

# Package ‘JUMPsem’

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**Type** Package

**Title** Enzyme Activity Inference with Structural Equation Modeling

**Version** 1.0

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**Depends** R (>= 3.5.0)

**Imports** lavaan (>= 0.6-11),  
tidyr (>= 1.2.0),  
stringr (>= 1.4.0),  
dplyr (>= 1.0.9),  
psych (>= 2.2.5),  
devtools (>= 2.4.5),  
EFAtools (>= 0.4.1)

**Suggests** knitr,  
rmarkdown

**Description** Provides tools to infer enzyme activity using structural equation modeling (SEM). The package integrates latent variable modeling with omics data to estimate pathway-level enzyme activity.

**License** GPL-3

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 7.3.2

**VignetteBuilder** knitr

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aceAdjacency	<i>aceAdjacency</i>
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**Description**

This script is used to build HUMAN HAT-substrate adjacency matrix.

**Usage**

aceAdjacency(databaseACE)

**Arguments**

databaseACE      Default database or customized database input.

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catchActRaw	<i>catchResults</i>
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**Description**

Catch Activity and Affinity. Activity contains Raw-Activity, Mean-Center, Z-scaled.

**Usage**

catchActRaw(x)

**Arguments**

x                      The results from modeling fitting.  
rawData              Input raw data (quantitative phospho-proteomics / ubiquitin-proteomics data)

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getFa	<i>Build MEME input</i>
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**Description**

script to extract MEME input .fa file

**Usage**

```
getFa(ss = NULL, es = NULL, filePath)
```

**Arguments**

ss	Vector; Substrate species; ("mouse", "rat", "human")
es	Vector; Kinase species; ("mouse", "rat", "human")
filePath	The path is used to save .fa files. Recommend to create a new folder to save all .fa.files.

**Examples**

```
getFa(ss = "human", es = c("human", "mouse", "rat"), filePath = getwd())
```

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inputTransformPSP	<i>inputTransform</i>
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**Description**

This script is used to log2 transform AND/OR whole proteome normalize input data.

**Usage**

```
inputTransformPSP(  
  input_raw,  
  input_log2_trans = FALSE,  
  relative.norm.p = TRUE,  
  whole_log2_trans = FALSE,  
  whole_proteome = NULL,  
  relative.norm.w = TRUE  
)
```

**Arguments**

<code>input_raw</code>	Input raw data (quantitative phospho-proteomics/ubiquitin-proteomics data)
<code>input_log2_trans</code>	FALSE or TRUE. Need program to do log2 transforming of the input file or not. Default is FALSE.
<code>relative.norm.p</code>	Logical. FALSE or TRUE. Need program to do relative normalization of the PTM input file or not. Default is TRUE.
<code>whole_log2_trans</code>	Need program to do log2 transforming of the whole proteome or not. Ignore if -whole.proteome is missing. Default is FALSE.
<code>whole_proteome</code>	Set up whole proteome used to normalize phospho-proteomics/ubiquitin-proteomics data by whole proteome. Default is NULL.
<code>relative.norm.w</code>	Logical. FALSE or TRUE. Need program to do relative normalization of the whole proteomic input file or not. Default is TRUE.

JUMPsem

*JUMPsem***Description**

Infer enzyme activity from phosphoralation, ubiquitination, acetylation

**Usage**

```
JUMPsem(
  input = NULL,
  datatype = NULL,
  organism = NULL,
  enzyme.organism = NULL,
  database = NULL,
  cor.off = 0.95,
  kmo.off = 0,
  mdsite = TRUE,
  enzyList = NULL,
  input.log2.norm = FALSE,
  relative.norm.p = TRUE,
  whole.log2.trans = FALSE,
  whole.proteome = NULL,
  relative.norm.w = TRUE,
  motif = NULL,
  output.folder = getwd()
)
```

**Arguments**

<code>input</code>	Dataframe. Your quantitative ubiquiti-proteomics, phospho-proteomic and acetyl-proteomics data.
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<code>datatype</code>	Factor. Data type: ubiquitination or phosphorylation or acetylation; ("ubi", "psp", "ace")
<code>organism</code>	Vector. Substrate species; ("human", "mouse", "rat")
<code>enzyme.organism</code>	Vector. Enzyme species; ("human", "mouse", "rat")
<code>database</code>	Dataframe. Customized database input. Default is NULL using internal database.
<code>cor.off</code>	Numeric. Set up correlation cutoff value 0-1 to remove high collinear variables. Default is 0.95.
<code>kmo.off</code>	Numeric. Set up KMO cutoff value 0-1. Default is 0.
<code>mdsite</code>	Logical. Mapping with precise phosphorylated modification sites or not. Default is TRUE.
<code>enzyList</code>	Vector. Program only calculate the enzyme in the enzyList. Default is to output ALL enzyme activities and affinities.
<code>input.log2.norm</code>	Logical. FALSE or TRUE. Need program to do log2 transforming of the input file or not. Default is FALSE.
<code>relative.norm.p</code>	Logical. FALSE or TRUE. Need program to do relative normalization of the PTM input file or not. Default is TRUE.
<code>whole.log2.trans</code>	Logical. FALSE or TRUE. Need program to do log2 transforming of the whole proteome or not. (Ignore if -whole.proteome is missing). Default is FALSE.
<code>whole.proteome</code>	Dataframe. Set up whole proteome used to normalize phosphor-proteome or ubiquitin-proteome whole proteome. Default is NULL.
<code>relative.norm.w</code>	Logical. FALSE or TRUE. Need program to do relative normalization of the whole proteomic input file or not. Default is TRUE.
<code>motif</code>	Matrix. Added kinase-substrate relationships from motif discovery.
<code>output.folder</code>	Character. Character vector of location to save files if desired. Default is current directory.

## Details

You can use JUMPsem package to get enzyme activity.

## Value

List

## Examples

```
result <- JUMPsem(input = input_ubi_example,
  datatype = "ubi",
  organism = "human",
  input.log2.norm = T)
```

```
result <- JUMPsem(input = input_psp_example,
  datatype = "psp",
  organism = "mouse",
```

```

enzyme.organism = c("human", "mouse", "rat"),
cor.off = 0.8,
input.log2.norm = TRUE,
whole.log2.trans = TRUE,
motif = motif_example,
whole.proteome = wholeProteome_example)

result <- JUMPsem(input = input_ace_example,
  datatype = "ace",
  organism = "human",
  input.log2.norm = FALSE)

```

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mstInput

*Build MAST input*


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### Description

Script to extract MAST input FASTA file.

### Usage

```
mstInput(input, savePath = getwd())
```

### Arguments

input	The same quantitative phosphoproteomics data as eSEM() input file.
savePath	The path to save built MAST input file. Default is the current directory.

### Examples

```
mstInput(input = input_psp_example)
```

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motifEx

*motifEx*


---

### Description

This script is used to extract kinase-substrate relationships from motif scanning and build motif\_input file for KSEM().

### Usage

```
motifEx(mastFile, savePath = getwd())
```

### Arguments

mastFile	The output file "extract_result.txt" from Step: 03step_ExtractMASTresult.sh.
savePath	Path to save motif_input.txt file. Default is current directory.

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pspAdjacency	<i>pspAdjacency</i>
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### Description

This script is used to build kinase-substrate adjacency matrix with only PSP database.

### Usage

```
pspAdjacency(sp, ep, databasePSP, mdsite)
```

### Arguments

sp	Substrate species; ("mouse", "rat", "human").
ep	Enzyme species; ("mouse", "rat", "human").
databasePSP	Default database or customized database input.
mdsite	Logical. Mapping with precise phosphorylated modification sites or not. Default is TRUE.

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pspMotifAdjacency	<i>pspMotifAdjacency</i>
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### Description

This script is used to build enzyme-substrate adjacency matrix with PSP database + Motif.

### Usage

```
pspMotifAdjacency(sp, ep, databasePSP, motif.ref, mdsite)
```

### Arguments

sp	Substrate species; ("mouse", "rat", "human").
ep	Enzyme species; ("mouse", "rat", "human").
databasePSP	Default database or customized database input.
motif.ref	Added enzyme-substrate relationships from motif discovery. Same as KSEM -motif parameter.
mdsite	Logical. Mapping with precise phosphorylated modification sites or not. Default is TRUE.

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singleEnzymeAff	<i>singleEnzymeAff</i>
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**Description**

Get single enzyme affinity.

**Usage**

```
singleEnzymeAff(enzyme, input, adj, cor.off, kmo.off, mdsite)
```

**Arguments**

enzyme	Single enzyme name.
input	Normalized and transformed phospho-proteomics / ubiquitin-proteomics data.
adj	Adjacency matrix.
cor.off	Set up correlation cutoff value 0-1 to remove high collinear variables. Default is 0.95.
kmo.off	Set up KMO cutoff value 0-1. Default is 0.
mdsite	Logical. Mapping with precise phosphorylated modification sites or not. Default is TRUE.

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singleHatAct	<i>singleHatAct</i>
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**Description**

Get single HAT activity.

**Usage**

```
singleHatAct(HAT, input, adj, cor.off, kmo.off)
```

**Arguments**

HAT	Single HAT name.
input	Normalized and transformed acetyl-proteomics data.
adj	Adjacency matrix.
cor.off	Set up correlation cutoff value 0-1 to remove high collinear variables. Default is 0.95.
kmo.off	Set up KMO cutoff value 0-1. Default is 0.



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singleKinaseAct	<i>singleKinaseAct</i>
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**Description**

Get single kinase activity.

**Usage**

```
singleKinaseAct(kinase, input, adj, cor.off, kmo.off, mdsite)
```

**Arguments**

kinase	Single kinase name.
input	Normalized and transformed phosphoproteomics data.
adj	Adjacency matrix.
cor.off	Set up correlation cutoff value 0-1 to remove high collinear variables. Default is 0.95.
kmo.off	Set up KMO cutoff value 0-1. Default is 0.
mdsite	Logical. Mapping with precise phosphorylated modification sites or not. Default is TRUE. description

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singleLigaseAct	<i>singleLigaseAct</i>
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**Description**

Get single ligase activity.

**Usage**

```
singleLigaseAct(ligase, input, adj, cor.off, kmo.off)
```

**Arguments**

ligase	Single ligase name.
input	Normalized and transformed ubiquiti-proteomics data.
adj	Adjacency matrix.
cor.off	Set up correlation cutoff value 0-1 to remove high collinear variables. Default is 0.95.
kmo.off	Set up KMO cutoff value 0-1. Default is 0.

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ubiAdjacency	<i>ubiAdjacency</i>
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**Description**

This script is used to build ligase-substrate adjacency matrix with only PSP database.

**Usage**

```
ubiAdjacency(sp, databaseUBI)
```

**Arguments**

sp	Substrate species; ("mouse", "rat", "human").
databaseUBI	Default database or customized database input.

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