

Walkthrough with SCC dataset

Load the Data

```
addpath('./data')  
load('SCC_dataset.mat')
```

Process data and select features

Remove low-quality cells and select features:

- filter cells don't express min_exp percentage of genes
- select top #gene_selected informative genes

```
min_exp = 0.95; gene_selected = 3000;  
[prodata,progene_name,procell_label] = preprocessing(data, gene_name, true_label, min_exp, ...  
gene_selected);
```

Consensus clustering

If choosing SC3 package to do the consensus clustering on the processed data, we can get the following cell-cell similarity matrix M:

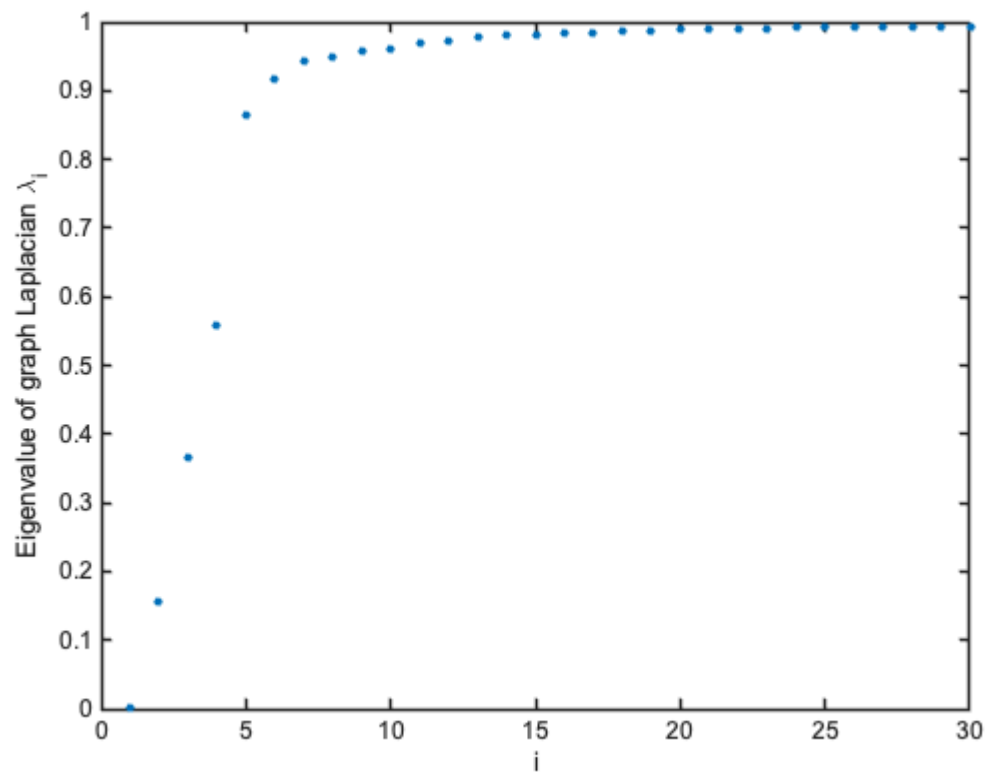
```
M = csvread('SCC_cell-cell.csv');
```

Number of clusters

The number of clusters is estimated by analyzing the largest gap of sorted eigenvalues of symmetric normalized graph Laplacian:

```
[eigenvalues] = plot_eigen_gap(M);
```

Number of cluster based on zero eigenvalues & Largest gap
1 4



```
No_cluster = 4;
```

Run QuanTC

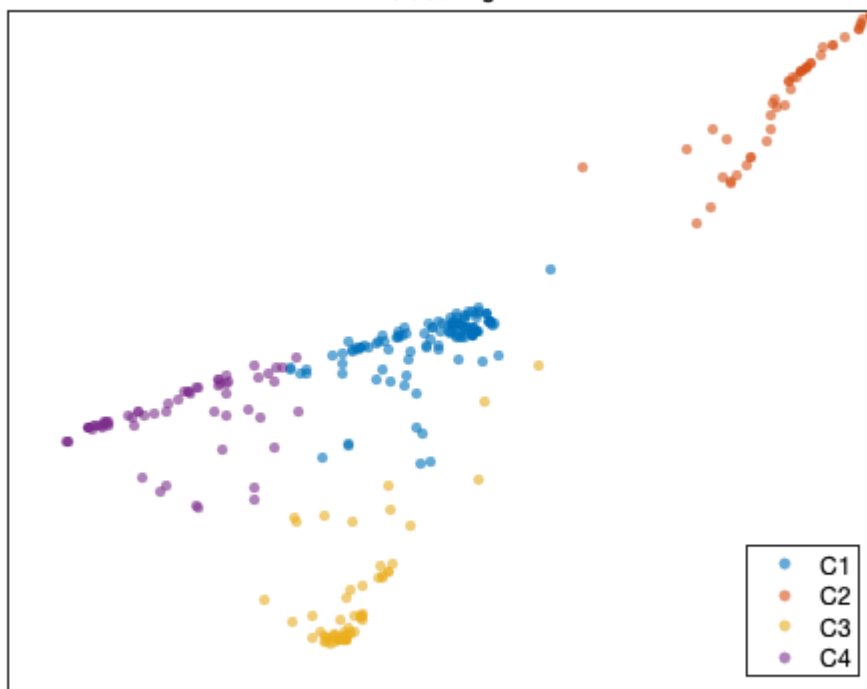
- **Soft clustering:** get the likelihoods of cells belonging to each cluster based on symmetric non-negative matrix factorization of M
- **CPI:** compute CPI value of each cell, use TC_cut to select cell with higher CPI values to be TC

```
TC_cut = 0.34;
[result] = run_QuanTC(prodata,M,No_cluster,TC_cut);
```

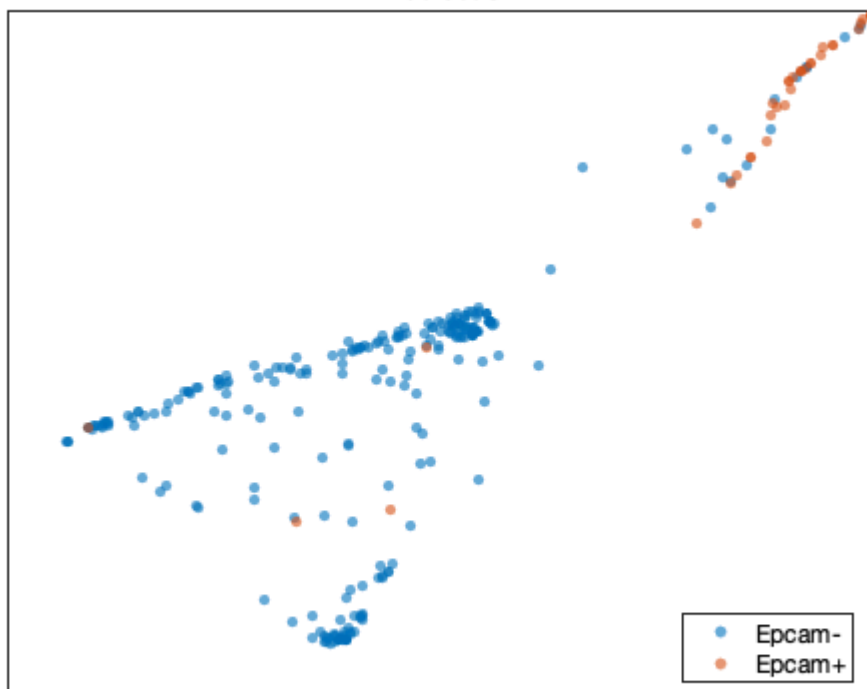
- **2d-visualization:** cells are visualized through the probabilistic regularized embedding (PRE) approach

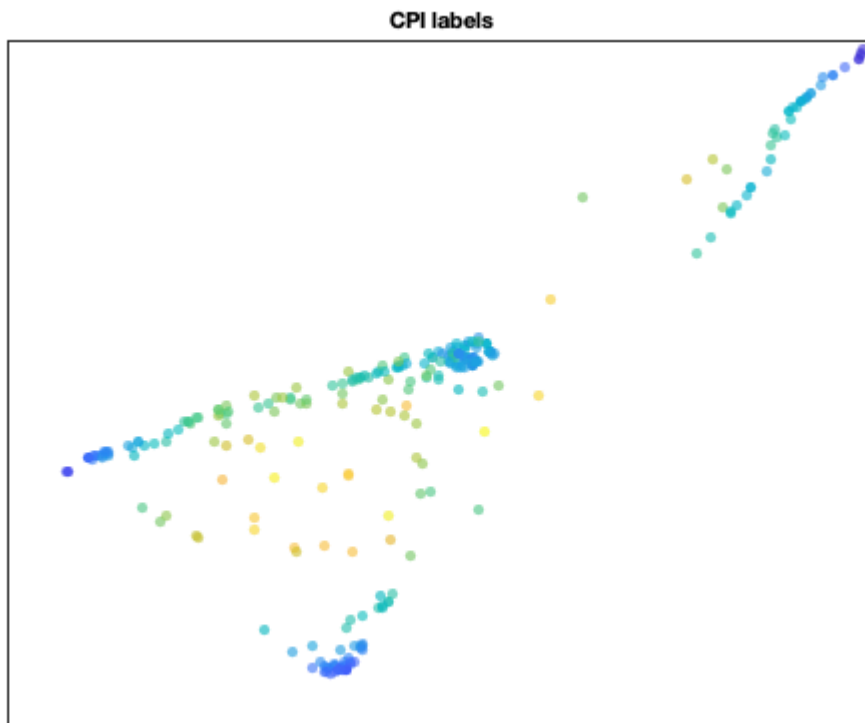
```
label_legend = {'Epcam-', 'Epcam+'};
cell_visu_PRE(result,procell_label, label_legend)
```

Clustering



True labels





Feature plot

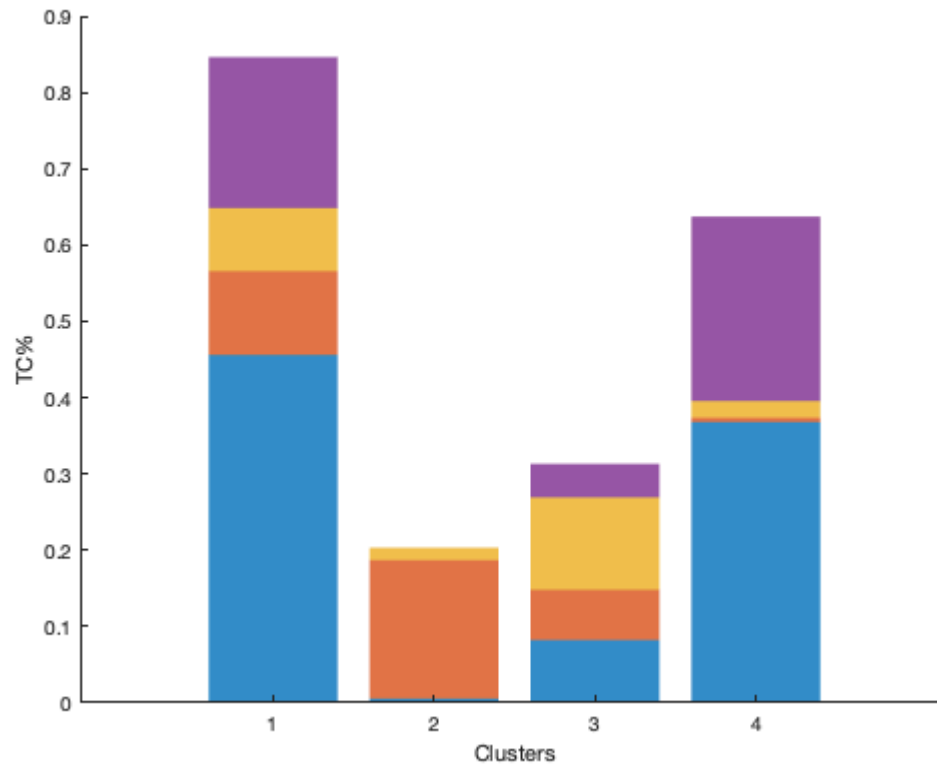
visualizes feature expressions on PRE

```
gene_plot = {'Epcam'};  
featureplot(result,prodata,progene_name,gene_plot)
```

Transition trajectory

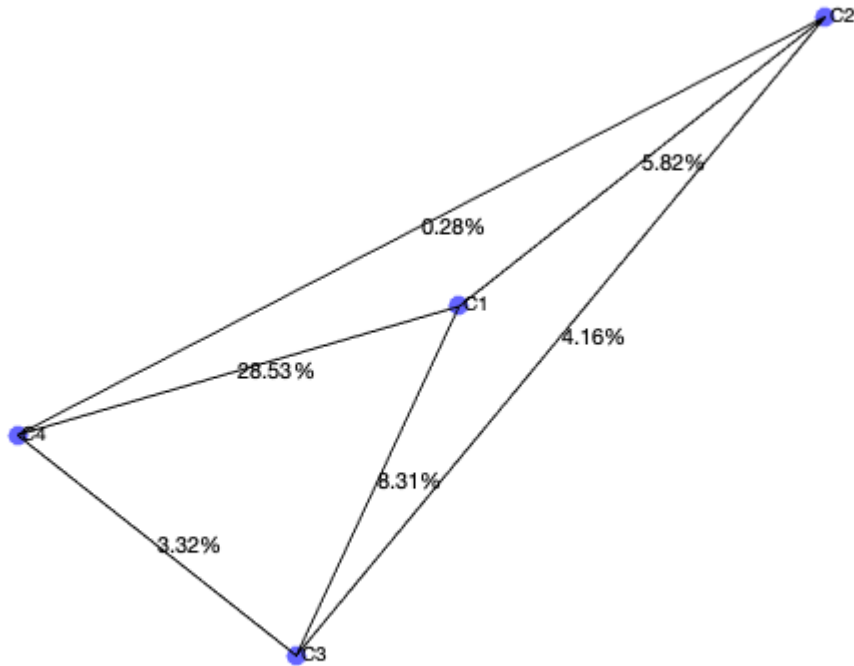
- **Start/ end cluster:** choose non-ICS based on the percentage of TC associated with each cluster relative to the total number of TC

```
pTC(result,No_cluster)
```



- **Potential transition trajectories:** list the percentage of cells over whole cell population to choose the potential transition trajectories

```
start_cluster = 2; %choose starting cluster based on the above graph
%plot TC% between clusters among all the cells
[path,ordered_cell] = traj(result,start_cluster,No_cluster);
```



```
trajectory: 2 1 3 4, percentage of cells involved: 0.50693
trajectory: 2 1 4, percentage of cells involved: 0.65651
trajectory: 2 3 1 4, percentage of cells involved: 0.84211
trajectory: 2 3 4, percentage of cells involved: 0.26039
trajectory: 2 4, percentage of cells involved: 0.1385
```

Finding cluster marker genes and the transition genes that mark transition from one trajectory

```
lam2 = 10;
[marker_gene,transition_gene] = markers(result,prodata,M,path{1},ordered_cell{1},lam2)
```

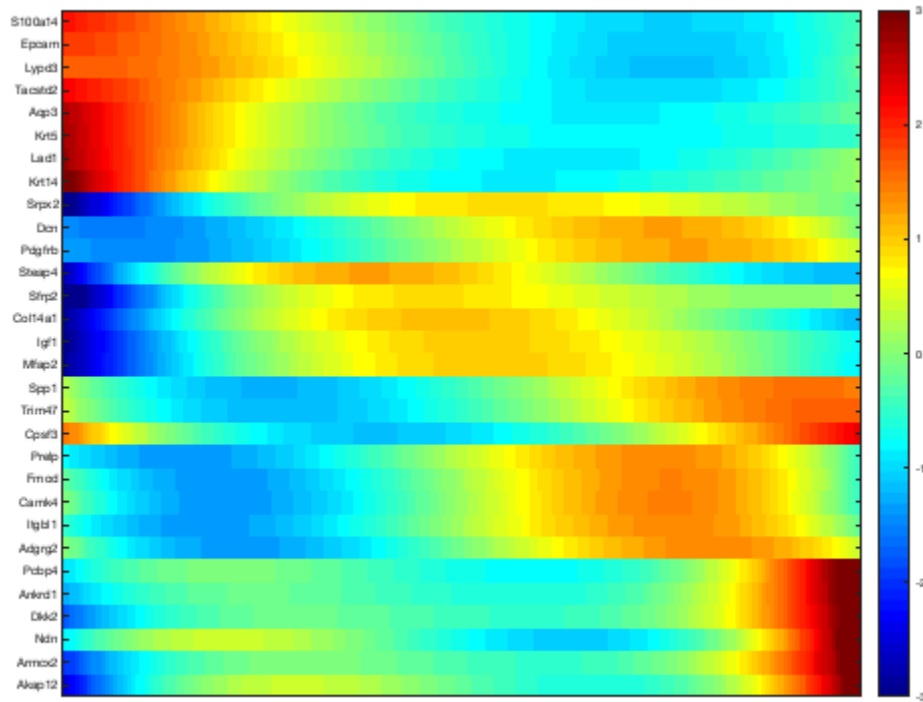
```
ans = 0.9783
Consistency of clustering: 0.97835
marker_gene = 4x1 cell array
    { 75x1 double}
    { 260x1 double}
    { 936x1 double}
    {1046x1 double}
transition_gene = 3x2 cell array
    {7x1 double}    {79x1 double}
    {4x1 double}    { 0x1 double}
    {[ 2263]}    { 0x1 double}
```

Heatmap

```

a = 5; b = 3; %plot top a marker genes and top 2b transition genes
[gene_plot, gene_cluster] = heatmap(prodata, marker_gene, transition_gene, path{1}, ...
ordered_cell{1}, progene_name, a, b);

```



Plot transition genes along the transition trajectory

```

trans_gene_plot = transition_gene{1,1}(1);
tran_gene_plot(result, prodata, trans_gene_plot, path{1}, ordered_cell{1}, progene_name)

```

