

## Customized Tool to visualize protein coverage after Maxquant search for MS data

Tool to show protein coverage across different samples/condition, using the Maxquant output file "peptides.txt"

```
library(tidyverse)
```

```
## Warning: package 'tidyverse' was built under R version 3.5.2
```

```
## -- Attaching packages ----- tidyverse 1.3.0 --
```

```
## v ggplot2 3.3.3    v purrr  0.3.4
## v tibble  3.0.5    v dplyr  0.8.5
## v tidyr   1.1.2    v stringr 1.4.0
## v readr   1.3.1    v forcats 0.5.0
```

```
## Warning: package 'dplyr' was built under R version 3.5.2
```

```
## Warning: package 'stringr' was built under R version 3.5.2
```

```
## Warning: package 'forcats' was built under R version 3.5.2
```

```
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()    masks stats::lag()
```

```
library(reshape2)
```

```
##
```

```
## Attaching package: 'reshape2'
```

```
## The following object is masked from 'package:tidyr':
```

```
##
```

```
## smiths
```

```
library(scales)
```

```
##
```

```
## Attaching package: 'scales'
```

```
## The following object is masked from 'package:purrr':
```

```
##
```

```
## discard
```

```
## The following object is masked from 'package:readr':
##
##   col_factor
```

```
library(splitstackshape)
```

```
## Warning: package 'splitstackshape' was built under R version 3.5.2
```

```
peptides<- as.data.frame(read.delim("peptides.txt", sep="\t",header=TRUE))

names(peptides)
```

```
## [1] "Sequence"          "N.term.cleavage.window" "C.term.cleavage.window"
## [4] "Amino.acid.before" "First.amino.acid"      "Second.amino.acid"
## [7] "Second.last.amino.acid" "Last.amino.acid"      "Amino.acid.after"
## [10] "A.Count"           "R.Count"              "N.Count"
## [13] "D.Count"           "C.Count"              "Q.Count"
## [16] "E.Count"           "G.Count"              "H.Count"
## [19] "I.Count"           "L.Count"              "K.Count"
## [22] "M.Count"           "F.Count"              "P.Count"
## [25] "S.Count"           "T.Count"              "W.Count"
## [28] "Y.Count"           "V.Count"              "U.Count"
## [31] "O.Count"           "Length"               "Missed.cleavages"
## [34] "Mass"              "Proteins"             "Leading.razor.protein"
## [37] "Start.position"    "End.position"         "Gene.names"
## [40] "Protein.names"     "Unique..Groups."      "Unique..Proteins."
## [43] "Charges"           "PEP"                  "Score"
## [46] "Fraction.Average"  "Fraction.Std.Dev."    "Fraction.1"
## [49] "Experiment.1"      "Experiment.2"         "Experiment.3"
## [52] "Experiment.4"      "Experiment.5"         "Experiment.6"
## [55] "Intensity"         "Intensity.1"          "Intensity.2"
## [58] "Intensity.3"       "Intensity.4"          "Intensity.5"
## [61] "Intensity.6"       "Reverse"              "Potential.contaminant"
## [64] "id"                "Protein.group.IDs"     "Mod..peptide.IDs"
## [67] "Evidence.IDs"      "MS.MS.IDs"           "Best.MS.MS"
## [70] "Oxidation..M..site.IDs" "MS.MS.Count"
```

Select the following columns into new dataframe, "Sequence", "Gene.names", "Start.position", "End.position" and the samples intensities (in this case will be columns between 56:61 )

```
select_col_peptides<- select(peptides, 1, 39,37,38,56:61)

names(select_col_peptides)
```

```
## [1] "Sequence"          "Gene.names"          "Start.position" "End.position"
## [5] "Intensity.1"       "Intensity.2"          "Intensity.3"   "Intensity.4"
## [9] "Intensity.5"       "Intensity.6"
```

Clean Up and Melting the dataframe

```

select_col_peptides[select_col_peptides == 0] <- NA
select_col_peptides[, 5:10] <- log(select_col_peptides[,5:10], 2)
select_col_peptides_melt<- melt(select_col_peptides,id.vars = c("Sequence","Gene.names", "Start.position")

#Calculate peptide lenght
select_col_peptides_melt$peptide_length <- (select_col_peptides_melt$End.position -select_col_peptides_

#remove peptides with NA intensities
select_col_peptides_melt<- subset(select_col_peptides_melt, Log2_Intesnsity>0 )

#Since we are only mapping true or false coverage, we remove redundant peptide
select_col_peptides_melt$duplicate_mark<-paste(select_col_peptides_melt$Samples,select_col_peptides_m
select_col_peptides_melt<-select_col_peptides_melt[!duplicated(select_col_peptides_melt$duplicate_mar]
#setting the bar height for visulization
select_col_peptides_melt$Bar_length = 1

#Finally breakdown the peptide into individula aa with corresponding corrdinates

select_col_peptides_melt= select_col_peptides_melt[complete.cases(select_col_peptides_melt), ]

select_col_peptides_melt_aa = select_col_peptides_melt %>%
group_by( Sequence, Gene.names, Samples, Start.position ) %>%
expandRows("peptide_length") %>%
mutate(Amino.acid.position = Start.position - 1 + cumsum(Bar_length))

```

Visualiza the protein coverage for any protein of interest (insert Gene name in Gene.names=="XX", in the code below)

```

ggplot( subset(select_col_peptides_melt_aa, Gene.names=="PABPN1" ), aes(x=Amino.acid.position,y= Bar_lei
geom_bar(stat = "identity", position = "fill")+
facet_wrap(~Samples,ncol = 1)+
#below you can set the full lenght of the protein of interest
xlim(1, 306)+
scale_y_continuous(expand = c(0,0))+
scale_fill_brewer(palette="Dark2")+
theme_classic()+
labs(title = "PABPN1")+
theme(axis.title.y=element_blank(),
axis.text.y=element_blank(),
axis.ticks.y=element_blank(),
axis.line = element_line(colour = 'black', size = 0.25),
axis.ticks = element_line(colour = 'black', size = 0.25),
strip.background = element_blank())+
annotate("segment", x=-Inf, xend=Inf, y=-Inf, yend=-Inf, color="black",size=0.25, linetype="solid")+
annotate("segment", x=-Inf, xend=Inf, y=Inf, yend=Inf, color="black",size=0.25, linetype="solid")+
annotate("segment", x=Inf, xend=Inf, y=-Inf, yend=Inf, color="black",size=0.25, linetype="solid")

```

## PABPN1

