Section 1: Microbiology to molecular biology 'Germs to Genes'

Molecular Approaches to Clinical Microbiology in Africa 2024

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Learning Outcomes

- 1. Overview of culture-based clinical microbiology.
- 2. The impact of molecular approaches.
- 3. Barriers to understanding molecular and genomic data.
- 4. How molecular data has affected our understanding of an exemplar disease, cholera.
- 5. 16S rRNA as an exemplar molecular approach, including an online exercise.

Questions in clinical microbiology

evolution

emergence

epidemiology

diagnosis

Centuries+

decades

years

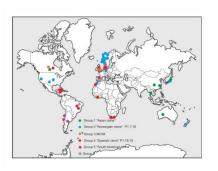
months

weeks

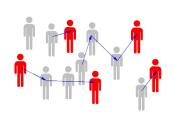
days

hours



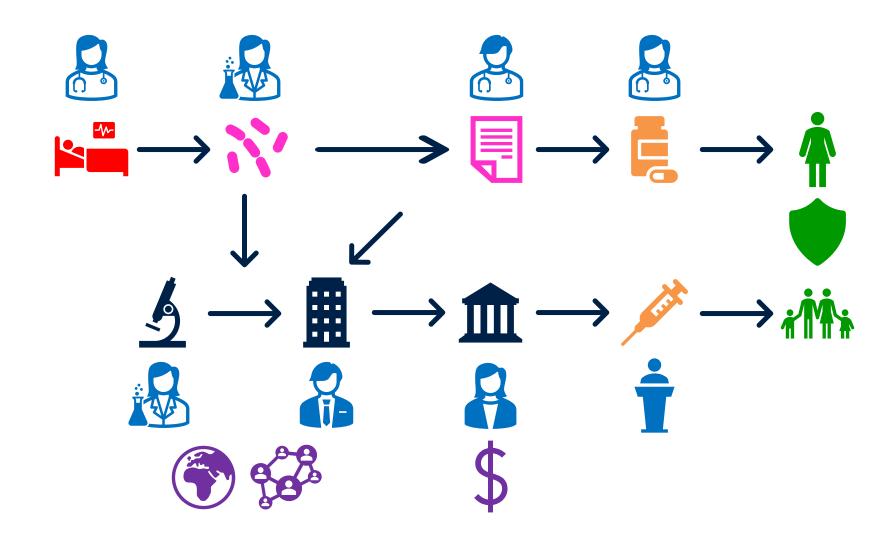








Clinical and public health microbiology without molecular approaches

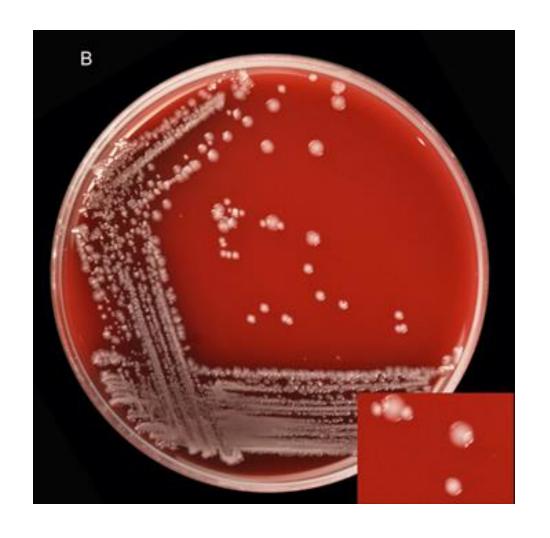


Microbiology: pure culture

- The ability to isolate pathogens was foundational to microbiology.
 - Koch's postulates.
- In addition to diagnosis, this provides material for further characterisation.
 - Isolates remain widely used.

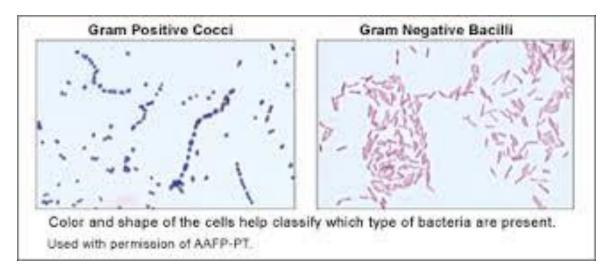
However;

- Culture requires sophisticated, often bespoke, reagents and equipment, and time;
- Many pathogens cannot be grown cultured:
 - lack of methods;
 - lack of suitable specimens.

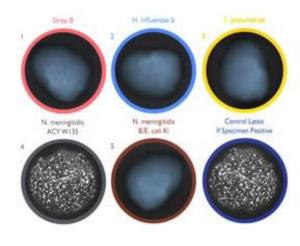


Phenotypic characterisation

Gram's Stain







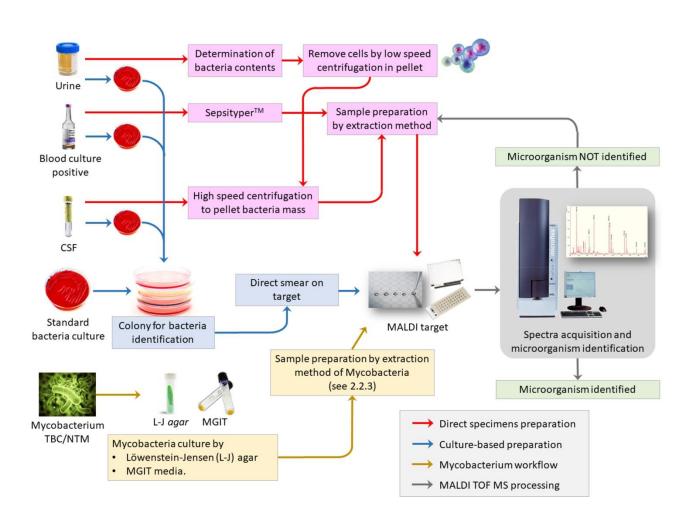


Anti-microbial resistance testing

Metabolic phenotyping For species identification



MALDI-TOF phenotyping



Matrix-assisted laser desorption/ionizationtime of flight (MALDI-TOF) mass spectrometry.

- Enables rapid and inexpensive (per sample) species identification
- Requires:
 - bacterial sample (usually an isolate);
 - the equipment;
 - a database linking spectra to bacterial species identification.
- Commercial systems are widely installed and used in clinical laboratories.

Hou, T. Y., Chiang-Ni, C. & Teng, S. H. (2019). Current status of MALDI-TOF mass spectrometry in clinical microbiology. *J Food Drug Anal.* 27, 404-414.

The molecular revolution and DNA sequences

Definitive:

- fundamental level of information;
- any part of the genome can be accessed.

Reproducible:

- nucleotide sequences are either right or wrong and can be checked;
- reverse mutations are (usually) rare.

Scalable:

 nucleotide sequencing technology can be conducted on one or many samples and on a few base pairs or a whole genome.

Manipulable:

nucleotide sequences can be analysed with model-based methods.

Can be done from a PCR reaction or microbiome sample:

an isolate is not necessarily required.



Frederick Sanger (1918-2013)

4081 otcettetca atcoccaatg ottgettec ataagcagat 4141 tggaagagat totgtotttt ogtatgoagg gogttgagtt 4201 ttgacggoca taaggotgot totgacgtto gtgatgagtt

Sanger, F., Air, G. M., Barrell, B. G., Brown, N. L., Coulson, A. R., Fiddes, C. A., Hutchison, C. A., Slocombe, P. M. & Smith, M. (1977). Nucleotide sequence of bacteriophage phi X174 DNA. *Nature* 265, 687-695.

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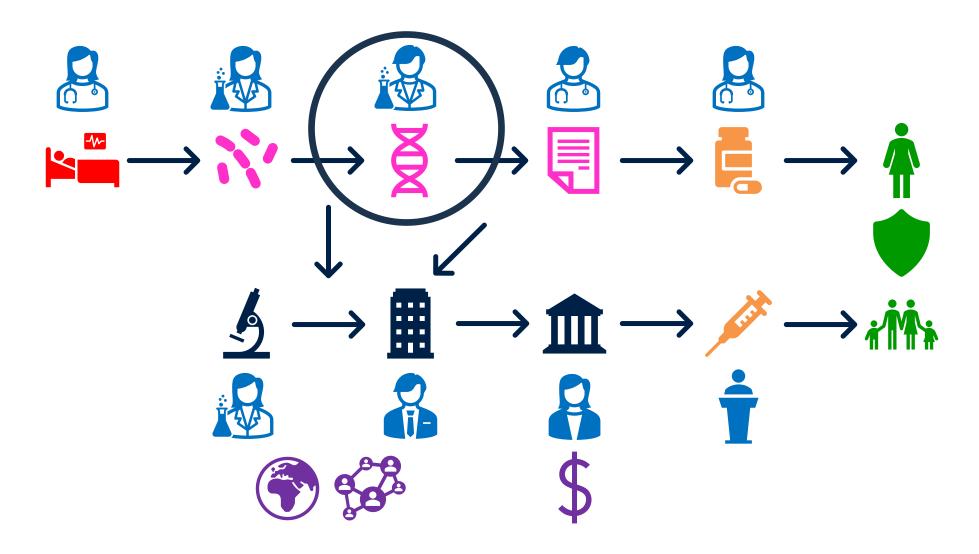
5281 tgg gaaaag t tactgtagce gacgttttg g cgg cgcaace tgtgacgaca aatctgctca 5341 aatttatgcg cgcttcgata aaaatgattg gcg tatccaa cctgca

61 aaattatett gataaageag gaattactae tgettgttta egaattaaat egaagtggae 121 tgetggegga aaatgagaaa attegaeeta teettgegea getegagaag etettaettt 181 gegaeettte geeateaaet aaegattetg teaaaaaetg aegegttgga tgaggagaag 241 tggettaata tgettggeae gttegteaag gaetggtta gatatgage aeatttigt

361 getgitcaac cactaatag taagaatca tgagtcaagt tactgaacaa tccgtacgtt 421 cccagaccg tittggcetci atlaagctca ticaggetic tgccgittig gatitaaccg 481 aagatgatti cgatitictg acgagtaaca aagitiggat tgctactgac cgcticcgtg 541 cctgtcgctg cgttgaagci tgcgtitatg gtacgtgga ctttgtggga taccctcgct 601 ittctgcccc ctottaagtit atloctoccg toattactta titatotcat cccorpaaca

661 ttcaaacggc ctgtctcatc atggaaggcg ctgaatttac ggaaaacatt attaatggcg 721 tcgagcgtcc ggttaaaggc gctgaattgt tcgcgtttac cttgcgtgta cgcgcagga a 781 acatchacgt tcttactdac gcacagaaga acotocgtca aaaattacgt cgcgaagag

Molecular approaches to clinical and public health microbiology



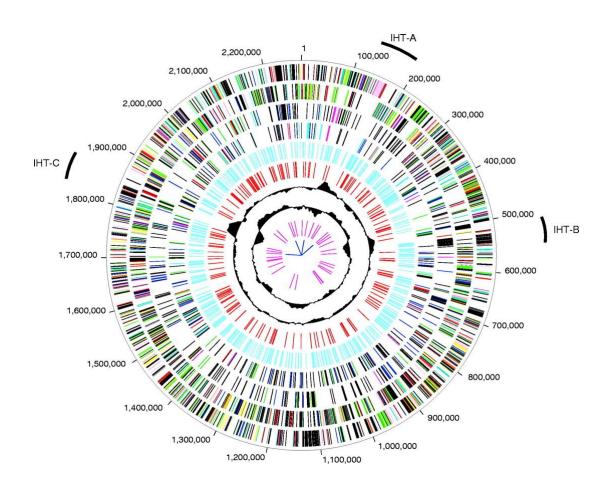
The role of genetics and genomics

- Nucleotide sequencing facilitates specimen characterisation,
 - sequence-based typing, up to the level of whole genome sequences (WGSs).

- Sequences are definitive, reproducible, and comparable.
 - easily stored, transported, and manipulated with analysis algorithms;

- However, datasets are large and complex,
 - requiring interpretation for the practitioner.

Translating molecular and genomic data



Barriers:

- unfamiliarity;
- complexity, apparent and real;
- applicability.

Solutions:

- explaining & summarising;
- structuring & curating;
- interpreting & synthesising.

Tettelin, H., Saunders, N. J., Heidelberg, J., Jeffries, A. C., Nelson, K. E., Eisen, J. A., Ketchum, K. A., Hood, D. W., Peden, J. F., Dodson, R. J., Nelson, W. C., Gwinn, M. L., DeBoy, R., Peterson, J. D., Hickey, E. K., Haft, D. H., Salzberg, S. L., White, O., Fleischmann, R. D., Dougherty, B. A., Mason, T., Ciecko, A., Parksey, D. S., Blair, E., Cittone, H., Clark, E. B., Cotton, M. D., Utterback, T. R., Khouri, H., Qin, H., Vamathevan, J., Gill, J., Scarlato, V., Masignani, V., Pizza, M., Grandi, G., Sun, L., Smith, H. O., Fraser, C. M., Moxon, E. R., Rappuoli, R. & Venter, J. C. (2000). Complete genome sequence of *Neisseria meningitidis* serogroup B strain MC58. *Science* 287, 1809-1815.

Impacts of molecular data on clinical microbiology principles and practice

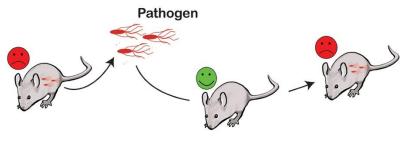
Koch's Postulates revisited

- Koch's postulates original formulation is, the:
 - microbe is found in all cases of disease and is absent in its absence;
 - microbe can be isolated and grown in pure culture;
 - cultured microbe can cause disease in a healthy host;
 - microbe can be re-isolated and cultured from this host.
- These ideas have been revisited in the light of knowledge on molecular pathology and the microbiome.
 - 'Molecular Koch's Postulates';
 - 'Ecological Koch's Postulates'.

Vonaesch, P., Anderson, M. & Sansonetti, P. J. (2018).

Pathogens, microbiome and the host: emergence of the ecological Koch's postulates. *FEMS Microbiol Rev.* **42**, 273-292.

Koch's postulate



«A pathogen, a disease»

Molecular Koch's postulate

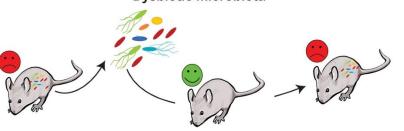
Virulence gene



«A virulence gene, a disease»

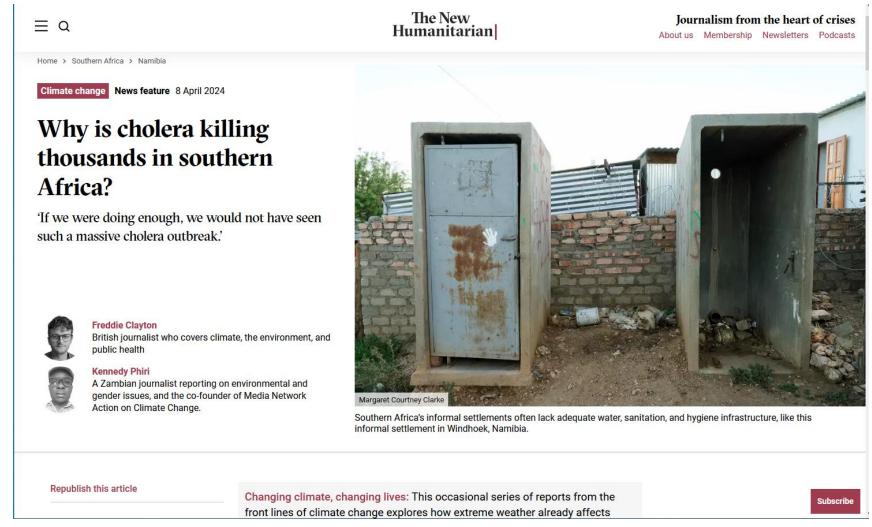
Ecological Koch's postulate

Dysbiotic microbiota



«A dysbiosis, a disease»

Case study: Cholera



Cholera: Koch's postulates

- Pacini 1854 presence of 'comma shaped' bacteria in stools.
- Snow 1854 cholera is water borne.
- Koch 1883 isolation of *Vibrio* cholerae.
- Gradual introduction of improved sanitation in Europe & North America.

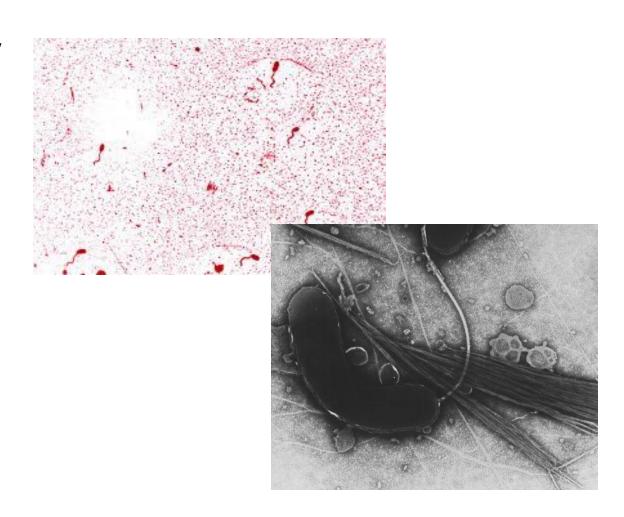


Robert Koch (third from the right) on a cholera research expedition in Egypt in 1884, one year after he identified *V. cholerae*, the microbe responsible for cholera.

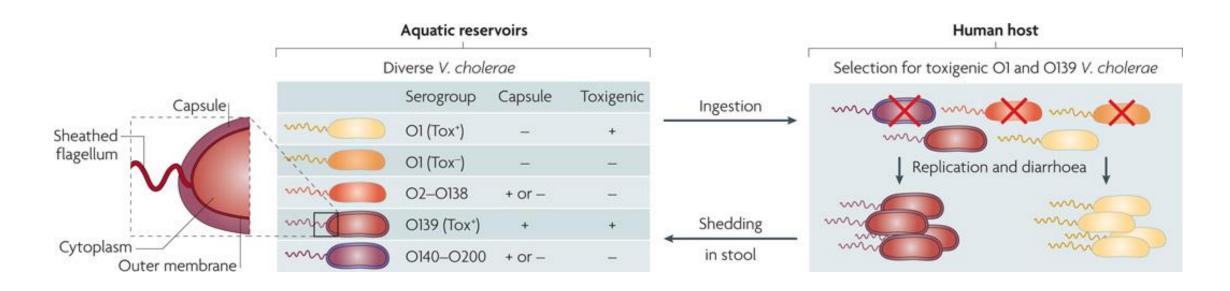
Hamlin, C. (2009). Cholera: the biography. Oxford, UK: Oxford University Press.

Cholera: the bacterium, Vibrio cholerae

- A Gram-negative bacterium, commonly found in brackish estuarine water.
- Differentiated serologically on the basis of the O antigen of its LPS.
- Cholera toxin-producing strains of the O1 and O139 serogroups cause the great majority of disease.
- All pathogenic *V. cholerae* express cholera toxin (Ctx) and toxin coregulated pilus (Tcp).



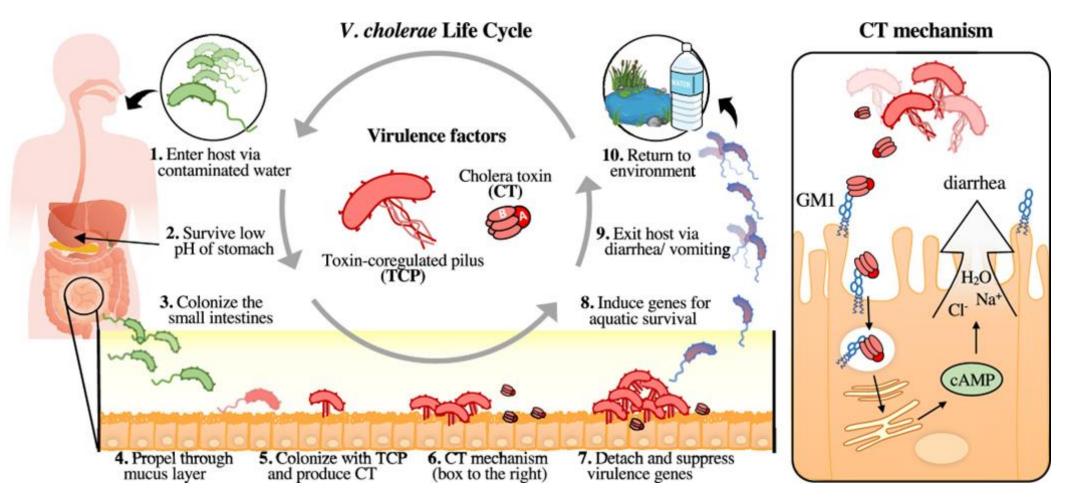
Cholera: the genes, encoding capsule and toxin



- More than 200 serogroups (O1- O200) of Vibrio cholerae exist.
- Only a subset of O1 and O139 are toxigenic (Tox⁺) and capable of causing cholera when ingested.
- The tcp gene is located on a pathogenicity island, the ctxA and ctxB genes on a filamentous phage.
- These variants are selected for in the host, Tox- are selected against

Nelson E.J. *et al* (2009). Cholera transmission: the host, pathogen and bacteriophage dynamic. *Nature reviews* **7**, 693-702

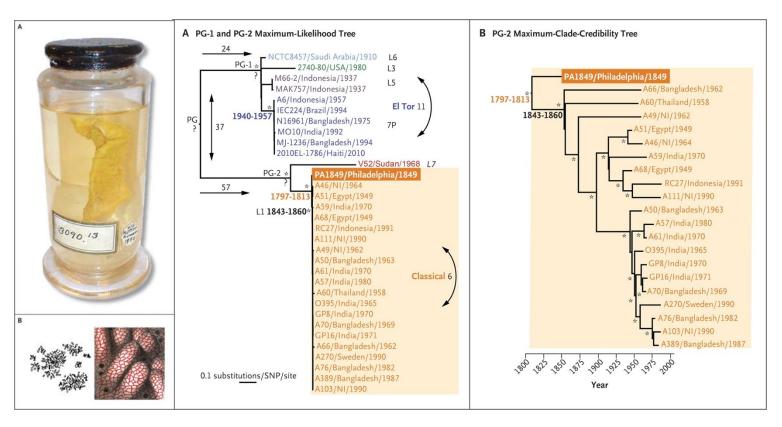
Cholera: the dysbiosis



Chac, D., Dunmire, C. N., Singh, J. & Weil, A. A. (2021). Update on Environmental and Host Factors Impacting the Risk of *Vibrio cholerae* Infection. *ACS Infect Dis.* **7**, 1010-1019

Cholera: the power of genomics, 2nd Pandemic strain, Philadelphia, 1849

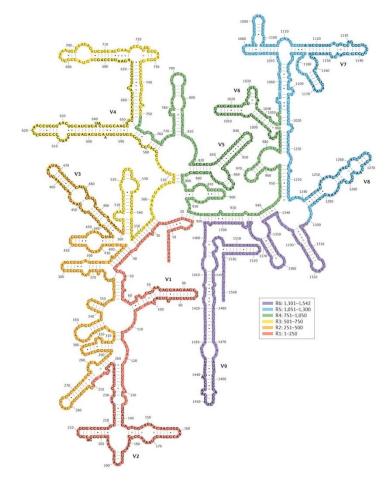
- A whole genome sequence was reconstructed
 - from preserved intestinal material from a cholera victim.
- The *V. cholerae* strain was O1 biotype,
 - 95-7% similarity to 'Classical'
 0395 genome.
- One or more tandem ctx phage sequences,
 - VPI-1 and VPI-2 present.



Devault, A. M., Golding, G. B., Waglechner, N., Enk, J. M., Kuch, M., Tien, J. H., Shi, M., Fisman, D. N., Dhody, A. N., Forrest, S., Bos, K. I., Earn, D. J., Holmes, E. C. & Poinar, H. N. (2014). Second-pandemic strain of *Vibrio cholerae* from the Philadelphia cholera outbreak of 1849. *N Engl J Med.* **370**, 334-340

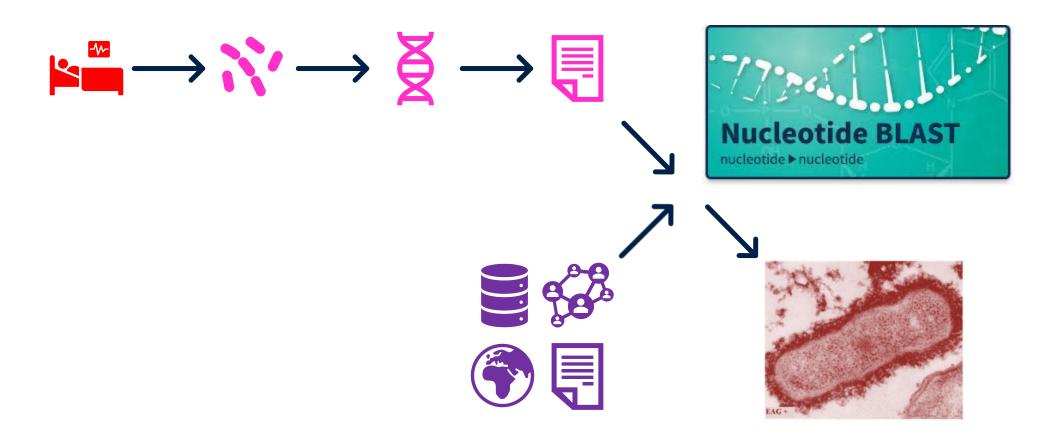
Single gene case study: 16S rRNA gene analysis

- 16S rRNA is a functionally essential structural component of the ribosome, highly conserved.
- Varies among organisms, although most variation is at the genus level.
- 16S rRNA gene, ~1.5Kbp, with:
 - universal, conserved regions, which can be used to generate primers;
 - nine variable regions which can be used as signatures to determine genus/species.



Yarza, P., Yilmaz, P., Pruesse, E., Glockner, F. O., Ludwig, W., Schleifer, K. H., Whitman, W. B., Euzeby, J., Amann, R. & Rossello-Mora, R. (2014). Uniting the classification of cultured and uncultured bacteria and archaea using 16S rRNA gene sequences. *Nat Rev Microbiol.* 12, 635-645.

16S rRNA process overview



16S rRNA sequencing in the clinic

Church, D. L., Cerutti, L., Gurtler, A., Griener, T., Zelazny, A. & Emler, **S.** (2020). Performance and Application of 16S rRNA Gene Cycle Sequencing for Routine Identification of Bacteria in the Clinical Microbiology Laboratory. *Clin* Microbiol Rev. 33, e00053-1

Sample Extraction

- · Verify method from various clinical samples/isolates being tested
- Use manual DNA extraction for up to 8-12 samples but automated method for high sample number
- Check DNA purity and concentration (i.e., NanoDrop)



FAST PCR

- Use High Tm (~60°C) 16S UMD primers + thermostable DNA polymerase
- Use FAST reagents used in FAST cycle-sequencing protocol (i.e., 1 h vs. 2.5 h for conventional PCR)
- Use FAST-ramping thermocycler



PCR Product Clean-Up

- ExoSAP-IT or equivalent enzymatic PCR product clean-up preferred
- Clean-up reaction removes unincorporated primers
- · Unincorporated nucleotides also degraded
- PCR product ready to use for sequencing without additional purification step (i.e., column purification kits)



FAST Cycle Sequencing

- Procedure similar to FAST PCR conditions
- Use BOTH forward (F) and reverse (R) primers = bidirectional read of rDNA
- · Reaction mix contains dNTPs and dye-labelled ddNTPs and decreased concentrations so extension is not inhibited
- Big Dye Xterminator extension product clean-up

Exercise, searching an NCBI database with 16SrRNA sequences

16s rRNA BLAST search online (1)

- Navigate to NCBI BLAST home page:
 - https://blast.ncbi.nlm.nih.gov/Blast.cgi
- Click on:







- https://github.com/WCSCourses/Molecular Approaches Clinical Microbiolog y 2024/blob/main/course data/bioinformatics/16S/16S samples.fas
- https://tinyurl.com/228hpuev

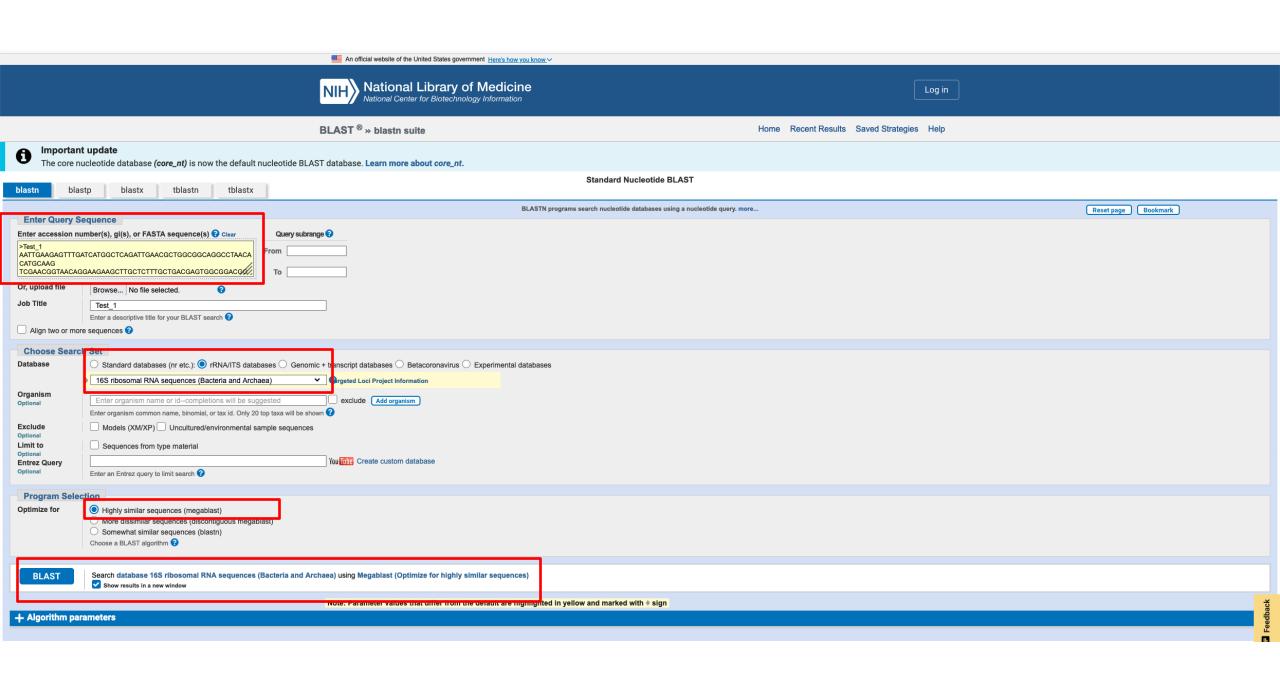












What bacterium is it?

- Look at the results of the search (scroll down the page as necessary).
- What is the likely organism that the 16S rRNA sequence came from?

Sample	Candidate Bacterium
Test 1	Escherichia/Shigella?
Test 2	Mycobacterium ?
Test 3	Vibrio cholerae ?

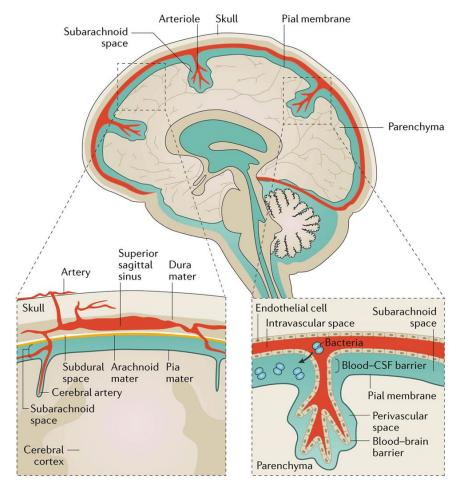
Encapsulated Theme: applications of molecular techniques

Molecular Approaches to Clinical Microbiology in Africa 2024 Martin Maiden, Keith Jolley



Meningitis

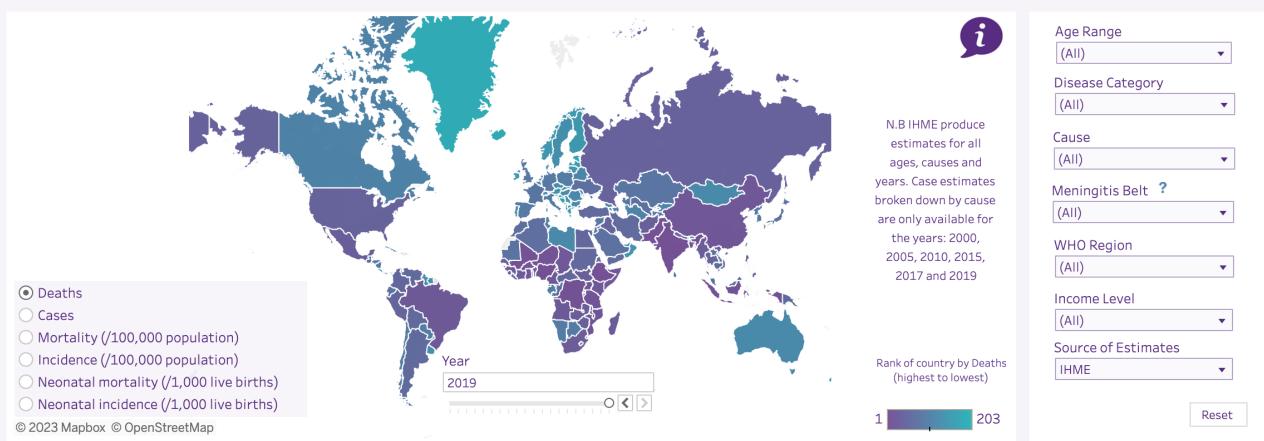
- Meningitis:
 - Inflammation of the meninges (tissues around the brain).
- Caused by invasion of the meninges by a pathogen:
 - Bacterial, viral, fungal, parasite;
 - Can be accompanied by other pathologies e.g. septicaemia (blood poisoning).
- Severe and frequently fatal especially when caused by bacteria:
 - Survivors frequently suffer sequalae e.g. digit or limb loss, brain damage, deafness.
- Diagnosis difficult
 - but essential for swift appropriate treatment.



Rodrigues, C. M. C. & Maiden, M. C. J. (2018). A world without bacterial meningitis: how genomic epidemiology can inform vaccination strategy. *F1000Res* **7,** 401.



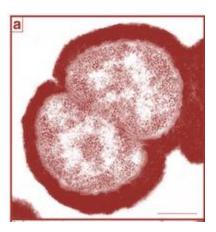
Meningitis and Neonatal Sepsis



Estimated Total Deaths 462,452

Estimated Total Cases 8,427,054

Meningitis causing organisms

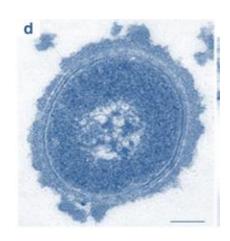


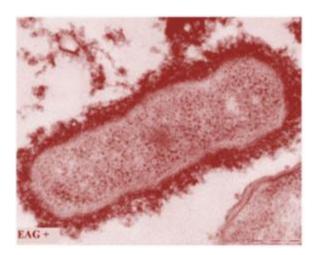
Neisseria meningitidis Meningococcus 12 capsular serogroups Genome: 2.27Mbp

Ganesh, K. *et al.* (2017) Molecular characterization of invasive capsule null *Neisseria meningitidis* in South Africa. *BMC Microbiol.* **17(1)**, 40.

Streptococcus pneumoniae
Pnuemococcus
>100 capsular serotypes
Genome: 2.04Mbp

Ndlangisa, K.M., et al. (2016) Two cases of serotypeable and non-serotypeable variants of *Streptococcus pneumoniae* detected simultaneously during invasive disease. *BMC Microbiol.* 16, 126.





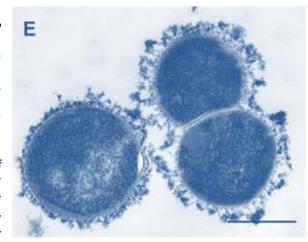
Haemophilus influenzae 'Hib' 6 capsular serotypes Genome: 1.83Mbp

Schouls L., et al. (2008) Two variants among *Haemophilus influenzae* serotype b strains with distinct *bcs4*, *hcsA* and *hcsB* genes display differences in expression of the polysaccharide capsule. BMC Microbiol. **8,** 35.

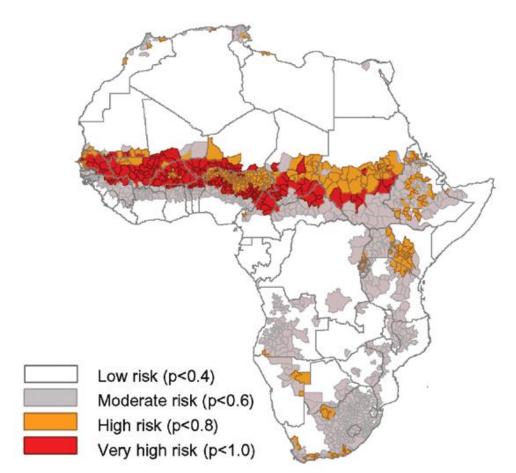
Group B Strep. (GBS)

10 capsular serotypes
Genome: 2.16Mbp

Lecours M.P., et al. (2012) Sialylation of Streptococcus suis serotype 2 is essential for capsule expression but is not responsible for the main capsular epitope. Microbes Infect. 14(11),



The meningitis belt



Meningitis belt extends from Ethiopia to Senegal.

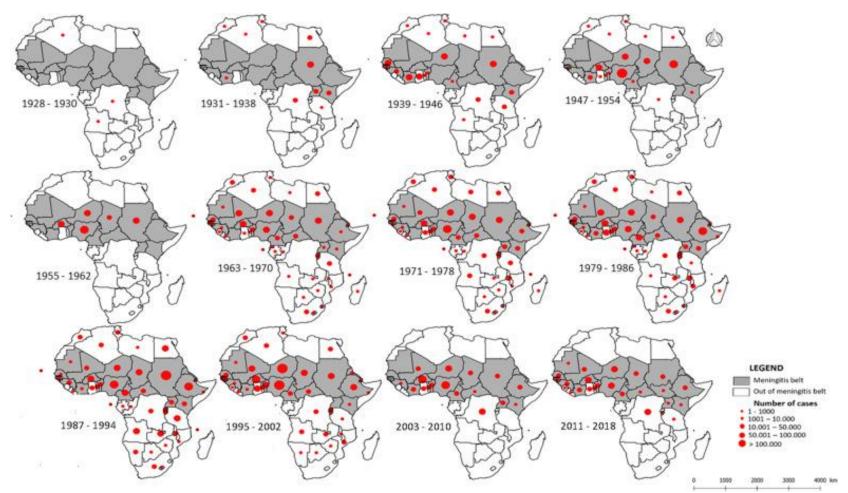
Sudan, Ethiopia, Chad, Niger, Northern Nigeria, Burkina Faso, and Mali are considered hyperendemic.

- 1905, first documented epidemic, Northern Nigeria.
- 1919-1924, second cycle, >45,000 deaths, Northern Nigeria.
- **1935-1937**, third cycle, Nigeria, 6,456 deaths.
- 1951-60, 340,000 cases, 53,000 deaths.
- 1996-1997, 300,000 cases, 30,000 deaths

Molesworth, A. M., Cuevas, L. E., Connor, S. J., Morse, A. P. & Thomson, M. C. (2003). Environmental risk and meningitis epidemics in Africa. *Emerg Infect Dis* **9**, 1287-1293.

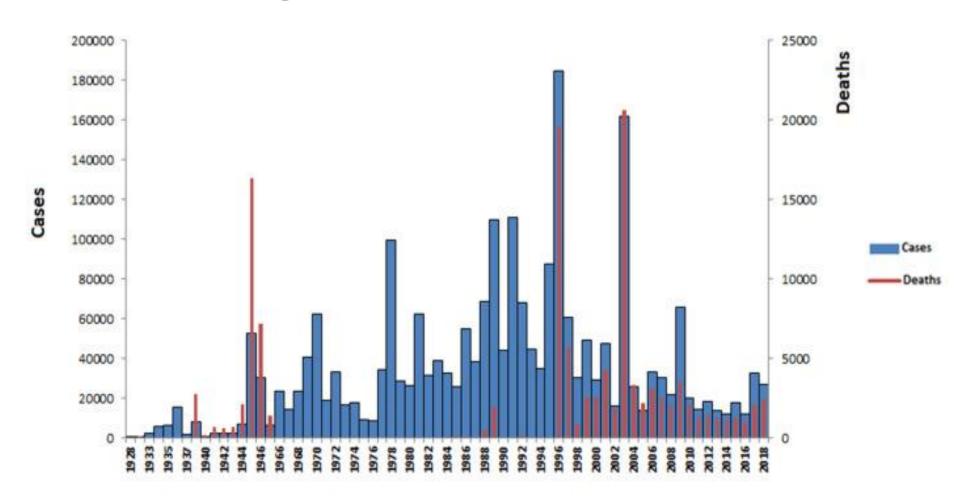
Greenwood, B. (1999). Manson Lecture. Meningococcal meningitis in Africa. Trans R Soc Trop Med Hyg 93, 341-353

Meningococcal disease in Africa



Mazamay, S., Guegan, J. F., Diallo, N., Bompangue, D., Bokabo, E., Muyembe, J. J., Taty, N., Vita, T. P. & Broutin, H. (2021). An overview of bacterial meningitis epidemics in Africa from 1928 to 2018 with a focus on epidemics "outside-the-belt". *BMC Infect Dis.* 21, 1027.

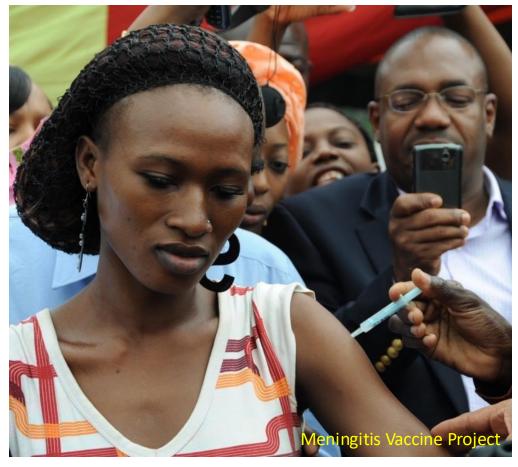
Meningococcal disease in Africa



Mazamay, S., Guegan, J. F., Diallo, N., Bompangue, D., Bokabo, E., Muyembe, J. J., Taty, N., Vita, T. P. & Broutin, H. (2021). An overview of bacterial meningitis epidemics in Africa from 1928 to 2018 with a focus on epidemics "outside-the-belt". *BMC Infect Dis.* 21, 1027.

MenAfriVac®: combatting epidemic meningococcal disease in Africa

- Meningitis Vaccine Project (MVP):
 - Gates funded, partnership with WHO and PATH;
 - Innovative development Northern technology, southern manufacturer;
 - Low cost (less than \$1 per dose, for sustainable use;
- MenAfriVac®, a polysaccharide-protein conjugate vaccine, introduced 2010 in Burkina Faso;
 - Everyone aged 1-29 years immunised;
 - Immediate reduction in disease levels.



Diomande, F. V. K., Djingarey, M. H., Daugla, D. M., Novak, R. T., Kristiansen, P. A., Collard, J. M., Gamougam, K., Kandolo, D., Mbakuliyemo, N., Mayer, L., Stuart, J., Clark, T., Tevi-Benissan, C., Perea, W. A., Preziosi, M. P., LaForce, F. M., Caugant, D., Messonnier, N., Walker, O. & Greenwood, B. (2015). Public Health Impact After the Introduction of PsA-TT: The First 4 Years. *Clinical Infectious Diseases* 61, S467-S472.





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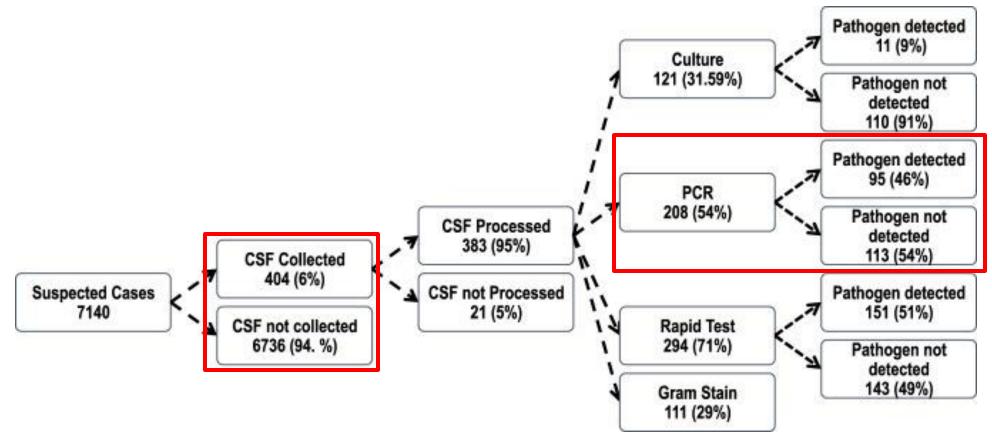


Global road map priorities for response, care and prevention

- Achievement of high immunization coverage, development of new affordable vaccines, and improved prevention strategies and outbreak response;
- Speedy diagnosis and optimal treatment for patients;
- Good data to guide prevention and control efforts;
- Care and support for those affected, focusing on early recognition and improved access to care and support for after-effects; and
- Advocacy and engagement, to ensure high awareness of meningitis, accountability for national plans, and affirmation of the right to prevention, care and after-care services.

https://www.who.int/news-room/events/detail/2024/04/26/default-calendar/the-first-high-level-meeting-to-defeat-meningitis---institut-pasteur-paris--france

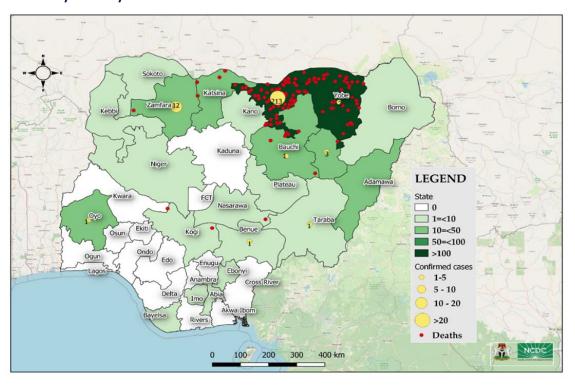
Increasing importance of molecular diagnosis and isolate characterisation in Africa



Kwambana-Adams, B. A., Amaza, R. C., Okoi, C., Rabiu, M., Worwui, A., Foster-Nyarko, E., Ebruke, B., Sesay, A. K., Senghore, M., Umar, A. S., Usman, R., Atiku, A., Abdullahi, G., Buhari, Y., Sani, R., Bako, H. U., Abdullahi, B., Yarima, A. I., Sikiru, B., Moses, A. O., Popoola, M. O., Ekeng, E., Olayinka, A., Mba, N., Kankia, A., Mamadu, I. N., Okudo, I., Stephen, M., Ronveaux, O., Busuttil, J., Mwenda, J. M., Abdulaziz, M., Gummi, S. A., Adedeji, A., Bita, A., Omar, L., Djingarey, M. H., Alemu, W., D'Alessandro, U., Ihekweazu, C. & Antonio, M. (2018). Meningococcus serogroup C clonal complex ST-10217 outbreak in Zamfara State, Northern Nigeria. *Scientific reports*. 8, 14194.

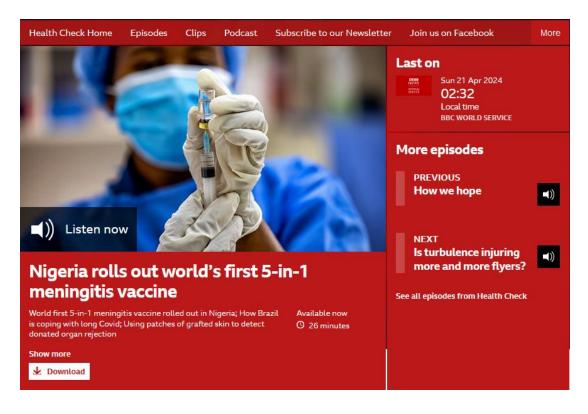
Meningitis in Nigeria 2022-2024

1686 suspected cases 1 October 2022 - 16 April 2023 https://www.who.int/emergencies/disease-outbreak-news/item/2023-DON454



5 in 1 meningitis vaccine introduced

https://www.bbc.co.uk/programmes/w3ct5t8d



Ashinze, P., Mafua, N. & Obafemi, E.(2024). Nigeria rolls out novel meningitis vaccine. Lancet. 403, 2373.

Exercise, searching an NCBI database with *Neisseria* 16S rRNA sequences

16s rRNA BLAST search online (2)

- Navigate to NCBI BLAST home page:
 - https://blast.ncbi.nlm.nih.gov/Blast.cgi
- Click on:









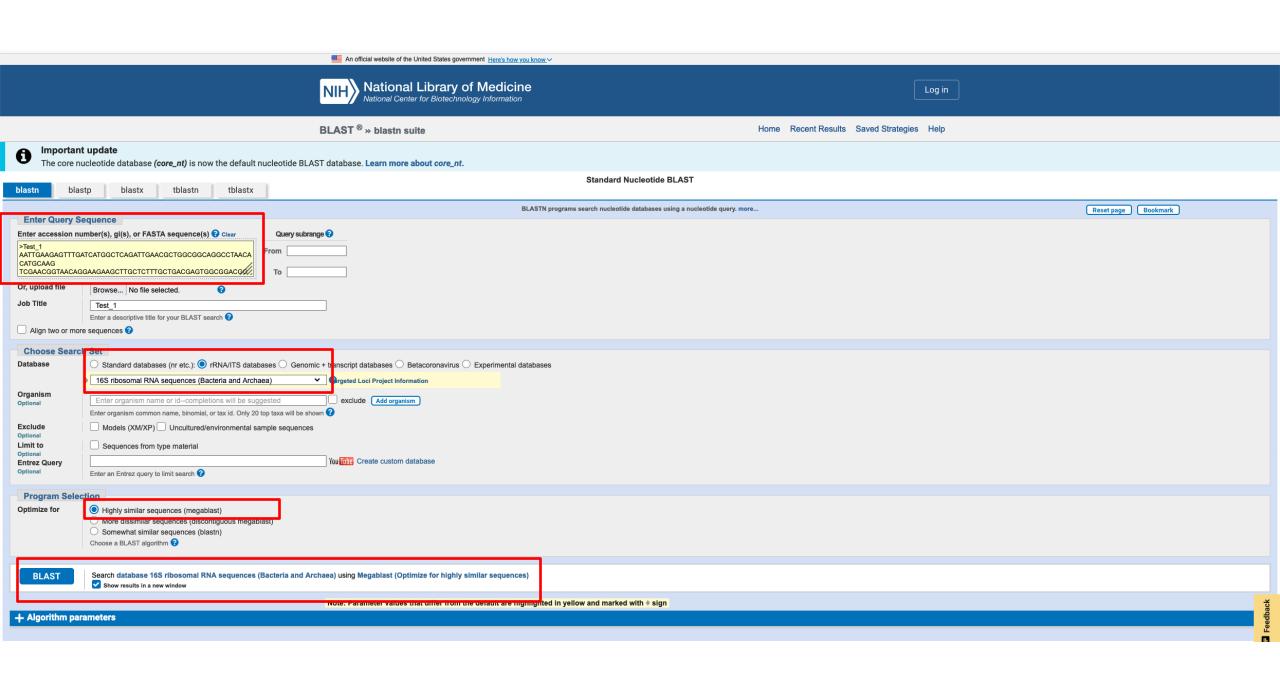


- Under 'Choose Search Set' click on 'rRNA/ITS databases'
- Check box 'Show results in a new window'. Click on







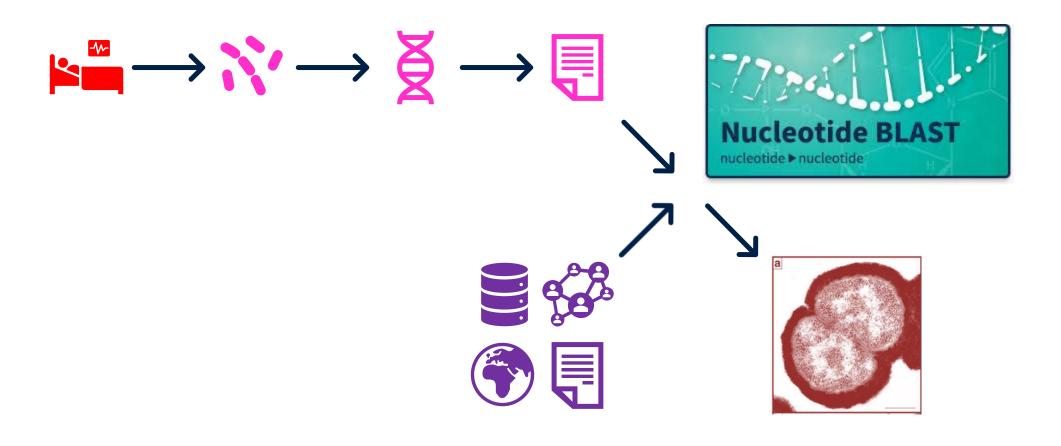


What bacterium is it?

- Look at the results of the search (scroll down the page as necessary).
- What is the likely organism that the 16S rRNA sequence came from?

Sample	Candidate Bacterium
Test 4	Neisseria mucosa
Test 5	Neisseria gonorrheoae
Test 6	Neisseria meningitidis

16S rRNA summary



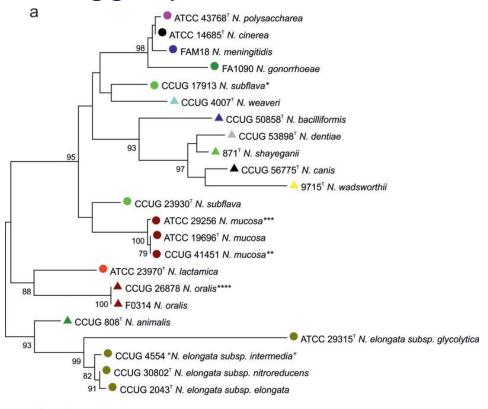
Case study: rplF assay

- **Problem:** rapid cost effective determination of *Neisseria* species from 1000s of isolates obtained in the MenAfriCar surveys.
 - 16s rRNA sequencing too cumbersome and insufficient discrimination or resolution.
- **Solution:** identify a short sequenceable gene fragment (~400bp in length) diagnostic for species.
- Implementation: Phylogeny of Neisseria species generated and compared to phylogenies of individual genes
 - A fragment of rplF gene was congruent with clusters in the phylogeny of all genes and used for the assay.

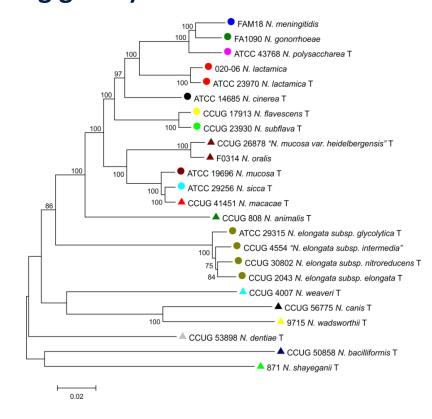
Bennett, J. S., Watkins, E. R., Jolley, K. A., Harrison, O. B. & Maiden, M. C. (2014). Identifying *Neisseria* species using the 50S ribosomal protein L6 (*rplF*) gene. *J Clin Microbiol* **52**, 1375-1381.

Comparison of 16s rRNA sequencing and *rplF* sequencing

16s rRNA gene fragment phylogeny (single rRNA encoding gene)



rMLST phylogeny (53 ribosomal protein encoding genes)



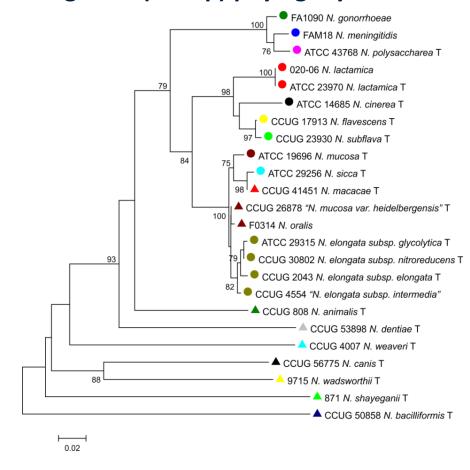
Bennett, J. S., Watkins, E. R., Jolley, K. A., Harrison, O. B. & Maiden, M. C. (2014). Identifying *Neisseria* species using the 50S ribosomal protein L6 (*rplF*) gene. *J Clin Microbiol* 52, 1375-1381.

Rapid species assignment: rplF sequence

rMLST phylogeny (53 genes)

FAM18 N. meninaitidis FA1090 N. gonorrhoeae ATCC 14685 N. cinerea T CCUG 17913 N. flavescens T – 🍑 CCUG 23930 N. subflava T 100 ▲ CCUG 26878 "N. mucosa var. heidelbergensis" T ▲ F0314 N. oralis ATCC 19696 N. mucosa T ATCC 29256 N. sicca T ATCC 29315 N. elongata subsp. glycolytica T 9715 N. wadsworthii T CCUG 50858 N. bacilliformis T 📤 871 N. shaveqanii T 0.02

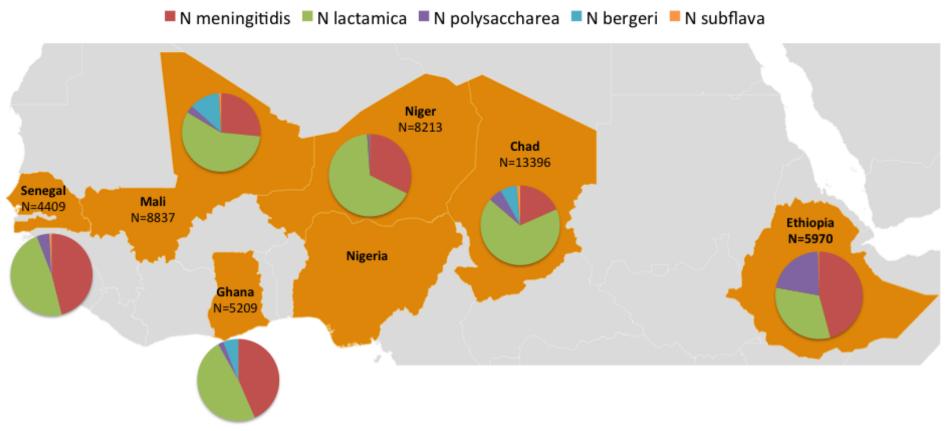
rplF fragment (413bp) phylogeny



Bennett, J. S., Watkins, E. R., Jolley, K. A., Harrison, O. B. & Maiden, M. C. (2014). Identifying *Neisseria* species using the 50S ribosomal protein L6 (*rplF*) gene. *J Clin Microbiol* 52, 1375-1381.

Neisseria species distribution menafricar





Diallo, K., et al., Greenwood, B. M. & Maiden, M. C. (2016). Pharyngeal carriage of *Neisseria* species in the African meningitis belt. *J Infect* **72**, 667-677.

