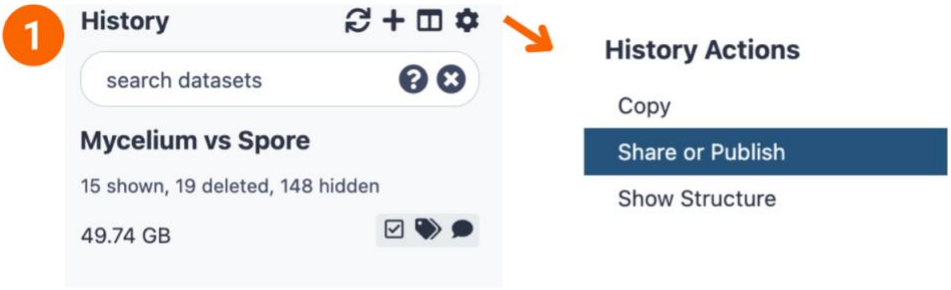
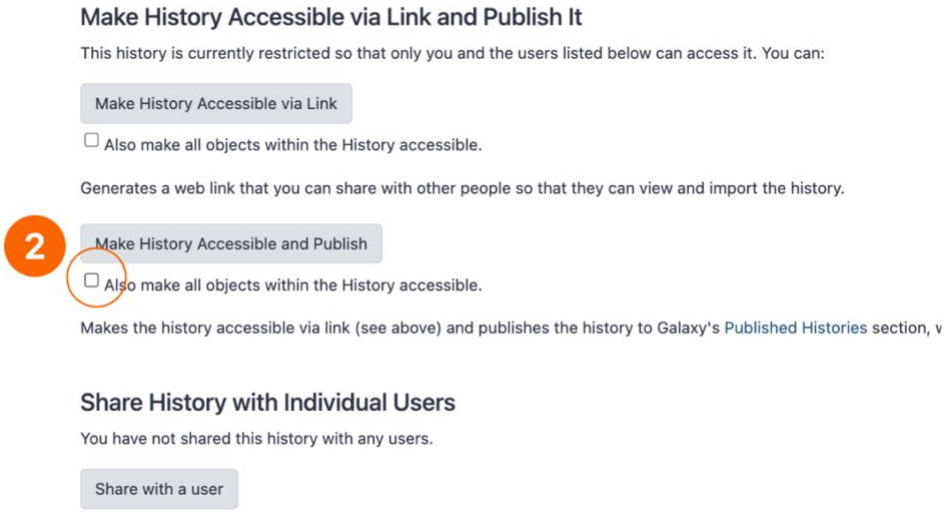
Variant Calling analysis, Part 2: Analyzing results (Group Exercise)

Learning objectives:

- Share and publish your workflow histories.
- Examine the outputs.
- View VCF files in JBrowse.
- Examine the filtered VFC file, extract Gene IDs, and create a Venny diagram.

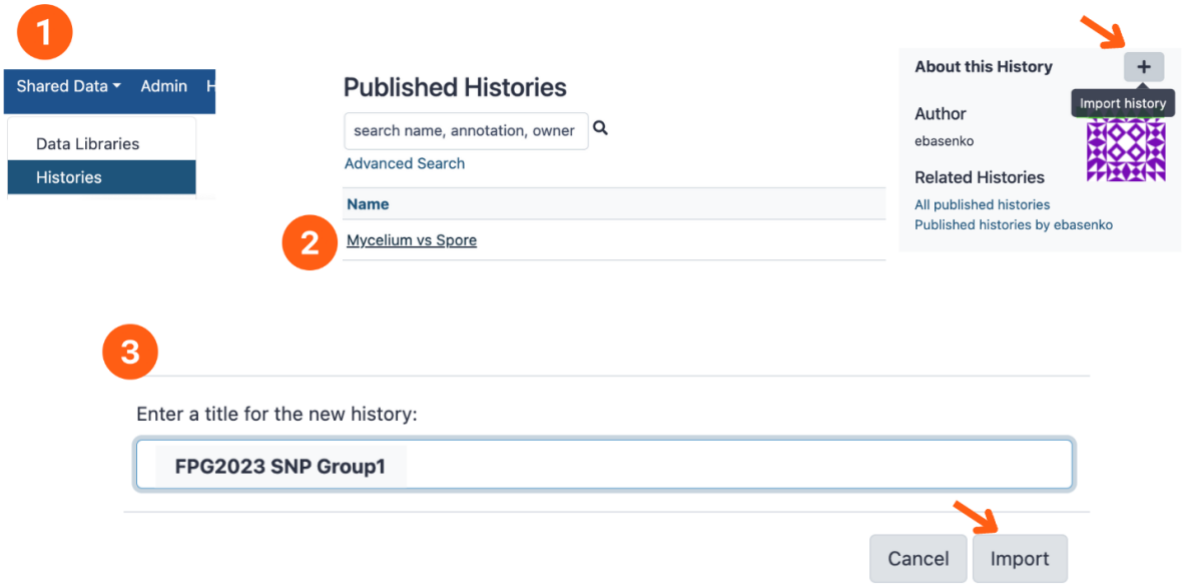
- **Share workflow histories with others.**
    1. Make sure your history has a useful name (e.g., Group3 SNPs, etc.) and click on the history action menu icon.
    2. Select the "Make History Accessible and Publish" option and check to make sure that all objects within History are accessible.





- **Importing workflow histories and output files into your own Galaxy workspace.**

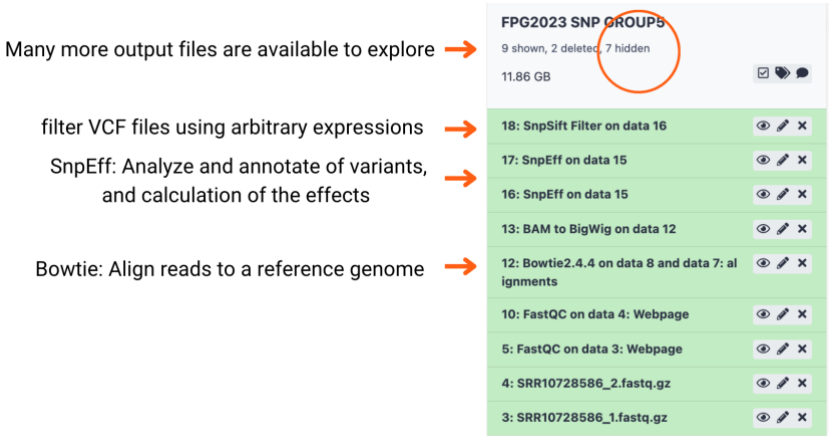
    1. Click on "Shared Data" at the top and select "Histories".
    2. Click on the history shared by your colleague, click on the plus icon on the far right and choose to import the history.
    3. You can give it a descriptive name if you prefer or leave it as is.



If everything worked out, you should see a list of completed workflow steps highlighted in green. The workflow generates many output files, however not all of the output files are visible. You can explore all the hidden files clicking on the word "hidden" (orange circle) – this will reveal all hidden files.

The Variant calling workflow has three major components: (1) mapping of raw reads to the reference genome, (2) calling variants, and (3) annotating variants. This workflow can be used to call single nucleotide polymorphisms, insertions and deletions (also defined as indels), and multiple nucleotide polymorphisms.

In this workflow, we used Bowtie2 to align and map sequences to a reference genome. Once they are aligned it may be worth checking the quality of this process because misalignments lead to false SNP calls. SAM or BAM files provide sore this information and you can find these files to export in the hidden workflow steps.



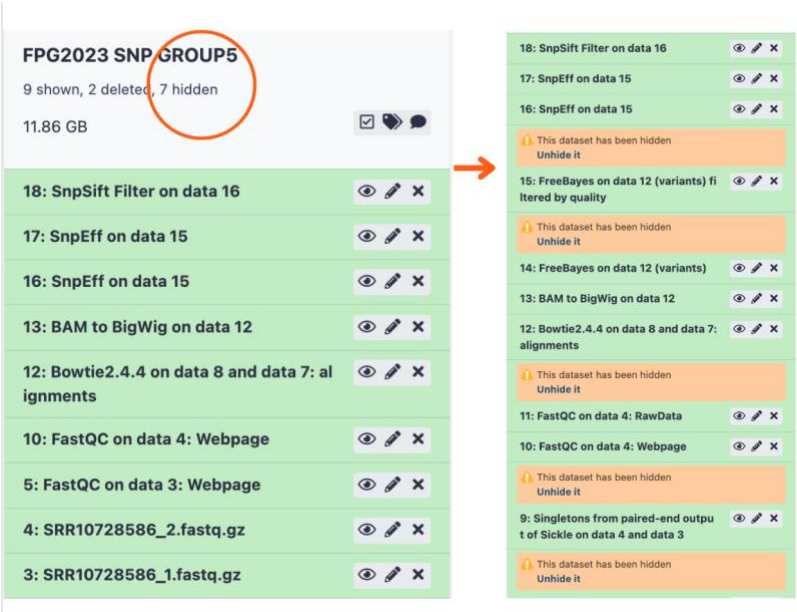
After reads have been aligned, they are sorted based on the chromosomal position. The tool that we are using is called Sort and it belongs to the suite of SAMtools. The sorted file is an input for downstream FreeBayes that calls SNPs and outputs into SnpEff that annotates variants.

Analysis and annotation of the genomic variants are carried out by the SnpEff tool. SnpEff is a variant annotation and effect prediction tool. It annotates and predicts the effects of variants on genes (such as amino acid changes). It uses reference genome to annotate genomic variants based on their genomic location and also predicts SNP coding effects. The genomic location features are intronic regions, 5' and 3' UTRs, and upstream, downstream, splice site and intergenic regions. SNP coding effects are categorized based on the effect of the amino

acid change and are classified into synonymous and non-synonymous, gain or loss of start codons, gain of loss of stop codon, and frame shifts.

The SnpSift tool annotates, filters, and manipulates genomic annotated variants. Once you annotated your files using SnpEff, you can use SnpSift to help you filter large genomic datasets (e.g. sort on high or moderate impact SNPs, etc.).

- **Examine your results.**

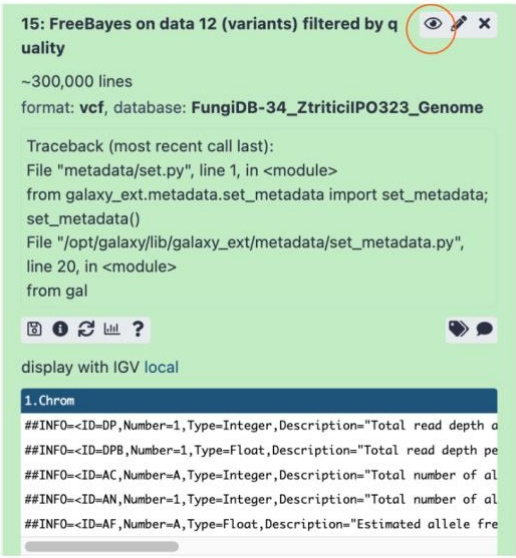


1. Click on the *hidden* files link in the history panel to reveal all workflow output files.
2. Examine the output files.
3. What does the tool FASTQC do?
4. What about Sickle?

The output of Sickle is used by a program called Bowtie2. Bowtie generates a file called a BAM file. Whenever dealing with sequence alignment files you will likely hear of file formats called SAM or BAM. SAM stands for Sequence Alignment/Map format, and BAM is the binary version of a SAM file.

Many of the downstream analysis programs that use BAM files require a sorted BAM file. This allows for more efficient analysis.

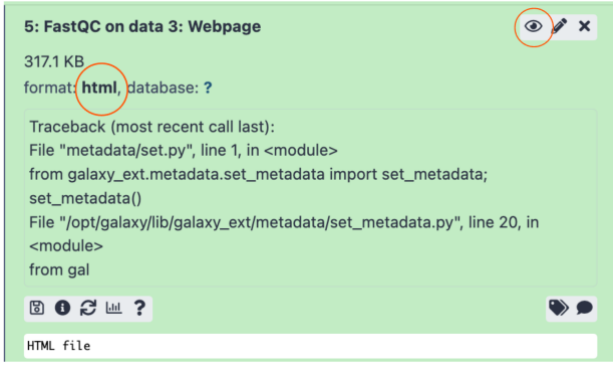
The sorted BAM file is the input for a program called FreeBayes. This program is a Bayesian genetic variant detector designed to find small polymorphisms, specifically SNPs (single-nucleotide polymorphisms), indels (insertions and deletions), MNPs (multi-nucleotide polymorphisms), and complex events (composite insertion and substitution events) smaller than the length of a short-read sequencing alignment. The output for many variant callers is a file called a VCF file. VCF stands for variant interchange format.

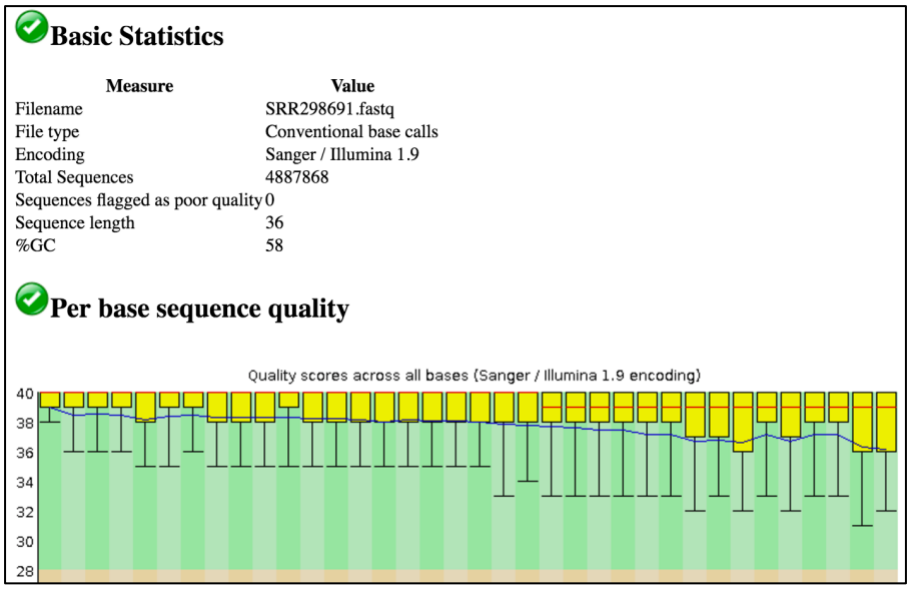


5. Examine the VCF file in your results (click on the *eye* icon to view its contents). Detailed information about VCF file content is available here: https://samtools.github.io/hts-specs/VCFv4.2.pdf

- **Examine sequence quality based on FastQC quality scores.**

FastQC provides an easy-to-navigate visual representation sequencing data quality and distribution of nucleotides per read position. What does the report tell you about the quality ?





Basic Statistics

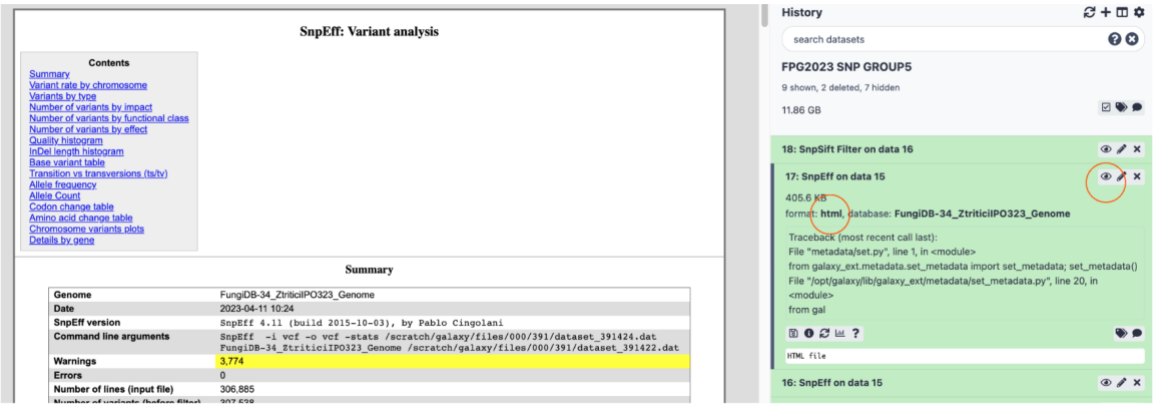
Measure	Value
Filename	SRR298691.fastq
File type	Conventional base calls
Encoding	Sanger / Illumina 1.9
Total Sequences	4887868
Sequences flagged as poor quality	0
Sequence length	36
%GC	58

Per base sequence quality

- **Examine SnpEff summaries (html)**

  - Click on the *View data icon* (eye) in the SnpEff output file that has the html format.

This will open the html file in Galaxy for your review.



The header contains a short summary and information about the run and it has several major components:

The Summary contains warnings about possible genomic annotation errors or inconsistencies identified in the reference genome. If there are many, use caution interpreting results and examine associated gff files for any issues (*e.g.* missing feature values in gff files, incomplete gene sequences, more than one stop codon per gene, *etc.*). Other components:

- Number of line (input file) - number of lines in vcf file
- Number of not variants: 0 - some packages report non-variant observations for nt positions between reference genome and vcf file generate.
- Number of known variants and multi-allelic VCF entries - if you work with a model organism where some variants were given an accession number (most commonly in mice and human projects) any recognised variants will be listed here

Summary

Genome	FungiDB-34_ZtriticiIPO323_Genome
Date	2023-04-11 10:24
SnpEff version	`SnpEff 4.11 (build 2015-10-03), by Pablo Cingolani`
Command line arguments	`SnpEff -i vcf -o vcf -stats /scratch/galaxy/files/000/391/dataset_391424.dat FungiDB-34_ZtriticiIPO323_Genome /scratch/galaxy/files/000/391/dataset_391422.dat`
Warnings	3,774
Errors	0
Number of lines (input file)	306,885
Number of variants (before filter)	307,538
Number of not variants (i.e. reference equals alternative)	0
Number of variants processed (i.e. after filter and non-variants)	307,538
Number of known variants (i.e. non-empty ID)	0 (0%)
Number of multi-allelic VCF entries (i.e. more than two alleles)	653
Number of effects	1,280,819
Genome total length	39,730,198
Genome effective length	39,730,198
Variant rate	1 variant every 129 bases

Variants rate details

Chromosome	Length	Variants	Variants rate
Ztri_MitoScaffold	43,947	18	2,441
Ztri_chr_1	6,088,797	44,156	137
Ztri_chr_10	1,682,575	15,039	111
Ztri_chr_11	1,624,292	14,012	115
Ztri_chr_12	1,462,624	12,767	114
Ztri_chr_13	1,185,774	10,694	110
Ztri_chr_14	773,098	2,064	374
Ztri_chr_15	639,501	7,821	81
Ztri_chr_16	607,044	5,094	119

- Number of effects - SNP effects summary by type and regions
- Genome total length - number of bp in the reference genome
- Genome effective length - how many nucleotides can be mapped back to the genome
- Variant rate - higher frequency of variants before samples can indicate selective pressure

Summary statistics for variant types

Here is an example of variant calls and what they mean in terms of nucleotide changes:

Number variantss by type

Type	Total
SNP	114,034
MNP	12,864
INS	6,907
DEL	7,304
MIXED	2,180
INTERVAL	0
Total	143,289

Type	What is means	Example
SNP	Single-Nucleotide Polymorphism	Reference = 'A', Sample = 'C'
Ins	Insertion	Reference = 'A', Sample = 'AGT'
Del	Deletion	Reference = 'AC', Sample = 'C'
MNP	Multiple-nucleotide polymorphism	Reference = 'ATA', Sample = 'GTC'
MIXED	Multiple-nucleotide and an InDel	Reference = 'ATA', Sample = 'GTCAGT'

Statistics for the variant effects and impacts:

- **High impact** normally refers to frame shift or new stop codon detections as those changes will generate profound effects on gene function.

- **Modifier SNPs** can affect promoter function, while low and moderate SNPs are most commonly identified inside genes and are either non-coding or non-synonymous SNPs.

- Base changes summary. SnpEff html files provide a breakdown of SNPs across gene features:

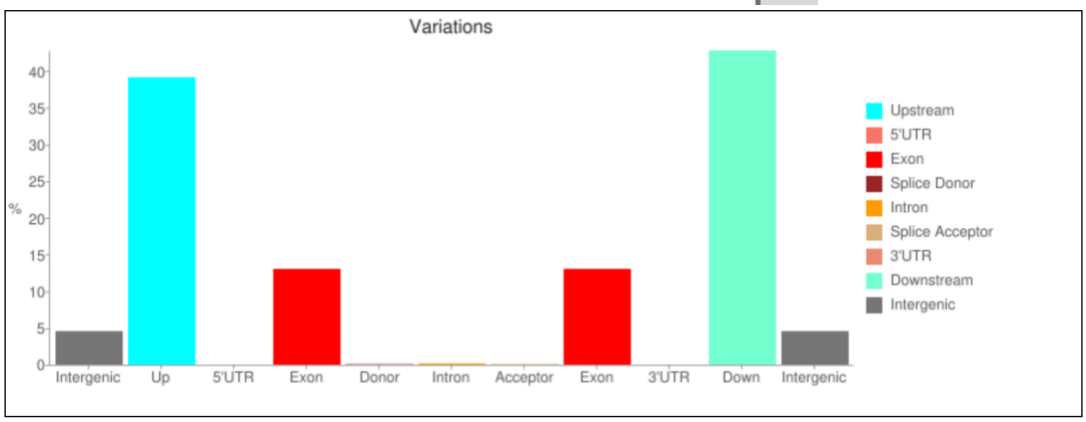
Number of effects by impact

Type (alphabetical order)	Count	Percent
HIGH	1,857	0.145%
LOW	87,874	6.861%
MODERATE	41,970	3.277%
MODIFIER	1,149,118	89.717%

Number of effects by functional class

Type (alphabetical order)	Count	Percent
MISSENSE	29,331	28.472%
NONSENSE	370	0.359%
SILENT	73,317	71.169%

Type (alphabetical order)	Count	Percent
DOWNSTREAM	321,858	40.292%
EXON	67,505	8.451%
INTERGENIC	74,749	9.358%
INTRON	1,064	0.133%
NONE	1	0%
SPLICE_SITE_ACCEPTOR	5	0.001%
SPLICE_SITE_DONOR	4	0.001%
SPLICE_SITE_REGION	176	0.022%
TRANSCRIPT	12	0.002%
UPSTREAM	333,432	41.741%



Additionally, you may see several SNPs being reported in several classes: missense variant + splice region variant. This means that some SNPs that are found within certain splice sites also contain a missense variant. SNPs in the splice sequences may affect intron splicing and lead to read through.

- Quality of reads is indicated in Phred's scale and is a good indicator of the quality of your datasets and results. Quality scores are normally represented by a bar graph where count = number of SNPs and X axis is quality score (higher score mean better p-values and high confidence of the results)

- Base changes: Reflects the frequency of base changes (purine-purine, purine-pyrimidine, pyrimidine-purine, pyrimidine-pyrimidine).

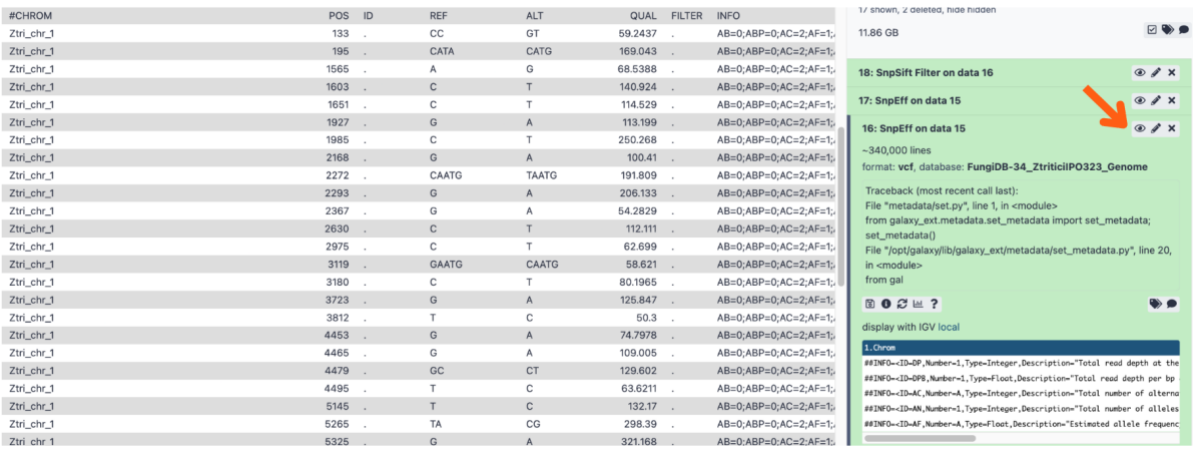
- Transition and transversion ratio help to identify if you may have a selective pressure on certain alleles (high ratio suggests that genes may be under selective pressure).

- Allele frequency statistics reports frequency of alleles and help to identify potential sequencing artifacts due to PCR enrichment step (generation of heterozygous counts in a haploid organism).

The vcf file generated by SnpEff contains information about SNPs and the genomic location. Post-processing of SNP data is normally required to make sense of thousands of SNPs and to decide which ones have biological and functional importance. Data processing can help you to extract SNP distribution and parse associated data including GeneIDs, protein-coding annotations, and effects in sequence ontology terms such as missense or synonymous variants, stop codon gain, etc. and also link changes to the genome model. SnpSift is among other programs that is often in SNP data post-processing. It can be installed and run locally to manipulate vcf files. Alternatively, you can also visualize vcf files in Artemis (additional steps are required to format the data).

Examining SNP information.

You can view the SNP information by clicking on the "eye" icon within the SnpEff vcf file.



#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO
Ztri_chr_1	133	.	CC	GT	59.2437	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	195	.	CATA	CATG	169.043	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	1565	.	A	G	68.5388	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	1603	.	C	T	140.924	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	1651	.	C	T	114.529	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	1927	.	G	A	113.199	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	1985	.	C	T	250.268	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	2168	.	G	A	100.41	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	2272	.	CAATG	TAATG	191.809	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	2293	.	G	A	206.133	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	2367	.	G	A	54.2829	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	2630	.	C	T	112.111	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	2975	.	C	T	62.699	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	3119	.	GAATG	CAATG	58.621	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	3180	.	C	T	80.1965	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	3723	.	G	A	125.847	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	3812	.	T	C	50.3	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	4453	.	G	A	74.7978	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	4465	.	G	A	109.005	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	4479	.	GC	CT	129.602	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	4495	.	T	C	63.6211	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	5145	.	T	C	132.17	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	5265	.	TA	CG	298.39	.	AB=0;ABP=0;AC=2;AF=1;.
Ztri_chr_1	5325	.	G	A	321.168	.	AB=0;ABP=0;AC=2;AF=1;.

The vcf file generated by SnpEff contains information about SNPs and the genomic location. Here is an example of a file opened in Excel:

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	unknown
CM001231	189057	.	AG	CT	787.449	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:143:0:0:143:5341:-207.887,-43.0473,0
CM001231	483825	.	G	A	64.8756	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:4:0:0:4:146:-10.0999,-1.20412,0
CM001231	518226	.	G	C	51.7908	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:8:0:0:7:276:-11.5007,-2.10721,0
CM001231	574021	.	C	G	237.265	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:17:0:0:17:583:-39.079,-5.11751,0
CM001231	609879	.	GAA	CAG	55.2785	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:32:8:277:22:861:-18.1711,-0.694735,0
CM001231	1090073	.	G	T	79.4156	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:8:2:75:6:238:-11.5539,-1.36362,0
CM001231	1090104	.	A	T	70.961	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:6:0:0:6:220:-12.5146,-1.80618,0
CM001231	1153611	.	CCTC	GCTG	111.123	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:8:5:188:3:97:-9.30616,-6.1461,0
CM001231	1159150	.	CT	GC	126.126	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:31:0:0:19:741:-29.7713,-5.71957,0
CM001231	1159438	.	C	G	82.3312	.	AB=0;ABP=0;	GT:DP:RO:QF	0/0:47:30:1092:17:640:0,-9.53002,-3.50705
CM001231	1159465	.	G	C	249.656	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:126:47:1770:79:3013:-53.8644,-25.2134,0
CM001231	1159499	.	T	C	124.95	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:143:32:1167:111:4248:-76.1575,-33.4865,0
CM001231	1181576	.	CC	TG	191.675	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:27:0:0:25:924:-41.7448,-7.52575,0
CM001231	1293309	.	C	G	51.22	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:2:0:0:2:78:-6.92763,-0.60206,0
CM001231	1323058	.	TT	GC	71.3001	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:6:0:0:6:223:-12.5485,-1.80618,0
CM001231	1485397	.	A	G	3558.42	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:499:0:0:497:18671:-804.678,-149.612,0
CM001231	1485429	.	G	A	3783.33	.	AB=0;ABP=0;	GT:DP:RO:QF	1/1:517:1:38:516:20010:-843.425,-151.978,0

Filtering VCF file data.

VCF files contain a lot of data about variants and their positions. SnpEff generates various analyses/summaries of VCF files (including GeneIDs that overlap variant positions). However, it is often necessary to filter VCF files further to obtain useful information for your specific question. For example, you may want to filter out SNP positions that have an impact on the coding sequence.

One tool that can be used is called SnpSift Filter (look at the last step of the pipeline you just ran). This tool allows you to write complex expressions to filter a VCF file. Your workflow is set up to use an expression that filters VCF files on moderate and high impact SNPs (this setting can be adjusted manually in the workflow editor). Here is the exact expression used:

(((ANN[*].IMPACT has 'HIGH') | (ANN[*].IMPACT has 'MODERATE')) & ((na FILTER) | (FILTER = 'PASS')))

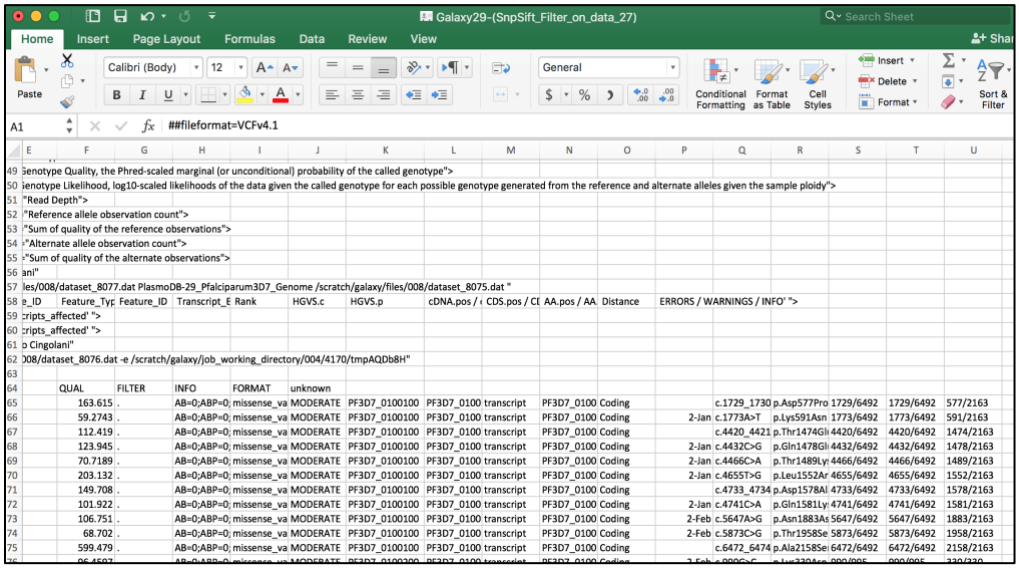
- **Extract filtered VCF file (SnpSift output) and convert into an Excel document.**

For this exercise, two groups with be sharing data SnpSift outputs: group 1 & 2, group 3 & 4, and group 5 & 6. <u>File manipulations should be performed on both SnpSift vcf files.</u>

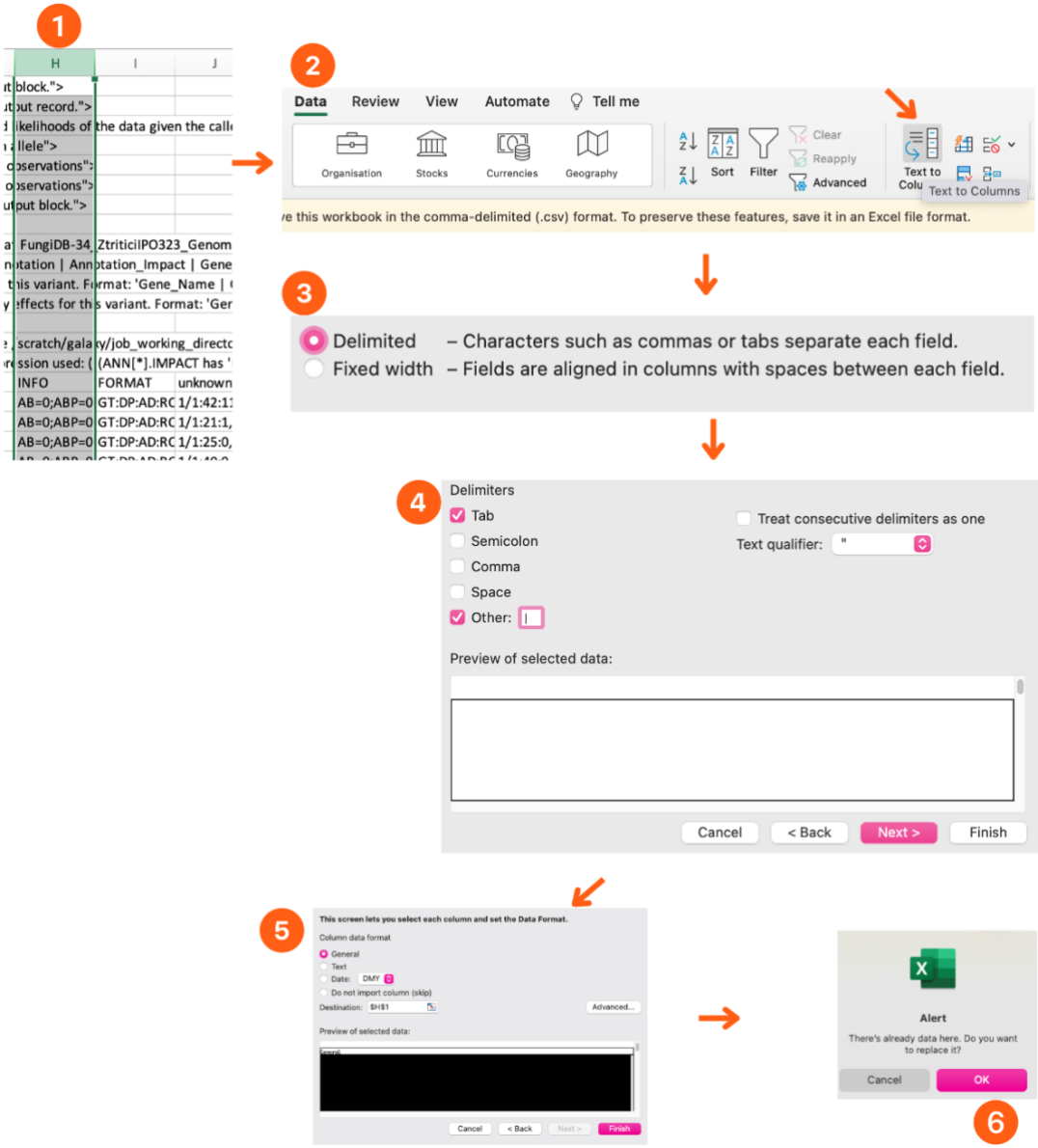
Look at the filtered vcf file in Galaxy. Notice that the Gene IDs are buried in the file, but the file has some structure which means you can extract them either programmatically or using a program like Excel.

Here are some steps you can take to extract Gene IDs from two VCF files then compare them to identify genes that are in common or that distinguish the two files.

1. Download the SnpSift Filter output by clicking on the save icon.
2. Right click and open this file with Excel.

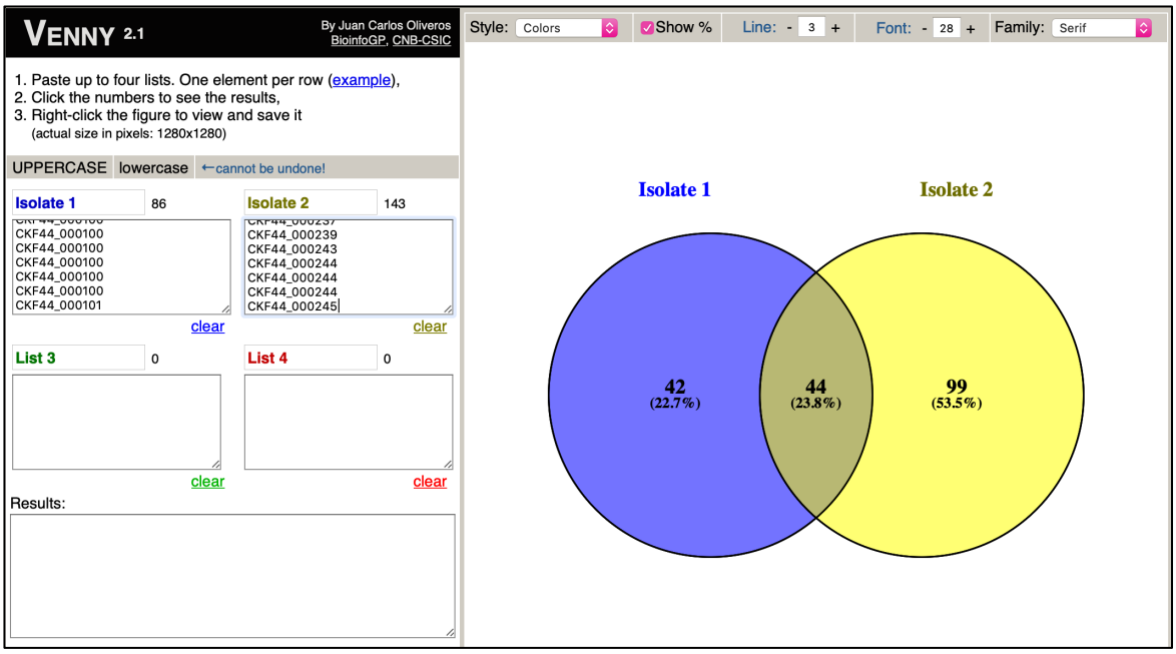


- **Manipulate Excel file to display SNP info in columns.**
  1. Select the “INFO” column.
  2. Navigate to the “Data” tab in Excel and choose “Text to Columns”.
  3. Use the “Delimited” option.
  4. Set delimiters to the “Tab” and “ | ” in the “Other” and click “Next”
  5. Leave other criteria at default and click on the “Finish” button.
  6. Click “OK” on the Alert pop-up.



Now you can look for Gene IDs of interest in the excel file. For example, if this is a known drug resistant line you can sort and examine SNPs based on their characteristics.

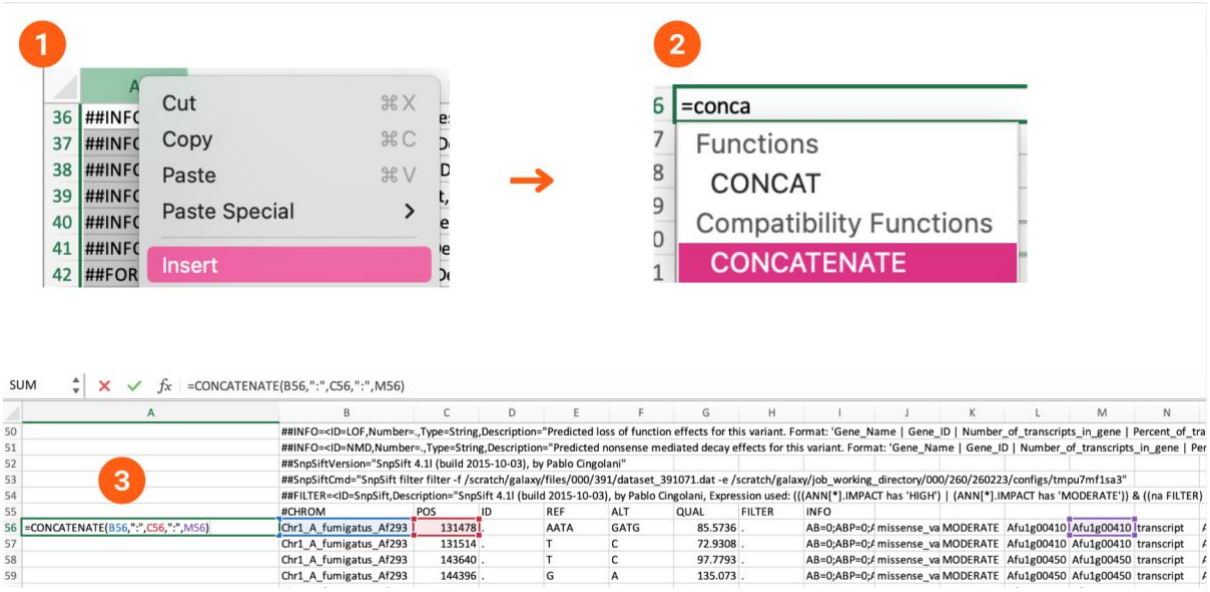
If you are comparing two or more strains, you may want to extract gene IDs from all VCF files and identify common signatures across isolates or strains. For this type of analysis, you can use http://bioinfogp.cnb.csic.es/tools/venny/ to generate a Venn diagram:



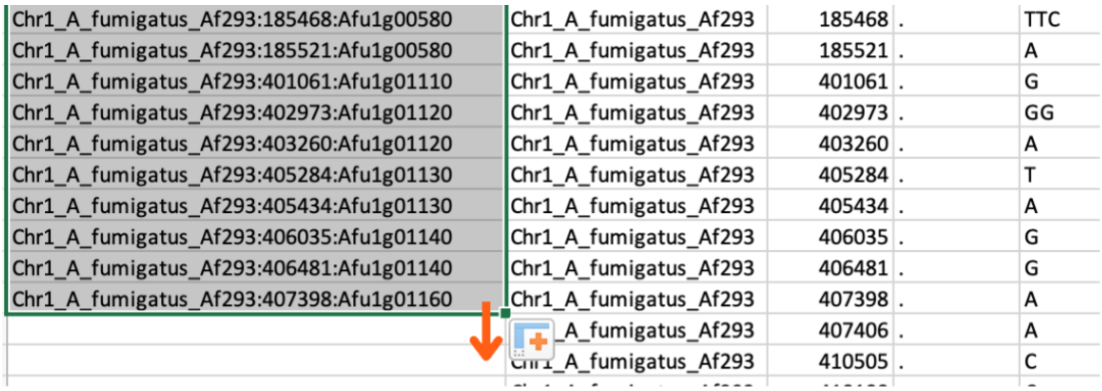
The screenshot above is showing comparison of between lists of GeneIDs. Is it possible to miss some important polymorphisms using this method? Of course, the answer is yes☺
For example, it is quite possible that a gene with a SNP in the WT and a SNP in the mutant that will be in the intersection of the two gene lists, contains different SNPs – you will miss this by doing the above steps. Below is a description of steps you can take to create a list of unique IDs for SNPs. This list of unique IDs can then be used in Venny.

- **Analyze your data in Venny.**

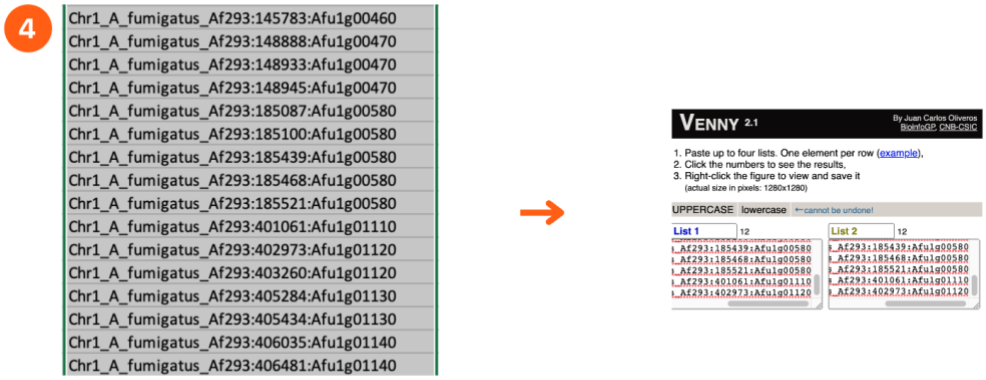
1. Start with the same excel files that you opened in the above section. Insert an empty column before the data.
2. Deploy the concatenate function in Excel.
3. Create a unique ID for SNPs by combining information from multiple columns to create something that looks like this: **chromosome:position:geneID**
   To do this you will use the concatenate function in Excel:
   =concatenate(cell#1,":",cell#2,":",cell#3)
   Cell#1 = cell with chromosome number
   Cell#2 = cell with position
   Cell#3 = cell with GeneID



You should get unique SNP IDs that look like this (for example): CP022321.1:15259:CKF44_000003. Copy this function for other entries:



4. Copy these newly generated unique IDs into List 1 and Lis2 on Venny http://bioinfogp.cnb.csic.es/tools/venny/ and examine the data.

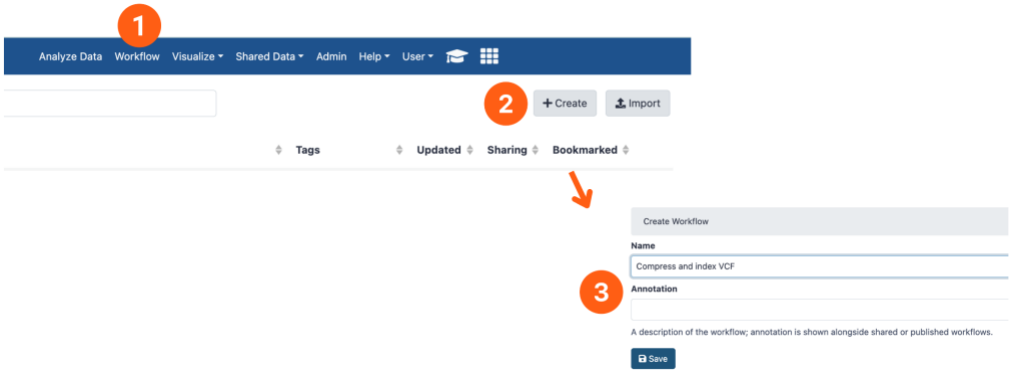


Viewing VCF file results in the JBrowse genome browser.

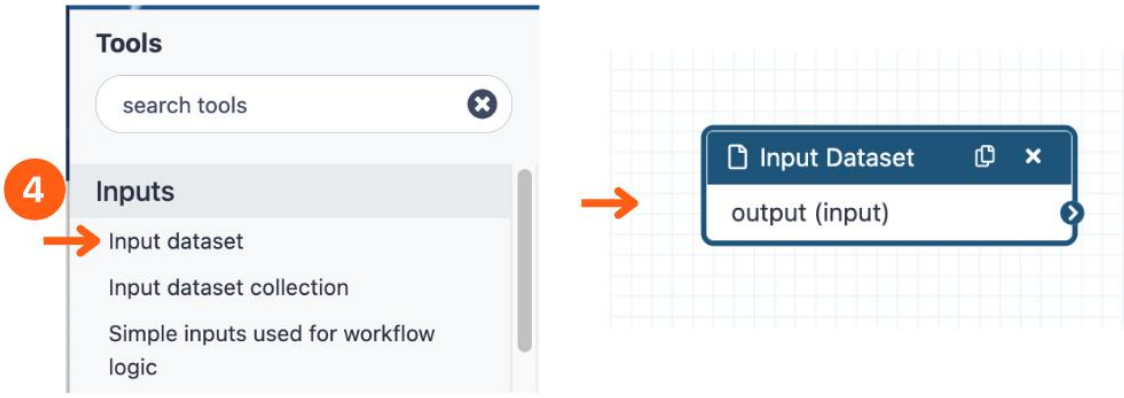
- **Create a workflow to generate a compressed vcf and index files for viewing your data in JBrowse.**

To view a VCF file in JBrowse, it first has to be indexed and compressed. This is done using two tools: bgzip and tabix, respectively. You can run these tools sequentially or you can set up a mini workflow and then run the workflow to generate the output files as follows:

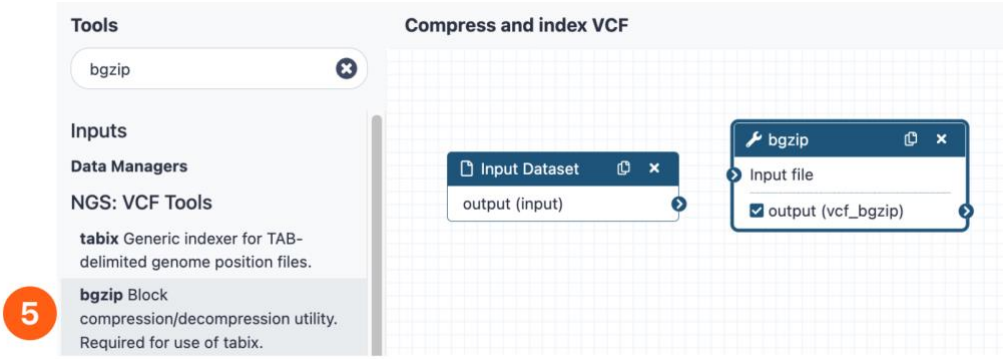
1. Click on the "Workflow" menu.
2. Click on the "Create" button to start a new workflow.
3. Give the workflow a name (e.g. Compress and index VCF) and click on the save button. This will open a workflow canvas.



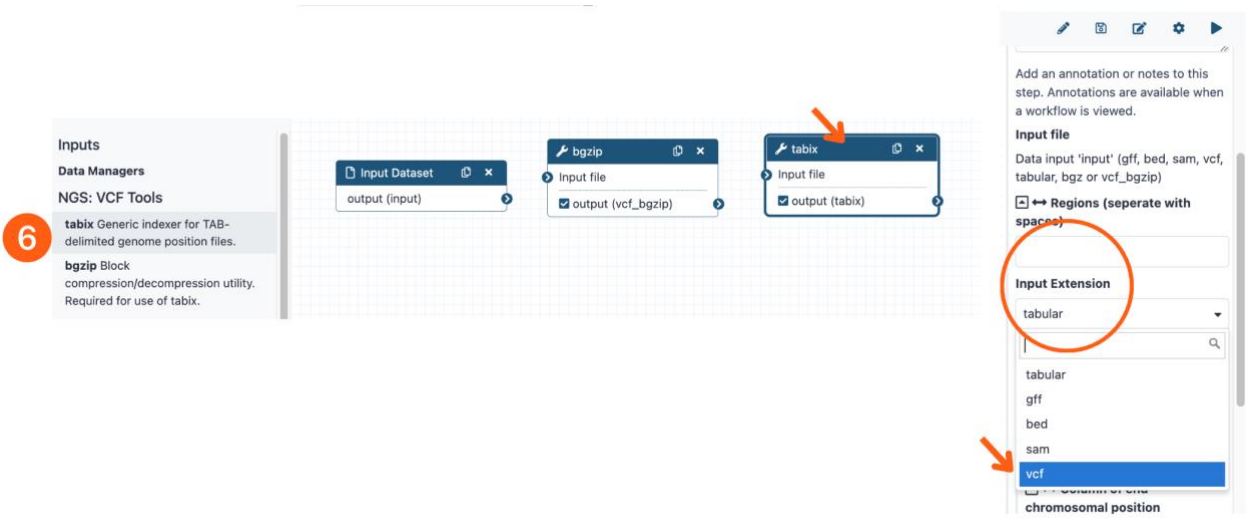
4. All workflows must start with an input file so add the "Input Dataset" step to the workflow using the menu on the left (you must click on the tool for it to appear in the workflow editor canvas).



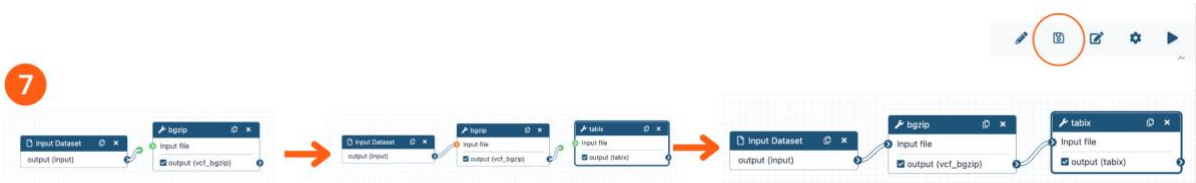
5. Using the menu on the left, search for and add the "bgzip" tool.



6. Using the menu on the left, search for and add the “tabix” tool. Left-click on the “tabix” icon and select “vcf” under “input selection” on the right (tool option section)



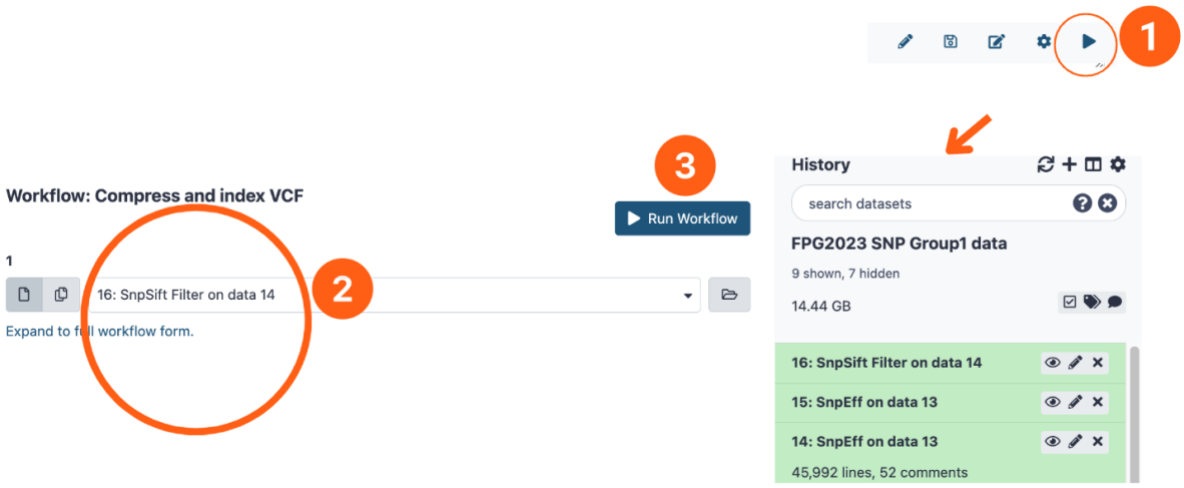
7. Connect each step/tool into a workflow and save it (the button is at the top of the screen)



- **Run the newly created workflow to generate a compressed vcf and index files.**
    1. Click on the "Play" button to start your workflow.
    2. Select the VCF file you want to process.

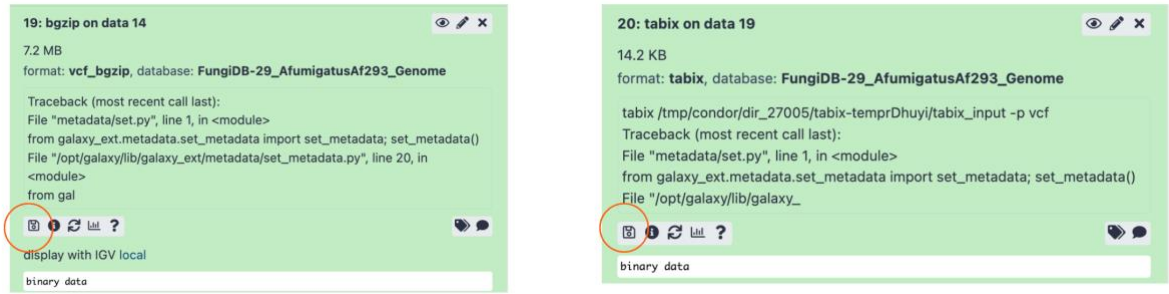
    Note that the workflow produced several vcf files - SnpSift, SnpEff.. In the screenshot below we will use a vcf filtered for high and moderate impact SNPs but if you are interested in all mutations, you may want to choose another file.

    3. Click on the "Run Workflow" button.



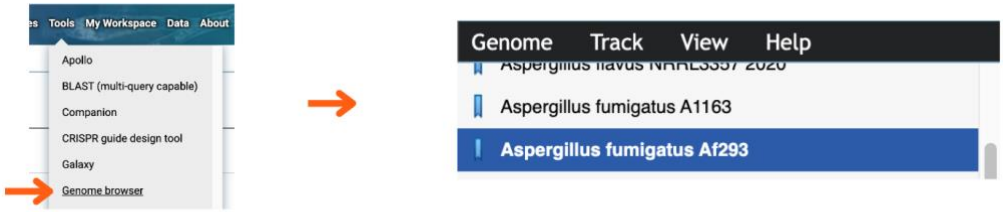
After the workflow completed running, you should have 2 new files in the history on the right (tabix and bgzip).

- **Download compressed vcf (vcf_bgzip) and index (tabix) files and view them in JBrowse.**
    1. Download both files by clicking on the download icon. You will need both files.

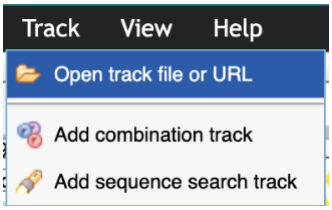


    2. After the files are downloaded, rename them as follows:
        a. The **vcf_bgzip** file to "**group#.vcf.gz"** (i.e. **group1.vcf.gz**)
        b. The **tabix** file to "**group#.vcf.gz.tbi**" (i.e. **group1.vcf.gz.tbi**)

3. Navigate to JBrowse in FungiDB and select the correct genome from the Genome drop-down menu.



4. Click on the Track menu, select "Open track file or URL".



5. Drag and drop your files in the window that appears. Notice that the file formats are autodetected. Click on the "Open" button at the bottom of the pop-up.

You should now be able to view the SNPs in JBrowse.