

# **Course Manual - Phylogenetic Analysis**

# Scenario 3 – Retrieval of sequences and phylogenetic analysis

From the metagenomic analysis carried out you have determined the pathogen responsible for the outbreak above and now wish to identify a likely source of infection. Using phylogenetic analysis, what is the likely source of the infection?

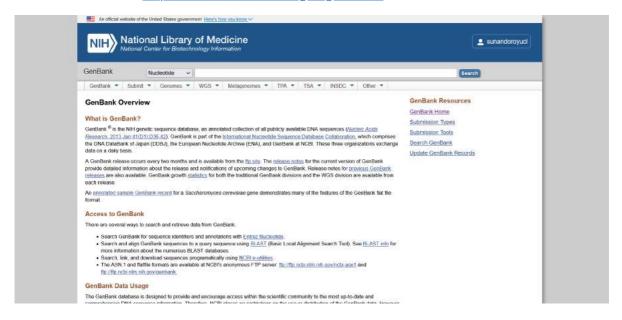
**Note**: All analysis in this section will be carried out on the viral spike protein sequence – NCBI Protein id - **QHD43416.1** from MN908947.3 (Wuhan-1 strain)

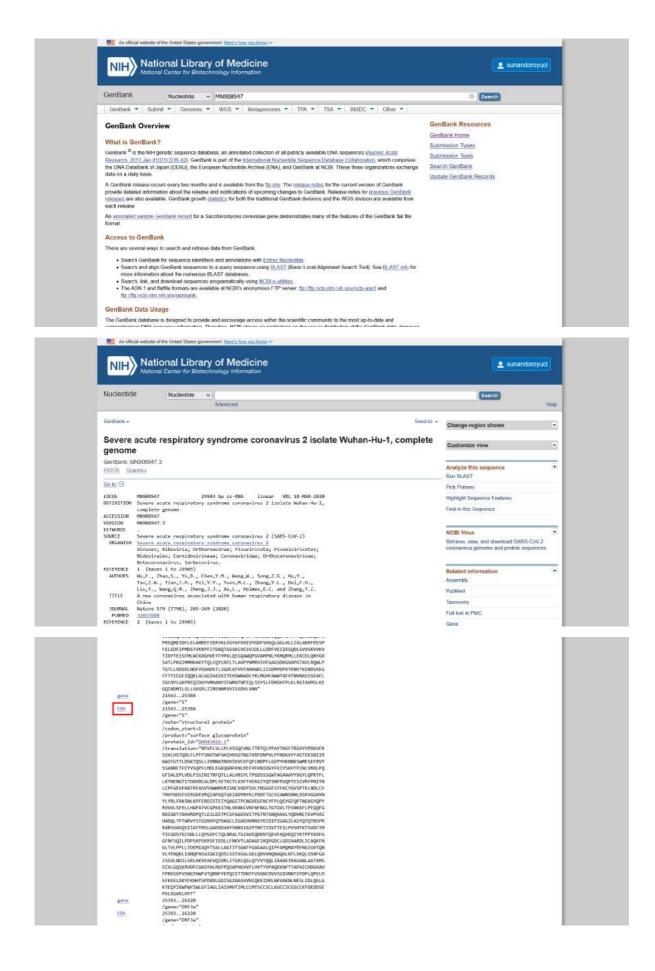
#### Software for the session:

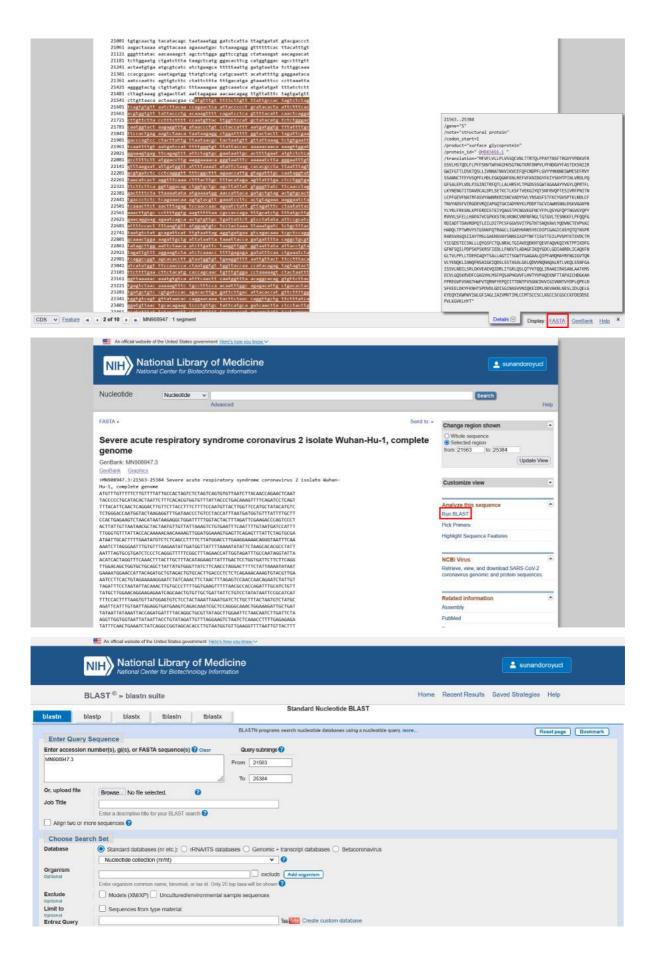
- 1. Mafft Alignment tool
- 2. MEGA Alignment Viewer and Editor
- 3. Modeltest-ng Model Testing
- 4. IQ-TREE Tree Building tool
- 5. Figtree/MEGA Tree Viewer and Editor

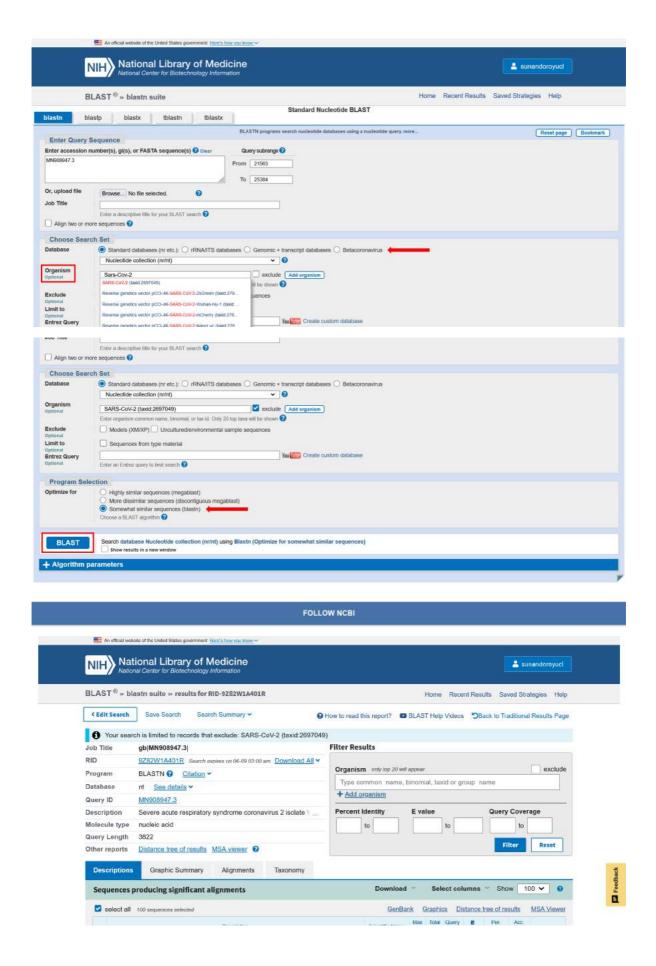
#### **Step 1 - Downloading related sequences**

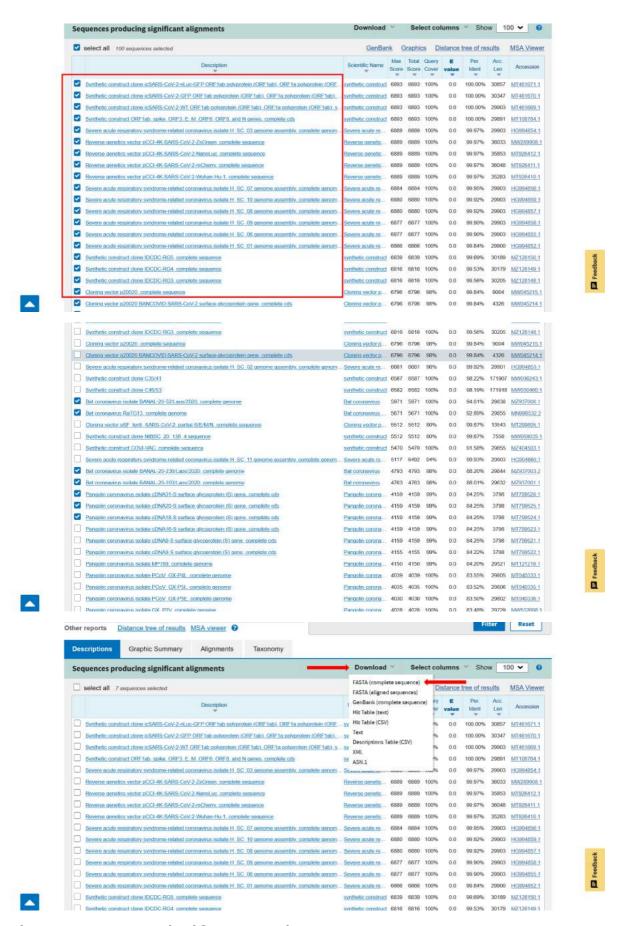
Website - GenBank (https://www.ncbi.nlm.nih.gov/genbank/)





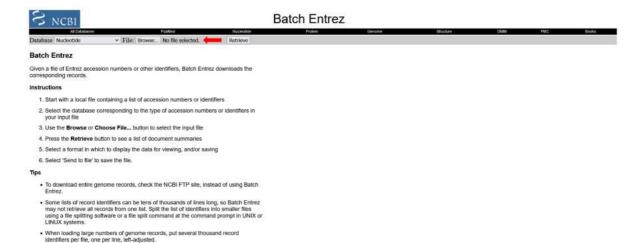






#### Alternate Ways to Download from GenBank

**Batch Entrez** (<a href="https://www.ncbi.nlm.nih.gov/sites/batchentrez">https://www.ncbi.nlm.nih.gov/sites/batchentrez</a>)



**Note:** If you have Accession from different databases you will have to run Batch Entrez multiple times each for a unique sequence database

File to Use ~/Sunando/Betacorona\_Accession.txt

 Ptease note that Batch Entrez will check for duplicate identifiers when reporting results from a list that you have imported.

When retrieving a list of Nucleotide accessions, you must select the specific component database from which the accessions or Gls were saved. For Nucleotide, choose either the CoreNucleotide, the EST or the GSS selection from the database menu. If you have a mixed list of nucleotide accessions or UIDs, you will need to run. It also that the property of the control of the co

Entrez E-utilities ftp://ftp.ncbi.nlm.nih.gov/entrez/entrezdirect/

Manual - https://www.ncbi.nlm.nih.gov/books/NBK179288/

QuickStart - http://bioinformatics.cvr.ac.uk/blog/ncbi-entrez-direct-unix-e-utilities/

#### **Browser**

```
https://eutils.ncbi.nlm.nih.gov/entrez/eutils/efetch.fcgi?
db=nucleotide&id=AY278488, AY304486, MN908947,
MT782115&rettype=fasta&retmode=text
```

#### **Command Line**

```
$ esearch -db "protein" -query "txid11270[Organism] AND L Protein Complete AND
refseq[filter]" | efetch -format fasta > outputfile.fasta
$ head outputfile.fasta
```

For the next step we will start with ~/Sunando/Spike.fas

Step 2 - Aligning Sequences.

#### **Software Used**

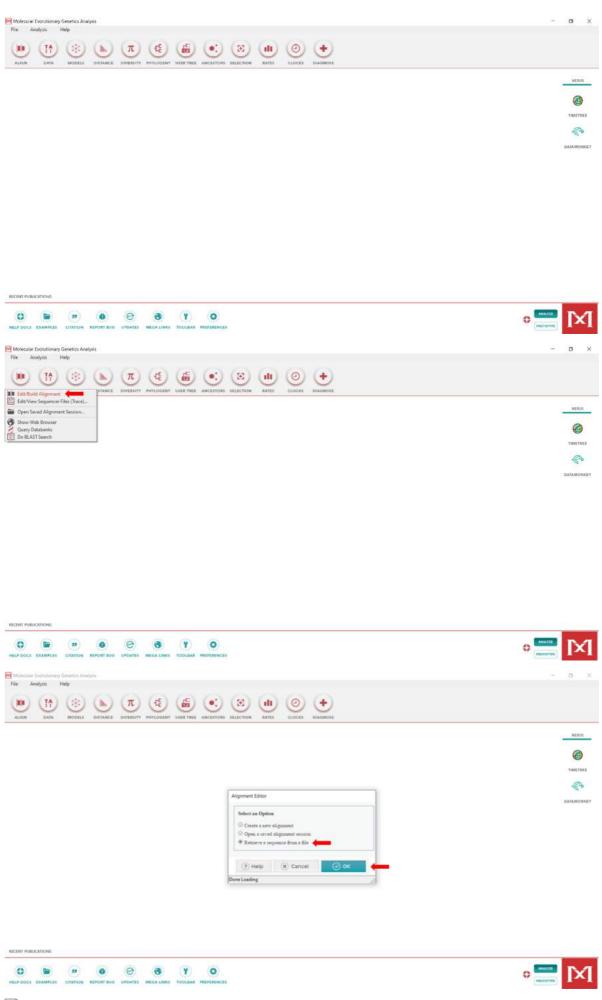
Mafft (https://mafft.cbrc.jp/alignment/software/)

MEGA (http://doua.prabi.fr/software/seaview)

Alternate Software – MUSCLE, CLUSTALW

To view/edit sequence files we will use MEGA. MEGA has a GUI and will launch as a standalone program

```
$ mega
```



```
$ mafft ~/Sunando/Spike.fas > ~/Sunando/outputfile.fas
```

Uses the default models to align the sequences. For highly divergent sequences this may produce inaccurate alignments.

Alternatives are if you have a curated alignment <u>mafft --add</u> works to add new sequences to existing alignments which puts more weightage on the existing alignment

We can also use the L-INS-i algorithm in Mafft that aligns more divergent sequences using pairwise local alignments

```
$ mafft --maxiterate 1000 --localpair ~/Sunando/Spike.fas > outputfilelinsi.fas
```

The final alignment file is ~/Sunando/Spike\_aln.fas

You can also use the alignment file you have generated.



Step 3 - Constructing a Phylogeny.

# **Software Used**

Modeltest-ng (https://github.com/ddarriba/modeltest)

IQ-TREE (http://www.iqtree.org/)

Alternate Software - PhyML,RAxML

For this session we will start with the aligned nucleotide sequences created in the last step.

#### **Model Testing**

To run model testing we will use Modeltest-ng

```
$ modeltest-ng -d aa -i ~/Sunando/Spike_aln.fas -o modeloutputfile -t ml -p 2
```

- -i: Input file
- -o: Output file
- **-t**: Sets the starting tree topology Genomics and Clinical Virology 2022 - Bioinformatics

# -p: Number of threads

We test three criteria to select the best fitting models BIC, AIC and AICc. The modeltest results for this alignment are in ~/Sunando/Spike\_model.out and ~/Sunando/Spike\_model.log

```
$ nano ~/Sunando/Spike_model.out
$ nano ~/Sunando/Spike_model.log
```

AICc	model	K	lnL	score	delta	weight
1	TVM+I+G4	9	-89441.2343	179094.4687	0.0000	0.5632
2	GTR+I+G4	10	-89440.4884	179094.9768	0.5081	0.4368
3	TVM+G4	8	-89467.5351	179145.0703	50.6016	0.0000
4	GTR+G4	9	-89467.0653	179146.1306	51.6620	0.0000
5	TPM3uf+I+G	4 7	-89514.2112	179236.4224	141.9537	0.0000
6	TIM3+I+G4	8	-89514.1910	179238.3821	143.9134	0.0000
7	TPM2uf+I+G	4 7	-89530.2483	179268.4965	174.0279	0.0000
8	TIM2+I+G4	8	-89530.0605	179270.1211	175.6524	0.0000
9	TPM3uf+G4	6	-89541.5959	179289.1919	194.7232	0.0000
10	TIM3+G4	7	-89541.6048	179291.2095	196.7409	0.0000
Model: lnL:	es: tes: s prop: pe:	 ГVM+I+G4 -89441.2343 9.2626 0.1931	0.1752 0.3691 1.7471 1.9174	5.3471 1.0000		

The model that we will use for tree building is TVM+I+G4

#### Tree building

To build a tree we are going to use IQ-TREE

```
$ iqtree -s ~/Sunando/Spike_aln.fas -bb 1000 -st DNA -nt 4 -alrt 1000 -pre
treeoutfile
```

-s: Input File

-bb: ultrafast bootstrap

-st: data type

-nt: Number of threads

-alrt: SH-like approximate likelihood ratio test

-pre: Prefix for output file

IQ-TREE outputs multiple files. The final tree file we will use has an extension of .contree

The final output we will take forward to the next step while IQ-TREE completes running will be ~/Sunando/Spike\_Tree.contree

# **Step 4 - Viewing and Modifying a Tree File.**

#### **Software Used**

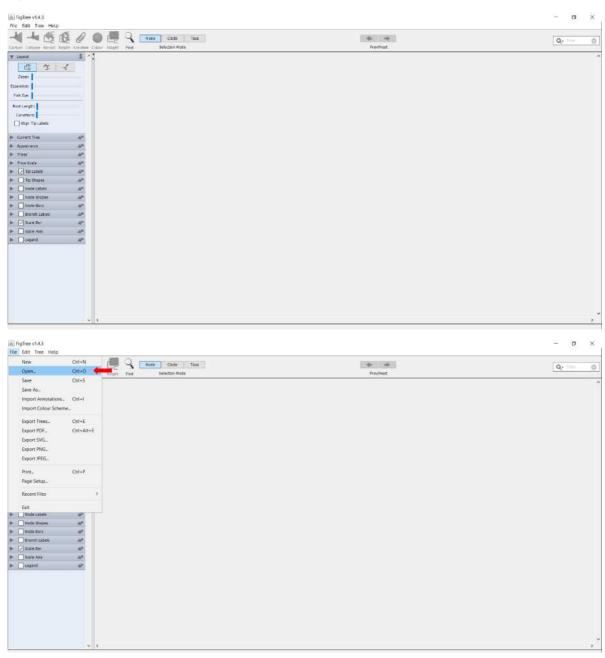
FigTree (http://tree.bio.ed.ac.uk/software/figtree/)

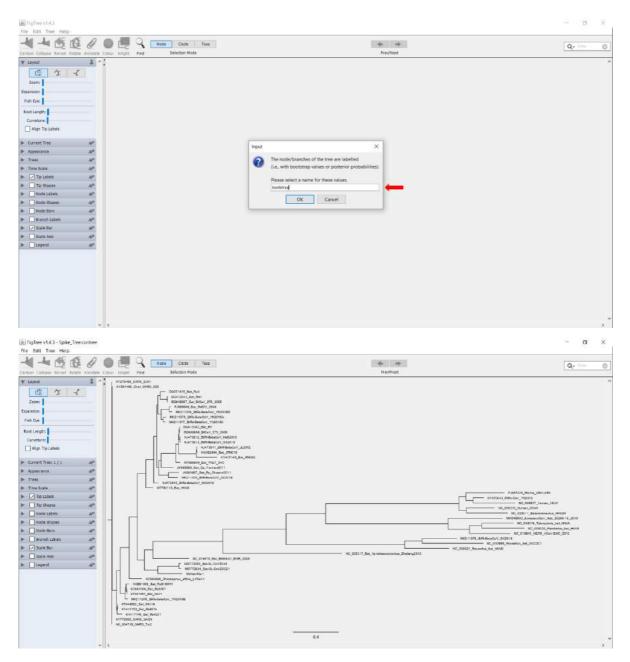
MEGA (<a href="https://www.megasoftware.net/">https://www.megasoftware.net/</a>)

# \$ figtree

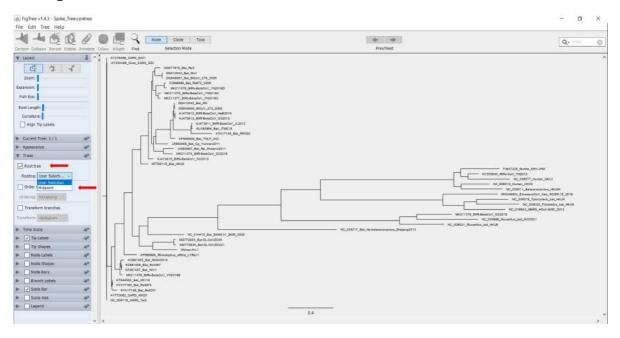
Figtree is Java based and will launch a GUI

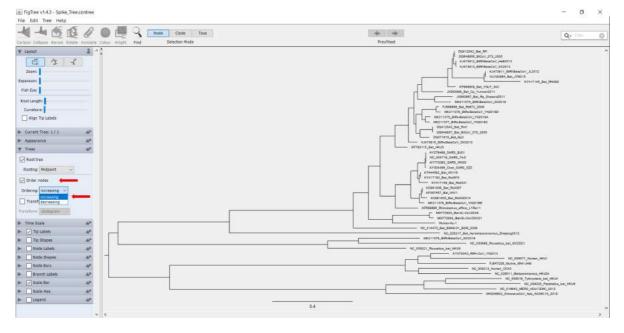
# **Open file -** ~/Sunando/Spike\_Tree.contree



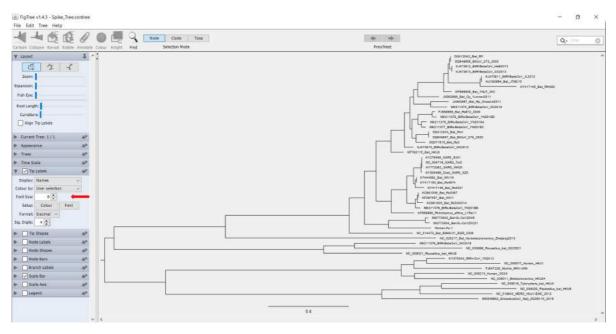


# **Rooting a Tree**

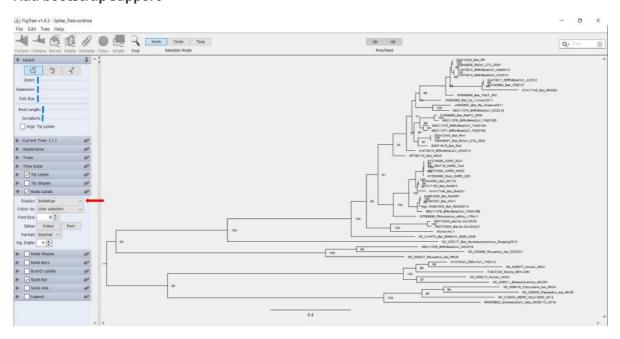


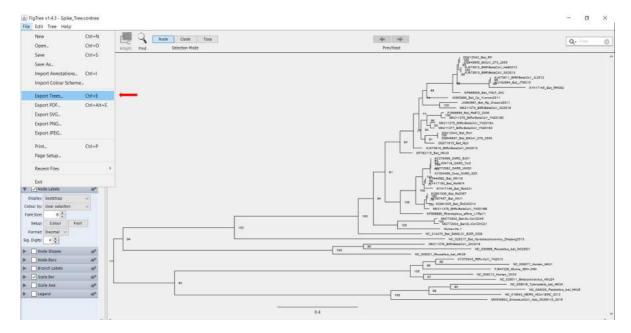


## **Modify Labels**



### Add bootstrap support

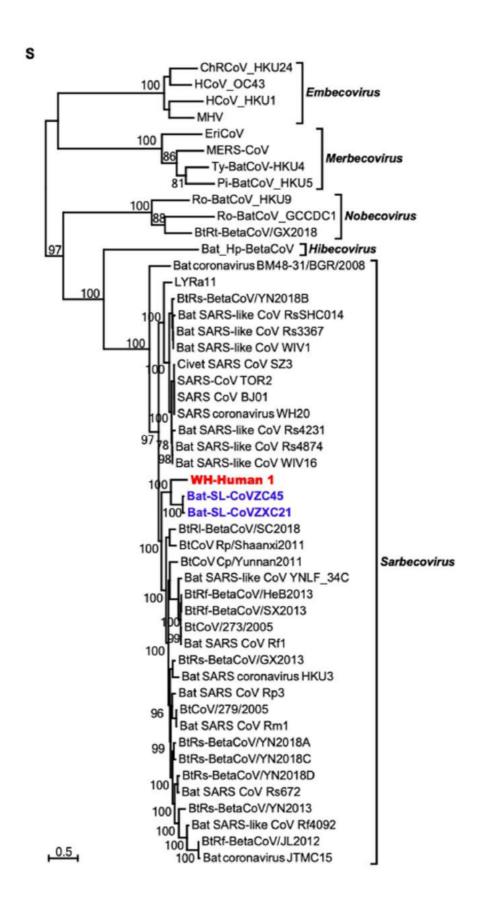




We can go through similar tree viewing in MEGA

### **Final Results**

We further process the trees in **PowerPoint/ggtree** or other image editing tools to add metadata for each sequence in the alignment. In this case we have added virus groups for each virus in the alignment.



Based on this data one would infer that the isolated virus most likely lies within the Sarbecovirus group which includes the original SARS coronavirus but is quite distinct from it. The closest related species both are from bats which could suggest a potential origin.