Variant annotation

VARIANT and HPG-VARIANT

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10th June 2014

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PRINCIPE FELIPE
CENTRO DE INVESTIGACION

ciberer
CENTRO DE INVESTIGACIÓN BIOMÉDICA EN RED DE ENFERMEDADES RARAS

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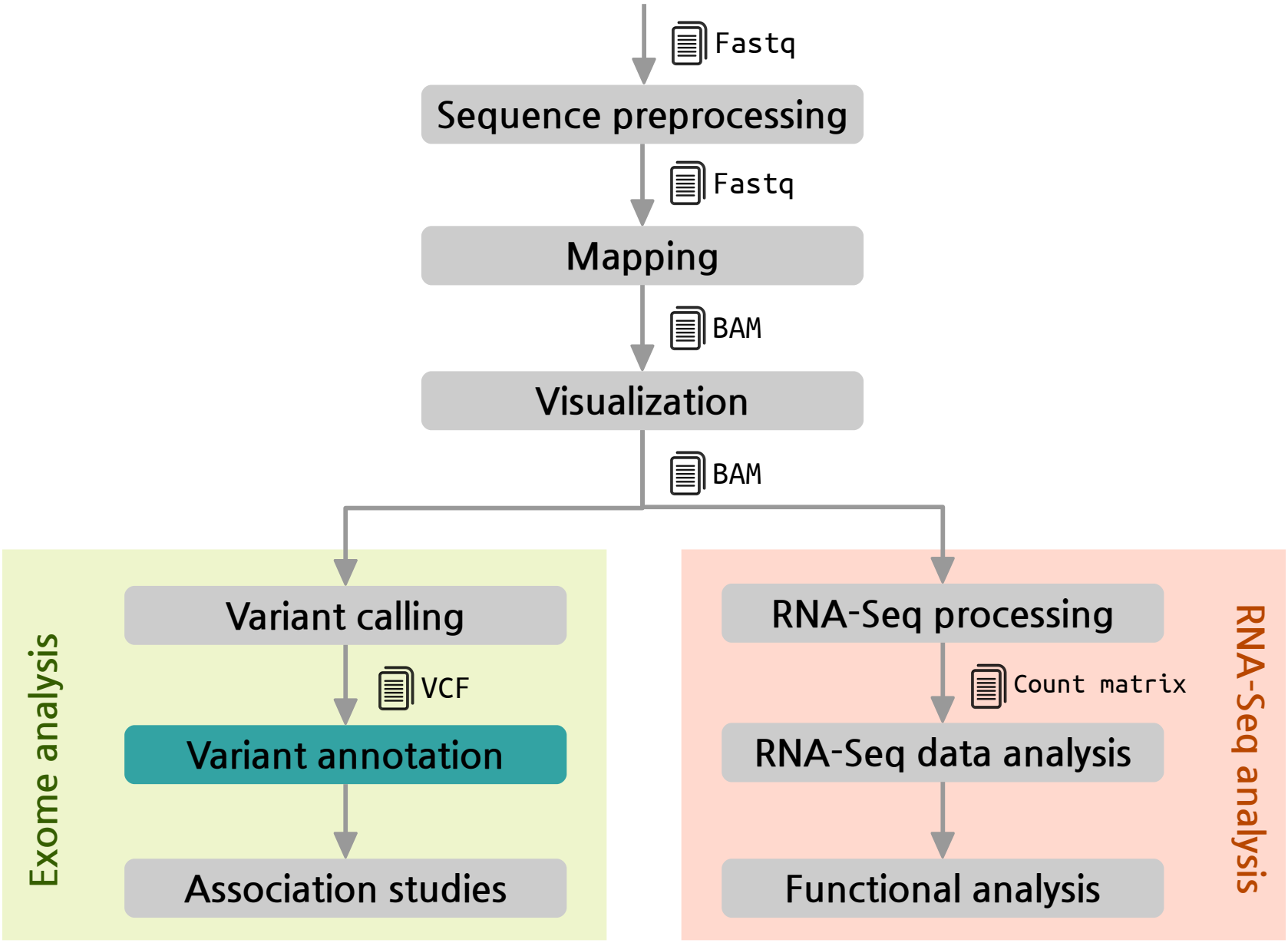
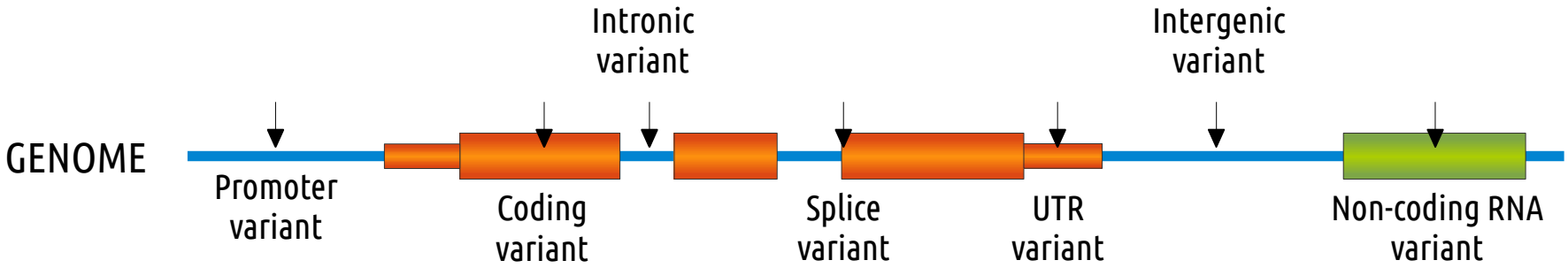
PLATAFORMA BIOINFORMÁTICA DE ENFERMEDADES RARAS

The pipeline



What is functional annotation?



Why we do that?

- Each individual exome carries ~25,000 variants → **PRIORITIZATION!**
- We want to identify a **small subset** of functionally important variants to pinpoint the putative disease causal variants
- We need strategies to **estimate the deleteriousness** of our variants to better identify disease-causal variants

CAUTION!

On average, each *normal* person is found to carry:

~11,000 synonymous variants

~11,000 non-synonymous variants

250 to 300 los-of-function variants in annotated genes

50 to 100 variants previously implicated in **inherited disorders**

1000 Genomes Project Consortium. *A map of human genome variation from population-scale sequencing.* **Nature**. 2010 Oct 28;467(7319):1061-73. PubMed PMID: 20981092

Sources of functional information

Table 1 Publicly available tools and databases for various tasks of genetic variant annotation and prioritization

Category	Database/tool/project	Description	URL
Genetic variant data sources	dbSNP[68]	Comprehensive, curated SNP and short indel database	http://www.ncbi.nlm.nih.gov/projects/SNP
	DbVar[69]	Comprehensive, curated database for structural variants	http://www.ncbi.nlm.nih.gov/dbvar
	DGV[70]	Human structural variants from samples with no phenotype	http://projects.tcag.ca/variation
Functional characterization of genomic elements	ENCODE[71]	High-throughput functional characterization of DNA elements, including noncoding regions	http://www.genome.gov/10005107
	SIFT[72], PolyPhen[73]	Prioritization of nonsynonymous SNPs	http://sift.jcvi.org, http://genetics.bwh.harvard.edu/pph2
Public gene–trait associations	dbGaP[34]	Comprehensive listing of genotype-to-phenotype mappings	http://www.ncbi.nlm.nih.gov/gap
	EGA[74]	Genotype–phenotype experiment archive	http://www.ebi.ac.uk/ega
Disease-associated mutations	HGMD[35]	Database for human disease mutations	http://www.hgmd.org
	OMIM[36]	Mendelian disease gene associations	http://www.ncbi.nlm.nih.gov/omim
	SwissVar[76]	Variant catalog of the UniProt knowledge bases	http://swissvar.expasy.org
	GAD[77]	NCBI source for genotype–disease associations	http://geneticassociationdb.nih.gov
	GWAS catalog from NHGRI[78]	SNP-phenotype associations found by GWAS	http://www.genome.gov/gwastudies
Whole-genome repositories	Complete genomics public genomes[79]	Complete genomics for 69 genomes from multiple ancestries (includes samples from the NHGRI and NIGMS repositories)	http://www.completegenomics.com/sequence-data/download-data
	1,000 Genomes[80]	Expanding resource currently housing three low-coverage whole genomes of multiple ancestries	http://www.1000genomes.org
Ancestry-focused variant data sources	HapMap[26]	Haplo-block mapping for diverse populations	http://www.hapmap.org
	HGDP[27]	SNP profiles of samples from several endogenous populations	http://hagsc.org/hgdp
Pharmacogenomic associations and data sources	PharmGKB[56]	Variant–pharmacokinetic/pharmacodynamic trait associations and gene–drug interactions	http://www.pharmgkb.org
	DrugBank[81]	Drug-target database with biochemical properties	http://drugbank.ca

Cordero P, Ashley EA. *Whole-genome sequencing in personalized therapeutics.* **Clin Pharmacol Ther**. 2012 Jun ;91(6):1001-9. PubMed PMID: 22549284

Computational method and tools

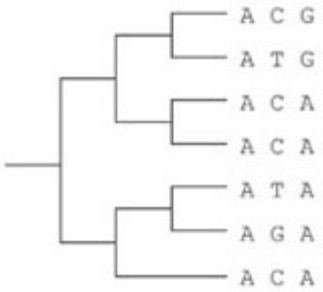
- **Annotated information** is sometimes **limited**, particularly for rare and complex traits
- Computational methods can measure deleteriousness by using **comparative genomics** and knowledge of **protein biochemistry and structure**

Comparative Genomics

Focus on sequences that have not been remove by **natural selection**.

Quantify evolutionary changes in genes or genomes and define conserved and neutral regions.

Variants observed in conserved sites are highly likely to be **deleterious**.



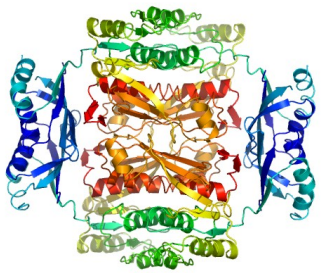
Effects in protein-coding variants

Can combine **evolutionary** and **biochemical** information.

Two types of approaches: first **principles** approaches and **trained** approaches.

Use **alignments of homologous proteins** to estimate mutational deleteriousness.

Use **biochemical data** such as amino acid properties, binding information and structural information to estimate the impact.



Effects in non-coding variants

The majority of the human genetic variation is in non-coding regions.

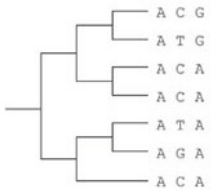
No detectable conservation outside vertebrates.

Main strategy for estimation is testing the **mammalian conservation** of the non-coding variants.

Cooper GM, Shendure J. *Needles in stacks of needles: finding disease-causal variants in a wealth of genomic data.* **Nature Reviews Genetics**. 2011 Aug 18;12(9):628-40. Pubmed PMID: 21850043

Computational methods and tools

Prediction scores for non-synonymous variants



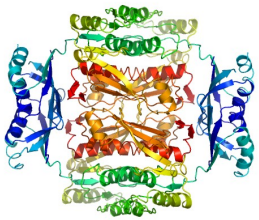


Table 1 | **Tools for protein-sequence-based prediction of deleteriousness**

Name	Type	Information	URL	Refs
MAPP	Constraint-based predictor	Evolutionary and biochemical	http://mendel.stanford.edu/SidowLab/downloads/MAPP/index.html	27
SIFT	Constraint-based predictor	Evolutionary and biochemical (indirect)	http://sift.bii.a-star.edu.sg/	39
PANTHER	Constraint-based predictor	Evolutionary and biochemical (indirect)	http://www.pantherdb.org/	41
MutationTaster*	Trained classifier	Evolutionary, biochemical and structural	http://www.mutationtaster.org/	40
nsSNP Analyzer	Trained classifier	Evolutionary, biochemical and structural	http://snpanalyzer.uthsc.edu/	44
PMUT	Trained classifier	Evolutionary, biochemical and structural	http://mmb2.pcb.ub.es:8080/PMut/	38
polyPhen	Trained classifier	Evolutionary, biochemical and structural	http://genetics.bwh.harvard.edu/pph2/	35
SAPRED	Trained classifier	Evolutionary, biochemical and structural	http://sapred.cbi.pku.edu.cn/	42
SNAP	Trained classifier	Evolutionary, biochemical and structural	http://www.rostlab.org/services/SNAP/	36
SNPs3D	Trained classifier	Evolutionary, biochemical and structural	http://www.snps3d.org/	51
PhD-SNP	Trained classifier	Evolutionary and biochemical (indirect)	http://gpcr2.biocomp.unibo.it/~emidio/PhD-SNP/PhD-SNP_Help.html	37

*Also makes predictions for synonymous and non-coding variant effects: for example, splicing. MAPP, Multivariate Analysis of Protein Polymorphism; polyPhen, polymorphism phenotyping.

Cooper GM, Shendure J. *Needles in stacks of needles: finding disease-causal variants in a wealth of genomic data.* **Nature Reviews Genetics**. 2011 Aug 18;12(9):628-40. Pubmed PMID: 21850043

Computational methods and tools

Prediction scores for non-coding variation

Table 2 | **Tools for nucleotide-sequence-based prediction of deleteriousness**

Name	Type	Information	URL	Refs
phastCons	Phylogenetic HMM	Evolutionary	http://compgen.bscb.cornell.edu/phast/	60
GERP	Single-site scoring	Evolutionary	http://mendel.stanford.edu/SidowLab/downloads/gerp/index.html	67
Gumby	Single-site scoring	Evolutionary	http://pga.jgi-psf.org/gumby/	21
phyloP	Single-site scoring	Evolutionary	http://compgen.bscb.cornell.edu/phast/	66
SCONE	Single-site scoring	Evolutionary	http://genetics.bwh.harvard.edu/scone/	68
binCons	Sliding-window scoring	Evolutionary	http://zoo.nhgri.nih.gov/binCons/index.cgi	69
Chai Cons	Sliding-window scoring	Evolutionary and structural	http://research.nhgri.nih.gov/software/chai	71
VISTA	Visualization tool (various scores)	Evolutionary	http://genome.lbl.gov/vista/index.shtml	70

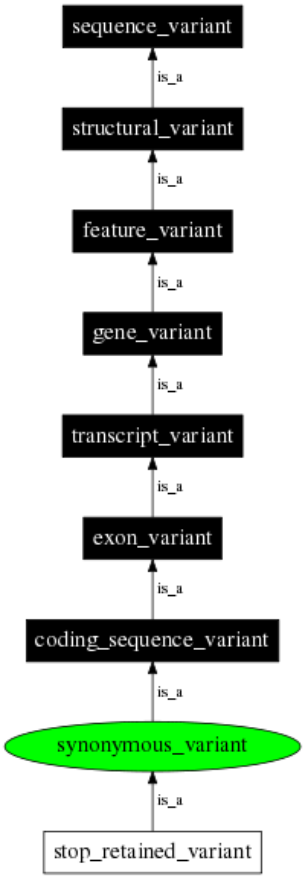
GERP, Genomic Evolutionary Rate Profiling; HMM, hidden Markov model; SCONE, Sequence Conservation Evaluation.

Cooper GM, Shendure J. *Needles in stacks of needles: finding disease-causal variants in a wealth of genomic data*. **Nature Reviews Genetics**. 2011 Aug 18;12(9):628-40. Pubmed PMID: 21850043

Consequence types

The standard: SO The Sequence Ontology Project Common vocabulary

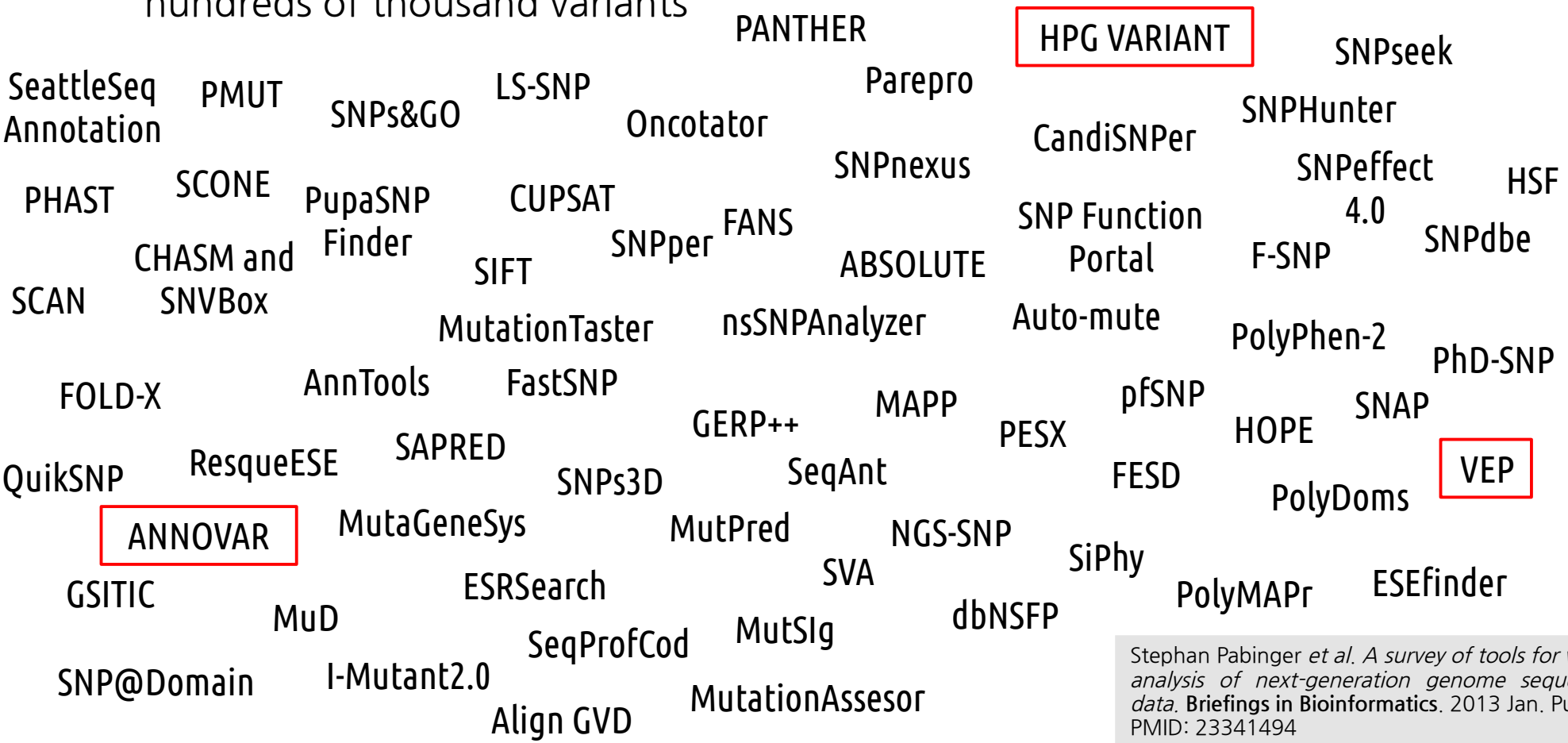
Label	SO accession	Description
Coding sequence	SO:0001580	In coding sequence with in determinate effect
Synonymous codon	SO:0001588	In coding sequence, not resulting in an amino acid change (silent mutation)
Non-synonymous codon	SO:0001583	In coding sequence and results in an amino acid change in the encoded peptide sequence
Stop gained	SO:0001587	In coding sequence, resulting in the gain of a stop codon
Stop lost	SO:0001587	In coding sequence, resulting in the gain of a stop codon
Splice site	SO:0001630	1-3bps in to an exon or 3-8bps into an intron
Splice acceptor	SO:0001574	A splice variant that changes the 2 base region at the 3' end of an intron
Splice donor	SO:0001575	A splice variant that changes the 2 base region at the 5' end of an intron
5' UTR	SO:0001623	In 5 prime untranslated region
3' UTR	SO:0001624	In 3 prime untranslated region
Upstream	SO:0001635	Within 5kb upstream of the 5 prime end of a transcript
Downstream	SO:0001633	Within 5kb downstream of the 3 prime end of a transcript
TFBS	SO:0001782	A sequence variant located with in a transcription factor binding site
miRNA target	SO:0000934	A binding site where the molecule is a microRNA
Intergenic	SO:0001628	More than 5 kb either upstream or downstream of a transcript



More information: http://www.ensembl.org/info/docs/variation/predicted_data.html

Tools for functional annotation

- We need to measure the **impact** of each variant in the genome
- We **cannot** annotate 25,000 variants **manually** checking more than 20 databases
- **Tools integrate** biological information and **ease** the functional annotation of hundreds of thousand variants



Stephan Pabinger *et al. A survey of tools for variant analysis of next-generation genome sequencing data.* **Briefings in Bioinformatics**. 2013 Jan. Pubmed PMID: 23341494

AnnoVar

ANNOVAR web site: http://www.openbioinformatics.org/annovar/

- Free and open source
- Can annotate SNV, insertions and deletions
- **Regulatory information**: Conserved genomic regions, TFBSs, miRNA targets and predicted miRNA secondary structures. ENCODE DNAse I hypersensitive sites, Histone methylations, ChIP and RNA-Seq peaks
- DbSNP, 1000 genomes SIFT and GERP filtering
- **Predictions**: Polyphen, LRT, MutationTaster, PhyloP
- Can handle **custom annotations** in GFF3
- Can handle 1 o 0-based coordinates
- **5 Species** (human, mouse, worm, fly, yeast)

- Accepts VCF4, GFF3-SOLiD and CSV BUT after conversion to their **particular input file**:

Chr	Start	End	Ref	Obs	Comments
1	161003	161003	C	T	comments: rs1000050

- **Perl** written program
- **Installation** required
- Users need to **download** every annotation database and save them locally (**~35GB** per assembly)
- Need to be **run several times**
- **Output**: several files depending on the query
- Does not use **Sequence Ontology** terms

Wang K, Li M, Hakonarson H. *ANNOVAR: Functional annotation of genetic variants from next-generation sequencing data.* **Nucleic Acids Research**. Sep;38(16):e164 Pubmed PMID: 20601685

AnnoVar

EXAMPLE of ANNOVAR usage

```
DOWNLOADING BIOLOGICAL DATA:

 user@computer:~$ annotate_variation.pl -buildver hg19 -downdb refgene humandb/

 user@computer:~$ annotate_variation.pl -buildver hg19 -downdb snp135 -webfrom annovar humandb/

 user@computer:~$ annotate_variation.pl -buildver hg19 -downdb phastConsElements46way humandb/

 user@computer:~$ annotate_variation.pl -buildver hg19 -downdb 1000g2012apr -webfrom annovar
humandb/

 user@computer:~$ annotate_variation.pl -buildver hg19 -downdb cytoBand humandb/


EXTRACTING THE EFFECT:

 user@computer:~$ annotate_variation.pl -geneanno example/ex1.human humandb/

 user@computer:~$ annotate_variation.pl -regionanno -dbtype band example/ex1.human humandb/

 user@computer:~$ annotate_variation.pl -filter -dbtype 1000g2012apr_eur example/ex1.human humandb/
```

Variant Effect Predictor (VEP)

VEP documentation site: http://www.ensembl.org/info/docs/variation/vep/index.html

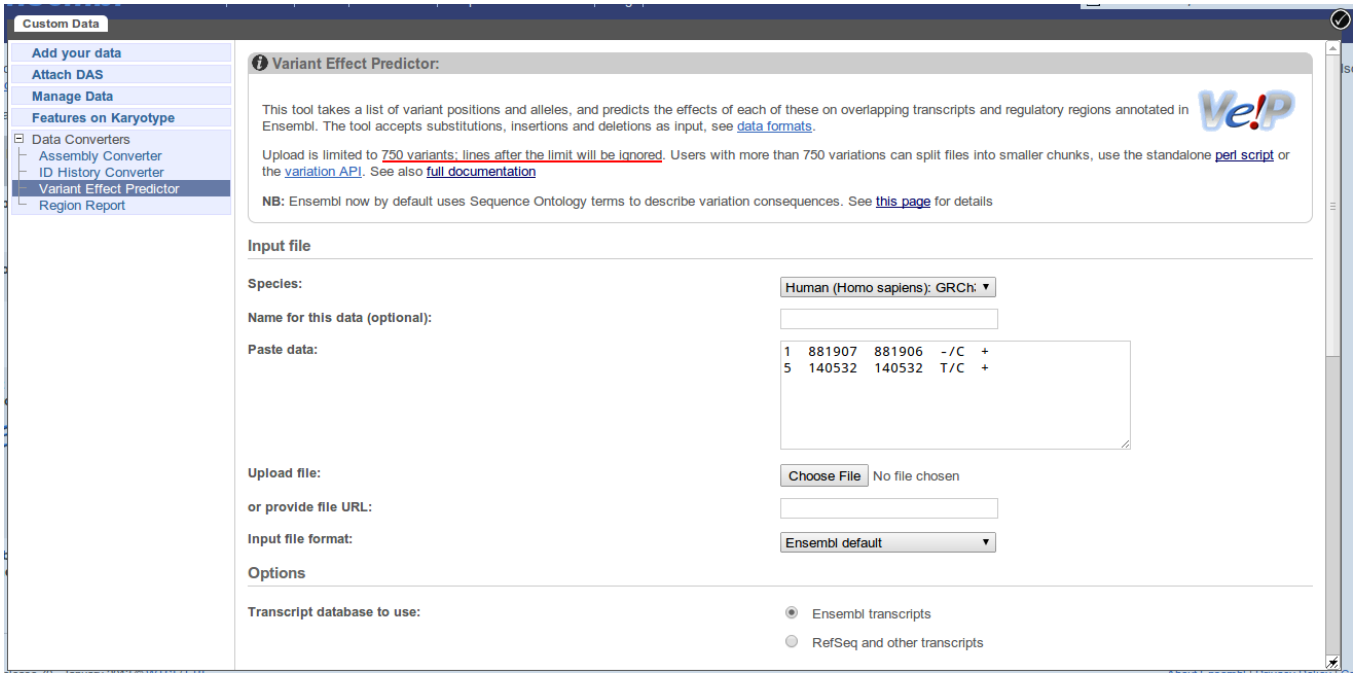
- Backed by **Ensembl**
- Free and open source
- **3 ways of functionality**: web interface, standalone Perl script and Ensembl's Perl API
- **Input** formats: CSV, VCF, Pileup and HGVS
- **Regulatory information**: TFBSs
- **Filtering** by coding regions and MAF
- **Predictions**: SIPF, PolyPhen
- 1000 genomes and dbSNP information
- Uses **Sequence Ontology**
- **Many species**

- Regulatory information does **not include miRNA targets**
- Web interface limit: **750 variants**
- The **standalone Perl script** needs:
  - **Perl** and **MySQL** support (more than 100GB of data)
  - **Download**, **install** and **update** every ~ 2 months
- Perl **API** requires:
  - **Installation** (Really, really hard!)
  - **Downloads** and **update**
  - API documentation → **Hard to understand**

McLaren W, Pritchard B, Rios D, Chen Y, Flicek P, Cunningham F. *Deriving the consequences of genomic variants with the Ensembl API and SNP Effect Predictor.* **BMC Bioinformatics** 26(16):2069-70(2010) Pubmed PMID: 20562413

Variant Effect Predictor (VEP)

VEP web interface: http://www.ensembl.org/tools.html



*The web interface to the VEP has a hard **limit of 750 variants** in your uploaded file. However, it is possible that the tool will not work with fewer variants than this, depending on the content of your data and the features you switch on. For example, a relatively small file (e.g. 100 variants) **may fail to return results if every variant in the file falls in a different gene and those genes are spread across many chromosomes**. Contrastingly, a file containing 500 variants may return results quickly if those variants all fall in just a few genes.*

*To mitigate this issue, users should consider **splitting up their input by chromosome** and uploading each chromosome's variants as a separate file. The problem can also be solved by using the VEP script - it is a command line tool, but not as hard to use as you might think! It also offers many more features than the web interface and is a generally much more powerful tool.*

http://www.ensembl.org/info/docs/variation/vep/index.html

McLaren W, Pritchard B, Rios D, Chen Y, Flicek P, Cunningham F. *Deriving the consequences of genomic variants with the Ensembl API and SNP Effect Predictor.* **BMC Bioinformatics** 26(16):2069-70(2010) Pubmed PMID: 20562413

Variant Effect Predictor (VEP)

EXAMPLE of API usage: Getting all variations in a particular human transcript and see what is the effect of that variation in the transcript

```perl
use strict;
use warnings;
use Bio::EnsEMBL::Registry;

my $registry = 'Bio::EnsEMBL::Registry';

$registry->load_registry_from_db(
    -host => 'ensembldb.ensembl.org',
    -user => 'anonymous'
);

my $stable_id = 'ENST00000393489'; #this is the stable_id of a human transcript
my $transcript_adaptor = $registry->get_adaptor('homo_sapiens', 'core', 'transcript'); #get the adaptor to get the Transcript from the database
my $transcript = $transcript_adaptor->fetch_by_stable_id($stable_id); #get the Transcript object

my $trv_adaptor = $registry->get_adaptor('homo_sapiens', 'variation', 'transcriptvariation'); #get the adaptor to get TranscriptVariation objects
my $trvs = $trv_adaptor->fetch_all_by_Transcripts([$transcript]); #get ALL effects of Variations in the Transcript

foreach my $tv (@{$trvs}) {
        my $tvas = $tv->get_all_alternate_TranscriptVariationAlleles();

        foreach my $tva(@{$tvas}) {
                my @ensembl_consequences;
                my @so_consequences;

                my $ocs = $tva->get_all_OverlapConsequences();

                foreach my $oc(@{$ocs}) {
                        push @ensembl_consequences, $oc->display_term;
                        push @so_consequences, $oc->SO_term;
                }

                my $sift = $tva->sift_prediction;
                my $polyphen = $tva->polyphen_prediction;

                print
                        "Variation ", $tv->variation_feature->variation_name,
                         " allele ", $tva->variation_feature_seq,
                         " has consequence ", join(",", @ensembl_consequences),
                         " (SO ", join(",", @so_consequences), ").";

                if(defined($sift)) {
                        print " SIFT=$sift";
                }
                if(defined($polyphen)) {
                        print " PolyPhen=$polyphen";
                }

                print "\n";
        }
}
```

McLaren W, Pritchard B, Rios D, Chen Y, Flicek P, Cunningham F. *Deriving the consequences of genomic variants with the Ensembl API and SNP Effect Predictor.* **BMC Bioinformatics** 26(16):2069-70(2010) Pubmed PMID: 20562413

HPG-VARIANT (aka VARIANT)

HPG-VARIANT web site: http://www.opencb.org/projects/hpg/doku.php?id=variant:overview

- Free and open source.
- **3 ways of functionality**: C CLI program, Web application and Java RESTful WS API
- **Cloud** variant annotator. Requires **no installation or updates**
- **Regulatory information**: Conserved genomic regions, TFBSs and miRNA targets. ENCODE DNAse I hypersensitive sites and Histone methylations
- dbSNP and 1000genomes information
- **Phenotypic information**: HGMD, COSMIC and OMIM
- **Cross-link** with many other DDBB (Ensembl, UniProt, PDB, etc)
- Polyphen and SIFT
- **Input**: VCF, GFF and BED. Accepts compressed files in ***tar.gz***
- **11 species** (human, mouse, rat, zebra fish, worm, fly, yeast, dog, pig, mosquito and plasmodium)
- HPG-VARIANT-GWAS to test for **association**

- **Young** program, many **new features coming**
  - PhastCons, GERP
  - Many more species (~25 new species)
  - Large structural variants annotation

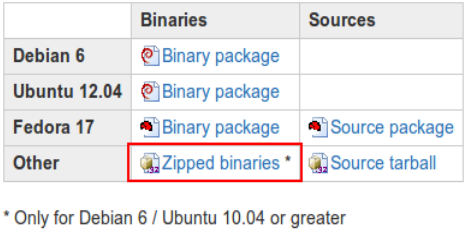
Medina I, De Maria A, Bleda M, Salavert F, Alonso R, Gonzalez CY, Dopazo J. *VARIANT: Command Line, Web service and Web interface for fast and accurate functional characterization of variants found by Next-Generation Sequencing.* **Nucleic Acids Research**. 2012 Jul;40(Web Server issue):W54-8 Pubmed PMID: 22693211

HPG-VARIANT (CLI Program)

- **Download** the program and save it into your `course/variant_annotation` folder :

  http://wiki.opencb.org/projects/hpg/doku.php?id=variant:downloads

- Extract the contents



	Binaries	Sources
Debian 6	Binary package	
Ubuntu 12.04	Binary package	
Fedora 17	Binary package	Source package
Other	Zipped binaries *	Source tarball

* Only for Debian 6 / Ubuntu 10.04 or greater

- Add the folder to your PATH

```
echo "export PATH=$PATH:/home/Desktop/hpg-variant-1.0" >> ~/.bashrc
source ~/.bashrc
```

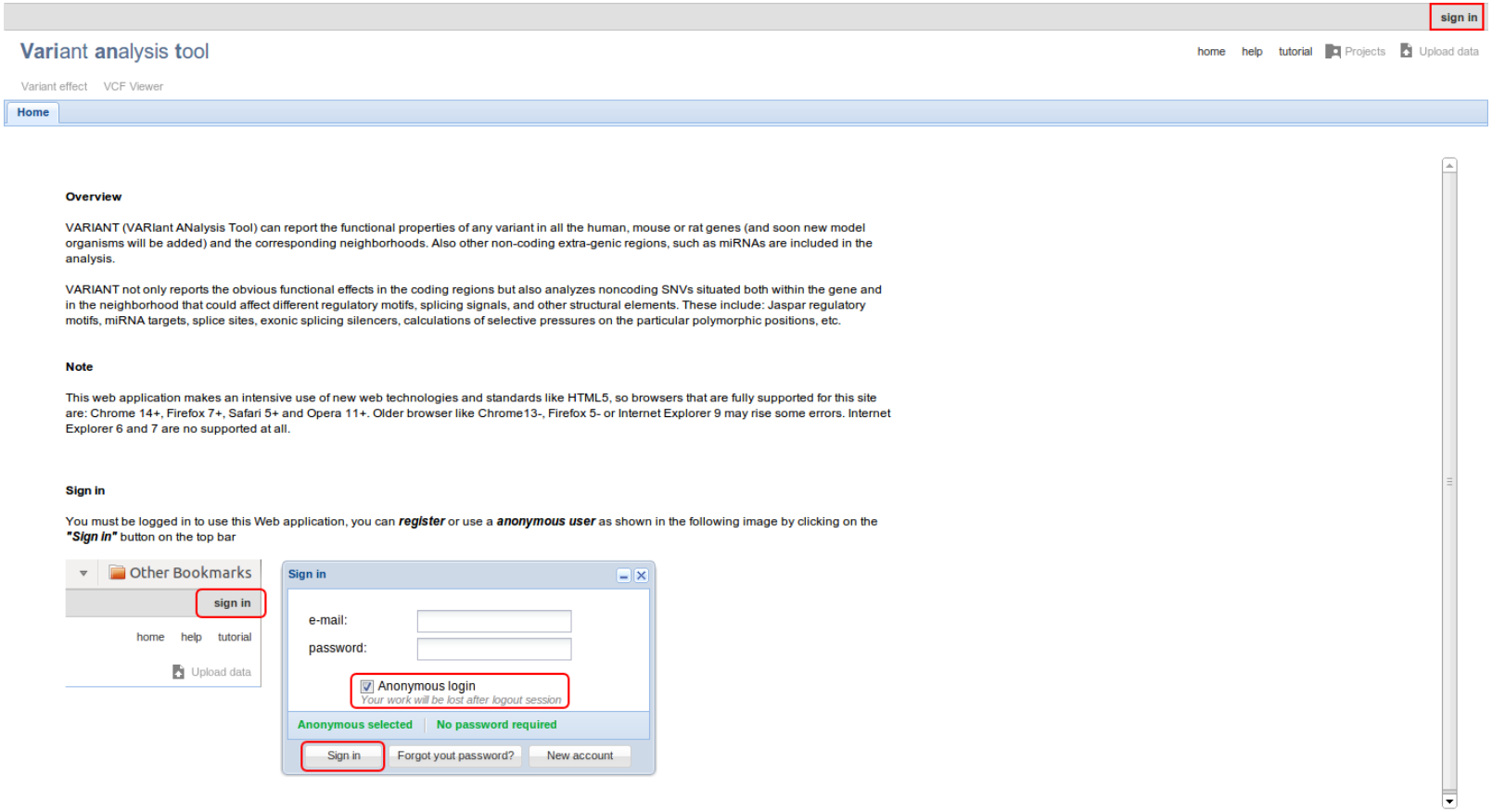
- Usage:

```
hpg-var-effect -v CHB.exon.2010_03.sites.vcf --outdir effect_output/
```

HPG-VARIANT
Web application

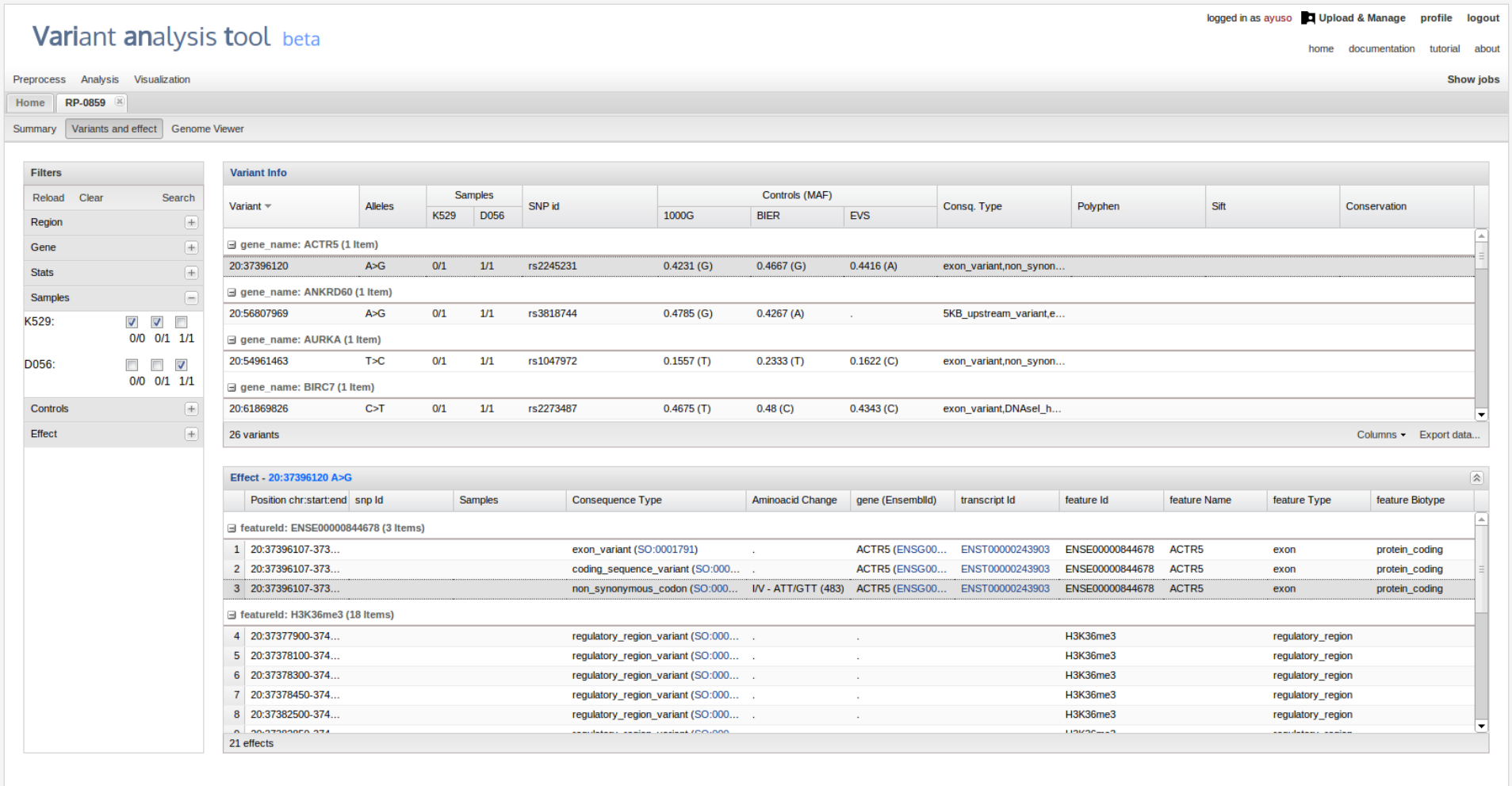
- Web application

  http://variant.bioinfo.cipf.es/



BierApp

Bierapp.babelomics.org



THANK YOU.