

Viewing the Companion output in Artemis

Learning objectives:

- Download embl files from Companion
- Open embl files in Artemis
- Viewing and interpreting the results of Companion in Artemis

In the next exercise we will examine the Companion output in more detail. We will use Artemis to have a closer look. First, we need to download the files. Go to the tab ‘Result files’ and download the embl file ‘Pseudochromosome level sequence and annotation’. The file is called embl.tar.gz.

Pcoa-Pkno (PCOA)

Completed

This job was submitted about 9 hours ago and ran for about 2 hours, finally finishing at 2022-10-09 16:00:19 UTC.

Genome statistics

Result files

Orthology

Phylogeny

Synteny

Job parameters

Pipeline logs

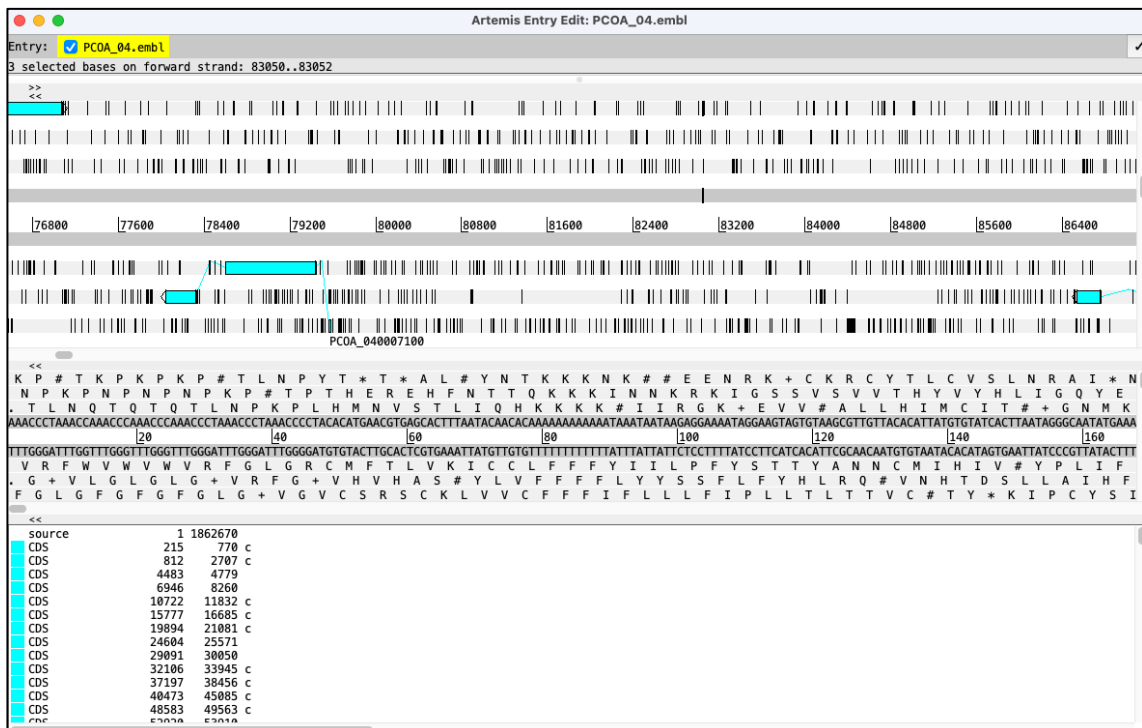
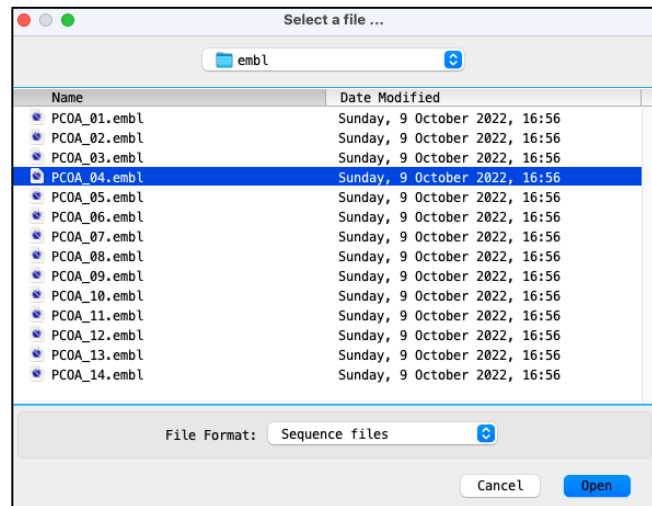
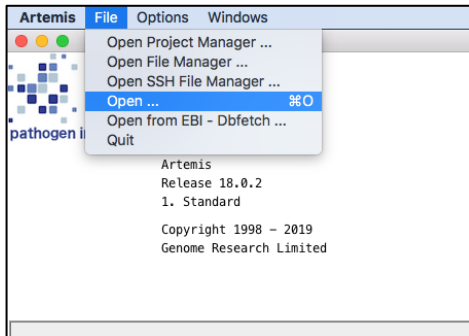
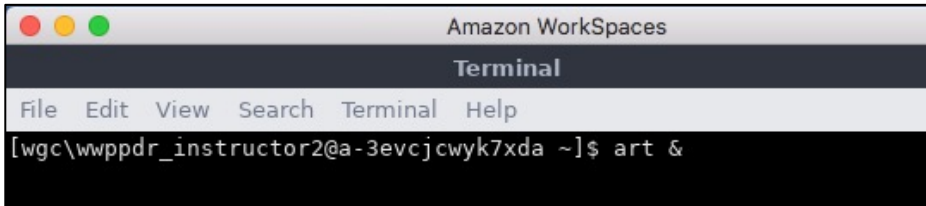
Validator report

	Format	MD5	Size
Pseudochromosome level genomic sequence	FASTA		7.79 MB
Pseudochromosome level gene annotations	GFF3		9.38 MB
Pseudochromosome layout	AGP		618 Bytes
Scaffold level genomic sequence	FASTA		7.79 MB
Scaffold level gene annotations	GFF3		9.43 MB
Scaffold layout	AGP		825 Bytes
Pseudochromosome level sequence and annotation	EMBL		13.9 MB
Gene Ontology function assignments	GAF1		710 KB
Protein sequences	FASTA		4.68 MB

Locate the file called embl.tar.gz in your download folder. Double click on the file. A new folder called ‘embl’ will be created that contains all embl files. Here is the output for *P. coatneyi*. The file contains all chromosomes that were assembled and annotated by Companion. Contigs that could not be placed on one of the chromosomes are in the file *00.embl

PCOA_01.embl	Yesterday at 16:56	1.8 MB	sequence
PCOA_02.embl	Yesterday at 16:56	1.5 MB	sequence
PCOA_03.embl	Yesterday at 16:56	1.8 MB	sequence
PCOA_04.embl	Yesterday at 16:56	3 MB	sequence
PCOA_05.embl	Yesterday at 16:56	2.4 MB	sequence
PCOA_06.embl	Yesterday at 16:56	2.1 MB	sequence
PCOA_07.embl	Yesterday at 16:56	3 MB	sequence
PCOA_08.embl	Yesterday at 16:56	3.2 MB	sequence
PCOA_09.embl	Yesterday at 16:56	4.1 MB	sequence
PCOA_10.embl	Yesterday at 16:56	2.8 MB	sequence
PCOA_11.embl	Yesterday at 16:56	4.2 MB	sequence
PCOA_12.embl	Yesterday at 16:56	7.8 MB	sequence
PCOA_13.embl	Yesterday at 16:56	3.1 MB	sequence
PCOA_14.embl	Yesterday at 16:56	3.2 MB	sequence

Artemis is a great tool to visualise your Companion output. Choose one of the chromosomes you've just downloaded and open it in Artemis. To do so, open a Terminal on your Workspace type “**art &**” and hit return. As an example chromosome 4 of *P. coatneyi* (PCOA_04.embl) is shown.



Once you have your Artemis window open, scroll along the chromosome. Do you find any problems in the annotation? Can you see any missing genes? How many pseudogenes can you find? (Hint: search for the qualifier: pseudo). Do you think all of the pseudogenes are real or are some misannotated? You can answer this question quite easily by using a tool called ACT, Artemis Comparison Tool. We will show you how to use it in the next step!

Comparative Genomics

Visualising the Companion output in ACT

Learning objectives:

- Opening ACT and loading files into ACT
- Viewing and interpreting Companion results in ACT
- Download of GFF files from VEuPathDB
- Creating ACT comparison files with NCBI blast
- Optional exercise: Open a 3-way comparison in ACT

Introduction

In the next part of the exercise we will explore the Companion output in more detail with a tool called Artemis Comparison Tool (ACT). ACT was written by Kim Rutherford and was designed to extract the additional information that can only be gained by comparing the growing number of sequences from closely related organisms (Carver *et al.* 2005). ACT is based on Artemis, so you will already be familiar with many of its core functions. It is essentially composed of three layers or windows. The top and bottom layers are mini Artemis windows (with their inherited functionality), showing the linear representations of the DNA sequences with their associated features. The middle window shows red and blue blocks, which span this middle layer and link conserved regions within the two sequences, in the forward and reverse orientation respectively. Consequently, if you were comparing two identical sequences in the same orientation you would see a solid red block extending over the length of the two sequences in this middle layer. If one of the sequences was reversed, and therefore present in the opposite orientation, there would be a blue 'hour glass' shape linking the two sequences. Unique regions in either of the sequences, such as insertions or deletions, would show up as breaks (white spaces) between the solid red or blue blocks.

In order to use ACT to investigate your own sequences of interest you will have to generate your own pairwise comparison files. Data used to draw the red or blue blocks that link conserved regions is generated by running pairwise BLASTN or TBLASTX comparisons of the sequences. ACT is written so that it will read the output of several different comparison file formats; these are outlined in Appendix III. Two of the formats can be generated using BLAST software freely downloadable from the NCBI, which can be loaded and run on a PC or Mac. You can also use the online BLAST web server from NIH-NCBI to produce an alignment file that can be loaded into ACT. This option can only be used with BLASTN. We will cover this option in this Module.

Aims

The aim of this Module is for you to become familiar with the basic functions of ACT.

In the first part of this exercise you will learn the basic functions of ACT by looking at the companion output of *P. coatneyi* compared to *P. knowlesi*. By comparing two chromosomes you will be able to study the degree of conservation of gene order and identify small and large synteny breaks. You can also look for incorrectly annotated genes.

Once you are familiar with ACT we will show you how to create your own comparison file and explore your Companion output in ACT.

Part 1: Starting up the ACT software

In the first part of this exercise we will all use the same files. Make sure you're in the **Module_2_Comparative_Genomics** directory.

Then type

act & [return]

A small start up window will appear.

To open ACT you can also double click the ACT icon on your Desktop.

The files that you are going to need are:

PCOA_04.embl

PCOA_04_comp_PKNH_04

PKNH_04_v2.embl

- embl file created by Companion
- tblastx comparison file
- *P. knowlesi* chr4 (reference used in Companion run)

Click 'File' then 'Open'

Use the File manager to drag and drop files.

Choose 'All Files'

For comparing more than two DNA files!

Click 'Apply' and wait

For more info on comparison files see Appendix III.

Instead of dragging and dropping the files, you can also choose them.

ACT File Options Windows

Open ...
Open SSH File Manager ...
Quit

institute
pathogen informatics

Artemis Comparison Tool
Release 18.0.2
1. Standard
Copyright 1998 - 2019
Genome Research Limited

File Manager

Name	Size	Modified
Module_2_Comparative_Genomics		Oct 17 10:55 2019
PCOA_04.embl	2875625	Oct 07 10:07 2019
PCOA_04_comp_PKNH_04	4231916	Oct 15 12:02 2019
PKNH_04_v2.embl	2798202	Oct 11 09:39 2019

Sequence file 1 arative_Genomics/PCOA_04.embl **Choose ...**

Comparison file 1 Genomics/PCOA_04_comp_PKNH_04 **Choose ...**

Sequence file 2 tive_Genomics/PKNH_04_v2.embl **Choose ...**

more files ...

Apply **Close**

Choose first sequence ...

Module_2_Comparative_G...

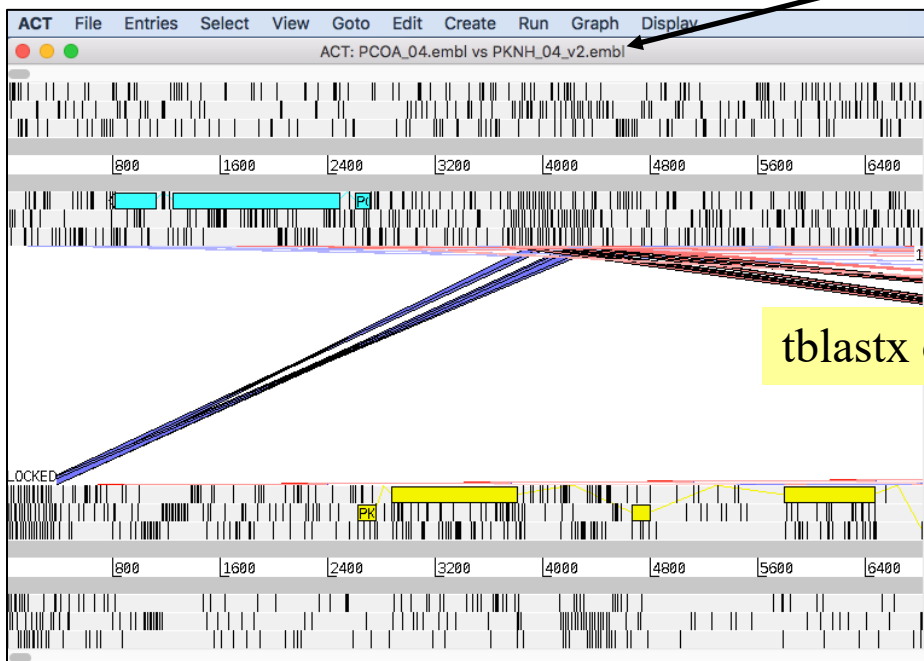
Name	Date Modified
optional_exercise	Thursday, 17 October 2019, 11:10
PCOA_04.embl	Monday, 7 October 2019, 10:07
PCOA_04_comp_PKNH_04	Tuesday, 15 October 2019, 12:02
PKNH_04_v2.embl	Friday, 11 October 2019, 09:39

File Format: All Files

Cancel **Open**

Once you have opened the files you will see a picture like this:

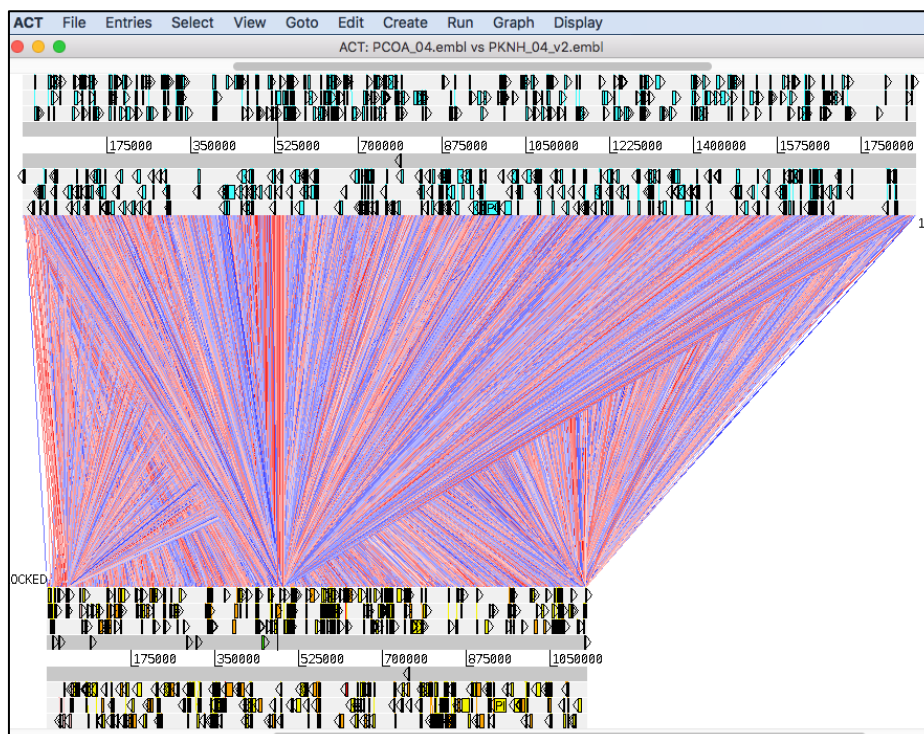
You can see the name of the genomes displayed in ACT on the top of the window.



tblastx comparison

P. coatneyi

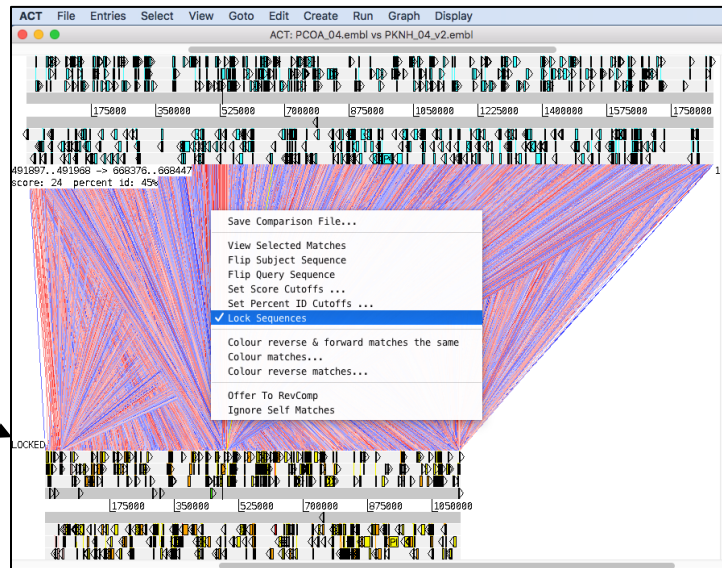
P. knowlesi chr4



1. Use the vertical sliders to zoom out. Drag or click the slider downwards from one of the genomes. The other genome will stay in synch.

When you scroll along with either slider both genomes move together. This is because they are 'locked' together. Right click over the middle comparison view panel. A small menu will appear, select Unlock sequences and then scroll one of the horizontal sliders. Notice that 'LOCKED' has disappeared from the comparison view panel and the genomes will now move independently.

LOCKED



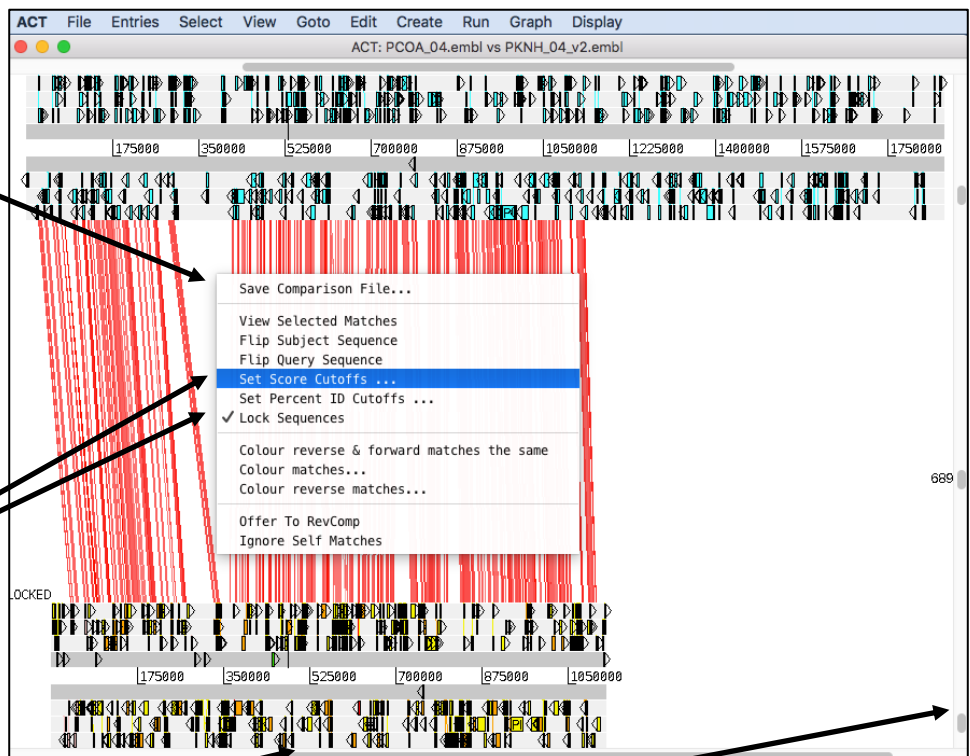
You can optimise your image by either removing 'low scoring' (or percentage ID) hits from view, as shown below 1-3 or by using the slider on the comparison view panel (4). The slider allows you to filter the regions of similarity based on the length of sequence over which the similarity occurs, sometimes described as the "footprint".

1

Right button click in the Comparison View panel

2

Select either 'Set Score Cutoffs' or 'Set Percent ID Cutoffs'

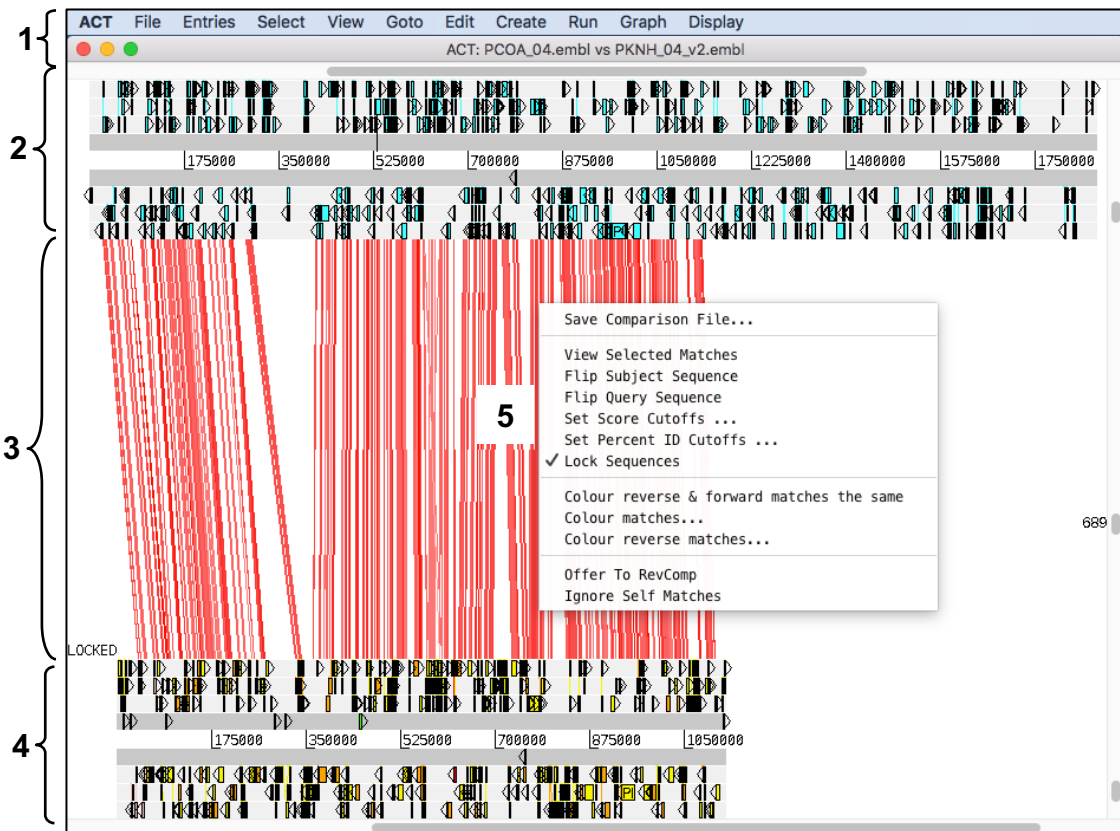


4

3

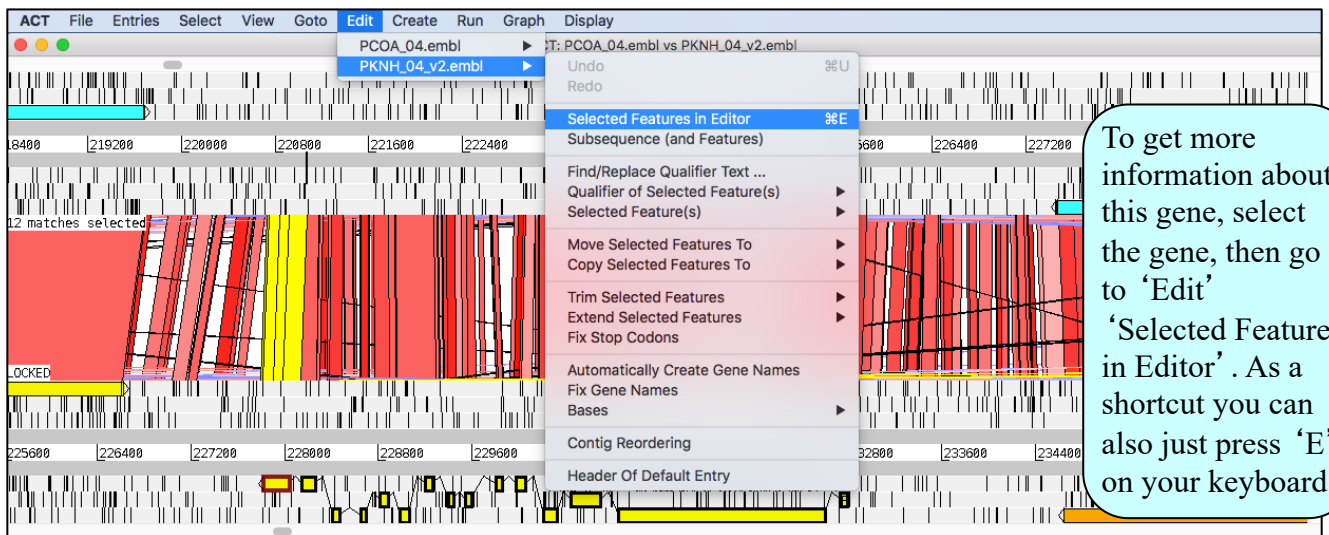
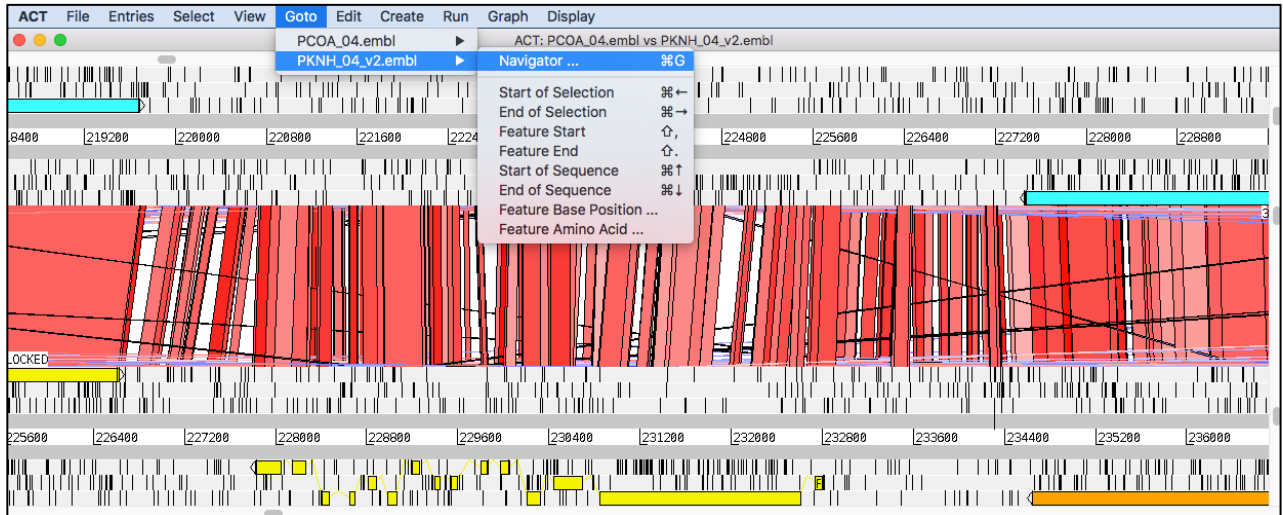
Move the sliders to manipulate the comparison view image.

Now that you have an ACT window open let's look what is in there.

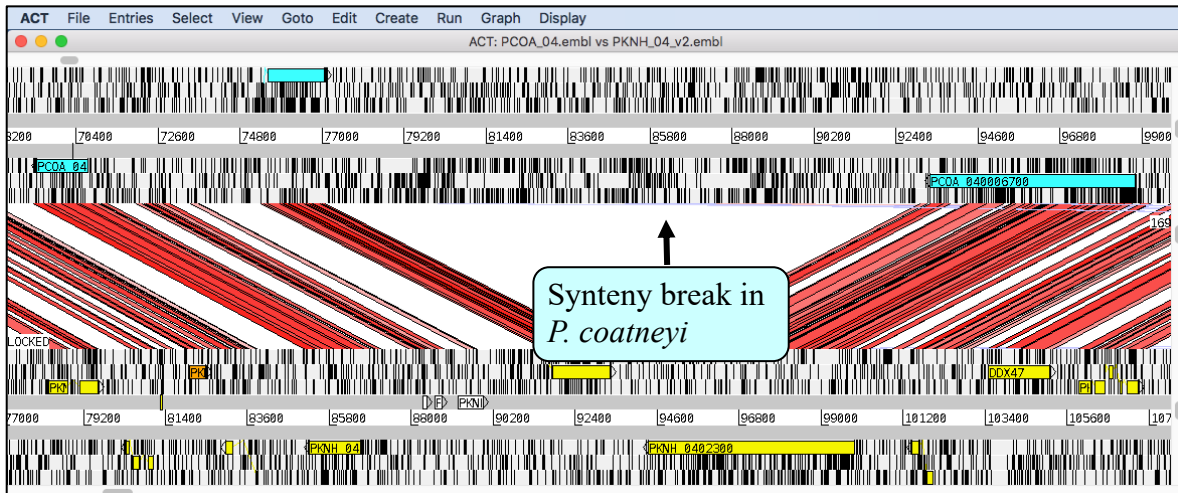


1. Drop-down menus. These are mostly the same as in Artemis. The major difference you will find is that after clicking on a menu header you will then need to select a DNA sequence before going to the full drop-down menu.
2. This is the Sequence view panel for 'Sequence file 1' (Subject Sequence) you selected earlier. It's a slightly compressed version of the Artemis main view panel. The panel retains the sliders for scrolling along the genome and for zooming in and out.
3. The Comparison View. This panel displays the regions of similarity between two sequences. Red blocks link similar regions of DNA with the intensity of red colour directly proportional to the level of similarity. Double clicking on a red block will centralise it. Blue blocks link regions that are inverted with respect to each other.
4. Artemis-style Sequence View panel for 'Sequence file 2' (Query Sequence).
5. Right button click in the Comparison View panel brings up this ACT-specific menu which we will use later.

ACT is a great tool to spot any problems in the automatic Companion annotation. Scroll along the genome to find genes that were missed by Companion. Go to the *P. knowlesi* gene PKNH_0405900 by using the Navigator. Compare it to the *P. coatneyi* annotation on the top. Can you see that this gene has been missed by Companion?



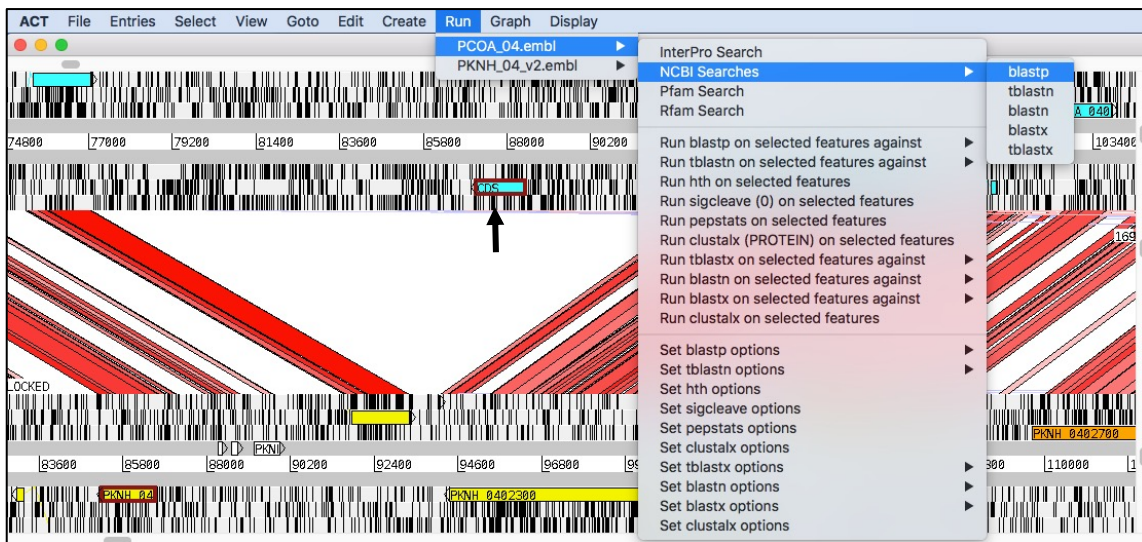
Scroll along the chromosome and try to get an estimate on the number of synteny breaks. Do you think there are any missing genes in the synteny breaks?



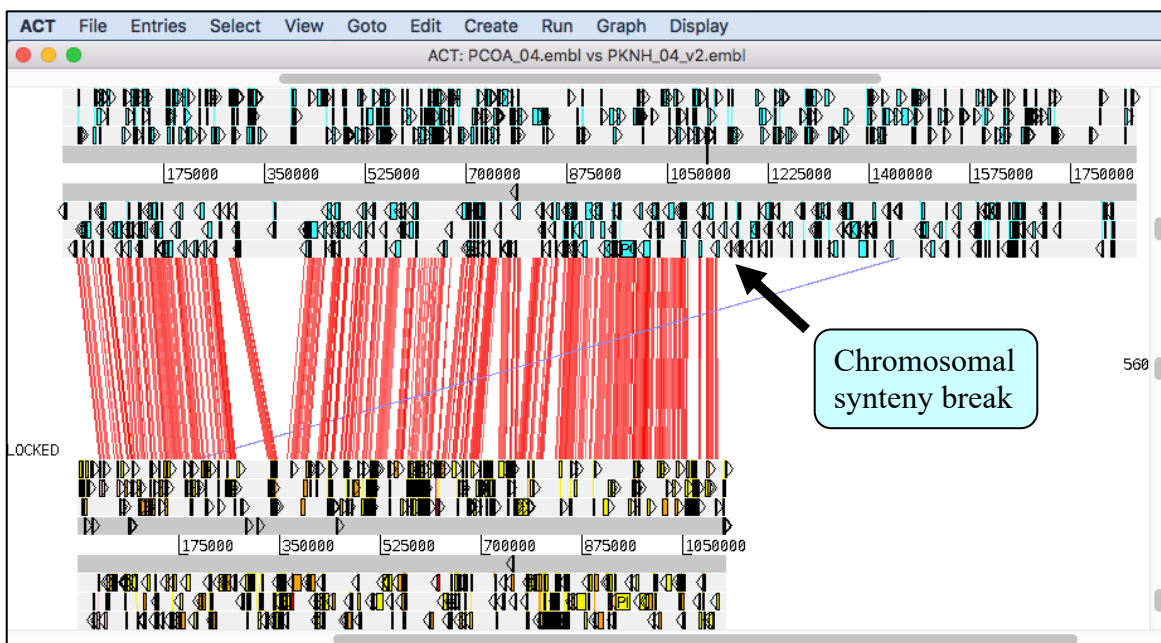
If you think there is a missing gene, you can just mark that area with your mouse. Then go to “Create” and choose “Feature From Base Range”. There is also a shortcut. Just press “C” on your keyboard.



Once you have created a feature, run blast to find out more about the possible missing gene. Can you assign a product?



Can you locate the region of a chromosomal synteny break point?



We will show you in the next part how to open a three-way comparison in ACT and explore the synteny break. This is an optional exercise. You can skip it and proceed to part 2, exploring your own Companion output.

Optional exercise

In this optional exercise we will show you how to open a three-way comparison in ACT and explore the chromosomal synteny break in *P. coatneyi*.

The files you are going to need are:

PKNH_13_v2.embl

- *P. knowlesi* chr13

PKNH_13_comp_PCOA_04

- tblastx comparison file

PCOA_04.embl

- *P. coatneyi* chr04

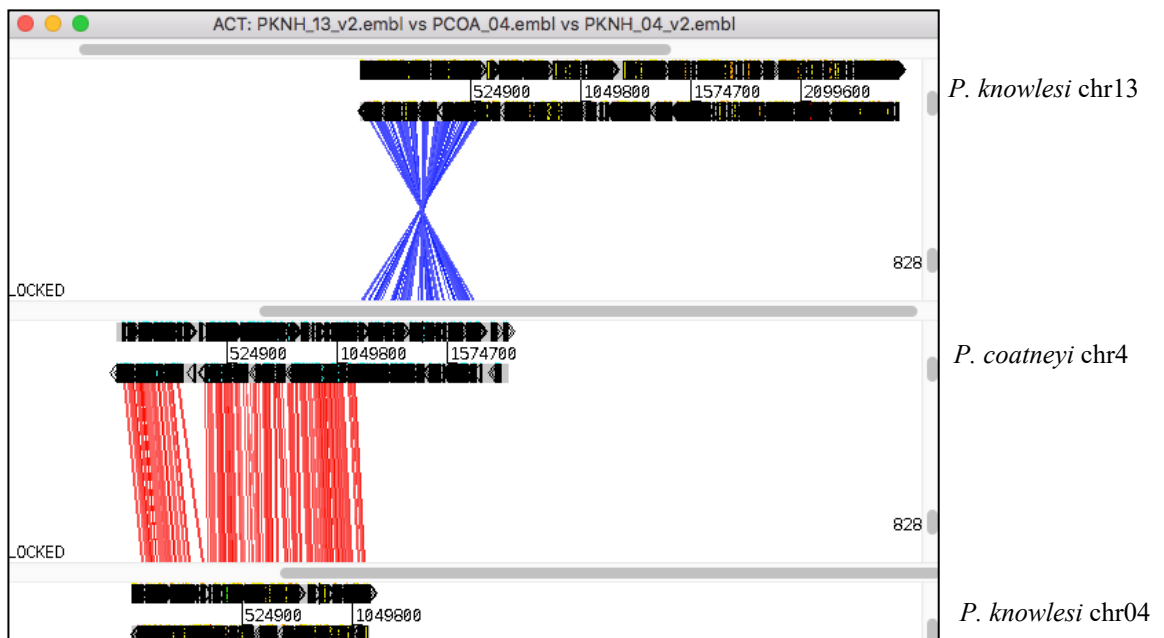
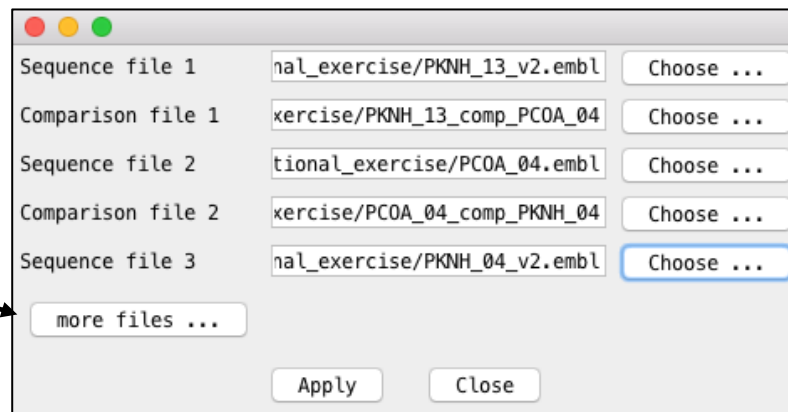
PCOA_04_comp_PKNH_04

- tblastx comparison file

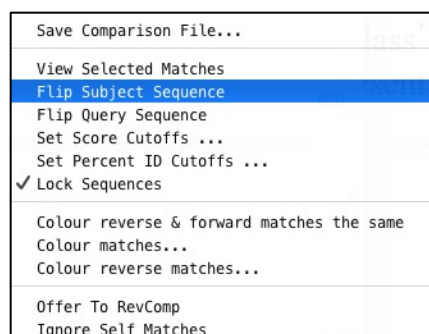
PKNH_04_v2.embl

- *P. knowlesi* chr4

Click on 'more files' to compare more than 2 files.



The blue 'hour glass' shape indicates that one of the chromosomes is reversed. With a right click in the middle area you can get an additional menu. Select 'Flip Subject Sequence' to flip one of the sequences.



Part 2: Explore your own Companion output in ACT

Now that you are familiar with the basic functions of ACT, let's explore your own Companion output. To do so, you need the Companion output (sequence and annotation), a comparison file and the reference (sequence and annotation). Have a look at your Companion output and choose one of the chromosomes you want to explore in more detail.

1. Download sequence and annotation of your Companion run

You've already downloaded the embl files in the first part of this exercise. To create the comparison file, you also need to download the sequence without annotation. Go to the 'Result files' and download the 'Pseudochromosome level genomic sequence'. This is a sequence file that contains all the chromosomes. Extract by copying and pasting the sequence of the chromosome you are interested in.

Cmel-Cpar (CMEL)
Completed

This job was submitted **2 days** ago and ran for **about 1 hour**, finally finishing at **2020-09-09 17:12:39 UTC**.

Genome statistics
Result files
Orthology
Phylogeny
Synteny
Job parameters
Pipeline logs
Validator report

	Format	MD5	Size
Pseudochromosome level genomic sequence	FASTA		2.5 MB
Pseudochromosome level gene annotations	GFF3		2.74 MB
Pseudochromosome layout	AGP		5.12 KB
Scaffold level genomic sequence	FASTA		2.5 MB
Scaffold level gene annotations	GFF3		2.83 MB
Scaffold layout	AGP		2.84 KB
Pseudochromosome level sequence and annotation	EMBL		4.95 MB
Gene Ontology function assignments	GAF1		1.44 MB
Protein sequences	FASTA		2.44 MB

Alternatively, open the embl file you've downloaded from Companion in Artemis and save the sequence as shown below.

2. Download sequence and annotation of your reference genome

Downloading the sequence and annotation of your reference, can either be done on VEuPathDB or on one of the main repositories like GenBank or ENA. How to download your sequence in GenBank is shown in the Appendix.

In VEuPathDB you need to download the annotation and sequence for the chromosome you are interested in separately. In the first part of the Companion exercise we showed you how to download a FASTA file. To download the annotation go to Genes on the left hand side, choose 'Genomic Location' and select 'Organism'. Now select the Chromosome and then click on 'Get Answer'.

Search for...

expand all | collapse all

Filter the searches below...

Genes

- ▶ Annotation, curation and identifiers
- ▶ Function prediction
- ▶ Gene models
- ▶ Genetic variation
- ▼ Genomic Location
 - Genomic Location
 - Proximity to Telomeres
- ▶ Immunology
- ▶ Orthology and synteny

Identify Genes based on Genomic Location

Search by Chromosome | Search by Sequence ID

Organism

Cryptosporidium parvum Iowa II

Chromosome

Choose chromosome

Start at

1

End Location (0 = end)

0

Get Answer

Genomic Loc
483 Genes
Step 1

483 Genes (470 ortholog groups) | Revise this search

Gene Results | Genome View | **Analyze Results**

Rows per page: 20

Download | Add to Basket | Add Columns

Gene ID	Transcript ID	Organism	Genomic Location(s)	Product Description
cgd5_10	cgd5_10-RA	Cryptosporidium parvum Iowa II	CM000433: 3,913 - 6,269 (-)	Uncharacterized Secreted Protein
cgd5_20	cgd5_20-RA	Cryptosporidium parvum Iowa II	CM000433: 6,771 - 9,053 (-)	Uncharacterized Secreted Protein
cgd5_30	cgd5_30-RA	Cryptosporidium parvum Iowa II	CM000433: 10,174 - 10,578 (-)	Uncharacterized protein
Cgd5_40	Cgd5_40-RA	Cryptosporidium parvum Iowa II	CM000433: 10,685 - 16,339 (-)	Uncharacterized Secreted Protein
cgd5_50	cgd5_50-RA	Cryptosporidium parvum Iowa II	CM000433: 16,600 - 19,535 (-)	Uncharacterized Secreted Protein

Download Genes

Results are from search: Genomic Location

Choose a Report:

- ☐ Tab- or comma-delimited (openable in Excel) - choose columns to make a custom table
- ☐ Tab- or comma-delimited (openable in Excel) - choose a pre-configured table
- ☐ FASTA - sequence retrieval, configurable
- ☒ GFF3 - gene models and optional sequences
- ☐ Standard JSON

Generate a report of your query result in GFF3 format

Download Type:

☒ GFF File

☐ Show in Browser

Get GFF3 file

2. Create the ACT comparison file

To create the comparison file, go to the following website:

<https://blast.ncbi.nlm.nih.gov/Blast.cgi> and select 'Nucleotide Blast', 'Align two or more sequences'.

BLAST® » blastn suite Home Recent Results Saved Strategies Help

Standard Nucleotide BLAST

blastn blasto blastx tblastn tblastx

BLASTN programs search nucleotide databases using a nucleotide query. [more...](#) [Reset page](#) [Bookmark](#)

Enter Query Sequence

Enter accession number(s), gi(s), or FASTA sequence(s) [Clear](#)

Query subrange [Clear](#)

From

To

Or, upload file No file chosen [Choose file](#)

Job Title

Enter a descriptive title for your BLAST search [Clear](#)

☒ Align two or more sequences [Clear](#) **←**

Upload the two sequences you would like to compare and then select a blast option.

BLAST® » blastn suite Home Recent Results Saved Strategies Help

Align Sequences Nucleotide BLAST

blastn blasto blastx tblastn tblastx

BLASTN programs search nucleotide subjects using a nucleotide query. [more...](#) [Reset page](#) [Bookmark](#)

Enter Query Sequence

Enter accession number(s), gi(s), or FASTA sequence(s) [Clear](#)

Query subrange [Clear](#)

From

To

Or, upload file mel_5.fasta [Choose file](#)

Job Title

Enter a descriptive title for your BLAST search [Clear](#)

☒ Align two or more sequences [Clear](#)

Enter Subject Sequence

Enter accession number(s), gi(s), or FASTA sequence(s) [Clear](#)

Subject subrange [Clear](#)

From

To

Or, upload file sequenceByS...celd.fasta [Choose file](#)

Program Selection

Optimize for

- ☐ Highly similar sequences (megablast)
- ☐ More dissimilar sequences (discontiguous megablast)
- ☒ Somewhat similar sequences (blastn) **←**

Choose a BLAST algorithm [Clear](#)

Megablast is intended for comparing a query to closely related sequences and works best if the target percent identity is 95% or more but is very fast. Discontiguous megablast uses an initial seed that ignores some bases (allowing mismatches) and is intended for cross-species comparisons. BlastN is slow, but allows a word-size down to seven bases.

[more...](#)

BLAST **←** Search nucleotide sequence using Blastn (Optimize for somewhat similar sequences)

☐ Show results in a new window

The BLAST run will take a few minutes. Once this is done, select the Download option 'Hit Table(text)'. This is the comparison file that you can open in ACT.

BLAST® » blastn suite-2sequences » results for RID-U93AK4KU114

Home Recent Results Saved Strategies Help

[< Edit Search](#) [Save Search](#) [Search Summary](#) [How to read this report?](#) [BLAST Help Videos](#) [Back to Traditional Results Page](#)

Job Title Cmel_5

RID [U93AK4KU114](#) Search expires on 10-15 22:59 pm [Download All](#)

Program Blast 2 sequences [Citation](#)

Query ID lcl|Query_4875 (dna)

Query Descr Cmel_5

Query Length 1074033

Subject ID lcl|Query_4877 (dna)

Subject Descr CM000433 | Cryptosporidium parvum Iowa II | 1 to 1080900

Subject Length 1080900

Other reports [MSA viewer](#)

Filter Results

Percent Identity to **E value** to [Filter](#) [Reset](#)

Descriptions [Graphic Summary](#) [Alignments](#)

Sequences producing significant alignments

☒ select all 1 sequences selected

Description	Max Score	Total Score	Query Cover	E value	Per. Ident	Accession
<input checked="" type="checkbox"/> CM000433 Cryptosporidium parvum Iowa II 1 to 1080900 (reverse-complement)	1.889e+05	1.566e+06	96%	0.0	92.56%	Query_4877

3. Open your file in ACT

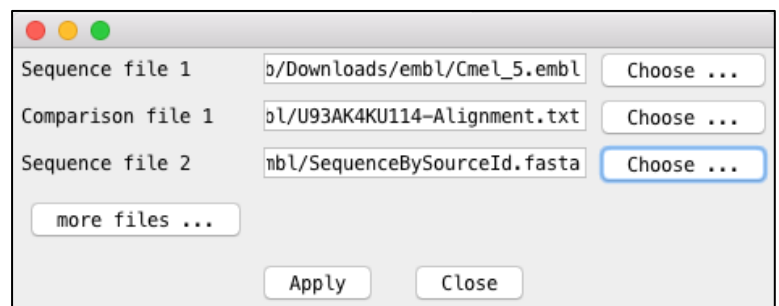
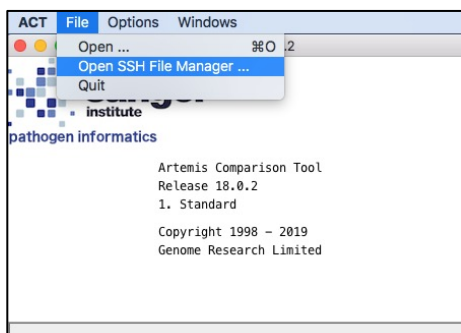
Let's open the two sequences in ACT.

The files that you are going to need are:

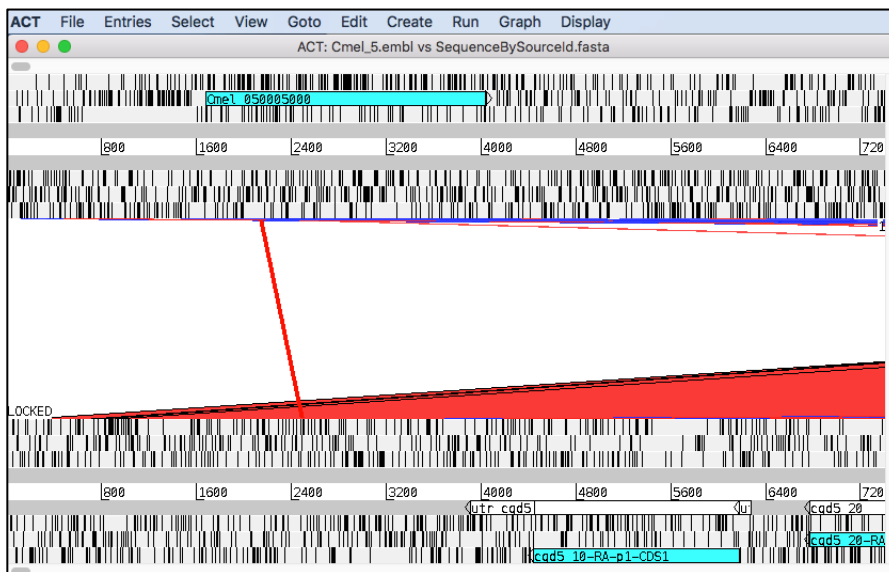
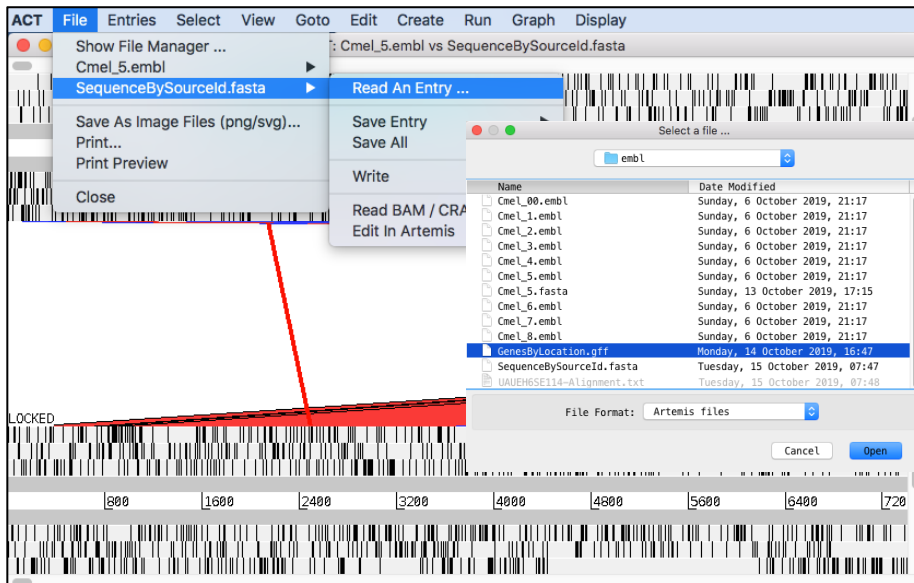
Companion embl file

Comparison file (Hit Table – downloaded from NCBI)

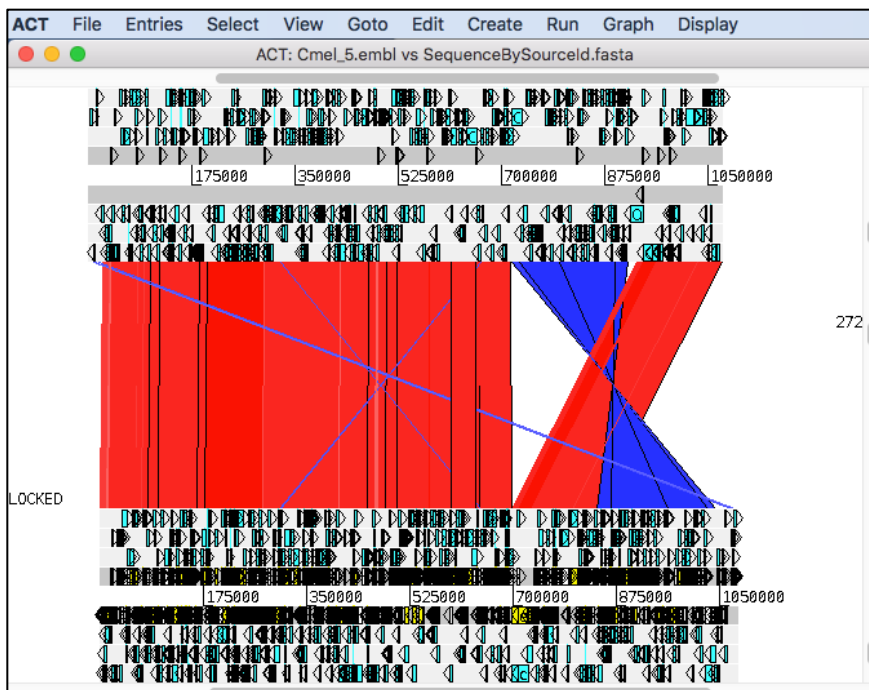
Reference (Fasta file and GFF file downloaded from PlasmoDB)



Once you've opened ACT load in the GFF file you've downloaded from VEuPathDB.



Use the vertical sliders to zoom out to get an overview. Drag or click the slider downwards from one of the genomes.



Once you've got an overview, zoom in to explore interesting areas, i.e. synteny breaks.