

Converting Raw LC-HRMS/MS Files into mzML files

Benjamin J. Place*

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Converting To The mzML Format

To use DIMSpec tools, raw data files produced by vendor software must be converted into *.mzML files. The easiest way to convert data files is to download and install the most recent version of ProteoWizard¹ from <https://proteowizard.sourceforge.io/> to use the MSConvert tool (Adusumilli, Ravali and Mallick, Parag 2017). Once installed, follow the next steps to convert the raw file(s) to *.mzML format.

1) Start MSConvert

From the Proteowizard main page, start the MSConvert program with Menu > Proteowizard > MSConvert²

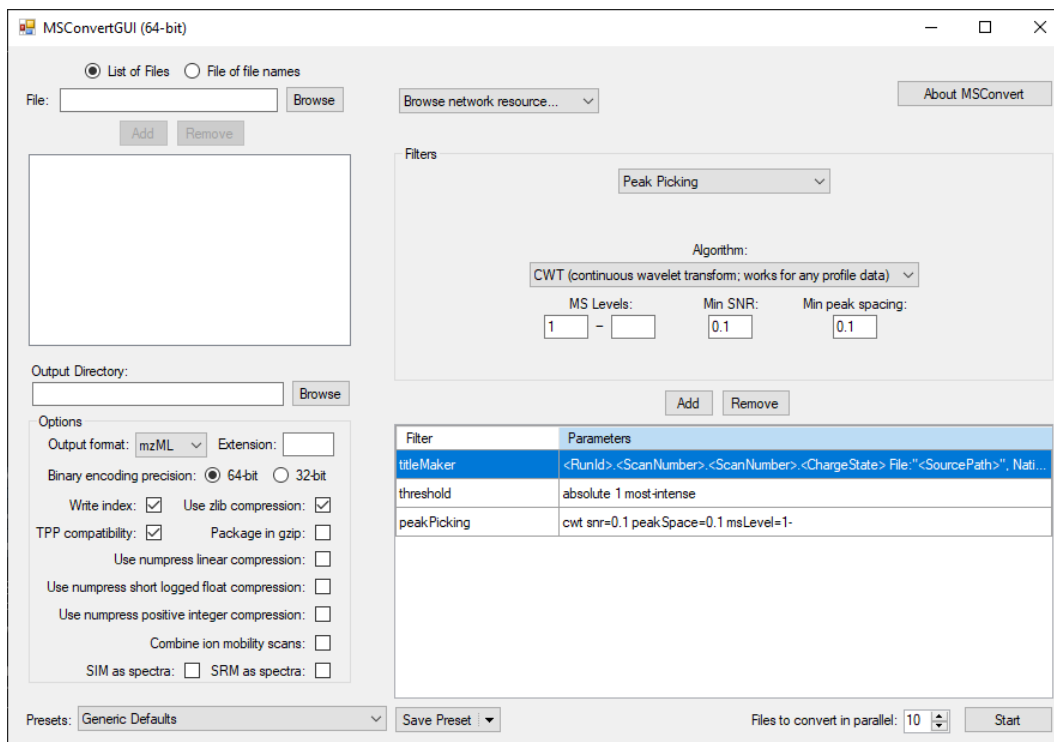


Figure 1: Initial screen when running MSConvert

*NIST | MML | CSD | benjamin.place@nist.gov

¹Any mention of commercial products is for information only; it does not imply recommendation or endorsement by NIST.

²MSConvert includes a command line interface for advanced use cases; settings must be the same as described in this guide.

2) Select the files to be converted

The MSConvert software can convert to mzML data files from the the following vendor formats at the time this guide was written: Thermo Fisher Scientific (*.raw); Waters (*.raw); SCIEX (*.wiff2); Agilent (*.D); Shimadzu (*.LCD); and Bruker (*.D)

1. Select the files to be converted using the **Browse** button at the top left (Fig 2A).

Note: If selecting only one file at a time, instead click the *Add* button to include the file in the list.

2. Select the *Output Directory* to save files by clicking the **Browse** button at the center left (Fig 2B).

Note: It will default to the same directory as the original file.

After loading the file, the program should look similar to Figure 2.

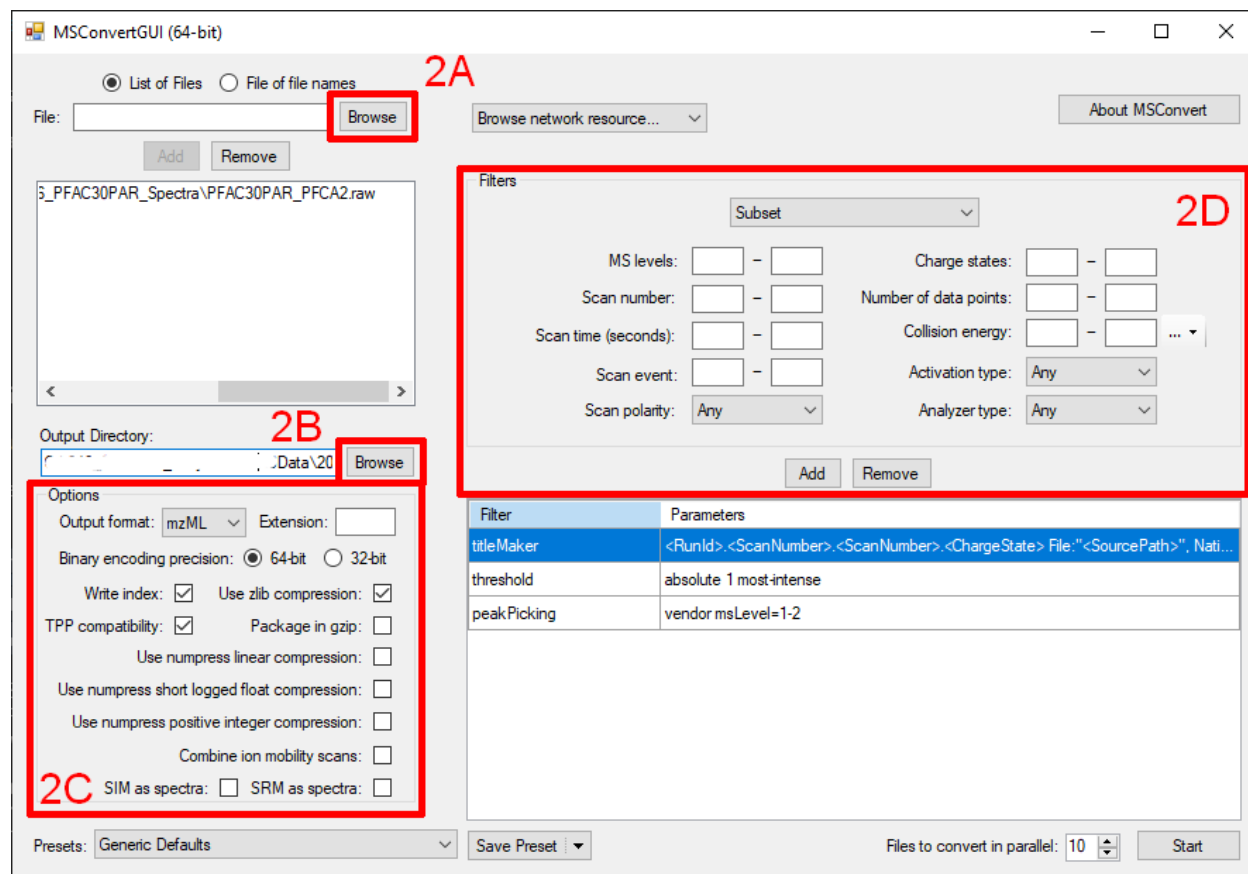


Figure 2: Interface for MSConvert after a data file has been loaded

3) Adjust the mzML file parameters in the Options box (Fig 2C).

1. Select **mzML** from the *Output format* input and make sure the *Extension* input is blank.
2. Select **64-bit** under *Binary encoding precision*.
3. Ensure boxes are checked next to *Write Index*, *TPP Compatibility*, and *Use zlib compression*; leave all other boxes unchecked.

The proper selections have been made in Figure 2.

4) Add conversion filters (Fig 2D)

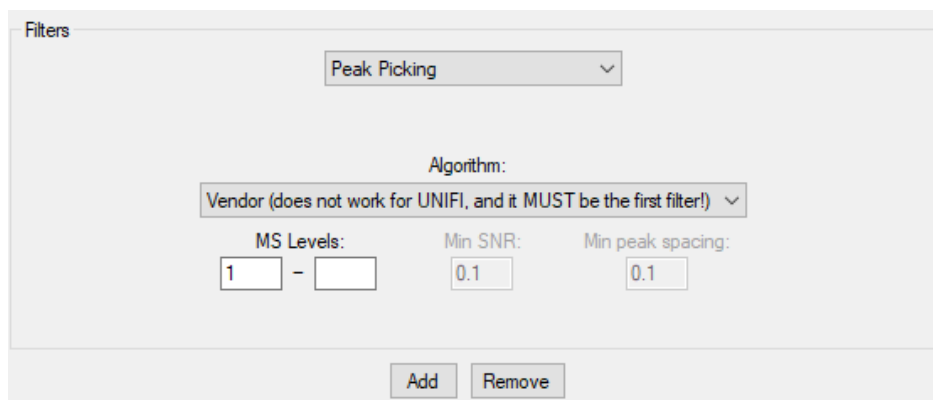
Apply the following data conversion filters; to adhere to current NIST practice the following settings *must* be used. Select the filter to be applied from the drop down menu at the top of the **Filters** box. Do this sequentially as described here. After setting each parameter click the **Add** button. To remove a parameter (in case of an entry or selection error), select that filter in the table below Fig 2D and click the **Remove** button.

1. Select the *Peak Picking* filter and apply the following parameters:

- **Algorithm:** Vendor or CWT
Note: Vendor filter does not work for Waters files, you must use CWT
- **MS Levels:** 1 - __ (leave the second box blank)
- If using CWT:
 - **Min. SNR:** 0.1
 - **Min. peak spacing:** 0.1

Parameters should match those in Figure 3.

Click the **Add** Button



The screenshot shows a 'Filters' window with a 'Peak Picking' filter selected. The 'Algorithm' dropdown is set to 'Vendor (does not work for UNIFI, and it MUST be the first filter!)'. The 'MS Levels' are set to '1' and an empty box. The 'Min SNR' is set to '0.1' and the 'Min peak spacing' is set to '0.1'. 'Add' and 'Remove' buttons are at the bottom.

Figure 3: Peak picking filter parameters

2. Select the *Threshold Peak Filter* and include the following parameters:

- **Threshold type:** Absolute intensity
- **Orientation:** Most intense
- **Value:** 1

Parameters should match those in Figure 4 (next page)

Click the **Add** Button

The screenshot shows a dialog box titled "Filters". Inside, there is a dropdown menu set to "Threshold Peak Filter". Below this, there are three settings: "Threshold type:" set to "Absolute intensity", "Orientation:" set to "Most intense", and "Value:" set to "1". At the bottom of the dialog are two buttons: "Add" and "Remove".

Figure 4: Threshold peak filter parameters

- If the raw data is from a **Waters Corporation** mass spectrometer, apply the *Lockmass Refiner* filter.
 - **Reference m/z :** use the mass-to-charge ratio of the lockmass, for Leucine-Enkephalin (most common), the negative ionization m/z is 554.2615 and the positive ionization m/z is 556.2771.
 - **m/z Tolerance:** 0.1

Parameters should match those in Figure 5

Click the **Add** Button

The screenshot shows a dialog box titled "Filters". Inside, there is a dropdown menu set to "Lockmass Refiner". Below this, there are two settings: "Reference m/z :" set to "554.2615" and " m/z Tolerance:" set to "0.1". At the bottom of the dialog are two buttons: "Add" and "Remove".

Figure 5: Lockmass refiner parameters

Selected filter parameters should be very similar to those in Figure 2 prior to conversion to *.mzML.

Note: The TitleMaker filter is automatically selected upon opening MSConvert; this can be included and will not affect the data analysis. However, peakPicking must still be the first line in the filters.

Note: Waters Corporation instruments should have the lockmass refiner filter in the as well.

5) Convert data files to the *.mzML format.

Once all settings are as required, click the **Start** button at the bottom right (Fig 2E). This will convert all added files to *.mzML format and put them in the assigned Output directory. Wait for the conversions to finish.

6) (Optional) Save the settings as a preset

Use the **Save Preset** drop down menu to use the same settings in the future (Fig 6) by selecting *Save Preset As...* To apply a preset profile to future files, select it from this same drop down menu.

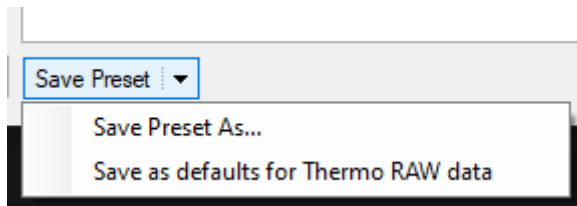


Figure 6: MSConvert presets menu

Once all files have been converted to the *.mzML format they are ready for use in other data tools.

This concludes the Quick Guide to Converting Raw LC-HRMS/MS Files into mzML files.

References

Adusumilli, Ravali and Mallick, Parag. 2017. "Data Conversion with ProteoWizard msConvert." *Methods in Molecular Biology (Clifton, N.J.)* 1550: 339–68. https://doi.org/10.1007/978-1-4939-6747-6_23.