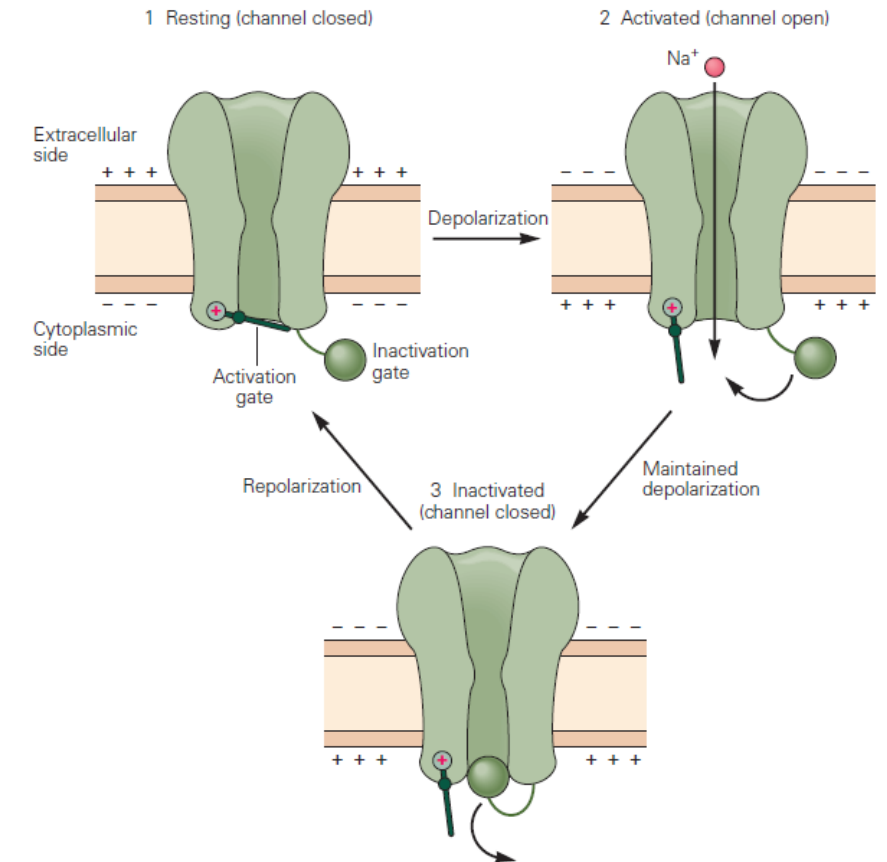
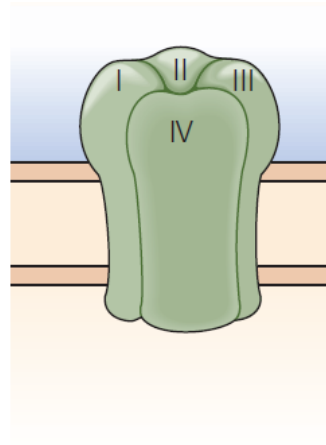
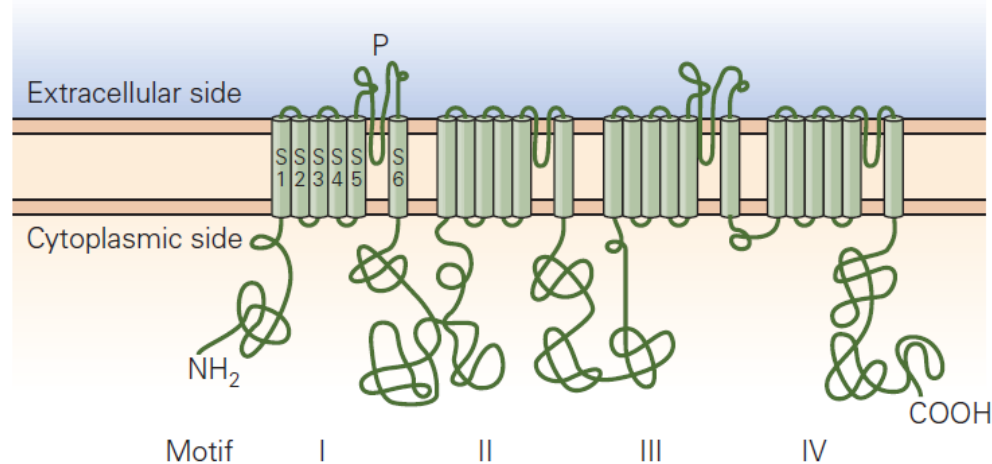
**1035** Accesses | **276** Citations | **10** Altmetric | [Metrics](#)



# Congenital insensitivity to pain (CIP)

Very often defects in SCN9A (Nav1.7)

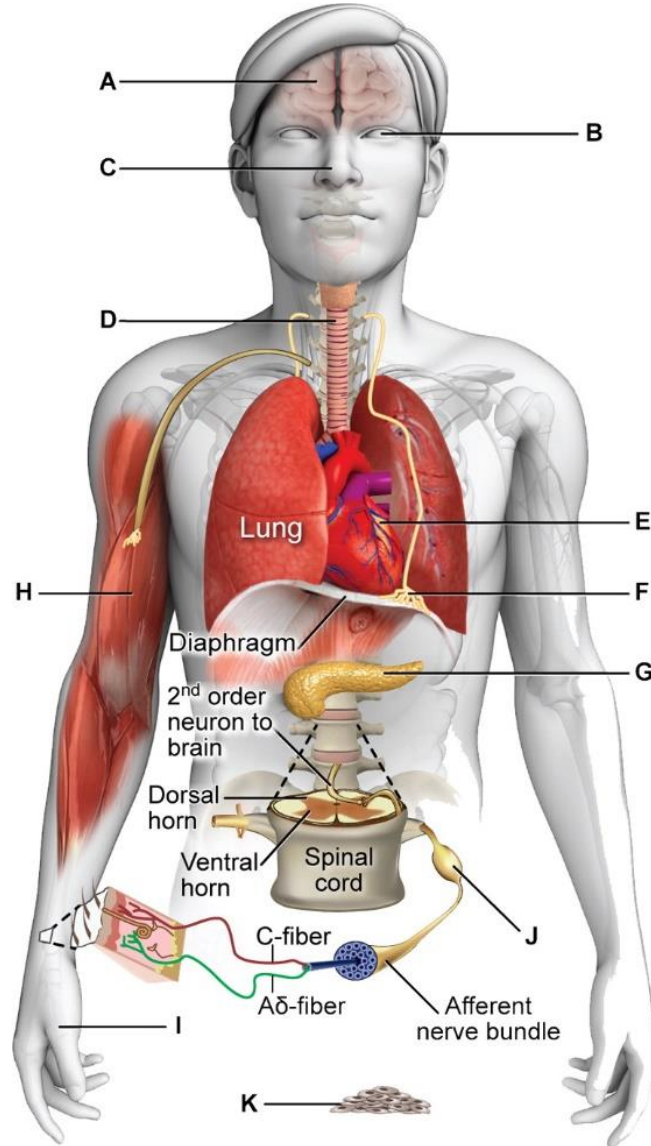
C Voltage-gated channel ( $\text{Na}^+$  channel)



→ Lack of Nav1.7 leads to no pain experience

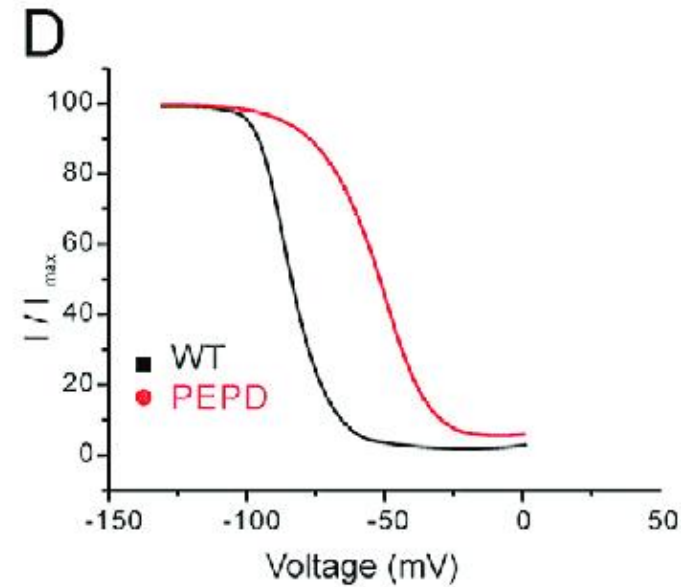
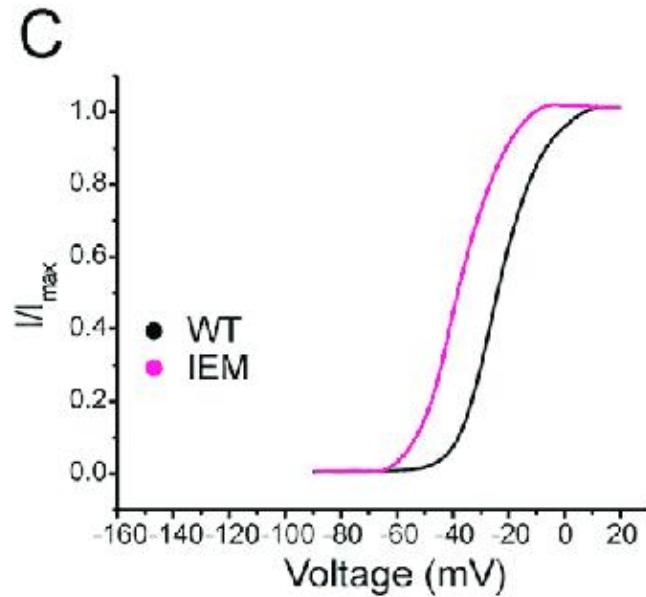
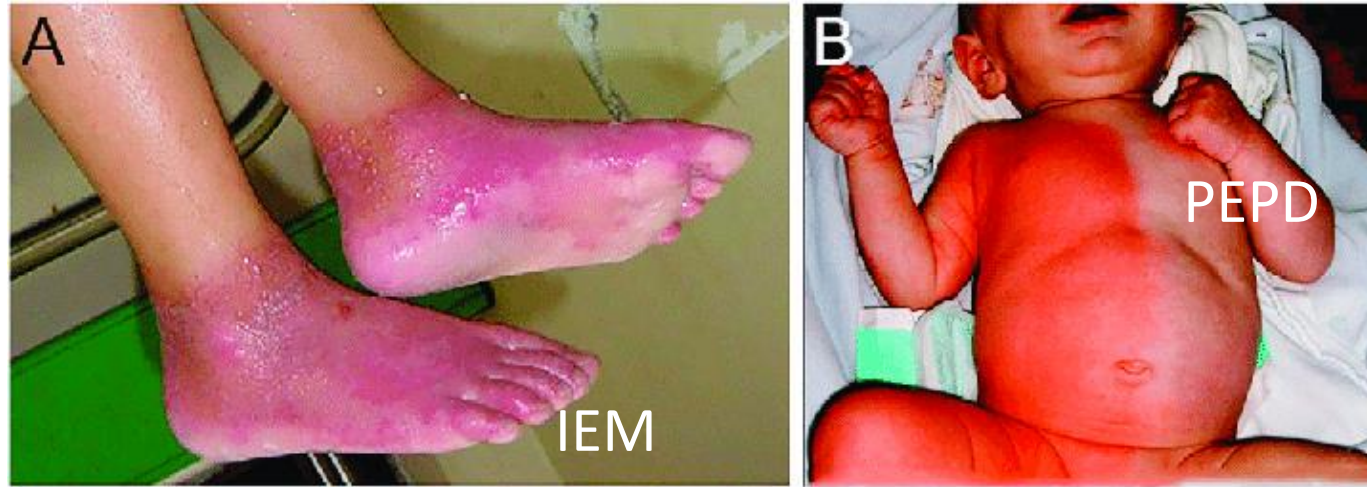


# Voltage-gated sodium channels



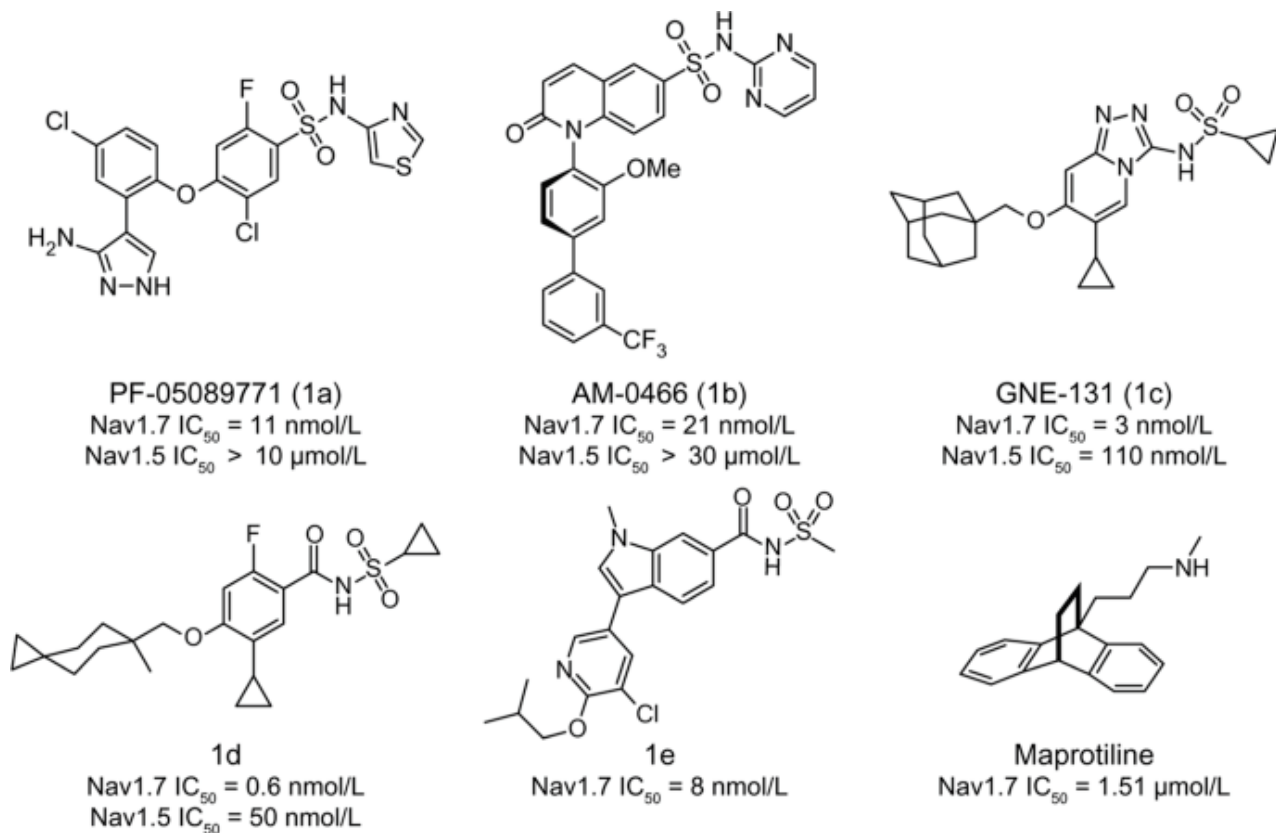
	Tissue	Na <sub>v</sub> Subtype	Effect of Na <sub>v</sub> Dysfunction on Physiology
<b>A</b>	Central nervous system	1.1, 1.2, 1.3, 1.6	Epilepsy, migraine, autism, ataxia <sup>36</sup>
<b>B</b>	Retina	1.8, 1.9	Altered visual processing <sup>62</sup>
<b>C</b>	Olfactory sensory neurons	1.7	Anosmia <sup>40,42</sup>
<b>D</b>	Sensory neurons and vagal sensory neurons innervating airways	1.7, 1.8, 1.9	Cough <sup>36,68</sup>
<b>E</b>	Heart muscle	1.5, 1.8	Brugada syndrome, QT syndrome, atrial fibrillation <sup>66,67</sup>
<b>F</b>	Nerves, musculature involved in ventilation	TTX-s Na <sub>v</sub> s	Respiratory cessation (TTX poisoning) <sup>69</sup>
<b>G</b>	Pancreatic $\beta$ -cells	1.7	Diabetes <sup>36</sup>
<b>H</b>	Skeletal muscle	1.4	Hyperkalaemic periodic paralysis, paramyotonia congenita, hypokalaemic periodic paralysis <sup>36</sup>
<b>I</b>	Skin	1.7, 1.8	Pain disorders, paroxysmal itch <sup>37,39</sup>
<b>J</b>	DRG neurons	1.6, 1.7, 1.8, 1.9	Pain disorders, paroxysmal itch <sup>37,39,51</sup>
<b>K</b>	Metastatic cancer cells	1.1-1.9 and $\beta$ -subunits	Ovarian, cervical, prostate, breast, colon, small cell lung cancer, melanoma, lymphoma <sup>35,70,71</sup>

# Hyperexcitability of Nav1.7



Waxman and Dib-Hajj, 2019

# Pain killer development for Nav1.7

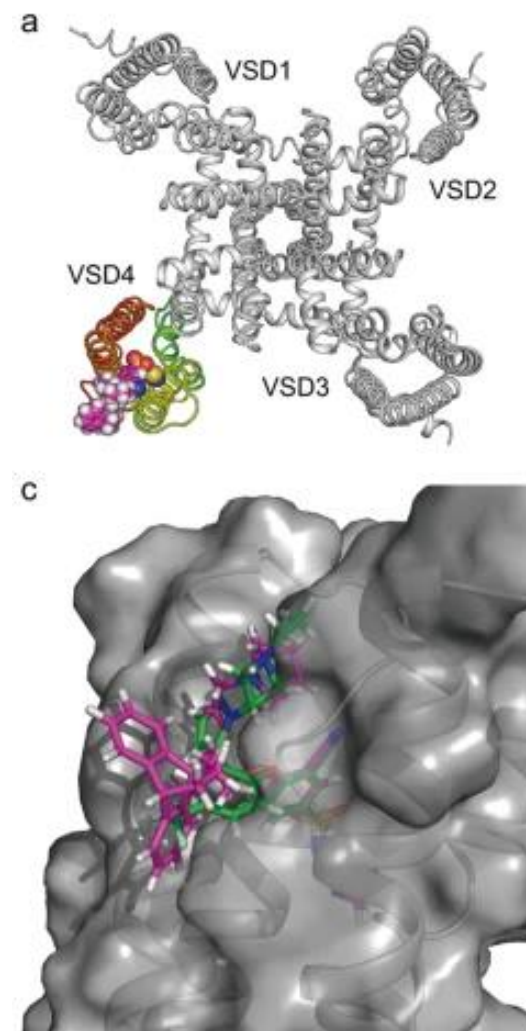


Article | Published: 17 July 2019

## Discovery of aryl sulfonamide-selective Nav1.7 inhibitors with a highly hydrophobic ethanoanthracene core

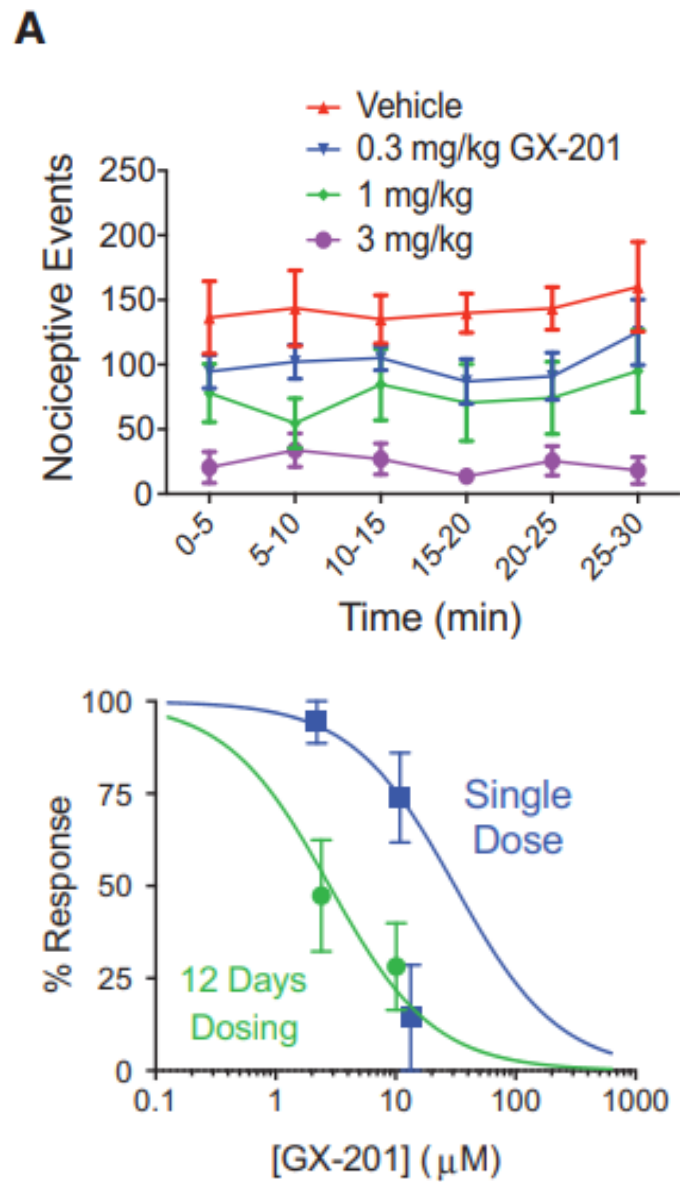
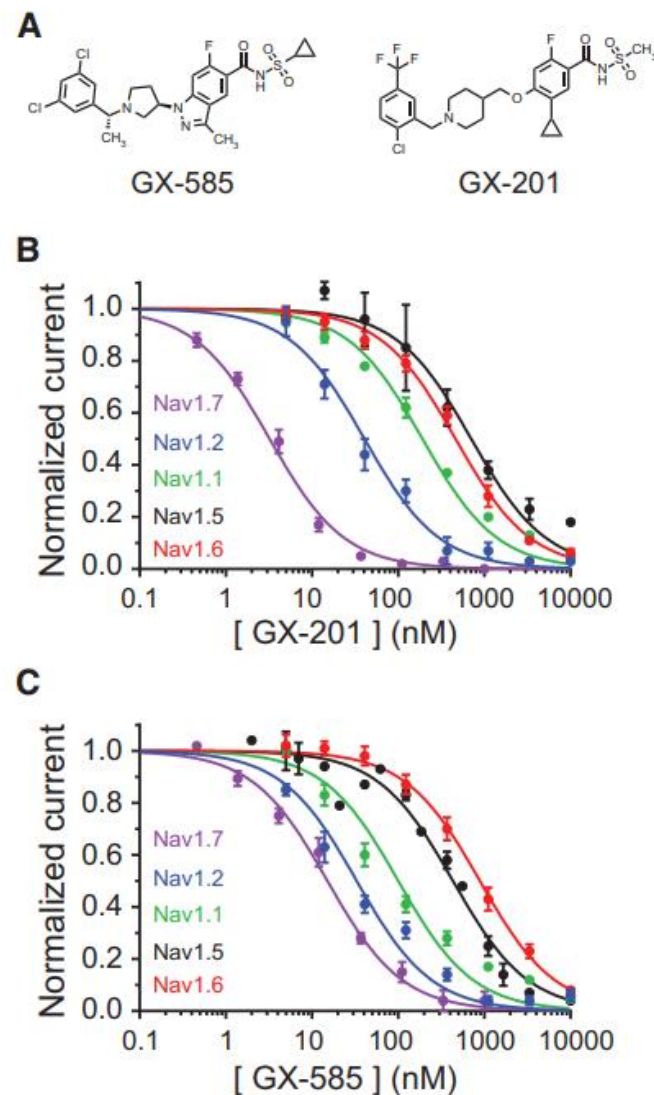
Jin-tao Wang, Yue-ming Zheng, Yue-ting Chen, Min Gu, Zhao-bing Gao & Fa-jun Nan

*Acta Pharmacologica Sinica* **41**, 293–302 (2020) | Cite this article





# Pain killer II



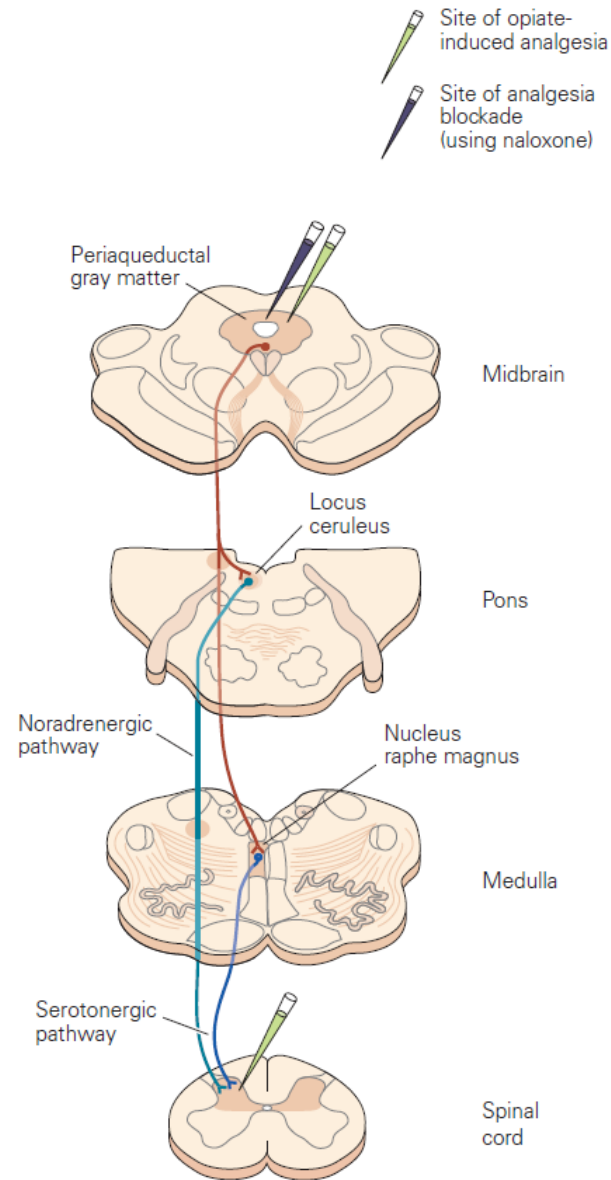
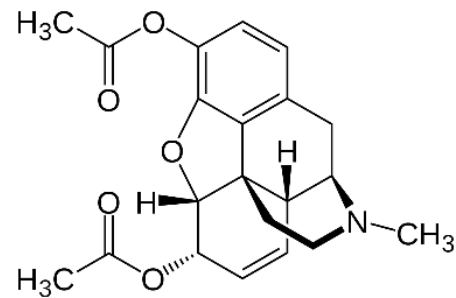
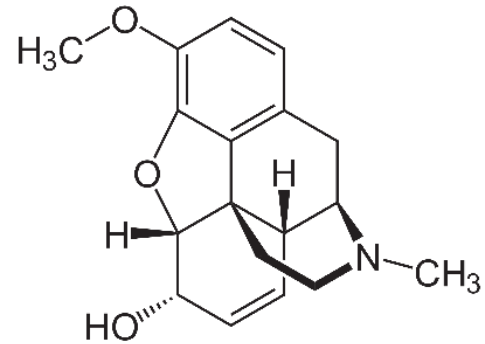
Cell Reports  
Article

## Selective Nav<sub>v</sub>1.7 Antagonists with Long Residence Time Show Improved Efficacy against Inflammatory and Neuropathic Pain

Girish Bankar,<sup>1</sup> Samuel J. Goodchild,<sup>1</sup> Sarah Howard,<sup>1</sup> Karen Nelkenbrecher,<sup>1</sup> Matthew Waldbrook,<sup>1</sup> Michelle Dourado,<sup>2</sup> Noah G. Shuart,<sup>1</sup> Sophia Lin,<sup>1</sup> Clint Young,<sup>1</sup> Zhiwei Xie,<sup>1</sup> Kuldip Khakh,<sup>1</sup> Elaine Chang,<sup>1</sup> Luis E. Sojo,<sup>1</sup> Andrea Lindgren,<sup>1</sup> Sultan Chowdhury,<sup>1</sup> Shannon Decker,<sup>1</sup> Michael Grimwood,<sup>1</sup> Jean-Christophe Andrez,<sup>1</sup> Christoph M. Dehnhardt,<sup>1</sup> Jodie Pang,<sup>2</sup> Jae H. Chang,<sup>2</sup> Brian S. Safina,<sup>2</sup> Daniel P. Sutherlin,<sup>2</sup> James P. Johnson, Jr.,<sup>1</sup> David H. Hackos,<sup>2</sup> C. Lee Robinette,<sup>1</sup> and Charles J. Cohen<sup>1,3,\*</sup>

<sup>1</sup>Xenon Pharmaceuticals, Burnaby, BC V5G 4W8, Canada  
<sup>2</sup>Genentech, South San Francisco, CA 94080, USA  
<sup>3</sup>Lead Contact  
\*Correspondence: ccohen@xenon-pharma.com  
<https://doi.org/10.1016/j.celrep.2018.08.063>

# Opioids as pain killer



## 2 Sensory input + opiates/opioids

