

# Package ‘eSEM’

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**Type** Package

**Title** Enzyme activity inference from Structural Equation Modeling

**Version** 1.0

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**Imports** lavaan(>= 0.6-11), tidyr(>= 1.2.0), stringr(>= 1.4.0), dplyr(>= 1.0.9), psych(>= 2.2.5), devtools(>= 2.4.5), EFAtools(>= 0.4.1)

**Description** eSEM is used to infer enzyme activity.

**Depends** R (>= 3.5.0)

**License** GPL-3

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 7.2.3

**Suggests** knitr,  
rmarkdown

**VignetteBuilder** knitr

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aceAdjacency	<i>aceAdjacency</i>
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### Description

This script is used to build HUMAN HAT-substrate adjacency matrix.

### Usage

```
aceAdjacency()
```

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catchActRaw	<i>catchResults</i>
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### Description

Catch Activity and Affinity. Activity contains Raw-Activity, Mean-Center, Z-scaled.

### Usage

```
catchActRaw(x)
```

### Arguments

x	The results from modeling fitting.
rawData	Input raw data (quantitative phospho-proteomics / ubiquitin-proteomics data)

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eSEM	<i>eSEM</i>
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### Description

Infer enzyme activity from phosphoralation, ubiquitination, acetylation

### Usage

```
eSEM(
  input = NULL,
  datatype = NULL,
  organism = NULL,
  cor.off = 0.95,
  kmo.off = 0,
  enzyList = NULL,
  input.log2.norm = FALSE,
  whole.log2.trans = FALSE,
  whole.proteome = NULL,
  motif = NULL,
  output.folder = getwd()
)
```

**Arguments**

<code>input</code>	Dataframe. Your quantitative ubiquitin-proteomics, phospho-proteomic and acetyl-proteomics data.
<code>datatype</code>	Factor. Data type: ubiquitination or phosphoralation or acetylation; ("ubi", "psp", "ace")
<code>organism</code>	Vector. Substrate species; ("human", "mouse", "rat")
<code>cor.off</code>	Numeric. Set up correlation cutoff value 0-1 to remove high collinear variables. Default is 0.95.
<code>kmo.off</code>	Numeric. Set up KMO cutoff value 0-1. Default is 0.
<code>enzyList</code>	Vector. Program only calculate the enzyme in the enzyList. Default is to output ALL enzyme activities and affinities.
<code>input.log2.norm</code>	Logical. FALSE or TRUE. Need program to do log2 transforming of the input file or not. Default is FALSE.
<code>whole.log2.trans</code>	Logical. FALSE or TRUE. Need program to do log2 transforming of the whole proteome or not. (Ignore if -whole.proteome is missing). Default is FALSE.
<code>whole.proteome</code>	Dataframe. Set up whole proteome used to normalize phosphor-proteome or ubiquitin-proteome whole proteome. Default is NULL.
<code>motif</code>	Matrix. Added kinase-substrate relationships from motif discovery.
<code>output.folder</code>	Character. Character vector of location to save files if desired. Default is current directory.

**Details**

You can use eSEM package to get enzyme activity.

**Value**

List

**Examples**

```
result <- eSEM(input = input_ubi_example,
               datatype = "ubi",
               organism = "human",
               input.log2.norm = T)

result <- eSEM(input = input_psp_example,
               datatype = "psp",
               organism = "mouse",
               input.log2.norm = TRUE,
               whole.log2.trans = TRUE,
               whole.proteome = wholeProteome_example)

result <- eSEM(input = input_ace_example,
               datatype = "ace",
               organism = "human",
               input.log2.norm = FALSE)
```

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`getFa`*Build MEME input*

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**Description**

script to extract MEME input .fa file

**Usage**

```
getFa(ks = NULL, filePath)
```

**Arguments**

<code>ks</code>	Vector; Kinase species; ("mouse", "rat", "human")
<code>filePath</code>	The path is used to save .fa files. Recommend to create a new folder to save all .fa.files.

**Examples**

```
getFa(ks = "human", filePath = getwd())
```

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`inputTransformPSP`*inputTransform*

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**Description**

This script is used to log2 transform AND/OR whole proteome normalize input data.

**Usage**

```
inputTransformPSP(  
  input_raw,  
  input_log2_trans = FALSE,  
  whole_log2_trans = FALSE,  
  whole_proteome = NULL  
)
```

**Arguments**

input_raw	Input raw data (quantitative phospho-proteomics/ubiquitin-proteomics data)
input_log2_trans	FALSE or TRUE. Need program to do log2 transforming of the input file or not. Default is FALSE.
whole_log2_trans	Need program to do log2 transforming of the whole proteome or not. Ignore if -whole.proteome is missing. Default is FALSE.
whole_proteome	Set up whole proteome used to normalize phospho-proteomics/ubiquitin-proteomics data by whole proteome. Default is NULL.

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mastInput	<i>Build MAST input</i>
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**Description**

Script to extract MAST input FASTA file.

**Usage**

```
mastInput(input, savePath = getwd())
```

**Arguments**

input	The same quantitative phosphoproteomics data as eSEM() input file.
savePath	The path to save built MAST input file. Default is the current directory.

**Examples**

```
mastInput(input = input_psp_example)
```

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motifEx	<i>motifEx</i>
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**Description**

This script is used to extract kinase-substrate relationships from motif scanning and build motif\_input file for KSEM().

**Usage**

```
motifEx(mastFile, savePath = getwd())
```

**Arguments**

mastFile	The output file "extract_result.txt" from Step: 03step_ExtractMASTresult.sh.
savePath	Path to save motif_input.txt file. Default is current directory.

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pspAdjacency	<i>pspAdjacency</i>
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### Description

This script is used to build kinase-substrate adjacency matrix with only PSP database.

### Usage

```
pspAdjacency(sp)
```

### Arguments

sp	species; ("mouse", "rat", "human").
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pspMotifAdjacency	<i>pspMotifAdjacency</i>
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### Description

This script is used to build enzyme-substrate adjacency matrix with PSP database + Motif.

### Usage

```
pspMotifAdjacency(sp, motif.ref)
```

### Arguments

sp	Species; ("mouse", "rat", "human").
motif.ref	Added enzyme-substrate relationships from motif discovery. Same as KSEM -motif parameter.

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singleEnzymeAff	<i>singleEnzymeAff</i>
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### Description

Get single enzyme affinity.

### Usage

```
singleEnzymeAff(enzyme, input, adj, cor.off, kmo.off)
```

**Arguments**

enzyme	Single enzyme name.
input	Normalized and transformed phospho-proteomics / ubiquitin-proteomics data.
adj	Adjacency matrix.
cor.off	Set up correlation cutoff value 0-1 to remove high collinear variables. Default is 0.95.
kmo.off	Set up KMO cutoff value 0-1. Default is 0.

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singleHatAct	<i>singleHatAct</i>
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**Description**

Get single HAT activity.

**Usage**

```
singleHatAct(HAT, input, adj, cor.off, kmo.off)
```

**Arguments**

HAT	Single HAT name.
input	Normalized and transformed acetyl-proteomics data.
adj	Adjacency matrix.
cor.off	Set up correlation cutoff value 0-1 to remove high collinear variables. Default is 0.95.
kmo.off	Set up KMO cutoff value 0-1. Default is 0.

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singleKinaseAct	<i>singleKinaseAct</i>
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**Description**

Get single kinase activity.

**Usage**

```
singleKinaseAct(kinase, input, adj, cor.off, kmo.off)
```

**Arguments**

kinase	Single kinase name.
input	Normalized and transformed phosphoproteomics data.
adj	Adjacency matrix.
cor.off	Set up correlation cutoff value 0-1 to remove high collinear variables. Default is 0.95.
kmo.off	Set up KMO cutoff value 0-1. Default is 0.

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singleLigaseAct	<i>singleLigaseAct</i>
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**Description**

Get single ligase activity.

**Usage**

```
singleLigaseAct(ligase, input, adj, cor.off, kmo.off)
```

**Arguments**

ligase	Single ligase name.
input	Normalized and transformed ubiquiti-proteomics data.
adj	Adjacency matrix.
cor.off	Set up correlation cutoff value 0-1 to remove high collinear variables. Default is 0.95.
kmo.off	Set up KMO cutoff value 0-1. Default is 0.

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ubiAdjacency	<i>ubiAdjacency</i>
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**Description**

This script is used to build ligase-substrate adjacency matrix with only PSP database.

**Usage**

```
ubiAdjacency(sp)
```

**Arguments**

sp	species; ("mouse", "rat", "human").
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